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A Squaraine-based Dipicolylamine Derivative Acting as a Turn-on Mercury(II) Fluorescent Probe in Water

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Reference	Authors	Medium
9a	G. Wang et al.	EtOH/water solutions and pH 7.0 PBS buffer solution
9b	X. Liu <i>et al.</i>	DMSO, AcOH and SDS solutions
9c	S. Lee <i>et al.</i>	MeCN
9d	H. Zhu <i>et al.</i>	0.005% TW-80 (Tween-80 or polysorbate 80) at pH 5.0 PB buffer solutions
9e	HS. So <i>et al.</i>	MeCN
9f	SY. Lin <i>et al.</i>	EtOH/water solutions
9g	B. A. Rao <i>et al.</i>	MeCN
9h	Q. Lin <i>et al.</i>	EtOH/water (30:70, v/v) solutions
9i	L. Hu <i>et al</i> .	DMSO/water (1:1, v/v) solutions and pH 8.0 PB buffer with 0.01 mol L^{-1} CTMAB (cetyltrimethylammonium bromide)
9j	K. M. Shafeekh <i>et al.</i>	МеОН
9k	C. Luo <i>et al.</i>	MeCN and MeCN/water (2:1, v/v) solutions
91	C. Chen <i>et al.</i>	AcOH and AcOH/water (10:90, v/v) solutions
9m	C. Chen <i>et al.</i>	AcOH/water (40:60, v/v) solution
9n	Y. Xu <i>et al.</i>	Aqueous solution with cucurbit[8]uril (CB8)
9р	M. C. Basheer et al.	DCM
9q	J. V. Ros-Lis et al.	MeCN/water (1:4, v/v) pH 9.6 CHES buffer solutions
9r	J. V. Ros-Lis et al.	MeCN/water (20:80, v/v) pH 6.9 HEPES buffer solutions

Table S1. Squaraine-based sensors for Hg²⁺ from the literature and the working medium used

