

Electronic Supplementary Information for

Synthesis and characterizations of guanidinate tin(II) complexes for ring-opening polymerizations of cyclic esters

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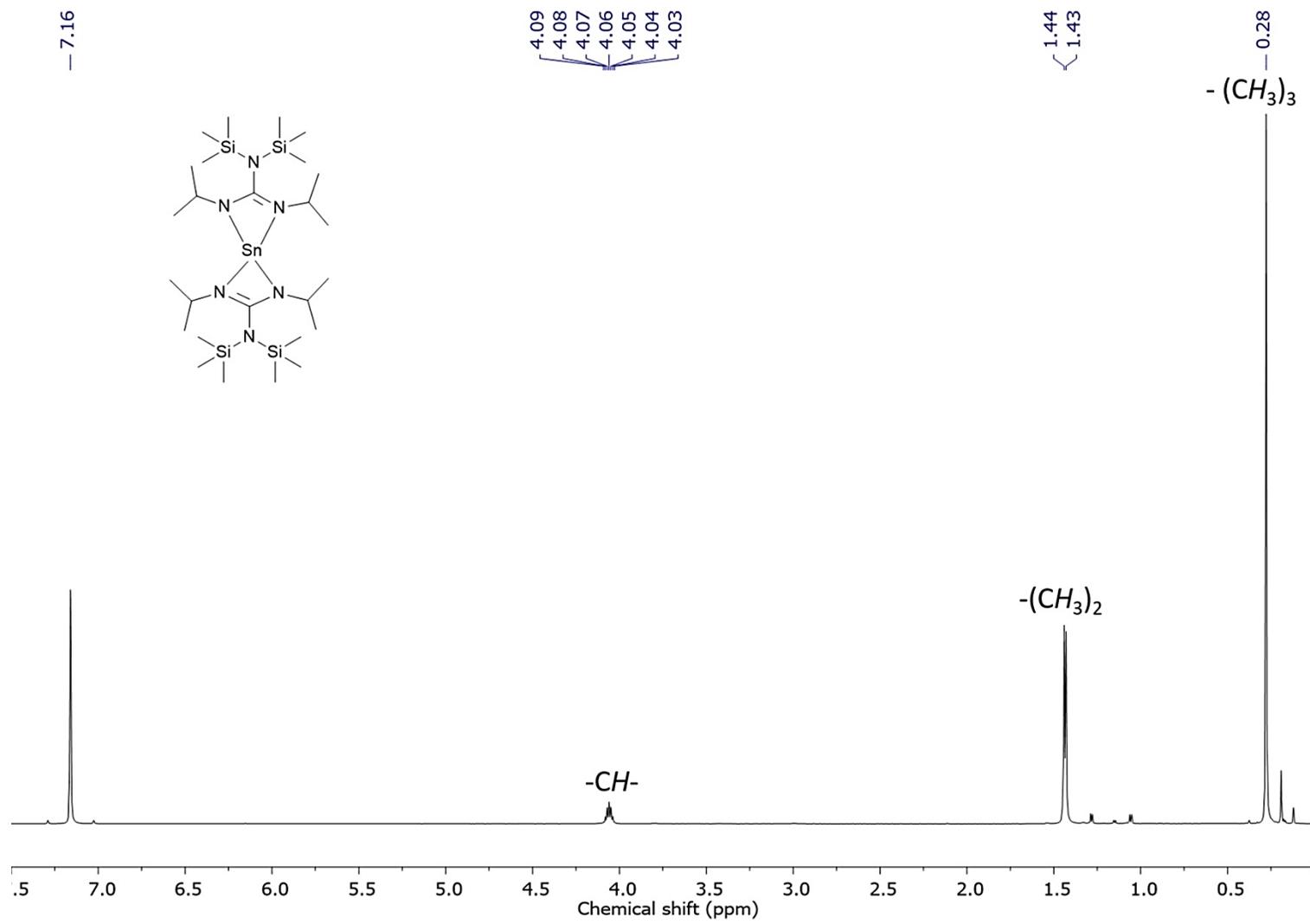


Figure S1. ¹H NMR spectra of **1** (600 MHz, C₆D₆, 30 °C).

