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Electronic Supplementary Information

A natural cyanobacterial protein C-phycoerythrin as an Hg²⁺ selective fluorescence probe

in aqueous systems

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Table of contents

S. No.	Торіс	Page No.
S1	Purification and characterization of CPE	S3
S2	Loss of CPE color with 7 μ M Hg ²⁺	S4
S 3	Decrease in fluorescence intensity w.r.t [Hg ²⁺]	S5
S4	Stern Volmer curve for CPE – Hg ²⁺ interaction	S6
S 5	Fluorescence life time analysis	S7
	References	S8

S1. CPE purification and characterization

The growth of the cyanobacterium *Lyngbya* sp. CCNM 2053 was carried out using previously published reports ¹. Briefly, fresh biomass was suspended in 0.1 M potassium phosphate buffer pH 7.0 at a solid liquid ratio of 75 mg ml⁻¹ (wet basis) and repeatedly frozen and thawed at -80 and 27 °C respectively. After 3 such cycles, the mixture was centrifuged (10,000 g, 10 °C, 10 minutes) and the supernatant was subjected to 25 % followed by 60 % ammonium sulphate precipitation. The pellet obtained after 60 % ammonium sulphate saturation was dissolved in minimal quantity of extraction buffer and extensively dialyzed against it. The partially purified CPE was further purified by ion-exchange (DEAE Sepharose, Sigma Aldrich, USA). The pink fractions were pooled together and brought to 60 % ammonium sulphate saturation and left overnight. After centrifugation, the pellet was redissolved in minimum quantity of the extraction buffer and dialyzed against it overnight. CPE quantification was carried out using the following equations ²:

CPC (mg ml - 1) = [A615 - 0.474A652]/5.34

(1)

APC (mg ml - 1) = [A652 - 0.208A615]/5.09(2)

CPE (mg ml - 1) = [A562 - 2.41(CPC) - 0.849(APC)]/9.62(3)

The purity ratio of CPE was calculated using the A_{562}/A_{280} ratio while its characteristics were verified using UV-visible and fluorescence spectroscopy.

S2. Loss of CPE color with 7 μ M Hg²⁺



Fig. S1. CPE (left) before and (right) after interaction with 7 μM Hg^{2+}.



Fig. S2. Decrease in fluorescence emission of CPE under increasing [Hg²⁺].



Fig. S3: Stern Volmer curve for CPE - Hg²⁺ interaction



Fig. S4. Fluorescence lifetime of CPE in a) absence of Hg²⁺ and, b) in presence of 3 μ M Hg²⁺

References

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