Supporting Information

Light-Induced Self-Assembly of Dithienylethene Bolaamphiphiles in water

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Scheme S1. Synthesis of monomers 1 and 2. (a) *N*-chlorosuccinimide, benzene/acetic acid (1:1), reflux, 70%; (b) glutaryl chloride, AlCl₃, CH₃NO₂, 54%; (c) TiCl₄-Zn, pyridine-THF, reflux, 72%; (d) i. *n*-C₄H₉Li, THF; ii. CO₂, 85%; (e) *tert*-butyl(2-aminoethyl)carbamate, HOBT, HBTU, DIPEA, CH₃CN, 47%; (f) CF₃CO₂H, CH₂Cl₂, 99%; (g) Fmoc-Lys(Boc)-OH or Fmoc-Lys-OCH₃; HOBT, HBTU, DIPEA, CH₃CN, 59% (8), 90% (9); (h) CF₃CO₂H, CH₂Cl₂, 89%; (i) (CH₃)₃SnOH, 1M CaCl₂ in 30% water/*i*-PrOH. 35% (2); 26% (10).

1. Experimental procedures

2-chloro-5-methyl thiophene



This compound was prepared as described in literature.¹ 2-Methyl thiophene (207 mmol, 20 mL, 1 eq) and *N*-chlorosuccinimide (227 mmol, 30 g, 1.1 eq) were added to a rigorously stirred mixture of benzene (125 mL) and glacial acetic acid (125 mL). This mixture was heated at reflux for 7 h, continuing to stir vigorously. The mixture was cooled to room temperature, then poured into ice-water (300 mL). Extracted with DCM (3×100 mL). The organic layers were combined, dried over Na₂SO₄, filtered, then purified by distillation to isolate a slightly brown liquid (143 mmol, 70%). ¹H NMR (400 MHz, CDCl₃): δ 6.58 (d, =CH, *J* = 3.6 Hz, 1H), 6.44 – 6.39 (m, =CH, 1H), 2.30 (s, CH₃, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃) δ 138.4, 126.4, 125.7, 124.3, 15.2 ppm. Data is in accordance with literature.¹

1,5-Bis(5-chloro-2-methylthiophen-3-yl)pentane-1,5-dione (3)



This compound was prepared as described in literature.¹ 2-chloro-5-methyl thiophene (30 mmol, 2.1 eq) and glutaryl dichloride (15 mmol, 1.8 mL, 1 eq) were added to dry MeNO₂ (120 mL) under N₂, then cooled to 0°C. AlCl₃ (30 mmol, 3.8 g, 2 eq) was carefully added to the mixture, and then it was stirred at room temperature for 4 h. The reaction mixture was poured into ice-water (100 mL). Crude product was extracted with CH₂Cl₂ (3 × 75 mL), dried over Na₂SO₄, filtered, and then concentrated in vacuo. Crude product was isolated with flash chromatography (10% EtOAc, Hexanes) to obtain a white solid of **3** (2.79 g, 7.7 mmol, 54%). ¹**H NMR** (400 MHz, CDCl₃): δ 7.19 (s, =CH, 1H), 2.86 (t, *J* = 7.5 Hz, CH₂, 4H), 2.67 (s, CH₃, 6H), 2.01 (quint, *J* = 7.5 Hz, CH₂, 2H) ppm. ¹³**C NMR** (100 MHz, CDCl₃): δ 194.7, 147.6, 134.7, 126.7, 125.2, 40.4, 18.1, 16.0 ppm. Data is in accordance with literature.¹

1,2-bis(5-chloro-2-methylthiophen-3-yl)cyclopent-1-ene(4)



