## **Supplementary Information**

## Selective electrocatalytic hydroboration of aryl alkenes

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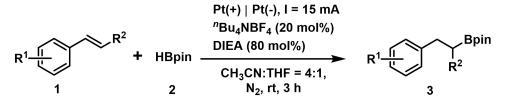
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#### **I. General Information**

**Materials**. All manipulations were carried out under argon atmosphere by using standard Schlenk techniques and Mikrouna argon-filled glove box. All solvents were dried and distilled over appropriate drying agents under argon. Various alkenes (aladdin), pinacolborane (HBpin, Aldrich), "Bu<sub>4</sub>NBF<sub>4</sub> (aladdin), NaO'Bu (aladdin), 2,2',6,6'-tetramethyl-1-piperidinyloxy (TEMPO, aladdin), galvinoxyl free radical (Aldrich),  $d_8$ -styrene (Aldrich), 5,5-dimethyl-1-pyrroline *N*-oxide (DMPO, aladdin), triethylamine (Et<sub>3</sub>N, aladdin), *N*,*N*-diisopropylethylamine (DIEA, aladdin), NaBD<sub>4</sub> (Heowns), pinacol (Heowns), allyl bromide (Allyl-Br, aladdin) and NaBO<sub>3</sub> • 4H<sub>2</sub>O (Energy Chemical) were obtained from commercial suppliers and used without further purification. THF- $d_8$  and CD<sub>3</sub>CN were purchased from Alfa Aesar and used with dried with 4Å molecular sieve. DBpin was prepared according to literature procedures.<sup>1-2</sup>

**Instruments.** Instrument for electrolysis was dual display potentiostat (CHI760E) (Shanghai Chenhua). The anode and cathode electrodes were all platinum electrodes (15 mm × 15 mm × 0.3 mm), which were purchased from Shanghai Vietnamese Magnetic Electronics. Gas chromatographic (GC) analyses were performed on an Agilent GC-8860 gas chromatography instrument with a FID detector and naphthalene was added as internal standard. Hydrogen detection experiments were recorded on a SHIMADZU GC-2014 gas chromatography instrument. <sup>1</sup>H, <sup>2</sup>D, <sup>11</sup>B, <sup>13</sup>C and <sup>19</sup>F NMR spectra were recorded on a Brüker 400 M Ultra Shield spectrometer. Chemical shifts ( $\delta$ ) were given in parts per million relative to CDCl<sub>3</sub> (7.26 ppm for <sup>1</sup>H; 77.16 ppm for <sup>13</sup>C). ESI-HRMS data were recorded on a HPLC/Q-Tof mass spectrometer. EPR spectra were recorded at room temperature on a JEOL JES-FE3AX spectrometer. Infrared spectra were recorded on a NICOLET iS50 ATR spectrometer. Molecular structure of compound **7t** was obtained on a Brüker SMART APEX CCD diffractometer with graphite monochromated Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å).

### **II. Experimental Procedures and Analytical Data**



General procedures for alkene hydroboration with pinacolborane (HBpin) (3a–3h). In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, alkenes (1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h, and the set-up diagrams are shown in Figure S1a–1c. After the solution of the crude products were concentrated in vacuum, the pure products (**3a–3h**) were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.

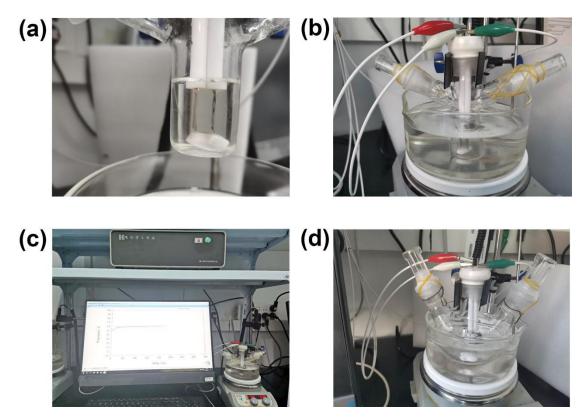
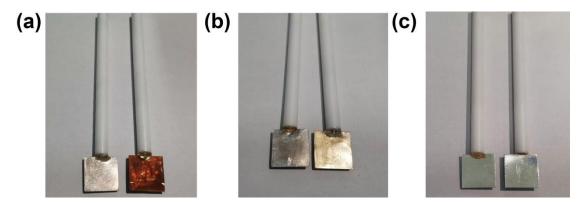
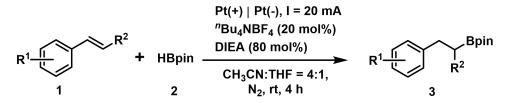


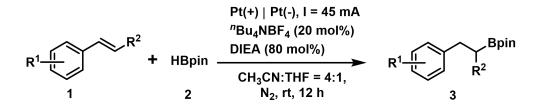
Figure S1. (a, b, c) Set-up diagrams of undivided cell electrolysis. (d) Device for gram scale synthesis.



**Figure S2.** Varying degrees of attachment of unknown yellow compounds on the anode surface after the electrolyzed reaction between styrene and HBpin under different conditions. (a) In the absence of THF and DIEA. (b) In the absence of DIEA. (c) Under the standard conditions. The cathode is in the left and the anode is on the right.

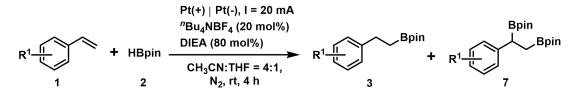


General procedures for alkene hydroboration with HBpin (3i–3r). In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm  $\times$  15 mm  $\times$  0.3 mm) as both anode and cathode, alkenes (1.0 mmol), HBpin (160 µL, 1.1 mmol), DIEA (132 µL, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 20 mA constant current under room temperature for 4 h, and the set-up diagrams are shown in Figure S1a–1c. After the solution of the crude products were concentrated in vacuum, the pure products (**3i–3r**) were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.

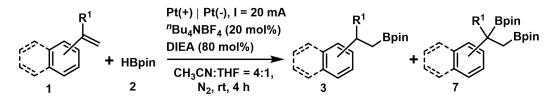


General procedures for gram scale synthesis (3a, 3b, 3d, 3e, 3g and 3h). In an

over-dried undivided three-neck flask (250 mL) equipped with two platinum electrodes (15 mm  $\times$  15 mm  $\times$  0.3 mm) as both anode and cathode, alkenes (10 mmol), HBpin (1.6 mL, 11 mmol), DIEA (1.32 mL, 8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (658 mg, 2 mmol), CH<sub>3</sub>CN (80 mL) and THF (20 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 45 mA constant current under room temperature for 12 h, and the set-up diagram is shown in Figure S1d. After the solution of the crude products were concentrated in vacuum, the pure products were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



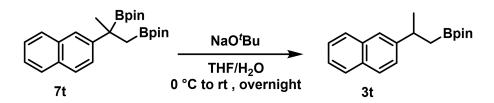
General procedures for alkene diborylation with HBpin (7a, 7b and 7g). In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm  $\times$  15 mm  $\times$  0.3 mm) as both anode and cathode, alkenes (1.0 mmol), HBpin (1280 µL, 8.8 mmol), DIEA (132 µL, 0.8 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 20 mA constant current under room temperature for 4 h, and the set-up diagrams are shown in Figure S1a-1c. After the solution of the crude products were concentrated in vacuum, the pure products were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



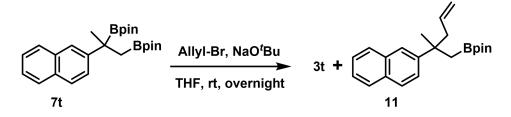
General procedures for alkene diborylation with HBpin (7r–7t). In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, alkenes (1.0 mmol), HBpin (640  $\mu$ L, 4.4 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF

(2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 20 mA constant current under room temperature for 4 h, and the set-up diagrams are shown in Figure S1a–1c. After the solution of the crude products were concentrated in vacuum, the pure products (7r-7t) were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.

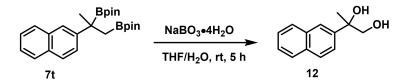
General procedures for the transformations of diboronate ester 7t.



A Schlenk tube (10 mL) equipped with a magnetic stir bar was added with diboronate ester **7t** (84 mg, 0.20 mmol), NaO'Bu (19 mg, 0.20 mmol) and THF (1.0 mL) at 0 °C, followed by three drops of water. The resulting solution was warming to room temperature overnight. Then the mixture was diluted with  $CH_2Cl_2$ , dried with anhydrous Na<sub>2</sub>SO<sub>4</sub> and filtered with diatomite. After the solution was concentrated in vacuum, the pure product **3t** (36 mg, 0.12 mmol, 61% yield) was obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



In an argon-filled glove-box, a Schlenk tube (10 mL) equipped with a magnetic stir bar was added with diboronate ester **7t** (84 mg, 0.20 mmol), NaO'Bu (192 mg, 2.00 mmol), Allyl-Br (86  $\mu$ L, 1.00 mmol) and anhydrous THF (1.0 mL) at room temperature, then the mixture was stirred overnight. Then the mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> and filtered with diatomite. After the solution was concentrated in vacuum, the pure product **3t** (17 mg, 0.06 mmol, 29% yield) and **11** (30 mg, 0.09 mmol, 45% yield) were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



A Schlenk tube (10 mL) equipped with a magnetic stir bar was added with diboronate ester **7t** (127 mg, 0.30 mmol), NaBO<sub>3</sub>•4H<sub>2</sub>O (370 mg, 2.40 mmol), THF (3.0 mL) and H<sub>2</sub>O (3.0 mL) at room temperature, then the resulting solution was stirred for 5 h and quenched with saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> aqueous solution. Then the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried with anhydrous Na<sub>2</sub>SO<sub>4</sub> and filtered with diatomite. After the solution was concentrated in vacuum, the pure product **12** (57 mg, 0.28 mmol, 94% yield) was obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.

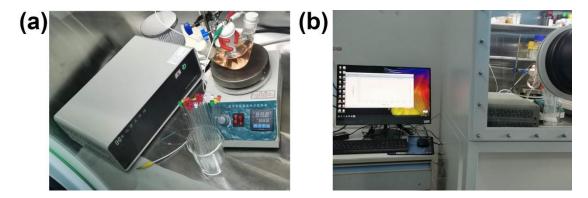
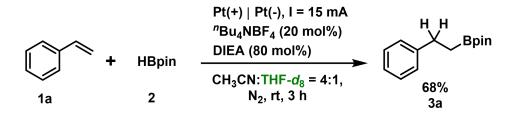
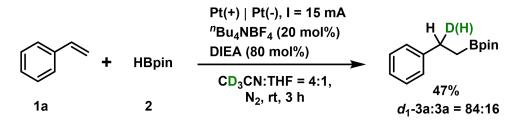


Figure S3. (a, b) Set-up diagrams for *in situ* <sup>1</sup>H NMR experiments.

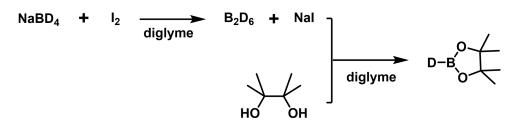
*In situ* <sup>1</sup>H NMR experiments to monitor the electrochemical reaction between styrene and HBpin. In an over-dried undivided three-neck flask (250 mL) equipped with two platinum electrodes (15 mm  $\times$  15 mm  $\times$  0.3 mm) as both anode and cathode, styrene (1.14 mL, 10 mmol), HBpin (11 mmol or 88 mmol), DIEA (1.32 mL, 8 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (658 mg, 2 mmol), CH<sub>3</sub>CN (80 mL) and THF (20 mL) were added in an argon-filled glove box. Then the reaction mixture was stirred and electrolyzed at a 45 mA constant current under room temperature for 12 h, and the set-up diagram is shown in Figure S3. The <sup>1</sup>H NMR spectra were collected every 0.5 h. The test samples were made up of 0.3 mL original samples and 0.2 mL CD<sub>3</sub>CN, and the sample collection was completed in the glove box.



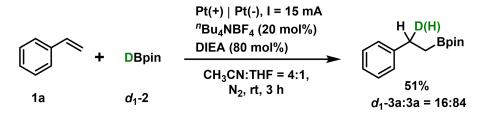
**Deuterium-labelling experiment in THF-***d*<sub>8</sub>. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF-*d*<sub>8</sub> (2 mL) were added under argon atmosphere. Then the mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. After the solution of the crude product was concentrated in vacuum, the pure product **3a** was obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



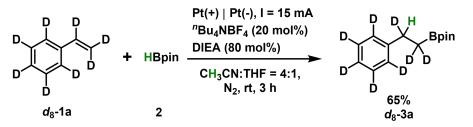
**Deuterium-labelling experiment in CD**<sub>3</sub>**CN.** In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CD<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. After the solution of the crude products were concentrated in vacuum, the mixed products *d*<sub>1</sub>-3a and 3a were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



**Preparation of DBpin.** The procedures were conducted the literature method.<sup>1-2</sup> A suspension solution of NaBD<sub>4</sub> (0.5 g, 12.2 mmol) in diglyme (10 mL) was added to the Schlenk tube equipped with a magnetic stir bar in an argon atmosphere. Then the above tube was connected to the second Schlenk tube with the cooled solution (0 °C) of pinacol (0.48 g, 4.08 mmol) in absolute THF (5 mL) *via* a plastic cannula, and the cannula submersed in THF solution. Iodine (1.55 g, 6.11 mmol) was dissolved in diglyme (6 mL) and the solution was slowly added to the NaBD<sub>4</sub> suspension over 1 h with a syringe. At the end of the addition of iodine, stream of N<sub>2</sub> was pass through the THF solution for about 2 h to remove the excess unreacted B<sub>2</sub>D<sub>6</sub> at room temperature. <sup>1</sup>H NMR data confirmed that the transformation of pinacol was completed. The 3.0 mL of THF solution was considered to be concentrated at 1.36 M DBpin.



**Deuterium-labelling experiment with DBpin.** In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), DBpin (0.8 mL, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (1.4 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. After the solution of the crude products were concentrated in vacuum, the mixed products *d*<sub>1</sub>-3a and 3a were obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.



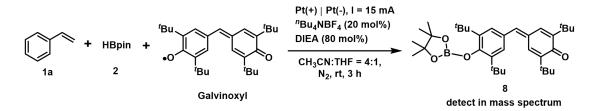
**Deuterium-labelling experiment with**  $d_8$ -styrene. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode,  $d_8$ -styrene (114 µL, 1.0 mmol), HBpin (160 µL, 1.1 mmol), DIEA (132 µL, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. After the solution of the crude product was concentrated in vacuum, the pure product  $d_8$ -3a was obtained by flash chromatography on silica gel using petroleum and ethyl acetate as eluent.

H<sub>2</sub> detection experiment under the standard conditions. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. A large number of tiny bubbles were observed during electrolysis and the release rate (mL·min<sup>-1</sup>) of gas with time was analyzed by SHIMADZU GC-2014 gas chromatography instrument.

H<sub>2</sub> detection experiment to monitor the electrochemical reaction in the absence of styrene and HBpin. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. The release rate (mL·min<sup>-1</sup>) of gas during the electrochemical reaction with time was analyzed by SHIMADZU GC-2014 gas chromatography instrument.  $H_2$  detection experiment to monitor the electrochemical reaction in the absence of styrene, HBpin, DIEA and THF. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol) and CH<sub>3</sub>CN (8 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. The release rate (mL·min<sup>-1</sup>) of gas during the electrochemical reaction with time was analyzed by SHIMADZU GC-2014 gas chromatography instrument.

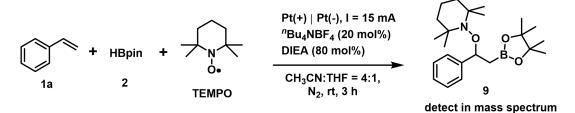
H<sub>2</sub> detection experiment to monitor the electrochemical reaction in the absence of DIEA. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. The release rate (mL·min<sup>-1</sup>) of gas during electrolysis with time was analyzed by SHIMADZU GC-2014 gas chromatography instrument.

H<sub>2</sub> detection experiment to monitor the electrochemical reaction in the absence of styrene, HBpin and DIEA. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, *n*Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h. The release rate (mL·min<sup>-1</sup>) of gas during the electrochemical reaction with time was analyzed by SHIMADZU GC-2014 gas chromatography instrument.





three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), galvinoxyl radical (422 mg, 1.0 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h, and the yield of product **3a** was determined by gas chromatography analysis. The adduct **8** of galvinoxyl radical and boron radical was determined by mass spectrometry.



Radical inhibition experiment with 2,2',6,6'-tetramethyl-1-piperidinyloxy (TEMPO). In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then different equivalents (0.25, 0.50, 0.75 or 1.0 mmol) of TEMPO were added to this system, respectively. The reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature for 3 h, and the yield of product **3a** was determined by gas chromatography analysis. The adduct **9** of carbon radical at the benzyl site with TEMPO was determined by mass spectrometry.

The radical inhibition experiment using 2-vinylphthalene as substrate was similar to the above method. The adducts of boron radical (namely compound **13**) and carbon radical (namely compound **14**) at the benzyl site with TEMPO were determined by gas chromatography-mass spectrometry and mass spectrometry, respectively.

**Procedures for cyclic voltammetry (CV) in CH<sub>3</sub>CN.** Cyclic voltammetry experiments were conducted in a 10 mL three-electrode cell equipped with a glassy carbon working electrode, a Ag/AgCl reference electrode and a platinum wire counter

electrode, and the reference electrode was submerged in a saturated aqueous KCl solution. The CV experiments were performed in CH<sub>3</sub>CN (10 mL) with "Bu<sub>4</sub>NBF<sub>4</sub> (32.9 mg, 0.1 mmol). The scan rate was 100 mV·S<sup>-1</sup> and the potential range was 0-5 V. The current was reported in mA and potential were reported in V against  $Fc^{+/0}$  redox couple.

**Procedures for cyclic voltammetry (CV) of DIEA in CH<sub>3</sub>CN.** Cyclic voltammetry experiments were conducted in a 10 mL three-electrode cell equipped with a glassy carbon working electrode, a Ag/AgCl reference electrode and a platinum wire counter electrode, and the reference electrode was submerged in a saturated aqueous KCl solution. The CV experiments were performed in CH<sub>3</sub>CN (10 mL) containing  $^{n}$ Bu<sub>4</sub>NBF<sub>4</sub> (32.9 mg, 0.1 mmol) and DIEA (16.5 µL, 0.1 mmol). The scan rate was 100 mV·S<sup>-1</sup> and the potential range was 0-5 V. The current was reported in mA and potential were reported in V against Fc<sup>+/0</sup> redox couple.

Procedures for cyclic voltammetry (CV) of HBpin in CH<sub>3</sub>CN. Cyclic voltammetry experiments were conducted in a 10 mL three-electrode cell equipped with a glassy carbon working electrode, a Ag/AgCl reference electrode and a platinum wire counter electrode, and the reference electrode was submerged in a saturated aqueous KCl solution. The CV experiments were performed in CH<sub>3</sub>CN (10 mL) containing  $^{n}$ Bu<sub>4</sub>NBF<sub>4</sub> (32.9 mg, 0.1 mmol) and HBpin (14.5 µL, 0.1 mmol). The scan rate was 100 mV·S<sup>-1</sup> and the potential range was 0-5 V. The current was reported in mA and potential were reported in V against Fc<sup>+/0</sup> redox couple.

**EPR experiment under the standard conditions.** In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), DMPO (113 mg, 1.0 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature. After 30 min, the solution was taken out and injected into a capillary tube with one end closed, then the solution was analyzed by EPR spectrometer at room temperature. Only one type of racial had been trapped by

DMPO: DMPO–H: g =2.0071, *A*<sub>N</sub> = 14.74 G, *A*<sub>H</sub> = 19.74 G.

#### EPR experiment to monitor the electrochemical reaction in the absence of HBPin.

In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, styrene (114  $\mu$ L, 1.0 mmol), DIEA (132  $\mu$ L, 0.8 mmol), DMPO (113 mg, 1.0 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature. After 30 min, the solution was taken out and injected into a capillary tube with one end closed, then the solution was analyzed by EPR spectrometer at room temperature. Two type of radicals had been trapped by DMPO: DMPO–H and DMPO–CH<sub>2</sub>CN (**10**, g = 2.0052,  $A_N = 14.47$  G,  $A_H = 20.82$  G).

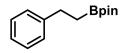
EPR experiment to monitor the electrochemical reaction in the absence of styrene. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, HBpin (160  $\mu$ L, 1.1 mmol), DIEA (132  $\mu$ L, 0.8 mmol), DMPO (113 mg, 1.0 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature. After 30 min, the solution was taken out and injected into a capillary tube with one end closed, then the solution was analyzed by EPR spectrometer at room temperature. Only DMPO–H was detected.

EPR experiment to monitor the electrochemical reaction in the absence of styrene and HBpin. In an over-dried undivided three-neck flask equipped with two platinum electrodes (15 mm × 15 mm × 0.3 mm) as both anode and cathode, DIEA (132  $\mu$ L, 0.8 mmol), DMPO (113 mg, 1.0 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature. After 30 min, the solution was taken out and injected into a capillary tube with one end closed, the the solution was analyzed by EPR spectrometer at room temperature. Two type of radicals had been trapped by DMPO: DMPO–H and **10**.

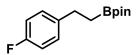
EPR experiment to monitor the electrochemical reaction in the absence of

**styrene, HBpin and DIEA.** In an over-dried undivided three-neck flask equipped with two platinum electrodes ( $15 \text{ mm} \times 15 \text{ mm} \times 0.3 \text{ mm}$ ) as both anode and cathode, DMPO (113 mg, 1.0 mmol), "Bu<sub>4</sub>NBF<sub>4</sub> (65.8 mg, 0.2 mmol), CH<sub>3</sub>CN (8 mL) and THF (2 mL) were added under argon atmosphere. Then the reaction mixture was stirred and electrolyzed at a 15 mA constant current under room temperature. After 30 min, the solution was taken out and injected into a capillary tube with one end closed, then the solution was analyzed by EPR spectrometer at room temperature. Two type of radicals had been trapped by DMPO: DMPO–H and **10**. The reduction state of the corresponding adduct **10** was detected by mass spectrometry.

