Electronic Supplementary Information

Dual-fixations of europium cation and TEMPO species on metal-organic frameworks for aerobic oxidation of alcohols

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General materials and methods

Concentration of solution was performed using rotary evaporator, and followed by followed by vacuum drying at 0.1-1 torr. All commercial reactants, reagents and solvents used without additional purification unless otherwise stated. All chemicals were purchased from Sigma-Aldrich, TCI, Alfa Aesar chemical company.

Thin layer chromatography (TLC) analysis was performed on pre-coated silica gel 60 F254 plates. Visualization on TLC was achieved by the use of UV light (254 and 365 nm). Flash column chromatography was undertaken on silica gel (400-630 mesh). ¹H and ¹³C NMR (nuclear magnetic resonance) spectra were recorded on a FT AM 400 (400 MHz for ¹H and 100 MHz for ¹³C) or FT AM 500 (500 MHz for ¹H and 125 MHz for ¹³C). Chemical shifts were quoted in parts per million (ppm) referenced to the appropriate solvent peak. The following abbreviations were used to describe peak patterns when appropriate: br = broad, s = singlet, d =doublet, t = triplet, q = quartet, quin = quintet and m = multiplet. Coupling constants, J, were reported in Hertz unit (Hz).

Approximately 10 mg of MOFs were dried under vacuum and digested with sonication in 590 μ L of DMSO-*d*₆ and 10 μ L of HF (48% aqueous solution). Powder X-ray diffraction (PXRD) data were collected on Bruker AXS D8 Discover (at 40 kV, 40 mA for CuKa (λ = 1.5406 Å)) and Rigaku Miniflex 600 (at 40 kV, 15 mA) with a scan speed of 1.0 min/step, a step size of 0.02° in 20, and a 20 range of 3-50°. N₂ sorption isotherms of samples were obtained using a BELSORP-max at 77K. Thermogravimetric analysis (TGA) were performed under N₂ atmosphere at a scan rate of 5 °C/min using Q50 from TA instruments. PL measurements (examined by Fluoromax-4P Luminescence Spectrophotometer from HORIBA, λ_{ex} = 395 nm) for Eu species, Eu-based MOFs and Eu-loaded MOFs were in degassed aqueous solution at ambient temperature. ICP-OES analysis were collected on Perkin Elmer Optima 7300DV, with pretreatment 20 mg of sample in HF-HNO₃(1:1=v/v) at 200 °C for 1 h. GC (Gas Chromatography) analysis for determination of conversions was performed on an Younglin YL-6500. All quantum chemical calculations were performed using the optimized molecular geometry functional B3LYP/6-311G+g(d,p) as implemented in the Gaussian 09 software suite.^{S1,S2}

Synthesis of Eu(Me-BPY)₂(NO₃)₃

To a solution of 5,5'-dimethyl-2,2'-bipyridine (37 mg, 0.20 mmol) in CH₃CN (2 mL) was added to a solution of Eu(NO₃)₃·5H₂O (86 mg, 0.20 mmol) in CH₃CN (2 mL). The clear solution mixture was heated 60 °C for 12 h, a colorless precipitate was collected, washed with 3 times CH₃CN (2 mL) then dried under high vacuum for 12 h yielded Eu(Me-BPY)₂(NO₃)₃ as a colorless solid (60 mg, 85%).



Synthesis of BPDC-TEMPO

BPDC-TEMPO was synthesized according to the reported literature.^{S3}

MOF characterizations

PXRD (Powder X-ray diffraction): PXRD data was collected at ambient temperature on a Bruker D8 Discover at 40 kV, 40 mA and a Rigaku Miniflex at 40 kV, 15 mA for CuKa (λ = 1.5406 Å), with a scan speed of 1 sec/step, a step size of 0.02° in 20, and a 20 range of 5-30°.

 N_2 full isotherm for UiO-67-bpy and UiO-67-bpy-Eu: Approximately 30-60 mg of MOF sample was evacuated under vacuum for a moment at room temperature. Samples were then transferred to a preweighed sample tube and degassed at 120 °C on a Micromeritics ASAP 2020 Adsorption Analyzer for a minimum of 6 h or until the outgas rate was <5 µmHg/min. The sample tube was re-weighted to obtain a consistent mass for the degassed MOF materials. BET surface area (m²/g) measurements were collected at 77 K by N₂ on a Micromeritics ASAP 2020 Adsorption Analyzer using a volumetric technique.

