Supplementary Material

Table S1. Data collection and refinement statistics

Data Collection ^a	FeCu-P19	FeCu-P19	Cu-P19 E44Q	Cu-P19 M88I	Cu-P19 D92H	Cu-FetP E46Q	Cu-FetP M90I
	(red)	(ox)					
Resolution Range (Å)	50.00-1.55	33.07-1.65	50.00-1.90	50.00-2.30	50.00-2.50	33.65-1.40	38.34-1.53
	(1.61-1.55)	(1.74-1.65)	(1.93-1.90)	(2.34-2.30)	(2.54-2.50)	(1.47-1.40)	(1.61-1.53)
Space group	$P2_{1}2_{1}2$	$P2_{1}2_{1}2$	$P2_{1}2_{1}2$	$P6_{2}$	P32	C2	$P2_{1}$
Cell dimensions (Å)	<i>a</i> = 53.91	<i>a</i> = 54.22	<i>a</i> = 55.35	a = b =	a = b =	<i>a</i> = 83.16	<i>a</i> = 49.16
	<i>b</i> = 73.23	<i>b</i> = 73.72	<i>b</i> = 73.34	132.97	132.46	<i>b</i> = 36.54	<i>b</i> = 51.61
	<i>c</i> = 74.93	<i>c</i> = 74.85	<i>c</i> = 78.96	<i>c</i> = 104.88	c = 104.59	c = 101.00	<i>c</i> = 66.14
						$\beta = 106.5$	$\beta = 93.3$
Wavelength (Å)	1.000	0.979	1.542	1.542	1.542	1.000	1.000
Unique Reflections	43367	36846	25709	45287	69955	50962	52557
Completeness (%)	98.9 (97.1)	100.0 (99.4)	99.2 (88.8)	97.5 (77.8)	99.0 (97.3)	87.4 (84.8)	99.4 (96.8)
Average I/oI	29.7 (4.1)	15.0 (3.1)	24.2 (1.7)	21.6 (1.4)	12.6 (1.63)	14.9 (5.0)	19.0 (2.6)
Redundancy	6.0	8.3	6.7	10.7	4.9	3.9	3.6
R _{merge}	0.050 (0.417)	0.082 (0.639)	0.086 (0.515)	0.104 (0.833)	0.109 (0.624)	0.053 (0.235)	0.058 (0.362)
Refinement							
R _{work}	0.175	0.178	0.174	0.235	0.270	0.186	0.176
R _{free}	0.207	0.211	0.222	0.277	0.299	0.212	0.208
No. of waters	286	260	256	385	541	274	348
Average <i>B</i> -value (Å ²)	19.9	21.9	34.0	47.0	51.0	15.4	17.1
r.m.s.d. bond lengths (Å)	0.011	0.010	0.013	0.003	0.003	0.010	0.011
Ramachandran plot							
Most-favorable (%)	91.7	92.1	97.7	95.8	96.9	93.1	91.9
Allowed (%)	8.3	7.5	2.3	4.0	2.9	6.5	8.1
Outliers (%)	0	0.4	0	0.2	0.2	0.4	0

^a Values for the highest resolution shell are shown in parenthesis

Strains and plasmids	Relevant genotype	Reference
<u>E. coli strains</u>		
XL1-Blue	<i>recA</i> 1, <i>endA</i> 1, <i>gyrA</i> 96, <i>thi</i> -1,	Stratagene
	hsdR17, relA1, supE44, lac (F¢, proAB,	
	<i>lacI</i> qZ_M15, Tn <i>10</i>)	
BL21	F - dcm , omp , $hsdS$, $(r_B m_B)$, lon , $gal\lambda$,	Novagen
	DE3, pLysS	
BL21-pLysS	F - dcm , omp , $hsdS$, $(r_B m_B)$, lon , $gal\lambda$,	1
	DE3	
	Genomic DNA of <i>E. coli</i> strain F11	Robert Koch-Institute, S2,
		Wernigerode
<u>C. jejuni strains</u>		
81-176 wild-type		2
reference strain		
81-176∆ <i>p19</i>		3
81-176Δ <i>p19</i>		3
complemented		
Vectors & plasmids		
pECD1098	pASK-IBA3:: <i>fetM</i>	1
	pASK-IBA6	IBA GmbH
pECD1138	pASK-IBA6:: <i>fetM</i> _PL	This study
pECD1101	pET22b(+):: <i>fetP</i>	1
pECD1128	pET22b(+):: <i>fetP</i> E46Q	This study
pECD1129	pET22b(+):: <i>fetP</i> M90I	This study

Table S2. Bacterial strains and plasmids used in this study.

