

Supplementary Information

# Biomimetic regioselective and high-yielding Cu(I)-catalyzed dimerization of sinapate esters in green solvent Cyrene™: towards sustainable antioxidant and anti-UV ingredients

Matthieu M. Mention,<sup>a</sup> Amandine L. Flourat,<sup>a</sup> Cédric Peyrot<sup>a</sup> and Florent Allais<sup>a\*</sup>

<sup>a</sup> URD Agro-Biotechnologies Industrielles (ABI), CEBB, AgroParisTech, 51110 Pomacle, France

\* Corresponding author: [florent.allais@agroparistech.fr](mailto:florent.allais@agroparistech.fr)

## Table of Contents

1. General Information .....	S3
2. Synthesis .....	S3
2.1. Synthesis of malonate mono-esters .....	S3
2.1.1. General Procedure (GP1) .....	S3
2.1.2. General Procedure (GP2) .....	S4
2.2. Synthesis of sinapate esters .....	S4
2.2.1. General Procedure (GP3) .....	S4
2.2.2. Ethyl sinapate .....	S4
2.2.3. Sinapoyl di- <i>tert</i> -butyl malate .....	S4
2.3. Synthesis of sinapate esters $\beta$ - $\beta'$ dehydrodimers .....	S4
2.3.1. Initial Procedure .....	S4
2.3.2. Optimized Procedure .....	S5
2.4. Methylation of ethyl sinapate $\beta$ - $\beta'$ dehydrodimer .....	S5
2.5. Acetylation of ethyl sinapate $\beta$ - $\beta'$ dehydrodimer .....	S5
2.6. Hydrogenation of ethyl sinapate $\beta$ - $\beta'$ dehydrodimer .....	S6
2.7. Ester reduction of ethyl sinapate $\beta$ - $\beta'$ dehydrodimer .....	S6
3. Compounds Characterizations .....	S6
3.1. Malonate mono-ester .....	S6
3.2. Sinapate esters .....	S7
3.3. Sinapate esters $\beta$ - $\beta'$ dimers .....	S9
4. Enzymatic dimerization of ethyl sinapate .....	S14
<b>Figure S1.</b> HPLC chromatogram for the dimerization of ethyl sinapate with laccase from <i>Trametes versicolor</i> (Zorbax Eclipse Plus C18 (2.1 mm*50 mm*1.8 $\mu$ m), $\lambda$ = 320 nm, flow rate set at 0.6 mL.min <sup>-1</sup> , oven temperature at 30 °C, and gradient applied: H <sub>2</sub> O/CH <sub>3</sub> CN from 75/25 to 70/30 in 18 min). .....	S14
5. Optimization of the sinapate ester $\beta$ - $\beta'$ dimerization .....	S14

5.1.	Screening of solvents .....	S14
	<b>Figure S2.</b> HPLC chromatograms of the dimerization of ethyl sinapate in pyridine (black), ethanol (pink), ethyl acetate (blue) and Cyrene™ (green) (Zorbax Eclipse Plus C18 (2.1 mm*50 mm*1.8 μm), λ = 320 nm, flow rate set at 0.6 mL.min <sup>-1</sup> , oven temperature at 30 °C, and gradient applied: H <sub>2</sub> O/CH <sub>3</sub> CN from 75/25 to 70/30 in 18 min).....	S14
5.2.	Design of experiments (D.O.E.) .....	S14
	<b>Figure S3  </b> Summary of fit for conversion (left) and yield (right) .....	S17
6.	Antiradical properties .....	S18
	<b>Figure S4  </b> Antiradical properties from the ABTS <sup>•+</sup> assay: comparison of EC <sub>50</sub> between sinapate esters and their β-β' dimers.....	S18
7.	Characterization Data .....	S19
7.1.	<sup>1</sup> H & <sup>13</sup> C NMR spectra .....	S19
7.2.	IR spectra.....	S65
7.3.	UV Spectra.....	S88
7.4.	ABTS inhibition .....	S98
8.	References .....	S108

# 1. General Information

Syringaldehyde, copper(I) bromide, aniline, pyridine, malonic acid, heptan-1-ol, *tert*-butanol, *iso*-propanol, gallicol, sodium carbonate, methyl iodine, acetic anhydride, DIBAL-H, palladium on carbon (10wt. % loading), potassium persulfate, Laccase from *Trametes versicolor* and Horseradish peroxidase (type II and IV) were purchased from Sigma Aldrich. Meldrum's acid, ABTS and 2-ethylhexan-1-ol were purchased from TCI. HCl<sub>conc</sub> and solvents were purchased from Fisher scientific and used as received. Cyrene was kindly offered by Circa Group (Australia). All chemicals were used directly without purification.

Chromatographic purifications of products were accomplished using a flash-prep LC system puriFlash® 4100 from Interchim with prepacked silica column (30 µm, Interchim PF-Si30-HP), dual wavelength collection ( $\lambda = 254$  and 320 nm) and a mixture of cyclohexane/ethyl acetate as eluant. <sup>1</sup>H NMR spectra were recorded on a Bruker Fourier 300 (300 MHz) and were calibrated with residual Acetone-*d*<sub>6</sub>, DMSO-*d*<sub>6</sub> or CDCl<sub>3</sub> protons signals at  $\delta$  2.05, 2.50 or 7.26 ppm respectively. Data are reported as follows: chemical shift ( $\delta$  ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, sept = septet, dd = doublet of doublets, td = triplet of doublets and m = multiplet), integration, coupling constant (Hz) and assignment. <sup>13</sup>C NMR spectra were recorded on a Bruker Fourier 300 (75 MHz) and were calibrated with Acetone-*d*<sub>6</sub>, DMSO-*d*<sub>6</sub> or CDCl<sub>3</sub> signals at  $\delta$  29.84, 39.52 or 77.16 ppm respectively. Data are reported as follows: chemical shift ( $\delta$  ppm) and attribution. All NMR assignments were made using COSY, HMBC and HSQC spectrum. Solvents were dried on a MBRAUN-SPS-800. IR spectra were recorded on an Agilent Cary 630 FTIR Spectrometer and are reported in frequency of absorption (cm<sup>-1</sup>). UV/Vis spectra were recorded in ethanol (C = 10<sup>-5</sup> mol/L) on an Agilent Cary 60 UV-Vis with 1 cm cuvette made of quartz and are reported in wavelength (nm). Melting points were recorded on a Mettler Toledo MP50 Melting Point System (T<sub>initial</sub>: 40 °C; Heating: 2 °C/min until 170 °C) with ME-18552 sample tubes. HPLC analysis were carried out on a Dionex UltiMate 3000 equipped with a Zorbax Eclipse Plus C18 column (2.1\*50 mm\*1.8 µm), a DAD and a Corona ultra RS both at  $\lambda = 320$  nm (flow rate set at 0.6 mL.min<sup>-1</sup>, oven temperature at 30 °C, and gradient applied: H<sub>2</sub>O/CH<sub>3</sub>CN from 75/25 to 70/30 in 18 min). High resolution mass spectrometries were performed by the PIAneT platform at URCA on a Micromass GC-TOF. Modde v.12.0 software (Umetrics AB, Sweden) was used to generate the design of experiments and analyze the experimental data by Response Surface Methodology (RSM), based on a 4 factors and 3 central points cubic centered face (CCF) design.

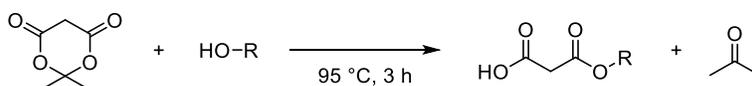
ABTS assays were adapted from Re et al.<sup>1</sup>. In summary, ABTS<sup>•+</sup> solution was prepared in water with ABTS (C = 7 mmol.L<sup>-1</sup>) and potassium persulfate (final concentration 2.45 mmol.L<sup>-1</sup>). Mixture was left to agitate in the dark for 16h and diluted by 50 with ethanol to reach around 0.7 of absorbance at 734 nm and 37 °C. In a 96 wells microplate, a mixture containing 10 µL of antiradical solution (final concentration in microwell were 2 to 60 µmol.L<sup>-1</sup>) and 190 µL of ABTS solution incubated at 37 °C. Absorbance was recorded at 734 nm until stabilization of the signal (6 min). To determine the exact value of inhibition, a reference (190 µL of ABTS solution with 10 µL of Ethanol) and a blank (200 µL of ethanol) were performed at the same time as each experiment. All experiments were carried out on an Epoch 2 from Biotek using 96-wells microplate.

## 2. Synthesis

### 2.1. Synthesis of malonate mono-esters

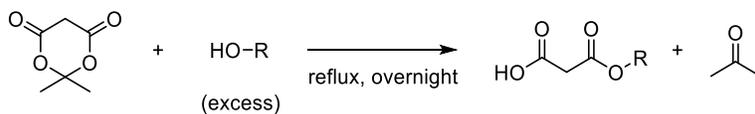
Malonate mono-esters were synthesized following methods described in the literature<sup>2,3</sup>. All malonate mono-esters were used without further purification for the synthesis of corresponding sinapate esters.

#### 2.1.1. General Procedure (GP1)



Meldrum's acid (5.0 g, 35 mmol) and the corresponding alcohol (35 mmol, 1 eq) were melted at 95 °C and agitated for 3 h. After cooling at r.t., the reaction mixture was partitioned between ethyl acetate and saturated NaHCO<sub>3</sub>. The aqueous layer was acidified until pH = 1 with concentrated HCl and extracted with ethyl acetate. The resulting organic layer was dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated.

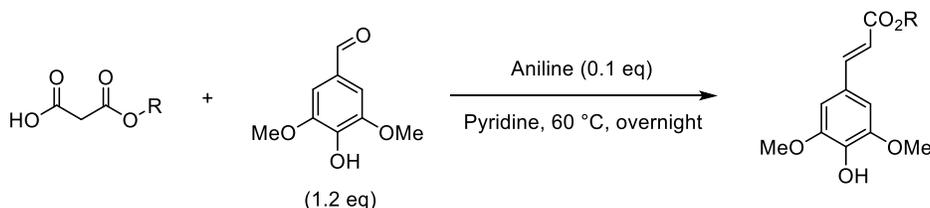
### 2.1.2. General Procedure (GP2)



Meldrum's acid (4.0 g, 27.8 mmol) and an excess of alcohol were heated at reflux overnight. After cooling at r.t., the excess of alcohol was evaporated under reduced pressure.

## 2.2. Synthesis of sinapate esters

### 2.2.1. General Procedure (GP3)



The corresponding malonate mono-esters (8.6 mmol, 1.5 M), syringaldehyde (1.89 g, 10.4 mmol) and aniline (79  $\mu$ L, 0.86 mmol) were mixed in pyridine (5.7 mL). The reaction mixture was stirred at 60 °C overnight. After cooling at r.t., the reaction was partitioned between ethyl acetate and 1M aqueous HCl. The organic layer was dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated before being purified by flash chromatography using cyclohexane/ethyl acetate.

### 2.2.2. Ethyl sinapate

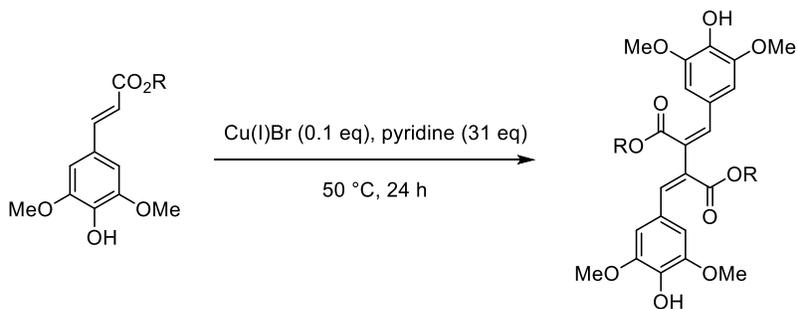
Ethyl sinapate was synthesized following method described by Jaufurally et al.<sup>4</sup>

### 2.2.3. Sinapoyl di-*tert*-butyl malate

Sinapoyl di-*tert*-butyl malate was synthesized following method described by Allais et al.<sup>5</sup>

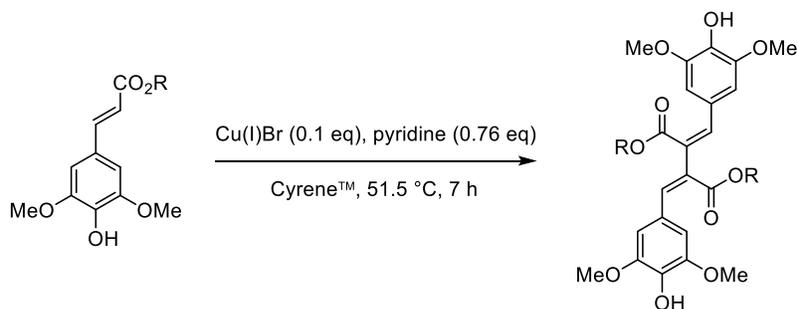
## 2.3. Synthesis of sinapate esters $\beta$ - $\beta'$ dehydrodimers

### 2.3.1. Initial Procedure



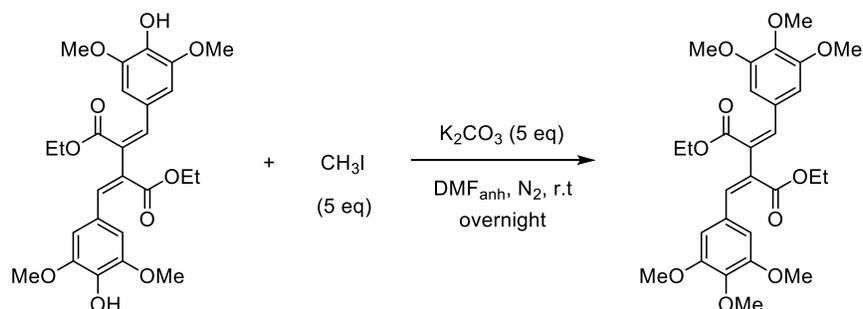
The corresponding sinapate ester (32 mmol) and Cu(I)Br (0.459 g, 3.2 mmol) were dissolved in pyridine (80 mL, 993 mmol) and warmed up at 50 °C. The reaction was stirred for 24 h in an opened flask. After cooling at r.t., the reaction mixture was diluted with ethyl acetate and washed with 1M aqueous HCl. The organic layer was dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated before being purified by flash chromatography using cyclohexane/ethyl acetate.

### 2.3.2. Optimized Procedure



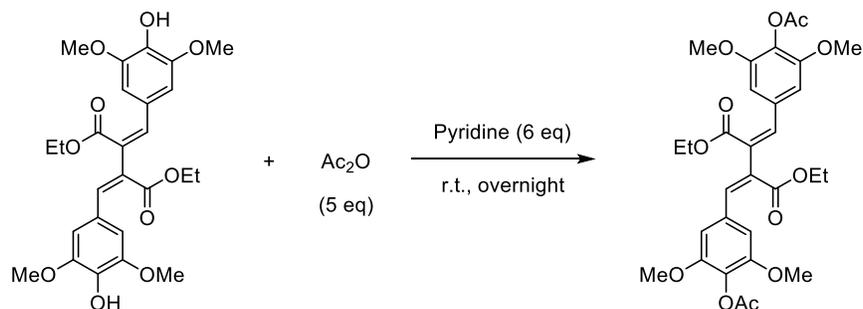
The corresponding sinapate ester (1 mmol, 1.81 M),  $\text{Cu(I)Br}$  (14.3 mg, 0.1 mmol) and pyridine (61  $\mu\text{L}$ , 0.76 mmol) were mixed in  $\text{Cyrene}^{\text{TM}}$  (552  $\mu\text{L}$ ). The reaction was stirred at  $51.5\text{ }^\circ\text{C}$  for 7 h in an opened flask. After cooling at r.t., the reaction mixture was diluted with ethyl acetate and washed with 1M aqueous HCl and water. The organic layer was dried over anhydrous  $\text{MgSO}_4$ , filtered and concentrated to afford the corresponding  $\beta$ - $\beta'$  dimer.

### 2.4. Methylation of ethyl sinapate $\beta$ - $\beta'$ dehydromer



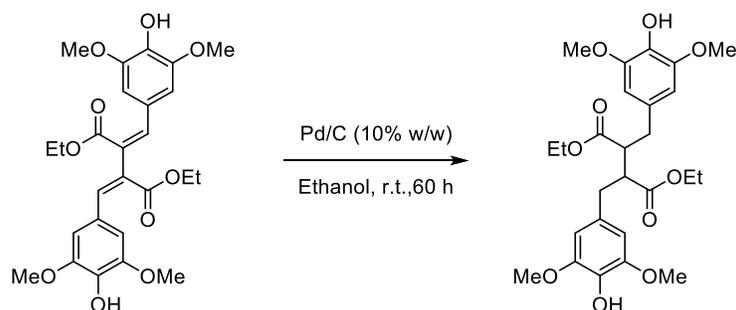
Ethyl sinapate  $\beta$ - $\beta'$  dehydromer (201 mg, 0.40 mmol, 0.5 M) was dissolved in anhydrous DMF (800  $\mu\text{L}$ ) and placed under  $\text{N}_2$ . After 10 min of stirring,  $\text{K}_2\text{CO}_3$  (276 mg, 2 mmol) and methyl iodide (125  $\mu\text{L}$ , 2 mmol) were added and the reaction was kept agitating under  $\text{N}_2$  at r.t. overnight. The mixture was then diluted in 1M aqueous HCl and partitioned between ethyl acetate and  $\text{H}_2\text{O}$ . The organic layer was dried over anhydrous  $\text{MgSO}_4$ , filtered and concentrated.

### 2.5. Acetylation of ethyl sinapate $\beta$ - $\beta'$ dehydromer



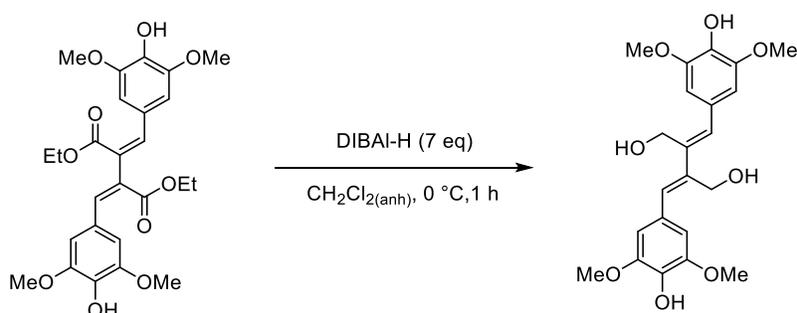
Ethyl sinapate  $\beta$ - $\beta'$  dehydromer (201 mg, 0.40 mmol) was dissolved in acetic anhydride (189  $\mu\text{L}$ , 2 mmol) and pyridine (193  $\mu\text{L}$ , 2.4 mmol) and stirred at r.t. overnight. The resulting precipitate was filtered and washed with 0.1M aqueous HCl and water, before being dried under vacuum.

## 2.6. Hydrogenation of ethyl sinapate $\beta$ - $\beta'$ dehydromer



Ethyl sinapate  $\beta$ - $\beta'$  dehydromer (1 g, 2 mmol) was dissolved in ethanol and stirred under  $N_2$  for 10 min. Palladium on carbon (100 mg, 10% w/w) was added and the mixture was stirred 10 more minutes before being placed under  $H_2$  for 60 h. The reaction was flushed with  $N_2$  and filtered over a pad of Celite<sup>®</sup> prior concentration under reduced pressure.

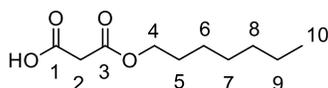
## 2.7. Ester reduction of ethyl sinapate $\beta$ - $\beta'$ dehydromer



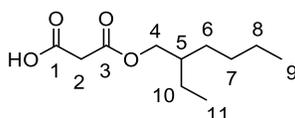
Ethyl sinapate  $\beta$ - $\beta'$  dehydromer (2 g, 4 mmol, 0.1 M) was dissolved in anhydrous dichloromethane under  $N_2$ . A solution of DIBAL-H (27.7 mL, 27.7 mmol, 1 M in  $CH_2Cl_2$ ) was added dropwise at 0 °C over 15 min. After 1 h, reaction was stopped by adding 15 mL of ethanol before being concentrated. The crude oil was partitioned between water and ethyl acetate. Organic layer was washed with brine, dried over anhydrous  $MgSO_4$  and concentrated before being purified by flash chromatography using cyclohexane/ethyl acetate.

## 3. Compounds Characterizations

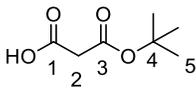
### 3.1. Malonate mono-ester



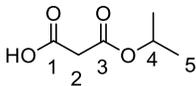
GP1 was followed with heptan-1-ol to obtain **monoheptyl malonate** as a colorless oil (77% yield).  $^1H$  NMR (300 MHz,  $CDCl_3$ ):  $\delta$  0.88 (t, 3H,  $J = 6$  Hz,  $H_{10}$ ), 1.29 (m, 8H,  $H_{6,7,8,9}$ ), 1.65 (m, 2H,  $H_5$ ), 3.44 (s, 2H,  $H_2$ ), 4.17 (t, 2H,  $J = 6.7$  Hz,  $H_4$ );  $^{13}C$  NMR (75 MHz,  $CDCl_3$ ):  $\delta$  14.2 ( $C_{10}$ ), 22.7 ( $C_9$ ), 25.8 ( $C_8$ ), 28.5 ( $C_5$ ), 29.0 ( $C_7$ ), 31.8 ( $C_6$ ), 40.7 ( $C_2$ ), 66.4 ( $C_4$ ), 167.5 ( $C_3$ ), 171.11 ( $C_1$ ); IR (FTIR): 2926, 2856, 1714, 1463, 1410, 1318, 1150  $cm^{-1}$ ; HRMS ( $m/z$ ) [ $M+Na$ ] $^+$  calcd for  $C_{10}H_{18}O_4Na$ , 225.1103; found, 225.1103.



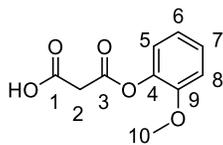
GP1 was followed with 2-ethylhexan-1-ol to obtain **mono-2-ethylhexyl malonate** as a light-yellow oil (83% yield).  $^1\text{H NMR}$  (300 MHz, DMSO-*d*6):  $\delta$  0.85 (m, 6H, H<sub>9,11</sub>), 1.29 (m, 8H, H<sub>6,7,8,10</sub>), 1.54 (m, 1H, H<sub>5</sub>), 3.36 (s, 2H, H<sub>2</sub>), 3.97 (s, 2H, H<sub>4</sub>), 12.76 (s, 1H, H<sub>COOH</sub>);  $^{13}\text{C NMR}$  (75 MHz, DMSO-*d*6):  $\delta$  10.8 (C<sub>9</sub>), 14.0 (C<sub>11</sub>), 22.4 (C<sub>8</sub>), 23.1 (C<sub>10</sub>), 28.3 (C<sub>7</sub>), 29.7 (C<sub>6</sub>), 38.1 (C<sub>5</sub>), 41.7 (C<sub>4</sub>), 66.6 (C<sub>2</sub>), 167.0 (C<sub>3</sub>), 168.1 (C<sub>1</sub>); IR (FTIR): 2958, 2928, 2861, 1717, 1461, 1410, 1382, 1318, 1267, 1149 cm<sup>-1</sup>; HRMS (*m/z*) [M+Na]<sup>+</sup> calcd for C<sub>11</sub>H<sub>20</sub>O<sub>4</sub>Na, 239.1259; found, 239.1262.



GP2 was followed with *tert*-butanol to obtain **mono-*tert*-butyl malonate** as a pale-yellow oil (97% yield). Characterization data were identical with those already described<sup>6</sup>.

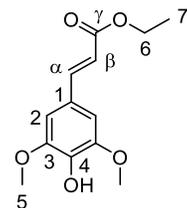


GP2 was followed with *iso*-propanol to obtain **mono-*iso*-propyl malonate** as a pale-yellow oil (76% yield). Characterization data were identical with those already described<sup>7</sup>.

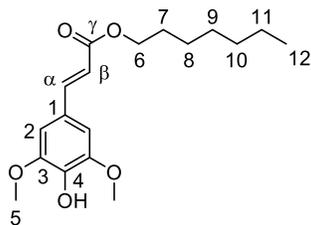


GP1 was followed with *g*aiacol to obtain **monogaiacol malonate** as a white crystal (60% yield). Characterization data were identical with those already described<sup>8</sup>.

### 3.2. Sinapate esters

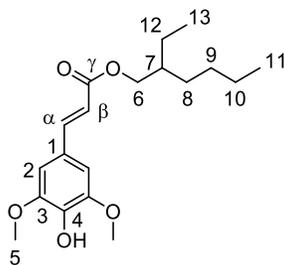


**Ethyl sinapate** was synthesized in two steps following methods from Jaufurally et al.<sup>4</sup> (80% global yield). Characterization data were identical with those already described<sup>4</sup>.

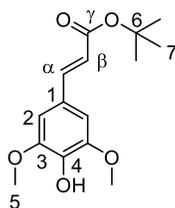


GP3 was followed with monoheptyl malonate to obtain **heptyl sinapate** as a brown oil (72% yield).  $^1\text{H NMR}$  (300 MHz, Acetone-*d*6):  $\delta$  0.77 (t, 3H, J = 6.8 Hz, H<sub>12</sub>), 1.23 (m, 8H, H<sub>8,9,10,11</sub>), 1.58 (m, 2H, H<sub>7</sub>), 3.79 (s, 6H, H<sub>5</sub>), 4.08 (t, 2H, J = 6.7 Hz, H<sub>6</sub>), 6.19 (d, 1H, J = 15.9 Hz, H<sub>β</sub>), 6.65 (s, 2H, H<sub>2</sub>), 7.47 (d, 1H, J = 15.9 Hz, H<sub>α</sub>);  $^{13}\text{C NMR}$  (75 MHz, CDCl<sub>3</sub>):  $\delta$  14.1 (C<sub>12</sub>), 22.6 (C<sub>11</sub>), 26.0 (C<sub>10</sub>), 28.8 (C<sub>7</sub>), 29.0 (C<sub>9</sub>), 31.8 (C<sub>8</sub>), 56.3 (C<sub>5</sub>), 64.7 (C<sub>6</sub>), 105.0 (C<sub>2</sub>), 116.0 (C<sub>β</sub>), 125.9 (C<sub>1</sub>), 137.1 (C<sub>4</sub>), 144.9 (C<sub>α</sub>), 147.2 (C<sub>3</sub>), 167.3 (C<sub>γ</sub>); IR (FTIR): 3406, 2925, 2853, 1700, 1631, 1601, 1511, 1455, 1424, 1375, 1337, 1279, 1252, 1214,

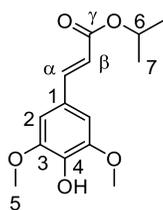
1149, 1109  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  202, 241, 330 nm; **HRMS** ( $m/z$ )  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{18}\text{H}_{27}\text{O}_5$ , 323.1858; found, 323.1855.



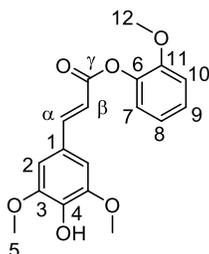
GP3 was followed with mono-2-ethylhexyl malonate to obtain **2-ethylhexyl sinapate** as a yellow-orange oil (66% yield).  **$^1\text{H}$  NMR** (300 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  0.88 (t, 6H,  $J = 7.4$  Hz,  $\text{H}_{11,13}$ ), 1.34 (m, 8H,  $\text{H}_{8,9,10,12}$ ), 1.59 (m, 1H,  $\text{H}_7$ ), 3.80 (s, 6H,  $\text{H}_5$ ), 4.05 (dd, 2H,  $J = 5.7, 2.0$  Hz,  $\text{H}_6$ ), 6.53 (d, 1H,  $J = 15.9$  Hz,  $\text{H}_\beta$ ), 7.03 (s, 1H,  $\text{H}_2$ ), 7.54 (d, 1H,  $J = 15.9$  Hz,  $\text{H}_\alpha$ ), 9.00 (s, 1H,  $\text{H}_{\text{phenol}}$ );  **$^{13}\text{C}$  NMR** (75 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  10.9 ( $\text{C}_{11}$ ), 14.0 ( $\text{C}_{13}$ ), 22.5 ( $\text{C}_{10}$ ), 23.2 ( $\text{C}_{12}$ ), 28.4 ( $\text{C}_9$ ), 29.8 ( $\text{C}_8$ ), 38.4 ( $\text{C}_7$ ), 56.1 ( $\text{C}_5$ ), 65.7 ( $\text{C}_6$ ), 106.2 ( $\text{C}_2$ ), 114.9 ( $\text{C}_\beta$ ), 124.4 ( $\text{C}_1$ ), 138.2 ( $\text{C}_4$ ), 145.4 ( $\text{C}_\alpha$ ), 148.0 ( $\text{C}_3$ ), 168.9 ( $\text{C}_\gamma$ ); **IR** (FTIR): 3407, 2928, 2858, 1699, 1631, 1598, 1511, 1455, 1424, 1377, 1337, 1278, 1252, 1214, 1148, 1109  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  202, 240, 332 nm; **HRMS** ( $m/z$ )  $[\text{M}+\text{Na}]^+$  calcd for  $\text{C}_{19}\text{H}_{28}\text{O}_5\text{Na}$ , 359.1834; found, 359.1835.



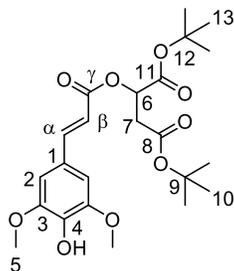
GP3 was followed with mono-*tert*-butyl malonate to obtain ***tert*-butyl sinapate** as a pale-yellow powder (56% yield). **mp**: 68 – 71  $^\circ\text{C}$ ;  **$^1\text{H}$  NMR** (300 MHz,  $\text{Acetone}-d_6$ ):  $\delta$  1.46 (s, 9H,  $\text{H}_7$ ), 3.85 (s, 6H,  $\text{H}_5$ ), 6.29 (d, 1H,  $J = 15.9$  Hz,  $\text{H}_\beta$ ), 6.95 (s, 2H,  $\text{H}_2$ ), 7.46 (d, 1H,  $J = 15.9$  Hz,  $\text{H}_\alpha$ );  **$^{13}\text{C}$  NMR** (75 MHz,  $\text{Acetone}-d_6$ ):  $\delta$  28.4 ( $\text{C}_7$ ), 56.6 ( $\text{C}_5$ ), 80.1 ( $\text{C}_6$ ), 106.5 ( $\text{C}_2$ ), 118.0 ( $\text{C}_\beta$ ), 126.2 ( $\text{C}_1$ ), 139.1 ( $\text{C}_4$ ), 145.0 ( $\text{C}_\alpha$ ), 148.8 ( $\text{C}_3$ ), 166.9 ( $\text{C}_\gamma$ ); **IR** (FTIR): 3388, 2970, 2933, 1706, 1631, 1598, 1511, 1463, 1424, 1367, 1333, 1308, 1248, 1222, 1133, 1096  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  202, 204, 238, 327 nm; **HRMS** ( $m/z$ )  $[\text{M}+\text{Na}]^+$  calcd for  $\text{C}_{15}\text{H}_{20}\text{O}_5\text{Na}$ , 303.1208; found, 303.1207.



GP3 was followed with mono-*iso*-propyl malonate to obtain ***iso*-propyl sinapate** as a pale-yellow oil (73% yield). Characterization data were identical to those already described<sup>9</sup>.

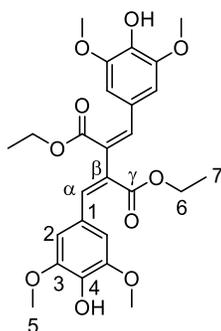


GP3 was followed with monogaïacol malonate to obtain **gaïacol sinapate** as a yellow-orange solid (55% yield). **mp**: 158 – 160 °C; **<sup>1</sup>H NMR** (300 MHz, DMSO-*d*6): δ 3.77 (s, 3H, H<sub>12</sub>), 3.82 (s, 6H, H<sub>5</sub>), 6.78 (d, 1H, *J* = 15.9 Hz, H<sub>β</sub>), 6.97 (td, 1H, *J* = 7.7, 1.5 Hz, H<sub>7</sub>), 7.14 (m, 4H, H<sub>2,8,9</sub>), 7.25 (td, 1H, *J* = 7.7, 1.5 Hz, H<sub>10</sub>), 7.73 (d, 1H, *J* = 15.9 Hz, H<sub>α</sub>), 9.08 (s, 1H, H<sub>phenol</sub>); **<sup>13</sup>C NMR** (75 MHz, DMSO-*d*6): δ 55.7 (C<sub>12</sub>), 56.1 (C<sub>5</sub>), 106.6 (C<sub>2</sub>), 112.8 (C<sub>8</sub>), 113.7 (C<sub>β</sub>), 120.6 (C<sub>7</sub>), 123.0 (C<sub>9</sub>), 124.3 (C<sub>1</sub>), 126.8 (C<sub>10</sub>), 138.7 (C<sub>4</sub>), 139.5 (C<sub>6</sub>), 147.3 (C<sub>α</sub>), 148.1 (C<sub>3</sub>), 151.1 (C<sub>11</sub>), 164.8 (C<sub>γ</sub>); **IR** (FTIR): 3377, 3062, 3021, 2936, 2842, 1716, 1603, 1496, 1458, 1426, 1380, 1338, 1307, 1286, 1248, 1213, 1154, 1106 cm<sup>-1</sup>; **UV/Vis** (Ethanol): λ<sub>max</sub> 201, 220, 242, 337 nm; **HRMS** (*m/z*) [M+Na]<sup>+</sup> calcd for C<sub>18</sub>H<sub>18</sub>O<sub>6</sub>Na, 353.1001; found, 353.1002.

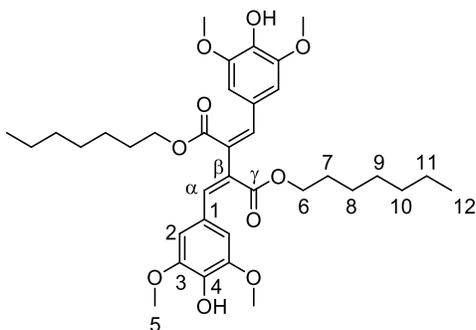


**Sinapoyl ditert-butyl malate** was synthesized as a yellow oil in 5 steps following methods from Allais et al.<sup>5</sup> (33% global yield). **<sup>1</sup>H NMR** (300 MHz, DMSO-*d*6): δ 1.41 (s, 18H, H<sub>10,13</sub>), 2.79 (m, 2H, H<sub>7</sub>), 3.81 (s, 6H, H<sub>2</sub>), 5.25 (m, 1H, H<sub>6</sub>), 6.60 (d, 1H, *J* = 15.8 Hz, H<sub>β</sub>), 7.06 (s, 2H, H<sub>2</sub>), 7.61 (d, 1H, *J* = 15.8 Hz, H<sub>α</sub>), 9.04 (s, 1H, H<sub>phenol</sub>); **<sup>13</sup>C NMR** (75 MHz, DMSO-*d*6) δ 27.5 (C<sub>10</sub>), 27.7 (C<sub>13</sub>), 37.0 (C<sub>7</sub>), 68.7 (C<sub>6</sub>), 80.9 (C<sub>9</sub>), 81.9 (C<sub>12</sub>), 106.4 (C<sub>2</sub>), 113.7 (C<sub>β</sub>), 124.2 (C<sub>1</sub>), 138.6 (C<sub>4</sub>), 146.7 (C<sub>α</sub>), 148.1 (C<sub>3</sub>), 165.8 (C<sub>γ</sub>), 167.7 (C<sub>8</sub>), 168.2 (C<sub>11</sub>); **IR** (FTIR): 3421, 2976, 2935, 1711, 1631, 1599, 1512, 1456, 1367, 1218, 1138, 1109; **UV/Vis** (Ethanol): λ<sub>max</sub> 242, 334 nm; **HRMS** (*m/z*) [M+Na]<sup>+</sup> calcd for C<sub>23</sub>H<sub>32</sub>O<sub>9</sub>Na, 475.1944; found, 475.1950.

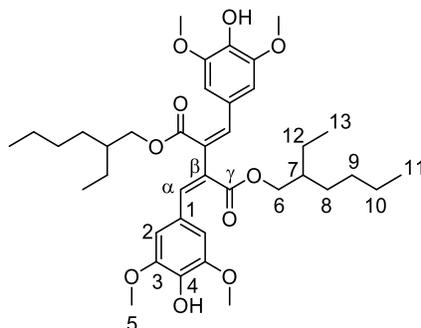
### 3.3. Sinapate esters β-β' dimers



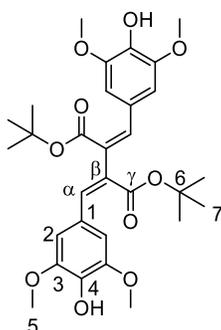
Dimerization of ethyl sinapate led to **ethyl sinapate β-β' dehydrodimer (8)** as a yellow powder (62% non-optimized yield / 89% optimized yield). **mp**: 98 – 101 °C; **<sup>1</sup>H NMR** (300 MHz, Acetone-*d*6): δ 1.12 (t, 6H, *J* = 7.1 Hz, H<sub>7</sub>), 3.76 (s, 12H, H<sub>5</sub>), 4.14 (q, 4H, *J* = 7.0 Hz, H<sub>6</sub>), 6.96 (s, 4H, H<sub>2</sub>), 7.82 (s, 2H, H<sub>α</sub>); **<sup>13</sup>C NMR** (75 MHz, Acetone-*d*6): δ 14.6 (C<sub>7</sub>), 56.6 (C<sub>5</sub>), 61.3 (C<sub>6</sub>), 108.8 (C<sub>2</sub>), 126.2 (C<sub>β</sub>), 126.6 (C<sub>1</sub>), 139.0 (C<sub>4</sub>), 142.7 (C<sub>α</sub>), 148.6 (C<sub>3</sub>), 167.6 (C<sub>γ</sub>); **IR** (FTIR): 3525, 3263, 2956, 1693, 1580, 1510, 1452, 1329, 1212, 1152, 1102, 1028 cm<sup>-1</sup>; **UV/Vis** (Ethanol): λ<sub>max</sub> 203, 243, 331 nm; **HRMS** (*m/z*) [M+Na]<sup>+</sup> calcd for C<sub>26</sub>H<sub>30</sub>O<sub>10</sub>Na, 525.1737; found, 525.1740.



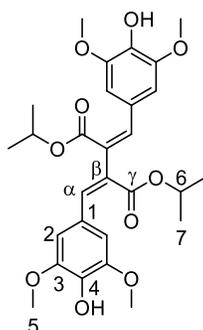
Dimerization of heptyl sinapate led to **heptyl sinapate  $\beta$ - $\beta'$  dehydrodimer (9)** as a yellow-brown oil (64% non-optimized yield / 90% optimized yield).  $^1\text{H NMR}$  (300 MHz, DMSO-*d*6):  $\delta$  0.82 (t, 6H, J = 6.9 Hz, H<sub>12</sub>), 1.16 (m, 16H, H<sub>8,9,10,11</sub>), 1.42 (m, 4H, H<sub>7</sub>), 3.66 (s, 12H, H<sub>5</sub>), 3.98 (m, 2H, H<sub>6</sub>), 4.10 (m, 2H, H<sub>6</sub>), 6.93 (s, 4H, H<sub>2</sub>), 7.78 (s, 2H, H <sub>$\alpha$</sub> ), 9.04 (s, 2H, H<sub>phenol</sub>);  $^{13}\text{C NMR}$  (75 MHz, DMSO-*d*6):  $\delta$  13.9 (C<sub>12</sub>), 22.0 (C<sub>11</sub>), 25.3 (C<sub>10</sub>), 28.2 (C<sub>9</sub>), 28.3 (C<sub>7</sub>), 31.2 (C<sub>8</sub>), 55.8 (C<sub>5</sub>), 64.5 (C<sub>6</sub>), 107.8 (C<sub>2</sub>), 124.0 (C <sub>$\beta$</sub> ), 124.6 (C<sub>1</sub>), 138.0 (C<sub>4</sub>), 141.6 (C <sub>$\alpha$</sub> ), 147.7 (C<sub>3</sub>), 166.7 (C <sub>$\gamma$</sub> ); IR (FTIR): 3464, 3168, 2924, 1707, 1582, 1513, 1453, 1329, 1214, 1107 cm<sup>-1</sup>; UV/Vis (Ethanol):  $\lambda_{\text{max}}$  201, 244, 333 nm; HRMS (*m/z*) [M+H]<sup>+</sup> calcd for C<sub>36</sub>H<sub>51</sub>O<sub>10</sub>, 643.3482; found, 643.3473.



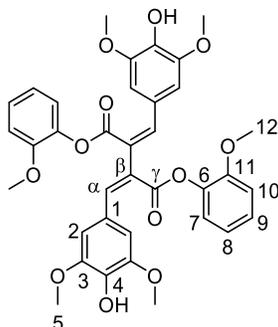
Dimerization of 2-ethylhexyl sinapate led to **2-ethylhexyl sinapate  $\beta$ - $\beta'$  dehydrodimer (10)** as a yellow-brown oil (55% non-optimized yield / 88% optimized yield).  $^1\text{H NMR}$  (300 MHz, CDCl<sub>3</sub>):  $\delta$  0.77 (m, 12H, H<sub>11,13</sub>), 1.15 (m, 16H, H<sub>8,9,10,12</sub>), 1.42 (m, 2H, H<sub>7</sub>), 3.79 (s, 12H, H<sub>5</sub>), 3.95 (m, 2H, H<sub>6</sub>), 4.03 (m, 2H, H<sub>6</sub>), 5.71 (s, 2H, H<sub>phenol</sub>), 6.84 (s, 4H, H<sub>2</sub>), 7.81 (s, 2H, H <sub>$\alpha$</sub> );  $^{13}\text{C NMR}$  (75 MHz, CDCl<sub>3</sub>):  $\delta$  11.0 (C<sub>11</sub>), 14.2 (C<sub>13</sub>), 23.0 (C<sub>10</sub>), 23.8 (C<sub>12</sub>), 29.0 (C<sub>9</sub>), 30.4 (C<sub>8</sub>), 38.9 (C<sub>7</sub>), 56.30 (C<sub>5</sub>), 67.3 (C<sub>6</sub>), 107.1 (C<sub>2</sub>), 125.5 (C <sub>$\beta$</sub> ), 126.3 (C<sub>1</sub>), 136.7 (C<sub>4</sub>), 142.1 (C <sub>$\alpha$</sub> ), 147.0 (C<sub>3</sub>), 167.6 (C <sub>$\gamma$</sub> ); IR (FTIR): 3407, 2927, 1698, 1588, 1510, 1454, 1329, 1213, 1152, 1104 cm<sup>-1</sup>; UV/Vis (Ethanol):  $\lambda_{\text{max}}$  202, 244, 333 nm; HRMS (*m/z*) [M+H]<sup>+</sup> calcd for C<sub>38</sub>H<sub>55</sub>O<sub>10</sub>, 671.3795; found, 671.3800.



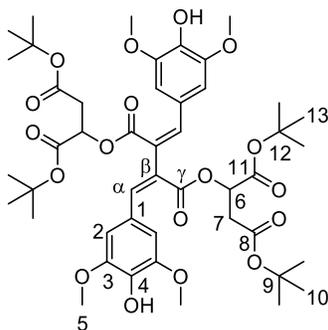
Dimerization of *tert*-butyl sinapate led to ***tert*-butyl sinapate  $\beta$ - $\beta'$  dehydrodimer (11)** as a yellow powder (58% non-optimized yield / 89% optimized yield). mp: 75 – 77 °C;  $^1\text{H NMR}$  (300 MHz, Acetone-*d*6):  $\delta$  1.36 (s, 18H, H<sub>7</sub>), 3.77 (s, 12H, H<sub>5</sub>), 6.92 (s, 4H, H<sub>2</sub>), 7.67 (s, 2H, H <sub>$\alpha$</sub> );  $^{13}\text{C NMR}$  (75 MHz, Acetone-*d*6):  $\delta$  28.0 (C<sub>7</sub>), 56.2 (C<sub>5</sub>), 80.4 (C<sub>6</sub>), 107.9 (C<sub>2</sub>), 108.3 (C<sub>2</sub>), 126.8 (C <sub>$\beta$</sub> ), 128.1 (C<sub>1</sub>), 138.1 (C<sub>4</sub>), 141.3 (C <sub>$\alpha$</sub> ), 148.3 (C<sub>3</sub>), 166.9 (C <sub>$\gamma$</sub> ); IR (FTIR): 3394, 2933, 1689, 1588, 1510, 1452, 1331, 1245, 1145, 1105 cm<sup>-1</sup>; UV/Vis (Ethanol):  $\lambda_{\text{max}}$  204, 244, 328 nm; HRMS (*m/z*) [M+Na]<sup>+</sup> calcd for C<sub>30</sub>H<sub>38</sub>O<sub>10</sub>Na, 581.2363; found, 581.2358.



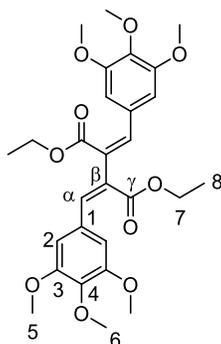
Dimerization of *iso*-butyl sinapate led to ***iso*-butyl sinapate  $\beta$ - $\beta'$  dehydrodimer (12)** as a yellow powder (58% non-optimized yield / 91% optimized yield). **mp**: 48 – 51 °C;  **$^1\text{H NMR}$**  (300 MHz, Acetone- $d_6$ ):  $\delta$  1.04 (d, 2H,  $J$  = 6.2 Hz,  $\text{H}_7$ ), 1.18 (d, 2H,  $J$  = 6.2 Hz,  $\text{H}_7$ ), 3.76 (s, 12H,  $\text{H}_5$ ), 4.98 (sept, 2H,  $J$  = 6.3 Hz,  $\text{H}_6$ ), 6.94 (s, 4H,  $\text{H}_2$ ), 7.78 (s, 2H,  $\text{H}_\alpha$ );  **$^{13}\text{C NMR}$**  (75 MHz, Acetone- $d_6$ ):  $\delta$  21.9 ( $\text{C}_7$ ), 22.1 ( $\text{C}_7$ ), 56.6 ( $\text{C}_5$ ), 68.6 ( $\text{C}_6$ ), 108.6 ( $\text{C}_2$ ), 126.8 ( $\text{C}_\beta$ ), 126.9 ( $\text{C}_1$ ), 138.8 ( $\text{C}_4$ ), 142.4 ( $\text{C}_\alpha$ ), 148.6 ( $\text{C}_3$ ), 167.2 ( $\text{C}_\gamma$ ); **IR** (FTIR): 3384, 2935, 1688, 1587, 1509, 1451, 1370, 1325, 1214, 1154, 1092  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  201, 244, 333 nm; **HRMS** ( $m/z$ ) [ $\text{M}+\text{Na}$ ] $^+$  calcd for  $\text{C}_{28}\text{H}_{34}\text{O}_{10}\text{Na}$ , 553.2050; found, 553.2049.



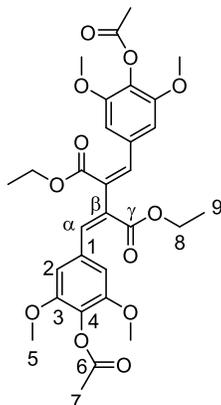
Dimerization of *gaïacol* sinapate led to ***gaïacol* sinapate  $\beta$ - $\beta'$  dehydrodimer (13)** as a yellow powder (42% non-optimized yield / 87% optimized yield). **mp**: 128 – 131 °C;  **$^1\text{H NMR}$**  (300 MHz, DMSO- $d_6$ ):  $\delta$  3.58 (s, 6H,  $\text{H}_{12}$ ), 3.72 (s, 12H,  $\text{H}_5$ ), 6.87 (dd, 2H,  $J$  = 7.9, 1.7 Hz,  $\text{H}_7$ ), 6.95 (td, 2H,  $J$  = 7.6, 1.4 Hz,  $\text{H}_8$ ), 7.11 (dd, 2H,  $J$  = 7.9, 1.7 Hz,  $\text{H}_{10}$ ), 7.15 (s, 4H,  $\text{H}_2$ ), 7.23 (m, 2H,  $\text{H}_9$ ), 8.04 (s, 2H,  $\text{H}_\alpha$ ), 9.20 (s, 2H,  $\text{H}_{\text{phenol}}$ );  **$^{13}\text{C NMR}$**  (75 MHz, DMSO- $d_6$ ):  $\delta$  55.6 ( $\text{C}_{12}$ ), 55.9 ( $\text{C}_5$ ), 108.3 ( $\text{C}_2$ ), 113.1 ( $\text{C}_{10}$ ), 120.7 ( $\text{C}_8$ ), 122.5 ( $\text{C}_7$ ), 122.6 ( $\text{C}_\beta$ ), 124.4 ( $\text{C}_1$ ), 127.0 ( $\text{C}_9$ ), 138.5 ( $\text{C}_4$ ), 139.6 ( $\text{C}_6$ ), 143.9 ( $\text{C}_\alpha$ ), 147.7 ( $\text{C}_3$ ), 151.1 ( $\text{C}_{11}$ ), 164.9 ( $\text{C}_\gamma$ ); **IR** (FTIR): 3398, 2935, 1712, 1585, 1498, 1453, 1329, 1211, 1148, 1105, 1021  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  202, 246, 338 nm; **HRMS** ( $m/z$ ) [ $\text{M}+\text{Na}$ ] $^+$  calcd for  $\text{C}_{36}\text{H}_{34}\text{O}_{12}\text{Na}$ , 681.1948; found, 681.1953.



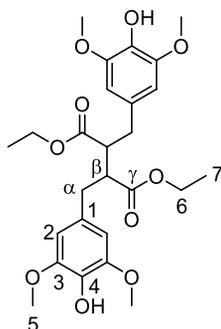
Dimerization of sinapoyl *di**tert*-butyl malate led to **sinapoyl *di**tert*-butyl malate  $\beta$ - $\beta'$  dehydrodimer (14)** as a yellow powder (42% non-optimized yield / 88% optimized yield). **mp**: 72 – 74 °C;  **$^1\text{H NMR}$**  (300 MHz, DMSO- $d_6$ ):  $\delta$  1.33 (m, 18H,  $\text{H}_{10}$ ), 1.37 (m, 18H,  $\text{H}_{13}$ ), 2.74 (m, 4H,  $\text{H}_7$ ), 3.66 (s, 12H,  $\text{H}_5$ ), 5.19 (m, 2H,  $\text{H}_6$ ), 6.88 (m, 4H,  $\text{H}_2$ ), 7.77 (s, 2H,  $\text{H}_\alpha$ ), 9.05 (s, 2H,  $\text{H}_{\text{phenol}}$ );  **$^{13}\text{C NMR}$**  (75 MHz, DMSO- $d_6$ ):  $\delta$  27.5 ( $\text{C}_{10,13}$ ), 36.8 ( $\text{C}_7$ ), 55.8 ( $\text{C}_5$ ), 69.2 ( $\text{C}_6$ ), 80.7 ( $\text{C}_9$ ), 81.7 ( $\text{C}_{12}$ ), 108.0 ( $\text{C}_2$ ), 122.3 ( $\text{C}_\beta$ ), 124.1 ( $\text{C}_1$ ), 138.2 ( $\text{C}_4$ ), 142.6 ( $\text{C}_\alpha$ ), 147.6 ( $\text{C}_3$ ), 165.5 ( $\text{C}_\gamma$ ), 167.4 ( $\text{C}_{11}$ ), 168.0 ( $\text{C}_8$ ); **IR** (FTIR): 3421, 2975, 1709, 1590, 1510, 1454, 1366, 1212, 1143, 1101  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  204, 243, 329 nm; **HRMS** ( $m/z$ ) [ $\text{M}+\text{Na}$ ] $^+$  calcd for  $\text{C}_{46}\text{H}_{62}\text{O}_{18}\text{Na}$ , 925.3834; found, 925.3835.



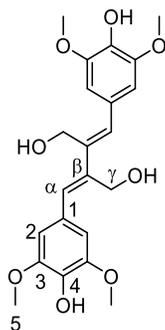
Methylation of ethyl sinapate  $\beta$ - $\beta'$  dehydrodimer (8) led to **methylated ethyl sinapate  $\beta$ - $\beta'$  dehydrodimer (15)** as an orange solid (93% yield). **mp**: 130 – 132 °C;  **$^1\text{H NMR}$**  (300 MHz, DMSO-*d*6):  $\delta$  1.10 (t, 6H,  $J$  = 7.1 Hz,  $\text{H}_8$ ), 3.77 (s, 12H,  $\text{H}_5$ ), 3.84 (s, 6H,  $\text{H}_6$ ), 4.15 (m, 4H,  $\text{H}_7$ ), 6.78 (s, 4H,  $\text{H}_2$ ), 7.83 (s, 2H,  $\text{H}_\alpha$ );  **$^{13}\text{C NMR}$**  (75 MHz, DMSO-*d*6):  $\delta$  14.2 ( $\text{C}_8$ ), 56.1 ( $\text{C}_5$ ), 61.1 ( $\text{C}_6$ ), 61.4 ( $\text{C}_7$ ), 107.1 ( $\text{C}_2$ ), 126.9 ( $\text{C}_\beta$ ), 130.4 ( $\text{C}_1$ ), 139.5 ( $\text{C}_4$ ), 142.3 ( $\text{C}_\alpha$ ), 153.2 ( $\text{C}_3$ ), 167.0 ( $\text{C}_\gamma$ ); **IR** (FTIR): 2936, 1695, 1573, 1500, 1459, 1415, 1333, 1230, 1115, 1029  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  202, 230, 310 nm; **HRMS** ( $m/z$ ) [ $\text{M}+\text{Na}$ ] $^+$  calcd for  $\text{C}_{28}\text{H}_{34}\text{O}_{10}\text{Na}$ , 553.2050; found, 553.2051.