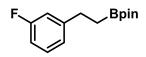
#### **III. Spectroscopic Data of Products**



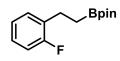
4,4',5,5'-Tetramethyl-2-phenethyl-1,3,2-dioxaborolane (3a)<sup>3</sup>: Colorless oil was obtained in 70% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.19-7.24 (m, 4H), 7.12-7.15 (m, 1H), 2.74 (t, J = 8.0 Hz, 2H), 1.20 (s, 12H), 1.13 (t, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 144.5, 128.3, 128.1, 125.6, 83.2, 30.1, 24.9.
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 34.02.



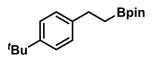
**2-(4-Fluorophenethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane** (3b)<sup>3</sup>: Colorless oil was obtained in 69% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.14-7.17 (m, 2H), 6.91-6.95 (m, 2H), 2.71 (t, *J* = 8.0 Hz, 2H), 1.21 (s, 12H), 1.11 (t, *J* = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  162.4 (d, *J* = 241.0 Hz), 140.1 (d, *J* = 3.0 Hz), 129.5 (d, *J* = 8.0 Hz), 115.0 (d, *J* = 21.0 Hz), 83.3, 29.3, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.61. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  – 118.37.



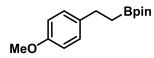
**2-(3-Fluorophenethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane** (3c)<sup>4</sup>: Colorless oil was obtained in 61% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.17-7.22 (m, 1H), 6.97-6.99 (m, 1H), 6.91-6.94 (m, 1H), 6.81-6.85 (m, 1H), 2.75 (t, *J* = 8.0 Hz, 2H), 1.22 (s, 12H), 1.13 (t, *J* = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  164.2 (d, *J* = 243.0 Hz), 147.1 (d, *J* = 7.0 Hz), 129.6 (d, *J* = 8.0 Hz), 123.8 (d, *J* = 2.0 Hz), 115.1 (d, *J* = 21.0 Hz), 112.5 (d, *J* = 21.0 Hz), 83.3, 29.8, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.77. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  – 114.20.



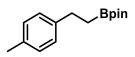
4,4',5,5'-Tetramethyl-2-(2-fluorophenylethyl)-1,3,2-dioxaborolane (3d)<sup>5</sup>: Colorless oil was obtained in 66% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.21-7.25 (m, 1H), 7.10-7.16 (m, 1H), 7.01-7.05 (m, 1H), 6.95-7.00 (m, 1H), 2.78 (t, J= 8.0 Hz, 2H), 1.22 (s, 12H), 1.15 (t, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  162.4 (d, J = 243.0 Hz), 131.3 (d, J = 16.0 Hz), 130.2 (d, J = 6.0 Hz), 127.3 (d, J = 8.0 Hz), 123.9 (d, J = 3.0 Hz), 115.2 (d, J = 22.0 Hz), 83.2, 24.9, 23.3 (d, J = 2.0 Hz). <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.77. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ -118.78.



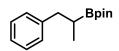
**2-(4-(Tert-Butyl)phenethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane**(3e)<sup>5</sup>:Colorless oil was obtained in 74% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.28-7.30 (m, 2H), 7.14-7.16 (m, 2H), 2.72 (t, J = 8.0 Hz, 2H), 1.30 (s, 9H), 1.22 (s, 12H), 1.14 (t, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  148.4, 141.5, 127.7, 125.2, 83.2, 34.5, 31.6, 29.5, 25.0. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.91.



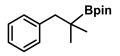
**2-(4-Methoxyphenethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (3f)<sup>3</sup>:** Colorless oil was obtained in 69% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.14 (d, *J* = 8.0 Hz, 2H), 6.82 (d, *J* = 8.0 Hz, 2H), 3.78 (s, 3H), 2.69 (t, *J* = 8.0 Hz, 2H), 1.22 (s, 12H), 1.12 (t, *J* = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  157.8, 136.7, 129.0, 113.7, 83.2, 55.4, 29.2, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.79.



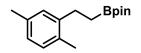
4,4',5,5'-Tetramethyl-2-(4-methylphenethyl)-1,3,2-dioxaborolane (3g)<sup>3</sup>: Colorless oil was obtained in 77% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ7.09 (q, J = 8.0 Hz, 4H), 2.73 (t, J = 8.0 Hz, 2H), 2.32 (s, 3H), 1.25 (s, 12H), 1.14 (t, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ141.5, 134.9, 129.0. 128.0, 83.2, 29.6, 24.9, 21.1. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ34.09.



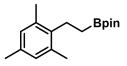
4,4',5,5'-Tetramethyl-2-(1-phenylpropan-2-yl)-1,3,2-dioxaborolane(3h)<sup>3</sup>:Colorless oil was obtained in 71% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.13-7.24 (m, 5H), 2.81 (dd, J = 8.0, 12.0 Hz, 1H), 2.55 (dd, J = 8.0, 12.0 Hz, 1H),1.33-1.38 (m, 1H), 1.19 (s, 6H), 1.18 (s, 6H), 0.97 (d, J = 8.0 Hz, 3H). <sup>13</sup>C NMR (100MHz, CDCl<sub>3</sub>, ppm):  $\delta$  142.5, 129.0, 128.1, 125.7, 83.1, 39.1, 24.8, 15.3. <sup>11</sup>B NMR(128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.31.



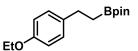
4,4',5,5'-Tetramethyl-2-(2-methyl-1-phenylpropan-2-yl)-1,3,2-dioxaborolane (3i)<sup>6</sup>: Colorless oil was obtained in 57% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.14-7.23 (m, 5H), 2.62 (s, 2H), 1.22 (s, 12H), 0.95 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): *δ* 140.6, 130.3, 127.8, 125.8, 83.2, 46.5, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): *δ* 34.60.



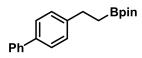
2-(2,5-Dimethylphenethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (3j)<sup>5</sup>: Colorless oil was obtained in 70% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.03 (d, J = 8.0 Hz, 2H), 6.90-6.91 (m, 1H), 2.70 (t, J = 8.0 Hz, 2H), 2.30 (s, 3H), 2.29 (s, 3H), 1.26 (s, 12H), 1.11 (t, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  142.4, 135.2, 132.6, 130.0, 129.1, 126.4, 83.2, 27.3, 24.9, 21.1, 18.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.03.



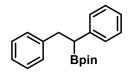
4,4',5,5'-Tetramethyl-2-(2,4,6-trimethylphenethyl)-1,3,2-dioxaborolane (3k)<sup>4</sup>: Pale yellow solid was obtained in 67% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  6.82 (s, 2H), 2.68 (t, *J* = 8.0 Hz, 2H), 2.30 (s, 6H), 2.25 (s, 3H), 1.28 (s, 12H), 0.96 (t, *J* = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  138.6, 135.8, 134.8, 128.9, 83.2, 25.0, 23.4, 20.9, 19.8. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.84.



**2-(4-Ethoxyphenethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (3l):** Colourless oil was obtained in 61% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.11 (d, *J* = 8.4 Hz, 2H), 6.79 (d, *J* = 8.8 Hz, 2H), 3.99 (q, *J* = 6.8 Hz, 2H), 2.70 (t, *J* = 8.0 Hz, 2H), 1.39 (t, *J* = 6.8 Hz, 3H), 1.22 (s, 12H), 1.13 (t, *J* = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  157.0, 136.5, 128.9, 114.3, 83.0, 63.4, 29.1, 24.9, 15.0. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.03. FT-IR (ATR): 3031, 2978, 2927, 2871, 1612, 1511, 1370, 1319, 1239, 1143, 1049, 967, 850, 804, 521. ESI-HRMS (*m/z*): cacld. for [**3l** + H]<sup>+</sup>: 276.2006; found: 276.2006.

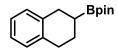


(2-([1,1'-Biphenyl]-4-yl)ethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (3m)<sup>4</sup>: White solid was obtained in 57% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.58-7.60 (m, 2H), 7.51-7.54 (m, 2H), 7.43 (t, J = 8.0 Hz, 2H), 7.30-7.35 (m, 3H), 2.81 (t, J = 8.0 Hz, 2H), 1.25 (s, 12H), 1.20 (t, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  143.7, 141.4, 138.6, 128.8, 128.6, 127.1, 127.1, 127.0, 83.3, 29.7, 25.0. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.01.



(1,2-Diphenylethyl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (3n)<sup>5</sup>: Colorless oil was obtained in 65% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.12-7.25 (m, 10H), 3.12-3.18 (m, 1H), 2.94-2.99 (m, 1H), 2.66-2.70 (m, 1H), 1.11 (s, 6H), 1.10 (m, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 142.7, 141.9, 129.0, 128.5, 128.5, 128.2, 125.9, 125.5, 83.5, 39.0, 24.7, 24.6. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 33.08.

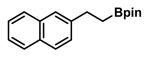
(2,3-Dihydro-1H-inden-2-yl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (30)<sup>3</sup>: Colorless oil was obtained in 65% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.21-7.23 (m, 2H), 7.11-7.13 (m, 2H), 2.95-3.11 (m, 4H), 1.84-1.94 (m, 1H), 1.27 (s, 12H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  144.5, 126.0, 124.3, 83.4, 35.3, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.56.



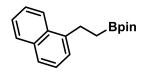
**4,4',5,5'-Tetramethyl-2-(1,2,3,4-tetrahydro-2-naphthalenyl)-1,3,2-dioxaborolane (3p)<sup>3</sup>:** Colorless oil was obtained in 64% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.09 (s, 4H), 2.75-2.92 (m, 4H), 2.04-2.07 (m, 1H), 1.64-1.74 (m, 1H), 1.35-1.41 (m, 1H), 1.29 (s, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 137.4, 136.9, 129.0, 128.9, 125.3, 83.0, 30.6, 29.7, 24.8, 24.7, 24.6. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 34.08.



(1,2-Dihydroacenaphthylen-1-yl)-4,4',5,5'-tetramethyl-1,3,2-dioxaborolane (3q)<sup>7</sup>: Pale yellow oil was obtained in 84% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.56-7.60 (m, 2H), 7.42-7.46 (m, 2H), 7.34-7.35 (m, 1H), 7.28-7.30 (m, 1H), 3.55 (d, J = 6.4 Hz, 2H), 3.30 (t, J = 6.8 Hz, 1H), 1.27 (d, J = 7.2 Hz, 12H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  147.5, 146.5, 139.2, 131.9, 128.0, 127.8, 122.2, 121.8, 119.2, 119.0, 83.8, 33.2, 25.1, 24.7, 24.7. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.34.

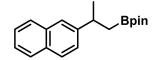


4,4',5,5'-Tetramethyl-2-(2-(naphthalen-2-yl)ethyl)-1,3,2-dioxaborolane (3r)<sup>8</sup>: Colourless oil was obtained in 6% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.74-7.80 (m, 3H), 7.45-7.65 (m, 1H), 7.36-7.45 (m, 3H), 2.92 (t, J = 8 Hz, 2H), 1.24 (m, 2H), 1.22 (s, 12H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  142.1, 133.8, 132.1, 127.8, 127.7, 127.6, 127.4, 125.9, 125.8, 125.0, 83.3, 30.4, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.79.

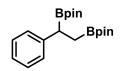


4,4',5,5'-Tetramethyl-2-(2-(naphthalen-1-yl)ethyl)-1,3,2-dioxaborolane (3s)<sup>3</sup>:

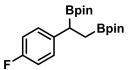
Colourless oil was obtained in 3% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 8.13 (d, *J* =8.4 Hz, 1H), 7.87 (d, *J* = 7.6 Hz, 1H), 7.56-7.73 (m, 1H), 7.42-7.48 (m, 2H), 3.27 (t, *J* = 8.0 Hz, 2H), 1.35 (t, *J* = 8.0 Hz, 2H), 1.28 (s, 12H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  140.5, 133.9, 131.9, 128.8, 126.4, 125.7, 125.7, 125.4, 125.1, 124.1, 83.2, 27.1, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.08.



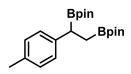
4,4',5,5'-Tetramethyl-2-(2-(naphthalen-2-yl)propyl)-1,3,2-dioxaborolane (3t)<sup>3</sup>: Colourless oil was obtained in 2% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 7.76-7.79 (m, 3H), 7.67 (s, 1H), 7.39-7.46 (m, 3H), 3.20-3.26 (m, 1H), 1.39 (d, J =6.8 Hz, 3H), 1.27-1.29 (m, 2H), 1.16 (s, 12H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 146.8, 133.7, 132.2, 127.9, 127.7, 127.7, 126.0, 125.8, 125.1, 124.5, 83.1, 36.0, 24.9, 24.9, 24.8. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.03.



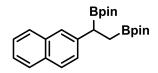
**2,2'-(1-Phenylethane-1,2-diyl)bis(4,4',5,5'-tetramethyl-1,3,2-dioxaborolane) (7a)**<sup>9</sup>: Colourless oil was obtained in 41% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.22 (d, J = 4.4 Hz, 4H), 7.07-7.11 (m, 1H), 2.51 (dd, J = 5.6, 10.8 Hz, 1H), 1.38 (dd, J = 11.2, 14.8 Hz, 1H), 1.20 (s, 12H), 1.19 (s, 6H), 1.17 (s, 6H), 1.13 (dd, J = 6.0, 16.4 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  145.5, 128.3, 128.0, 125.0, 83.3, 83.2, 25.1, 24.8, 24.8, 24.6. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.03.



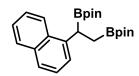
2,2'-(1-(4-Fluorophenyl)ethane-1,2-diyl)bis(4,4',5,5'-tetramethyl-1,3,2-dioxaborol ane) (7b)<sup>10</sup>: Colourless oil was obtained in 43% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.15-7.18 (m, 2H), 6.88-6.92 (m, 1H), 2.50 (dd, J = 6.0, 10.4 Hz, 1H), 1.32 (dd, J = 10.8, 17.2 Hz, 1H), 1.19 (s, 12H), 1.18 (s, 6H), 1.17 (s, 6H), 1.07 (dd, J= 10.0, 16.0 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  162.1 (d, J =240.4 Hz), 141.1 (d, J = 3.0 Hz), 129.3 (d, J = 7.6 Hz), 115.0 (d, J = 20.8 Hz), 83.4, 83.2, 25.1, 24.8, 24.6. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.70.



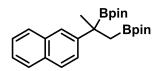
**2,2'-(1-p-Tolylethane-1,2-diyl)bis(4,4',5,5'-tetramethyl-1,3,2-dioxaborolane) (7g)**<sup>9</sup>: White solid was obtained in 20% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.11 (d, J = 8.0 Hz, 2H), 7.03 (d, J = 7.6 Hz, 2H), 2.47 (dd, J = 5.2, 10.8 Hz, 1H), 2.28 (s, 3H), 1.33-1.37 (m, 1H), 1.21 (s, 12H), 1.20 (s, 6H), 1.18 (s, 6H), 1.06-1.10 (m, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  142.4, 134.2, 129.0, 127.9, 83.3, 83.1, 25.1, 24.8, 24.8, 24.6, 21.1. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  33.78.



2,2'-(1-(naphthalen-2-yl)ethane-1,2-diyl)bis(4,4',5,5'-tetramethyl-1,3,2-dioxaboro lane) (7r): White solid was obtained in 71% isolated yield, m.p. 145-146 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  7.70-7.77 (m, 3H), 7.65 (s, 1H), 7.34-7.42 (m, 3H), 2.69 (dd, *J* = 6.0, 11.2 Hz, 1H), 1.23-1.24 (m, 2H), 1.20 (s, 12H), 1.19 (s, 6H), 1.18 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  147.0, 133.8, 131.7, 128.0, 127.4, 127.2, 126.6, 125.6, 124.9, 123.9, 83.5, 83.2, 25.3, 24.9, 24.8, 24.7, 24.6. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  34.04. FT-IR (ATR): 3052, 2977, 2930, 1469, 1369, 1389, 1271, 1139, 969, 845, 761, 675, 477. ESI-HRMS (*m/z*): cacld. for [**7r** + H]<sup>+</sup>: 407.2789; found: 407.2794.



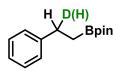
**2,2'-(1-(naphthalen-1-yl)ethane-1,2-diyl)bis(4,4',5,5'-tetramethyl-1,3,2-dioxaboro lane) (7s):** Colourless oil was obtained in 84% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 8.17-8.19 (m, 1H), 7.80-7.82 (m, 1H), 7.63-7.65 (m, 1H), 7.38-7.46 (m, 4H), 3.22 (dd, *J* = 6.0, 10.0 Hz, 1H), 1.24-1.25 (m, 2H), 1.21 (s, 6H), 1.20 (s, 12H), 1.16 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 142.2, 134.1, 132.1, 128.7, 125.9, 125.9, 125.3, 125.2, 125.0, 124.7, 83.5, 83.2, 25.1, 24.8, 24.8, 24.7. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 33.78. FT-IR (ATR): 3062, 2978, 2931, 1736, 1509, 1447, 1453, 1329, 1143, 981, 850, 779, 673. ESI-HRMS (*m/z*): cacld. for [**7s** + H]<sup>+</sup>: 407.2789; found: 407.2772.



**2,2'-(2-(naphthalen-2-yl)propane-1,2-diyl)bis(4,4',5,5'-tetramethyl-1,3,2-dioxabo rolane) (7t):** White solid was obtained in 88% isolated yield, m.p. 84-85 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.72-7.79 (m, 4H), 7.59-7.61 (m, 1H), 7.36-7.43 (m, 2H), 1.51 (s, 3H), 1.26-1.27 (m, 2H), 1.22 (s, 12H), 1.21 (s, 6H), 1.19 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 147.0, 133.8, 131.7, 128.0, 127.4, 127.2, 126.6, 125.6, 125.0, 124.0, 83.5, 83.2, 25.2, 24.9, 24.8, 24.7, 24.6. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 34.24. FT-IR (ATR): 3055, 2976, 2929, 1476, 1380, 1312, 1139, 1112, 970, 859, 747, 475. ESI-HRMS (*m/z*): cacld. for [**7t** + H]<sup>+</sup>: 421.2945; found: 421.2929.

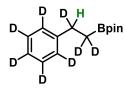
*d*<sub>1</sub>-pinacolborane (DBPin)<sup>1-2</sup>: DBpin was obtained of 1.36 M in THF. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  1.19 (s, 12H). <sup>2</sup>D NMR (62 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  2.86-4.50 (br). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  83.1, 24.8. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>,

ppm): δ28.08.

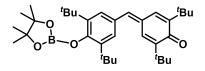


**monodeuterium boronic ester**  $d_1$ -**3a**:**3a** = **84**:**16**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.24-7.25 (m, 2H), 7.20-7.21 (m, 2H), 7.13-7.15 (m, 1H), 2.71-2.76 (m, 1.16H), 1.21 (s, 12H), 1.13-1.14 (m, 2H). <sup>2</sup>D NMR (62 MHz, CHCl<sub>3</sub>, ppm): δ 2.74. <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 144.5, 128.3, 128.1, 125.6, 83.2, 29.8, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 33.71.

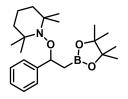
monodeuterium boronic ester *d*<sub>1</sub>-3a:3a = 16:84: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.21-7.25 (m, 4H), 7.13-7.17 (m, 1H), 2.76 (t, *J* = 8.0 Hz, 1.84H), 1.22 (s, 12H), 1.15 (t, *J* = 8.0 Hz, 2H). <sup>2</sup>D NMR (62 MHz, CDCl<sub>3</sub>, ppm): δ 2.88 (d, *J* = 4.0 Hz). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 144.6, 128.3, 128.1, 125.6, 83.2, 30.0, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 33.91.



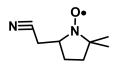
**mono-proteo boronic ester** *d*<sub>8</sub>-3a: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 2.72 (s, 1H), 1.22 (s, 12H). <sup>2</sup>D NMR (62 MHz, CHCl<sub>3</sub>, ppm): δ 7.20-7.30 (m, 5D), 2.74 (s, 1D), 1.12 (s, 2D). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 144.3, 128.0, 127.8, 127.5, 125.1, 83.2, 29.5, 24.9. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 33.91.



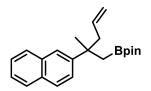
8: ESI-HRMS (*m*/*z*): cacld. for [8 + H]<sup>+</sup>: 548.4146; found: 548.4141.



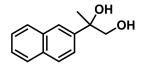
**9**: ESI-HRMS (*m*/*z*): cacld. for [**9** + H]<sup>+</sup>: 387.3054; found: 387.3053.



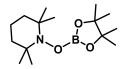
**10**: ESI-HRMS (m/z): cacld. for [**10** + 2H]<sup>+</sup>: 155.1179; found: 155.1182.



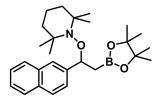
**4,4',5,5'-tetramethyl-2-(2-methyl-2-(naphthalen-2-yl)pent-4-en-1-yl)-1,3,2-dioxab orolane (11):** Colourless liquid was obtained in 45% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.72-7.79 (m, 4H), 7.54-7.57 (m, 1H), 7.39-7.45 (m, 2H), 5.47-5.58 (m, 1H), 4.90-4.99 (m, 2H), 2.47-2.63 (m, 2H), 1.53 (s, 3H), 1.25-1.28 (m, 2H), 1.05 (s, 6H), 1.02 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 146.9, 135.8, 133.4, 131.8, 128.1, 127.5, 127.4, 125.7, 125.4, 125.2, 124.4, 117.1, 82.9, 49.5, 39.6, 27.1, 24.8, 24.7. <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>, ppm): δ 33.17. FT-IR (ATR): 3058, 2976, 2925, 2855, 1734, 1355, 1325, 1143, 969, 848, 816, 746. ESI-HRMS (*m/z*): cacld. for [**11** + H]<sup>+</sup>: 336.2370; found: 336.2375.



