

Supplementary information

Biocatalytic microgels (μ -Gelzymes): synthesis, concepts, and emerging applications

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Enzyme class	Enzyme	Section	Immobilisation technique	Microgel	Enzyme loading [mg/g microgel/ copolymer]	Enzyme loading efficiency [%]	Residual activity after loading* [%]	Performance of μ -Gelzyme	Ref.
Hydrolase	Cellubiase	1.1	Covalent immobilization (pre-synthesised microgels)	PAAm-AA-GMA	35.1	80	78	Reusability: 71% after 8 reaction cycles	1
	Cellulase	1.1	Covalent immobilization (pre-synthesised microgels)	PAAm-AA-GMA	37.9	100	90	Reusability: 72% after 10 reaction cycles	1
	Cellulase	2.2	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains)	PVPS	-	-	24	-	2
	Cellulase	2.2	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains)	PVPS	41.9	87	~10-40	Organic solvent stability: 70% against acetone (free enzyme 37.5%) Chaotropic agent stability: 51.5% against urea (free enzyme 37.9%)	3
	Cellulase	2.2	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains)	PVGMA	-	-	39	Organic solvent stability: 87% against acetone (free enzyme 37%) Chaotropic agent stability: 81% against urea (free enzyme 38%)	4
	Cellulase	3.1	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains)	PVP-PDS	15.5	77	4.5-8.8	Stimuli-responsive enzyme release: 55% activity of free enzyme reconstituted after release	5
	Galactosidase (β -galactosidase)	3.2	Encapsulation during microgel synthesis (APS/TEMED initiation)	PNIPAAm-AAm	-	-	-	Regulation of enzyme activity by temperature-induced collapse and swelling of microgels in packed bed reactor. Increased mass transfer rate due to collapse/swelling cycles "pumping process"	6
	Galactosidase (β -galactosidase)	3.2	Encapsulation during microgel synthesis (APS/TEMED initiation)	PNIPAAm-AAm	-	-	-	Regulation of enzyme activity by temperature-induced collapse and swelling of microgels in packed bed reactor. Increased mass transfer rate due to collapse/swelling cycles "pumping process"	7
	Galactosidase (β -galactosidase)	3.2	Encapsulation during microgel synthesis (APS/TEMED initiation)	PNIPAAm	-	-	-	Regulation of enzyme activity by pressure-induced collapse and swelling of microgels in packed bed reactor. Increased mass transfer rate due to collapse/swelling cycles "pumping process"	8

Hydrolase	Galactosidase (β -galactosidase)	3.1	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains, enzymatic auto-oxidation of thiol groups)	Poly (glycidol) thiol-functionalised	21-41	40-44	83-85	Co-immobilisation with HRP	9
	Glucoamylase	3.1	Hydrophobic adsorption in the collapsed state of microgels followed by crosslinking with glutaraldehyde after adsorption	PNIPAAm-HEMA (aminated)	14.2	29	4.6	Storage stability: up to 100% activity after 30 days	10
	Glucosidase (β -D-glucosidase)	3.1	Adsorption (Hydrogenbond formation)	PNIPAAm with polystyrene core	620	-	325	-	11
	Lipase (CalB)	2.2	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains)	Poly(glycidol)	0.4 mg/mL	75	25	Esterification activity: 87 % (free enzyme 13%) Molecular weight of polymer: 4900 M_n /Da (free enzyme 1140 M_n /Da)	12
	Lipase (CalB)	3.1	Immobilisation by solvent exchange from polar to unpolar media	PNIPAAm	3	2.5	-	Esterification activity: ~0.03 U in hexane (free enzyme~0.001 U); ~30-fold improvement	13
	Lipase (LCR)	1.1	Covalent immobilization (pre-synthesised microgels)	PAAm functionalized with amine groups	1.13 mg/m ² of PVDF membrane	-	3.2	-	14
	Lysozyme	3.1	Adsorption (electrostatic interaction)	PNIPAAm-AA with polystyrene core	675	-	350	-	15
	Trypsin	1.1	Covalent immobilization (pre-synthesised microgels)	PNIPAAm-AA	493	-	155	Thermal stability: 70% after 3 h at 45 °C (free enzyme ~40%) Storage stability: 80% after 60 days (55% free enzyme)	16
	Trypsin	3.1	Hydrophobic adsorption in the collapsed state of microgels followed by crosslinking with glutaraldehyde after adsorption	PNIPAAm-HEMA (aminated)	0.3	44.2	18.6	Storage stability: up to 50% activity after 30 days	10
	Urease	4	Encapsulation during microgel synthesis (APS initiation)	PNIPAAm-VIm	16	-	-	Control of membrane permeability by urease induced microgel swelling collapse.	17

