

## Electrochemical N-demethylation of tropane alkaloids

Ali Alipour Najmi <sup>a</sup>, Zhangping Xiao <sup>b</sup>, Rainer Bischoff <sup>a</sup>, Frank J. Dekker <sup>b</sup>, Hjalmar P. Permentier <sup>a\*</sup>

<sup>a</sup> Department of Analytical Biochemistry, Groningen Research Institute of Pharmacy, University of Groningen, A Deusinglaan 1, 9713 AV Groningen, The Netherlands

<sup>b</sup> Chemical and Pharmaceutical Biology, Groningen Research Institute of Pharmacy (GRIP), University of Groningen, Groningen, the Netherlands

### Supporting Information

Introduction.....	2
Experimental section.....	3
LC-MS chromatograms.....	6
<sup>1</sup> H, <sup>13</sup> C NMR and HRMS data.....	10
<sup>1</sup> H and <sup>13</sup> C NMR spectra.....	20

## Introduction

**Table S1 Conditions and advantages/disadvantages of different catalytic methods for the N-demethylation of tropane alkaloids (References order is based on the main manuscript).**

Ref.	Catalyst	Temp.	Solvent	Yield %	Oxidant	Time	Disadvantages	Advantages
[9]	NA	RT	Chloroform	90.5 (noratropine)	2,2,2-Trichloroethyl chloroformate 5 equiv.	20 h	1) A carbamate intermediate is produced which needs to be cleaved. 2) Very toxic oxidant. 3) Toxic solvent. 4) Long time. 5) Requires 8.8 equiv. of Zn dust	1) High yield
[10]	NA	RT	Chloroform	100 (noratropine)	$\alpha$ -chloroethyl chloroformate 5 equiv.	4 h	1) A carbamate intermediate is produced which needs to be cleaved. 2) Very toxic oxidant. 3) Toxic solvent. 4) Requires 10.2 equiv. of sodium hydrogen carbonate	1) Excellent yield
[10]	NA	30	Water (0.5 M sulfuric acid)	79.8 (norscopolamine)	KMnO <sub>4</sub> 2.5 equiv.	1 h	1) Toxic and hazardous oxidant. 2) Requires 2.5 equiv. of oxidants	1) Only water as solvent
[11]	NA	RT	Water	16.8 (noratropine)	KMnO <sub>4</sub> 2 equiv.	10 min	1) Toxic oxidant, 2) Very low yield	1) Quick reaction. 2) Only water as solvent
[12]	Light, Bengal rose, TPP (Photochemistry)	RT	Dichloromethane	66 (noratropine)	Photochemical reaction, oxygen and light	6 h	1) Toxic solvent. 2) Complex reaction mixture	1) No toxic oxidants. 2) Medium yield.
[15]	FeSO <sub>4</sub> ·7H <sub>2</sub> O 1.5 equiv.	RT	Methanol	51 (noratropine)	Magnesium bis (monoperoxyphthalate) hexahydrate 1.1 equiv.	45 min	1) N-oxide formation. 2) HCl isolation. 3) TPPS for iron chelating/separation. 4) Toxic solvent. 5) Low yield	1) No toxic oxidants
[17]	Fe(II)TPPS	-80-0 °C	Chloroform or methanol	42 (noratropine)	H <sub>2</sub> O <sub>2</sub> or m-CPBA	NA	1) N-oxide formation. 2) Hazardous oxidant, 3) HCl salt isolation, 4) Catalyst separation, 5) Low temperature. 6) Toxic solvent. 7) Low yield	1) Stabilizing the catalyst by acetate buffer
[8]	Ferrocene 0.1 equiv.	80 °C	Chloroform and isopropanol	60 (noratropine)	H <sub>2</sub> O <sub>2</sub> or m-CPBA	24 h	1) Toxic catalyst. 2) Hazardous oxidants. 3) Low yield. 4) Toxic solvents in part of the process. 5) Catalyst separation	1) Application of green solvent in part of the process
[2]	Iron powder 1 equiv.	RT	Chloroform	81 (noratropine)	H <sub>2</sub> O <sub>2</sub> or m-CPBA	2 h	1) N-oxide formation 2) HCl salt isolation. 3) hazardous oxidants. 4) toxic solvents. 5) catalyst separation	1) High yield
[18] Green Chemistry	Iron nanoparticles (FeSO <sub>4</sub> and NaBH <sub>4</sub> ) 1 equiv.	RT	Chloroform and isopropanol	85 (noratropine)	m-CPBA 1.2 equiv.	3 h	1) toxic sodium borohydride for nanoparticle synthesis. 2) N-oxide formation 3) HCl salt isolation 4) hazardous oxidant. 5) Catalyst separation. 6) Toxic solvent in part of the process.	1) High yield. 2) Application of isopropanol in part of the process
[18] Green Chemistry	Fe <sub>3</sub> (CO) <sub>12</sub> 0.05 equiv.	RT	Chloroform and isopropanol/ethyl acetate	79 (noratropine)	m-CPBA 1.2 equiv.	19 h	1) Dangerous catalyst. 2) N-oxide formation. 3) HCl salt isolation. 4) Hazardous oxidants 3) Long reaction time. 4) Toxic solvent in part of the reaction	1) Medium yield. 2) Application of isopropanol in part of the reaction. 3) Low catalyst load
[3] and [6] Green Chemistry	Fe(II)-TAML 0.1 equiv.	RT	Ethanol Acetone DMSO	75 (noratropine) 77 (norscopolamine)	H <sub>2</sub> O <sub>2</sub> , 50 equiv.	1 h	1) Deactivating of H <sub>2</sub> O <sub>2</sub> by MnO <sub>2</sub> . 2) Very high equivalent of H <sub>2</sub> O <sub>2</sub>	1) Application of green solvent. 2) High yield. 3) Low catalyst load

## Experimental section

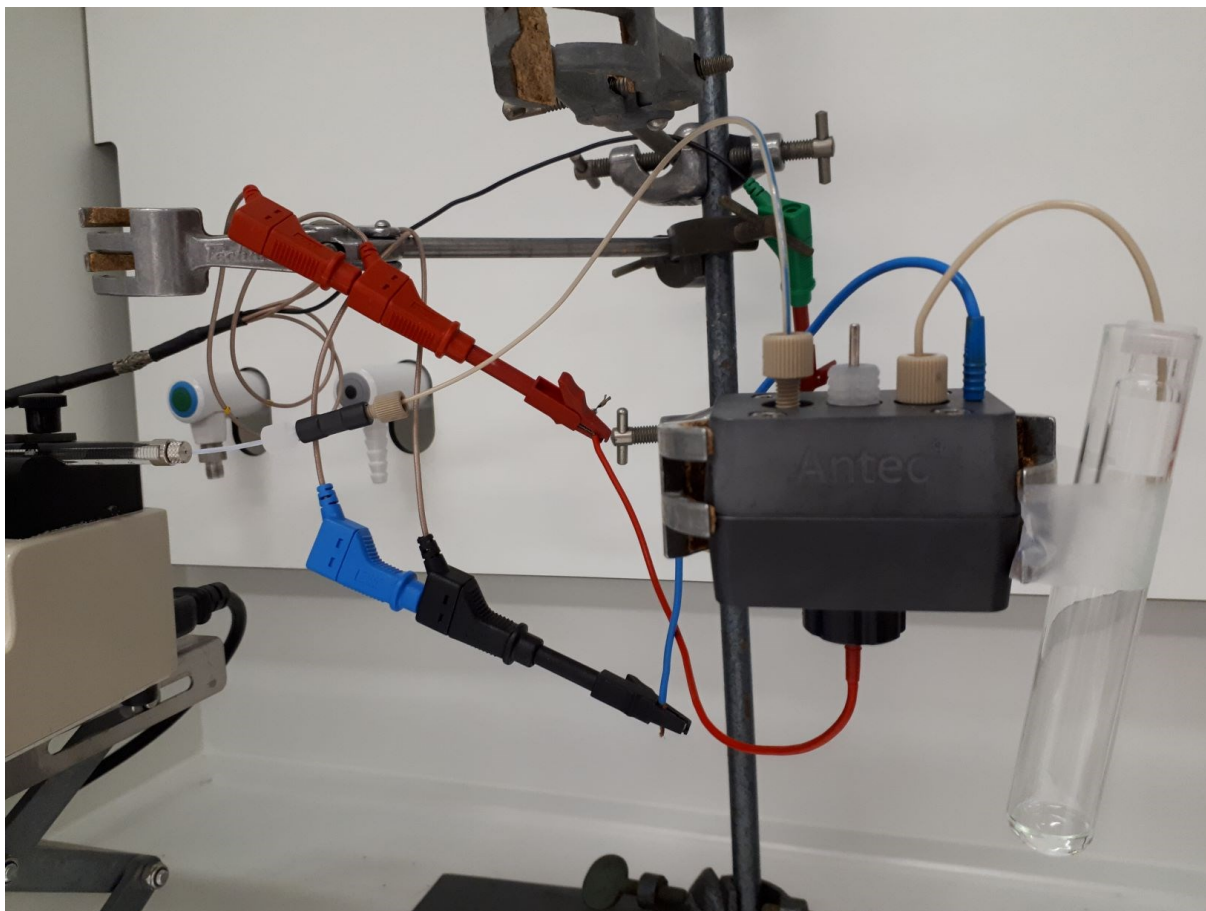


Fig S1, Set-up of the analytical electrochemical flow-cell ( $\mu$ -PrepCell, Antec Scientific)

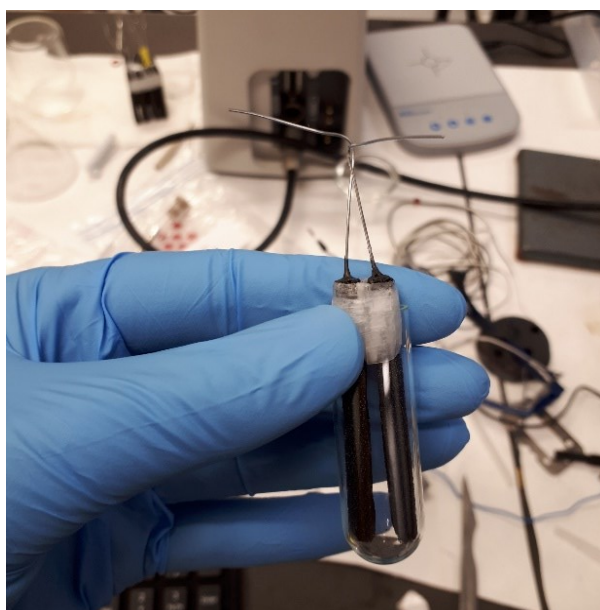
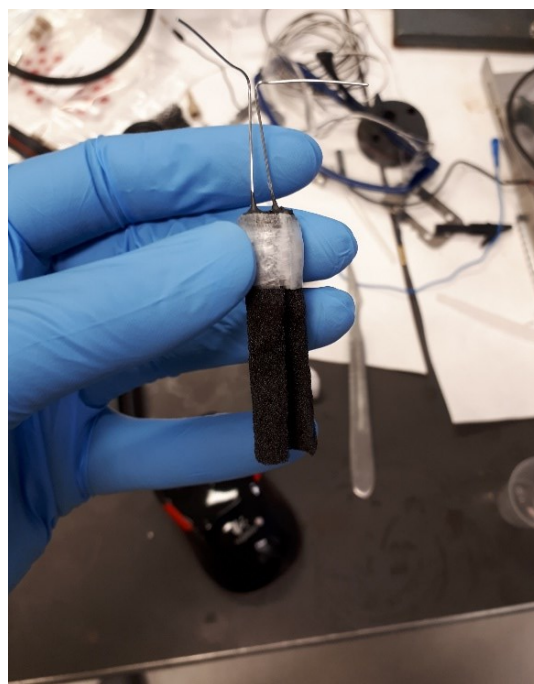


Fig S2, Set-up of the home-made electrochemical cell using a 100 PPI GC electrode, a stainless steel wire and a half-cut glass test tube

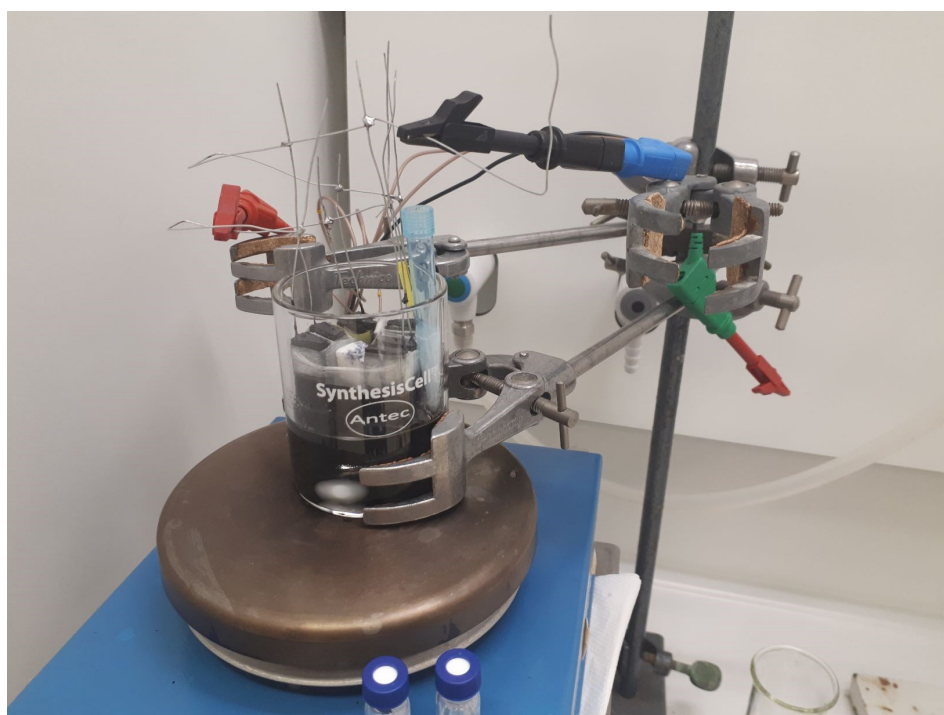
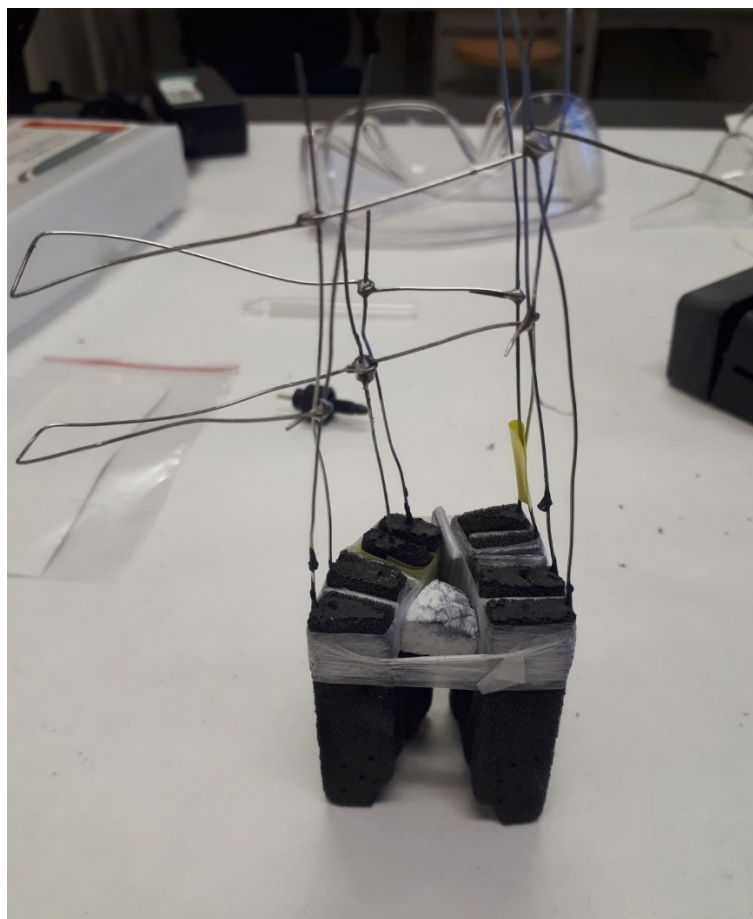


Fig S3, Home-made electrochemical cell with a stack of four paired anode-cathode electrodes for gram-scale synthesis

### LC-MS chromatograms

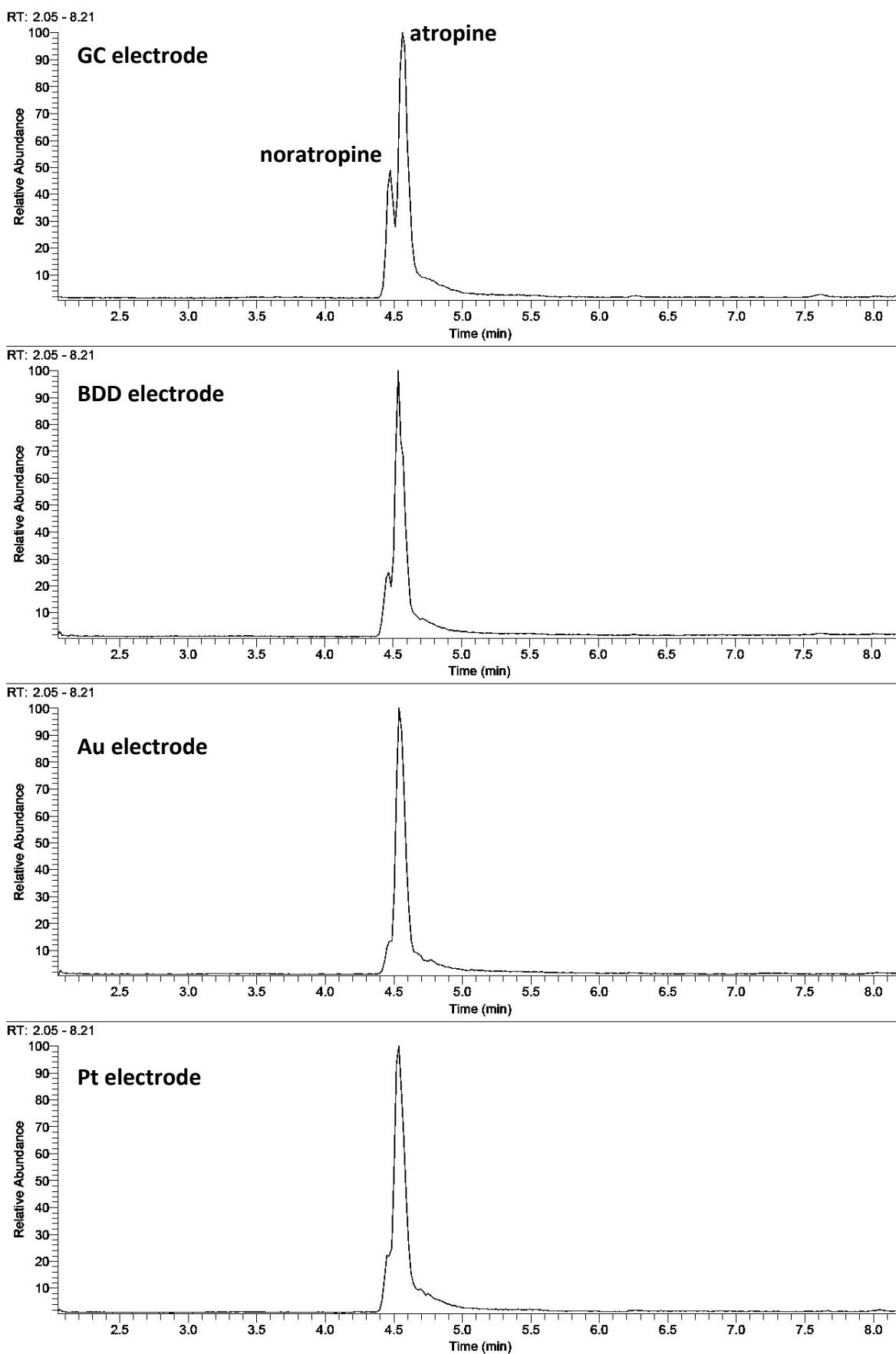


Fig S4, LC-MS chromatogram of flow-cell experiment with different electrode materials. Reaction condition: 10 mM, 5  $\mu$ L/min, 200  $\mu$ A, ethanol, NaClO<sub>4</sub> (4 equivalent).

