Electronic Supplementary Material (ESI) for New Journal of Chemistry. This journal is © The Royal Society of Chemistry and the Centre National de la Recherche Scientifique 2020

Electronic Supplementary Information (ESI)

Greener Synthesis of 1,2,3-Triazoles using Copper (I)-Exchanged Magnetically Recoverable β-Zeolite as Catalyst

Elizama R. Costa,^a Floyd C. D. Andrade,^a Danilo Yano de Albuquerque,^a Luanne E. M. Ferreira,^b Thiago M. Lima,^{*b} Carolina G. S. Lima,^c Domingos S. A. Silva,^d Ernesto A. Urquieta-González,^d Márcio W. Paixão,^a and Ricardo S. Schwab^{*a}

^a Centre of Excellence for Research in Sustainable Chemistry (CERSusChem), Departamento de Química, Universidade Federal de São Carlos - UFSCar, Rodovia Washington Luís, km 235 - SP-310, São Carlos, São Paulo, 13565-905, Brazil. E-mail: rschwab@ufscar.br

^b Departamento de Química Inorgânica, Universidade Federal Fluminense, Campus do Valonguinho, Outeiro São João Batista s/n, Centro, Niterói-RJ, 24020-150, Brazil. E-mail:tmlima@id.uff.br

^c Departamento de Química Orgânica, Universidade Federal Fluminense, Campus do Valonguinho, Outeiro São João Batista s/n, Centro, Niterói-RJ, 24020-150, Brazil.

^d Research Center on Advanced Materials and Energy, Universidade Federal de São Carlos (DEQ), C. Postal 676, 13565-905, São Carlos, SP, Brazil

Table of Contents

1. Experimental	S 3
1.1 Materials and Methods	S 3
1.2 General Procedure for the 1,3-Dipolar Cycloaddition Reaction	S 4
2. Characterization data	S5
2.1 Catalyst characterization	S5
2.2 Characterization data of 1,4-dissubstituted 1,2,3-triazoles	S 10
3. ¹ H and ¹³ C NMR Spectra of the 1,4-dissubstituted 1,2,3-triazoles	S14

1. Experimental

1.1 Materials and Methods

Hydrogen nuclear magnetic resonance spectra (¹H NMR) were obtained, on Bruker Advance 400 MHz spectrometer. ¹H NMR spectra were referenced to the residual hydrogen signal in CDCl₃ at δ = 7.26 ppm. Coupling constants (*J*) are reported in Hertz [Hz] using the standard notation to described the multiplicity of signals. Carbon-13 nuclear magnetic resonance spectra (¹³C NMR) were obtained at 101 MHz. ¹³C spectra were referenced to the CDCl₃ triplet signal at δ = 77.0 ppm. Flash column chromatography was performed using Merck Silica Gel (230-400 mesh). Thin layer chromatography (TLC) was performed using Merck Silica Gel GF254, 0.25 mm thickness. For visualization, TLC plates were either placed under short wave ultraviolet light (254 nm), stained with iodine vapor, or acidic vanillin. The yields of the products included in all tables refer to isolated yields. All commercially available reagents were purchased at the highest commercial quality and used without further purification, unless otherwise noted.

The particle size and morphological studies of the Fe_3O_4 microspheres used as support and the catalysts were performed by Transmission Electron Microscopy (TEM), which was carried out using a FEI TECNAI G2 F20 microscope. For the TEM analyses, the powder samples were dispersed in ethanol and sonicated for 5 min. One drop of this solution was placed on a 400 mesh nickel grid with carbon film and the sample was dried at room temperature.

The XRD measurements were performed in a Shimadzu XRD 6000 Diffractometer using the K α radiation of a Cu source (λ =1.54056 Å), 40 kV, 30 mA and 2 θ in the range of 5° to 80° and a rate of 2° min⁻¹.

