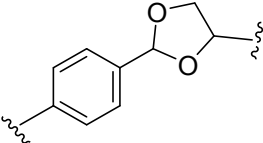
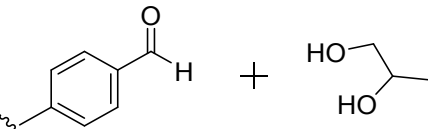
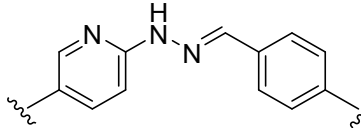
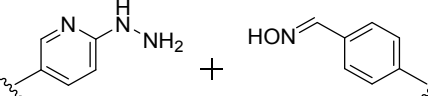
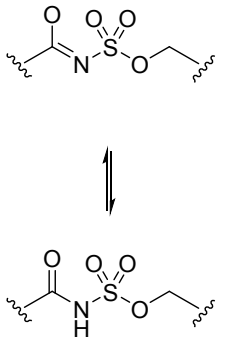
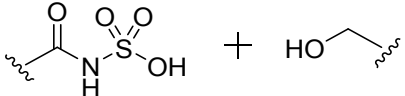
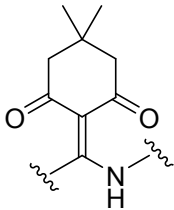
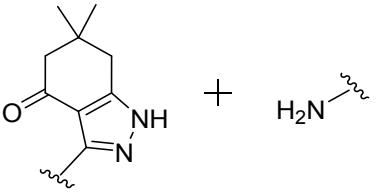
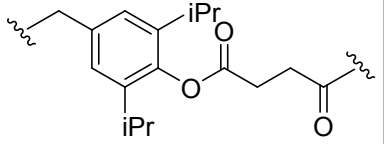
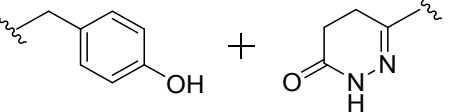
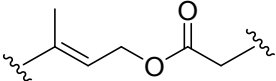
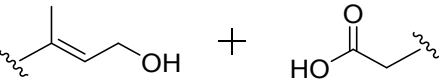
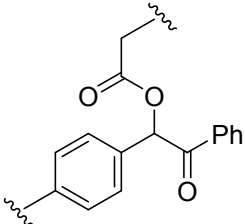
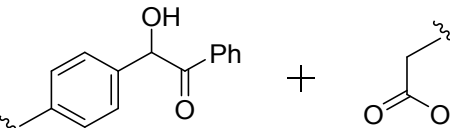
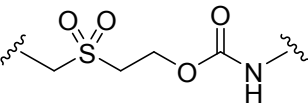
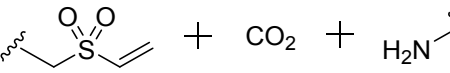


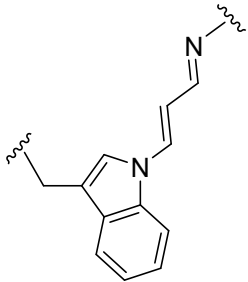
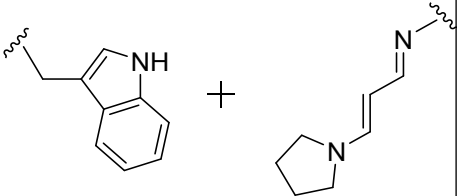
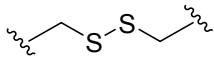
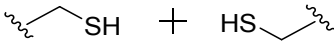
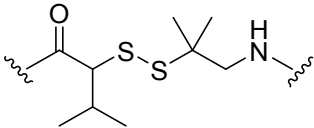
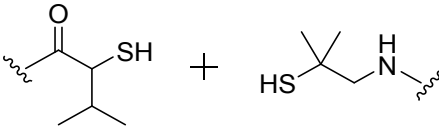
TABLE S1: Chemically cleavable linkers

