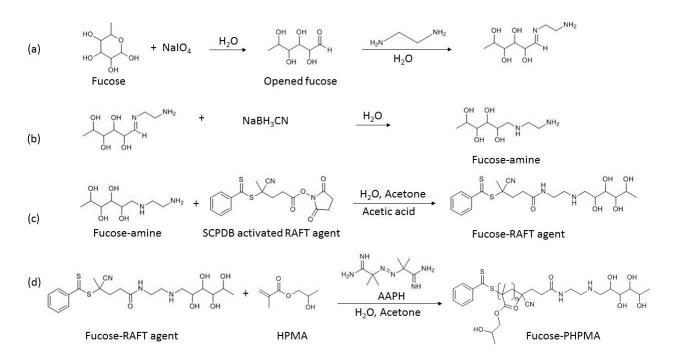
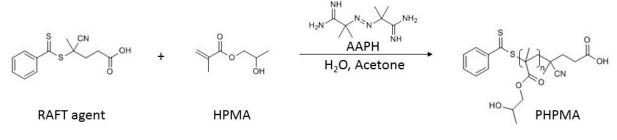
Supporting information: Fucose-Modified Thermoresponsive Poly(2-Hydroxypropyl Methacrylate) Nanoparticles for Controlled Doxorubicin Release from an Injectable Depot

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Scheme S1. The synthesis steps to fucose-PHPMA. a) The fucose conjugation to ethylenediamine. b) Reduction of the imine to fucose-amine. c) SCPDB RAFT agent conjugation to fucose-amine. d) HPMA polymerization from the fucose-RAFT agent.



Scheme S2. The synthesis of PHPMA.

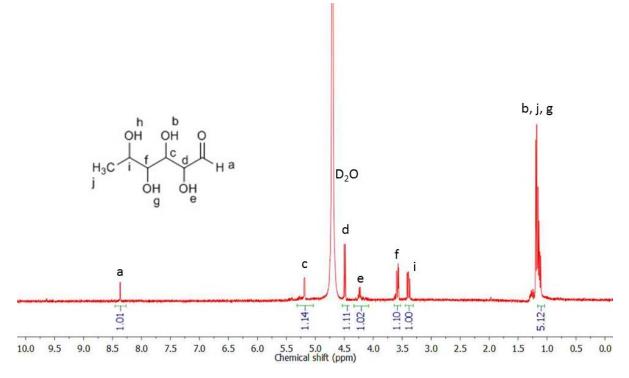


Figure S1. The 500 MHz ¹H-NMR spectrum of opened fucose in D_2O at 25 °C.

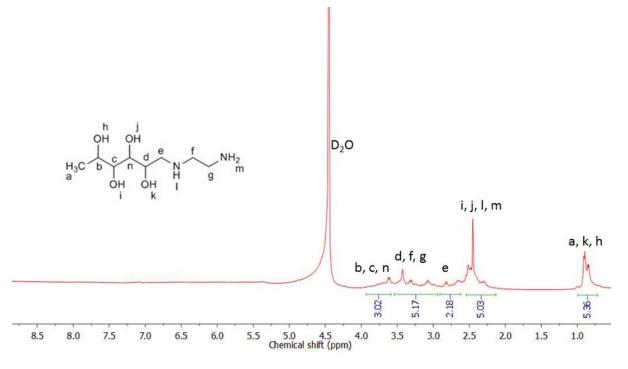


Figure S2. The 500 MHz ¹H-NMR spectrum of fucose-amine in D₂O at 25 °C.

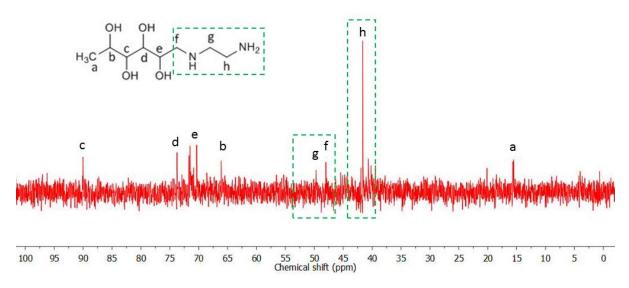


Figure S3. The 125 MHz 13 C-NMR spectrum of fucose-amine in D₂O at 25 °C.

