

Supporting Information:

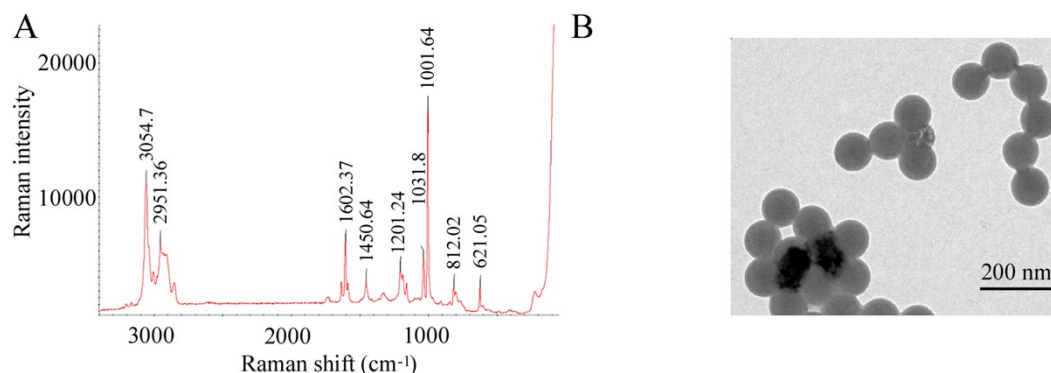


Figure S1. Properties of nanopolystyrene. (A) Raman spectroscopy of nanopolystyrene particles. The Raman spectroscopy analysis indicated that the nanopolystyrene showed the peaks at 1001.64 cm⁻¹ (breathing vibration of benzene ring), at 1031.8 cm⁻¹ (symmetric extension vibration of carbon atoms in benzene ring), at 1201.24 cm⁻¹ (stretching vibration of carbon atoms between benzene ring and polyethylene group), at 1450.64 cm⁻¹ (asymmetric bending vibration of hydrogen atoms), and at 1602.37 cm⁻¹ (asymmetric stretching vibration of benzene ring carbon atoms). (B) TEM image of nanopolystyrene particles in K medium before the sonication.