This compound was prepared as described in literature¹ with modification.² With all glassware flame-dried and in N₂ environment, TiCl₄ (4.2 mL, 4.2 mmol, 1M/toluene, 1.5 eq) was added carefully to 0°C freshlydistilled THF (20 mL) whereby a suspension formed. Zinc powder (0.55 g, 8.4 mmol, 3 eq) and dry pyridine (0.11 mL, 1.4 mmol, 0.5 eq) were added and the mixture brought to reflux (80°C) for 1 h. A solution of **3** (1.0 g, 2.8 mmol, 1 eq) in dry THF (3 mL) was added dropwise to the mixture and heated at reflux (80°C) for 5 h. The mixture was cooled to rt, added saturated K₂CO₃ solution (several drops) to quench, and filtered reaction over a celite cake while washing with EtOAc. Organic solvent was dried with Na₂SO₄, concentrated in vacuo, and crude product was purified with flash chromatographed (hexanes, 100%) to isolate a white crystalline solid (0.66 g, 2.0 mmol, 72% yield) ¹**H NMR** (400 MHz, CDCl₃): δ 6.57 (s, =CH, 2H), 2.72 (t, J = 7.5 Hz, CH₂, 4H), 2.02 (quint, J = 7.5 Hz, CH₂, 2H), 1.89 (s, CH₃, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 135.8, 134.4, 133.3, 126.7, 125.2, 38.2, 22.8, 14.2 ppm. Data is in accordance with literature.¹

4,4'-(Cyclopent-1-ene-1,2-diyl)bis(5-methylthiophene-2-carboxylic acid) (5)



This compound was prepared as described in literature.¹ In a round-bottomed flask equipped with a stirring bar, **4** (2.0 g, 6.0 mmol, 1.0 equiv) was dissolved in dry THF (60 mL). Then *n*-BuLi (6.1 mL, 2.5 M in hexanes, 2.5 equiv) was added dropwise and the black mixture was stirred for 1 h. Solid CO₂ (dry ice, excess) was added, after which a brown suspension was formed which was stirred for 30 min. Water (60

mL) was added slowly and the aqueous layer was washed with Et₂O (60 mL) and subsequently acidified to pH = 1 with HCl (1M). After extraction with CH₂Cl₂ (3 × 60 mL), the combined organic layers were dried over Na₂SO₄, filtered and concentrated to yield the title compound as a brown solid (1.8 g, 85%). The product was used without further purification. ¹H NMR (400 MHz, DMSO-d₆): δ 12.94 (bs, COOH, 2H), 7.42 (s, =CH, 2H), 2.78 (t, *J* = 7.5 Hz, CH₂, 4H) 2.02 (quint, *J* = 7.5 Hz, CH₂, 2H), 1.93 (s, CH₃, 6H) ppm. ¹³C NMR (100 MHz, DMSO-d₆): δ 162.6, 141.7, 136.4, 134.3, 133.8, 130.4, 37.8, 22.3, 14.3 ppm. Data is in accordance with literature.¹

tert-Butyl (2-aminoethyl)carbamate

tert-Butyl (2-aminoethyl)carbamate was prepared as described in literature.³ A solution of di-*tert*-butyl dicarbonate (5.0 g, 23 mmol) in 1,4-dioxane (7 mL) was added dropwise over 4 h min to a solution of ethane-1,2-diamine (4.6 mL, 69 mmol) in 1,4-dioxane (30 mL). A white precipitate formed slowly as the reaction was stirred at rt. After 19 h, the mixture was quenched with K₂CO₃ solution, and then extracted with DCM (3×100 mL). The combined organic layers were washed once with NaHCO₃ (saturated), dried with Na₂SO₄, then concentrated to yield *tert*-butyl (2-aminoethyl)carbamate as a white solid (3.20 g, 87%). ¹H NMR (400 MHz, CDCl₃): δ 4.93 (bs, NH, 1H), 3.17 (q, *J* = 5.9 Hz, CH₂, 2H), 2.80 (t, *J* = 5.9 Hz, CH₂, 2H), 1.45 (s, CH₃, 6H), 1.22 (bs, NH₂, 2H) ppm. ¹³C-NMR (100 MHz, CDCl₃): δ 156.2, 79.2, 43.5, 41.9, 28.4 ppm. Data is in accordance with literature.³

