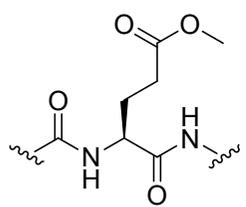
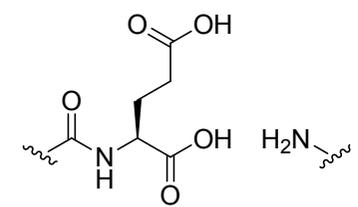


**TABLE S3: Enzymatically cleavable linkers**

Cleavable linker	Structure	Cleavage products	Cleavage conditions	Advantages	Disadvantages
TEV-cleavable linker <sup>1,2</sup>	$\text{---ENLYFQG---}$	$\text{---ENLYFQ-COOH}$ $\text{H}_2\text{N-G---}$	TEV protease	High specificity (only cleaves between Gln-Gly of sequence on LHS). => low background	Can get varying cleavage efficiencies, dependent on activity and stability of the protease. <sup>a</sup>
Trypsin-cleavable linker <sup>3-5</sup>	$\text{---XXXK/RXXX---}$ X = any amino acid other than K or R	$\text{---XXXK/R-COOH}$ $\text{H}_2\text{N---}$ X = any amino acid other than K or R	Trypsin	Cleave linker and obtain tryptic peptides in a single step.	Protein or peptide cannot be specifically eluted
V8-cleavable linker <sup>6</sup>			NaOH then V8 protease (aka <b>endoproteinase Glu-C</b> )	Will only cleave after alkaline ester hydrolysis, and therefore orthogonal to protease digestion of target proteins	Not yet been applied to whole proteome analysis

<sup>a</sup> activity and stability of the involved protease is applicable to all protease cleavable linkers. Overall, this should not represent a bottleneck.

## References

- 1 A. E. Speers and B. F. Cravatt, *J. Am. Chem. Soc.*, 2005, **127**, 10018–10019.
- 2 E. Weerapana, A. E. Speers and B. F. Cravatt, *Nat. Protoc.*, 2007, **2**, 1414–1425.
- 3 E. E. Carlson and B. F. Cravatt, *Nat. Methods*, 2007, **4**, 429–435.
- 4 M. Broncel, R. A. Serwa, P. Ciepla, E. Krause, M. J. Dallman, A. I. Magee and E. W. Tate, *Angew. Chemie - Int. Ed.*, 2015, **54**, 5948–5951.
- 5 M. Broncel, R. A. Serwa, T. D. Bunney, M. Katan and E. W. Tate, *Mol. Cell. Proteomics*, 2016, **15**, 715–725.
- 6 M. Hashimoto, S. Okamoto, K. Nabeta and Y. Hatanaka, *Bioorg Med Chem Lett*, 2004, **14**, 2447–2450.