- 1 Koch, D. *et al.* Characterization of a dipartite iron-uptake system from uropathogenic *Escherichia coli* strain F11. *J Biol Chem* **286**, 25317-25330, doi:10.1074/jbc.M111.222745 (2011).
- 2 Korlath, J. A. *et al.* A point-source outbreak of campylobacteriosis associated with consumption of raw milk. *J Infect Dis* **152**, 592-596 (1985).
- 3 Chan, A. C. K. *et al.* Structure and function of P19, a high-affinity iron transporter of the human pathogen *Campylobacter jejuni*, *J Mol Biol* **401**, 590-604 (2010).

Table S3. Primers used in this study.

Primer	Sequence
fetM_PD BsaI d	ATGGTA <u>GGTCTC</u> AGCGCACCTCGACGAATTATGCGCCTTT
fetM_PD BsaI u	ATGGTA <u>GGTCTC</u> ATATCAACTATAGAGCAGCAGACTCCACT
<i>fetP</i> E46Q d	AAAGCCGATGTTCACCTTCAGGCGGATATCCACGCTGTA
<i>fetP</i> E46Q u	TACAGCGTGGATATCCGCCTGAAGGTGAACATCGGCTTT
fetP M90I d	GGCACCTTCATGCCGATCGTTGCCAGCGATGGC
fetP M90I u	GCCATCGCTGGCAACGATCGGCATGAAGGTGCC
p19 E44Q phos Cj	GCAGATATTCATCTACAAGCTGACATTCACGC
p19 E44Q phos Rec	CTTTAGCAGATATTCACCTACAAGCTGACATTCACGCAC
<i>p19</i> M88I phos Cj and Rec	GGAACTTTGATGCCTATCGTGGCTGATGATGG
p19 D92H phos Cj and Rec	CCTATGGTGGCTGATCATGGTCCTCACTATGGTG

Underlined: restriction endonuclease recognition site

Bold letters: altered bases leading to amino acid exchange

Table S4. Taxonomic assignment of FetP-like sequences. The number (N) of predicted protein sequences in each cluster identified from a BLAST search is shown with the taxonomic assignment. The ε value compared to the FetP sequence of least related sequence of the cluster is indicated. Changes are truncations, non-conserved amino acid residues in metal binding site (FetP numbering) and a conserved cysteine residue in the leader of the sequences from Gram-positive bacteria, which might allow anchoring of the protein in the cytoplasmic membrane. The ratio of the sequences with these changes is also indicated. The clusters are shown in Supplementary Figure S6.