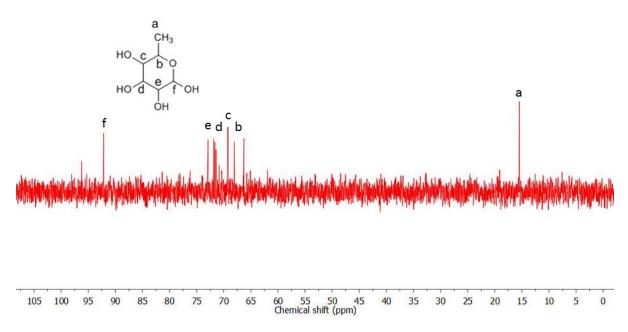


Figure S4. The 125 MHz 13 C-NMR spectrum of fucose in D₂O at 25 °C.

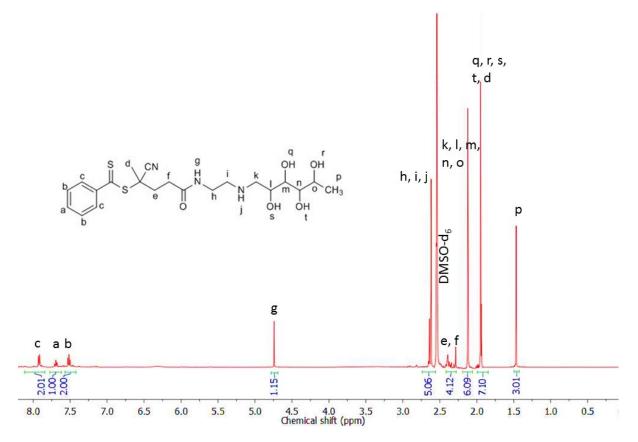


Figure S5. The 500 MHz ¹H-NMR spectrum of fucose-RAFT agent in DMSO-d₆ at 25 °C.

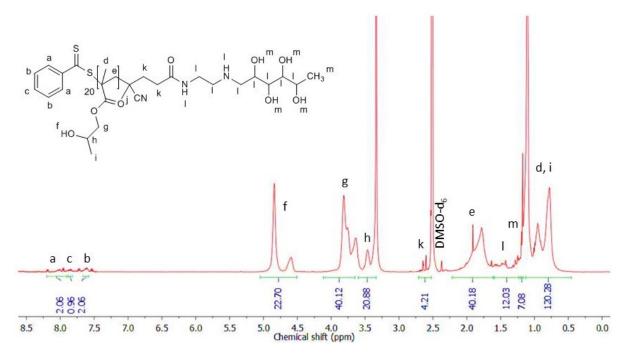


Figure S6. The 500 MHz ¹H-NMR spectrum of fucose-PHPMA₂₀ in DMSO-d₆ at 25 °C.

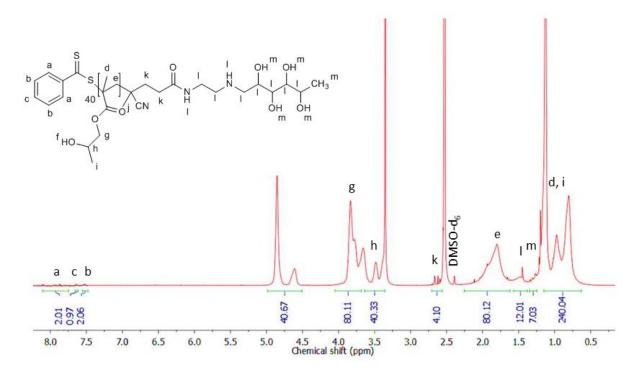


Figure S7. The 500 MHz ¹H-NMR spectrum of fucose-PHPMA₄₀ in DMSO-d₆ at 25 °C.

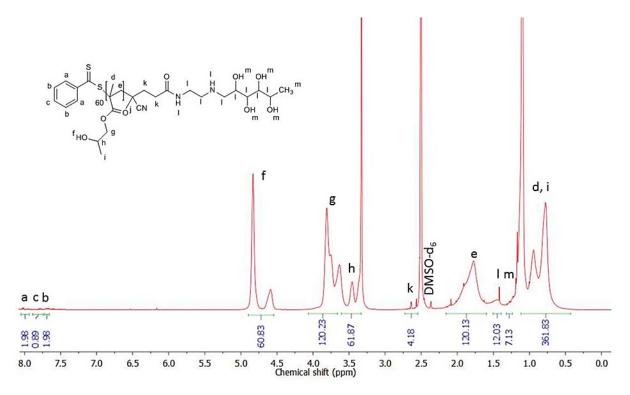


Figure S8. The 500 MHz ¹H-NMR spectrum of fucose-PHPMA₆₀ in DMSO-d₆ at 25 °C.

