

## Electronic Supporting Material

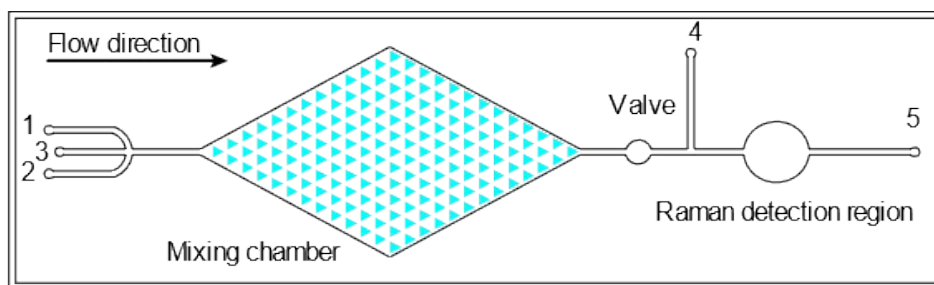


Fig. S1 CAD diagram of the microfluidic Raman chip.

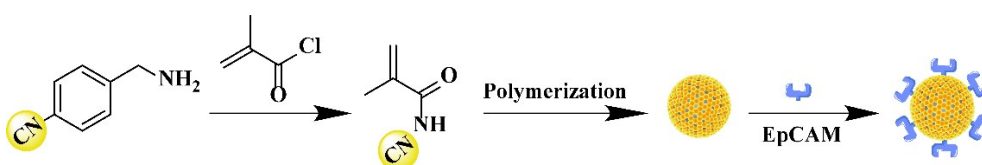


Fig. S2 Process for Raman bead synthesis.

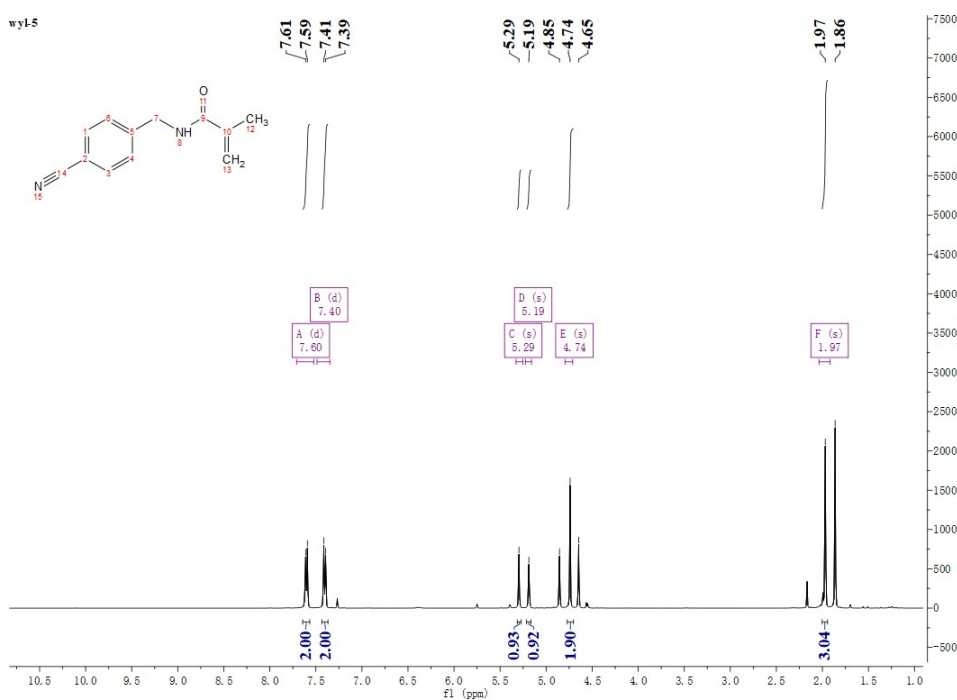
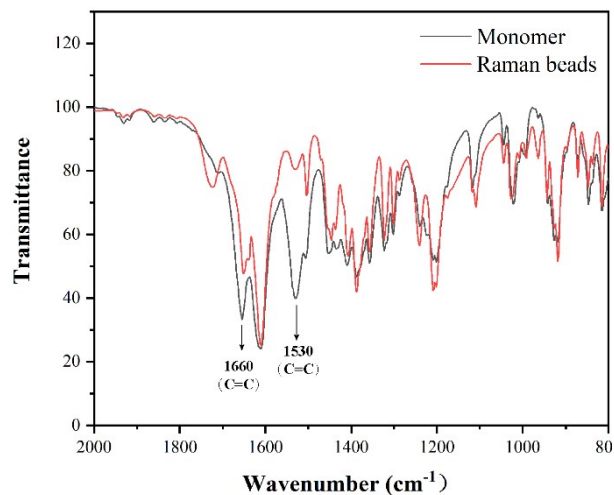
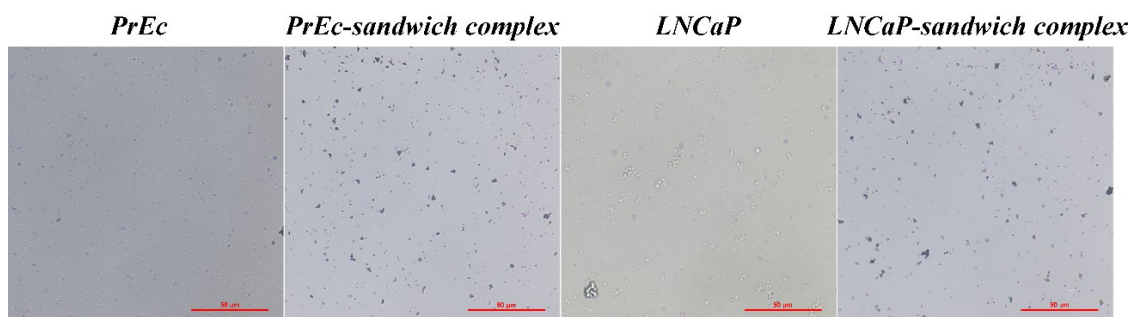