Hydrolase	α -Chymotrypsin	2.1	Encapsulation during microgel synthesis (APS/TEMED initiation)	PNIPAAm with Calcium-alginate as the polymerisation mould	558-2991	75-80	~10	Reusability: 75% after 30 reaction cycles (1 month)	18
Ligase	Acetyl-CoA synthetase	1.2	Covalent immobilization (pre-synthesised microgels)	PNIPAAm-AEMA	20	68	61	Storage stability: 80% after 9 days (63 % free enzyme) Thermal stability: 75% after 15 min at 60 °C (60 % free enzyme) Reusability: 50% after 4 cycles	19
	Acetyl-CoA synthetase	4	Adsorption (electrostatic interaction)	PNIPAAm-PEI	278	97	120	Reusability: 70 % after 7 cycles	20
Lyase	DERA	2.1	Encapsulation during microgel synthesis (enzyme also as crosslinker; covalent bonds to microgel network)	PNIPAAm-TIaAm	-	44	Strongly impaired	Storage stability: 100 % activity after lyophilisation	21
Oxidoreductase	Alcohol dehydrogenase	4	Encapsulation during microgel synthesis	PNIPAA-MAA PNIPAAm- PNIPMAAm	-	-	-	Stabilisation and heat-induced breaking of emulsions for catalysis	22
	Glucose oxidase	2.1	Encapsulation during microgel synthesis (APS/TEMED initiation)	PAAm-AA	-	-	-	Storage stability: 100% after 8 months	23
	Glucose oxidase	2.1	Encapsulation during microgel synthesis (enzyme initiated polymerisation)	PVCL	-	-	40	-	24
	Glucose oxidase	2.1	Encapsulation during microgel synthesis (enzyme initiated polymerisation, enzyme also as crosslinker; covalent bonds to microgel network)	PNIPAAm-TIaAm	-	31	4	-	21
	HRP	1.2	Covalent immobilization (pre-synthesised microgels)	PHEAA	22	-	87-96	Thermal stability: 76 % activity after 5.5 h at 50° C (free enzyme 20%)	25
	HRP	1.2	Covalent immobilization (pre-synthesised microgels)	PEGMA	44	-	33	Thermal stability 50 °C: 75% after 5 h (free enzyme 40%)	26
	HRP	2.1	Encapsulation during microgel synthesis (enzyme initiated polymerisation)	PDMAA	-	-	102	Storage stability: 98 % activity after 3 months Thermal stability: 33% after 30 min at 70°C (14% free enzyme)	27

Oxidoreductase	HRP	2.2	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains)	dPG	1.6	40	-	Thermal stability: 70% after 42 h at 50°C (free enzyme 10%) Reuseability: 50% after 6 cycles	28
	HRP	3.1	Non-covalent immobilisation Immobilisation by solvent exchange from polar to unpolar media	PNIPAAm	12	-	-	Organic solvent stability: 0.016 $\mu\text{mol min}^{-1} \mu\text{g}^{-1}$ in isopropanol (free enzyme 6.15*10 ⁻⁵ $\mu\text{mol min}^{-1} \mu\text{g}^{-1}$); ~260-fold improvement	29
	HRP	3.1	Encapsulation during microgel synthesis (crosslinking of reactive polymer chains, enzymatic auto-oxidation of thiol groups)	Poly(glycidol) thiol-functionalised	-	-	90	-	9
	Laccase (CueO)	1.2	Covalent immobilization (pre-synthesised microgels)	PVCL-GMA	64.2	-	126	Organic solvent stability: up to 3.8-fold improved activity in DMSO compared with free enzyme Decolorisation of indigo carmine : 79% decolorisation efficiency after 8 cycles Reusability: 55% after 11 cycles	30
	Laccase (Trametes vesicolor)	2.1	Encapsulation during microgel synthesis (APS/TEMED initiation)	PNIPAAm-VIm	-	35	-	-	31
	Laccase (Trametes vesicolor)	2.1	Encapsulation during microgel synthesis (APS/TEMED initiation)	PNIPAAm-AAm	-	96.4	100	Storage stability: 91% activity after 56 days (free enzyme 42%) Reusability: 78 % after 10 cycles	32
	Lactic acid dehydrogenase	1.1	Covalent immobilization (pre-synthesised microgels)	PNIPAAm-PEI	-	37	78	Thermal stability	33
	Cytochrome P450 BM3	1.2	Covalent immobilization (pre-synthesised microgels)	PVCL-GMA	89.6	-	22	Organic solvent stability: up to 4-fold improved activity in DMSO compared with free enzyme	30
Transferase	Pyruvate kinase	1.1	Covalent immobilisation (pre-synthesised microgels)	PNIPAm-PEI	-	20	223	Thermal stability Storage stability: ~70% after 14 days (free enzyme ~40 %)	33