N₂ full isotherm for UiO-67-TEMPO, UiO-67-(bpy&TEMPO), UiO-67-(bpy-Eu&TEMPO): These MOFs were measured according to activation at 50 °C for 24 h.

TGA (Thermogravimetric analysis): Approximately 10 mg of MOF was used for TGA measurements, after BET analysis (activated). Sample was analyzed under a stream of N_2 using a TGA/DSC 1 running from room temperature to 700 °C with a scan rate of 5 °C/min.

PL (Photoluminescence) Measurements: PL measurements (examined by Fluoromax-4P Luminescence Spectrophotometer from HORIBA, λ_{ex} = 395 nm) for Eu species, Eu-based MOFs and Eu-loaded MOFs were in degassed aqueous solution at ambient temperature.

Chemical stability of MOFs: Approximately 20 mg of MOF was placed in 4 mL vials containing 2 mL of acetone, DCM, pyridine, H₂O, 0.1 M HCl, 0.1 M NaOH. Samples were allowed to sit statically at 25 °C for 24 h. The samples were filtered and recovered for analysis.

Procedures for aerobic oxidations

Condition I in Tables 2 and 3 (Eu-fixed): Alcohol (0.25 mmol), UiO-67-bpy-Eu (2 mg, 0.0025 mmol), TEMPO (0.4 mg, 0.0025 mmol), sodium nitrate (0.2 mg, 0.0025 mmol) and toluene (1 mL) were added to a scintillation vial. Acetic acid (0.1 mL) was added to solution, and then the mixture was stirred at 60 °C under air. Resulting mixture was filtered by syringe filter and checked by conversion ratio with GC column or the desired product was isolated by a silica gel column chromatography.

Condition II in Table 3 (TEMPO-fixed): Alcohol (0.25 mmol), Eu(NO₃)₃ (1 mg, 0.0025 mmol), UiO-67-TEMPO (3 mg, 0.0025 mmol), sodium nitrate (0.2 mg, 0.0025 mmol) and toluene (1 mL) were added to a scintillation vial. Acetic acid (0.1 mL) was added to solution, and then the mixture was stirred at 60 °C under air. Resulting mixture was filtered by syringe filter and checked by conversion ratio with GC column or the desired product was isolated by a silica gel column chromatography.

Condition III in Table 3 (Eu & TEMPO-fixed): Alcohol (0.25 mmol), UiO-67-bpy-Eu (2 mg, 0.0025 mmol), UiO-67-TEMPO (3 mg, 0.0025 mmol), sodium nitrate (0.2 mg, 0.0025 mmol) and toluene (1 mL) were added to a scintillation vial. Acetic acid (0.1 mL) was added to solution, and then the mixture was stirred at 60 °C for 18 h under air. Resulting mixture was filtered by syringe filter and checked by conversion ratio with GC column or the desired product was isolated by a silica gel column chromatography.

Condition IV in Table 3 (Eu-fixed + TEMPO-fixed): Alcohol (0.25 mmol), UiO-67-(bpy-Eu)(TEMPO) (6 mg, 0.0025 mmol), sodium nitrate (0.2 mg, 0.0025 mmol) and toluene (1 mL) were added to a scintillation vial. Acetic acid (0.1 mL) was added to solution, and then the mixture was stirred at 60 °C for 18 h under air. Resulting mixture was filtered by syringe filter and checked by conversion ratio with GC column or the desired product was isolated by a silica gel column chromatography.

General GC conditions

GC-Condition #1: GsBP-5 column, 30m x 0.32mm (id); FID detector, 300 °C; injection: 200 °C; carrier gas: nitrogen; carrier gas rate: 1 mL / min; area normalization:

- Reaction condition optimization experiments (Table 1), and the detection of benzyl alcohol (**1a**), 4chlorobenzylalcohol (**1d**), 3-chlorobenzylalcohol (**1e**), 2-chlorobenzylalcohol (**1f**), 2-thiophene methanol (**1k**), and 3-thiophene methanol (**1I**) were performed under a Condition Is column temperature: 120 °C for 2 minutes, increasing to 190 °C in a rate of 10 °C / min.