Acetylation of ethyl sinapate  $\beta$ - $\beta'$  dehydrodimer (8) led to **acetylated ethyl sinapate  $\beta$ - $\beta'$  dehydrodimer (16)** as a light-yellow solid (99% yield). **mp**: 99 – 101 °C;  **$^1\text{H NMR}$**  (300 MHz, DMSO-*d*6):  $\delta$  1.10 (t, 6H,  $J$  = 7.1 Hz,  $\text{H}_9$ ), 2.31 (s, 6H,  $\text{H}_7$ ), 3.73 (s, 12H,  $\text{H}_5$ ), 4.15 (m, 4H,  $\text{H}_8$ ), 6.72 (s, 4H,  $\text{H}_2$ ), 7.81 (s, 2H,  $\text{H}_\alpha$ );  **$^{13}\text{C NMR}$**  (75 MHz, DMSO-*d*6):  $\delta$  14.2 ( $\text{C}_9$ ), 20.6 ( $\text{C}_7$ ), 56.2 ( $\text{C}_5$ ), 61.5 ( $\text{C}_8$ ), 106.4 ( $\text{C}_2$ ), 128.0 ( $\text{C}_\beta$ ), 129.8 ( $\text{C}_4$ ), 133.2 ( $\text{C}_1$ ), 142.4 ( $\text{C}_\alpha$ ), 152.2 ( $\text{C}_3$ ), 166.7 ( $\text{C}_\gamma$ ), 168.6 ( $\text{C}_6$ ); **IR** (FTIR): 2939, 1767, 1695, 1584, 1502, 1460, 1416, 1227, 1192, 1125, 1010  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  205, 222, 299 nm; **HRMS** ( $m/z$ ) [ $\text{M}+\text{Na}$ ] $^+$  calcd for  $\text{C}_{30}\text{H}_{34}\text{O}_{12}\text{Na}$ , 609.1948; found, 609.1946.

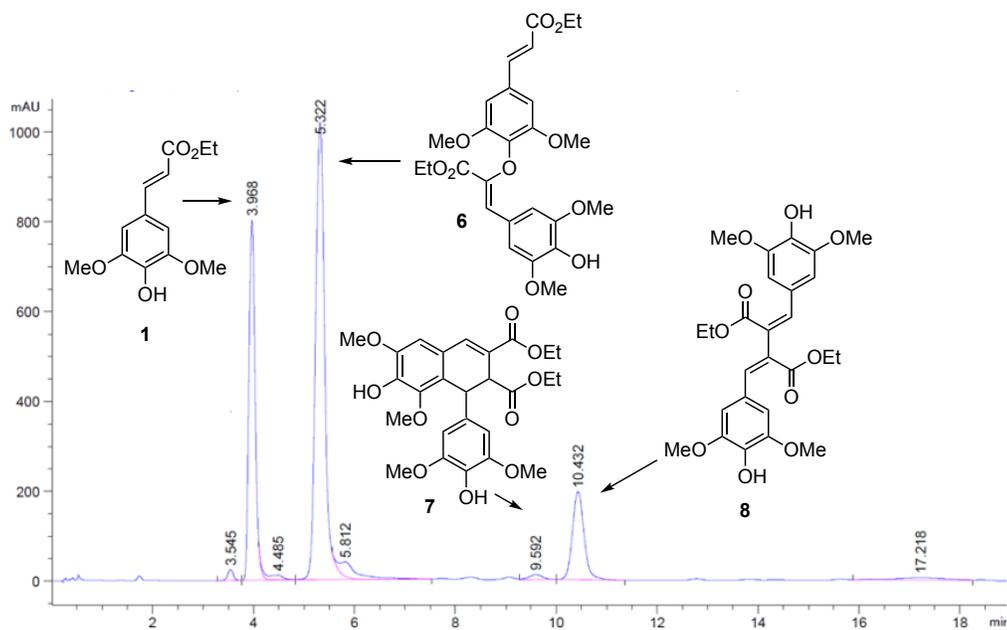


Hydrogenation of ethyl sinapate  $\beta$ - $\beta'$  dehydrodimer (8) led to **ethyl sinapate  $\beta$ - $\beta'$  dihydrodimer (17)** as a light-yellow solid (95% yield). **mp**: 73 – 76 °C;  **$^1\text{H NMR}$**  (300 MHz, DMSO-*d*6):  $\delta$  1.08 (t, 6H,  $J$  = 7.2 Hz,  $\text{H}_7$ ), 2.79 (m, 4H,  $\text{H}_\alpha$ ), 2.89 (m, 2H,  $\text{H}_\beta$ ), 3.68 (s, 12H,  $\text{H}_5$ ), 3.96 (q, 4H,  $J$  = 7.2 Hz,  $\text{H}_6$ ), 6.34 (s, 4H,  $\text{H}_2$ ), 8.16 (s, 2H,  $\text{H}_{\text{phenol}}$ );  **$^{13}\text{C NMR}$**  (75 MHz, DMSO-*d*6):  $\delta$  14.0 ( $\text{C}_7$ ), 35.5 ( $\text{C}_\alpha$ ), 48.6 ( $\text{C}_\beta$ ), 55.8 ( $\text{C}_5$ ), 59.9 ( $\text{C}_6$ ), 106.3 ( $\text{C}_2$ ), 128.4 ( $\text{C}_1$ ), 134.0 ( $\text{C}_4$ ), 147.7 ( $\text{C}_3$ ), 173.0 ( $\text{C}_\gamma$ ); **IR** (FTIR): 3443, 2937, 1720, 1609, 1515, 1457, 1369, 1323, 1207, 1105, 1031  $\text{cm}^{-1}$ ; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  207, 273 nm; **HRMS** ( $m/z$ ) [ $\text{M}+\text{Na}$ ] $^+$  calcd for  $\text{C}_{26}\text{H}_{34}\text{O}_{10}\text{Na}$ , 529.2050; found, 529.2048.



DIBAL-H reduction of ethyl sinapate  $\beta$ - $\beta'$  dehydrodimer (**8**) led to **sinapic alcohol  $\beta$ - $\beta'$  dehydrodimer (18)** as a yellow powder (46% yield). **mp**: 84 – 86 °C;  **$^1\text{H NMR}$**  (300 MHz, DMSO-*d*6):  $\delta$   $\delta_{\text{H}}$  3.63 (s, 12H, H<sub>5</sub>), 3.92 (dd, 4H,  $J$  = 14.4, 3.9 Hz, H <sub>$\gamma$</sub> ), 5.10 (t, 2H,  $J$  = 5.1 Hz, H<sub>alcohol</sub>), 6.56 (s, 2H, H <sub>$\alpha$</sub> ), 6.81 (s, 4H, H<sub>2</sub>), 8.38 (s, 2H, H<sub>phenol</sub>);  **$^{13}\text{C NMR}$**  (75 MHz, DMSO-*d*6):  $\delta$  55.7 (C<sub>5</sub>), 63.6 (C <sub>$\gamma$</sub> ), 105.3 (C<sub>2</sub>), 124.1 (C <sub>$\alpha$</sub> ), 127.2 (C<sub>1</sub>), 134.7 (C<sub>4</sub>), 139.0 (C <sub>$\beta$</sub> ), 147.7 (C<sub>3</sub>); **IR** (FTIR): 3352, 2935, 1595, 1511, 1451, 1320, 1210, 1108, 1018 cm<sup>-1</sup>; **UV/Vis** (Ethanol):  $\lambda_{\text{max}}$  210, 274 nm; **HRMS** ( $m/z$ ) [M+Na]<sup>+</sup> calcd for C<sub>22</sub>H<sub>26</sub>O<sub>8</sub>Na, 441.1525; found, 441.1519

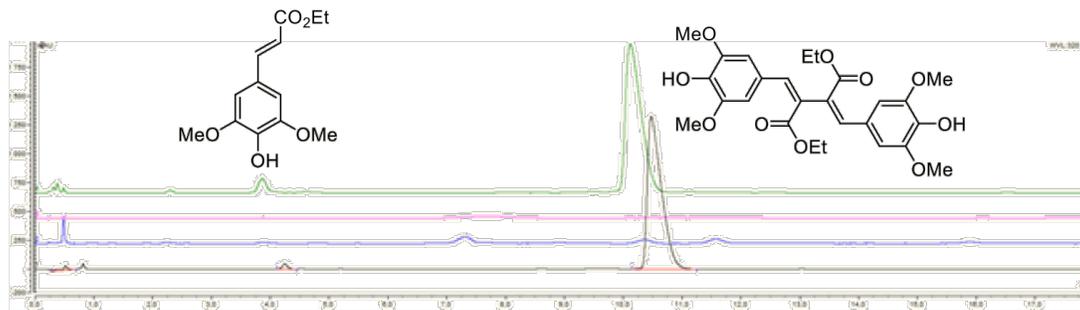
## 4. Enzymatic dimerization of ethyl sinapate



**Figure S1.** HPLC chromatogram for the dimerization of ethyl sinapate with laccase from *Trametes versicolor* (Zorbax Eclipse Plus C18 (2.1 mm\*50 mm\*1.8  $\mu$ m),  $\lambda$  = 320 nm, flow rate set at 0.6 mL.min<sup>-1</sup>, oven temperature at 30 °C, and gradient applied: H<sub>2</sub>O/CH<sub>3</sub>CN from 75/25 to 70/30 in 18 min).

## 5. Optimization of the sinapate ester $\beta$ - $\beta'$ dimerization

### 5.1. Screening of solvents



**Figure S2.** HPLC chromatograms of the dimerization of ethyl sinapate in pyridine (black), ethanol (pink), ethyl acetate (blue) and Cyrene™ (green) (Zorbax Eclipse Plus C18 (2.1 mm\*50 mm\*1.8  $\mu$ m),  $\lambda$  = 320 nm, flow rate set at 0.6 mL.min<sup>-1</sup>, oven temperature at 30 °C, and gradient applied: H<sub>2</sub>O/CH<sub>3</sub>CN from 75/25 to 70/30 in 18 min).

### 5.2. Design of experiments (D.O.E.)

The effect of the four independent variables  $X_1$  (Temperature),  $X_2$  (ratio Catalyst/Substrate),  $X_3$  (Ratio Amine/Catalyst) and  $X_4$  (Concentration) on responses  $Y_1$  (yield) and  $Y_2$  (conversion) was modelled using a polynomial response:

$$Y_i = \beta_0 + \sum_{i=1}^4 \beta_{ki}x_i + \sum_{i=1}^4 \beta_{kii}x_i^2 + \sum_{i=1}^4 \sum_{j=i+1}^4 \beta_{kij}x_i x_j$$

where  $Y_i$  represents the response  $i$ ,  $x_i$  are the coded independent variables,  $\beta_0$  is a constant coefficient, and  $\beta_{ki}$ ,  $\beta_{kii}$ , and  $\beta_{kij}$  are the linear, quadratic, and interaction coefficients, respectively.

The variable levels  $X_i$  were scaled and centered (coded) according to the equation below such that  $X_0$  corresponded to the central value:

$$x_i = \frac{X_i - X_0}{\Delta X_i} \text{ where } i = 1, 2, 3, \dots, k$$

where  $x_i$  is the dimensionless value of an independent variable,  $X_i$  is the real value of an independent variable,  $X_0$  is the real value of an independent variable at the centre point, and  $\Delta X_i$  is the step change.

**Table S1** | Design of experiments for the  $\beta$ - $\beta'$  dimerization of ethyl sinapate (experimental values obtained by HPLC)

Exp No	Exp Name	Run Order	Incl/Excl	Temperature (°C)	Ratio Cat/Substrate	Ratio amine/cat	Concentration (mol.L <sup>-1</sup> )	Yield (%)	Conversion (%)
1	N1	3	Incl	25	2.01	1	0.249	0.01	0.01
2	N2	14	Incl	70	2.03	1	0.247	0.01	7.58
3	N3	15	Incl	25	10.1	1	0.249	0.01	0.01
4	N4	27	Incl	70	10	1	0.25	0.01	0.01
5	N5	17	Incl	25	2.01	10	0.249	0.01	0.01
6	N6	4	Incl	70	2.01	10.1	0.246	1.63	3.26
7	N7	7	Incl	25	9.95	10.02	0.251	2.84	5.68
8	N8	23	Incl	70	10.2	10	0.246	16.83	20.71
9	N9	9	Incl	25	1.99	1.01	2.49	0.01 <sup>a</sup>	33.31
10	N10	10	Incl	70	2.01	1.02	2.447	3.41	59.34
11	N11	1	Incl	25	10.1	1	2.49	12.27	81.05
12	N12	11	Incl	70	10.2	1	2.447	4.22	83.89
13	N13	24	Incl	25	2.02	10	2.48	3.88	27.71
14	N14	5	Incl	70	2	10.07	2.487	34.49	97.08
15	N15	12	Incl	25	9.91	10.02	2.52	35.49	94.8
16	N16	20	Incl	70	10.3	9.99	2.428	26.23	99.99
17	N17	13	Incl	25	6.02	5.5	1.358	34.99	56.7
18	N18	26	Incl	70	6.06	5.49	1.358	40.78	98.37
19	N19	21	Incl	47.5	1.99	5.51	1.374	55.79	64.59
20	N20	16	Incl	47.5	10.47	5.3	1.358	69.77	99.99
21	N21	25	Incl	47.5	5.99	1	1.368	7.32	88.61
22	N22	22	Incl	47.5	6.02	10.02	1.363	53.29	79.64
23	N23	8	Incl	47.5	6.06	5.5	0.248	5.41	11.18
24	N24	6	Incl	47.5	6	5.51	2.497	49.87	98.69
25	N25	18	Incl	47.5	5.98	5.51	1.374	89.05	98.02
26	N26	19	Incl	47.5	6.02	5.51	1.363	83.3	95.4
27	N27	2	Incl	47.5	6.03	5.51	1.363	81.46	97.01
28	N28	28	Incl	70	2.01	10.1	0.246	0.01	28.68
29	N29	29	Incl	70	2	10.07	2.487	35.7	88.56
30	N30	30	Incl	25	9.91	10.02	2.52	61.11	95
31	N31	31	Incl	25	6.02	5.5	1.358	57.5	75.11
32	N32	32	Incl	47.5	6.02	10.02	1.363	53.02	60.59

<sup>a</sup> abnormal value excluded of the D.O.E for the yield optimization

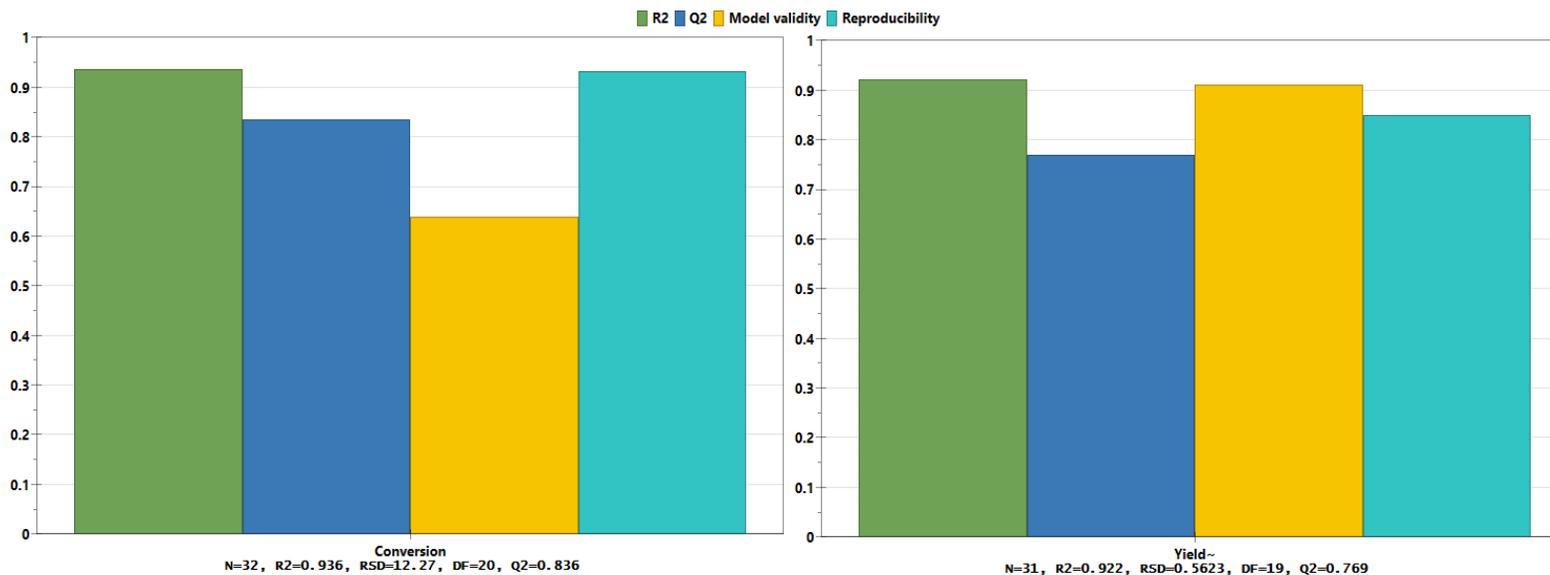


Figure S3 | Summary of fit for conversion (left) and yield (right)

Table S2 | Analysis of the variance (ANOVA) for the logit model of the conversion

Source of variation	Degrees of freedom	Sum of squares	Mean square	Standard deviation	Significance
Regression	11	44155	40414	63.357	0 <sup>a</sup>
Residual	19	3012	150.6	12.272	
Lack of Fit	12	2298	176.8	13.298	0.237 <sup>b</sup>
Pure error	7	713.25	101.89	10.094	
R <sup>2</sup> /R <sup>2</sup> adj <sup>c</sup>	0.933/0.876				
Q <sup>2</sup>	0.839				

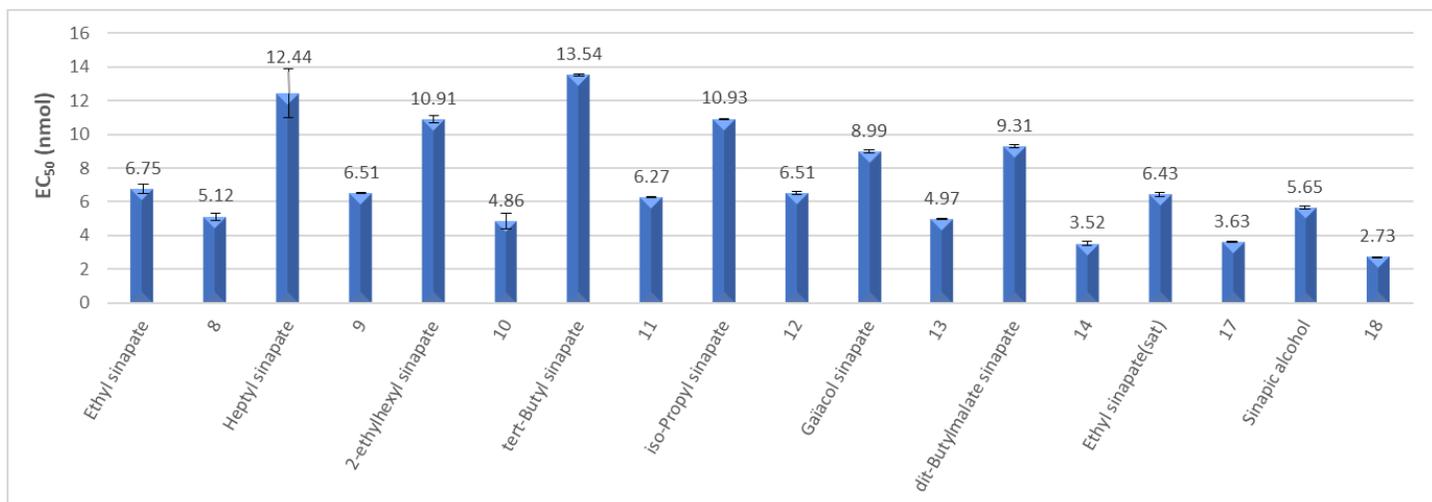
<sup>a</sup> Significance at the 95 % level; <sup>b</sup> lack of fit; <sup>c</sup> R<sup>2</sup> adjusted for degree of freedom

Table S3 | Analysis of the variance (ANOVA) for the logit model of the yield

Source of variation	Degrees of freedom	Sum of squares	Mean square	Standard deviation	Significance
Regression	11	70.675	6.425	2.5348	0 <sup>a</sup>
Residual	19	6.0076	0.3162	0.5623	
Lack of Fit	12	3.3232	0.2769	0.5262	0.704 <sup>b</sup>
Pure error	7	2.6844	0.3839	0.6193	
R <sup>2</sup> /R <sup>2</sup> adj <sup>c</sup>	0.933/0.876				
Q <sup>2</sup>	0.769				

<sup>a</sup> Significance at the 95 % level; <sup>b</sup> lack of fit; <sup>c</sup> R<sup>2</sup> adjusted for degree of freedom

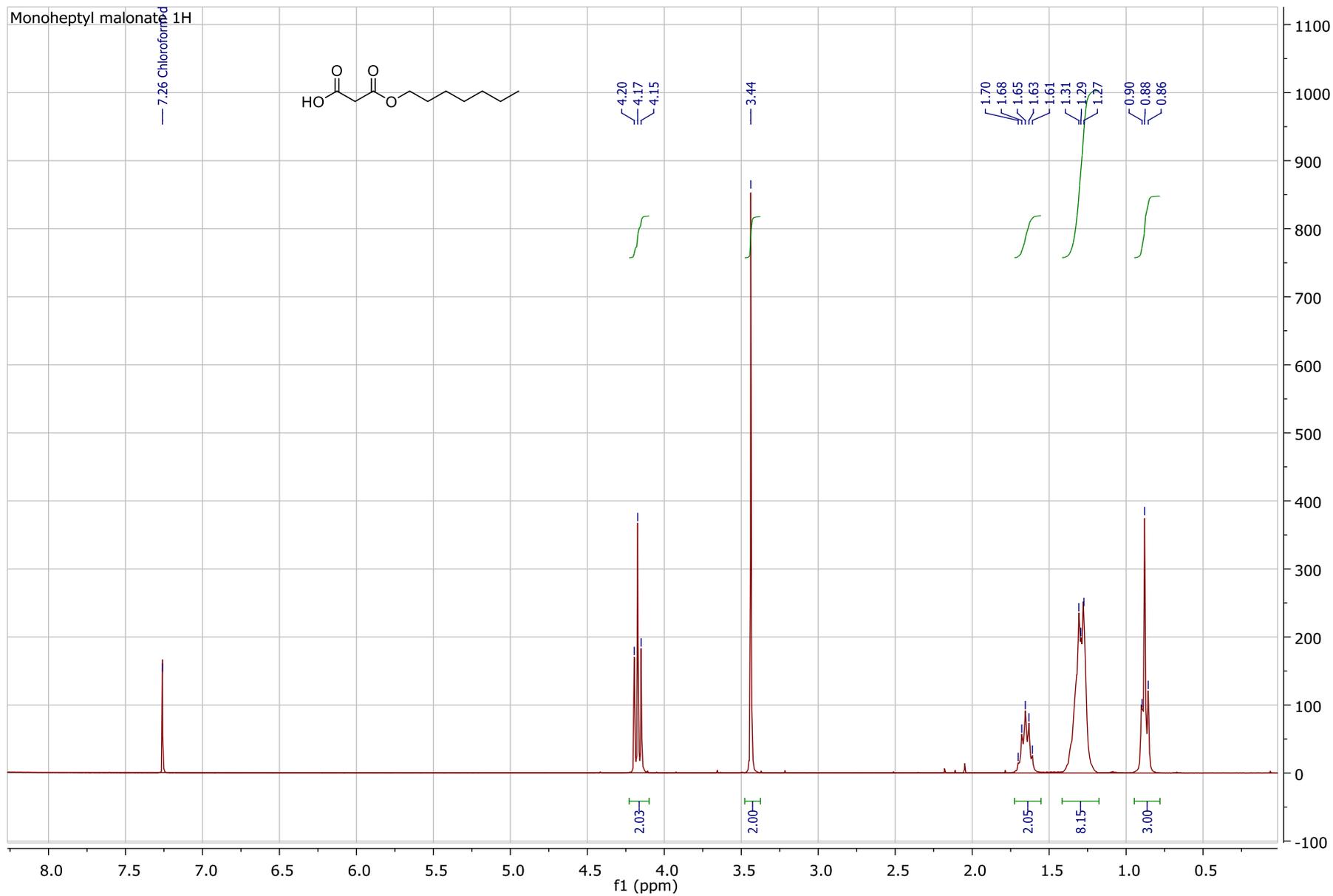
## 6. Antiradical properties



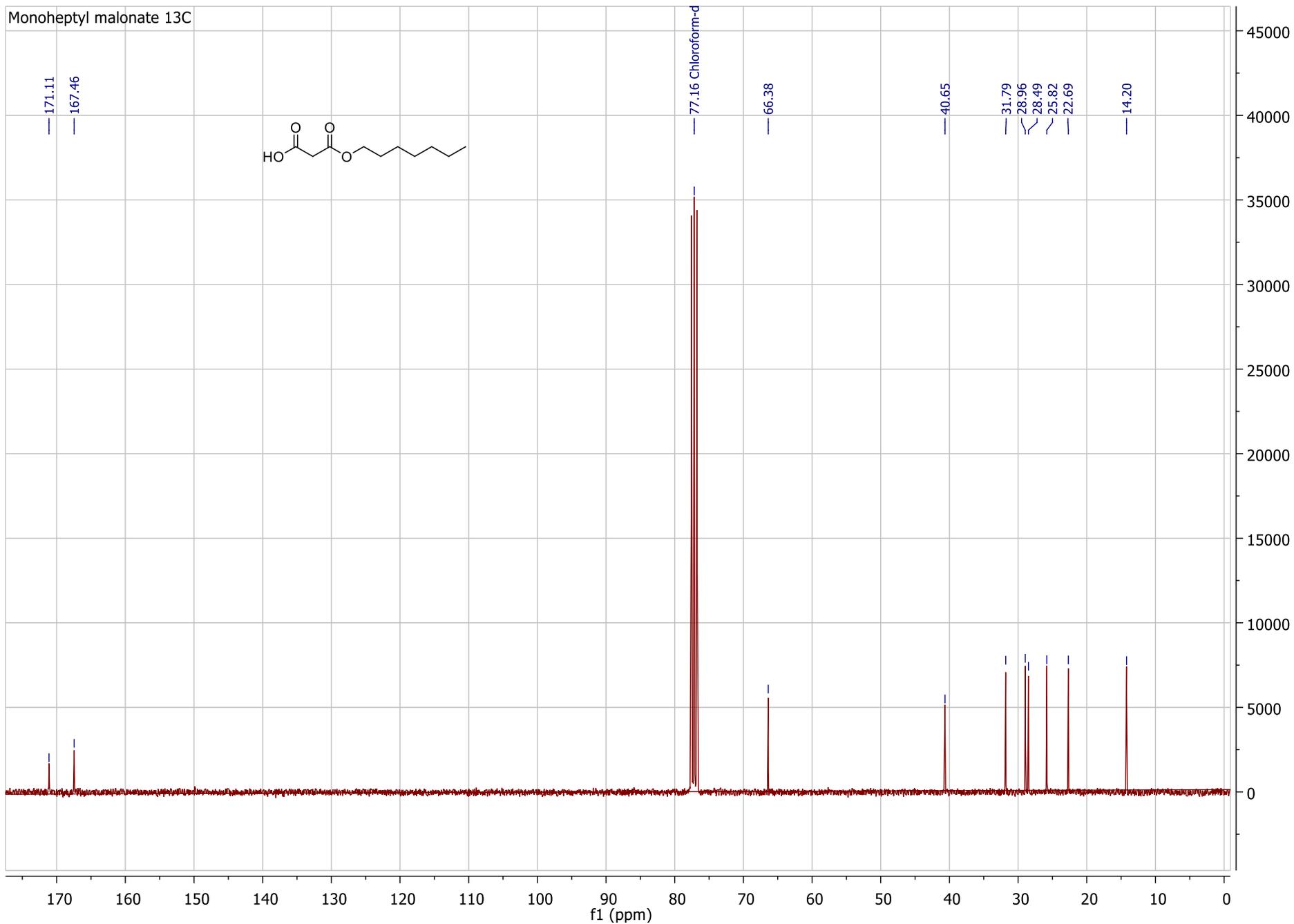
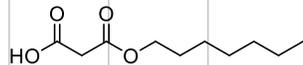
**Figure S4** | Antiradical properties from the ABTS<sup>•+</sup> assay: comparison of EC<sub>50</sub> between sinapate esters and their  $\beta$ - $\beta'$  dimers.

## 7. Characterization Data

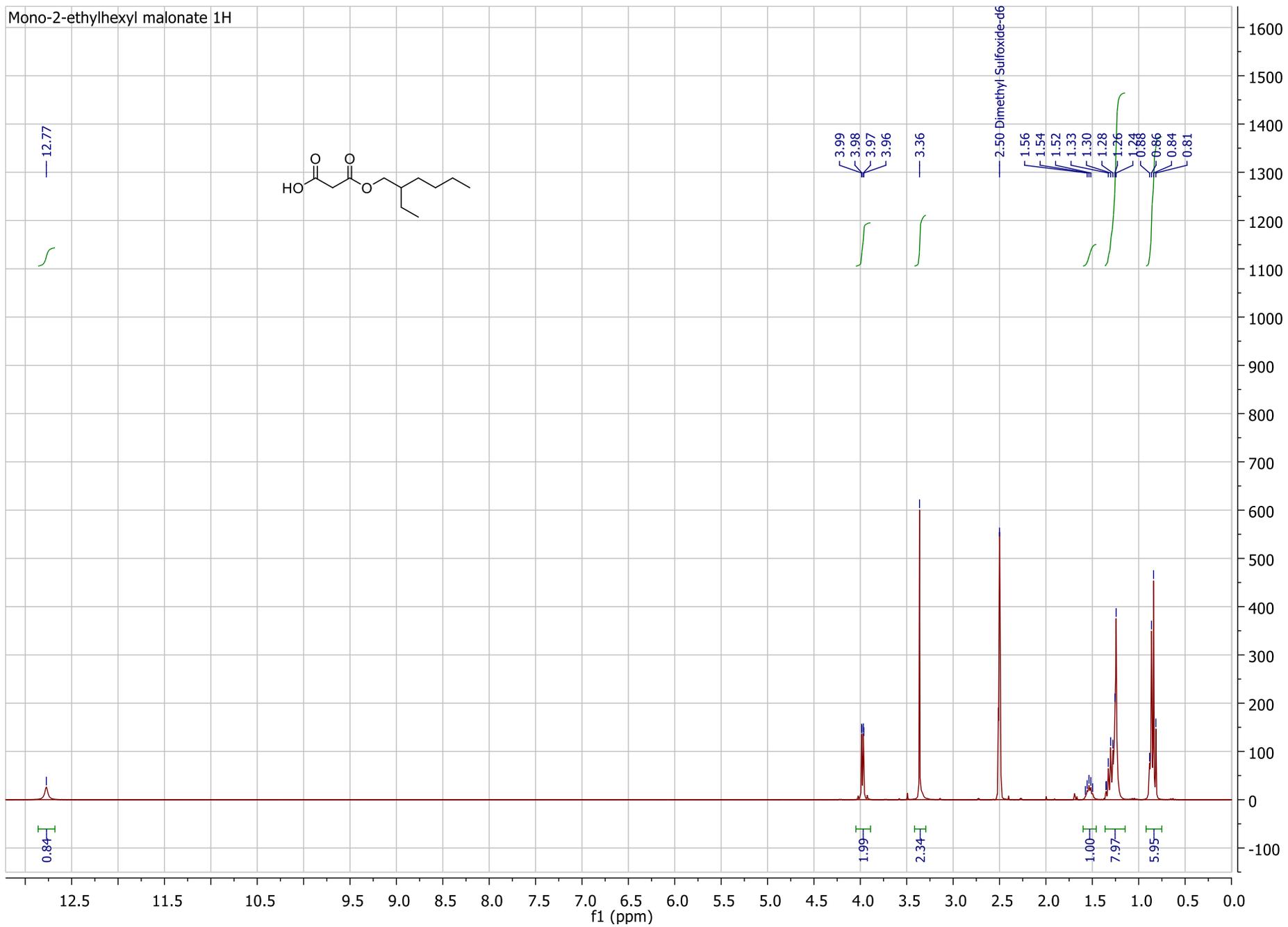
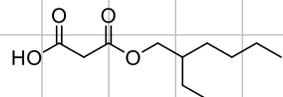
### 7.1. $^1\text{H}$ & $^{13}\text{C}$ NMR spectra



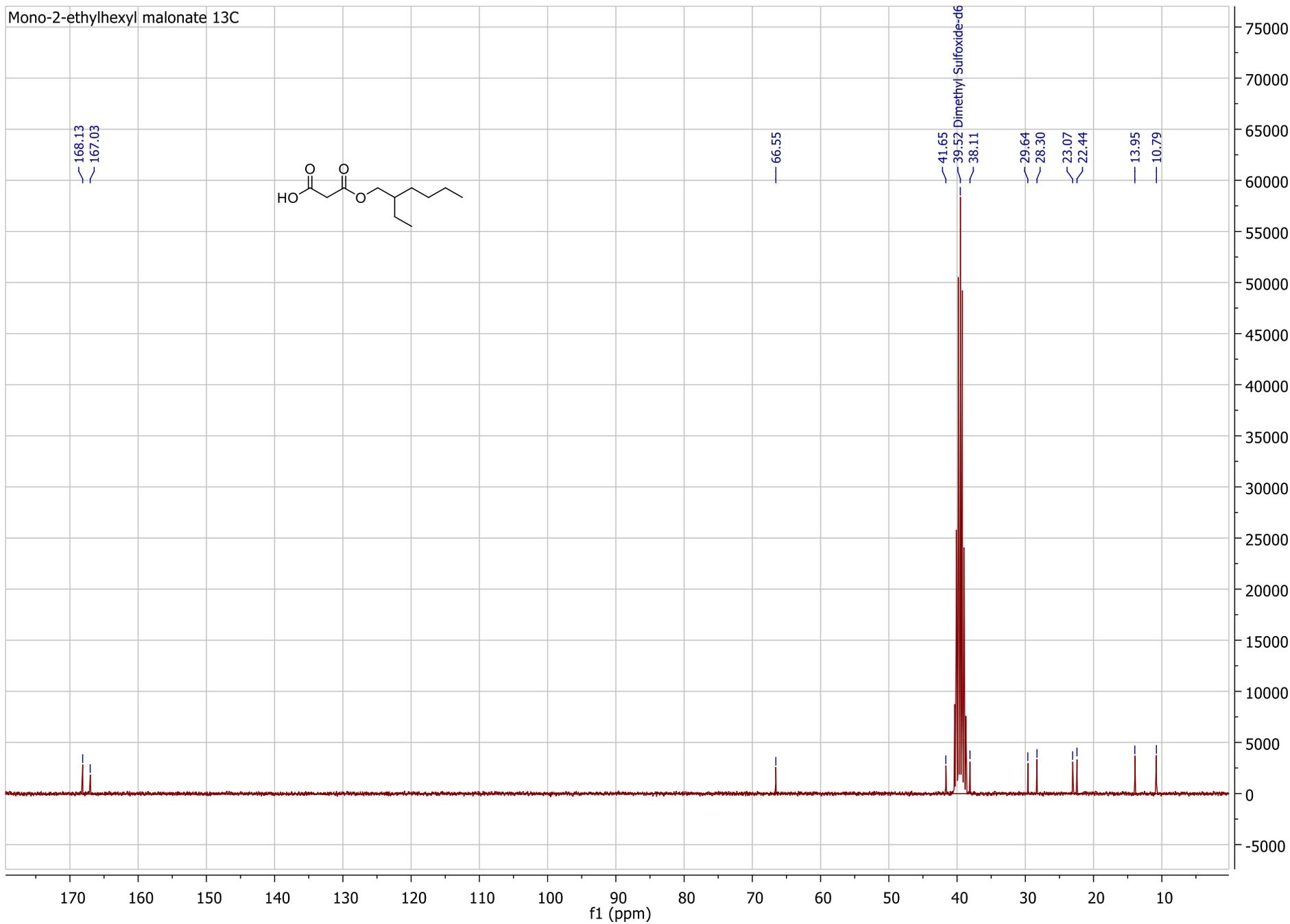
Monoheptyl malonate 13C



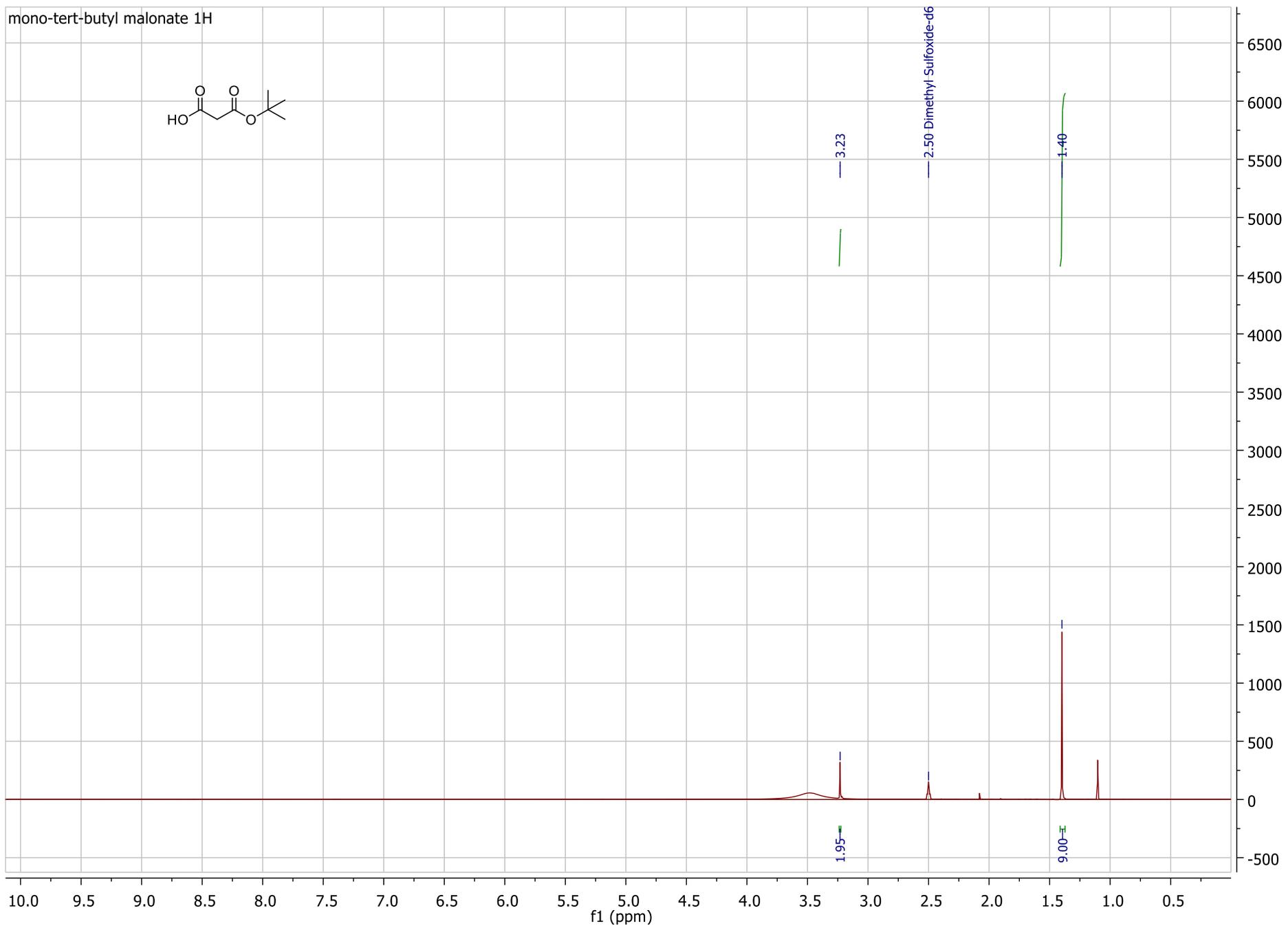
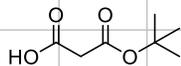
Mono-2-ethylhexyl malonate 1H



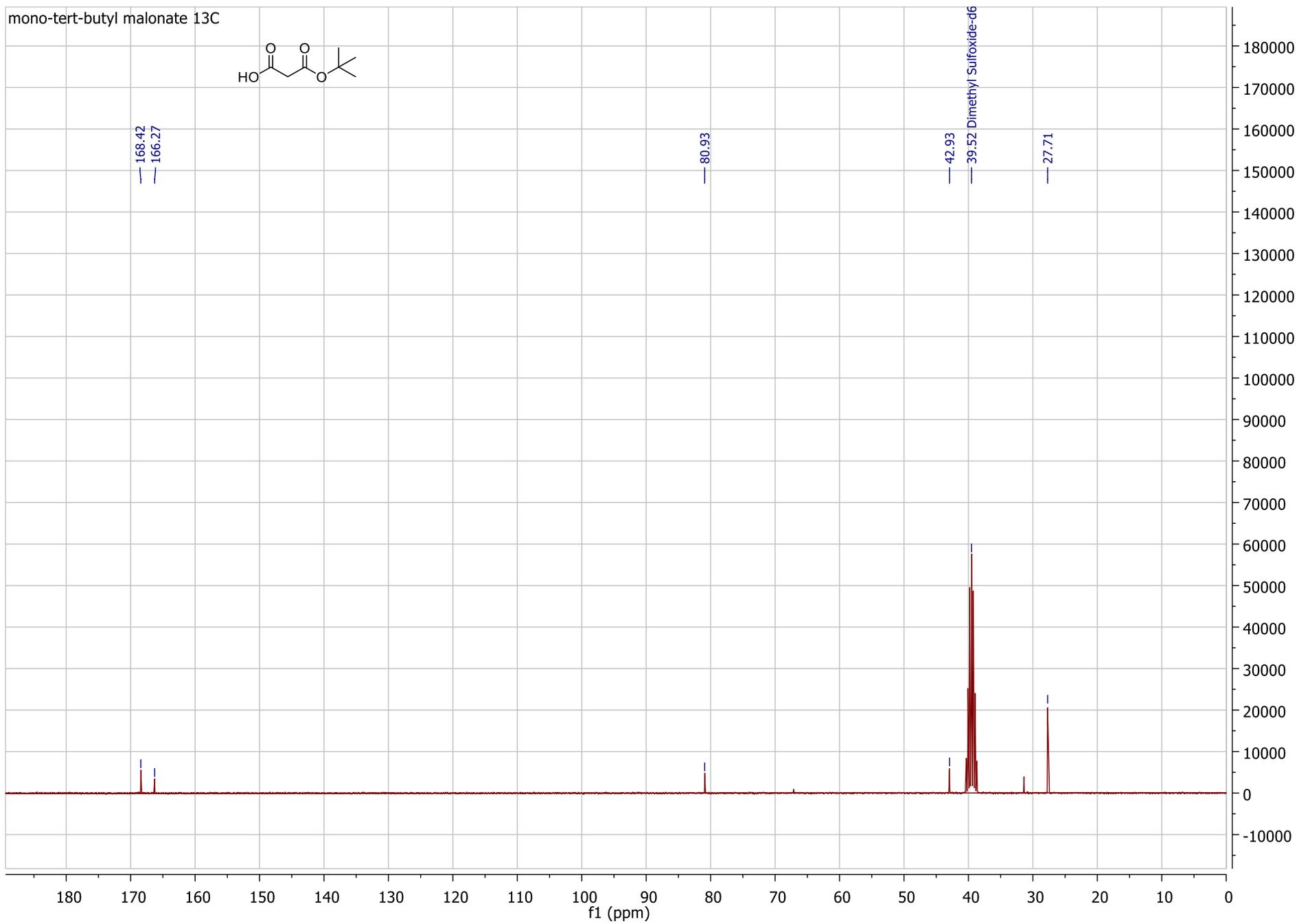
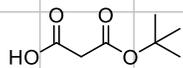
Mono-2-ethylhexyl malonate 13C



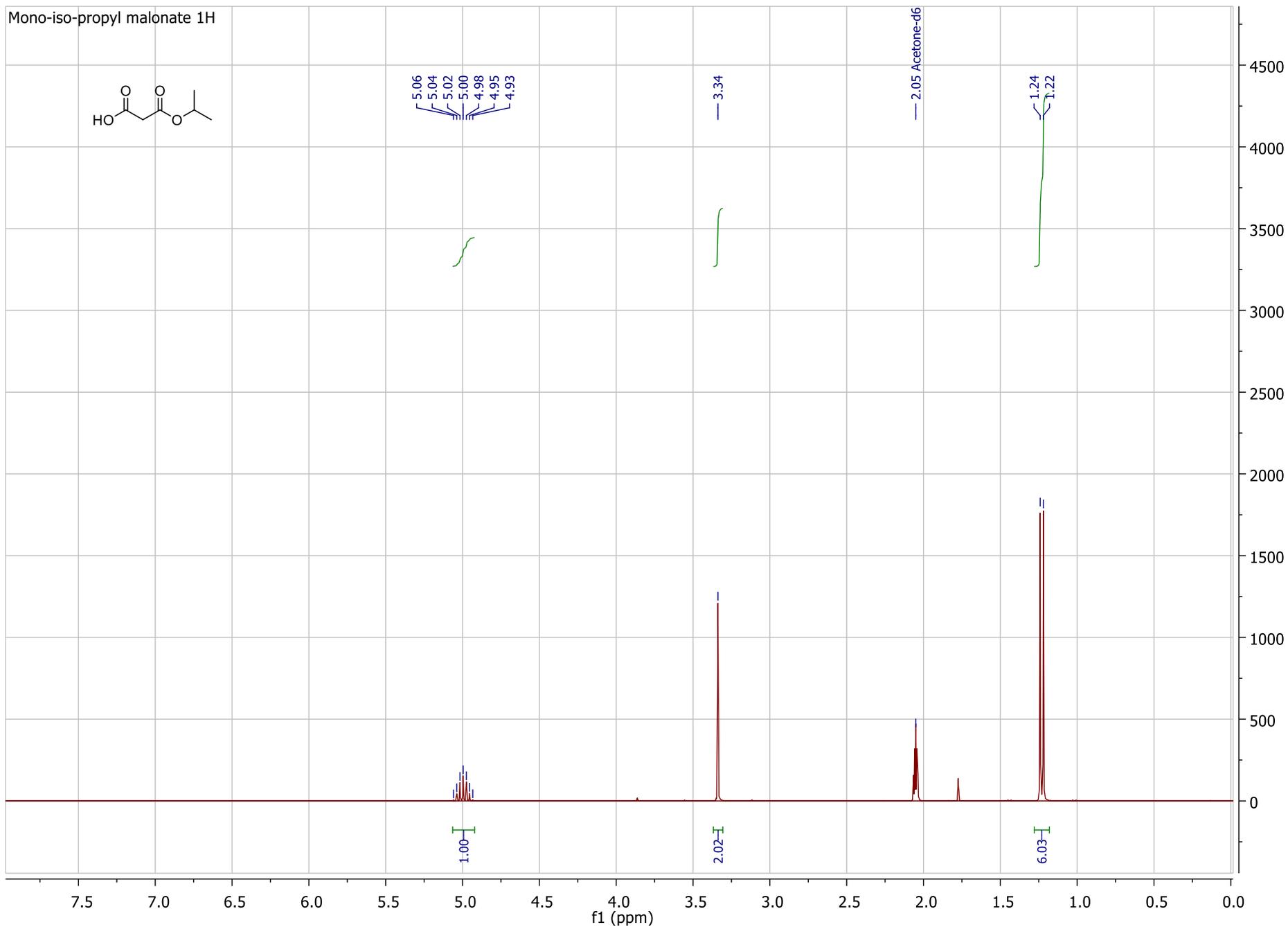
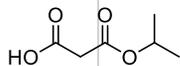
mono-tert-butyl malonate 1H



mono-tert-butyl malonate 13C

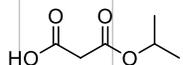


Mono-iso-propyl malonate 1H



Mono-iso-propyl malonate 13C

168.14  
167.04

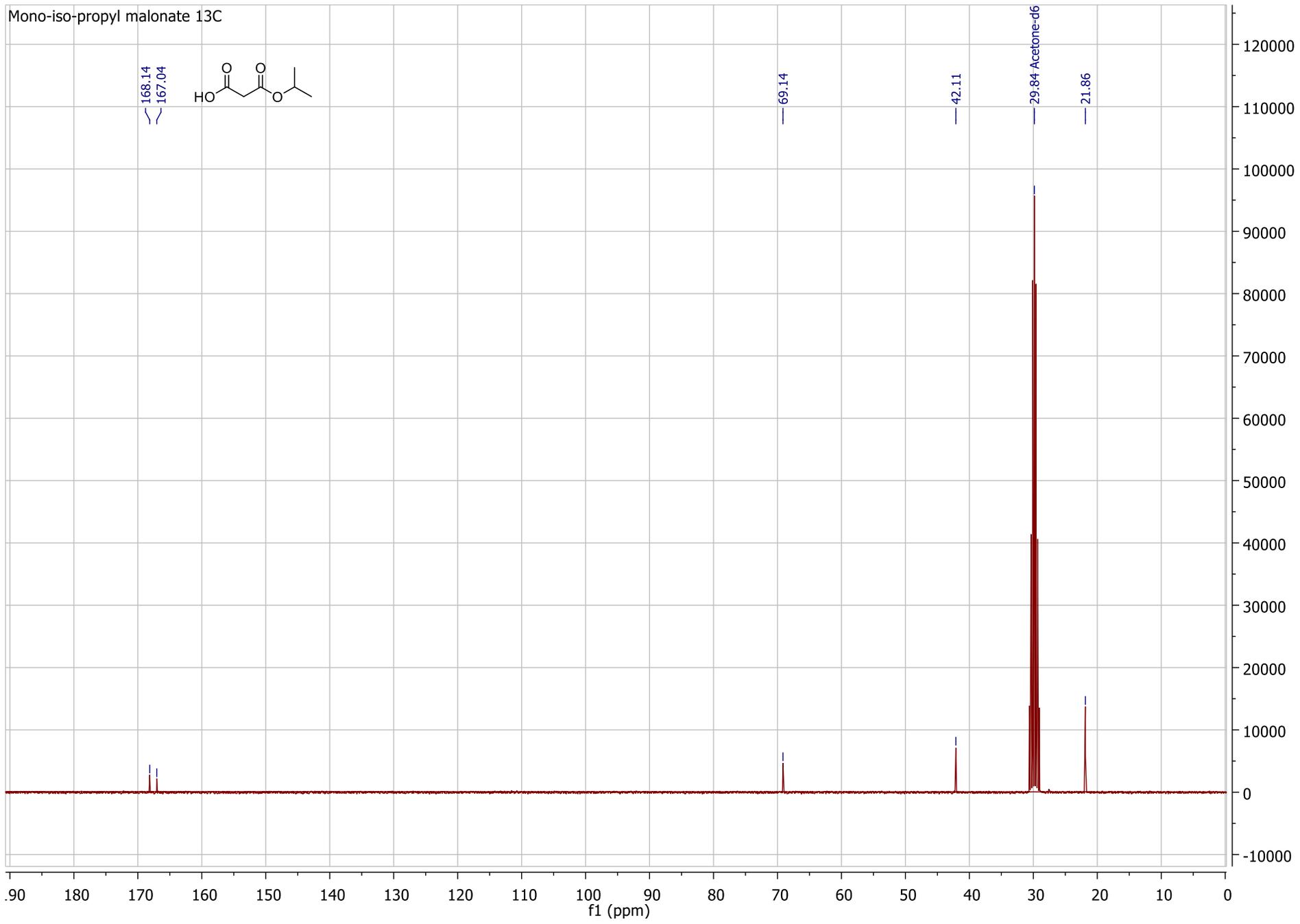


69.14

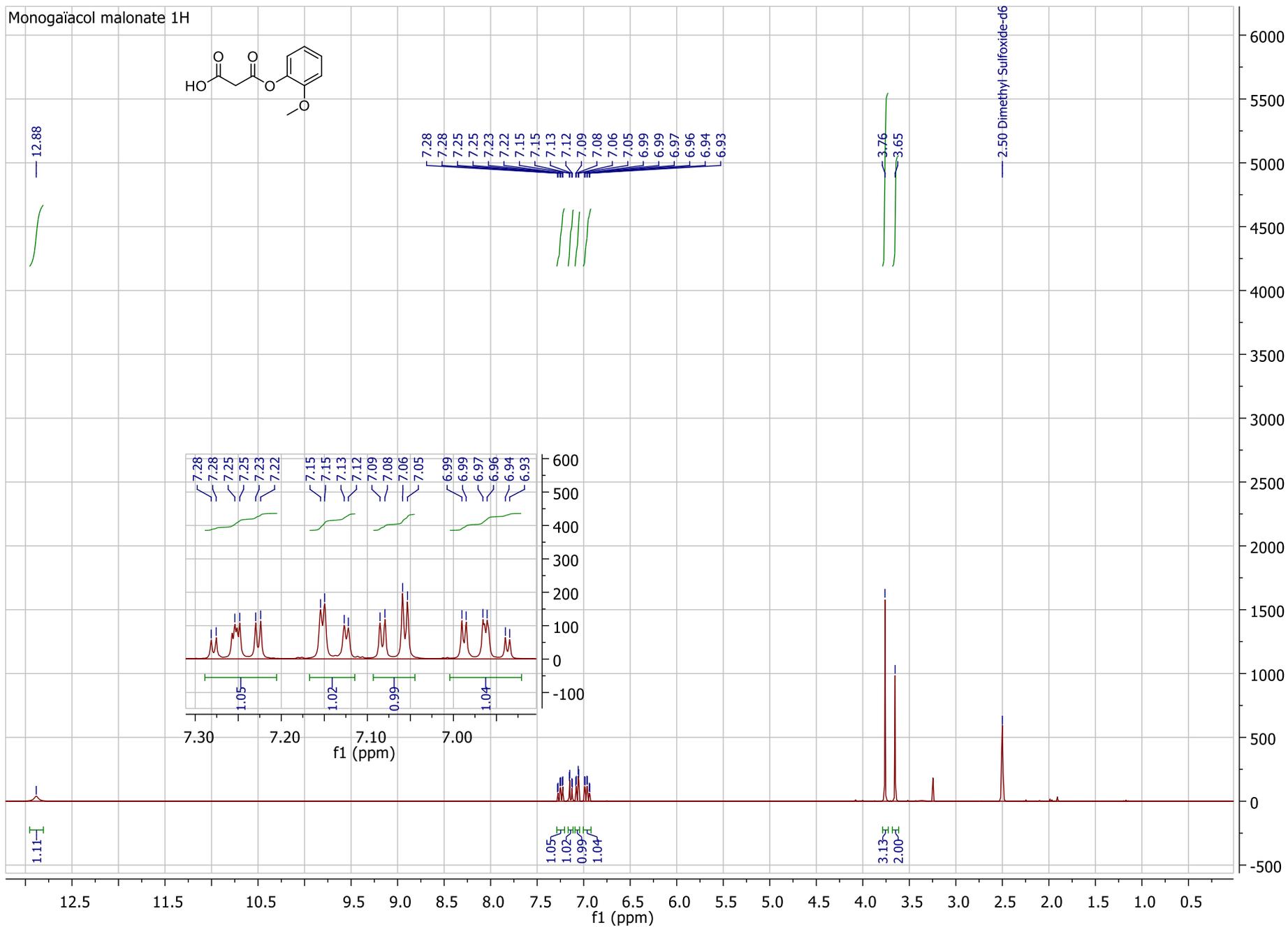
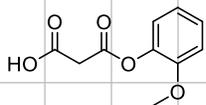
42.11