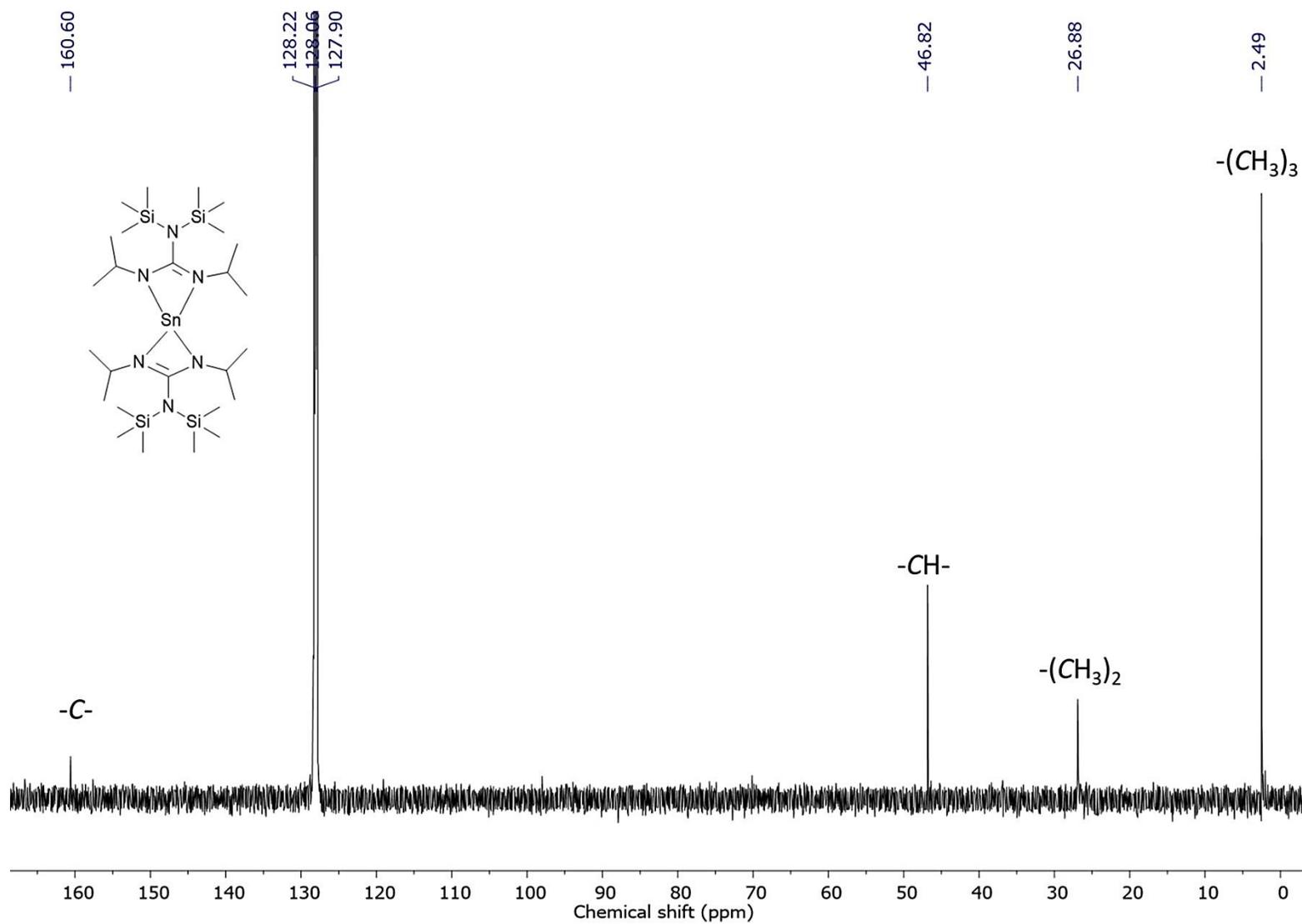


Figure S2. $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of **1** (150 MHz , C_6D_6 , 30°C).