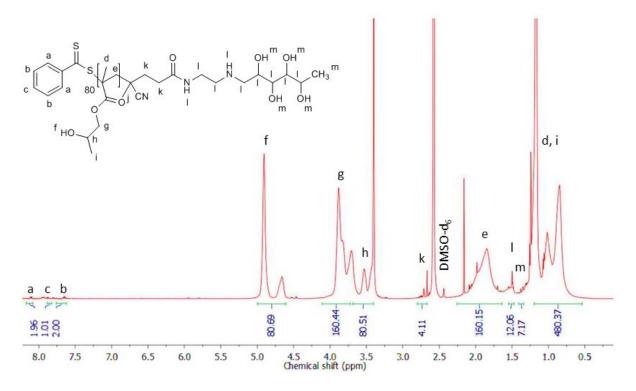


Figure S9. The 500 MHz ¹H-NMR spectrum of fucose-PHPMA₈₀ in DMSO-d₆ at 25 °C.

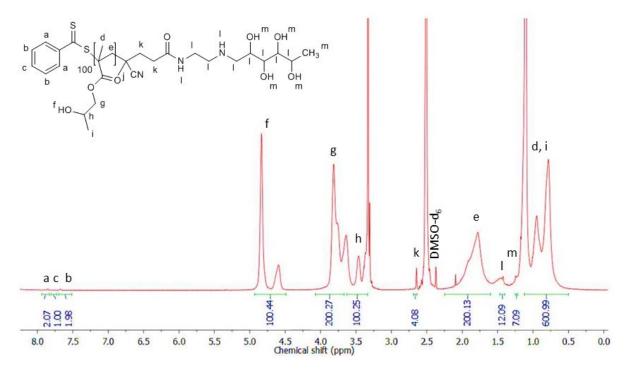


Figure S10. The 500 MHz ¹H-NMR spectrum of fucose-PHPMA₁₀₀ in DMSO-d₆ at 25 °C.

| Table S1. Advanced Polymer Chromatography | (APC) data for Fucose-PHPMA polymers. |
|---|---------------------------------------|
|---|---------------------------------------|

| Polymer | Theoretical M _n (g.mol ⁻¹) ^a | Average M _n (g.mol⁻¹) ^b | Dispersity |
|-----------------------------|--|---|------------|
| Fucose-PHPMA ₂₀ | 3348 | 3700 | 1.38 |
| Fucose-PHPMA ₄₀ | 6228 | 7600 | 1.48 |
| Fucose-PHPMA ₆₀ | 9108 | 11400 | 1.69 |
| Fucose-PHPMA ₈₀ | 11988 | 15500 | 1.55 |
| Fucose-PHPMA ₁₀₀ | 14868 | 16000 | 1.82 |

^a Theoretical molecular weight of synthesised polymers is calculated via the relative atomic mass of each element.

^b Average M_n was determined by advanced polymer chromatography – see *Materials and methods*.

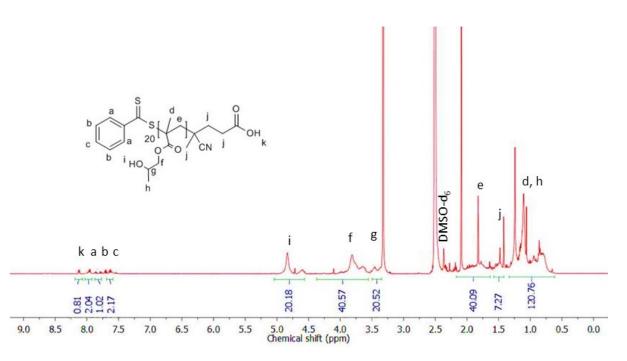


Figure S11. The 500 MHz 1 H-NMR spectrum of PHPMA₂₀ in DMSO-d₆ at 25 °C.

