

A Novel Antimicrobial Technology to Enhance Food Safety and Quality of Leafy Vegetables using Engineered Water Nanostructures

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Fig. S1 Inactivation of *E. coli* on stainless steel coupons from various EWNS based nano-sanitizers synthesized with single and combined active ingredients.

Fig. S2 Inactivation of various microorganisms on stainless steel coupon from EWNS based nano-sanitizers synthesized with a combination of AIs (10% hydrogen peroxide, 1% citric acid, 0.1% lysozyme and 0.0025% nisin).

Table S1. Physio-chemical characterization of the EWNS based nano-sanitizer synthesized with 10% hydrogen peroxide, 1% citric acid, 0.1% lysozyme and 0.0025% nisin.

Fig. S1 Inactivation of *E. coli* on stainless steel coupons from various EWNS based nano-sanitizers synthesized with (a) single and (b) combined active ingredients. Microorganisms were inoculated on stainless steel coupon and exposed to the EWNS based nano-sanitizers synthesized with various single and combined active ingredients. The coupon was inoculated with $\sim 10^6$ CFU/coupon of *E. coli*. The coupons were then exposed to the EWNS based nano-sanitizers for 30 s as shown in Fig. 1. The temperature and relative humidity were ~ 24 °C and $\sim 40\%$, respectively. The error bar represents 1 standard deviation. (Contains data adapted with permission from Huang, R., Vaze, N., Soorneedi, A., Moore, M. D., Xue, Y., Bello, D., & Demokritou, P. (2019). Inactivation of Hand Hygiene-Related Pathogens Using Engineered Water Nanostructures. ACS Sustainable Chemistry & Engineering, 7(24), 19761-19769. Copyright 2019, American Chemical Society).

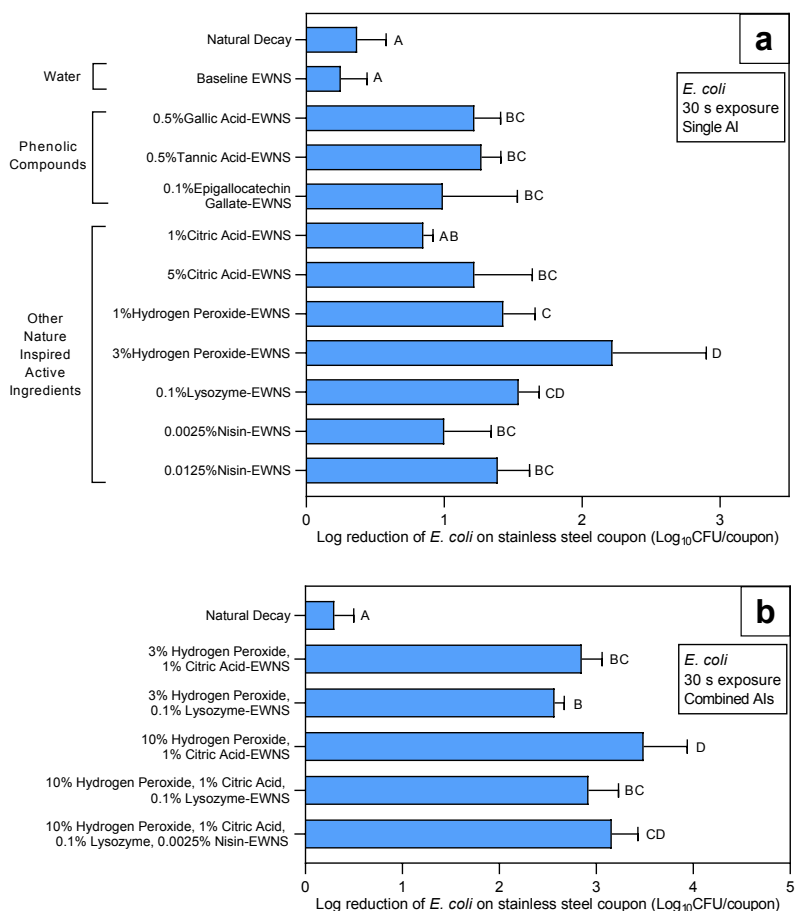


Fig. S2 Inactivation of various microorganisms on stainless steel coupon from EWNS based nano-sanitizers synthesized with an AI cocktail: (a): *Escherichia coli*, (b) *Listeria innocua*, (c) *Penicillium italicum* and (d) *Pseudomonas fluorescens*. Microorganisms were inoculated on stainless steel coupon and exposed to the EWNS based nano-sanitizers synthesized with a combination of AIs (10% hydrogen peroxide, 1% citric acid, 0.1% lysozyme and 0.0025% nisin). The coupon was inoculated with $\sim 10^6$ CFU/coupon of *E. coli*, $\sim 10^6$ CFU/coupon of *L. innocua*, $\sim 10^6$ PFU/coupon of bacteriophage MS2, $\sim 10^5$ CFU/coupon of *P. italicum* or $\sim 10^6$ CFU/coupon of *Ps. fluorescens*. The coupons were then exposed to the EWNS based nano-sanitizers for 0 – 15 min as shown in Fig. 1. The temperature and relative humidity were ~ 24 °C and $\sim 40\%$, respectively. The error bar represents 1 standard deviation.

