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- Supplementary Information -

A quantitative empirical directing group scale for selectivity

in iridium-catalysed hydrogen isotope exchange reactions

Daria S. Timofeeva, David M. Lindsay, David J. Nelson*, William J. Kerr*

WestCHEM Department of Pure and Applied Chemistry, Thomas Graham Building, 295

Cathedral Street, Glasgow, G1 1XL, UK

david.nelson@strath.ac.uk

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1. General Experimental Details

General

For the synthetic procedures, standard Schlenk techniques under an inert gas atmosphere (Ar or N_2) were used, unless otherwise stated. Materials obtained from commercial sources were used without further purification. All glassware was flame dried and cooled under a stream of nitrogen.

Materials

Dichloromethane, tetrahydrofuran, diethyl ether and toluene were obtained from a PureSolv SPS-400-5 Solvent Purification System. Ethyl acetate was dried over K_2CO_3 , then distilled under a nitrogen atmosphere and stored over 3 Å molecular sieves. Dry organic solvents and distilled water used for cross-coupling reactions were additionally degassed by bubbling argon through the solvent for 30 min. For reactions/work-up procedures in air, p.a. grade solvents were used. Petroleum ether refers to alkanes with a boiling point range of 40-60°C.

(1,3-Bis-(2,4,6-trimethylphenyl)imidazolium chloride,^{S1} phenylthiazoline,^{S2} and 1-methyl-2phenylimidazole^{S3} were synthesised according to literature procedures. 2-Phenylthiazole, 2phenylpyrimidine, 2-(4-acetyl)phenylpyridine, and 2-(4-cyano)phenylpyridine were prepared by cross-coupling reactions of corresponding phenylboronic acids and heterobromides as described in section 2. 2-(4-Acetyl)phenyloxazoline and 2-(4-(pyridin-2-yl)phenyl)-4,5dihydrooxazole were obtained from the reaction between corresponding aryl nitriles and amino alcohols catalysed by [Cu(Cl)(IPr)].^{S4} Anhydrous Na[BArF₂₄] (BArF₂₄ = tetrakis[3,5bis(trifluoromethyl)phenyl]borate)) was obtained following Bergman's synthesis,^{S5} followed by recrystallising the crude Na[BArF₂₄]·*x*(solvent) prior to drying.^{S6} Phosphine/NHC monodentate complex **Ir-1** with the BArF₂₄ counterion was synthesised from neutral chlorocarbene complex **Ir-2**^{S7} in a procedure adapted from that published before for preparation of corresponding complexes with BF₄ and OTf counterions.^{S8}

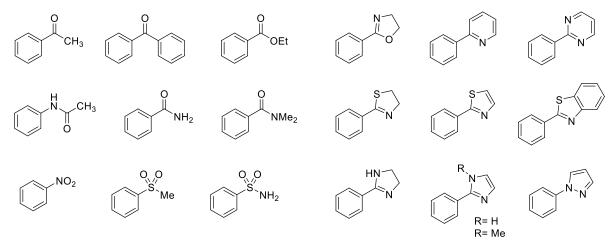
Flash column chromatography was carried out using silica gel (230-400 mesh). Thin layer chromatography (TLC) was performed using Merck silica plates coated with fluorescent indicator and visualised by UV light (254 nm)

Analysis

NMR Spectroscopy: ¹H (400 MHz), ¹³C{¹H} (101 MHz), ¹¹B (128 MHz) ¹⁹F (376 MHz) and ³¹P{¹H} (162 MHz) NMR spectra were obtained on a Bruker AV3-400 instrument with a liquid nitrogen Prodigy cryoprobe. The chemical shifts (δ) are reported in ppm relative to the residual protonated solvent for ¹H NMR or solvent signal for ¹³C{¹H} NMR (CDCl₃: δ_H 7.26 ppm and δ_C 77.16 ppm; DMSO-*d*₆: δ_H 2.50 ppm and δ_C 39.51 ppm; C₆D₆: δ_H 7.16 ppm; acetone-*d*₆: δ_H 2.05 ppm).^{S9} Coupling constants (*J*) are reported in Hz and refer to ³*J*_{H-H} couplings, unless otherwise stated. Multiplicities are expressed with s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad signal). If no multiplicity is given for ¹³C{¹H} data, the signal is a singlet. NMR assignments were made using additional 2D NMR experiments where necessary. *Infrared Spectroscopy*: Infrared (IR) spectra were collected on a Shimadzu IRAffinity-1 Spectrophotometer with only major peaks being reported.

Elemental analysis was performed using a Perkin-Elmer CH2400 instrument.

a) For intermolecular competition experiments



b) For intramolecular competition experiments

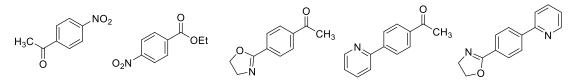


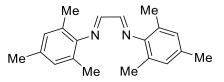
Figure S1. Scope of the substrates used in the study

2. Synthesis and Characterisation

2.1. Synthesis of Iridium (I) Complexes

Synthesis of 1,3-Bis-(2,4,6-trimethylphenyl)imidazolium chloride (IMes·HCl)^{S1}

N,*N*'-dimesitylethanediimine



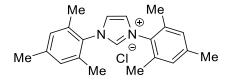
2,4,6-Trimethylaniline (84.2 mL, 0.60 mol, 2.0 equiv.) was dissolved in methanol (200 mL) and cooled to 0 °C, and a solution of 40% glyoxal in water (34.4 mL, 0.30 mol, 1.0 equiv.) with one or two drops of formic acid was added. The solution was warmed to room temperature and stirred for two days. The yellow suspension was filtrated and washed with a minimum volume of methanol and diethyl ether to afford N,N-dimesitylethanediimine (78.5 g, 0.27 mol, 90%) as a yellow powder, which was used immediately in the next step.

¹**H NMR** (400 MHz, CDCl₃) δ = 8.11 (s, 2H, CH=N), 6.92 (s, 4H, ArH), 2.30 (s, 6H, *p*-CH₃), 2.17 (s, 12H, *o*-CH₃).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 163.6, 147.6, 134.4, 129.1, 126.7, 20.9, 18.3.

NMR data are consistent with the literature.^{S10}

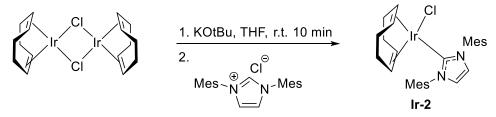
1,3-Bis-(2,4,6-trimethylphenyl)imidazolium chloride (IMes·HCl)^{S1}



Paraformaldehyde (8.05g, 0.27 mol, 1.0 equiv.) was suspended in a solution of 4M hydrochloric acid in dioxane (94 mL, 0.38 mol, 1.4 equiv.) and stirred until complete dissolution of the white solid. THF (300 mL) was added, followed by the slow addition of N,N'-dimesitylethanediimine (78.5 g, 0.27 mol, 1.0 equiv.). The resulting solution was stirred at 40 °C for 2 days. The suspension was cooled to room temperature and the white precipitate was collected by filtration, and washed with THF (100 mL) and diethyl ether (100 mL) to afford the crude product, which was recrystallized from a DCM/Et₂O mixture to afford 1,3-bis-(2,4,6-trimethylphenyl)imidazolium chloride (36.6 g, 0.11 mol, 40%) as a white powder.

¹**H NMR** (400 MHz, CDCl₃) δ = 10.64 (s, 1H, N-CH=N), 7.61 (d, *J* = 0.9 Hz, 2H), 7.02 (s, 4H, Ar), 2.33 (s, 6H, *p*-CH₃), 2.17 (s, 12H, *o*-CH₃).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 141.4, 139.2, 134.2, 130.7, 130.0, 124.8, 21.2, 17.7. NMR data are consistent with the literature.^{S1}



Bis(1,5-cyclooctadiene)diiridium(I) dichloride (500 mg, 0.75 mmol, 1 equiv.) and potassium *tert*-butoxide (167 mg, 1.50mmol, 2 equiv.) were added to a flame-dried Schlenk tube under argon and stirred under vacuum for 5 min. THF (12.5 mL) was added and the mixture was stirred under argon for 10 min. IMes·HCl (508 mg, 1.50 mmol, 2 equiv.) was then added and the resulting reaction mixture was stirred for 4 h. The solvent was removed *in vacuo*, and column chromatography (50% ethyl acetate in petroleum ether) afforded the title compound (730 mg, 1.14 mmol, 76 %) as yellow solid. The isolated catalyst was dried in a vacuum oven (40 °C, 1 mbar) for 24 h before use. This process was repeated batch-wise to obtain a of the quantity of **Ir-2** necessary for all competition studies and synthesis of **Ir-1** catalyst.

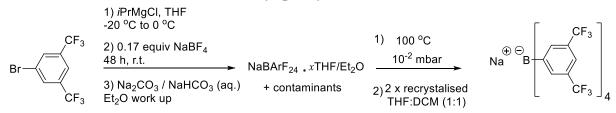
m.p. > 190 °C (decomposition)

¹**H** NMR (400 MHz, CDCl₃) δ = 7.04 – 6.96 (m, 4H, Ar-H), 6.95 (s, 2H, NCH=CHN), 4.19 – 4.12 (m, 2H, COD CH), 3.01 – 2.94 (m, 2H, COD CH), 2.36 (s, 12H, *o*-CH₃Ar), 2.16 (s, 6H, *p*-CH₃Ar), 1.78 – 1.59 (m, 4H, COD CH₂), 1.39 – 1.20 (m, 4H, COD CH₂).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 180.9, 138.8, 137.5, 136.2, 134.5, 129.7, 128.3, 123.4, 82.7, 51.6, 33.6, 29.1, 21.3, 19.8, 18.4.

NMR data are consistent with the literature.^{S7}

Sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate [Na(BArF₂₄)]^{S5,S6}



A 2.0 M solution of *i*-PrMgCl in THF (100 mL, 0.20 mol, 6.6 equiv.) was added dropwise over 45 min to a stirred solution of 1-bromo-3,5-bistrifluoromethylbenzene (30 mL, 0.17 mol, 5.8 equiv.) in THF (150 mL) chilled to -20 °C. After the reaction was allowed to warm from -20 °C to 0 °C over 1 h, NaBF₄ (3.3 g, 0.03 mol, 1.0 equiv.) was quickly added as a solid under a stream of N₂. The mixture then stirred for 48 h at 23 °C (under N₂). All work-up and purification procedures were then carried out under air. The contents of the flask were poured into a solution of Na₂CO₃ (44 g) and NaHCO₃ (22 g) in water (600 mL). This mixture was stirred vigorously for 1 h and then extracted with diethyl ether (4×200 mL). The combined organic layers were washed with brine and dried over anhydrous Na₂SO₄. After filtration of the mixture and rotary evaporation of the filtrate, the crude Na[BArF₂₄]·xTHF/Et₂O was dried at 100 °C/10⁻² mbar for 10 h to yield a tacky brown/yellow solid. Dissolving the resulting oily crude material in a 1:1 mixture of dichloromethane and tetrahydrofuran (30 mL) and cooling the mixture at -23 °C for 48 h yielded an off-white crystalline solid, which was then recrystallised again under the same conditions. Anhydrous Na[BArF₂₄]·THF (12.7 g, 0.014 mmol, 48%) was obtained as white solid by drying the resulting crystalline solid under vacuum ($< 10^{-2}$ mbar) for 10 h. Anhydrous Na[BArF₂₄] was stored under an atmosphere of argon.

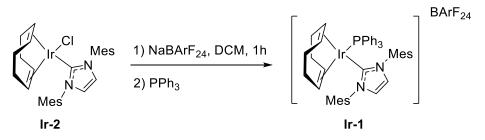
¹**H** NMR (400 MHz, acetone- d_6) δ = 7.79 (br, 8H, ortho-Ar-H), 7.67 (br, 4H, para-Ar-H).

¹³C{¹H} NMR (101 MHz, DMSO-*d*₆) δ = 161.0 (q, ¹*J*_{C-B} = 49.9 Hz, *ipso*-C), 134.0 (br, *ortho*-C), 128.5 (qq, ³*J*_{C-B} = 2.8 Hz, ²*J*_{C-F} = 32.2 Hz, *meta*-C), 124.0 (q, ¹*J*_{C-F} = 272 Hz, CF₃), 117.5 (br, *para*-C).

¹¹**B** NMR (128 MHz, acetone- d_6) $\delta = -6.65$.

¹⁹**F** NMR (376 MHz, acetone- d_6) $\delta = -63.3$.

NMR data are consistent with the literature.^{S5}



[Ir(COD)Cl(IMes)], Ir-2, (200 mg, 0.31 mmol, 1.0 equiv.) and NaBArF₂₄ (275 mg, 0.31 mmol, 1.0 equiv.) were added to a flame-dried Schlenk tube under an argon atmosphere. The solids were then dissolved in anhydrous DCM (10 mL) and stirred for 30 min. The triphenylphosphine ligand (82 mg, 0.31 mmol, 1.0 equiv.) was then added slowly, initiating an orange to red colour change. After a further 30 min stirring, the reaction mixture was filtered through celite and concentrated *in vacuo*, resulting in a red oil. This residue was purified by column chromatography (50% DCM in petroleum ether) to afford the title compound as a red crystalline solid (365 mg, 0.21 mmol, 68 %). The isolated catalyst was dried in a vacuum oven (40 °C, 1 mbar) for 24 h before use. This process was repeated batch wise to obtain the quantities of Ir-1 necessary for all competition studies.

m.p.: >150 °C (decomposition)

¹**H** NMR (400 MHz, CDCl₃) $\delta = 7.73 - 7.69$ (m, 8H, Ar- BArF₂₄), 7.51 (br, 4H, Ar-BArF₂₄), 7.45 - 7.39 (m, 3H, Ar-H), 7.31 - 7.24 (m, 8H, Ar-H and NCH=CHN), 7.15 - 7.07 (m, 6H, Ar-H), 7.02 (s, 2H, Ar-H), 6.66 (s, 2H, Ar-H), 4.39 - 4.32 (m, 2H, COD-CH), 3.38 - 3.31 (m, 2H, COD-CH), 2.34 (s, 6H, Ar-CH₃), 2.08 (s, 6H, Ar-CH₃), 1.75 (s, 6H, Ar-CH₃), 1.68 - 1.45 (m, 6H, COD-CH₂), 1.31 - 1.24 (m, 2H, COD-CH₂).

¹³C{¹H} NMR (101 MHz, CDCl₃) $\delta = 178.1$ (d, ${}^{2}J_{C-P} = 8.4$ Hz), 161.9 (q, ${}^{1}J_{C-B} = 49.9$ Hz), 140.2, 135.6, 135.2, 134.9, 134.8, 132.3, 132.2, 131.4, 131.3, 130.9, 130.6, 129.9, 129.5, 129.2, 128.9, 128.7, 128.6, 126.2, 124.7 (q, ${}^{1}J_{C-F} = 272$ Hz), 117.6, 80.6, 80.5, 78.7, 31.9, 30.3, 30.2, 21.2, 20.9, 19.0.

¹¹**B** NMR (128 MHz, CDCl₃) δ = - 6.64.

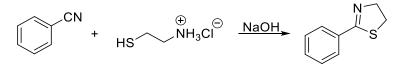
¹⁹**F NMR** (376 MHz, CDCl₃) δ = - 62.4.

³¹**P**{¹**H**} **NMR** (162 MHz, CDCl₃) δ = 16.4.

NMR data are consistent with the literature.^{S8}

2.2. Synthesis of Substrates

2-Phenylthiazoline^{S2}



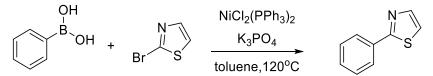
In air, a 10 mL round-bottom flask was charged with benzonitrile (0.50 g, 4.85 mmol), cysteamine hydrochloride (0.83 g, 7.28 mmol) and NaOH (40 mg, 0.97 mmol). The reaction was stirred at 80 °C for 2 hours under solvent-free conditions. The crude product was dissolved in ethyl acetate (2 mL) and water (10 mL) was added. The layers were separated and the aqueous layer was then extracted with ethyl acetate (3×10 mL). The combined organic layers were dried over MgSO₄, filtered and dried under vacuum to give the title compound (0.76 g, 4.66 mmol, 96%).

¹**H** NMR (400 MHz, CDCl₃) δ = 7.88 – 7.80 (m, 2H, Ar-H), 7.48 – 7.37 (m, 3H, Ar-H), 4.46 (t, *J* = 8.3 Hz, 2H, CH₂), 3.41 (t, *J* = 8.3 Hz, 2H, CH₂).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 169.0, 133.2, 131.4, 128.6, 128.5, 65.1, 33.7.

NMR data are consistent with the literature.^{S2}

2-Phenylthiazole

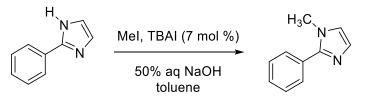


Phenylboronic acid (762 mg, 6.25 mmol, 1.25 equiv.), $[NiCl_2(PPh_3)_2]$ (262 mg, 0.40 mmol, 8 mol%), K₃PO₄ (1.59 g, 7.50 mmol, 1.5 equiv.) were added to a flame-dried two-necked roundbottom flask equipped with a stirrer bar and condenser. The system was evacuated for 5 minutes and filled with N₂, then dry toluene (10 mL) and 2-bromothiazole (820 mg, 5.0 mmol, 1 equiv.) were added. The reaction mixture was heated at reflux overnight (16 h) and was then cooled to room temperature before water (50 mL) and Et₂O (50 mL) were added. The aqueous layer was extracted with diethyl ether (3 × 50 mL). The combined organic layers were dried over MgSO₄, filtered, and the solvent was removed under vacuum. The residue was purified by column chromatography (10 % Et₂O in hexane) to afford the title compound as a yellow oil (163 mg, 1.01 mmol, 20 %).

¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.01 - 7.94$ (m, 2H, Ar-H), 7.87 (d, J = 3.3 Hz, 1H, Ar-H), 7.48 - 7.41 (m, 3H, Ar-H), 7.33 (d, J = 3.3 Hz, 1H, CH).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 168.6, 143.8, 133.8, 130.1, 129.1, 126.7, 118.9.

NMR data are consistent with the literature.^{S11}



Methyl iodide (0.48 mL, 7.6 mmol) was added to the biphasic mixture obtained from 2phenylimidazole (1.0 g, 6.9 mmol), tetra-*n*-butylammonium iodide (0.19 g, 0.51 mmol), 50% aqueous NaOH (30 mL) and toluene (30 mL). After stirring for 15 min at room temperature, the mixture was diluted with toluene (30 mL) and H₂O (30 mL). The organic phase was separated, dried (MgSO₄), and concentrated under reduced pressure to give pale yellow oil (1.03 g, 6.51 mmol, 97 %).

¹**H** NMR (400 MHz, CDCl₃) δ = 7.65 – 7.60 (m, 2H, Ar-H), 7.48 – 7.37 (m, 3H, Ar-H), 7.12 (d, *J* = 1.2 Hz, 1H, CH), 6.97 (d, *J* = 1.2 Hz, 1H, CH), 3.19 (s, 3H, CH₃).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 147.9, 130.5, 128.8, 128.8, 128.7, 128.3, 122.5, 34.6. NMR data are consistent with the literature. ^{S12}

2-Phenylpyrimidine

$$\begin{array}{c} OH \\ B \\ OH \\ Br \\ N \\ \end{array}$$

$$\begin{array}{c} PdCl_2(dppf) \\ Na_2CO_3 \\ \hline THF/H_2O (1:1) \\ 80 \ ^{\circ}C 24 \ h \end{array}$$

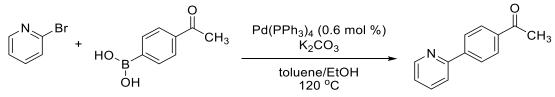
A flame dried 50 mL round-bottom flask was charged with $[PdCl_2(dppf)]$ (40 mg, 0.05 mmol), phenylboronic acid (611 mg, 5.0 mmol), Na₂CO₃ (1.23 g, 12.0 mmol) and suspended in premixed 1:1 solution of THF/H₂O (60 mL). Subsequently, 2-bromopyrimidine (646 mg, 4.0 mmol) was added and the reaction mixture was stirred at 80 °C overnight (16 h), and then cooled to room temperature. H₂O (30 mL) was added to the reaction mixture. The aqueous layer was extracted with diethyl ether (3 × 50 mL). The combined organic layers were dried over MgSO₄, filtered, and the solvent was removed under vacuum. The residue was purified by column chromatography (20 % Et₂O in hexane) to afford the title compound as a yellow oil (331 mg, 2.12 mmol, 53 %).

¹**H** NMR (400 MHz, CDCl₃) δ = 8.81 (d, *J* = 4.9 Hz, 2H, Ar-H), 8.49 – 8.42 (m, 2H, Ar-H), 7.52 – 7.47 (m, 3H, Ar-H), 7.18 (t, *J* = 4.8 Hz, 1H, Ar-H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 164.9, 157.4, 137.7, 130.9, 128.7, 128.3, 119.2.

NMR data are consistent with the literature.^{S11}

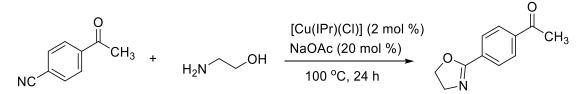
2-(4-Acetyl)phenylpyridine



A flame dried 50mL round-bottom flask was charged with [Pd(PPh₃)₄] (21 mg, 0.018 mmol), 4acetylphenylboronic acid (738 mg, 4.50 mmol), potassium carbonate (1.21 g, 9.0 mmol). The solids were suspended in premixed toluene/ethanol (3:2) (30 mL). Subsequently, 2-bromopyridine (0.28 mL, 3.0 mmol) was added and the reaction mixture was stirred for 24 h at 120 °C before being allowed to cool to room temperature. Water (100 mL) was added, the layers were separated, and the aqueous layer was extracted with diethyl ether (3 x 50 mL). The combined organic layers were dried over MgSO₄, filtered, and then the solvent was removed under vacuum yielding the crude yellow solid. The crude product was dissolved in DCM (5 mL) and passed through a short pad of silica which was then washed with further DCM. The solvent was evaporated, and the residue was crystallised from a DCM/pentane mixture to afford the product as a white solid (517 mg, 2.62 mmol, 87%).

¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.77 - 8.71$ (m, 1H, Ar-H), 8.13 - 8.09 (m, 2H, Ar-H), 8.09 - 8.04 (m, 2H, Ar-H), 7.82 - 7.78 (m, 2H, Ar-H), 7.33 - 7.27 (m, 1H, Ar-H), 2.65 (s, 3H, CH₃). ¹³C{¹H} NMR (101 MHz, CDCl₃) $\delta = 198.0$, 150.1, 137.1, 129.0, 127.2, 123.1, 121.2, 26.9. NMR data are consistent with the literature.^{S13}

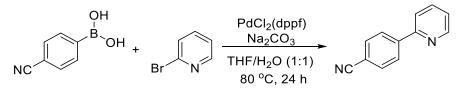
2-(4-Acetyl)phenyloxazoline^{S5}



A flame dried vial was charged with [Cu(IPr)(Cl)] (10 mg, 0.02 mmol), 4-acetylbenzonitrile (145 mg,1.0 mmol) and NaOAc (16 mg, 0.2 mmol), and evacuated and backfilled with N₂. Ethanolamine (0.24 mL, 4.0 mmol) was added and the reaction was stirred at 100 °C for 16 h under solvent-free conditions. The reaction mixture was cooled to room temperature, dissolved in DCM and passed through a short pad of silica with DCM as the eluent. The solvent was removed under vacuum yielding a pale yellow solid (60 mg, 0.32 mmol, 32 %).

¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.06 - 7.97$ (m, 4H, Ar-H), 4.47 (t, $J = 9.6, 2H, CH_2$), 4.10 (t, $J = 9.6, 2H, CH_2$), 2.63 (s, 3H, CH₃).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 197.7, 139.1, 132.0, 128.5, 128.4, 68.0, 55.3, 26.9. NMR data are consistent with the literature.^{S14} 2-(4-Cyano)phenylpyridine



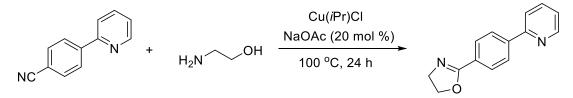
A flame dried 50 mL round-bottom flask was charged with $[PdCl_2(dppf)]$ (31 mg, 0.03 mmol), 4cyanophenylboronic acid (856 mg, 5.83 mmol), Na₂CO₃ (1.30 g, 12.0 mmol). The solids were suspended in premixed THF/H₂O (1:1) (40 mL). Subsequently, 2-bromopyridine (0.38 mL, 4.0 mmol) was added, the reaction mixture was stirred at 80 °C for 16 h and then cooled to room temperature. H₂O (30 mL) was added to the reaction mixture, the layers were separated, and the aqueous layer was extracted with EtOAc (2 × 30 mL). The combined organic solution was dried over MgSO₄. The solvent was removed under reduced pressure, and the residue was purified by recrystallisation from a DCM/pentane mixture to afford the title compound as yellow solid (554 mg, 3.07 mmol, 77%).

¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.75 - 8.67$ (m, 1H), 8.12 - 8.07 (m, 2H), 7.82 - 7.72 (m, 4H), 7.33 - 7.27 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ = 155.2, 150.1, 143.5, 137.2, 132.6, 127.5, 123.4, 121.1, 118.9, 112.5.

NMR data are consistent with the literature.^{S15}

2-(4-(Pyridin-2-yl)phenyl)-4,5-dihydrooxazole^{S4}



A flame dried vial was charged with [Cu(Cl)(IPr)] (135 mg, 0.27 mmol), 2-(4cyano)phenylpyridine (500 mg, 2.77 mmol) and NaOAc (114 mg, 1.39 mmol), and evacuated and backfilled with N₂. Ethanolamine (0.34 mL, 4.0 mmol) was added and the reaction was stirred at 100 °C for 16 h under solvent-free conditions. The reaction mixture was cooled to room temperature, dissolved in DCM and transferred to a separating funnel containing brine (200 mL). The product was extracted with DCM (2×50 mL) and dried over MgSO₄, filtered, and the solvent was removed under vacuum. The residue was purified by column chromatography (70 % EtOAc in hexane) to afford the title compound as pale pink solid (248 mg, 1.10 mmol, 40 %). **m.p.**: 120-125 °C

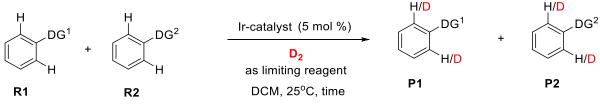
¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.76 - 8.68$ (m, 1H), 8.11 - 8.01 (m, 4H, Ar-H), 7.81 - 7.74 (m, 2H), 7.30 - 7.23 (m, 1H), 4.46 (t, J = 9.5, 2H, CH₂), 4.09 (t, J = 9.5, 2H, CH₂).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ = 164.5, 156.5, 149.9, 142.0, 136.9, 128.7, 128.2, 126.9, 122.7, 120.9, 67.7, 55.1.

Anal. Calculated for C₁₄H₁₂N₂O: C, 74.98; H, 5.39; N, 12.49. Found: C, 74.28; H, 5.35; N, 12.34.

FTIR (neat): 3286, 3047, 2970, 2931, 2990, 2877, 1642, 1572, 1465, 1256, 1071, 1016, 940, 862, 785,731, 699, 615 cm⁻¹.

3. Intermolecular Competition Experiments 3.1. General Information



Reaction conditions

The relative rates of hydrogen-deuterium exchange reactions have been determined by competition experiments, where equimolar quantities of each of the two substrates bearing different DGs and catalytic amounts of iridium complexes were treated with a limiting amount of D_2 in DCM at 25°C. As a limiting amount of D_2 gas should be used to avoid full conversion of the substrates, its volume was controlled by adding the required amount of solvent.

The volume of 0.10 mmol of the D_2 gas can be calculated according to the ideal gas law (eq. S-1).

$$PV = nRT \tag{S-1}$$

 $n_{(max)} = 0.10 \text{ mmol}$ T = 298 K (25 °C) $R = 0.0821 \text{ L} \cdot \text{atm} \cdot \text{K}^{-1} \cdot \text{mol}^{-1}$ P = 1 atm $V_{(max)} = nRT/P = 0.10 \times 0.0821 \times 298 / 1 = 2.45 \text{ mL}$

As the volume of J. Young Schlenk flasks (Figure S2) used for the competition experiments is approximately 8 mL, the use of 6.0 mL of the solvent will lead to less than 1 equivalent of deuterium gas. None of the substrates in the study undergo complete deuteration under these conditions, which are quite different for those used in most HIE experiments where complete deuteration at one site is desired.



Figure S2. The J. Young Schlenk flasks used for competition experiments.

General Procedure (GP1)

The two substrates of interest (0.10 mmol each) were added to one J. Young Schlenk flask, along with the catalyst of choice (0.005 mmol, unless otherwise noted) in air. DCM (6 mL) was added in such a way to rinse the inner walls of the flask. The flask was then sealed (with the gas inlet left open) under air before being cooled in a dry ice-acetone bath. The flask was evacuated and flushed with deuterium three times via a balloon. The gas inlet was then closed with fast thread tap, creating a sealed atmosphere of deuterium. After sealing the flask, it was placed in the thermostated water bath, and the reaction timer was started. The reaction mixture was stirred at 25 °C (1 h for catalyst Ir-1 and 16 h for catalyst Ir-2) before the removal of the excess deuterium and the opening of the flask to air. The reaction mixture was quenched with few drops of MeCN and transferred to a single necked flask together with washings (DCM) before removing the solvent under reduced pressure. For NH-containing substrates (benzamide, benzenesulfonamide, acetanilide, phenylimidazol(in)e) the residue was directly analysed by ¹H NMR. For other substrates, the residue was dissolved in a small portion of 1:1 mixture of petroleum ether and diethyl ether (or EtOAc) and passed through a short plug of silica, eluting with a 1:1 mixture of petroleum ether and diethyl ether (or EtOAc) $(3 \times 2 \text{ mL})$. The solvent was evaporated again under reduced pressure and the residue was analysed by ¹H NMR.

NMR spectrometer parameters are as follows: the relaxation delay was set to 20 s and the number of scans to four or higher when needed. After careful phasing and baseline correction, the integration of the signals was carried out manually.

Determination of Competition Rate Constants

The level of deuterium incorporation in the substrates was determined from the obtained ¹H NMR spectra (eq. S-2). The integrals were calibrated against a peak corresponding to a position which does not undergo labelling. In addition, the calibration signal was chosen to have as little overlap as possible with other peaks.

%D = 100 -
$$\left(\frac{\text{residual integral}}{\text{number of labelling sites}} \times 100 \% \right)$$
 (S-2)

Based on the ratio between the initial and remaining concentrations of non-deuterated substrates **R**, the competition constants κ were determined (eq. S-3).

$$\kappa = \frac{k_1}{k_2} = \frac{\log([\mathbf{R1}]_0 / [\mathbf{R1}]_1)}{\log([\mathbf{R2}]_0 / [\mathbf{R2}]_1)}$$
(S-3)

Initial concentrations of the substrates are defined by mass balance (eq. S-4).

$$[\mathbf{R}]_0 = [\mathbf{R}]_t + [\mathbf{P}]_t \tag{S-4}$$

The relative concentrations $[\mathbf{R}]_0$ and $[\mathbf{R}]_t$ were derived using equation (S-5) from the residual integral $I_{R(t)}$ of the peak corresponding to H/D positions, and the integral $I_{R(0)}$ of the peak used for calibration, which corresponds to both remaining starting material and deuterated product.

$$\frac{[\mathbf{R}]_{0}}{[\mathbf{R}]_{t}} = \frac{\mathbf{I}_{R(0)} / \mathbf{N}}{\mathbf{I}_{R(t)} / \mathbf{N}}$$
(S-5)

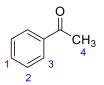
where N is the number of protons contributing to the corresponding peak.

Each combination was analysed three times, and the competition constants are the average of all runs. Standard error of the mean was determined for each averaged competition constant and are summarised in Tables S42-43).

The tables given below for each competition experiment (Table S1 to Table S41) summarize the amounts of the reagents used, integrals from the NMR spectra (Figures S3-S175) used to calculate the relative concentrations of substrates and the values of the competition constants κ .

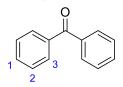
Spectral details for unlabelled substrates used in intermolecular competition studies

Acetophenone

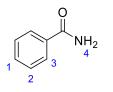


¹**H** NMR (400 MHz, CDCl₃) δ = 7.99 – 7.93 (m, 2H, H-3), 7.60 – 7.53 (m, 1H, H-1), 7.51 – 7.42 (m, 2H, H-2), 2.61 (s, 3H, H-4). Incorporation expected at δ 7.99 – 7.93 ppm (H-3) Determined against integral at δ 2.61 ppm (H-4) ¹**H** NMR (400 MHz, DMSO-*d*₆) δ = 7.98 – 7.94 (m, 2H, H-1), 7.67 – 7.61 (m, 1H, H-3), 7.55 – 7.50 (m, 2H, H-2), 2.58 (s, 3H, H-4). Incorporation expected at δ 7.98 – 7.96 ppm (H-3) Determined against integral at δ 2.58 ppm (H-4)

Benzophenone



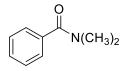
Benzamide



¹**H NMR** (400 MHz, CDCl₃) δ = 7.84 – 7.78 (m, 4H, H-3), 7.63 – 7.53 (m, 2H, H-1), 7.52 – 7.45 (m, 4H, H-2). Incorporation expected at δ 7.84 – 7.78 ppm (H-3) Determined against integral at δ 7.63 – 7.53 ppm (H-1)

¹**H NMR** (400 MHz, CDCl₃) δ = 7.86 – 7.77 (m, 2H, H-3), 7.57 – 7.48 (m, 1H, H-1), 7.48 – 7.39 (m, 2H, H-2), 6.24 (bs, 2H, H-4) Incorporation expected at δ 7.86 – 7.77 ppm (H-3) Determined against integral at δ 7.48 – 7.39 ppm (H-2) ¹**H NMR** (400 MHz, DMSO- *d*₆) δ = 7.96 (bs, 1H, H-4), 7.90 – 7.85 (m, 2H, H-3), 7.54 – 7.48 (m, 1H, H-1), 7.48 – 7.41 (m, 2H, H-2), 7.35 (s, 1H, H-4) Incorporation expected at δ 7.90 – 7.85 ppm (H-3) Determined against integral at δ 7.48 – 7.41 ppm (H-2)

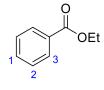
N,N-Dimethylbenzamide



¹**H NMR** (400 MHz, CDCl₃) δ = 7.42 – 7.36 (m, 5H, Ar-H), 3.17 – 2.88 (m, 6H, 2 × CH₃). Incorporation expected at δ 7.43 – 7.36 ppm

Determined against integral at δ 3.17 – 2.88 ppm

Ethylbenzoate



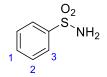
Nitrobenzene



¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.08 - 8.02$ (m, 2H, H-3), 7.58 - 7.52 (m, 1H, H-1), 7.47 - 7.40 (m, 2H, H-2), 4.38 (q, J = 7.1 Hz, 2H, CH₂), 1.40 (t, J = 7.1 Hz, 3H, CH₃). Incorporation expected at $\delta 8.07 - 8.03$ ppm (H-3) Determined against integral at $\delta 4.38$ ppm (OCH₂CH₃)

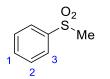
¹**H NMR** (400 MHz, CDCl₃) $\delta = 8.26 - 8.20$ (m, 2H, H-3), 7.73 - 7.66 (m, 1H, H-1), 7.58 - 7.51 (m, 2H, H-2). Incorporation expected at δ 8.26 - 8.20 ppm (H-3) Determined against integral at δ 7.73 - 7.66 ppm (H-1) ¹**H NMR** (400 MHz, DMSO- *d*₆) $\delta = 8.30 - 8.16$ (m, 2H, H-3), 7.90 - 7.79 (m, 1H, H-1), 7.73 - 7.61 (m, 2H, H-2). Incorporation expected at δ 8.30 - 8.16 ppm (H-3) Determined against integral at δ 7.73 - 7.61 ppm (H-2)

Benzenesulfonamide



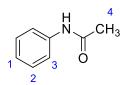
¹**H NMR** (400 MHz, DMSO- d_6) $\delta = 7.87 - 7.80$ (m, 2H, H-3), 7.64 - 7.54 (m, 3H, H-1 and H-2), 7.36 (bs, 2H, NH₂) Incorporation expected at δ 7.87 – 7.80 ppm (H-3) Determined against integral at δ 7.64 – 7.54 ppm (H-1+H-2)

(Methylsulfonyl)benzene

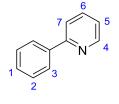


¹**H NMR** (400 MHz, DMSO- d_6) $\delta = 7.95 - 7.92$ (m, 2H, H-3), 7.76 -7.72 (m, 1H, H-1), 7.68 – 7.64 (m, 2H, H-2), 3.21 (s, 3H, H-4).

Acetanilide



2-phenylpyridine



Incorporation expected at δ 7.95 – 7.92 ppm (H-3) Determined against integral at δ 7.68 – 7.64 ppm (H-2)

¹**H** NMR (400 MHz, DMSO- d_6) $\delta = 9.90$ (bs, 1H, NH), 7.60 – 7.55 (m, 2H, H-3), 7.30 – 7.24 (m, 2H, H-2), 7.04 – 6.99 (m, 1H, H-1), 2.04 (s, 3H, H-4). Incorporation expected at δ 7.60 – 7.55 ppm (H-3) Determined against integral at δ 2.04 ppm (H-4)

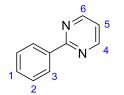
¹**H NMR** (400 MHz, CDCl₃) $\delta = 8.73 - 8.67$ (m, 1H, H-4), 8.02 - 7.98 (m, 2H, H-3), 7.78 – 7.70 (m, 2H, H-6 and H-7), 7.51 – 7.45 (m, 2H, H-2), 7.45 -7.39 (m, 1H, H-1), 7.25 - 7.21 (m, 1H, H-5).

Incorporation expected at $\delta 8.02 - 7.98$ ppm (H-3)

Determined against integral at δ 8.73 – 8.67 ppm (H-4) or at δ 7.78 – 7.70 (H-6+H-7) depending on the competition partner.

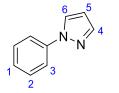
¹**H NMR** (400 MHz, DMSO- d_6) $\delta = 8.69 - 8.65$ (m, 1H, H-4), 8.11 - 8.06(m, 2H, H-3), 7.97 - 7.93 (m, 1H, H-7), 7.90 - 7.84 (m, 1H, H-6), 7.52 -7.46 (m, 2H, H-2), 7.46 – 7.41 (m, 1H, H-1), 7.38 – 7.32 (m, 1H, H-5). Incorporation expected at $\delta 8.11 - 8.06$ ppm (H-3) Determined against integral at δ 7.90 – 7.84 ppm (H-6)

2-Phenylpyrimidine

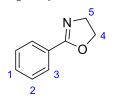


¹**H** NMR (400 MHz, CDCl₃) $\delta = 8.81$ (d, J = 4.9 Hz, 2H, H-4), 8.48 - 8.43(m, 2H, H-3), 7.52 – 7.48 (m, 3H, H-1 and H-2), 7.18 (t, J = 4.9 Hz, 1H, H-5)

1-phenylpyrazole



2-phenyloxozoline



Incorporation expected at δ 8.48 – 8.43 ppm (H-3) Determined against integral at δ 7.18 ppm (H-5) ¹**H NMR** (400 MHz, CDCl₃) δ = 7.92 (d, J = 2.2 Hz, 1H, H-6), 7.75 – 7.68

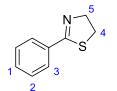
(m, 3H, H-3 and H-4), 7.48 – 7.43 (m, 2H, H-2), 7.32 – 7.26 (m, 1H, H-1), 6.49 - 6.45 (m, 1H, H-5). Incorporation expected at δ 7.75 – 7.68 ppm (H-3)

Determined against integral at δ 7.92 ppm (H-6)

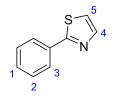
¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.97 - 7.93$ (m, 2H, H-3), 7.50 - 7.44 (m, 1H, H-1), 7.43 – 7.37 (m, 2H, H-2), 4.43 (t, J = 9.5 Hz, 2H, H-4), 4.06 (t, J = 9.5 Hz, 2H, H-5).

Incorporation expected at δ 7.97 – 7.93 ppm (H-3) Determined against integral at δ 4.43 ppm (H-4)

2-phenylthiazoline



2-phenylthiazole



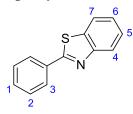
¹**H NMR** (400 MHz, CDCl₃) δ = 7.86 – 7.81 (m, 2H, H-3), 7.48 – 7.37 (m, 3H, H-1 and H-2), 4.46 (t, *J* = 8.3 Hz, 2H, H-4), 3.41 (t, *J* = 8.3 Hz, 2H, H-5).

Incorporation expected at δ 7.86 – 7.81 ppm (H-3) Determined against integral at δ 4.46 ppm (H-4)

¹**H NMR** (400 MHz, CDCl₃) $\delta = 8.00 - 7.94$ (m, 2H, H-3), 7.87 (d, J = 3.3 Hz, 2H, H-4), 7.48 - 7.42 (m, 3H, H-2 and H-1), 7.33 (d, J = 3.3 Hz, 2H, H-5).

Incorporation expected at δ 8.00 – 7.94 ppm (H-3) Determined against integral at δ 7.87 or 7.33 ppm (H-4 or H-5)

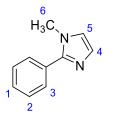
2-phenylbenzothiazole



¹**H** NMR (300 MHz, CDCl₃) $\delta = 8.14 - 8.06$ (m, 3H, H-3 and H-4), 7.91 (d, J = 8.0 Hz, 1H, H-7), 7.53 - 7.48 (m, 4H, H-2 and H-5), 7.41-7.37 (m, 1H, H-6).

Incorporation expected at $\delta 8.14 - 8.06$ ppm (H-3) Incorporation determined against $\delta 7.91$ ppm (H-7)

1-methyl-2-phenylimidazole



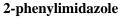
¹**H NMR** (400 MHz, CDCl₃) $\delta = 7.65 - 7.60$ (m, 2H, H-3), 7.47 - 7.34 (m, 3H, H-1 and H-2), 7.12 (d, J = 1.2 Hz, 1H, H-5), 6.97 (d, J = 1.2 Hz, 1H, H-4), 3.19 (s, 3H, H-6).

¹**H NMR** (400 MHz, DMSO- d_6) δ = 12.49 (bs, 1H, NH), 7.98 – 7.92 (m, 2H, H-3), 7.48 – 7.39 (2H, H-2), 7.36 – 7.29 (m, 1H, H-1), 7.13 (s, 2H, H-4 and

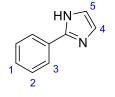
Incorporation expected at δ 7.65 – 7.60 ppm (H-3) Incorporation determined against δ 7.12 ppm (H-5)

Incorporation expected at δ 7.98 – 7.92 ppm (H-3)

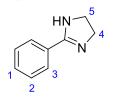
Incorporation determined against δ 7.13 ppm (H-4+H-5)



H-5).



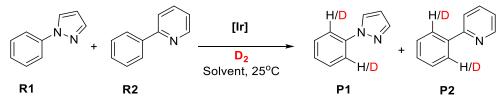
2-phenylimidazole



¹**H NMR** (400 MHz, DMSO-*d*₆) δ = 7.86 – 7.80 (m, 2H, H-3), 7.49 – 7.36 (m, 3H, H-1 and H-2), 3.60 (s, 4H, H-4 and H-5). Incorporation expected at δ 7.86 – 7.80 ppm (H-3) Incorporation determined against δ 3.60 ppm (H-4+H-5)

3.2. Effects of the Reaction Conditions on Competition Rate Constants

The competition labelling of 2-phenylpyridine and 1-phenylpyrazole was chosen as a model reaction to test the influence of catalyst loading, reaction times and solvent on the competition rate constants κ .



Mass of reagents: 1-Phenylpyrazole (14.4 mg, 0.1 mmol); 2-Phenylpyridine (15.5 mg, 0.1 mmol); for catalyst **Ir-2** (2.1 mg for 2.5 mol %, 4.3 mg, 5 mol. %, 6.4 mg for 7.5 mol %, 8.6 mg for 10 mol %); for catalyst **Ir-1** (8.7 mg, 5 mol %); Volume (solvent) = 6.0 mL Deuteration expected at δ (**R1**) = 7.78 – 7.67 ppm and at δ (**R2**) = 8.04 – 7.97 ppm Determined against integral at δ 6.50 – 6.42 for **R1** and at δ 8.72 – 8.68 for **R2** *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, CDCl₃) $\delta = 8.72 - 8.68$ (m, 1H, **R2**), 8.04 - 7.97 (m, 2H/D **R2**), 7.92 (d, J = 2.4 Hz, 1H, **R1**), 7.78 - 7.67 (m, 2H, **R2**, 1H, **R1**, 2H/D **R1**), 7.52 - 7.38 (3H, **R2** and 2H, **R1**), 7.31 - 7.26 (m, 1H, **R1**), 7.24 - 7.20 (m, 1H, **R2**), 6.50 - 6.42 (m, 1H, **R1**)

Catalyst loading

Competition experiments between 2-phenylpyridine and 1-phenylpyrazole with different loadings of the catalyst **Ir-2** (2.5 to 10 mol %) were performed in DCM (6.0 mL) following General Procedure GP1 for intermolecular competition experiments (time (t) = 16 h).