Cluster	N N	/in ε value	Changes	Taxon	Genera
ΙA	2	2.00E-70	none	Spirochaeta	Spirochaeta
I B1	1	6.00E-72	none	δ -Proteobacteria	Geobacter
I B2	1	1.00E-69	none	α -Proteobacteria	Rhodospirillum
I B3	17	6.00E-58	none	e-Proteobacteria	Campylobacter, Wollinella
I C1	1	6.00E-83	none	g-Proteobacteria	Vibrio
I C2	1	1.00E-88	E5, truncated	g-Proteobacteria	Psychromonas
I C3	2	2.00E-87	$1/2 = \Delta LEADIH$	Chlamydia	Opitutaceae
I C4	18	1.00E-88	none	g-Proteobacteria	Aggregatibacter, Haemophilus, Pasteurella, Actinobacillus, Cardiobacterium
I C5	22	4.00E-91	none	Enterobacteriaceae	Escherichia, Citrobacter, Shigella, Klebsiella, Yersinia, Brenneria, Pectobacterium (=Seratia)
II A1	9	2.00E-49	none	Enterobacteriaceae	Providencia, Proteus, Morganella
II A2	1	2.00E-50	none	α -Proteobacteria	Candidatus Liberibacter solanacearum
II A3	2	3.00E-53	none	γ -Proteobacteria	Vibrio, Photobacterium
II B1	16	3.00E-42	none	γ -Proteobacteria	Pseudomonas
II B2	1	3.00E-52	none	α -Proteobacteria	Rhodopseudomonas
II B3	48	3.00E-43	1/48 E5/truncated	β -Proteobacteria	Burkholderia, Herbaspirillum
II B4	3	2.00E-46	none	α -Proteobacteria	Rhodopseudomonas. Rhodovulum
II B5	17	2.00E-27	none	α -Proteobacteria	Beijerinckia, Brucella, Rhodopseudomonas, Rhodomicrobium, Methylobacterium
II B6	7	1.00E-44	1/7 E5/truncated	α -Proteobacteria	Hyphomicrobium, Mesorhizobium, Sinorhizobium, Rhizobium
II C1,	11	6.00E-47	none	β -Proteobacteria	Achromobacter, Bordetella
II C2	4	2.00E-45	1/4 E5D	β -Proteobacteria	Pseudogulbenkiamia, Chromobacterium, Verminephrobacter
II C3	1	4.00E-50	none	β -Proteobacteria	Dechloromonas
II C4	2	2.00E-47	none	β -Proteobacteria	Aromatoleum, Azoarcus
II C5	1	3.00E-47	none	γ -Proteobacteria	gamma proteobacterium HdN1
II C6	1	5.00E-46	none	β -Proteobacteria	Dechlorosoma
II C7	2	4.00E-37	none	α -Proteobacteria	Magnetospirillum
II C8	2	2.00E-44	none	β -Proteobacteria	Rubrivivax
III A	1	5.00E-51	none	γ -Proteobacteria	gamma proteobacterium IMCC2047
III B	1	2.00E-54	none	Deferribacteres	Flexistipes
III C	16	2.00E-13	2/16: H44, E5, E46, truncated	Firmicutes	Centipeda, Selenomonas, Dialister, Pelosinus, Megasphaera,
III D1	3	1.00E-17	1/3: H44, E5, E46; truncated	Actinobacteria	Mobiluncus
III D2	3	1.00E-27	none, Leader-Cys?	Firmicutes	Parvimonas

III D3	18	1.00E-06	Cys-Leader, 2/18 M90; 1/18 M90, H97, H127, D94	Fusobacteria	Fusobacterium
III D4	1	1.00E-21	none, leader-Cys?	Firmicutes	Clostridiales genomosp. BVAB3 str. UPII9-5
III D5	3	4.00E-25	none, Cys-Leader	Synergistetes	Synergistetes, Jonquetella, Pyramidobacter
III E1	10	6.00E-28	Cys-Leader, 1/10 H127, 1/10 M90,H97	Actinobacteria	Actinomyces
III E2,	34	3.00E-34	1/34 M90, H127, E5, D94	Actinobacteria	Actinomaces, Parascardovia, Scardovia, Bifidobacterium, Gardnerella
III E3	2	1.00E-26	none	Firmicutes	Abiotrophia, Ruminococcus
III F1	1	6.00E-44	none, leader-Cys	Firmicutes	Clostridium
III F2	3	9.00E-40	none	Spirochaeta	Treponema
III F3	7	7.00E-26	1/7: M90, H97	Spirochaeta	Treponema
III F4	11	2.00E-26	none, Leacer-Cys?	Firmicutes	Filifactor, Johnsonella, Peptinophilus, Finegoldia, Anaerococcus, Stomatobaculum
IV A	1	2.00E-46	none	Planctomyces	Singulisphaera
IV B	1	7.00E-46	none	α -Proteobacteria	Phaeospirillum
IV C	1	2.00E-55	none, TwinR-leader?	lpha -Proteobacteria	magnetite-containing magnetic vibrio
V A	2	8.00E-35	1/2 E5/truncated	α -Proteobacteria	Magnetospirillum
V B	1	5.00E-24	none	BACTERIA	uncultured bacterium Bio2
VI A	2	5.00E-42	none	γ-Proteobacteria	Pseudomonas
VI B	11	4.00E-29	none, Leacer-Cys?	lpha -Proteobacteria	Gluconobacter, Acetobacter
VII A	1	3.00E-11	H44, E5, leader too long	Bacteroides/Chlorobi	Salinibacter
VII B	1	5.00E-05	H44, E5, E46, truncated	BACTERIA	uncultured marine bacterium EB0_39H12

Supplementary Figures



Figure S1. P19 metal site variants are unable to fully restore growth under iron limitation. *C. jejuni* wild-type 81-176, 81-176 $\Delta p19$, and trans-complemented strains grown under 0-12 μ M DFO in a stationary 96-well plate incubated in a tri-gas incubator at 37 °C. After 24 h of growth, the plates were resuspended and OD₆₀₀ readings were taken. The two plots here and the one in Fig. 4 represent experiments performed on three separate occasions. Plot points represent the amount of bacterial growth averaged from three separate wells with standard deviation error bars drawn.