29.84 Acetone-d6

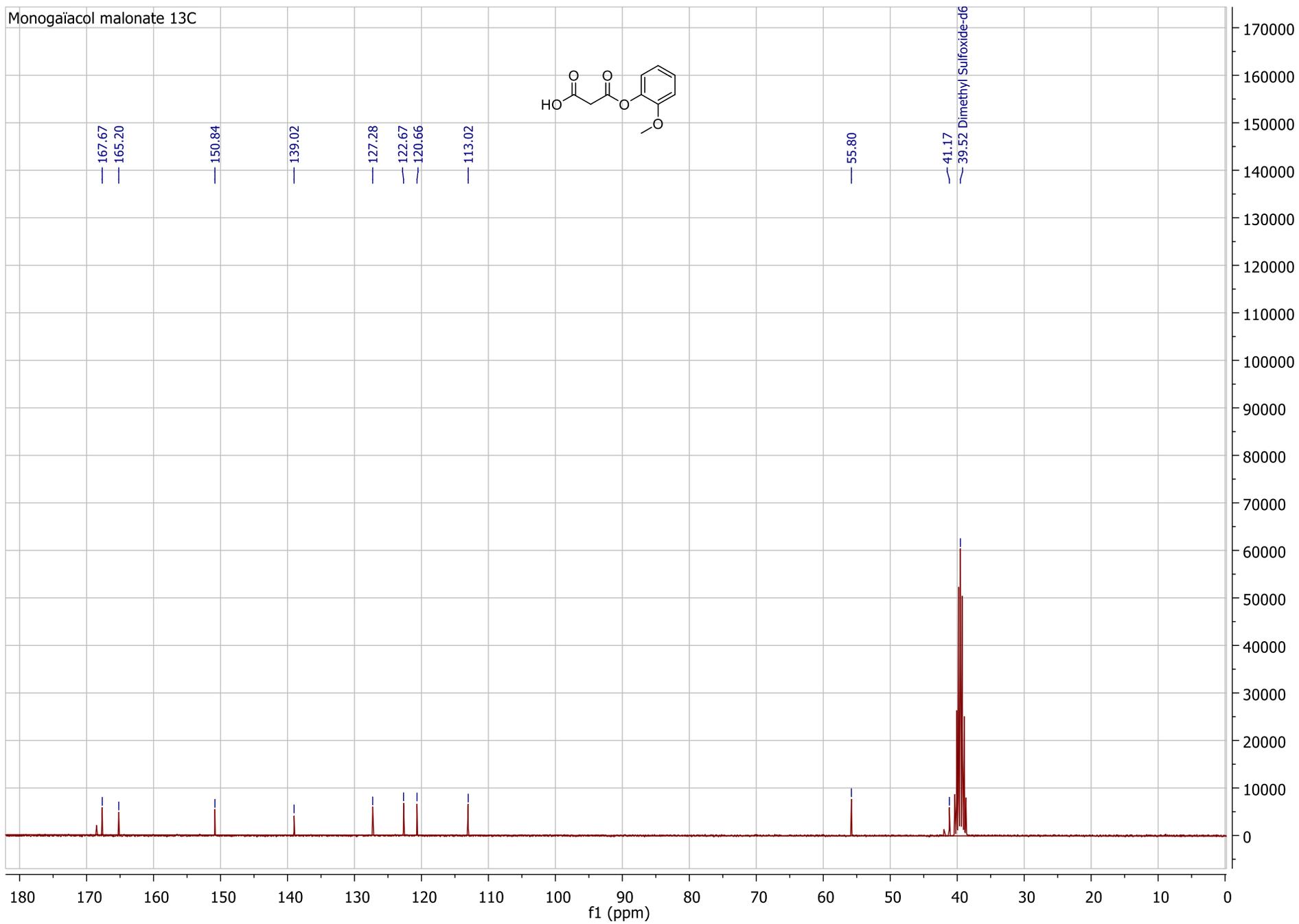
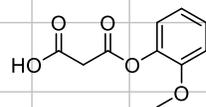
21.86



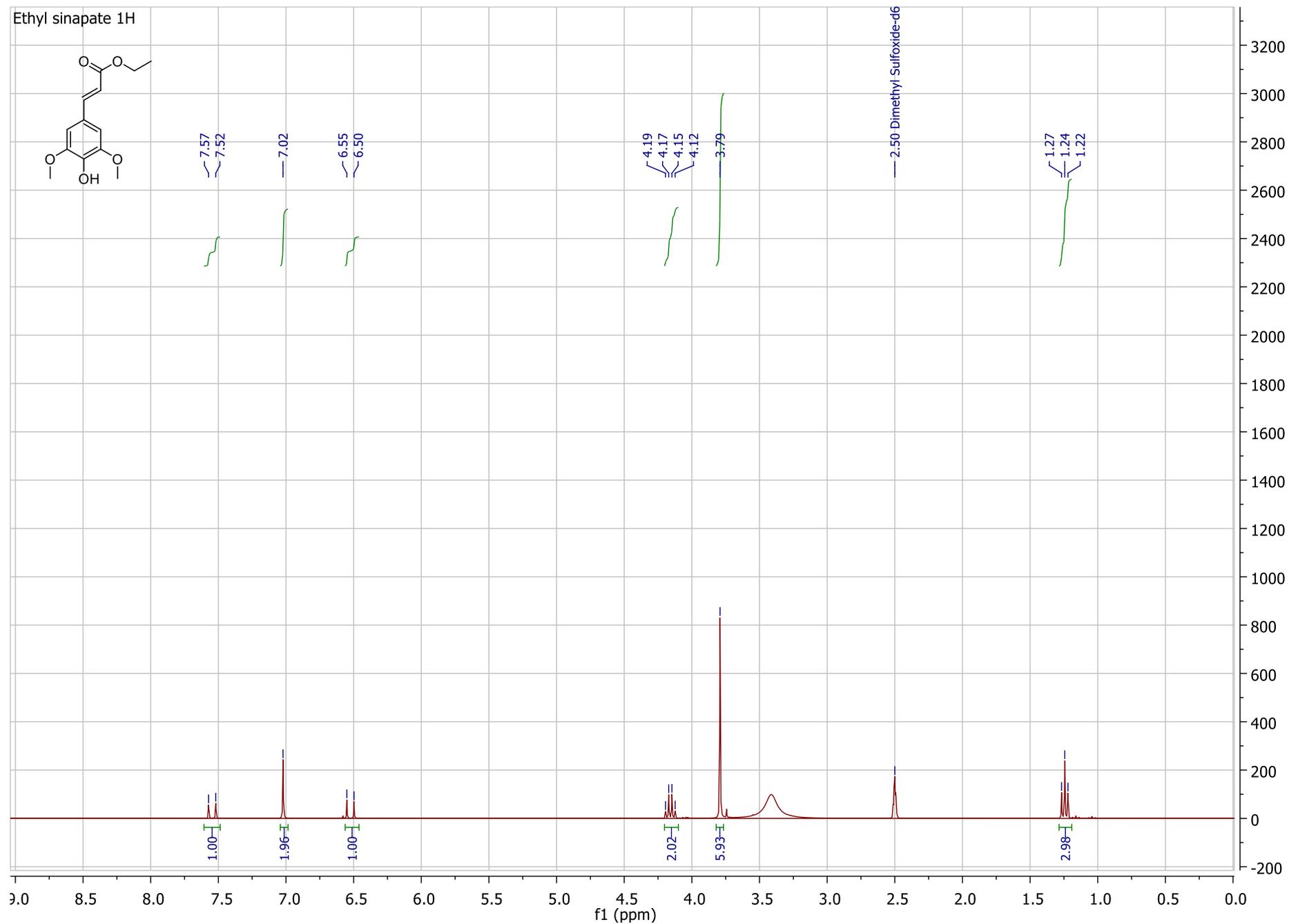
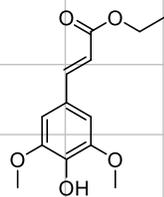
Monogäicöcol malonate 1H



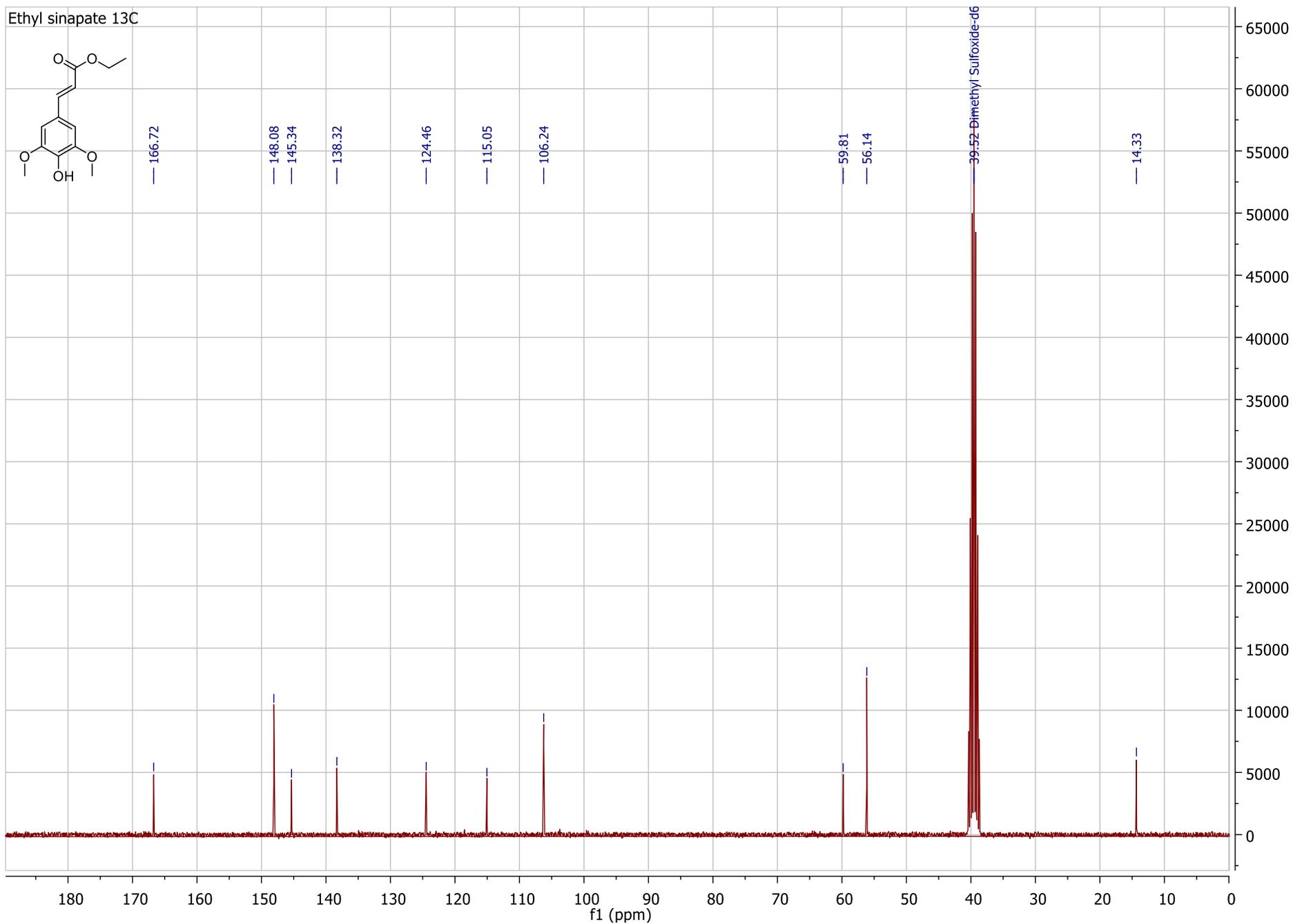
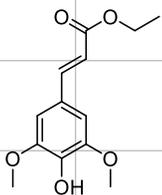
Monogaiacol malonate 13C



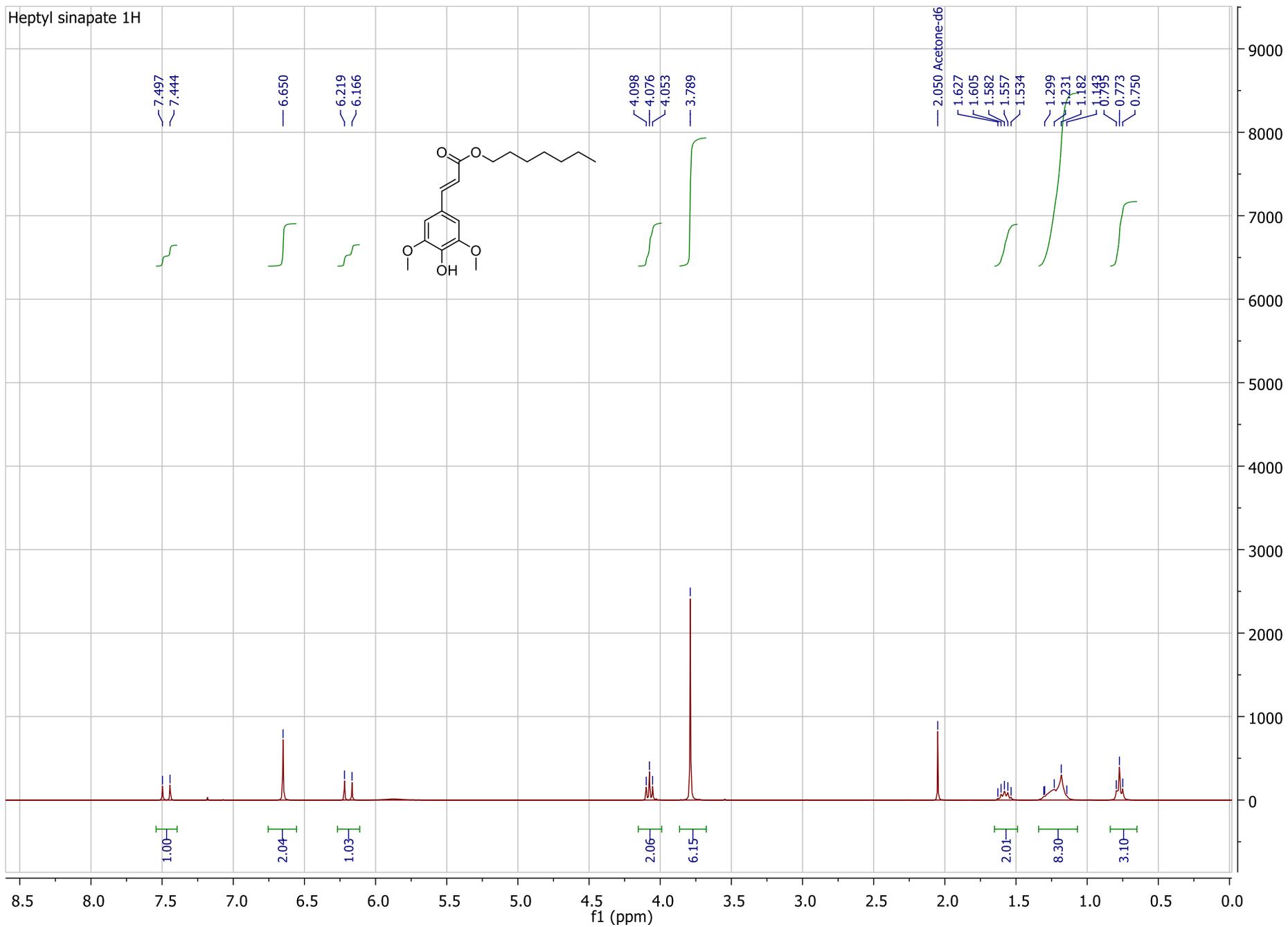
Ethyl sinapate 1H



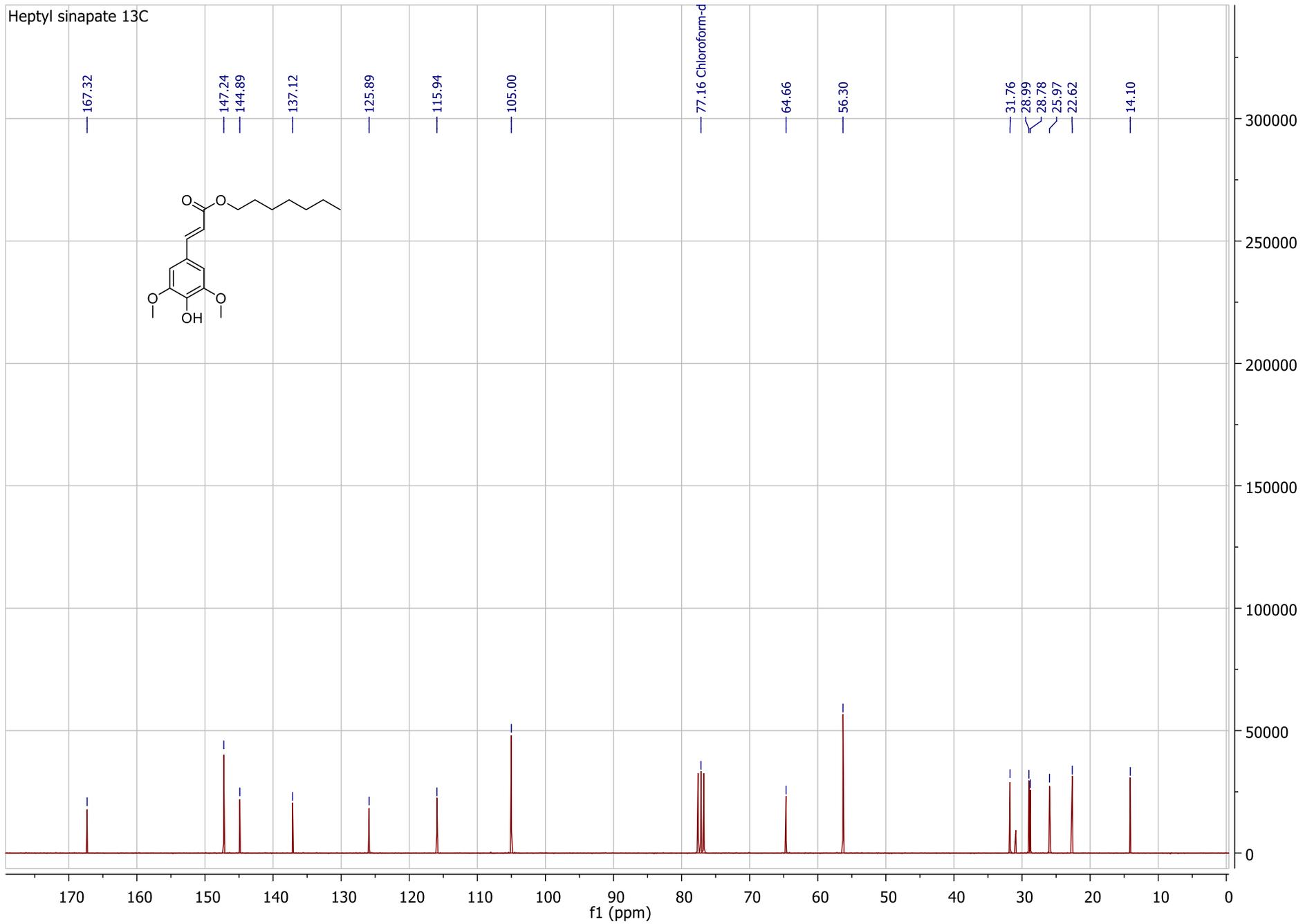
Ethyl sinapate 13C



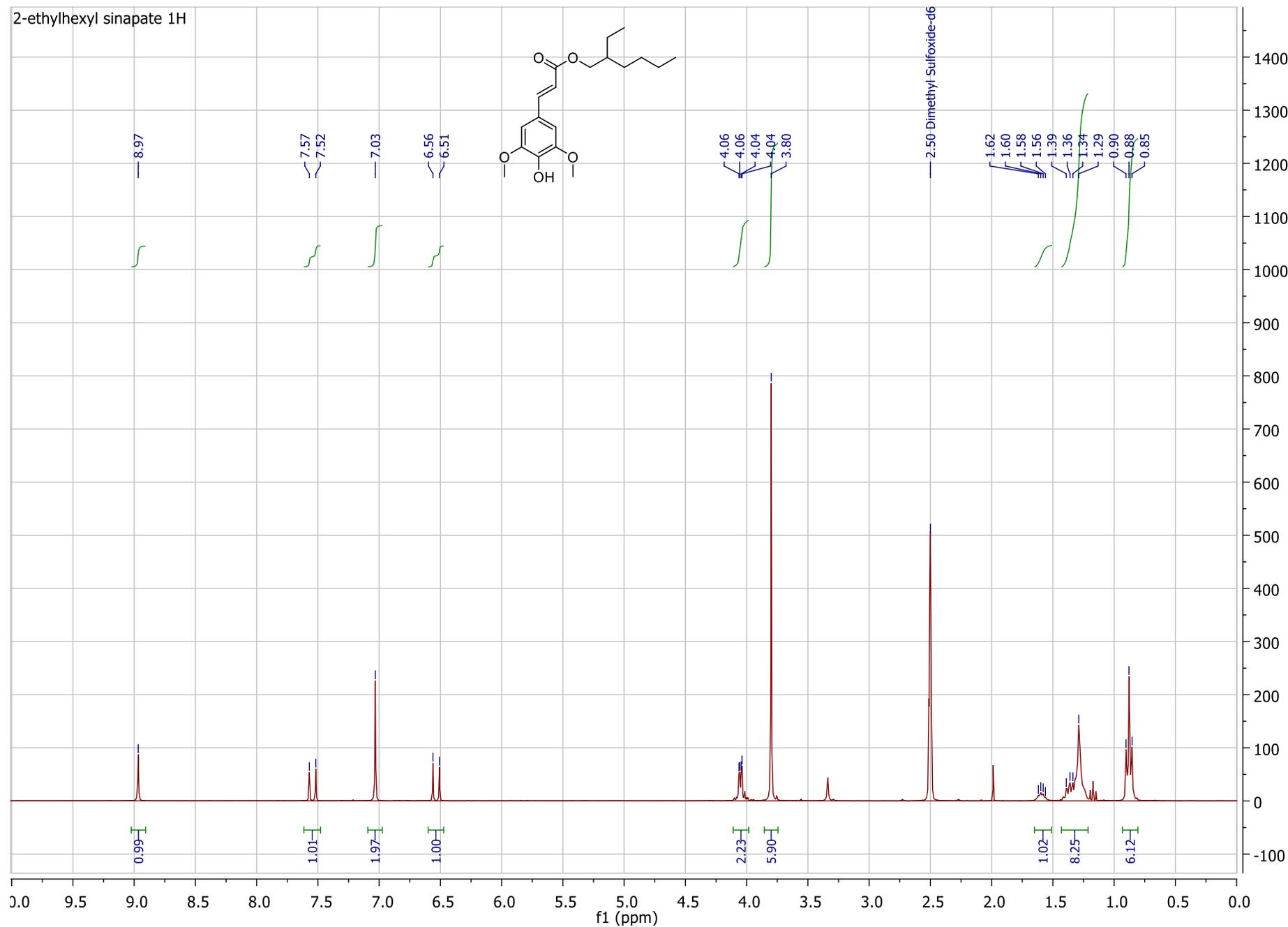
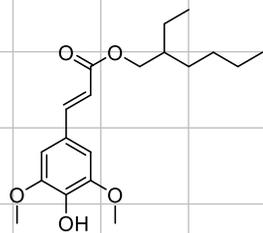
Heptyl sinapate 1H



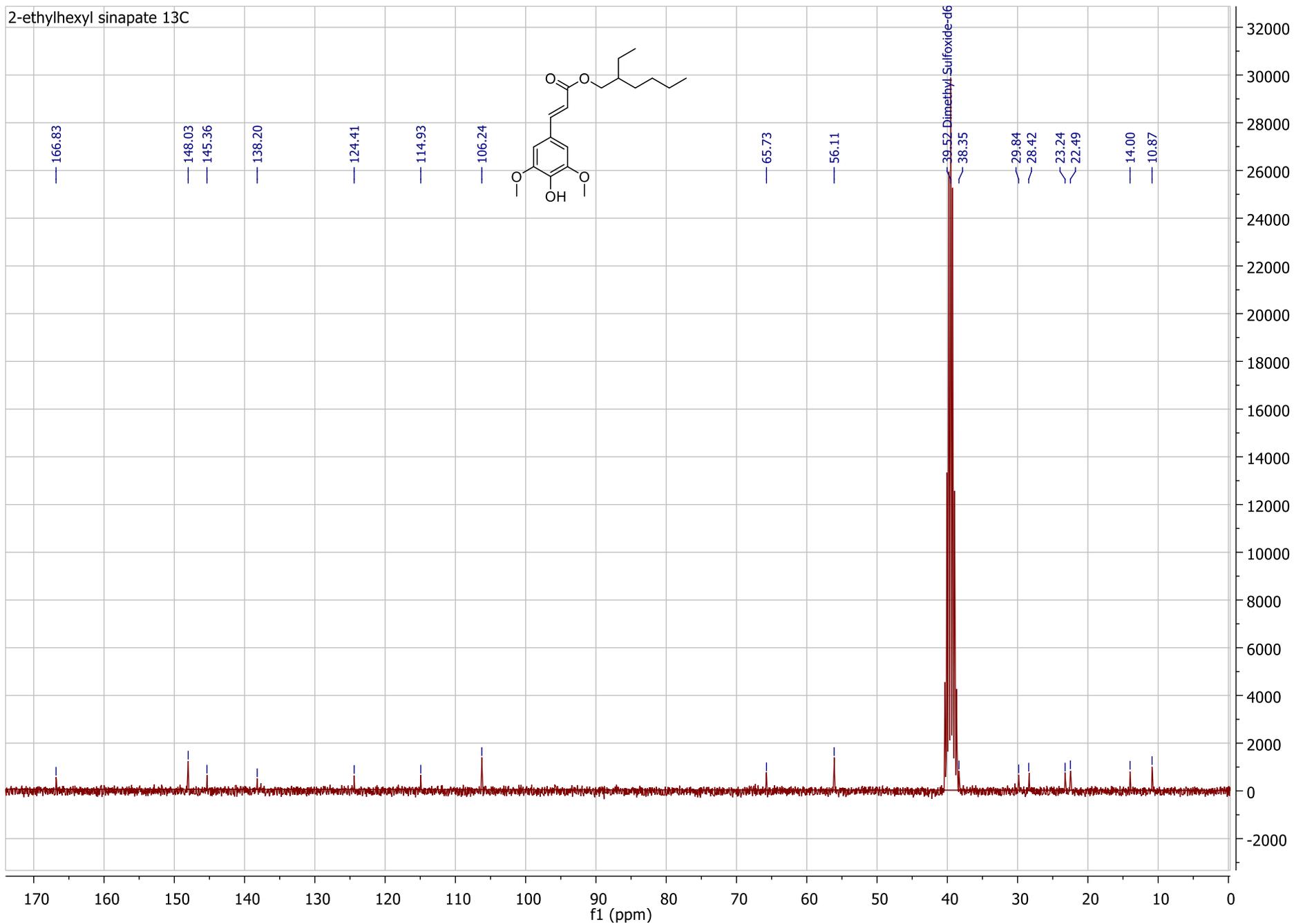
Heptyl sinapate 13C



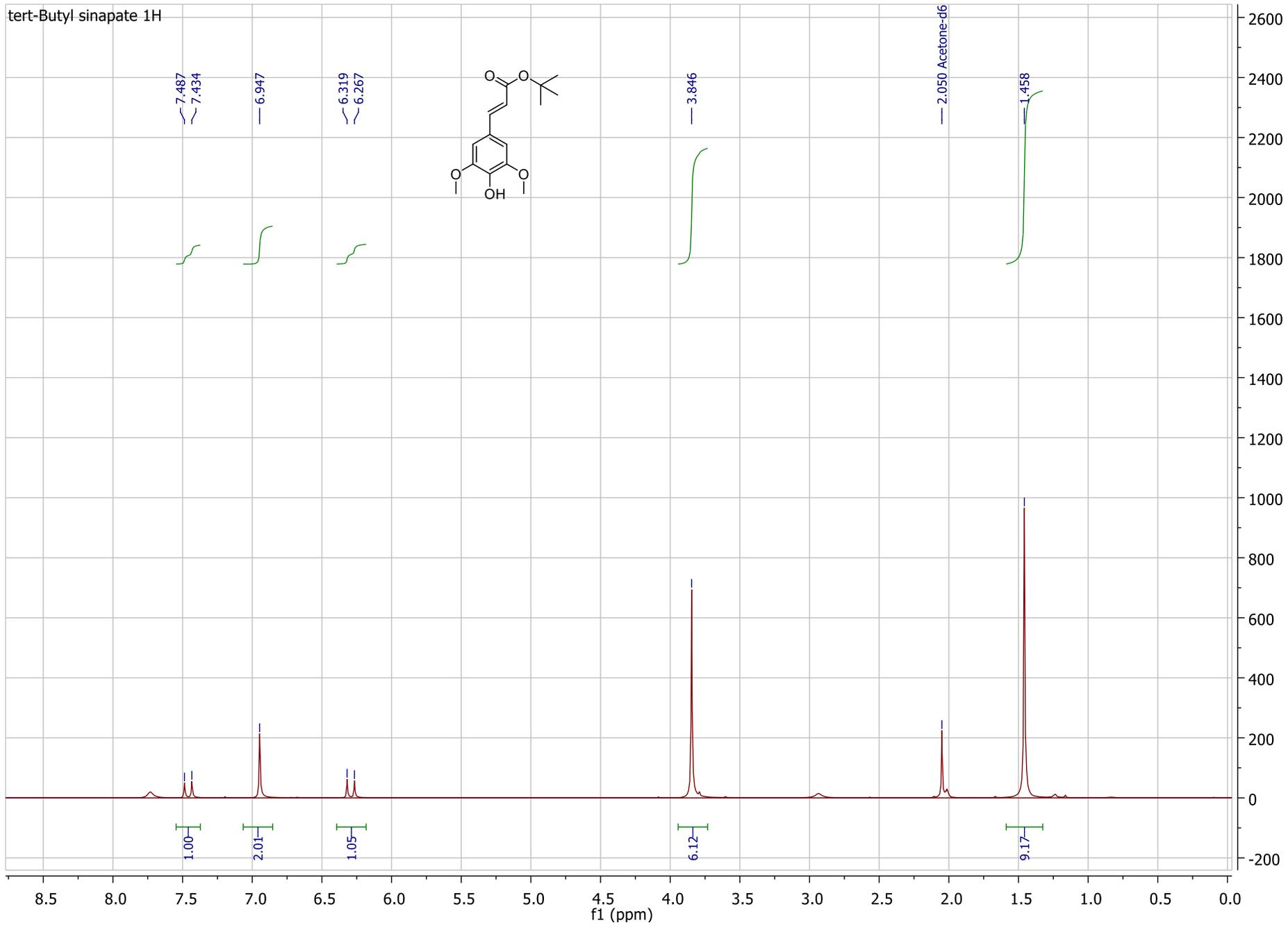
2-ethylhexyl sinapate 1H



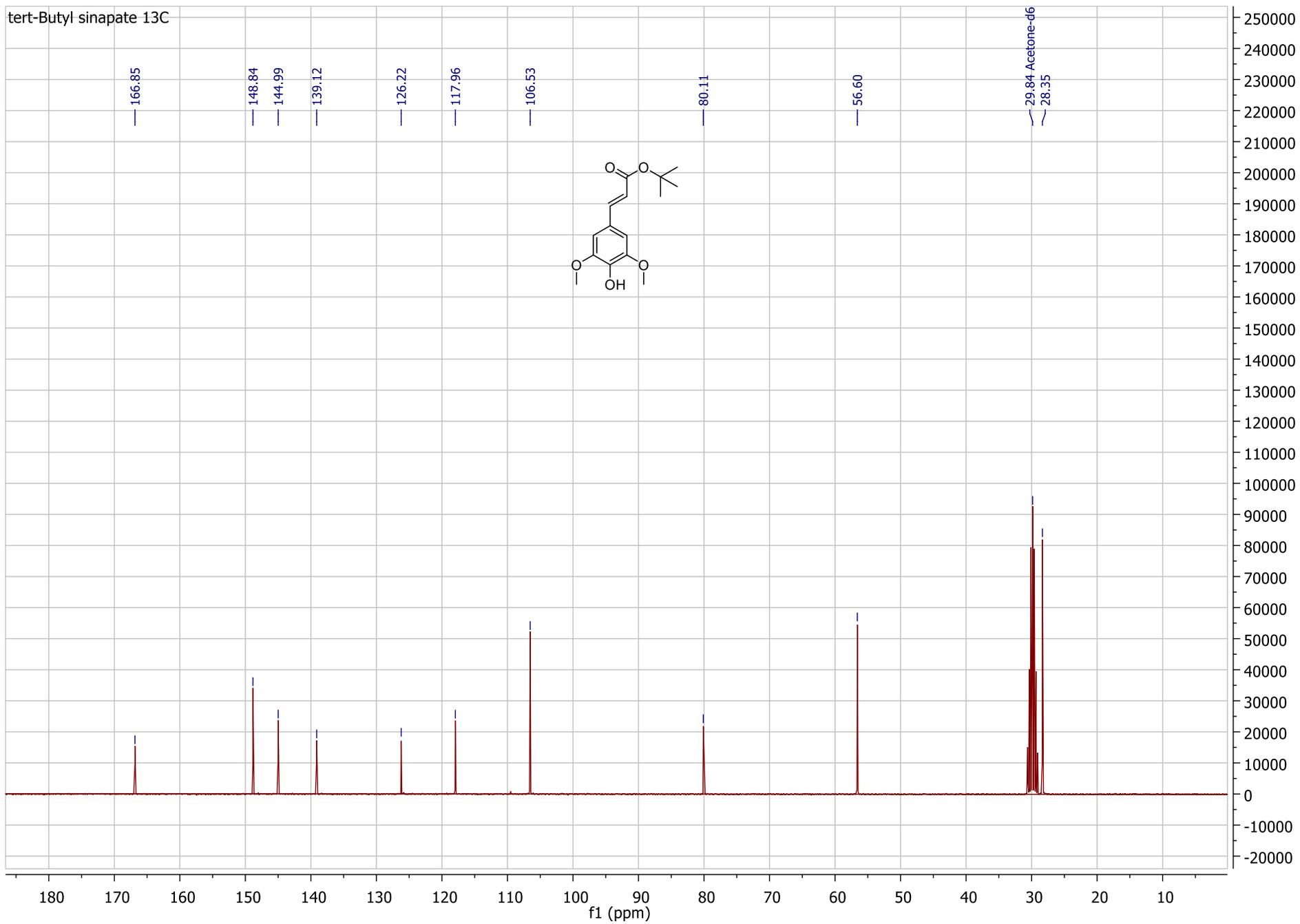
2-ethylhexyl sinapate 13C



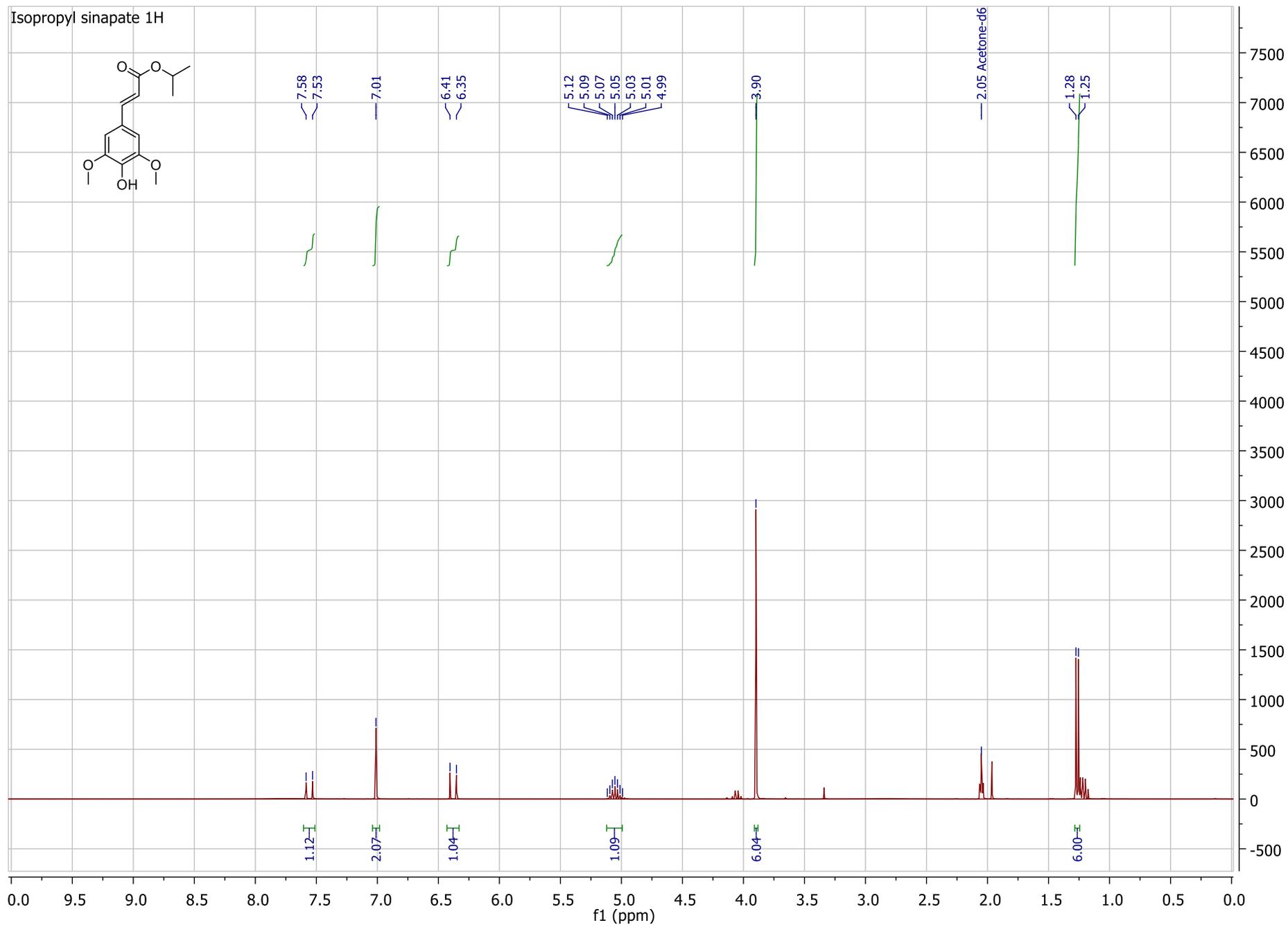
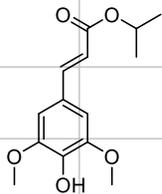
tert-Butyl sinapate 1H