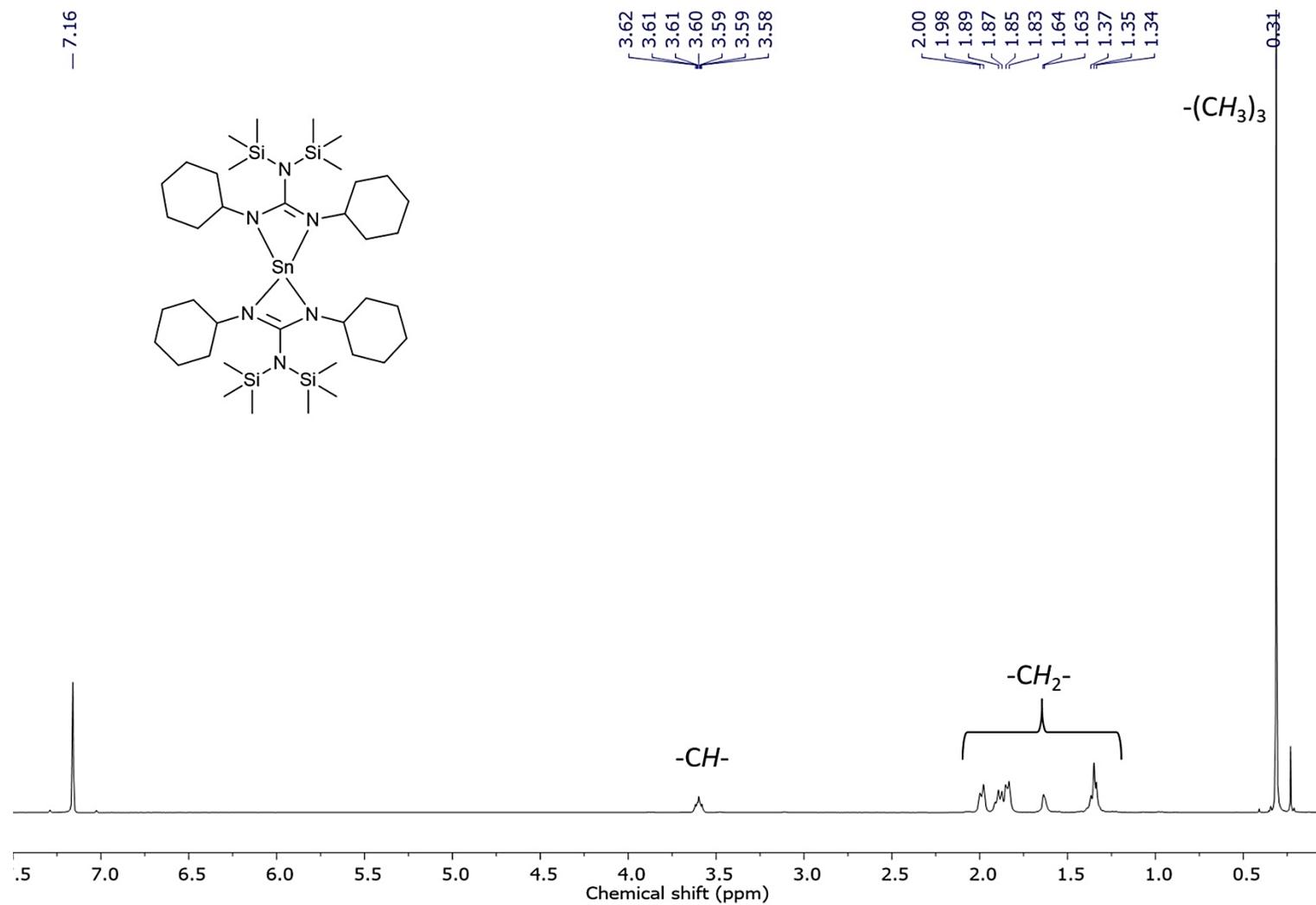


Figure S3. ^1H NMR spectra of **2** (600 MHz, C_6D_6 , 30 $^\circ\text{C}$).