RT: 1.92 - 9.36

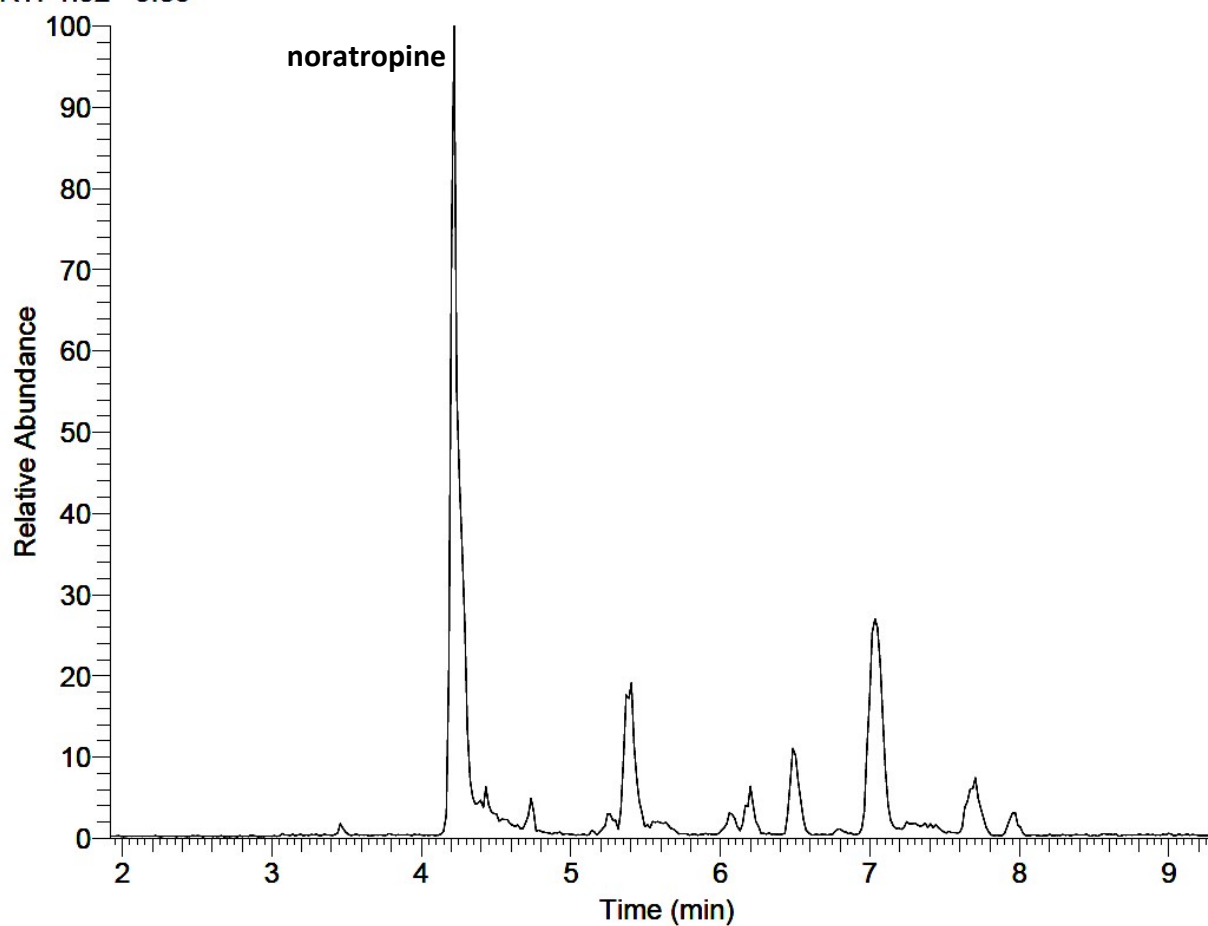


Fig S5, LC-MS chromatogram of final reaction mixture of entry 5, Table 1. 57 mM atropine, ethanol/water 2:1, 8 mA/3 h.

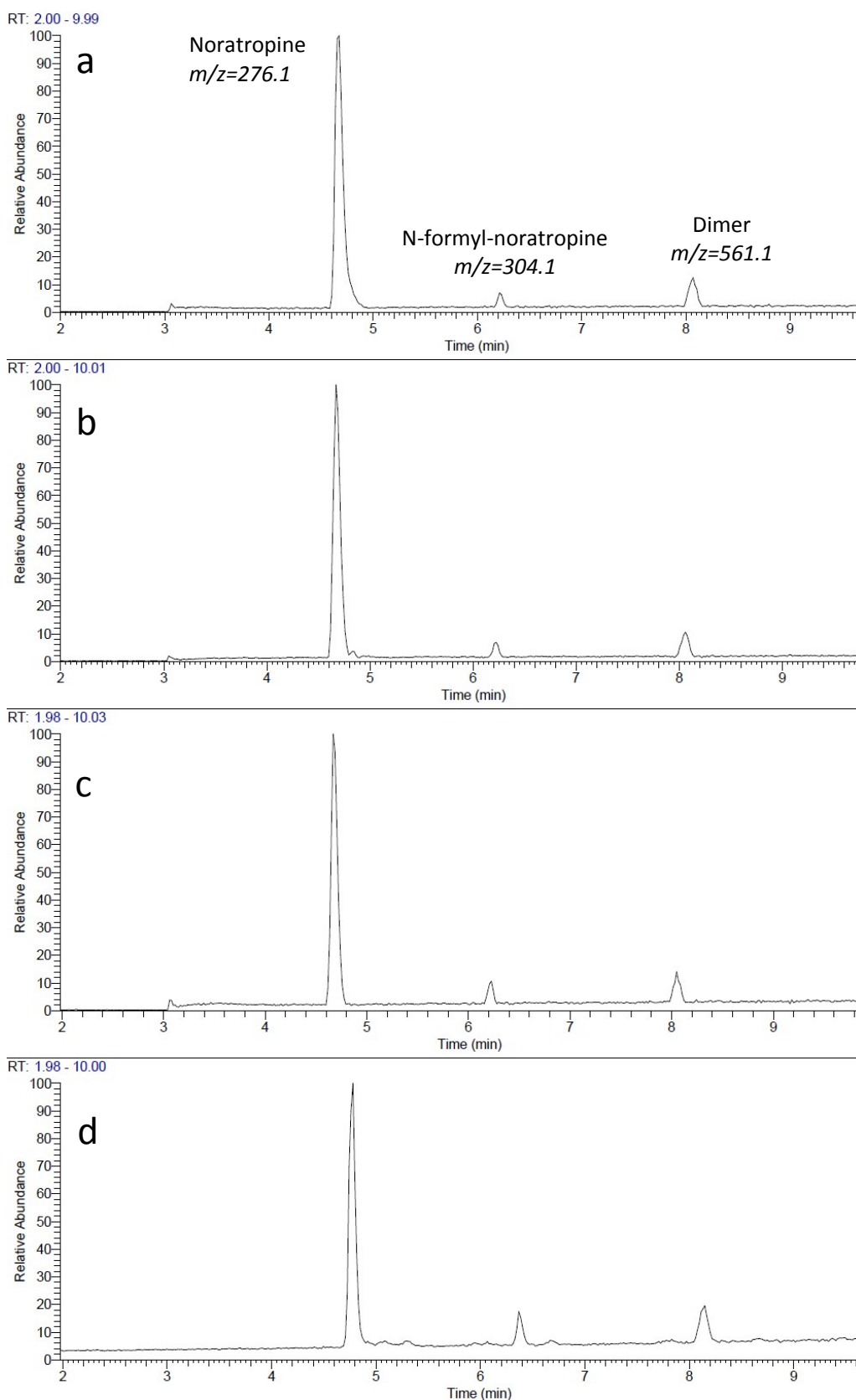


Fig S6, LC-MS chromatogram of final reaction mixture in four different conditions, entry 9-12, Table 1 (panels a-d, respectively).

57 mM atropine, ethanol/water 2:1, 4 mA/4.5 h.



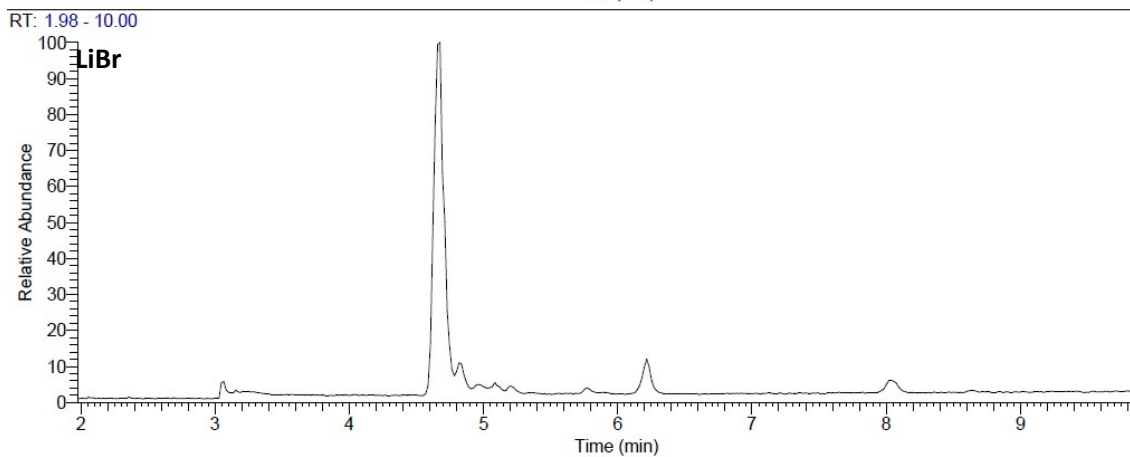
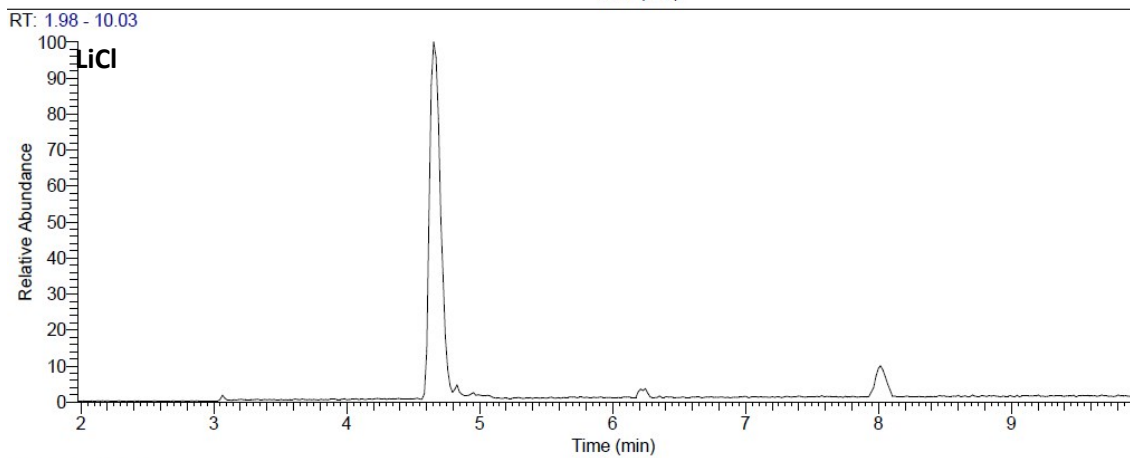
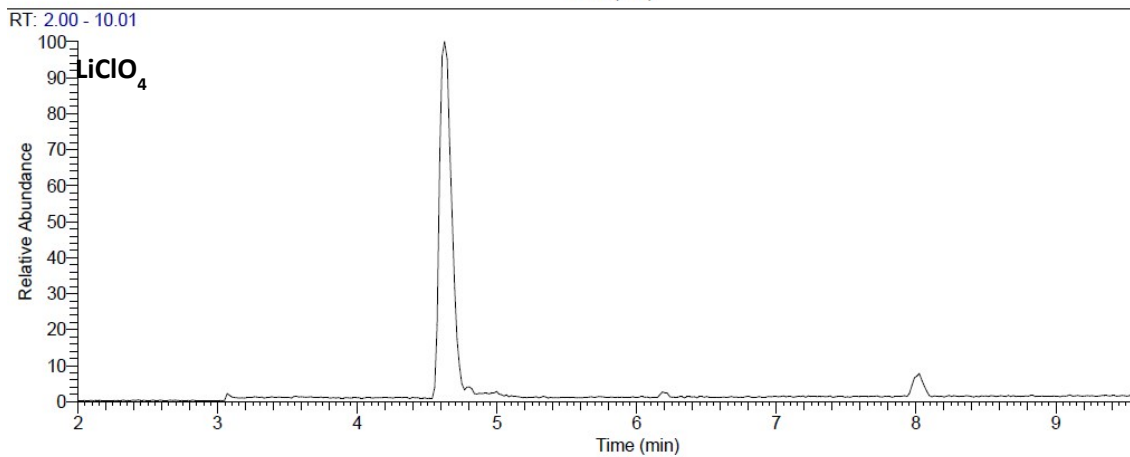
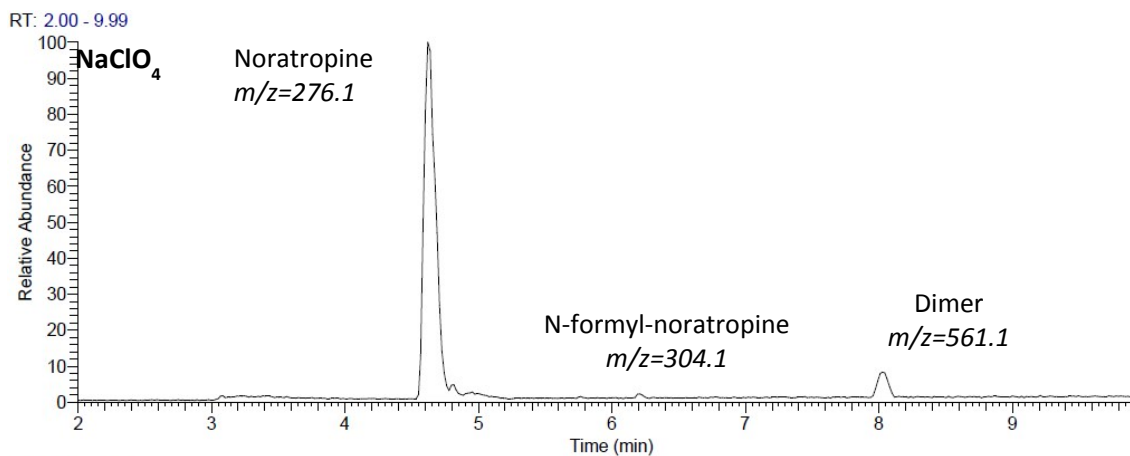


Fig S7, LC-MS chromatogram of final reaction mixture with four different supporting electrolytes, entry 1-4, Table 3. 57 mM atropine, methanol/water 2:1.

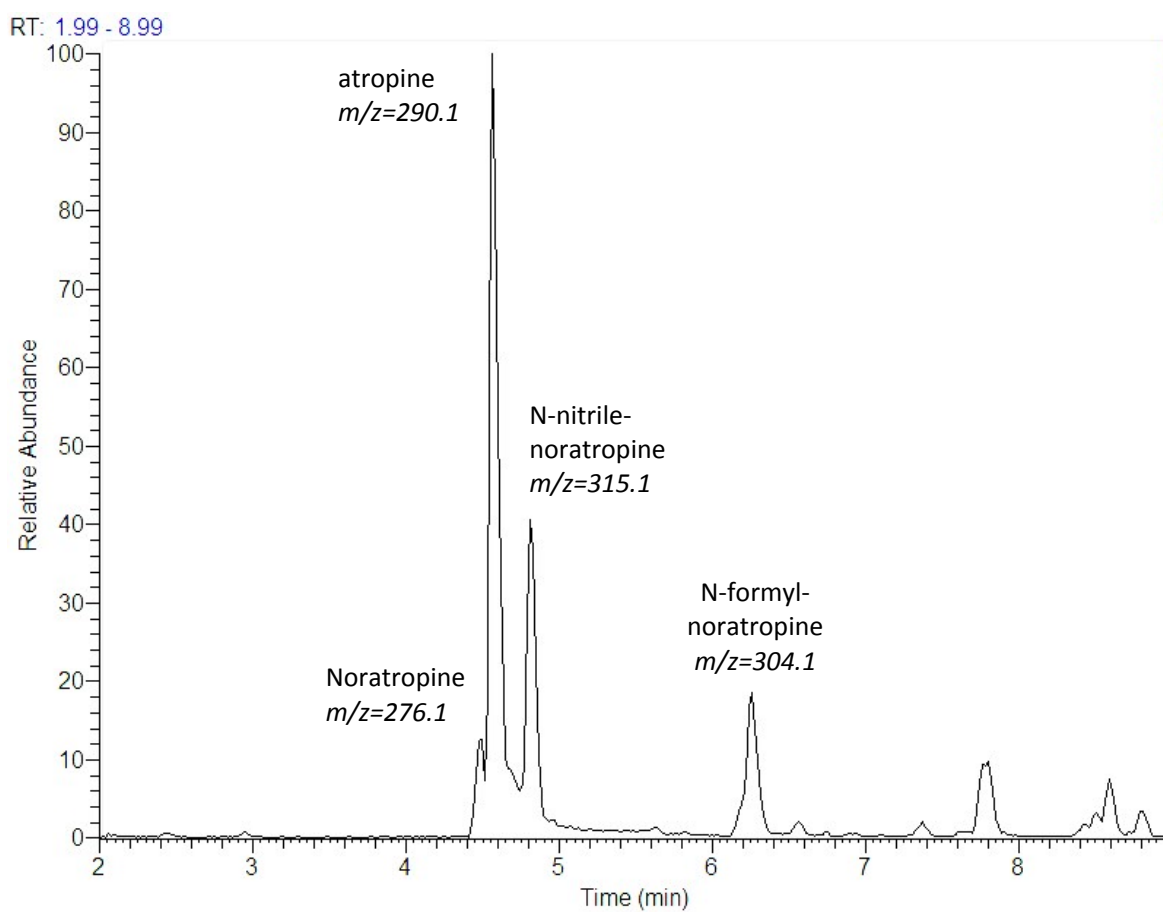


Fig S8, LC-MS chromatogram of flow-cell experiment with added KCN. Reaction condition: 10 mM atropine, 1  $\mu$ L/min, 200  $\mu$ A, ethanol/water 1:1, NaClO<sub>4</sub> (4 equivalent), KCN (4 equivalent).