Entry	catalyst loading (mol %)	$I_{R1(t)} \\ N = 2H$	$\begin{split} I_{R1(0)} \\ N = 1 H \end{split}$	$%D_{R1}$	$I_{R2(t)} \\ N = 2H$	$\begin{split} I_{R2(0)} \\ N = 1 H \end{split}$	%D _{R2}	κ
1	2.5	1.40 ^a	1.00	30	1.32	0.86	23	1.35
2	5.0	1.29 ^b	1.00	36	1.22	0.85	28	1.32
3	7.5	1.48 °	1.00	26	1.43	0.94	24	1.10
4	10	1.17^{d}	1.00	42	1.09	0.91	40	1.05

Table S1. Competition labelling of 1-phenylpyrazole and 2-phenylpyridine using different loadings ofcatalyst Ir-2.

 ${}^{a}I_{R1(t)} = 4.12 - 1.00 - (0.86 \times 2); {}^{b}I_{R1(t)} = 3.99 - 1.00 - (0.85 \times 2);$

^c $I_{R1(t)} = 4.36 - 1.00 - (0.94 \times 2);$ ^d $I_{R1(t)} = 3.99 - 1.00 - (0.91 \times 2)$

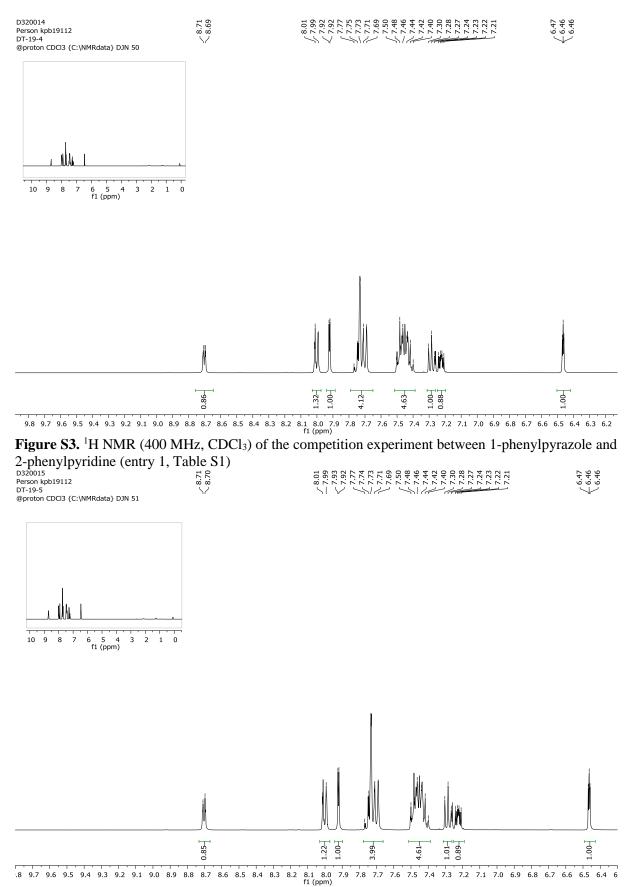


Figure S4. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 2, Table S1)

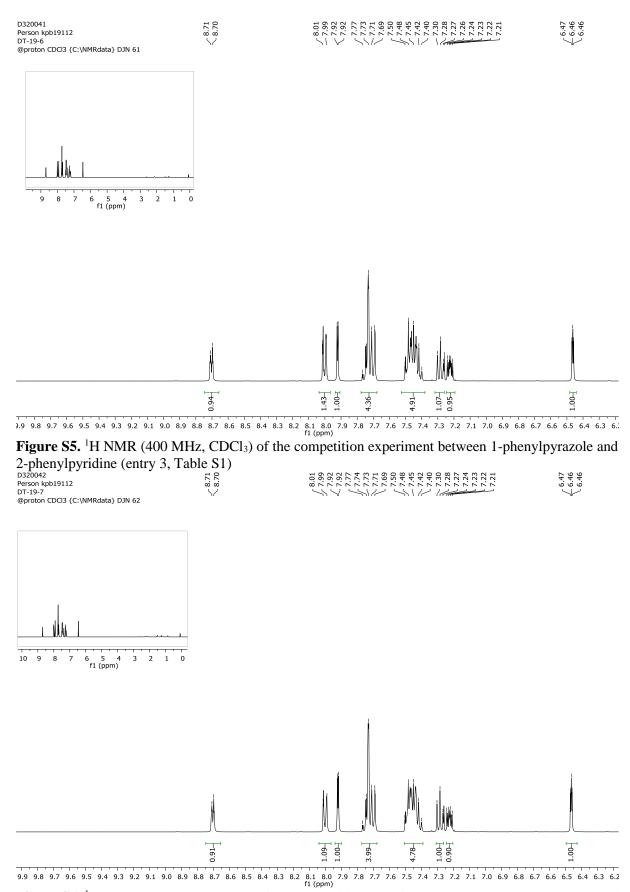


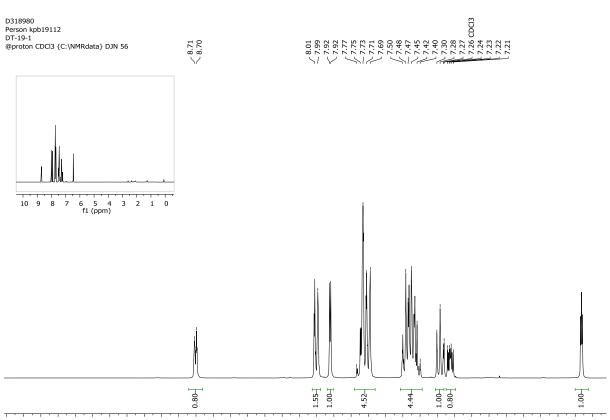
Figure S6. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 4, Table S1)

Time-dependence

Competition experiments between 1-phenylpyrazole and 2-phenylpyridine with the catalyst **Ir-2** (5 mol %) were performed in DCM (6.0 mL) over different time periods following the General Procedure GP1 for intermolecular competition experiments.

Table S2. Competition labelling of 1-phenylpyrazole and 2-phenylpyridine using catalyst **Ir-2** over different time periods.

Entry	reaction time	$\begin{split} I_{R1(t)} \\ N = 2H \end{split}$	$\begin{split} I_{R1(0)} \\ N = 1 H \end{split}$	%D _{R1}	$\begin{split} I_{R2(t)} \\ N = 2H \end{split}$	$\begin{split} I_{R2(0)} \\ N = 1 H \end{split}$	$%D_{R2}$	κ
1	1h	1.92ª	1.00	4	1.55	0.80	3	1.29
2	2h	1.89 ^b	1.00	6	1.75	0.91	4	1.44
3	16h	1.65 ^c	1.00	18	1.75	1.00	13	1.44
${}^{a}I_{R1(t)} = 4$.52-1.00-(0	0.80×2); ^b I _F	$a_{1(t)} = 4.71 - 1$.00-(0.91	×2); ^c $I_{R1(t)}$ =	= 4.65-1.00	-(1.00×2);	



^{3,8} 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 6.7 6.6 6.5 6.4 **Figure S7.** ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 1, Table S2)

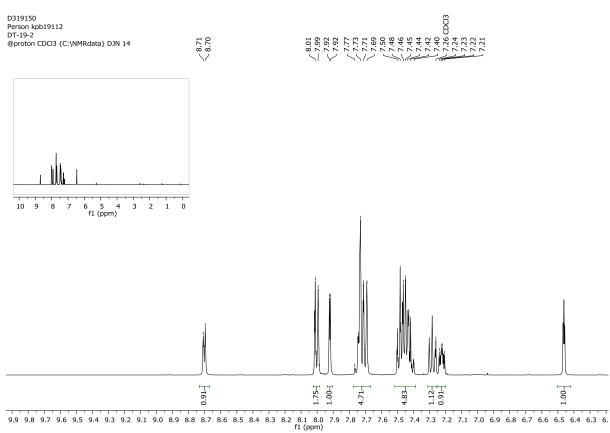


Figure S8. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 2, Table S2)

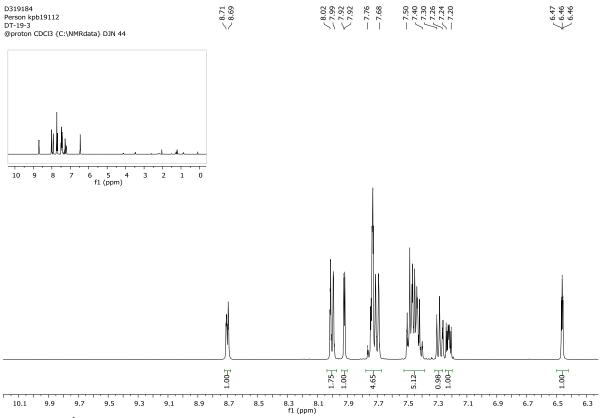


Figure S9. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 3, Table S2)

Solvent effects

Competition experiments between 1-phenylpyrazole and 2-phenylpyridine with the catalyst **Ir-1** (5 mol %) were performed in various solvents (6.0 mL) following the General Procedure GP1 for intermolecular competition experiments (time (t) = 1 h).

Solvent	Entry	$\begin{split} I_{R1(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R1(0)} \\ N = 1 H \end{split}$	$%D_{R1}$	$\begin{split} I_{R2(t)} \\ N = 2 H \end{split}$	$I_{R2(0)}$ $N = 1H$	$%D_{R2}$	κ
DCM	1	1.26 ^a	1.00	37	1.74	1.18	26	1.49
	2	0.90 ^b	1.00	55	1.38	1.07	36	1.78
	3	0.92 ^c	1.00	54	1.40	1.08	35	1.84
							Avera	ge <i>κ</i> = 1.71
$a \mathbf{I}_{R1(t)}$	= 4.62 - 1	.00-(1.18×2)); ${}^{b}I_{R1(t)} = 4.0$	4-1.00-(1.07×2); ° l	$I_{R1(t)} = 4.08 -$	-1.00-(1.08	3×2);
THF	1	1.15 ^a	1.00	43	1.32	1.01	35	1.30
	2	1.22 ^b	1.00	39	1.36	1.00	32	1.28
	3	0.95°	1.00	53	1.13	0.96	41	1.40
							Avera	ge $\kappa = 1.33$
^a I _{R1(t}	= 4.17 - 1	.00-(1.01×2)); ${}^{b}I_{R1(t)} = 4.2$	2-1.00-(1.00×2); ° l	$I_{R1(t)} = 3.87 -$	-1.00-(0.96	5×2);
Et ₂ O	1	1.17 ^a	1.00	42	1.39	1.05	34	1.30
	2	0.73 ^b	1.00	64	0.91	0.97	53	1.33
	3	0.78°	1.00	61	1.03	0.96	46	1.51
							Avera	ge <i>κ</i> = 1.38
$a \mathbf{I}_{\mathbf{R}1(t)}$	= 4.27-1.	00-(1.05×2)	; ${}^{b}I_{R1(t)} = 3.6$	67-1.00-(0.97×2); °	$I_{R1(t)} = 3.66$	-1.00-(0.9	6×2);
Toluene	1	0.98 ^a	1.00	51	1.45	1.09	33	1.75
	2	0.98^{b}	1.00	51	1.42	1.06	33	1.78
	3	1.11 ^c	1.00	45	1.39	0.98	29	1.71
							Avera	ge <i>κ</i> = 1.75
$a \mathbf{I}_{R1(t)}$	= 4.16–1.	00-(1.09×2)	; ${}^{b}I_{R1(t)} = 4.1$	0-1.00-(1.06×2); °	$I_{R1(t)} = 4.07$ -	-1.00-(0.9	8×2);
EtOAc	1	1.38 ^a	1.00	31	1.28	0.98	35	0.87
	2	0.90 ^b	1.00	55	0.84	1.00	58	0.92
	3	0.88 ^c	1.00	56	0.82	1.00	59	0.92
							Avera	ge $\kappa = 0.90$
$^{a}I_{R1(t)}$	= 4.34–1.	00-(0.98×2)	; ^b $I_{R1(t)} = 3.9$	0-1.00-((1.00×2); °	$I_{R1(t)} = 3.88$	-1.00-(1.0	0×2);

Table S3. Competition labelling of 1-phenylpyrazole and 2-phenylpyridine using catalyst**Ir-1** indifferent solvents.

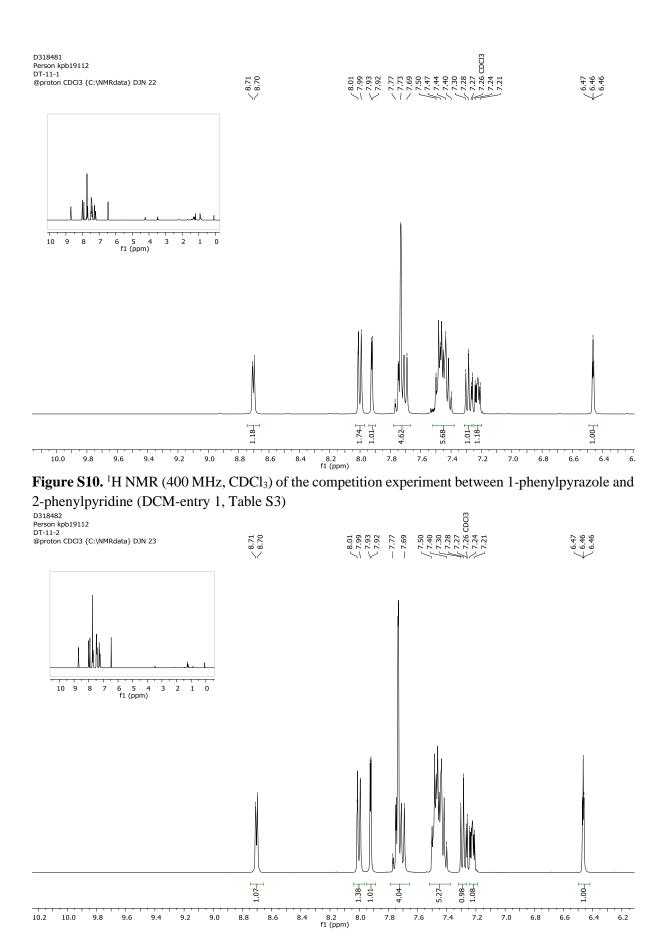


Figure S11. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (DCM-entry 2, Table S3)

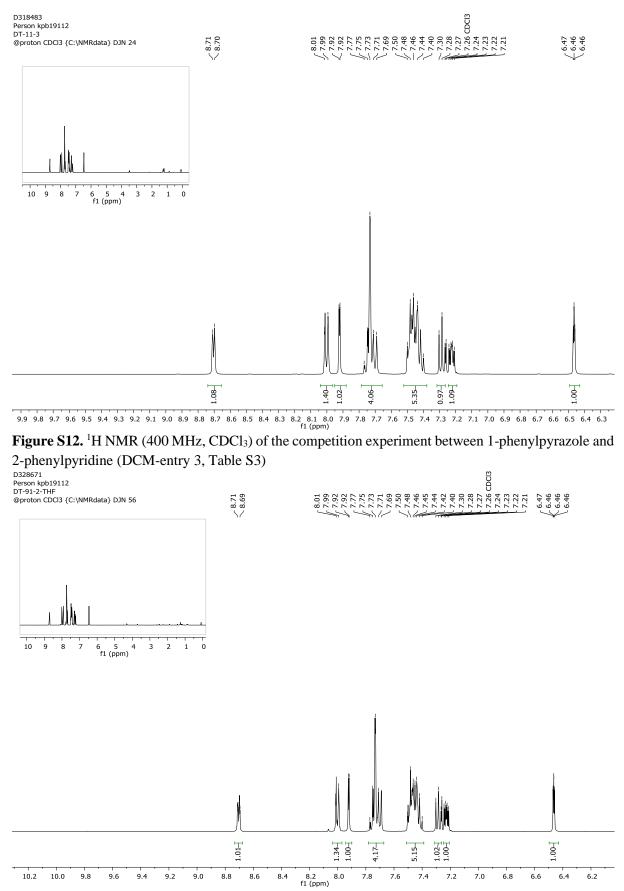
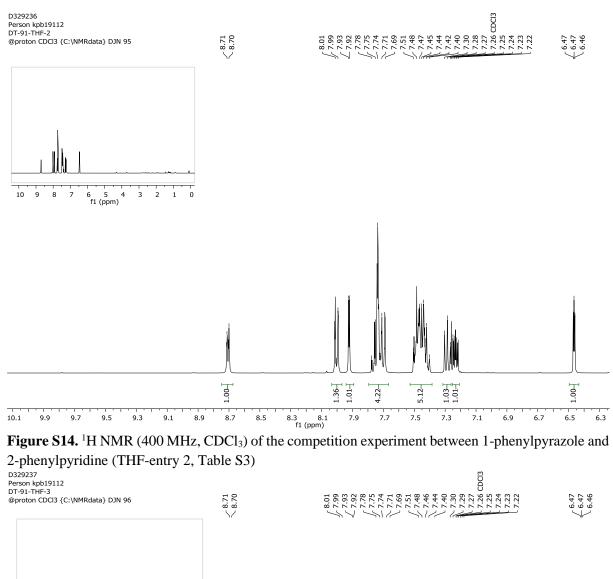


Figure S13. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (THF-entry 1, Table S3)



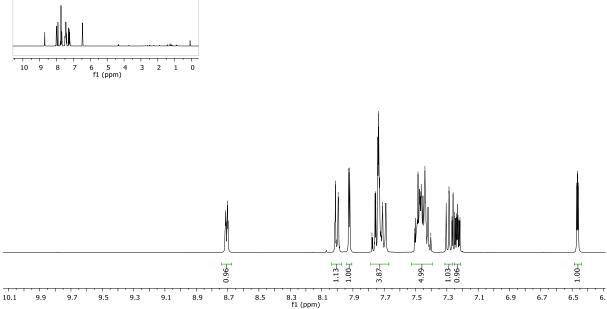


Figure S15. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (THF-entry 3, Table S3)

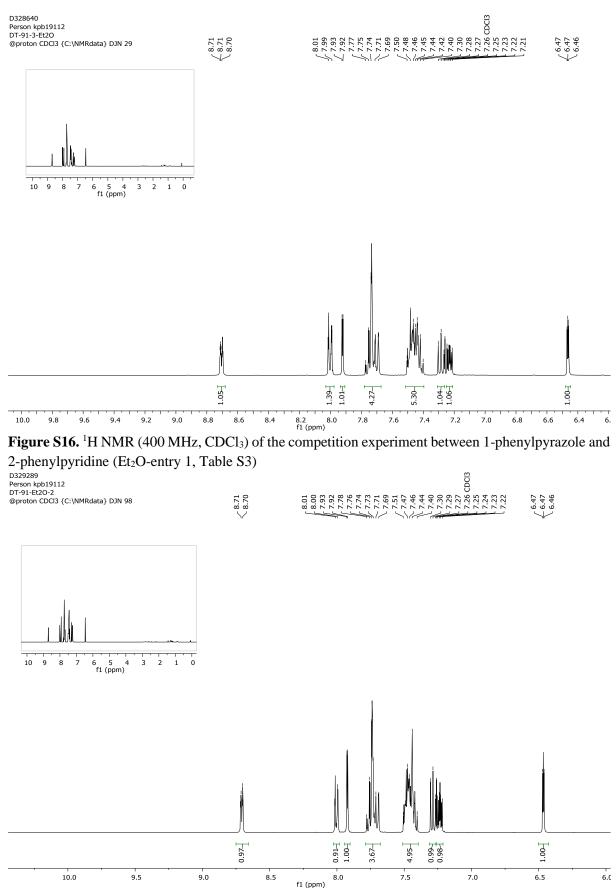


Figure S17. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (Et₂O-entry 2, Table S3)

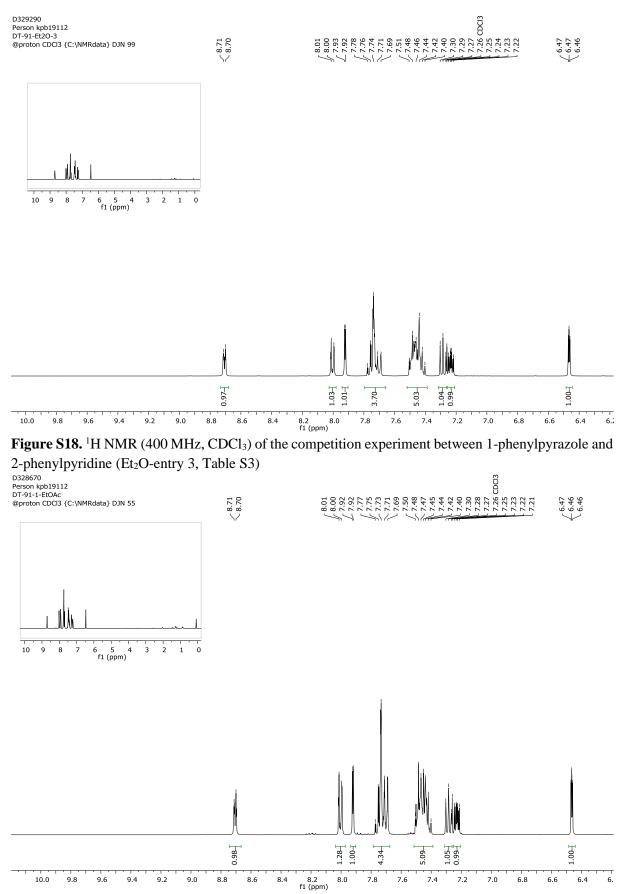


Figure S19. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (EtOAc-entry 1, Table S3)

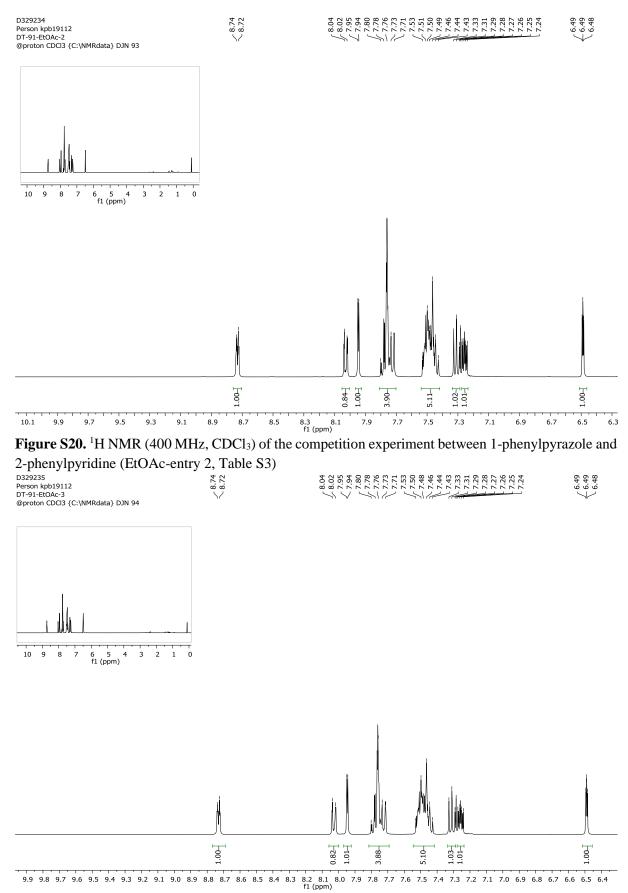


Figure S21. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (EtOAc-entry 3, Table S3)

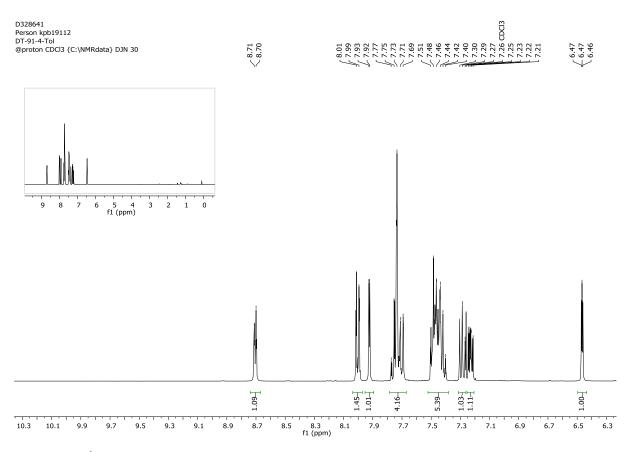


Figure S22. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (Toluene-entry 1, Table S3)

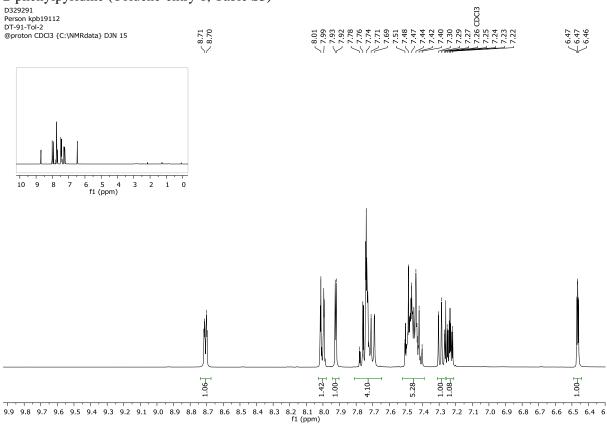


Figure S23. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (Toluene-entry 2, Table S3)

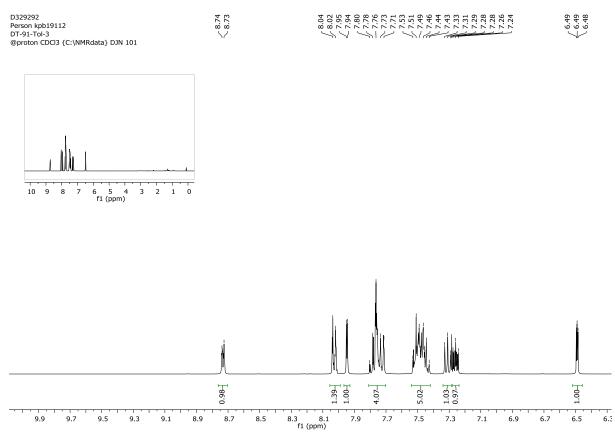
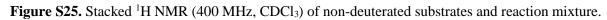


Figure S24. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (Toluene-entry 3, Table S3)

3.3. Competition Experiments with [(COD)Ir(IMes)PPh₃]BArF₂₄(Ir-1)

Table S4. Determination of the competition rate constant κ from the labelling experiment between acetophenone and benzophenone

	Substrate R1		Substra	te R2	Catalyst				
					Ir-1 [(COD)Ir(IMes)PPh ₃][BArF ₂₄]				
Mass	12.0 n	ng	18.2	mg	8.7 mg				
	-) = 7.84 - 7.7				
			• •		= 7.63 - 7.5	3 ppm			
-	e	$C(x) \delta = 7.99$			84 – 7.77 (m,	4H/D R2) 7	63 – 7 5		
		,			60 (s, 3H, R1		.05 7.5		
Entry	$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 3H$	%D _{R1}	$I_{R2(t)}$ $N = 4H$	$\frac{I_{R2(0)}}{N = 2H}$	%D _{R2}	к		
1	1.39	3.00	31	3.07	2.22 ^a	31	0.99		
2	1.08	3.00	46	2.65	2.44 ^b	46	1.01		
3	1.01	3.00	50	2.18	2.12 ^c	49	1.04		
			Average	$\kappa = 1.01$					
$I_{R2(0)} = 3.22$	2-(3.00/3); ^t	$^{b}I_{R2(0)} = 3.44$	- (3.00/3); ^c	$I_{R2(0)} = 3.12 -$	- (3.00/3);				
non-deuterate	ed acetophenone					1			
non-deuterate	ed acetophenone						- 3		
non-deuterate							3		
	ed acetophenone								
	Juh								
	Juh								
	Juh								
	ed benzophenone								
non-deuterate	ed benzophenone								
non-deuterate	ed benzophenone								
non-deuterate	ed benzophenone								



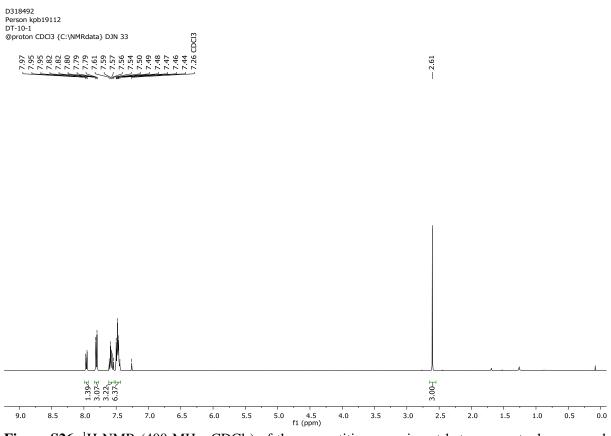


Figure S26. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzophenone (entry 1, Table S4).

D318493 Person kpb19112 DT-10-2 @proton CDCI3 {C:\NMRdata} DJN 34

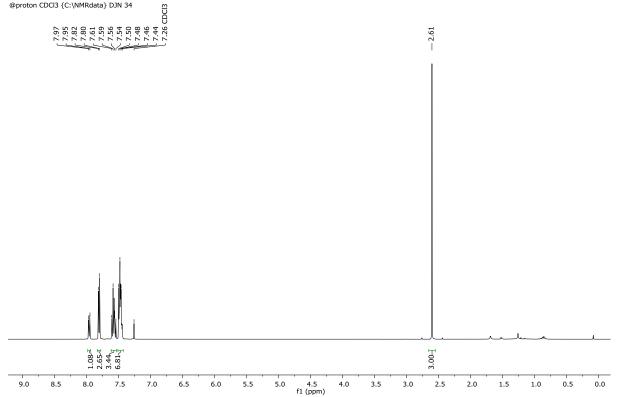


Figure S27. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzophenone (entry 2, Table S4).

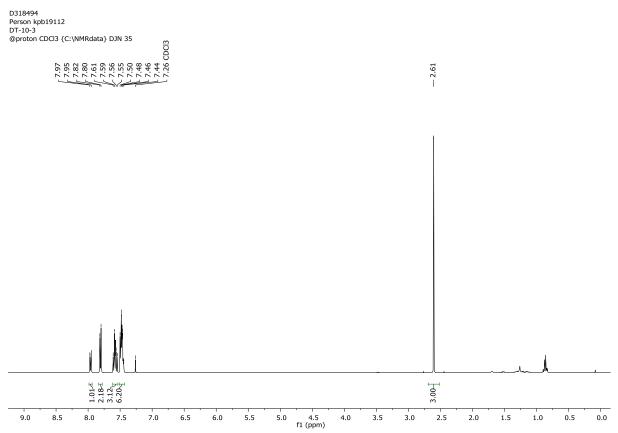


Figure S28. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzophenone (entry 3, Table S4).

	Substrate R1		Substra	ate R2		Catalyst			
O N		NH ₂	CH3			Ir-1 [(COD)Ir(IMes)PPh3][BArF24			
Mass	12.1 mg		12.0	mg	8.7 mg				
	n expected at	· ·				••			
	d against integ	-			$t\delta(\mathbf{R2}) = 2.6$	0 ppm			
-	etails of the de 400 MHz, CD				R6 = 7.77 (m	2H/D R1) 7	50 _ 7 30		
	1 and 3H, R2)	,			50 - 7.77 (III,	211/ D K1), 7	.59 - 7.5		
Entry	$\frac{I_{R1(t)}}{N = 2H}$	$\frac{I_{R1(0)}}{N = 3H}$	%D _{R1}	$I_{R2(t)}$ $N = 2H$	$I_{R2(0)} \\ N = 3H$	% D _{R2}	κ		
1	2.09	3.78 ^a	17	1.94	3.00	3	6.14		
2	1.57	3.84 ^b	39	1.85	3.00	8	6.27		
3	1.65	3.93°	37	1.86	3.00	7	6.37		
			Average	$\kappa = 6.26$					
$^{a}I_{R1(0)} = 6.7$	$78 - 3.00; {}^{b}I_{R1}$	$_{(0)} = 6.84 - 3.$	00; ^c $I_{R1(0)} =$	6.93 – 3.00;					
non-deut	terated acetophenon	e					-3		
							- 2		
/									
deuterati	ed reaction mixture						- 1		
U									

Table S5. Determination of the competition rate constant κ from the labelling experiment between benzamide and acetophenone

Figure S29. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

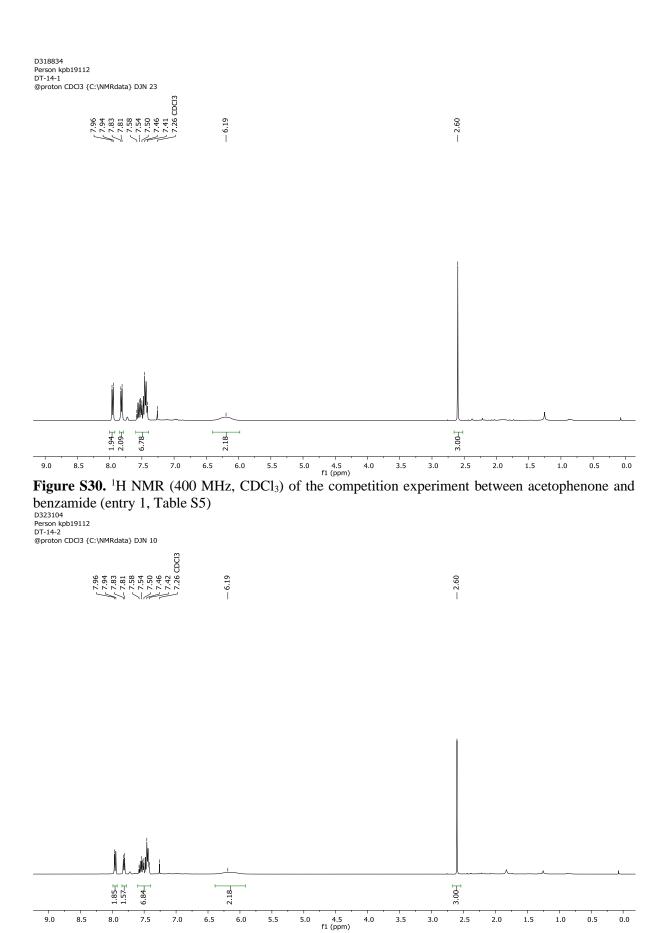


Figure S31. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzamide (entry 2, Table S5)

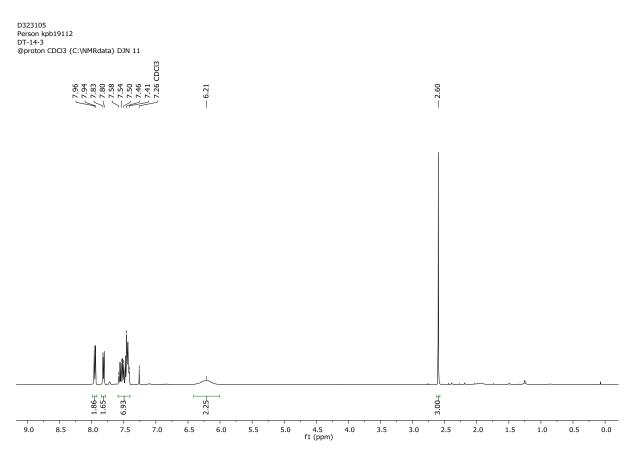


Figure S32. 1 H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzamide (entry 3, Table S5)

	Substrat	e R1	Su	bstrate R2		Catalys	t
	O II)		0 		Ir-1	
		CH₃		NMe ₂	[(CC	D)Ir(IMes)PF	hallBArF24
					[(00	<i>D</i>)II(IIII05)I I	115][[21 111 24]
Mass	12.0 r	ng	-	14.9 mg		8.7 mg	,
	-				$(\mathbf{R2}) = 7.42 - 7$		
	-	-		~ ~	at δ (R2) = 3	.17 – 2.88 ppr	n
•	etails of the a					111 D4\ 7	
		<i>,</i>				n, 1H, R1), 7.4	
(m, 2H, K			$\mathbf{K}\mathbf{Z}$ and \mathbf{SH} ,			R2), 2.60 (s, 3)	\mathbf{H}, \mathbf{K}
Entry	$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 3H$	D_{R1}	$I_{R2(t)}$ $N = 2H$	$I_{R2(0)}$ $N = 6H$	$%D_{R2}$	κ
1	1.50	$\frac{N - 311}{3.00}$	25	1.62^{a}	5.32	9	2 1 9
2	1.50	3.00	23 23	1.02 ^a 1.96 ^b	5.52 6.29	9 7	3.18 3.74
3	1.34	3.00	23 28	2.06 ^c	6.59	6	4.82
	1.15	5.00		$\frac{2.00}{\text{ge }\kappa = 3.91}$	0.57	0	1.02
$^{\rm L}$ IP2(1) = 4 2	$28 - (5.32)/6 \times$	$(3)^{\circ}$ $[n_{2}(n) - 5]$	10 - (6.29)	6 4 3 1 1	535 - (659)	/6×3	
1 K2(t) 112		$(3, I_{R2(t)} - 3)$	(0.27)	$0 \times 3, I_{R2(t)} =$	5.55 (0.57)	,0,5	
	ed acetophenone	$\frac{1}{1}$ (t) - 3.	.10 (0.29)	0^{-5} , $1_{R2(t)}$ –			
		(1, 1) = 0	10 (0.25)/	0×5 , $1_{R2(t)} - $			
		(1, 1) = 0		0×5 , $1_{R2(t)} =$			- 3
		(1, 1) = 0		0×3, 1 _{R2(t)} –			-3
		(J, 1 _{K2(t)} – J.		0×5 , $1_{R2(t)} =$			-3
				0×3, 1 _{R2(t)} –			-3
non-deuterat				0×3, 1 _{R2(t)} –			-3
non-deuterat				0×3, 1 _{R2(t)} –			-3
non-deuterat				0×3, 1 _{R2(t)} –			-3
non-deuterat				0×3, 1 _{R2(t)} –			-3
non-deuterat				0×3, 1 _{R2(t)} –			-3
non-deuterat				0×3, 1 _{R2(t)} –	M		-3
non-deuterat	ed acetophenone			0×3, 1 _{R2(t)} –	M		-3
non-deuterat	ed acetophenone			0×3, 1 _{R2(t)} –	M		-2
non-deuterat	ed acetophenone			0×3, 1 _{R2(t)} –			-3
non-deuterat	ed acetophenone			0×3, 1 _{R2(t)} –	M		-2
non-deuterat	ed acetophenone			0×3, 1 _{R2(t)} –			-2

Table S6. Determination of the competition rate constant κ from the labelling experiment between acetophenone and *N*,*N*-dimethylbenzamide.

Figure S33. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

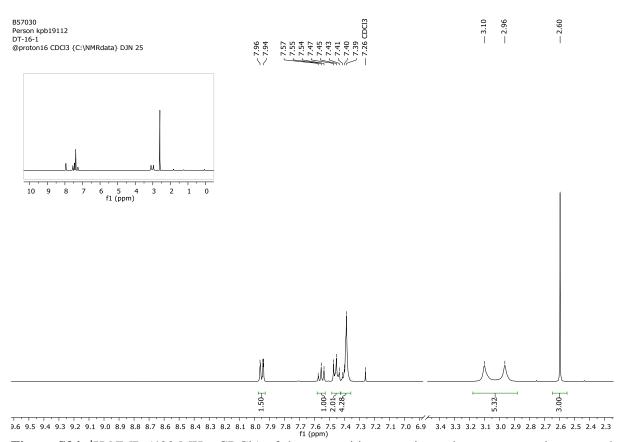


Figure S34. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and N,N-dimethylbenzamide (entry 1, Table S6). D323115 Person kpb19112 DT-16-3 @proton CDCl3 {C:\NMRdata} DJN 20

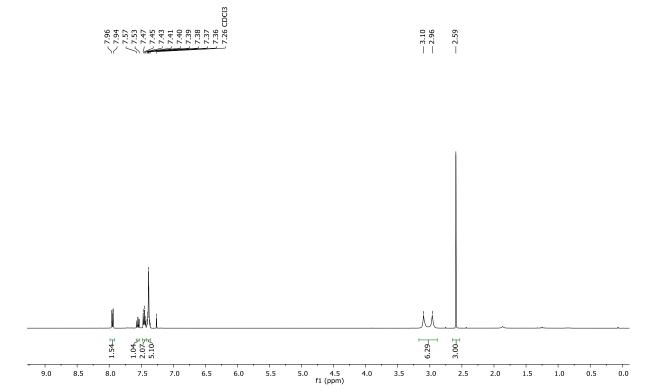


Figure S35. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and N,N-dimethylbenzamide (entry 2, Table S6).

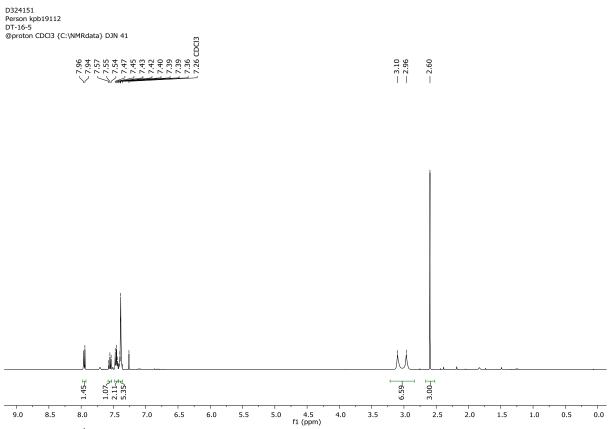


Figure S36. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and *N*,*N*-dimethylbenzamide (entry 3, Table S6).

Table S7. Determination of the competition rate constant κ from the labelling experiment between acetophenone and nitrobenzene.

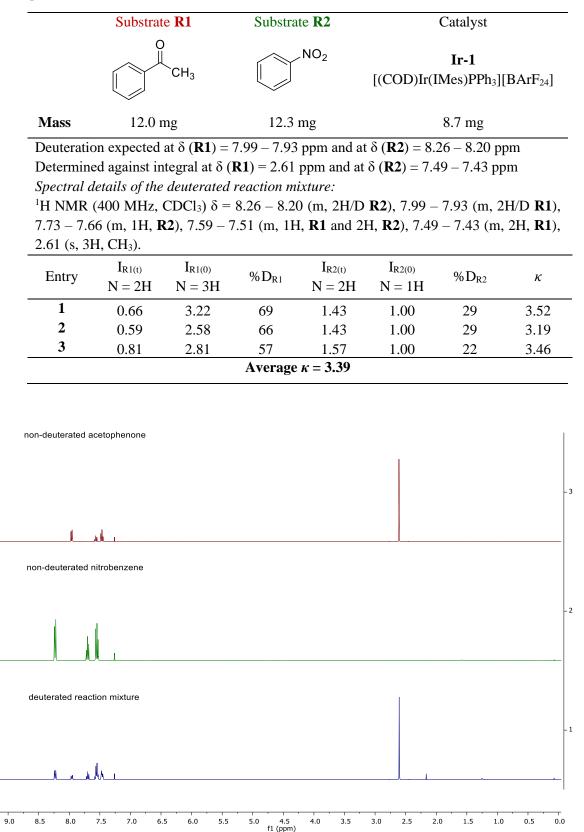


Figure S37. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

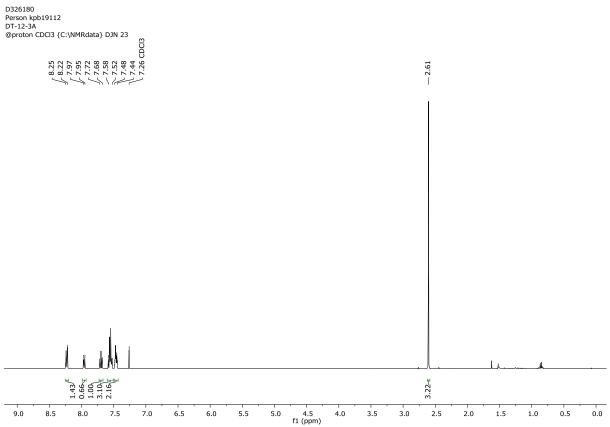


Figure S38. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and nitrobenzene (entry 1, Table S7).

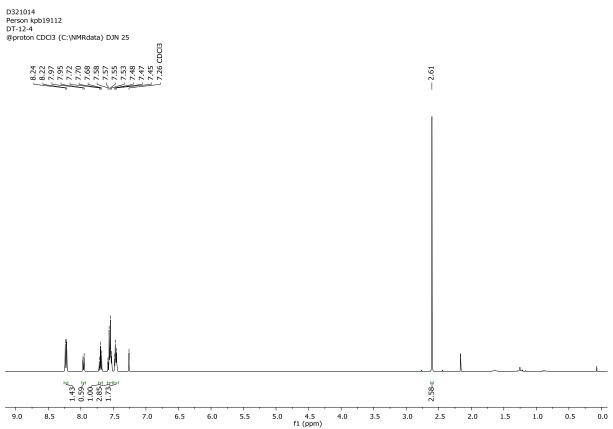


Figure S39. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and nitrobenzene (entry 2, Table S7).