X-ray Photoelectron Spectroscopy (XPS) was performed using a Scienta Omicron ESCA+ spectrometer with a high-performance hemispheric analyzer (EA 125) with monochromatic Al K α (hv = 1486.6 eV) radiation as the excitation source. The operating pressure in the ultrahigh vacuum chamber (UHV) during analysis was 2x10⁻⁹ mbar. Energy steps of 50 and 20 eV were used for the survey and high-resolution spectra, respectively. TPD measurements were conducted in a Micromeritics Autochem II 2920 Chemisorption Analyzer equipment with a TCD (Thermal Conductivity Detector) detector. Fifty miligrams of the sample were pre-treated under 30 mL min⁻¹ of He flow at a heating rate of 10 °C min⁻¹ until 500 °C and kept at this temperature for 30 minutes. After this period, the reactor was cooled to 120 °C, and the He flow was kept for 60 minutes.

ICP-OES measurements were conducted in a Thermo Fisher Scientific, iCAP 6300 Duo, with a CID (Charge Injection Device) detector.

The magnetic measurements were carried out using and apparatus combining a superconducting quantum interference device and a vibrating sample magnetometer model Quantum Design MPMS SQUID-VSM. The magnetization was measured as function of the applied magnetic field (MxH) at room temperature and up to 70 kOe.

Thermogravimetric analyses were performed in a Shimadzu, thermogravimetric module TGA-50.

The textural properties of the materials were determined by nitrogen physisorption measurements at -196°C at a Micromeritics equipment (ASAP-2420). Previously to the analyses, the samples were treated under vacuum at 90 °C for 30 minutes and 200 °C for 6 h in order to eliminate water and physically adsorbed gases. The pore size distribution was calculated using the Barrett-Joyner-Halenda (BJH) model, and the specific surface area was calculated using the BET equation. The micropore volume and external area were estimated through the t-plot method using Halsey's equation. The total micropore volume was calculated at $p/p_0 = 0.98$.

The FT-IR analyses were conducted in the transmission mode using KBr as a dillutant. The pellet containing 1% (m/m) of the sample was analyzed in a Bruker Vertex 70 spectrometer equipped with a L-alanine and deuterium-dopped triglycine sulfate dectector. The spectrum was collected in the range of 400 to 4000 cm⁻¹ with a resolution of 4 cm⁻¹.

1.2. General Procedure for the 1,3-Dipolar Cycloaddition Reaction

Typical procedure:



In an Ace® pressure tube, the γ -Fe₂O₃- β -Cu(I) catalyst (9.0 mol%), alkyl halide (0.5 mmol), sodium azide (0.5 mmol), alkyne (0.5 mmol), and 3.0 mL of water were added. The reaction tube was sealed and heated in oil bath at 110°C and magnetically stirred for 12 h. After the completion of the reaction and separation of catalyst with the aid of a permanent magnet, the product was extracted from the catalyst by washing with ethyl

acetate. The solvents and volatiles were completely removed under vacuum to give the crude product. The resulting crude compound was purified by flash column chromatography on silica gel (hexane:ethyl acetate, 7:3). The catalyst was then washed with ethyl acetate, water and ethanol, dried at 100°C for 12 h and reused in a subsequent reaction run. The isolated product was then subjected to ¹H NMR and ¹³C NMR.

2. Characterization data

2.1 Catalyst characterization



Figure S1. XRD diffractograms of Fe₃O₄ nanoparticles and magnetically recoverable copperexchanged beta zeolite catalysts (F: Fe₃O₄/ γ -Fe₂O₃; B: β -zeolite; Z: ZSM-12 zeolite; C: CuO)



Figure S2. TEM images of (a) pure β -zeolite, (b) γ -Fe₂O₃- β -Na⁺ and (c) γ -Fe₂O₃- β -Cu(I).



Figure S3. XPS spectra of a) the fresh, b) the deactivated and c) the reactivated catalysts γ -Fe₂O₃- β -Cu(I).

Catalyst	Component	Position (eV)	FWHM	Concentration (%)
Fresh catalyst	Cu ⁺ species	932.39	2.45	59.8
	Cu ²⁺ species	934.15	5.18	29.1
	Cu ²⁺ satellite	942.04	2.09	11.1
Catalyst	Cu ⁺ species	932.19	2.58	42.6
after 7 th	Cu ²⁺ species	934.02	5.23	31.5
reaction run	Cu ²⁺ satellite	940.69	3.20	25.8
Recalcined . catalyst	Cu ⁺ species	932.489	3.19	51.7
	Cu ²⁺ species	936.68	5.79	27.0
	Cu ²⁺ satellite	941.24	4.32	21.3

Table S1. Relative quantitative amount of each Cu components in the XPS spectrum of copper

 exchanged magnetically recoverable beta zeolite.