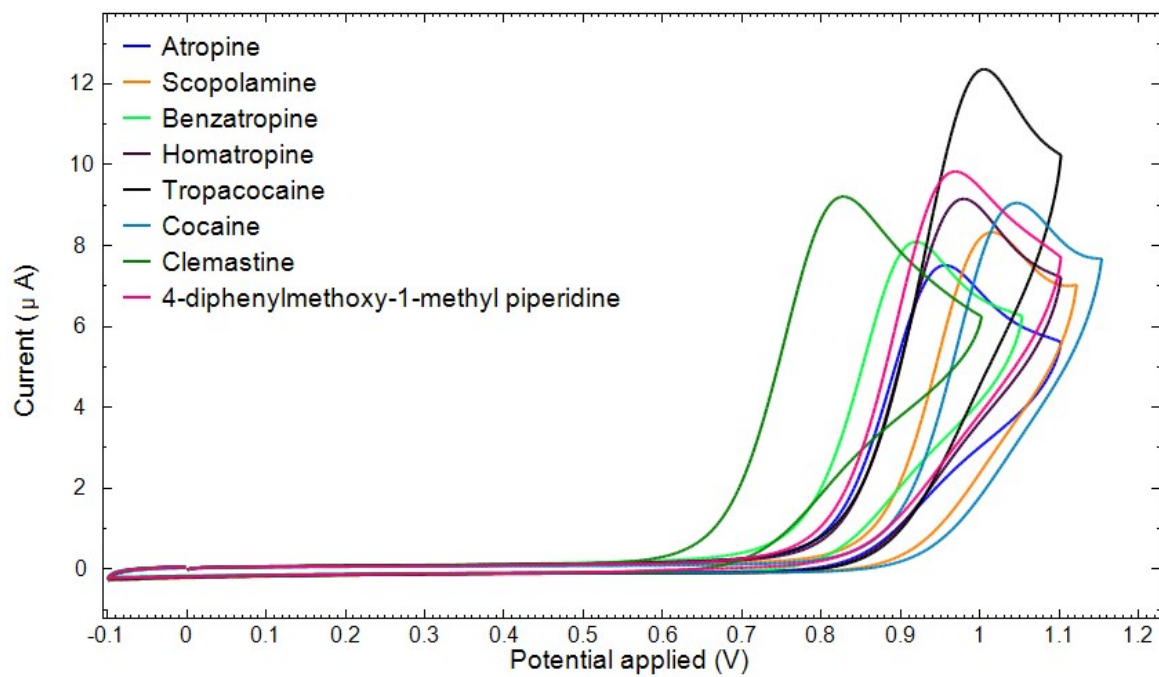


Fig S9, Cyclic voltammetry of different compounds (5 mM) in ethanol/water (2:1) with  $\text{NaClO}_4$  (0.1 M) as supporting electrolyte, glassy carbon both as working and counter electrodes, scan rate = 100 mV/s.

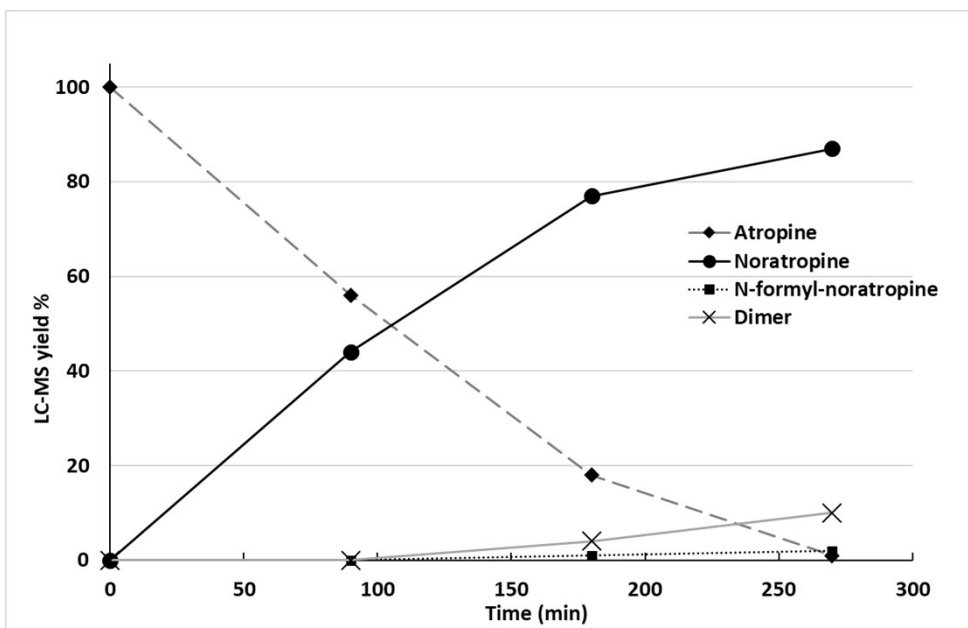


Fig S10, Product distribution during the reaction period for the electrochemical N-demethylation of atropine (Table 2, entry 10).

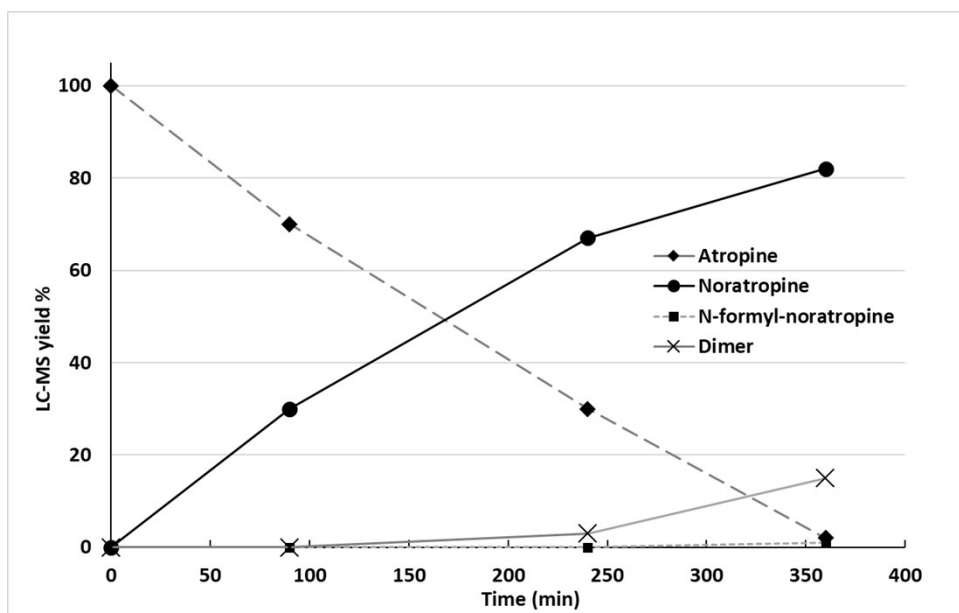


Fig S11, Product distribution during the reaction period for the electrochemical N-demethylation of atropine in gram-scale (Table 4, entry 2).

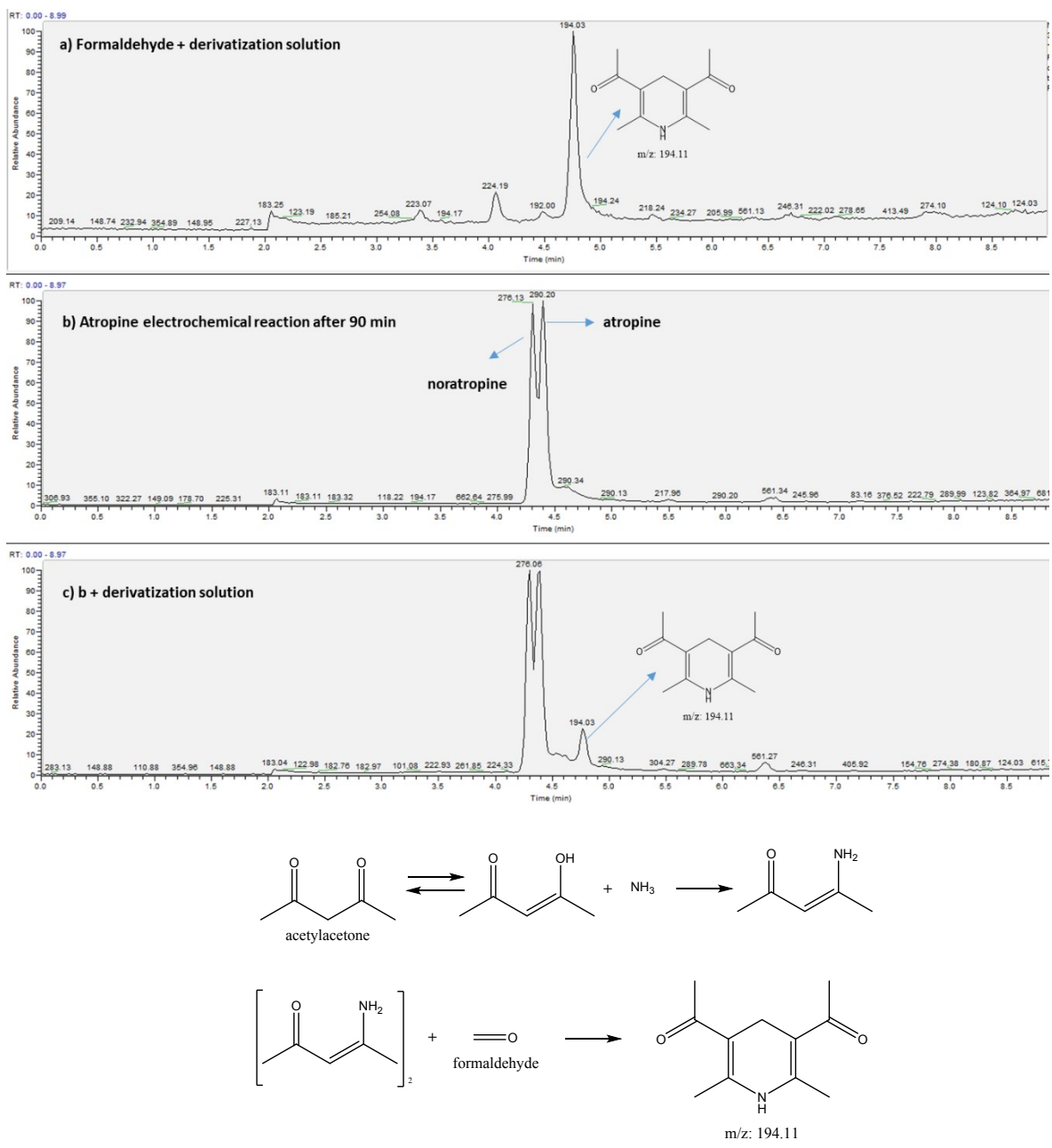


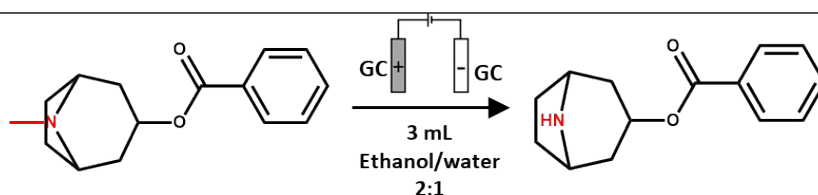
Fig S12, LC-MS chromatogram and reaction scheme of the derivatization reaction of formaldehyde in the reaction solution.

Table S2, electrochemical N-demethylation of atropine to noratropine using another set of electrodes. 50 mg atropine, 3 mL, ethanol, NaClO<sub>4</sub> (4 equiv.) (the same condition as Table 2, entry 9-12)

Entry	Current/time	Added water	Conversion (%) <sup>a</sup>	Yield (%)
1	4 mA / 4.1 h	18.5 M (33% v/v)	99	74 <sup>b</sup>
2	4 mA / 4.1 h	18.5 M (33% v/v)	99	70 <sup>b</sup>
3	4 mA / 4.1 h	18.5 M (33% v/v)	99	70 <sup>b</sup>
4	4 mA / 4.1 h	18.5 M (33% v/v)	99	76 <sup>b</sup>

<sup>a</sup> determined by LC-MS. <sup>b</sup> Isolated yield after 2-step-NH<sub>4</sub>OH-LLE

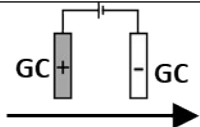
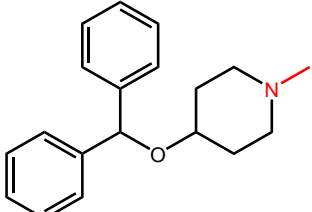
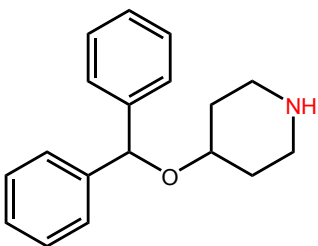
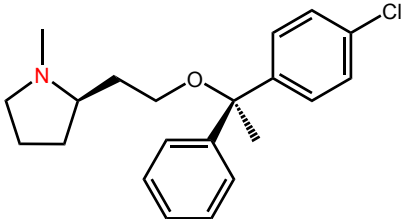
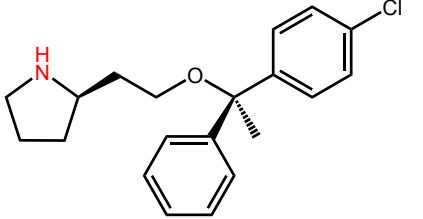
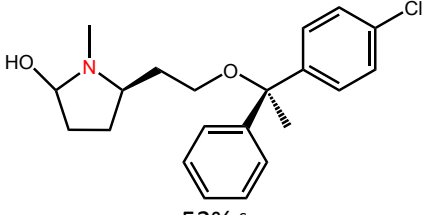
Table S3, electrochemical N-demethylation of tropacocaine at different pH values.



Entry	Current/time	Supporting electrolyte (4 equiv.)	Conversion (%) <sup>a</sup>	Yield (%)	pH <sup>b</sup>
1 <sup>c, d</sup>	4 mA / 7 h	NaClO <sub>4</sub>	99	70 <sup>e</sup>	11
2 <sup>c</sup>	4 mA / 4 h	LiCl	0	-	4
3 <sup>f</sup>	4 mA / 1 h	NaClO <sub>4</sub>	2	-	5
4 <sup>f, g</sup>	4 mA / 1 h	NaClO <sub>4</sub>	95	81 <sup>h</sup>	10

<sup>a</sup> determined by LC-MS. <sup>b</sup> measured by pH paper. <sup>c</sup> 54 mM. <sup>d</sup> reaction with free amine. <sup>e</sup> isolated yield by 2-step-NH<sub>4</sub>OH LLE. <sup>f</sup> 20 mM. <sup>g</sup> pH increased by adding equimolar amount of sodium carbonate. <sup>h</sup> yield determined by LC-MS.

Table S4, electrochemical N-demethylation of N-methylpiperidine and N-methylpyrrolidine examples.

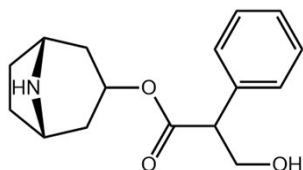
Starting compound	 60 mg 3 mL Ethanol/water (2:1) NaClO <sub>4</sub> (4 equiv.)	Synthesized compound
 4-diphenylmethoxy-1-methyl piperidine <b>13</b>	0.96 V <sup>a</sup> 4 mA / 12 h	 44 % <sup>b</sup>
 Clemastine <b>14</b>	0.82 V <sup>a</sup> 4 mA / 9 h	 19% <sup>c</sup> +  52% <sup>c</sup>

<sup>a</sup> electrochemical oxidation potential versus Ag/AgCl. <sup>b</sup> Isolated yield after silica-gel chromatography.

<sup>c</sup> Isolated yield after C18 chromatography.

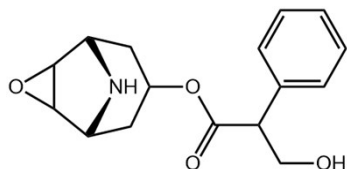
## $^1\text{H}$ and $^{13}\text{C}$ NMR of compounds

### Noratropine



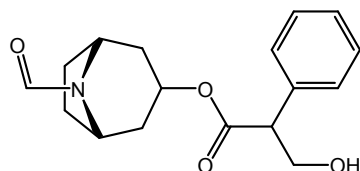
$^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.41 – 7.25 (m, 5H), 5.07 (t,  $J = 5.2$  Hz, 1H), 4.21 – 4.12 (m, 1H), 3.86 – 3.74 (m, 2H), 3.43 (dt,  $J = 6.8, 3.1$  Hz, 1H), 3.31 (dt,  $J = 6.7, 3.0$  Hz, 1H), 2.45 (br.s, 2H), 2.00 (dddd,  $J = 15.1, 5.3, 3.7, 1.5$  Hz, 1H), 1.96 – 1.83 (m, 2H), 1.74 (dt,  $J = 15.1, 1.8$  Hz, 1H), 1.70 – 1.59 (m, 1H), 1.53 (dq,  $J = 15.0, 1.8$  Hz, 1H), 1.52 – 1.40 (m, 1H), 1.31 (ddd,  $J = 13.1, 9.2, 4.5$  Hz, 1H).  $^{13}\text{C NMR}$  (126 MHz,  $\text{CDCl}_3$ )  $\delta$  172.36 (C=O), 135.92 (C), 128.94 (CH), 128.25 (CH), 127.79 (CH), 68.73 (CH), 64.03 ( $\text{CH}_2$ ), 54.66 (CH), 53.17 (CH), 53.09 (CH), 37.20 ( $\text{CH}_2$ ), 36.92 ( $\text{CH}_2$ ), 28.98 ( $\text{CH}_2$ ), 28.59 ( $\text{CH}_2$ ). **HRMS**; Calculated for  $\text{C}_{16}\text{H}_{22}\text{NO}_3$   $[\text{M}+\text{H}^+]=276.1597$ , observed = 276.1594. Data are consistent with literature <sup>1,2</sup>.