**2-(naphthalen-2-yl)propane-1,2-diol** (12)<sup>11</sup>: White solid was obtained in 94% isolated yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): δ 7.93 (s, 1H), 7.82-7.85 (m, 3H), 7.46-7.51 (m, 3H), 3.85-3.88 (m, 1H), 3.67-3.69 (m, 1H), 2.93 (br, 1H), 2.17 (br, 1H), 1.59 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm): δ 142.4, 133.3, 132.6, 128.3, 127.6, 126.4, 126.1, 124.1, 123.5, 75.1, 71.0, 26.1.



**13**: GC-MS (*m/z*): cacld. for [**13**]<sup>+</sup>: 282.2355; found: 282.2385.



14: ESI-HRMS (m/z): cacld. for [14 + H]<sup>+</sup>: 437.3210; found: 437.3218.

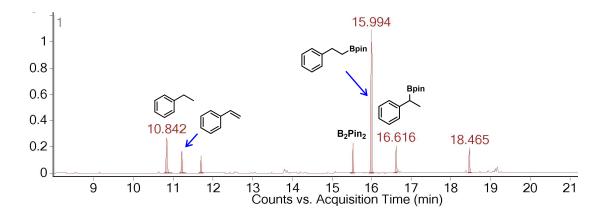
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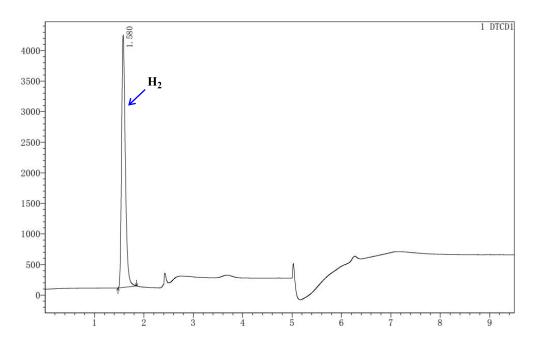
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#### V. Gas Spectra



**Figure S4.** GC–MS spectrum of the hydroboration reaction between styrene and HBpin in the CH<sub>3</sub>CN/THF mixture under the standard conditions.



**Figure S5.** GC spectrum of gases produced in the hydroboration reaction between styrene and HBpin in the CH<sub>3</sub>CN/THF mixture under the standard conditions.

## **VI. Electrochemical Conditions Screening**

la la	Pt(+)   Pt(-), I = 15 <sup>n</sup> Bu <sub>4</sub> NBF <sub>4</sub> (20 mo + HBpin 2 2	l%)	$\begin{array}{c} \text{Bpin} \\ + \\ + \\ + \\ B_2 \text{Pin}_2 \\ 6 \end{array}$
Entry	Solvent	Yield of 3a (%) <sup>b</sup>	3a/4a/5/6 <sup>c</sup>
1	$CH_3CN:THF = 7:3$	49	59:8:21:12
2	<b>CH</b> <sub>3</sub> <b>CN:THF</b> = 4:1	56	62:9:17:12
3	$CH_3CN:THF = 9:1$	43	57:9:24:10
4	$CH_{3}CN:THF = 9.5:0.5$	49	60:6:20:14
5	$CH_3CN:Et_2O = 4:1$	45	77:0:13:10
6	$CH_3CN:1,4$ -dioxane = 4:1	48	62:0:28:10

Table S1. Screening of solvent<sup>a</sup>

<sup>*a*</sup>Reaction conditions: **1a** (1.0 mmol), **2** (1.1 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=15 mA, solvent, the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 3 h.

<sup>b</sup>The yields of **3a** were determined by gas chromatography (GC) analysis using naphthalene as internal standard.

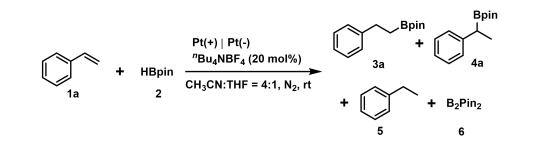
Table S2. Screening of electrode and temperature<sup>a</sup>

la 1a	+ HBpin 2 HBpin 2 HBpin 2 HBpin 2 H3CN:THF = 4:1, N <sub>2</sub> , rt, 3 h	í VÝ V	$\begin{array}{c} \text{Bpin} \\ + \\ + \\ + \\ + \\ + \\ B_2 \text{Pin}_2 \\ 6 \end{array}$
Entry	Variation from the standard conditions	Yield of 3a(%) <sup>b</sup>	3a/4a/5/6 <sup>c</sup>
1	C(+)   Pt(-), 40 °C	20	59:0:36:5
2	Pt(+)   Ni(-), 40 °C	40	56:27:9:8
3	C(+)   Pt(-)	31	63:0:30:7
4	Pt(+)   Ni(-)	51	57:9:24:10
5	60 °C	38	59:6:27:8
6	40 °C	42	60:8:22:10
7	none	56	62:9:17:12
8	0 °C	53	61:10:17:12

<sup>*a*</sup>Reaction conditions: **1a** (1.0 mmol), **2** (1.1 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=15 mA, CH<sub>3</sub>CN:THF=4:1 ( $\nu/\nu$ ), the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 3 h.

<sup>b</sup>The yields of **3a** were determined by gas chromatography (GC) analysis using naphthalene as internal standard.

 Table S3. Screening of time, current and electrolyte<sup>a</sup>



Entry	Variation from the standard conditions	Yield of $3a (\%)^b$	3a/4a/5/6°
1	2 h	39	59:9:23:9
2	4 h	55	68:7:10:15
3	10 mA	37	68:8:8:16
4	13 mA	49	65:9:14:12
5	none	56	67:7:15:11
6	20 mA	51	57:8:25:10
7	30 mA	53	55:14:27:4
8	0.5 eq. "Bu4NBF4	51	58:10:20:12

<sup>*a*</sup>Reaction conditions: **1a** (1.0 mmol), **2** (1.1 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=15 mA, CH<sub>3</sub>CN:THF=4:1 ( $\nu/\nu$ ), the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 3 h.

<sup>b</sup>The yields of **3a** were determined by gas chromatography (GC) analysis using naphthalene as internal standard.



Ta 1a	+ HBpin 2 + HBpin Pt(+)   Pt(-) "Bu <sub>4</sub> NBF <sub>4</sub> Additives CH <sub>3</sub> CN:TI N <sub>2</sub> , rt	(20 mol%) HF = 4:1, 3a	Bpin      +
Entry	Additives	Yield of $3a (\%)^b$	3a/4a/5/6 <sup>c</sup>
$1^a$	Et <sub>3</sub> N (1.1 eq.)	44	65:10:23:2
2 <sup><i>c</i></sup>	Et <sub>3</sub> N (1.1 eq.)	51	75:10:13:2
3 <sup><i>a</i></sup>	DIEA (1.1 eq.)	60	73:10:13:4
4 <sup><i>c</i></sup>	DIEA (1.1 eq.)	67	76:11:12:1
5 <sup>c</sup>	DIEA (0.9 eq.)	71	76:7:14:3
<b>6</b> <sup><i>c</i></sup>	DIEA (0.8 eq.)	72	82:5:11:2
7 <sup>c</sup>	DIEA (0.65 eq.)	69	79:7:11:3
8 <sup>c</sup>	DIEA (0.55 eq.)	70	81:5:12:2
9 <sup>c</sup>	DIEA (0.4 eq.)	67	74:7:15:4
10 <sup>c</sup>	DIEA (0.25 eq.)	63	69:7:19:5

<sup>*a*</sup>Reaction conditions: **1a** (1.0 mmol), **2** (1.1 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=15 mA, additives, CH<sub>3</sub>CN (10 mL), N<sub>2</sub>, rt, 3 h.

<sup>b</sup>The yields of **3a** were determined by gas chromatography (GC) analysis using naphthalene as internal standard.

<sup>*c*</sup>Reaction conditions: **1a** (1.0 mmol), **2** (1.1 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=15 mA, additives, CH<sub>3</sub>CN:THF=4:1 ( $\nu/\nu$ ), the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 3 h.

	Pt(+)   Pt(-) <sup>//</sup> Bu <sub>4</sub> NBF <sub>4</sub> (20 mol%) DIEA (80 mol%)	Bpin
1a	<ul> <li>HBpin</li> <li>CH<sub>3</sub>CN:THF = 4:1,</li> <li>N<sub>2</sub>, rt, 12 h</li> </ul>	3a
Entry	Current	<b>Yield of 3a (%)</b> <sup>b</sup>
1	15 mA	31
2	30 mA	46
3	45 mA	61

Table S5. Screening of current in gram-scale experiments<sup>a</sup>

<sup>*a*</sup>Reaction conditions: **1a** (10 mmol), **2** (11 mmol), DIEA (80 mol%), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current, CH<sub>3</sub>CN:THF=4:1 ( $\nu/\nu$ ), the total volume of the solvent is 100 mL, N<sub>2</sub>, rt, 12 h.

<sup>b</sup>Isolated yields are shown.

**Table S6.** Screening of the molar ratio of 2-vinylphthalene to HBpin to prepare diboronate ester  $7r^a$ 

+ HBpin 1r 2	Pt(+)   Pt(-), I = 20 mA <sup>n</sup> Bu <sub>4</sub> NBF <sub>4</sub> (20 mol%) DIEA (80 mol%) CH <sub>3</sub> CN:THF = 4:1, N <sub>2</sub> , rt, 4 h	Bpin 3r	Bpin Bpin Bpin 7r
Entry	1r:2 (mol:mo	ol)	3r:7r <sup>b</sup>
1	1:1.1		65:35
2	1:2.2		38:61
3	1:3.3		14:86
4	1:4.4		9:91

<sup>*a*</sup>Reaction conditions: **1r** (1.0 mmol), **2**, DIEA (80 mol%), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=20 mA, CH<sub>3</sub>CN:THF=4:1 ( $\nu/\nu$ ), the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 4 h.

<sup>b</sup>The yield ratios of  $3\mathbf{r}$  to  $7\mathbf{r}$  were determined by the peak area ratios of gas chromatography (GC).

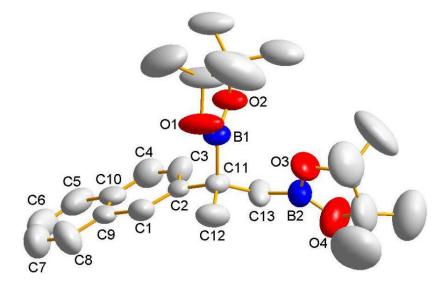
+ 1a	$\begin{array}{c} Pt(+) \mid Pt(-), \ I = 20 \ mA \\ {}^{n}Bu_{4}NBF_{4} \ (20 \ mol\%) \\ \hline \\ DIEA \ (80 \ mol\%) \\ \hline \\ CH_{3}CN:THF = 4:1, \\ 2 \\ N_{2}, \ rt, \ 4 \ h \end{array}$	Bpin Bpin + 7a
Entry	1a:2 (mol:mol)	3a:7a <sup>b</sup>
1	1:4.4	65:35
2	1:6.6	53:47
3	1:8.8	40:60
4	1:10.0	40:60

**Table S7.** Screening of the molar ratio of styrene to HBpin to prepare diboronate ester $7a^a$ 

*a*Reaction conditions: **1a** (1.0 mmol), **2**, DIEA (80 mol%), *<sup>n</sup>*Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), Pt(+)|Pt(-), constant current (I)=20 mA, CH<sub>3</sub>CN:THF=4:1 ( $\nu/\nu$ ), the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 4 h.

<sup>b</sup>The yield ratios of 3a to 7a were determined by the peak area ratios of gas chromatography (GC).

## **VII.** Crystal Information



**Figure S6.** ORTEP diagram of compound **7t**. Thermal ellipsoids are shown at 50% probability level. All hydrogen atoms on carbons are omitted for the sake of clarity.

Compound	7t
Formula	$C_{25}H_{36}B_2O_4$
Formula weight	422.16
Crystal dimensions (mm <sup>3</sup> )	$0.26 \times 0.25 \times 0.25$
Crystal system	monoclinic
Space group	'P 21/n'
a (Å)	6.5462(6)
b (Å)	27.280(2)
c (Å)	14.4106(12)
α (°)	90.00
eta (°)	90.549(3)
γ (°)	90.00
Volume (Å <sup>3</sup> )	2573.4(4)
Ζ	4
T (K)	293(2)
$D_{calcd} (g \ cm^{-3})$	1.090
$\mu ~(\mathrm{mm}^{-1})$	0.070
F (000)	912
No. of rflns. collected	21174
No. of indep. rflns. $/R_{int}$	3893 / 0.0551
No. of obsd. rflns. $[I_0 > 2\sigma(I_0)]$	2526
Data / restraints / parameters	3893 / 103 / 308
$R_1 / wR_2 [I_0 > 2\sigma(I_0)]$	0.1499 / 0.3733
$R_1 / wR_2$ (all data)	0.1952 / 0.3965
GOF (on $F^2$ )	1.001
Largest diff. peak and hole (e $Å^{-3}$ )	0.974 / -0.359
CCDC No.	2022111

Table S8. Crystal data and structural refinement for compound 7t

Distances (Å)			
B1–O1	1.319(8)	B2–O3	1.355(9)
B1–O2	1.347(8)	B2–O4	1.320(9)
B1-C11	1.588(8)	B2-C13	1.585(11)
Angles (°)			
C2C11B1	107.1(5)	C12C11B1	109.4(5)
C13-C11-B1	111.0(5)	C11-C13-B2	111.3(6)

Table S9. Selected bond distances and angles for 7t

## VIII. NMR Spectra

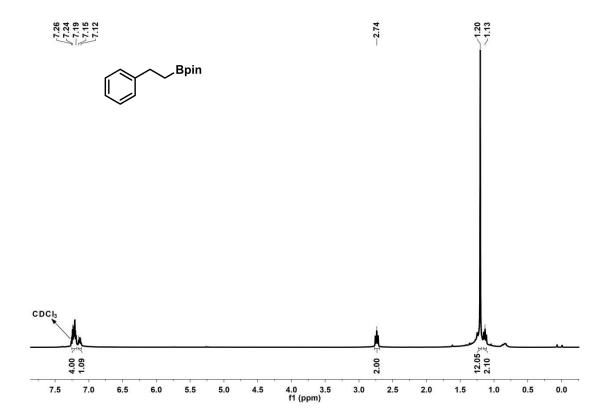


Figure S7. <sup>1</sup>H NMR spectrum of 3a in CDCl<sub>3</sub>.

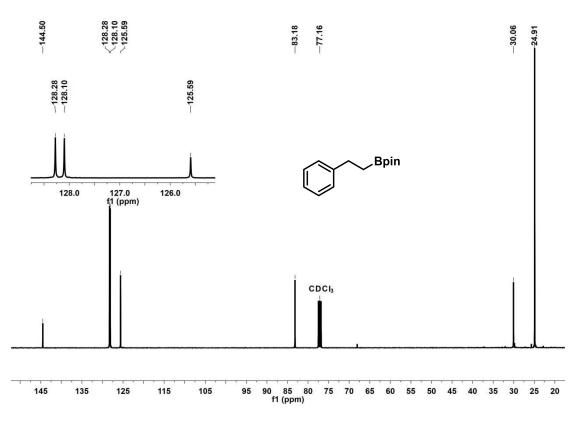


Figure S8. <sup>13</sup>C NMR spectrum of 3a in CDCl<sub>3</sub>.

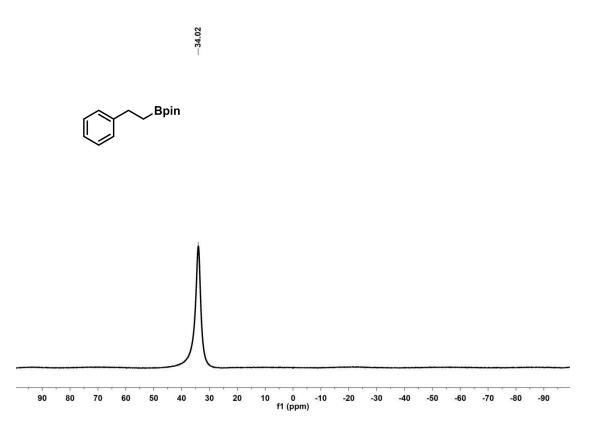


Figure S9. <sup>11</sup>B NMR spectrum of 3a in CDCl<sub>3</sub>.

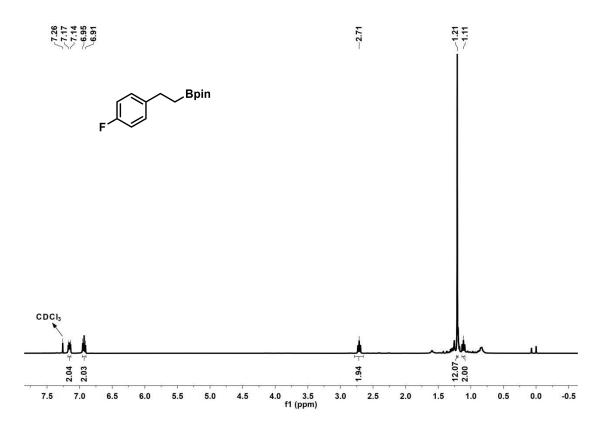


Figure S10. <sup>1</sup>H NMR spectrum of 3b in CDCl<sub>3</sub>.

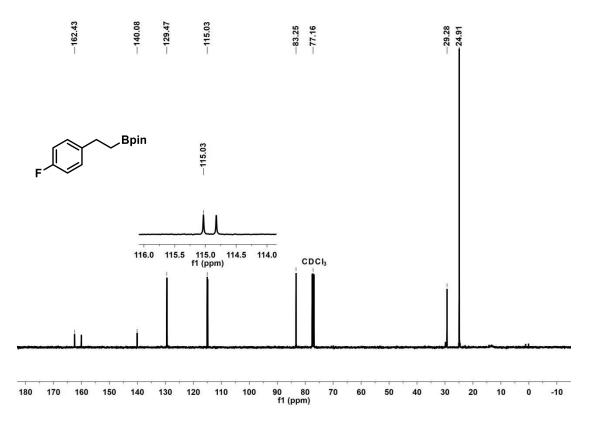


Figure S11. <sup>13</sup>C NMR spectrum of **3b** in CDCl<sub>3</sub>.

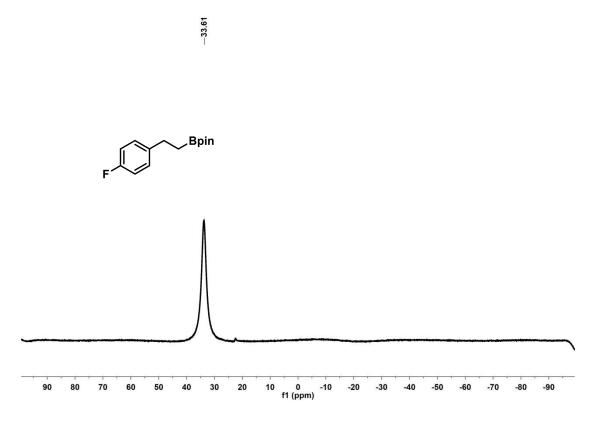


Figure S12. <sup>11</sup>B NMR spectrum of 3b in CDCl<sub>3</sub>.

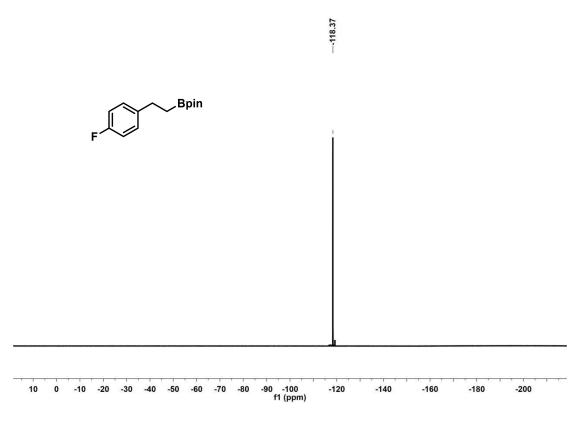


Figure S13. <sup>19</sup>F NMR spectrum of 3b in CDCl<sub>3</sub>.

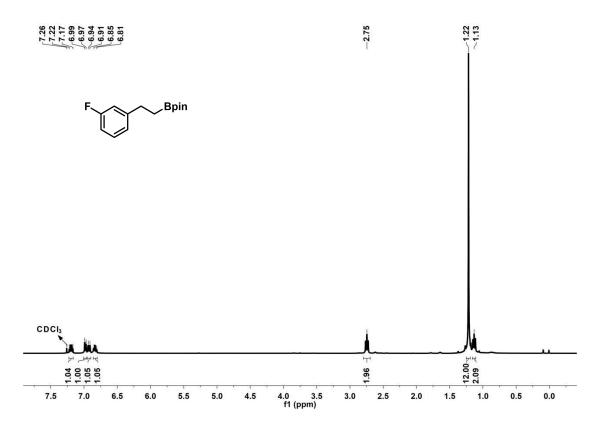


Figure S14. <sup>1</sup>H NMR spectrum of 3c in CDCl<sub>3</sub>.

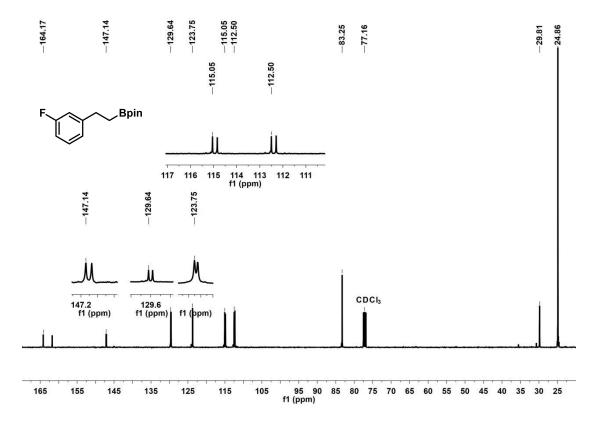


Figure S15. <sup>13</sup>C NMR spectrum of 3c in CDCl<sub>3</sub>.