- 4-Nitrobenzylalcohol (1b), 4-bromobenzylalcohol (1c), 4-methylbenzylalcohol (1g), 4-

methoxybenzylalcohol (**1h**), 3-methoxybenzylalcohol (**1i**), 2-methoxybenzylalcohol (**1j**) were detected under a Condition Is column temperature: 150 °C for 2 minutes, raising to 210 °C in a rate of 10 °C / min.

- 1-Hexanol (1m) and 2-heptanol (1n) were detected under a Condition Is column temperature: 80 °C for 1 minutes, raising to 100 °C in a rate of 1 °C / min

- Diphenyl ketone (**2o**) was detected under a Condition Is column temperature: 120 °C for 1 minutes, raising to 190 °C in a rate of 2 °C / min.

GC-Condition #2: DB-5 column, 30m x 0.32mm (id); FID detector, 350 °C; injection: 250 °C; carrier gas: nitrogen; carrier gas rate: 1 mL / min; area normalization.

- Pyrenyl aldehyde (**2p**) was detected under a Condition Is column temperature: 120 °C for 2 minutes, raising to 330 °C in a rate of 15 °C / min.

Characterizations of aldehydes or ketones from aerobic oxidations



Benzaldehyde (**2a**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate/*n*-hexane (1/30, v/v) as the eluent. ¹H NMR (400 MHz, CDCl₃) δ 10.03 (s, 1H), 7.90-7.88 (m, 2H), 7.64-7.62 (m, 1H), 7.56-7.52 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 192.6, 136.5, 134.6, 129.9, 129.1. Condition I in Tables 1 and 2: Isolation yield (98%, 26 mg), obtained as a colorless liquid. Condition II in Table 3: GC conversion (99%, 9 h). Condition III in Table 3: GC conversion (65%, 18 h). Condition IV in Table 3: GC conversion (70%, 18 h).

O₂N

4-Nitrobenzaldehyde (**2b**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate/*n*-hexane (1/10, v/v) as the eluent, obtained as a pale yellow liquid. ¹H NMR (500 MHz, CDCl₃) δ 10.15 (s, 1H), 8.40-8.37 (m, 2H), 8.09-8.06 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 190.5, 151.3, 140.2, 130.6, 124.4.

Condition I: GC conversion (99%, 9 h).

4-Bromobenzaldehyde (**2c**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:20 v/v) as the eluent, obtained as a colorless solid. ¹H NMR (500 MHz, CDCl₃) δ 9.97 (s, 1H), 7.75-7.73 (m, 2H), 7.69-7.67 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 191.2, 135.2, 132.6, 131.1, 129.9.

Condition I: GC conversion (99%, 11 h).



4-Chlorobenzaldehyde (2d)^{S3}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:20 v/v) as the eluent, obtained as a colorless solid. ¹H NMR (400 MHz, CDCl₃) δ

9.98 (s, 1H), 7.84-7.82 (m, 2H), 7.53-7.51 (m, 2H); ^{13}C NMR (100 MHz, CDCl₃) δ 191.0, 141.1, 134.8, 131.1, 129.6.

Condition I: GC conversion (99%, 11 h).

CI

3-Chlorobenzaldehyde (**2e**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate/*n*-hexane (1/30 v/v) as the eluent, obtained as a light yellow liquid. ¹H NMR (500 MHz, CDCl₃) δ 10.47 (s, 1H), 7.92-7.90 (m, 1H), 7.78-7.76 (m, 1H), 7.45-7.43 (m, 1H), 7.39-7.36 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 189.9, 138.0, 135.2, 132.6, 130.7, 129.5, 127.4. Condition I: GC conversion (99%, 8 h).



2-Chlorobenzaldehyde (**2f**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:30 v/v) as the eluent, obtained as a light yellow liquid. ¹H NMR (500 MHz, CDCl₃) δ 9.97 (s, 1H), 7.85-7.85 (m, 1H), 7.77-7.75 (m, 1H), 7.61-7.58 (m, 1H), 7.50-7.47 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 190.1, 137.9, 135.6, 134.5, 130.5, 129.4, 128.1. Condition I: GC conversion (99%, 13 h).