Figure S4. SQUID magnetization curves of (a) Fe_3O_4 microspheres and (b) copper exchanged magnetically recoverable beta zeolite [γ -Fe₂O₃- β -Cu(I)].



Figure S5. Thermogravimetric curves of (a) deactivated catalyst after 7th reuse (b) reactivated catalyst after calcination under 550 °C for 3 h and (c) catalyst after the 8th reaction run.

Catalyst	Weight loss	Temperature	Weight loss
Catalyst	step	range	%
Catalyst after	1 st step	25-150 °C	5.896
7 th reaction run	2 nd step	150-550 °C	6.407
Reactivated	1 st step	25-150 °C	6.231
catalyst	2 nd step	150-550 °C	2.000
Catalyst after	1 st step	25-150 °C	6.232
8 th reaction run	2 nd step	150-550 °C	6.247

Table S2. Weight loss assigned to the steps in the curves of the Figure S4.



Figure S6. Nitrogen physisorption isotherms of the fresh, deactivated after 7th reaction run and reactivated catalyst after thermal treatment (550 °C for 3 h).

Sample	$S_{External}$ (m ² g ⁻¹)	$S_{BET} (m^2 g^{-1})$	V _{Micropore} (m ³ g ⁻¹)	V _{Toral} (m ³ g ⁻¹)*
Fresh catalyst	52	196	0.06	0.24
Deactivated after 7 th run	78	202	0.05	0.25
Reactivated after thermal treatment	71	210	0.05	0.24

Table S3. Textural properties of the fresh, deactivated after 7th run and reactivated after thermal treatment.



Figure S7. FT-IR spectra of a) β -zeolite in the sodium form and the magnetically recoverable β -zeolite in the sodium form and b) the fresh and spent catalysts.



Figure S8. TEM image of the reactivated catalyst γ -Fe₂O₃- β -Cu(I) after the calcination procedure.



Figure S9. TEM image and chemical mapping of the reactivated catalyst γ -Fe₂O₃- β -Cu(I) after the calcination procedure.

2.2 Characterization data of 1,4-dissubstituted 1,2,3-triazoles



1-benzyl-4-phenyl-1H-1,2,3-triazole (**3a e 3b**): ¹H NMR (400 MHz, CDCl₃) δ 7.86-7.78 (m, 2H), 7.66 (s, 1H), 7.50-7.38 (m, 5H), 7.36-7.29 (m, 3H), 5.58 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 148.22, 134.73, 130.48, 129.26, 128.92, 128.33, 128.17, 125.83, 119.70, 54.37.



1-(4-bromobenzyl)-4-phenyl-1H-1,2,3-triazole (**3c**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.80 (d, J = 8.3 Hz, 1H), 7.67 (s, 1H), 7.51 (d, J = 8.3 Hz, 1H), 7.40 (t, J = 7.4 Hz, 1H), 7.32 (t, J = 7.3 Hz, 1H), 7.17 (d, J = 8.2 Hz, 1H), 5.52 (s, 1H). ¹³**C NMR** (101 MHz, CDCl₃) δ 148.40,

137.84, 133.66, 132.37, 130.26, 129.68, 128.87, 128.35, 125.75, 123.01, 119.47, 53.61.



1-(4-chlorobenzyl)-4-phenyl-1H-1,2,3-triazole (**3d**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.82-7.78 (m, 2H), 7.66 (s, 1H), 7.45-7.30 (m, 6H), 7.25 (d, J = 9.1 Hz, 3H), 5.55 (s, 2H). ¹³**C NMR** (101 MHz, CDCl₃) δ

148.52, 134.98, 133.31, 130.47, 129.50, 128.98, 128.42, 125.84, 119.56, 53.61.