Di-*t*-butyl (((4,4'-(cyclopent-1-ene-1,2-diyl)bis(5-methylthiophene-4,2-diyl-2-carbonyl))bis(azanediyl))bis(ethane-2,1-diyl))dicarbamate (6)



A solution of *tert*-butyl (2-aminoethyl)carbamate (290 mg, 1.80 mmol) in dry MeCN (5 mL) was added to a solution of HOBt (231 mg, 1.70 mmol), HBTU (653 mg, 1.70 mmol), DTE-diacid (**5**) (300 mg, 0.90 mmol), and DIEA (0.60 mL, 3.40 mmol) in dry MeCN (15 mL) under N₂ atmosphere. The reaction was stirred at room temperature for 12 h and then concentrated in vacuo. The residue was dissolved in EtOAc (100 mL) and washed with NaHCO₃ (saturated) (2 × 30 mL), citric acid (10%; 2 × 30 mL). The organic solution was dried (Na₂SO₄), filtered, and concentrated in vacuo. The residue was purified by silica gel chromatography (methanol: DCM; 3:97) to give the title compound as a pink solid (258 mg, 47% yield). M.p.: 123–124 °C. (acetone/hexane) ¹**H NMR** (400 MHz, CD₂Cl₂): δ 7.14 (s, =CH, 2H), 7.02 (bs, NH, 2H), 5.20 (bs, NH, 2H), 3.34 (dd, *J* = 10.9, 5.4 Hz, CH₂, 4H), 3.22 (dd, *J* = 10.9, 5.4 Hz, CH₂, 4H), 2.66 (t, *J* = 7.5 Hz, CH₂, 4H), 1.99 – 1.90 (quint, *J* = 7.5 Hz, CH₂, 2H), 1.83 (s, CH₃, 6H), 1.33 (s, CH₃, 18H) ppm. ¹³C **NMR** (100 MHz, CD₂Cl₂): δ 162.8, 157.7, 140.5, 136.8, 135.2, 134.9, 129.6, 79.9, 41.9, 40.5, 38.8, 28.5, 23.3, 14.8 ppm. **ESI-MS** calcd. for C₃₁H₄₄N₄NaO₆S₂: [M+Na]⁺: 655.2600, found: 655.2610. 4,4'-(Cyclopent-1-ene-1,2-diyl)bis(N-(2-aminoethyl)-5-methylthiophene-2-carboxamide) (7)



TFA (4 mL) was added to a solution of **7** (324 mg, 0.5 mmol) at room temperature in CH₂Cl₂ (4 mL). The reaction mixture was stirred at rt for 2 h. TFA was removed with addition of diethyl ether (3 × 25 mL) to the residue (red oil) followed by solvent removal in vacuo. The final product was obtained as a light red oil (220 mg, 99% yield). ¹H NMR (250 MHz, CD₃OD): δ 7.39 (s, =CH, 2H), 3.50 (t, *J* = 5.9 Hz, CH₂, 4H), 3.04 (t, *J* = 5.9 Hz, CH₂, 4H), 2.72 (t, *J* = 7.4 Hz, CH₂, 4H), 2.07 – 1.93 (m, CH₂, 2H), 1.83 (s, CH₃, 6H) ppm. ¹³C NMR (100 MHz, DMSO-d₆): δ 162.6, 140.2, 137.1, 136.1, 135.1, 130.3, 80.1, 38.0, 31.7, 27.8, 15.2 ppm. ESI-MS calcd. for C₂₁H₂₉N₄O₂S₂: [M+H]⁺: 433.1732, found: 433.1754.