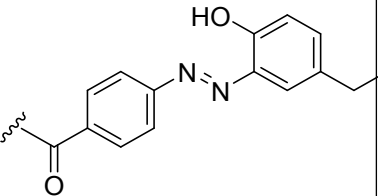
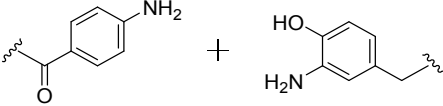
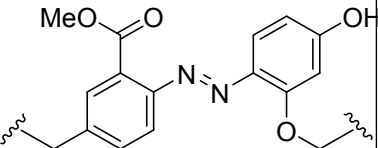
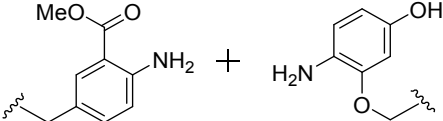
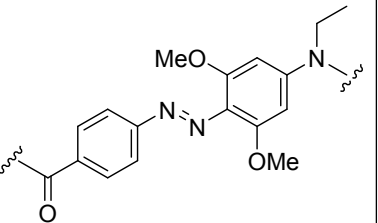
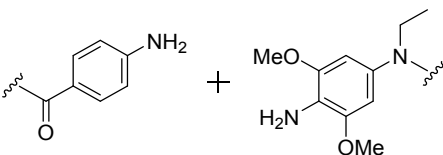
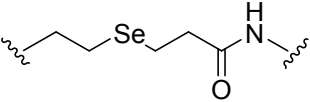
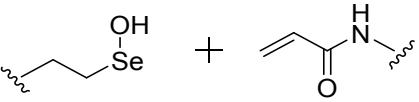
	Cleavable linker	Structure	Cleavage products	Cleavage conditions	Advantages	Disadvantages
Acid-labile	Dialkoxydiphenylsilane (DADPS) ¹			10% HCOOH	mild conditions Stable in aqueous KF solutions of concentrations up to 5 M	
	4-hydroxybenzyl-alcohol ²			TFA	Based on well established solid phase chemistry	Strong acid may lead to release of non-specifically bound proteins
	Boc (t-butyl-oxy-carbonyl) group ^{3,4}			TFA	Based on well established protecting group	Strong acid may lead to release of non-specifically bound proteins
	Acyl hydrazones ₅			Acyl hydrazine pH 4–5	Stable at basic pH (pH 8–10).	Tends to prematurely hydrolyze at neutral pH

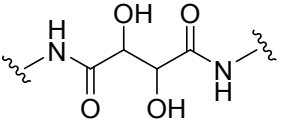
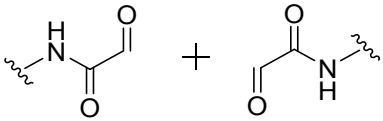
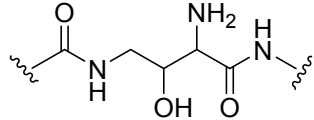
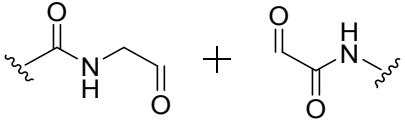
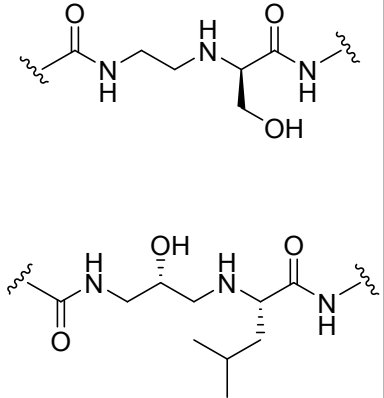
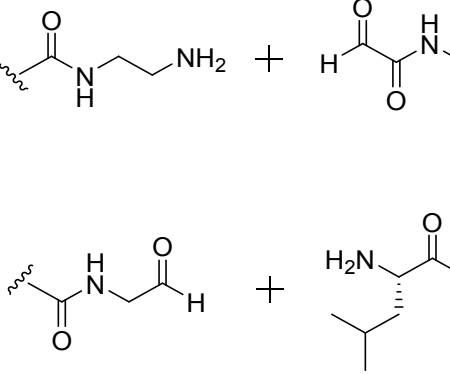
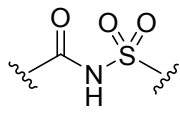
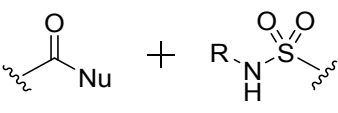
Cyclic acetals ⁶			1M guanidine in 1% TFA at 37 °C for 1 hour	Possibility to reintroduce a label to the released biomolecule	Only tested on lysates spiked with model proteins
Bisaryl hydrazones ⁷			NH ₂ OH, aniline pH 4–5 Also by ETD (see Table S4)	stable at neutral pH possibility to reintroduce a label to the released biomolecule	high concentrations of aniline (100 mM) are required and full cleavage is not always achieved
Acyl-sulfamate ⁸	 pH 7.6 Stable pH 2.0 Less stable		pH 2, 5h @ 37°C	Stable even at 95 °C with 10 mM hydrazine (5 min)	