### Norscopolamine



$^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.43 – 7.32 (m, 2H), 7.36 – 7.29 (m, 1H), 7.32 – 7.20 (m, 2H), 5.03 (t,  $J = 5.2$  Hz, 1H), 4.17 (dd,  $J = 11.0, 8.8$  Hz, 1H), 3.81 (dd,  $J = 11.0, 5.3$  Hz, 1H), 3.76 (dd,  $J = 8.8, 5.2$  Hz, 1H), 3.27 (d,  $J = 3.0$  Hz, 1H), 3.22 (dd,  $J = 3.9, 2.3$  Hz, 1H), 3.07 (dd,  $J = 3.9, 2.2$  Hz, 1H), 2.61 (br.s, 1H), 2.72 (s, 1H), 2.52 (d,  $J = 3.0$  Hz, 1H), 2.13 (ddd,  $J = 15.2, 5.4, 3.9$  Hz, 1H), 2.04 (ddd,  $J = 15.3, 5.2, 3.9$  Hz, 1H), 1.67 (dq,  $J = 15.3, 1.8$  Hz, 1H), 1.44 (dq,  $J = 15.5, 1.8$  Hz, 1H).  $^{13}\text{C NMR}$  (126 MHz,  $\text{CDCl}_3$ )  $\delta$  171.86 (C=O), 135.82 (C), 129.13 (CH), 128.17 (CH), 128.11 (CH), 66.94 (CH), 63.95 ( $\text{CH}_2\text{-OH}$ ), 54.40 (CH), 53.97 (CH), 53.47 (CH), 51.92 (CH), 51.80 (CH), 31.34 ( $\text{CH}_2$ ), 31.14 ( $\text{CH}_2$ ). **HRMS**: Calculated for  $\text{C}_{16}\text{H}_{20}\text{NO}_4$   $[\text{M}+\text{H}^+]=290.1387$ , observed = 290.1388. Data are consistent with literature <sup>1-3</sup>.

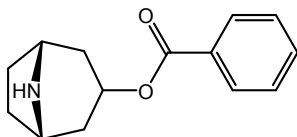
### N-formyl-noratropine



$^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$  8.08 (s, 0.5H), 8.06 (s, 0.5H), 7.40 – 7.25 (m, 5H), 5.21 – 5.15 (m, 1H), 4.56 (dt,  $J = 6.8, 3.0$  Hz, 0.5H), 4.43 (dt,  $J = 6.8, 2.9$  Hz, 0.5H), 4.22 (ddd,  $J = 9.9, 7.9, 1.3$  Hz, 1H), 4.01 (dt,  $J = 6.7, 2.8$  Hz, 0.5H), 3.91 – 3.80 (m, 2.5H), 2.46 (broad s, 1H), 2.20 – 1.55 (m, 7H), 1.28 (m, 1H).  $^{13}\text{C NMR}$  (126 MHz,  $\text{CDCl}_3$ )  $\delta$  172.16 (OC=O), 172.14 (OC=O), 157.37 (CH=O), 135.41 (C), 135.33 (C), 129.06 (CH), 129.04 (CH), 128.11 (CH), 128.08 (CH), 128.00 (CH), 127.96 (CH), 68.23 (HC-O), 64.12 ( $\text{CH}_2\text{-OH}$ ), 64.08 ( $\text{CH}_2\text{-OH}$ ), 54.34 (CH), 54.31 (CH), 53.37 (CH), 53.27 (CH), 48.52 (CH), 48.44 (CH), 38.47 ( $\text{CH}_2$ ), 38.14 ( $\text{CH}_2$ ), 35.82 ( $\text{CH}_2$ ), 35.57 ( $\text{CH}_2$ ), 27.88 ( $\text{CH}_2$ ), 27.40 ( $\text{CH}_2$ ), 27.11 ( $\text{CH}_2$ ), 26.68 ( $\text{CH}_2$ ). **HRMS**: Calculated for  $\text{C}_{17}\text{H}_{22}\text{NO}_4$   $[\text{M}+\text{H}^+]=304.1543$ , observed = 304.1544. Data are consistent with literature <sup>2</sup>.

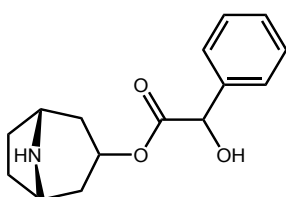


### Nortropacocaine



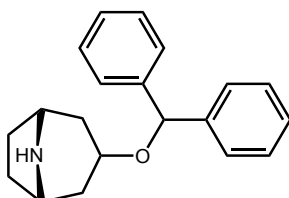
**<sup>1</sup>H NMR** (500 MHz, CD<sub>3</sub>OD) δ 8.05 – 7.94 (m, 2H), 7.59 (dd, J = 8.3, 6.5 Hz, 1H), 7.46 (t, J = 7.8 Hz, 2H), 5.29 (tt, J = 11.2, 6.1 Hz, 1H), 3.61 (p, J = 3.0 Hz, 2H), 2.10 (ddd, J = 13.6, 6.1, 2.9 Hz, 2H), 1.94 – 1.79 (m, 4H), 1.82 – 1.69 (m, 2H). **<sup>13</sup>C NMR** (126 MHz, CDCl<sub>3</sub>) δ 166.56 (C=O), 133.36 (C), 130.58 (CH), 129.53 (CH), 128.27 (CH), 67.50 (CH), 54.31 (CH), 38.98 (CH<sub>2</sub>), 29.15 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>14</sub>H<sub>18</sub>NO<sub>2</sub> [M+H<sup>+</sup>]=232.1332, observed = 232.1333. Data are consistent with literature <sup>4,5</sup>.

### Norhomatropine



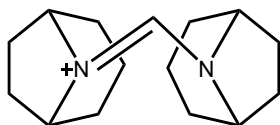
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d) δ 7.47 – 7.27 (m, 5H), 5.14 – 5.00 (m, 2H), 3.45 – 3.38 (m, 1H), 3.32 (s, 1H), 3.29 – 3.24 (m, 1H), 2.12 – 1.95 (m, 1H), 1.94 – 1.86 (m, 1H), 1.85 – 1.71 (m, 2H), 1.64 (tq, J = 12.3, 6.4, 5.9 Hz, 1H), 1.49 – 1.35 (m, 2H), 1.14 – 0.95 (m, 1H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d) δ 172.91 (C=O), 138.54 (C), 128.62 (CH), 128.49 (CH), 126.72 (CH), 73.29 (CH), 70.12 (CH), 52.93 (CH), 52.83 (CH), 36.97 (CH<sub>2</sub>), 36.74 (CH<sub>2</sub>), 28.87 (CH<sub>2</sub>), 28.36 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>15</sub>H<sub>20</sub>NO<sub>3</sub> [M+H<sup>+</sup>] = 262.1438, observed = 262.1438.

### Norbenzatropine



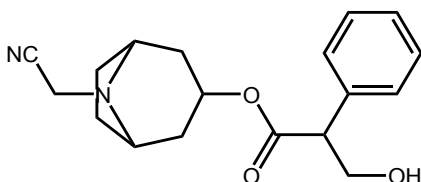
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d) δ 7.37 – 7.22 (m, 10H), 5.42 (s, 1H), 4.75 (s, 3H), 4.06 – 4.01 (m, 2H), 3.84 – 3.75 (m, 1H), 2.50 – 2.36 (m, 4H), 2.26 – 2.17 (m, 2H), 2.06 (d, J = 15.3 Hz, 2H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d) δ 141.98 (C), 128.55 (CH), 127.65 (CH), 126.71 (CH), 81.60 (CH), 67.66 (CH), 53.99 (CH), 33.42 (CH<sub>2</sub>), 26.31 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>20</sub>H<sub>24</sub>NO [M+H<sup>+</sup>] = 294.1852, observed = 294.1852.

### 8-((8-azabicyclo[3.2.1]octan-8-yl)methylene)-8-azabicyclo[3.2.1]octan-8-ium



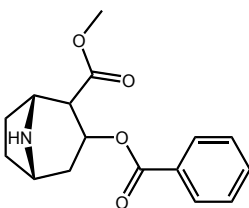
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d)  $\delta$  8.18 (s, 1H), 4.52 – 4.37 (m, 4H), 2.29 – 2.17 (m, J = 5.6, 4.7 Hz, 4H), 2.04 – 1.92 (m, 4H), 1.91 – 1.72 (m, 11H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d)  $\delta$  143.22 (CH), 63.85 (CH), 56.54 (CH), 33.21 (CH<sub>2</sub>), 32.09 (CH<sub>2</sub>), 28.35 (CH<sub>2</sub>), 25.68 (CH<sub>2</sub>), 16.22 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>15</sub>H<sub>25</sub>N<sub>2</sub><sup>+</sup> [M] = 233.2012, observed = 233.2012.

### 8-(cyanomethyl)-8-azabicyclo[3.2.1]octan-3-yl 3-hydroxy-2-phenylpropanoate



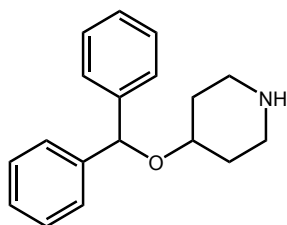
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d)  $\delta$  7.37 – 7.23 (m, 5H), 5.05 (t, J = 5.3 Hz, 1H), 4.18 (dd, J = 11.0, 8.6 Hz, 1H), 3.86 – 3.76 (m, 2H), 3.27 (d, J = 13.7 Hz, 3H), 3.19 – 3.12 (m, 1H), 2.44 – 2.24 (br.s, 1H), 2.21 – 2.05 (m, 2H), 1.87 – 1.74 (m, 3H), 1.66 – 1.51 (m, 2H), 1.26 – 1.20 (m, 1H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d)  $\delta$  172.20 (CO), 135.46 (C), 129.01 (CH), 128.14 (CH), 127.90 (CH), 116.95 (CN), 67.54 (CH), 64.15 (CH<sub>2</sub>-OH), 58.95 (CH), 58.86 (CH), 54.28 (CH), 40.81 (CH<sub>2</sub>), 36.65 (CH), 36.45 (CH), 25.25 (CH), 24.70 (CH). **HRMS:** Calculated for C<sub>18</sub>H<sub>23</sub>N<sub>2</sub>O<sub>3</sub> [M+H<sup>+</sup>] = 315.1703, observed = 315.1703. Data are consistent with literature <sup>6,7</sup>.

### Norcocaine



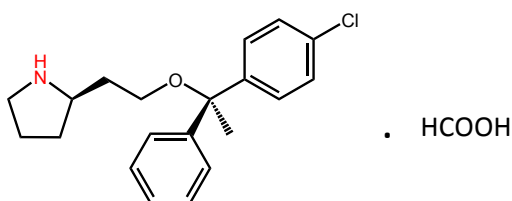
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d)  $\delta$  7.96 – 7.91 (m, 2H), 7.55 (td, J = 7.3, 1.4 Hz, 1H), 7.45 – 7.38 (m, 2H), 5.44 (dt, J = 11.6, 6.5 Hz, 1H), 3.82 (td, J = 7.6, 2.6 Hz, 2H), 3.64 (s, 3H), 3.47 (s, 1H), 3.11 (dd, J = 6.8, 2.2 Hz, 1H), 2.20 – 1.95 (m, 4H), 1.80 – 1.70 (m, 2H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d)  $\delta$  172.83 (C=O), 165.64 (C=O), 133.32 (CH), 129.94 (C), 129.64 (CH), 128.52 (CH), 66.75 (CH), 56.07 (CH), 53.40 (CH), 51.96 (CH<sub>3</sub>), 48.02 (CH), 34.82 (CH<sub>2</sub>), 28.10 (CH<sub>2</sub>), 27.20 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>16</sub>H<sub>20</sub>NO<sub>4</sub> [M+H<sup>+</sup>] = 290.1386, observed = 290.1386.

### 4-(benzhydryloxy)piperidine



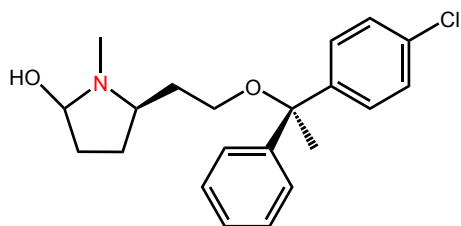
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d) 7.32 (d,  $J = 5.5$  Hz, 8H), 7.26 – 7.23 (m, 2H), 5.44 (s, 1H), 3.81 – 3.67 (m, 2H), 3.45 (ddd,  $J = 14.0, 10.5, 3.9$  Hz, 2H), 3.27 (dt,  $J = 12.7, 4.6$  Hz, 2H), 1.99 (qt,  $J = 14.8, 4.0$  Hz, 4H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d)  $\delta$  142.01 (C), 128.72 (CH), 127.92 (CH), 127.01 (CH), 81.52 (CH), 67.49 (CH), 41.68 (CH<sub>2</sub>), 27.59 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>18</sub>H<sub>22</sub>NO [M+H<sup>+</sup>]=268.1696, observed = 268.1696 .

**(R)-2-(2-((R)-1-(4-chlorophenyl)-1-phenylethoxy)ethyl)pyrrolidine formate**



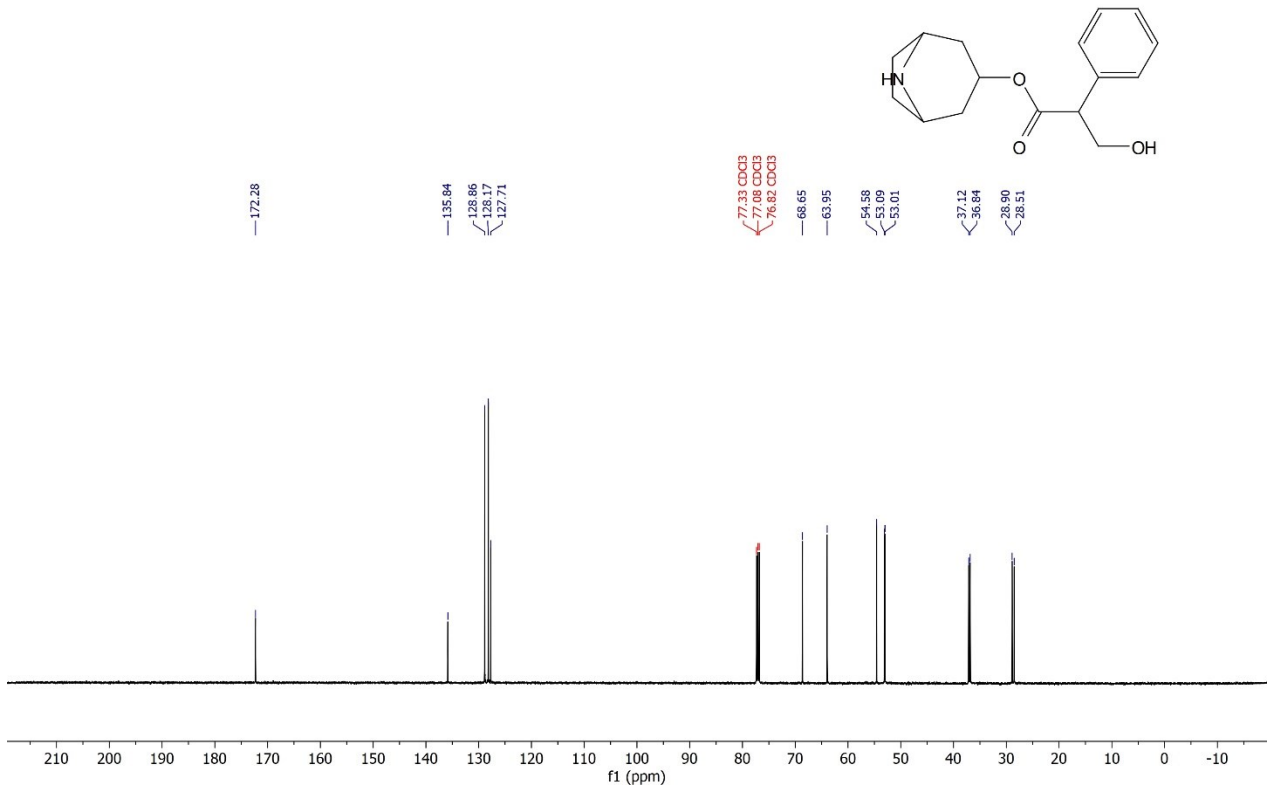
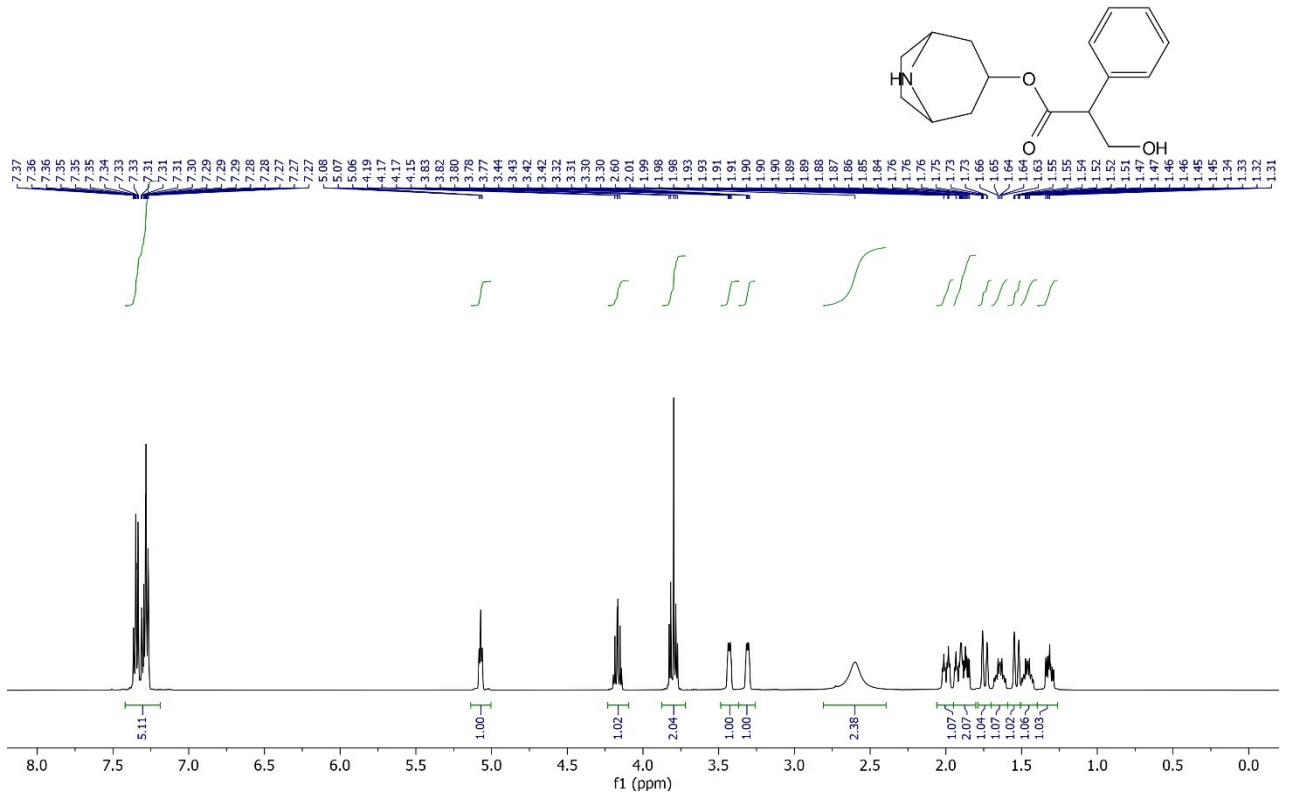
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d) 8.58 (s, 1H), 7.35 – 7.23 (m, 10H), 3.66 (tt,  $J = 8.9, 6.5$  Hz, 1H), 3.43 – 3.17 (m, 4H), 2.25 – 1.87 (m, 6H), 1.86 (d,  $J = 4.4$  Hz, 3H), 1.68 (dq,  $J = 12.8, 8.8$  Hz, 1H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d)  $\delta$  169.07 (C=O), 145.84 (C), 145.10 (C), 132.95 (CCl), 128.32 (CH), 128.30 (CH), 128.27 (CH), 127.34 (CH), 126.72 (CH), 80.70 (C), 59.76 (CH<sub>2</sub>), 57.78 (CH), 44.15 (CH<sub>2</sub>), 32.72 (CH<sub>2</sub>), 30.56 (CH<sub>2</sub>), 25.63 (CH<sub>3</sub>), 23.84 (CH<sub>2</sub>). **HRMS:** Calculated for C<sub>20</sub>H<sub>25</sub>NOCl[M+H<sup>+</sup>] = 330.1619, observed = 330.1616 .