((9H-Fluoren-9-yl)methoxy)carbonyl)-L-lysine (Fmoc-Lys-OH)



This compound was prepared as described in literature.³ Fmoc-Lys(Boc)-OH (1.0 g, 2.10 mmol) was dissolved in a solution of TFA/CH₂Cl₂ (1:1, 50 mL) and stirred for 2 h at room temperature. Following evaporation and diethyl ether extraction of the residue, Fmoc-Lys-OH was obtained as a white powder (0.75 g, 95 % yield). ¹**H-NMR** (400 MHz, DMSO-d₆): δ 7.99 (d, *J* = 7.5 Hz, =CH, 2H), 7.76-7.71 (m, =CH, 2H), 7.62 (d, *J* = 8.1 Hz, NH, 1H), 7.42 (t, *J* = 7.5 Hz, =CH, 2H), 7.33 (t, *J* = 7.5 Hz, =CH, 2H), 4.34-4.26 (m, CH₂, 2H), 4.23 (t, *J* = 6.5 Hz, CH, 1H), 3.97-3.91 (m, CH, 1H), 2.78 (dd, *J* = 12.6, 6.5 Hz, CH₂, 2H), 1.76-1.72 (m, CH₂, 2H), 1.64-1.51 (m, NH₂, CH₂, 4H), 1.39-1.35 (m, CH₂, 2H); ¹³C-NMR (100 MHz, DMSO-d₆): δ 173.8, 156.2, 143.8, 140.7, 127.6, 127.0, 125.2, 120.1, 65.6, 53.6, 46.6, 38.6, 30.1, 26.5, 22.5. Data is in accordance with literature.³

Methyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-lysinate (Fmoc-Lys-OMe)



This compound was prepared as described in literature.³ Thionyl chloride (0.5 mL, 5.1 mmol) was slowly added to a suspension of Fmoc-Lys-OH (500 mg, 1.4 mmol) in dry methanol (20 mL). The resulting clear solution was kept at room temperature for 2 h and then evaporated to dryness. Upon re-dissolving in methanol (10 mL) and dropwise addition into diethyl ether, the ester precipitated. A white solid (503 mg, 97 %) was obtained. Data is in accordance with literature.³ **¹H NMR** (400 MHz, DMSO-d₆): δ 7.91-7.87 (m, =CH, NH, 3H), 7.78-7.70 (m, =CH, 2H), 7.43 (t, *J* = 7.5 Hz, =CH, 2H), 7.34 (t, *J* = 7.5 Hz, =CH, 2H), 4.38 – 4.26 (m, CH₂, 2H), 4.23 (t, *J* = 6.8 Hz, CH, 1H), 4.05 – 3.97 (m, CH, 1H), 3.63 (s, CH₃, 3H), 2.75 (bs, NH₂, 2H), 1.76 – 1.48 (m, CH₂, 6H), 1.43 – 1.27 (m, CH₂, 2H); ¹³C-NMR (100 MHz, DMSO-d₆): 172.8, 156.1, 143.8, 140.7, 127.6, 127.0, 125.2, 120.1, 66.6, 53.6, 51.9, 46.6, 38.4, 30.0, 26.4, 22.4.

Fmoc-N-BocLys-DTE-N-BocLys-Fmoc (8)