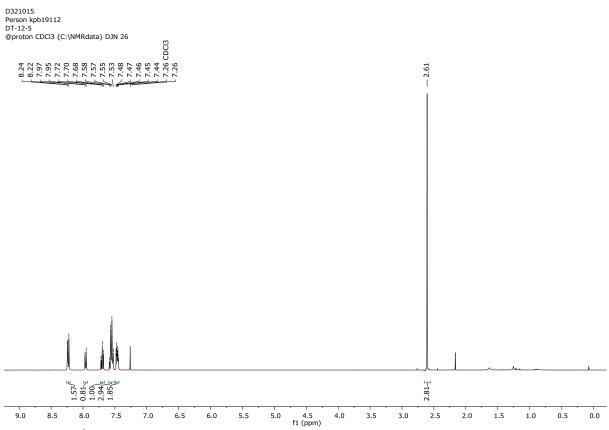


Figure S40. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and nitrobenzene (entry 3, Table S7).

Table S8. Determination of t	ne competition rate	e constant κ from	the labelling	experiment between
nitrobenzene and ethyl benzoa	te.			

	Substr	ate R1	Substra	ate R2		Catalyst	
		∕NO₂		O OEt	[(COD)Ir(I	Ir-1 Mes)PPh ₃][]	BArF ₂₄]
Mass	12.3	5 mg	15.0	mg		8.7 mg	
Deuterati	on expected	at δ (R1) = 8	8.26 - 8.20	ppm and at δ	(R2) = 8.08	- 8.02 ppm	
		x			$(\mathbf{R2}) = 4.38$		
	e	deuterated					
. *	b	$CDCl_3) \delta = 8$.26 - 8.20 (1	m, 2H/D, R1), $8.08 - 8.02$	2 (m, 2H/D)	R2), 7.70
¹ H NMR	(400 MHz, 0	· · · · · · · · · · · · · · · · · · ·	-), 8.08 – 8.02 R2), 7.43 (t,	-	
1 H NMR (t, J = 7.4	(400 MHz, 0 4 Hz, 1H, R	· · · · · · · · · · · · · · · · · · ·	51 (m, 2H,	R1 and 1H, 1), 8.08 – 8.02 R2), 7.43 (t,	-	
¹ H NMR $(t, J = 7.4)$	(400 MHz, 0 4 Hz, 1H, R	1), 7.59 – 7.5	51 (m, 2H,	R1 and 1H, 1		-	
1 H NMR (t, $J = 7.4$ 4.38 (q, J	(400 MHz, 0 4 Hz, 1H, R $\frac{1}{I} = 7.1$ Hz, 2 $\frac{1}{I_{R1(t)}}$	$\begin{array}{c} \textbf{1}), \ 7.59 - 7.3 \\ \textbf{H}, \ \textbf{R2}), \ 1.39 \\ \hline I_{\text{R1}(0)} \end{array}$	51 (m, 2H, $(t, J = 7.1 \text{ H})$	R1 and 1H, 1 Hz, 3H, R2). I _{R2(t)}	R2), 7.43 (t, $I_{R2(0)}$	J = 7.6 Hz,	2H, R2)
¹ H NMR (t, $J = 7.4$ 4.38 (q, J Entry	(400 MHz, 0) 4 Hz, 1H, R V = 7.1 Hz, 22 $I_{R1(t)}$ N = 2H	1), $7.59 - 7.5$ H, R2), 1.39 $I_{R1(0)}$ N = 1H	51 (m, 2H, $\frac{1}{2}$ (t, $J = 7.1$ H $\% D_{R1}$	R1 and 1H, $I_{z, 3H, R2}$. $I_{R2(t)}$ N = 2H	R2), 7.43 (t, $I_{R2(0)}$ $N = 2H$	$J = 7.6 \text{ Hz},$ $\% D_{\text{R2}}$	2H, R2), κ
1 H NMR (t, <i>J</i> = 7.4 4.38 (q, <i>J</i> Entry 1	$(400 \text{ MHz}, 0)$ $(400 \text{ MHz}, 0)$ $(4 \text{ Hz}, 1\text{ H}, \mathbf{R})$ $(7 = 7.1 \text{ Hz}, 2)$ $I_{R1(t)}$ $N = 2\text{H}$ 0.88	$I), 7.59 - 7.5$ $H, R2), 1.39$ $I_{R1(0)}$ $N = 1H$ 1.00	$51 (m, 2H, 1) (t, J = 7.1 H) \% D_{R1} 56$	R1 and 1H, I Hz, 3H, R2). $I_{R2(t)}$ N = 2H 1.83	R2), 7.43 (t, $I_{R2(0)}$ $N = 2H$ 2.12	J = 7.6 Hz, % D _{R2} 14	2H, R2), κ 5.58
$\frac{1}{1}$ H NMR (t, J = 7.4 4.38 (q, J) Entry 1 2	$(400 \text{ MHz}, 0)$ $(400 \text{ MHz}, 0)$ $4 \text{ Hz}, 1\text{H}, \mathbf{R}$ $7 = 7.1 \text{ Hz}, 22$ $\overline{I_{R1(t)}}$ $N = 2\text{H}$ 0.88 1.04	$\begin{array}{c} \textbf{1}), \ 7.59 \ - \ 7.5 \\ \textbf{H}, \ \textbf{R2}), \ 1.39 \\ \hline I_{R1(0)} \\ \textbf{N} = 1 \\ \hline 1.00 \\ 1.00 \end{array}$	51 (m, 2H, $\frac{1}{2}$ (t, $J = 7.1$ H %D _{R1} 56 48 62	R1 and 1H, $I_{z, 3H, R2}$. $I_{R2(t)}$ N = 2H 1.83 1.87	R2), 7.43 (t, $I_{R2(0)}$ <u>N = 2H</u> 2.12 2.06	J = 7.6 Hz, $\% D_{R2}$ 14 9	2H, R2), κ 5.58 6.76
$\frac{1}{1}$ H NMR (t, J = 7.4 4.38 (q, J) Entry 1 2	$(400 \text{ MHz}, 0)$ $(400 \text{ MHz}, 0)$ $4 \text{ Hz}, 1\text{H}, \mathbf{R}$ $7 = 7.1 \text{ Hz}, 22$ $\overline{I_{R1(t)}}$ $N = 2\text{H}$ 0.88 1.04	$\begin{array}{c} \textbf{1}), \ 7.59 \ - \ 7.5 \\ \textbf{H}, \ \textbf{R2}), \ 1.39 \\ \hline I_{R1(0)} \\ \textbf{N} = 1 \\ \hline 1.00 \\ 1.00 \end{array}$	51 (m, 2H, $\frac{1}{2}$ (t, $J = 7.1$ H %D _{R1} 56 48 62	R1 and 1H, I Hz, 3H, R2). $I_{R2(t)}$ N = 2H 1.83 1.87 1.74	R2), 7.43 (t, $I_{R2(0)}$ <u>N = 2H</u> 2.12 2.06	J = 7.6 Hz, $\% D_{R2}$ 14 9	2H, R2), κ 5.58 6.76
$\frac{1}{H} \text{ NMR}$ $(t, J = 7.4$ $4.38 (q, J)$ Entry 1 2	$(400 \text{ MHz}, 0)$ $(400 \text{ MHz}, 0)$ $4 \text{ Hz}, 1\text{H}, \mathbf{R}$ $7 = 7.1 \text{ Hz}, 22$ $\overline{I_{R1(t)}}$ $N = 2\text{H}$ 0.88 1.04	$\begin{array}{c} \textbf{1}), \ 7.59 \ - \ 7.5 \\ \textbf{H}, \ \textbf{R2}), \ 1.39 \\ \hline I_{R1(0)} \\ \textbf{N} = 1 \\ \hline 1.00 \\ 1.00 \end{array}$	51 (m, 2H, $\frac{1}{2}$ (t, $J = 7.1$ H %D _{R1} 56 48 62	R1 and 1H, I Hz, 3H, R2). $I_{R2(t)}$ N = 2H 1.83 1.87 1.74	R2), 7.43 (t, $I_{R2(0)}$ <u>N = 2H</u> 2.12 2.06	J = 7.6 Hz, $\% D_{R2}$ 14 9	2H, R2), κ 5.58 6.76

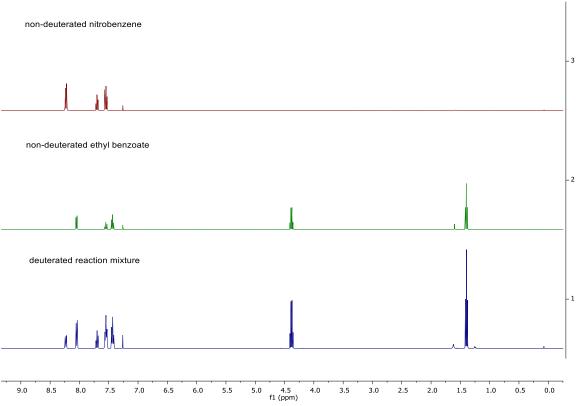
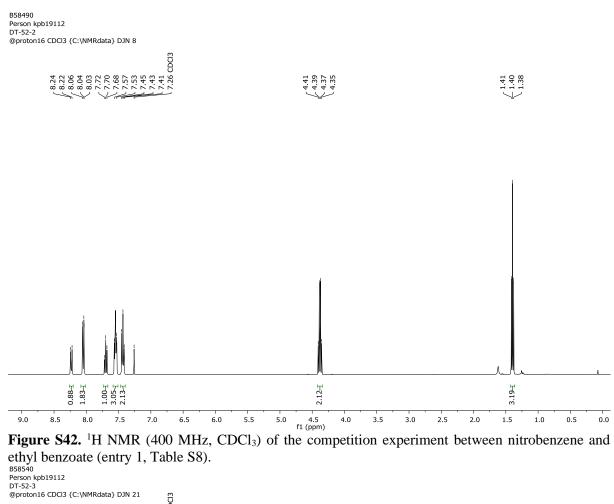
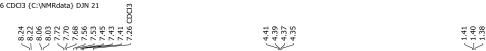
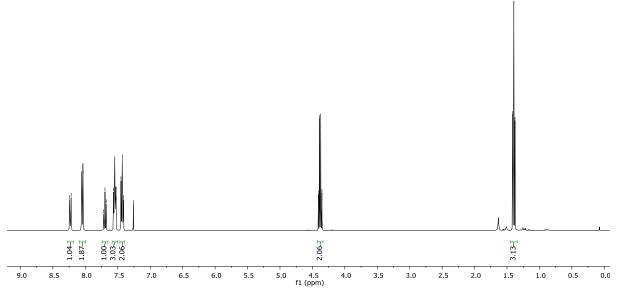
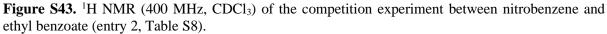


Figure S41. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.









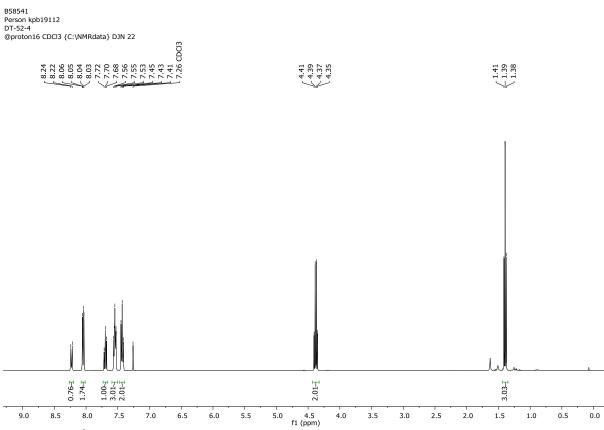
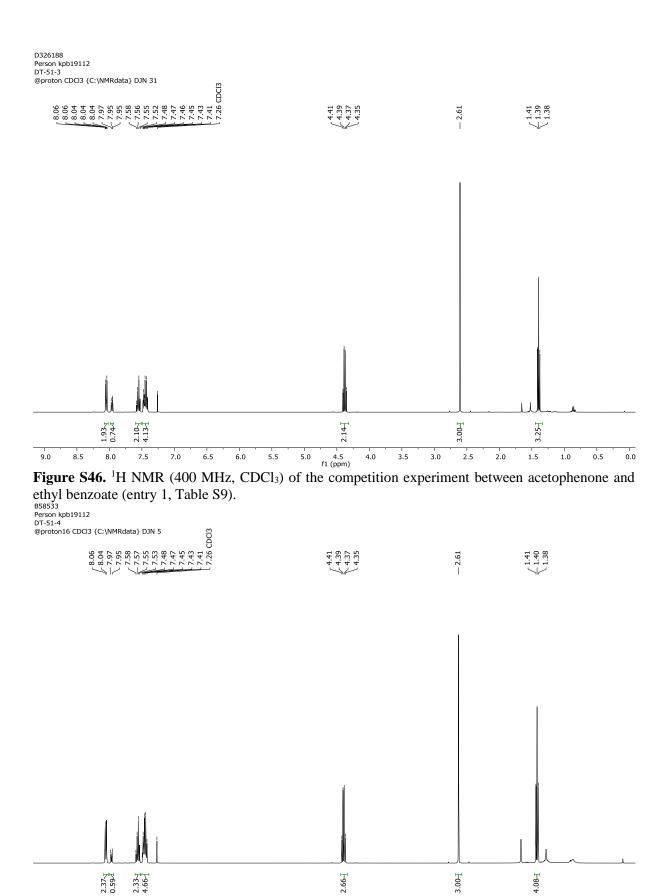


Figure S44. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between nitrobenzene and ethyl benzoate (entry 3, Table S8).

Table S9. Determination of the competition rate constant κ from the labelling experiment between acetophenone and ethyl benzoate.

	Substrate	e R1	Substrate	e R2		Catalyst	
	0	`CH₃	0 I	`OEt	[(COD)Ir(I	Ir-1 Mes)PPh ₃][BArF ₂₄]
Mass	12.0 m	ng	15.0 n	ng		8.7 mg	
	-		-	-	δ (R2) = 8.08		ı
	-	-	· -	-	$(\mathbf{R2}) = 4.38$	ppm	
-	e e	deuterated r $(DCl_3) \delta = 8$			2), 7.99 – 7.9	93 (m. 2H/E) R1). 7.
– 7.52 (m,	1H, R1 and	<i>,</i>	7.49 - 7.40	(m, 2H, R1	and 2H, R2		
Entry	$\frac{I_{R1(t)}}{I_{R1(t)}}$ $N = 2H$	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	%D _{R1}	$\frac{I_{R2(t)}}{I_{R2(t)}}$ $N = 2H$	$\begin{split} I_{R2(0)} \\ N = 2H \end{split}$	% D _{R2}	κ
1	0.74	3.00	63	1.93	2.14	10	9.63
2	0.59	3.00	71	2.37	2.66	11	10.5
3	0.54	3.00	73	$\frac{1.89}{\kappa = 10.38}$	2.13	11	10.9
non-deuterated a	acetophenone						
non-deuterated a	acetophenone						
I	acetophenone						
I							
non-deuterated							

Figure S45. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.



4.5 f1 (ppm) 5.5 Figure S47. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and ethyl benzoate (entry 2, Table S9).

5.0

4.0

3.5

3.0

2.5

1.5

1.0

0.5

0.0

2.0

2.334 4.664

7.5

7.0

6.5

6.0

2.37₄ 0.594

8.0

8.5

9.0

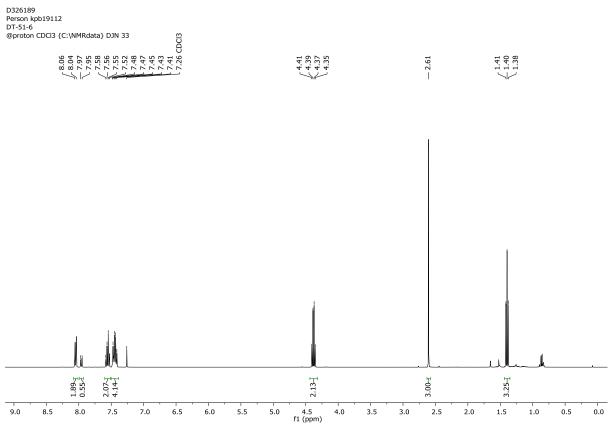


Figure S48. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and ethyl benzoate (entry 3, Table S9).

Table S10. Determination of the competition rate constant κ from the labelling experiment between acetanilide and benzamide.

	Substrate	R 1	Substrate	R2		Catalyst	
	0 I	NH ₂	HN N O	∠CH₃	[(COD)Ir(II	Ir-1 Mes)PPh ₃][B	SArF ₂₄]
Mass	12.1 m	g	13.5 mg	,	4.3 m	g (2.5 mol %)
Determine Spectral da H NMR (+ R1), 7.60 -	n expected at d against inte etails of the a 400 MHz, DI - 7.55 (m, 2H - 7.24 (m, 2H	egral at δ (R leuterated re MSO- d_6) δ = I/D, R2), 7.5	1) = 7.47 - 7 eaction mixte = 9.90 (br, 1) 54 - 7.41 (m	7.41 ppm and <i>ure:</i> H, R2), 7.96 , 1H, R1), 7.	d at δ (R2) = (br, 1H, R1) 47 - 7.41 (m	7.30 - 7.24), 7.91 - 7.84	(m, 2H
Entry	$\frac{I_{R1(t)}}{I_{R1(t)}}$ $N = 2H$	I, I(2), 7.04 $I_{R1(0)}$ N = 2H	%D _{R1}	$I_{R2(t)}$ $I_{R2(t)}$ $N = 2H$	$\frac{I_{R2(0)}}{I_{R2(0)}}$ N = 2H	%D _{R2}	κ
1	1.75	1.93	9	1.86	2.00	7	1.35
2	1.76	1.98	11	1.86	2.00	7	1.62
3	1.72	1.96	12	$\frac{1.85}{\kappa = 1.55}$	2.00	8	1.68
		h			l		
non-deuterat	ed acetanilide						
۸	l					J	
deuterated re	eaction mixture						
۸	J	ht_i				ļ	
0.0 9.5 9.C) 8.5 8.0 7	.5 7.0 6.5	6.0 5.5 5.0 f1 (ppm)	4.5 4.0 3	.5 3.0 2.5	2.0 1.5 1.0	0.5 0

Figure S49. Stacked ¹H NMR (400 MHz, DMSO-*d*₆) of non-deuterated substrates and reaction mixture.

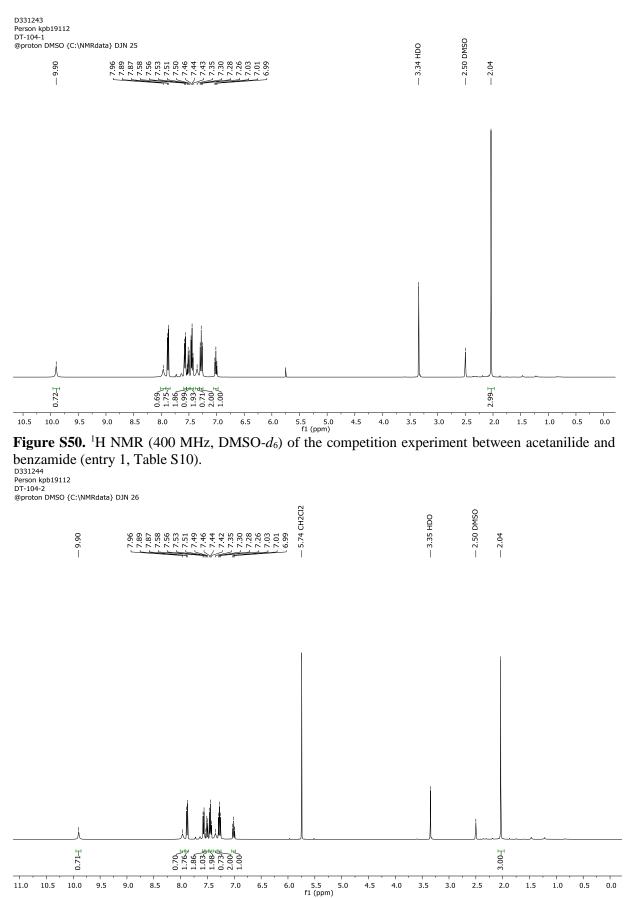


Figure S51. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between acetanilide and benzamide (entry 2, Table S10).

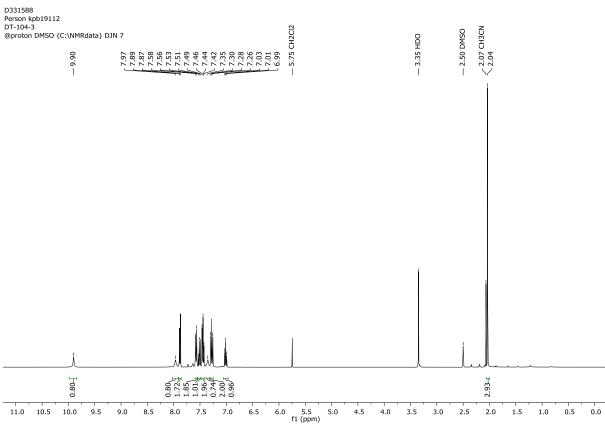


Figure S52. ¹H NMR (400 MHz, DMSO-*d*₆) of the competition experiment between acetanilide and benzamide (entry 3, Table S10).

Table S11. Determination of the competition rate constant κ from the labelling experiment between acetanilide and nitrobenzene.

	Substr	ate R1	Subs	trate R2		Catalyst	
	L L	H V ⊂ H ₃ O		NO ₂	[(COD)Ir(Ir-1 IMes)PPh ₃]	[BArF ₂₄]
Mass	13.5	5 mg	12	.3 mg	4.3 n	ng (2.5 mol	%)
Determi Spectrat ¹ H NMI 7.81 (m	ined against l details of th R (400 MHz	integral at 8 <i>he deuterate</i> , DMSO- <i>d</i> ₆) 7.70 – 7.65 ($\delta (\mathbf{R1}) = 7.5$ ed reaction $\delta = 9.90$ ((m, 2H, R 2)	30 – 7.24 pp: <i>mixture:</i> (br, 1H, R1), 2), 7.60 – 7.5	at δ (R2) = 8.2 m and at δ (R 8.26 - 8.20 (55 (m, 2H/D,	(2) = 7.87 - 100 (m, 2H/D, R	7.81 ppn 2), 7.87
Entry	$I_{R1(t)}$ N = 2H	$I_{R1(0)}$ N = 2H	$%D_{R1}$	$I_{R2(t)}$ $N = 2H$	I _{R2(0)} N =1H	%D _{R2}	к
1	1.11	2.00	45	1.75	0.91	4	15.01
2	0.81	2.00	60	1.68	0.89	6	15.63
3	0.94	2.00	53	$\frac{1.73}{\text{ge }\kappa = 15.18}$	0.91	5	14.89
n-deuteratec	d acetanilide	1.1.			н _о о		
n-deuterated	d acetanilide				H2O		

11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 f1 (ppm)

Figure S53. Stacked ¹H NMR (400 MHz, DMSO-*d*₆) of non-deuterated substrates and reaction mixture.

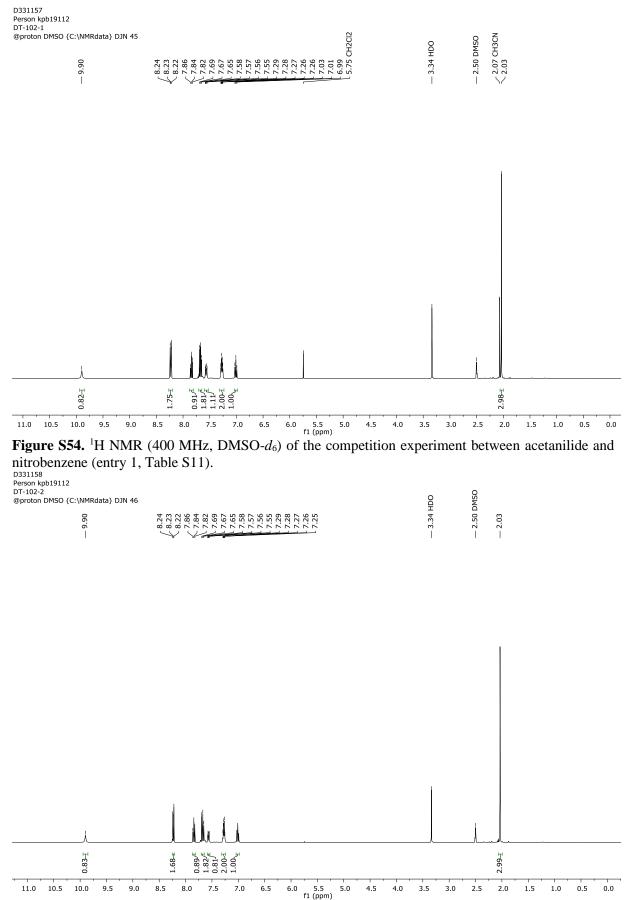


Figure S55. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between acetanilide and nitrobenzene (entry 2, Table S11).

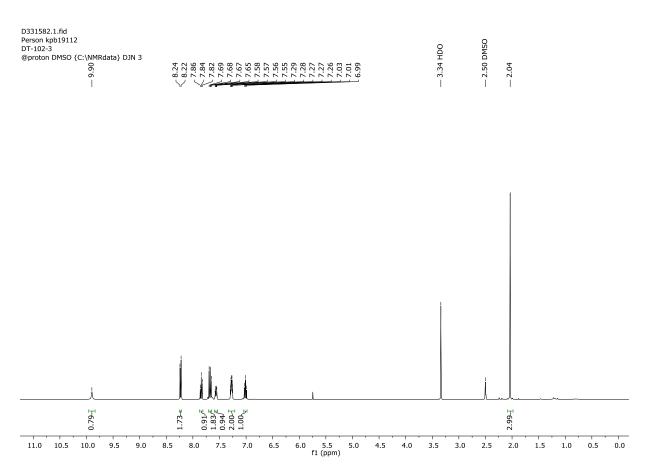


Figure S56. ¹H NMR (400 MHz, DMSO-*d*₆) of the competition experiment between acetanilide and nitrobenzene (entry 3, Table S11).

Table S12. Determination of the competition rate constant κ from the labelling experiment between 2-phenylpyrimidine and benzamide.

	Substr	ate R1	Subs	strate R2		Catalyst	
	N Ph	N N		NH ₂	[(COD)Ir(Ir-1 IMes)PPh ₃]	[BArF ₂
Mass	15.6	mg	12	2.1 mg		8.7 mg	
	-			••	at δ (R2) = 7.		•
	-	-			at δ 7.55 – 7.3	39 ppm for I	R2
-	details of th				H, R1), 8.48 -	8/13 (m 2)	
					d 3H, R2), 7.		
	0 (br, 2H, R		<i>c</i> (1	ii, 511, 111 ui	u 311, 112), 7.	10 (0,0 11	0112,
	I _{R1(t)}	I _{R1(0)}	0/ D	I _{R2(t)}	I _{R2(0)}	0/ D	
Entry	N = 2H	N = 2H	$%D_{R1}$	N = 2H	N = 3H	$%D_{R2}$	κ
1	0.48	2.00	76	2.05	3.61 ^a	15	8.9
2	1.10	2.00	45	2.01	3.40 ^b	11	4.9
3	0.77	2.00	62	1.96	3.34 °	12	7.4
	6.61 — (2.00) d 2-phenylpyrimid			age $\kappa = 7.12$ (2.00/2×3); °	$I_{R2(0)} = 6.34$ -	- (2.00/2×3)	;
				0	$I_{R2(0)} = 6.34$ -	- (2.00/2×3)	;
on-deuterated				0	$I_{R2(0)} = 6.34$ -	- (2.00/2×3)	;
on-deuterated	d 2-phenylpyrimid			0	I _{R2(0)} = 6.34 -	- (2.00/2×3)	;
non-deutera	d 2-phenylpyrimid			0	I _{R2(0)} = 6.34 -	- (2.00/2×3)	;
non-deutera	d 2-phenylpyrimid			0	I _{R2(0)} = 6.34 -	- (2.00/2×3)	;

Figure S57. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

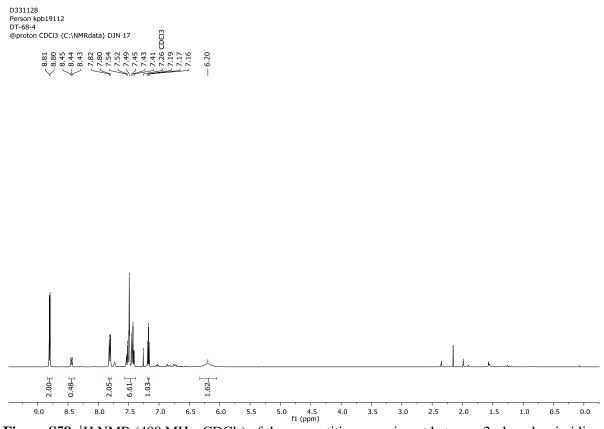


Figure S58. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyrimidine and benzamide (entry 1, Table S12). D331185 Person kpb19112 DT-103-3 @proton CDCI3 {C:\NMRdata} DJN 14



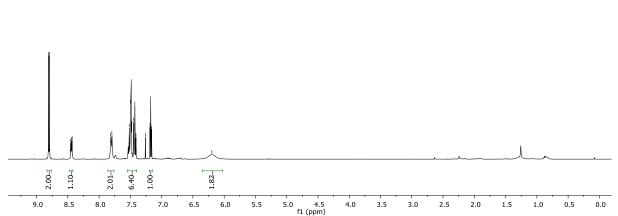


Figure S59. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyrimidine and benzamide (entry 2, Table S12).

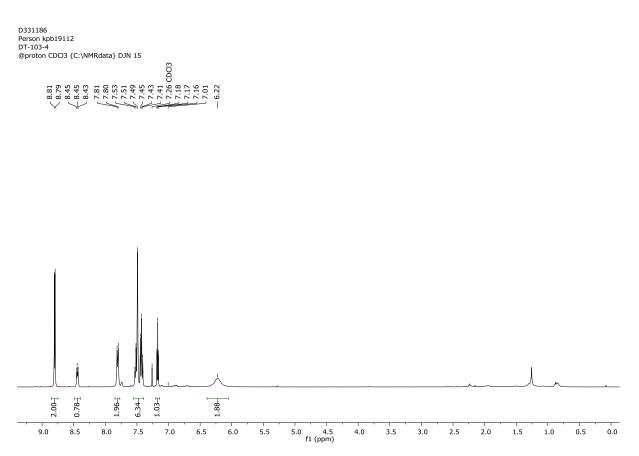


Figure S60. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyrimidine and benzamide (entry 3, Table S12).

Table S13. Determination of the competition rate constant κ from the labelling experiment between 2-phenylpyridine and acetophenone.

	Substr	ate R1	Subs	trate R2		Catalyst	
	Ph	N		CH3	[(COD)Ir	Ir-1 (IMes)PPh	3][BArF
Mass	15.5	mg	12	.0 mg		8.7 mg	
	n expected a						
	d against int etails of the d	-			and at δ (R2)) = 2.60 pp	m
¹ H NMR (4 – 7.94 (m,	400 MHz, C 2H/D, R2), d 3H, R1), 2	DCl ₃) $\delta = 8$. 7.78 - 7.70	74 – 8.66 ((m, 2H, R	(m, 1H, R1),			
Entry	$\frac{I_{R1(t)}}{N = 2H}$	$\frac{I_{R1(0)}}{N = 2H}$	%D _{R1}	$\begin{split} I_{R2(t)} \\ N = 2H \end{split}$	$I_{R2(0)}$ $N = 3H$	%D _{R2}	κ
1	1.75	2.26	23	1.97	3.00	2	16.92
2	1.62	2.26	28	1.96	3.00	2	16.48
3			20	1.96	3.00	2	16.88
	1.60	2.25	29 Average		5.00		10.00
	1.60	2.25			5.00	2	10.00
	2-phenylpyridine	2.25					10.00
on-deuterated 2	2-phenylpyridine	2.25					
on-deuterated 2	2-phenylpyridine	2.25					

9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 f1 (ppm)

Figure S61. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

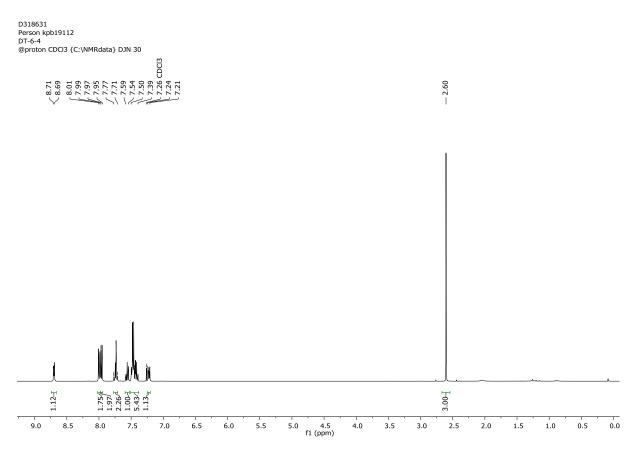


Figure S62. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between of 2-phenylpyridine and acetophenone (entry 1, Table S13).

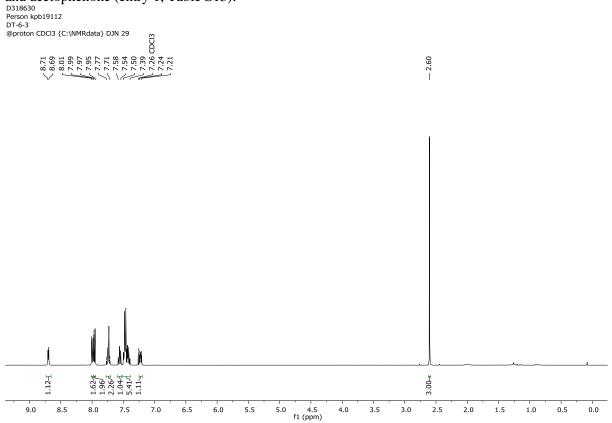


Figure S63. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between of 2-phenylpyridine and acetophenone (entry 2, Table S13).

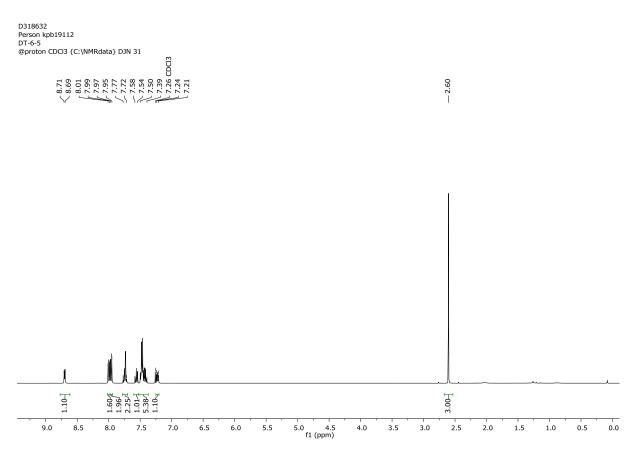


Figure S64. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between of 2-phenylpyridine and acetophenone (entry 3, Table S13).

Table S14. Determination of the competition rate constant κ from the labelling experiment between 2-phenyloxazoline and 2-phenylpyridine.

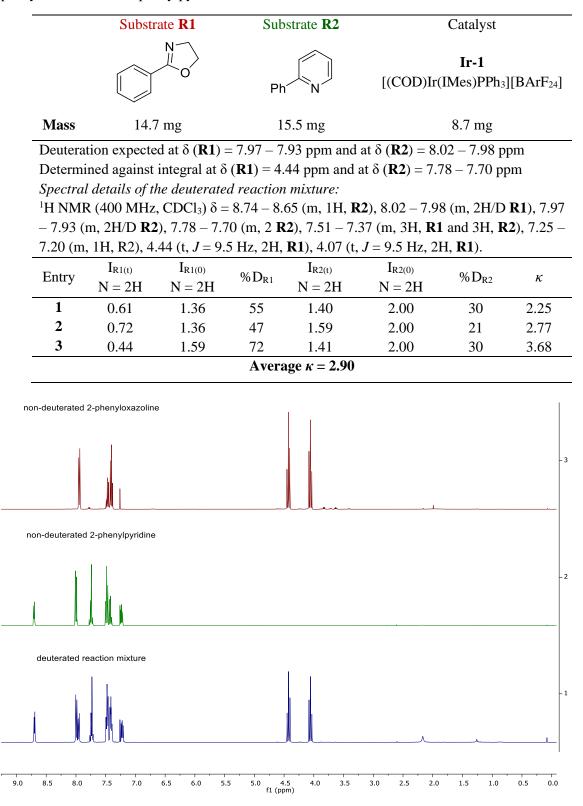


Figure S65. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

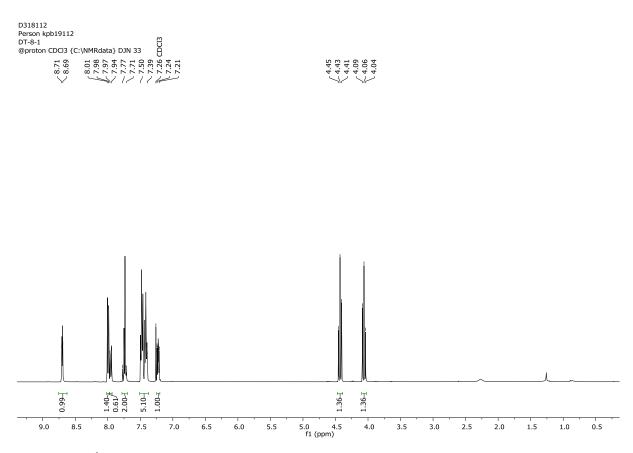
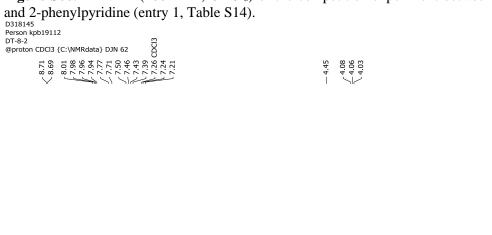


Figure S66. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylpyridine (entry 1, Table S14).



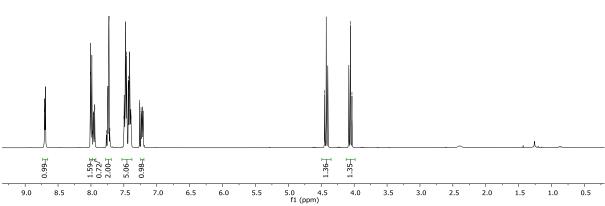


Figure S67. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylpyridine (entry 2, Table S14).

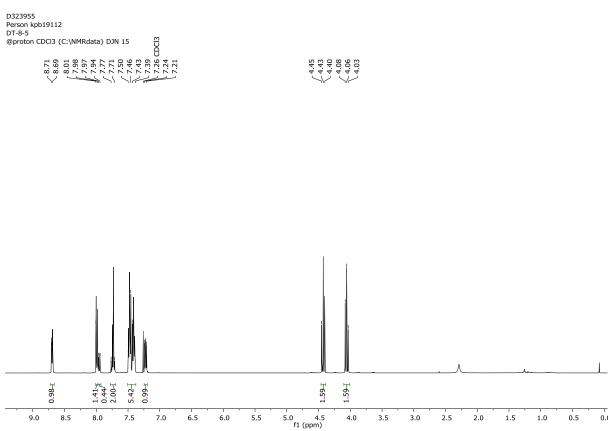


Figure S68. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylpyridine (entry 3, Table S14).

Table S15. Determination of the competition rate constant κ from the labelling experiment between 2-phenylthiazole and 1-phenylpyrazole.

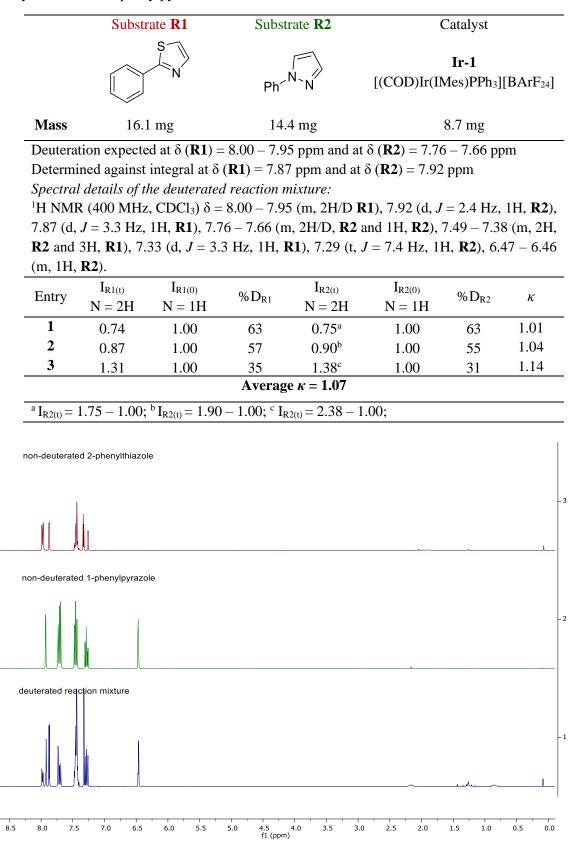


Figure S69. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D323641 Person kpb19112 DT-15-4 @proton CDCl3 {C:\NMRdata} DJN 11

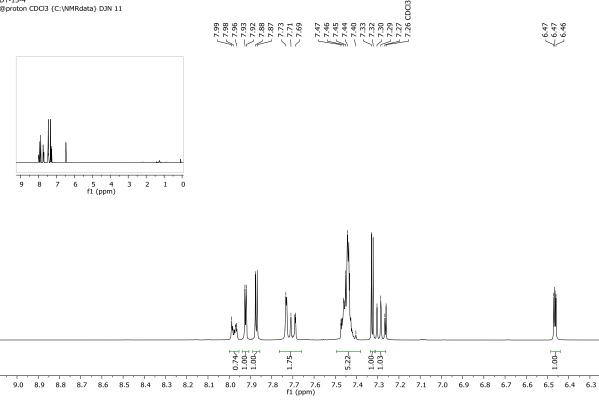


Figure S70. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazole and

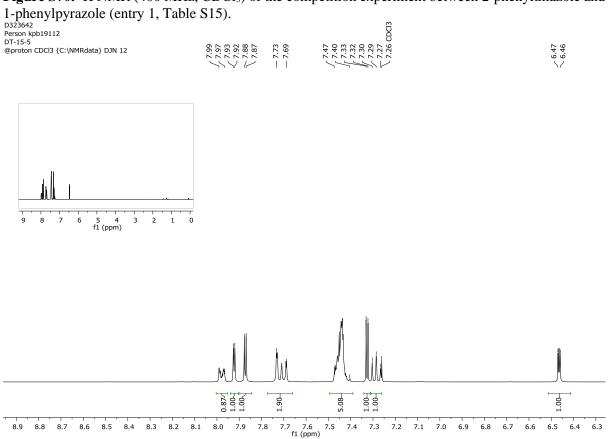


Figure S71. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazole and 1-phenylpyrazole (entry 2, Table S15).

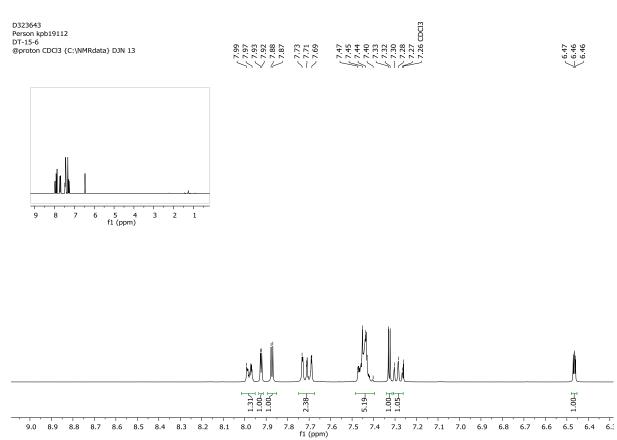


Figure S72. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazole and 1-phenylpyrazole (entry 3, Table S15).

Table S16. Determination of the competition rate constant κ from the labelling experiment between 2-phenyloxazoline and 2-phenylthiazoline.