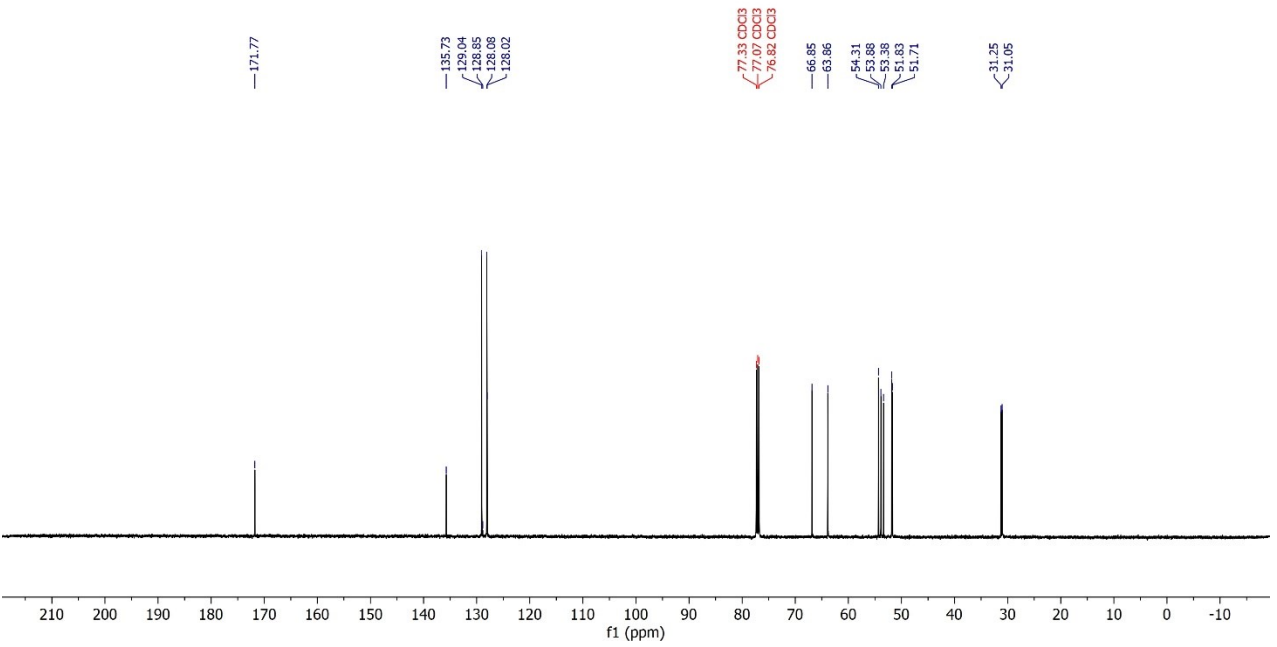
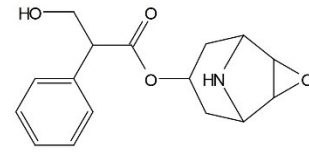
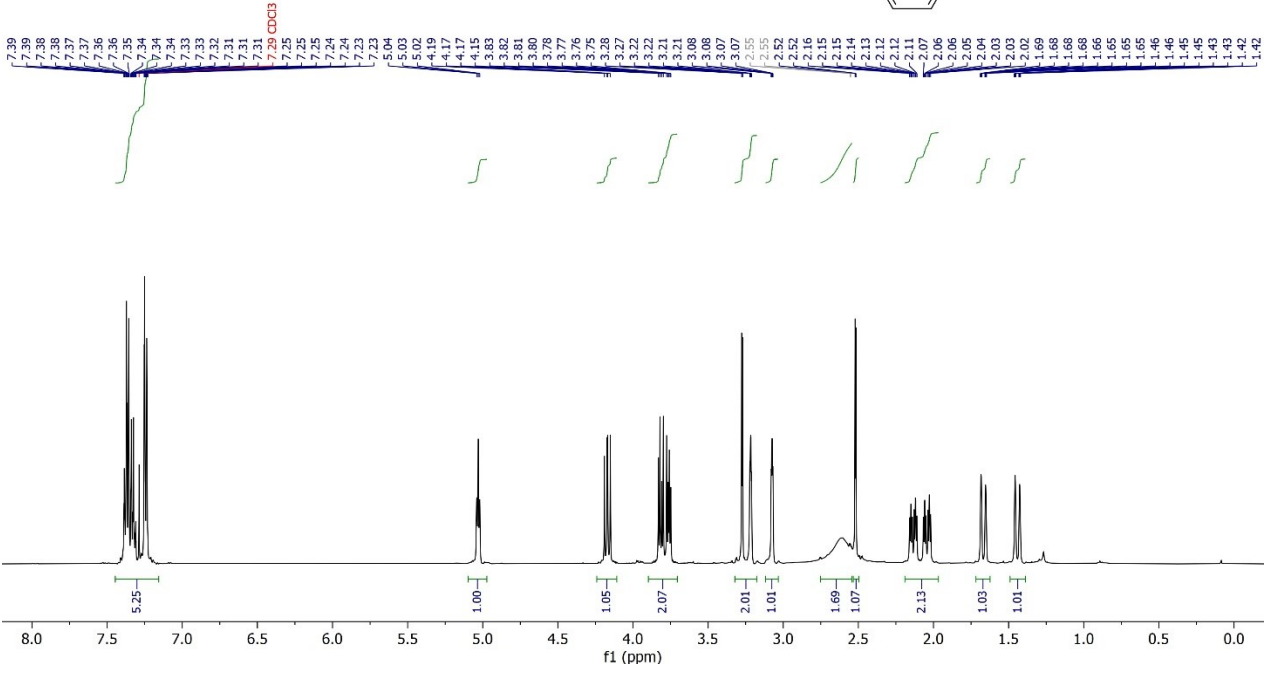
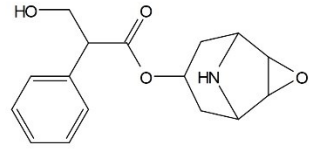
**(5R)-5-(2-((R)-1-(4-chlorophenyl)-1-phenylethoxy)ethyl)-1-methylpyrrolidin-2-ol**

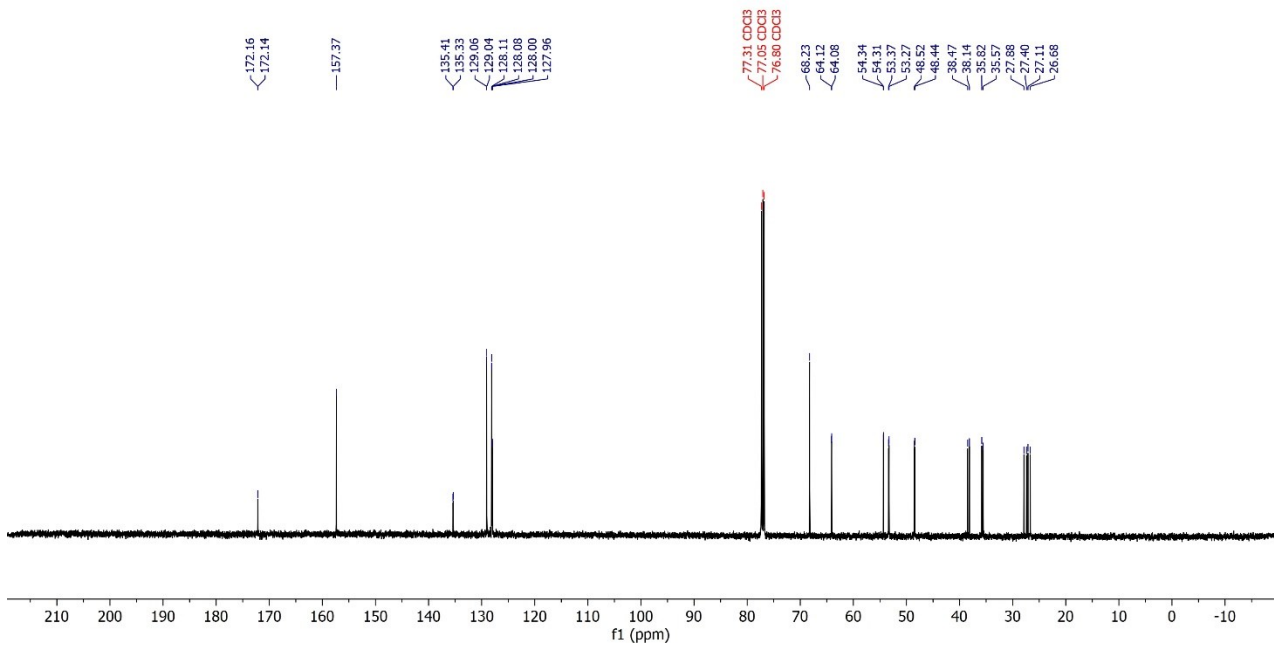
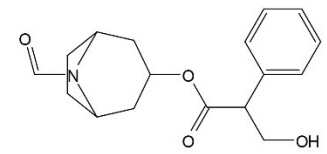
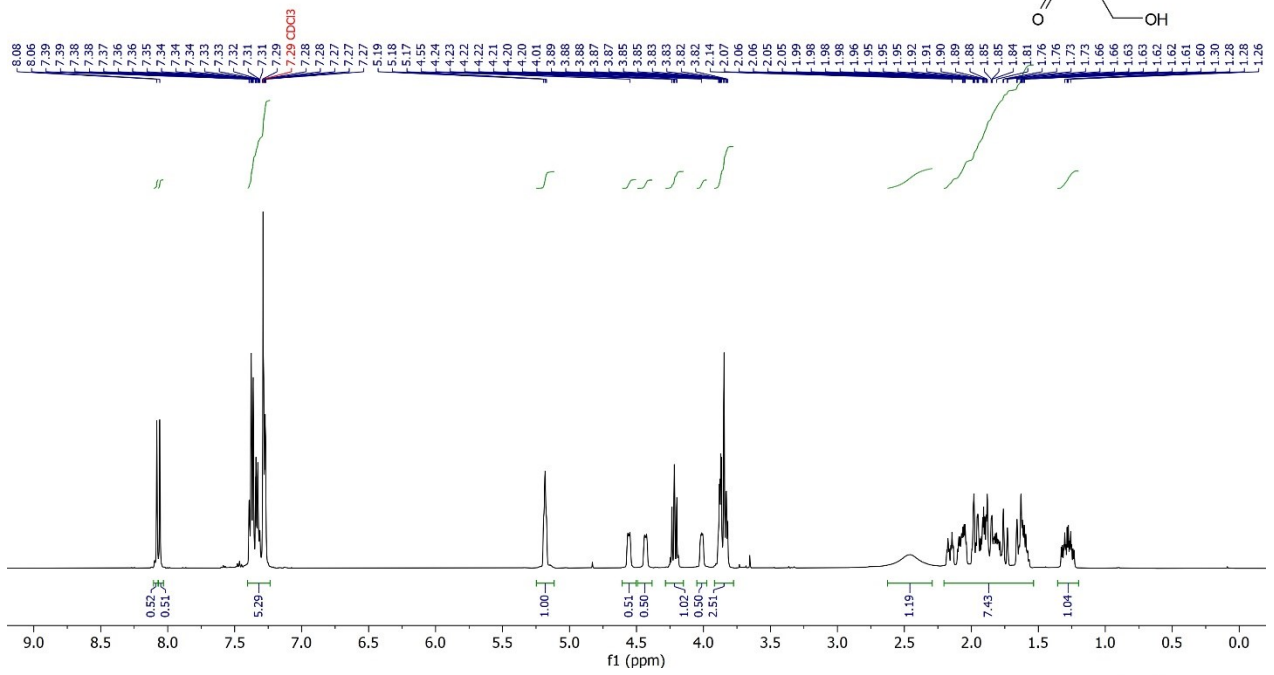
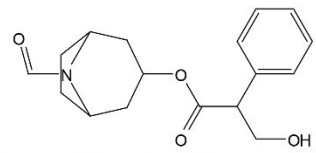


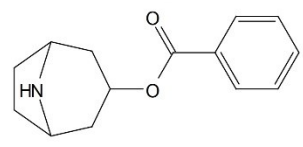
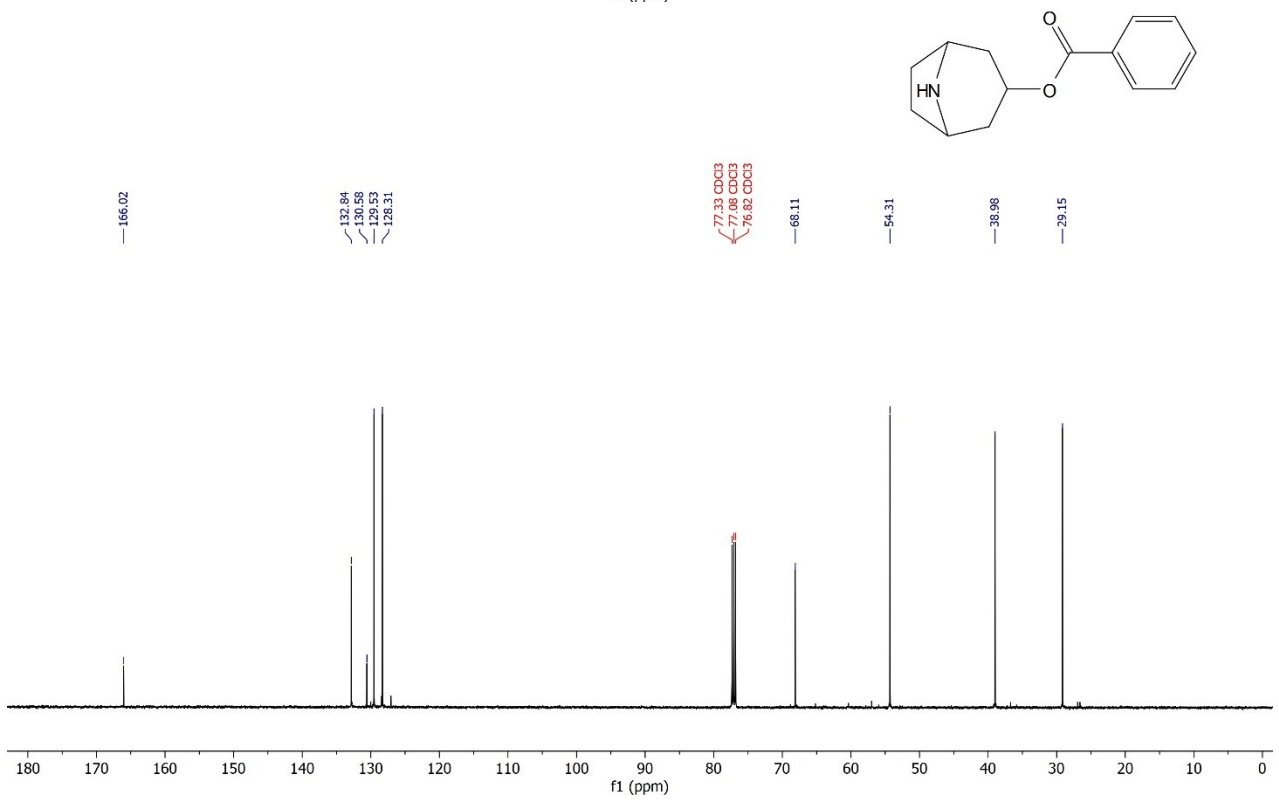
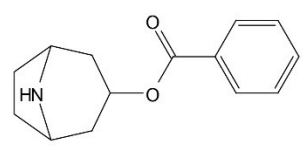
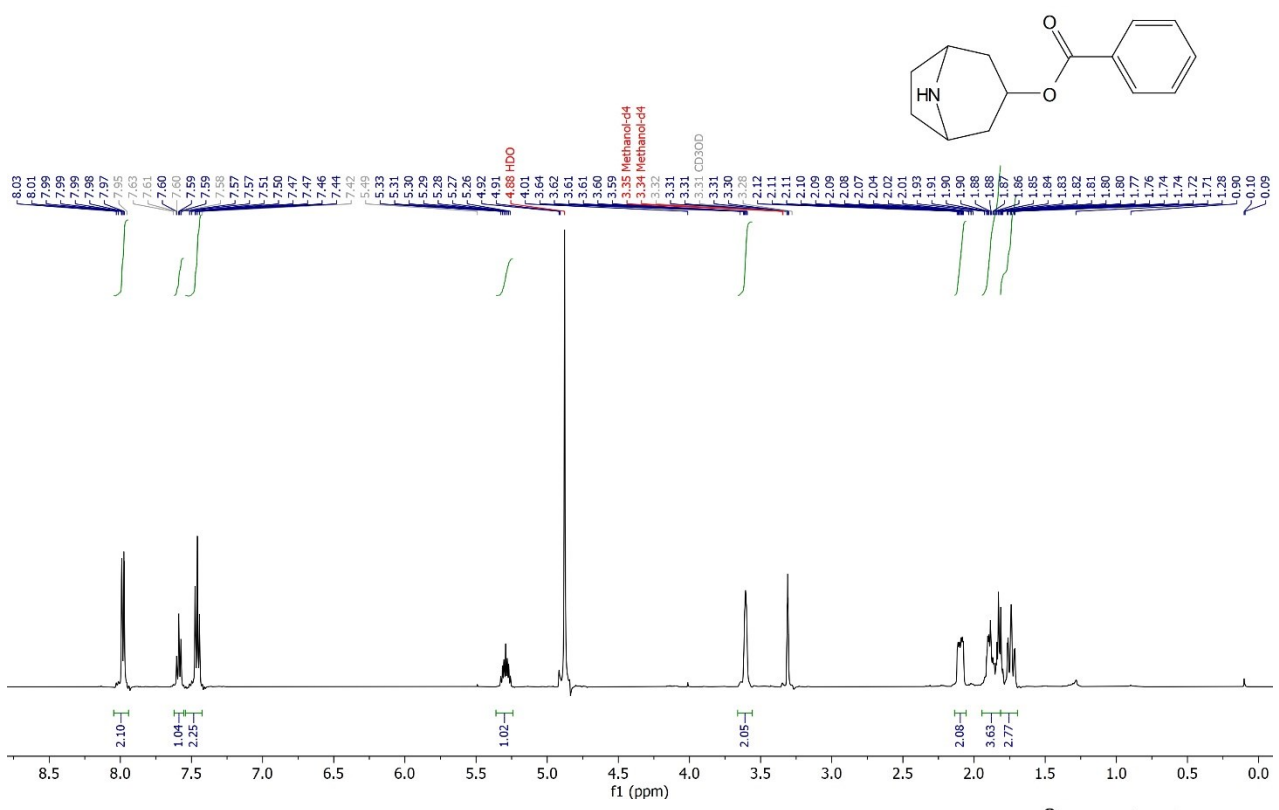
**<sup>1</sup>H NMR** (500 MHz, Chloroform-d)  $\delta$  7.36 – 7.24 (m, 9H), 3.99 (ddd,  $J = 11.4, 8.4, 2.6$  Hz, 1H), 3.81 (dq,  $J = 14.2, 8.8, 8.1$  Hz, 2H), 3.45 (s, 3H), 3.39 (dq,  $J = 8.3, 4.2, 3.1$  Hz, 1H), 3.29 (td,  $J = 9.0, 4.0$  Hz, 1H), 2.33 – 2.09 (m, 4H), 2.02 – 1.88 (m, 2H), 1.85 (s, 3H). **<sup>13</sup>C NMR** (126 MHz, Chloroform-d)  $\delta$  145.15 (C), 145.01 (C), 132.85 (CCl), 128.30 (CH), 128.26 (CH), 128.09 (CH), 127.41 (CH), 126.71 (CH), 80.80 (CH), 76.39 (C), 69.18 (CH), 58.77 (CH<sub>2</sub>), 51.82 (CH<sub>3</sub>), 28.24 (CH<sub>2</sub>), 27.76 (CH<sub>2</sub>), 25.49 (CH<sub>2</sub>), 19.56 (CH<sub>3</sub>). **HRMS:** Calculated for C<sub>21</sub>H<sub>27</sub>ClNO<sub>2</sub> [M+H<sup>+</sup>] = 360.1725, observed = 360.1720.