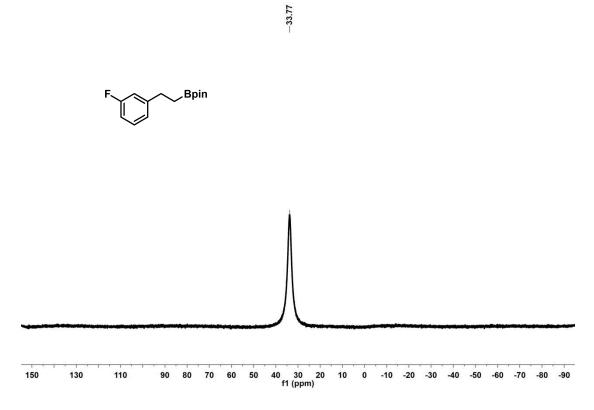


Figure S16. <sup>11</sup>B NMR spectrum of 3c in CDCl<sub>3</sub>.

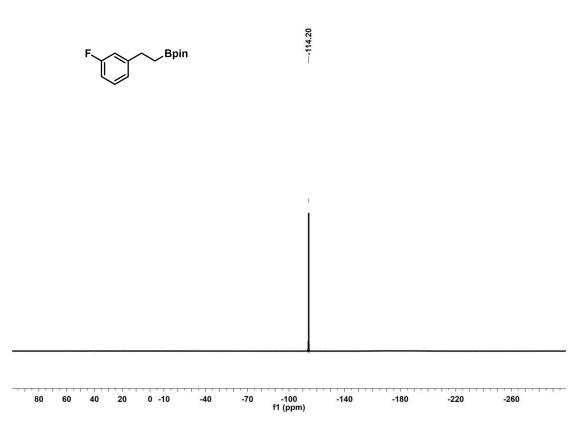


Figure S17. <sup>19</sup>F NMR spectrum of 3c in CDCl<sub>3</sub>.

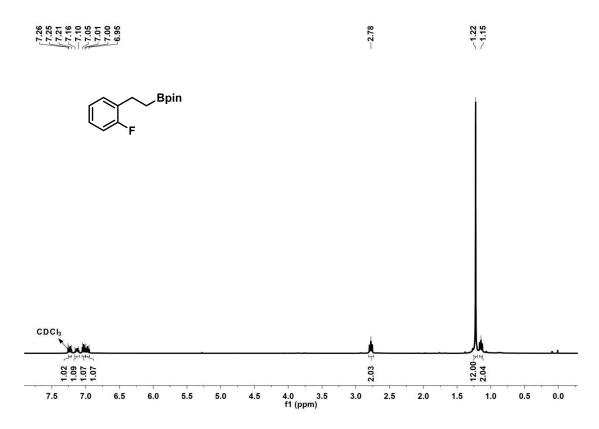


Figure S18. <sup>1</sup>H NMR spectrum of 3d in CDCl<sub>3</sub>.

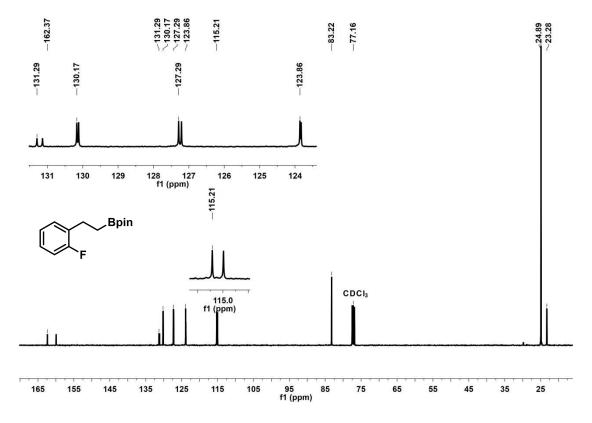


Figure S19. <sup>13</sup>C NMR spectrum of 3d in CDCl<sub>3</sub>.

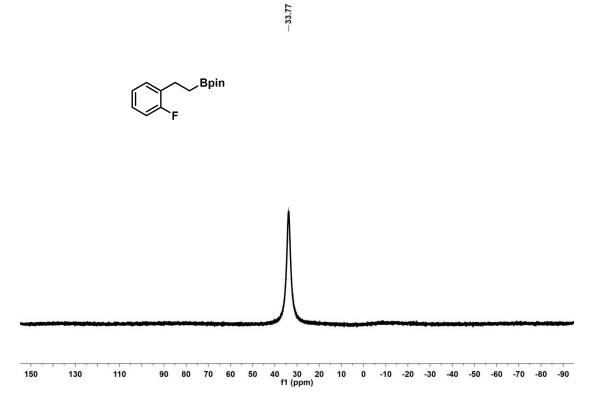


Figure S20. <sup>11</sup>B NMR spectrum of 3d in CDCl<sub>3</sub>.

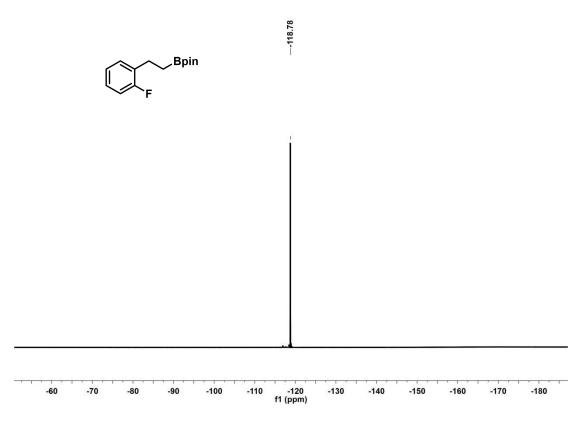


Figure S21. <sup>19</sup>F NMR spectrum of 3d in CDCl<sub>3</sub>.

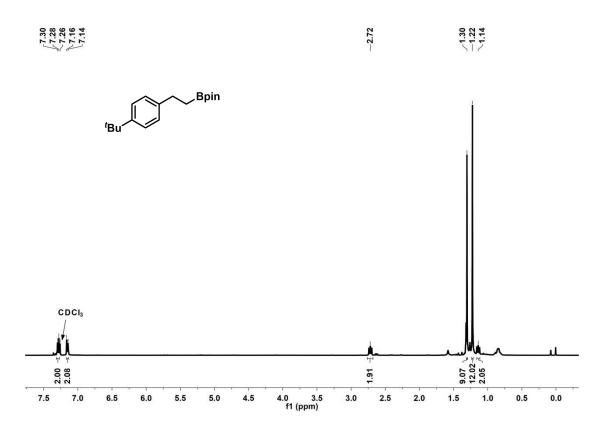


Figure S22. <sup>1</sup>H NMR spectrum of 3e in CDCl<sub>3</sub>.

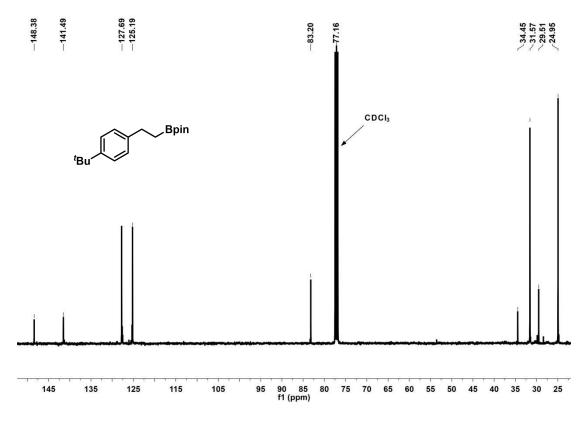


Figure S23. <sup>13</sup>C NMR spectrum of 3e in CDCl<sub>3</sub>.

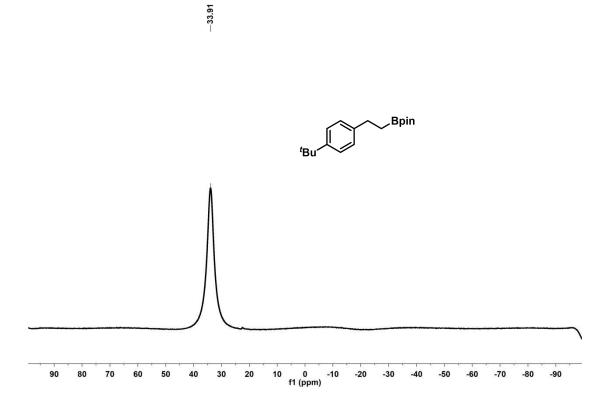


Figure S24. <sup>11</sup>B NMR spectrum of 3e in CDCl<sub>3</sub>.

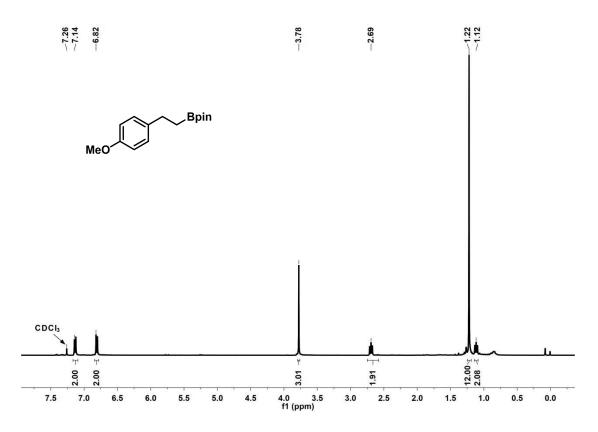


Figure S25. <sup>1</sup>H NMR spectrum of 3f in CDCl<sub>3</sub>.

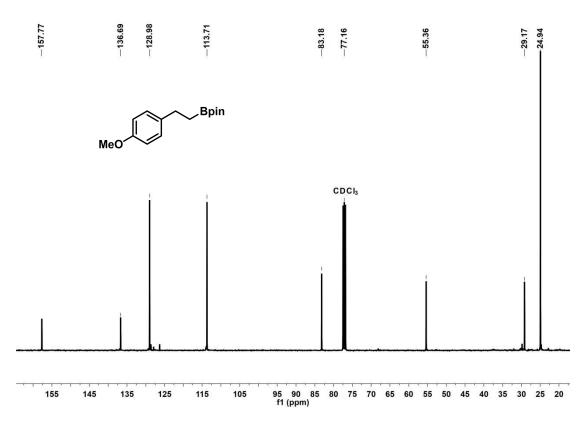


Figure S26. <sup>13</sup>C NMR spectrum of 3f in CDCl<sub>3</sub>.

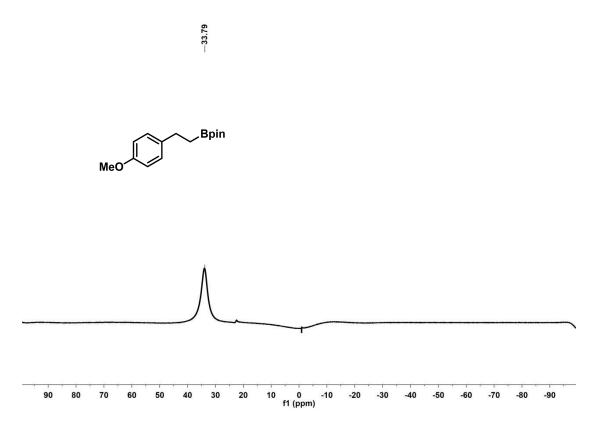


Figure S27. <sup>11</sup>B NMR spectrum of 3f in CDCl<sub>3</sub>.

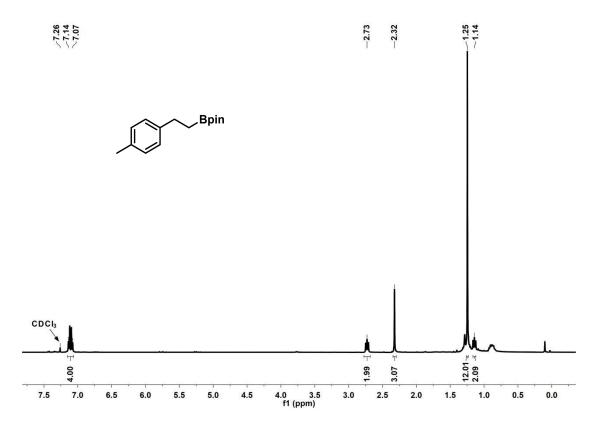


Figure S28. <sup>1</sup>H NMR spectrum of 3g in CDCl<sub>3</sub>.

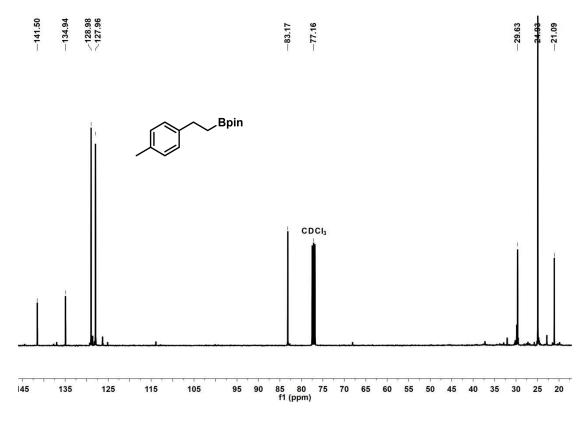


Figure S29. <sup>13</sup>C NMR spectrum of 3g in CDCl<sub>3</sub>.

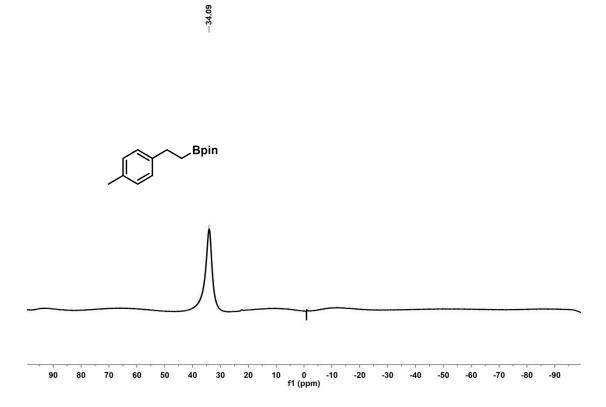


Figure S30. <sup>11</sup>B NMR spectrum of 3g in CDCl<sub>3</sub>.

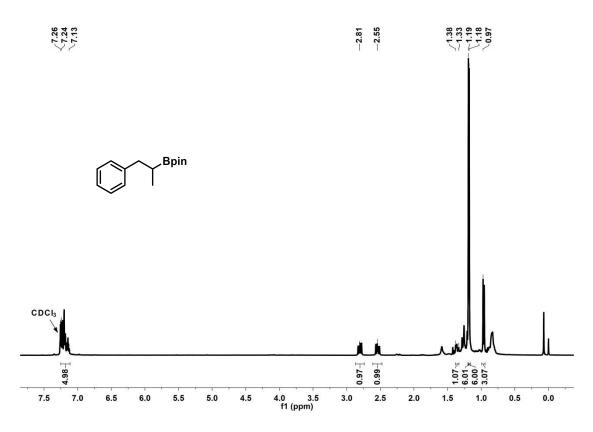


Figure S31. <sup>1</sup>H NMR spectrum of 3h in CDCl<sub>3</sub>.

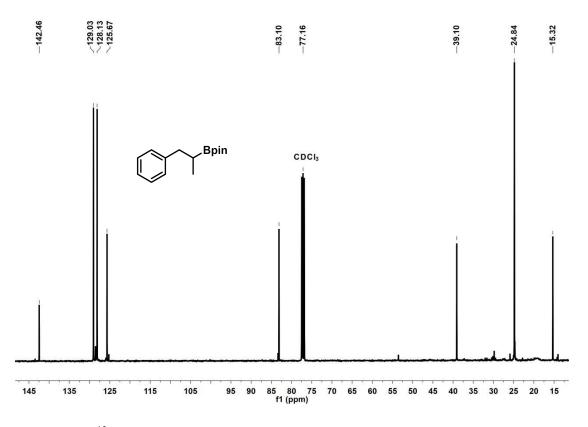


Figure S32. <sup>13</sup>C NMR spectrum of 3h in CDCl<sub>3</sub>.

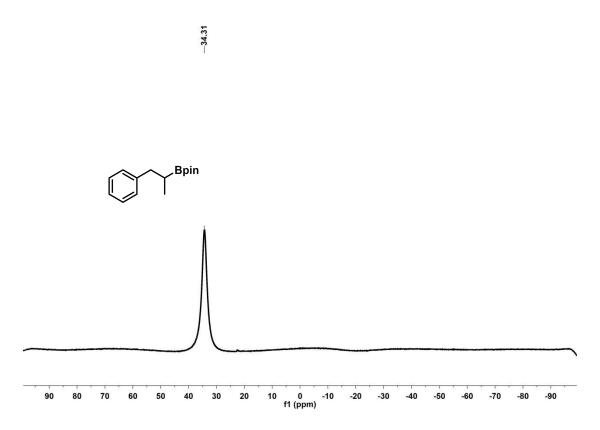


Figure S33. <sup>11</sup>B NMR spectrum of 3h in CDCl<sub>3</sub>.

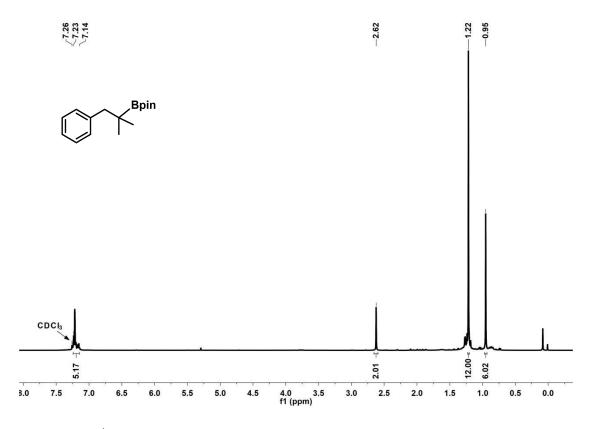


Figure S34. <sup>1</sup>H NMR spectrum of 3i in CDCl<sub>3</sub>.

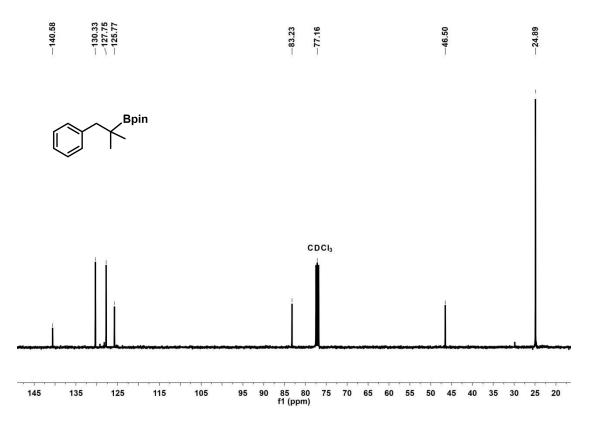
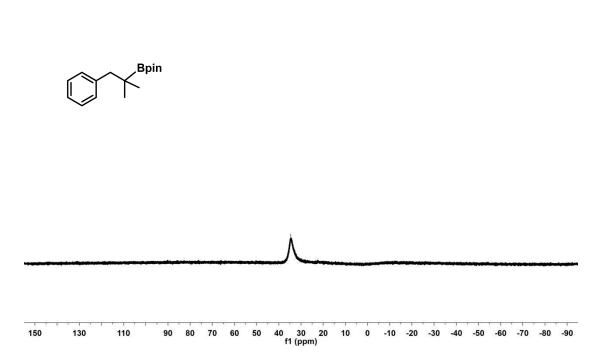


Figure S35. <sup>13</sup>C NMR spectrum of 3i in CDCl<sub>3</sub>.



-34.60

Figure S36. <sup>11</sup>B NMR spectrum of 3i in CDCl<sub>3</sub>.

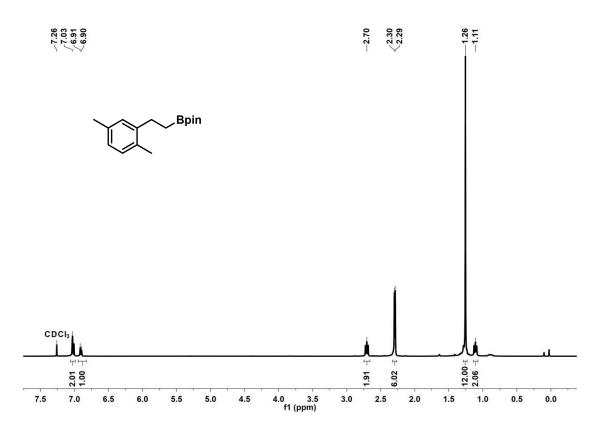


Figure S37. <sup>1</sup>H NMR spectrum of 3j in CDCl<sub>3</sub>.

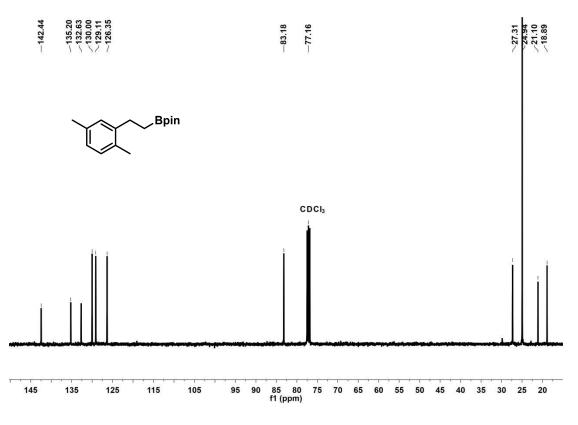


Figure S38. <sup>13</sup>C NMR spectrum of 3j in CDCl<sub>3</sub>.