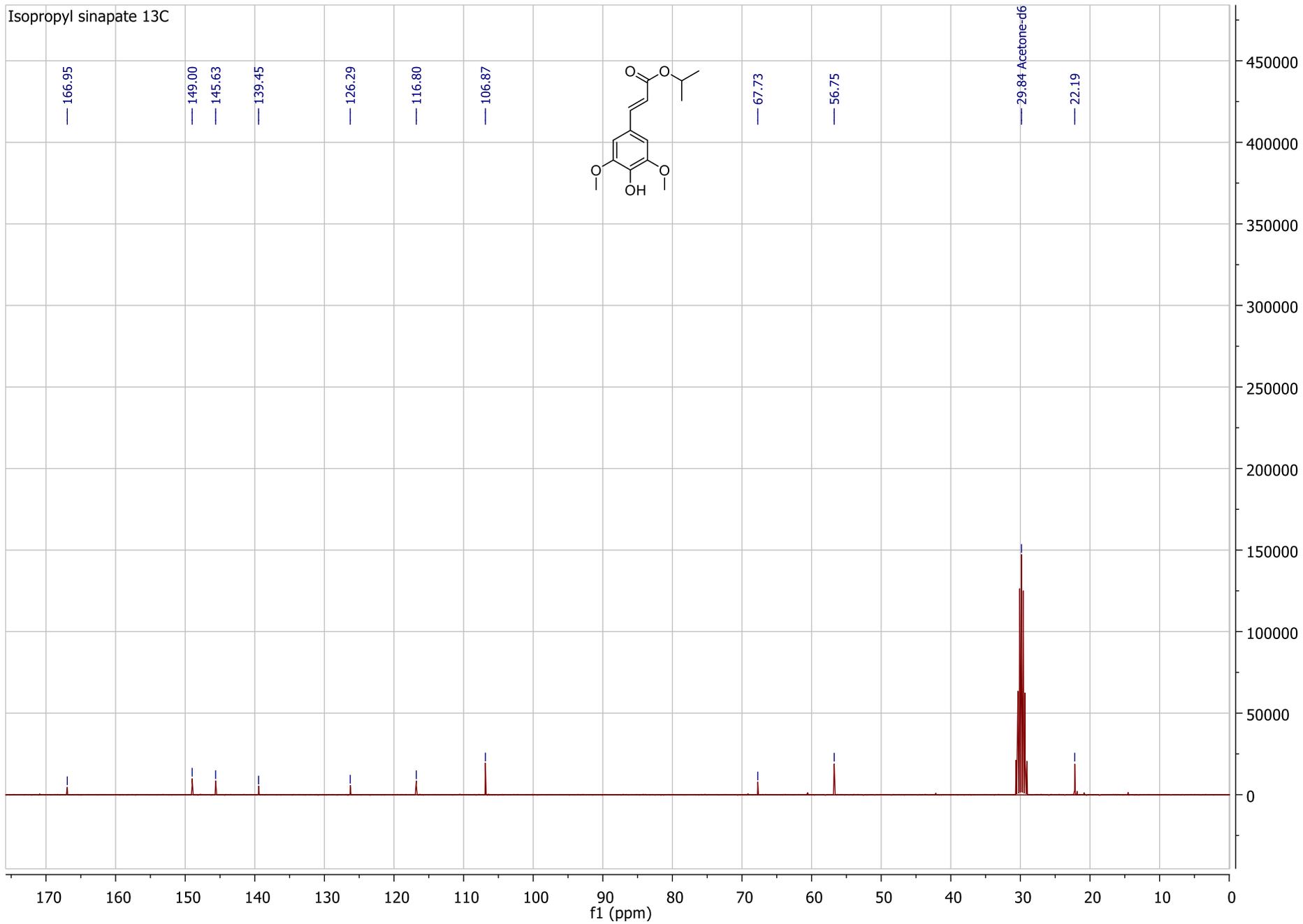
tert-Butyl sinapate 13C



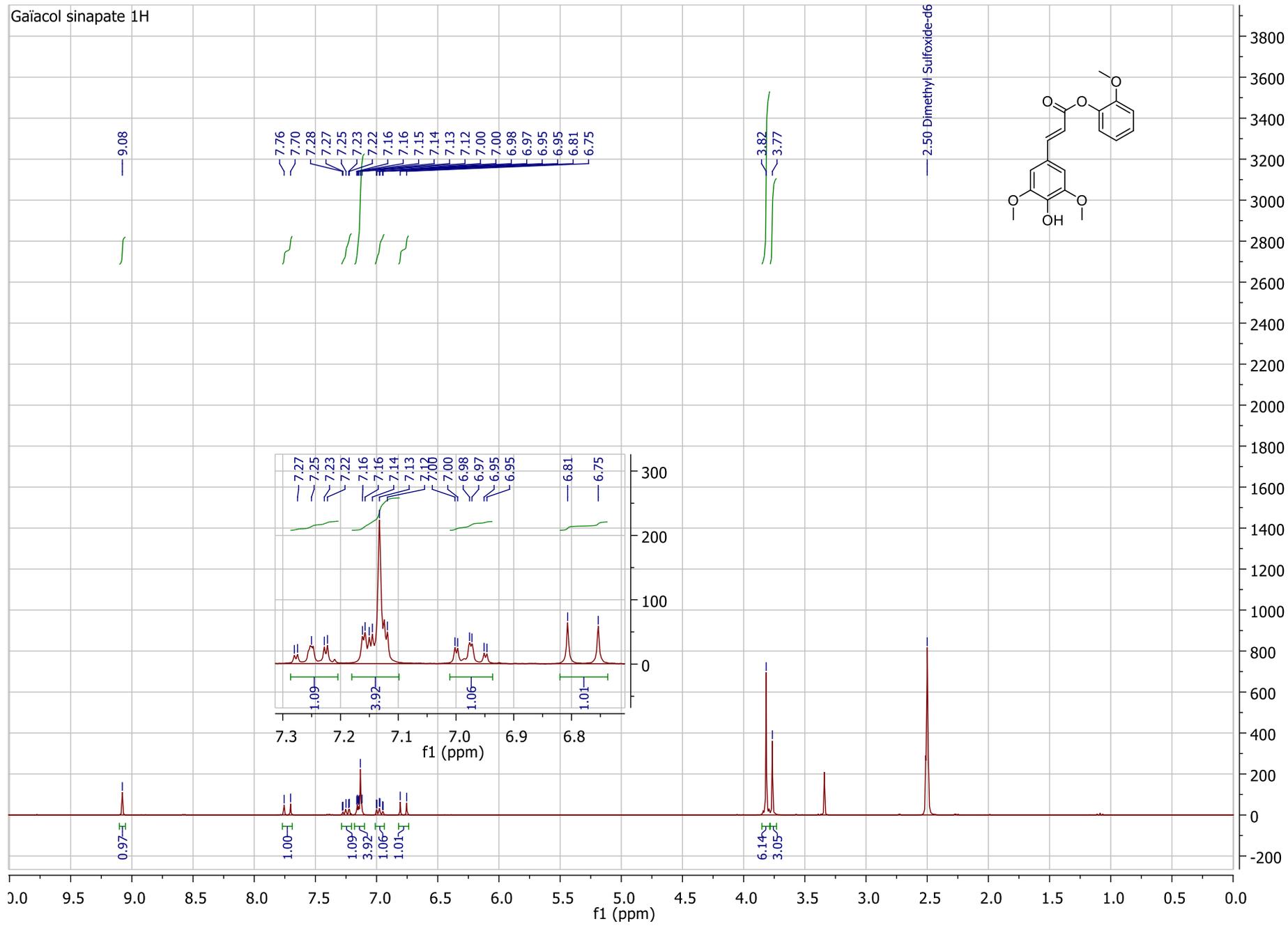
Isopropyl sinapate 1H



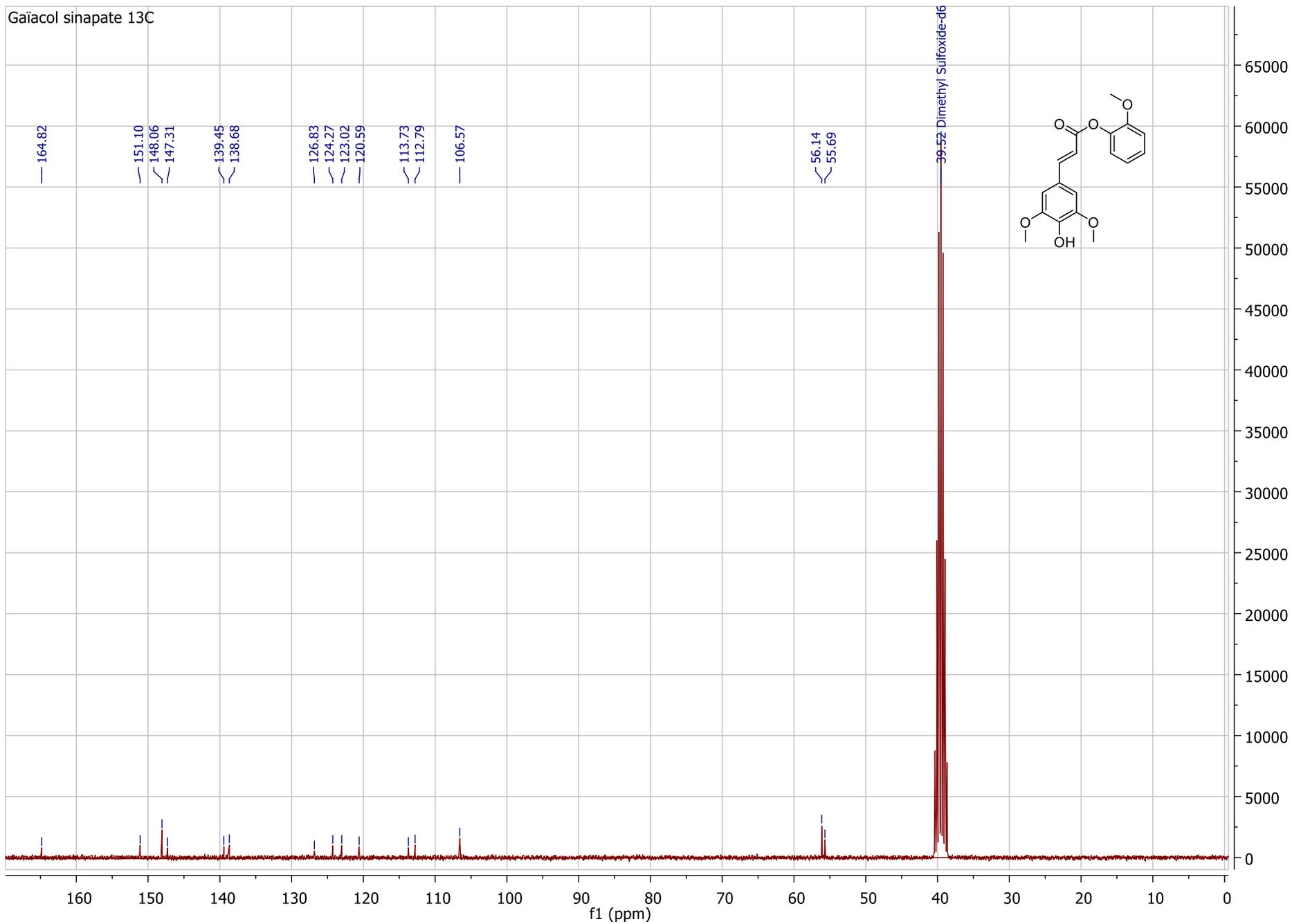
Isopropyl sinapate 13C



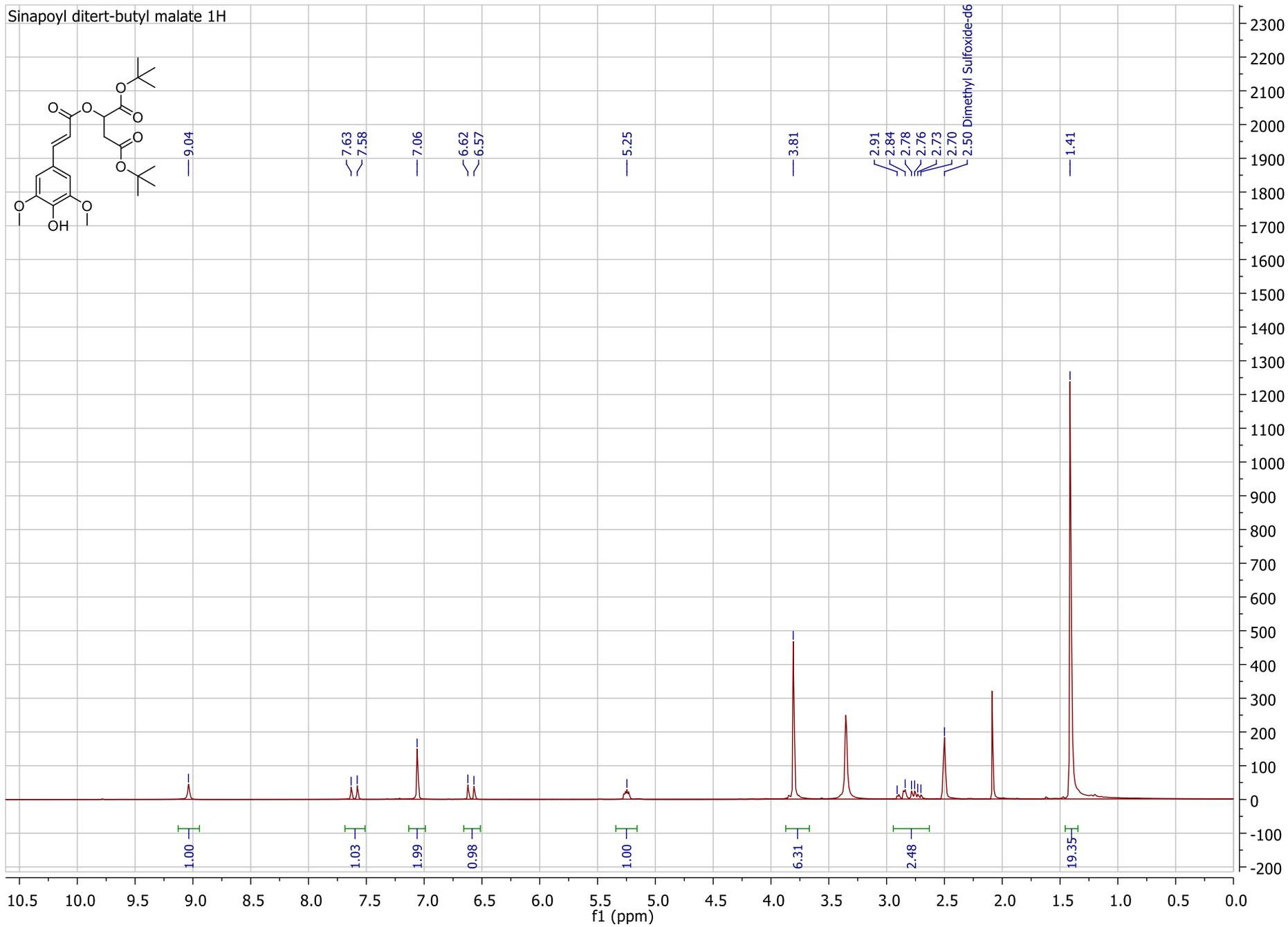
Gaiacol sinapate 1H



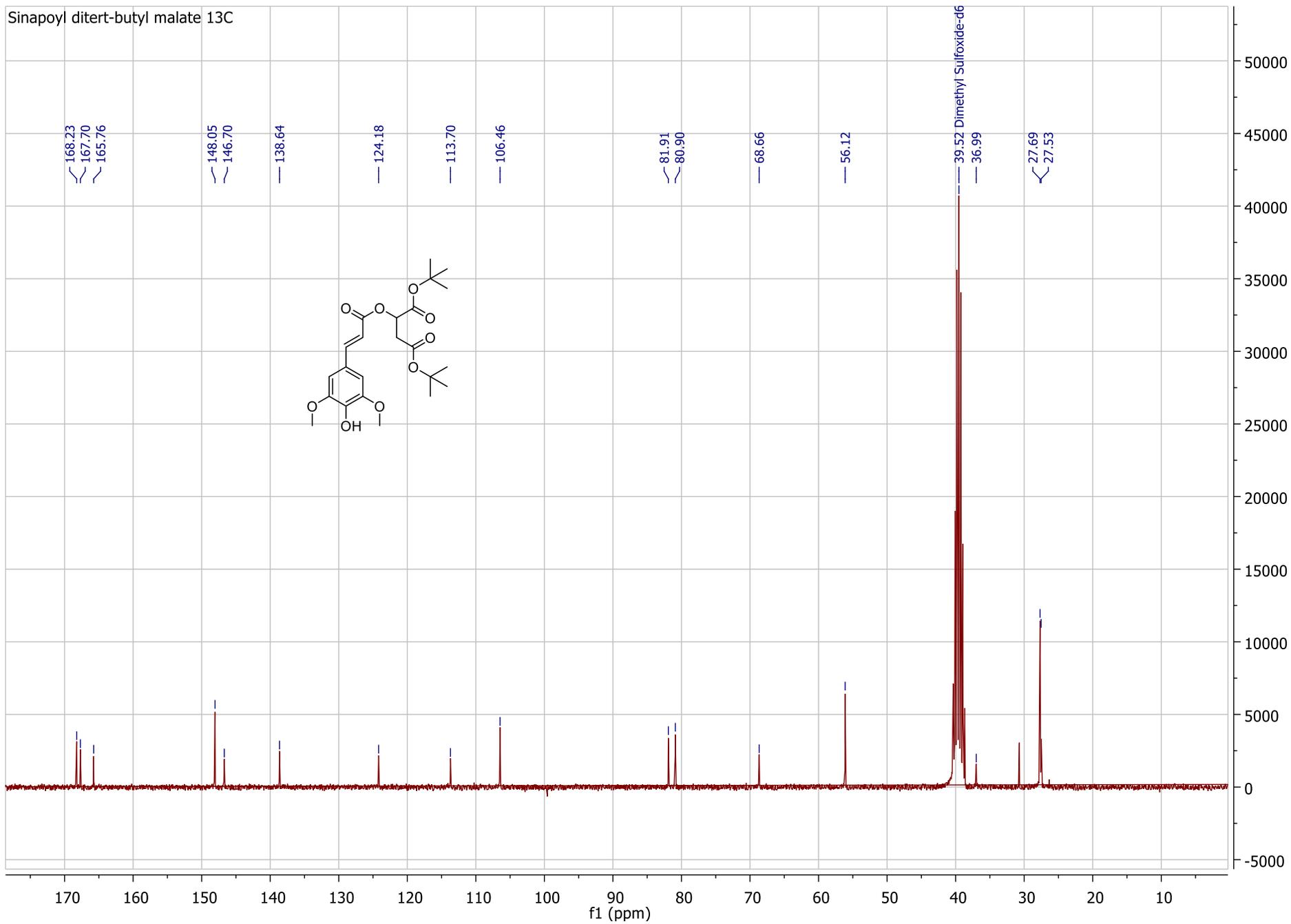
Gaiacol sinapate 13C



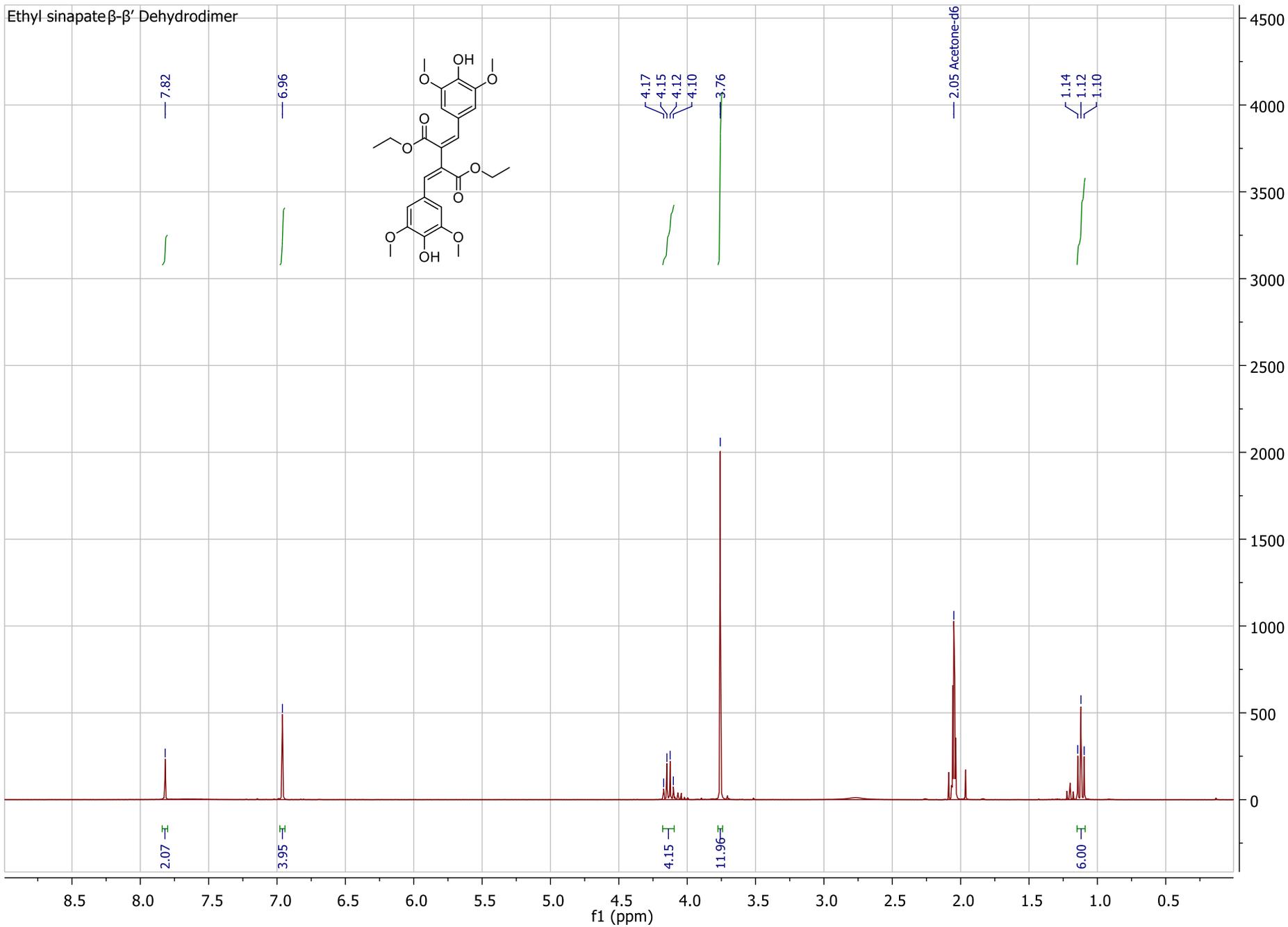
Sinapoyl ditert-butyl malate 1H



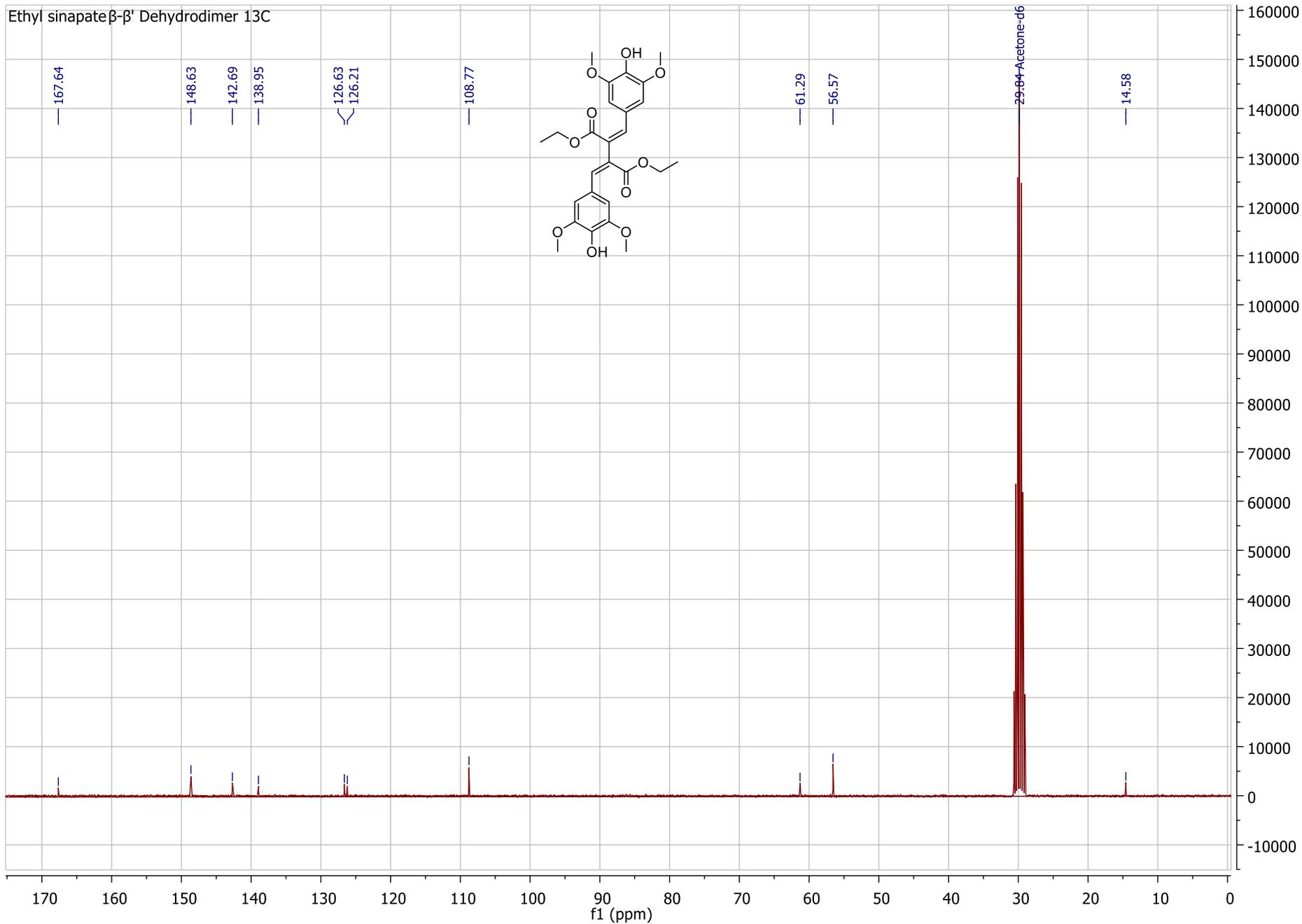
Sinapoyl di-tert-butyl malate 13C



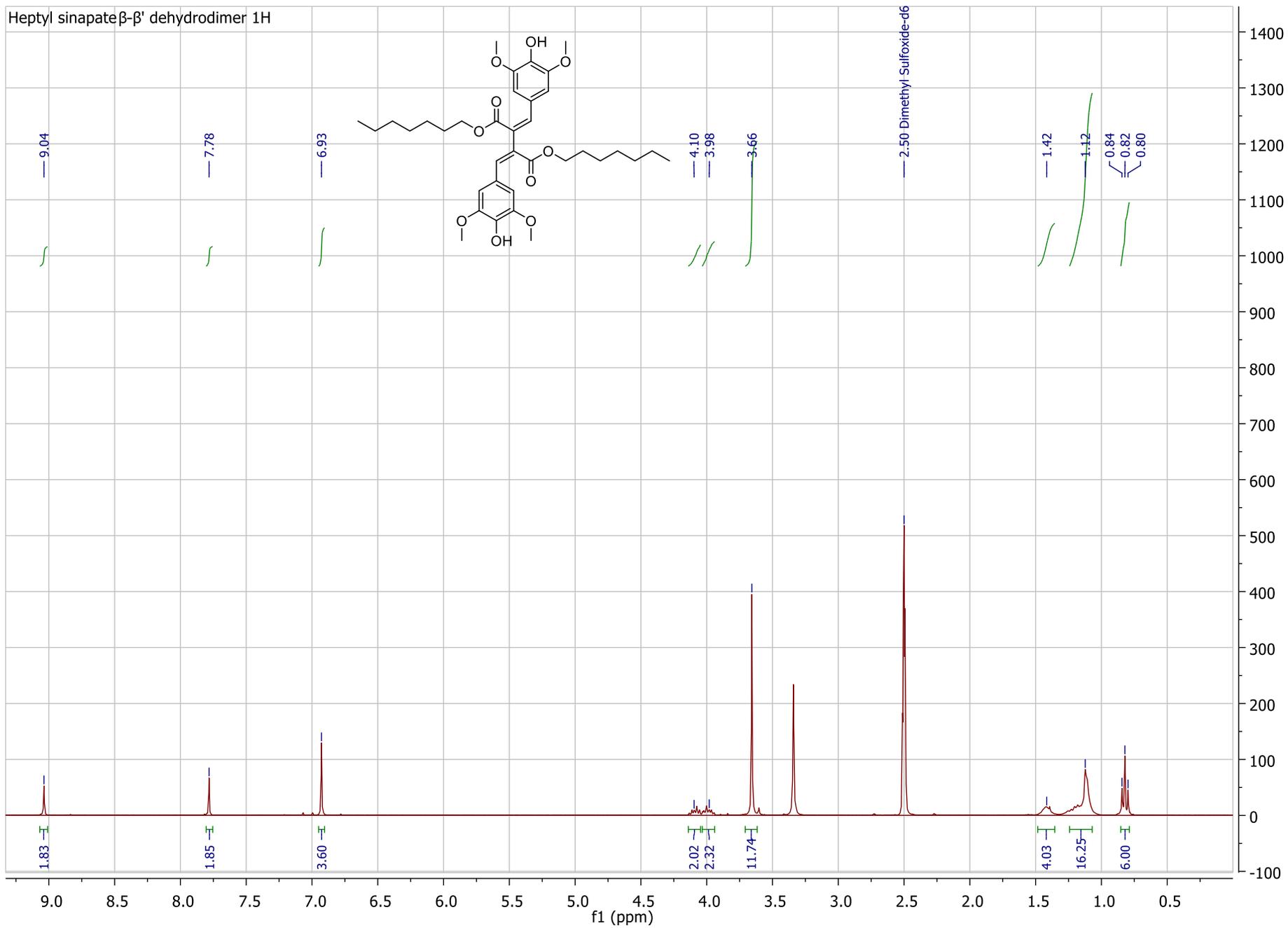
Ethyl sinapate- $\beta$ - $\beta'$  Dehydrodimer



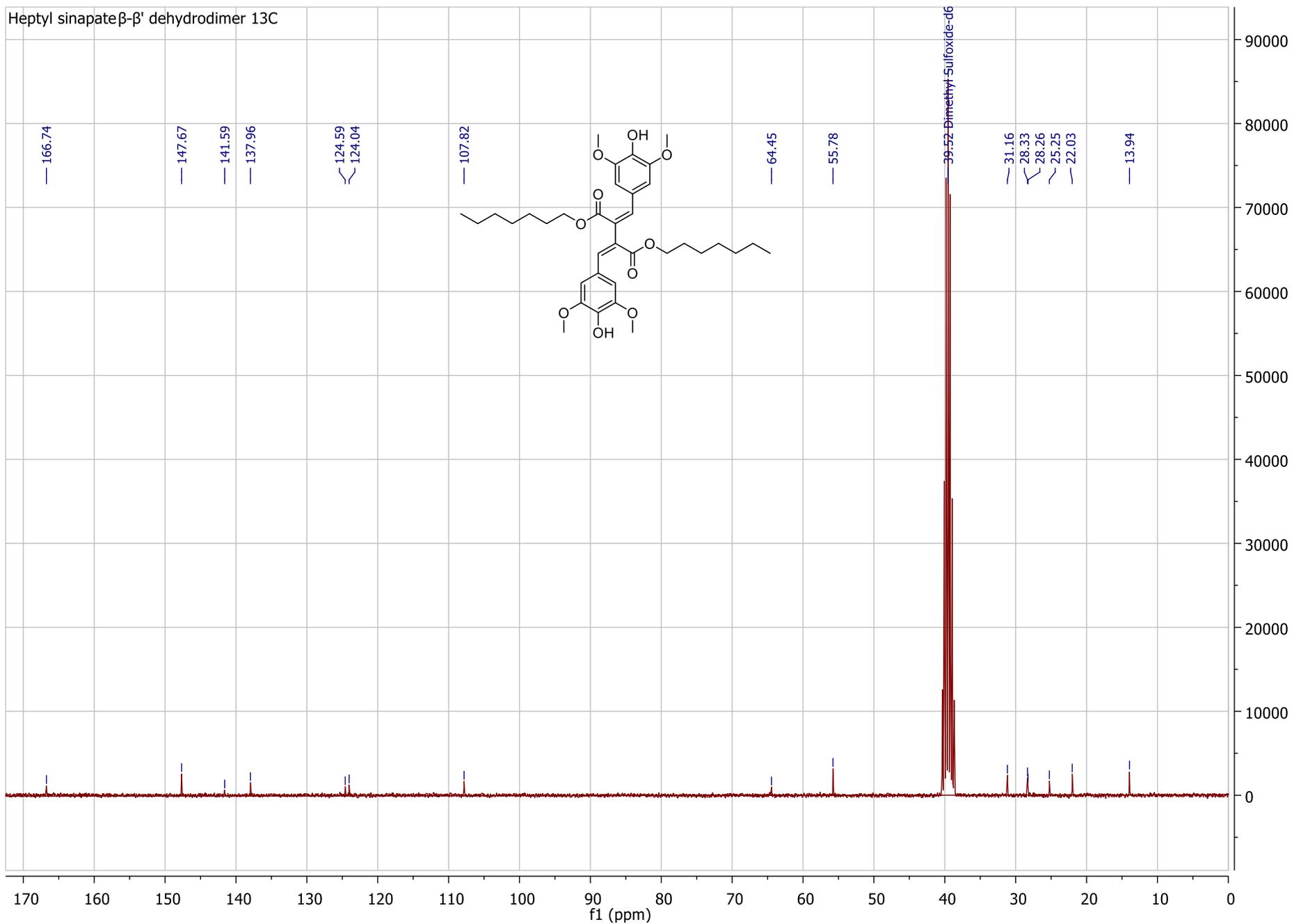
Ethyl sinapate  $\beta$ - $\beta'$  Dehydromer 13C



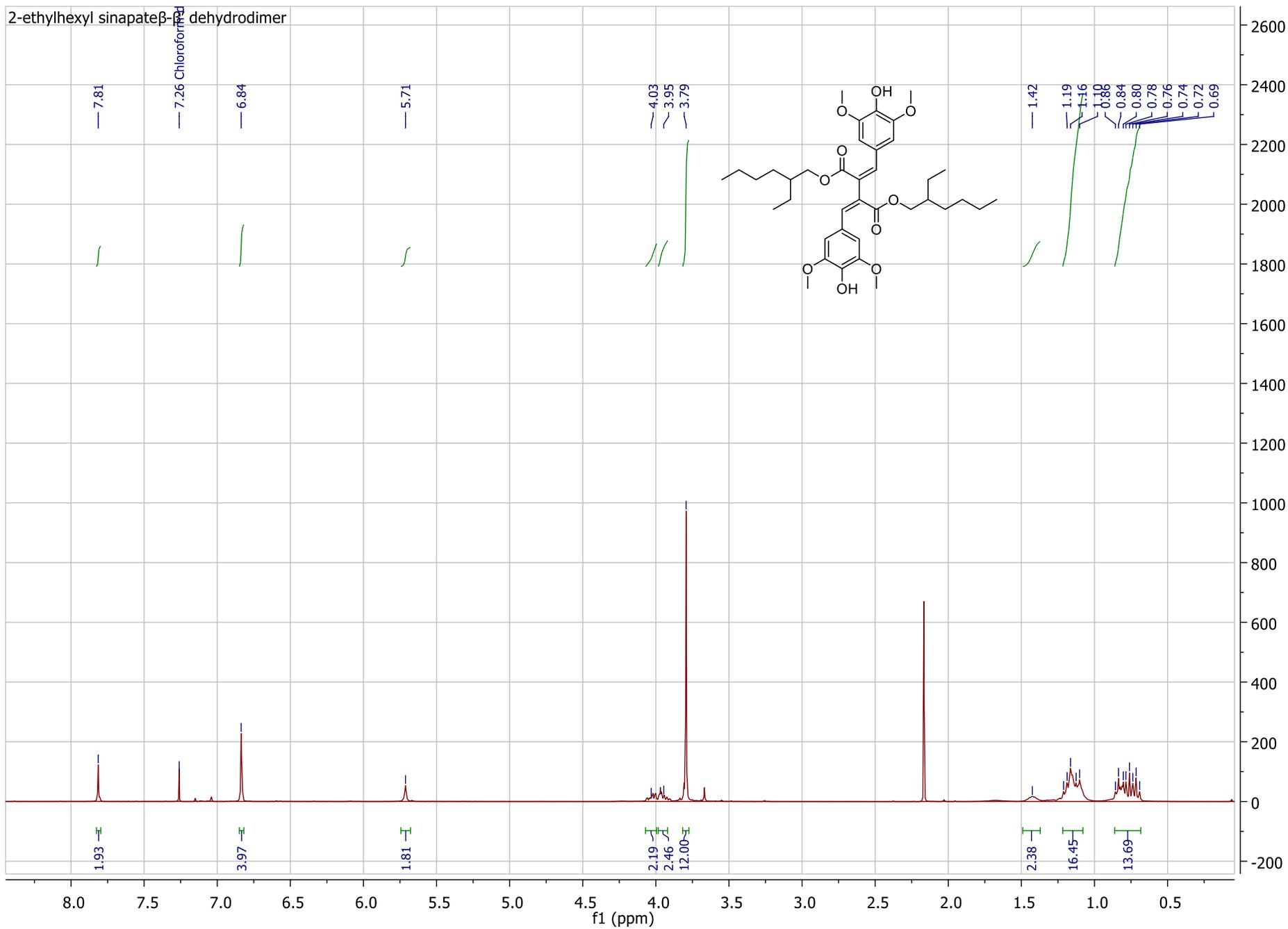
Heptyl sinapate-β-β' dehydromer 1H



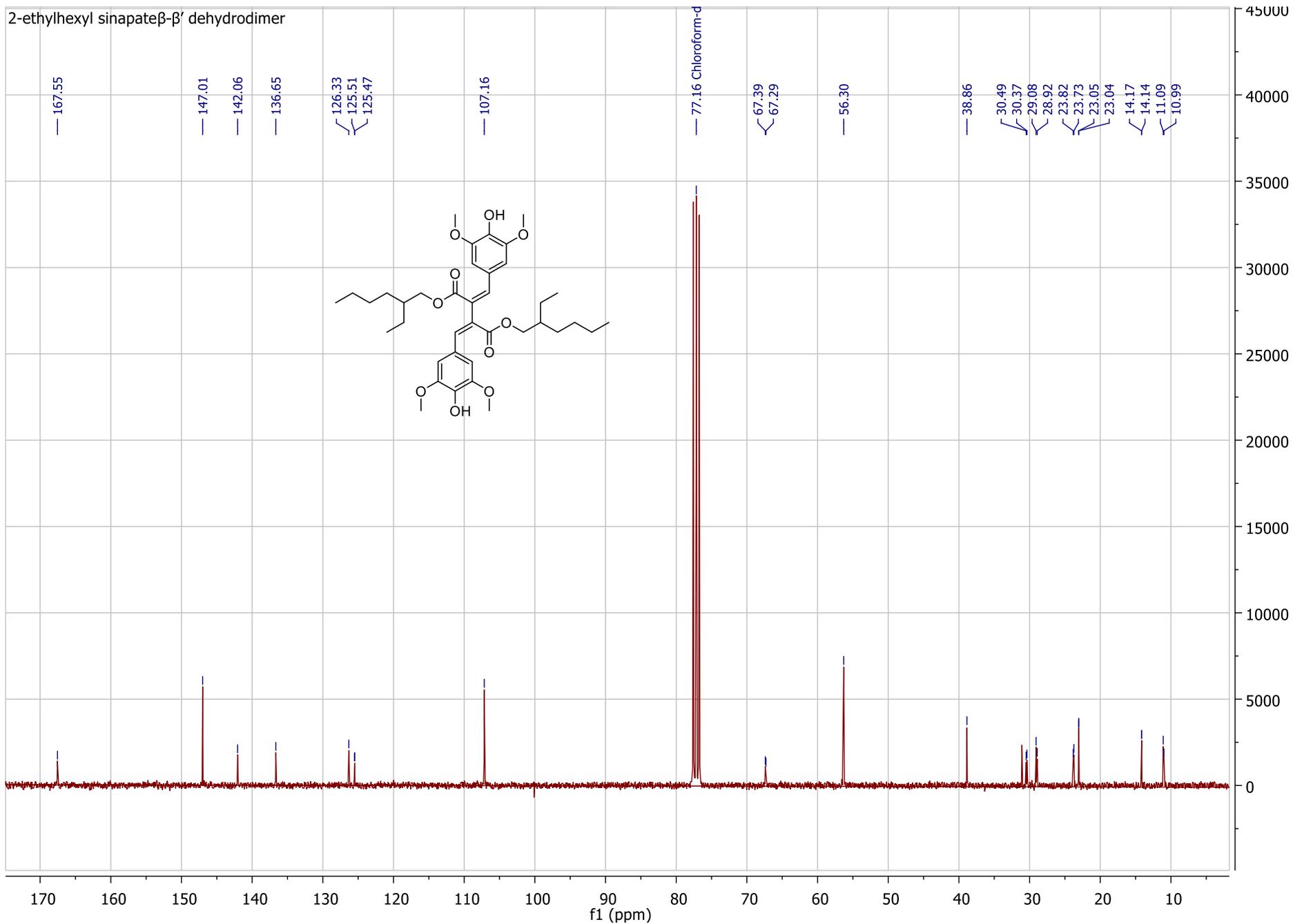
Heptyl sinapate-β-β' dehydromer 13C

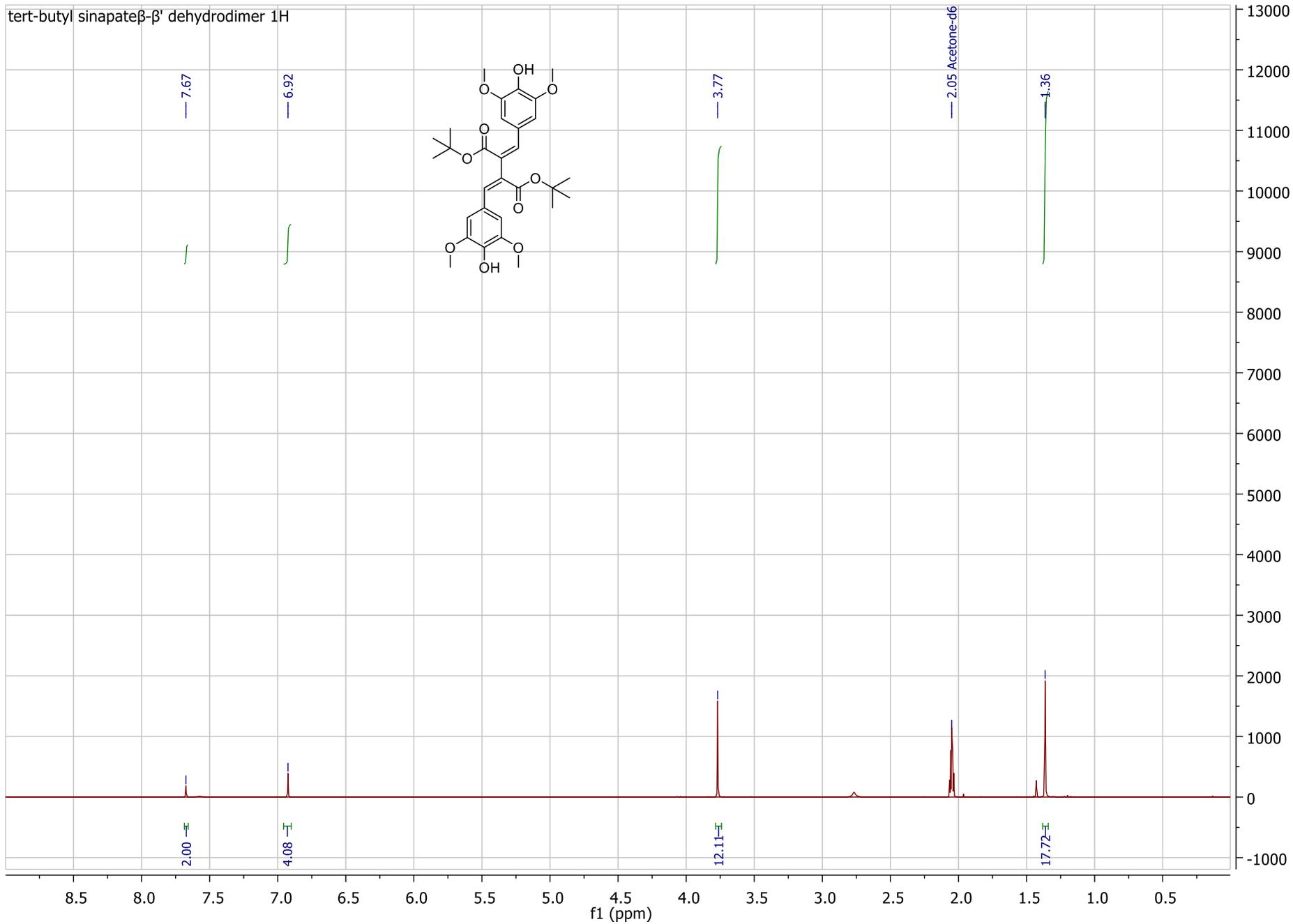


2-ethylhexyl sinapate $\beta$ -D dehydrodimer

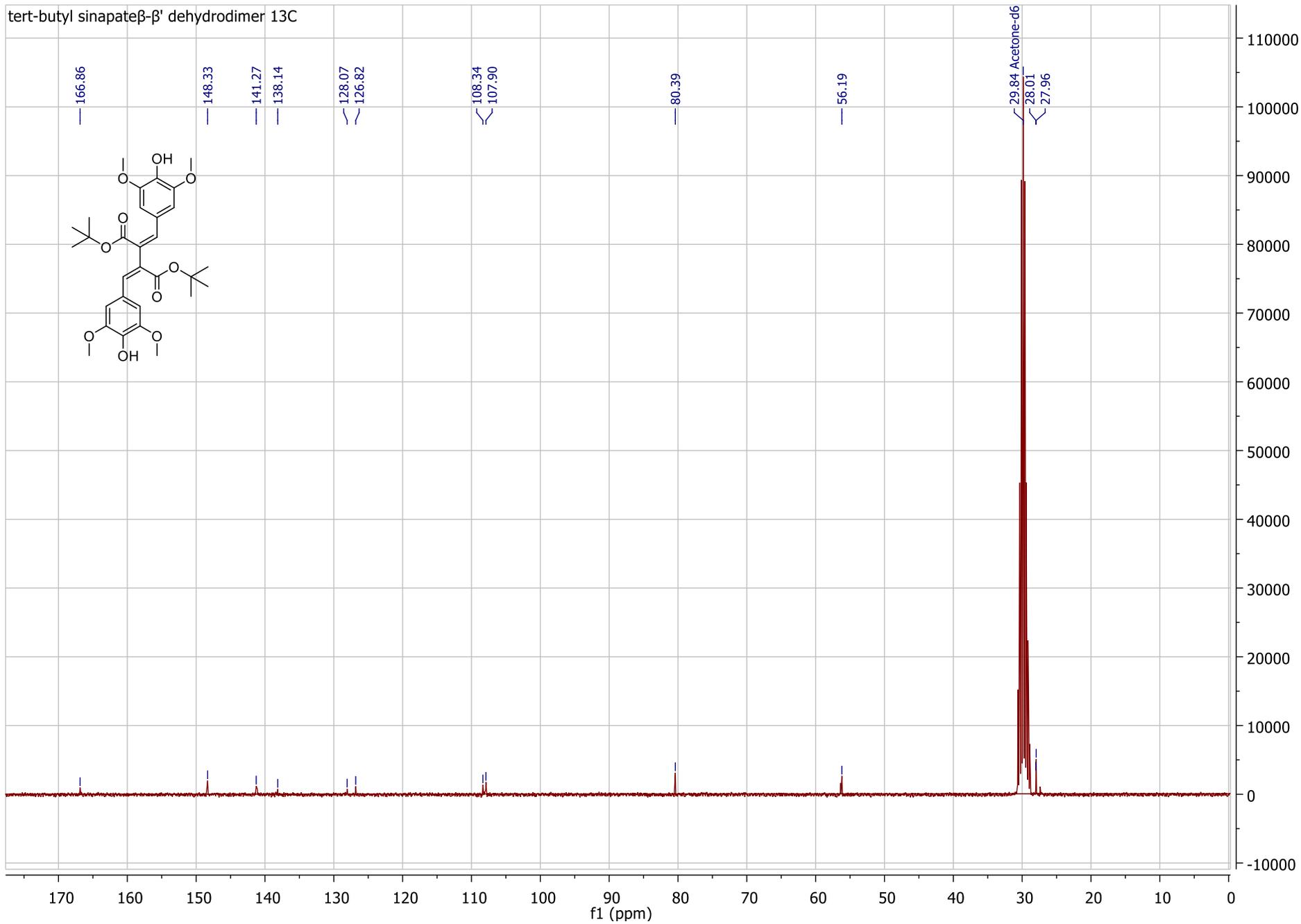


2-ethylhexyl sinapate-β-β' dehydromer

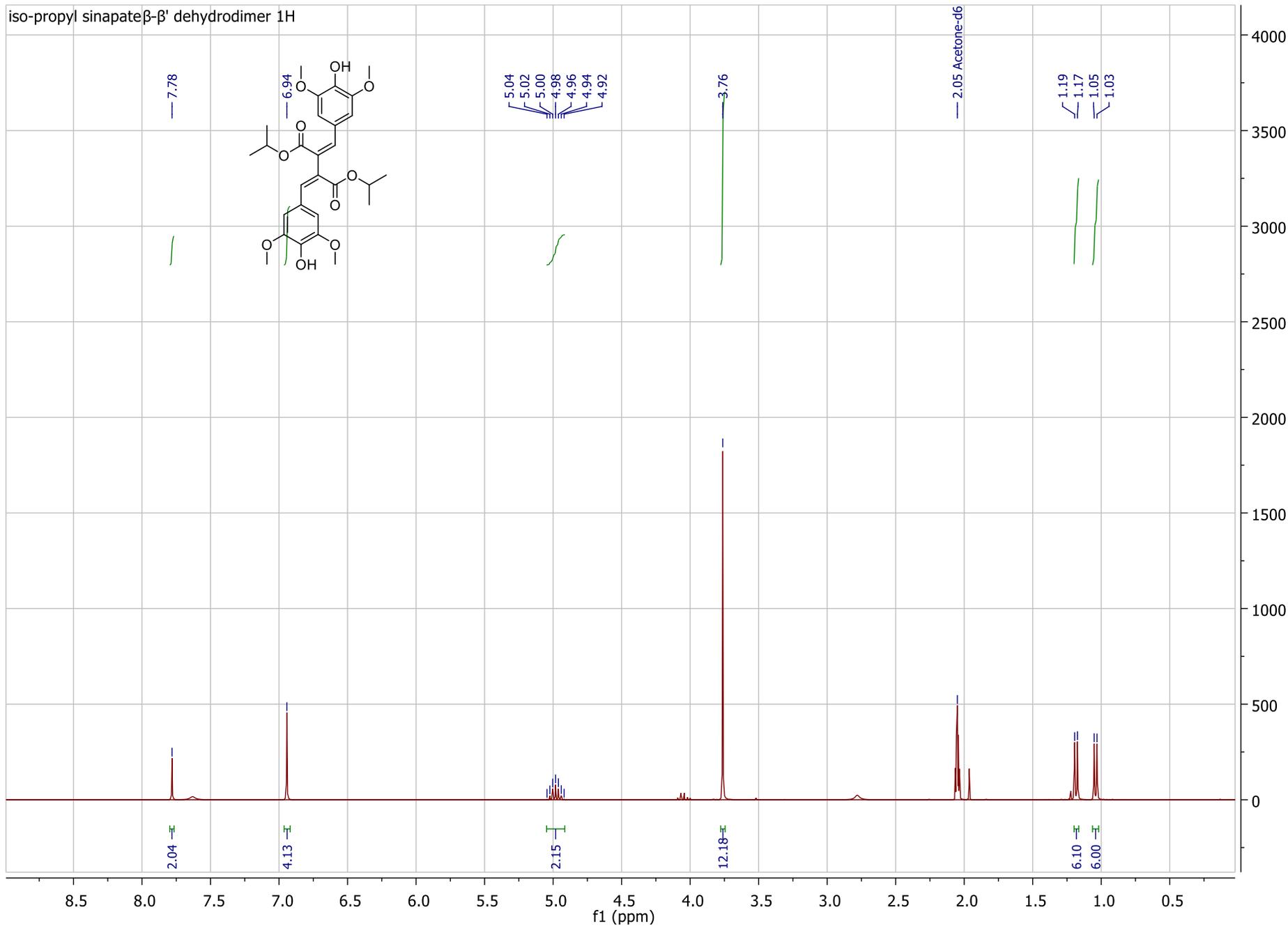




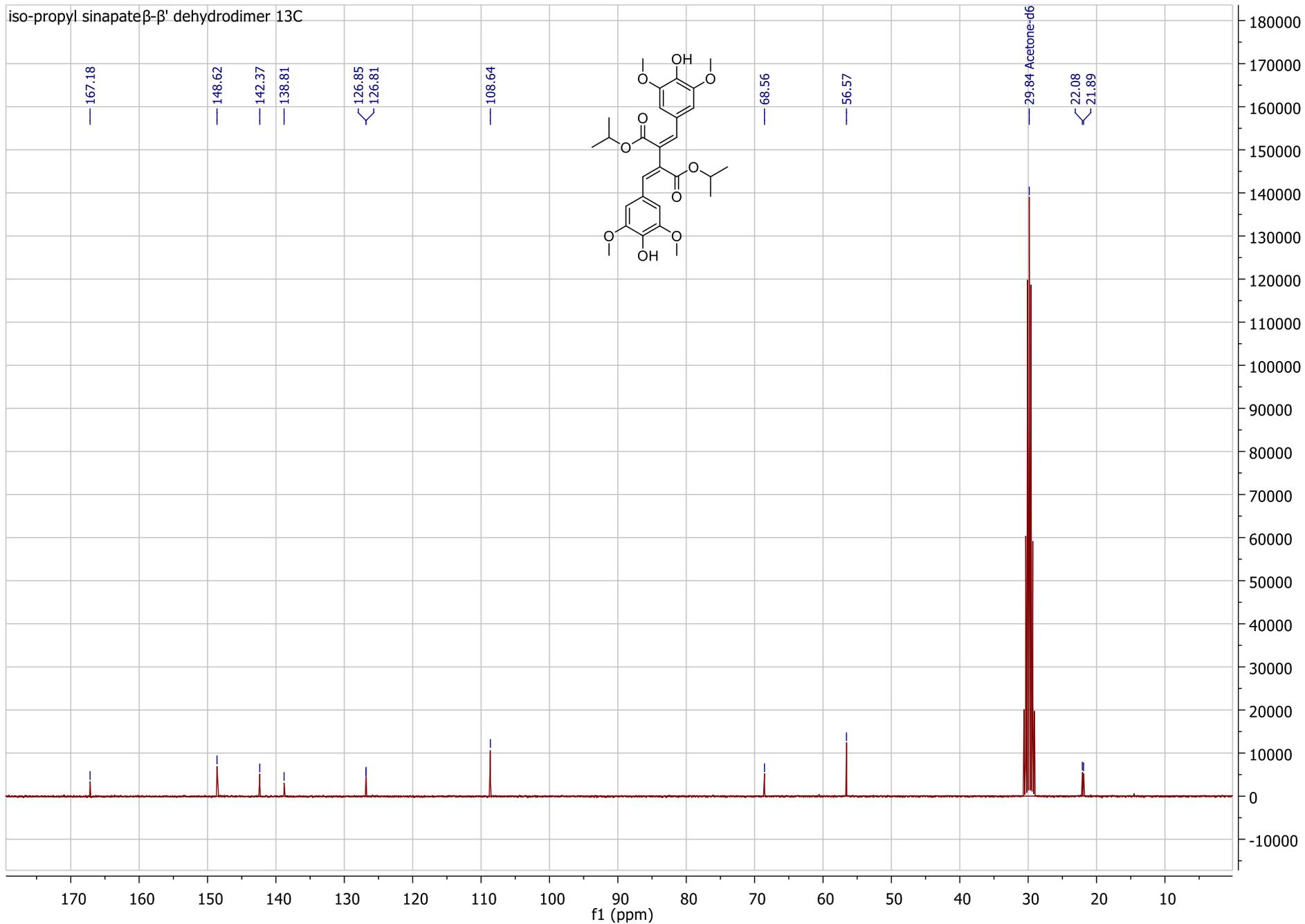
tert-butyl sinapate-β-β' dehydrodimer 13C



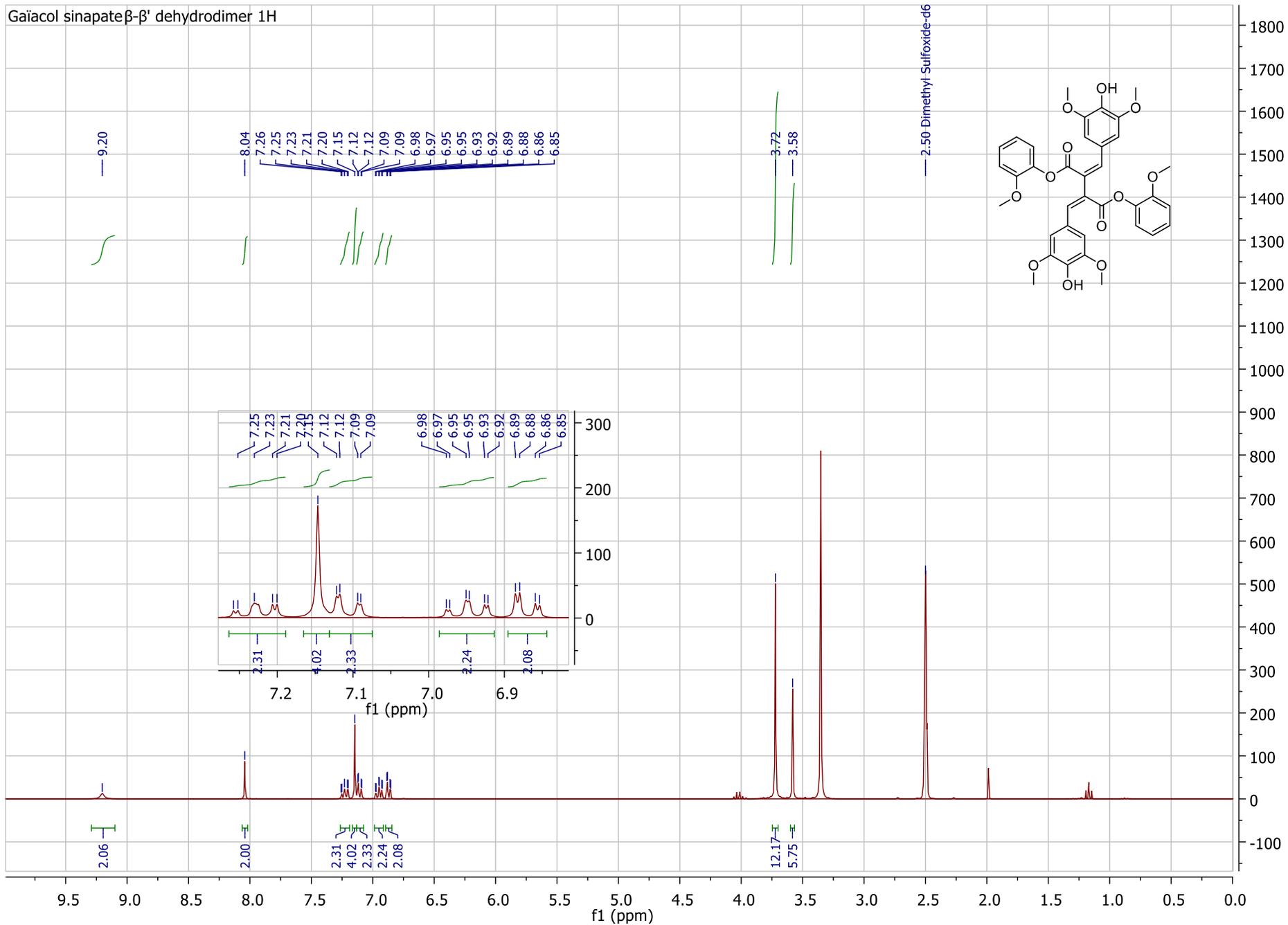
iso-propyl sinapate $\beta$ - $\beta'$  dehydromer 1H



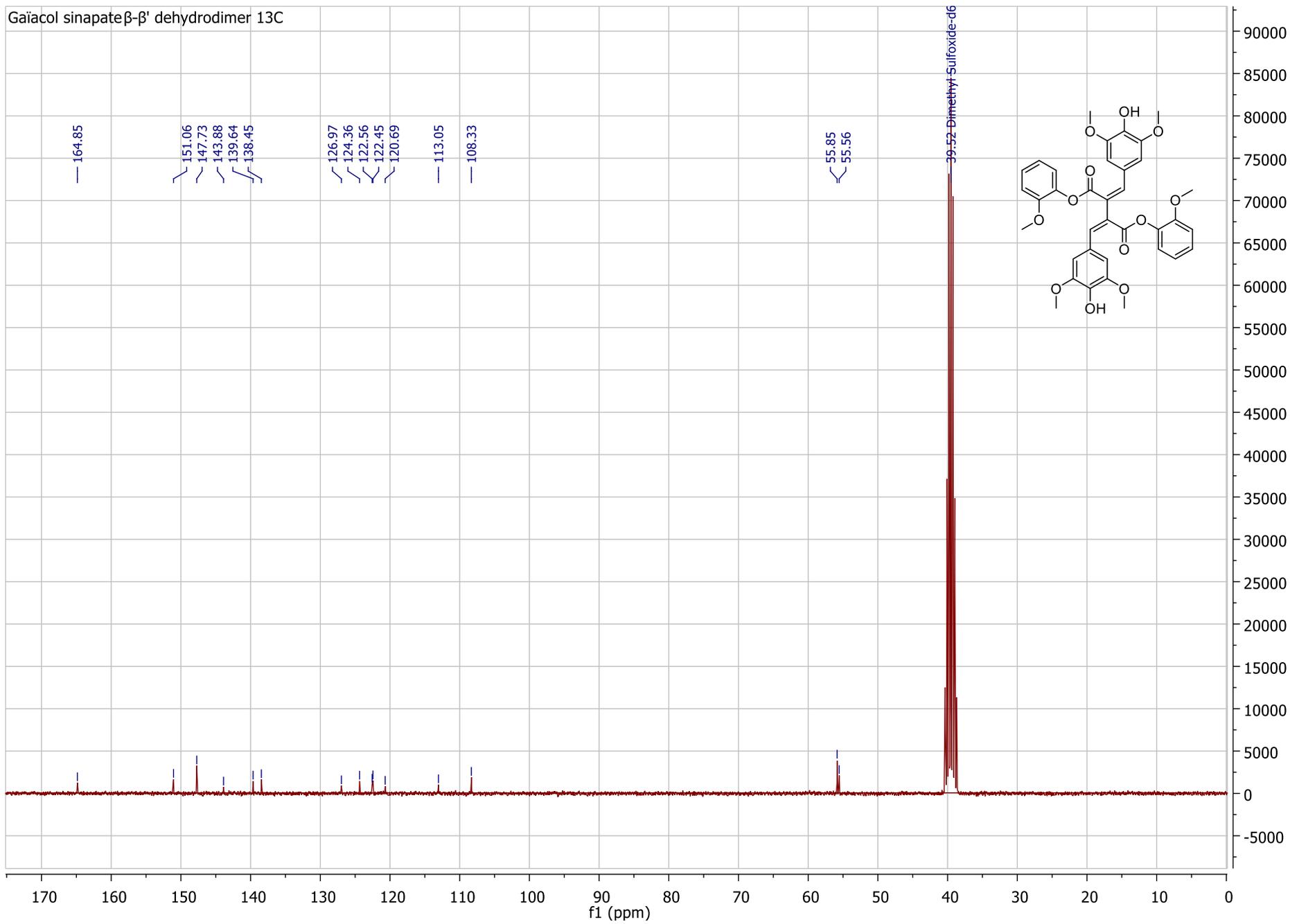
iso-propyl sinapate $\beta$ - $\beta'$  dehydromer 13C