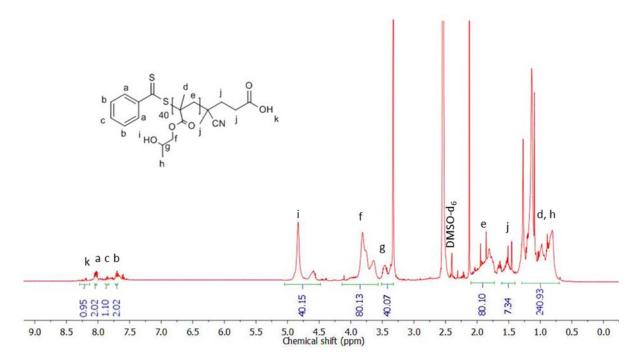


Figure S12. The 500 MHz ¹H-NMR spectrum of PHPMA₄₀ in DMSO-d₆ at 25 °C.

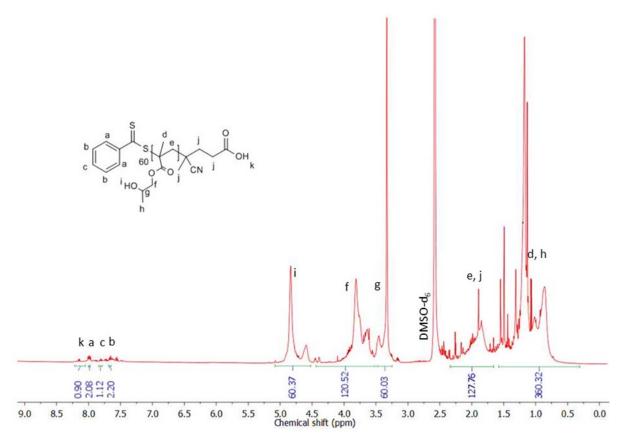


Figure S13. The 500 MHz 1 H-NMR spectrum of PHPMA₆₀ in DMSO-d₆ at 25 °C.

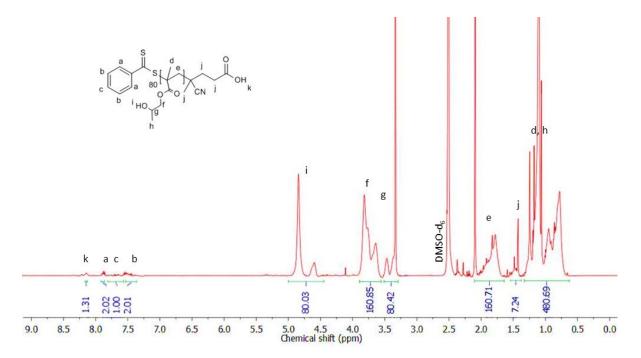


Figure S14. The 500 MHz 1 H-NMR spectrum of PHPMA₈₀ in DMSO-d₆ at 25 °C.

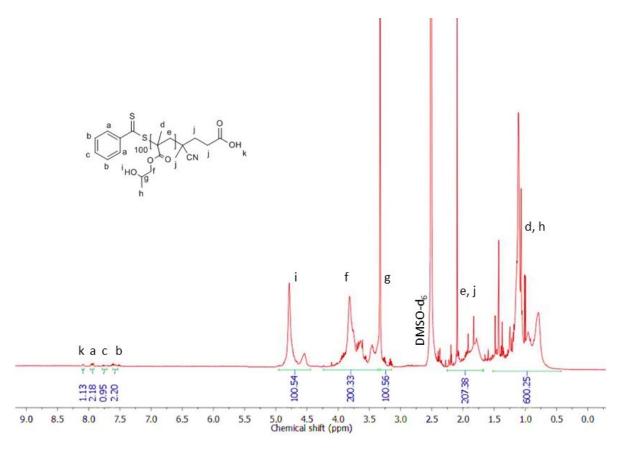


Figure S15. The 500 MHz ¹H-NMR spectrum of PHPMA₁₀₀ in DMSO-d₆ at 25 °C.

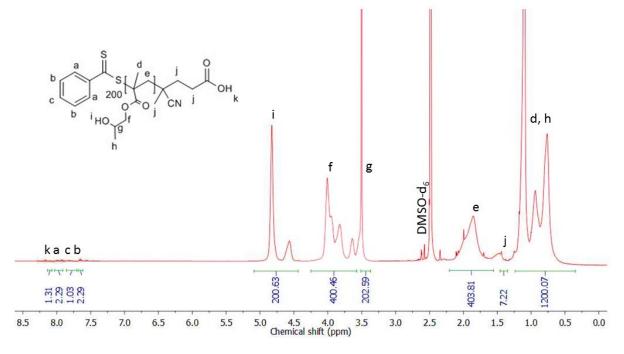


Figure S16. The 500 MHz ¹H-NMR of PHPMA₂₀₀ in DMSO-d₆ at 25 °C.