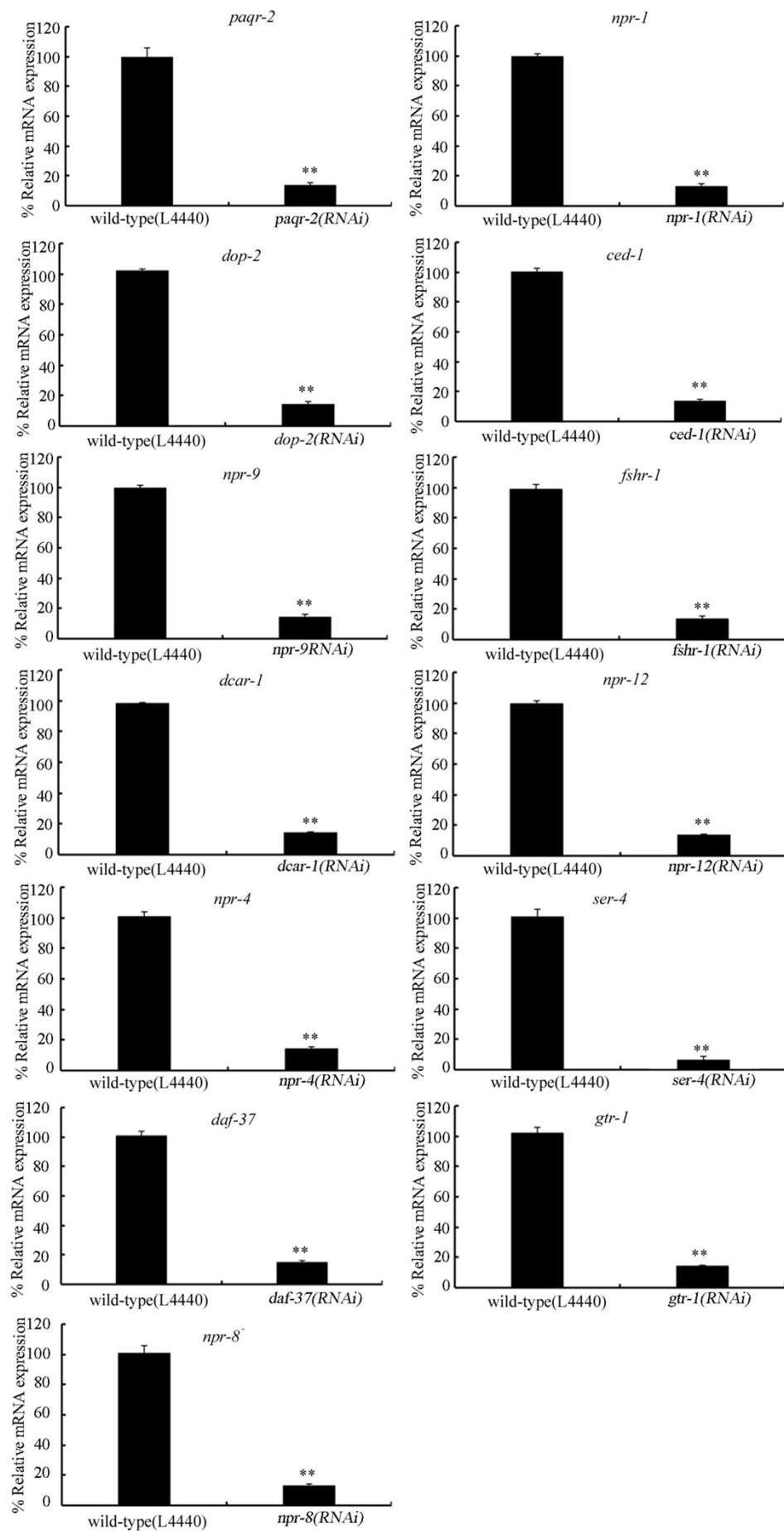


Figure S2. qRT-PCR analysis of the efficiency of RNAi knockdown of *paqr-2*, *npr-1*, *dop-2*, *ced-1*, *npr-9*, *fshr-1*, *dcar-1*, *npr-12*, *npr-4*, *ser-4*, *daf-37*, *gtr-1*, and *npr-8* in wild-type nematodes. L4440, empty vector. Bars represent means $\pm$ SD. $**P < 0.01$ vs wild-type.

