

Electronic Supplementary Material (ESI)

**Identification of P218 as a potent inhibitor of *Mycobacterium ulcerans* DHFR**

Gustavo Pelicioli Riboldi,<sup>ab</sup> Rachael Zigweid,<sup>c</sup> Peter J. Myler,<sup>cd</sup> Stephen J. Mayclin,<sup>cd</sup> Rafael Miguez Couñago,<sup>ab\*</sup> Bart L. Staker<sup>\*c</sup>

a. Centro de Química Medicinal (CQMED), Centro de Biologia Molecular e Engenharia Genética (CBMEG), Universidade Estadual de Campinas (UNICAMP), Campinas, SP, 13083-875, Brazil.

b. Structural Genomics Consortium, Departamento de Genética e Evolução, Instituto de Biologia, UNICAMP, Campinas, SP, 13083-886, Brazil.

c. Center for Infectious Disease Research, Seattle Children's Research Institute, Seattle, Washington, 98109, USA.

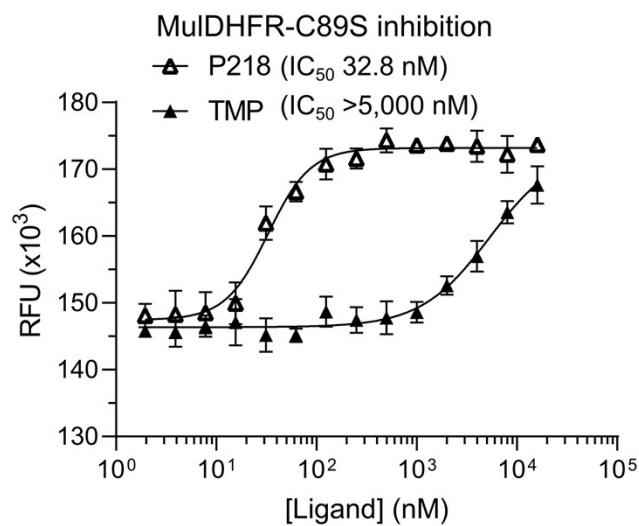
d. Department of Pediatrics, University of Washington, Seattle, Washington, 98195, USA.

\* rafael.counago@unicamp.br; bart.staker@seattlechildrens.org

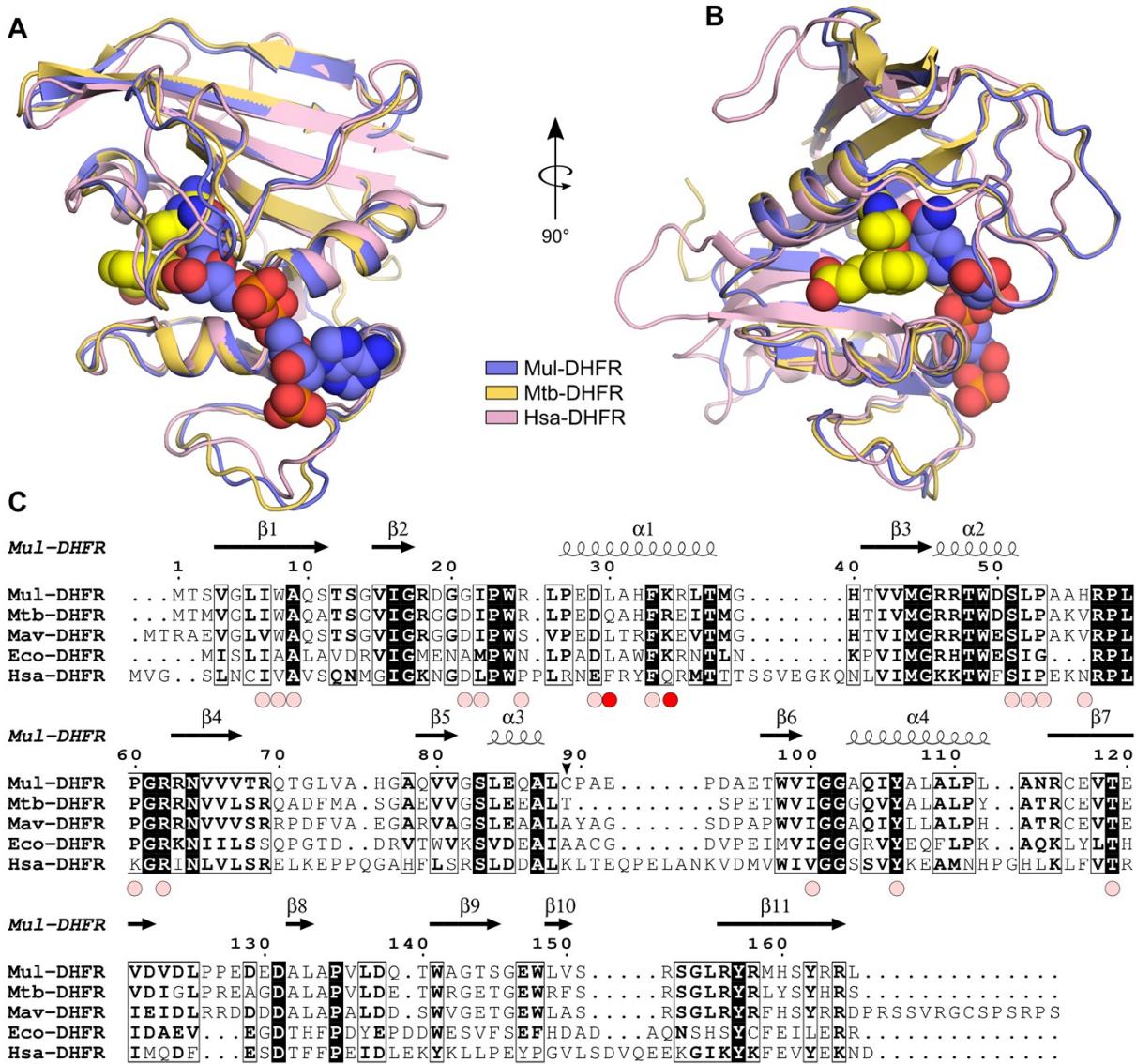
**Supplementary Table S1. Crystallographic data and refinement statistics for MuLDHFR-C89S crystals.**