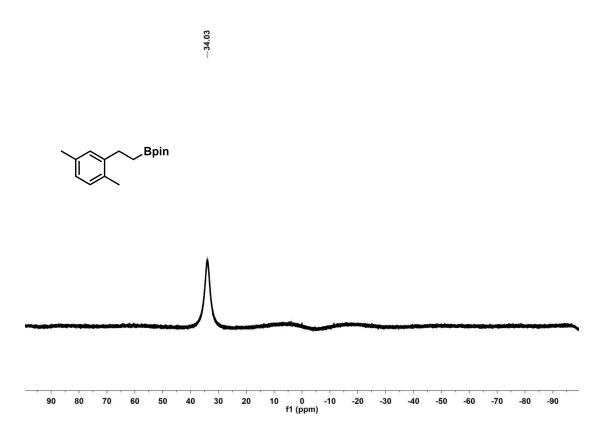


Figure S39. <sup>11</sup>B NMR spectrum of 3j in CDCl<sub>3</sub>.

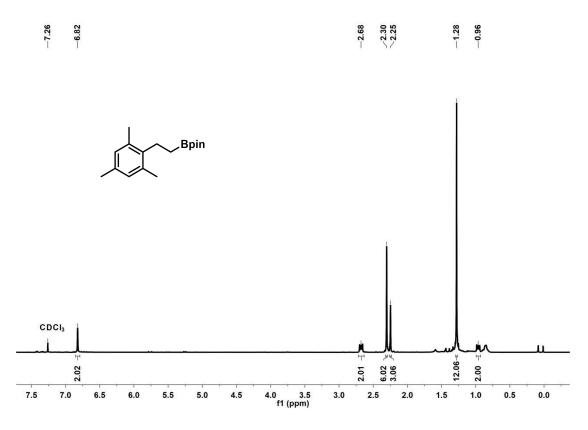


Figure S40. <sup>1</sup>H NMR spectrum of 3k in CDCl<sub>3</sub>.

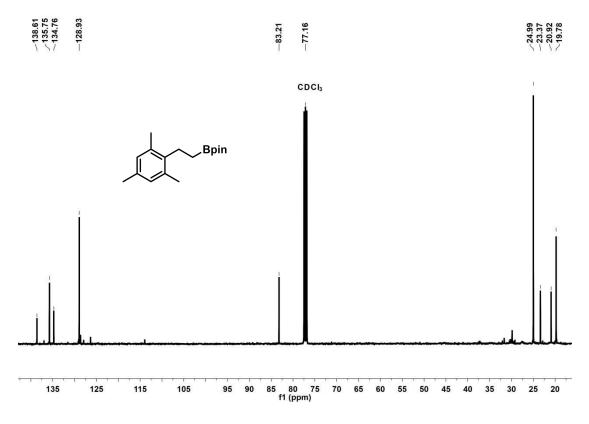


Figure S41. <sup>13</sup>C NMR spectrum of 3k in CDCl<sub>3</sub>.

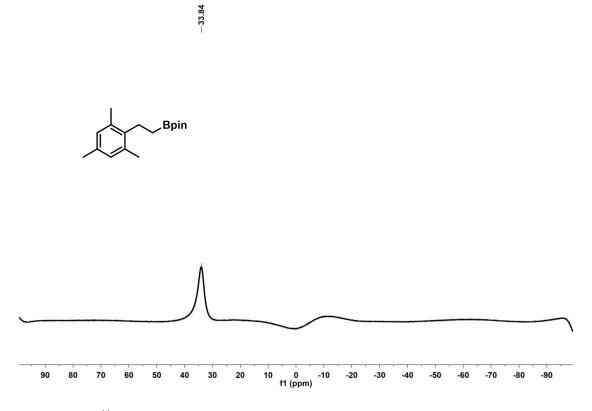


Figure S42. <sup>11</sup>B NMR spectrum of 3k in CDCl<sub>3</sub>.

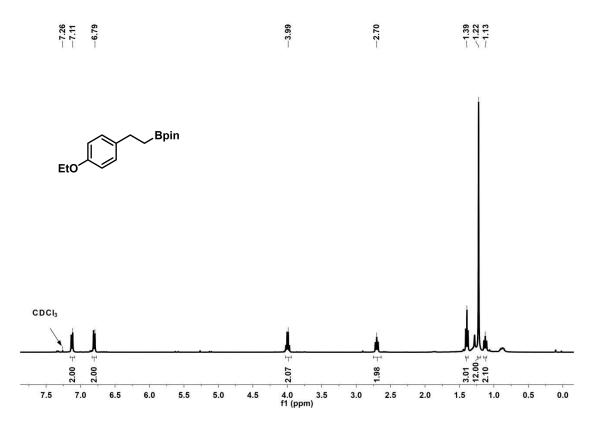


Figure S43. <sup>1</sup>H NMR spectrum of 31 in CDCl<sub>3</sub>.

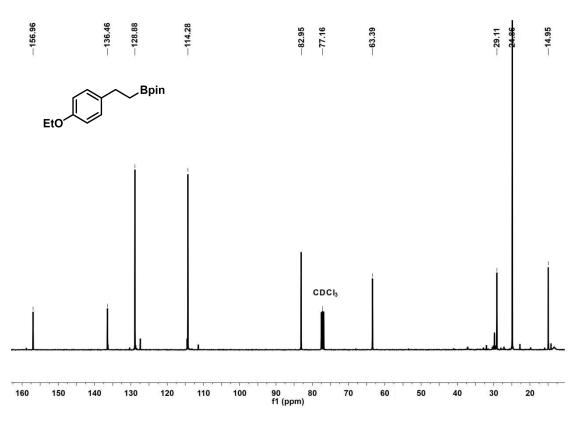


Figure S44. <sup>13</sup>C NMR spectrum of 3l in CDCl<sub>3</sub>.

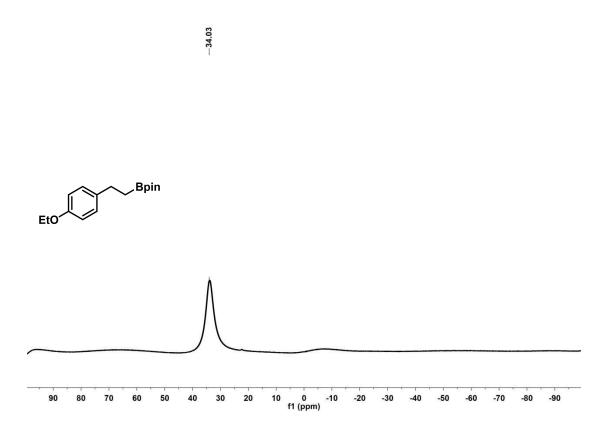


Figure S45. <sup>11</sup>B NMR spectrum of 3l in CDCl<sub>3</sub>.

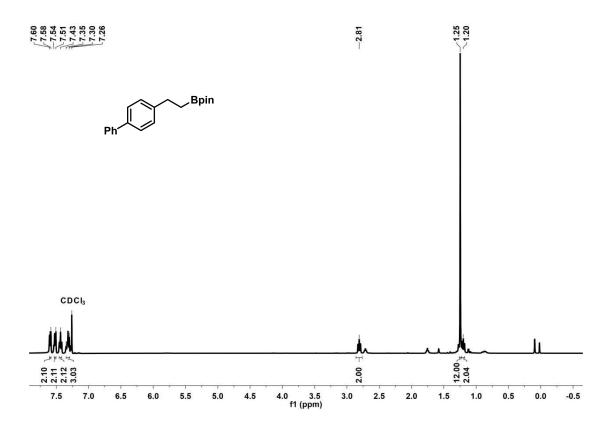


Figure S46. <sup>1</sup>H NMR spectrum of 3m in CDCl<sub>3</sub>.

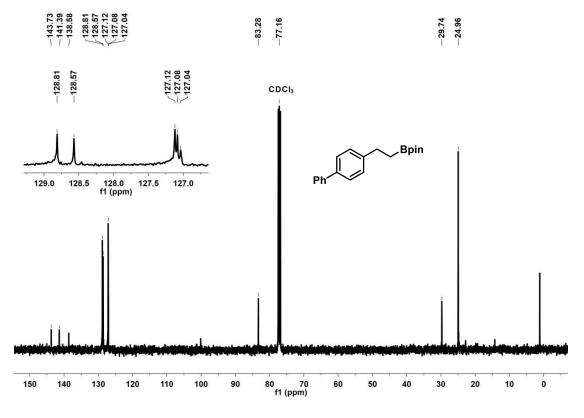


Figure S47. <sup>13</sup>C NMR spectrum of **3m** in CDCl<sub>3</sub>.

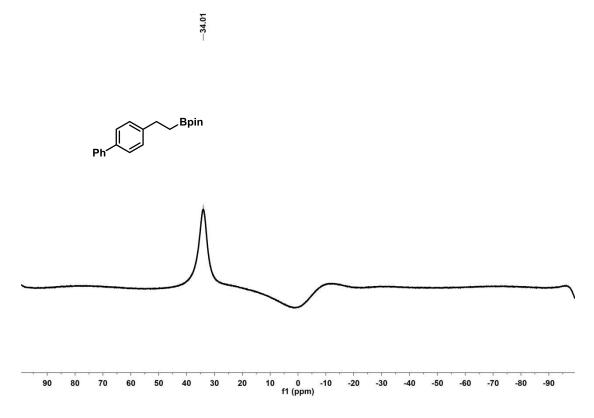


Figure S48. <sup>11</sup>B NMR spectrum of **3m** in CDCl<sub>3</sub>.

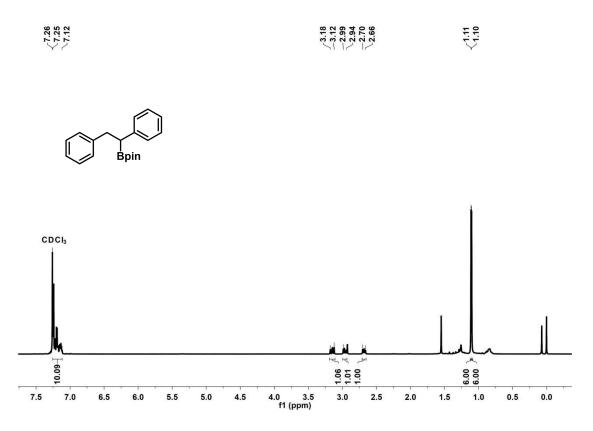


Figure S49. <sup>1</sup>H NMR spectrum of 3n in CDCl<sub>3</sub>.

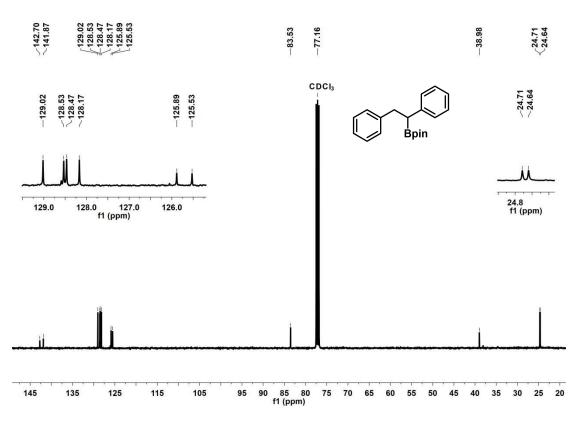


Figure S50. <sup>13</sup>C NMR spectrum of **3n** in CDCl<sub>3</sub>.

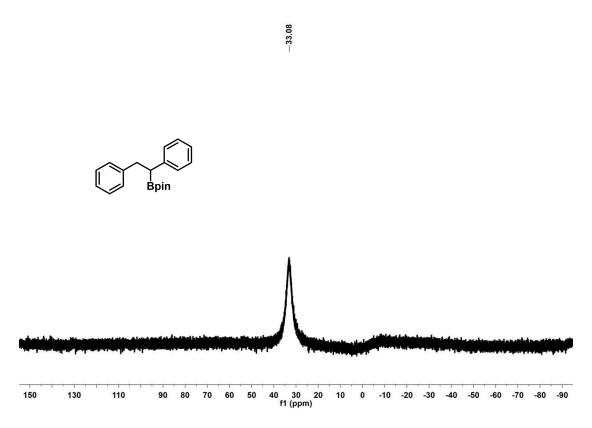


Figure S51. <sup>11</sup>B NMR spectrum of 3n in CDCl<sub>3</sub>.

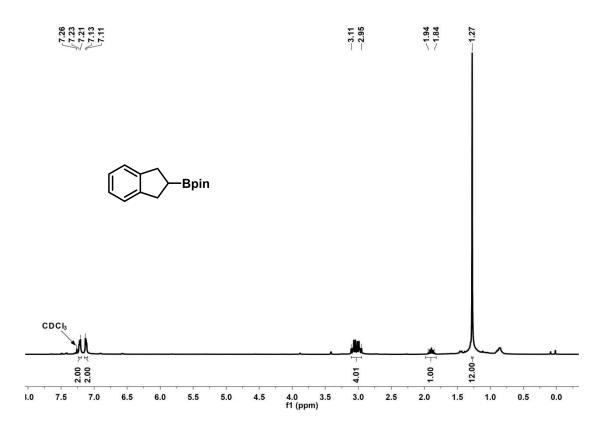


Figure S52. <sup>1</sup>H NMR spectrum of 30 in CDCl<sub>3</sub>.

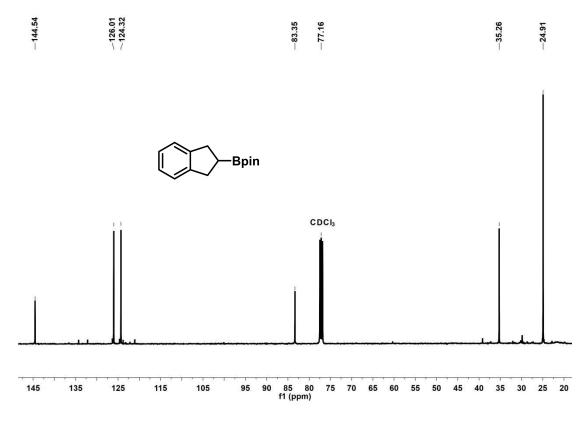


Figure S53. <sup>13</sup>C NMR spectrum of **30** in CDCl<sub>3</sub>.

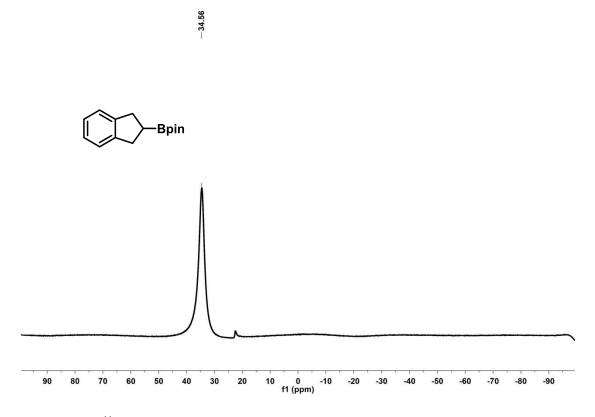


Figure S54. <sup>11</sup>B NMR spectrum of 30 in CDCl<sub>3</sub>.

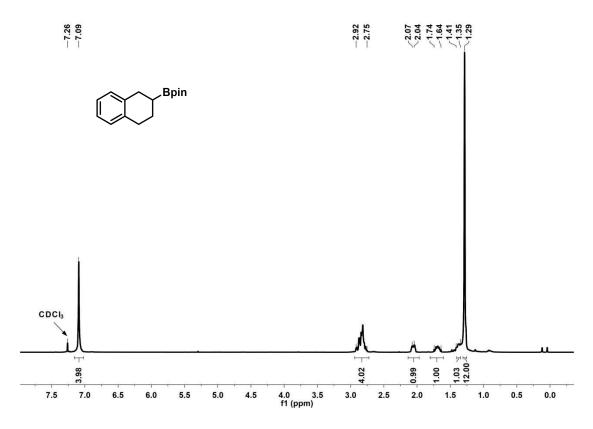


Figure S55. <sup>1</sup>H NMR spectrum of **3p** in CDCl<sub>3</sub>.

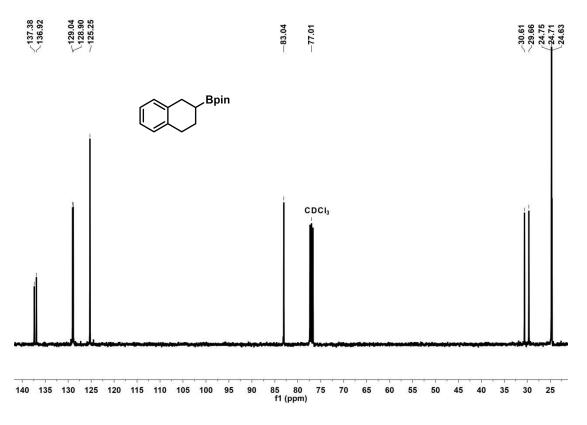


Figure S56. <sup>13</sup>C NMR spectrum of **3p** in CDCl<sub>3</sub>.

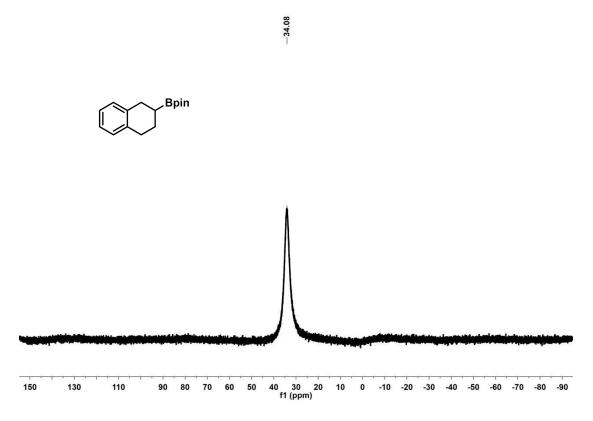


Figure S57. <sup>11</sup>B NMR spectrum of 3p in CDCl<sub>3</sub>.

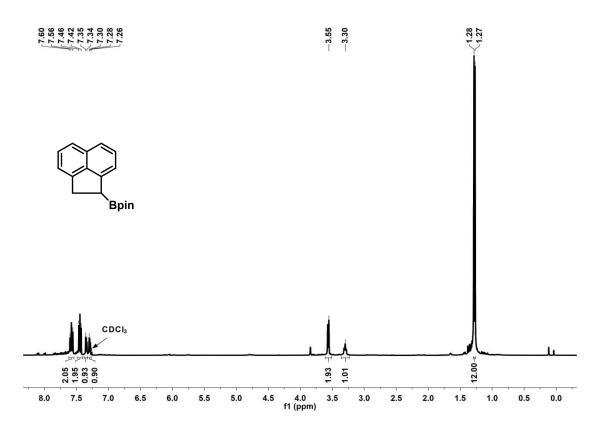


Figure S58. <sup>1</sup>H NMR spectrum of 3q in CDCl<sub>3</sub>.

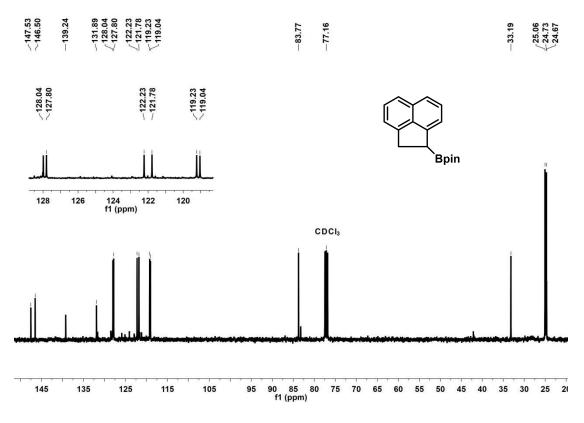
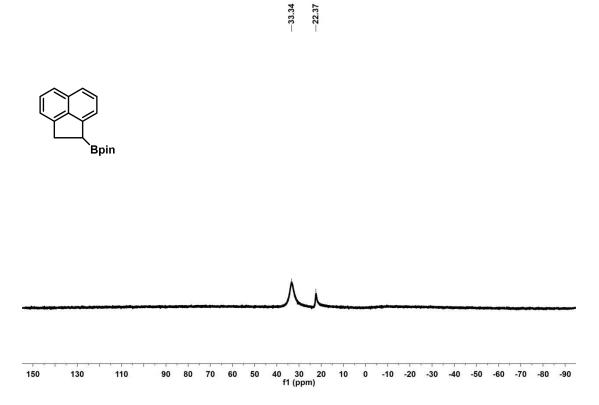


Figure S59. <sup>13</sup>C NMR spectrum of 3q in CDCl<sub>3</sub>.



-22.37

Figure S60. <sup>11</sup>B NMR spectrum of 3q in CDCl<sub>3</sub>.

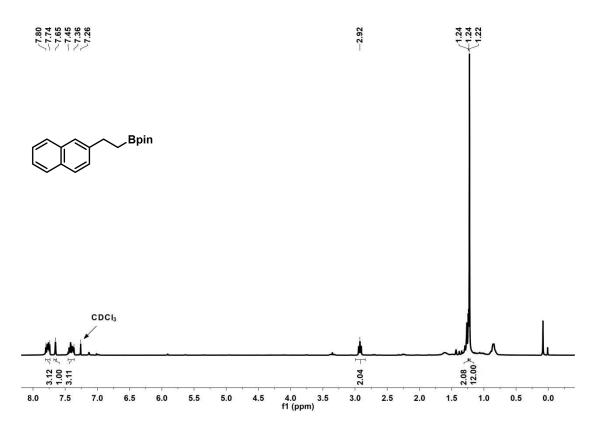


Figure S61. <sup>1</sup>H NMR spectrum of 3r in CDCl<sub>3</sub>.