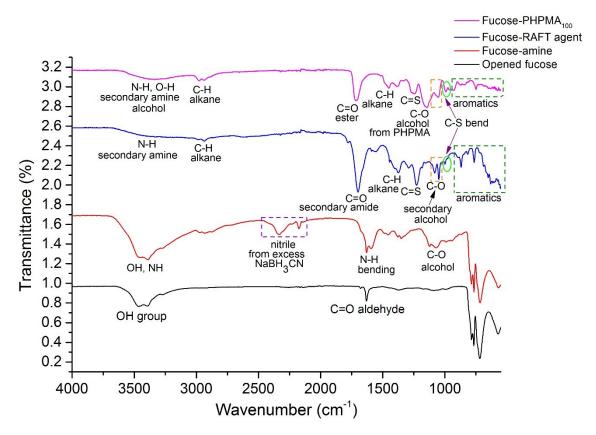


Figure S17. FT-IR spectra of opened fucose, fucose-amine, fucose-RAFT agent, and fucose-PHPMA₁₀₀.

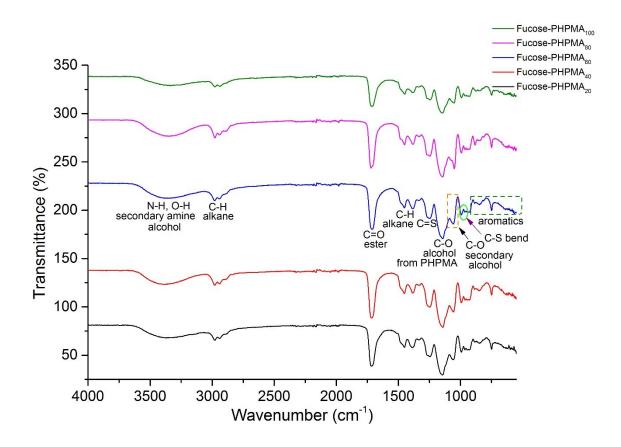


Figure S18. FT-IR spectra of fucose-PHPMA consisting of 20, 40, 60, 80 and 100 repeat units.

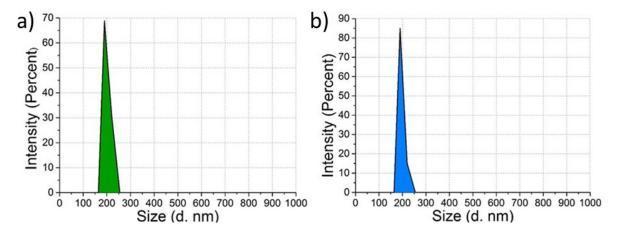
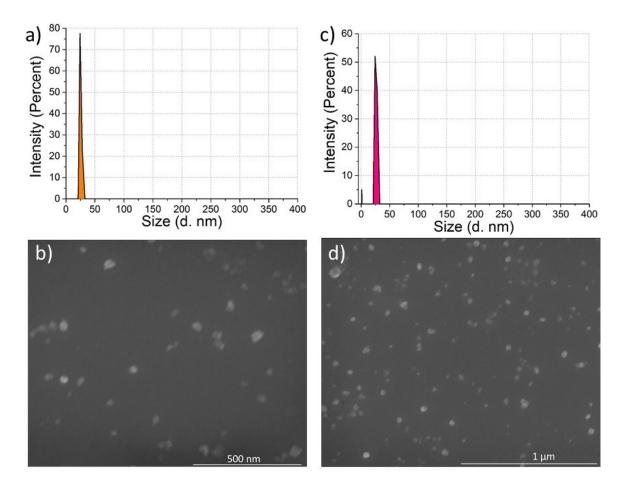
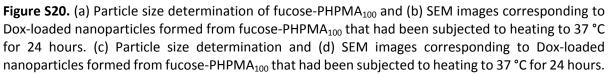


Figure S19. (a) Particle size determination of fucose-PHPMA₁₀₀ nanoparticles (a) and Dox-loaded fucose-PHPMA₁₀₀ nanoparticles (b).