1. P. A. Limadinata, A. Li and Z. Li, *Green Chem.*, 2015, **17**, 1194-1203.
2. H. Peng, M. Kather, K. Rübsam, F. Jakob, U. Schwaneberg and A. Pich, *Macromolecules*, 2015, **48**, 4256-4268.
3. H. Peng, K. Rübsam, F. Jakob, U. Schwaneberg and A. Pich, *Biomacromolecules*, 2016, **17**, 3619-3631.
4. H. Peng, K. Rübsam, C. Hu, F. Jakob, U. Schwaneberg and A. Pich, *Biomacromolecules*, 2019, **20**, 992-1006.
5. H. Peng, K. Rübsam, F. Jakob, P. Pazdziior, U. Schwaneberg and A. Pich, *Macromol. Rapid Commun.*, 2016, **37**, 1765-1771.
6. T. G. Park and A. S. Hoffman, *Appl. Biochem. Biotechnol.*, 1988, **19**, 1-9.
7. T. G. Park and A. S. Hoffman, *J. Biomed. Mater. Res.*, 1990, **24**, 21-38.
8. Y. Wang, X. Zhong and S. Wang, *J. Chem. Technol. Biotechnol.*, 1996, **67**, 243-247.
9. S. Singh, F. Topuz, K. Hahn, K. Albrecht and J. Groll, *Angew. Chem. Int. Ed.*, 2013, **52**, 3000-3003.
10. A. Hamerska-Dudra, J. Bryjak and A. W. Trochimczuk, *Enzyme Microb. Technol.*, 2007, **41**, 197-204.
11. N. Welsch, A. Wittemann and M. Ballauff, *J. Phys. Chem. B*, 2009, **113**, 16039-16045.
12. S. Engel, H. Höck, M. Bocola, H. Keul, U. Schwaneberg and M. Möller, *Polymers*, 2016, **8 (10)**, 372.
13. K. Gawlitza, C. Wu, R. Georgieva, M. Ansorge-Schumacher and R. von Klitzing, *Z. Phys. Chem.*, 2012, **226**, 749-759.
14. G. Vitola, D. Buning, J. Schumacher, R. Mazzei, L. Giorno and M. Ulbricht, *Macromol. Biosci.*, 2017, **17**, 1600381.
15. N. Welsch, A. L. Becker, J. Dzubiella and M. Ballauff, *Soft Matter*, 2012, **8**, 1428-1436.
16. E. Lai, Y. Wang, Y. Wei, G. Li and G. Ma, *J. Appl. Polym. Sci.*, 2016, **133**, 43343.
17. K. Ogawa, B. Wang and E. Kokufuta, *Langmuir*, 2001, **17**, 4704-4707.
18. M. Bayhan and A. Tuncel, *J. Appl. Polym. Sci.*, 1998, **67**, 1127-1139.
19. N. C. Dubey, B. P. Tripathi, M. Stamm and L. Ionov, *Biomacromolecules*, 2014, **15**, 2776-2783.
20. N. C. Dubey, B. P. Tripathi, M. Muller, M. Stamm and L. Ionov, *ACS Appl. Mater. Interfaces*, 2015, **7**, 1500-1507.
21. S. Reinicke, T. Fischer, J. Bramski, J. Pietruszka and A. Böker, *RSC Adv.*, 2019, **9**, 28377-28386.
22. S. Wiese, A. C. Spiess and W. Richtering, *Angew. Chem. Int. Ed.*, 2013, **52**, 576-579.
23. E. L. Cabarcos, J. R. Retama and B. Lopez-Ruiz, in *Trends in Colloid and Interface Science XVII. Progress in Colloid and Polymer Science*, Springer, Berlin, Heidelberg, 2004, vol. 126, ch. 42, pp. 194-196.
24. E. Gau, F. Flecken, A. N. Ksiazkiewicz and A. Pich, *Green Chem.*, 2018, **20**, 431-439.
25. M. S. Tehrani, Y. Lu, G. Guerin, M. Soleimani, D. Pichugin and M. A. Winnik, *Biomacromolecules*, 2015, **16**, 3134-3144.
26. S. M. Tehrani, Y. Lu and M. A. Winnik, *Macromolecules*, 2016, **49**, 8711-8721.
27. S. Bao, D. Wu, T. Su, Q. Wu and Q. Wang, *RSC Adv.*, 2015, **5**, 44342-44345.
28. C. Wu, C. Böttcher and R. Haag, *Soft Matter*, 2015, **11**, 972-980.
29. K. Gawlitza, R. Georgieva, N. Tavraz, J. Keller and R. von Klitzing, *Langmuir*, 2013, **29**, 16002-16009.
30. Z. Zou, E. Gau, I. El-Awaad, F. Jakob, A. Pich and U. Schwaneberg, *Bioconj. Chem.*, 2019, **30**, 2859-2869.
31. S. Schachschal, H.-J. Adler, A. Pich, S. Wetzel, A. Matura and K.-H. van Pee, *Colloid Polym. Sci.*, 2011, **289**, 693-698.
32. O. Yamak, N. A. Kalkan, S. Aksoy, H. Altinok and N. Hasirci, *Process Biochem.*, 2009, **44**, 440-445.
33. N. C. Dubey, B. P. Tripathi, M. Muller, M. Stamm and L. Ionov, *Biomacromolecules*, 2016, **17**, 1610-1620.