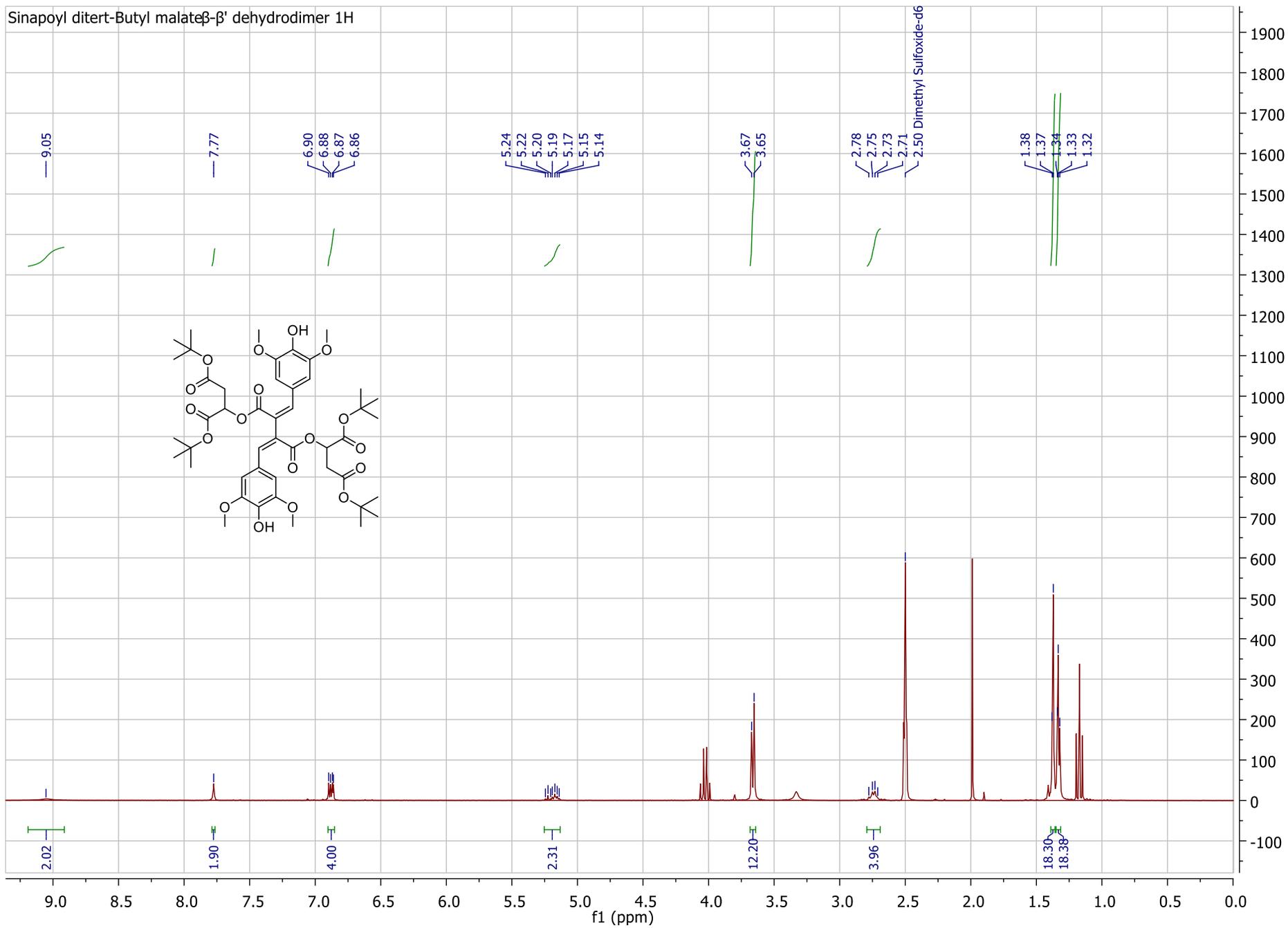
Gaïacol sinapate $\beta$ - $\beta'$  dehydromer 1H



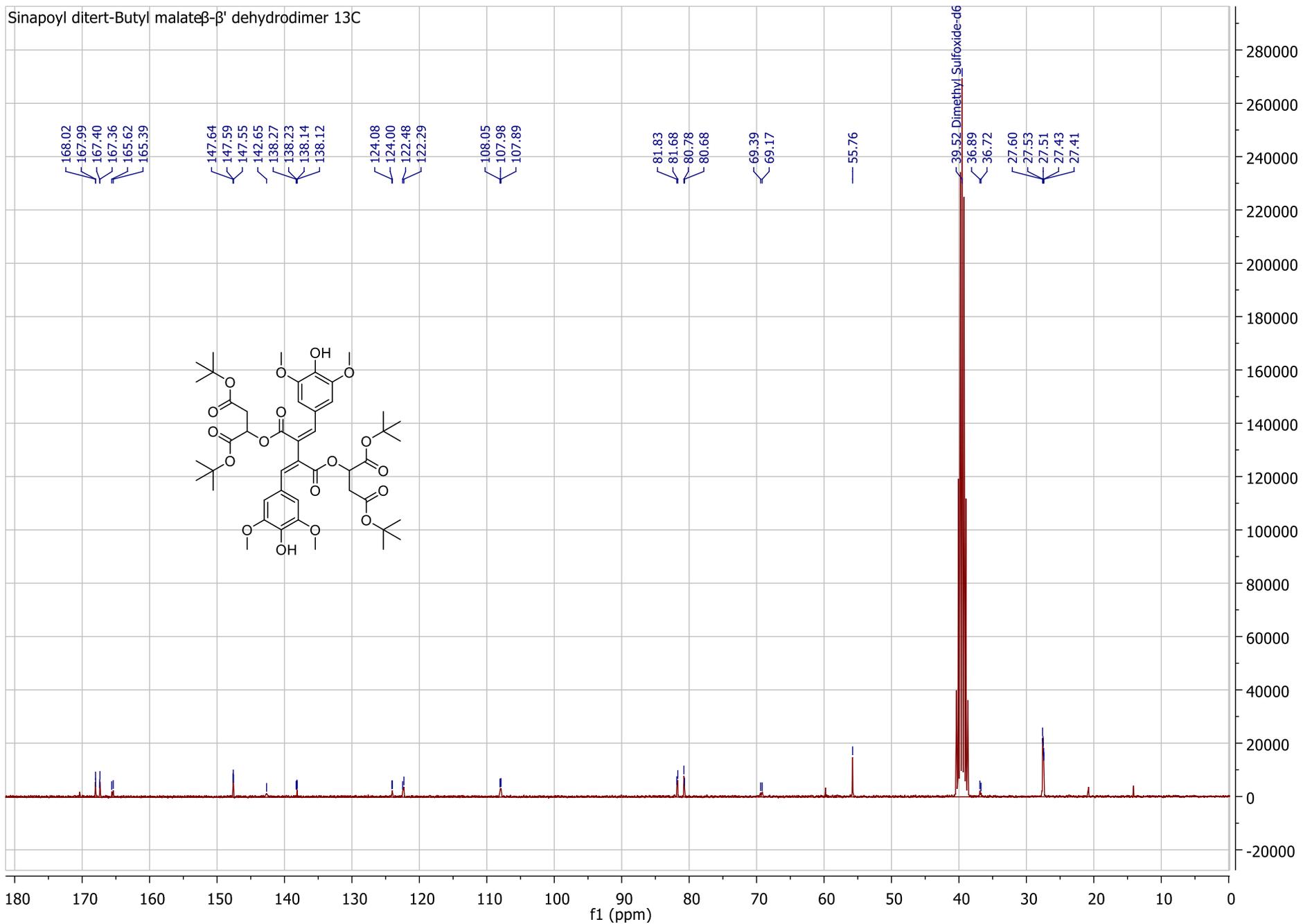
Gaiacol sinapate  $\beta$ - $\beta'$  dehydrodimer 13C



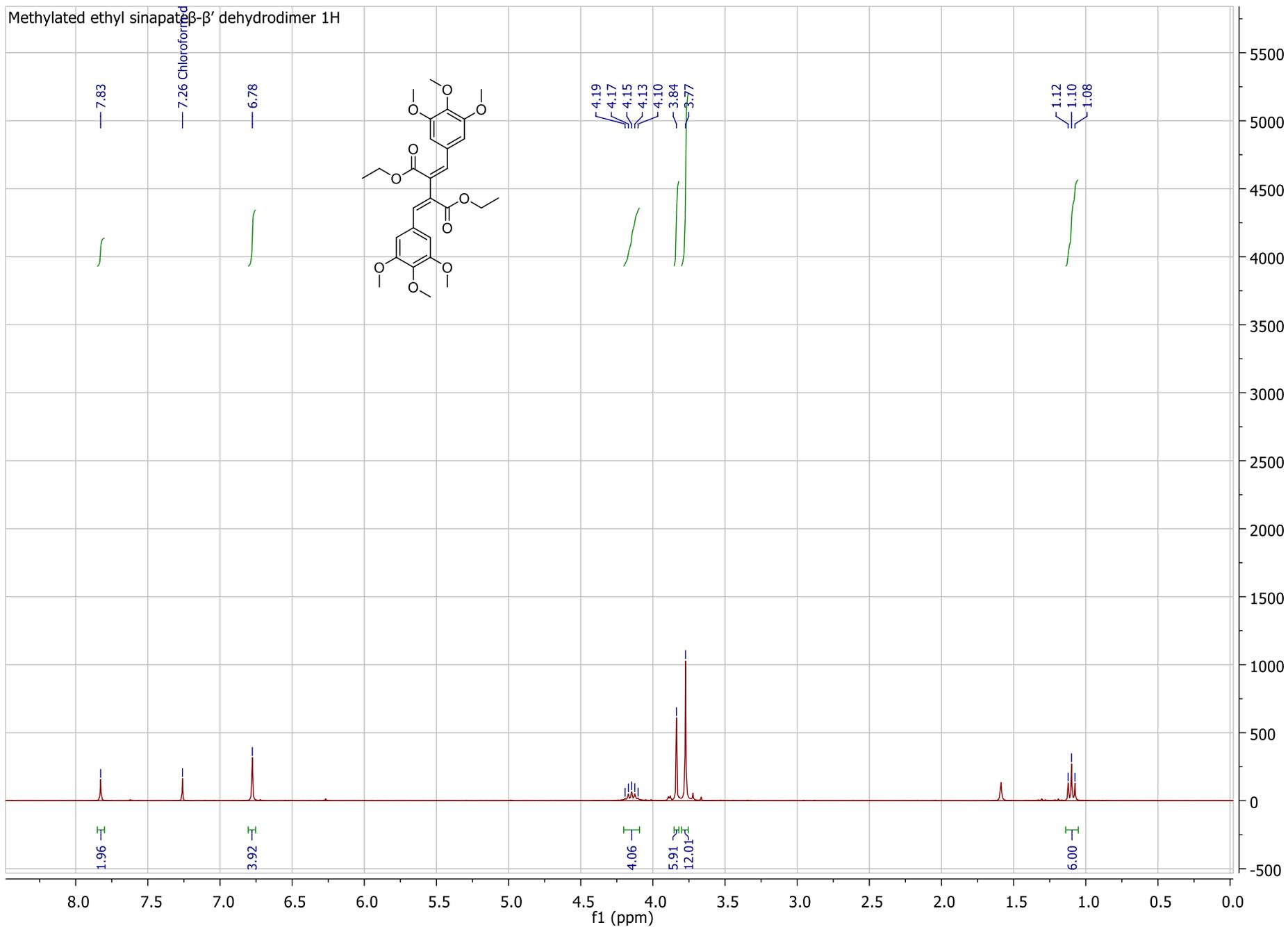
Sinapoyl ditert-Butyl malate-β' dehydrodimer 1H



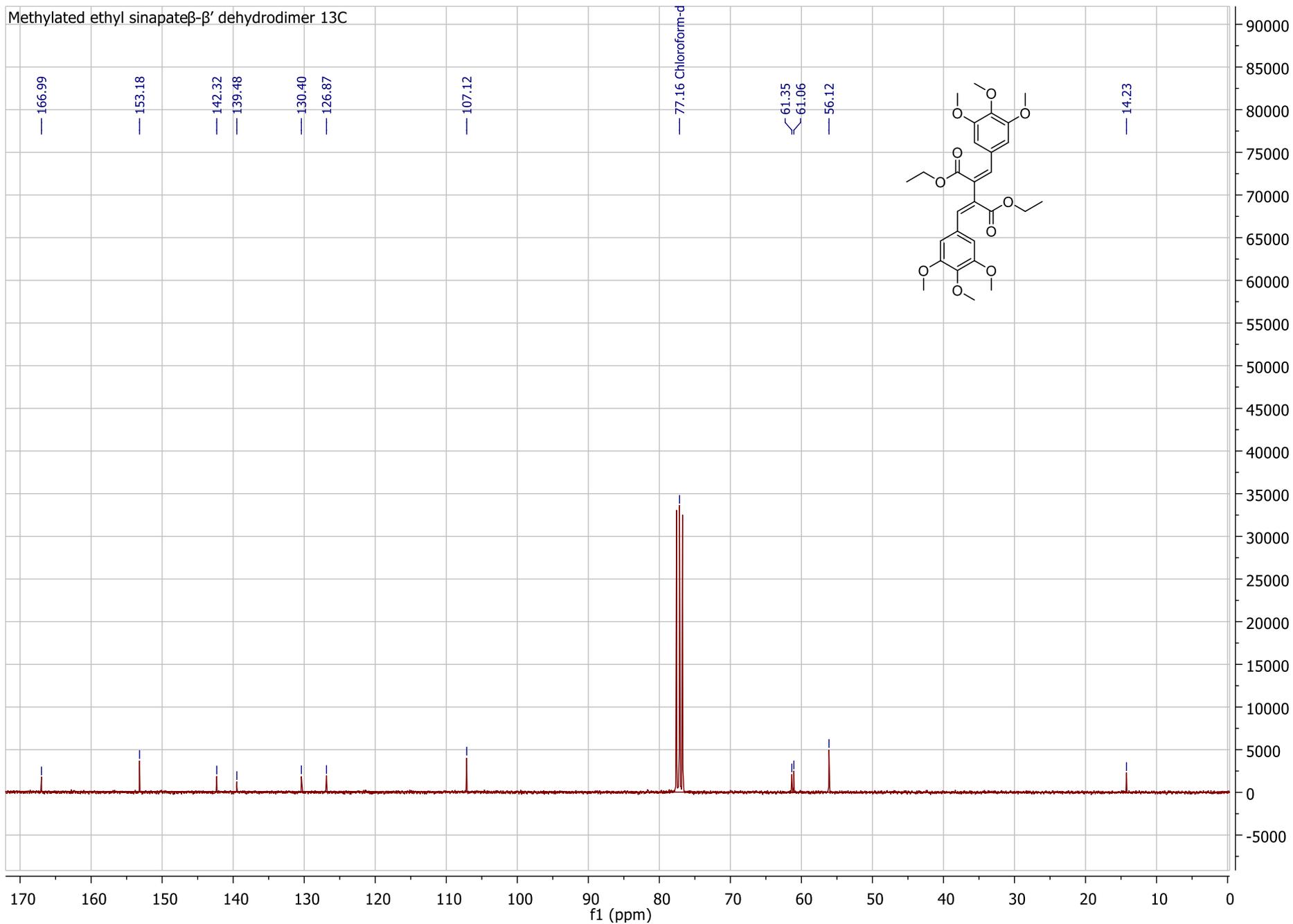
Sinapoyl di-tert-Butyl malate-β-β' dehydromer 13C



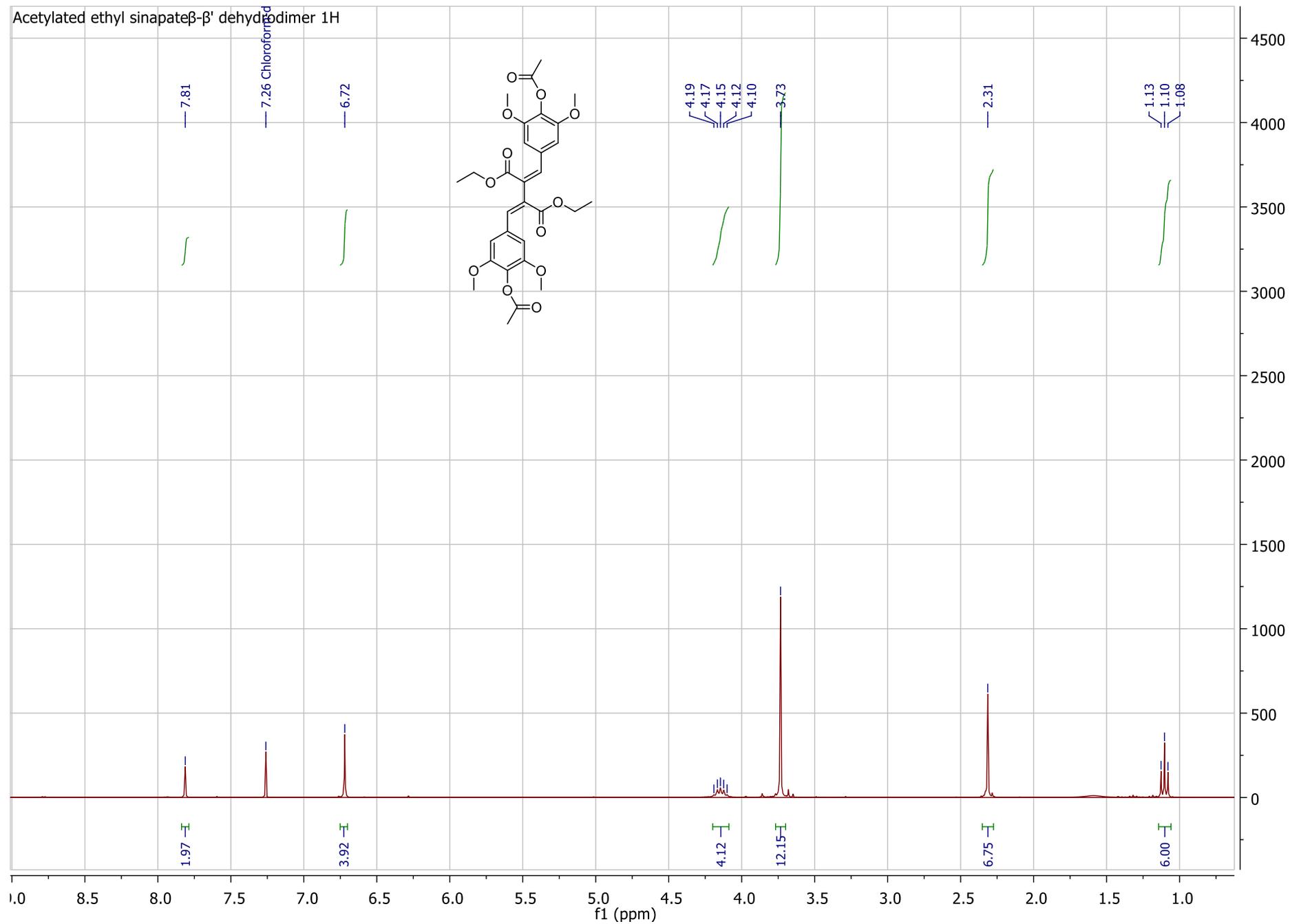
Methylated ethyl sinapate- $\beta$ - $\beta'$  dehydromer 1H



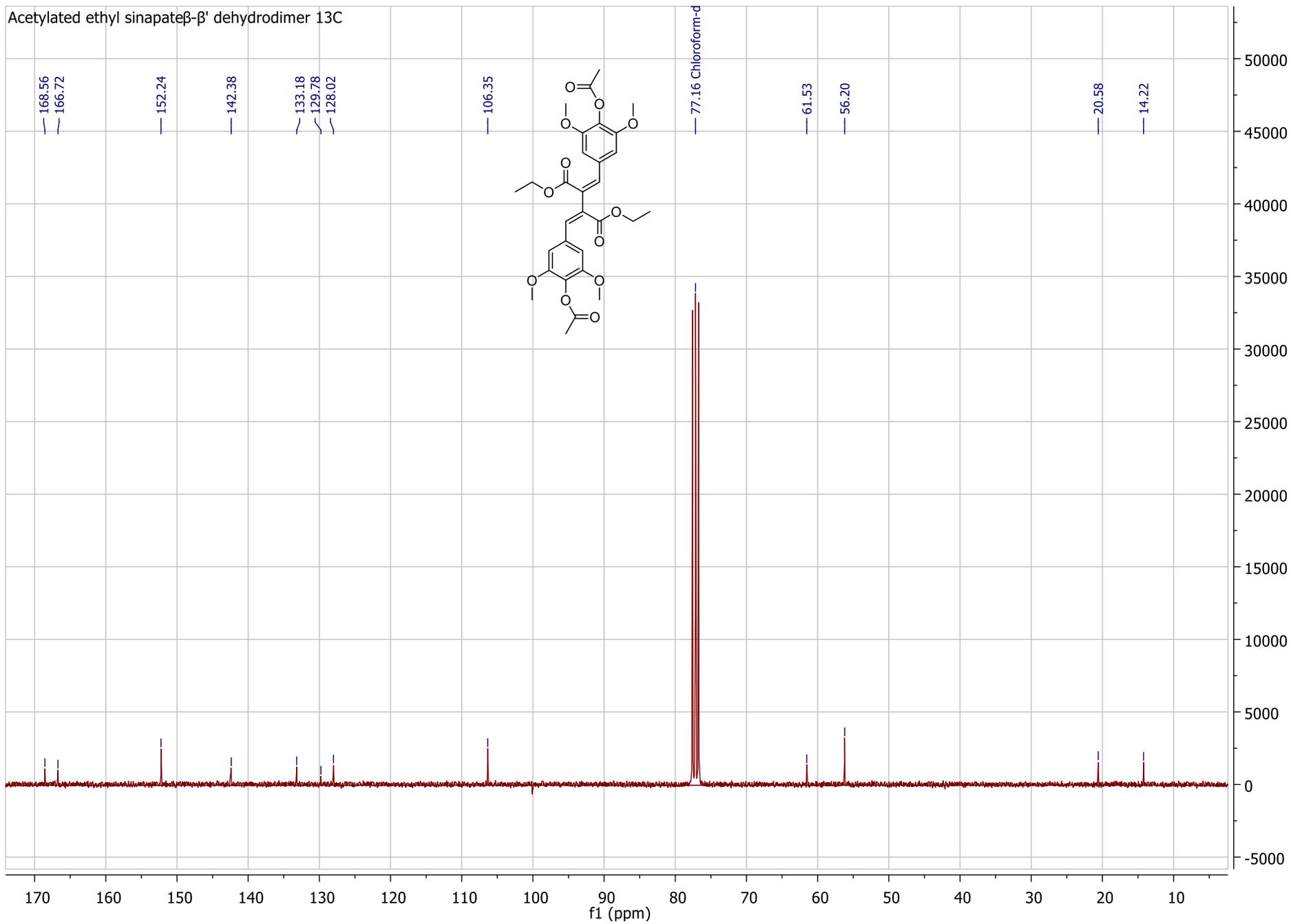
Methylated ethyl sinapate $\beta$ - $\beta'$  dehydrodimer 13C



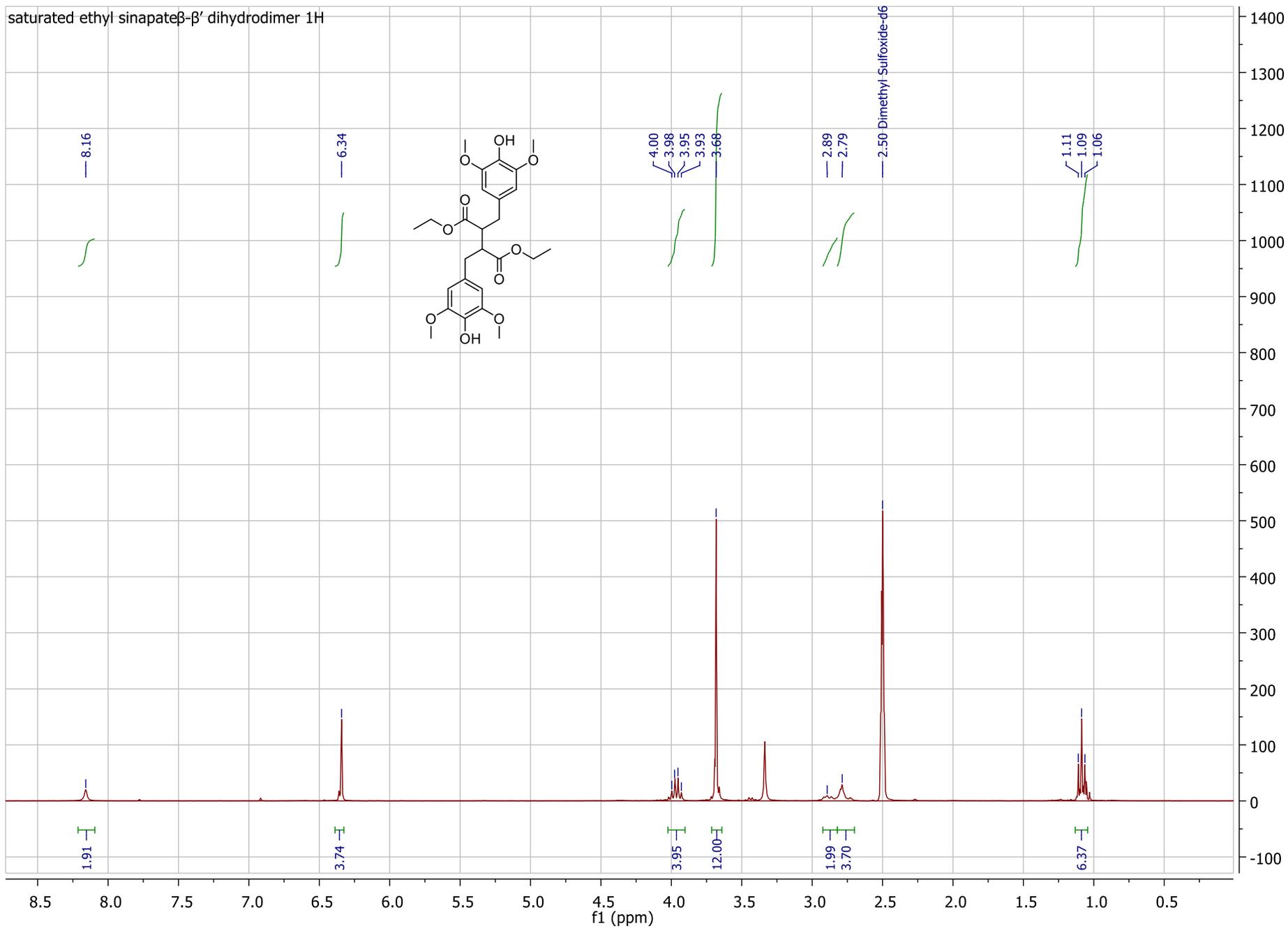
Acetylated ethyl sinapate-β' dehydromer 1H



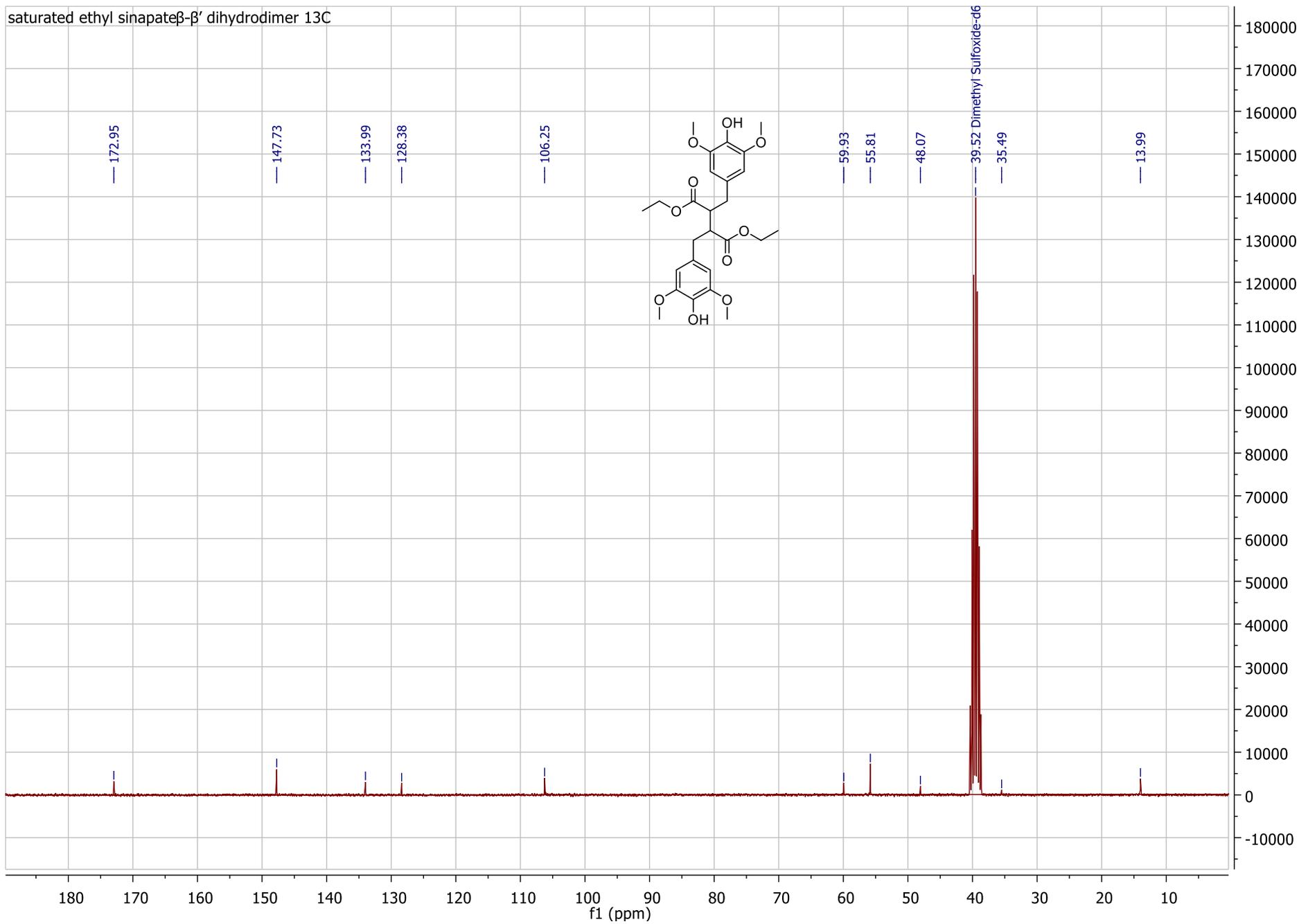
Acetylated ethyl sinapate-β' dehydrodimer 13C



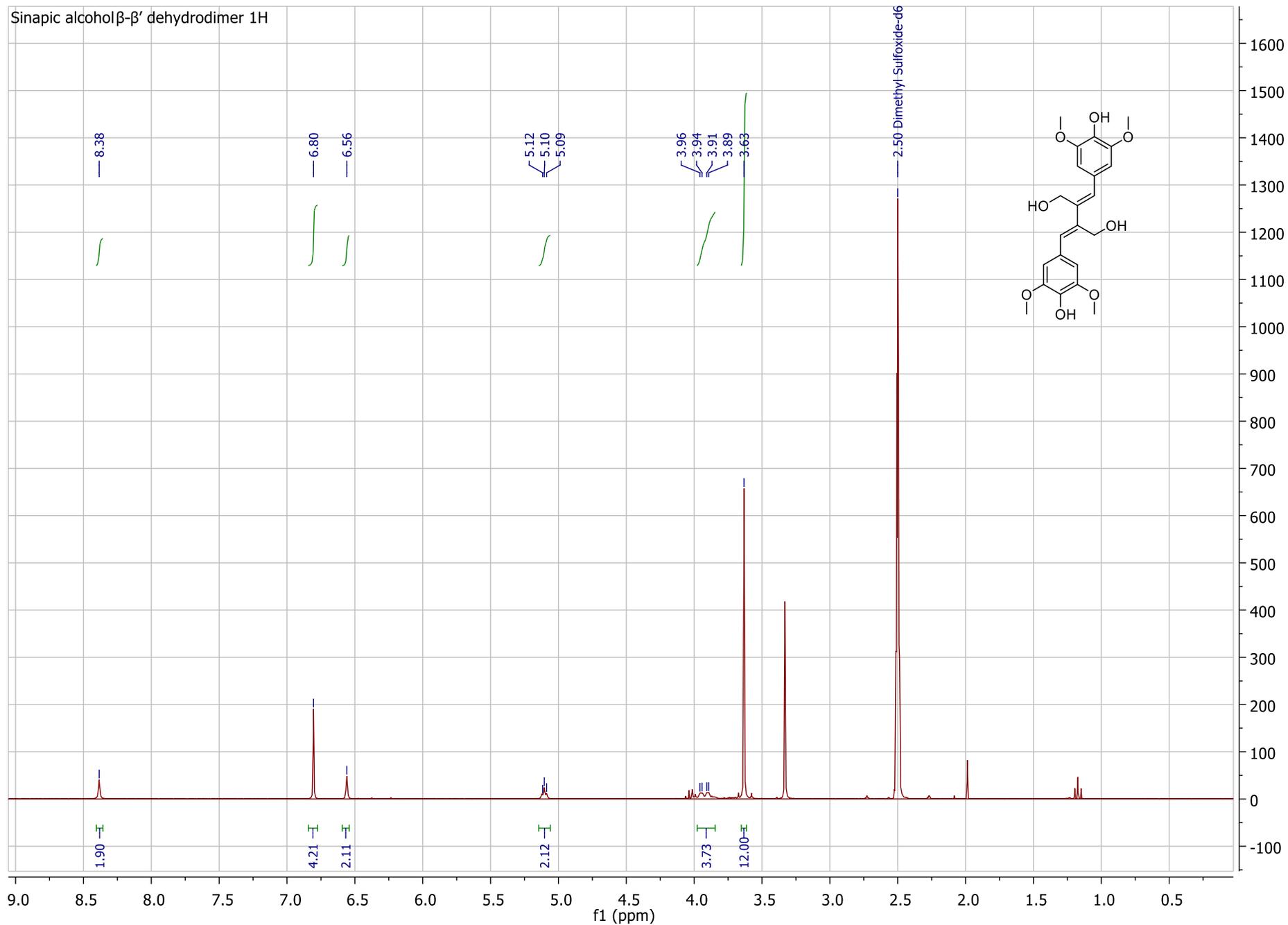
saturated ethyl sinapate $\beta$ - $\beta'$  dihydrodimer 1H



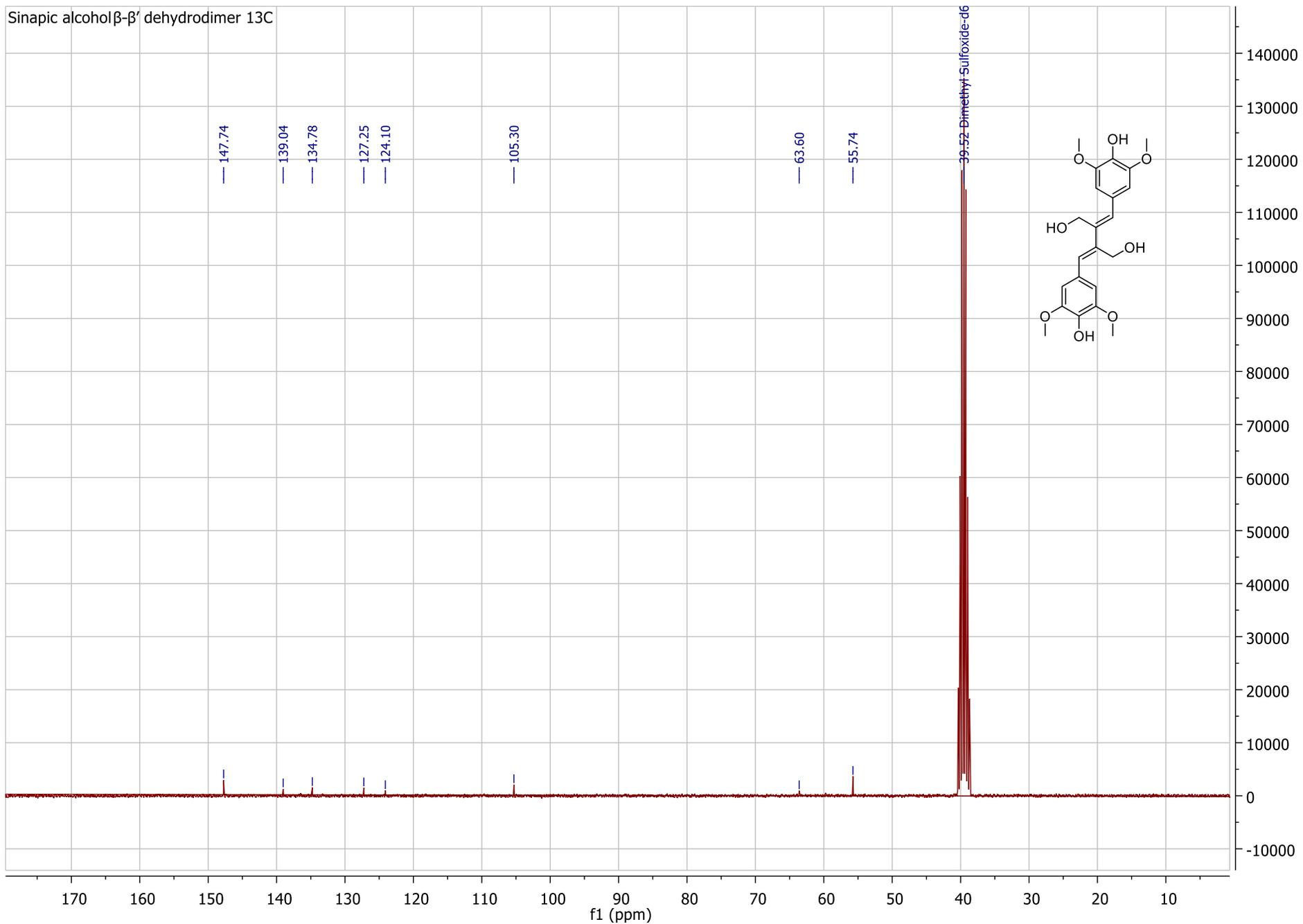
saturated ethyl sinapate $\beta$ - $\beta'$  dihydrodimer 13C



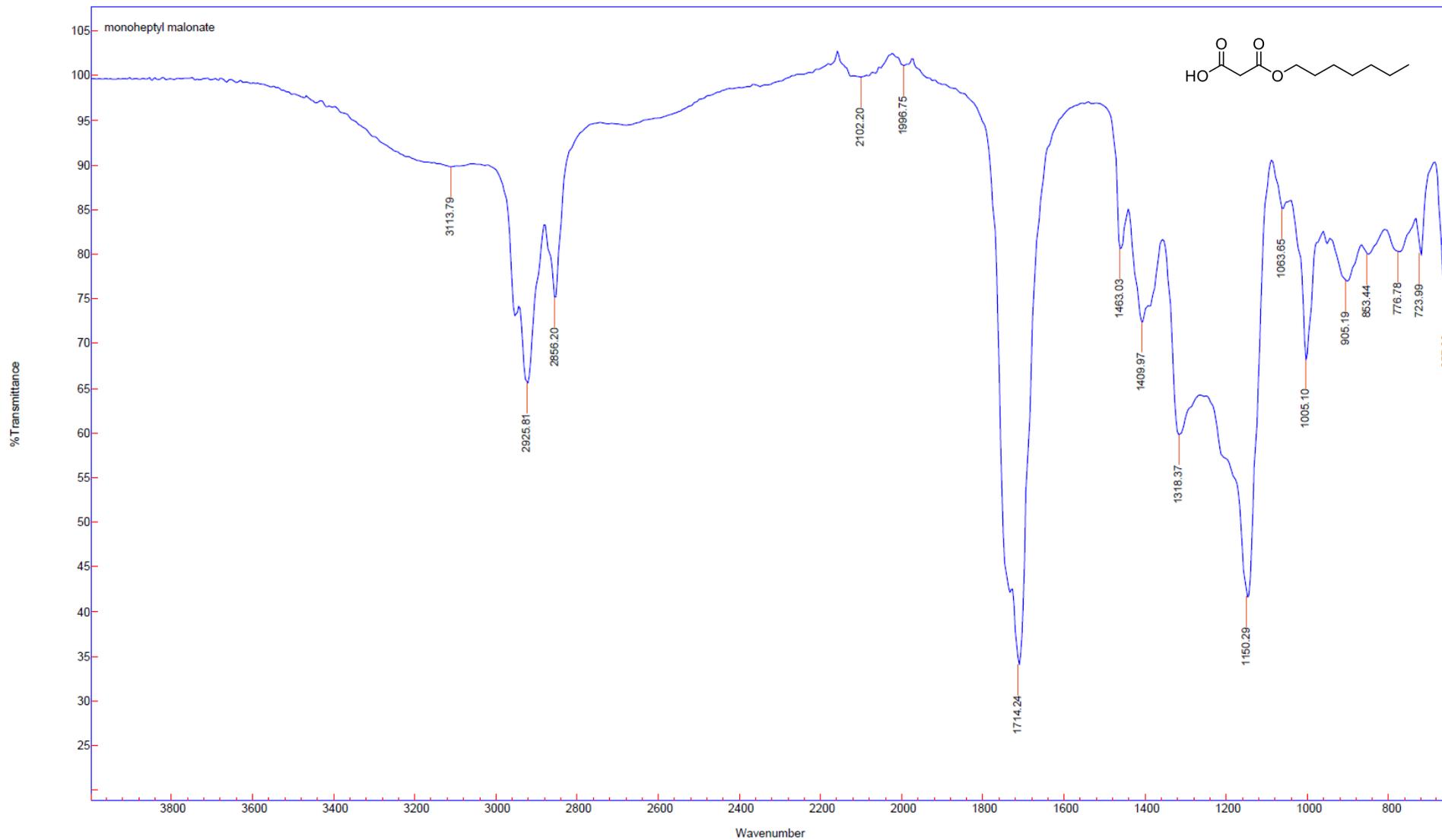
Sinapic alcohol-β-β' dehydrodimer 1H

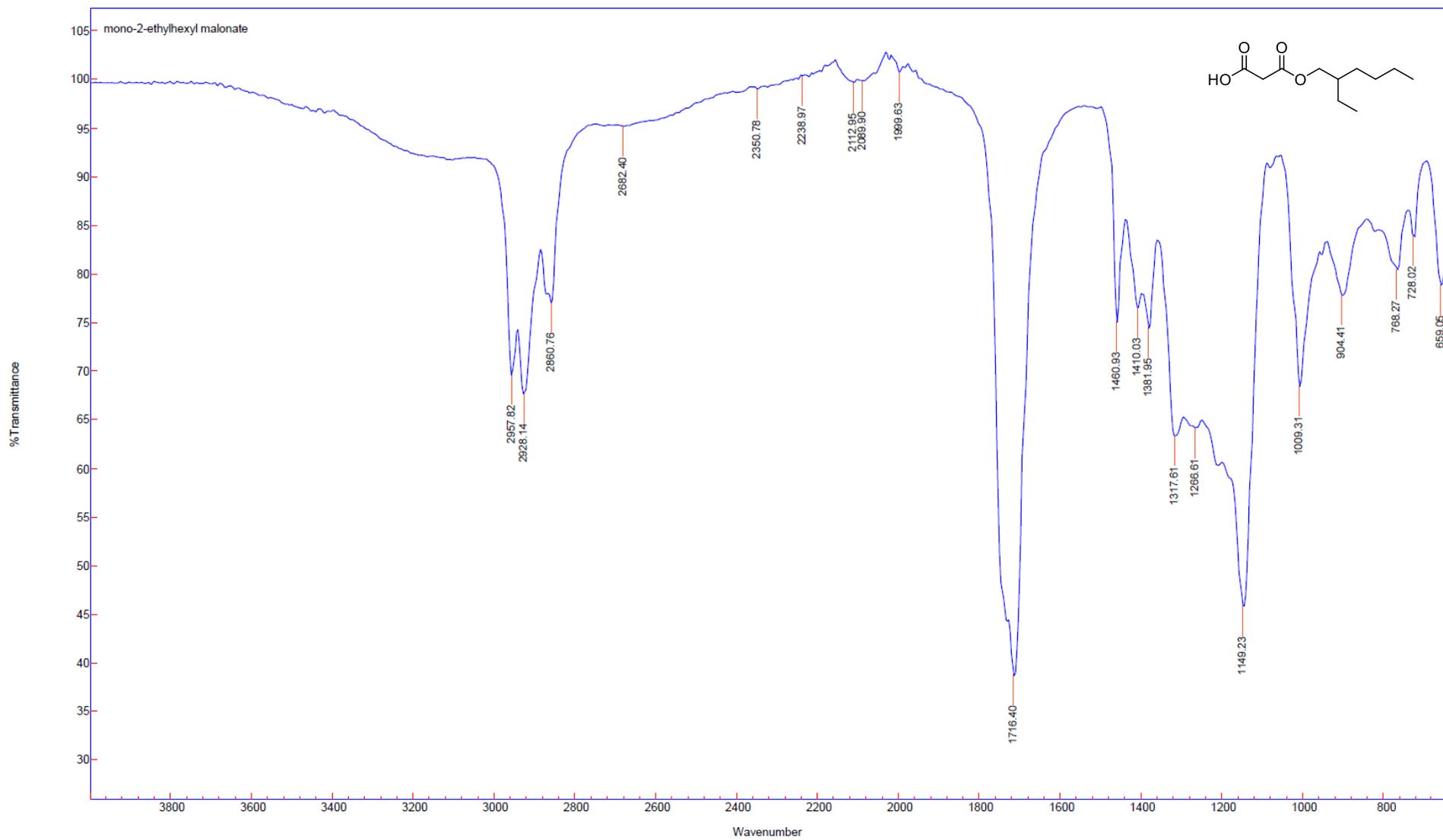


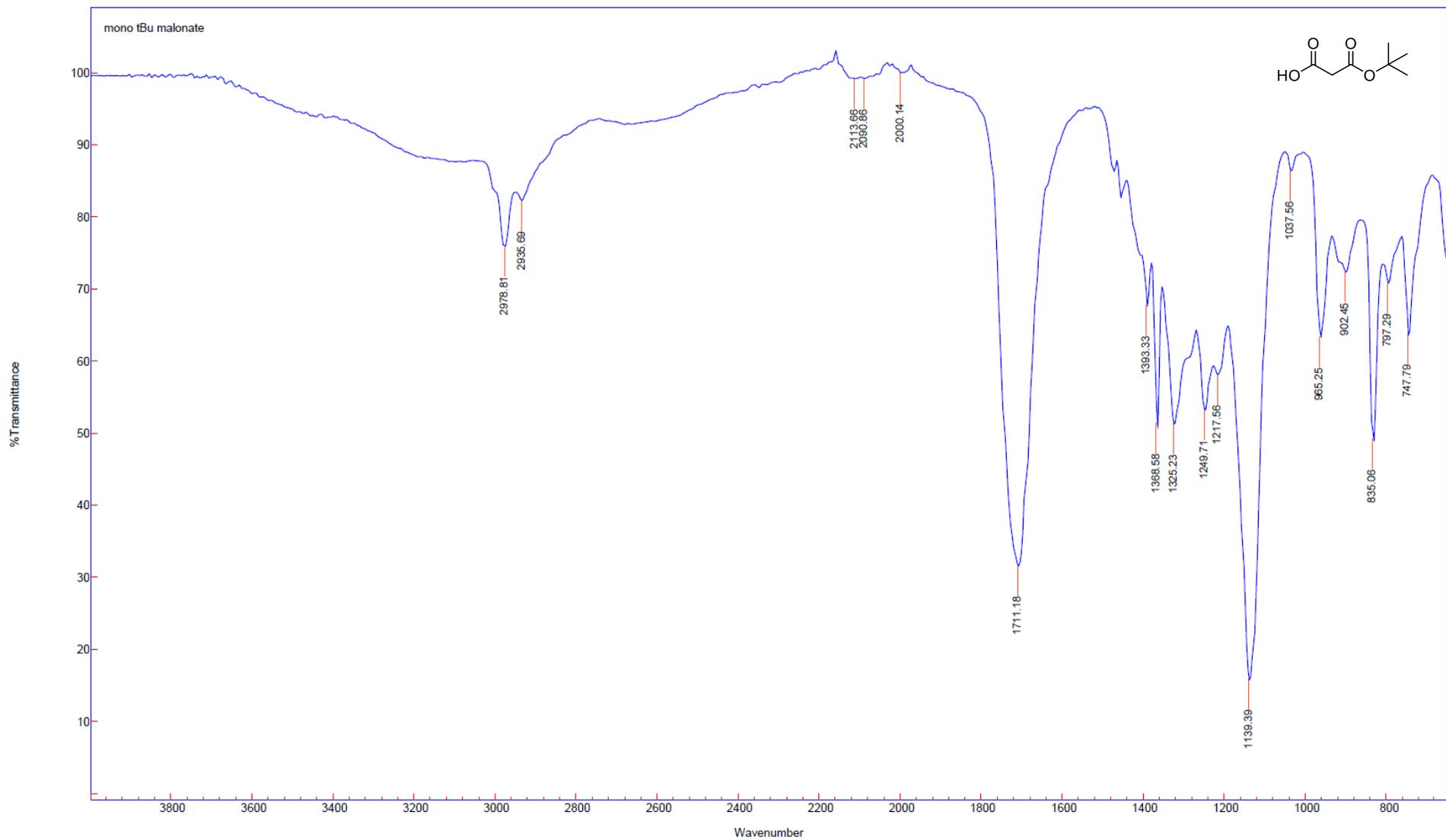
Sinapic alcohol- $\beta$ - $\beta'$  dehydrodimer 13C