Data collection	
Crystal	Native
PDB ID	6UWW
X-ray source	APS LS-CAT 21-ID-D
Wavelength (Å)	0.8666
Space group	P 1 2 <sub>1</sub> 1
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	28.73, 66.20, 44.52
$\alpha$ , $\beta$ , $\gamma$ (°)	90.000, 91.614, 90.000
Resolution (Å)	50-0.92 (0.94-0.92)
No. of unique reflections*	112,854 (8,042)
R <sub>merge</sub> (%)	3.6 (49.1)
Mean I/σI	14.66 (2.01)
Mean CC <sub>1/2</sub>	99.9 (74.8)
Completeness (%)	98.1 (94.7)
Redundancy	3.5 (2.9)
Refinement Statistics	
Resolution (Å)	50-0.92 (0.94-0.92)
R <sub>work</sub> / R <sub>free</sub> (%)	13.15 (21.59) / 14.54 (22.67)
No. of atoms / Mean B-factor (Å)	
Protein atoms	1,331 / 12.2
Solvent atoms	271 / 29.8
NADPH/P218 atoms	84 / 13.9
RMSD bond lengths	0.009 Å
RMSD bong angles	1.29°
Ramachandran plot (%)	
Favored	98.8
Allowed	1.2
Outliers	0

Data for the outmost shell are given in parentheses.



**Supplementary Figure S1** - Enzyme inhibition of mutant MulDHFR-C89S by TMP (filled symbols) and P218 (empty symbols). The half-maximal inhibitory concentration ( $IC_{50}$ ) for each compound is shown in parenthesis. Data shown are mean  $\pm$  SD of triplicates.



**Supplementary Fig. S2 - The structure of MulDHFR-C89S bound to P218 and NADPH reveals a conserved DHFR architecture.** (A-B) Superposition of MulDHFR-C89S (blue cartoon) onto the P218-NADPH-bound structures of *M. tuberculosis* (yellow cartoon, PDB ID 5U26) and human (pink cartoon, PDB ID 4DDR)<sup>1</sup> enzymes. P218 and NADPH, as seen in the MULDHFR-C89S co-structure, are shown as van der Waal spheres. (C) Structure-based sequence alignment of *M. ulcerans* (Mul-), *M. tuberculosis* (Mtb-), *M. avium* (Mav-), *E. coli* (Eco-), and human (Hsa-) DHFR. Pink circles indicate structurally-equivalent residues within a 4 Å radius of ligand P218, as seen in our MulDHFR-C89S structure. Red circles indicate structurally-equivalent residues thought responsible for sterically preventing P218 binding to the human enzyme. The black arrowhead indicates the position of Cys89 in MulDHFR mutated to a serine to improve protein

crystallization. Absolutely conserved residues are indicated by a black background. Similar residues are shown in bold and framed in a box. The secondary structure ( $\alpha$ -helices, shown as coils; and  $\beta$ -sheets, shown as arrows), and the numbering shown in the top line are for MulDHFR. Protein sequence / structures used in were: Mul-DHFR (UniProt ID A0PQG8, PDB ID 6UWW) (this work), Mtb-DHFR (UniProt ID P9WNX1, PDB ID 5U26), Mav-DHFR (UniProt ID O30463, PDB ID 2W3W), Eco-DHFR (UniProt ID P0ABQ4, PDB ID 1RF7)<sup>2</sup>, and Hsa-DHFR (UniProt ID P00374, PDB ID 4DDR)<sup>1</sup>. Structural alignment by PROMALS3D <sup>3</sup>.

**Supplementary References:**

- 1 Y. Yuthavong, B. Tarnchompoon, T. Vilaivan, P. Chitnumsub, S. Kamchonwongpaisan, S. A. Charman, D. N. McLennan, K. L. White, L. Vivas, E. Bongard, C. Thongphanchang, S. Taweechai, J. Vanichtanankul, R. Rattanajak, U. Arwon, P. Fantauzzi, J. Yuvaniyama, W. N. Charman and D. Matthews, *Proc. Natl. Acad. Sci.*, 2012, **109**, 16823–16828.
- 2 M. R. Sawaya and J. Kraut, *Biochemistry*, 1997, **36**, 586–603.
- 3 J. Pei and N. V. Grishin, *Bioinformatics*, 2007, **23**, 802–808.