Figure S2. Cu-bound *C. jejuni* P19 (a) E44Q and (b) M88I variants do not exhibit positional plasticity. The copper cation is colored in brown. A 2Fo-Fc map contoured at 1 σ (blue) is overlaid the active site. Sulfur, oxygen and nitrogen atoms are colored yellow, red and dark blue, respectively. One protomer of the FetP dimer is shown in purple and the other in pink.



Figure S3. In source decay (ISD) MALDI-MS spectrum of ecFetM. ecFetM carrying a C-terminal streptag was solubilized from the membrane fraction after expression in *E. coli* and purified. In source decay led to a series of peptides and the molecular masses of these peptides (peaks) were determined with the matrixassisted laser desorption mass spectrometer (MALDI-MS). The mass difference between the peptides identified the respective amino acid just removed from the protein with the sequence indicated above. This sequence, LDKTAELYQQQHADE, followed that of the amino acids within the smallest peptide identified with a mass of 1648.102 (left-most peak) which corresponded to the peptide STNYAPLIEDIEQR. This sequence was therefore identified as the amino-terminal sequence of the mature ecFetM-strep-tag protein as isolated from the membrane.



Figure S4. CD spectrum of ecFetM-PD. A circular dichroism spectrum (θ_{MRW}) of the purified periplasmic region of ecFetM, ecFetM-PD, is shown. Protein concentration was 15 μ M in 25 mM Bis-Tris/HCl, pH 7.2, 25°C. Shown is a difference spectrum plus vs. minus protein.



Figure S5. Original Far western blots of ecFetM-PD-FetP and cjFetM-PD-P19 interactions. Control blots (e.g. –P19) were prepared alongside non-controls (e.g. +P19) and were imaged together. The P19 and FetP experiments were performed separately.



Figure S6. Clusters of FetP-like sequences. The data resulting from a BLAST search (1) was further analyzed by COBALT (2) and Geneious (www.geneious.com) to sort the sequences into clusters (I to VII), sub-clusters (A, B, C,...) and sub-sub-clusters (1, 2, 3, ...). The predicted FetP-like proteins within each sub-sub-cluster are listed in Table S4. Filled circles and squares are single sequences, arrowheads are two or more sequences with the size of the arrowhead indicating the number of sequences. Enterobacteria in dark blue, γ -proteobacteria in green, α -proteobacteria in red, β -proteobacteria in yellow, spirochetes in magenta, actinobacteria in grey and firmicutes in brown.

- 1. Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucl Acid Res 25:3389-3402.
- Papadopoulos JS, Agarwala R. 2007. COBALT: constraint-based alignment tool for multiple protein sequences. Bioinformatics 23:1073-1079.

Figure S7. Genomic environment of genes encoding P19-like proteins (orange, 'COG-Tpd') and genes encoding FetM-like transporters (light blue, 'COG-FTR1'). The scale is in kbp and the organism is indicated above the gene symbols, which are in the direction of transcription.