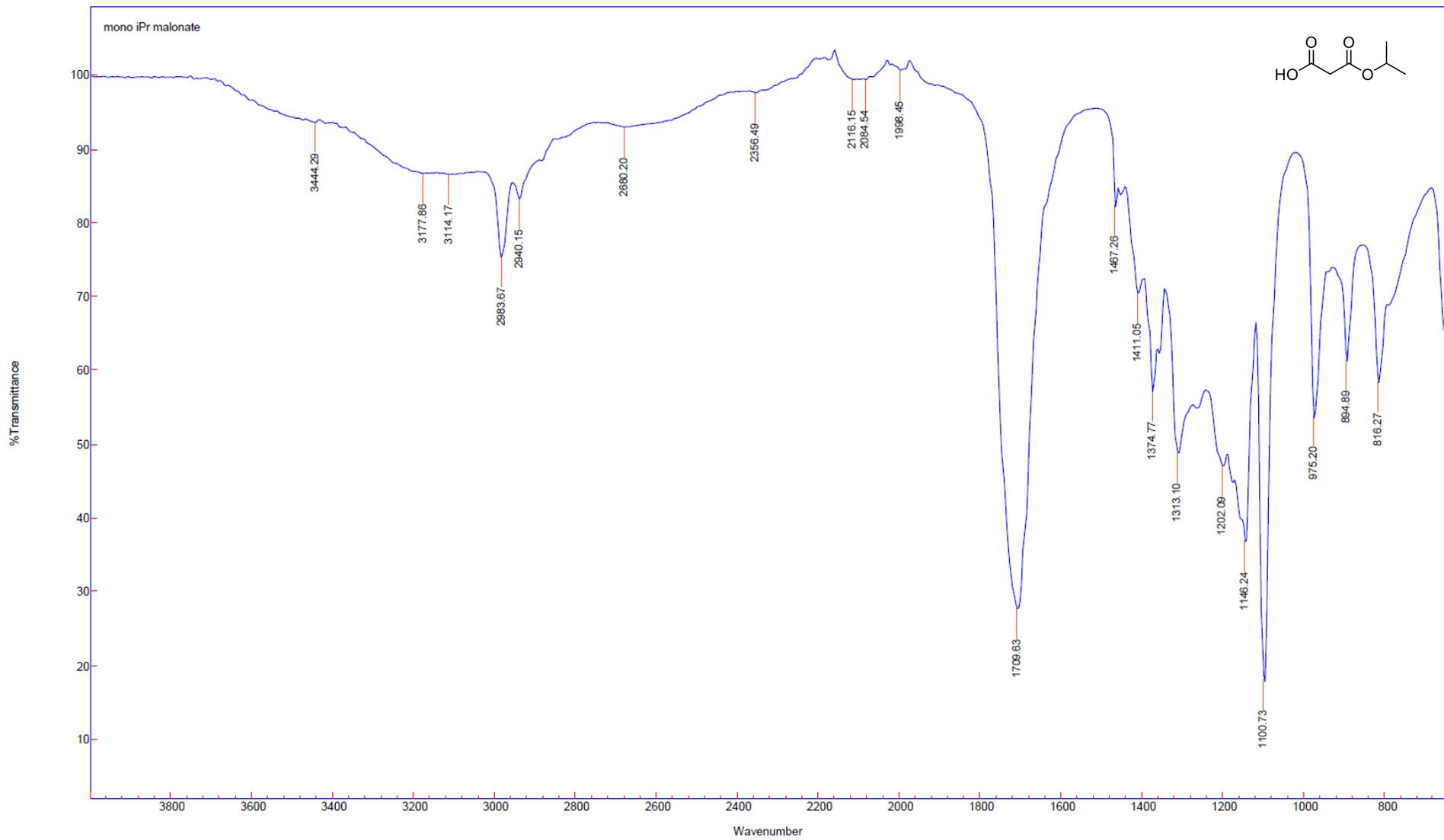


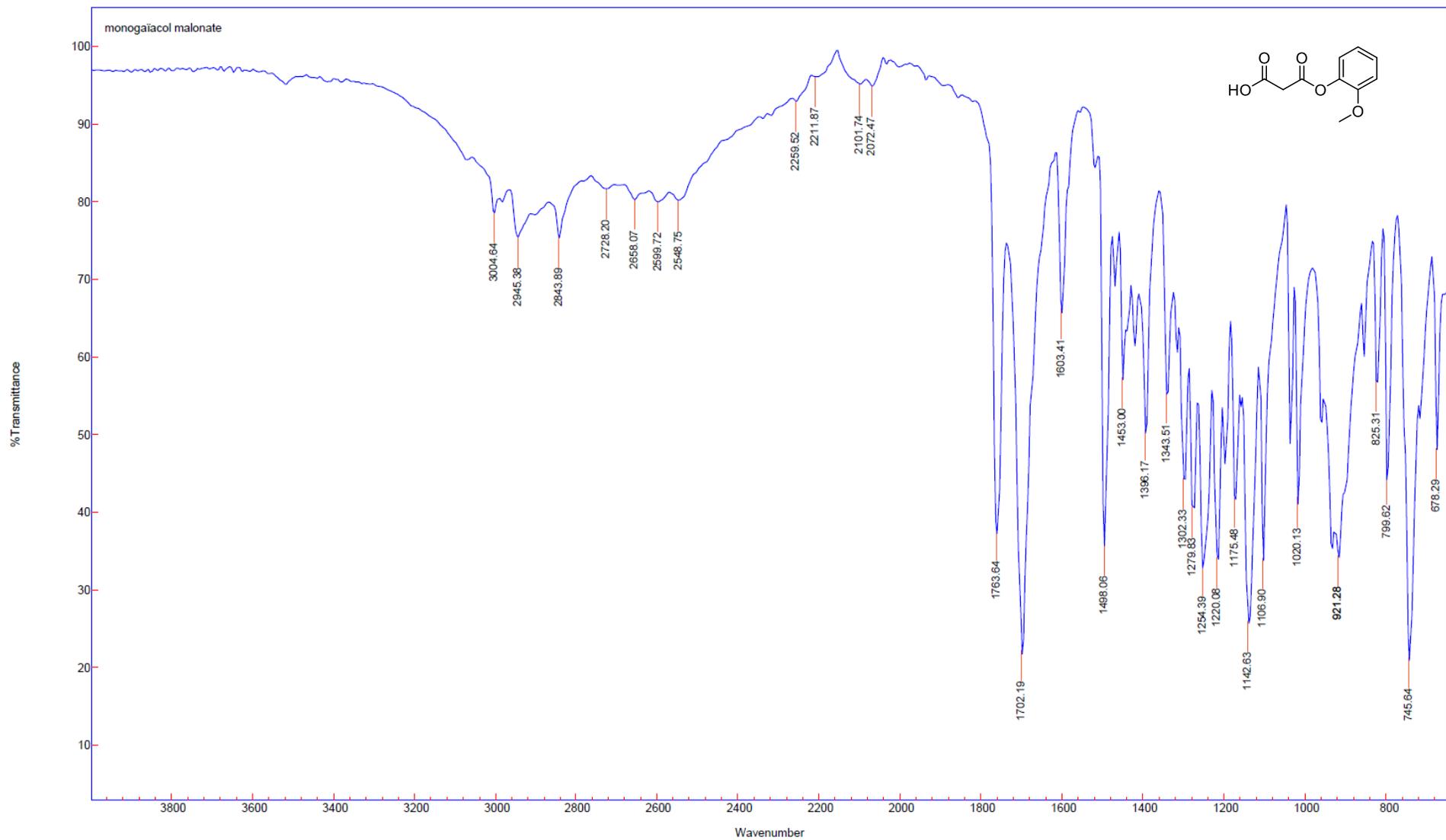
## 7.2. IR spectra

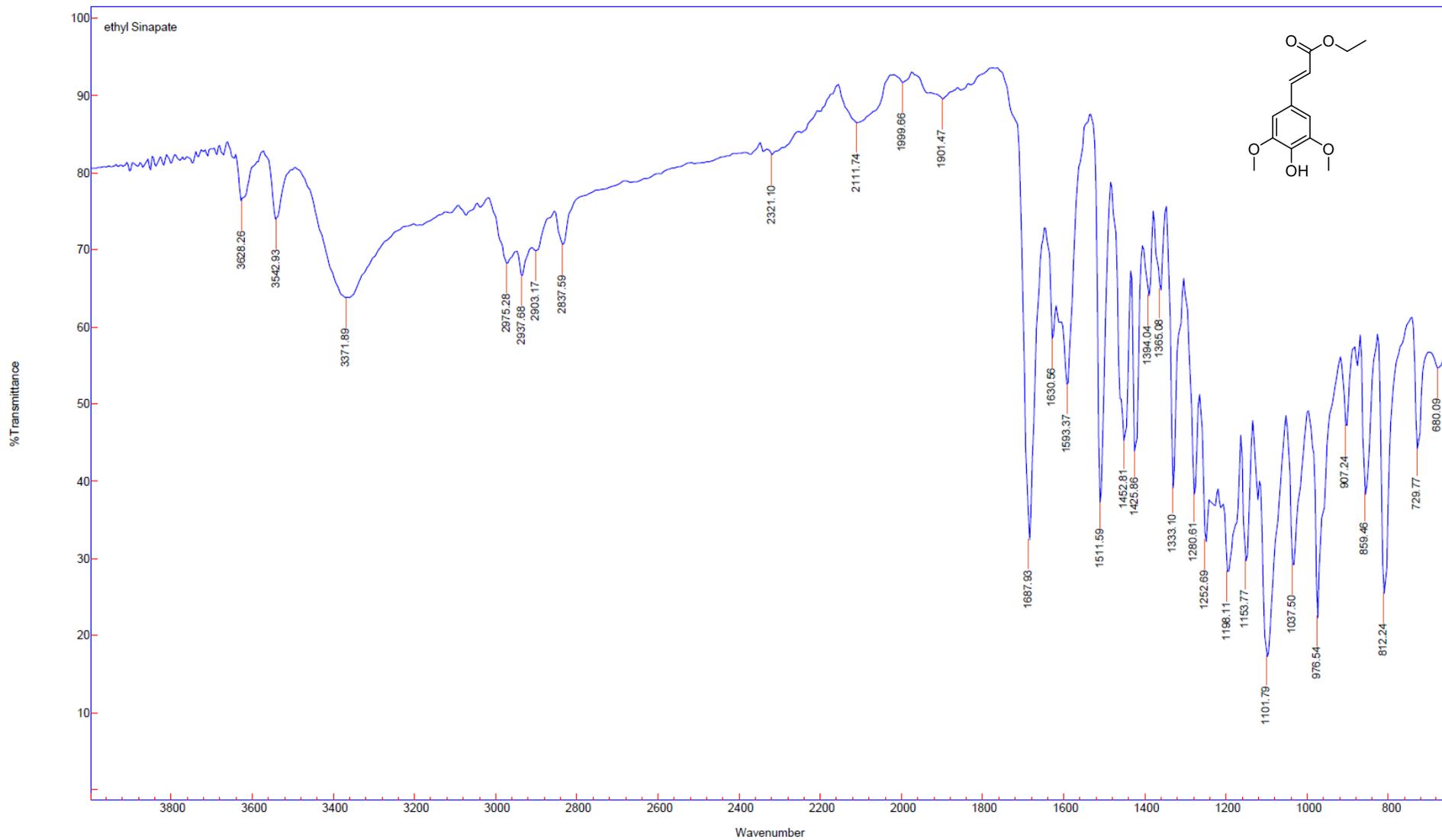


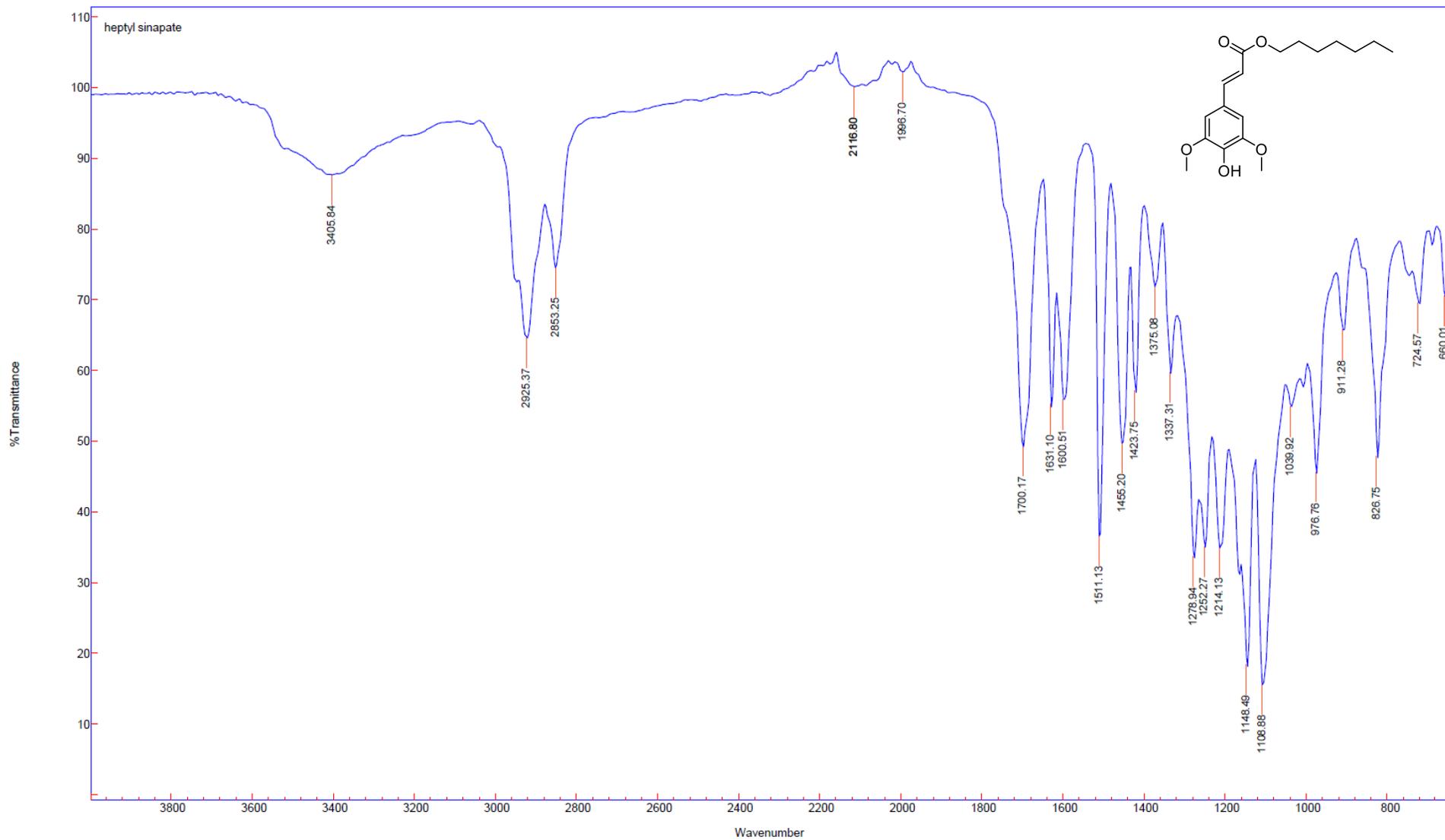


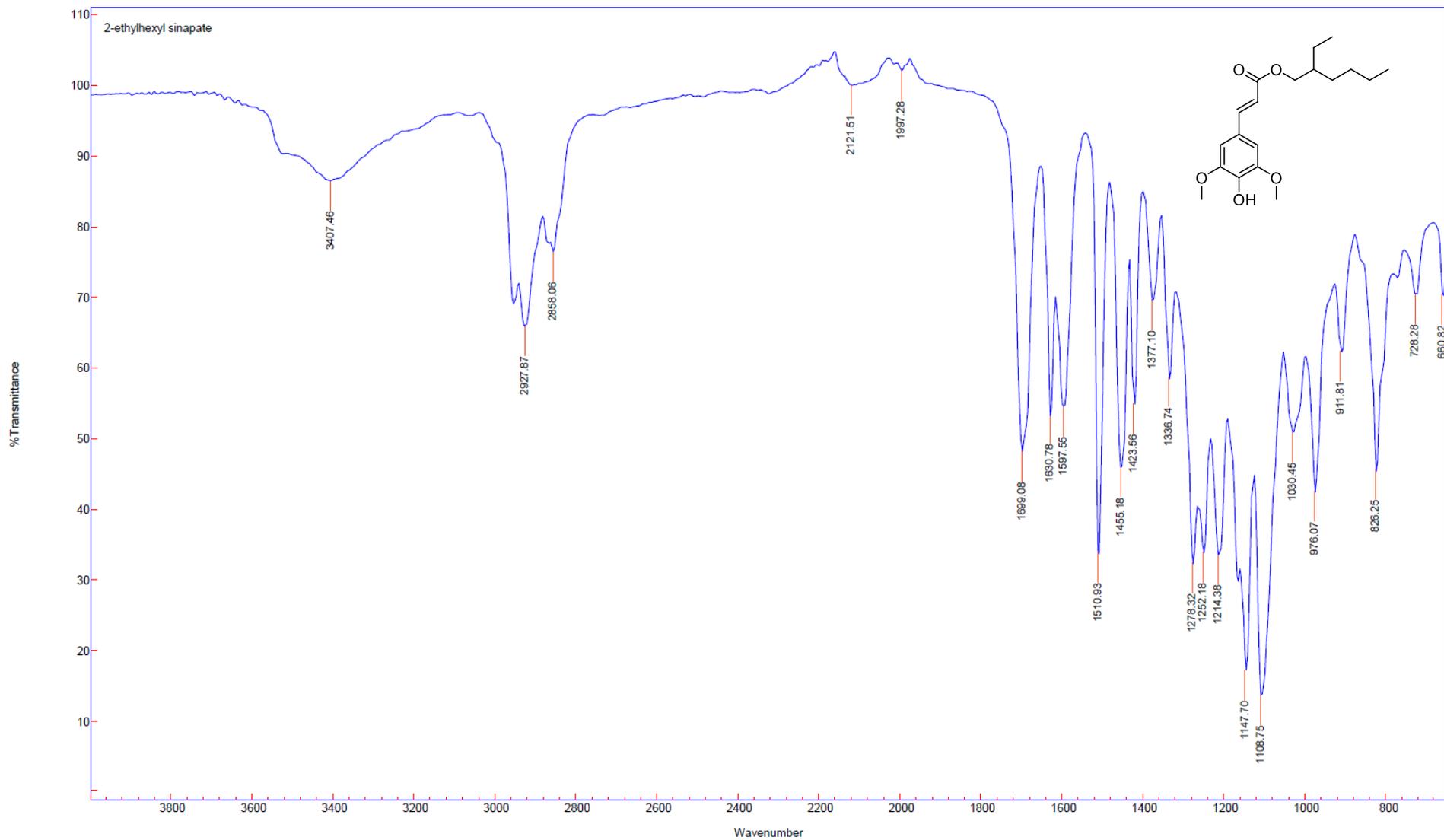


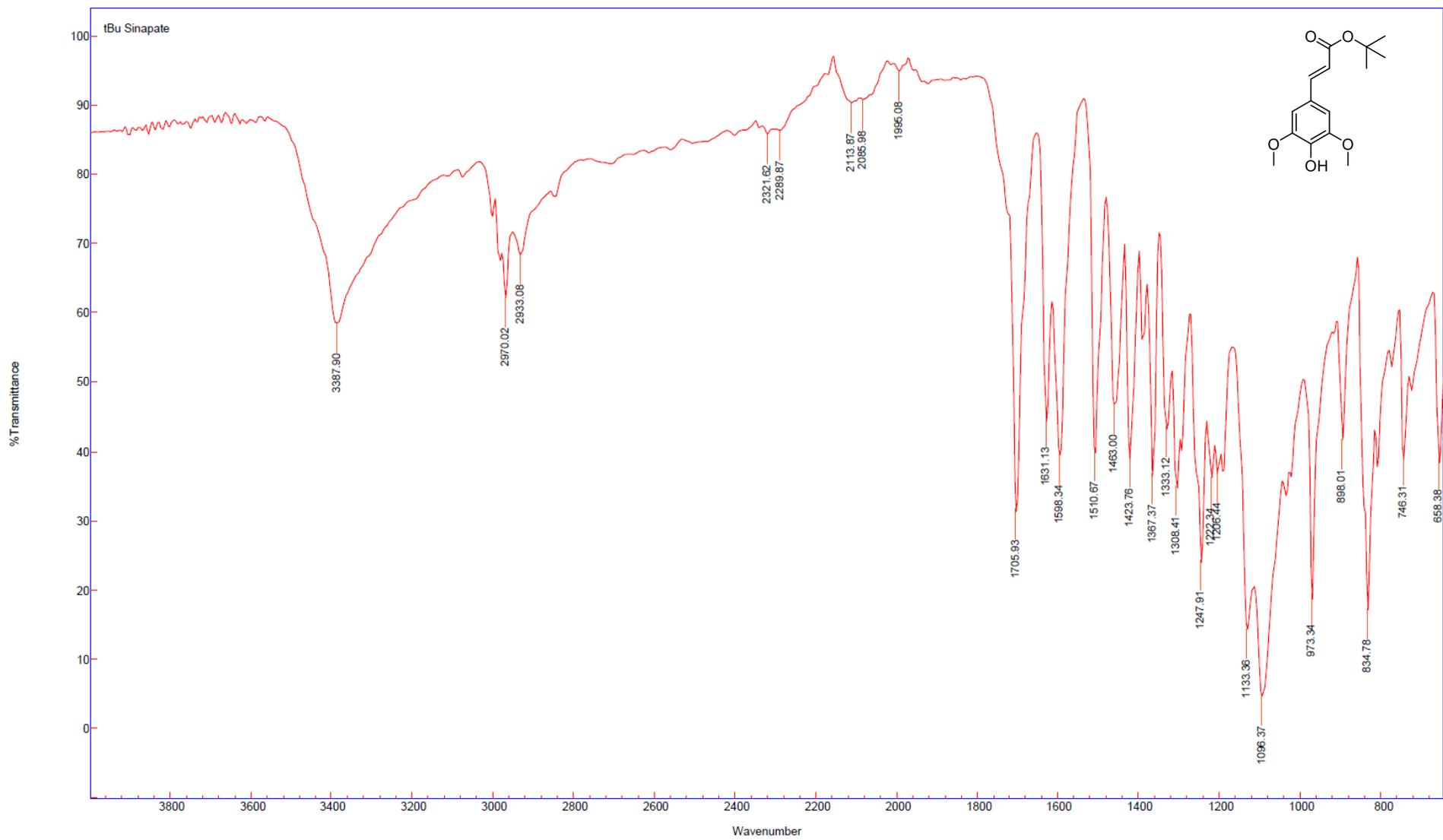


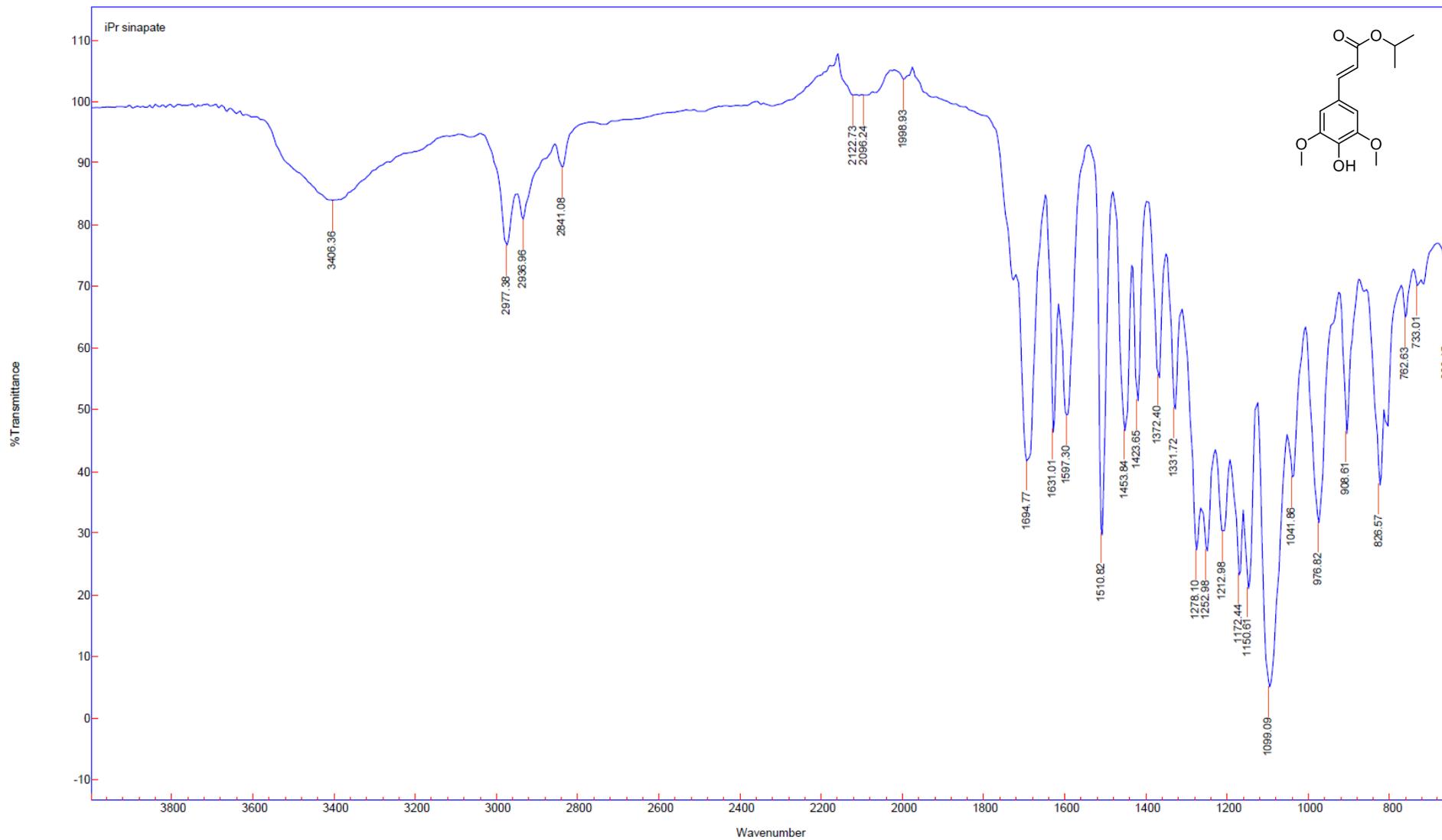


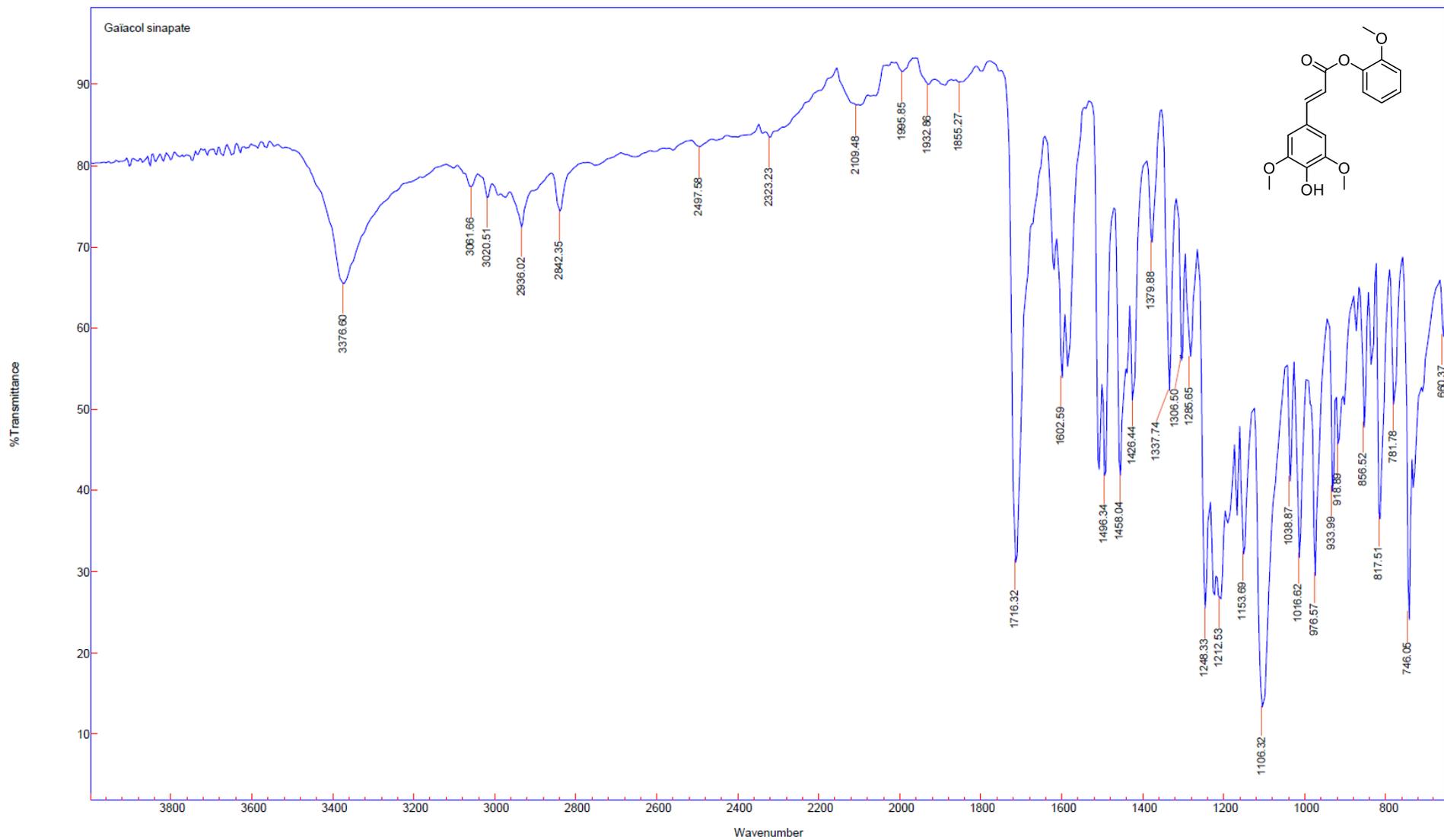


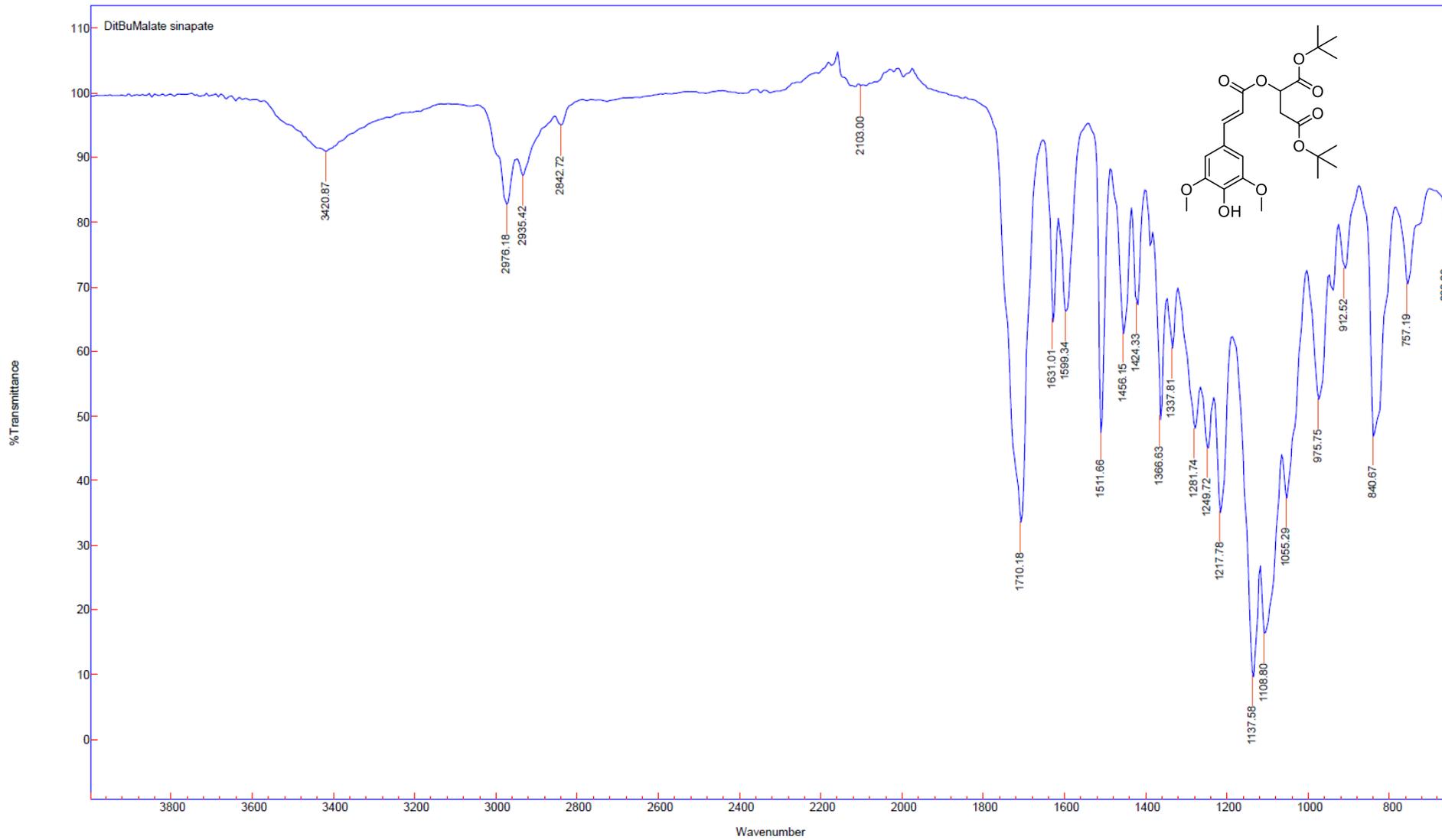


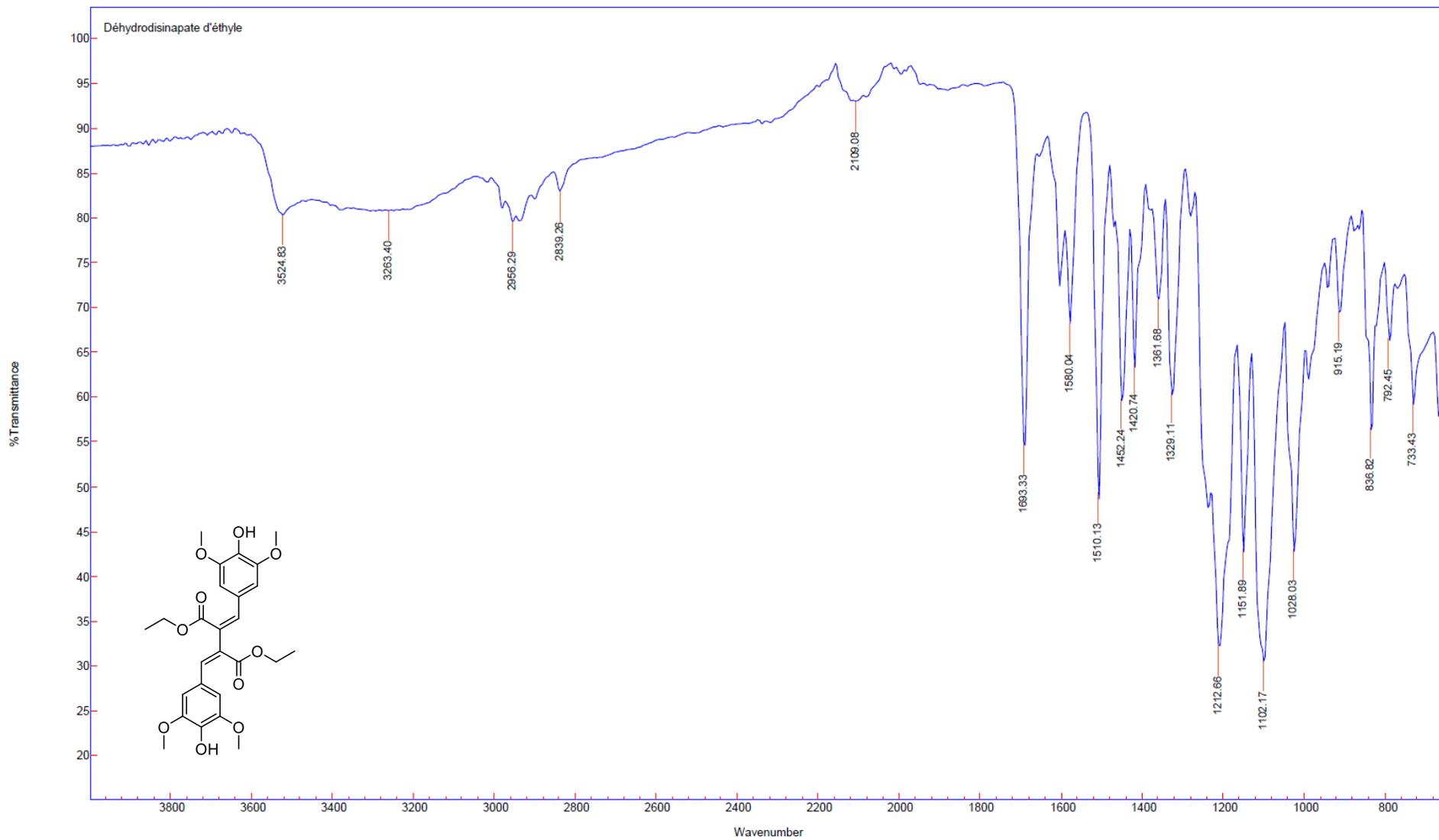


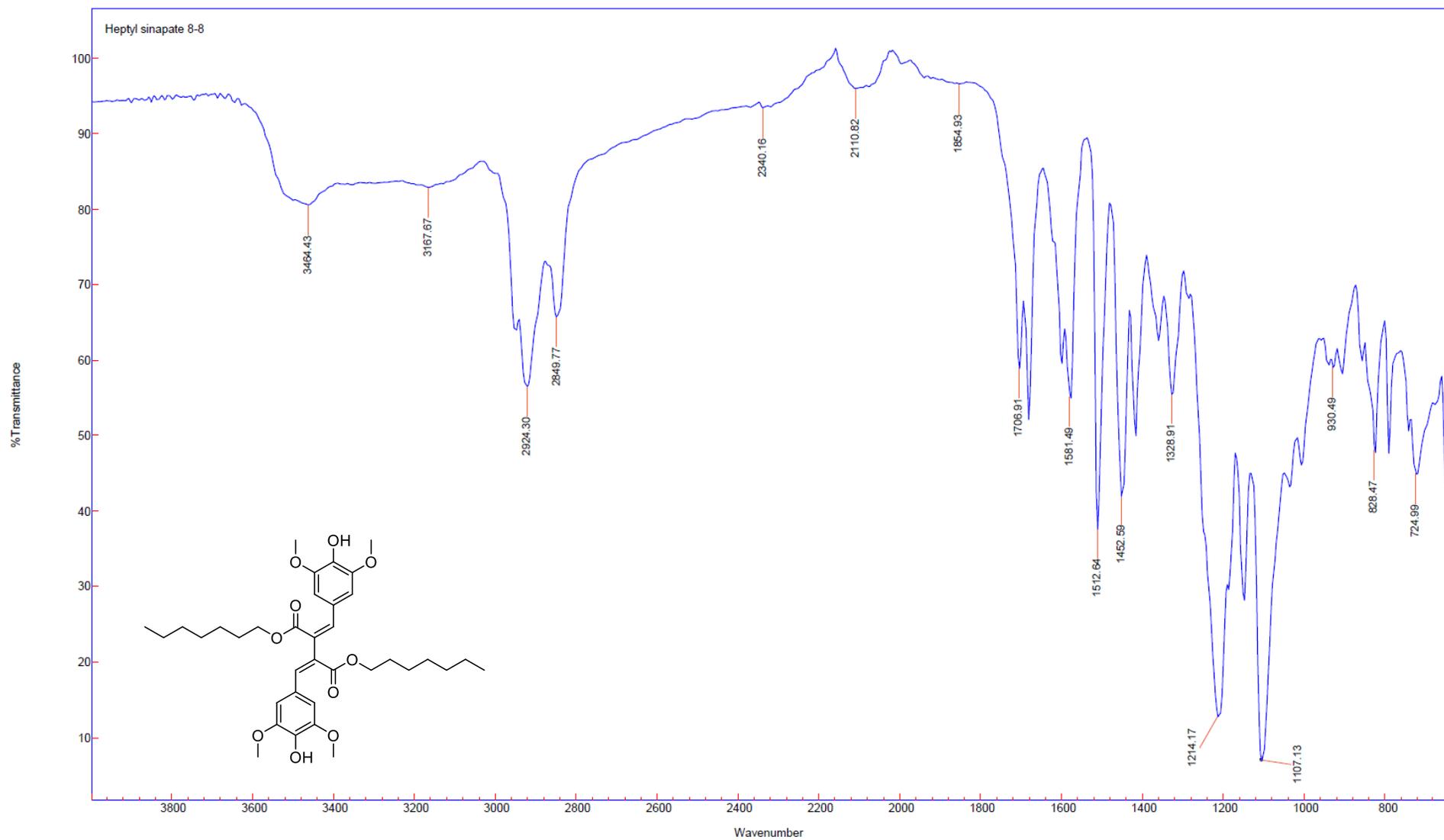


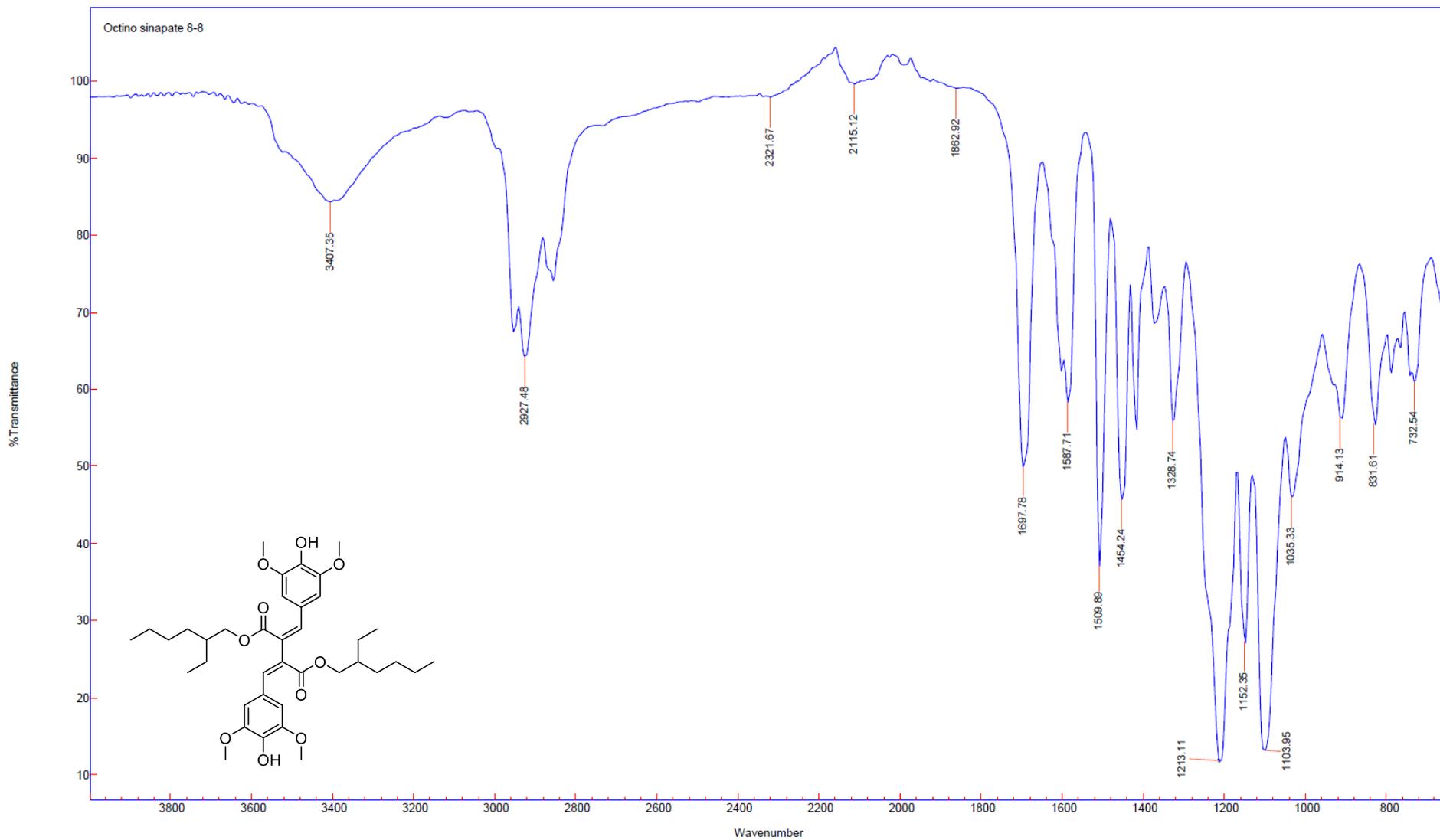


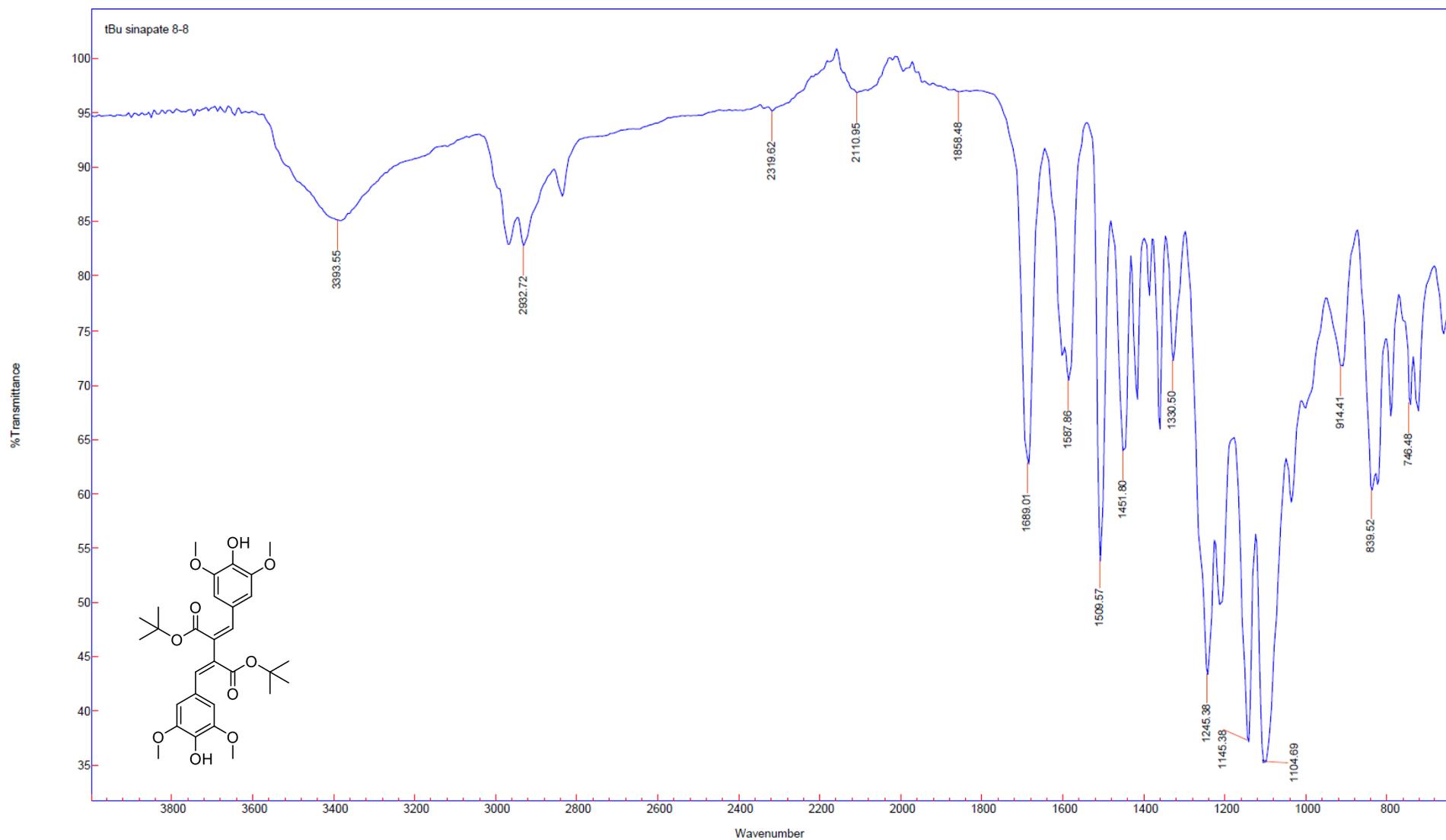


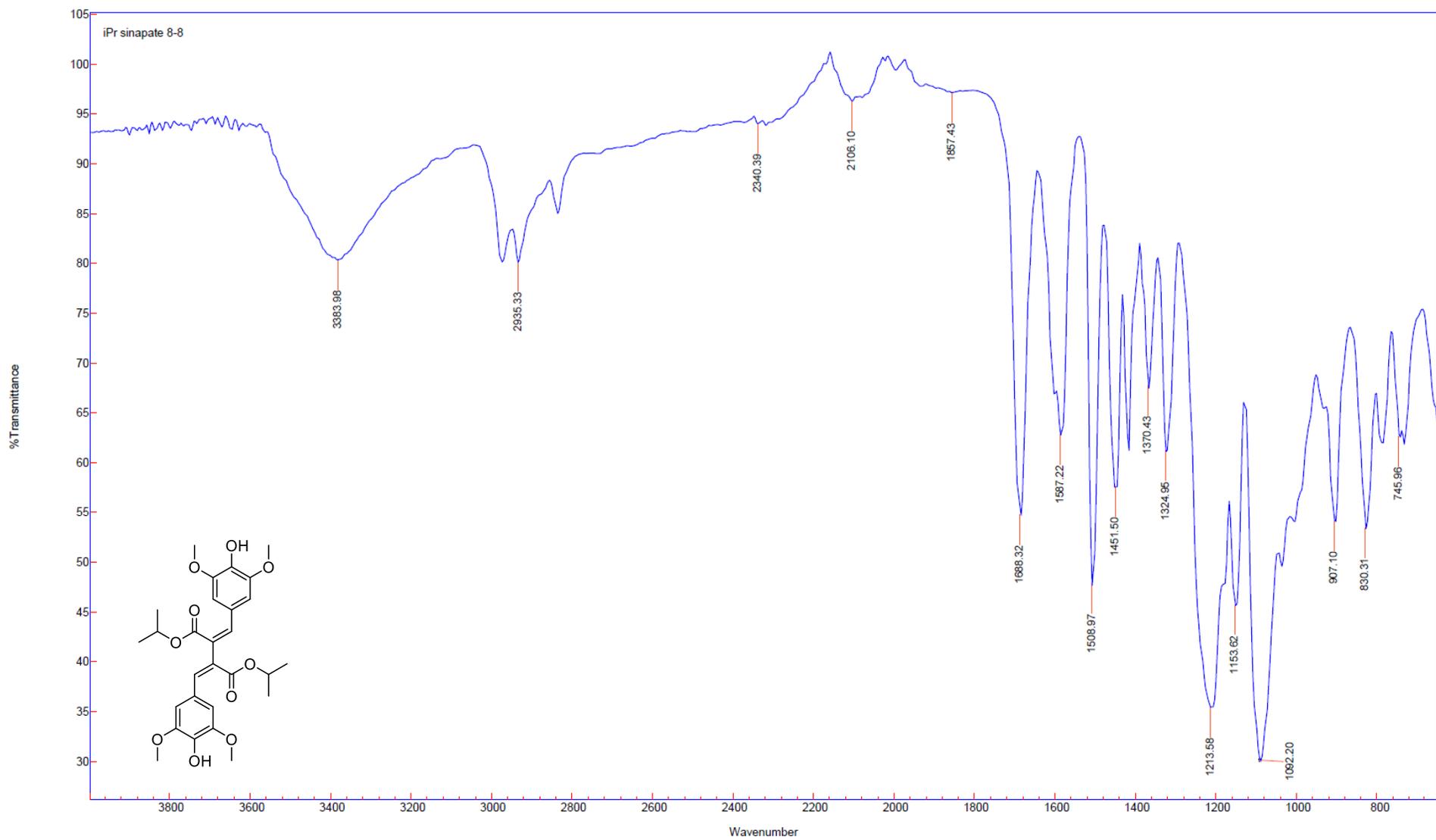


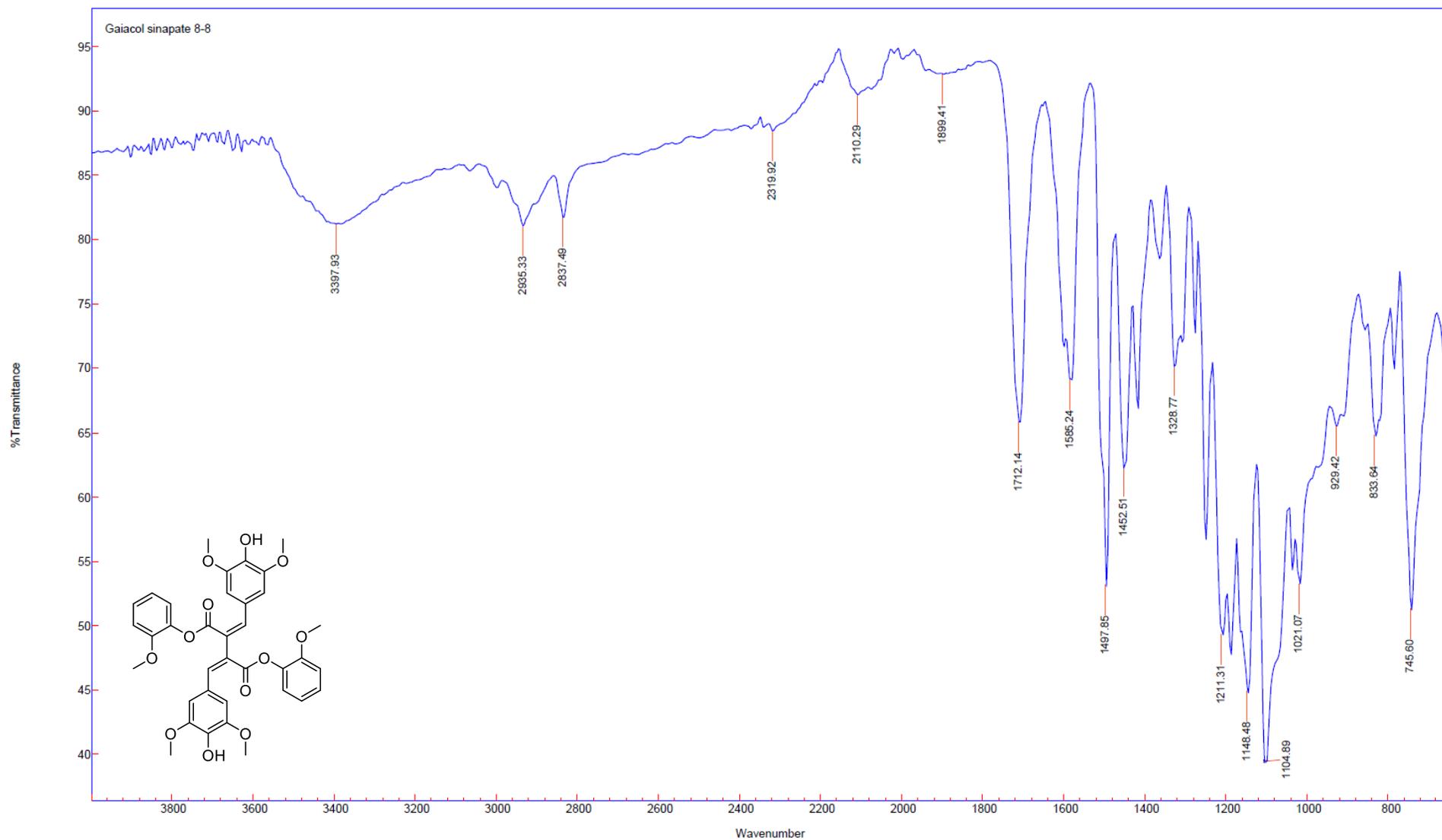


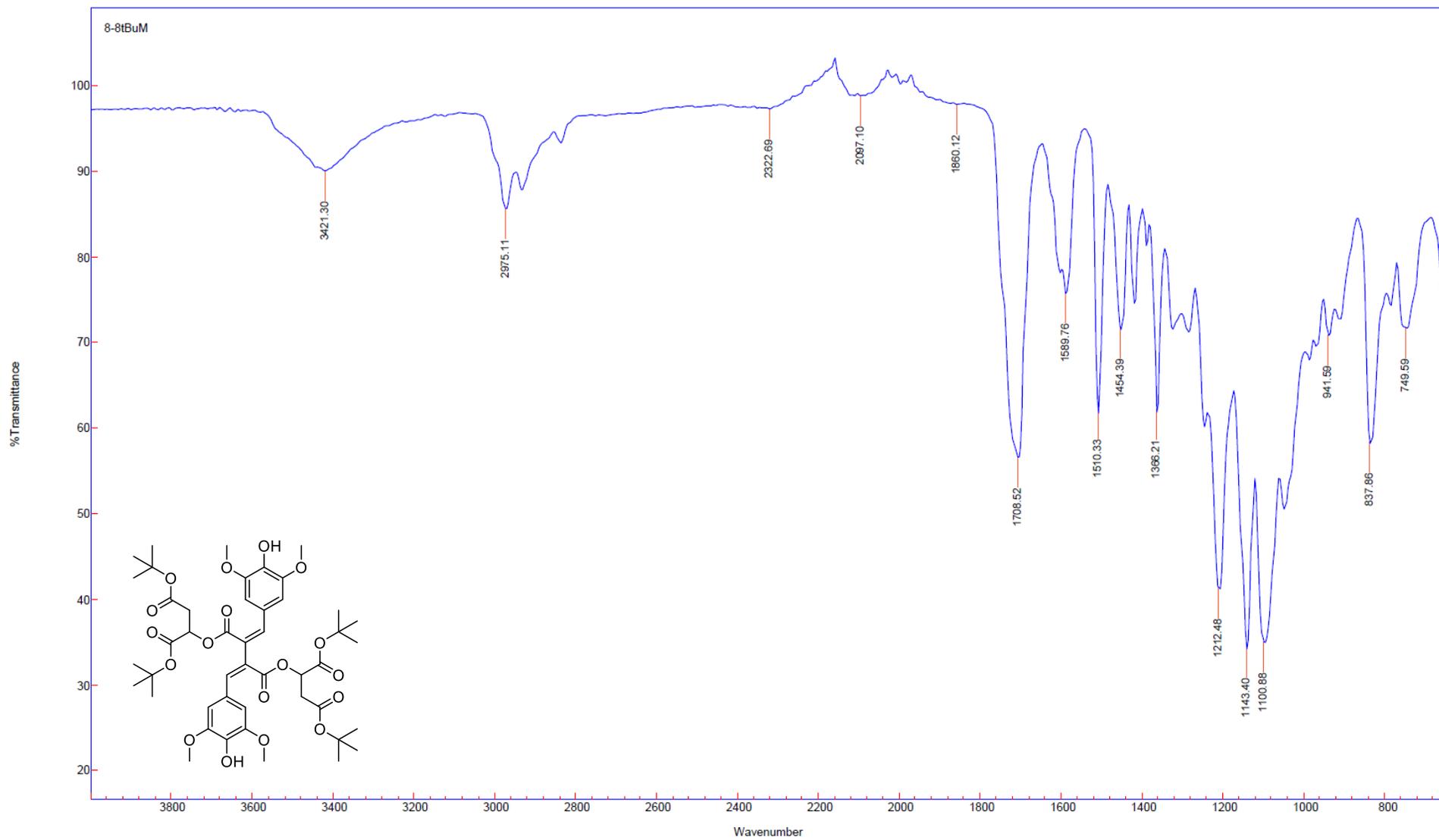


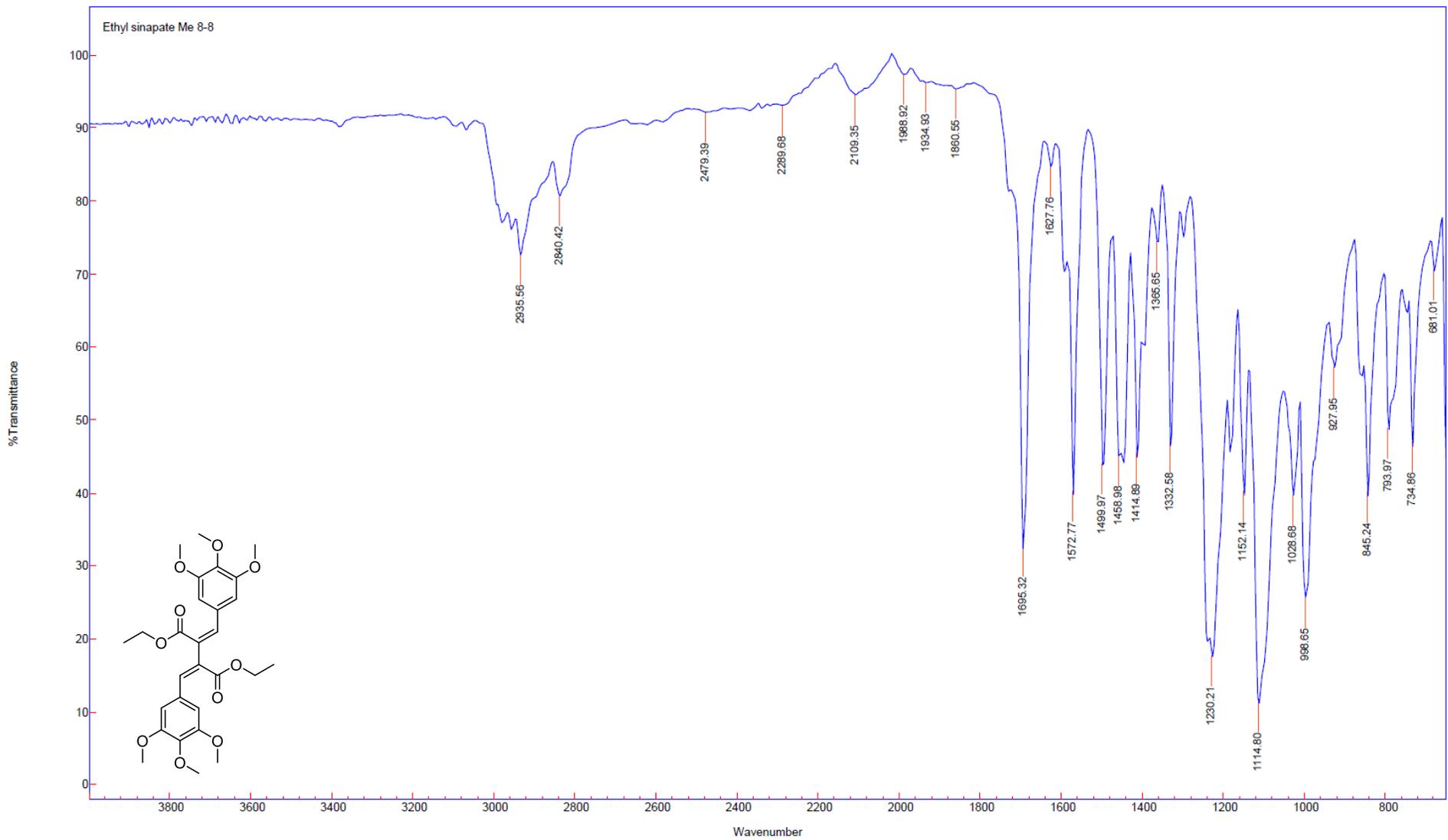