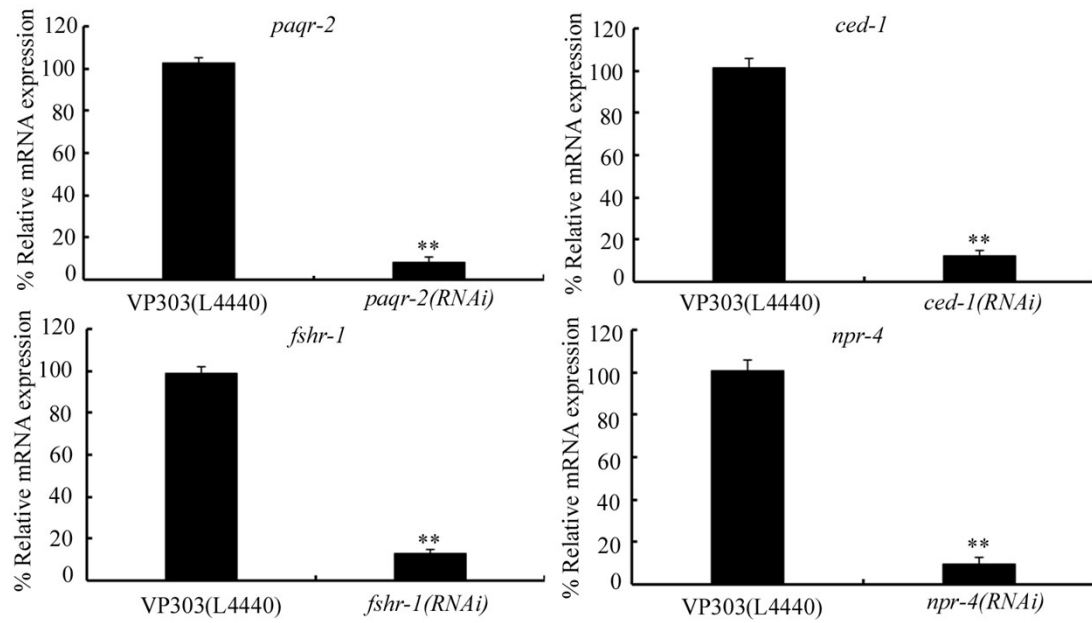


Figure S3. qRT-PCR analysis of the efficiency of intestine-specific RNAi knockdown of *paqr-2*, *ced-1*, *fshr-1*, and *npr-4*. L4440, empty vector. Bars represent means $\pm$ SD. ** $P < 0.01$ vs VP303.

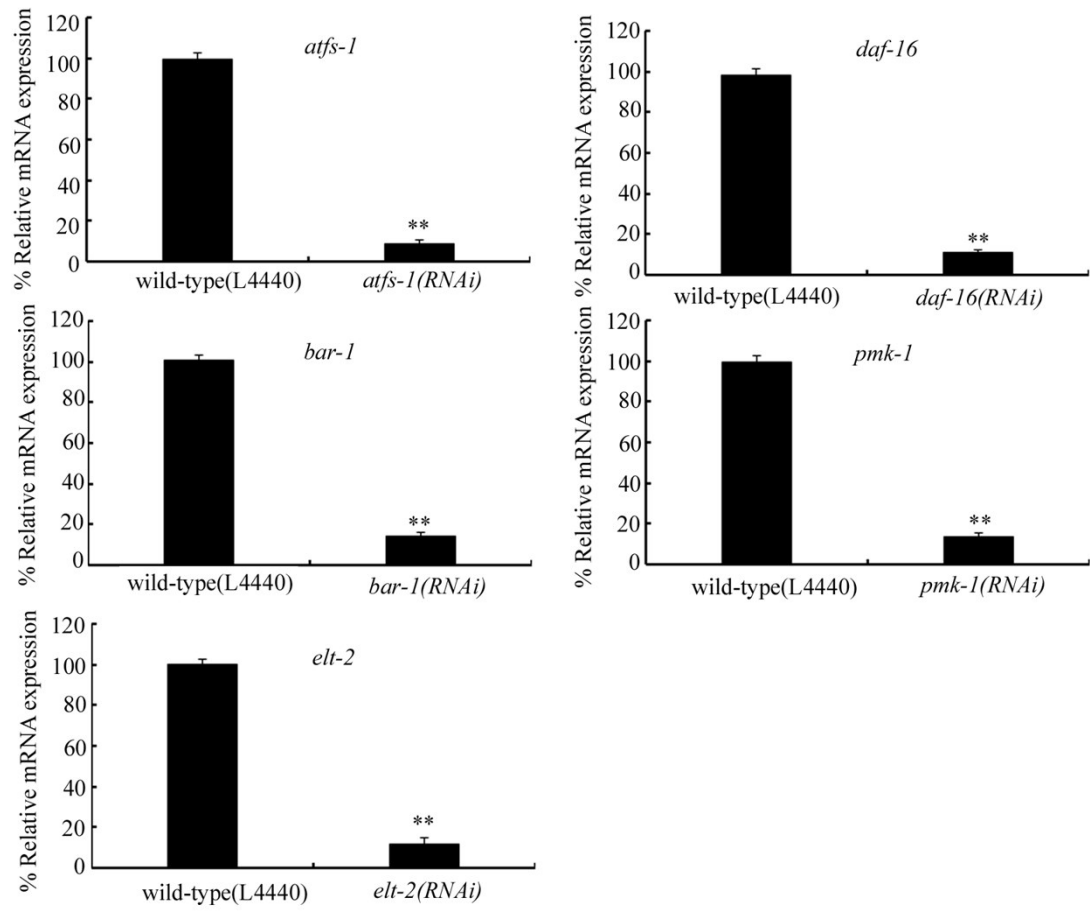


Figure S4. qRT-PCR analysis of the efficiency of RNAi knockdown of *bar-1*, *elt-2*, *pmk-1*, *daf-16*, or *atfs-1*. Bars represent means $\pm$ SD. ** $P < 0.01$ vs wild-type.