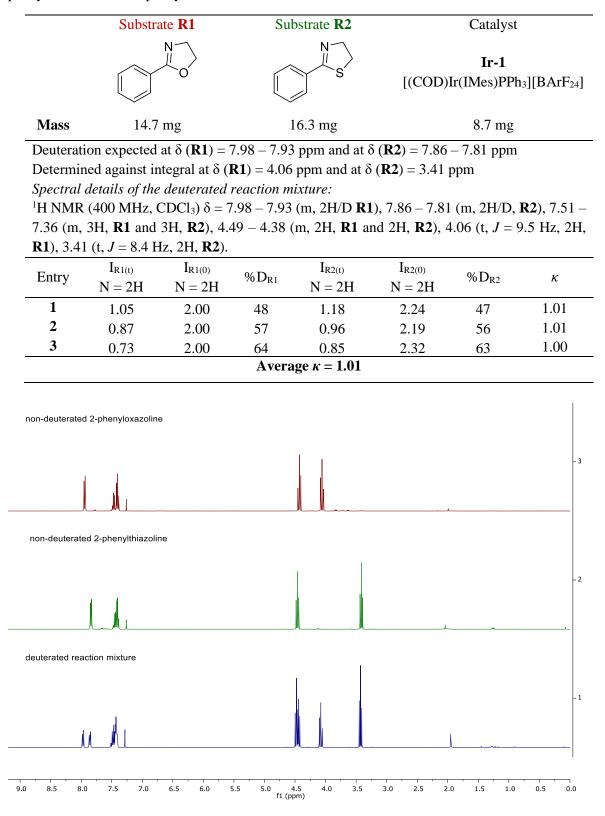


Figure S73. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

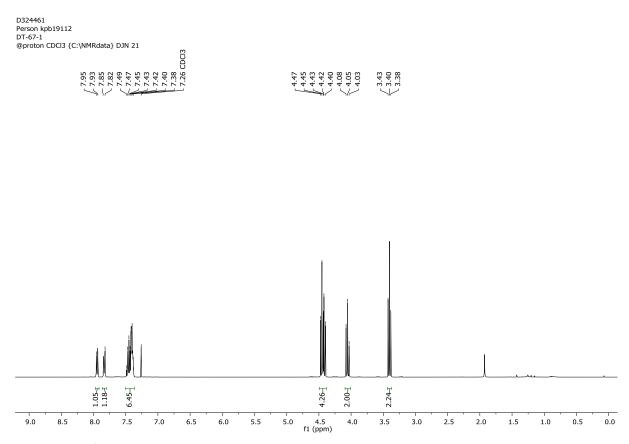


Figure S74. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylthiazoline (entry 1, Table S16). D324462 Person kpb19112 DT-67-2 @proton CDCI3 {C:\NMRdata} DJN 22

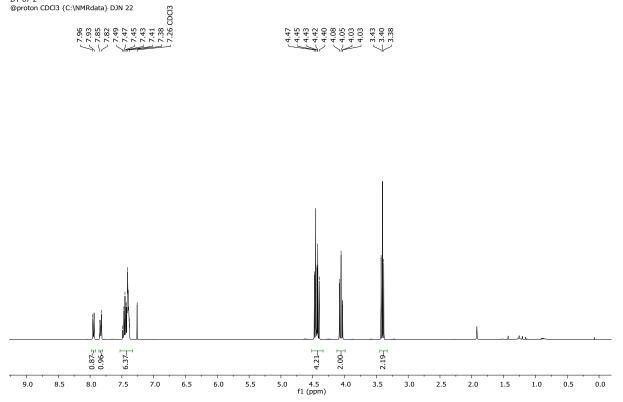


Figure S75. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylthiazoline (entry 2, Table S16).

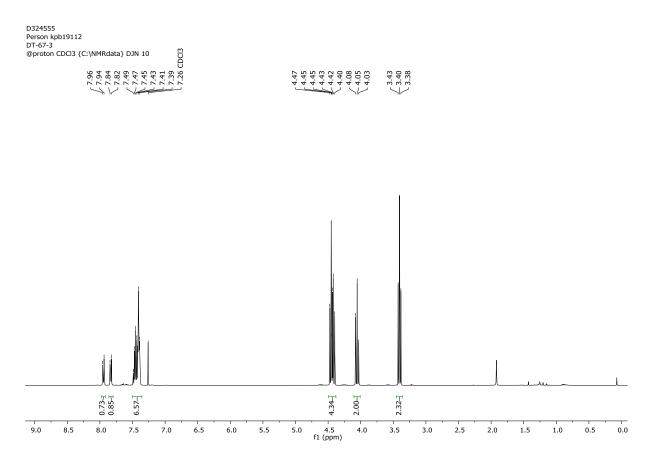


Figure S76. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylthiazoline (entry 3, Table S16).

Table S17. Determination of the competition rate constant κ from the labelling experiment between 2-phenylthiazoline and 2-phenylthiazole.

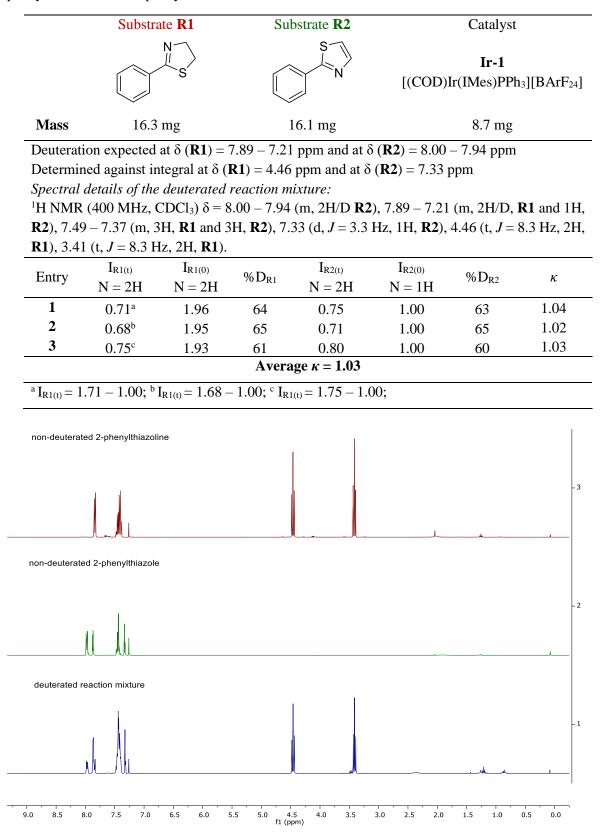
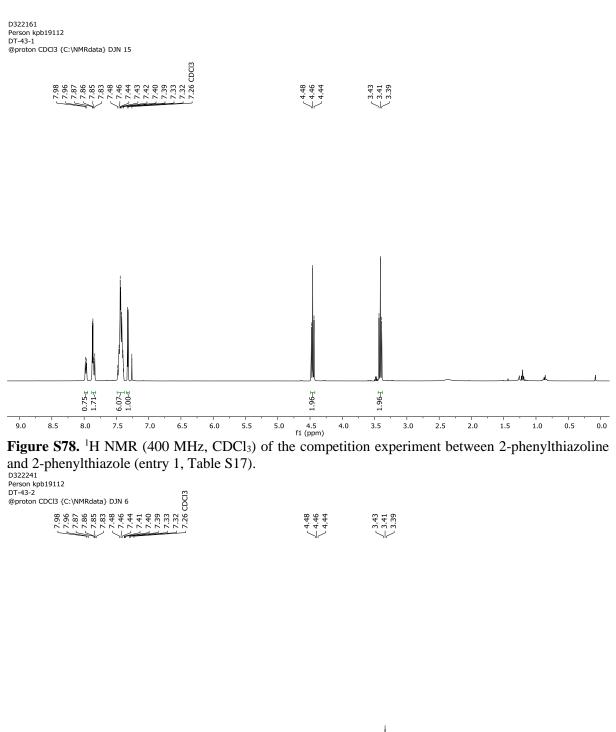


Figure S77. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.



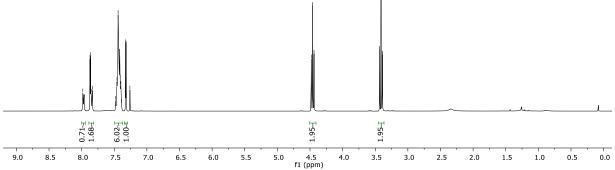


Figure S79. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazoline and 2-phenylthiazole (entry 2, Table S17).

D322242 Person kpb19112 DT-43-3 @proton CDCl3 {C:\NMRdata} DJN 7

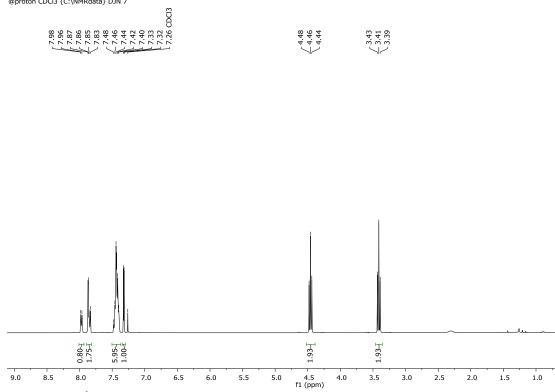
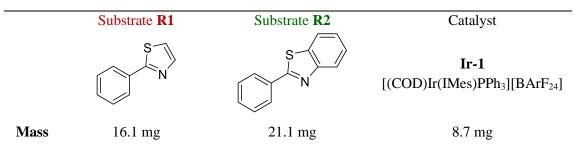


Figure S80. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazoline and 2-phenylthiazole (entry 3, Table S17).

0.5

0.0

Table S18. Determination of the competition rate constant κ from the labelling experiment between 2-phenylthiazole and 2-phenylbenzothiazole.



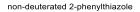
Deuteration expected at δ (**R1**) = 8.00 – 7.94 ppm and at δ (**R2**) = 8.14 – 8.06 ppm Determined against integral at δ (**R1**) = 7.33 ppm and at δ (**R2**) = 7.93 – 7.85 ppm *Spectral details of the deuterated reaction mixture:*

¹H NMR (400 MHz, CDCl₃) δ = 8.14 – 8.06 (m, 2H/D, **R2** and 1H, **R2**), 8.00 – 7.94 (m, 2H/D **R1**), 7.93 – 7.85 (m, 1H, **R1** and 1H, **R2**), 7.54 – 7.35 (m, 3H, **R1** and 4H, **R2**), 7.33 (d, *J* = 3.3 Hz, 1H, **R1**).

Entry	$\begin{split} I_{R1(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R1(0)} \\ N = 1 H \end{split}$	%D _{R1}	$\begin{split} I_{R2(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R2(0)} \\ N = 1 H \end{split}$	$%D_{R2}$	κ
1	0.60	1.00	70	1.83 ^a	1.08 ^d	15	7.26
2	0.80	1.00	60	2.06 ^b	1.16 ^e	11	7.71
3	0.57	1.00	72	1.86 ^c	1.10 ^f	15	7.48
			Avera	age $\kappa = 7.48$			

 ${}^{a}I_{R2(t)} = 2.91 - 1.00; {}^{b}I_{R2(t)} = 3.22 - 1.00; {}^{c}I_{R2(t)} = 2.96 - 1.00;$

$${}^{d}I_{R2(0)} = 2.08 - 1.00; {}^{e}I_{R2(0)} = 2.16 - 1.00; {}^{f}I_{R2(0)} = 2.10 - 1.00;$$



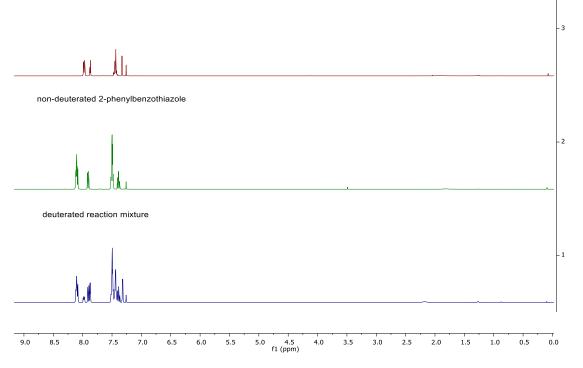
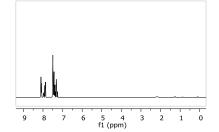


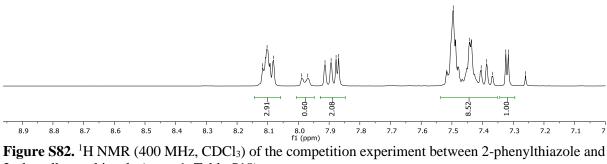
Figure S81. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D322296 Person kpb19112 DT-44-2 @proton CDCl3 {C:\NMRdata} DJN 1





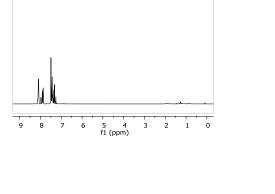




2-phenylbenzothiazole (entry 1, Table S18) D32297 Person kpb19112 DT-44-3 @proton CDCI3 {C:\NMRdata} DJN 2







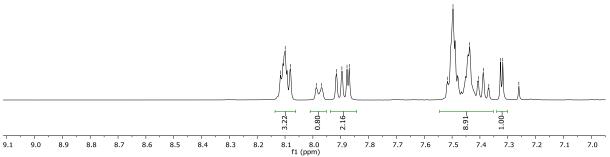


Figure S83. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazole and 2-phenylbenzothiazole (entry 2, Table S18).

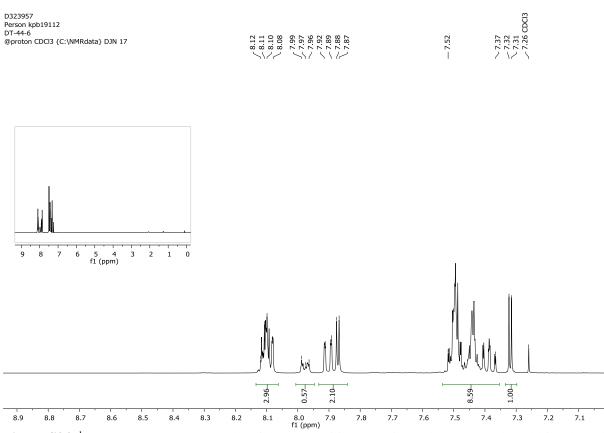


Figure S84. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazole and 2-phenylbenzothiazole (entry 3, Table S18).

Table S19. Determination of the competition rate constant κ from the labelling experiment between 2-phenylpyridine and 2-phenylbenzothiazole.

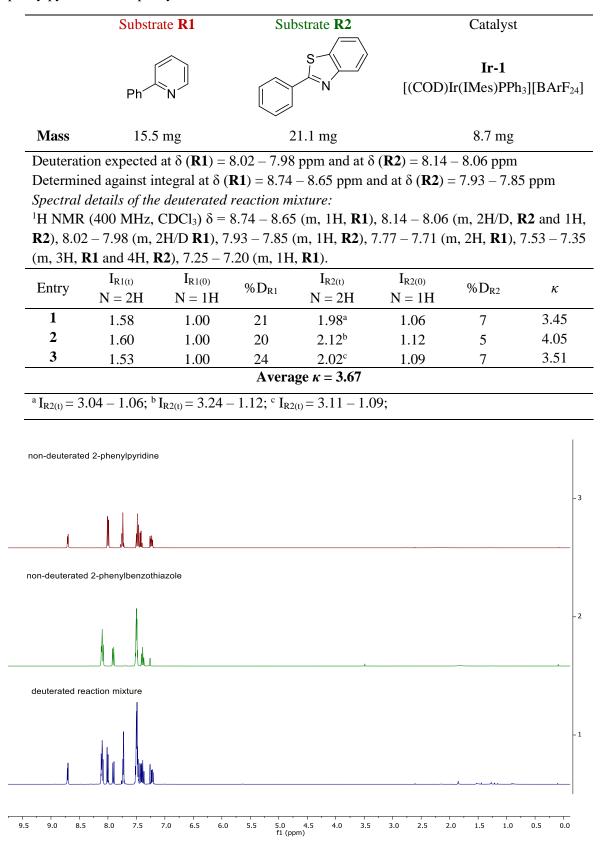


Figure S85. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

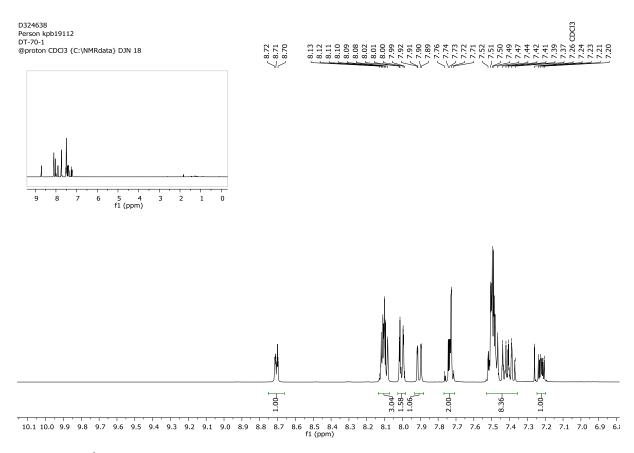


Figure S86. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyridine and

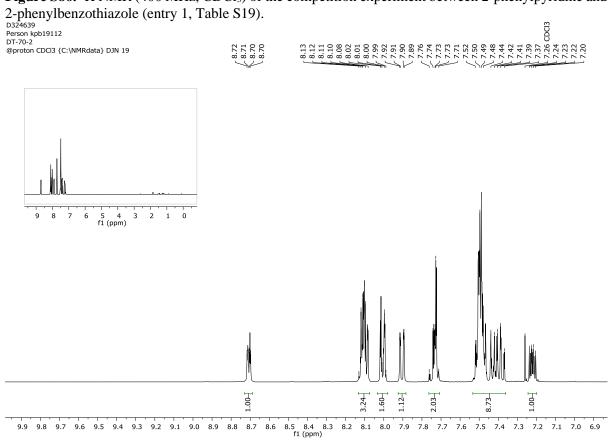


Figure S87. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyridine and 2-phenylbenzothiazole (entry 2, Table S19).

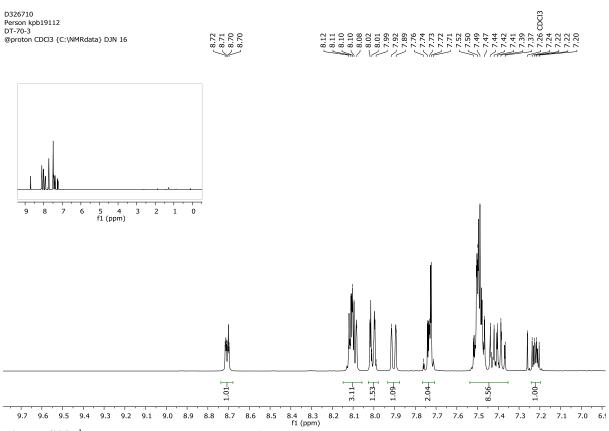


Figure S88. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyridine and 2-phenylbenzothiazole (entry 3, Table S19).

	Substrat	e R1	Substrate R2			Catalyst	
	Me	N		N S	[(COD	Ir-1)Ir(IMes)PPI	n3][BArF ₂₄]
Mass	15.8 r	ng	1	6.3 mg		8.7 mg	
¹ H NMR ((m, 3H, R		$DCl_3) \delta = 7.8$), 7.12 (d, $J=$	5 – 7.80 (m 1.2 Hz, 1H,	, 2H/D R2), 7. R1), 6.95 (d, J			
(211, 102), .	I	т		Inter	Inner		
Entry	$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 1H$	$%D_{R1}$	$I_{R2(t)}$ $N = 2H$	$I_{R2(0)}$ $N = 2H$	$%D_{R2}$	κ
			%D _{R1}			%D _{R2}	к 4.43
Entry	N = 2H	N = 1H		N = 2H	N = 2H		-
Entry 1	N = 2H 0.62	N = 1H 1.00	69	N = 2H 1.72	N = 2H 2.24	23	4.43

Table S20. Determination of the competition rate constant κ from the labelling experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazoline.

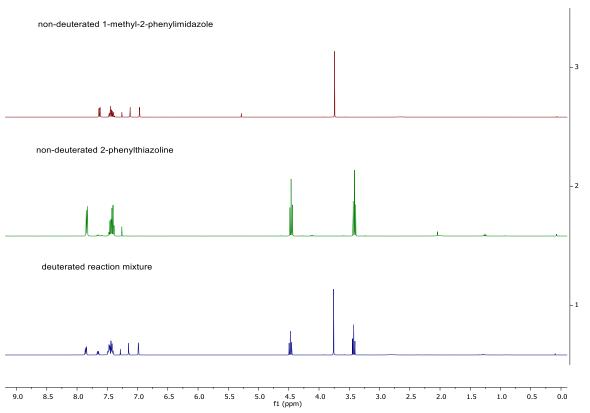


Figure S89. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

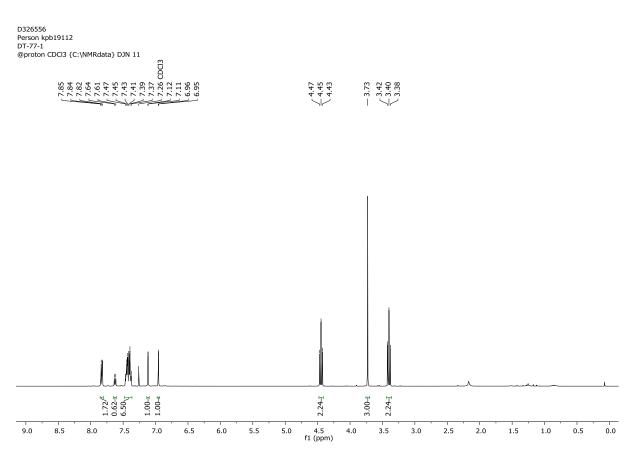
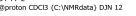


Figure S90. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2phenylimidazole and 2-phenylthiazoline (entry 1, Table S20). Person kpb19112 DT-77-2 @proton CDCl3 {C:\NMRdata} DJN 12



g		
7.85 7.86 7.82 7.61 7.45 7.45 7.45 7.45 7.42 6.95 6.95 6.95 6.95 6.95 6.95 6.95	4.47 4.45 4.43	3.73 3.42 3.40 3.38
	\leq	

m

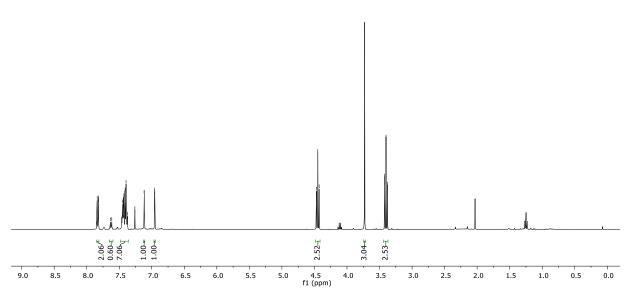


Figure S91. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2phenylimidazole and 2-phenylthiazoline (entry 2, Table S20).

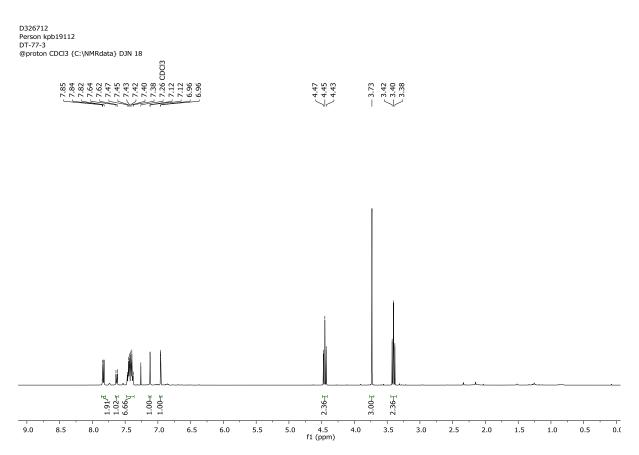
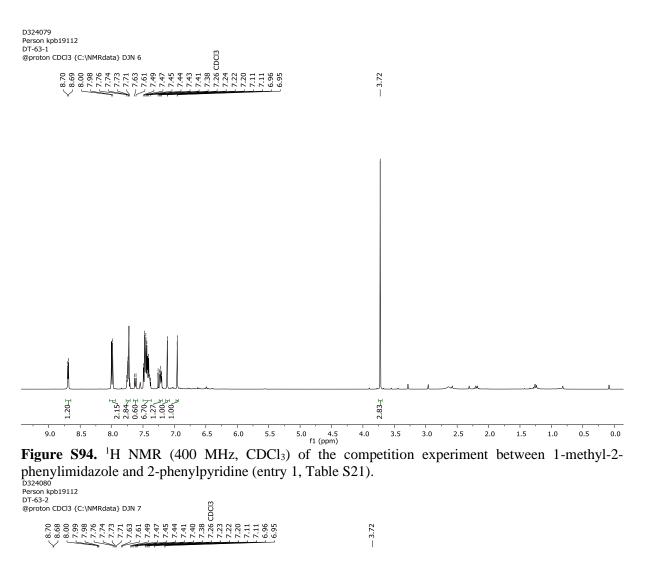


Figure S92. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazoline (entry 3, Table S20).

	Substrat	e R1	Sul	ostrate R2		Catalyst	t		
	Me			~					
						Ir-1			
		Ν.	PI		[(COD)Ir(IMes)PP	h ₃][BArF ₂₄		
Mass	15.8 r	ng]	5.5 mg		8.7 mg			
	•		**	m and at δ (R2		**			
	-	•		m and at δ (R2)) = 8.73 - 8.6	6 ppm			
-	letails of the a								
				1H, R2), 8.02					
				5 (m, 3H, R1 a		7.24 – 7.18 (m, 1H, R2		
7.12 (d, J			J=1.2 Hz, 1	H, R1), 3.73 (s					
Entry	$\begin{split} I_{R1(t)} \\ N = 2H \end{split}$	$I_{R1(0)}$ $N = 1H$	$%D_{R1}$	$I_{R2(t)}$ $N = 2H$	$\begin{split} I_{R2(0)} \\ N = 1 H \end{split}$	$%D_{R2}$	К		
1	0.59	1.00	70	2.15	1.20	10	10.95		
2	0.51	1.00	75	1.88	1.10	15	8.69		
3	0.67	1.00	67	2.04	1.13	10	10.68		
			Averag	ge $\kappa = 10.11$					
non-deuter:	ated 1-methyl-2-phe	nylimidazole	I				-3		
non-deutera	ated 2-phenylpyridin	9					- 2		
(
A	ed reaction mixture								
K	ed reaction mixture						-1		
deuterat							1		

Table S21. Determination of the competition rate constant κ from the labelling experiment between 1-methyl-2-phenylimidazole and 2-phenylpyridine.

Figure S93. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.



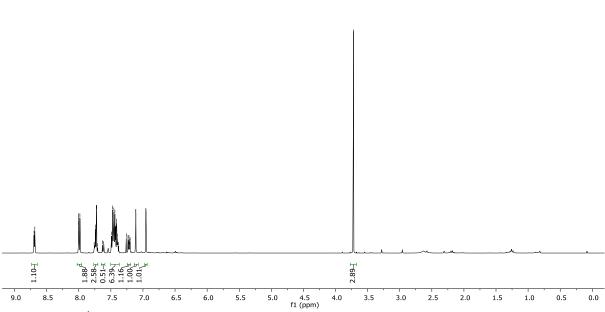


Figure S95. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylpyridine (entry 2, Table S21).

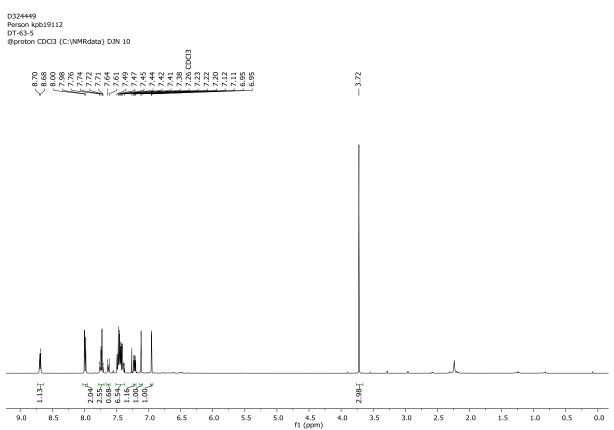


Figure S96. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylpyridine (entry 3, Table S21).

Table S22. Determination of the competition rate constant κ from the labelling experiment between 2phenylpyridine and 2-phenylpyrimidine.

Determined Spectral de ¹ H NMR (4 8.49 – 8.42 7.39 (m, 31 Entry	d against i <i>etails of th</i> 400 MHz, 2 (m, 2H/I	$\begin{array}{c} 1 \text{ at } \delta (\mathbf{R1}) = \\ \text{ntegral at } \delta \\ e \text{ deuterate} \\ \text{CDCl}_3) \delta = \\ \text{CDCl}_3) \delta = \\ \text{D, } \mathbf{R2}), 8.0 \\ \text{3H, } \mathbf{R2}), 7 \\ \hline \text{I}_{\text{R1}(0)} \\ \text{N} = 2\text{H} \\ \hline 1.94 \\ 1.88 \end{array}$	= 8.02 - 7 (R1) = 7. <i>d reaction</i> = 8.80 (d, 2 - 7.97 (5.6 mg .97 ppm and 77 – 7.70 pp <i>mixture:</i> J = 4.8 Hz, m, 2H/D R1	[(COD)Ir(at δ (R2) = 8. m and at δ (R2) 2H, R2), 8.73), 7.77 - 7.70 , 7.17 (t, $J = 4$ I _{R2(0)} N = 2H	2) = 8.80 pp 6 - 7.67 (m, 6 (m, 2H, R)	om om 1H, R 1), 7.52
Deuteration Determined Spectral de ¹ H NMR (4 8.49 – 8.42 7.39 (m, 3H Entry 1 2 3	n expected d against i etails of th 400 MHz, 2 (m, 2H/I H, R1 and $I_{R1(t)}$ N = 2H 1.39 1.58	$\begin{array}{c} 1 \text{ at } \delta (\mathbf{R1}) = \\ \text{ntegral at } \delta \\ e \text{ deuterate} \\ \text{CDCl}_3) \delta = \\ \text{CDCl}_3) \delta = \\ \text{D, } \mathbf{R2}), 8.0 \\ \text{3H, } \mathbf{R2}), 7 \\ \hline \text{I}_{\text{R1}(0)} \\ \text{N} = 2\text{H} \\ \hline 1.94 \\ 1.88 \end{array}$	= 8.02 - 7 (R1) = 7. <i>d reaction</i> = 8.80 (d, 2 - 7.97 (.25 - 7.20 %D _{R1}	.97 ppm and 77 – 7.70 pp mixture: J = 4.8 Hz, m, 2H/D R1 (m, 1H, R1) $I_{R2(t)}$	m and at δ (R 2), 8.73), 7.77 - 7.70 , 7.17 (t, $J = 4$ $I_{R2(0)}$	49 - 8.42 pp 2) = 8.80 pp - 7.67 (m, 2H, R) 4.8 Hz, 1H, 1	m 1H, R 1), 7.52 R2).
Determined <i>Spectral de</i> ¹ H NMR (4 8.49 – 8.42 7.39 (m, 3H Entry N 1 2 3	d against i etails of th 400 MHz, 2 (m, 2H/J H, R1 and $I_{R1(t)}$ N = 2H 1.39 1.58	ntegral at δ e deuterate CDCl ₃) δ D, R2), 8.0 3H, R2), 7 I _{R1(0)} N = 2H 1.94 1.88	$(\mathbf{R1}) = 7.$ d reaction = 8.80 (d, 2 - 7.97 (.25 - 7.20 %D _{R1}	77 - 7.70 pp mixture: J = 4.8 Hz, m, 2H/D R1 (m, 1H, R1) $I_{R2(t)}$	m and at δ (R 2), 8.73), 7.77 - 7.70 , 7.17 (t, $J = 4$ $I_{R2(0)}$	$2) = 8.80 \text{ pp}$ -7.67 (m, $(m, 2H, \mathbf{R})$ $1.8 \text{ Hz}, 1H, 1$	m 1H, R 1), 7.52 R2).
Entry N 1 2 3	$I_{R1(t)} = 2H$ 1.39 1.58	$I_{R1(0)} \\ N = 2H \\ 1.94 \\ 1.88$	$%D_{R1}$	I _{R2(t)}	I _{R2(0)}		
1 2 3	N = 2H 1.39 1.58	N = 2H 1.94 1.88				$%D_{R2}$	1/
2 3	1.58	1.88	28		13 - 211		ĸ
3				1.54	2.00	23	1.2
	1.59		16	1.71	2.00	15	1.1
non-deuterated		1.85	14	1.76	2.00	12	1.13
non-deuterate	ed 2-phenylpyr	imidine					
deuterated re	eaction mixture						

Figure S97. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

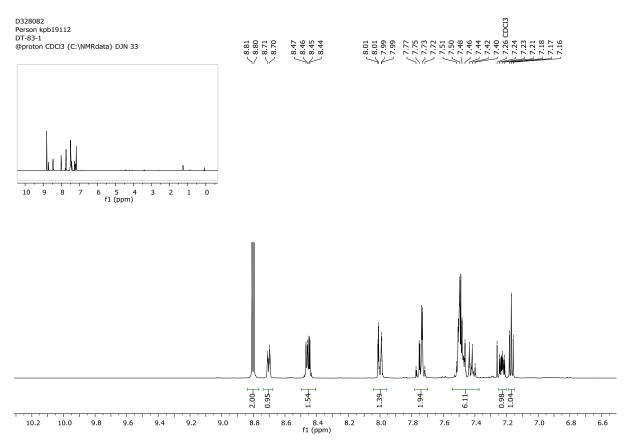


Figure S98. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyridine and

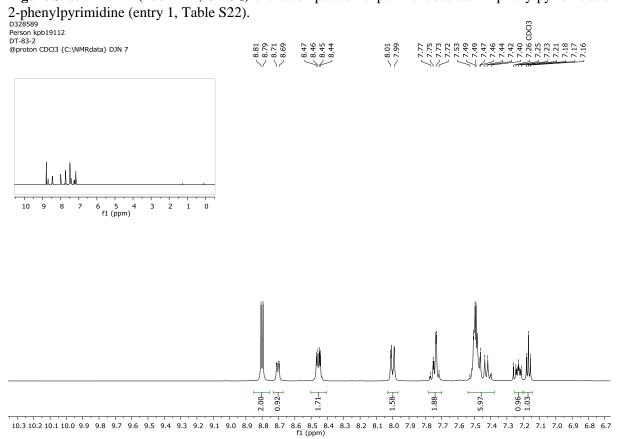


Figure S99. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyridine and 2-phenylpyrimidine (entry 2, Table S22).

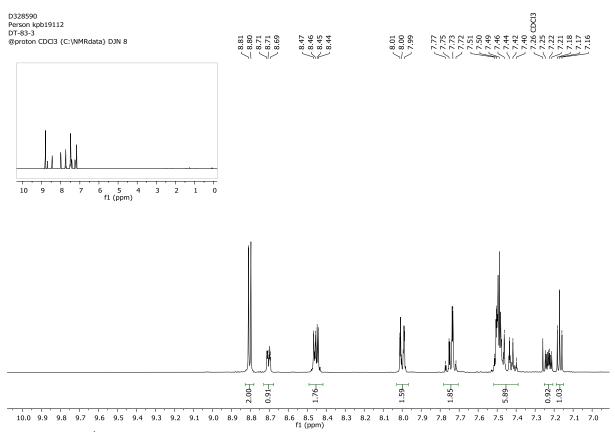


Figure S100. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyridine and 2-phenylpyrimidine (entry 3, Table S22).

3.4. Competition Experiments with (COD)Ir(IMes)Cl (Ir-2)

Table S23. Determination of the competition rate constant κ from the labelling experiment between acetophenone and benzamide.

	Substrate R1		Su	bstrate R2		Catalyst		
	O C	℃Н₃		NH ₂ Ir-2 [(COD)Ir(IMes		es)Cl]		
Mass	12.0 n	ng		12.1 mg		3.2 mg		
Determined Spectral det ¹ H NMR (4	against integ tails of the de 00 MHz, CD	gral at δ (R1) euterated real OCl_3) $\delta = 7.95$	= 2.60 ppm <i>ction mixtur</i> (d, J = 7.5)	t δ (R2) = 7.8 and at δ (R2 e: Hz, H/D R1), R2), 2.60 (s,	= 7.63 - 7.3 7.82 (d, $J = 7$		R2), 7.63	
Entry	$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 3H$	%D _{R1}	$I_{R2(t)}$ $N = 2H$	$I_{R2(0)}$ $N = 3H$	$%D_{R2}$	κ	
1	1.76	3.00	12	1.95	3.20 ^a	9	1.42	
2	1.71	3.00	15	1.76	2.93 ^b	10	1.50	
3	1.41	3.00	30	2.22	4.11 ^c	19	1.66	
	2 00. bI	$e_{(0)} = 5.93 - 3.00$	0	k = 1.53				
non-deuterated a							-3	
deuterated react	l,ull							

8.6 8.4 8.2 8.0 7.8 7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 **Figure S101.** Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture

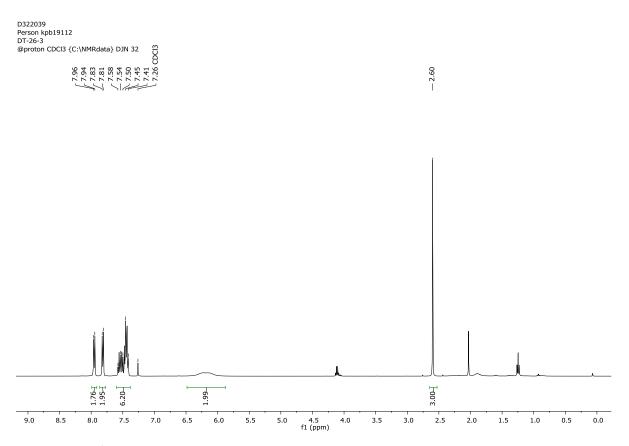


Figure S102. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzamide (entry 1, Table S23).

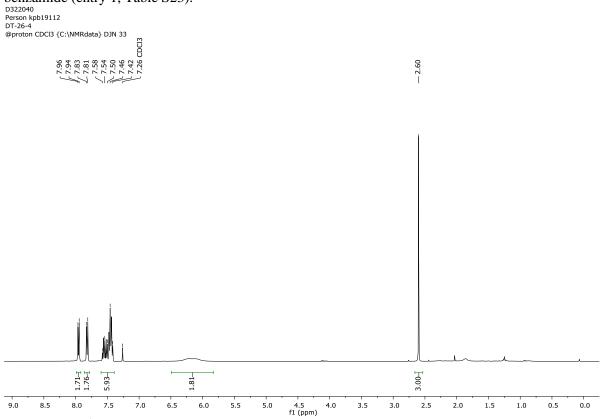


Figure S103. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzamide (entry 2, Table S23).

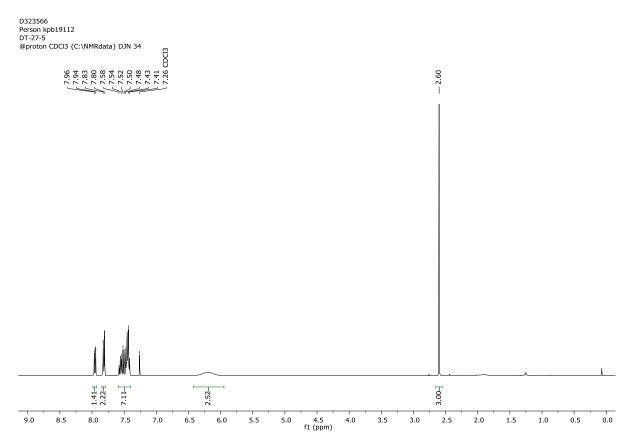


Figure S104. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and benzamide (entry 3, Table S23).

Table S24. Determination of the competition rate constant κ from the labelling experiment between benzenesulfonamide and acetophenone.

	Substrat	e R1	Su	bstrate R2		Catalyst	t
	C S	2 NH ₂		CH3	[(Ir-2 (COD)Ir(IM	es)Cl]
Mass	15.7 r	ng		12.0 mg		3.2 mg	
		$\delta (\mathbf{R1}) = 7.87$					
		gral at($\mathbf{R1}$) =			$(\mathbf{R2}) = 2.58 \text{ p}$	opm	
-	U	euterated real (180 d) = 7			7 97 7 90		1) 767
		$ASO-d_6) \delta = 7$ R2), 7.35 (br				$(m, 2H/D \mathbf{K})$	L), /.0/
	I KI and JII I _{R1(t)}	$\frac{II}{I_{R1(0)}}$		$\frac{JS(\mathbf{S}, \mathbf{SH}, \mathbf{K})}{\mathbf{I}_{R2(t)}}$	I _{R2(0)}		
Entry	N = 2H	N = 3H	$%D_{R1}$	N = 2H	N = 3H	$%D_{R2}$	κ
1	2.01	4.26 ^a	29	1.45	3.00	28	1.07
2	1.50	3.80 ^b	41	1.37	3.00	32	1.39
3	1.66	5.39°	54	1.09	3.00	46	1.27
		$_{(t)} = 6.80 - 3.0$	Average				
non-deuterated b	penzenesulfonamid	le					
non-deuterated t	penzenesulfonamid	le					- 3
non-deuterated t	benzenesulfonamid	le			I		- 3
/		le					- 3
/		le			I.		-3
)(le					-3
/		le					
non-deuterated b		le					
)(le			[
non-deuterated	acetophenone	le			I		
non-deuterated	acetophenone	le			I.	^	-2
)(acetophenone	le			I		
non-deuterated	acetophenone	le					-2
non-deuterated	acetophenone	le					-2

Figure S105. Stacked ¹H NMR (400 MHz, DMSO-*d*₆) of non-deuterated substrates and reaction mixture.



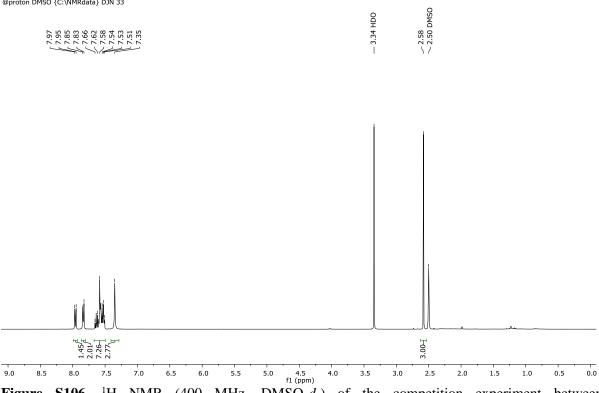
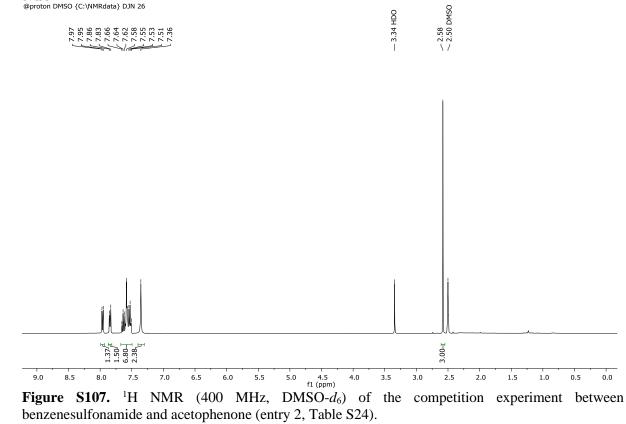


Figure S106. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between benzenesulfonamide and acetophenone (entry 1, Table S24).

D320308 Person kpb19112 DT-23-3 @proton DMSO {C:\NMRdata} DJN 26



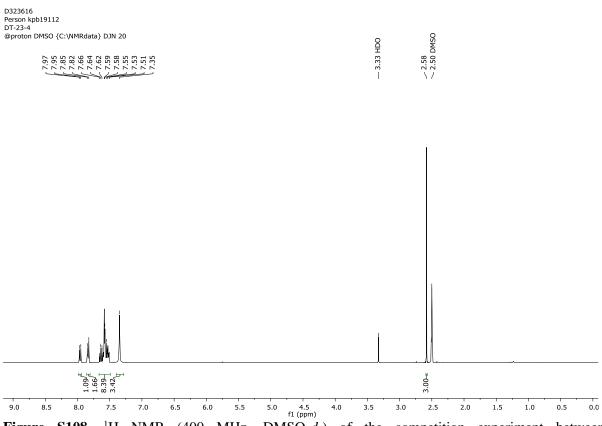


Figure S108. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between benzenesulfonamide and acetophenone (entry 3, Table S24).

Table S25. Determination of the competition rate constant κ from the labelling experiment between benzenesulfonamide and benzamide.