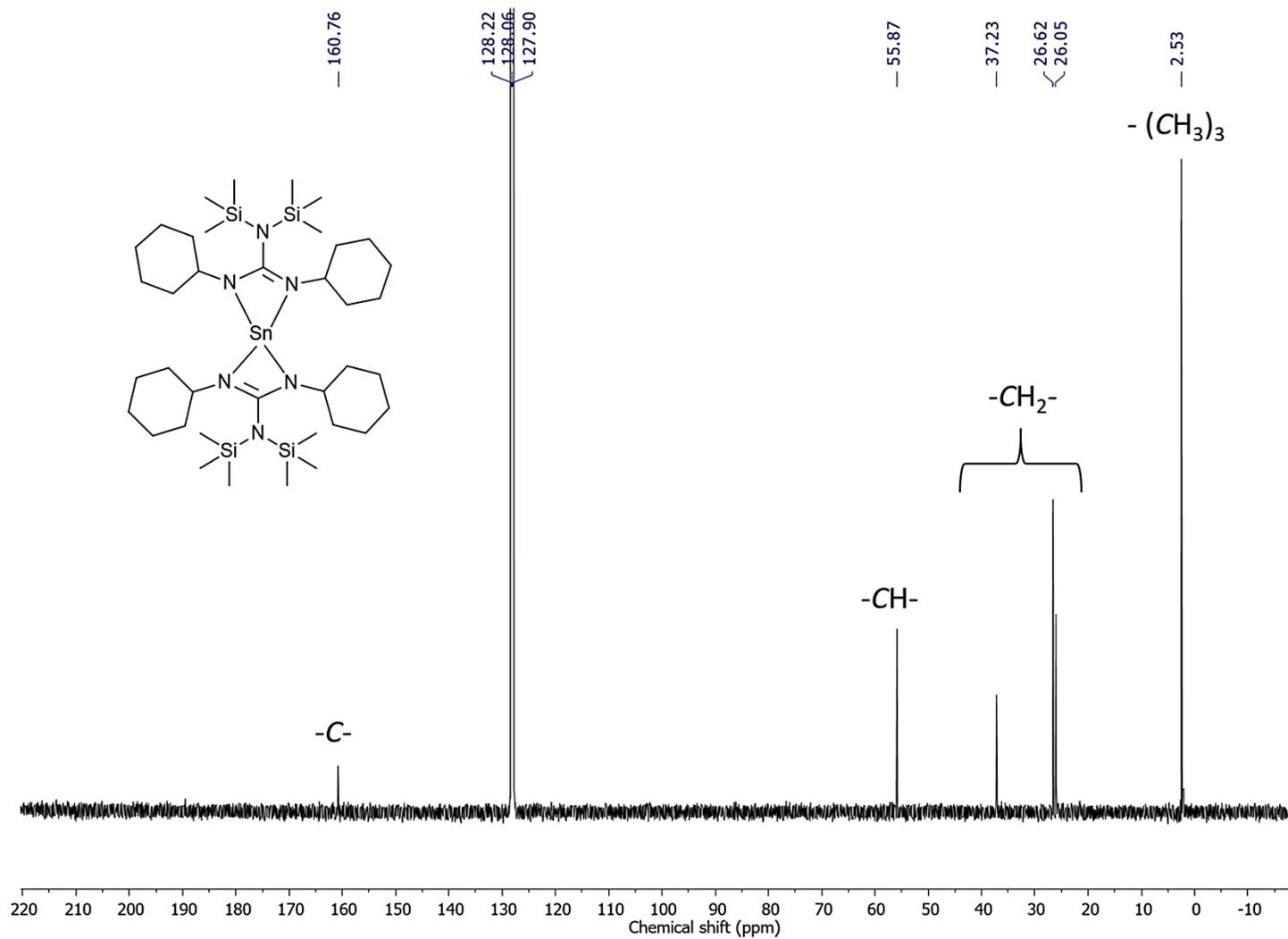


Figure S4. ¹³C{¹H} NMR spectra of **2** (150 MHz, C₆D₆, 30 °C).

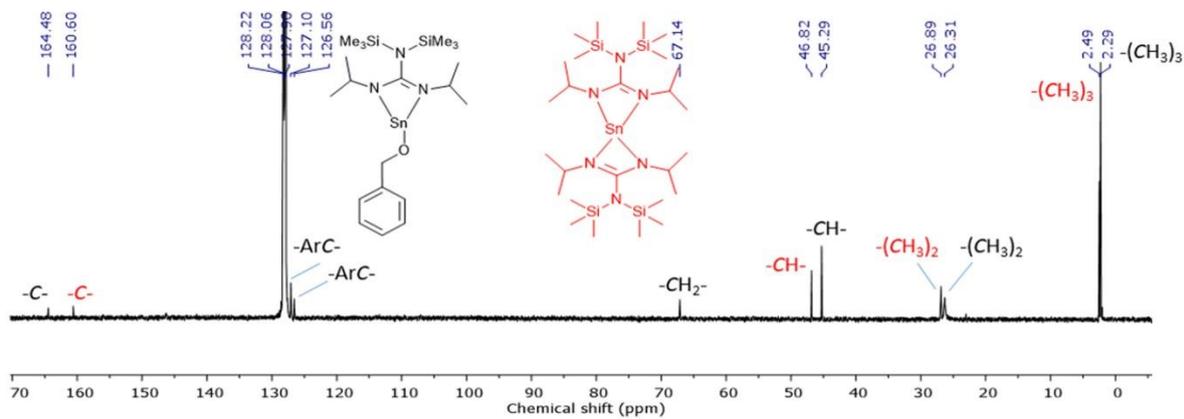


Figure S5. $^{13}\text{C}\{^1\text{H}\}$ (150 MHz, C_6D_6 , 30 °C) of **5** upon standing at room temperature for 3 h (600 MHz, C_6D_6 , 30 °C).