A solution of **7** (209 mg, 0.50 mmol) in dry THF (5 mL) was added to a solution of HOBt (131 mg, 1.0 mmol), HBTU (379 mg, 1.0 mmol), Fmoc-Lys(Boc)-OH (453 mg, 1.0 mmol), and DIEA (0.60 mL, 3.40 mmol) in dry MeCN under and N₂ atmosphere. The reaction was stirred at room temperature for 12 h then concentrated in vacuo. The residue was dissolved in EtOAc (100 mL) and washed with NaHCO₃ (saturated) solution (2 × 30 mL), citric acid (10 %; 2 × 30 mL). The organic solution was dried (Na₂SO₄), filtered, and concentrated in vacuo. The residue was purified by silica gel chromatography (3% MeOH in DCM) to give **8** as a pink solid (379 mg, 59 % yield). M.p.: 156–157 °C. (acetone/hexane). ¹**H NMR** (400 MHz, CDCl₃): δ 7.73 (d, *J* = 7.3 Hz, =CH, 4H), 7.56 (d, *J* = 7.3 Hz, =CH, 4H), 7.36 (t, *J* = 7.4 Hz, =CH, 4H), 7.30 – 7.23 (m, =CH, 6H), 7.17 (bs, NH, 2H), 6.09 (bs, NH, 2H), 4.87 (bs, NH, 2H), 4.41 – 4.07 (m, NH, CH, CH₂, 8H), 3.56 – 3.35 (m, CH₂, 8H), 3.03 – 3.02 (m, CH₂, 4H), 2.64 – 2.61 (m, CH₂, 4H), 1.96 (s, CH₃, 6H), 1.94 – 1.84 (m, CH₂, 2H), 1.83 – 1.50 (m, CH, CH₂, 8H), 1.41 (s, CH₃, 18H), 1.37 – 1.30 (m, CH₂, 4H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 173.4, 162.7, 156.5, 156.3, 143.8, 141.2, 140.3, 136.3, 135.0, 134.0, 130.0, 127.7, 127.1, 125.1, 120.0, 79.2, 67.1, 55.2, 54.0, 47.1, 42.2, 37.9, 28.4, 22.9, 22.7, 18.5, 17.4, 14.6, 11.9 ppm. **ESI-MS** calcd. for C₇₃H₈₈N₈NaO₁₂S₂: [M+Na]⁺: 1355.5861, found: 1355.5884.

Fmoc-Lys-DTE-Lys-Fmoc (1)



To a solution of **8** (330 mg, 0.3 mmol) in CH₂Cl₂ (10 mL), was added TFA (10 mL) at rt. The reaction mixture was stirred at rt for 2 h. The solvent was removed under reduced pressure and cold diethyl ether (25 mL) was added to the red oil. A red precipitate formed and was collected by centrifugation. The crude sediment was dissolved in water and dried by lyophilization, then further purified by reversed-phase HPLC to give **1** as light red solid (249 mg, 89 %). M.p.: 138–140 °C. (methanol). ¹H NMR (500 MHz, CD₃OD): δ 7.80 (d, *J* = 7.5 Hz, =CH, 4H), 7.67 – 7.64 (m, =CH, 4H), 7.43 (s, =CH, 2H), 7.40 (t, *J* = 7.5 Hz, =CH, 4H), 7.31 (t, *J* = 7.5 Hz, =CH, 4H), 4.45 – 4.33 (m, CH₂, 4H), 4.21 (t, *J* = 6.6 Hz, CH, 2H), 4.08 – 4.00 (m, CH, 2H), 3.55 – 3.37 (m, CH₂, 8H), 2.93 (t, *J* = 6.6 Hz, CH₂, 4H), 2.72 – 2.69 (m, CH₂, 4H), 2.04 – 1.92 (m, CH₂, 2H), 1.84 (s, CH₃, 6H), 1.73 – 1.62 (m, CH₂, 8H), 1.52 – 1.37 (m, CH₂, 4H) ppm. ¹³C NMR (100 MHz, CD₃OD): δ 175.3, 164.7, 145.4, 145.3, 142.8, 141.8, 138.0, 136.3, 135.9, 131.1, 129.0, 128.3, 126.3, 121.1, 68.1, 56.6, 48.6, 40.6, 40.6, 40.4, 39.6, 32.5, 28.3, 24.0, 23.9, 14.9 ppm. ESI-MS calcd. for C₆₃H₇₃N₈O₈S₂: [M+H]⁺: 1133.4993, found: 1133.4987.



¹**H** NMR (400 MHz, CD₃OD): δ 7.82 (d, J = 7.5 Hz, =CH, 4H), 7.69 (t, J = 7.5, =CH, 4H), 7.43-7.40 (m, =CH, 4H), 7.33 (dd, J = 4.5, 1.0 Hz, =CH, 4H), 6.64 (d, J = 2.8, =CH, 2H), 4.48-4.37 (m, CH₂, 4H), 4.25 (t, J = 6.7, CH, 2H), 4.06 – 4.03 (m, CH, 2H), 3.52 – 3.36 (m, CH₂, 8H), 2.95 (t, J = 6.8, CH₂, 4H), 2.35 (t, J = 6.9, CH₂, 4H), 1.91-1.63 (m, CH₂, CH₃, 16H), 1.52-1.39 (m, CH₂, 4H) ppm. ¹³C NMR (100 MHz, CD₃OD): δ 173.8, 163.9, 163.2, 157.1, 143.9, 143.8, 141.8, 138.9, 127.5, 126.8, 124.8, 119.6, 66.6, 65.6, 55.1 (2C), 39.1, 31.0 (2C), 29.2, 26.7, 26.3, 24.6, 22.5, 13.3 ppm.