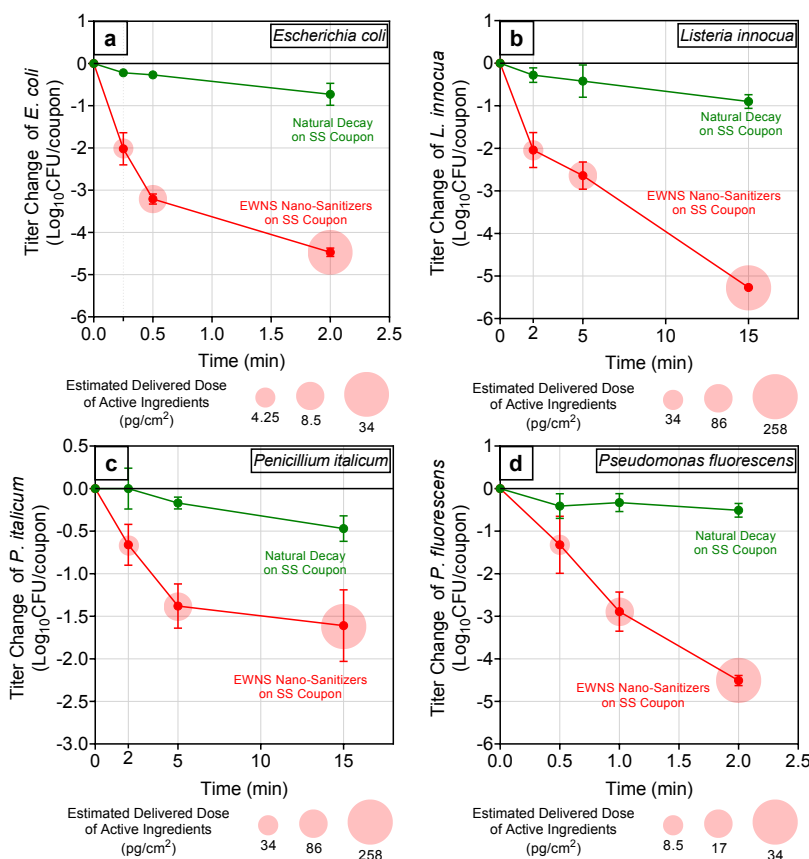


Table S1. Physio-chemical characterization of the EWNS based nano-sanitizer synthesized with 10% hydrogen peroxide, 1% citric acid, 0.1% lysozyme and 0.0025% nisin. (Adapted with permission from Huang, R., Vaze, N., Soorneedi, A., Moore, M. D., Xue, Y., Bello, D., & Demokritou, P. (2019). Inactivation of Hand Hygiene-Related Pathogens Using Engineered Water Nanostructures. ACS Sustainable Chemistry & Engineering, 7(24), 19761-19769.

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<i>Physical Characterization</i>		
SMPS-measured mean diameter (nm)		21.0 ± 2.6
CPC-measured concentration (number of particles/cm ³)		181090 ± 45371
Electric charge (e ⁻ /particle)		10.4 ± 1.6
<i>Chemical Characterization</i>		
	Estimated Concentration	Measured Concentration
Short-lived ROS (mol H ₂ O ₂ equivalent unit/particle)	N/A	$1.88 \pm 0.72 \times 10^{-14}$
Total ROS (mol H ₂ O ₂ equivalent unit/particle)	N/A	$3.41 \pm 0.33 \times 10^{-14}$
Citric acid (ng/particle)	$4.86 \pm 1.55 \times 10^{-11}$	$< 2.47 \pm 0.41 \times 10^{-9} *$

N/A: Not applicable.

*: 1 replicate was below the limit of detection out of a total of 3 replicates.

The detection limit of short-lived ROS and total ROS is 1.38×10^{-10} nmol H₂O₂ equivalent unit/particle.

The detection limit of citric acid is 1.84×10^{-9} ng/particle.

Details on the plaque assay utilized from Su and D'Souza:

E. coli B-15597 was grown in TSB containing 0.1% glucose, 2 mM CaCl₂, and 10 µg/ml thiamine for 6 h. MS2 treated with GSE or water after neutralization with TSB containing 3% beef extract was serially diluted in TSB, and 0.7 ml of diluted phage was mixed with 0.3 ml of 6-

h E. coli host. The 1-ml host-virus combination was then added to 8 ml of 0.6% molten top agar, mixed and poured on tryptic soy agar (TSA) bottom agar plates, and incubated at 37°C overnight before counting (Su & D'Souza, 2011).

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