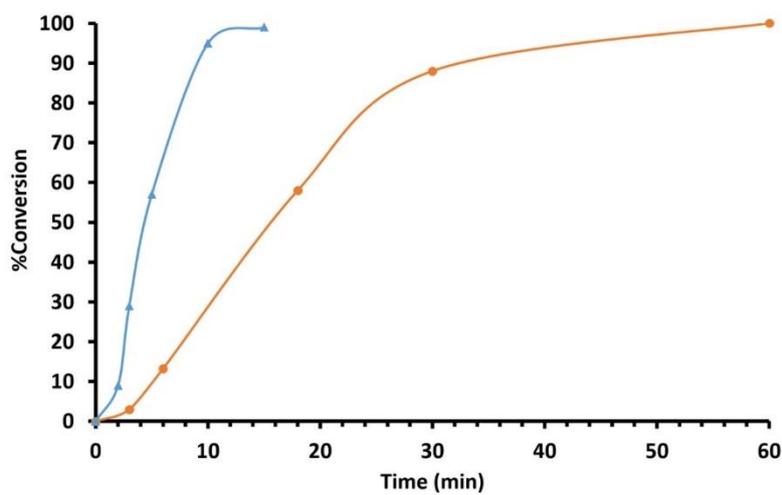


Figure S6. Neat polymerization at 120 °C.; ▲ = using ϵ -CL: 1 molar ratio of 200: 1, ● = using ϵ -CL: 2 molar ratio of 200: 1 (Table 2, entries 4-5).

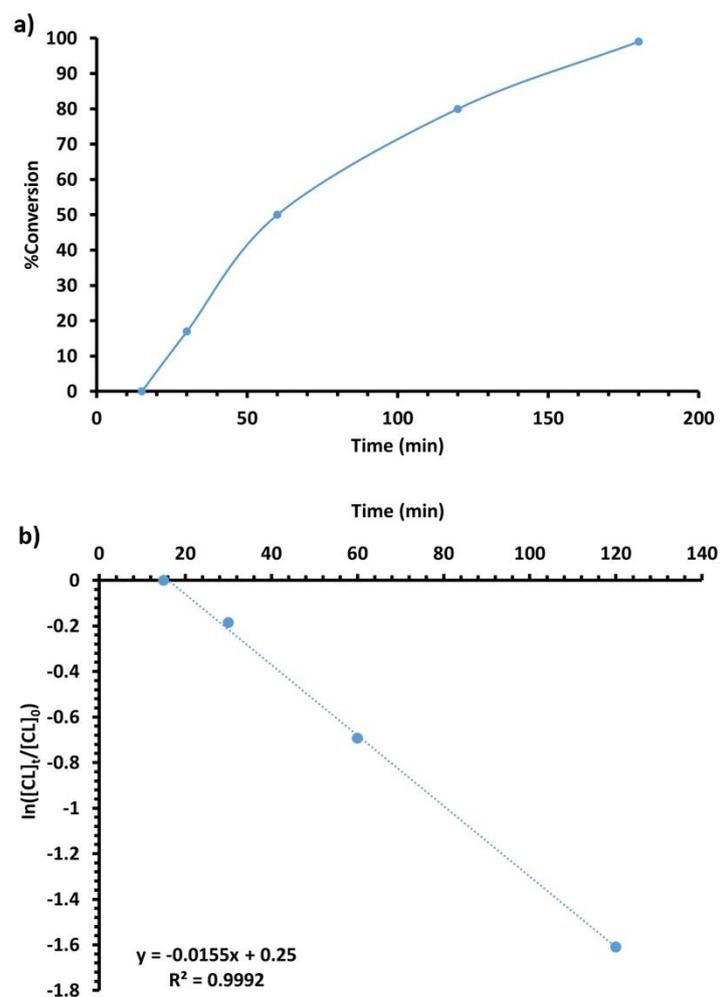


Figure S7. Solution polymerization using ϵ -CL: **1** molar ratio of 100: 1 at 100 °C in toluene (Table 2, entry 8); a) plots of conversion vs. time, b) linear plots of $\ln([\epsilon\text{-CL}]_t/[\epsilon\text{-CL}]_0)$ vs. time.