	Substrate R1	Substrate R2	Catalyst
	SNH ₂	NH ₂	Ir-2 [(COD)Ir(IMes)Cl]
Mass	15.7 mg	12.1 mg	3.2 mg

Deuteration expected at δ (**R1**) = 7.86 – 7.81 ppm and at δ (**R2**) = 7.90 – 7.86 ppm Determined against integral at δ (**R1**) = 7.63 – 7.54 ppm and at δ (**R2**) = 7.48 – 7.41 ppm *Spectral details of the deuterated reaction mixture:*

¹H NMR (400 MHz, DMSO- d_6) δ = 7.96 (bs, 1H, **R2**), 7.90 – 7.86 (m, 2H/D **R2**), 7.86 – 7.81 (m, 2H/D **R1**), 7.63 – 7.54 (m, 3H, **R1**), 7.54 – 7.49 (m, 1H, **R2**), 7.48 – 7.41 (m, 2H, **R2**), 7.35 (bs, 1H, **R2** and 2H, **R1**)

Entry	$\begin{split} I_{R1(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R1(0)} \\ N &= 3H \end{split}$	$%D_{R1}$	$\begin{split} I_{R2(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R2(0)} \\ N &= 2H \end{split}$	$%D_{R2}$	К
1	1.30	3.07	36	1.84	2.00	8	5.44
2	1.44	3.19	32	1.88	2.00	6	6.30
3	1.55	2.98	22	1.90	2.00	5	4.84
			Average	$\kappa = 5.53$			

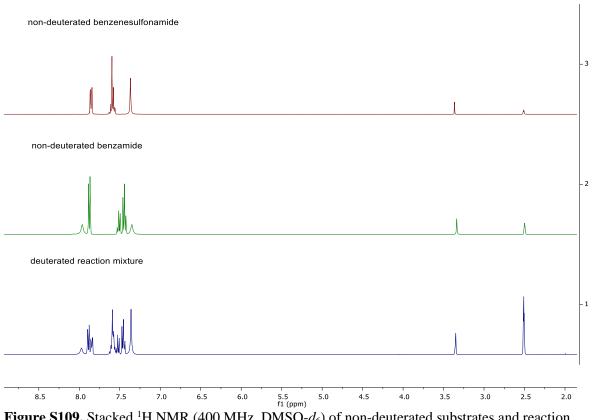


Figure S109. Stacked ¹H NMR (400 MHz, DMSO-*d*₆) of non-deuterated substrates and reaction mixture.

D323618 Person kpb19112 DT-27-5 @proton DMSO {C:\NMRdata} DJN 22

- 3.34 HDO

4.5 f1 (ppm) 0.0 9.0 8.5 8.0 . 7.5 . 7.0 6.5 6.0 5.5 5.0 4.0 3.5 . 3.0 2.5 2.0 1.5 1.0 0.5 Figure S110. ¹H NMR (400 MHz, DMSO-d₆) of the competition experiment between benzenesulfonamide and benzamide (entry 1, Table S25). B58397 Person kpb19112 dt-27-3 @proton16 DMSO {C:\NMRdata} DJN 23 - 3.34 HDO

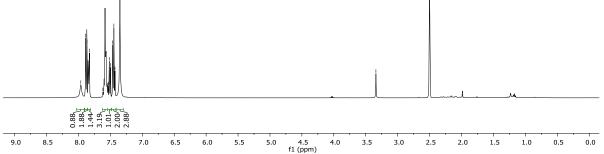


Figure S111. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between benzenesulfonamide and benzamide (entry 2, Table S25).

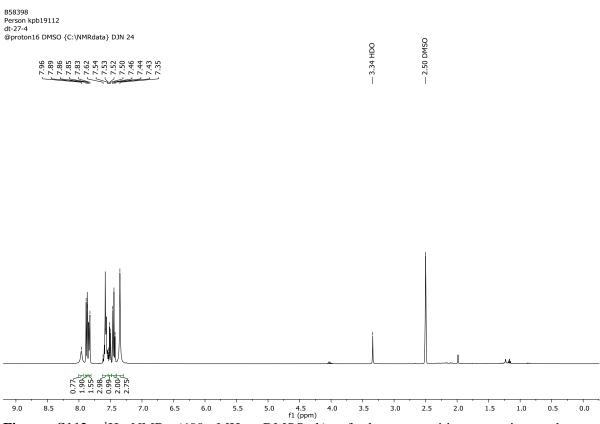


Figure S112. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between benzenesulfonamide and benzamide (entry 3, Table S25).

$\overline{)} = 7.42 - 7.3$ = 3.18 - 2.83 8 - 7.53 (m, 8 (m, 6H, R2 I _{R2(0)} N = 6H 9.79 6.12 6.77 0 = 5.50 - (6.7)	³ ppm 1H, R1), 7.4), 2.60 (s, 3 H %D _{R2} 9 8 6	g 8 - 7.43
	(COD)Ir(IM 3.2 mg 5 ppm 8 ppm 1H, R1), 7.4), 2.60 (s, 3H % D _{R2} 9 8 6	8 – 7.43 I , R1) κ 9.87 9.24
	3.2 mg 5 ppm 8 ppm 1H, R1), 7.4), 2.60 (s, 3H %D _{R2} 9 8 6	8 – 7.43 I , R1) κ 9.87 9.24
$= 3.18 - 2.88$ $8 - 7.53 (m, 6H, R2)$ $I_{R2(0)}$ $N = 6H$ 9.79 6.12 6.77	5 ppm 5 ppm 1 H, R1), 7.4), 2.60 (s, 3H % D _{R2} 9 8 6	8 – 7.43 I , R1) κ 9.87 9.24
$= 3.18 - 2.88$ $8 - 7.53 (m, 6H, R2)$ $I_{R2(0)}$ $N = 6H$ 9.79 6.12 6.77	³ ppm 1H, R1), 7.4), 2.60 (s, 3 H %D _{R2} 9 8 6	I , R1) κ 9.87 9.24
8 - 7.53 (m, 8 (m, 6H, R2) IR2(0) N = 6H 9.79 6.12 6.77	1H, R1), 7.4), 2.60 (s, 3 H) $%D_{R2}$ 9 8 6	I , R1) κ 9.87 9.24
$\frac{8 \text{ (m, 6H, R2)}}{I_{R2(0)}}$ $\frac{N = 6H}{9.79}$ 6.12 6.77), 2.60 (s, 3E %D _{R2} 9 8 6	I , R1) κ 9.87 9.24
$\frac{8 \text{ (m, 6H, R2)}}{I_{R2(0)}}$ $\frac{N = 6H}{9.79}$ 6.12 6.77), 2.60 (s, 3E %D _{R2} 9 8 6	I , R1) κ 9.87 9.24
$I_{R2(0)} \\ N = 6H \\ 9.79 \\ 6.12 \\ 6.77 \\$	%D _{R2} 9 8 6	к 9.87 9.24
N = 6H 9.79 6.12 6.77	9 8 6	9.87 9.24
6.12 6.77	8 6	9.24
6.77	6	
		10.09
$_{0} = 5.50 - (6.7)$	77) / 6 × 3	
$_{0} = 5.50 - (6.7)$	77) / 6 × 3	
		- 3
		- 2
		- 1
	u.	
	lab	<u> </u>
	1	l

Table S26. Determination of the competition rate constant κ from the labelling experiment between acetophenone and *N*,*N*-dimethylbenzamide.

Figure S113. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

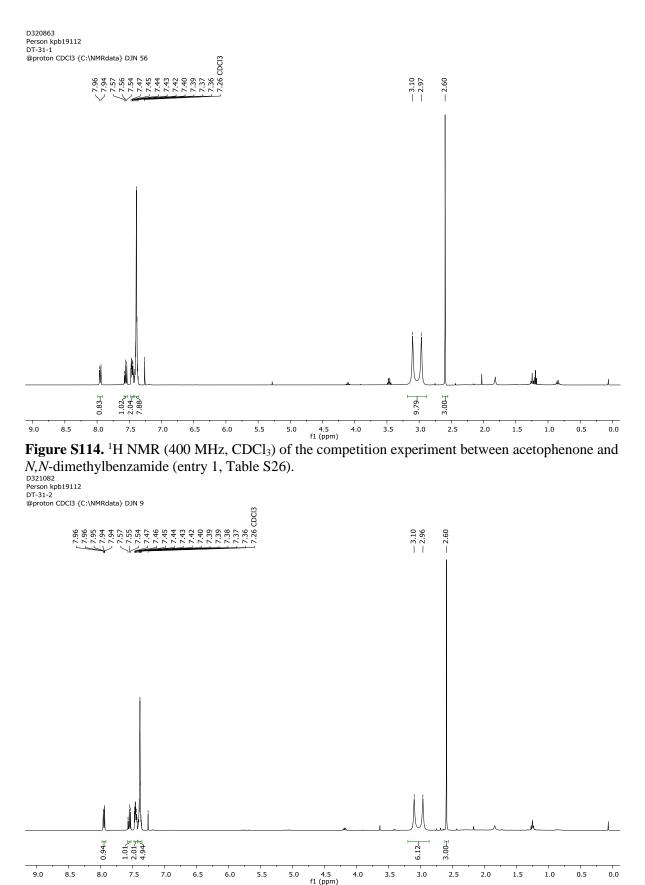


Figure S115. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and *N*,*N*-dimethylbenzamide (entry 2, Table S26).

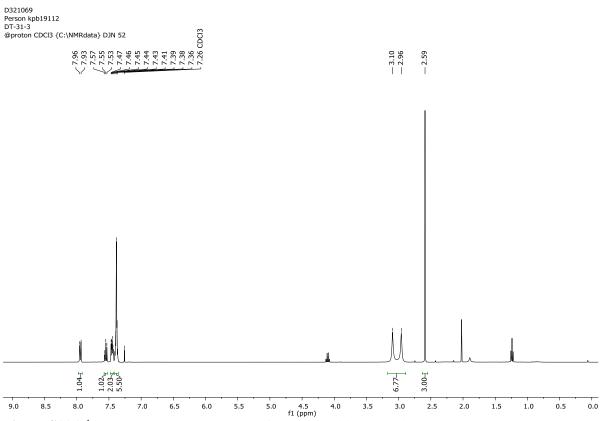


Figure S116. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between acetophenone and *N*,*N*-dimethylbenzamide (entry 3, Table S26).

Table S27. Determination of the competition rate constant κ from the labelling experiment between benzenesulfonamide and methylphenylsulfone.

	Substrat	e R1	Su	bstrate R2		Catalyst		
	S ^{O2} NH ₂			O ₂ S_Me		Ir-2 [(COD)Ir(IMes)0		
Mass	15.7 r	ng		15.6 mg		3.2 m	g	
Determined Spectral de ¹ H NMR (4	n expected at d against integ etails of the de 400 MHz, DM 2), 7.64 – 7.62	gral at δ (R1) euterated read ISO- d_6) $\delta = 7$	= 7.60 – 7.5 ction mixtur .95 – 7.93 (n	55 ppm and a <i>e:</i> n, 2H/D R2),	t δ (R2) = 7.6 7.85 - 7.83 (54 – 7.62 ppn m, 2H/D R1)		
Entry	$\frac{I_{R1(t)}}{I_{R1(t)}}$ $N = 2H$	$\frac{I_{R1(0)}}{N = 3H}$	%D _{R1}	$\frac{I_{R2(t)}}{N = 2H}$	$\frac{I_{R2(0)}}{I_{R2(0)}}$ N = 2H	%D _{R2}	κ	
1	1.42	2.94	28	1.98	2.00	1	32.07	
2	0.64	2.68	64	1.95	2.00	3	40.55	
3	1.42	3.30	35	1.98	2.00	1	43.56	
	1.+∠ d benzenesulfonam			$e \kappa = 38.73$		-		
					Λ		- 3	
non-deuterate		iide			λ			
non-deuterate	d benzenesulfonam	iide					-3	

4.5 4.0 f1 (ppm) 0.0 9.0 8.5 8.0 7.5 7.0 5.0 3.5 3.0 2.5 1.0 0.5 6.5 6.0 5.5 2.0 1.5

Figure S117. Stacked ¹H NMR (400 MHz, DMSO- d_6) of non-deuterated substrates and reaction mixture.

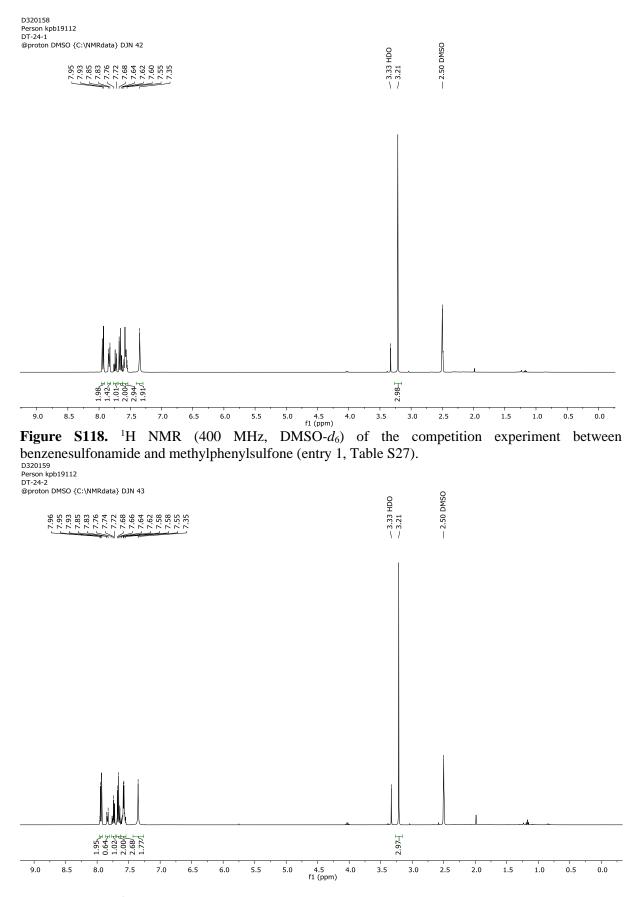


Figure S119. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between benzenesulfonamide and methylphenylsulfone (entry 2, Table S27).

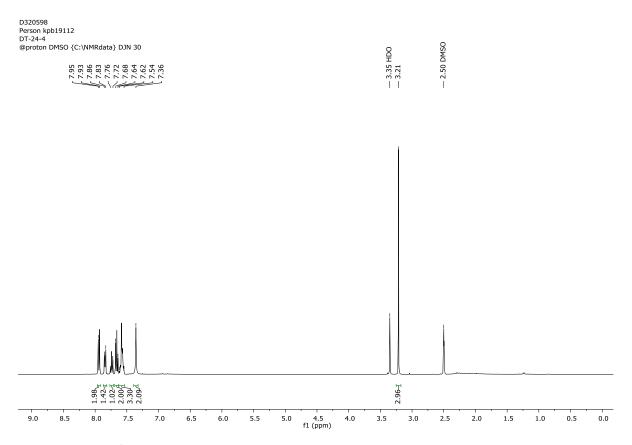
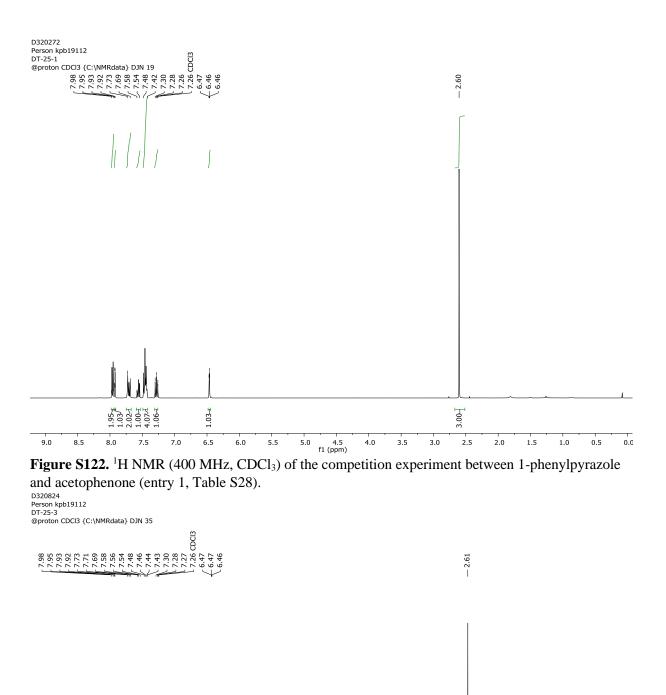


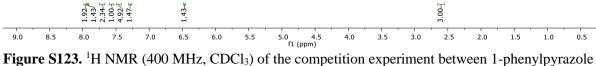
Figure S120. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between benzenesulfonamide and methylphenylsulfone (entry 3, Table S27).

	Substrat	e R1	Su	bstrate R2		Cataly	Catalyst	
	آ Ph ^{-N}	Ň		CH3		Ir-2 [(COD)Ir(IN		
Mass	14.4 ı	ng		12.0 mg		3.2 m	g	
	•	$\delta (\mathbf{R1}) = 7.73$				**		
	-	gral at δ (R1)			$t \delta (\mathbf{R2}) = 2.6$	60 ppm		
-	0	euterated read OCl_3) $\delta = 7.98$			$D^{2}(A I - 2A)$	H ₇ 1H D1)	775 76	
		1, 7.60 - 7.53			-			
		5 (m, 1H, R1)	-		2 (111, 211, 111	una 211, 112)	, 7.31 7.2	
	I _{R1(t)}	I _{R1(0)}		I _{R2(t)}	I _{R2(0)}	0/ D		
Entry	N = 2H	N = 1H	$%D_{R1}$	N = 2H	N = 3H	$%D_{R2}$	κ	
1	0.99 ^a	1.03	52	1.95	3.00	3	28.94	
2	0.91 ^b	1.43	68	1.92	3.00	4	28.05	
3	0.93 ^c	1.05	56	1.95	3.00	3	32.17	
			0	$\kappa = 29.72$				
$I_{R1(t)} = 2.0$	$02 - 1.03; {}^{b}I_{R1}$	$_{(t)} = 2.34 - 1.4$	$3; ^{c} I_{R1(t)} = 1$.98 – 1.05;				
non-deuterate	ed 1-phenylpyrazole						1	
							-3	
non doutora	ited acetophenone							
non-deutera								
							- 2	
/	mh_i							
deuterate	d reaction mixture					1		
							- 1	
de .	"M_M							
MM							1	
MM							I	

Table S28. Determination of the competition rate constant κ from the labelling experiment between 1-phenylpyrazole and acetophenone.

Figure S121. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.





and acetophenone (entry 2, Table S28).

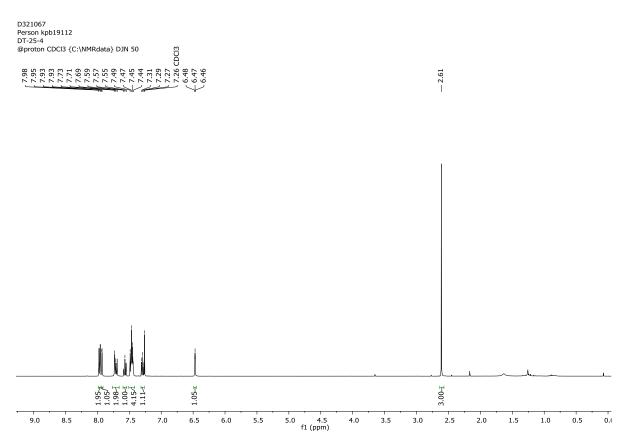


Figure S124. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and acetophenone (entry 3, Table S28).

Substrate R1		Substrate R2			Catalyst			
	Ph ^{-N} N		Ph			Ir-2 [(COD)Ir(IMes)Cl]		
Mass	14.4 mg		15.5 mg		3.2 mg			
Deuteratio	n expected at	$\delta(\mathbf{R1}) = 7.78$	8 – 7.67 ppn	n and at δ (R 2	(2) = 8.04 - 7.9	97 ppm		
Determine	d against inte	gral at δ (R1)	= 6.50 - 6.4	42 ppm and at	$t \delta\left(\mathbf{R2}\right) = 8.7$	2 – 8.68 ppm	l	
Spectral de	etails of the d	euterated read	ction mixtur	e:				
-	400 MHz, CE				– 7.97 (m, 21	H/D R2), 7.92	2 (d, $J = 2$	
Hz, 1H, R	1), 7.78 – 7.6	7 (m, 2H, R2	, 1H, R1 , 2	H/D R1), 7.5	2 – 7.38 (3H,	R2 and 2H,	R1), 7.31	
	H, R1), 7.24 -	-						
Entry	I _{R1(t)}	I _{R1(0)}	$%D_{R1}$	I _{R2(t)}	I _{R2(0)}	%D _{R2}	к	
	N = 2H	N = 1H	/0D _{R1}	N = 2H	N = 1H			
1	1.65 ^a	1.00	18	1.75	1.00	13	1.44	
2	1.29 ^b	1.00	36	1.22	0.85	28	1.32	
3	1.23 ^c	1.00	39	1.32	0.94	30	1.37	
			Averag	e <i>κ</i> = 1.38				
$a I_{P1(4)} = 4.6^{4}$	5-1.00-(1.00)	$(x^2)^{\cdot b} I_{P_1(x)} = 3$	99-1 00-(0	85×2) · · In1($= 4 \ 11 - 1 \ 00 -$	$-(0.94 \times 2)$		
I KI(t) 110.		(2), IKI(t) 5.	<u>,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,</u>	1007(2) , 1 KI(t)		(0.9 1/2)		
non-deutera	ated 1-phenylpyrazo	le						

Table S29. Determination of the competition rate constant κ from the labelling experiment between 1-phenylpyrazole and 2-phenylpyridine.

non-deuterated 2-phenylpyridine

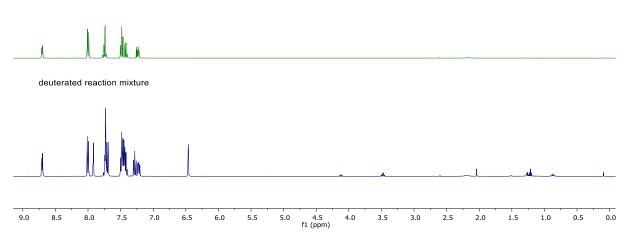
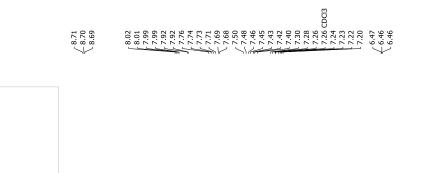


Figure S125. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D319184 Person kpb19112 DT-19-3 @proton CDCI3 {C:\NMRdata} DJN 44



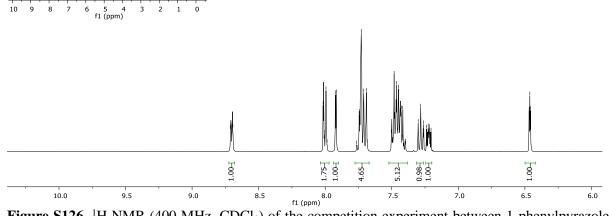


Figure S126. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 1, Table S29).

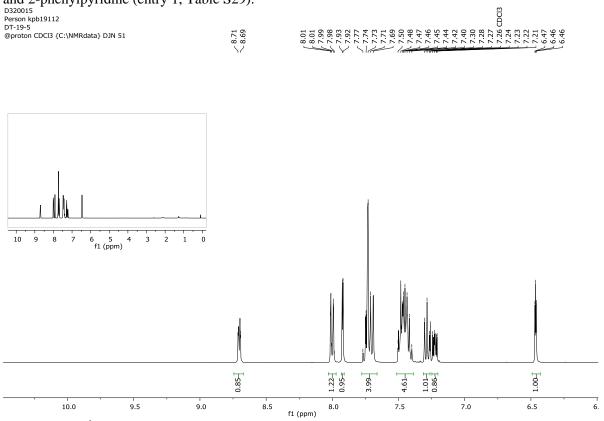


Figure S127. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 2, Table S29).

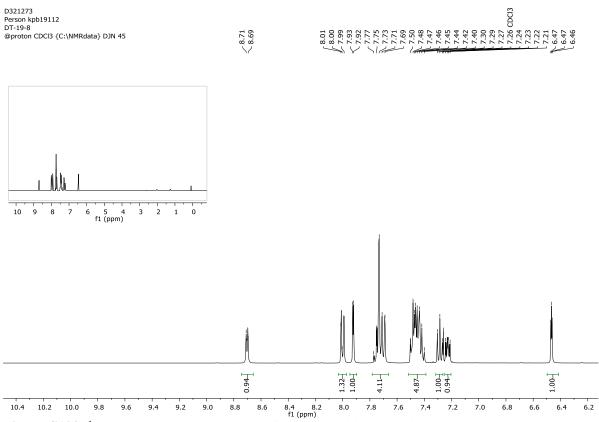


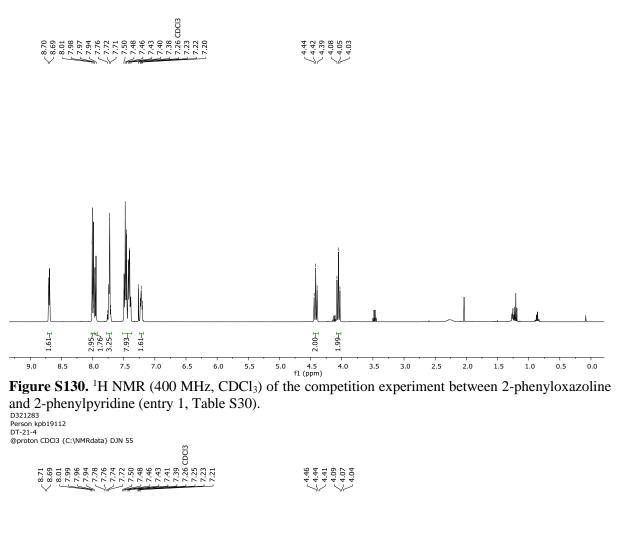
Figure S128. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylpyridine (entry 3, Table S29).

	Substrate	e R1	Su	bstrate R2		Catalys	t	
	N O			hN		Ir-2 [(COD)Ir(IMes)Cl]		
Mass	14.7 n	ng		15.5 mg		3.2 mg	5	
Determine Spectral de H NMR (4 (m, 2H/D)	n expected at d against integ etails of the de 400 MHz, CD R1), 7.77 – 7.0 (t, $J = 9.5$ Hz,	gral at δ (R1) euterated read (Cl ₃) δ = 8.73 69 (m, 2H R2)	= 4.44 ppm <i>ction mixtur</i> - 8.66 (m, 2), 7.52 - 7.2	and at δ (R2 <i>e:</i> 1H, R2), 8.02 37 (m, 3H, R 1	(x = 8.73 - 8.6)	6 ppm H/D R2), 7.9'		
Entry	$\frac{I_{R1(t)}}{N = 2H}$	$\frac{I_{R1(0)}}{I_{R1(0)}}$ $N = 2H$	%D _{R1}	$\frac{I_{R2(t)}}{I_{R2(t)}}$ $N = 2H$	$I_{R2(0)}$ $N = 1H$	$%D_{R2}$	κ	
1	1.76	2.00	12	2.95	1.61	8	1.46	
2	1.52	2.00	24	1.66	1.02	19	1.33	
3	1.28	2.00	36	2.02 e $\kappa = 1.41$	1.38	27	1.43	
non-deutera	ated 2-phenyloxazoli	ne			. 4		-3	
non-deuter	rated 2-phenylpyridin	e						
							- 2	
deuterat	ted reaction mixture							
							- 1	
9.0 8.5	8.0 7.5 7.0	6.5 6.0 5.	5 5.0 4.5 f1 (ppm)	4.0 3.5 3	.0 2.5 2.0	1.5 1.0 0.5	0.0	

Table S30. Determination of the competition rate constant κ from the labelling experiment between 2-phenyloxazoline and 2-phenylpyridine.

Figure S129. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D319185 Person kpb19112 DT-21-2 @proton CDCI3 {C:\NMRdata} DJN 45



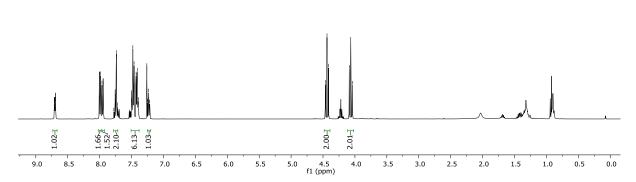


Figure S131. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylpyridine (entry 2, Table S30).

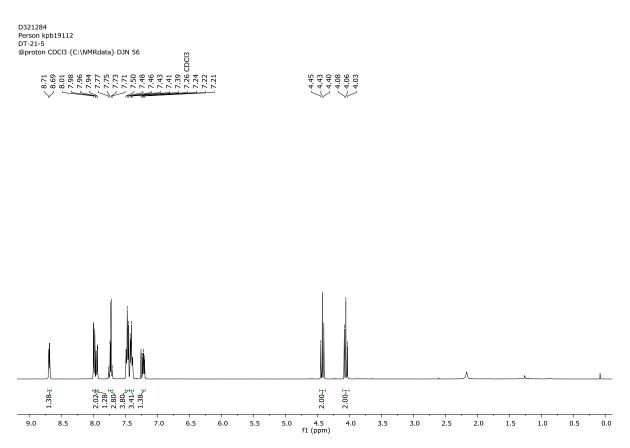


Figure S132. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenyloxazoline and 2-phenylpyridine (entry 3, Table S30).

Table S31. Determination of the competition rate constant κ from the labelling experiment between 1-phenylpyrazole and 2-phenylthiazole.

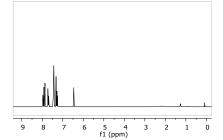
	Substrate R1			bstrate R2		Catalyst Ir-2 [(COD)Ir(IMes)Cl]		
	Ph ^N N			S N				
Mass	14.4 n	ng		16.1 mg		3.2 mg		
Deuteratio	n expected at	$\delta (\mathbf{R1}) = 7.78$	8 – 7.67 ppn	n ppm and at a	δ (R2) = 8.02	- 7.96 ppm		
	d against integ				$t \delta (\mathbf{R2}) = 7.8$	8 ppm		
-	etails of the de							
	400 MHz, CD							
	Hz, 1H, R2), $(d, J = 3.3 \text{ Hz}, d)$							
N 2), 7.33 ($\frac{(u, J - 5.5 \text{ HZ},}{I_{R1(t)}}$	$I\Pi, K2), 7.2$ $I_{R1(0)}$	9 (l, J – 7.4	$I_{R2(t)}$	$I_{R2(0)} = 0.43$ (ш, 1 п , кт <i>)</i> .		
Entry	N = 2H	N = 1H	$%D_{R1}$	N = 2H	N = 1H	$%D_{R2}$	κ	
1	0.82ª	1.00	59	1.25	1.10	43	1.5	
2	1.12 ^b	1.00	44	1.52	1.05	28	1.7	
3	1.19 ^c	1.00	41	1.41	1.00	30	1.4	
$I_{R1(t)} = 1.8$	$2-1.00; {}^{b}I_{R1(t)}$	= 2.12 - 1.00;	$I_{R1(t)} = 2.1$	9–1.00;				
non-deuterate	d 1-phenylpyrazole							
	d 1-phenylpyrazole							
non-deutera	ted 2-phenylthiazole							
non-deutera	ted 2-phenylthiazole							

Figure S133. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D321928 Person kpb19112 DT-30-2 @proton CDCI3 {C:\NMRdata} DJN 22

7.199 7.197 7.192 7.192 7.188 7.188 7.188 7.188 7.188 7.173 7.144 7.144 7.144 7.144 7.144 7.144 7.129 7.133 7.130 7.132 7.133 7.132 7.133 7.132 7.133 7.133 7.133 7.133 7.133 7.133 7.133 7.133 7.133 7.133 7.1447 7.1447 7.1447 7.1447 7.1447 7.1447 7.1447 7.1447 7.1447 7.1447 7.

 $\frac{6.47}{6.46}$



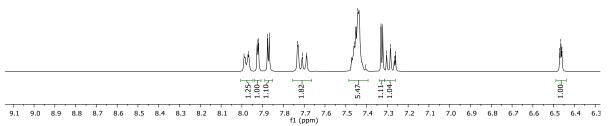


Figure S134. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylthiazole (entry 1, Table S31).

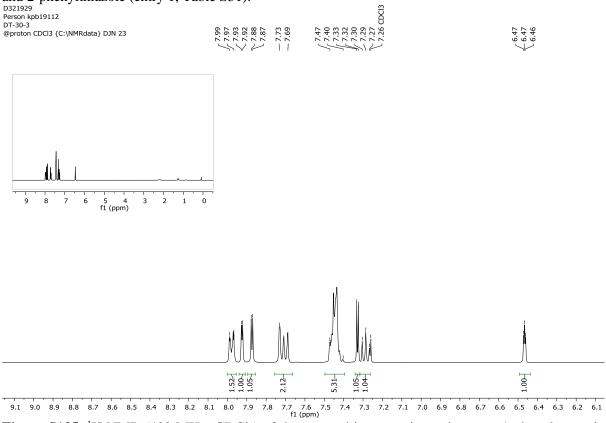


Figure S135. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylthiazole (entry 1, Table S31).

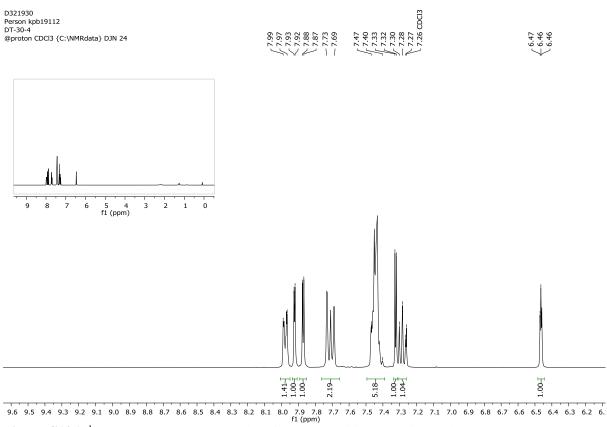
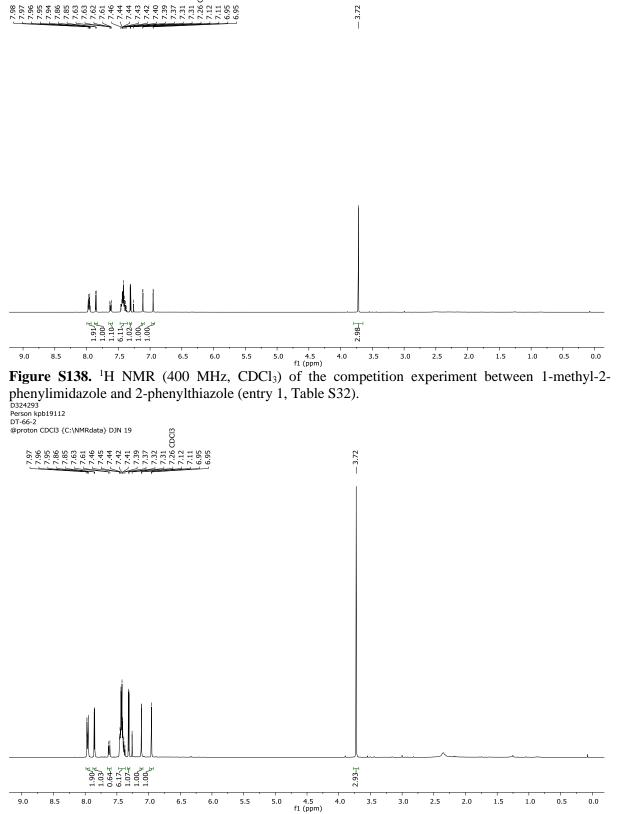


Figure S136. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-phenylpyrazole and 2-phenylthiazole (entry 1, Table S31).

	Substrate R1		Su	bstrate R2		Catalyst		
	Me			ș-1				
			<u></u>			Ir-2		
		N				[(COD)Ir(IMes)Cl]		
Mass	15.8 r	ng		16.1 mg		3.2 mg	5	
Deuteratio	n expected at	δ (R1) = 7.67	7 – 7.61 ppm	and at δ (R 2	(3) = 8.02 - 7.9	6 ppm	-	
	d against integ					- 11		
	etails of the de							
¹ H NMR (4	400 MHz, CD	Cl_3) $\delta = 8.02$	- 7.96 (m, 2	2H/D, R2), 7.3	88 (d, $J = 3.3$	Hz, 1H, R2),	7.67 – 7.	
(m, 2H/D,	R1), 7.50 – 7	7.38 (m, 3H, 1	R1 and 3H,	R2), 7.33 (d,	J = 3.3 Hz,	1H, R2), 7.12	2 (d, $J = 1$	
Hz, 1H, R	1), 6.95 (d, <i>J</i> =	= 1.2 Hz, 1H,	R1), 3.74 (s	s, 3H, R1)				
Entry	$\begin{split} I_{R1(t)} \\ N &= 2H \end{split}$	$I_{R1(0)}$ $N = 1H$	$\%D_{R1}$	$\begin{split} I_{R2(t)} \\ N = 2H \end{split}$	$\begin{split} I_{R2(0)} \\ N &= 1 H \end{split}$	$%D_{R2}$	κ	
1	1.10	1.00	45	1.91	1.00	5	12.98	
2	0.63	1.00	69	1.90	1.03	8	14.29	
3	0.71	1.00	65	1.95	1.06	8	12.39	
			Average	$\kappa = 13.22$				
non-deutera	ated 1-methyl-2-phe	nylimidazole						
			1				- 3	
	rated 2-phenylthiazo						- 3	
non-deuter							- 3	
non-deute	rated 2-phenylthiazo		_ I					
non-deute	rated 2-phenylthiazo							
non-deute	rated 2-phenylthiazo		_I				-2	
non-deute	rated 2-phenylthiazo						-2	

Table S32. Determination of the competition rate constant κ from the labelling experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazole.

Figure S137. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.



D324290 Person kpb19112 DT-66-1 @proton CDCI3 {C:\NMRdata} DJN 16

CDC13

Figure S139. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazole (entry 2, Table S32).

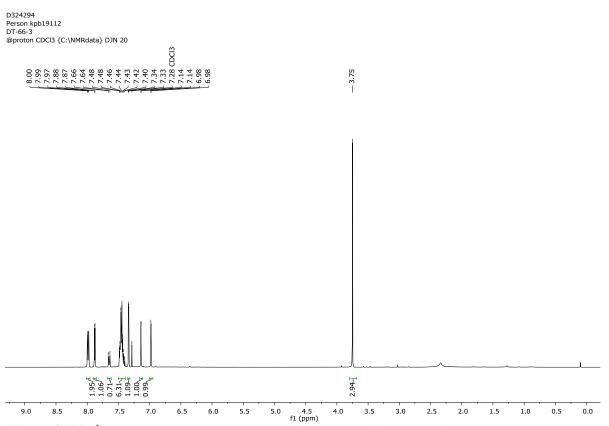


Figure S140. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazole (entry 3, Table S32).

Table S33 Determination of the competition rate constant κ from the labelling experiment between 1methyl-2-phenylimidazole and 2-phenylpyridine.

Substrate	e R1	Su	bstrate R2		Catalyst		
Me	N	Ρ	h	[((Ir-2 [(COD)Ir(IMes)Cl]		
15.8 mg			15.5 mg		3.2 mg		
n expected at d against integ etails of the de 400 MHz, CD	δ (R1) = 7.67 gral at δ (R1) euterated read (Cl ₃) δ = 8.73	7 – 7.61 ppm =7.12 ppm <i>ction mixtur</i> – 8.66 (m, 1	and at δ (R2 and at δ (R2) e: H, R2), 8.02) =8.02 - 7.9 = 7.77 - 7.69 - 7.96 (m, 2H	6 ppm 9 ppm I/D R2), 7.7	7-7.	
						7.18 (
$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 1H$	%D _{R1}	$I_{R2(t)}$ $N = 2H$	$I_{R2(0)}$ $N = 2H$	%D _{R2}	к	
1.58	1.00	21	2.09	2.32	10	2.2	
1.60	1.00	20	2.02	2.28	11	1.8	
1.36	0.87			2.00	12	1.9	
	L						
!-phenylpyridine						_	
d_h_							
action mixture							
			1				
	Me 15.8 m 15.8 m 15.8 m 15.8 m 15.8 m 15.8 m 15.8 m 15.8 m 16.0 m 1.2 (d, $J=1.2$ m 1.58 m 1.60 m 1.36	$\frac{15.8 \text{ mg}}{15.8 \text{ mg}}$ The of DCM was increased in expected at δ (R1) = 7.67 and against integral at δ (R1) etails of the deuterated reaction of the deuterated reactio	Me f(x) = f(x) = f(x	$ \begin{array}{c} Me \\ j \\ $	$ \begin{array}{c} Me \\ if if it is increased in the integrated integrated in the integrated int$	$\frac{Me}{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{\zeta_{$	

Figure S141. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

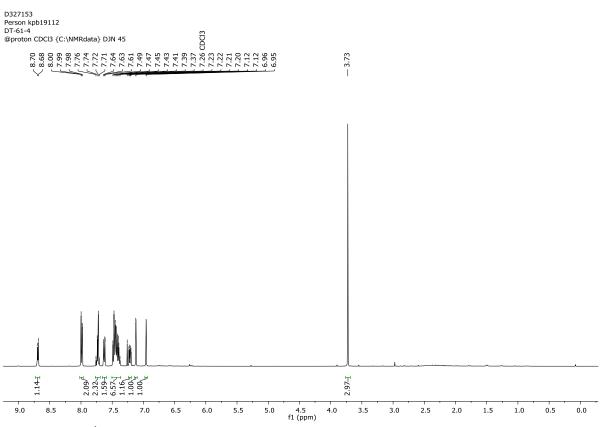


Figure S142. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2phenylimidazole and 2-phenylpyridine (entry 1, Table S33).

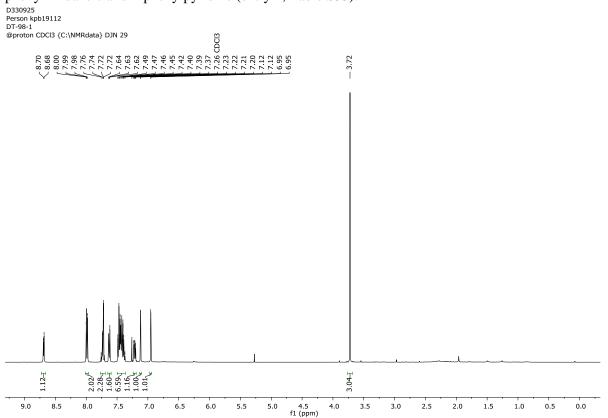


Figure S143. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2phenylimidazole and 2-phenylpyridine (entry 2, Table S33).

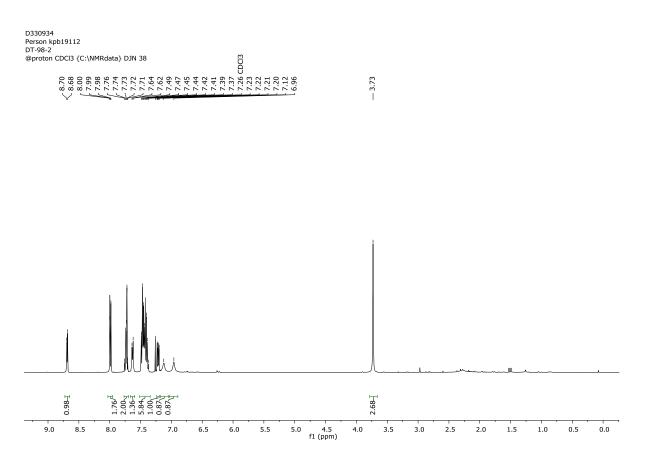
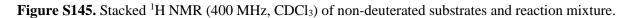
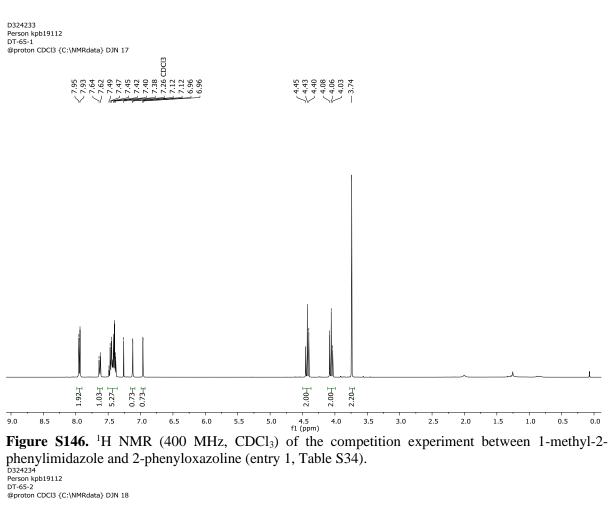


Figure S144. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylpyridine (entry 3, Table S33).

Table S34. Determination of the competition rate constant κ from the labelling experiment between 1-methyl-2-phenylimidazole and 2-phenyloxazoline.

	Substrate R1 Me N		Su	bstrate R2		Catalyst Ir-2 [(COD)Ir(IMes)Cl]		
				N				
Mass	15.8 n	ng		14.7 mg		3.2 mg		
Determined Spectral der ¹ H NMR (4 (m, 3H, R1	l against integ tails of the de 00 MHz, CD and 3H, R2)	gral at δ (R1) euterated read (Cl ₃) δ = 7.98	= 7.12 ppm ction mixtur - 7.90 (m, 2 .2 Hz, 1H, 1	2H/D R2), 7.6 R1), 6.96 (d, .	() = 4.44 ppm (57 - 7.61 (m,)	2H/D, R1), 7		
Entry	$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 1H$	%D _{R1}	$I_{R2(t)}$ $N = 2H$	$\begin{split} I_{R2(0)} \\ N &= 2H \end{split}$	$%D_{R2}$	к	
1	1.03	0.73	29	1.92	2.00	4	8.55	
2	1.31	0.93	30	1.91	2.00	5	7.61	
3	1.76	1.05	16	1.96 e $\kappa = 8.30$	2.00	2	8.74	
							- 3	
non-deutera	ited 2-phenyloxazo	line						
							- 2	
deuterated	reaction mixture							
							- 1	





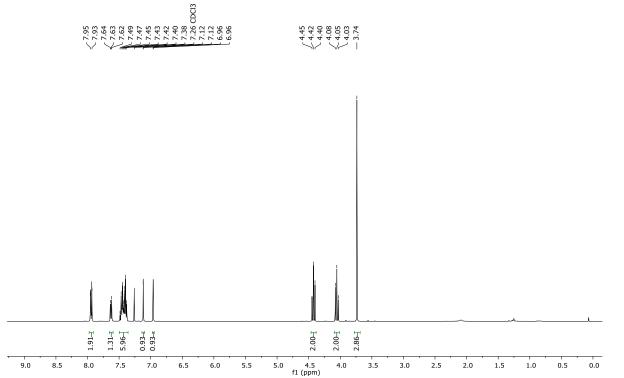


Figure S147. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2phenylimidazole and 2-phenyloxazoline (entry 2, Table S34).