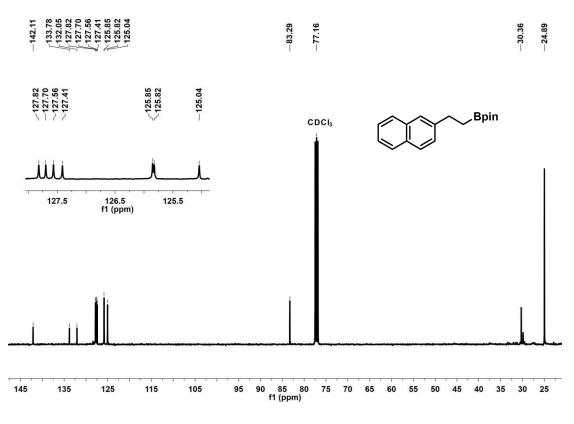


Figure S62. <sup>13</sup>C NMR spectrum of 3r in CDCl<sub>3</sub>.

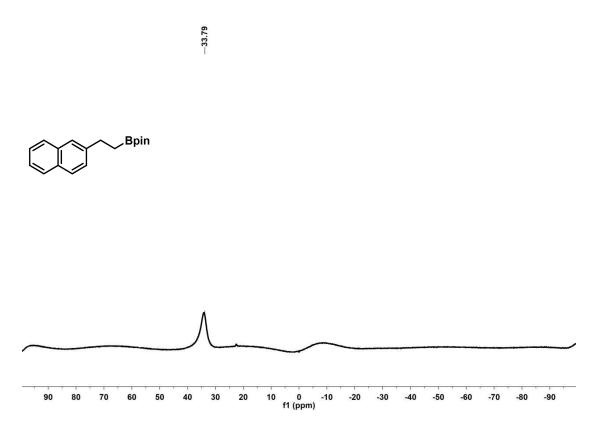


Figure S63. <sup>11</sup>B NMR spectrum of 3r in CDCl<sub>3</sub>.

13	87	13	56	48	42	26
8	-	2	2	2	5	5

-3.27

71.35

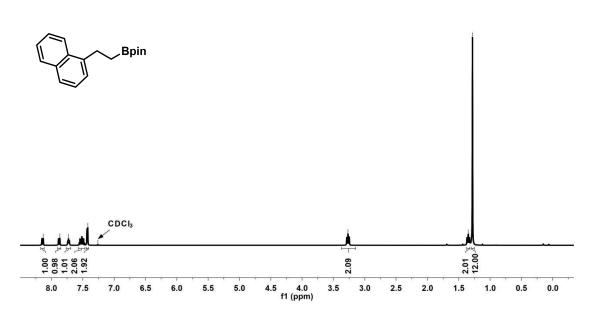


Figure S64. <sup>1</sup>H NMR spectrum of 3s in CDCl<sub>3</sub>.

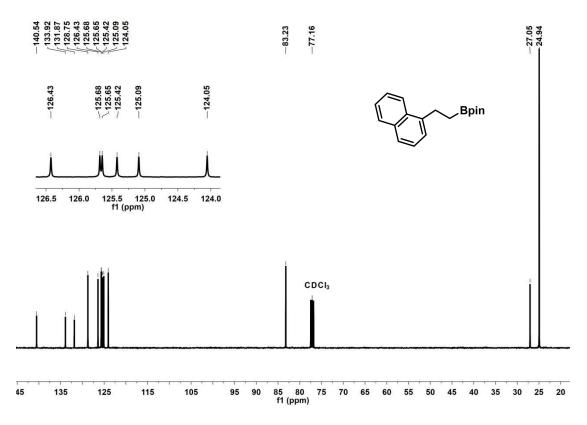
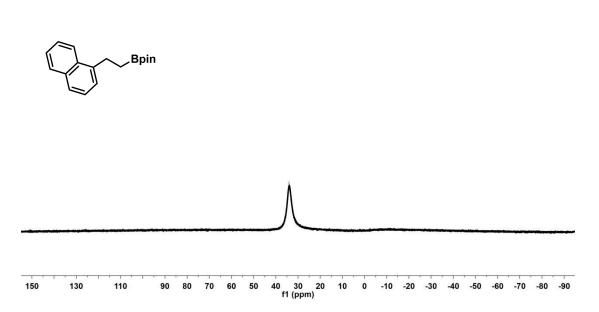


Figure S65. <sup>13</sup>C NMR spectrum of 3s in CDCl<sub>3</sub>.



-34.08

Figure S66. <sup>11</sup>B NMR spectrum of 3s in CDCl<sub>3</sub>.

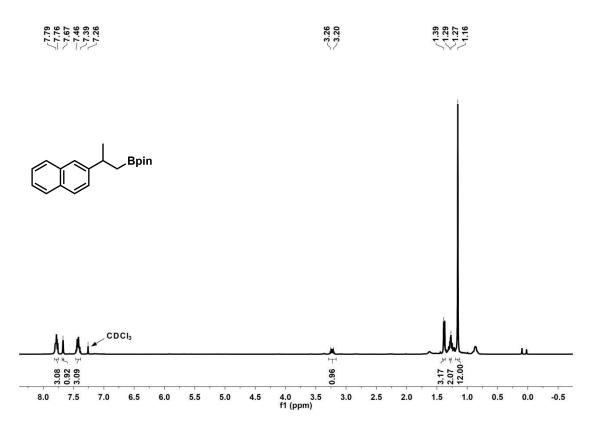


Figure S67. <sup>1</sup>H NMR spectrum of 3t in CDCl<sub>3</sub>.

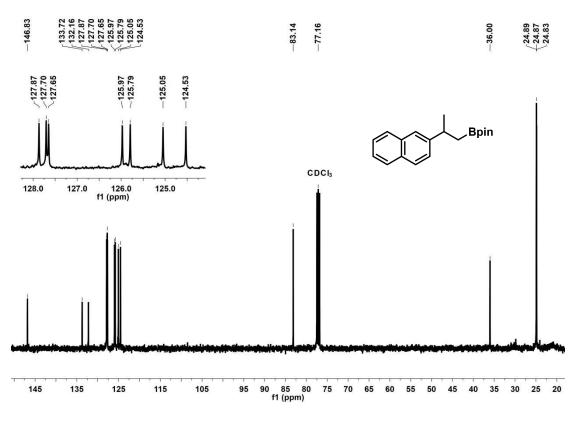


Figure S68. <sup>13</sup>C NMR spectrum of 3t in CDCl<sub>3</sub>.

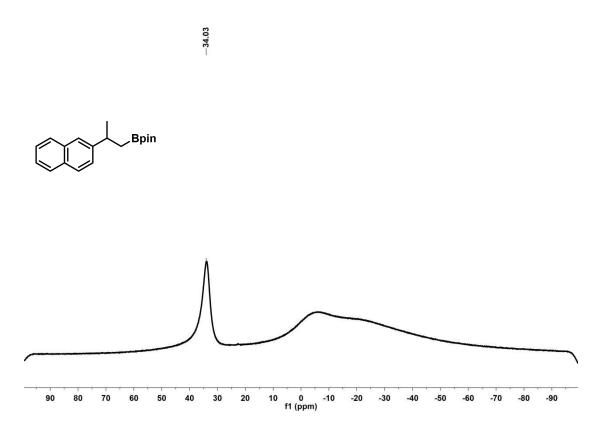


Figure S69. <sup>11</sup>B NMR spectrum of 3t in CDCl<sub>3</sub>.

-2.51 1.20 1.17 1.17

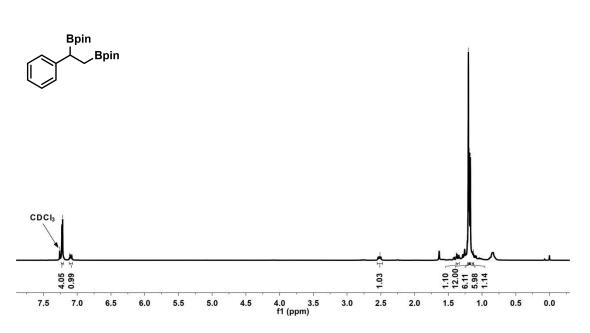


Figure S70. <sup>1</sup>H NMR spectrum of 7a in CDCl<sub>3</sub>.

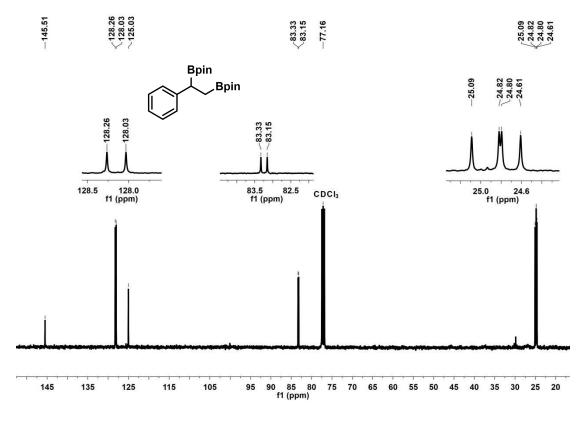


Figure S71. <sup>13</sup>C NMR spectrum of 7a in CDCl<sub>3</sub>.

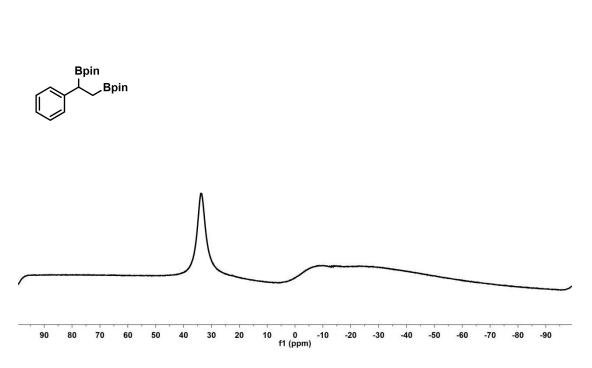


Figure S72. <sup>11</sup>B NMR spectrum of 7a in CDCl<sub>3</sub>.

-34.03

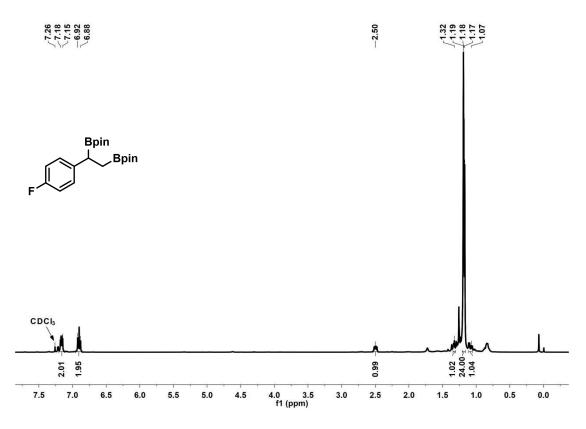


Figure S73. <sup>1</sup>H NMR spectrum of 7b in CDCl<sub>3</sub>.

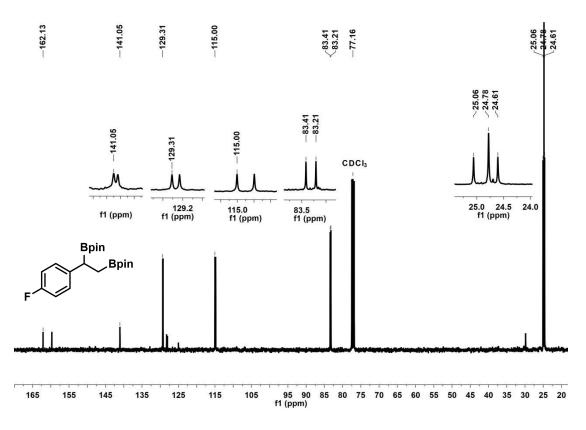


Figure S74. <sup>13</sup>C NMR spectrum of 7b in CDCl<sub>3</sub>.

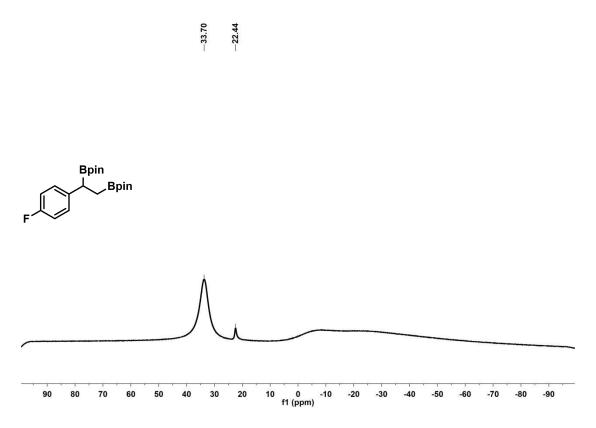


Figure S75. <sup>11</sup>B NMR spectrum of 7b in CDCl<sub>3</sub>.

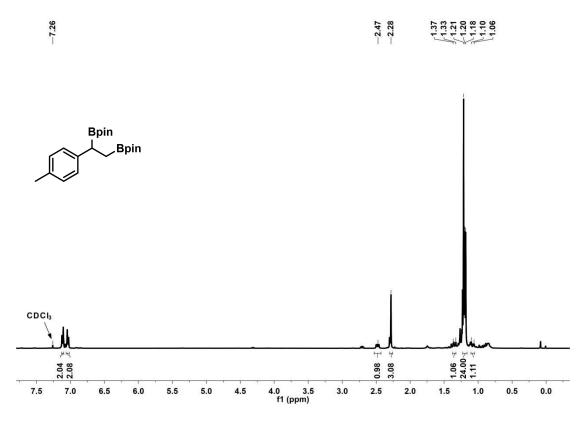


Figure S76. <sup>1</sup>H NMR spectrum of 7g in CDCl<sub>3</sub>.

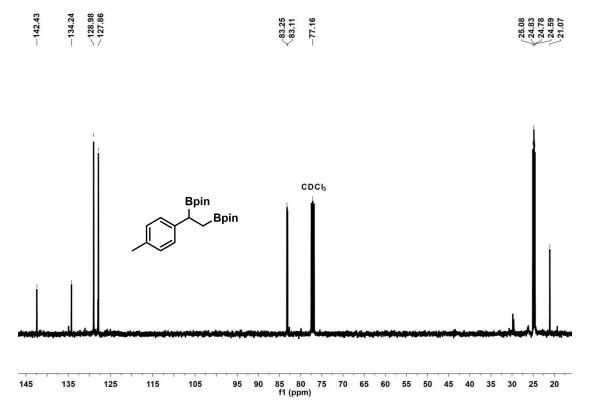


Figure S77. <sup>13</sup>C NMR spectrum of 7g in CDCl<sub>3</sub>.

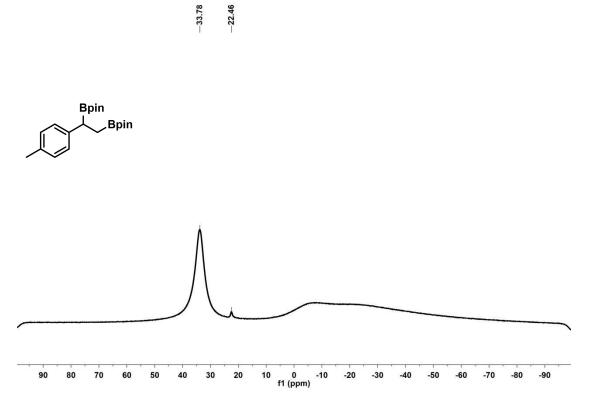


Figure S78. <sup>11</sup>B NMR spectrum of 7g in CDCl<sub>3</sub>.

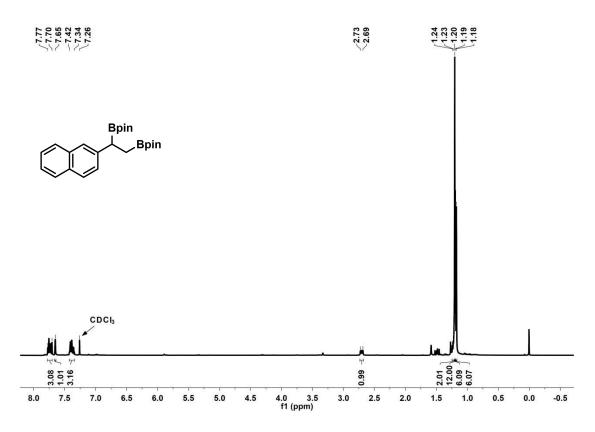


Figure S79. <sup>1</sup>H NMR spectrum of 7r in CDCl<sub>3</sub>.

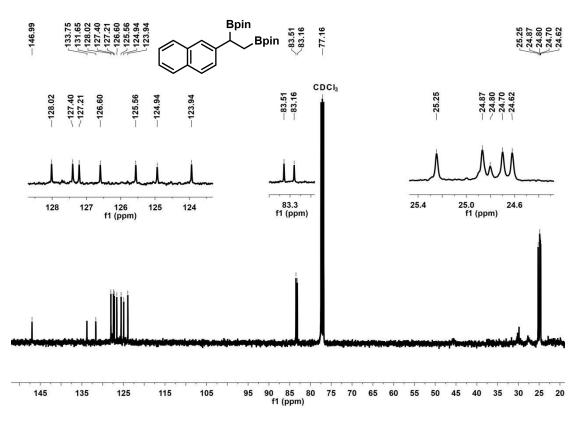


Figure S80. <sup>13</sup>C NMR spectrum of 7r in CDCl<sub>3</sub>.

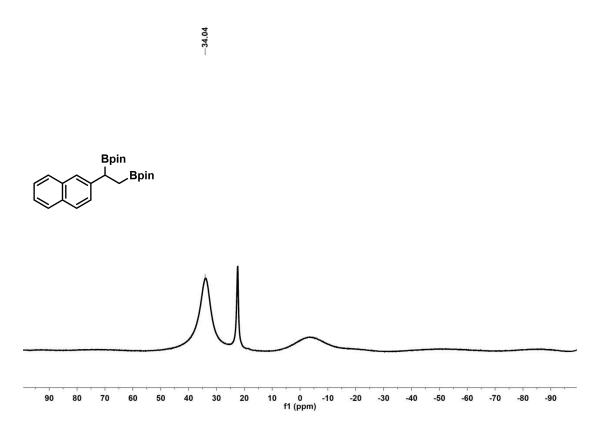


Figure S81. <sup>11</sup>B NMR spectrum of 7r in CDCl<sub>3</sub>.

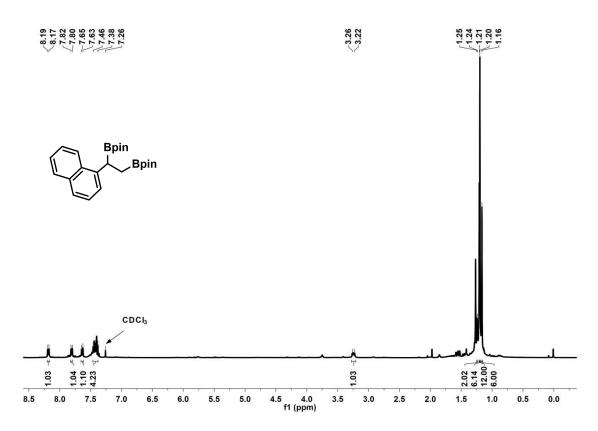


Figure S82. <sup>1</sup>H NMR spectrum of 7s in CDCl<sub>3</sub>.

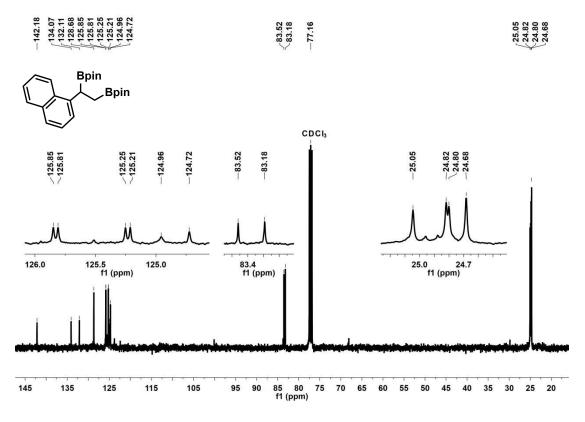
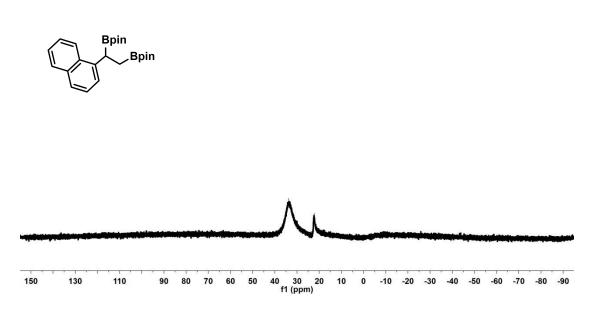


Figure S83. <sup>13</sup>C NMR spectrum of 7s in CDCl<sub>3</sub>.



-33.78 -22.26

Figure S84. <sup>11</sup>B NMR spectrum of 7s in CDCl<sub>3</sub>.

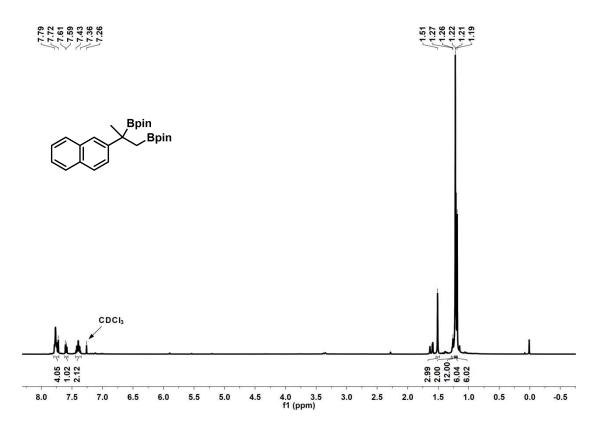


Figure S85. <sup>1</sup>H NMR spectrum of 7t in CDCl<sub>3</sub>.

