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## **Supporting Information For:**

## Post-Synthetic Modification of a Metal–Organic Framework with Chemodosimeter for Rapid Detection of Lethal Cyanide via Dual Emission

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## **Materials and Methods:**

The preparation of thieno[2,3-b]thiophene-2,5-dicarboxylic acid (H<sub>2</sub>TDC) and 2-cyano-3-(pyren-1-yl)acrylic acid (CPAA) chemodosimeter was carried out by following literature protocols.<sup>1, 2</sup> The <sup>1</sup>H NMR spectra of these ligands are shown in Figures S1-S2. All other reagent grade starting materials were used as received from the commercial suppliers. Perkin Elmer Spectrum Two FT-IR spectrometer was used to record fourier transform infrared (FT-IR) spectra in the region of 400-4000 cm<sup>-1</sup>. The following indications are used to characterize absorption bands: very strong (vs), strong (s), medium (m), weak (w), shoulder (sh) and broad (br). Thermogravimetric analyses (TGA) were carried out with a Mettler-Toledo TGA/SDTA 851e thermogravimetric analyzer in a temperature range of 30-700 °C under air atmosphere at a heating rate of 5 °C min<sup>-1</sup>. Ambient temperature X-Ray powder diffraction (XRPD) patterns were measured on a Bruker D2 Phaser X-ray diffractometer operated at 30 kV, 10 mA using Cu-K $\alpha$  ( $\lambda$  = 1.5406 Å). The nitrogen sorption isotherms up to 1 bar were recorded using a Quantachrome Autosorb iQ-MP gas sorption analyzer at -196 °C. Before the sorption measurements, the compounds were degassed for 12 h under dynamic vacuum at 120 °C. Steady state fluorescence studies were performed with a HORIBA JOBIN YVON Fluoromax-4 spectrofluorometer. DFT calculations were carried out with Gaussian 09 package using 6-31+G (d,p) basis set with B3LYP method.<sup>3</sup>



Figure S1. <sup>1</sup>H NMR spectrum of of H<sub>2</sub>TDC ligand.



Figure S2. <sup>1</sup>H NMR spectrum of of CPAA ligand.



Figure S3. XRPD patterns of as-synthesized (a), methanol exchanged (b) and thermally activated (c) forms of 1.



Figure S4. XRPD patterns of 1' before (a) and after post-synthetic ligand exchange (b).



Figure S5. FT-IR spectra of as-synthesized 1 (red) and thermally activated 1' (black).



Figure S6. FT-IR spectra of P-1' before (black) and after (red) treatment with cyanide.



**Figure S7.** <sup>1</sup>H NMR spectra of **1'** (a) and **P-1'** (b) after framework digestion in  $K_3PO_4/D_2O$ . In <sup>1</sup>H NMR spectrum of digested **P-1'**, presence of the new peaks were observed for the pyrene protons (blue shaded area) along with the proton peaks of  $H_2TDC$  ligand (red shaded area).

**Digestion protocol:** 10 mg of MOF sample was added to 0.5 mL of DMSO- $d_6$ . To this solution was added 0.3 mL of saturated K<sub>3</sub>PO<sub>4</sub> in D<sub>2</sub>O. After shaking for 10 min, the MOF sample was dissolved and the organic phase was collected and analyzed by <sup>1</sup>H NMR spectroscopy.



**Figure S8.** <sup>1</sup>H NMR spectrum of **P-1'** after digestion in  $K_3PO_4/D_2O$ . To calculate the percent of conversion, the peaks corresponding to  $H_2TDC$  ligand was set to an integration of 1 and all new peaks were integrated accordingly. For **P-1'**, these new peaks are all approx. ~0.30 with respect to protons of  $H_2TDC$  ligand, corresponding to ~23% incorporation of CPAA ligand in the framework.



**Figure S9.** TG curves of as-synthesized **1** (black) and activated **1'** (red) recorded in an air atmosphere in the temperature range of 25-700 °C with a heating rate of 5 °C min<sup>-1</sup>.