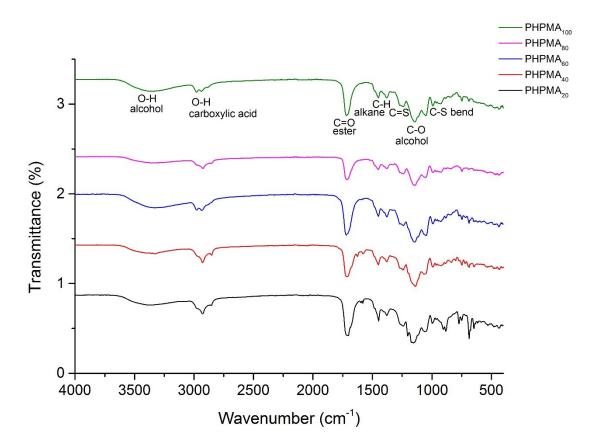


Figure S21. FT-IR spectra of PHPMA consisting of 20, 40, 60, 80 and 100 repeat units.

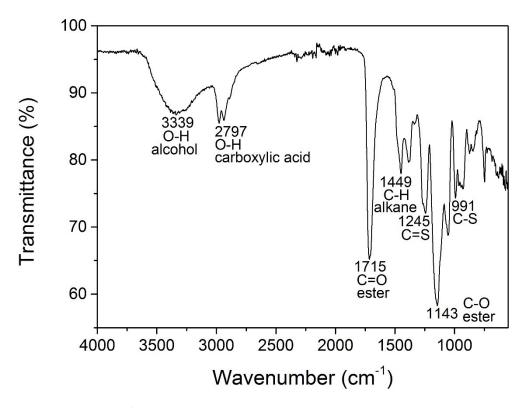


Figure S22. FTIR spectrum of PHPMA₂₀₀.

Nanoparticle creation for DLS analysis

2.0 mg of the synthesised polymer was dissolved in 1.0 mL of DMF and the solution added dropwise to 1.0 mL of deionised water whilst stirring. The solvent mixture of DMF and deionised water is in a 1:1 volume ratio.

Table S2. Nanoparticle size and PDI values for nanoparticles formed from $PHPMA_{20}$, $PHPMA_{40}$, $PHPMA_{60}$, $PHPMA_{80}$ and $PHPMA_{100}$ at room temperature.

| Polymer | Size (nm) | PDI |
|----------------------|--------------------|-------|
| PHPMA ₂₀ | 85 ±11 (51.2 %) | 0.683 |
| | 440.9 ±56 (48.8 %) | |
| PHPMA ₄₀ | 800.6 ±100 | 0.837 |
| PHPMA ₆₀ | 722 ±92 | 0.786 |
| PHPMA ₈₀ | 1992 ±265 | 0.644 |
| PHPMA ₁₀₀ | 488 ±284 | 0.653 |
| | | |

Table S3. Water and polymer content of each depot formed, as determined gravimetrically.

| Polymer | Percentage of polymer in | Percentage of water in Depot |
|--------------------------------------|--------------------------|------------------------------|
| | Depot | |
| PHPMA ₈₀ | 36.4 | 63.6 |
| PHPMA ₈₀ formed in water | 12.9 | 87.1 |
| PHPMA ₂₀₀ | 65.3 | 34.7 |
| PHPMA ₂₀₀ formed in water | 17.5 | 82.5 |

Dox loading of fucose-PHPMA₁₀₀ Nanoparticles

2.5 mg of Dox was dissolved in 20 μ L of trimethylamine and 3.0 mL of chloroform, and stirred for 4 hours in dark (bright red solution). 3.0 mg of fucose-PHPMA₁₀₀ was dissolved in 1.0 mL of DMF (colourless solution). The polymer solution was then added dropwise into 7.0 mL mL of PBS buffer (colourless solution). Dox solution was added dropwise into the polymer solution to yield a final volume of 8.0 mL (bright red solution).

The Dox concentration for each sample was

 $\frac{2.5 \ mg}{8.0 \ mL} \approx 0.3125 \ mg \ mL^{-1}$

After three days dialysis for each sample, the concentration for fucose-PHPMA₁₀₀ in PBS buffer solution was 0.2562 mg mL⁻¹, as determined by UV-vis spectroscopy.