4-Methylbenzaldehyde (**2g**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate/*n*-hexane (1/20, v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ 9.95 (s, 1H), 7.78-7.77 (m, 2H), 7.34-7.32 (m, 2H), 2.44 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 192.2, 145.7, 134.4, 130.0, 129.9, 22.0.

Condition I: GC conversion (99%, 11 h).



4-Methoxybenzaldehyde (**2h**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate/*n*-hexane (1/30, v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ

9.88 (s, 1H), 7.84-7.82 (m, 2H), 7.00-6.99 (m, 2H), 3.88 (s, 3H); ^{13}C NMR (125 MHz, CDCl₃) δ 190.9, 164.7, 132.1, 130.1, 114.4, 55.7. Condition I: GC conversion (99%, 18 h).

MeO

3-Methoxybenzaldehyde (**2i**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate/*n*-hexane (1/30, v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ 9.98 (s, 1H), 7.46-7.44 (m, 2H), 7.40-7.39 (m, 1H), 7.20-7.17 (m, 1H) 3.87 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.3, 160.3, 137.9, 130.2, 123.7, 121.7, 112.2, 55.6. Condition I: GC conversion (99%, 8 h).



2-Methoxybenzaldehyde (**2j**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:30 v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ 10.47 (s, 1H), 7.84-7.82 (dd, 1H, *J* = 9.6, 2.3 Hz), 7.58-7.53 (ddd, 1H, *J* = 10.6, 9.2, 2.3 Hz), 7.05-6.98 (m, 2H) 3.93 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 190.0, 162.0, 136.1, 128.7, 125.0, 120.8, 111.7, 55.8.

Condition I: GC conversion (99%, 14 h).



2-Thiophenecarboxaldehyde (**2k**)^{S4}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:30 v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ 9.95 (s, 1H), 7.79-7.78 (dd, 1H, *J* = 3.8, 1.2 Hz), 7.78-7.78 (m, 1H), 7.23-7.21 (dd, 1H, *J* = 4.9, 3.8 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 183.1, 144.2, 136.4, 135.3, 128.4. Condition I: GC conversion (99%, 9 h).

3-Thiophenecarboxaldehyde (**2I**)^{S4}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:30 v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ 9.94 (s, 1H), 8.13-8.12 (dd, 1H, *J* = 2.9, 1.2 Hz), 7.56-7.54 (dd, 1H, *J* = 5.1, 1.1 Hz), 7.39-7.37 (ddd, 1H, *J* = 5.1, 2.9, 0.9 Hz); ¹³C NMR (125 zMHz, CDCl₃) δ 185.1, 143.2, 136.8, 127.5, 125.5. Condition I: GC conversion (99%, 12 h).



1-Hexanal (**2m**)^{S4}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:40 v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ 9.76 (s, 1H), 2.43-2.40 (td, 2H, *J* = 7.4, 1.9 Hz), 1.66-1.61 (m, 2H), 1.35-1.29 (m, 4H) 0.91-0.88 (t, 3H, *J* = 7.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 203.2, 44.0, 31.5, 22.5, 21.9, 14.0. Condition I: GC conversion (99%, 15 h).



2-Heptanone (**2n**)^{S4}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:40 v/v) as the eluent, obtained as a colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ 2.43-2.40 (t, 3H, *J* = 7.5 Hz), 2.13 (s, 3H), 1.60-1.54 (quint, 2H, *J* = 7.6 Hz) 1.32-1.24 (m, 4H), 0.90-0.87 (t, 3H, *J* = 7.3 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 209.6, 43.9, 34.5, 30.0, 23.7, 22.6, 14.1. Condition I: GC conversion (99%, 18 h).



Diphenyl ketone (**2o**)^{S3}: Desire product was isolated by flash column purification with ethyl acetatehexane (1:40 v/v) as the eluent, obtained as a colorless solid. ¹H NMR (400 MHz, CDCl₃) δ 7.82-7.80 (m, 4H), 7.59-7.57 (m, 2H), 7.51-7.47 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 196.9, 137.7, 132.6, 130.2, 128.4.