Burkholderia pseudomallei 9 c COG-UorB	contig (+ strand <u>CDG-Ted</u> <u>Bese</u> u9_1	, 1826118 10100025090 <u>ci</u>	25) 0G-NapH	COG-G1cG	COG-HemP	C0G1741	COG-LysR	Bp <u>se</u> u9_010100
		DG-FTR1		>=>	Bpseu9_0101	00025115		COG-ExbD
					Bpseu9_	010100025120		
Burkholderia pseudomallei 91 ^{COG-UorB}	contig (+ stran COG-Ted Bese9 03	d, 190911	908) 0G-NapH	C0G-G1cG	COG-HemP	Bpse9 01010002595	51	Bese9 0101000:
		0G-FTR1			C Bpse9_01010	0025941	COG-LysR	COG-ExbD
	-				Bpse9_0	10100025946		
					_	C0G1741		
Burkholderia pseudomallei B72 COG-UurB	210 contig (+ st	rand, 1833.	.11832) 09-NacH	006-61-6	COG-HewP	0061741	COG-Luck	BoceB 0101000
		BpseB	_010100024706		BpseB_01010	0024726		COG-ExbD
					BpseB_0	10100024731		
Burkholderia pseudomallei BCC	215 contig (+ s	trand, 1826						
	CDG-TPId BPSeBC_I	010100024128 C	OG-NapH		COG-HemP	00024153	COG-LysR	COG-ExbD
	Ē	<u> </u>			BpseBC_	010100024158		
Burkholderia pseudomallei DM9	98 contig (+ str	and. 1827	11826)		<			
COG-Uvr8	COG-Tpd BpseD_0:	L0100025699 C	OG-NapH		COG-HemP	C0G1741	COG-LysR	BpseD_0101000:
	c L	OG-FTR1			BpseD_01010	10100025724		COG-ExbD
Dominia Informática a constructor a 11 dá 1404	-242	2 (1) (1)	407066 50706	~				
	COG-TPd BPSS 035	2 (+ suranu 8 C	0G-NapH		COG-HemP	C0G1741	COG-LysR	COG-ExbD
		0G-FTR1			BPSS 036	3		
Burkholderia pseudomallei NCT COG-UvrB	C 13177 contig COG-Ted BeseN_0:	(+ strand, 10100024509 <u>c</u>	181611815) OG-NapH	COG-G1cG	COG-HemP	C0G1741	COG-LysR	BpseN_0101000:
		DG-FTR1			BpseN_01010	0024534		COG-ExbD
	-				BpseN_0	10100024539		
Burkholderia pseudomallei Pas cog-UorB	steur contig (+ COG-Ted BeseP 03	strand, 717	417174) 0G-NapH	COG-G1cG	BeseP 0	3004871	COG-LysR	COG-ExbD
				>=		C0G1741		
Burkholderia pseudomallei S13	L 3 contig (+ stra	nd, 71961	7196)					10000000
C0G-UvrB	CDG-TPd BpseS_0	3000786 C	OG-NapH		BpseS_0	3000791	COG-LysR	C0G3666
Burkholderia sp. 383 chromoso	OME 1 (- strand, COG-Ted Boep181	26228062 94_A5536 C	632806) 0G-NapH	COG-Her	MP C0G174	1 COG-Lysk	2000	-Ехьр
	Ê				Bcep18194_A5532			C0G-To10
Burkholderia thailandensis Bt cog-UorB	4 contig (- str CDG-Ted BthaB_0:	and, 5875	15875) 0G-Naph	COG-G1cG	COG-HemP	C0G1741	COG-LysR	COG-ExbD
		0G-FTR1			BthaB_01010	0011480		
					BthaB_0	10100011475		51
Burkholderia thailandensis E2	264 chromosome I COG-Ted BTH II2	I (- strand	, 247388324838 Юд-Марн	383) C0G-61c6	COG-HemP	C0G1741		COG-ExbD
		0G-FTR1			BTH_112	:029	COG-LysR	
Burkholderia thailandensis MS	[SMB43 contig (+	strand. 176	2.11761)					
	COG-Tpd Bpse38_	010100020017 C	OG-NapH		COG-HemP	C061741	COG-LysR	COG-ExbD
	Ĕ			Des	e30_01010002003 Bpse38_0101	,00020047		
					Bpse38	010100020052		
Burkholderia thailandensis T>	(DOH contig (- s	trand, 4910	3.,59103)			3		
	COG-TPd BthaT_0:	L0100011883 C	OG-NapH		COG-HemP	COG1741	COG+Ly\$R	COG-ExbD
	Č				BthaT_0	10100011853		чĩ
Burkholderia ubonensis Bu cor	ntig (- strand.	57010570)						
	COG-Ted BuboB_01	.0100005606 CC	DG-NapH		P C06174:	1 COG-LysR	BuboB_0:	10100005571
				E		00		C0G-To19
Punkhal dan ini uriate ani unai - an	L obnorroome de l	- otherstand	540040 05500401					
COG-UorB	COG-Ted Bcep180:	- surand, 2 8_2293 C	06-NapH	Сод-не Сод-не	EMP C0617	41 COG-Lys	R C0	G-ExbD
	Ê	0G-FTR1			Bcep 18 08_2289			C0G-To10
Burkholderia xenovorans LB400 COG-UorB	chromosome 1 (<u>COG-TPd</u> <u>Bxe_</u> A3:	+ strand, 1 ¹⁵¹	45394814639482 COG-Naph) Bxe_A31/	48 B <u>×e_A314</u> 6	C0G1741	COG-LysR	COG-E:
		COG-FTR1			-HemP COG	1765		
				11111117-				