Fmoc-Lys-OMe-DTE-OMe-Lys_Fmoc (9)



A solution of Fmoc-Lys-OMe (439 mg, 1.20 mmol) in dry DMF (3 mL) was added to a solution of HOBt (155 mg, 1.20 mmol), HBTU (435 mg, 1.20 mmol), DTE-diacid (200 mg, 0.60 mmol), and DIEA (0.40 mL, 2.30 mmol) in dry THF (15 mL) under N₂ atmosphere. The reaction was stirred at room temperature overnight and then concentrated in vacuo. The residue was dissolved in EtOAc (100 mL) and washed with NaHCO₃ (saturated) solution (2×30 mL), citric acid (10 %; 2×30 mL). The organic solution was dried over MgSO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel chromatography (Acetone/hexane, 3:7) to give title compound as a light pink solid (555 mg, 90 % yield). M.p.: 137–138 °C. (acetone/hexane)

¹**H** NMR (400 MHz, Acetone-d₆): δ 7.86 (d, J = 7.5 Hz, =CH, 4H), 7.71 (d, J = 7.5 Hz, =CH, 4H), 7.62 (t, J = 5.6 Hz, NH, 2H), 7.46 (s, =CH, 2H), 7.41 (t, J = 7.5 Hz, =CH, 4H), 7.35 – 7.29 (m, =CH, 4H), 6.88 (d, J = 8.2 Hz, NH, 2H), 4.39 – 4.19 (m, CH, CH₂, 6H), 3.68 (s, CH₃, 6H), 3.38 – 3.33 (m, CH₂, 4H), 2.73 (t, J = 7.4 Hz, 4H), 2.01 – 1.94 (m, CH₂, 2H), 1.89 (s, CH₃, 6H), 1.86 – 1.71 (m, CH, CH₂, 6H), 1.67 – 1.41 (m, CH₂, 8H) ¹³C NMR (100 MHz, Acetone-d₆): δ 173.7, 162.2, 157.1, 145.1, 142.1, 140.1, 137.2, 137.0, 135.5, 129.4, 128.6, 128.0, 126.2, 120.8, 67.2, 55.0, 52.3, 48.0, 39.9, 39.7, 32.0, 30.0, 23.9, 23.5, 14.7. **ESI-MS** calcd. for C₆₁H₆₄N₄NaO₁₀S₂: [M+Na]⁺: 1099.3962, found: 1099.3956.

Fmoc-Lys-OMe-DTE-OH-Lys_Fmoc (10) and Fmoc-Lys-DTE-Lys and Lys-DTE-Lys (2)

Trimethyltin chloride (370 mg, 1.9 mmol) was dissolved in MeOH (5 mL) treated with KOH (104 mg, 1.9 mmol) was added to the solution. Then, the mixture was stirred for 1 h at ambient temperature at which time potassium chloride precipitated as a white solid. The reaction mixture was subsequently filtered and the filtrate containing (CH₃)₃SnOH was evaporated under vacuum. The solid (CH₃)₃SnOH was added to a mixture of CaCl₂ in water and *i*-PrOH (20 mL) and. Fmoc-Lys-OMe-DTE-OMe-Lys-Fmoc (**9**; 100 mg, 0.1 mmol) was added to the (CH₃)₃SnOH solution was stirred for 12 h at 100°C. The solvent was evaporated in vacuo and the crude residue was dissolved in water: CH₃CN (1:1) (40 mL) and purified by reversed-phase HPLC on preparative Varian Dynamax C18 column eluting with a linear gradient of acetonitrile/water with 0.1 % TFA (20/80 to 90/10 over 7 h) producing three products were obtained: Lys-DTE-Lys (**2**) (20 mg, 35 %), Fmoc-Lys-DTE-Lys (10 mg, 13 %) and Fmoc-Lys-DTE-Lys-Fmoc (**10**) (25 mg, 26 %) were obtained as white solids. M.p. for **10**: 135–136 °C. (acetone/hexane), M.p. for **2**: 125–126 °C. (acetone/hexane).