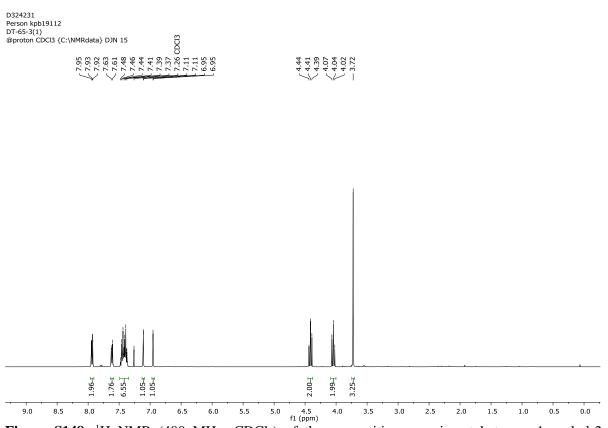


Figure S148. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenyloxazoline (entry 3, Table S34).

	Substrate	e R1	Sul	bstrate R2		Catalys	st	
	Me N			N				
		× ×		<\s∕		Ir-2		
		IN				[(COD)Ir(IN	les)CI]	
Mass	15.8 n	ng]	16.3 mg		3.2 mg	5	
	n expected at	•						
	d against integ) = 4.45 ppm			
-	etails of the de							
	400 MHz, CD	,						
	1 and 3H, R2)				<i>J</i> =1.2 Hz, 1H	(, R1), 4.45 (t	J = 8.3	
Iz, 2H, R	2), 3.73 (s, 3H		, J = 8.3 Hz,		T			
Entry	$I_{R1(t)}$ $N = 2H$	$I_{R1(0)}$ $N = 1H$	$%D_{R1}$	$I_{R2(t)}$ $N = 2H$	$I_{R2(0)}$ $N = 2H$	$%D_{R2}$	κ	
1	0.87	1.00	57	1.72	2.32	26	2.78	
2	0.90	1.00	55	1.51	2.34	35	1.82	
3	0.99	1.00	51	1.66	2.26	27	2.28	
			Average	$\kappa = 2.29$				
							- 3	
non-deut	erated 2-phenylthiaz	oline						
				I.				
					L		- 2	
deutera	ted reaction mixture							
							- 1	
	al a co							
			M	/M				

Table S35. Determination of the competition rate constant κ from the labelling experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazoline.

Figure S149. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

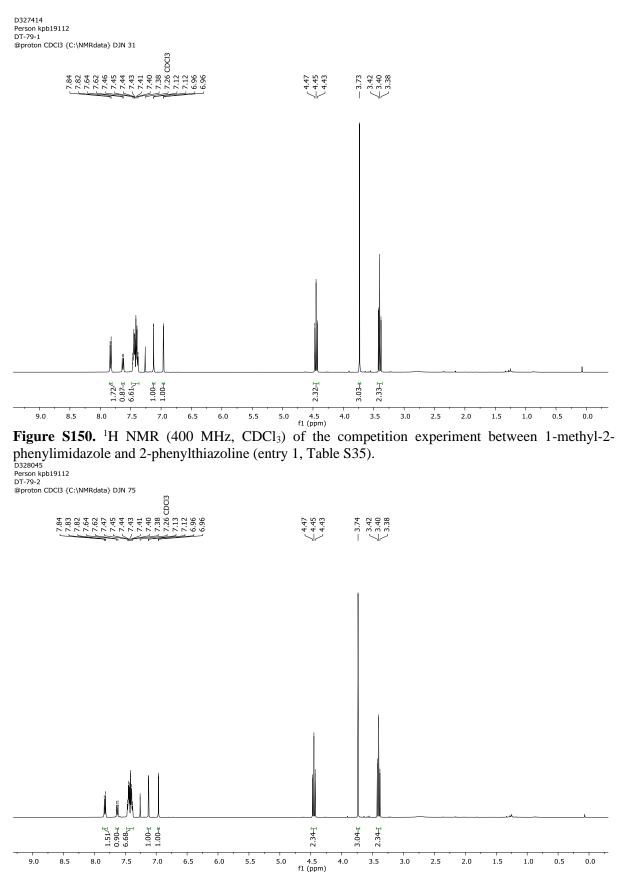


Figure S151. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazoline (entry 2, Table S35).

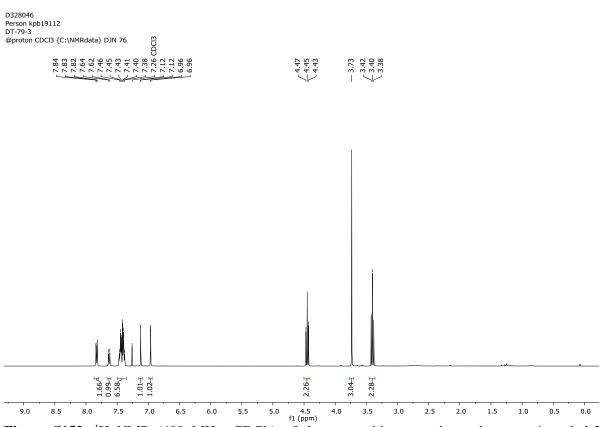


Figure S152. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 1-methyl-2-phenylimidazole and 2-phenylthiazoline (entry 3, Table S35).

Table S36. Determination of the competition rate constant κ from the labelling experiment between 2-phenylthiazoline and 2-phenyloxazoline.

	Substrate R1	Substrate R2	Catalyst
	N S		Ir-2 [(COD)Ir(IMes)Cl]
Mass	16.3 mg	14.7 mg	3.2 mg

Deuteration expected at δ (**R1**) = 7.86 – 7.81 ppm and at δ (**R2**) = 7.98 – 7.93 ppm Determined against integral at δ (**R1**) = 3.41 ppm and at δ (**R2**) = 4.06 ppm

Spectral details of the deuterated reaction mixture:

¹H NMR (400 MHz, CDCl₃) δ = 7.98 – 7.93 (m, 2H/D **R2**), 7.86 – 7.81 (m, 2H/D, **R1**), 7.51 – 7.36 (m, 3H, **R1** and 3H, **R2**), 4.49 – 4.40 (m, 2H, **R1** and 2H, **R2**), 4.06 (t, *J* = 9.5 Hz, 2H, **R2**), 3.41 (t, *J* = 8.4 Hz, 2H, **R1**).

Entry	$\begin{split} I_{R1(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R1(0)} \\ N &= 2 H \end{split}$	$%D_{R1}$	$\begin{split} I_{R2(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R2(0)} \\ N &= 2H \end{split}$	$%D_{R2}$	к
1	1.47	2.45	40	1.50	2.00	25	1.78
2	1.88	2.76	32	1.62	2.00	19	1.82
3	1.47	2.49	41	1.50	2.00	25	1.83
			Average	е <i>к</i> = 1.81			

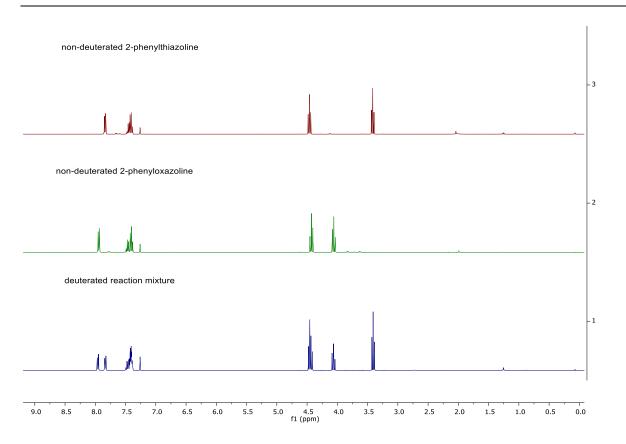
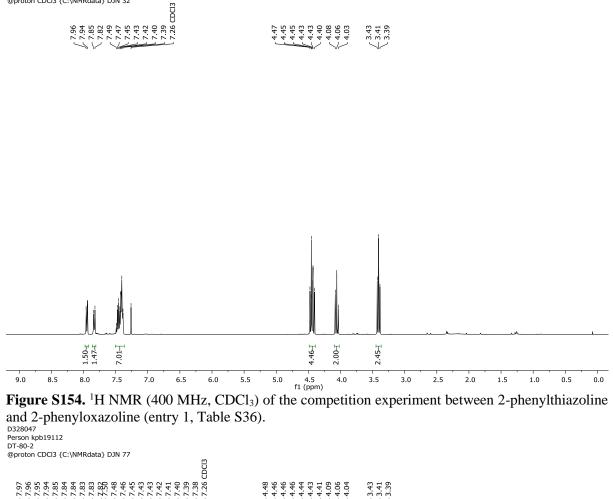
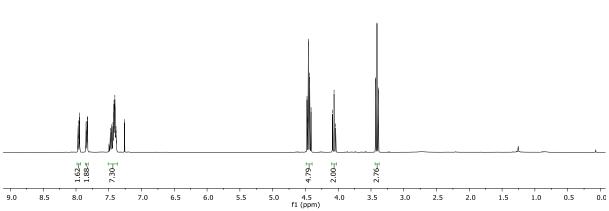
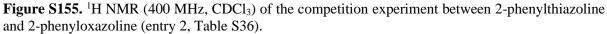


Figure S153. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D327415 Person kpb19112 DT-80-1 @proton CDCI3 {C:\NMRdata} DJN 32







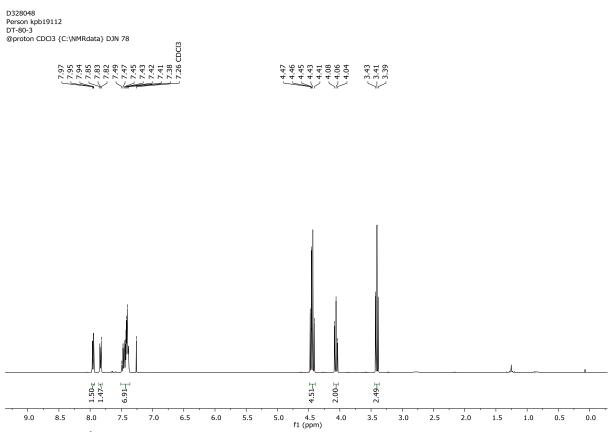


Figure S156. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylthiazoline and 2-phenyloxazoline (entry 3, Table S36).

Table S37. Determination of the competition rate constant κ from the labelling experiment between 2-
phenylpyrimidine and 2-phenylpyridine.

	Substrat	e R1	Substrate R2			Catalyst		
	N ² Ph	N	Ρ	hN		Ir-2 [(COD)Ir(IN	/les)Cl]	
Mass	15.6 r	ng	15.5 mg			3.2 mg		
Deuteratio	n expected at	$\delta(\mathbf{R1}) = 8.49$	9 – 8.42 ppm	and at δ (R 2	() = 8.02 - 7.9	97 ppm		
Determine	d against integ	gral at δ (R1)	= 8.80 ppm	and at δ (R2) = 7.77 - 7.7	'0 ppm		
Spectral de	etails of the de	euterated rea	ction mixtur	e:				
¹ H NMR (4	400 MHz, CE	δCl_3 $\delta = 8.80$) (d, $J = 4.8$	Hz, 2H, R1),	8.73 - 7.67	(m, 1H, R2),	8.49 - 8.42	
(m, 2H/D,	R1), $8.02 - 7$.97 (m, 2H/D	R2), 7.77 –	7.70 (m, 2H	R2), 7.52 –	7.39 (m, 3H,	R1 and 3H,	
R2), 7.25 -	- 7.20 (m, 1H	, R2), 7.17 (t	J = 4.8 Hz,	1H, R1).				
Entry	$I_{R1(t)} \\ N = 2H$	$\begin{split} I_{R1(0)} \\ N &= 2H \end{split}$	$%D_{R1}$	$\begin{split} I_{R2(t)} \\ N = 2H \end{split}$	$I_{R2(0)}$ $N = 2H$	%D _{R2}	κ	
1	1.70	2.00	15	1.75	2.01	13	1.17	
2	1.52	2.00	24	1.58	1.96	19	1.27	
3	1.56	2.00	22	1.66	2.00	17	1.33	
			Average	$\kappa = 1.26$				

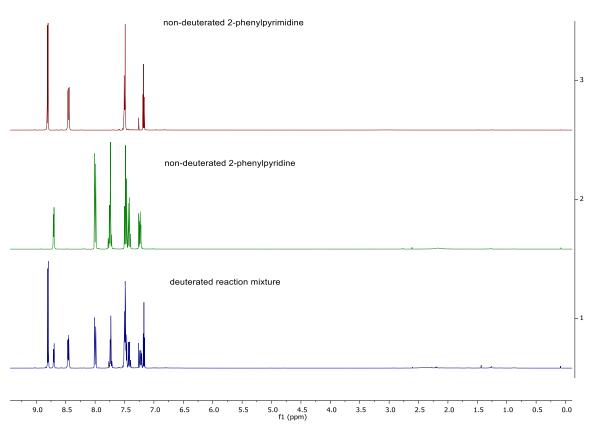
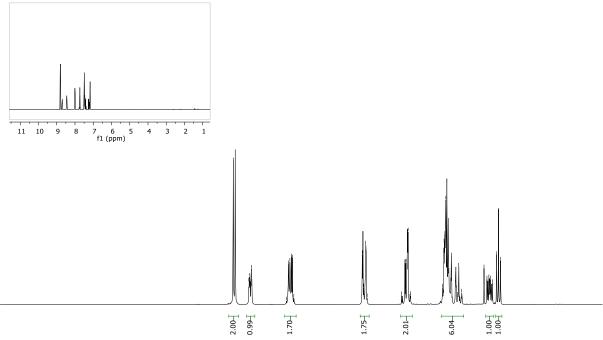


Figure S157. Stacked ¹H NMR (400 MHz, CDCl₃) of non-deuterated substrates and reaction mixture.

D328079 Person kpb19112 DT-86-1 @proton CDCl3 {C:\NMRdata} DJN 30



10.2 10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 6.7 6.6 f1 (ppm)

Figure S158. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyrimidine and 2-phenylpyridine (entry 1, Table S37).

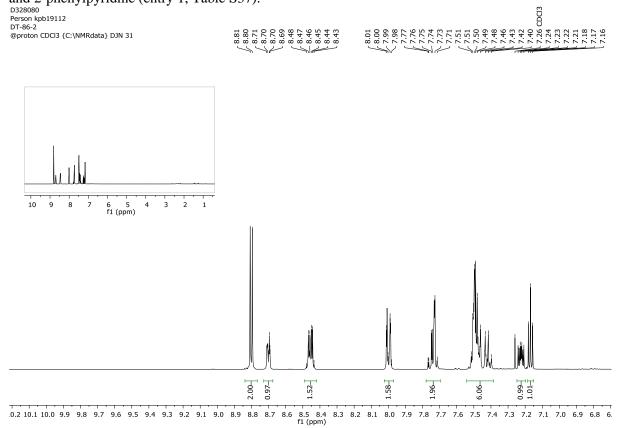


Figure S159. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyrimidine and 2-phenylpyridine (entry 2, Table S37).

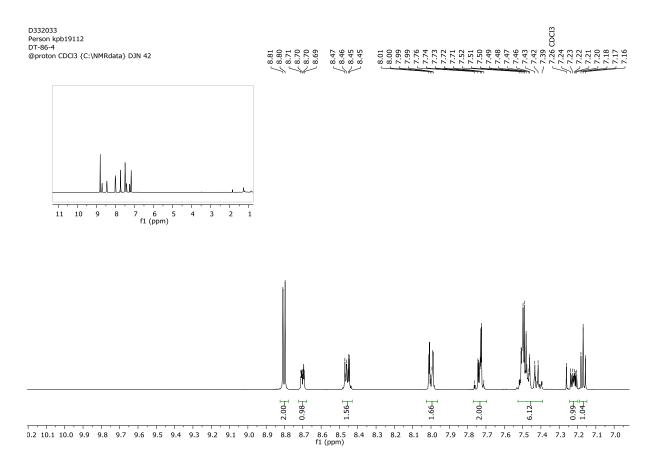


Figure S160. ¹H NMR (400 MHz, CDCl₃) of the competition experiment between 2-phenylpyrimidine and 2-phenylpyridine (entry 3, Table S37).

	Substrate R1		Su	Substrate R2			Catalyst		
						Ir-2			
		N H	F	Ph-N-N		[(COD)Ir(IN	/les)Cl]		
Mass	14.4 r	ng		14.4 mg		3.2 m	2		
	on expected at	· ·	• •			• •			
	d against inte				$t\delta\left(\mathbf{R2}\right)=8.4$	49 ppm			
-	etails of the do				4 1 2511-	111 D3) 7 (0 7 00		
	400 MHz, DN R1), 7.87 – 7		,						
	2), $7.36 - 7.29$					-			
	$I_{R1(t)}$	I _{R1(0)}		$\frac{I_{R2(t)}}{I_{R2(t)}}$	$\frac{III, III), 0.00}{I_{R2(0)}}$				
Entry	N = 2H	N = 2H	$%D_{R1}$	N = 2H	N = 1H	$%D_{R2}$	κ		
1	1.08	2.23	52	1.87	1.00	6	10.79		
2	1.04	2.11	51	1.87	1.00	6	10.53		
3	1.06	2.15	51	1.88	1.00	6	11.43		
			Average	$\kappa = 10.91$					
$I_{R1(t)} = 4.2$	23–1.00×2; ^b I	$R_{1(t)} = 4.11 - 1.0$	00×2 ; ^c I _{R1(t)}	$= 4.15 - 1.00 \times$:2;				
			1						
non-deut	erated 2-phenylimic	dazole							
						1			
non-deur	terated 1-phenylpyr	22010							
non-deu	terated 1-prienyipyi	azole							
deuterate	ed reaction mixture								
			a 1						
٨									

Table S38. Determination of the competition rate constant κ from the labelling experiment between 2-phenylimidazole and 1-phenylpyrazole.

Figure S161. Stacked ¹H NMR (400 MHz, DMSO-*d*₆) of non-deuterated substrates and reaction mixture.

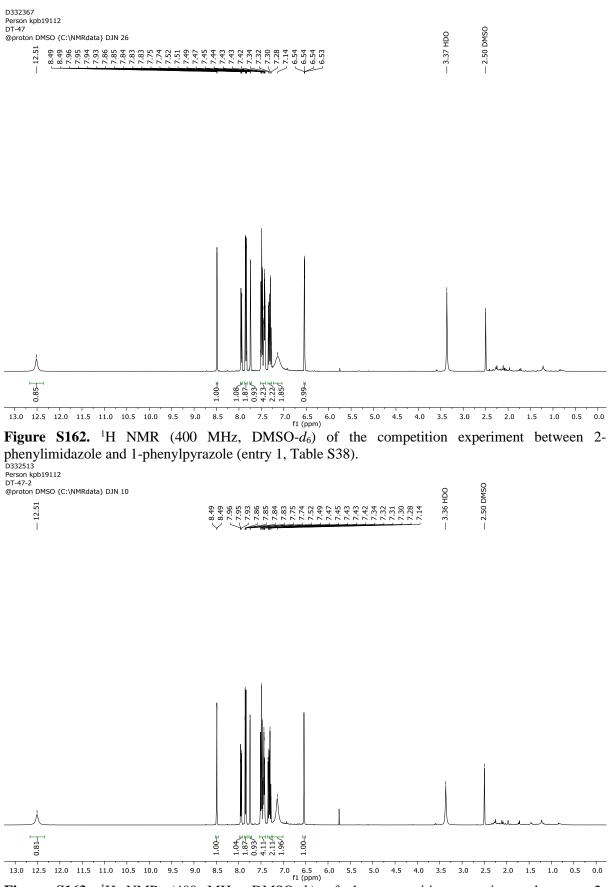
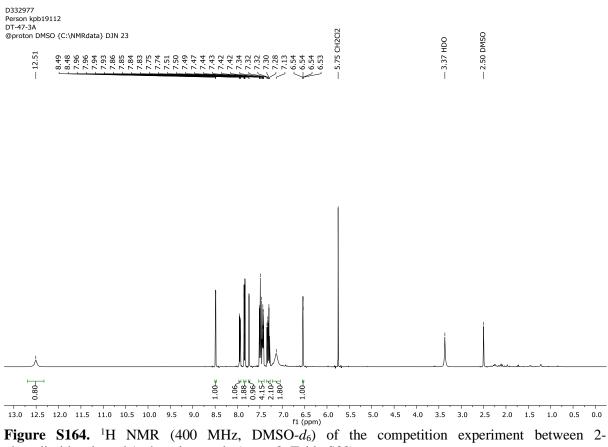


Figure S163. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between 2-phenylimidazole and 1-phenylpyrazole (entry 2, Table S38).



phenylimidazole and 1-phenylpyrazole (entry 3, Table S38).

	Substrat	e R1	Substrate R2			Catalyst Ir-2 [(COD)Ir(IMes)Cl]		
				Ph				
Mass	14.4 r	ng		15.5 mg		3.2 m	g	
Deuteration ex	spected at	$\delta (\mathbf{R1}) = 7.98$	- 7.92 pp	m and at δ (R 2	k) = 8.11 - 8.0	6 ppm		
	-			.40 ppm and at	$t\delta\left(\mathbf{R2}\right)=7.9$	0 – 7.84 ppm		
-	e e	euterated reac						
		,		H, R1), 8.70 –				
				-7.84 (m, 1H	, R2), 7.53 –	7.40 (m, 2H,	R1 and 3E	
R2), 7.37 – 7.3 Entry	$\frac{1}{I_{R1(t)}}$ $N = 2H$	$\frac{\mathbf{RI} \text{ and } \mathbf{IH}, \mathbf{I}}{\mathbf{I}_{\mathbf{R}1(0)}}$ $\mathbf{N} = 2\mathbf{H}$	(K2), 7.14	$\frac{\text{br, 2H, R1}}{I_{R2(t)}}$ $N = 2H$	$I_{R2(0)}$ $N = 1H$	$%D_{R2}$	к	
1	0.74 ^a	2.06 ^d	64	1.73	1.00	14	7.06	
2	0.80^{b}	2.06 ^e	61	1.78	1.00	11	8.12	
3	0.72 °	1.94 ^f	64	1.74	1.00	13	7.30	
			Avera	ge $\kappa = 7.49$				
$I_{R1(t)} = 1.74-1$.00; ^b I _{R1(t)}	= 1.80-1.00; °	$I_{R1(t)} = 1.7$	2-1.00;				
d I _{R1(0)} = 5.06-	1.00×3: ° I	$F_{R1(0)} = 5.06-1.0$	00×3: ^f I _{R10}	$_{(0)} = 4.99 - 1.00$	×3:			
(*)		(*)	,					
non-deuterated 2	2-phenylimidaz	ole						
							- 3	
non-deuterated	2-phenvlpvridi	ne			I			
					Ӊѻ		- 2	
		1	I.		-			
			<u>k</u> .					
			/*N/A		JIJ	·		
deuterated rea	ction mixture							
		1					- 1	

Table S39. Determination of the competition rate constant κ from the labelling experiment between 2-phenylimidazole and 2-phenylpyridine.

13.0 12.5 12.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 fl (ppm)

Figure S165. Stacked ¹H NMR (400 MHz, DMSO-*d*₆) of non-deuterated substrates and reaction mixture.

н₂о

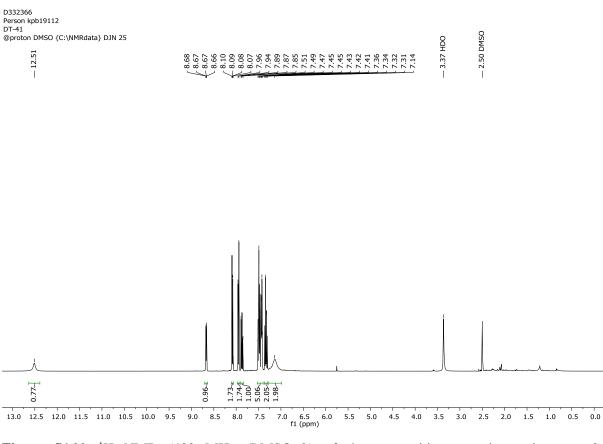


Figure S166. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between 2-phenylimidazole and 1-phenylpyridine (entry 1, Table S39).

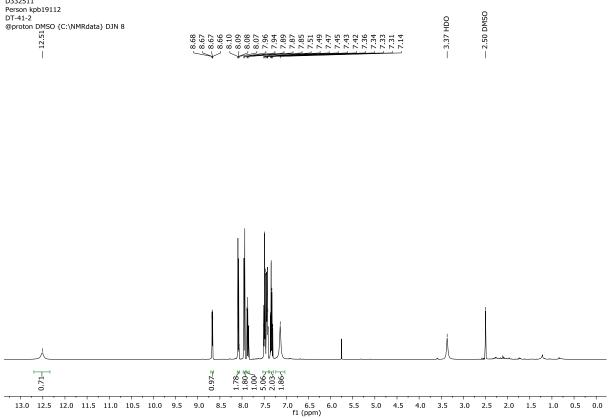
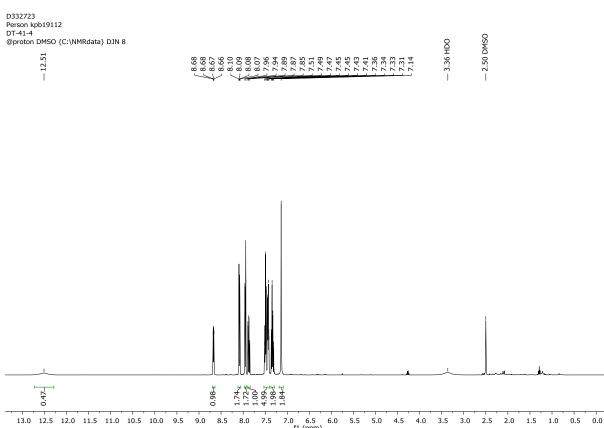


Figure S167. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between 2-phenylimidazole and 1-phenylpyridine (entry 2, Table S39).



^{13.0} ^{12.5} ^{12.0} ^{11.5} ^{11.0} ^{10.5} ^{10.0} ^{9.5} ^{9.0} ^{8.5} ^{8.0} ^{7.5} ^{7.0} ^{6.5} ^{6.0} ^{6.5} ^{5.0} ^{4.5} ^{4.0} ^{3.5} ^{3.0} ^{2.5} ^{2.0} ^{1.5} ^{1.0} ^{0.5} ^{0.0} ^{Figure S168. ¹H NMR (400 MHz, DMSO-*d*₆) of the competition experiment between 2-phenylimidazole and 1-phenylpyridine (entry 3, Table S39).}

Table S40. Determination of the competition rate constant κ from the labelling experiment between 2-phenylimidazoline and 2-phenylpyridine.

	Substrate	e R1	Su	bstrate R2		Catalyst			
			Ρ	h		Ir-2 [(COD)Ir(IMes)Cl]			
Mass	14.6 n	ng	15.5 mg			3.2 mg			
Determine Spectral de ¹ H NMR (4 (m, 1H, R 2	n expected at d against integ etails of the de 400 MHz, DM 2), 7.90 – 7.8	gral at δ (R1) euterated read ISO- d_6) $\delta = 8$ 1 (m, 2H/D,	= 3.61 ppm etion mixtur .70 - 8.64 (1	and at δ (R2 e: m, 1H, R2), 8) = 7.97 - 7.9 3.11 - 8.06 (m	93 ppm n, 2H/D, R2),			
Entry	$\frac{\text{H, R2}, 3.61 \text{ (}}{\text{I}_{\text{R1(t)}}}$ N = 2H	$\frac{III, 4H, KI}{I_{R1(0)}}$ N = 4H	$%D_{R1}$	$I_{R2(t)} \\ N = 2H$	$I_{R2(0)}$ $N = 1H$	%D _{R2}	ĸ		
1	0.86 ª	3.48	51	1.72	1.00	14	4.67		
2	1.17^{b}	3.37	31	1.81	1.00	10	3.65		
3	1.14 °	3.41	33	<u>1.84</u> е <i>к</i> = 4.38	1.00	8	4.83		
non-deutera	ted 2-phenylimidazo						-3		
non-deutera	ated 2-phenylpyridin	e							
					щo		-2		
deuterated r	reaction mixture						- 1		
		l.ln_M_				•			

12.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 f1 (ppm)

Figure S169. Stacked ¹H NMR (400 MHz, DMSO- d_6) of non-deuterated substrates and reaction mixture.

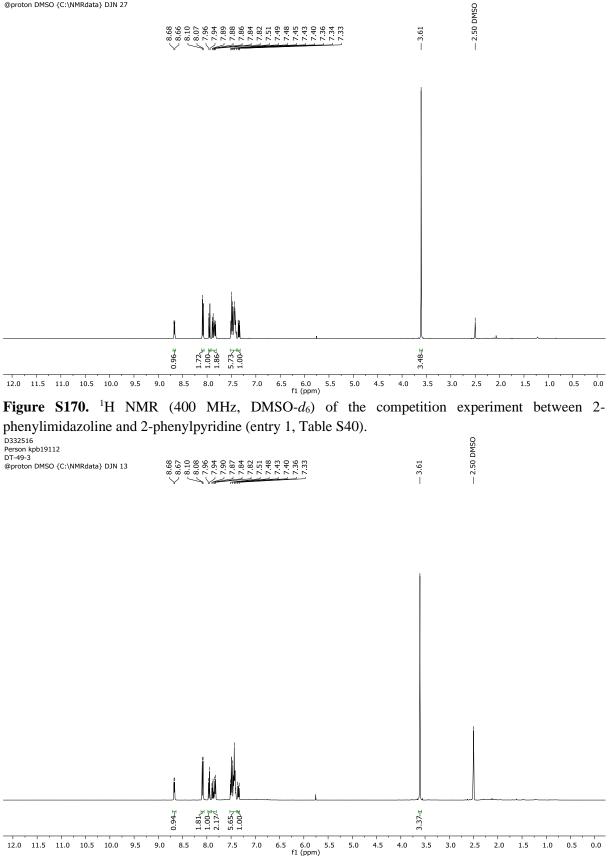


Figure S171. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between 2-phenylimidazoline and 2-phenylpyridine (entry 2, Table S40).

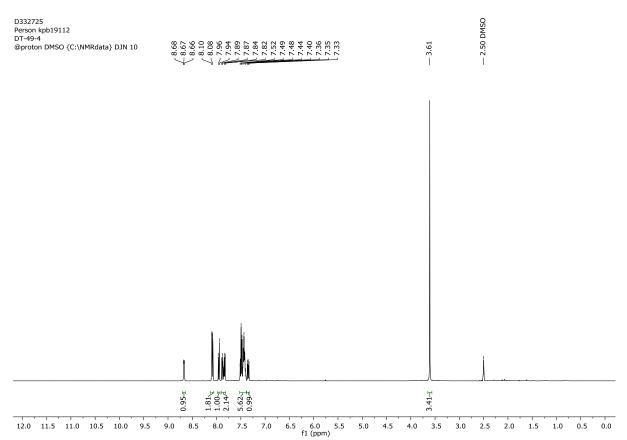


Figure S172. ¹H NMR (400 MHz, DMSO-*d*₆) of the competition experiment between 2-phenylimidazoline and 2-phenylpyridine (entry 3, Table S40).

Table S41. Determination of the competition rate constant κ from the labelling experiment between 2-
phenylimidazoline and 2-phenylimidazole.

	Substrate R1			Substrate R2			Catalyst	
				N N N N N N N N N N N N N N N N N N N	Ir-2 [(COD)Ir(IMes)Cl]			
Mass	14.6 mg		14.4 mg		3.2 mg			
Determine Spectral de	on expected at ad against integetails of the de 400 MHz, DM	gral at $\delta = 3.6$ euterated read	51 ppm for F <i>ection mixtur</i>	R1 and at $\delta = \delta$	7.36 – 7.29 pj	pm for R2	R1), 7.50	
7.38 (m, 3	H, R1 and 3H	, R2), 7.36 –	7.29 (m, 1H	l, R2),7.13 (b	r, 2H, R2), 3.	61 (s, 4H, R 1	l).	
Entry	$\begin{split} I_{R1(t)} \\ N &= 2H \end{split}$	$\begin{split} I_{R1(0)} \\ N = 4 H \end{split}$	$%D_{R1}$	$\begin{split} I_{R2(t)} \\ N = 2H \end{split}$	$\begin{split} I_{R2(0)} \\ N = 1 H \end{split}$	$%D_{R2}$	κ	
1	1.09	3.41	36	1.39	1.00	31	1.23	
2	1.09	3.47	37	1.26	1.00	37	1.01	
3	1.17	3.58	35	1.33	1.00	34	1.04	
			Averag	e $\kappa = 1.09$				
non-deuterat	ed 2-phenylimidazol	ine	L		,		-3	
non-deuterat	ed 2-phenylimidazol	ie 					- 2	

^{13.0} ^{12.5} ^{12.0} ^{11.5} ^{11.0} ^{10.5} ^{10.0} ^{9.5} ^{9.0} ^{8.5} ^{8.0} ^{7.5} ^{7.0} ^{6.5} ^{6.0} ^{6.5} ^{6.0} ^{5.5} ^{5.0} ^{4.5} ^{4.0} ^{3.5} ^{3.0} ^{2.5} ^{2.0} ^{1.5} ^{1.0} ^{0.5} ^{0.0} ^{14.5} ¹

ulu

- 1

deuterated reaction mixture

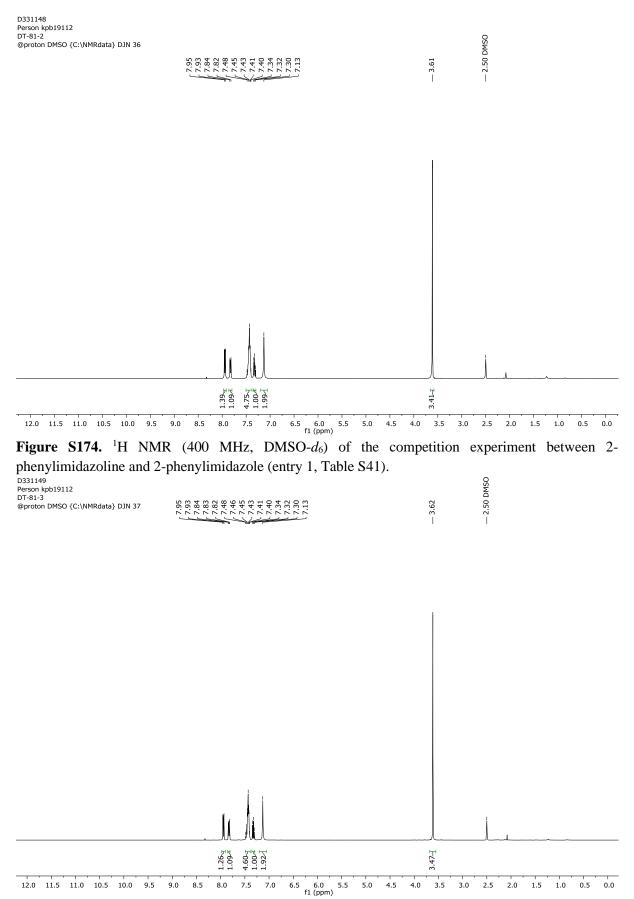


Figure S175. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between 2-phenylimidazoline and 2-phenylimidazole (entry 2, Table S41).

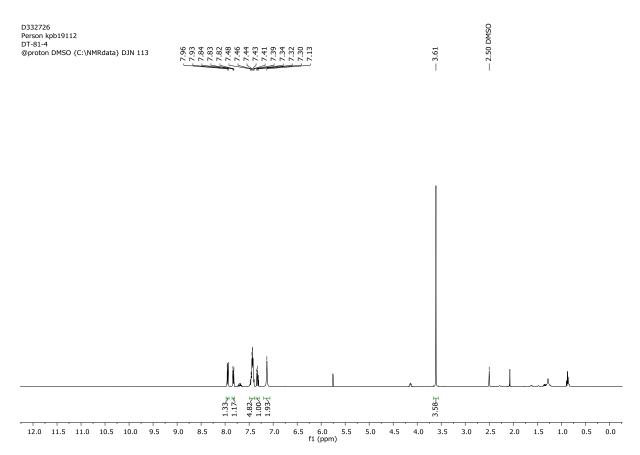


Figure S176. ¹H NMR (400 MHz, DMSO- d_6) of the competition experiment between 2-phenylimidazoline and 2-phenylimidazole (entry 3, Table S41).

3.5. Linear Regression Analysis.

Linear regression analysis of the experimentally obtained data set (κ^{exptl}) was performed to find the optimised k_{rel} values, which represent the relative rates of reaction of the different substrates, by minimising the sum of the squares of the errors $\Sigma\Delta^2$, where $\Delta^2 = \Sigma(\kappa^{\text{exptl}} - \kappa^{\text{calcd}})^2$ using Excel's Solver.

Calculated competition rate constants κ^{calcd} are expressed by equation (S-6)

$$\log \kappa = \log k_1 - \log k_2 \tag{S-6}$$

The pyridine directing group was assigned as the reference substrate with $k_{rel} = 1$. All k_{rel} were constrained to be >= 0.001.

Solving the resulting overdetermined set of linear equations (eq. S-6) by least squares minimization yielded the k_{rel} values listed in Tables S42-S43.

$H = DG_1$ $H = K_1$ K_1 $H/D = DG_1$ $H/D = H/D$ H/D $P1$	$\begin{array}{c} & H \\ H \\ H \\ R \\ R \\ H^{-1} \\ D_2 (limiting reagent) \\ H/D \\ DG_2 \\ H/D \\ D/H \\ P \\ P \\ \end{array}$
Substrates	k _{rel}
1-methyl-2-phenylimidaz	ole 10.13
2-phenyloxazoline	2.80

2.66

2.26

2.04

1.77 1.00

0.48

0.38

0.27 0.06

0.06

0.03

0.02

0.01

		Competition Rate Constants									
	Catalyst: [(COD)ir(IMes)PPh ₃]BArF ₂₄	competition rate constants						Standard	Standard		
			imental	Calculated fr		Err	ors	Averaged	Deviation	Error	
		K opt	log & ^{expt}	log & ^{caled}	K calcd	Δ (exptl - calcd)	$(\Delta (expt - calcd))^2$	κ^{expt}	(SD)	(SE)	$\kappa^{calcd} \kappa^{expt}$
DT-10	acetophenone vs benzophenone	0.99 1.01 1.03	-0.01 0.00 0.01	0.003	1.01	-0.02 0.00 0.02	0.00 0.00 0.00	1.01	0.02	0.01	1.00
DT-14	benzamide vs acetophenone	6.14 6.27 6.37	0.79 0.80 0.80	0.80	6.31	-0.17 -0.04 0.05	0.03 0.00 0.00	6.26	• 0.11	0.07	1.01
DT-16	acetophenone vs N, N-dimethylbenzamide	3.18 3.74 4.82	0.50 0.57 0.68	0.59	3.91	-0.73 -0.18 0.91	0.54 0.03 0.83	3.91	0.84	0.48	1.00
DT-52	nitrobenzene vs ethylbenzoate	5.58 6.76 6.71	0.75 0.83 0.83	0.76	5.75	-0.17 1.01 0.95	0.03 1.02 0.92	6.35	• 0.67	0.38	0.91
DT-12	acetophenone vs nitrobenzene	3.52 3.19 3.46	0.55 0.50 0.54	0.27	1.86	1.65 1.33 1.60	2.73 1.76 2.56	3.39	• 0.18	0.10	0.55
DT-51	acetophenone vs ethylbenzoate	9.63 10.58	0.98	1.03	10.71	-1.08 -0.13	1.17 0.02	10.38	0.68	0.39	1.03
DT-104	benzamide vs acetanilide	10.95 1.35 1.62	1.04 0.13 0.21	-0.11	0.78	0.25 0.57 0.85	0.06 0.33 0.72	1.55	• 0.18	0.10	0.50
DT-102	acetanilide vs nitrobenzene	1.68 15.01 15.63	0.22 1.18 1.19	1.18	15.14	0.90 -0.13 0.49	0.81 0.02 0.24	15.18	• 0.40	0.23	1.00
DT-6	2-phenylpyndine vs acetophenone	14.89 16.92 16.48	1.17 1.23 1.22	1.22	16.78	-0.25 0.14 -0.30	0.06 0.02 0.09	16.76	• 0.24	0.14	1.00
DT-103	2-phenylpyrimidine vs benzamide	16.88 8.90 4.97	1.23 0.95 0.70	0.85	7.07	0.10 1.82 -2.10	0.01 3.32 4.41	7.12	1.99	1.15	0.99
DT-11	1-phenylpyrazole vs 2-phenylpyndine	7.48 1.52 1.82	0.87 0.18 0.26	0.25	1.77	0.41 -0.25 0.05	0.17 0.06 0.00	1.71	• 0.17	0.10	1.03
DT-8	2-phenyloxazoline vs 2-phenylpyndine	1.79 2.25 2.77	0.25 0.35 0.44	0.45	2.80	0.02 -0.55 -0.02 0.88	0.00 0.30 0.00	2.90	0.72	0.42	0.96
DT-15	2-phenyithiazole vs 1-phenyipyrazole	3.68 1.01 1.04 1.14	0.57 0.01 0.02 0.06	0.05	1.15	-0.14 -0.11 -0.01	0.77 0.02 0.01 0.00	1.07	• 0.07	0.04	1.08
DT-43	2-phenyithiazoline vs 2-phenyithiazole	1.04	0.05	0.04	1.11	-0.01 -0.07 -0.09 -0.08	0.00 0.01 0.01 0.01	1.03	• 0.01	0.01	1.08
DT-67	2-phenyloxazoline vs 2-phenylthiazoline	1.03 1.01 1.01 1.00	0.01 0.00 0.00	0.09	1.24	-0.08 -0.23 -0.23 -0.23	0.05 0.05 0.05	1.01	• 0.00	0.00	1.23
DT-44	2-phenylthiazole vs 2-phenylbenzothiazole	7.26 7.71 7.48	0.86 0.89 0.87	0.87	7.48	-0.25 -0.22 0.23 0.00	0.05 0.05 0.00	7.48	0.22	0.13	1.00
DT-70	2-phenylpyridine vs 2-phenylbenzothiazole	3.45 4.05 3.51	0.54 0.61 0.55	0.57	3.68	-0.23 0.38 -0.16	0.05 0.14 0.03	3.67	0.33	0.19	1.00
DT-77	1-methyl-2-phenylimidazole vs 2-phenylthiazoline	4.43 5.97	0.65 0.78	0.65	4.49	-0.05 1.49	0.00 2.21	4.53	1.40	0.81	0.99
DT-63	1-methyl-2-phenylimidazole vs 2-phenylpyridine	3.18 10.95 8.69	0.50 1.04 0.94	1.01	10.13	-1.30 0.82 -1.43	1.70 0.67 2.05	10.11	1.23	0.71	1.00
DT-83	2-phenylpyridine vs 2-phenylpyrimidine	10.68 1.28 1.11	1.03 0.11 0.05	-0.42	0.38	0.55 0.90 0.73	0.31 0.81 0.54	1.19	0.08	0.05	0.32
		1.18	0.07			0.81	0.65				

Table S42. Linear regression analysis for k_{rel} determination for catalyst Ir-1.