Yersinia pestis KIM (- strand COG-FTR1	d, 26049132614913) 92369	COG-Selly	C06-SalY	C0G+BcP x C0G2863	tehB	hmsH
Yersinia pestis Pestoides F (YPDSF_1182 YPDSF_1181	(- strand, 13229851332 C00-Tpd C004393	2985)	C0G-Lo1E		COG-TehB	COG-PepP
Yersinia pestis biovar Antiqu COG-FTR1	ua str. B42003004 contig C0G-TPd C0G4393	(+ strand, 222707232	707) C0G-Lo1E C0G-Sali	x cog2863	tehB>	YpB42003
Yersinia pestis biovar Antiqu COG-FTR1	ua str. E1979001 contig COG-TPId COG4393	(+ strand, 3046540465	COG-LoIE	x = = = = = = = = = = = = = = = = = = =	tehB	COG-PepP
Yersinia pestis biovar Antiqu COG-FTR1	ua str. UG05-0454 contig COG-TPd COG4393	g (- strand, 419545195	(4) COG-Lo1E COG-Sal:		tehB	COG-PepP
Yersinia pestis biovar Mediev YP1684 fTR12	valis 91001 (+ strand, 1 COG-TPd COG4393	L8586451868645)	COG-SalY	trxR1	tehB	PepP1
Yersinia pestis biovar Orient Y⊧F1991016_3889 ➡ COG-FTR1	COG-Trid	cog-Selly	437) YPF1991016_3894 COG-Sa1		3 tehB	COG-PepF
Versinia pestis biovar Urient	LD 247E9 (strand 24E5	coe-saly	305065) C06-Lo1E C06-Sali	x cog2863	tehB	COG-PepP
COG-FTR1	COG-Tpd COG4393	C06-541Y	C0G-Lo1E C0G-Sa1	x C06286	3 tehB	COG-PepF
Actinobacillus succinogenes 1	COG-Ted COG4393	COG-SalY	C0G-Lo1E	X C0G-BCP C0G2863	3 tehB	COG-PepF
COG-FTR1	multocida str. Pm70 (-	cog-Saly strand, 518685528685)		C061999 Asuc	R0071 COG-GutQ	COG177
PepE COG-FTRI	+ strand, 379226838022	C0G-SalY	C0G-Lo1E C0G-Sa1X	resA PHO	445 minC	
PST_3501 PST_3502 Photobacterium sp. SKA34 cont	COG-TR4 PST_3504 tig (+ strand, 7710076		S-Pf18 PST_3508		dnr:S COG4	273
SKA34_07738 SKA34_07748 COG-Io1E Vibrio shilonii AK1 contig (-	GG-Tpd SKR34_07758 COG-FTR1 COG-FTR1 - strand, 136811146811	C062837			5KA34_07788	
Compylobacter coli RM2228 cor	ntig (+ strand, 447175	54717)	COG-SalX	C0G-BCP C0G- C0G2847	NarC USAK1_19	COG-Link
Compylobacter concisus 13826	C064393 C06-Ted (+ strand, 3053593153	C0G-5a1V			~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	νΑ ΓΡ10
Çampylobacter curvus 525.92 (COG4393	C0G+SalY		CCC13826_0769 C9 CCC13826_077	nT	>
COG-FTR1 Campylobacter fetus subsp. fe	> etus 82-40 (+ strand, 52	29696539696)		CCV52592_0427	6 COG+C1PH	
Campylobacter hominis ATCC B4	COG-Ted AA-381 (- strand, 397942	C06-5a1Y				
	COG-Ted		COG-SalX	CHAB381_	10443 feoB	