Fmoc-Lys-DTE-Lys-Fmoc (10)



¹**H** NMR (400 MHz, CDCl₃/CD₃OD (1/1)): δ 7.96 – 7.92 (m, =CH, 4H), 7.81 – 7.74 (m, =CH, NH, 8H), 7.59 – 7.52 (m, =CH, 4H), 7.50 – 7.45 (m, =CH, 4H), 4.62 – 4.35 (m, CH, CH₂, 10H), 3.00 – 2.87 (m, CH₂, 4H), 2.28 – 2.15 (m, CH₂, 4H), 2.06 (s, CH₃, 6H), 1.99 – 1.36 (m, CH₂, 12H); ¹³C-NMR (100 MHz, CDCl₃/CD₃OD (1/1)): δ 162.5, 156.6, 143.4, 143.3, 140.8, 139.7, 136.0, 134.2, 134.0, 129.0, 127.0, 126.5, 124.5, 119.3, 66.4, 53.3, 46.6, 38.9, 37.9, 31.0, 28.4, 22.4, 22.2, 13.7. **ESI-MS** calcd. for C₅₉H₆₁N₄O₁₀S₂: [M+H]⁺: 1049.3829, found: 1049.3824.

Lys-DTE-Lys (2)



¹**H** NMR (400 MHz, D₂O/CD₃OD (1/1)): δ 7.40 (s, =CH, 2H), 4.01 – 3.83 (m, CH, CH₂, 6H), 3.02 - 2.95 (m, CH₂, 4H), 2.87 - 2.72 (m, CH₂, 4H), 2.11 - 1.89 (m, CH₂, 2H), 1.87 (s, CH₃, 6H), 1.71 - 1.36 (m, 8H); ¹³C-NMR (100 MHz, D₂O/CD₃OD (1/1)): δ 172.6, 164.7, 141.9, 138.1, 136.3, 135.7, 131.0, 54.1, 40.4, 39.6, 31.2, 30.1, 23.9, 23.4, 14.8. **ESI-MS** calcd. for C₂₉H₄₁N₄O₆S₂: [M+H]⁺: 605.2468, found: 605.2256.

2. Circular Dichroism (CD) and Ultraviolet-Visible (UV-Vis) Spectroscopy.

CD spectra were recorded on a JASCO spectrometer under a nitrogen atmosphere. Experiments were performed in a quartz cell with a 1 mm or 1 cm path length over the range of 190-650 nm at 25 °C. UV spectra were recorded on a Shimadzu UV-3200 spectrometer under an atmospheric condition. Experiments were performed in a quartz cell with a 1 mm or 1 cm path length over the range of 190-650 nm at 25 °C.



Figure S1: Photochromic behavior of **1**. 3 mM stock solutions of **1-open** in **A**) water, **B**) methanol, **C**) THF, and **D**) DMF were irradiated with 254 nm UV light over time, then dilute to 0.3 mM prior to analysis by UV-Vis. **E**) Comparison of all four solvents at the PSS (40 min in 254 nm UV light). UV-Vis spectroscopy was conducted on a Shimadzu UV-2450 Spectrometer with a TCC-240A temperature-controlled cell holder using a 1 mm path length quartz cuvette.



Figure S2. Photochromic behavior of **2**. UV/Vis absorption spectra in water (1 mM) during the course of irradiation at 254 nm from the open state to the photostationary state. Inset: time-dependent absorption changes at 524 nm.