All k rel	constrained	>= 0.001
-----------	-------------	----------

N,N-dimethylbenzamide

2-phenylpyrimidine

2-phenyithiazoline 2-phenyithiazole

1-phenylpyrazole

2-phenylpyndine acetanilide

2-phenylbenzothiazole

benzamide

acetophenone benzophenone

nitrobenzene

ethylbenzoate

Sum of squares of errors: 32,51

H	H
DG ₁	H
H	K2
R1	K2
	-2 (limiting reagent)
H/D	H/D
DG ₁	DG ₂
H/D	D/H
P1	P2
Substrates	k _{sel}
Substrates	k _{rel}
2-phenylimidazole	7.11
2-phenylimidazole	7.11 4.45
2-phenylimidazole	7.11
2-phenylimidazoline	4.45
2-phenylimidazole	7.11
2-phenylimidazoline	4.45
1-methyl-2-phenylimidazol	e 3.08
2-phenylimidazole	7.11
2-phenylimidazoline	4.45
1-methyl-2-phenylimidazol	e 3.08
2-phenylpyrimidine	1.26
2-phenylimidazole	7.11
2-phenylimidazoline	4.45
1-methyl-2-phenylimidazol	= 3.08
2-phenylpyrimidine	1.26
2-phenylthiazoline	1.02
2-phenylimidazole	7.11
2-phenylimidazoline	4.45
1-methyl-2-phenylimidazol	e 3.08
2-phenylpyrimidine	1.26
2-phenylthiazoline	1.02
2-phenylthiazoline	1.00
2-phenylimidazole	7.11
2-phenylimidazoline	4.45
1-methyl-2-phenylimidazol	e 3.08
2-phenylpyrimidine	1.26
2-phenylthiazoline	1.02
2-phenylpyridine	1.00
1-phenylpyrazole	0.64

acetophenone

N,N-dimethylbenzamide

All k_{ml} constrained >= 0.001

methylphenylsulfone

benzamide

0.02

0.01

0.002

0.001

Catalyst: [(COD)Ir(IMes)Cl]		Competitio	on Constants		Errors			Standard	Standard	d	
		rimental	Calculated f	rom Solver			Averaged	Deviation	Error		
	€ exptl	$\log \kappa^{expt}$	$\log \kappa^{calcd}$	κ^{calcd}	Δ (expti - calcd)	$\left(\Delta\left(exptl-calcd\right)\right)^2$	κ^{exptl}	(SD)	(SE)	$\kappa^{\text{calcd}/}\kappa^{\text{exptl}}$	
acetophenone vs benzamide	1.42 1.50 1.66	0.15 0.18 0.22	0.38	2.37	-0.95 -0.87 -0.71	0.90 0.75 0.51	1.53	0.12	0.07	1.55	
benzenesulfonamide vs acetophenone	1.07 1.39 1.27	0.03 0.14 0.10	0.34	2.17	-1.09 -0.78 -0.89	1.19 0.61 0.80	1.24	0.16	0.09	1.74	
acetophenone vs N,N-dimethylbenzamide	9.87 9.24 10.09	0.99 0.97 1.00	0.99	9.73	0.13 -0.49 0.35	0.02 0.24 0.13	9.73	0.44	0.25	1.00	
benzenesulfonamide vs benzamide	5.44 6.30 4.84	0.74 0.80 0.68	0.71	5.14	0.30 1.16 -0.30	0.09 1.35 0.09	5.53	0.74	0.42	0.93	
1-phenylpyrazole vs acetophenone	28.94 28.05 32.17	1.46 1.45 1.51	1.47	29.72	-0.78 -1.67 2.45	0.61 2.79 6.00		2.17	1.25	1.00	
benzenesulfonamide vs methylphenylsulfone	32.07 40.55 43.56	1.51 1.61 1.64	1.59	38.73	-6.66 1.82 4.83	44.34 3.33 23.37	56.75	5.96	3.44	1.00	
1-phenylpyrazole vs 2-phenylpyridine	1.44 1.32 1.37	0.16 0.12 0.14	-0.19	0.64	0.80 0.68 0.74	0.64 0.47 0.54	1.38	0.06	0.03	0.46	
2-phenyloxazoline vs 2-phenylpyridine	1.46 1.33 1.43	0.16 0.12 0.16	-0.41	0.39	1.07 0.95 1.04	1.15 0.89 1.09	1.41	0.07	0.04	0.27	
1-phenylpyrazole vs 2-phenylthiazole	1.58 1.79 1.49	0.20 0.25 0.17	0.43	2.70	-1.12 -0.90 -1.21	1.25 0.81 1.46		0.16	0.09	1.66	
2-Phenylthiazoline vs 2-phenyloxazoline	1.78 1.82 1.83	0.25 0.26 0.26	0.42	2.64	-0.87 -0.82 -0.81	0.75 0.67 0.65	1.81	0.03	0.02	1.46	
1-methyl-2-phenylimidazole vs 2-phenylpyridine	2.26 1.84 1.93	0.35 0.27 0.28	0.49	3.08	-0.82 -1.24 -1.15	0.67 1.53 1.33	2.01	0.22	0.13	1.53	
1-methyl-2-phenylimidazole vs 2-phenyloxazoline	8.55 7.61 8.74	0.93 0.88 0.94	0.90	7.98	0.57 -0.36 0.77	0.32 0.13 0.59	0.50	0.60	0.35	0.96	
1-methyl-2-phenylimidazole vs 2-phenylthiazole	12.98 14.29 12.39	1.11 1.15 1.09	1.11	13.00	-0.01 1.29 -0.61	0.00 1.66 0.37	13.22	0.97	0.56	0.98	
1-methyl-2-phenylimidazole vs 2-phenylthiazoline	2.78 1.82 2.28	0.44 0.26 0.36	0.48	3.02	-0.24 -1.20 -0.74	0.06 1.43 0.55	2.29	0.48	0.28	1.32	
2-Phenylpyrimidine vs 2-phenylpyridine	1.17 1.27 1.33	0.07 0.10 0.12	0.10	1.26	-0.09 0.01 0.07	0.01 0.00 0.01	1.26	0.08	0.05	1.00	
2-phenylimidazoline vs 2-phenylimidazole	1.23 1.01 1.04	0.09 0.00 0.02	-0.20	0.63	0.60 0.38 0.42	0.36 0.14 0.17	1.09	0.12	0.07	0.57	
2-phenylimidazole vs 2-phenylpyridine	7.06 8.12 7.30	0.85 0.91 0.86	0.85	7.11	-0.05 1.01 0.19	0.00 1.02 0.04	7.45	0.55	0.32	0.95	
2-phenylimidazole vs 1-phenylpyrazole	10.79 10.53 11.43	1.03 1.02 1.06	1.05	11.13	-0.35 -0.61 0.30	0.12 0.37 0.09	10.91	0.46	0.27	1.02	
2-phenylimidazoline vs 2-phenylpyridine	4.67 3.65 4.83	0.67 0.56 0.68	0.65	4.45	0.22 -0.80 0.38	0.05 0.63 0.14	4.38	0.64	0.37	1.01	

Table S43. Linear regression analysis for k_{rel} determination for catalyst Ir-2.

Sum of squares of	
errors:	109.301

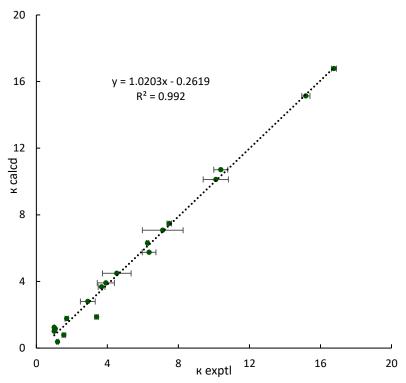


Figure S177. Plot of experimental *versus* calculated (from linear regression) competition constants κ for catalyst **Ir-1**. The error bars correspond to standard error on the mean.

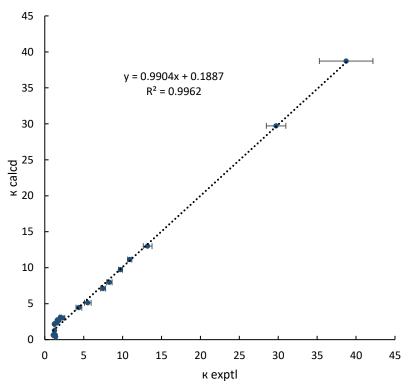
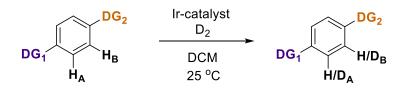


Figure S178. Plot of experimental *versus* calculated (from linear regression) competition constants κ for catalyst **Ir-2**. The error bars correspond to standard error on the mean.

4. Intramolecular Competition Experiments

4.1. General Information



General Procedure (GP2)

The substrate (0.10 mmol) and the catalyst (0.005 mmol) of choice were added to one J. Young Schlenk flask under air. The solvent, DCM (6 mL), was added in such a way to rinse the inner walls of the flask. The flask was then sealed (with gas inlet left open) under air before being cooled in a dry ice–acetone bath. The flask was evacuated and flushed with deuterium three times *via* a balloon. The gas inlet was then closed with fast thread tap, creating a sealed atmosphere of deuterium. After sealing the flask was placed in the thermostated water bath at 25 °C and the reaction timer was started. The reaction mixture was stirred for 1 h (for catalyst **Ir-1**) or 16 h (for catalyst **Ir-2**) before removing excess deuterium and replacing it with air. The reaction mixture was quenched with few drops of MeCN and transferred to a single necked flask together with washings (DCM) before removing the solvent under reduced pressure. The residue was dissolved in a small portion of 1:1 mixture of petroleum ether with diethyl ether (or ethyl acetate) and passed through a short plug of silica, eluting with a 1:1 mixture of petroleum ether with ethyl acetate where necessary, depending on the substrates used. The solvent was evaporated under reduced pressure and the residue was analysed directly by ¹H NMR spectroscopy.

Determination of Competition Rate Constants

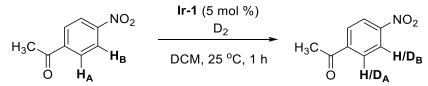
The level of deuterium incorporation (%D) in the substrates was determined by ¹H NMR. The integrals were calibrated against a peak corresponding to a position not expected to be labelled. Equations S-7 and S-8 were used to calculate the extent of labelling and competition rate constant κ' :

$$\%D = 100 - \left(\frac{\text{residual integral}}{\text{number of labelling sites}} \times 100\%\right)$$
(S-7)

$$\kappa' = \frac{\% D_{A}}{\% D_{B}} = \frac{2 - \text{residual integral } H_{A}}{2 - \text{residual integral } H_{B}}$$
(S-8)

4.1. Competition Experiments with [(COD)Ir(IMes)PPh3]BArF24 (Ir-1)

Labelling of *p*-nitroacetophenone



According to GP2: 16.5 mg of substrate and 8.7 mg of catalyst

Spectral details of the reaction mixture:

¹H NMR (400 MHz, CDCl₃) δ = 8.33 – 8.30 (m, 2H, H/D_B), 8.13 – 8.09 (m, 2H, H/D_A), 2.68 (s, 3H, CH₃)

Deuteration expected at δ (H_A) = 8.13 – 8.09 ppm and δ (H_B) = 8.33 – 8.30 ppm.

Determined against integral at $\delta = 2.68$ ppm.

Table S44. Determination of the competition rate constant κ' from the labelling of *p*-nitroacetophenone.

Average	0.97	52 51	1.22	39	1.32 1.32
3	0.97	52	1.22	39	1.32
2	1.12	44	1.32	34	1.29
1	0.84	58	1.13	44	1.33
Entry	(H/D_A)	$\% D_A$	(H/D_B)	$\% D_B$	κ΄
	residual integral		residual integral		



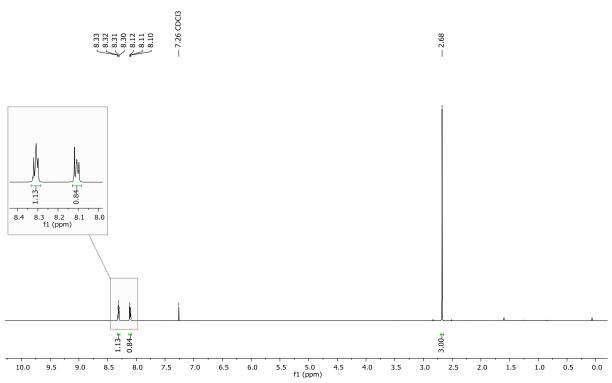
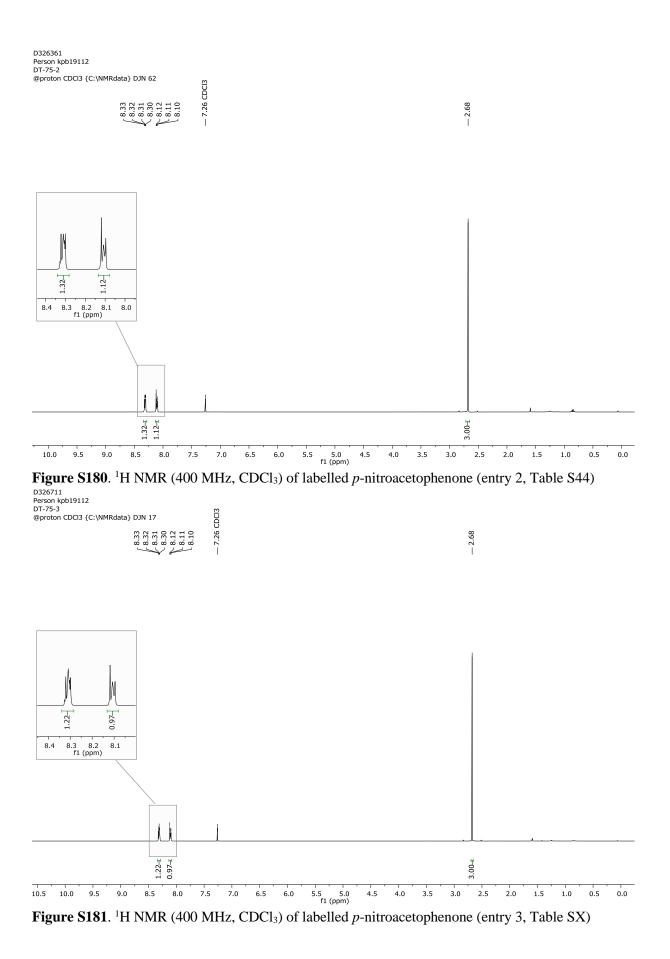
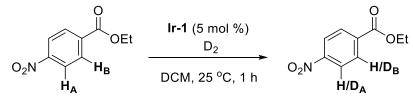


Figure S179. ¹H NMR (400 MHz, CDCl₃) of labelled *p*-nitroacetophenone (entry 1, Table S44)



S153

Labelling of ethyl 4-nitrobenzoate



According to GP2:19.5 mg of substrate and 8.7 mg of catalyst *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, CDCl₃) δ 8.30 – 8.26 (m, 2H, H/D_A), 8.23 – 8.18 (m, 2H, H/D_B), 4.43 (q, *J* = 7.1 Hz, 2H, CH₂), 1.42 (t, *J* = 7.1 Hz, 3H, CH₃).

Deuteration expected at δ (H_A) = 8.30 – 8.26 ppm and δ (H_B) = 8.23 – 8.18 ppm. Determined against integral at δ = 4.43 ppm.

Table S45. Determination of the competition rate constant κ' from the labelling of ethyl 4-nitrobenzoate.

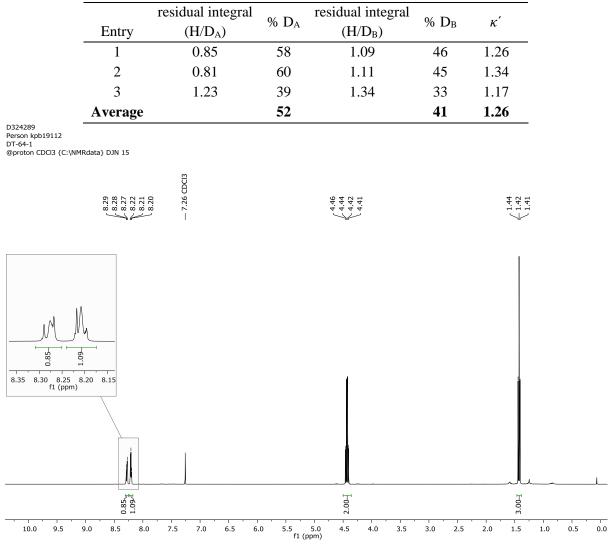


Figure S182. ¹H NMR (400 MHz, CDCl₃) of labelled ethyl 4-nitrobenzoate (entry 1, Table S45)



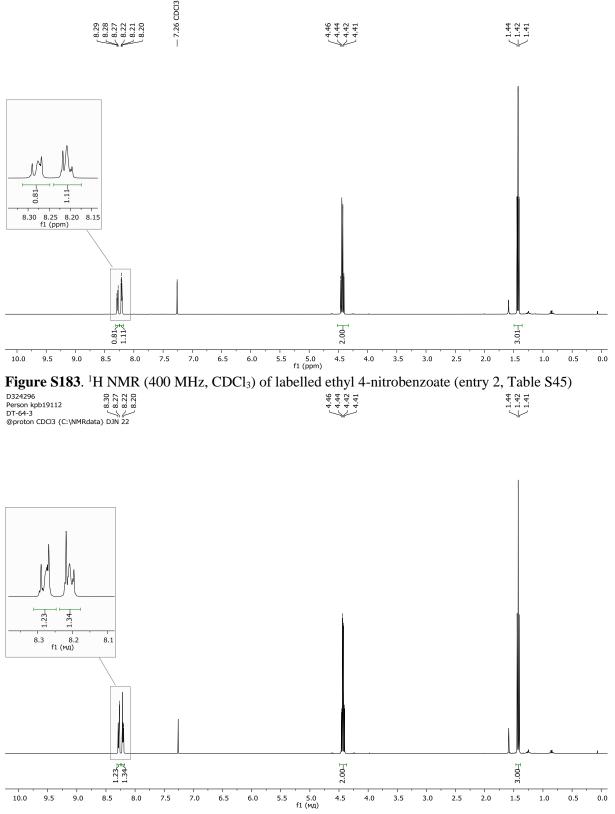
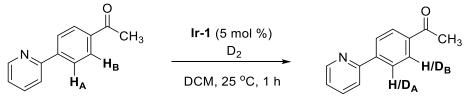


Figure S184. ¹H NMR (400 MHz, CDCl₃) of labelled ethyl 4-nitrobenzoate (entry 3, Table S45)

Labelling of 4-acetyl-1-(pyridin-2-yl)benzene



According to GP2: 19.7 mg of substrate and 8.7 mg of catalyst *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, CDCl₃) δ = 8.76 – 8.68 (m, 1H, Ar), 8.12 – 8.08 (m, 2H, H_A), 8.07 – 8.03 (m, 2H, H_B), 7.80 – 7.75 (m, 2H, Ar), 7.31 – 7.26 (m, 1H, Ar), 2.65 (s, 3H, CH₃).

Deuteration expected at δ (H_A) = 8.12 – 8.08 ppm and δ (H_B) = 8.07 – 8.03 ppm.

Determined against integral at $\delta = 2.65$ ppm.

Note: Small signals, corresponding to the **Ir-1** catalyst, were present in the ¹H NMR spectra, however there is no overlap with the substrate signals.

Table S46. Determination of the competition rate constant κ' from the labelling of 4-acetyl-1-(pyridin-2-yl)benzene.

Entry	residual integral (H/D _A)	% D _A	residual integral (H/D _B)	% D _B	ĸ
1	1.51	25	1.97	1.5	16.33
2	1.66	17	1.98	1.0	17.00
3	1.55	23	1.97	1.5	15.00
Average		21		1.3	16.11

D324682 Person kpb19112 DT-68-1 @proton CDCl3 {C:\NMRdata} DJN 14

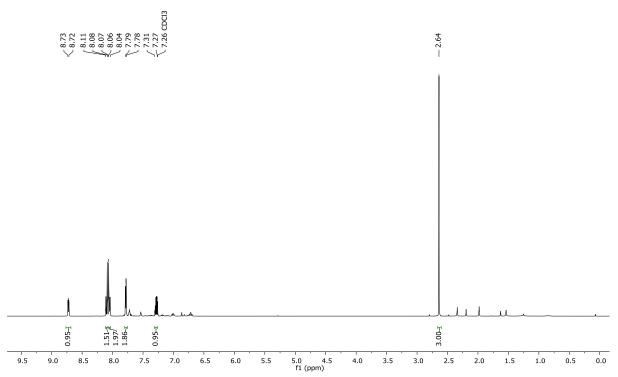


Figure S185. ¹H NMR (400 MHz, CDCl₃) of labelled 4-acetyl-1-(pyridin-2-yl)benzene (entry 1, Table S46)

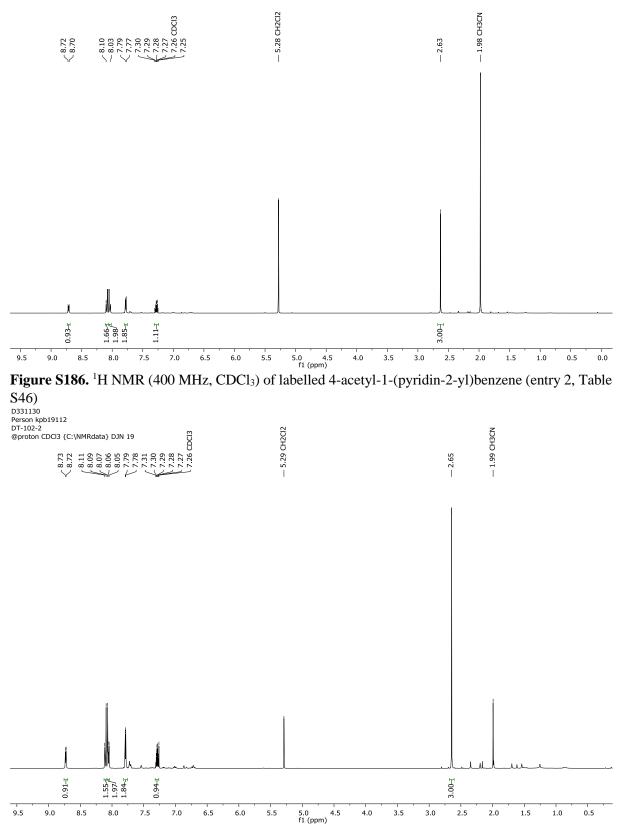
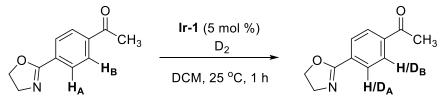


Figure S187. ¹H NMR (400 MHz, CDCl₃) of labelled 4-acetyl-1-(pyridin-2-yl)benzene (entry 3, Table S46)

Labelling of 2-(4-acetyl)phenyloxazoline



According to GP2: 18.9 mg of substrate and 8.7 mg of catalyst *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, CDCl₃) δ = 8.06 – 8.01 (m, 2H, H_A), 8.01 – 7.96 (m, 2H, H_B), 4.47 (t, *J* = 9.6 Hz, 3H, CH₂), 4.10 (t, *J* = 9.6 Hz, 2H, CH₂), 2.62 (s, 3H, CH₃).

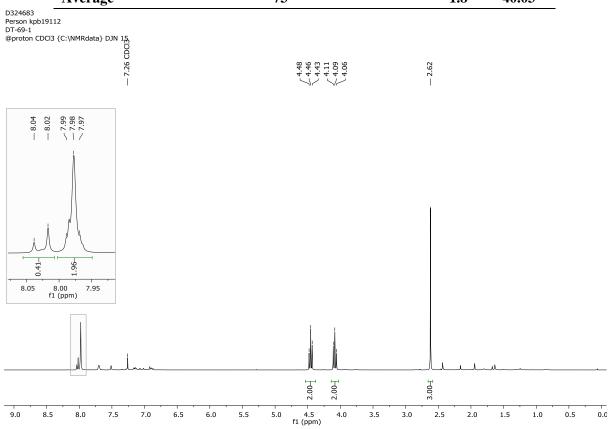
Deuteration expected at δ (H_A) = 8.06 - 8.01 ppm and δ (H_B) = 8.01 - 7.96 ppm.

Determined against integral at $\delta = 4.47$ ppm.

Note: Small signals, corresponding to the **Ir-1** catalyst, were present in the ¹H NMR spectra, however they do not overlap with the substrate signals.

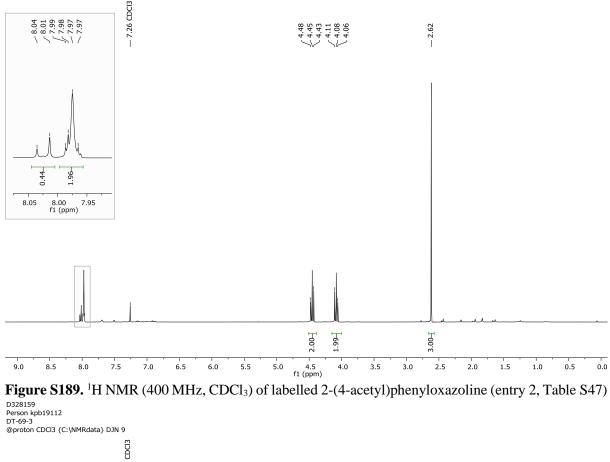
Table S47. Determination of the competition rate constant κ' from the labelling of 2-(4-acetyl)phenyloxazoline.

	residual integral		residual integral		
Entry	(H/D_A)	$\% D_A$	(H/D_B)	$\% D_B$	ĸ
1	0.41	80	1.96	2.0	39.75
2	0.44	78	1.96	2.0	39.00
3	0.76	62	1.97	1.5	41.33
Average		73		1.8	40.03









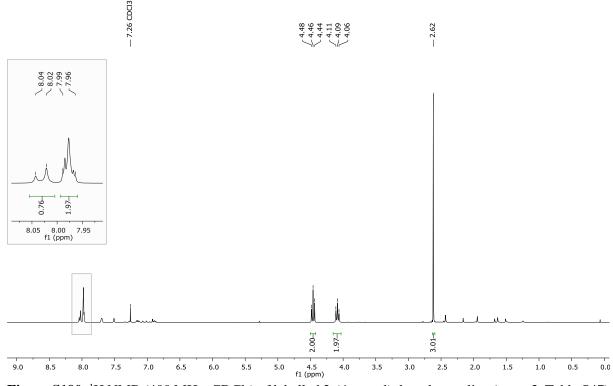
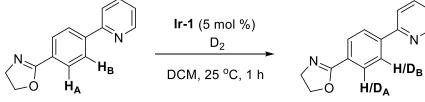


Figure S190. ¹H NMR (400 MHz, CDCl₃) of labelled 2-(4-acetyl)phenyloxazoline (entry 3, Table S47)

Labelling of 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole



According to GP2: 22.4 mg of substrate and 8.7 mg of catalyst *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, C_6D_6) $\delta = 8.56 - 8.51$ (m, 1H, Ar-H), 8.36 - 8.32 (m, 1H, H_A), 8.17 - 8.11 (m, 2H, H_B), 7.25 - 7.21 (m, 1H, Ar-H), 7.11 - 7.04 (m, 1H, Ar-H), 6.66 - 6.61 (m, 1H, Ar-H), 3.77 - 3.69 (m, 2H, CH₂), 3.67 - 3.58 (m, 2H, CH₂).

Deuteration expected at δ (H_A) = 8.36 - 8.32 ppm and δ (H_B) = 8.17 - 8.11 ppm. Determined against integral at δ = 6.66 - 6.61 ppm.

Note: Small signals, corresponding to the **Ir-1** catalyst, were present in the ¹H NMR spectra, however they do not overlap with the substrate signals.

Table S48. Determination of the competition rate constant κ' from the labelling of 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole.

	residual integral		residual integra	1	
Entry	(H/D_A)	$\% D_A$	(H/D_B)	$\% D_B$	ĸ
1	0.80	60	1.45	28	2.18
2	0.42	79	1.57	22	3.67
3	0.48	76	1.62	19	4.00
Average		72		23	3.29

D328680

Person kpb19112 DT-88-1

DT-88-1 @proton C6D6 {C:\NMRdata} DJN 4

	ŏ
$ \begin{array}{c} 8.54 \\ 8.53 \\ 8.35 \\ 8.34 \\ 8.34 \\ 8.33 \\ 8.15 \\ 8$	7.24 7.09 7.07 7.07 6.65 6.65 6.65

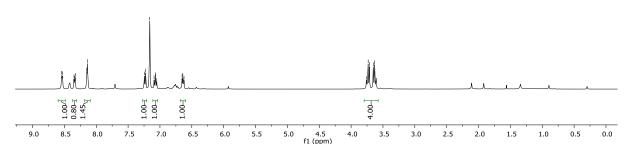


Figure S191. ¹H NMR (400 MHz, C₆D₆) of labelled 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole (entry 1, Table S48)

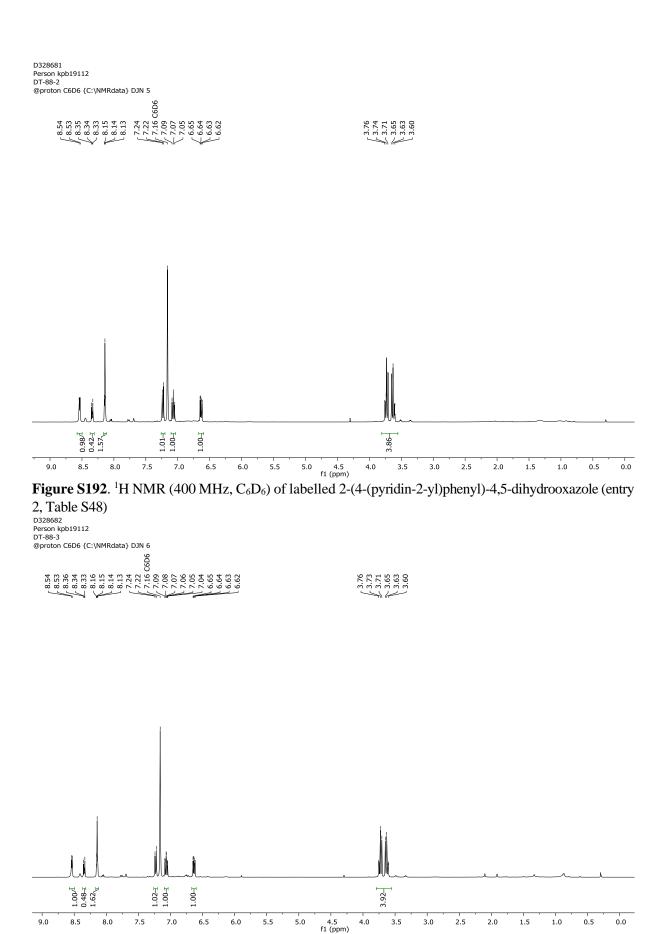
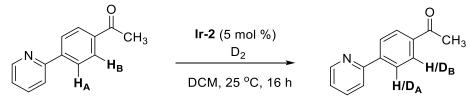


Figure S193. ¹H NMR (400 MHz, C₆D₆) of labelled 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole (entry 3, Table S48)

4.2. Competition Experiments with (COD)Ir(IMes)Cl (Ir-2)

Labelling of 4-acetyl-1-(pyridin-2-yl)benzene



According to GP2: 19.7 mg of substrate and 3.2 mg of catalyst *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, CDCl₃) $\delta = 8.76 - 8.72$ (m, 1H), 8.13 - 8.09 (m, 2H, H_A), 8.09 - 8.04 (m, 2H, H_B), 7.82 - 7.78 (m, 2H, Ar), 7.33 - 7.27 (m, 1H, Ar), 2.65 (s, 3H, CH₃).

Deuteration expected at δ (H_A) = 8.13 – 8.09 ppm and δ (H_B) = 8.09 – 8.04 ppm. Determined against integral at δ = 2.65 ppm.

Table S49. Determination of the competition rate constant κ' from the labelling of 4-acetyl-1-(pyridin-2-yl)benzene.

Entry	residual integral	% D _A	residual integral	% D _B	ĸ
	(H/D_A)		(H/D_B)		
1	1.32	34	1.96	2	17.00
2	1.52	24	1.97	2	16.00
3	1.21	40	1.95	3	15.80
Average		33		2	16.27



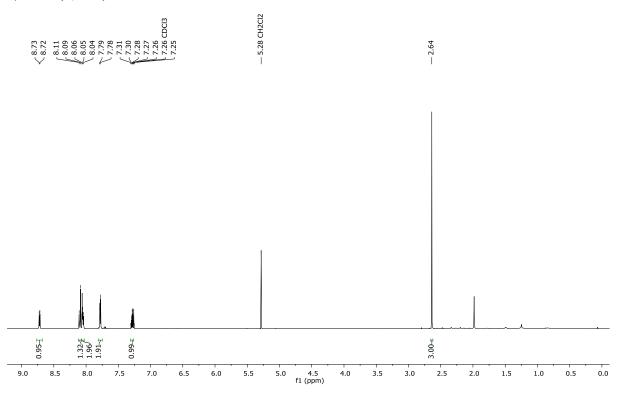
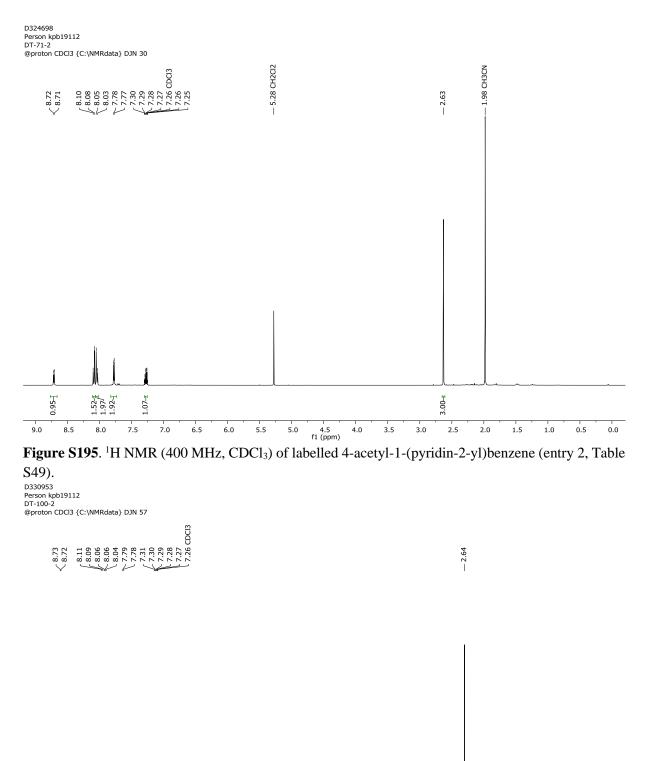


Figure S194. ¹H NMR (400 MHz, CDCl₃) of labelled 4-acetyl-1-(pyridin-2-yl)benzene (entry 1, Table S49).



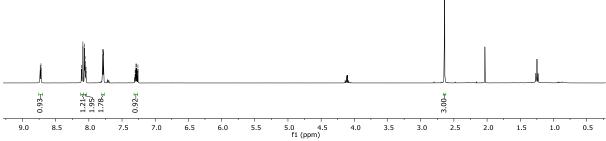
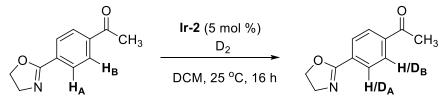


Figure S196. ¹H NMR (400 MHz, CDCl₃) of labelled 4-acetyl-1-(pyridin-2-yl)benzene (entry 3, Table S49).

Labelling of 2-(4-acetyl)phenyloxazoline



According to GP2: 18.9 mg of substrate and 3.2 mg of catalyst Spectral details of the reaction mixture: ¹H NMR (400 MHz, CDCl₃) $\delta = 8.06 - 8.01$ (m, 2H, H_A), 8.01 - 7.96 (m, 2H, H_B), 4.47 (t, J = 9.6 Hz, 3H, CH₂), 4.10 (t, J = 9.6 Hz, 2H, CH₂), 2.62 (s, 3H, CH₃). Deuteration expected at δ (H_A) = 8.06 - 8.01 ppm and δ (H_B) = 8.01 - 7.96 ppm. Determined against integral at $\delta = 4.47$ ppm.

Table S50. Determination of the competition rate constant κ' from the labelling of 2-(4-acetyl)phenyloxazoline.

Entry	residual integral	% D _A	residual integral	% D _B	ĸ
-	(H/D_A)		(H/D_B)		
1	1.41	30	1.93	4	8.43
2	1.27	37	1.93	4	10.43
3	0.48	76	1.88	6	12.67
Average		47		4	10.51

D324699 Person kpb19112 DT-72-1

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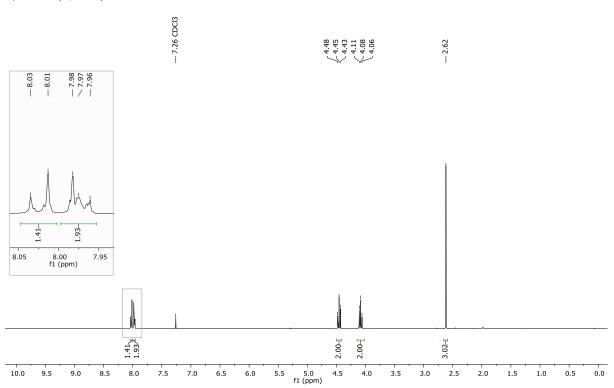
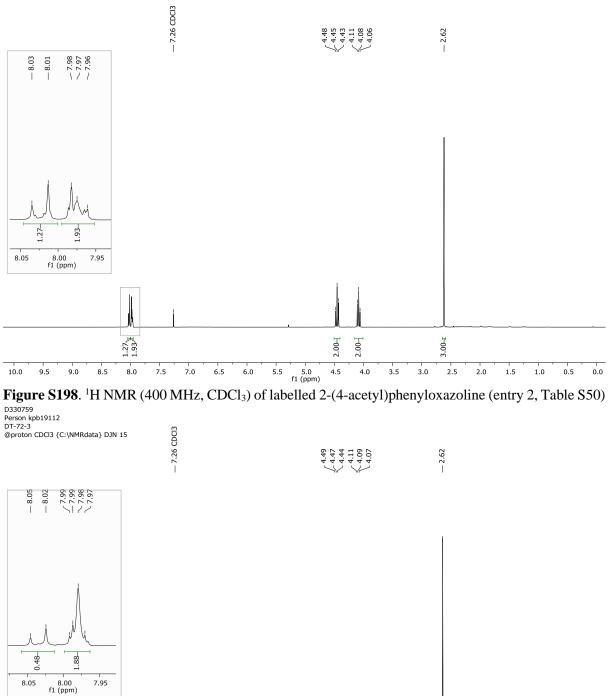


Figure S197. ¹H NMR (400 MHz, CDCl₃) of labelled 2-(4-acetyl)phenyloxazoline (entry 1, Table S50)





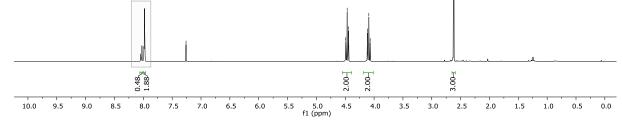
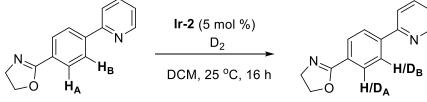


Figure S199. ¹H NMR (400 MHz, CDCl₃) of labelled 2-(4-acetyl)phenyloxazoline (entry 3, Table S50)

Labelling of 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole



According to GP2: 22.4 mg of substrate and 3.2 mg of catalyst *Spectral details of the reaction mixture:*

¹H NMR (400 MHz, C_6D_6) δ = 8.56 – 8.51 (m, 1H, Ar-H), 8.36 – 8.32 (m, 1H, H_A), 8.17 – 8.11 (m, 2H, H_B), 7.25 – 7.21 (m, 1H, Ar-H), 7.11 – 7.04 (m, 1H, Ar-H), 6.66 – 6.61 (m, 1H, Ar-H), 3.77 – 3.58 (m, 4H, 2×CH₂).

Deuteration expected at δ (H_A) = 8.36 – 8.32 ppm and δ (H_B) = 8.17 – 8.11 ppm.

Determined against integral at $\delta = 6.66 - 6.61$ ppm.

Table S51. Determination of the competition rate constant κ ' from the labelling of 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole.

Entry	residual integral	% D _A	residual integral	$\% D_B$	к'
	(H/D_A)		(H/D_B)		
1	1.53	24	1.74	13	1.81
2	1.59	21	1.79	11	1.95
3	1.53	24	1.79	11	2.24
Average		23		11	2.00

D328065

Person kpb19112 DT-87-1

@proton C6D6 {C:\NMRdata} DJN 17

	C6D6
8.55 8.54 8.54 8.54 8.53 8.35 8.35 8.35 8.33 8.33 8.15 8.15 8.15	7.25 7.23 7.16 7.09 7.07 7.05 6.65 6.65 6.65

.76	74	23	Ę	.66	63	61	60	
m.	m.	m.	m.	\dot{m}	\dot{m}	\dot{m}	\dot{m}	
5		5	-	4	2	_	_	

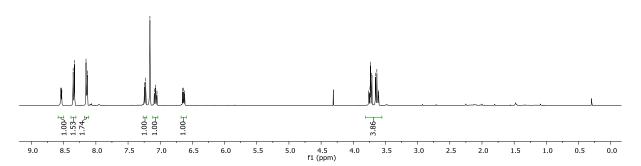


Figure S200. ¹H NMR (400 MHz, C₆D₆) of labelled 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole (entry 1, Table S51)

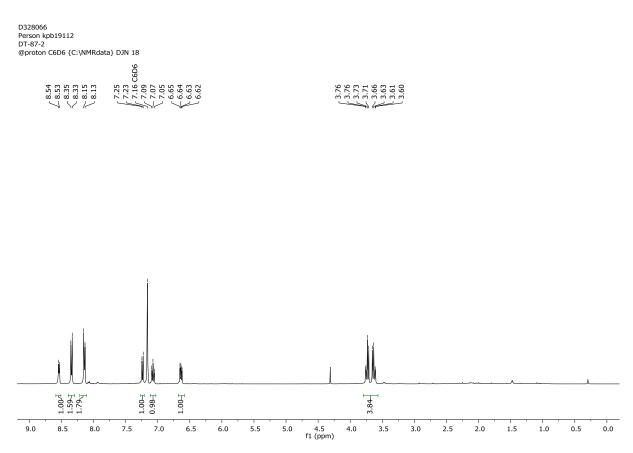


Figure S201. ¹H NMR (400 MHz, C₆D₆) of labelled 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole (entry 2, Table S51)



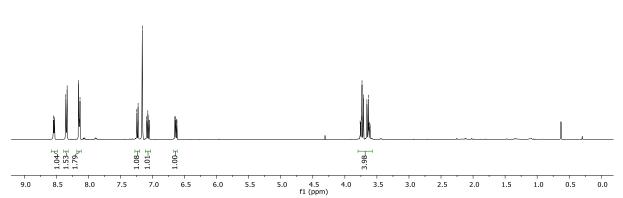


Figure S202. ¹H NMR (400 MHz, C₆D₆) of labelled 2-(4-(pyridin-2-yl)phenyl)-4,5-dihydrooxazole (entry 3, Table S51)

5. References.

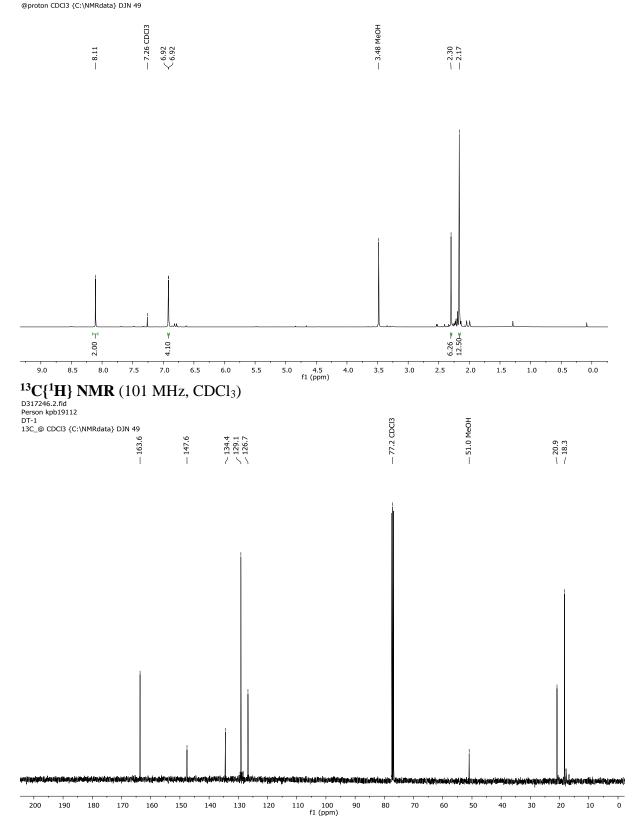
- [S1] A. Beillard, X. Bantreil, T.-X. Métro, J. Martinez and F. Lamaty, *Dalton Trans.*, 2016, 45, 17859-17866.
- [S2] M. Trose, F. Lazreg, M. Lesieur and C. S. J. Cazin Green Chem., 2015, 17, 3090-3092.
- [S3] M. Debdab, F. Mongin and J. P. Bazureau, Synthesis, 2006, 23, 4046-4052.
- [S4] M. Trose, F. Lazreg, M. Lesieur and C. S. J. Cazin, J. Org. Chem. 2015, 80 (20), 9910-9914.
- [S5] N. A. Yakelis and R. G. Bergman, Organometallics, 2005, 24, 3579–3581.
- [S6] A. J. Martínez-Martínez and A. S. Weller, Dalton Trans., 2019, 48, 3551-3554.
- [S7] A. R. Cochrane, S. Irvine, W. J. Kerr, M. Reid, S. Andersson and G. N. Nilsson, J. Label. Compd. Radiopharm., 2013, 56, 451-454.
- [S8] A. R. Kennedy, W. J. Kerr, R. Moir and M. Reid, Org. Biomol. Chem., 2014, 12, 7927-7931.
- [S9] G. R. Fulmer, A. J. M. Miller, N. H. Sherden, H. E. Gottlieb, A. Nudelman, B. M. Stoltz, J. E. Bercaw and K. I. Goldberg, *Organometallics*, 2010, 29, 2176-2179.
- [S10] J. H. Kim, R. T. Mertens, A. Agarwal, S. Parkin, G. Berger and S. G. Awuah, *Dalton Trans.*, 2019, 48, 6273-6282.
- [S11] M. R. Kumar, K. Park and S. Lee, Adv. Synth. Catal, 2010, 352, 3255-3266.
- [S12] A. S. B. Prasad, T. M. Stevenson, J. R. Citineni, V. Nyzam and P. Knochel, *Tetrahedron* 1997, 53, 7237-7254.
- [S13] C. M. Zinser, K. G. Warren, R. E. Meadows, F. Nahra, A. M. Al-Majid, A. Barakat, M. S. Islam, S. P. Nolan and C. S. J. Cazin, *Green Chem.*, 2018, 20, 3246-3252
- [S14] J. B. Badiang and J. Aubé, J. Org. Chem., 1996, 61 (7), 2484-2487.
- [S15] W.-C. Chen, Y.-C. Hsu, W.-C. Shih, C.-Y. Lee, W.-H. Chuang, Y.-F. Tsai, P. P.-Y. Chen and T.-G. Ong, *Chem. Commun.*, 2012, 48, 6702-6704.