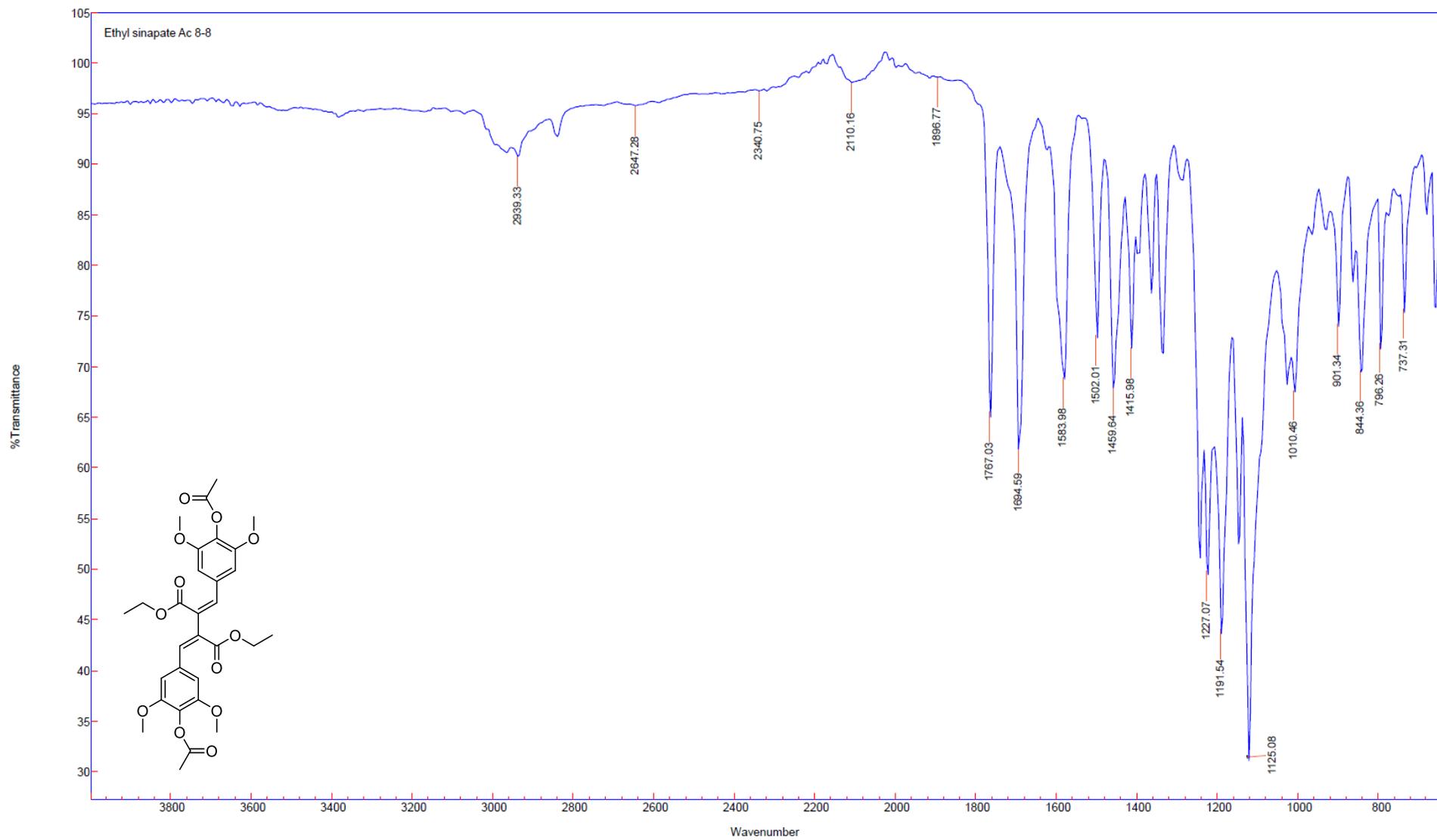


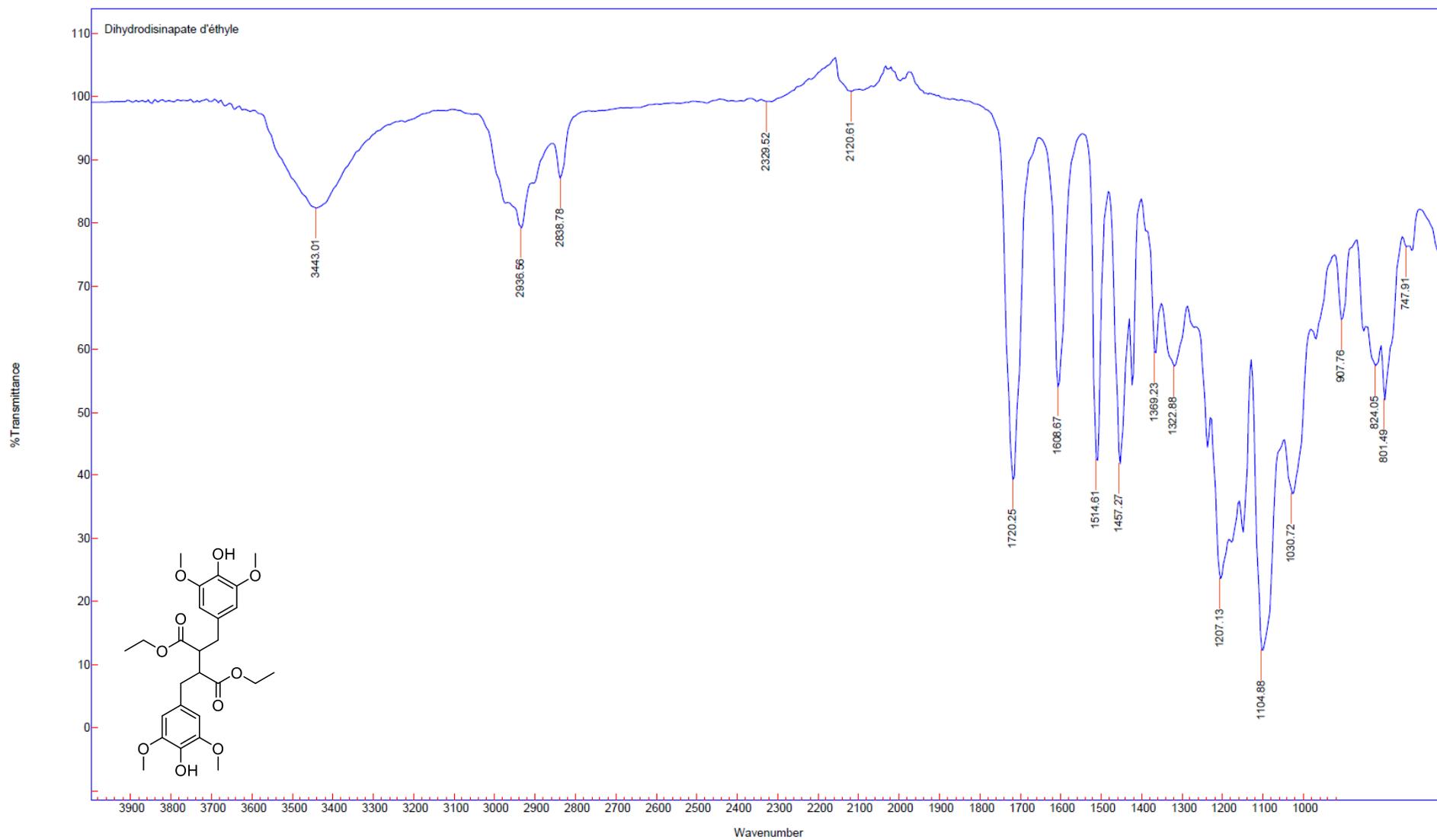


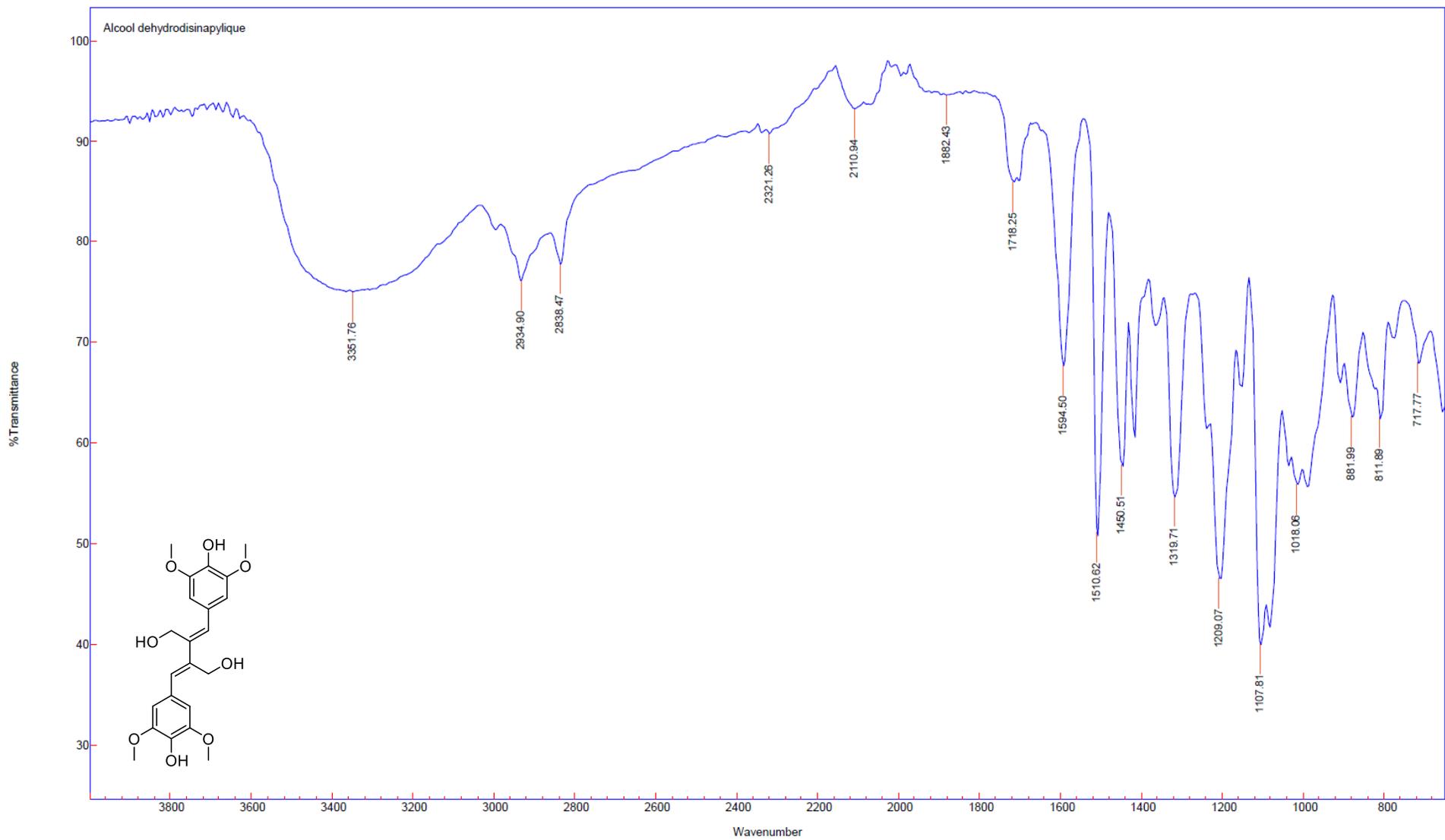




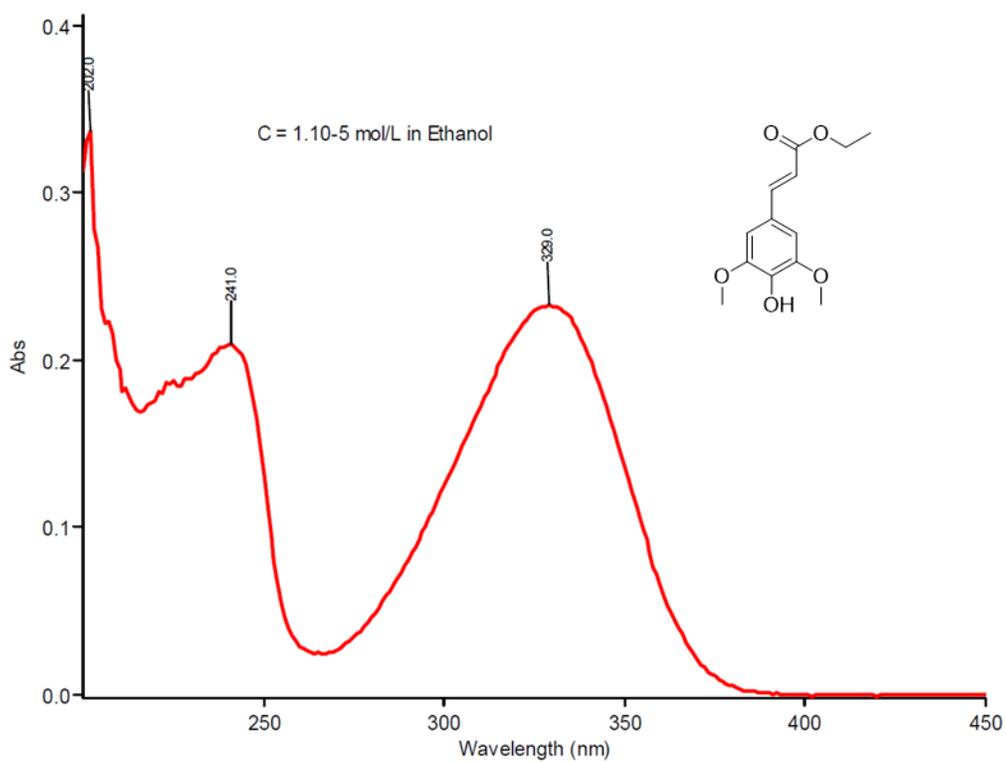
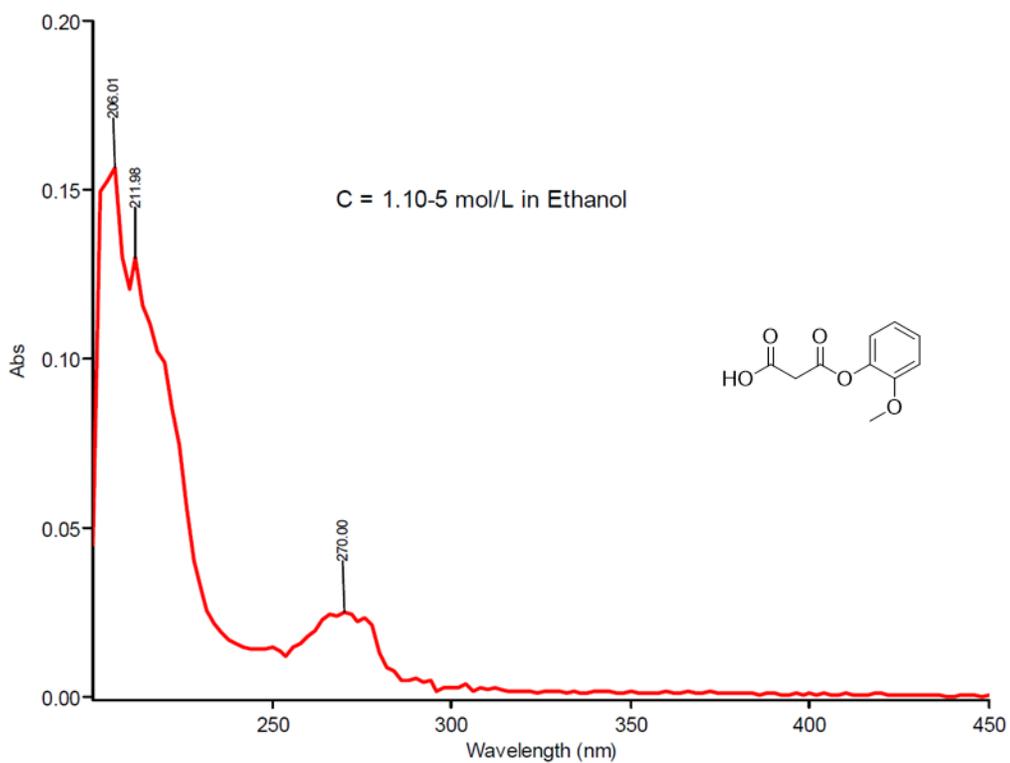


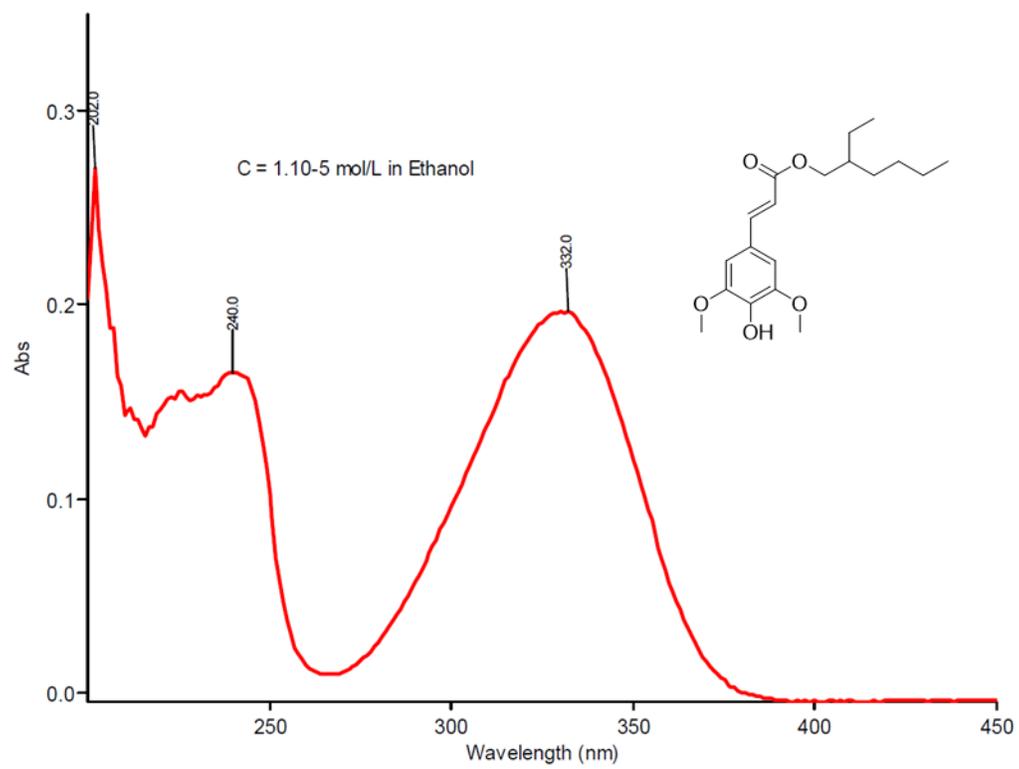
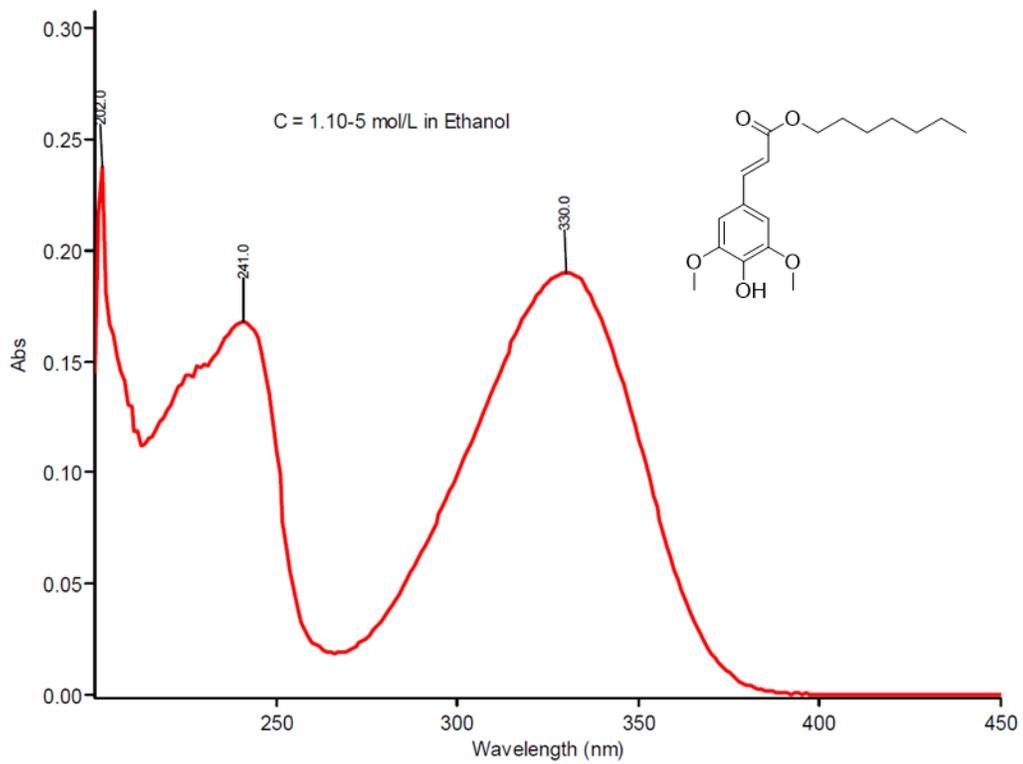


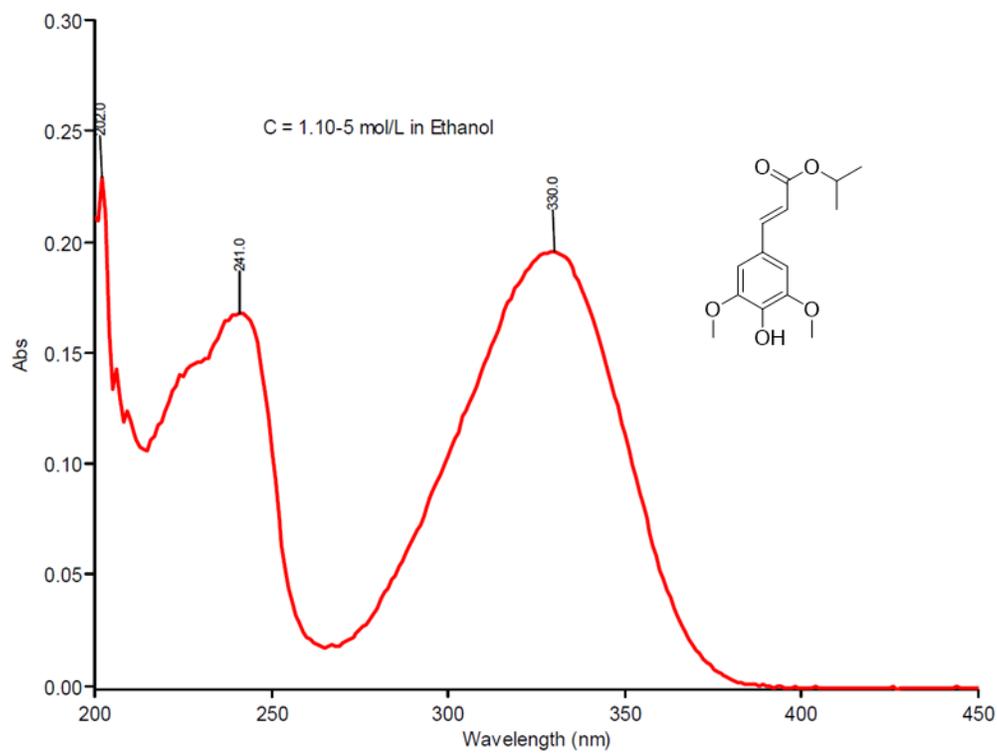
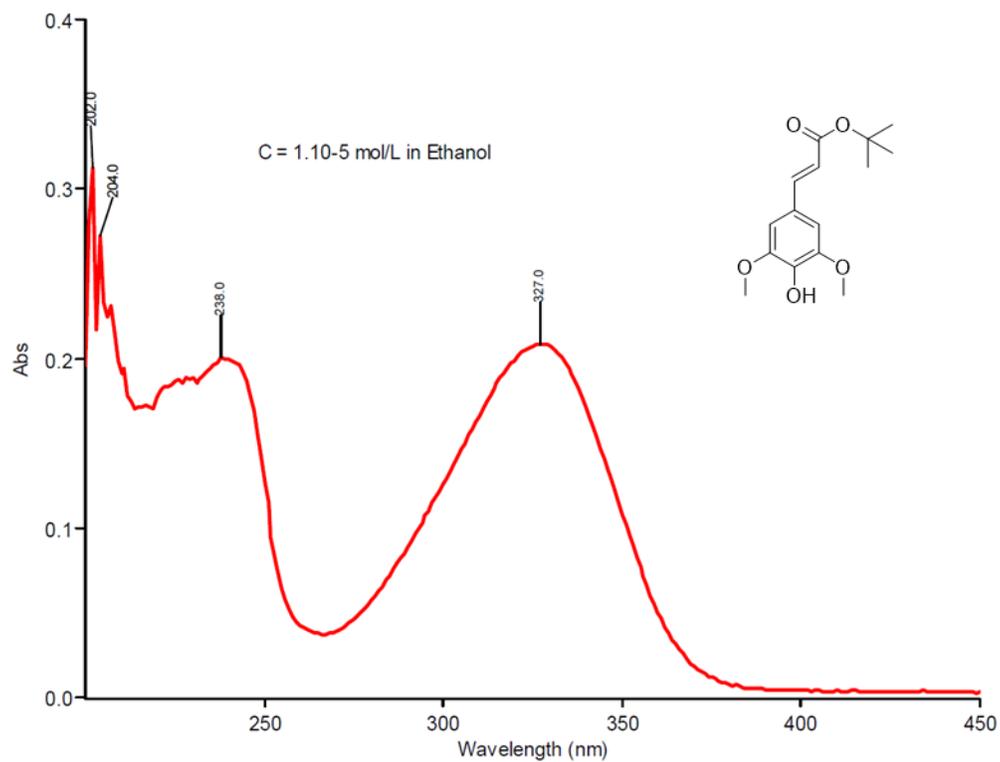




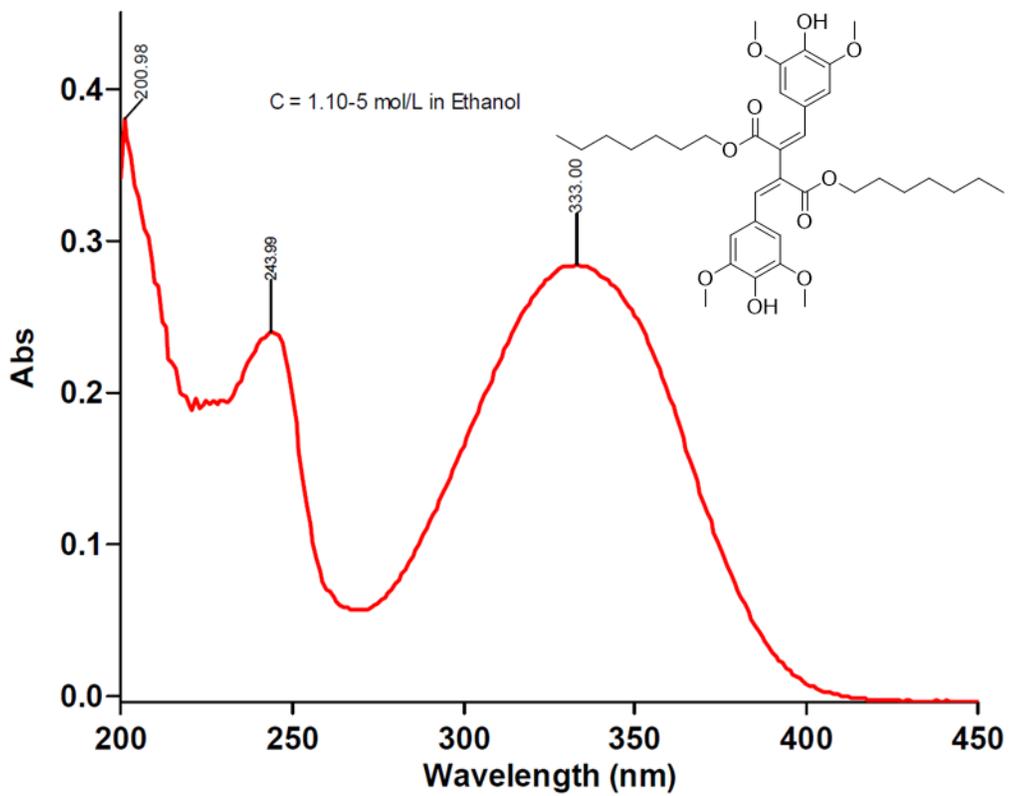
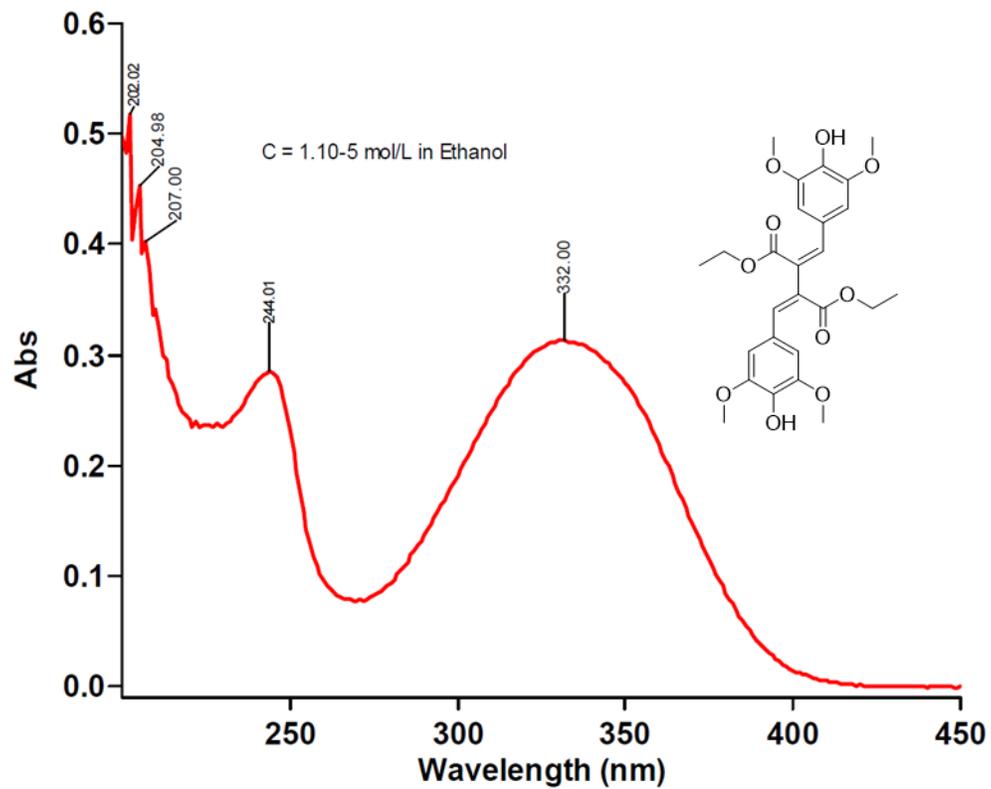
### 7.3. UV Spectra

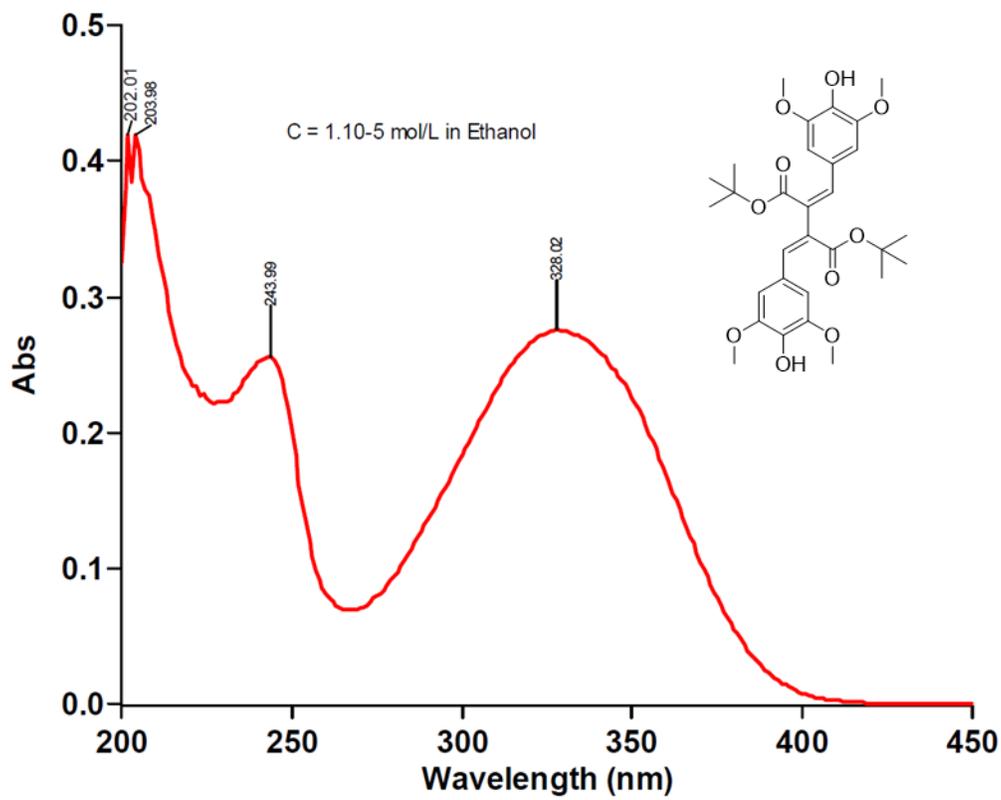
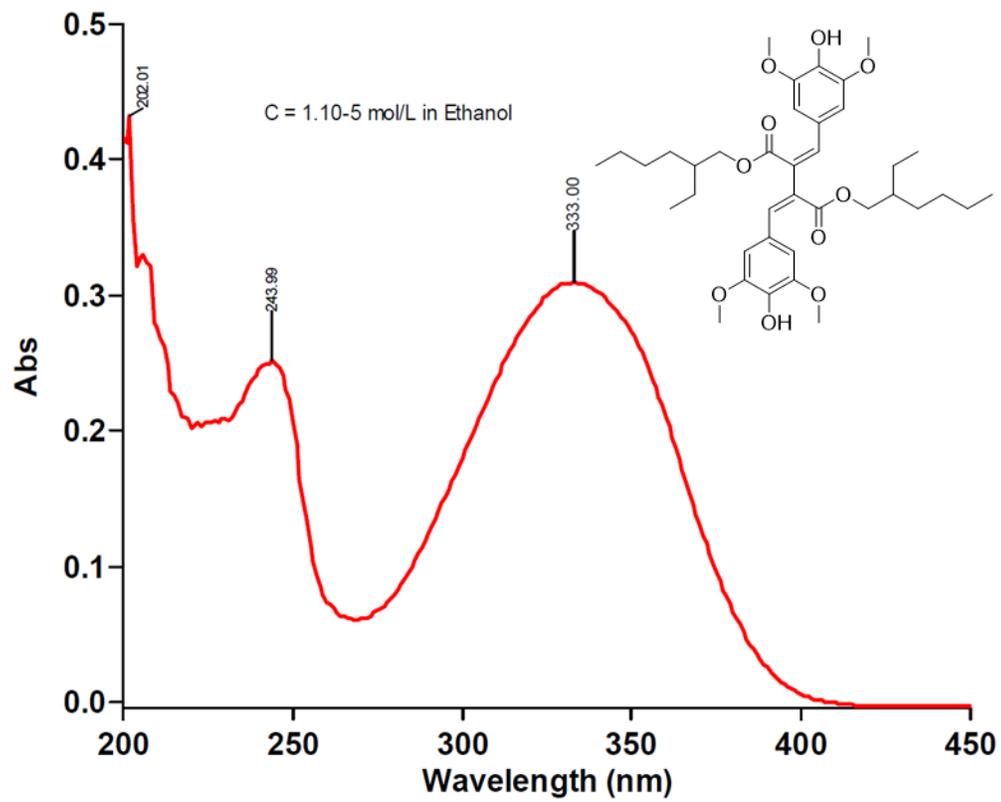


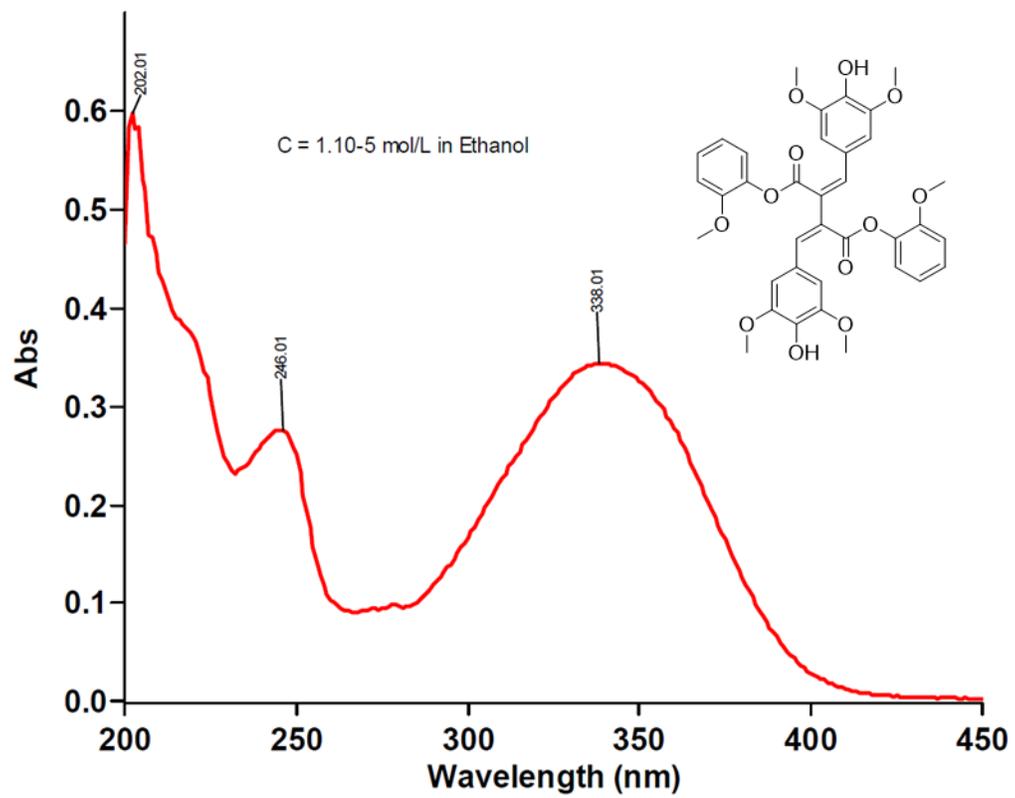
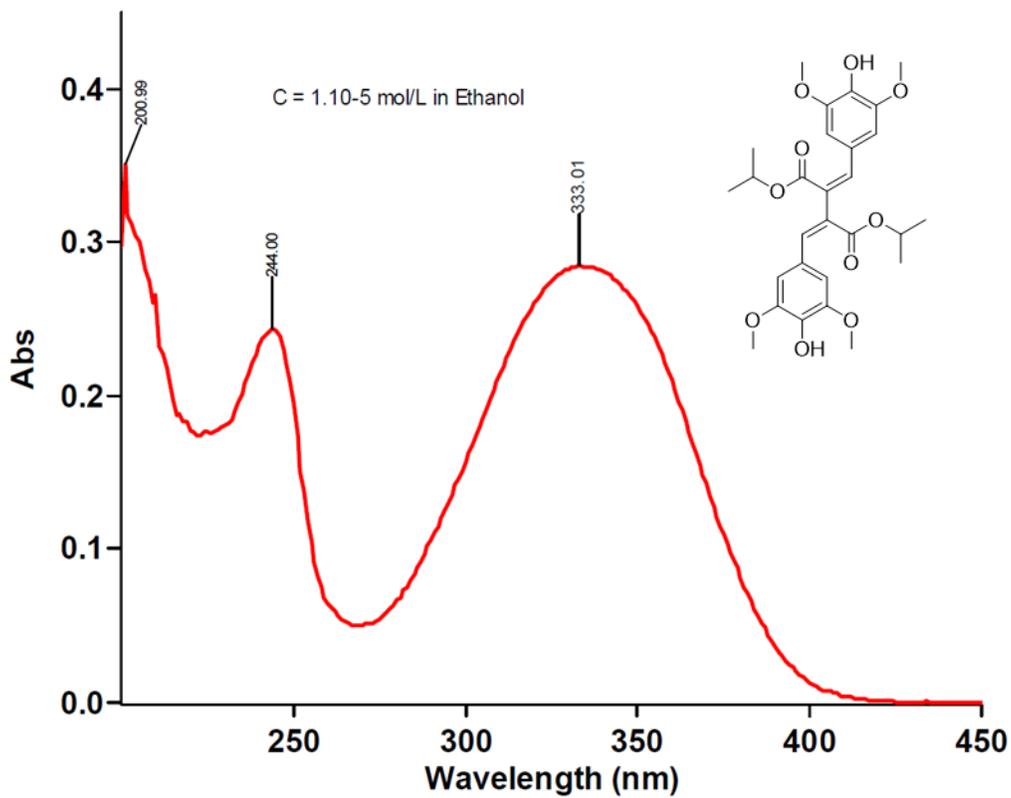


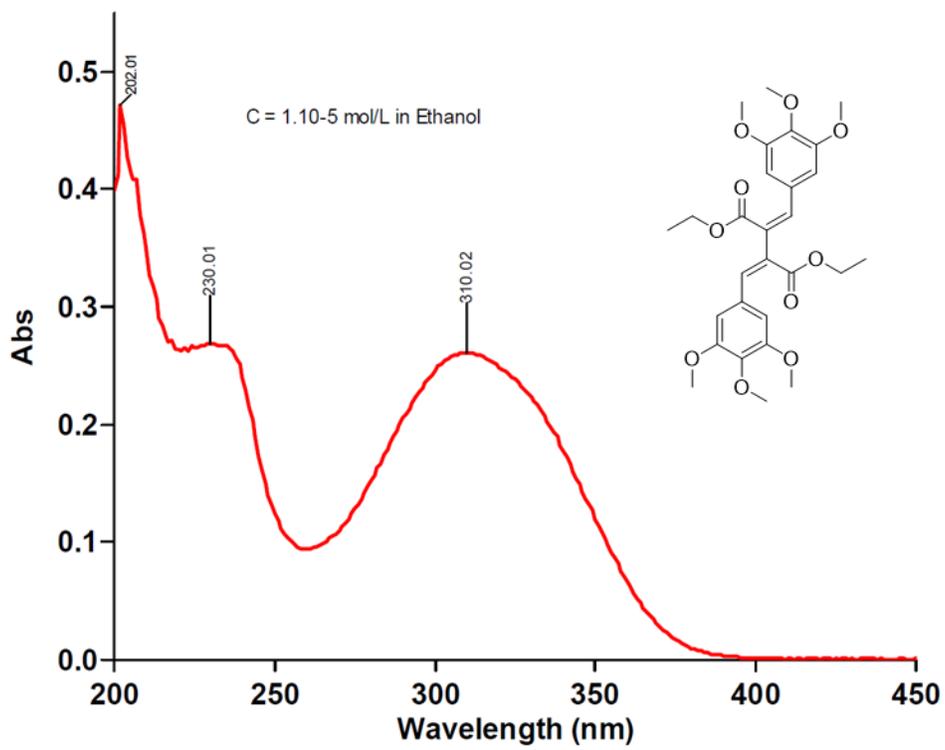
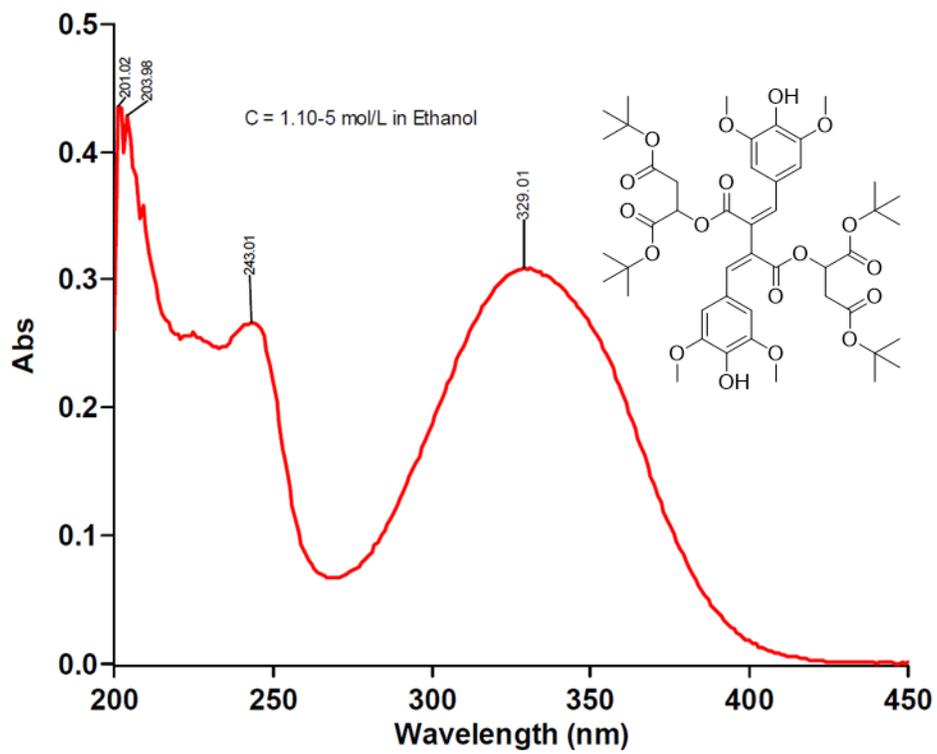