Condition I: GC conversion (99%, 11 h).

Condition II: GC conversion (99%, 11 h).

Condition III: GC conversion (49%, 18 h).

Condition IV: GC conversion (48%, 18 h).



1-Pyrenecarboxaldehyde (**2p**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:40 v/v) as the eluent, obtained as a light yellow solid. ¹H NMR (500 MHz, CDCl₃) δ 10.62 (s, 1H), 9.18-9.16 (d, 1H, *J* = 9.3 Hz), 8.21-8.20 (d, 1H, *J* = 7.9 Hz), 8.15-8.13 (d, 1H, *J* = 7.6 Hz), 8.09-8.07 (d, 1H, *J* = 9.25 Hz), 8.04-8.02 (d, 1H, J = 8.9 Hz), 8.00-7.96 (m, 2H), 7.87-7.85 (d, 1H, J = 8.9 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 192.9, 135.2, 131.0, 130.8, 130.7, 130.5, 130.5, 130.2, 127.1, 127.0, 126.9, 126.7, 126.4, 124.3, 124.3, 123.8, 122.7.

Condition I: GC conversion (99%, 11 h).

Condition II: GC conversion (68%, 18 h).

Condition III: GC conversion (42%, 18 h).

Condition IV: GC conversion (45%, 18 h).



phenyl(pyren-1-yl)methanone (**2q**)^{S3}: Desire product was isolated by flash column purification with ethyl acetate-hexane (1:40 v/v) as the eluent. Isolation yield (85%, 18 h), obtained as a light yellow solid. ¹H NMR (500 MHz, DMSO) δ 8.40-8.09 (m, 9H), 7.81-7.79 (dd, 2H, *J* = 8.1, 1.3 Hz), 7.72-7.69 (dt, 1H, *J* = 8.6, 1.1 Hz), 7.58-7.54 (t, 2H, *J* = 7.9 Hz), ¹³C NMR (125 MHz, CDCl₃) δ 197.6, 138.1, 133.7, 132.9, 132.5, 130.8, 130.1, 130.1, 129.1, 128.9, 128.9, 128.7, 127.3, 126.9, 126.7, 126.4, 126.1, 124.2, 124.2, 123.9, 123.6.

Condition I: Isolation yield (85%, 65 mg, 18 h), obtained as a light yellow solid. Condition II: Isolation yield (60%, 46 mg, 18 h), obtained as a light yellow solid. Condition III: Isolation yield (30%, 23 mg, 18 h), obtained as a light yellow solid. Condition IV: Isolation yield (28%, 21 mg, 18 h), obtained as a light yellow solid.

Table S1. Crystal data and structure refinement for $Eu(Me-BPY)_2(NO_3)_3$.

Identification code	Eu(Me-BPY) ₂ (NO ₃) ₃	
Empirical formula	C ₂₄ H ₂₄ EuN ₇ O ₉	
Formula weight	706.47	
Temperature	293(2) K	
Wavelength	0.71073Å	
Crystal system	Orthorhombic	
Space group	Pbcn	
Unit cell dimensions	a = 16.643(3) Å	α = 90° .
	b = 9.4200(19)Å	β = 90°.
	c = 17.491(4)Å	γ = 90°.
Volume	2742.2(9)Å ³	
Z	4	
Density (calculated)	1.745Mg/m ³	
Absorption coefficient	2.354mm ⁻¹	
F(000)	1436	
Crystal size	0.3 x 0.2 x 0.1mm ³	
Theta range for data collection	3.379 to 27.497°.	
Index ranges	-21<=h<=21,-12<=k<=12, -	22<= <=22
Reflections collected	25638	
Independent reflections	3154[R(int) = 0.0642]	
Completeness to theta= 25.242°	99.8 %	
Absorption correction	Semi-empirical from equiva	alents
Max. and min. transmission	0.790 and 0.574	
Refinement method	Full-matrixleast-squares or	n F ²
Data / restraints / parameters	3154 / 0 / 189	
Goodness-of-fit on F ²	1.008	
Final R indices [I>2sigma(I)]	R1 = 0.0357, wR2 = 0.0986	6
R indices (all data)	R1 = 0.0579, wR2 = 0.122	7
Extinction coefficient	n/a	
Largest diff. peak and hole	1.020 and -1.123 e.Å ⁻³	

Table S2. Calculated BET surface area, pore volume, and pore size of UiO-67-bpy, UiO-67-bpy-Eu, UiO-67-(bpy&TEMPO), and UiO-67-(bpy-Eu&TEMPO).