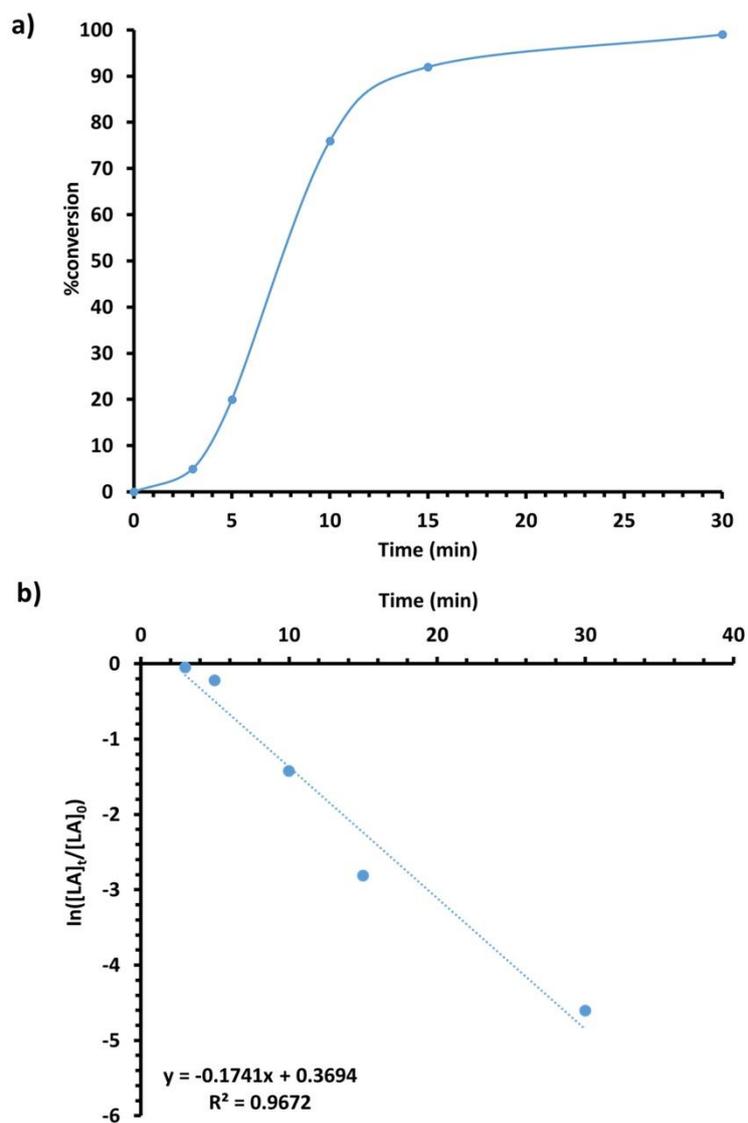


Figure S8. Solution polymerization using L-LA: 1 molar ratio of 100: 1 at 100 °C in toluene (Table 2, entry 11); a) plots of conversion vs. time, b) linear plots of $\ln([LA]_t/[LA]_0)$ vs. time.

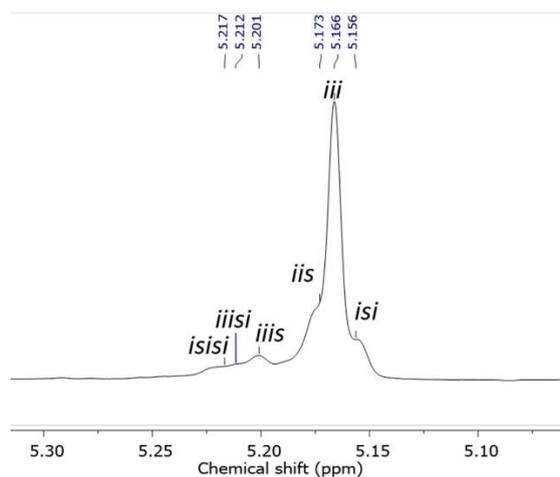


Figure S9. Homonuclear decoupled ^1H NMR spectra of PLAs obtained from *L*-LA (600 MHz, CDCl_3 , 30 $^\circ\text{C}$). NMR signals were assigned according to literature.^{1,2}