(a) G. Wang, W. Xu, H. Yang and N. Fu, Highly Sensitive and Selective Strategy for Imaging Hg²⁺ Using Near-Infrared 9 Squaraine Dye in Live Cells and Zebrafish, Dyes and Pigments, 2018, 157, 369-376; (b) X. Liu, N. Li, M. M. Xu, C. Jiang, J. Wang, G. Song and Y. Wang, Dual Sensing Performance of 1,2-Squaraine for the Colorimetric Detection of Fe³⁺ and Hg²⁺ ions, Materials, 2018, 11, 1998; (c) S. Lee, B. A. Rao and Y.-A. Son, A Highly Selective Fluorescent Chemosensor for Hg²⁺ Based on a Squaraine–bis(rhodamine-B) Derivative: Part II, Sensors and Actuators B: Chemical, 2015, 210, 519– 532; (d) H. Zhu, Y. Lin, G. Wang, Y. Chen, X. Lin and N. Fu, A Coordination Driven Deaggregation Approach toward Hg²⁺specific Chemosensors Based on Thioether Linked Squaraine-Aniline Dyads, Sensors and Actuators B: Chemical, 2014, **198**, 201–209; (e) H.-S. So, H.-S. So, B. A. Rao, J. Hwang, K. Yesudas and Y.-A. Son, Synthesis of Novel Squaraine– bis(rhodamine-6G): A Fluorescent Chemosensor for the Selective Detection of Hg²⁺, *Sensors and Actuators B: Chemical*, 2014, 202: 779–787; (f) S.-Y. Lin, H.-J. Zhu, W.-J. Xu, G.-M. Wang and N.-Y. Fu, A Squaraine Based Fluorescent Probe for Mercury Ion via Coordination Induced Deaggregation Signaling, Chinese Chem. Lett., 2014, 25, 1291-1295; (g) B. A. Rao, H. Kim and Y.-A. Son, Synthesis of Near-Infrared Absorbing Pyrylium-Squaraine Dye for Selective Detection of 2⁴, Sensors and Actuators B: Chemical, 2013, **188**, 847–856; (h) Q. Lin, Y. Huang, J. Fan, R. Wang and N. Fu, A Hgʻ Squaraine and Hg²⁺-based Colorimetric and "Turn on" Fluorescent Probe for Cysteine, Talanta, 2013, 114, 66–72; (i) L. Hu, Y. Zhang, L. Nie, C. Xie and Z. Yan, Colorimetric Detection of Trace Hg²⁺ with Near-Infrared Absorbing Squaraine Functionalized by Dibenzo-18-crown-6 and its Mechanism, Spectrochim. Acta A Mol. Biomol. Spectrosc., 2013, 104, 87-91; (j) K. M. Shafeekh, M. K. A. Rahim, M. C. Basheer, C. H. Suresh and S. Das, Highly Selective and Sensitive Colourimetric Detection of Hg²⁺ lons by Unsymmetrical Squaraine Dyes, Dyes and Pigments, 2013, 96, 714-721; (k) C. Luo, Q. Zhou, B. Zhang and X. Wang, A New Squaraine and Hg²⁺-based Chemosensor with Tunable Measuring Range for Thiol-Containing Amino Acids, New J. Chem., 2011, 35, 45-48; (I) C. Chen, H. Dong, Y. Chen, L. Guo, Z. Wang, J. J. Sun and N. Fu, Dual-mode Unsymmetrical Squaraine-based Sensor for Selective Detection of Hg²⁺ in Aqueous Media. Org. Biomol. Chem., 2011, 9, 8195-8201; (m) C. Chen, R. Wang, L. Guo, N. Fu, H. Dong, Y. Yuan, A Squaraine-based Colorimetric and "Turn on" Fluorescent Sensor for Selective Detection of Hg^{2+} in an Aqueous Medium, Org. Lett., 2011, 13, 1162–1165; (n) Y. Xu, M. J. Panzner, X. Li, W. J. Youngs and Y. Pang, Host-guest Assembly of Squaraine Dye in Cucurbit[8]uril: Its Implication in Fluorescent Probe for Mercury Ions, Chem. Commun., 2010, 46, 4073-4075; (o) E. M. Nolan and S. J. Lippard, Tools and Tactics for the Optical Detection of Mercuric Ion, Chem. Rev., 2008, 108, 3443–3480; (p) M. C. Basheer, S. Alex, K. G. Thomas, C. H. Suresh and S. Das, A Squaraine-based Chemosensor for Hg^{2+} and Pb²⁺ Tetrahedron, 2006, 62, 605-610; (q) J. V. Ros-Lis, M. D. Marcos, R. Martínez-Máñez, K. Rurack and J. Soto, A Regenerative Chemodosimeter Based on Metal-Induced Dye Formation for the Highly Selective and Sensitive Optical Determination of Hg^{2+} lons, *Angew. Chem. Inter. Ed.*, 2005, **44**, 4405–4407; (r) J. V. Ros-Lis, R. Martínez-Máñez, K. Rurack, F. Sancenón, J. Soto and M. Spieles, Highly Selective Chromogenic Signaling of Hg^{2+} in Aqueous Media at Nanomolar Levels Employing a Squaraine-Based Reporter, Inorg. Chem., 2004, 43, 5183-5185.

Equilibrium reaction	sbdpa	dpa ^b	
	$\log \beta_i^{H}$		
$L + H^{+} \rightleftharpoons HL^{+}$	10.85(2)	7.11	
$L + 2 H^+ \rightleftharpoons H_2 L^{2+}$	14.80(6)	9.59	
$L + 3 H^{+} \rightleftharpoons H_{3}L^{3+}$	18.12(4)	-	

Table S2. Overall (β_i^{H}) protonation constants of **sbdpa** and dpa in aqueous solution at 298.2±0.1 K and in 0.10±0.01 M KNO₃

^a This work; L denotes the ligand in general; values in parenthesis are standard deviations in the last significant figures. ^b T = 293.2 K, I = 0.1 M in KNO₃.²⁸

Table S3. Overall ($\beta_{M_mH_hL_l}$) stability constants of the complexes of **sbdpa** and dpa with Hg²⁺, Cu²⁺ and Zn²⁺ in aqueous solution at 298.2 K±0.1 in 0.10±0.01 M KNO₃