1-(4-nitrobenzyl)-4-phenyl-1H-1,2,3-triazole (**3e**): ¹**H NMR** (400 MHz, CDCl₃) δ 8.24 (d, J = 8.7 Hz, 2H), 7.81 (d, J = 8.6 Hz, 2H), 7.76 (s, 1H), 7.48-7.38 (m, 4H), 7.34 (tt, J = 6.3, 1.4 Hz, 1H), 5.70 (s, 2H). ¹³**C NMR** (101 MHz, CDCl₃) δ 148.88, 148.24, 141.86, 130.21, 129.06, 128.68, 128.64, 125.88, 124.50, 119.80, 53.33.



1-(3-methoxybenzyl)-4-phenyl-1H-1,2,3-triazole (**3f**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.81-7.78 (m, 2H), 7.67 (s, 1H), 7.41-7.37 (m, 2H), 7.33-7.26 (m, 2H), 6.90-6.88 (m, 2H), 6.83-6.82 (m, 2H), 5.53 (s, 2H), 3.77 (s, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 160.24, 148.32, 136.23, 130.63, 130.34, 128.91, 128.27, 125.80, 120.35, 119.64, 114.34, 113.75, 55.42,

54.26.



1-(2-bromobenzyl)-4-phenyl-1H-1,2,3-triazole (**3g**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.84-7.80 (m, 2H), 7.79 (s, 1H), 7.63 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.44-7.36 (m, 2H), 7.36-7.28 (m, 2H), 7.26 – 7.17 (m, 2H), 5.71 (s, 2H). ¹³**C NMR** (101 MHz, CDCl₃) δ 134.21, 133.25, 130.46, 130.31, 128.85, 128.29, 125.77, 123.44, 119.86, 53.92.



1-(2,6-dichlorobenzyl)-4-phenyl-1H-1,2,3-triazole (**3h**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.80 (d, *J* = 7.3 Hz, 2H), 7.70 (s, 1H), 7.46-7.36 (m, 4H), 7.35-7.28 (m, 2H), 5.91 (s, 2H). ¹³**C NMR** (101 MHz, CDCl₃) δ 147.68, 136.87, 131.15, 130.40, 130.12, 128.96, 128.79, 128.19, 125.76, 119.28, 49.14.



13.96.



1-hexyl-4-phenyl-1H-1,2,3-triazole (**3i**): ¹**H** NMR (400 MHz, CDCl₃) δ 7.86 – 7.81 (m, 1H), 7.76 (s, 1H), 7.44-7.38 (m, 1H), 7.35-7.29 (m, 1H), 4.38 (t, *J* = 7.2 Hz, 1H), 2.00-1.88 (m, 1H), 1.38-1.27 (m, 3H), 0.95-0.82 (m, 2H). ¹³**C** NMR (101 MHz, CDCl₃) δ 147.66, 130.59, 128.85, 128.14, 125.72, 119.50, 50.51, 31.17, 30.31, 26.17, 22.42,

1-benzyl-4-(4-nitrophenyl)-1H-1,2,3-triazole (**3j**): ¹**H NMR** (400 MHz, CDCl₃) δ 8.27 (d, J = 9.0 Hz, 2H), 7.97 (d, J = 9.0 Hz, 2H), 7.79 (s, 1H), 7.45-7.38 (m, 3H), 7.36-7.31 (m, 2H), 5.61 (s, 2H). ¹³**C NMR** (101 MHz, CDCl₃) δ 147.43, 146.15, 136.90, 134.27, 129.44, 129.21, 128.33, 126.26, 124.41, 121.09, 54.62.



1-benzyl-4-(4-bromophenyl)-1H-1,2,3-triazole (**3k**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.67 (d, *J* = 8.7 Hz, 1H), 7.65 (s, 1H), 7.52 (d, *J* = 8.7 Hz, 1H), 7.43-7.35 (m, 2H), 7.34-7.29 (m, 1H), 5.57 (s, 1H). ¹³**C NMR** (101 MHz, CDCl₃) δ 147.14, 134.43, 131.98, 129.37, 129.24, 128.93, 128.15, 127.26, 122.14, 119.61, 54.39.