VP303(L4440)



paqr-2(RNAi)



Figure S5. Effect of intestinal RNAi knockdown of *paqr-2* on intestinal permeability based on the erioglaucine disodium staining. The nematodes were stained by 5% erioglaucine disodium (a blue dye) for 3 h. L4440, empty vector.

Table S1. Primer information for qRT-PCR

Gene	Forward primer (5'-3')	Reverse primer (5'-3')
<i>paqr-2</i>	TGTCGAGGACAGCACAACTG	GCCCTCCCTTTTTACGACGA
<i>seb-3</i>	TAACTCCGCTGCCTCTTTGG	CAAATGGAGTCCCTGGGCAT
<i>avr-14</i>	TCATCGTCTCGTGGGTCTCT	TGTACCCTTCTGTTGGGAGC
<i>avr-15</i>	ATAGGTCGATTGCGGAGAGG	CAAAACCCGCCAGAAGCAAT
<i>glc-1</i>	AGTCACCACGCTGCTTACAA	GTTCTGCTGGCCAAAACGAG
<i>npr-1</i>	GGAGACCGGCGTAAAAGTGA	ATTCTTCCGGCTCAGCAACA
<i>dop-2</i>	AGCAGAAGTACGACGGCAAA	CTTGCGAAACACGTCGACTC
<i>dop-3</i>	ATGTTGGCTGGACAACACCA	AGTGCTTGATACACGTGGCA
<i>ser-7</i>	GCGGGCCAATTCAACAACT	AGCTGGAAGATTGGGTTCGG
<i>mgl-1</i>	TTGGCTTGCATCAGAGTCGT	TGGCGTGGCATATTTTCAGGT
<i>mgl-2</i>	GGTAGGAGCACTTCTGGCTG	GTAGCTTGAGACAGCGCGAG
<i>ced-1</i>	GCTGGACGGGGAAATAACGA	GTCGTTACTGCATGTGCTGG
<i>npr-9</i>	CTTTCCCGAGTGGACATGCT	GCACGTTGACCGTTTTTCCA
<i>fshr-1</i>	TTGGTGGATGGCTTTTTGCG	ACGGTCTTCTTCGCGAGTTT
<i>dcar-1</i>	TGGATGGGCAATGCTTCCAT	ACCACCATGTCGCGGTTTAT
<i>glp-1</i>	GAAGTGCCCTTCATGAGGCT	TCGGTGTCTTCCGTCTTCG
<i>npr-12</i>	TCTACCACTTGCTGCCTTGG	GCTGCTGTACCGGTAGTTGT
<i>unc-63</i>	TTCTCATGACACGACCGCAA	GGCAACACTTTCGACAGCAG
<i>srh-220</i>	AGTCGTCGGGTGCTCAATTA	TTCGATGTTCAAGTCCAGGC
<i>npr-4</i>	GTAGTCGCATTCGCTTGCAG	TCATTGCCGACTTTTTGCGG
<i>npr-2</i>	ATACACGTGTTGAGGCTCCG	GCAGGGAGAATGTGACAAGGA
<i>frpr-4</i>	ATACACGTGTTGAGGCTCCG	GCAGGGAGAATGTGACAAGGA
<i>unc-68</i>	AGGAGGGCAACAGGATGATG	CCGCGTGTCCGTATAACAGT
<i>ser-4</i>	CAGACCGTAATCCTCGCCTC	AATTGAGGCAGTGCAGACGA
<i>daf-37</i>	ATCACCGTTGCAGTCACGAT	CGAGGACCAGTGTAGCTTCC
<i>gtr-1</i>	CGTCGCAAACCTCGCTTTAG	CGGCACAATTGCGCATATCA
<i>lin-12</i>	CGGATCATACGGCTCGTCAA	TCGGAGTATCGCGTCATTGG