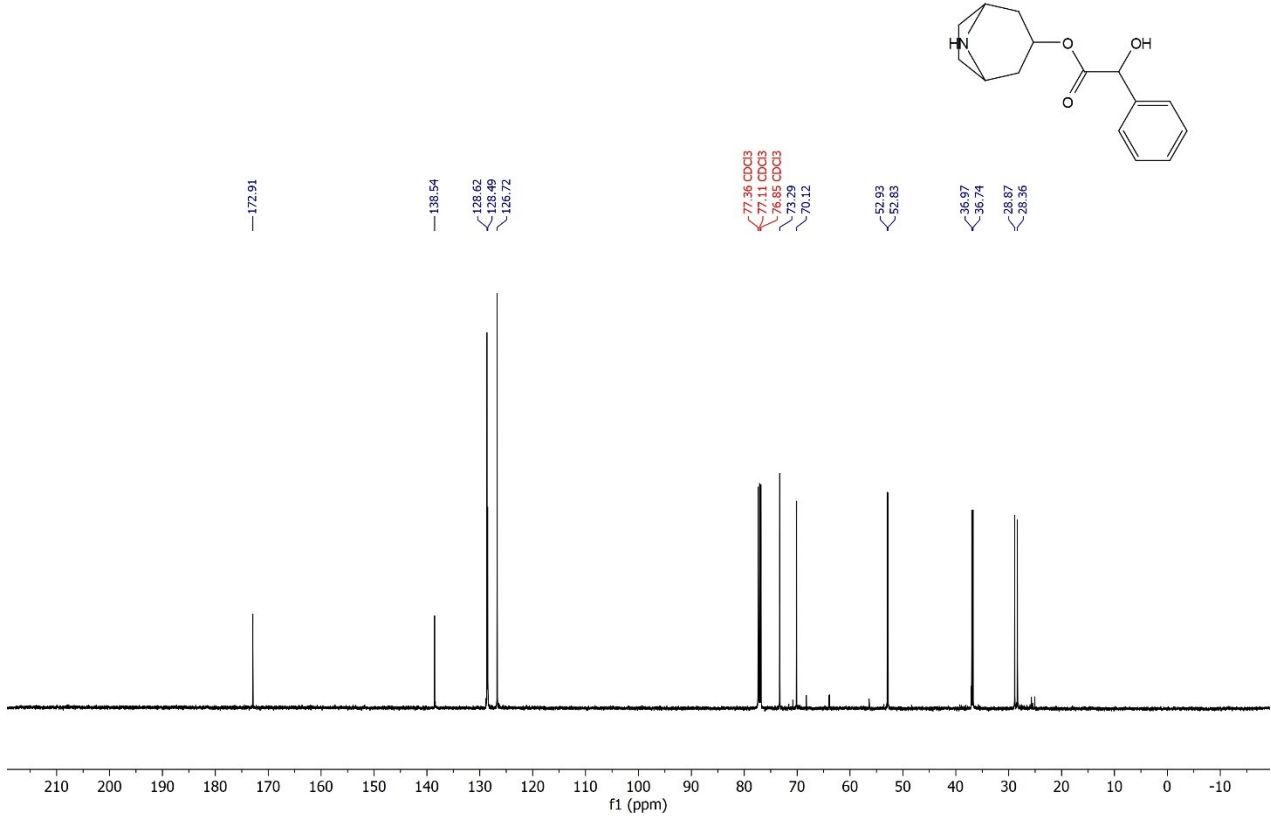
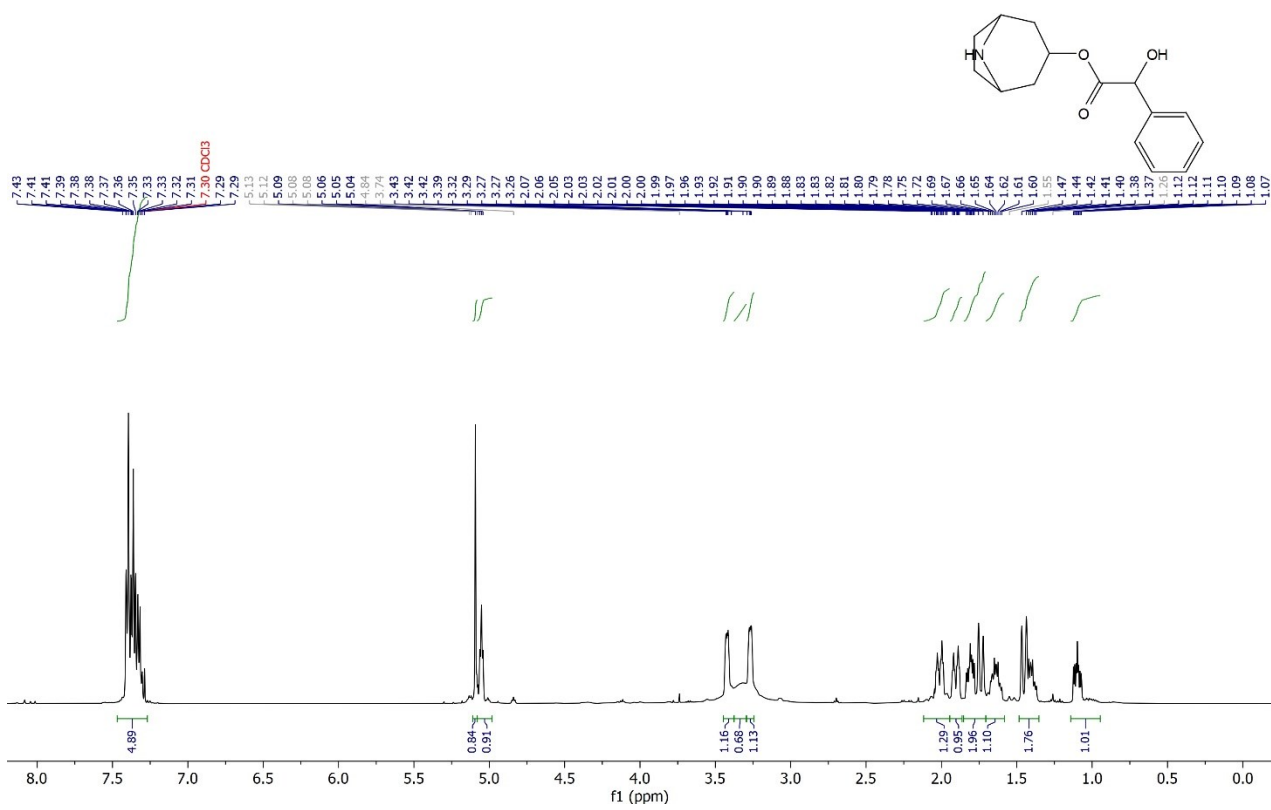
# $^1\text{H}$ and $^{13}\text{C}$ NMR spectra



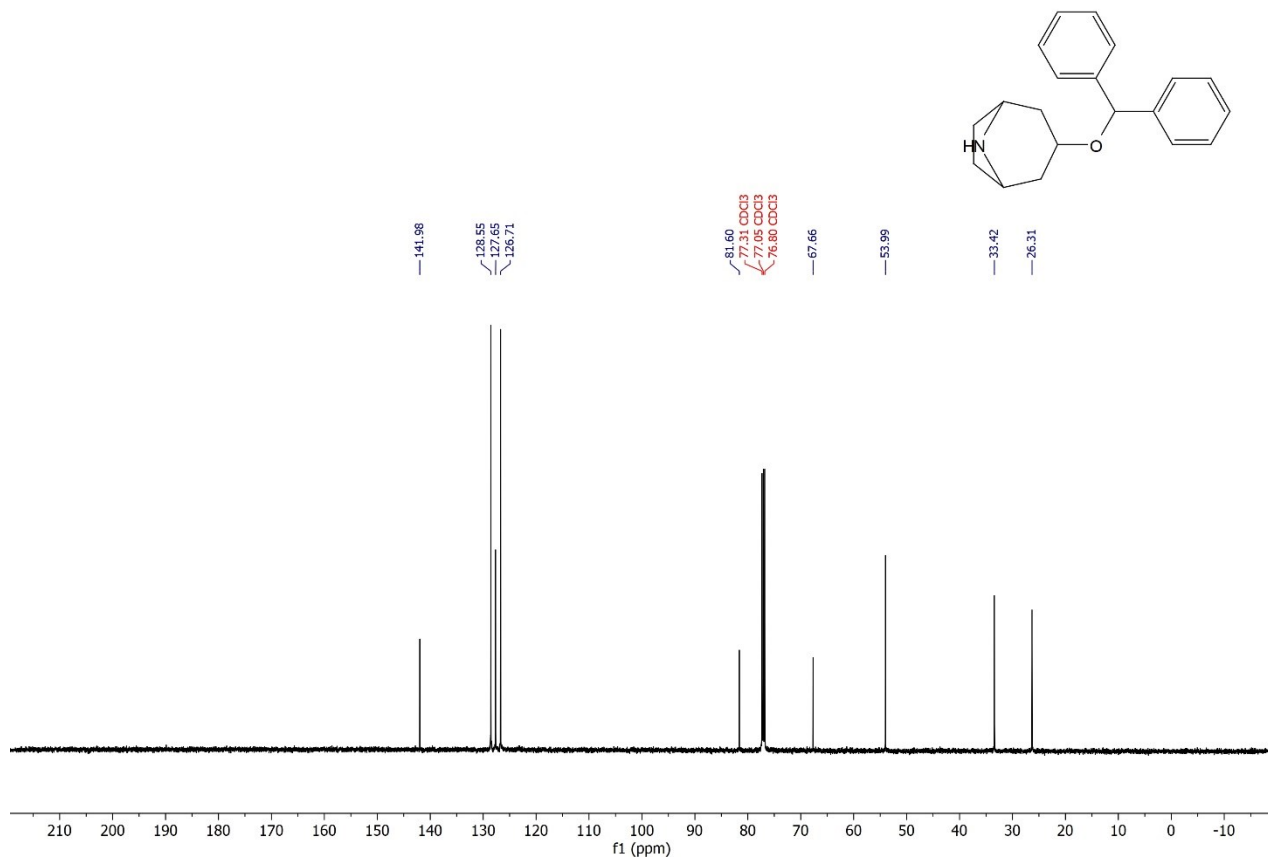
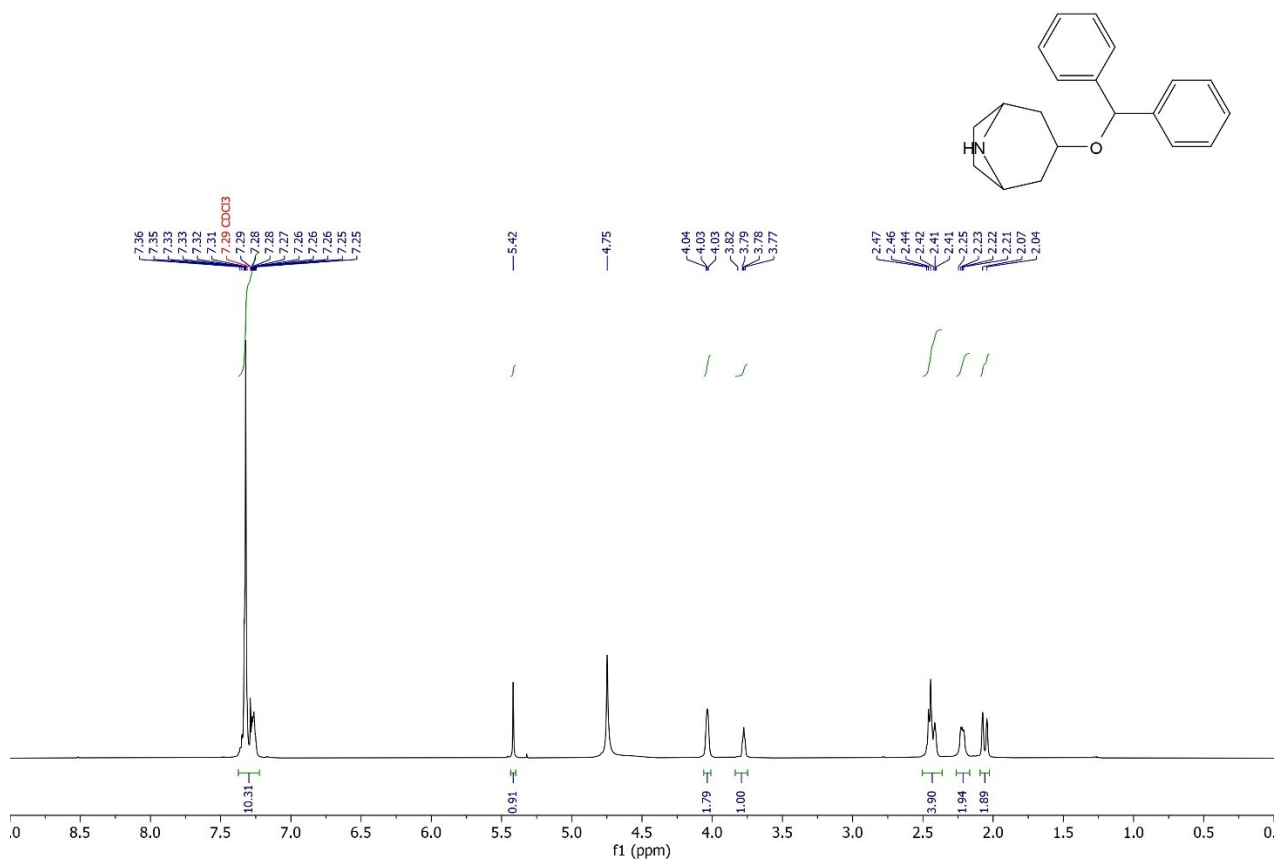


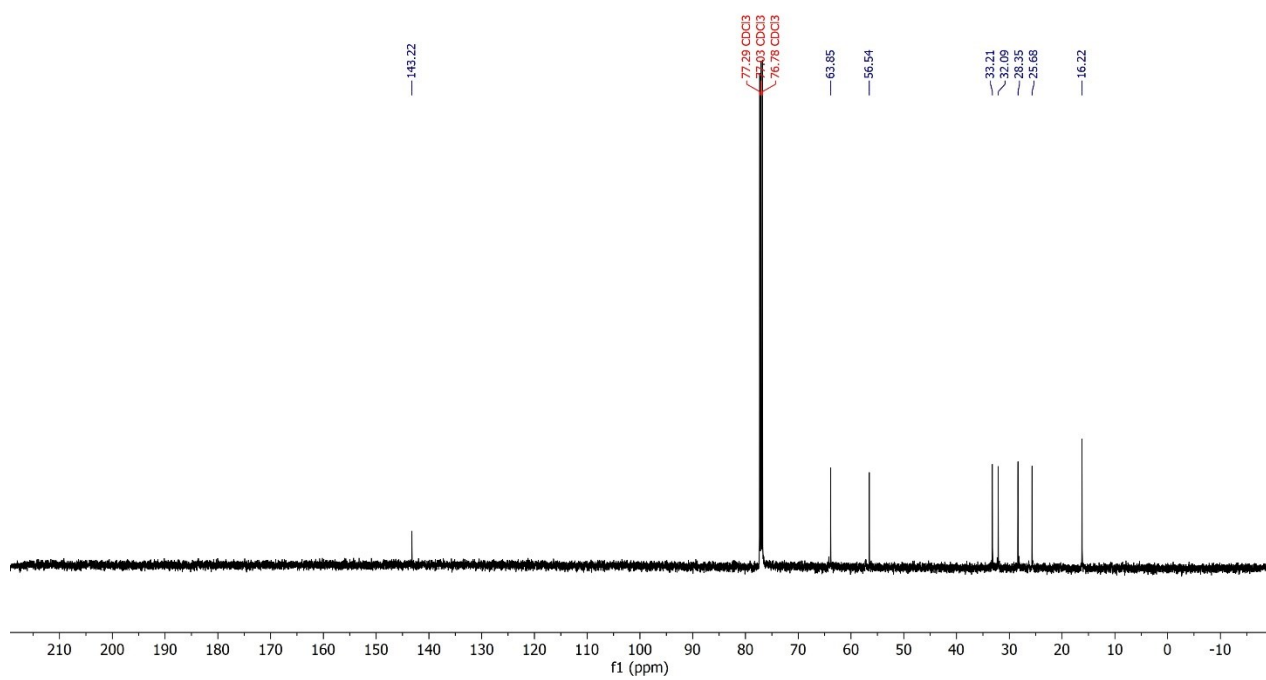
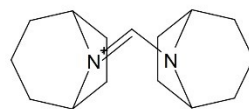
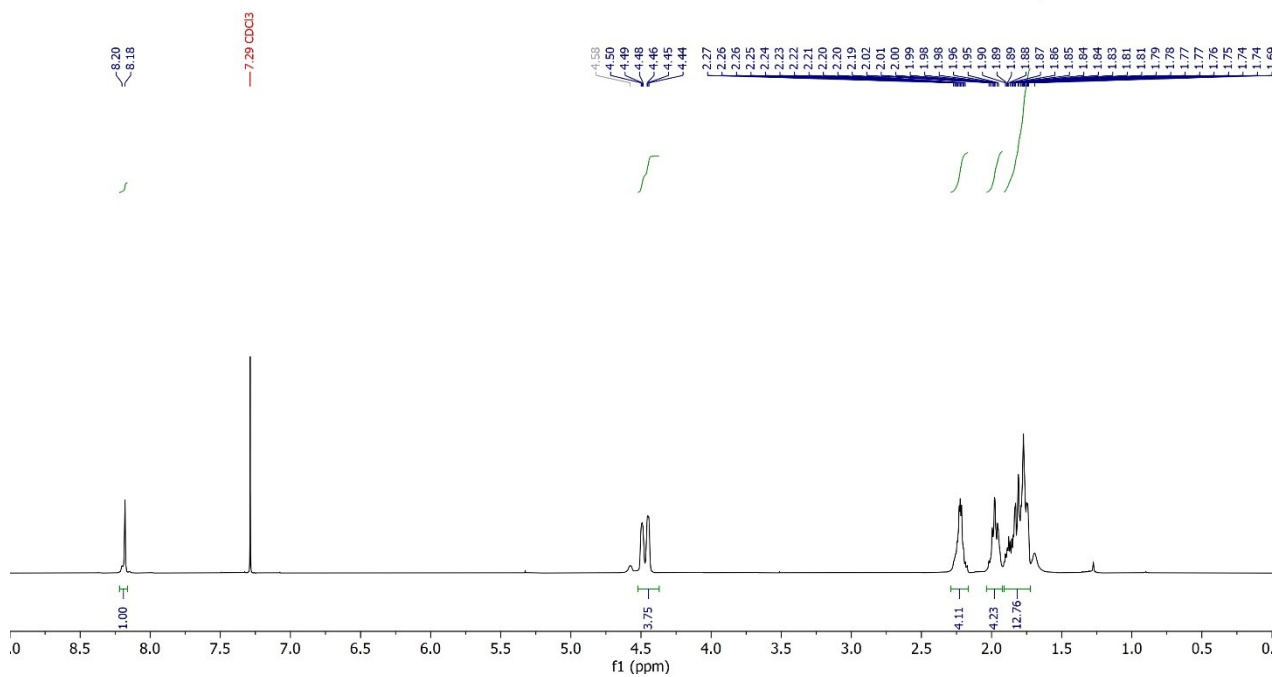
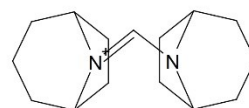