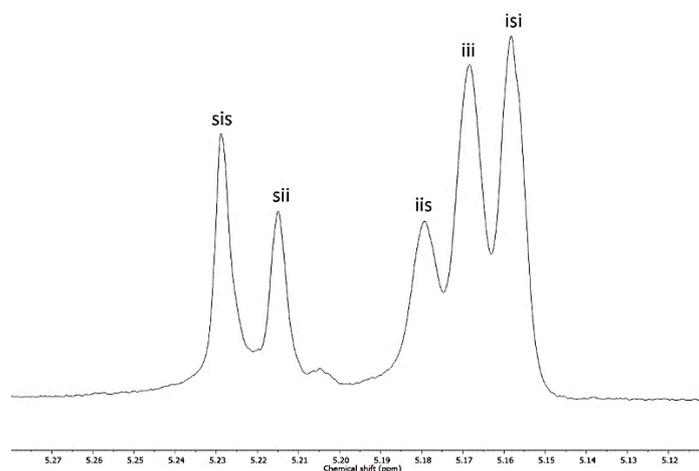


Figure S10. Homonuclear decoupled ^1H NMR spectra of PLAs obtained from *rac*-LA (600 MHz, CDCl_3 , 30 $^\circ\text{C}$). NMR signals were assigned according to literature.³ The probability of racemic enchainment (P_r) was calculated as shown below.

The determination of probability of racemic enchainment (P_r)

Generally, the atactic PLA shows five tetrads in its homonuclear decoupled ^1H NMR spectra including $[sis]$, $[sii]$, $[iis]$, $[iii]$, and $[isi]$. According to tetrad probabilities based on Bernoullian statistics⁴, The P_r value can be calculated using following equation;

$$[sis] = P_r^2/2 \quad (1)$$

$$2[sis] = P_r^2 \quad (2)$$

$$P_r = \sqrt{2[sis]} \quad (3)$$

$$P_m = 1 - P_r \quad (4)$$

Where P_m equals the probability of *meso* enchainment.

$[sis]$ value equals the integrated intensity ratio of *sis* tetrads divided by that of total methine proton region. For heterotactic PLAs, the P_r value is close to 1, on the other hand the P_r value of isotactic PLAs is close to 0. For atactic PLAs, the P_r value is 0.5.

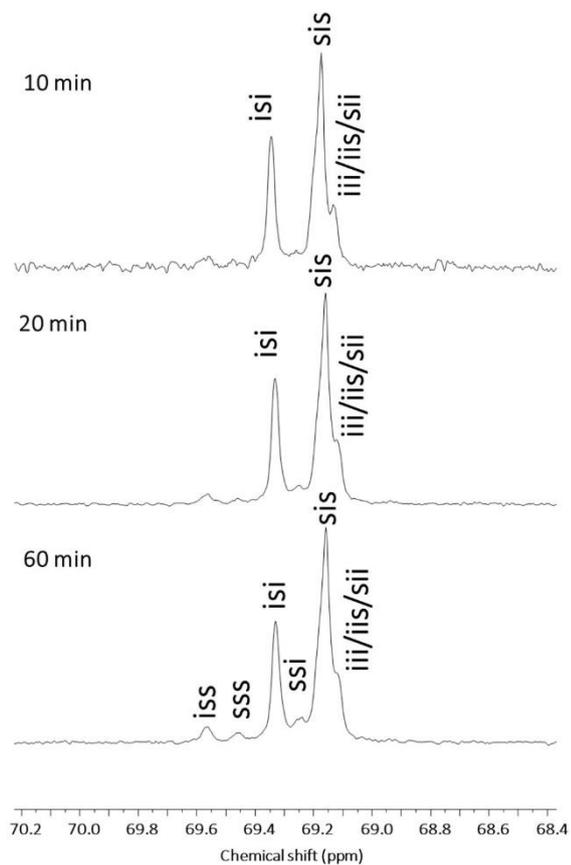


Figure S11. $^{13}\text{C}\{^1\text{H}\}$ spectra (150 MHz, CDCl_3 , 30 °C) of PLA obtained from the polymerization of *rac*-LA at 10 min (43 % conv.), 20 min (80% conv.), and 60 min (95% conv.).