Therefore, the percentage that was encapsulated by the polymer fucose-PHPMA₁₀₀ was,

At pH 7.4, $\frac{0.2562}{0.3125} \times 100\% \approx 82.0\%$

The mass of Dox in the above sample was 0.2562 mg mL⁻¹ \times 8.0 mL = 2.0496 mg

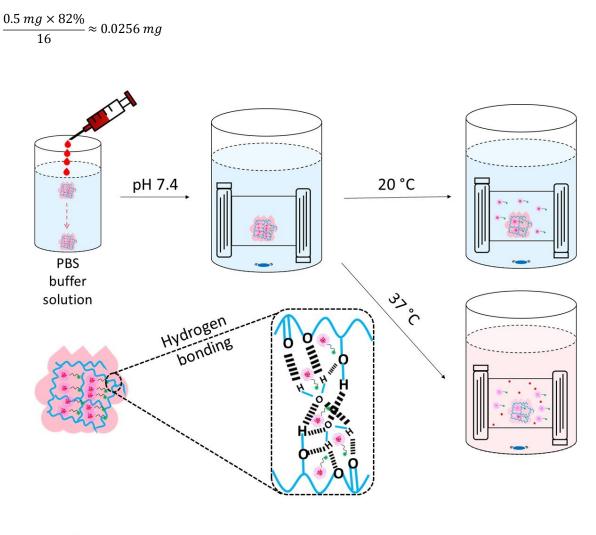
All the Dox release samples were prepared in PBS buffer.

The same Dox loading procedures were applied to fucose-PHPMA₂₀, fucose-PHPMA₄₀, fucose-PHPMA₆₀ and fucose-PHPMA₈₀ nanoparticles.

Loading Dox fucose-PHPMA₁₀₀ nanoparticles in PHPMA₂₀₀ Depot

0.5 mg freeze dried Dox encapsulated fucose-PHPMA₁₀₀ was dissolved in 0.8 mL of DMSO. 0.550 g of PHPMA₂₀₀ gel was dissolved in the DMSO solution. 0.05 mL of the red mixture was syringed and injected in 14.0 mL of PBS buffer solution, producing Dox fucose-PHPMA₁₀₀ in PHPMA₂₀₀ gel.

The mass of Dox for each sample was



PHPMA₂₀₀ and Dox loaded fucose-PHPMA₁₀₀ nanoparticles in DMSO

PHPMA₂₀₀ gel holds Dox loaded fucose-PHPMA₁₀₀ nanoparticles

Figure S23. Dox release from fucose-PHPMA₁₀₀ nanoparticles encapsulated in PHPMA₂₀₀ gel in PBS buffer solution at 20 °C and at 37 °C; inside structure of Dox-loaded fucose-PHPMA₁₀₀ nanoparticles encapsulated in PHPMA₂₀₀ gel in PBS buffer solution.

| Polymer | Theoretical M _n (g.mol ⁻¹) ^a | Average M _n (g.mol ⁻¹) ^b | Dispersity |
|----------------------|--|--|------------|
| PHPMA ₂₀ | 3159 | 3100 | 1.22 |
| PHPMA ₄₀ | 6039 | 6000 | 1.37 |
| PHPMA ₆₀ | 8919 | 8800 | 1.12 |
| PHPMA ₈₀ | 11799 | 11700 | 1.15 |
| PHPMA ₁₀₀ | 14679 | 14400 | 1.33 |
| PHPMA ₂₀₀ | 29079 | 28500 | 1.24 |
| | | | |

Table S4. Advanced Polymer Chromatography data revealing the average polymers molecular weight and dispersity values for PHPMA polymers used to create the depots.

^a Theoretical molecular weight of synthesised polymers is calculated via the relative atomic mass of each element.

^b Average M_n was determined by advanced polymer chromatography – see *Materials and methods*.

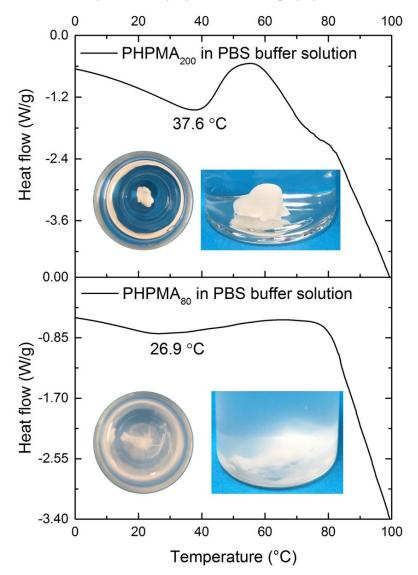


Figure S24. DSC thermograms of PHPMA₂₀₀ and PHPMA₈₀ depots formed in PBS buffer solution.