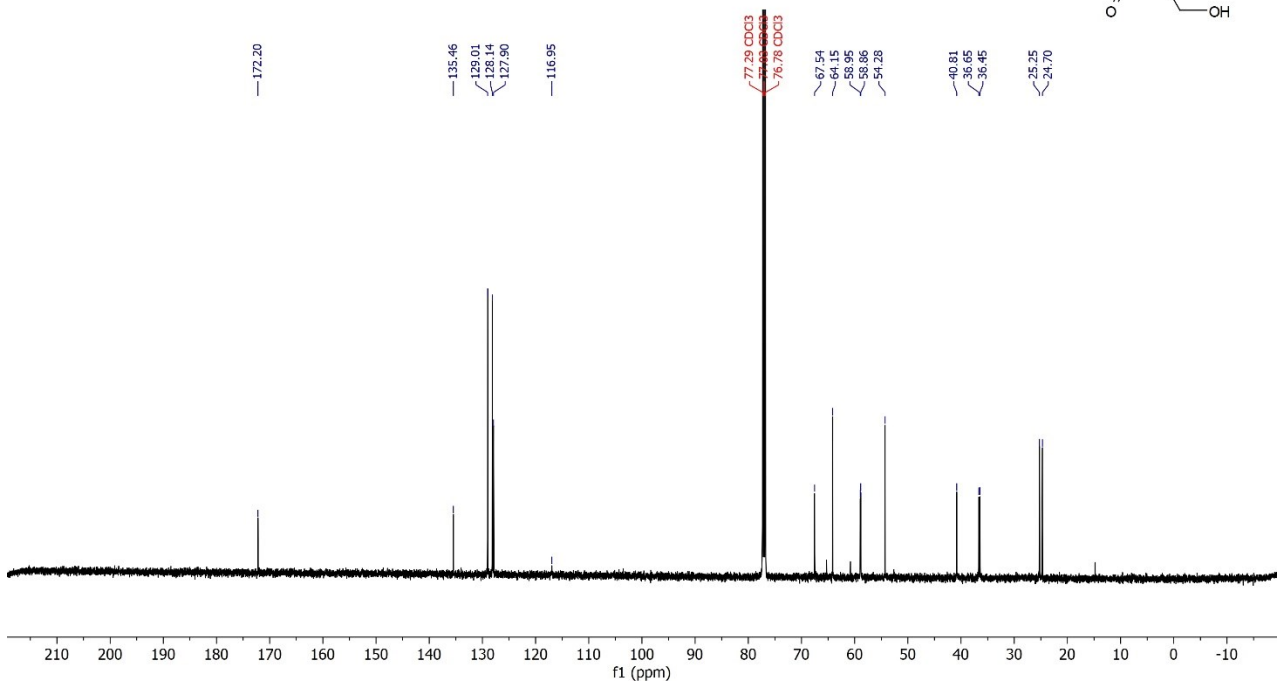
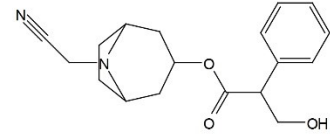
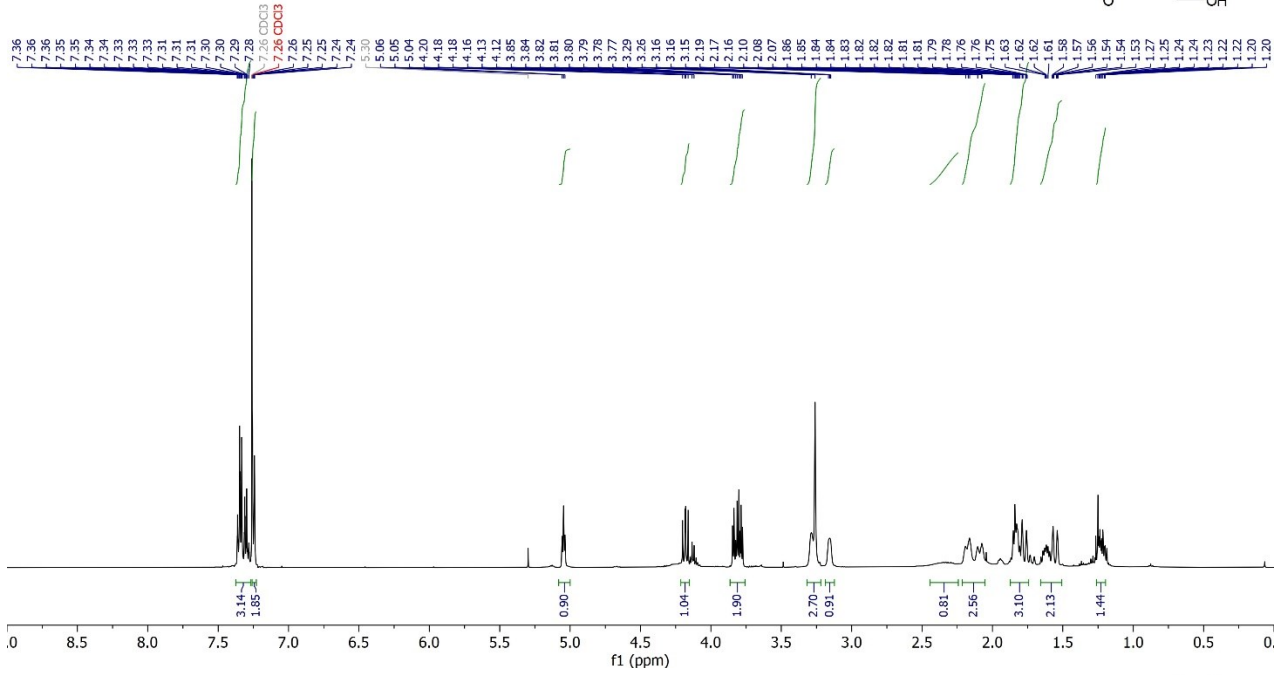
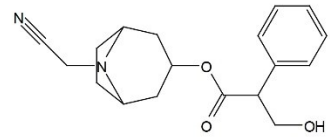


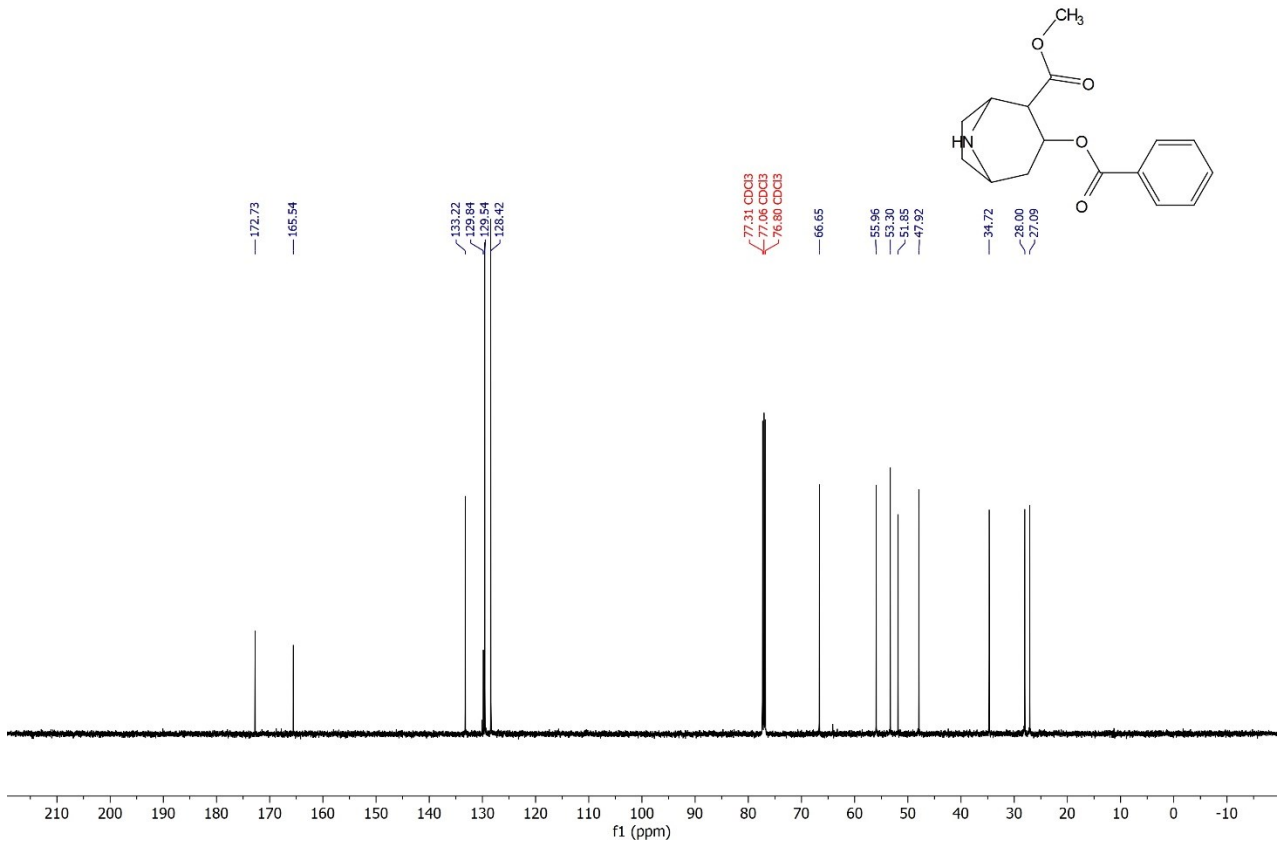
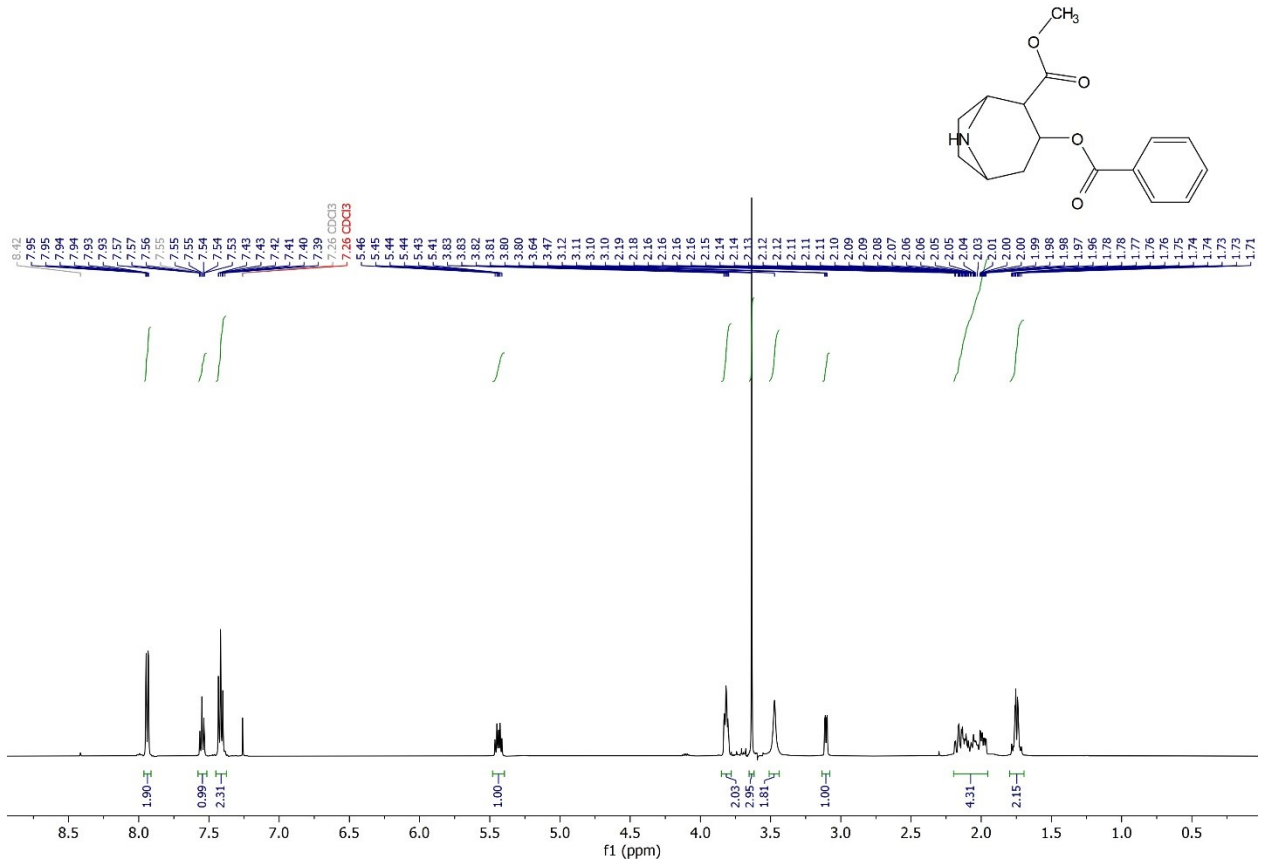


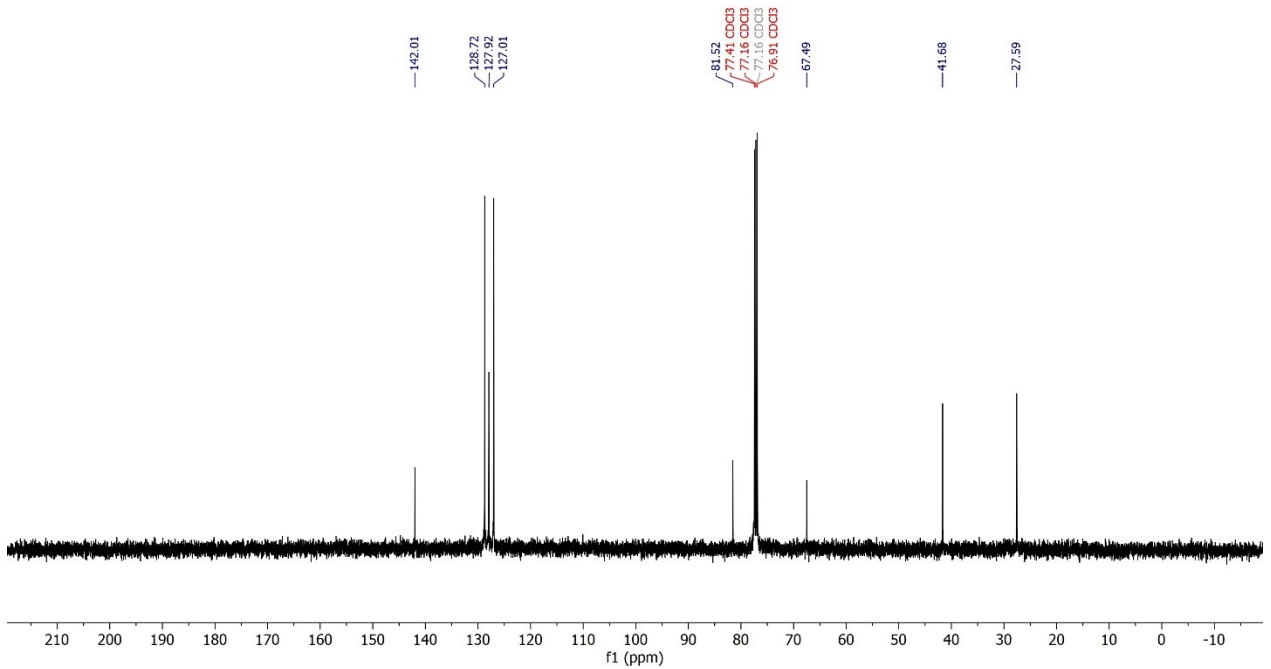
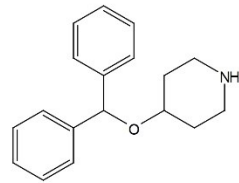
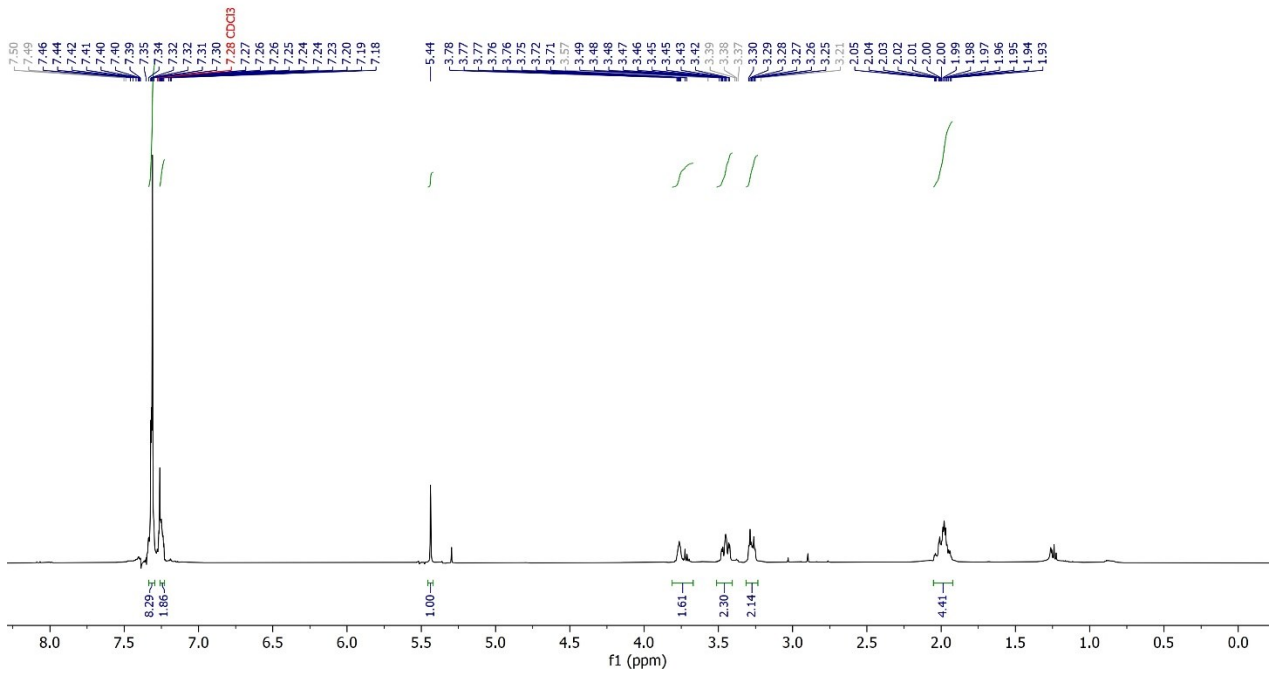
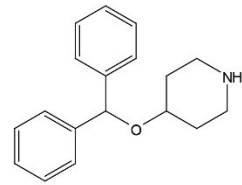