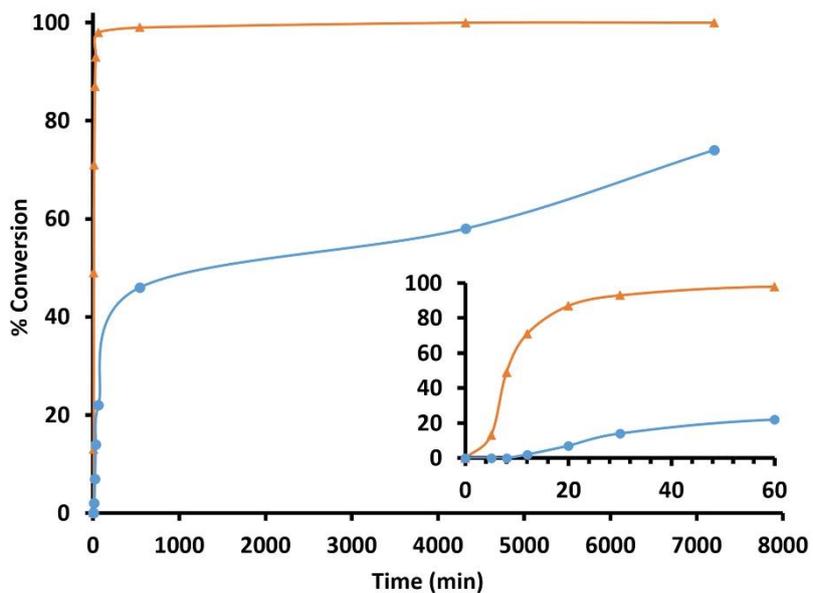


Figure S12. Solution copolymerization of *L*-LA and ϵ -CL using *L*-LA: ϵ -CL: 1 molar ratio of 100: 100: 1 at 100 °C (Table 2, entry 13); \blacktriangle = PLA, \bullet = PCL.

Determination of average sequence length

For a quantitative analysis according to Bernoulli statistics, $^{13}\text{C}\{^1\text{H}\}$ NMR spectroscopy was used to analyse the copolymer chains by the integrated intensity ratio of resonance signals. Here in, the transesterification reaction was observed in polymerization using **1** and **Sn(Oct)₂** as a catalyst. This phenomenon leads to generate CLC signal which are the second mode of transesterification implying the chains were cut during copolymerization. The experimental-average lengths of lactidyl blocks l_{LL} and l_C can be calculated by the following equation.⁵⁻⁸

$$l_{LL} = \frac{1}{2} (LLL + LLC + CLL + CLC) / (CLC + \frac{1}{2} (LLC + CLL)) \quad (1)$$

$$LLL = \frac{1}{2} [CLLLL] + \frac{1}{2} [LLLLC] + \frac{1}{3} [CLLLC] + [LLLLL] \quad (2)$$

$$LLC = \frac{1}{2} [CLLC] + \frac{1}{2} [LLLLC] + \frac{1}{3} [CLLLC] \quad (3)$$

$$CLL = \frac{1}{2} [CLLC] + \frac{1}{2} [CLLLL] + \frac{1}{3} [CLLLC] \quad (4)$$

$$CLC = [CLC] \quad (5)$$

$$l_C = (LCL + CCL + LCC + CCC) / (LCL + \frac{1}{2} (CCL + LCC)) \quad (6)$$

$$LCL = [LLCLL] + [LLCLC] + [CLCLL] + [CLCLC] \quad (7)$$

$$CCL = [CCLC] + [CCLL] \quad (8)$$

$$LCC = [CLCC] + [LLCC] \quad (9)$$

$$CCC = [CCC] \quad (10)$$

References

- 1 E. J. Shin, A. E. Jones and R. M. Waymouth, *Macromolecules*, 2012, **45**, 595–598.
- 2 K. A. M. Thakur, R. T. Kean, E. S. Hall, J. J. Kolstad, T. A. Lindgren, M. A. Doscotch, J. I. Siepmann and E. J. Munson, *Macromolecules*, 1997, **30**, 2422–2428.
- 3 N. Nimitsiriwat, V. C. Gibson, E. L. Marshall, A. J. P. White, S. H. Dale and M. R. J. Elsegood, *Dalton Trans.*, 2007, 4464–4471.
- 4 B. M. Chamberlain, M. Cheng, D. R. Moore, T. M. Ovitt, E. B. Lobkovsky and G. W. Coates, *J. Am. Chem. Soc.*, 2001, **123**, 3229–3238.
- 5 J. Kasperczyk and M. Bero, *Die Makromol. Chemie*, 1991, **192**, 1777–1787.
- 6 M. Bero, J. Kasperczyk and G. Adamus, *Die Makromol. Chemie*, 1993, **194**, 907–912.
- 7 M. Bero and J. Kasperczyk, *Macromol. Chem. Phys.*, 1996, **197**, 3251–3258.
- 8 J. Kasperczyk and M. Bero, *Die Makromol. Chemie*, 1993, **194**, 913–925.