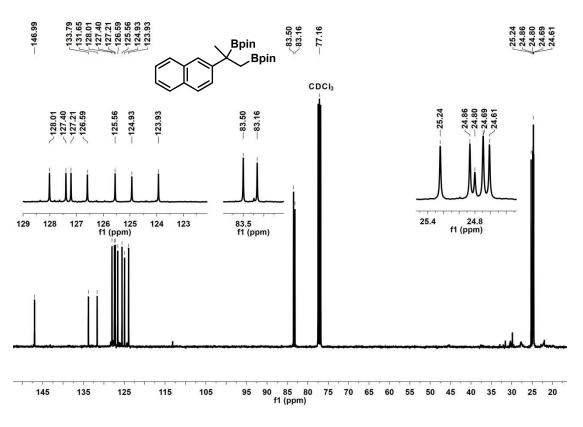


Figure S86. <sup>13</sup>C NMR spectrum of 7t in CDCl<sub>3</sub>.

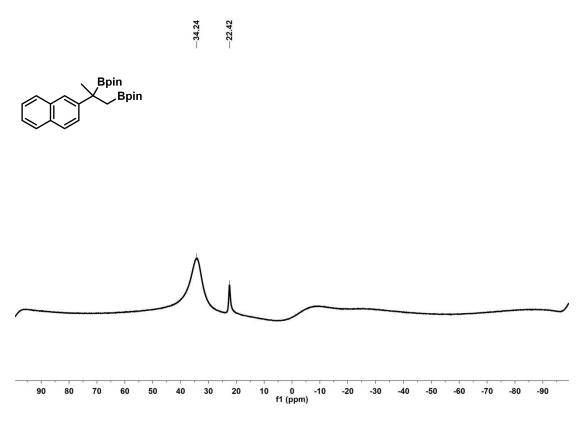


Figure S87. <sup>11</sup>B NMR spectrum of 7t in CDCl<sub>3</sub>.

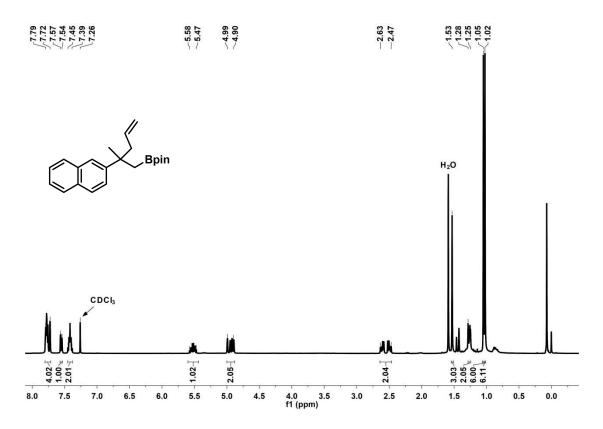


Figure S88. <sup>1</sup>H NMR spectrum of 11 in CDCl<sub>3</sub>.

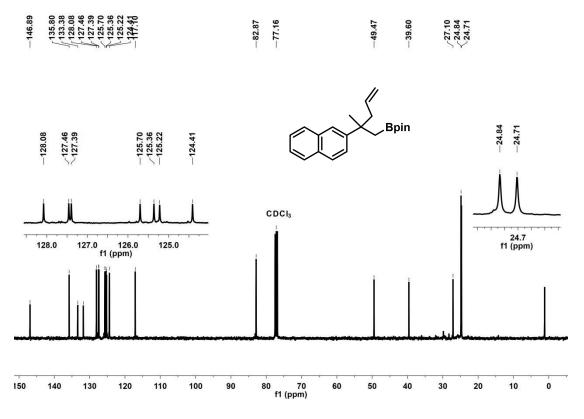
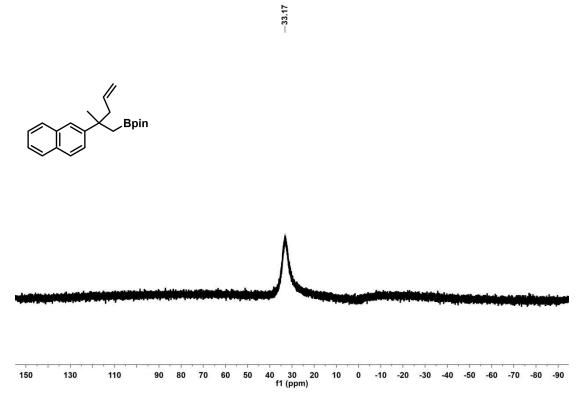


Figure S89. <sup>13</sup>C NMR spectrum of 11 in CDCl<sub>3</sub>.



SFigure S90. <sup>11</sup>B NMR spectrum of 11 in CDCl<sub>3</sub>.

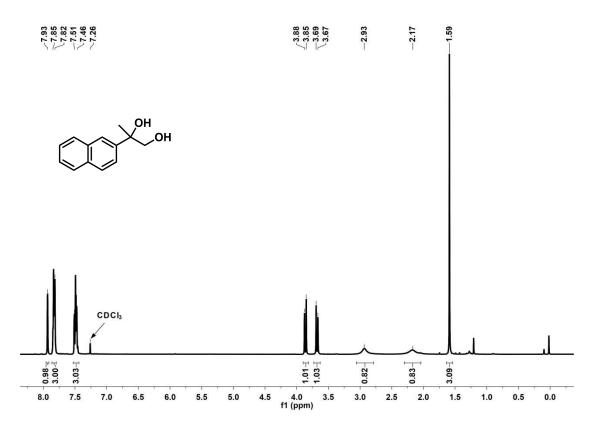


Figure S91. <sup>1</sup>H NMR spectrum of 12 in CDCl<sub>3</sub>.

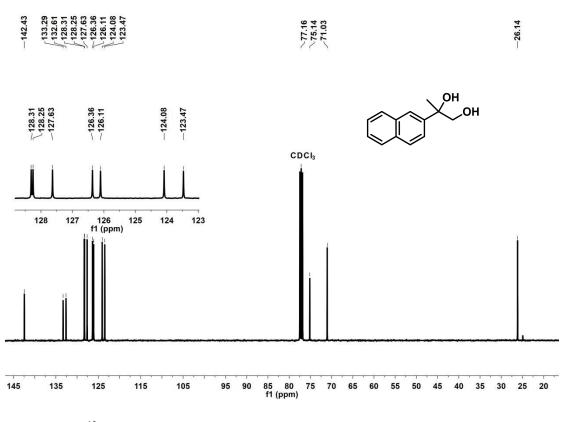
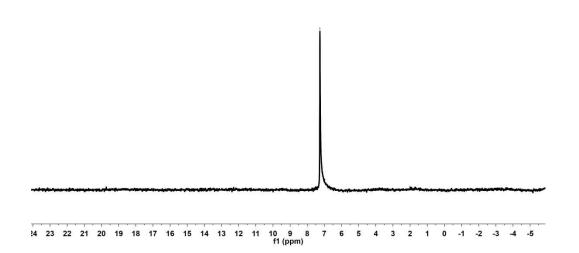
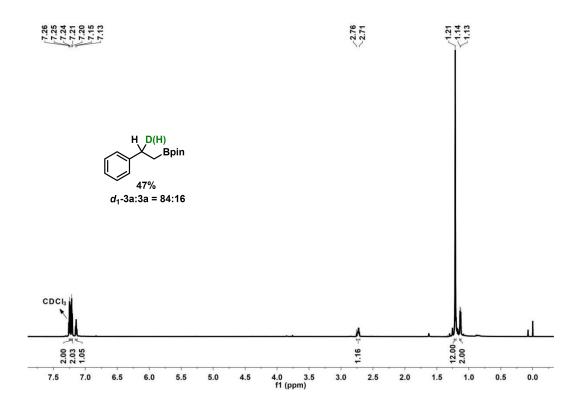


Figure S92. <sup>13</sup>C NMR spectrum of 12 in CDCl<sub>3</sub>.



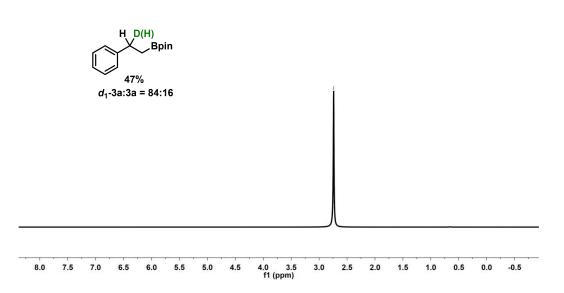


**Figure S93.** <sup>2</sup>D NMR spectrum (CDCl<sub>3</sub>) of deuterium-labelling experiment in THF- $d_8$ . No deuterium product was detected.

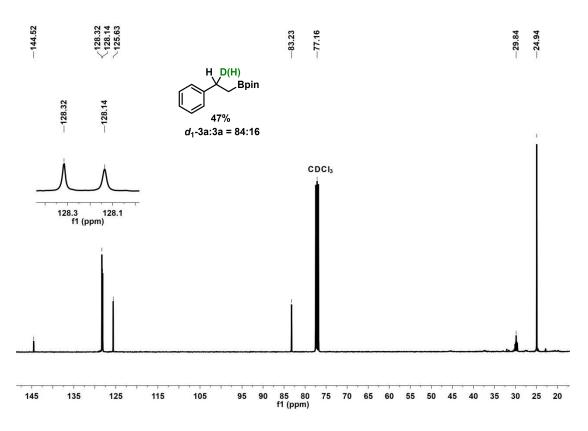


**Figure S94.** <sup>1</sup>H NMR spectrum of the mixture of  $d_1$ -**3a** and **3a** in a ratio of 84:16 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted in CD<sub>3</sub>CN solvent.





**Figure S95.** <sup>2</sup>D NMR spectrum of the mixture of  $d_1$ -**3a** and **3a** in a ratio of 84:16 in CHCl<sub>3</sub>. The deuterium-labelling experiment was conducted in CD<sub>3</sub>CN solvent.



**Figure S96.** <sup>13</sup>C NMR spectrum of the mixture of  $d_1$ -**3a** and **3a** in a ratio of 84:16 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted in CD<sub>3</sub>CN solvent.

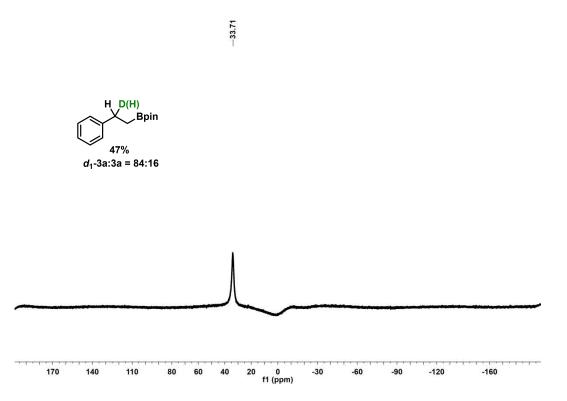


Figure S97. <sup>11</sup>B NMR spectrum of the mixture of  $d_1$ -3a and 3a in a ratio of 84:16 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted in CD<sub>3</sub>CN solvent.

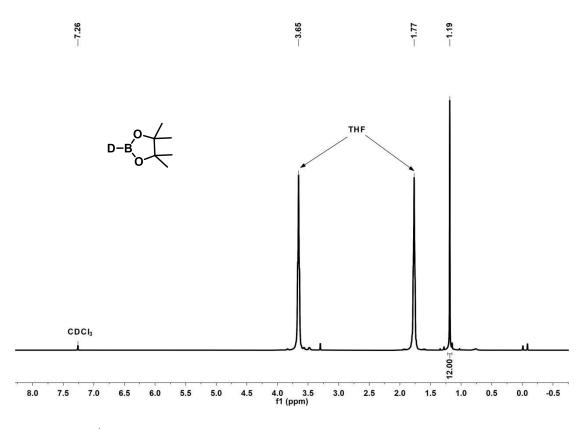


Figure S98. <sup>1</sup>H NMR spectrum of DBpin in CDCl<sub>3</sub>.

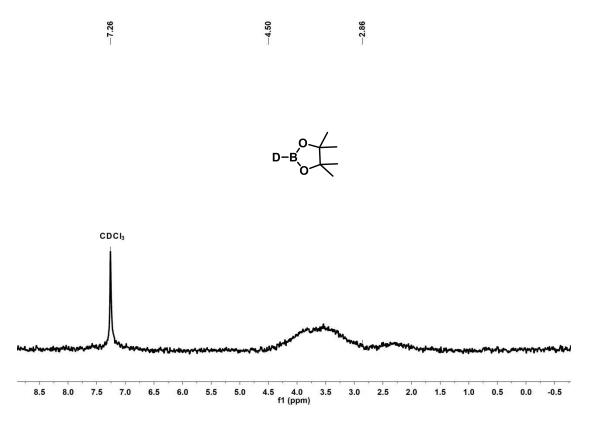


Figure S99. <sup>2</sup>D NMR spectrum of DBpin in CDCl<sub>3</sub>.

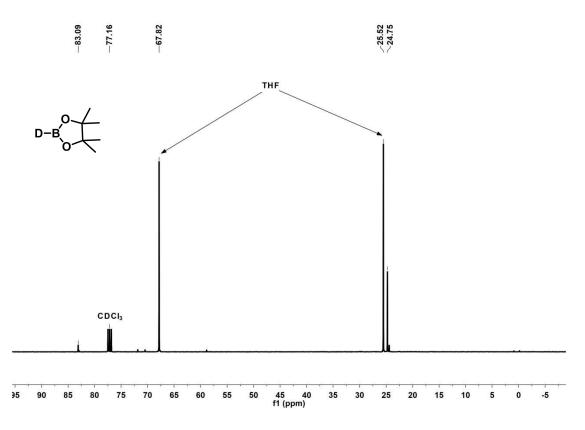
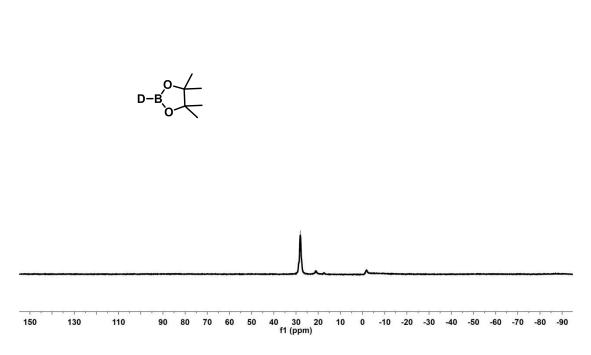


Figure S100. <sup>13</sup>C NMR spectrum of DBpin in CDCl<sub>3</sub>.



-28.08

Figure S101. <sup>11</sup>B NMR spectrum of DBpin in CDCl<sub>3</sub>.

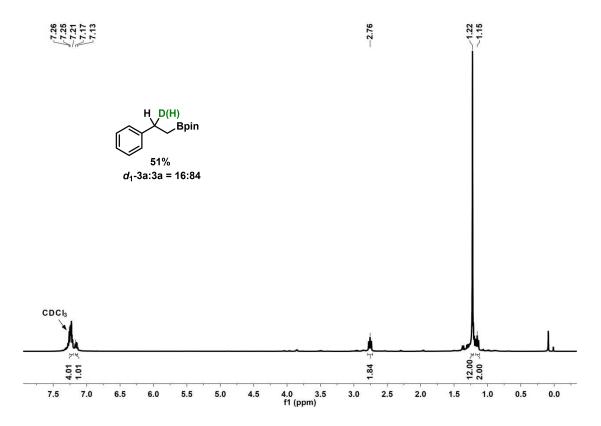


Figure S102. <sup>1</sup>H NMR spectrum of the mixture of  $d_1$ -3a and 3a in a ratio of 16:84 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with DBpin as substrate.

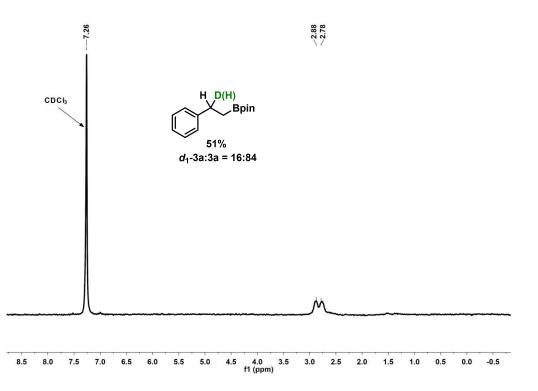
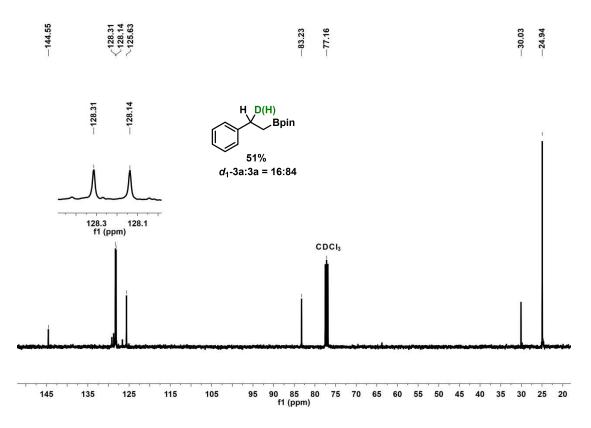
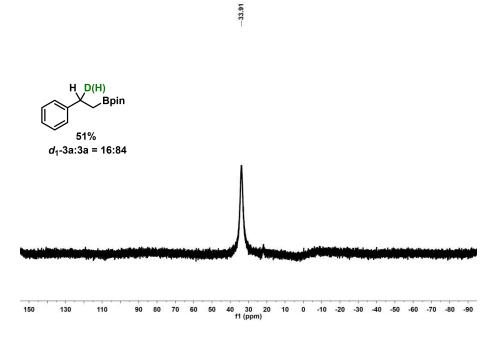


Figure S103. <sup>2</sup>D NMR spectrum of the mixture of  $d_1$ -3a and 3a in a ratio of 16:84 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with DBpin as substrate.



**Figure S104.** <sup>13</sup>C NMR spectrum of the mixture of  $d_1$ -**3a** and **3a** in a ratio of 16:84 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with DBpin as substrate.



**Figure S105.** <sup>11</sup>B NMR spectrum of the mixture of  $d_1$ -**3a** and **3a** in a ratio of 16:84 in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with DBpin as substrate.

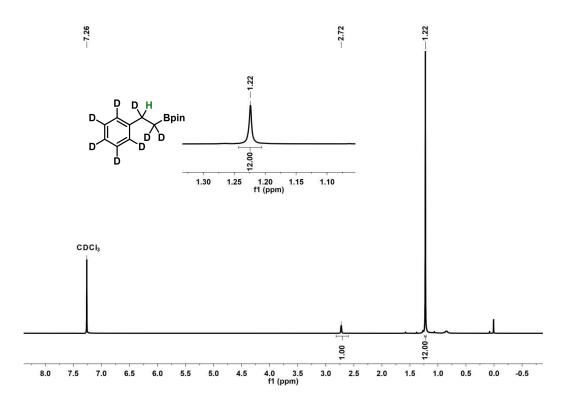


Figure S106. <sup>1</sup>H NMR spectrum of  $d_8$ -3a in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with  $d_8$ -styrene as substrate.

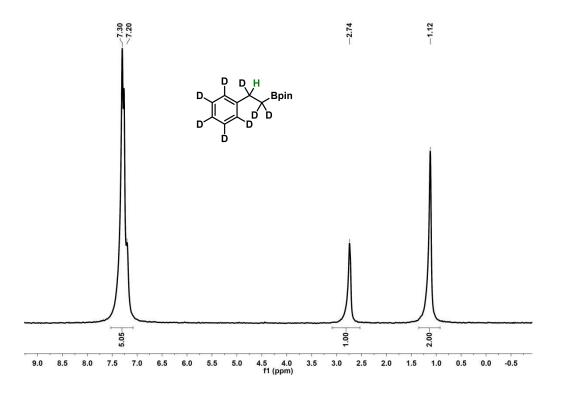


Figure S107. <sup>2</sup>D NMR spectrum of  $d_8$ -3a in CHCl<sub>3</sub>. The deuterium-labelling experiment was conducted with  $d_8$ -styrene as substrate.

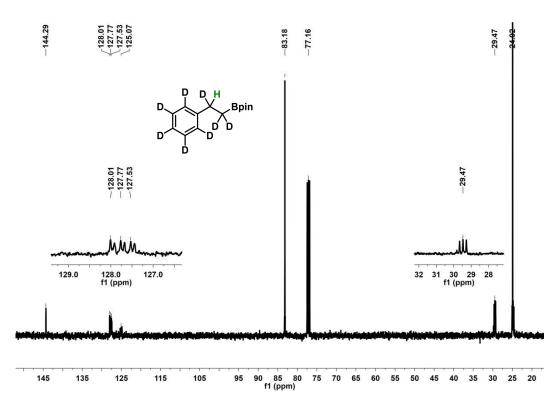


Figure S108. <sup>13</sup>C NMR spectrum of  $d_8$ -3a in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with  $d_8$ -styrene as substrate.