	$\log eta_{M_{m}H_{h}L_{l}}$					
Equilibrium reaction ^a	sbdpa			dpa ^b		
	Hg ²⁺	Cu ²⁺	Zn ²⁺	Hg ²⁺	Cu ²⁺	Zn ²⁺
$M^{2+} + H^{+} + L \rightleftharpoons [M^{II}HL]$	18.06(5)	16.01(3)				
$M^{2+}+L \rightleftharpoons [M^{II}L]$	14.60(4)	11.75(4)	8.4(1)		13.85	7.63
$M^{2+}+L \rightleftharpoons [M^{II}LOH] + H^{+}$		3.18(5)				
$M^{II} + L \rightleftarrows [M^{II}L(OH)_2] + 2H^+$		-7.28(6)				
M^{2+} + 2 L \rightleftharpoons [$M^{II}L_2$]				22.25	18.5	12.15
$2 \operatorname{M}^{2^+} + \operatorname{H}^+ + \operatorname{L} \rightleftarrows [\operatorname{M}_2^{ {}^{II}}\operatorname{HL}]$		19.76(7)	17.84(4)			
$2 M^{\parallel} + L \rightleftharpoons [M_2^{\parallel}L]$	18.97(9)	16.30(7)	13.14(7)			
$2 M^{II} + L \rightleftarrows [M_2^{II}LOH] + H^+$		9.43(9)	5.9(1)			
$2 M^{II} + L \rightleftarrows [M_2^{II} L(OH)_2] + 2H^+$		1.64(9)	-1.4(1)			
$2M^{\parallel} + L \rightleftharpoons [M_2^{\parallel}L(OH)_3] + 3H^+$		-7.2(1)	-9.5(1)			
$2M^{II} + L \rightleftarrows [M_2^{II}L(OH)_4] + 4H^+$		-17.0(1)	-19.6(1)			

^a This work; L denotes the ligand in general; values in parenthesis are standard deviations in the last significant figures. ^b T = 293.2 K, I = 0.1 M in KNO₃, G. Anderegg, E. Hubmann, N. G. Podder and F. Wenk, XI. Pyridinderivate als Komplexbildure. XI. Die Thermodynamik der Metallkomplexbildung mit Bis-, Tris- und Tetrakis [(2-pyridyl)methyl]-aminen, *Helv. Chim. Acta*, 1977, **60**, 123–140.



Fig. S1 Absorption and emission spectra of **sbdpa** in aqueous solution with pH variations at $C_L = 1.3 \times 10^{-3}$ M and T = 298.2 K. $\lambda_{exc} = 420$ nm.



Fig. S2 Absorption and emission spectra of the complex of **sbdpa** with Hg²⁺ 1:1 M:L in aqueous solution at pH 3.1; $C_L = 1.3 \times 10^{-3}$ M; T = 298.2 K. $\lambda_{exc} = 340$ nm.



Fig. S3 Fluorescence intensity change of **sbdpa** (2.5×10^{-5} M) upon titration with Hg(NO₃)₂ in aqueous buffered solution with MES (2.5×10^{-3} M) at pH 5.0 and *T* = 298.2 K. λ_{exc} = 340 nm.



Fig. S4 Species distribution diagrams calculated for the complexes of **sbdpa** with Cu^{2+} and Zn^{2+} cations at 2:1 M:L ratio. $C_M = 2C_L = 2.0 \times 10^{-3}$ M. L denotes the ligand.



Fig. S5 Competition distribution diagram of the overall amounts of the **sbdpa** in function of pH in presence of Hg^{2+} , Cu^{2+} , and Zn^{2+} in the 1:1:5:5 ratio. $C_{Hg^{2+}} = C_L = 2.5 \times 10^{-5}$ M, $C_{Cu^{2+}} = C_{Zn^{2+}} 1.25 \times 10^{-4}$ M. L denotes the **sbdpa** ligand.



Fig. S6 1 H-NMR spectrum of sbdpa in CDCl₃.



Fig. S7 ¹³C-NMR spectrum of sbdpa in CDCl₃.











Fig. S10 ESI mass spectrum of sbdpa in H₂O/MeOH.



Fig. S11 ATR-FTIR spectrum from **sbdpa** recorded at room temperature; no characteristic $v_{C=0}$ bands found at \cong 1700 cm⁻¹, nor typical water bands at 3408, 1644, and 700 cm⁻¹, supporting the fact that sbdpa is in its zwitterionic form, and that it is not an hydrate although it is recrystallized from water.



Fig. S12 Evolution of ¹H-NMR spectra of **sbdpa** in D_2O with time at pD 3.39 (top) and pD 7.63 (bottom).