<i>tkr-1</i>	CTGCTGGCTTCCATACCACA	TCACCGTTCATGGCAACTCA
<i>npr-8</i>	CCGAAGCTACGGATGGAGAC	ACCAAAGAGCCGCATAGCTT
<i>age-1</i>	ATGGAAACCGCCGAGTGT	ATTGGCAGTCGGTTCAGG
<i>daf-16</i>	AGGTGTTACACGTGGCCAAT	TGGCTTCTTACGACAACGCT
<i>sod-3</i>	TCTCCAACCAGCGCTGAAAT	CAAGTAGTAGGCGTGCTCCC
<i>bar-1</i>	CCTAATTTGCACGCTACGGC	TATGACCATCGCCCATTGCG
<i>pmk-1</i>	CGACTCCACGAGAAGGAT	ATATGTACGACGGGCATG
<i>mdt-15</i>	AGCGAAGAGGATTGGCCTTC	CCAGCCGGAGTGTTTCCTAA
<i>sbp-1</i>	ACCGTACCCGGAACCAATTC	TCCCTTCCCAGAAAAGCGTC
<i>atf-7</i>	GCTGAAAAGAATCAATTACTGACA C	CGTGCTGCTCATCACTCATTG
<i>skn-1</i>	AGTGCTTCTCTTCGGTAGCC	ATCCGTTGATCATCACGCCA
<i>elt-2</i>	GGAAGTGCAAAAGCAGACCG	TCATCAACCCAGCTGTAGCG
<i>nhr-8</i>	ACAACAGCTGTGAACAGTCC	CGCCGGAGAAACTGTGTAGT
<i>hsp-4</i>	TGGCAAACGCGTACTGTGAT	ATCTTGAACGGCCAGTGCTT
<i>hsp-6</i>	CTTGAAGCTCACCAGAGCCA	GGCGATTTGGGAGGACTTGA
<i>lgg-1</i>	CCGAAGGAGACAAGATCCGC	TTCTTCGTGATGGTCCTGGT
<i>hlh-30</i>	ATTTCCAACGAGACGCTTGC	GGAGATGGTGACTTGCTGCT
<i>atfs-1</i>	CACAACTTTTGGTGCCAGG	TAGGATCTCTGCCGCAAACC
<i>tba-1</i>	TCAACACTGCCATCGCCGCC	TCCAAGCGAGACCAGGCTTCAG

Table S2. Primer information for DNA constructs

	Forward primer (5'-3')	Reverse primer (5'-3')
<i>Pges-1</i>	ATATCTAGAAGCCACTCAGCCACTT CA	ATAGGATCCCATCTGAATTCAA AGATA
<i>paqr-2/Y32H12A.5</i>	ATACCCGGGATGGAGGAAGATGAC GTG	ACGGGTACCTCATAAACCAACA TCCGC
<i>.1</i>		
<i>fshr-1/C50H2.1a</i>	ATACCCGGGATGACATGCTCAACAT AT	GCGGGTACCCTAAGAACAATCA GAAAC
<i>1</i>		

Table S3. Expression patterns of some genes encoding GPCRs (<https://wormbase.org>)

Gene	Intestine	Neurons	Germline
<i>pqar-2</i>	+	+	+
<i>npr-1</i>		+	
<i>ced-1</i>	+	+	+
<i>fshr-1</i>	+	+	
<i>dcar-1</i>		+	
<i>ser-4</i>		+	
<i>daf-37</i>		+	
<i>dop-2</i>		+	
<i>npr-9</i>		+	
<i>npr-12</i>		+	
<i>npr-4</i>	+	+	
<i>gtr-1</i>		+	
<i>npr-8</i>		+	