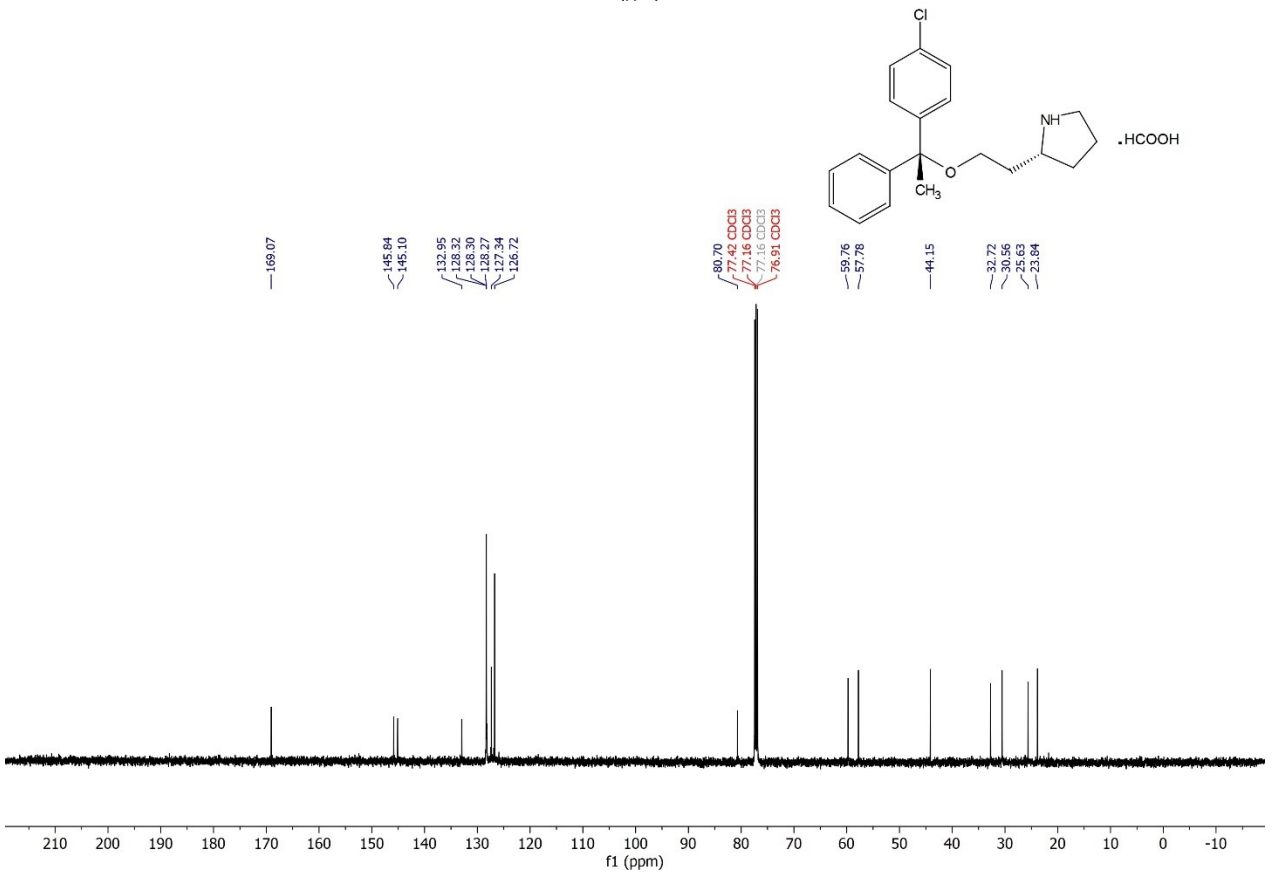
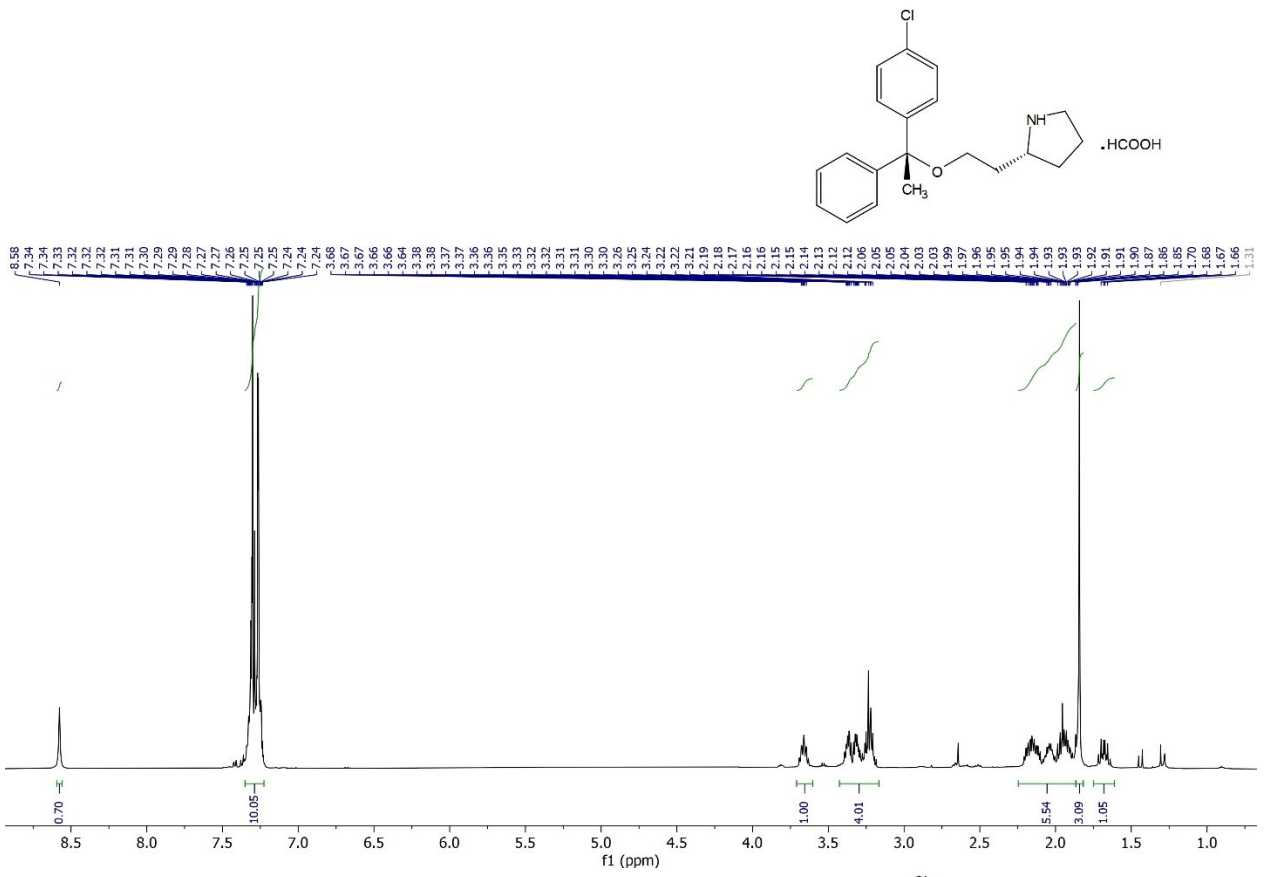


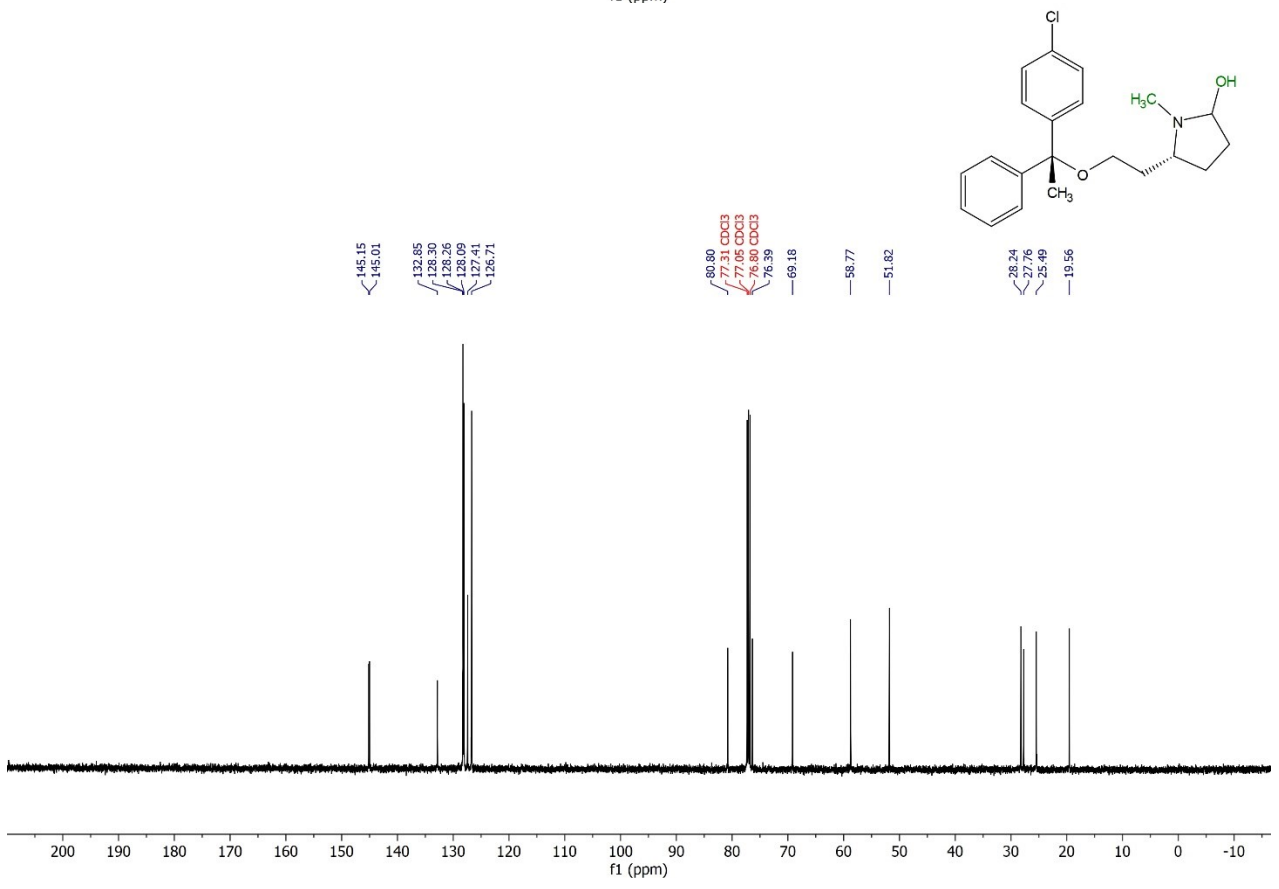
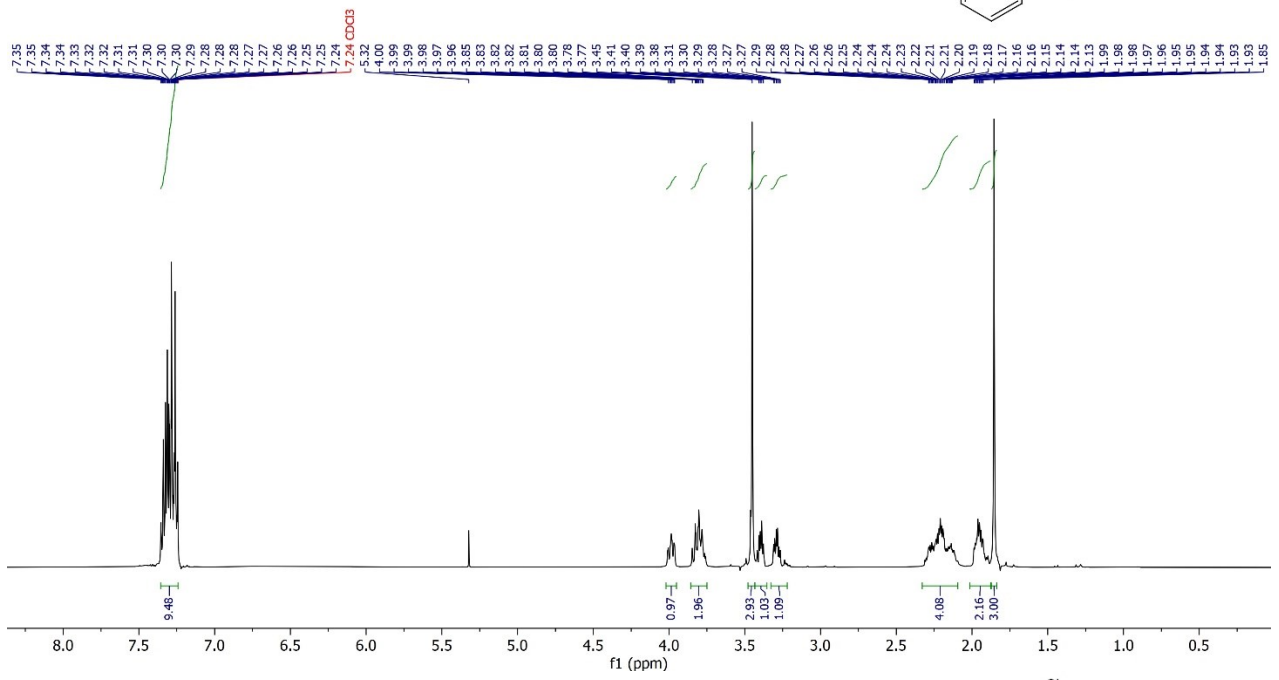




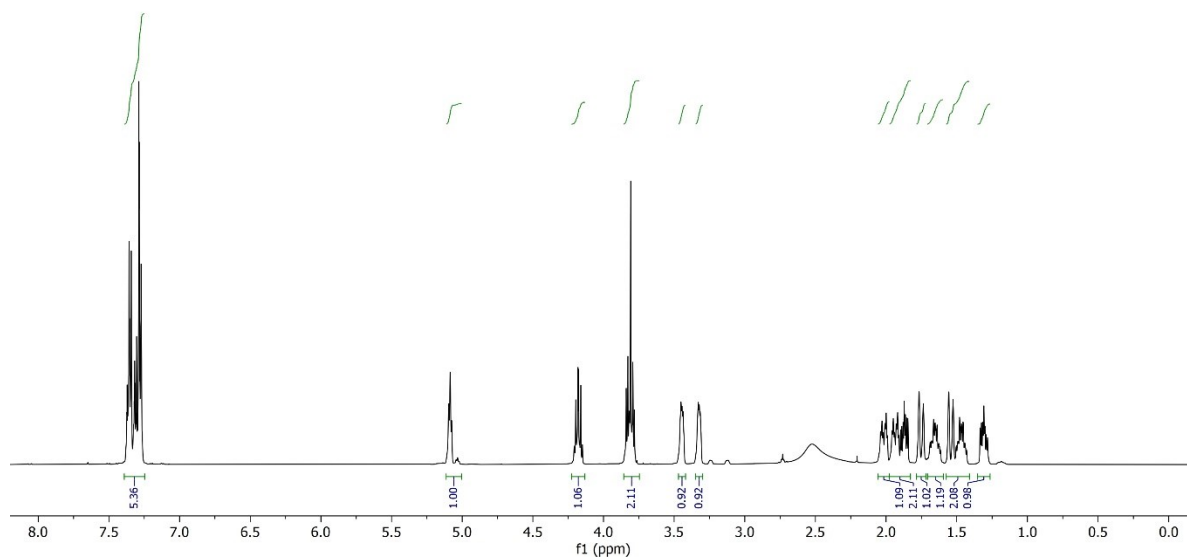
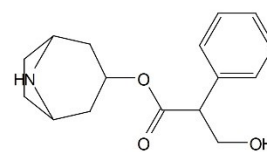








## <sup>1</sup>H NMR of gram-scale synthesized noratropine (Table 5, entry 1)



## References

- 1 D. D. Do Pham, G. F. Kelso, Y. Yang and M. T. W. W. Hearn, *Green Chem.*, 2012, **14**, 1189–1195.
- 2 D. D. Do Pham, G. F. Kelso, Y. Yang and M. T. W. Hearn, *Green Chem.*, 2014, **16**, 1399–1409.
- 3 J. P. Barham, M. P. John and J. A. Murphy, *J. Am. Chem. Soc.*, 2016, **138**, 15482–15487.
- 4 E. Edink, A. Akdemir, C. Jansen, R. Van Elk, O. Zuiderveld, F. J. J. De Kanter, J. E. Van Muijlwijk-Koezen, A. B. Smit, R. Leurs and I. J. P. De Esch, *Bioorganic Med. Chem. Lett.*, 2012, **22**, 1448–1454.
- 5 G. Maksay, P. Nemes and T. Biró, *J. Med. Chem.*, 2004, **47**, 6384–6391.
- 6 A. M. Nauth, A. Lipp, B. Lipp and T. Opatz, *European J. Org. Chem.*, 2017, **2017**, 2099–2103.
- 7 J. C. Orejarena Pacheco, A. Lipp, A. M. Nauth, F. Acke, J. P. Dietz and T. Opatz, *Chem. - A Eur. J.*, 2016, **22**, 5409–5415.