Entry	MOF	BET surface area (m²/g)	Pore volume (cm ³ /g)	NLDFT pore size (nm)
1	UiO-67-bpy	1828	0.96	1.12
2	UiO-67-bpy-Eu	671	0.36	1.05
3	UiO-67-(bpy&TEMPO)	1372	0.73	1.12
4	UiO-67-(bpy-Eu&TEMPO)	609	0.33	0.97

 Table S3.
 Molecular formula and ICP-OES data of UiO-67-bpy-Eu and UiO-67-(bpy-Eu&TEMPO).

Entry	MOF	Molecular formula of MOFs from ideal structures	ICP-OES element	Amount (ppm [µmol])
1	UiO-67-bpy-Eu	Zr ₆ O ₄ (bpydc-Eu) _{4.5} (bpydc) _{1.5}	Zr	113050 [1239]
			Eu	144710 [952]
2	UiO-67-(bpy-Eu&TEMPO)	Zr ₆ O ₄ (bpydc-Eu) _{2.5} (bpydc) _{1.1} (bpdc- TEMPO) _{2.4}	Zr	142672 [1564]
			Eu	99167 [653]



Fig. S1 PXRD of Eu-MOF-1, Eu-MOF-2, UiO-67-bpy and UiO-67-bpy-Eu.



Fig. S2 PL spectra of Eu species, Eu-based MOFs and Eu-loaded MOFs.



Fig. S3 X-ray structure of Eu(Me-BPY)₂(NO₃)₃.



Fig. S4 PXRD patterns and N2 full isotherms of Eu-loaded or/and TEMPO-functionalized UiO-67s



Fig. S5 IR spectra of UiO-67-bpy and UiO-67-Eu after metalation with $Eu(NO_3)_3$ to confirm the existence of NO_3^- in the framework (reference IR peaks of nitrate anion: 1410-1340 for antisymmetric stretch.^{S15}



Fig. S6 TGA of UiO-67-bpy (top, black line) and UiO-67-bpy-Eu (bottom, blue line).



Fig. S7 Reusable test and PXRD of recovered UiO-67-bpy-Eu after aerobic oxidation reaction.



Fig. S8 Hot filtration test for the aerobic oxidation of benzyl alcohol with UiO-67-bpy-Eu.



Fig. S9 TGA of UiO-67-(bpy&TEMPO) (middle, pink line) and UiO-67-(bpy-Eu&TEMPO) (bottom, purple line).



Fig. S10 ¹H NMR spectra of UiO-67-(bpy-Eu&TEMPO) after acid digestion.



Fig. S11 PXRD patterns of UiO-67-bpy (**B0**), UiO-67-bpy-Eu (**B**), UiO-67-TEMPO (**D**), UiO-67-(bpy&TEMPO) (**E**), and UiO-67-(bpy-Eu&TEMPO) (**E**) after exposure to a variety of chemicals.



Fig. S12 B3LYP/6-311G(d,p) optimized molecular geometry of substrates and molecular sizes.



Scheme S1. General mechanism for transition metal-catalyzed aerobic oxidation with TEMPO moiety

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Appendix

Spectral copies of ¹H and ¹³C NMR of the obtained compounds



4-Nitrobenzaldehyde (2b)







4-Chlorobenzaldehyde (2d)



S29

3-Chlorobenzaldehyde (2e)



2-Chlorobenzaldehyde (2f)



4-Methylbenzaldehyde (2g)



S32

4-Methoxybenzaldehyde (2h)



3-Methoxybenzaldehyde (2i)



S34

2-Methoxybenzaldehyde (2j)



S35

2-Thiophenecarboxaldehyde (2k)



3-Thiophenecarboxaldehyde (2I)





210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)



S39



S40





Phenyl pyrenyl ketone (2q)