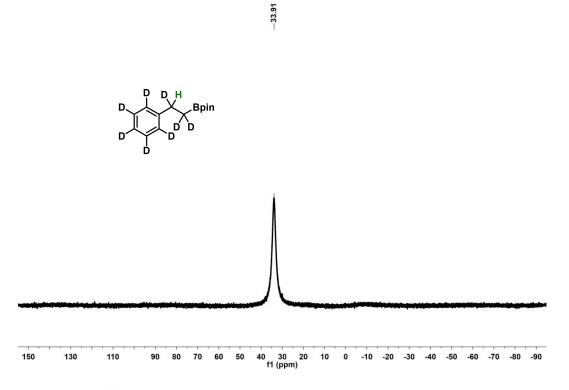
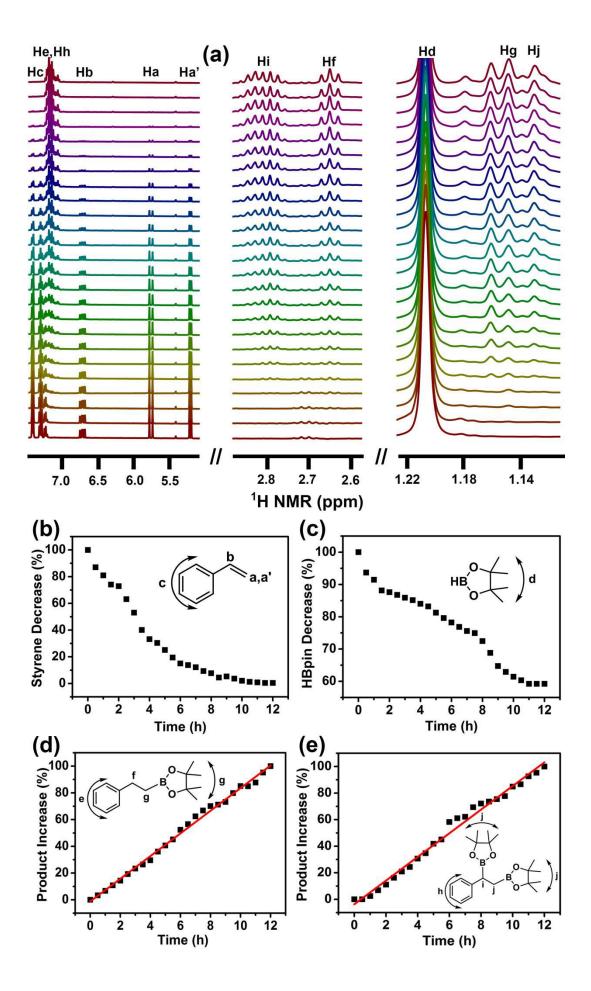
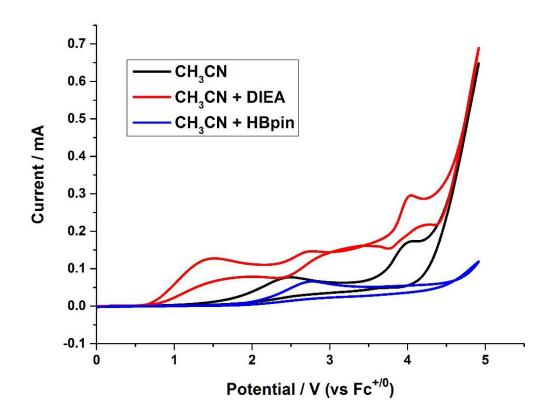


Figure S109. <sup>11</sup>B NMR spectrum of  $d_8$ -3a in CDCl<sub>3</sub>. The deuterium-labelling experiment was conducted with  $d_8$ -styrene as substrate.



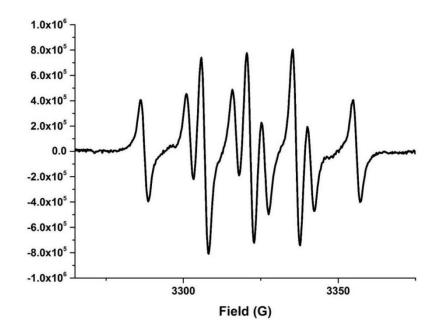
**Figure S110.** *In situ* <sup>1</sup>H NMR data of electrochemical hydroboration of styrene (10 mmol) with HBpin (88 mmol). **a** <sup>1</sup>H NMR spectra were collected every 0.5 h. The test samples were made up of 0.3 mL original samples and 0.2 mL CD<sub>3</sub>CN. Total volume of the reaction solution was 100 mL. **b** Decreased trend of styrene under electrolysis, determined by the *in situ* <sup>1</sup>H NMR data. Initial peak area of CH group at the benzyl site of styrene was defined as 100%. **c** Decreased trend of HBpin under electrolysis, determined by the *in situ* <sup>1</sup>H NMR data. Initial peak area of CH<sub>3</sub> group of HBpin was defined as 100%. **d** Increased trend of product **3a** under electrolysis, determined by the *in situ* <sup>1</sup>H NMR data. Final peak area of CH<sub>2</sub> at the benzyl site of **3a** was defined as 100%. **e** Increased trend of product **7a** under electrolysis, determined by the *in situ* <sup>1</sup>H NMR data. Final peak area of CH<sub>2</sub> at the benzyl site of **3a** was defined as 100%. **e** Increased trend of product **7a** under electrolysis, determined by the *in situ* <sup>1</sup>H NMR data. Final peak area of CH<sub>2</sub> at the benzyl site of **3a** was defined as 100%. **e** Increased trend of product **7a** under electrolysis, determined by the *in situ* <sup>1</sup>H NMR data. Final peak area of CH<sub>2</sub> at the benzyl site of **3a** was defined as 100%.



# IX. Cyclic Voltammogram

**Figure S111.** Cyclic voltammogram of DIEA (10 mM) and HBpin (10 mM) in <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub>/CH<sub>3</sub>CN at room temperature.

#### X. EPR Spectra



**Figure S112.** EPR spectrum under the standard conditions. Standard conditions: styrene (1.0 mmol), HBpin (1.1 mmol), <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%), DIEA (0.8 mmol), DMPO (1.0 mmol), Pt(+)|Pt(-), constant current (I) = 15 mA, CH<sub>3</sub>CN:THF = 4:1 (v/v), the total volume of the solvent is 10 mL, N<sub>2</sub>, rt, 0.5 h. One type of radical had been trapped by DMPO: DMPO–H.

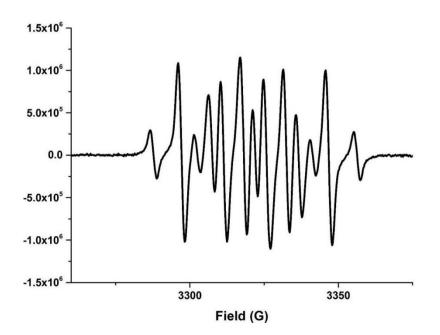
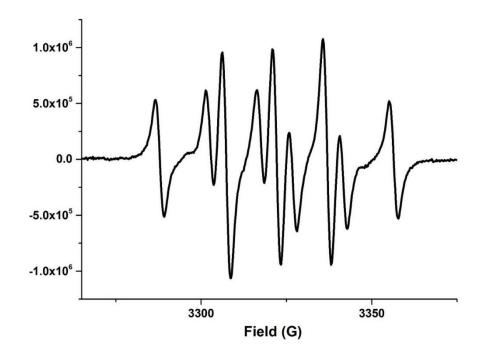
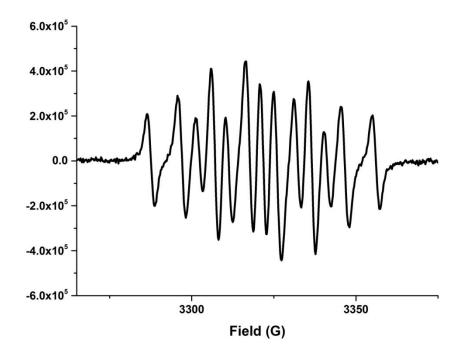


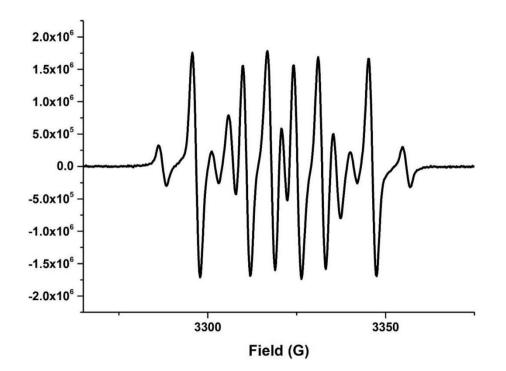
Figure S113. EPR spectrum to monitor the electrochemical reaction in the absence of HBPin. Two type of radicals had been trapped by DMPO: DMPO-H and DMPO- $CH_2CN$  (10).



**Figure S114.** EPR spectrum to monitor the electrochemical reaction in the absence of styrene. One type of radical had been trapped by DMPO: DMPO–H.



**Figure S115.** EPR spectrum to monitor the electrochemical reaction in the absence of styrene and HBpin. Two type of radicals had been trapped by DMPO: DMPO–H and DMPO–CH<sub>2</sub>CN (**10**).



**Figure S116.** EPR spectrum to monitor the electrochemical reaction in the absence of styrene, HBpin and DIEA. Two type of radicals had been trapped by DMPO: DMPO–H and DMPO–CH<sub>2</sub>CN (**10**).

### **XI. Mass Spectra**

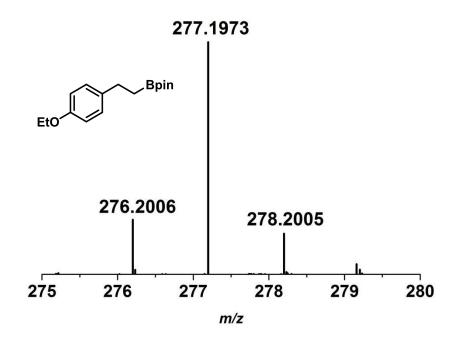


Figure S117. ESI-HRMS spectrum of 31 in CH<sub>3</sub>CN.

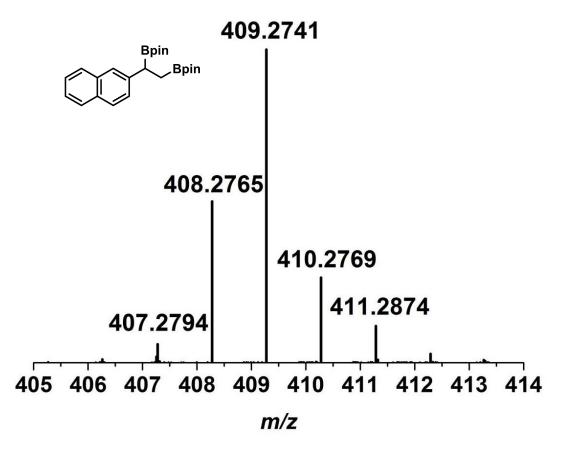


Figure S118. ESI-HRMS spectrum of 7r in CH<sub>3</sub>CN.

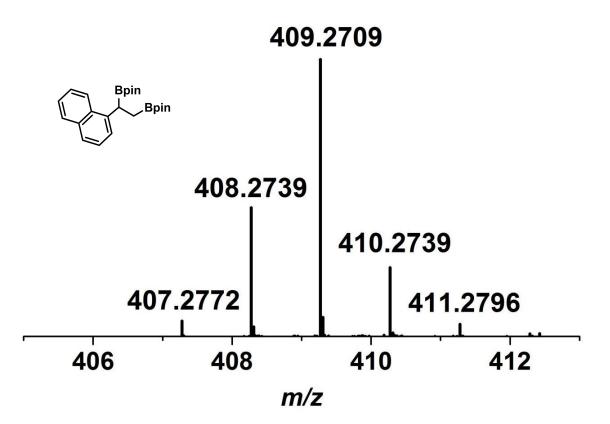


Figure S119. ESI-HRMS spectrum of 7s in CH<sub>3</sub>CN.

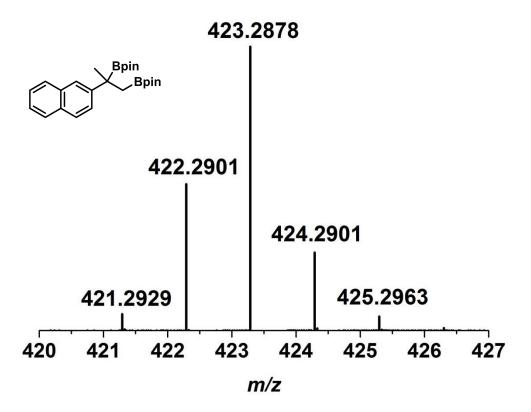
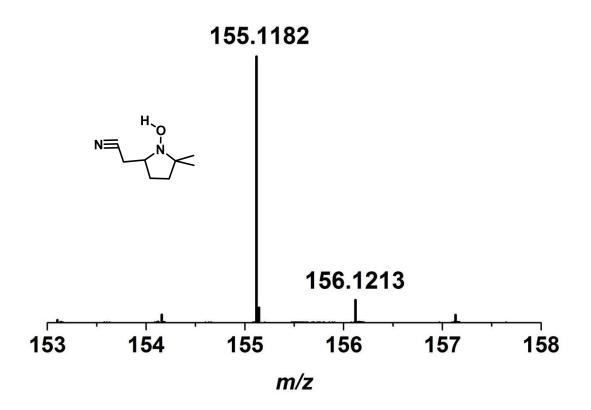


Figure S120. ESI-HRMS spectrum of 7t in CH<sub>3</sub>CN.



**Figure S121.** ESI-HRMS spectrum of reduction state of the corresponding adduct DMPO–CH<sub>2</sub>CN (**10**) in CH<sub>3</sub>CN/THF mixture.

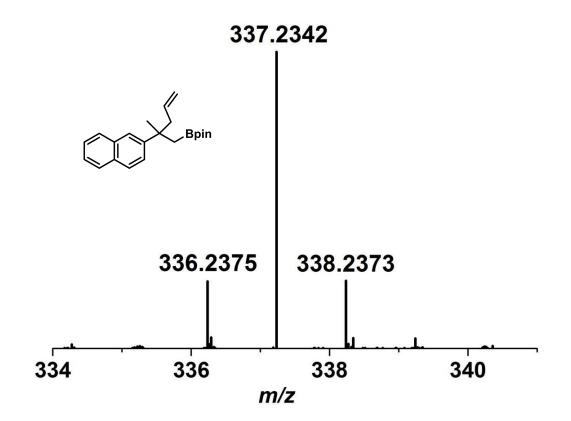


Figure S122. ESI-HRMS spectrum of 11 in CH<sub>3</sub>CN.

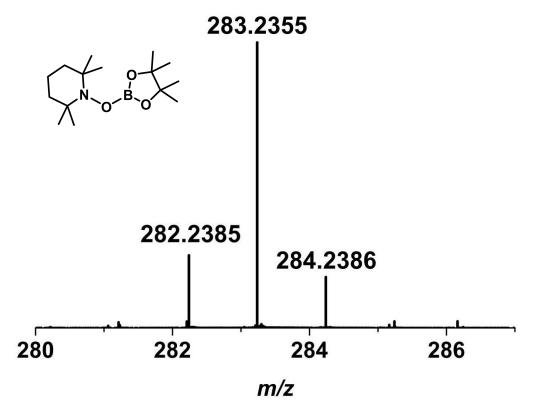


Figure S123. GC-MS spectrum of 13 in CH<sub>3</sub>CN.

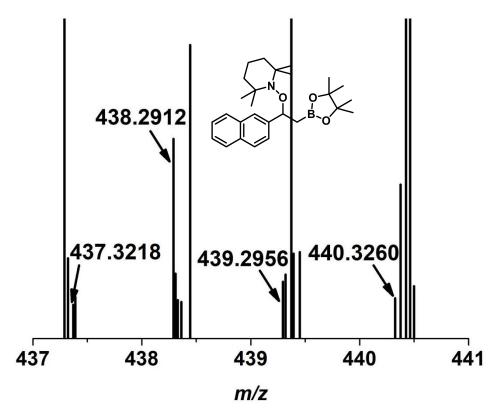


Figure S124. ESI-HRMS spectrum of 14 in CH<sub>3</sub>CN.

# XII. FT-IR (ATR) Spectra

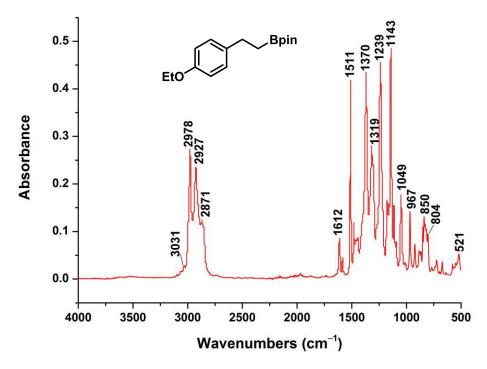


Figure S125. FT-IR (ATR) spectrum of 3l.

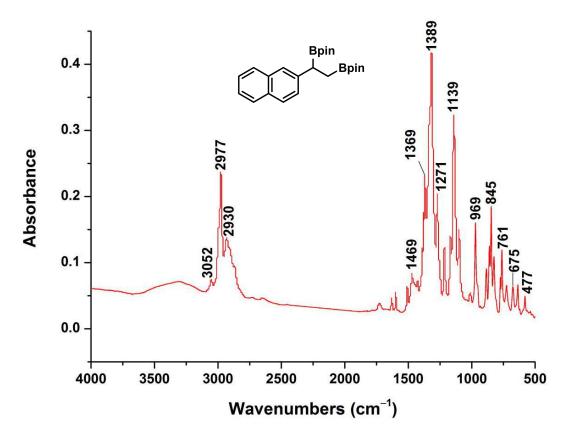


Figure S126. FT-IR (ATR) spectrum of 7r.

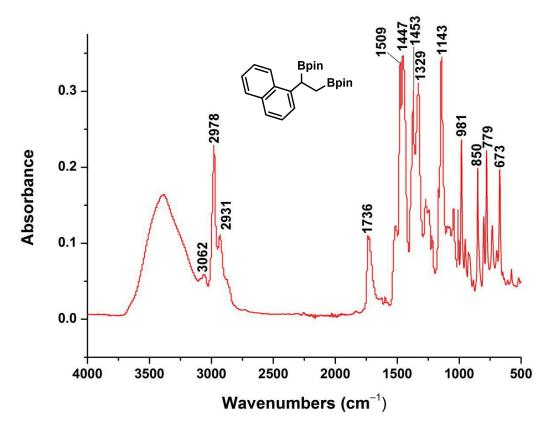


Figure S127. FT-IR (ATR) spectrum of 7s.

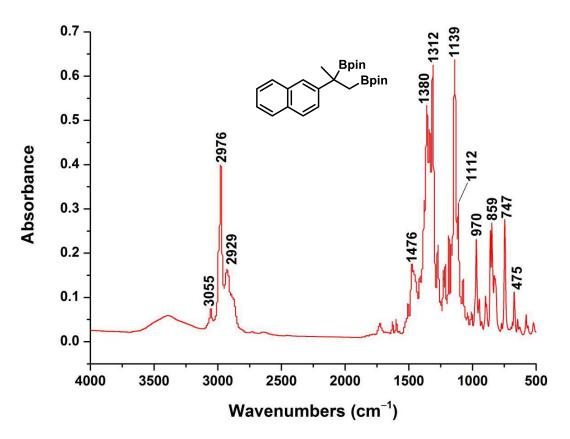


Figure S128. FT-IR (ATR) spectrum of 7t.

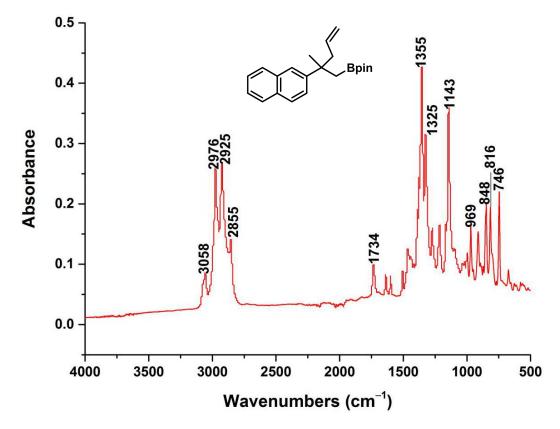
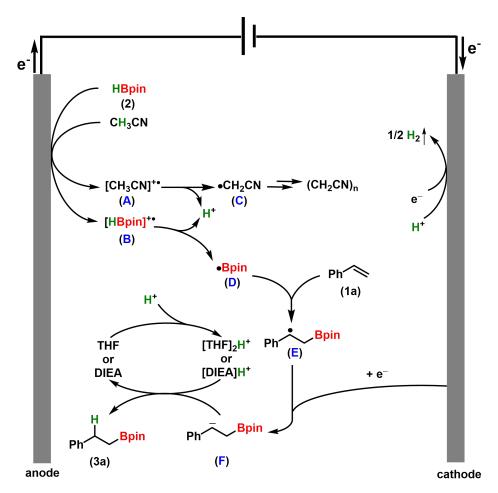


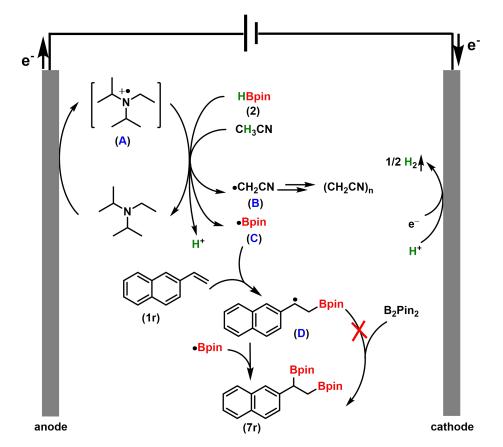
Figure S129. FT-IR (ATR) spectrum of 11.

XIII. Proposed Electrochemical Cycle in the Absence of DIEA



**Figure S130.** A possible mechanism of electrochemical hydroboration with styrene as substrate in the absence of DIEA.

# XIV. Proposed Electrochemical Cycle for Diboration



**Figure S131.** A possible mechanism of electrochemical hydroboration with 2-vinylphthalene as substrate.

Using <sup>*n*</sup>Bu<sub>4</sub>NBF<sub>4</sub> (20 mol%) as the supporting electrolyte and CH<sub>3</sub>CN:THF (*v:v* = 4:1) as the mixed solvent, with the addition of 0.8 equivalent of DIEA, and 15 mA constant current at room temperature for 3 h under inert atmosphere, both hydroboration product (**3r**) and diboronate ester (**7r**) were detected with 1.0 equivalent of 2-vinylphthalene and 1.1 equivalents of HBpin as substrates (Scheme 1 in manuscript). The yield of product **7r** did not increase when additional 3.3 equivalents of B<sub>2</sub>Pin<sub>2</sub> were added to this system. In addition, no diboronate esters **7r** was observed when B<sub>2</sub>Pin<sub>2</sub> as the only source of boron. These results indicated that the formation of diboronate ester was not associated with B<sub>2</sub>Pin<sub>2</sub> and hydroboration product could not transfer to diboronate ester in the presence of HBpin or B<sub>2</sub>Pin<sub>2</sub>.