**Figure S10.** TG curves of **P-1** recorded in an air atmosphere in the temperature range of 25-700 °C with a heating rate of 5 °C min<sup>-1</sup>.



Figure S11. FE-SEM images of 1' (a, b) and P-1'(c, d) in different magnifications.



**Figure S12.** N<sub>2</sub> adsorption (solid symbols) and desorption (empty symbols) isotherms of 1' (weed green squares) and **P-1'** (black circles) measured at -196 °C.



Figure S13. Hydrolytic stability of P-1' in different pH media.



Figure S14. Structure of CPAA probe and its different potential sites.



Figure S15. Change in the fluorescence intensity of P-1' upon addition of 2 mM aqueous solution (150  $\mu$ L) of various anions.



**Figure S16.** Time-dependent fluorescence enhancement of **P-1'** at 433 nm upon addition of different concentrations of CN<sup>-</sup> ion.



**Figure S17.** Change in the fluorescence intensity of **P-1'** upon addition of 2 mM CN<sup>-</sup> solution (150  $\mu$ L) in presence of other competitive anions (150  $\mu$ L).



**Concentration of CN<sup>-</sup> (mM) Figure S18.** Change in the fluorescence intensity of P-1' as a function of CN<sup>-</sup> concentration.

Sl No.	Sensor Material	Type of material	Sensing Medium	LOD	Ref.
1	P-1'	MOF	THF/ H <sub>2</sub> O	0.35 µM	this work
2	CAU-10-N <sub>2</sub> H <sub>3</sub>	MOF	Water	0.48 µM	4
3	M-ZIF-90	MOF	DMSO/ H <sub>2</sub> O	2 μΜ	5
4	carbazole- functionalized Zr(IV) MOF	MOF	Water	0.14 μΜ	6
5	Bio-MOF-1⊃DAAC	MOF	HEPES buffer	5.2 ppb	7
6	Tb-ADP-Bipy MOF	MOF	Water	30 nM	8
7	PAT-TFBE	gel	Water	1.82 µM	9
8	BP	Benzo-pyrylium– pheno-thiazine conjugate	CH <sub>3</sub> CN/H <sub>2</sub> O	0.13 μΜ	10
9	NBD-SSH-Cu <sup>2+</sup>	peptide-based ensemble	Water	24.9 nM	11
10	pyridyl azo-based chemosensor <b>2</b>	gel	Sol-gel medium	9.36 µM	12
11	pyridinium-fused	Chemo-dosimeter	THF/ H <sub>2</sub> O	54 nM	13

Table S1. Comparison of the various existing fluorescent materials for the sensing of CN<sup>-</sup>.

	pyridinone iodide				
12	3T-2CN	Oligo-thiophene chemo-sensor	DMSO/ H <sub>2</sub> O	0.19 μΜ	14
13	AuAgNCs@ ew	bimetallic gold–silver nanoclusters	Water	0.138 μM	15
14	{Ru <sup>II</sup> ( <sup>t</sup> Bubpy)(CN) <sub>4</sub> - [Cu <sup>II</sup> (dien)] <sub>3</sub> }(ClO <sub>4</sub> ) <sub>2</sub>	trinuclear hetero- bimetallic Ru(II)-Cu(II) complex	DMF/ H <sub>2</sub> O	1.2 μΜ	16
15	receptor 1	DMN conjugated benzo-thiazole	DMF/H <sub>2</sub> O	0.16 µM	17
16	PNA⊃GBP·I <sub>2</sub>	Supra-molecular polymer	DMSO-H <sub>2</sub> O	41 nM	18



Figure S19. XRPD patterns of P-1' before (a) and after (b) cyanide sensing.



**Figure S20.** Suppression of ICT (intramolecular charge transfer) due to addition of cyanide ion to the CPAA probe.



**Figure S21.** <sup>1</sup>H NMR spectrum of (a) **P-1'** and (b) cyanide treated **P-1'** digested in K<sub>3</sub>PO<sub>4</sub>/D<sub>2</sub>O. The appearance of the new peak at 5.79 ppm in the <sup>1</sup>H NMR spectrum of cyanide treated **P-1'** supports the proposed nucleophilic addition of cyanide to the vinyl group of the pyrene moiety.