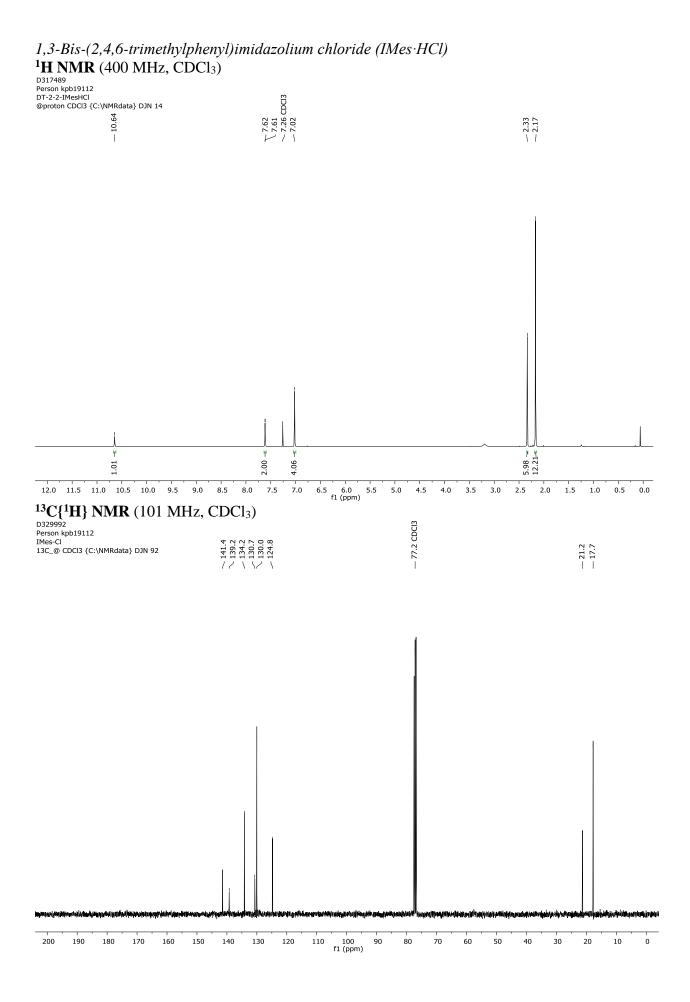
6. Copies of NMR Spectra 6.1. ¹H and ¹³C{¹H} NMR of synthesised substrates

N,N'-dimesitylethanediimine

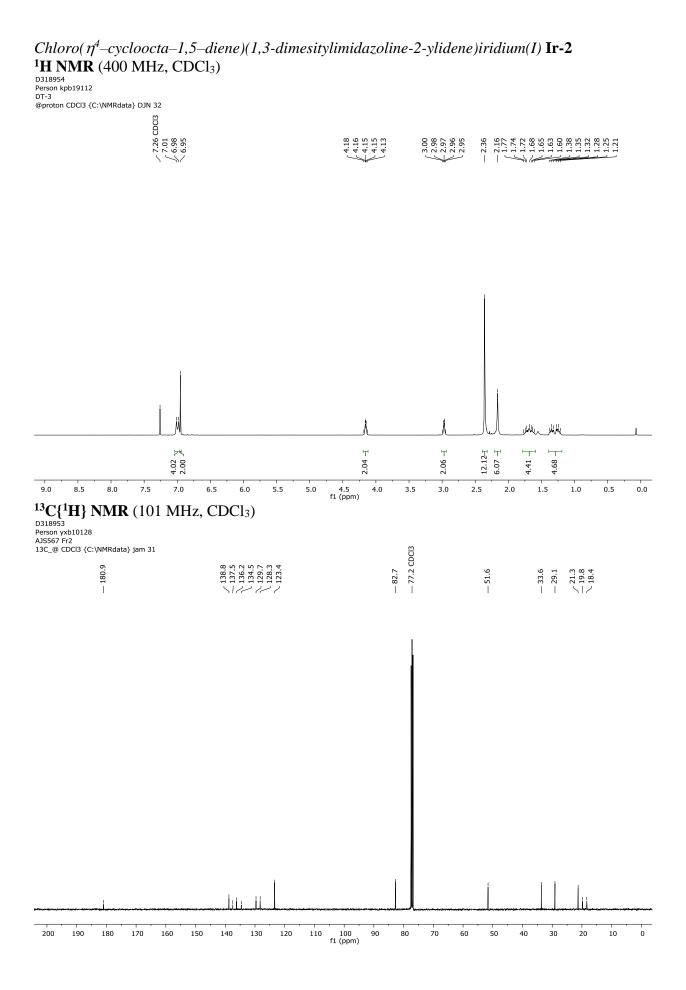
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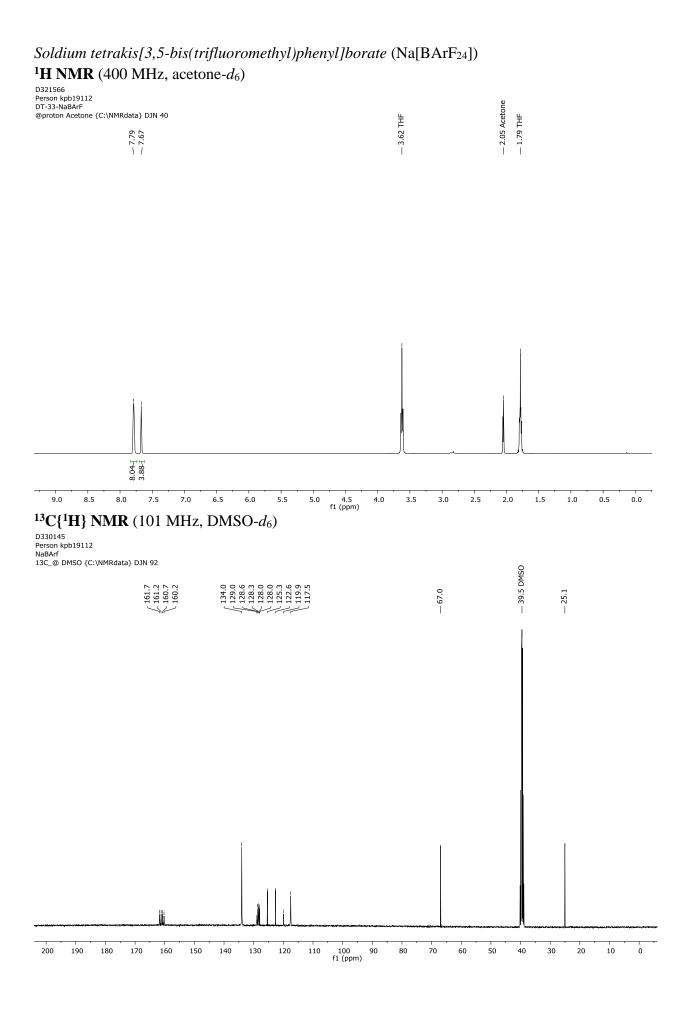




S170

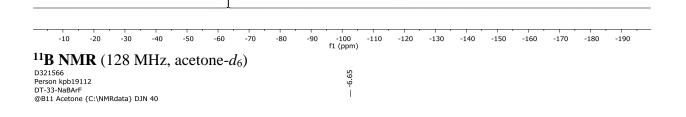


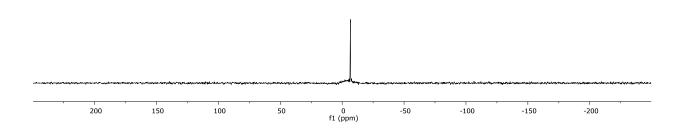
S171



¹⁹**F NMR** (376 MHz, acetone-*d*₆)

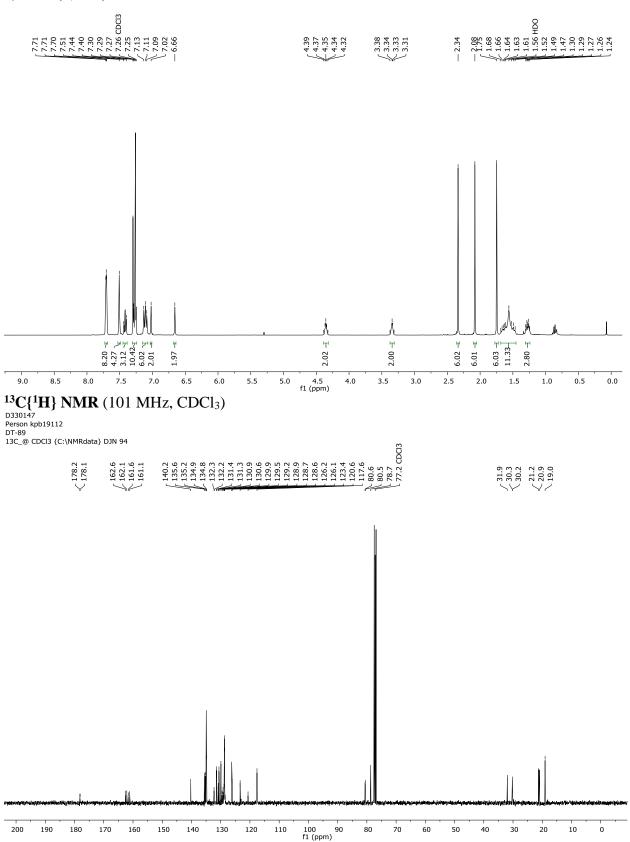
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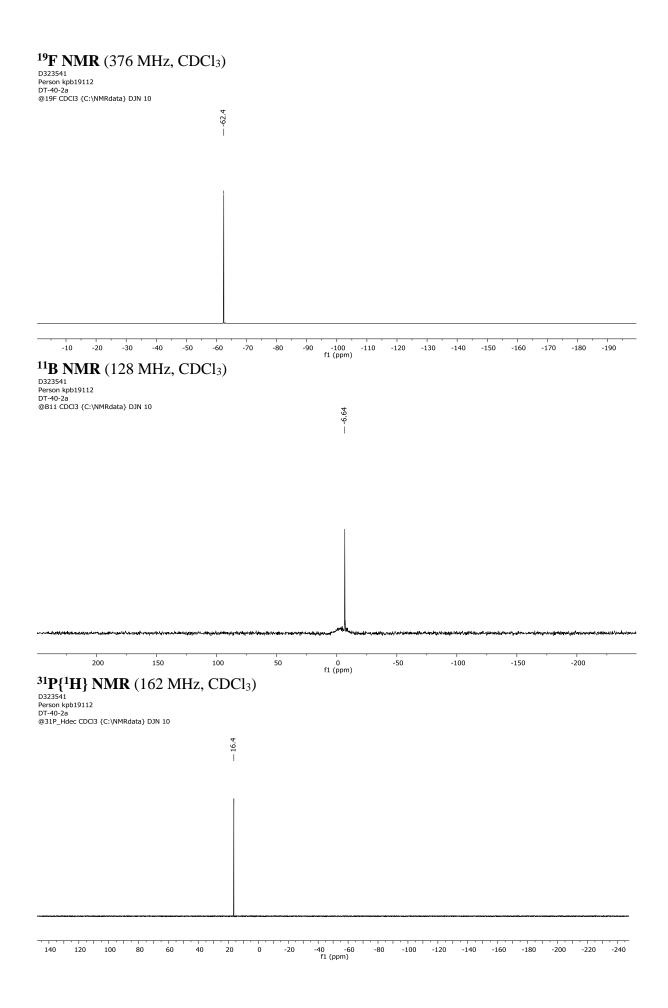


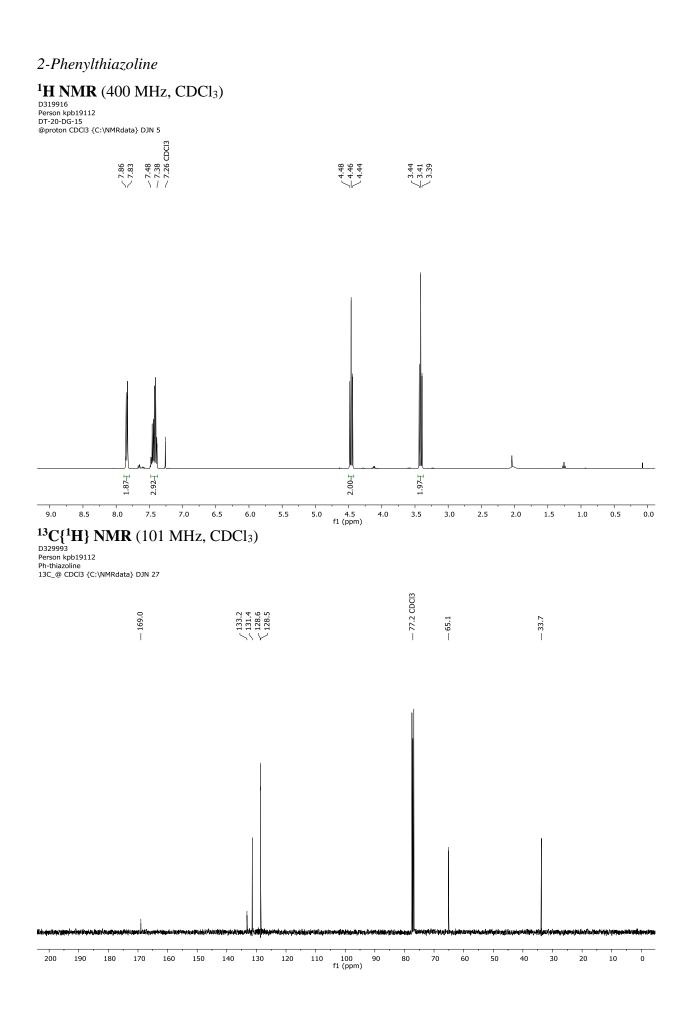


[(COD)Ir(IMes)(PPh₃)]BArF₂₄ Ir-1 ¹H NMR (400 MHz, CDCl₃)

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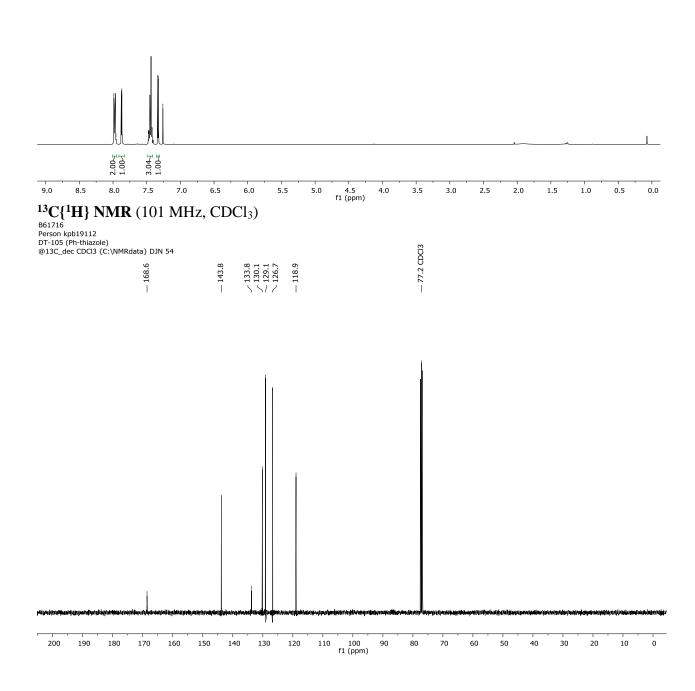


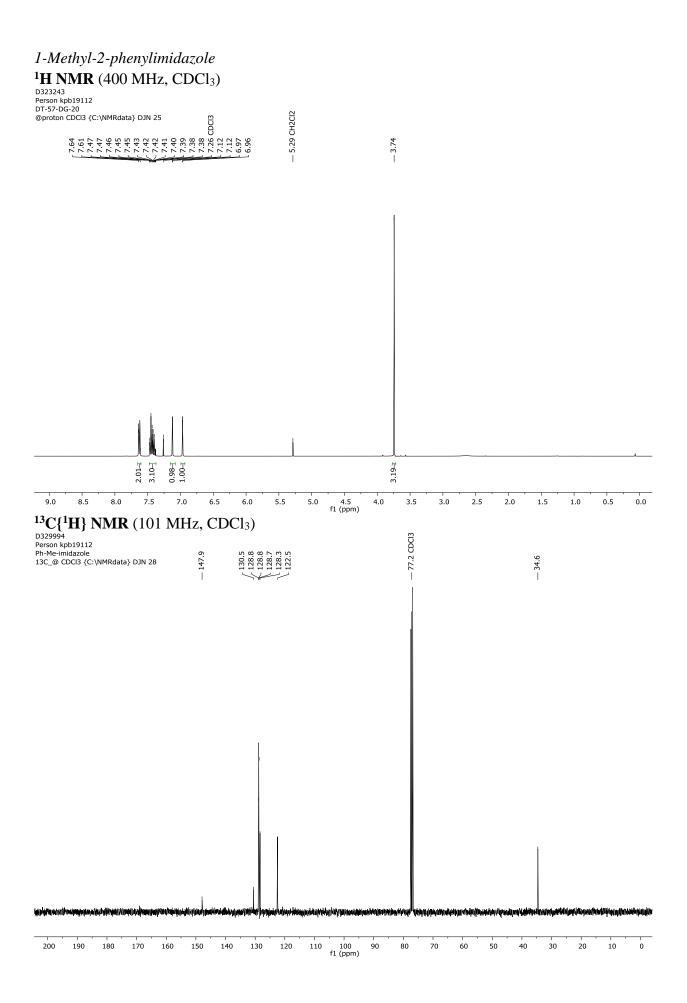


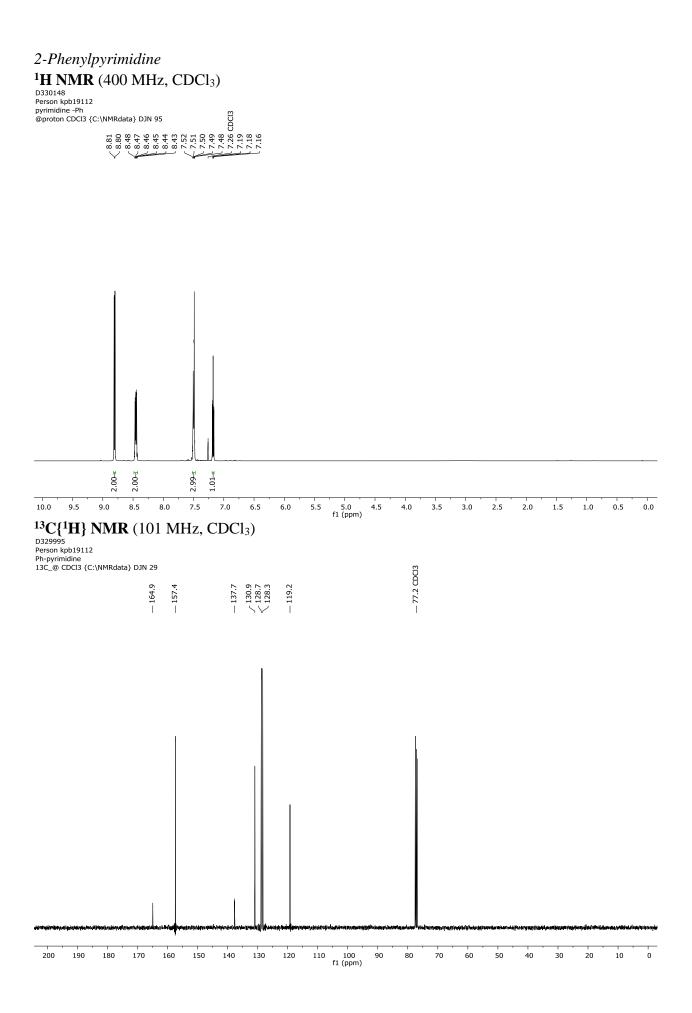
2-Phenylthiazole

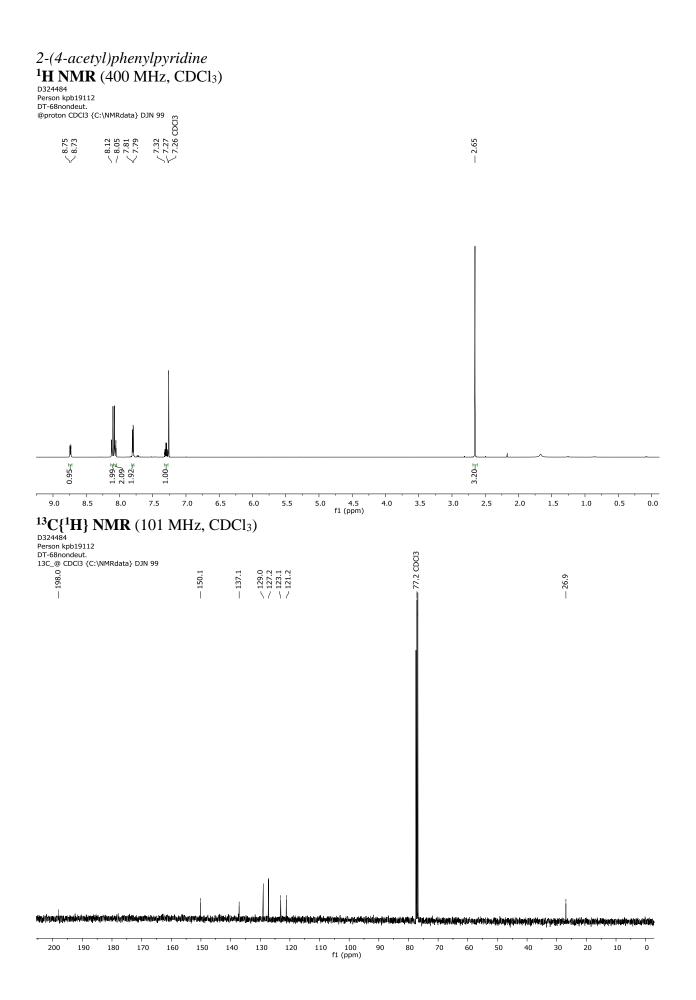
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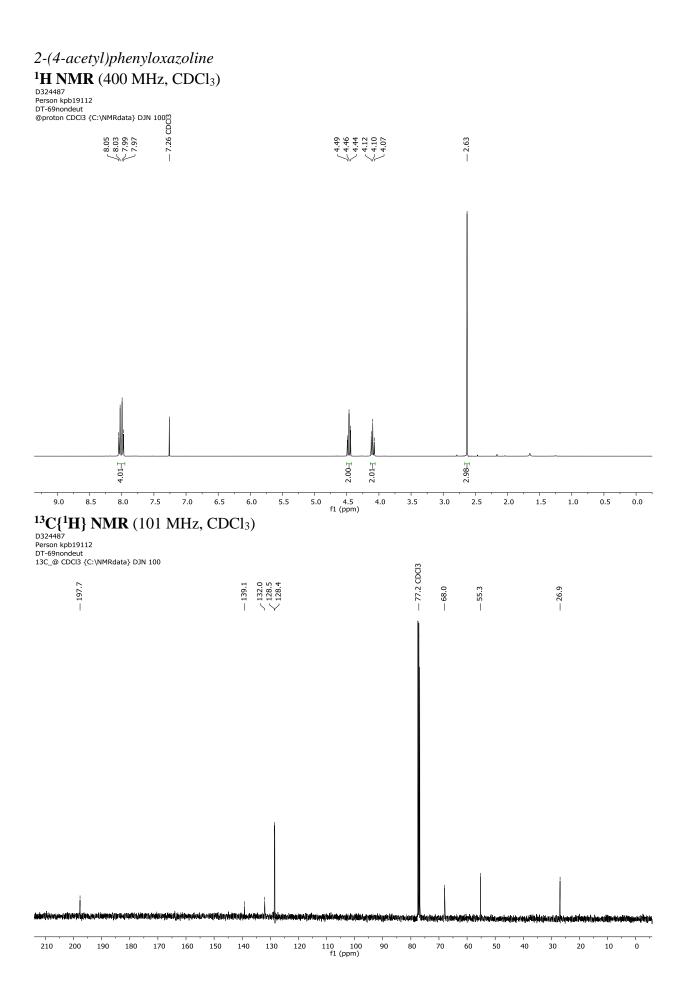
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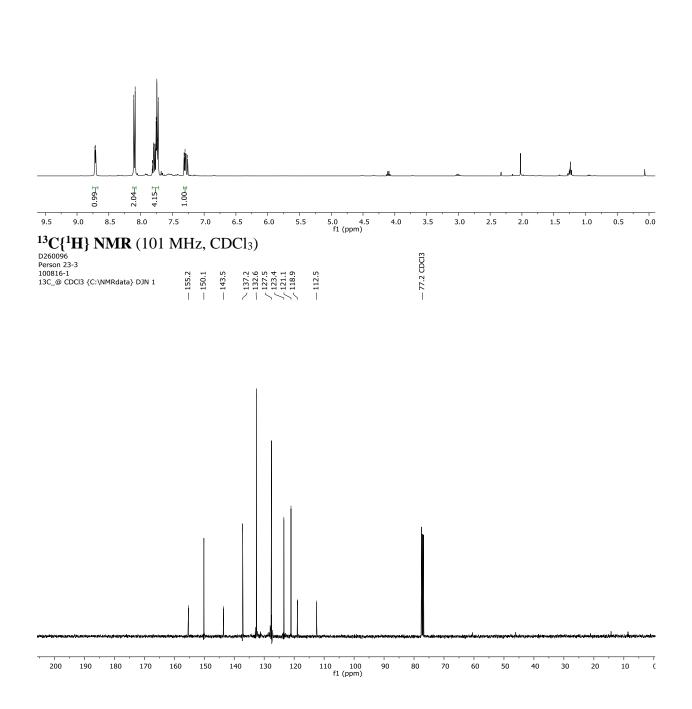


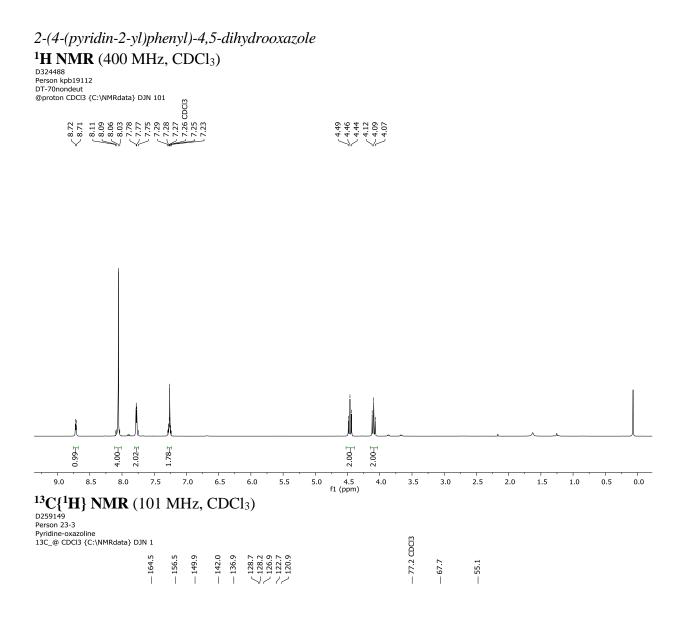


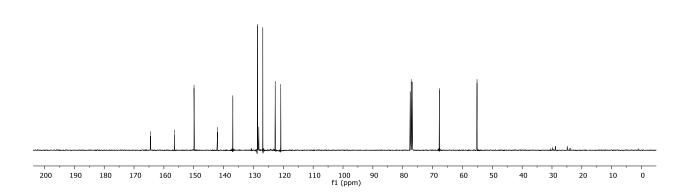


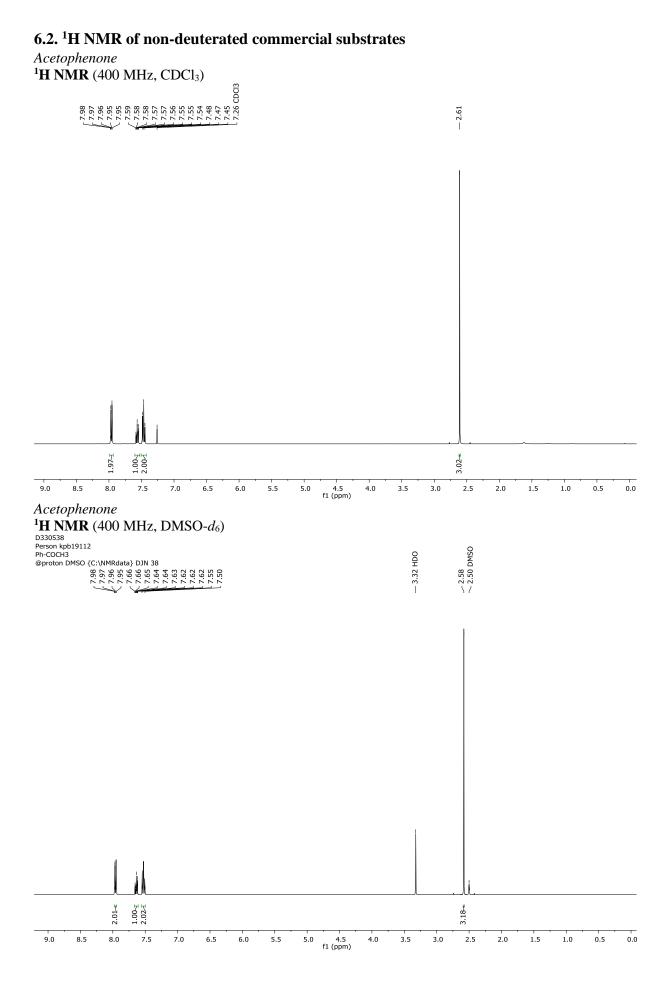


2-(4-cyano)phenylpyridine ¹H NMR (400 MHz, CDCl₃) D260096 Person 23-3 100816-1 @proton CDCl3 {C:\NMRdata} DJN 1



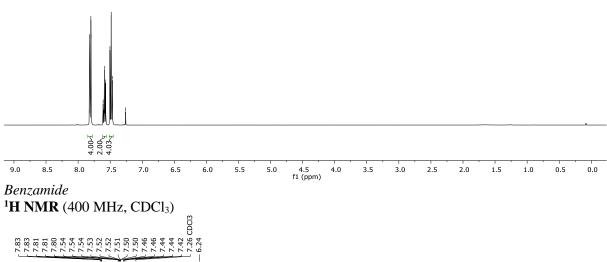


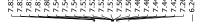


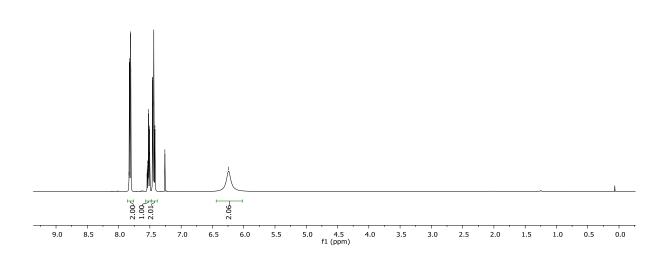


S184

Benzophenone ¹**H NMR** (400 MHz, CDCl₃) 7.83 7.82 7.80 7.80 7.80 7.51 7.51 7.55 7.57 7.56 7.47 7.26 CDCl3

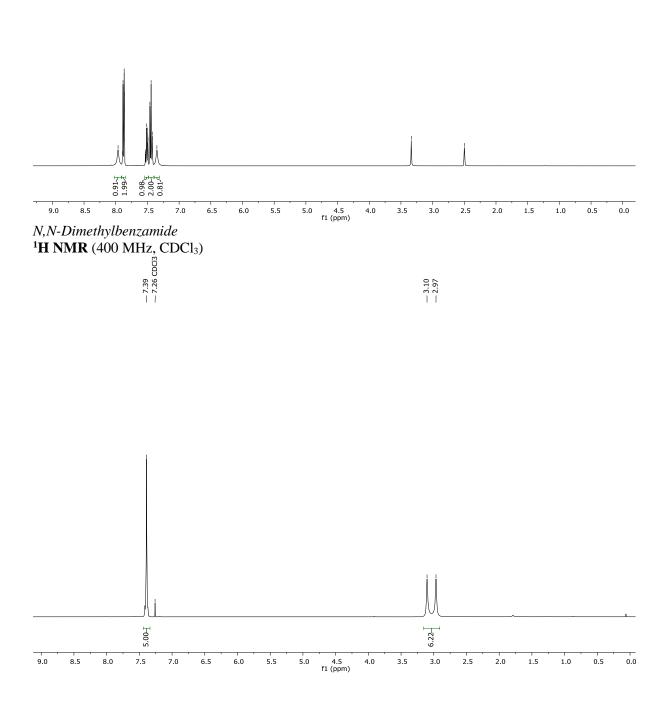


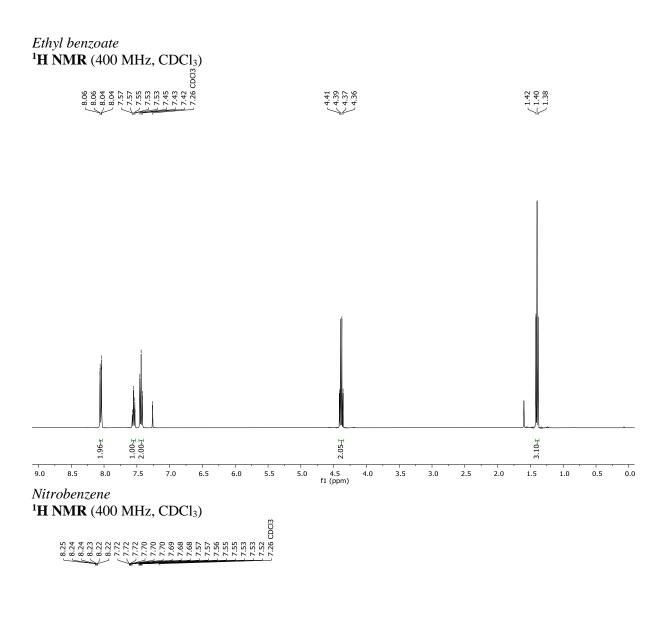


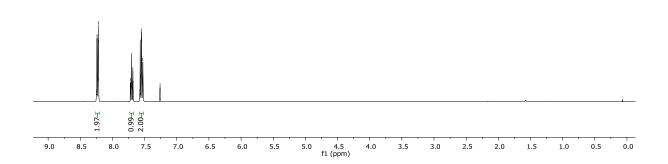


Benzamide ¹**H NMR** (400 MHz, DMSO-*d*₆)

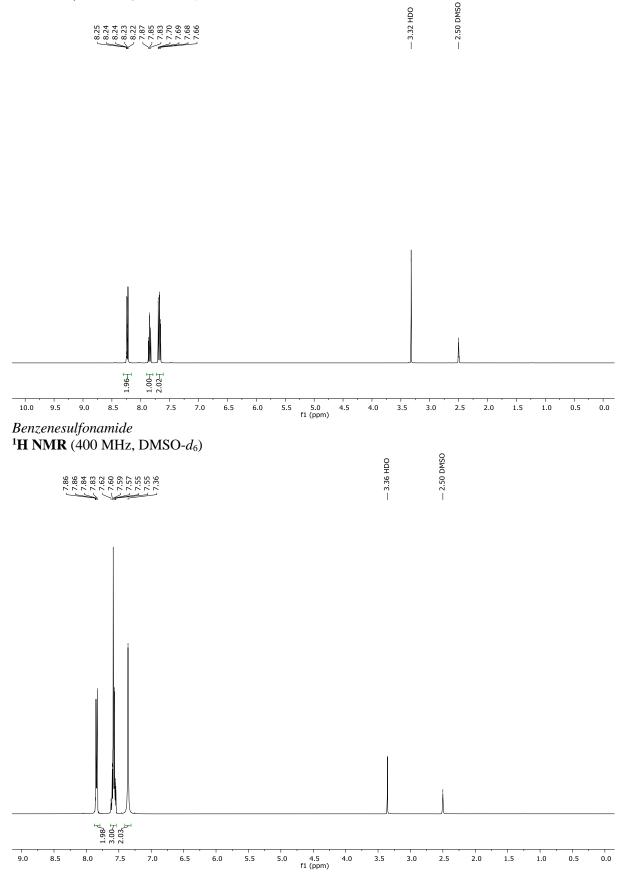


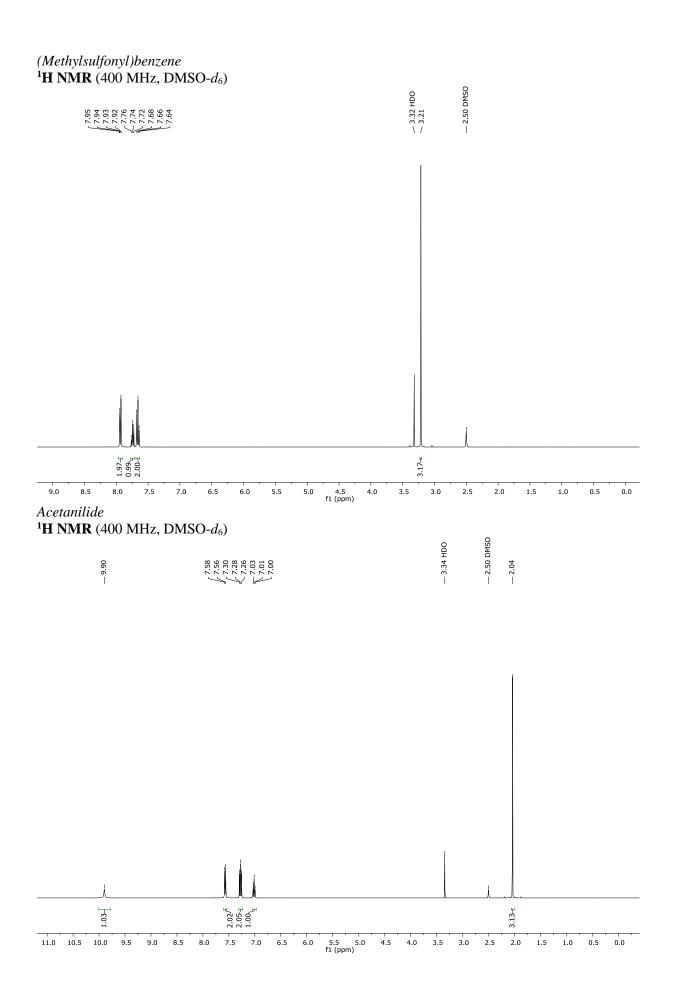


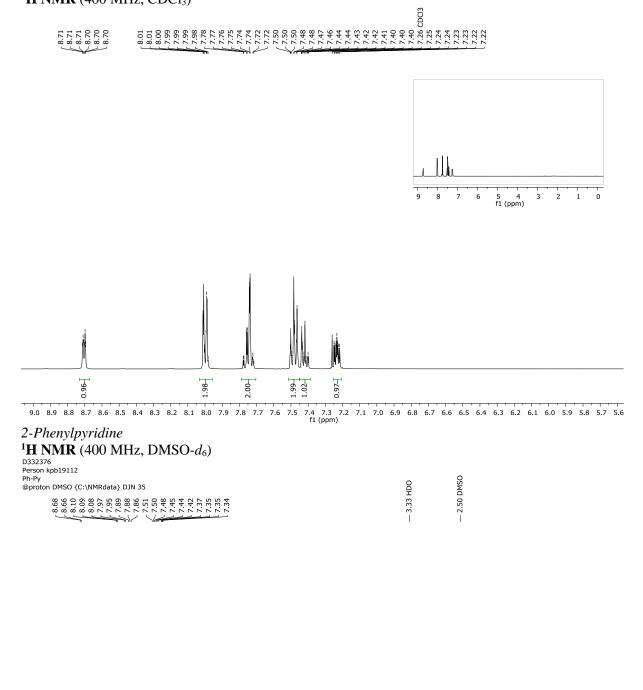


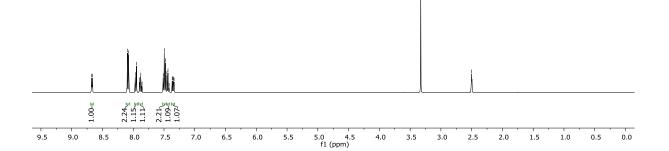


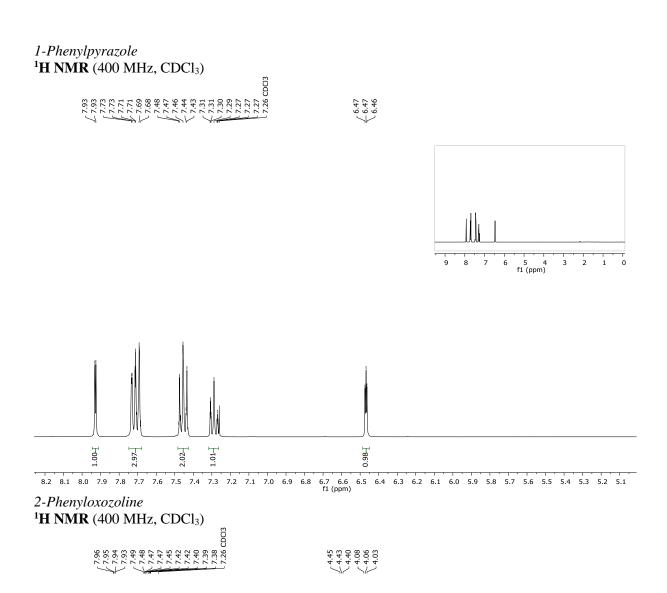
Nitrobenzene ¹**H NMR** (400 MHz, DMSO-*d*₆

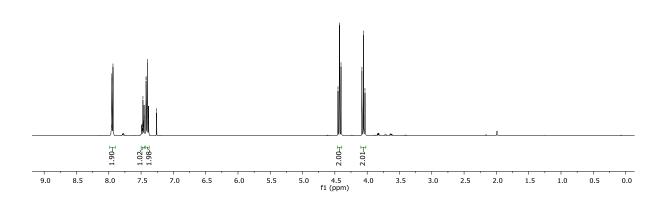




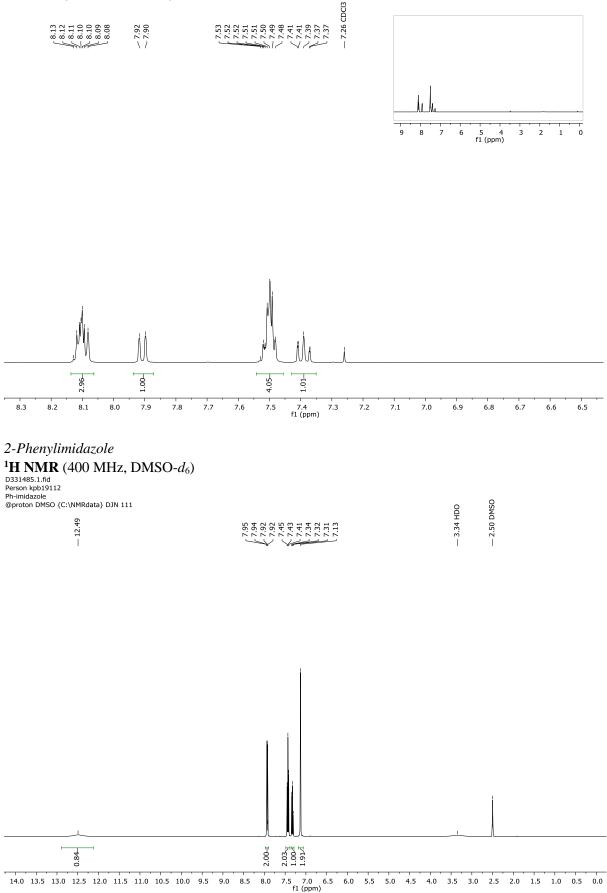








2-Phenylbenzothiazole ¹H NMR (400 MHz, CDCl₃)



S192

2-Phenylimidazoline ¹**H NMR** (400 MHz, DMSO-d₆