Treponema pallidum subsp. pallidum str. Nichols (- strand, 10469401056940)		A
Acetobacter pasteurianus IFO 3283-01 (+ strand, 27360072746007) thic APR01_25240 ftr1	lysR COG-GabT	
Aggregatibacter actinomycetemcomitans D11S-1 (+ strand, 14787261488726)	C06-Bcp C06637 D115_15	67
Aggregatibacter aphrophilus NJ8700 (- strand, 566887576887)	C0622663 D115_1566	
COG-FTR1 COG-FTR1 COG-FTR1 COG-Salv COG-Sa	Bind_0394 C06535	
Bifidobacterium angulatum DSM 20098 contig (+ strand, 4315853158) BIFING_00636 COG-Ted COG4393 COG-Salv COG-Salv COG-Salv	Bind_0395	Π
Bifidobacterium bifidum NCIMB 41171 contig (+ strand, 99469) BbifNu 01000009251Y cog-LoIE cog-Salv	C061511	1010(
COG-FTR1 Bifidobacterium breve DSM 20213 contig (+ strand, 3956249562)	BbitN4_010100006958	4_010 ≥
Bifidobacterium catenulatum DSM 16992 contig (+ strand, 147516157516)	-SalX	
C06-FTR1 C06-Te4 BIF6R_00142 C06-SalV C	BIFCAT_00147 C061511	-
	BDP_0169 COG-PurR	cog [→
COG-FTRI	Blon_0202	-
Bifidobacterium longum subsp. infantis CCUG 52486 contig (- strand, 7307883078) COG-TR1 COG-TR1 COG-TR4 COG-SalX COG-	BloniC5_010100002317 C061511	
Bordetella avium 197N (+ strand, 13159861325986) phbC phbB phbF C06-FTR1 C06-NapH C06384 BAV1229 C06323 BAV1225 BAV1225 BAV1229 C06323	7 cph81	-
Brucella abortus S19 chromosome 2 (- strand, 819341829341) COG-PFs COG-GuaA BRbS19_II07770 COG-NapH	C0G-ackA C0G	G-Cyp
Brucella melitensis ATCC 23457 chromosome II (+ strand, 340880350880) COG-Pfs guaA COG-Tpid COG-FTR1		30368 G-Cyf
Brucella microti CCM 4915 chromosome 2 (+ strand, 361154371154) C065293 BHI_II376 BHI_II377 BHI_II379 BHI_II374 C00-Tpd C06-FTR1		[384 G-Cyr
BHI_II375 Burkholderia ambifaria IOP40-10 contig (+ strand, 168511684) C000-Uref C000-Ted Bam10P4010DRAFT_5519 C00-HerP C001741	COG-LysR COG-ExbD	
C0G-FTR1	3 3	o 1Q
Burkholderia ambifaria MEX-5 contig (+ strand, 171911718) COG-UurB COG-FTRI COG-FTRI COG-FTRI	COG-LysR COG-ExbD	010
Burkholderia cenocepacia J2315 chromosome 1 (- strand, 25442872554287)	C0G-LysR C0G-ExbD	010
Burkholderia glumae BGR1 chromosome 1 (- strand, 29037632913763) COG-UwrB COG-Ted belu_122520 COG-FTR1 COG-FTR1 COG-FTR1	COG-LysR COG-ExbD	19



Providencia alcalifaciens DSM 30120 c COG-Hiab PROVALCAL_02412 COG-Ted	ontig (+ strand, 1213901 PROVALCAL_02414 co62822	.31390)	PROVALCAL_02418	COG-LysR	COG-AdeC
	COG-FTR1	C0G2837	PROVALCAL_02419		
Providencia rettgeri DSM 1131 contig COG-MiaB PROVRETT_01162 COG-Tpd	(- strand, 4538255382) PROVRETT_01160 C062822 > □		PROVRETT_01156 PROVRETT_01154	COG-LysR	COG-AdeC
			>		
Providencia rustigianii DSM 4541 cont COG-HiaB PROVRUST_01722 COG-TPd	ig (+ strand, 4360253602 PROVRUST_01720 COG2822 COG-FTR1	2) C062837	PROURUST_01716PROVRUST_01715	COG-LysR	COG-AdeC
Salinibacter ruber (- strand, 2614444 co63002	2624444) SRM_02289 C062353	SRH_02267	SRH_02265 SRH_02264		К
Selenomonas sputigena ATCC 35185 cont coG-ExbD coG-FTR1 COG-Ted COG-TonB	ig (+ strand, 3306343063 co64393 ≻	3) C0G-Lo1E	C0G-Sa1X C0G-AP	DE COG534	COG4769
Treponema pallidum subsp. pallidum SS phes COG-FTR1 tpd	14 (- strand, 1048385105 ▼PASS_0969 ▼PASS_0970 ▼	38385) ASS_0968	TPRSS_0967 TPRSS_0	966	COG-AcrA
Yersinia pseudotuberculosis PB1/+ (+ COG-FTR1	strand, 21965872206587) YPTS_1990 C064393	-SalY		0G2863 C0G-TehB	
Yersinia pseudotuberculosis YPIII (- COG-FTR1	strand, 24902932500293) YPK_2250 C004393	Saly COG-Lol		0G2863 C0G-TehB	