**Figure S22.** <sup>1</sup>H NMR spectrum of cyanide treated **P-1'** after digestion. The peaks corresponding to H<sub>2</sub>TDC ligand were set to an integration of 1 and all new peaks were integrated accordingly. For cyanide treated **P-1'**, the new peak (H<sub>b</sub>) at 5.79 ppm is approx. 0.16 with respect to protons of H<sub>2</sub>TDC ligand. Due to nucleophilic attack of cyanide, the integration value of vinylic proton (H<sub>a</sub>) decreases from ~ 0.30 to ~0.14. Hence, percentage conversion of incorporated CPAA ligand to its cyanide adduct is ~53% under sensing conditions.

## **References:**

- 1. W.-W. He, G.-S. Yang, Y.-J. Tang, S.-L. Li, S.-R. Zhang, Z. M. Su and Y.-Q. Lan, *Chem. Eur. J.*, 2015, **21**, 9784-9789.
- 2. A. Kathiravan, M. Panneerselvam, K. Sundaravel, N. Pavithra, V. Srinivasan, S. Anandand and M. Jaccob, *Phys. Chem. Chem. Phys.*, 2016, **18**, 13332-13345
- M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. J. A. Montgomery, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 09, Revision A.1.; Gaussian, Inc.: , Wallingford, CT*, 2009.
- 4. R. Dalapati, S. Nandi, H. Reinsch, B. K. Bhunia, B. B. Mandal, N. Stock and S. Biswas, *CrystEngComm*, 2018, **20**, 4194-4201.

- A. Karmakar, N. Kumar, P. Samanta, A. V. Desai and S. K. Ghosh, *Chem. Eur.J.*, 2016, 22, 864 -868.
- 6. A. Das and S. Biswas, *Sens. Actuators, B*, 2017, **250**, 121-131.
- 7. A. Karmakar, B. Joarder, A. Mallick, P. Samanta, A. V. Desai, S. Basu and S. K. Ghosh, *Chem. Commun.*, 2017, **53**, 1253-1256.
- 8. L. Wang, S. Wang and Y. Chen, *Microchim. Acta*, 2017, **184**, 4597-4602.
- 9. H. Fang, G. Cai, Y. Hu and J. Zhang, *Chem. Commun.*, 2018, **54**, 3045-3048.
- 10. S. Mondal, S. S. Ali, S. Manna, K. Maiti, M. R. Uddin, S. Mandal, D. Mandal and A. K. Mahapatra, *New J. Chem.*, 2017, **41**, 12581-12588.
- 11. K. H. Jung and K.-H. Lee, Anal. Chem., 2015, 87, 9308-9314.
- 12. A. Panja and K. Ghosh, *ChemistrySelect*, 2018, **3**, 1809-1814.
- 13. J. Li, J. Gao, W. W. Xiong, P. Z. Li, H. Zhang, Y. Zhao and Q. Zhang, *Chem. Asian J.*, 2014, **9**, 121-125.
- 14. L. Lan, T. Li, T. Wei, H. Pang, T. Sun, E. Wang, H. Liu and Q. Niu, *Spectrochim. Acta, Part A*, 2018, **193**, 289-296.
- 15. L. Tian, Y. Li, T. Ren, Y. Tong, B. Yang and Y. Li, *Talanta*, 2017, **170**, 530-539.
- 16. C.-F. Chow, M. H. Lam and W.-Y. Wong, *Inorg. Chem.*, 2004, **43**, 8387-8393.
- 17. K. Keshav, P. Torawane, M. K. Kumawat, K. Tayade, S. K. Sahoo, R. Srivastava and A. Kuwar, *Biosens. Bioelectron.*, 2017, **92**, 95-100.
- 18. Q. Lin, K.-P. Zhong, J.-H. Zhu, L. Ding, J.-X. Su, H. Yao, T.-B. Wei and Y.-M. Zhang, *Macromolecules*, 2017, **50**, 7863-7871.