Figure S3: **A)** CD Spectra of **1** in water (1 mM) (solid-open, dash-closed). **B) 1-closed** extended UV-Vis and CD spectra showing a lack of CD signal correlating with DTE ring-closed form.

3. Photoisomerization

Photoisomerization of **1** was performed in quartz cuvettes by irradiation with 254 nm light in a Rayonet UV Chamber. The open:closed ratio of **1** was measured by ¹H NMR (in CD₃OD) by the ratio of the protons (4H, $2 \times -CH_2$ - (cyclopentenyl) at ~2.35 ppm (closed-form) and 2.70 ppm (open-form). Samples were prepared by evaporation of the isomerization solvent and dissolution in CD₃OD for NMR analysis. The open/closed ratios were best determined by analytical reverse-phase HPLC, which provided more accurate ratios due to the ability to minimize sample handling prior to analysis.



Figure S4: ¹H NMR of **A**) (blue arrow) **1-open** and (red arrow) **1-closed** irradiated with 254 nm UV light (45 min) in water (switched to CD_3OD for ¹H NMR). Reverse-phase analytical HPLC of **B**) **1-open** prior to irradiation and **C**) **1**, after irradiation at 254 nm at the PSS (closed:open 4:1).



Figure S5: **A**) ¹H NMR of (blue arrow) **1-open** and (red arrow) **1-closed** via 254 nm UV light (45 min) in CD₃OD. **B**) Reverse-phase analytical HPLC of **1** at the PSS after irradiation at 254 nm light (45 min) in CD₃OD [closed:open = 18:1].

4. Atomic Force Microscopy (AFM)

AFM images were collected on a NanoScope IIIa device at ambient temperature in tapping mode using silicon tips (NSC14/AIBS, MikroMasch). 10 μ L of the sample solution (250 μ M) was diluted and placed on freshly cleaved mica. After adsorption for 30 min under moist conditions, excess solution was removed by absorption onto filter paper. The sample was allowed to dry fully before being scanned. The scanning speed was at a line frequency of 1.0 Hz, and the original images were sampled at a resolution of 512 x 512 pixels.

5. Transmission Electron Microscopy Measurement (TEM)

Using clean parafilm for aqueous samples, a copper grid (200 mesh) was floated on 50 uL drops of diluted samples for 3-5 min. After removal of the excess solution with filter paper by tapping the edge of the grid to a chem-wipe, the grid was floated on 50 μ L drops of 2 % wt uranyl acetate solution for negative stain for 30 seconds. The excess solution was removed by tapping the edge of the grid to a chem-wipe. The dried specimen was observed with the Technai G2 Spirit instrument operating at 80 kV. The data were analyzed with Image-Pro Software.



Figure S6: TEM images of aqueous solutions (10 mM) of **1-open: A**) after 24 h and **B**) 24 h after sonication (20% 1 min, pulsed 10s on, 5s off) and of **1-closed**: **C**) after 24 h in darkness and **D**) 24 h (in darkness) after sonication (20% 5 min, pulsed 10s on, 5s off). TEM images were recorded after diluting in water (1 mM) and evaporating onto a carbon-coated copper grid with uranyl acetate as a negative stain. Fiber diameters were 11.9 nm \pm 2.3 nm (analyzed with ImageJ Pro Software).



Figure S7. TEM images of aqueous solutions (10 mM) of **1-closed** after irradiation in MeOH, then evaporated and dispersed in water. **A**) 24 h/darkness w/o ultrasonication, **B**) 10 min/darkness after 5 min ultrasonication (20% pulsed, 10s on, 5s off), and **C**) 24 h/darkness after 5 min ultrasonication (20% pulsed, 10s on, 5s off). TEM images were recorded after diluting in water (1 mM) and evaporating onto a carbon-coated copper grid with uranyl acetate as a negative stain.

¹H, ¹³C NMR and HRMS Spectra



15





















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