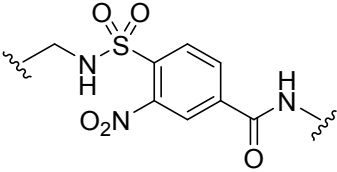
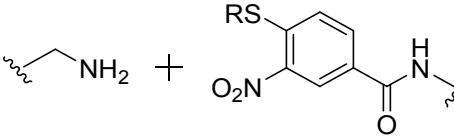
Base-labile	Dde ⁹			2% hydrazine	Based on well established protective group chemistry	(DDE) linker has been demonstrated to be unstable as no modified peptides could be detected upon application of a cleavage reagent, due to instability of the linker during on-bead trypsin digestion ¹⁰ The conventional way of deprotecting the Dde group in peptide synthesis consists of using 2% hydrazine. This turned out to be not compatible with fluorescent scanning of SDS-PAGE ⁹
	Levulinoyl ester ¹¹			100 mM hydrazine	acid stable and can be removed selectively with respect to other esters	

	Allylic ester ¹²			pH 11		
	Benzoin ester ¹³			pH 8	Also usable as a photocleavable linker	
	Sulfonyl-based reversible amine ¹⁴			100 mM ammonium bicarbonate, pH 9, overnight at 4 °C	Traceless linker when reacted with amines of proteins.	Long cleavage time

	Indole-based linker ¹⁵			50mM or 100mM pyrrolidine@4°C		
Reductive cleavage	Disulfide ^{1,9}			DTT TCEP or BME	Commercial building block available	Application in click chemistry is limited (due to reducing conditions during CuAAC); disulfide exchange may occur. Incompatible with reduction/alkylation workflow of MS sample preparation
	Hindered disulfide ¹⁶ 17			10mM TCEP		

	Diazobenzene ¹⁸			25mM Na ₂ S ₂ O ₄ (several elutions)	Short synthesis of building block; linker changes color upon cleavage	Difficult purification + multiple elution steps necessary
	HAZA ¹⁹			1mM Na ₂ S ₂ O ₄ (< 10 s)	Fast cleavage; linker changes color upon cleavage	Multistep synthesis of linker
	Highly labile diazobenzene ²⁰			0.5mM Na ₂ S ₂ O ₄ (< 5 s)	Very fast cleavage; linker changes color upon cleavage	
Oxidative cleavage	Selenium ether ²¹⁻²⁴			8 mM H ₂ O ₂ for 1 h		

	Vicinal diol ²⁵			10 mM periodate Neutral pH		
	1,2-Amino alcohols ²⁶			10 µM periodate		
	Seramox / iso-seramox ²⁷			1 mM periodate Within 5mins		
Others	Acyl-sulfonamide ²⁸⁻³⁰			ICH2CN pH 9, NH4OH or pH 5 ICH2COO ⁻ or 200mM iodoacetimide pH 8		

	Nitro-sulfonamide ³¹			RSH pH 9.2		
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