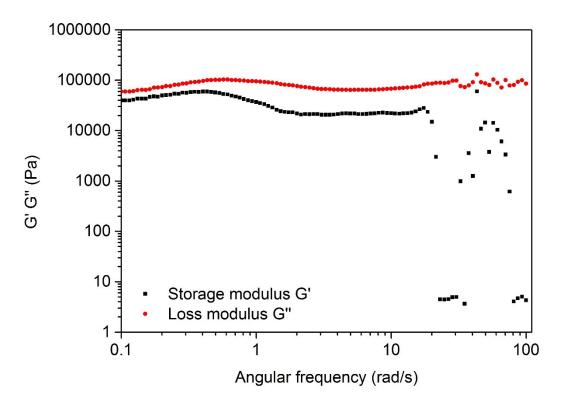


Figure S25. Rheogram of PHPMA₈₀ depot formed in PBS buffer solution.

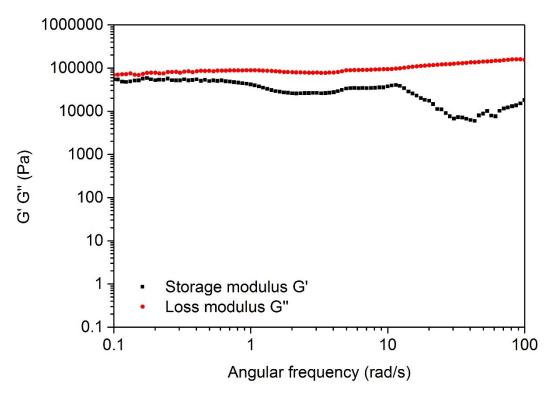


Figure S26. Rheogram of PHPMA₂₀₀ depot formed in PBS buffer solution.

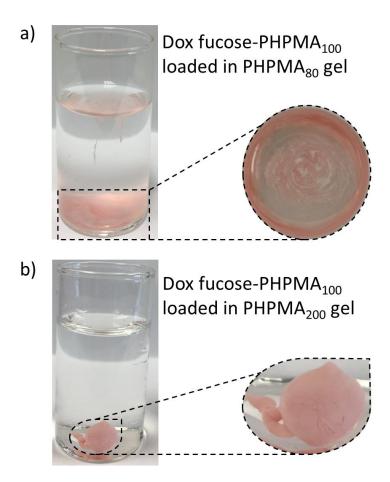


Figure S27. a) Dox fucose-PHPMA₁₀₀ nanoparticles loaded in PHPMA₈₀ gel dissolved in DMSO injected in PBS buffer solution; b) Dox fucose-PHPMA₁₀₀ nanoparticles loaded in PHPMA₂₀₀ gel dissolved in DMSO injected in PBS buffer solution.

Calibration curve for Dox free base release

0.003 g of doxorubicin hydrochloride was dissolved in 20 μ L of triethylamine and 3.0 mL of chloroform, and the solution mixed with PBS buffer (pH 7.4) to obtain the desired concentrations (Table S5). The absorbance values were measured using a Perkin Elmer Lambda 35 UV/VIS Spectrometer.

| Concentration of doxorubicin at pH 7.4 (mg mL ⁻¹) | Absorbance at 498 nm |
|---|----------------------|
| 0.01 | 0.07282 |
| 0.005 | 0.03569 |
| 0.003 | 0.02024 |
| 0.001 | 0.00388 |

Table S5. Absorbance of doxorubicin with different concentrations

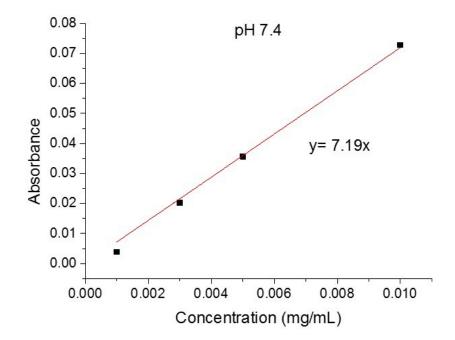


Figure S28. Calibration with line of best fit for doxorubicin free base.