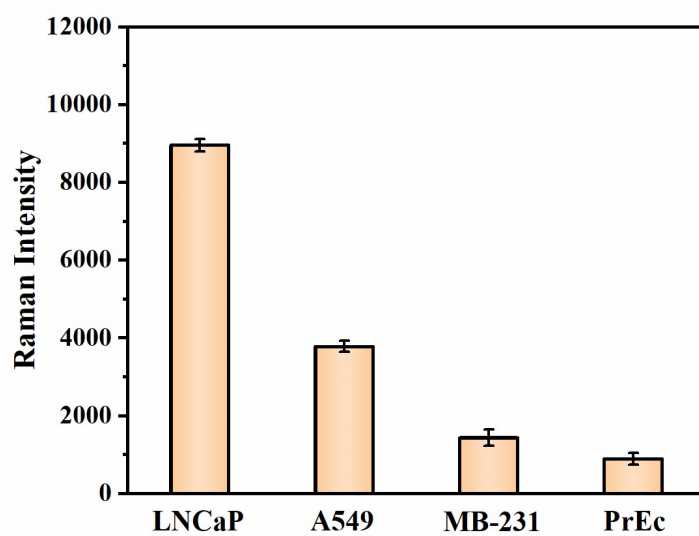
Fig. S3 NMR spectrum of the Ethylene-based Raman monomer.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.61, 7.59, 7.41, 7.39, 5.29, 5.19, 4.85, 4.74, 4.65, 1.97, 1.86.



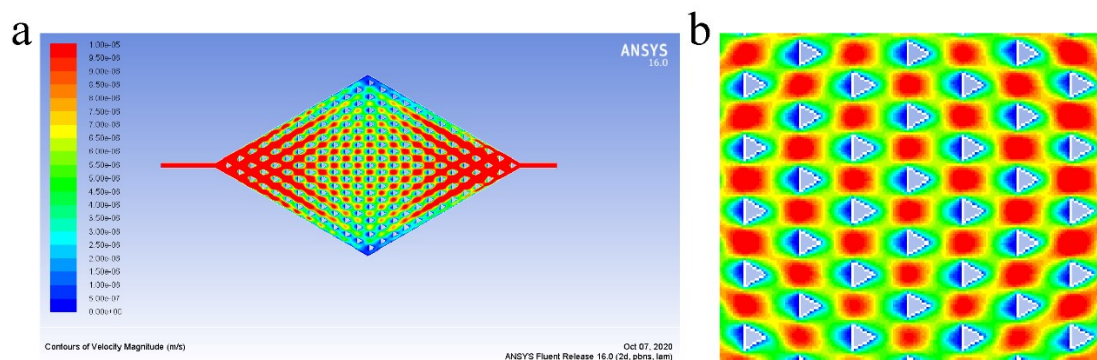
**Fig. S4** FT-IR spectra of the monomer and Raman beads.



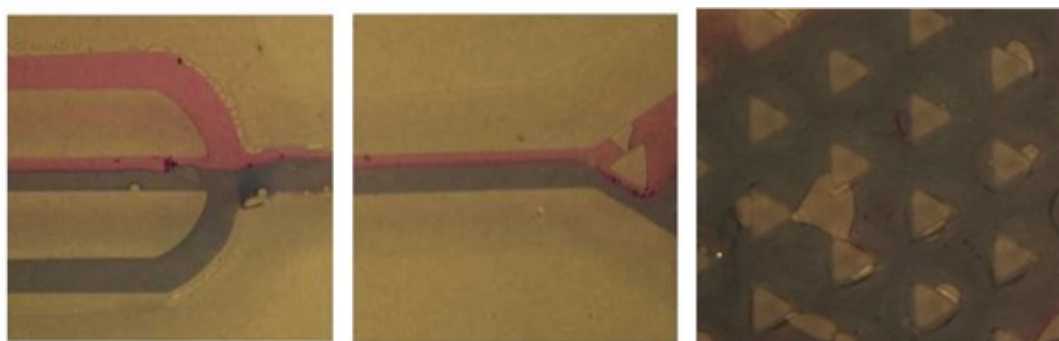
**Fig. S5** Bright-field images of PrEC exosomes, CD63-Mag/PrEC exosome/Raman beads, LNCaP exosomes, and CD63-Mag/LNCaP exosome/Raman beads. Scale bars, 50  $\mu\text{m}$ .



