Supplementary Information

Energy transfer from WT-LH2 to ΔcrtB RCLH1 under physiological conditions

In contrast to the data in Fig. 5, obtained with surface-patterned LH complexes that were partially dehydrated and then sealed under a protective Argon atmosphere, the data for Fig S1 were obtained with surface-attached complexes sealed in Argon-sparged imaging buffer. Fig. S1A shows merged wide field false colour fluorescence image of complexes illuminated by a 470 nm LED recorded in epifluorescence mode. The green lines correspond to fluorescence emission filtered by a 857/30 nm bandpass filter, which indicates the distribution of LH2 complexes. The red squares correspond to fluorescence filtered by a 900/32 nm bandpass filter, which identifies the location of $\Delta crtB$ RCLH1 complexes. In the absence of coupled LH2 complexes the $\Delta crtB$ RCLH1 complexes cannot harvest energy because of the absence of carotenoids and the 470 nm excitation, so only the $\Delta crtB$ RCLH1 complexes in the intersecting cross-over area fluoresce because nearby LH2 complexes deliver excitation energy. This is confirmed when analysing the fluorescence spectral images when the sample is excited with a 485 nm laser. The fluorescence intensity images at 857 nm and 890 nm



Fig. S1. Spectral and lifetime fluorescence data of cross-patterned LH2 and $\Delta crtB$ RCLH1 complexes on a functionalised glass substrate imaged under physiological conditions (in imaging buffer). A – False colour fluorescence image (470 nm wide field excitation), showing the LH2 (green) and $\Delta crtB$ RCLH1 (red) lines, filtered by 857/30 nm and 900/32 nm bandpass filters, respectively. B - Spectral map showing the emission intensity at 860 nm (LH2 emission); the excitation source is 485 nm pulsed laser, scan size 32 μ m. C – Simultaneously recorded spectral map showing the emission intensity at 890 nm ($\Delta crtB$ RCLH1 emission), scan size 32 μ m. There is increased emission in the areas where LH2 is present as the energy transfer donor. The positions of three pixels used for acquiring spectral data are marked. D – Individual emission spectra recorded in the pixels of the images in panels A and B marked with 1 (LH2 only, green line), 2 (cross-over area, orange line) and 3 ($\Delta crtB$ RCLH1 only, red line), respectively. E- Intensity-averaged lifetime image obtained at 485 nm excitation and 857 nm emission (LH2 emission), clearly showing a decrease in the lifetime in the cross-over areas, where the two complexes are in close proximity, scan size 32 μ m. F – Individual decay curves recorded in the pixels of the lifetime image in panel E marked with 4 (orange, average lifetime of 395 ps) and 5 (green, average lifetime of 653 ps), respectively.

are shown in (Figs. S1, B and C, respectively) reveal $\Delta crtB$ RCLH1 emission arising only from the crossover areas of the pattern.

The fluorescence lifetime results also provided evidence for energy transfer between LH2 and $\Delta crtB$ RCLH1. Fig. S1, E represents the amplitude weighted average lifetime image of LH2 at 857 nm with typical lifetime decay curves are shown in Fig. S1, F. Lifetime decay curves were fitted by a biexponential decay function, where the green curve represents lifetime decay in LH2 only area (marked 5) and the orange curve represents the lifetime decay in the cross-over area (marked 4). In the LH2 only area, the measured lifetime is around 640 ps (a faster component of 550ps and a slower component of 750 ps). In the cross-over area, the lifetime of LH2 emission is shortened to 440 ps (a faster component of 320ps and a slower component of 500 ps). Such a decrease in LH2 lifetime in the cross-over area indicates that EET indeed occurs between LH2 and $\Delta crtB$ RCLH1.



Fig. S2. Spectral and lifetime fluorescence data of cross-patterned LH2 and *AcrtB* RCLH1 complexes on a functionalised glass substrate obtained 60 days after the sample preparation showing a long-term stability of the cross-patterned LH2 and *AcrtB* RCLH1 complexes. The sample was sealed in Argon protective atmosphere and stored at 4°C. A - False colour fluorescence image (470 nm wide field excitation) showing the LH2 (green) and AcrtB RCLH1 (red) lines, filtered by 857/30 nm and 900/32 nm bandpass filters, respectively. B - Spectral map showing the emission intensity at 860 nm (LH2 emission); the excitation source is a 485 nm pulsed laser, scan size 32 μ m. C – Simultaneously acquired spectral map showing the emission intensity at 890 nm (Δ crtB RCLH1 emission), scan size 32 μ m. D – Individual emission spectra recorded in the pixels of the images in panels A and B marked with 1 (LH2 only, green line), 2 (cross-over area, orange line) and 3 ($\Delta crtB$ RCLH1 only, red line), respectively. The spectral deconvolution shows a clear drop in the LH2 emission (olive peak fit) and an increase in the $\Delta crtB$ RCLH1 emission (pink peak fit), compared to the LH2 emission and the $\Delta crtB$ RCLH1 emission outside the cross-over area. E- Intensity-averaged lifetime image obtained at 485nm excitation and 857nm emission (LH2 emission), clearly showing a decrease in the lifetime in the cross-over areas, where the two complexes are in close proximity, scan size 32 µm. F – Individual decay curves recorded in the pixels of the lifetime image in panel E marked with 4 (orange, average lifetime of 382 ps) and 5 (green, average lifetime of 683 ps), respectively.