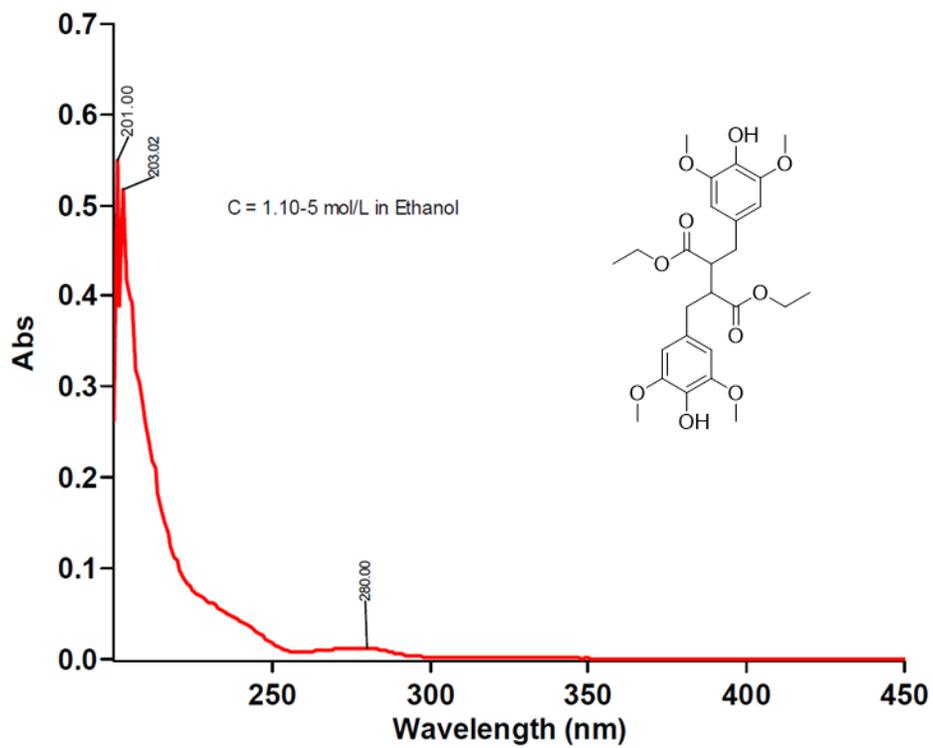
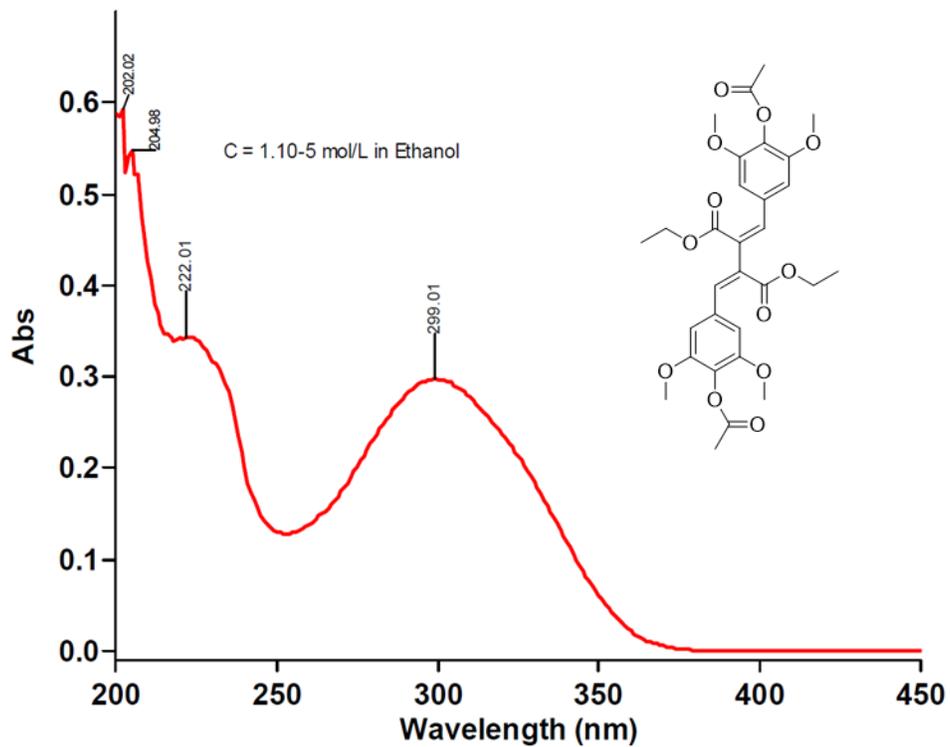


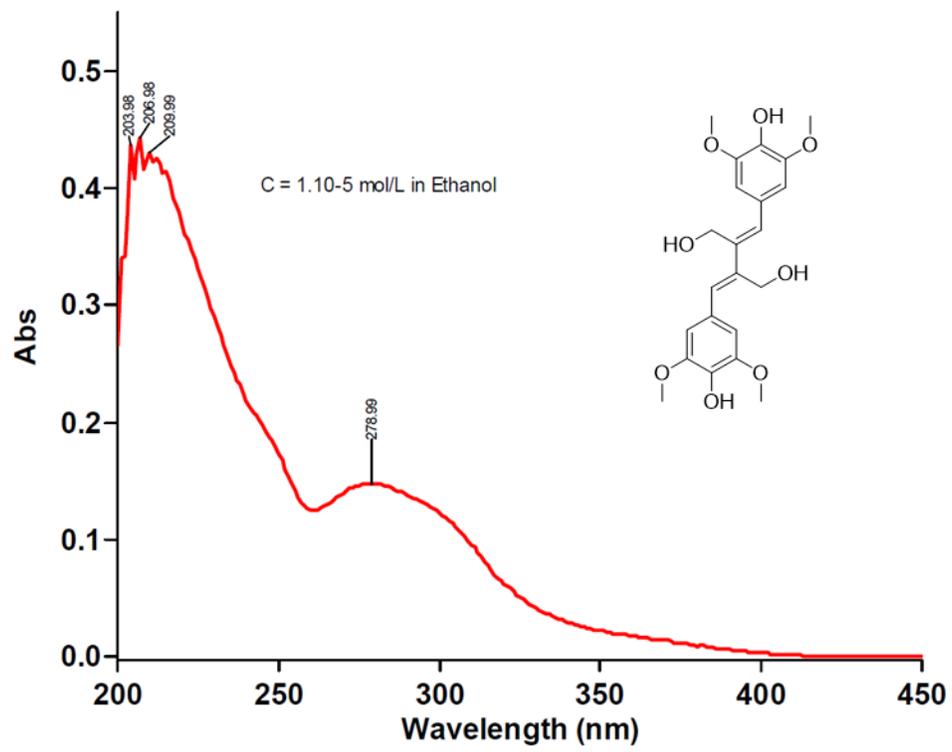




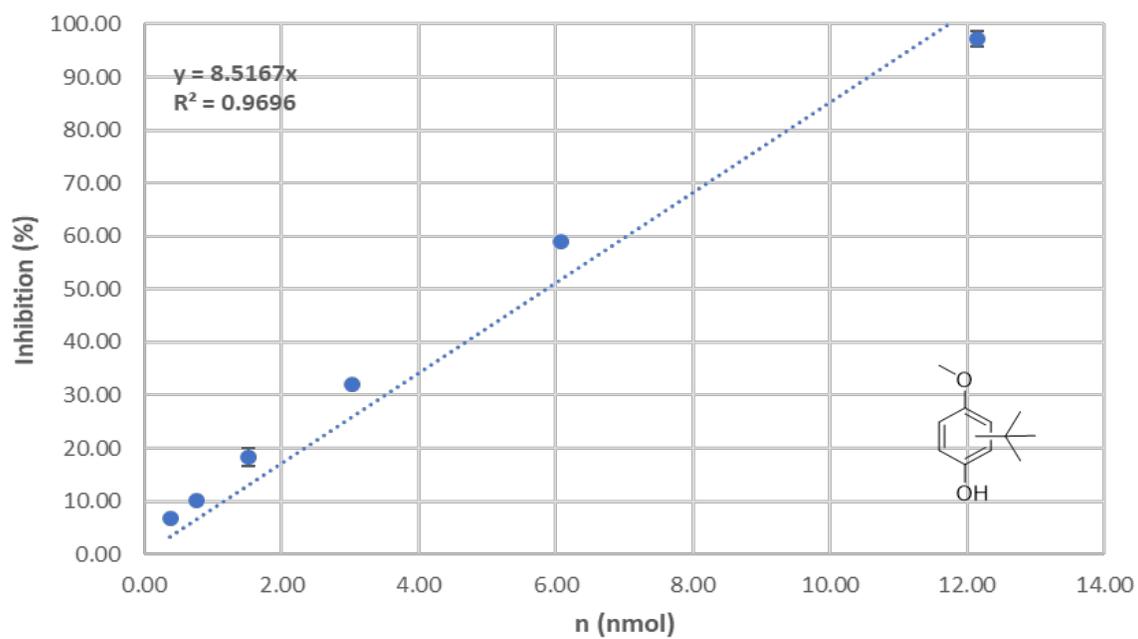
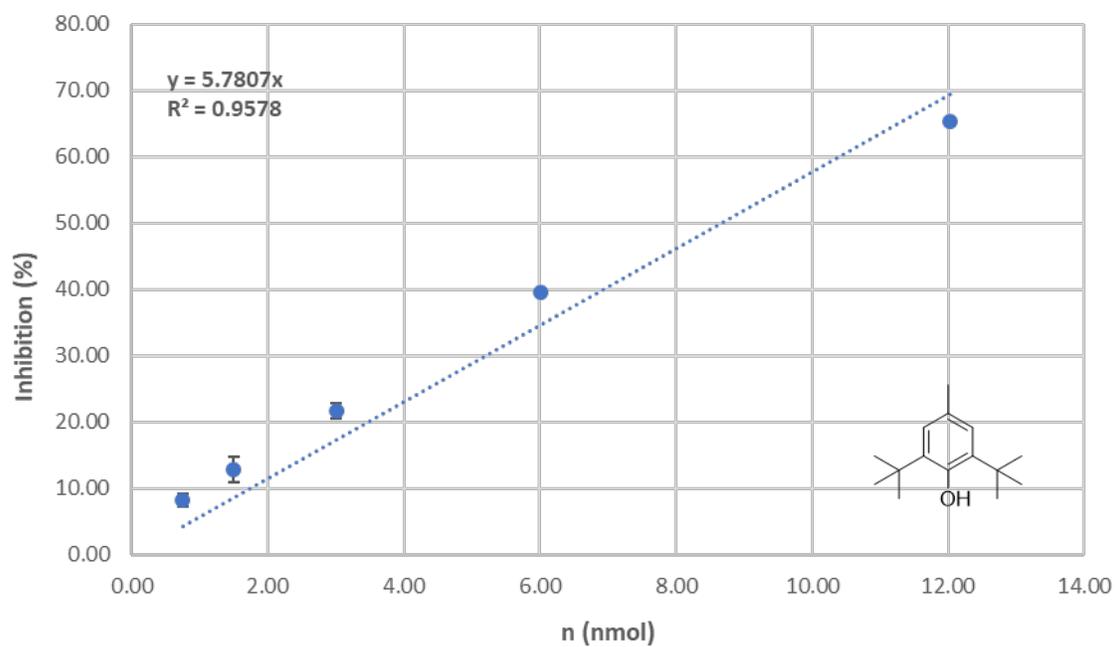


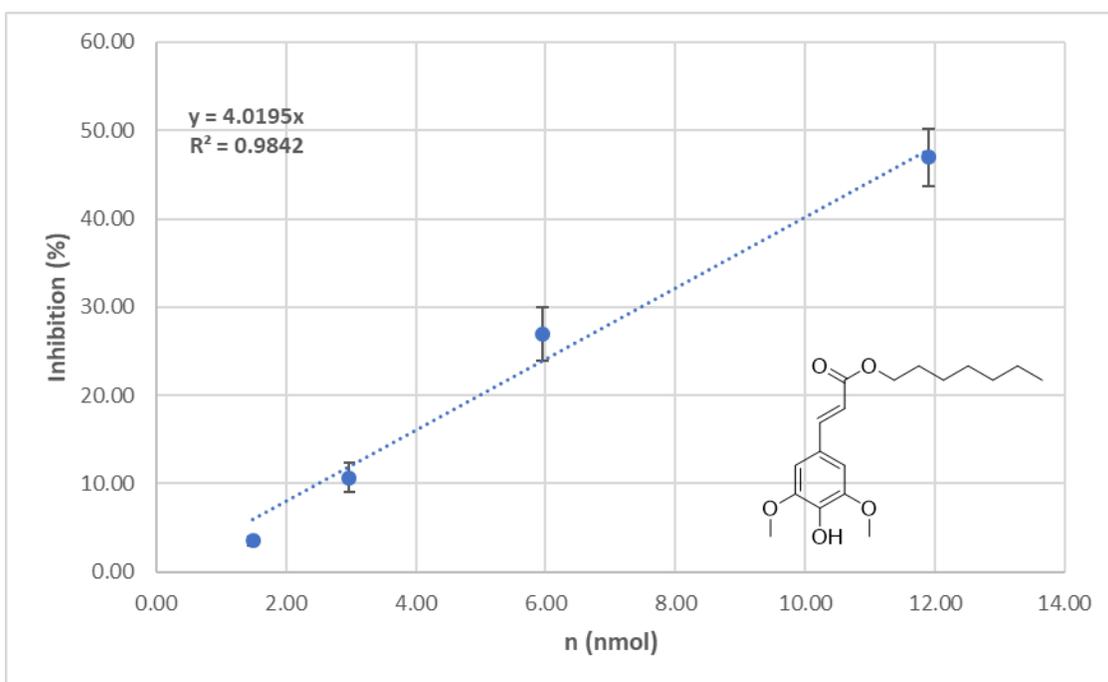
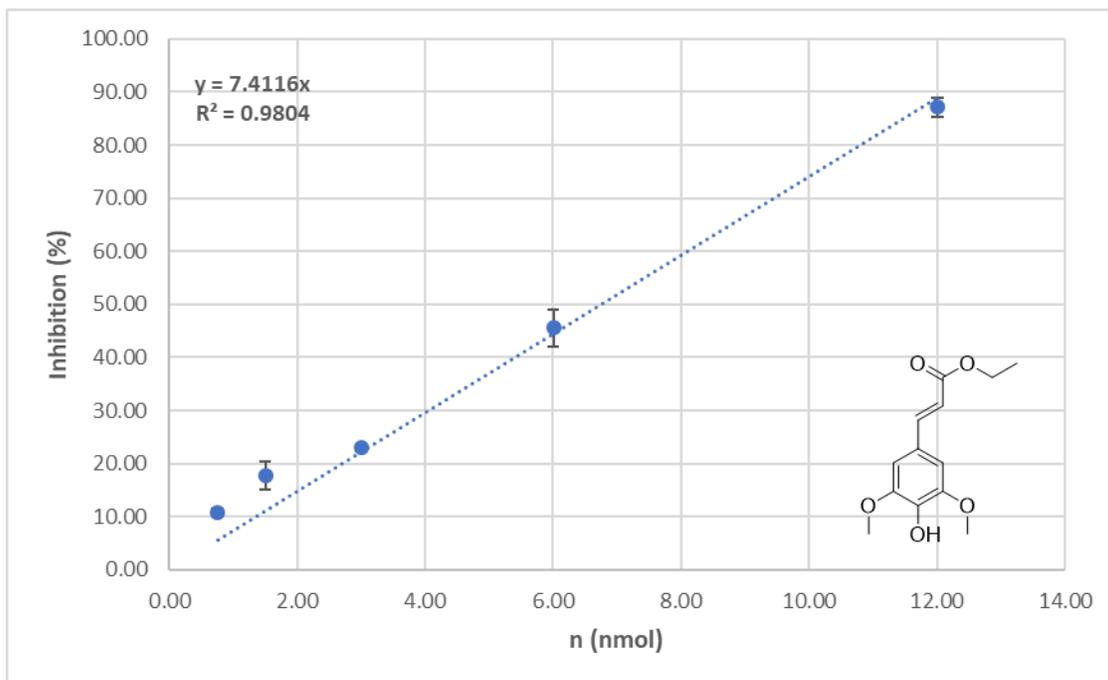


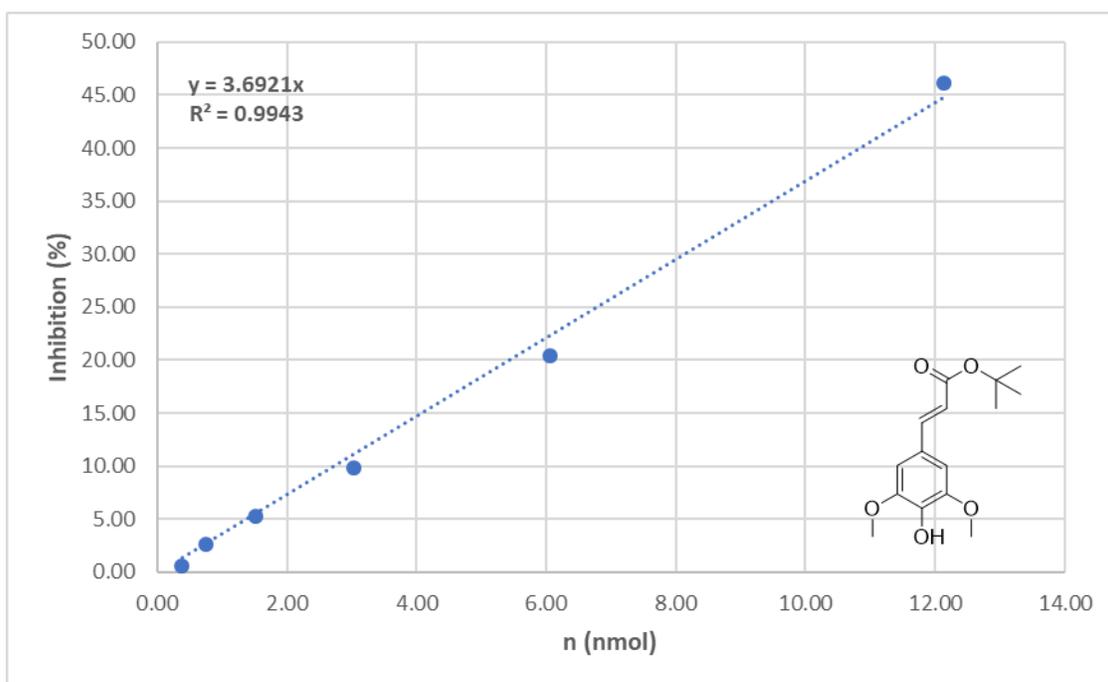
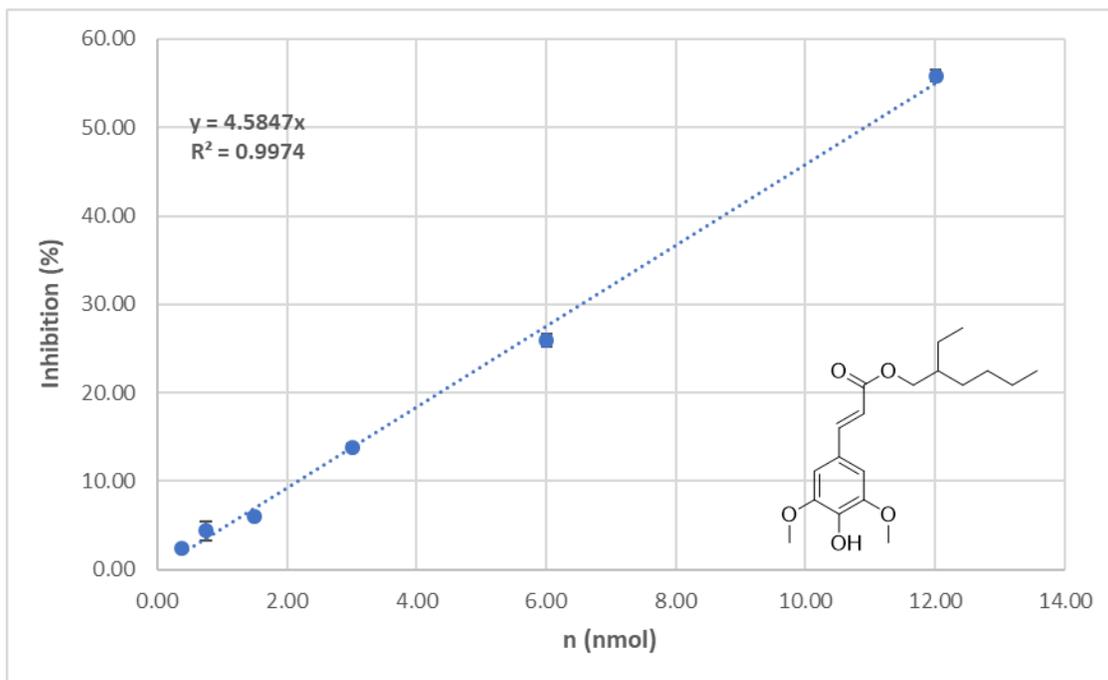


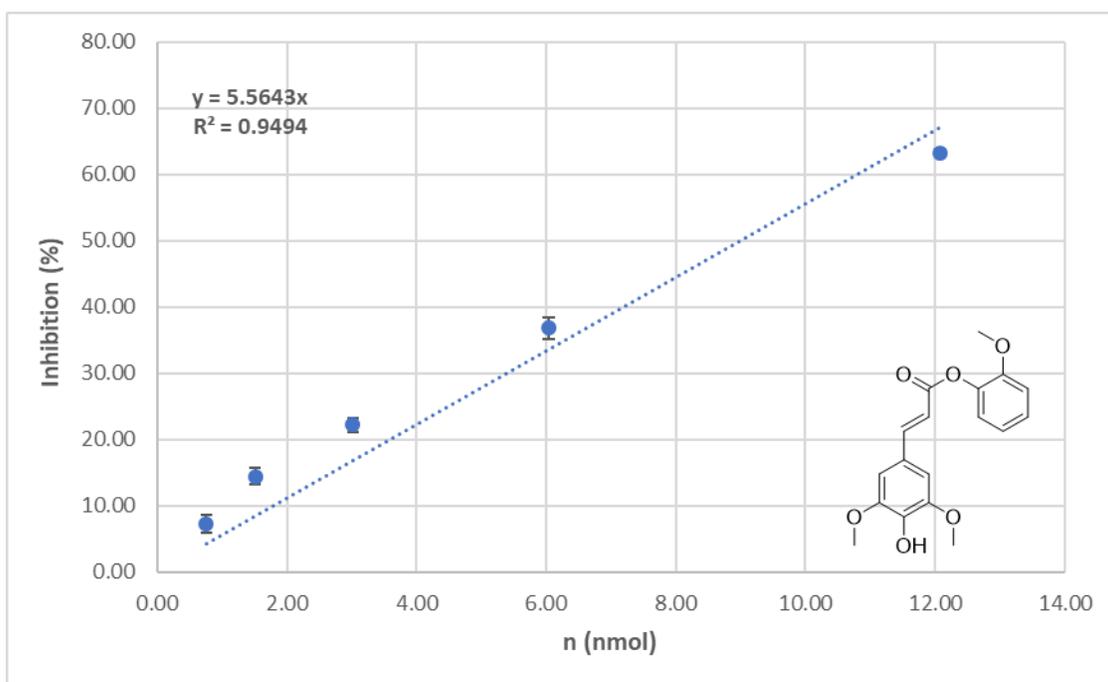
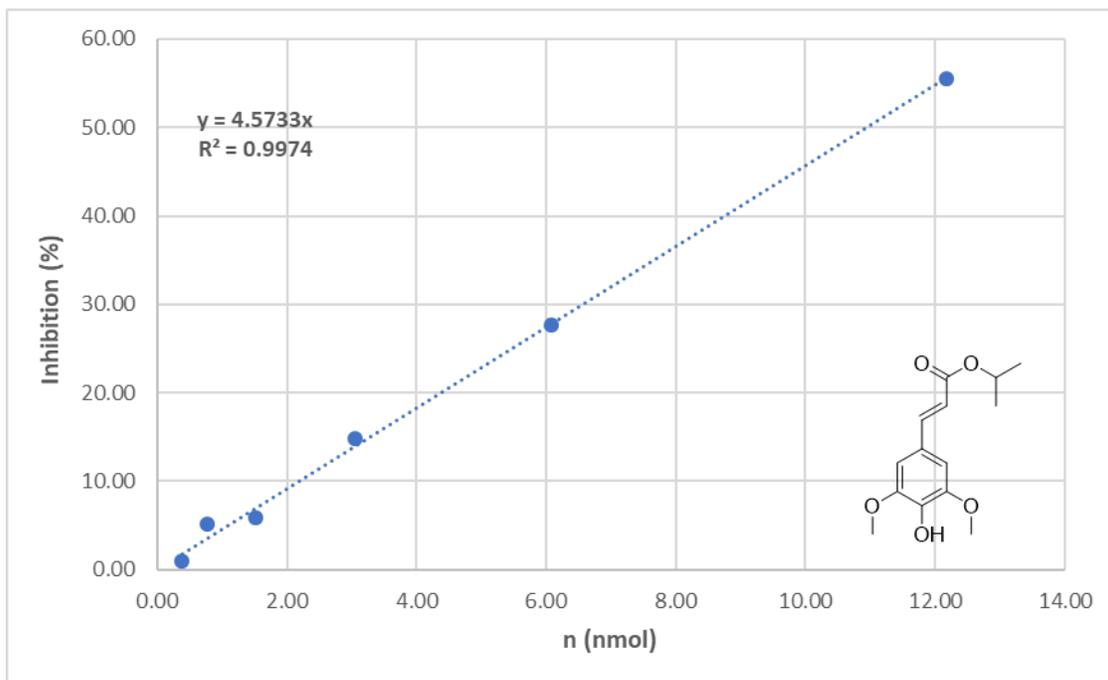


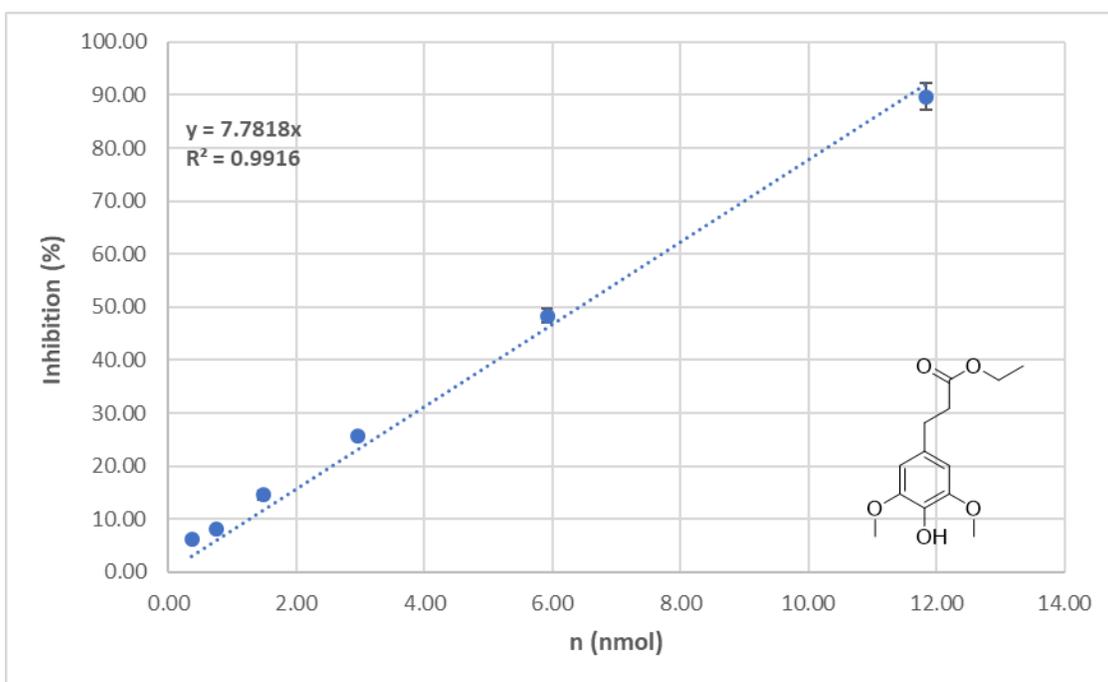
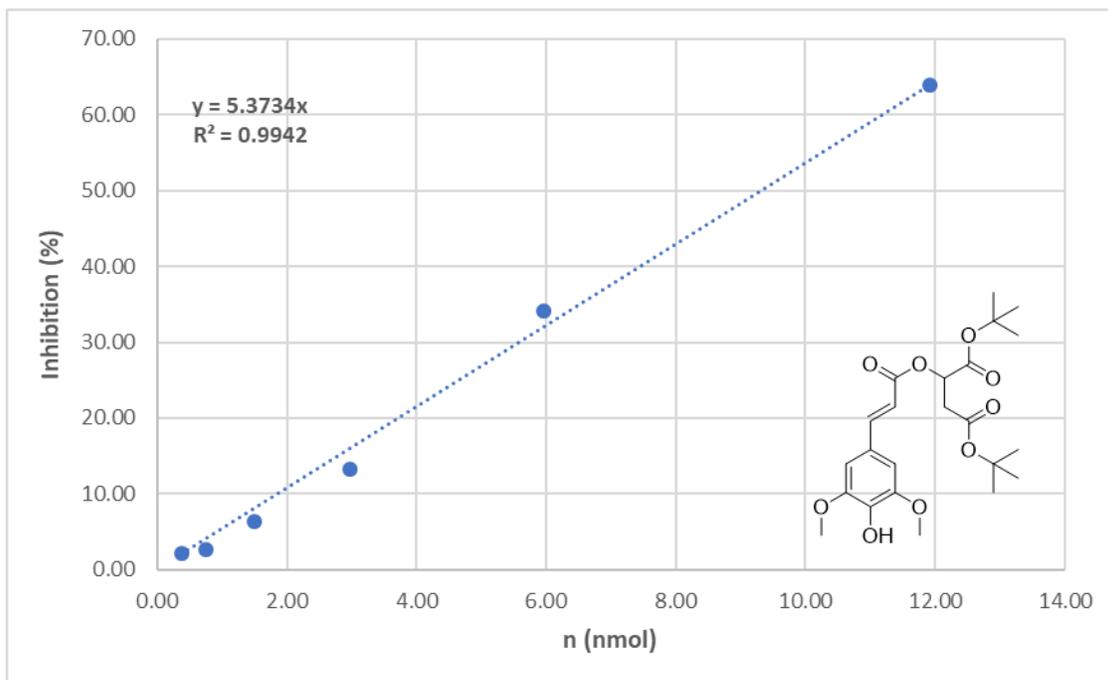
#### 7.4. ABTS inhibition

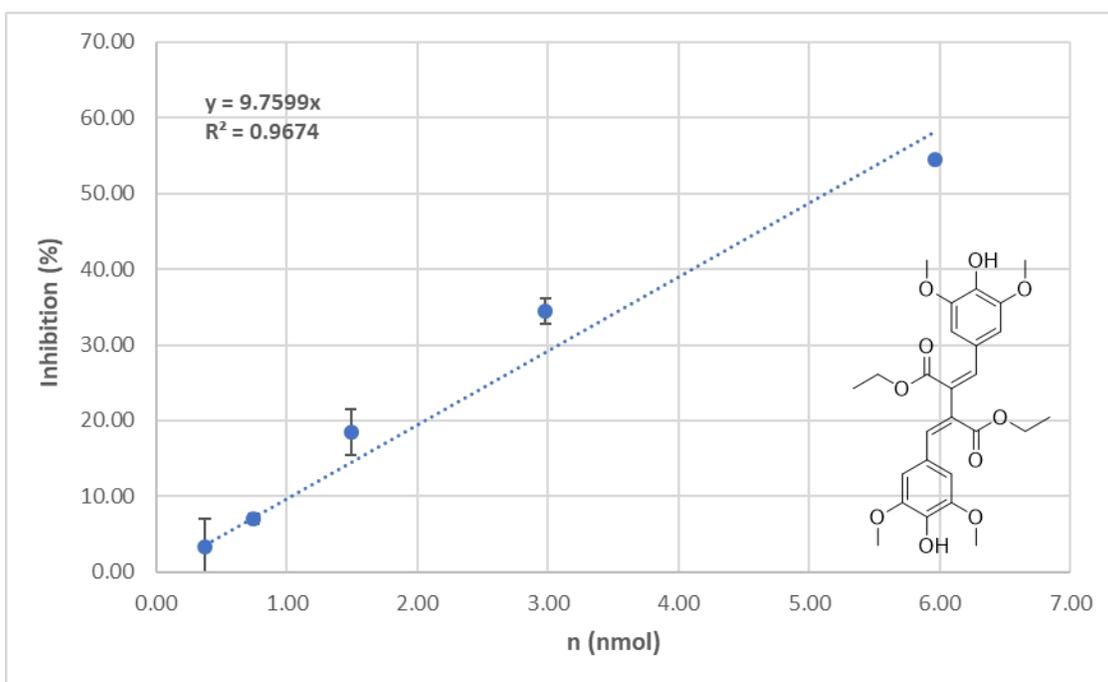
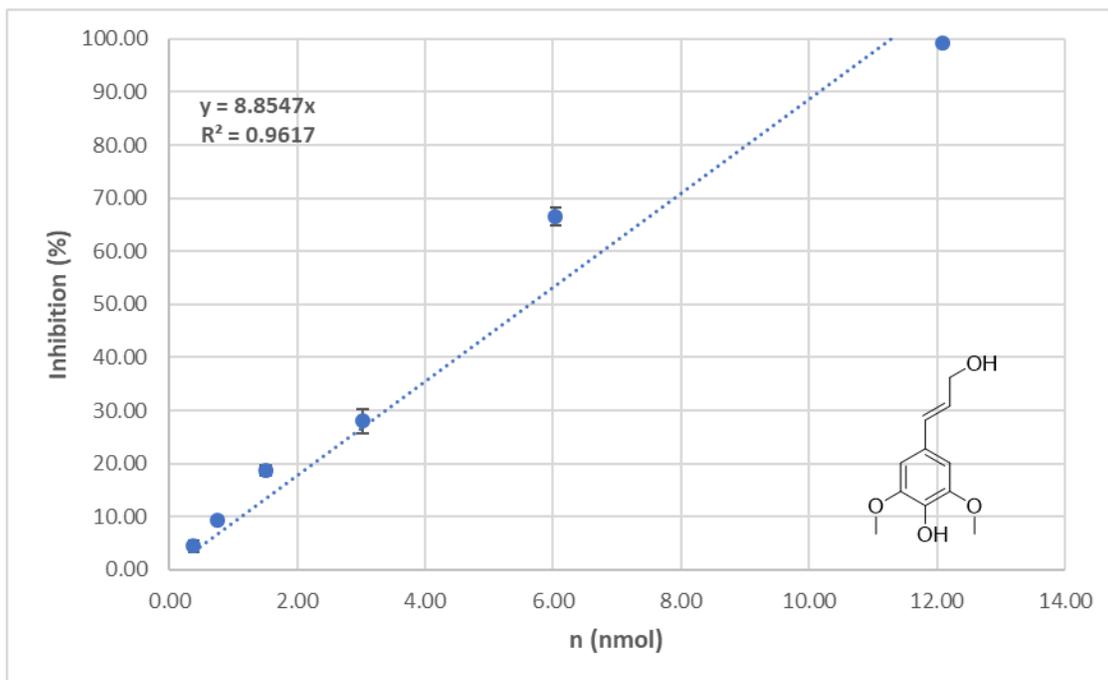


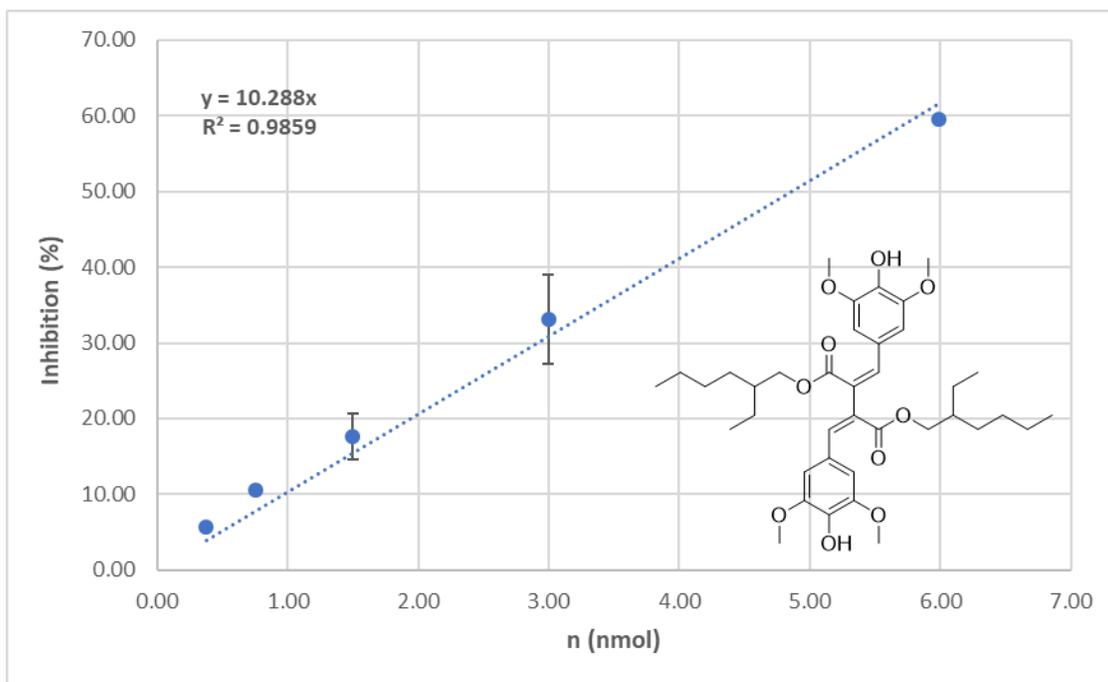
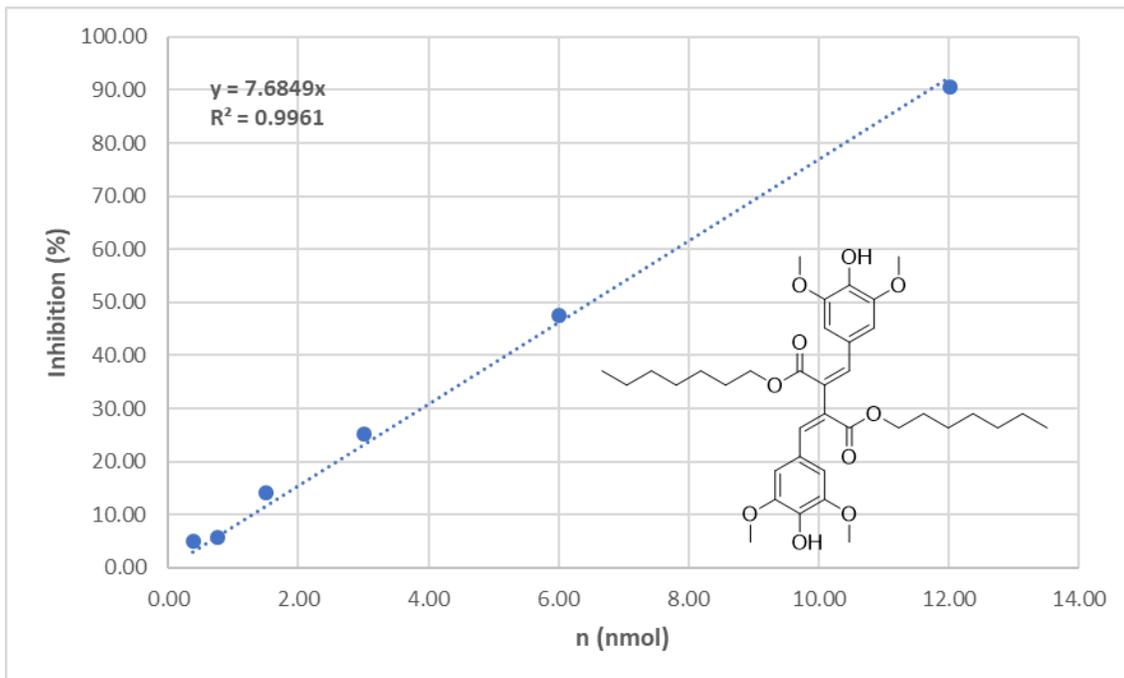


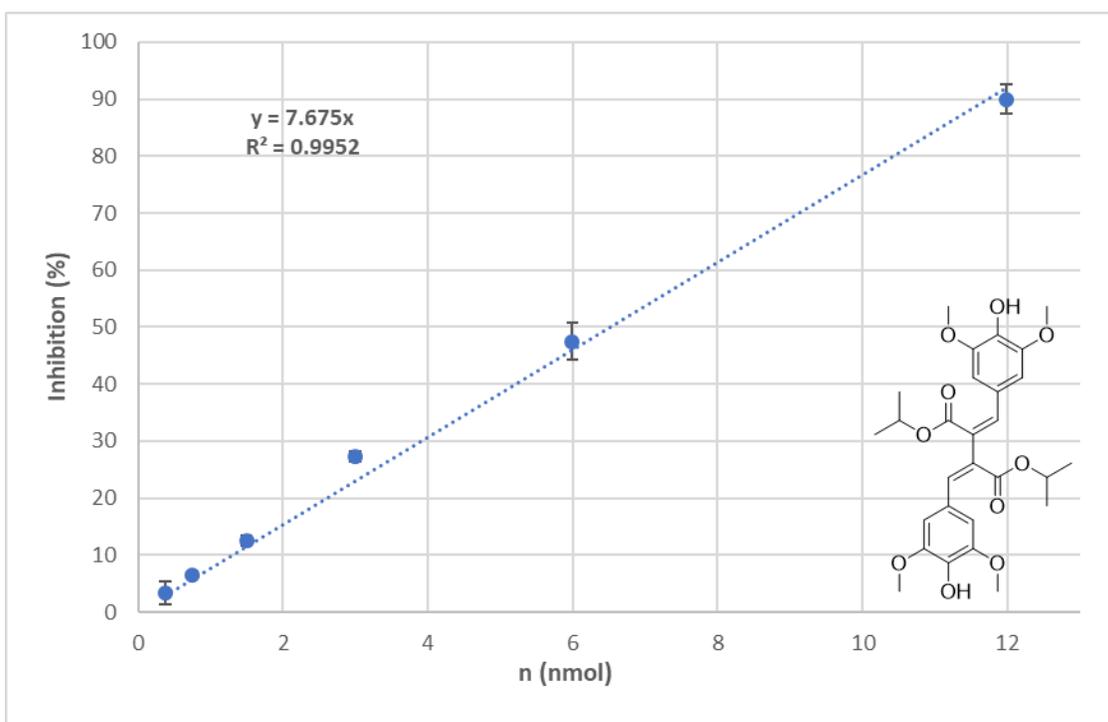
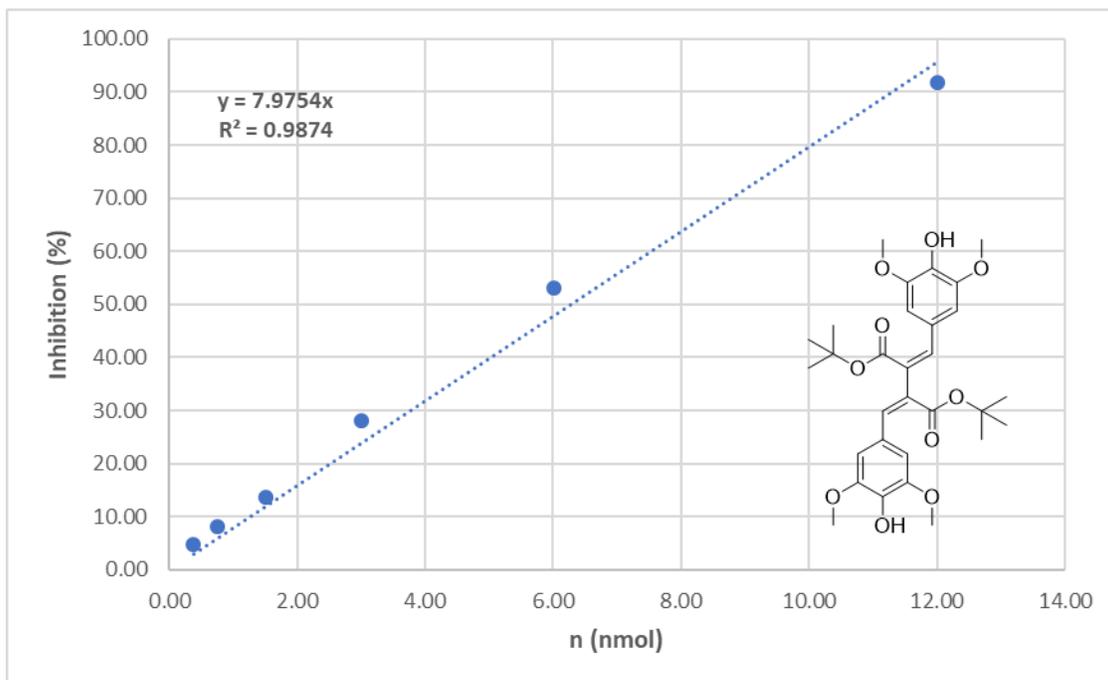




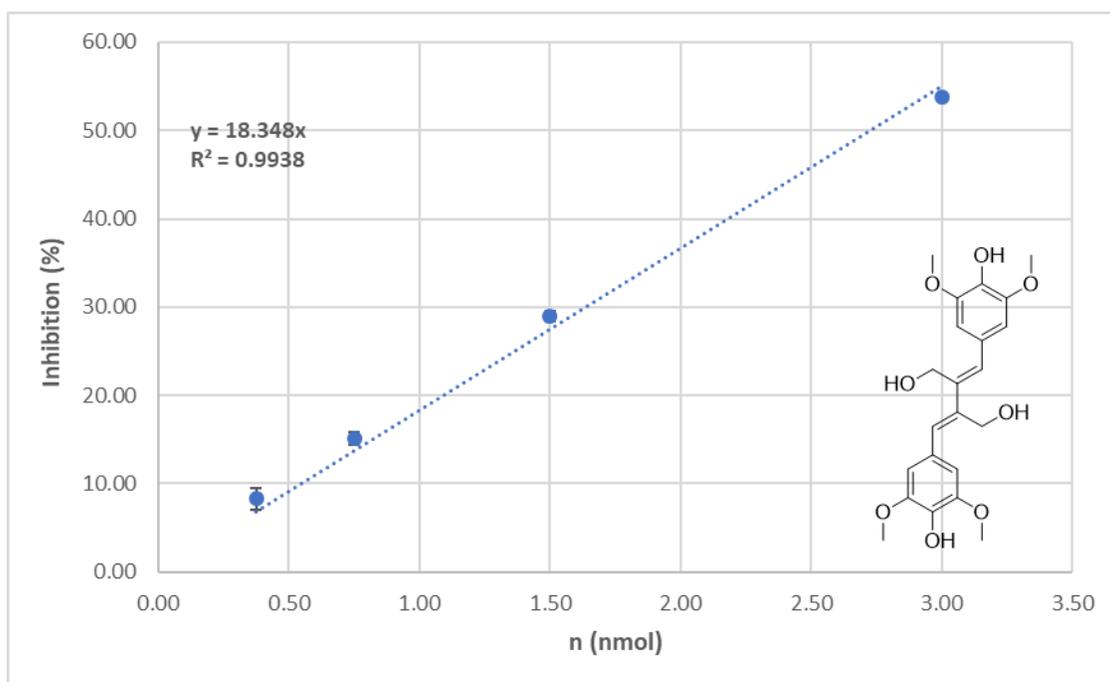
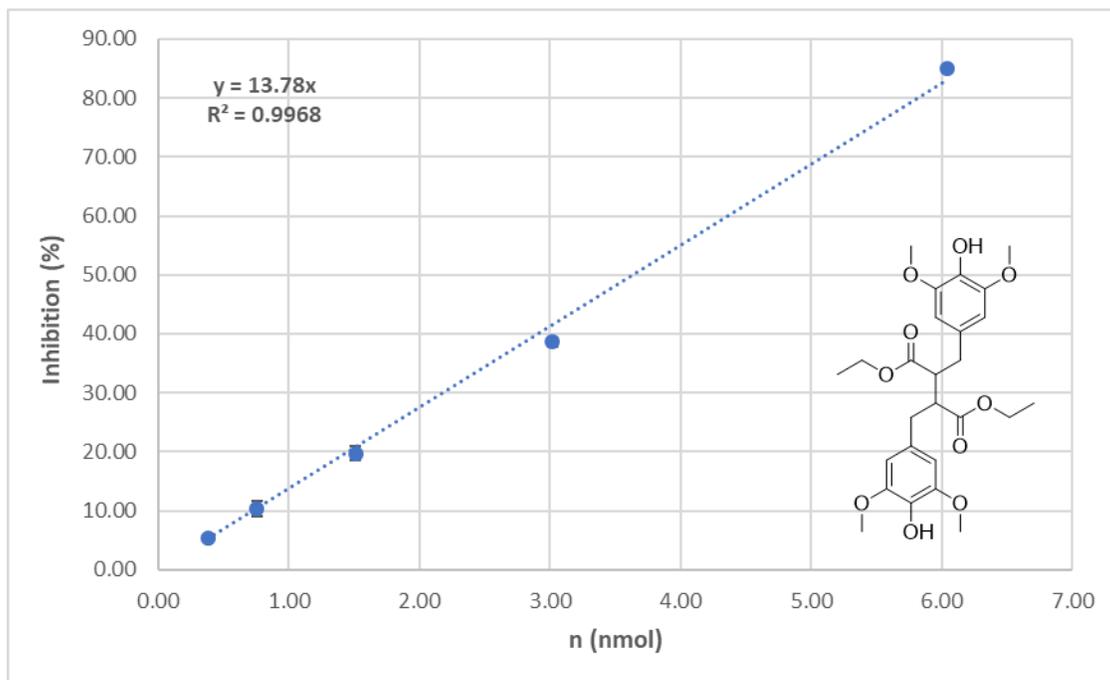












## 8. References

1. Re R., Pellegrini N., Proteggente A., Pannala A., Yang M., Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biol. Med.* 1999, **26**(9/10): 1231-1237.
2. Menezes J. C. J. M. D. S., Kamat S. P., Cavaleiro J. A. S., Gaspar A., Garrido J., Borges F. Synthesis and antioxidant activity of long chain alkyl hydroxycinnamates. *Eur. J. Med. Chem.* 2011, **46**(2): 773-777.
3. Padwal J., Lewis W., Moody C. J. Synthesis of the reported structure of crassiflorone, a naturally occurring quinone isolated from the African ebony *Diospyros crassiflora*, and regioisomeric pentacyclic furocoumarin naphthoquinones. *Org. Biomol. Chem.* 2011, **9**(9): 3484-3493.
4. Jaufurally A. S., Teixeira A. R. S., Hollande L., Allais F., Ducrot P.-H. Optimization of the laccase-catalyzed synthesis of (±)-syringaresinol and study of its thermal and antiradical activities. *ChemistrySelect* 2016, **1**(16): 5165-5171.
5. Allais F., Martinet S., Ducrot P.-H. Straightforward total synthesis of 2-O-feruloyl-L-malate, 2-O-sinapoyl-L-malate and 2-O-5-hydroxyferuloyl-L-malate. *Synthesis* 2009(21): 3571-3578.
6. Kanemitsu T., Furukoshi S., Miyazaki M., Nagata K., Itoh T. Application of asymmetric alkylation of malonic diester with phase-transfer catalysis: synthesis of LFA-1 antagonist BIRT-377. *Tetrahedron: Asymmetry* 2015, **26**(4): 214-218.
7. Page P. C. B., Moore J. P. G., Mansfield I., McKenzie M. J., Bowler W. B., Gallagher J. A. Synthesis of bone-targeted oestrogenic compounds for the inhibition of bone resorption. *Tetrahedron* 2001, **57**(9): 1837-1847.
8. Padilha G., Birmann P. T., Domingues M., Kaufman T. S., Savegnago L., Silveira C. C. Convenient Michael addition/β-elimination approach to the synthesis of 4-benzyl- and 4-aryl-selenyl coumarins using diselenides as selenium sources. *Tetrahedron Lett.* 2017, **58**(10): 985-990.
9. Hayat S., Atta ur R., Choudhary M. I., Khan K. M., Abbaskhan A. Two new cinnamic acid esters from marine brown alga *Spatoglossum variabile*. *Chem. Pharm. Bull.* 2002, **50**(9): 1297-1299.