1-benzyl-4-(p-tolyl)-1H-1,2,3-triazole (**3l**): ¹**H** NMR (400 MHz, CDCl₃) δ 7.69 (d, *J* = 8.1 Hz, 1H), 7.62 (s, 1H), 7.42-7.35 (m, 1H), 7.33-7.29 (m, 1H), 7.21 (d, *J* = 7.9 Hz, 1H), 5.57 (s, 1H), 2.36 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 148.11, 138.22, 134.57, 129.53, 129.18, 128.84, 128.12, 127.33, 125.70, 119.28, 54.38, 21.29.



1-benzyl-4-(4-methoxyphenyl)-1H-1,2,3-triazole (**3m**): ¹**H NMR** (400 MHz, CDCl₃) δ 7.72 (d, *J* = 8.9 Hz, 2H), 7.57 (s, 1H), 7.40-7.35 (m, 3H), 7.33-7.28 (m, 2H), 6.93 (d, *J* = 8.9 Hz, 2H), 5.56 (s, 2H), 3.83 (s, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 159.73, 148.25, 134.90, 129.28, 128.89, 128.19, 127.14, 123.40, 118.81, 114.34, 55.46, 54.35.



1-benzyl-4-(o-tolyl)-1H-1,2,3-triazole (**3n**): ¹**H** NMR (400 MHz, CDCl₃) δ 7.77-7.72 (m, 1H), 7.56 (s, 1H), 7.42-7.35 (m, 3H), 7.33-7.29 (m, 2H), 7.25 (dd, *J* = 3.4, 2.3 Hz, 3H), 5.61 (s, 2H), 2.43 (s, 3H). ¹³**C** NMR (101 MHz, CDCl₃) δ 147.69, 135.61, 134.92, 130.97, 129.98, 129.26, 129.00, 128.86, 128.27, 1285.09, 126.17, 121.78, 54.27, 21.50.



1-benzyl-4-butyl-1H-1,2,3-triazole (**3o**): ¹H NMR (400 MHz, CDCl₃) δ 7.32-7.27 (m, 1H), 7.20-7.17 (m, 1H), 7.13 (s, 1H), 5.42 (s, 1H), 2.71-2.53 (m, 1H), 1.55 (ddd, *J* = 12.9, 8.5, 6.5 Hz, 1H), 1.29 (dq, *J* = 14.6, 7.4 Hz, 1H), 0.84 (t, *J* = 7.4 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ

148.82, 134.87, 129.08, 128.66, 127.99, 120.60, 54.09, 31.47, 25.32, 22.31, 13.80.



(1-benzyl-1H-1,2,3-triazol-4-yl)(4-methoxyphenyl)methanone (**3q**): ¹**H NMR** (400 MHz, CDCl₃) δ 8.50 (d, J = 9.0 Hz, 2H), 8.14 (s, 1H), 7.44-7.38 (m, 3H), 7.35-7.31 (m, 2H), 6.99 (d, J = 9.0 Hz, 2H), 5.60 (s, 2H), 3.89 (s, 3H). ¹³**C NMR** (101 MHz,

CDCl₃) δ 184.04, 163.95, 148.94, 133.88, 133.25, 129.46, 129.28, 128.51, 128.25, 113.81, 55.64, 54.58.

3. ¹H and ¹³C NMR Spectra of the 1,4-dissubstituted 1,2,3-triazoles



Figure 1. ¹H and ¹³C NMR spectra in CDCl₃ of compound **3a**.



Figure 2. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3c.



Figure 3. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3d.

0



Figure 4. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3e.



Figure 5. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3f.



Figure 6. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3g.



Figure 7. ¹H and ¹³C NMR spectra in CDCl₃ of compound **3h**.



Figure 8. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3i.



Figure 9. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3j.



Figure 10. ¹H and ¹³C NMR spectra in CDCl₃ of compound **3**k.



Figure 11. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3l.



Figure 12. ¹H and ¹³C NMR spectra in CDCl₃ of compound **3m**.



Figure 13. ¹H and ¹³C NMR spectra in CDCl₃ of compound 3n.



Figure 14. ¹H and ¹³C NMR spectra in CDCl₃ of compound 30.



Figure 15. ¹H and ¹³C NMR spectra in CDCl₃ of compound **3q**.