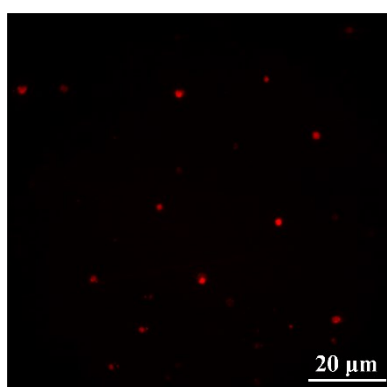
**Fig. S6** Specificity of the microfluidic Raman chip, error bars indicate standard deviation of measurements (n=3).



**Fig. S7** (a) Finite element simulations of the flow velocity profile inside the staggered triangular pillars array. (b) A larger view of the middle mixing chamber.



**Fig. S8** Photographs of red and blue inks flowing in the microfluidic channel.



**Fig. S9** The Mag-CD63-Exo complex fluorescence images on the triangular micropillar area of the chip.

**Table S1.** Information on clinical serum samples.

Patient ID	TNM	Sex	Age, years	PLT, $10^9 L^{-1}$	Pathology
------------	-----	-----	------------	--------------------	-----------

1	NA	M	53	-	Control
2	NA	M	56	-	Control
3	NA	M	52	-	Control
4	NA	M	52	-	Control
5	NA	M	51	-	Control
6	NA	M	53	-	Control
7	NA	M	55	-	Control
8	NA	M	54	-	Control
9	T2N0M0	M	71	127	Malignant
10	T1N0M0	M	75	223	Malignant
11	T3N0M0	M	74	118	Malignant
12	T2N0M0	M	72	188	Malignant
13	T4N0M1	M	78	180	Malignant
14	T1N0M0	M	76	197	Malignant
15	T2N0M0	M	75	198	Malignant
16	T2N0M0	M	77	140	Malignant
17	T2N0M0	M	59	268	Malignant
18	T2N0M0	M	79	177	Malignant

**Table S2.** Comparison of different microfluid chip methods for detection of exosomes

Target	Detection method	LOD	Volume	Detection time	Ref
PSA	Immunoassay	0.01 ng mL <sup>-1</sup>	50 µL	5 min	1
Hepatocellular exosomes	Electrochemical assay	4.39×10 <sup>3</sup> particles mL <sup>-1</sup>	30 µL	3.5 h	2
Ovarian cancer exosomes	Fluorogenic	---	>20 µL	40 min	3

	ELISA				
Ovarian cancer exosomes	Fluorogenic ELISA	$5 \times 10^4$ particles $\text{mL}^{-1}$	20 $\mu\text{L}$	>2 h	4
SKBR3, T84, and LNCaP exosomes	SERS	$3.2 \times 10^4$ particles $\text{mL}^{-1}$ $7.3 \times 10^4$ particles $\text{mL}^{-1}$ $2.03 \times 10^5$ particles $\text{mL}^{-1}$	2 $\mu\text{L}$	>2 h	5
HepG2 exosomes	SERS	$2.7 \times 10^4$ particles $\text{mL}^{-1}$	200 $\mu\text{L}$	>10 h	6
Breast cancer exosomes	SERS	$2 \times 10^6$ particles $\text{mL}^{-1}$	15 $\mu\text{L}$	2 h	7
Prostate cancer exosomes	Raman spectroscopy	$1.6 \times 10^2$ particles $\text{mL}^{-1}$	20 $\mu\text{L}$	1 h	This work

LOD, limit of detection

#### References:

1. R. Gao, Z. Lv, Y. Mao, L. Yu, X. Bi, S. Xu, J. Cui and Y. Wu, *ACS Sens*, 2019, **4**, 938-943.
2. H. Xu, C. Liao, P. Zuo, Z. Liu and B. C. Ye, *Anal Chem*, 2018, **90**, 13451-13458.
3. Z. Zhao, Y. Yang, Y. Zeng and M. He, *Lab Chip*, 2016, **16**, 489-496.
4. P. Zhang, M. He and Y. Zeng, *Lab Chip*, 2016, **16**, 3033-3042.
5. Z. Wang, S. Zong, Y. Wang, N. Li, L. Li, J. Lu, Z. Wang, B. Chen and Y. Cui, *Nanoscale*, 2018, **10**, 9053-9062.
6. Y. F. Tian, C. F. Ning, F. He, B. C. Yin and B. C. Ye, *Analyst*, 2018, **143**, 4915-4922.
7. E. A. Kwizera, R. O'Connor, V. Vinduska, M. Williams, E. R. Butch, S. E. Snyder, X. Chen and X. Huang, *Theranostics*, 2018, **8**, 2722-2738.