

Comparison of Molecularly Imprinted Plasmonic Nanosensors Performances for Bacteriophage Detection

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Supplementary file

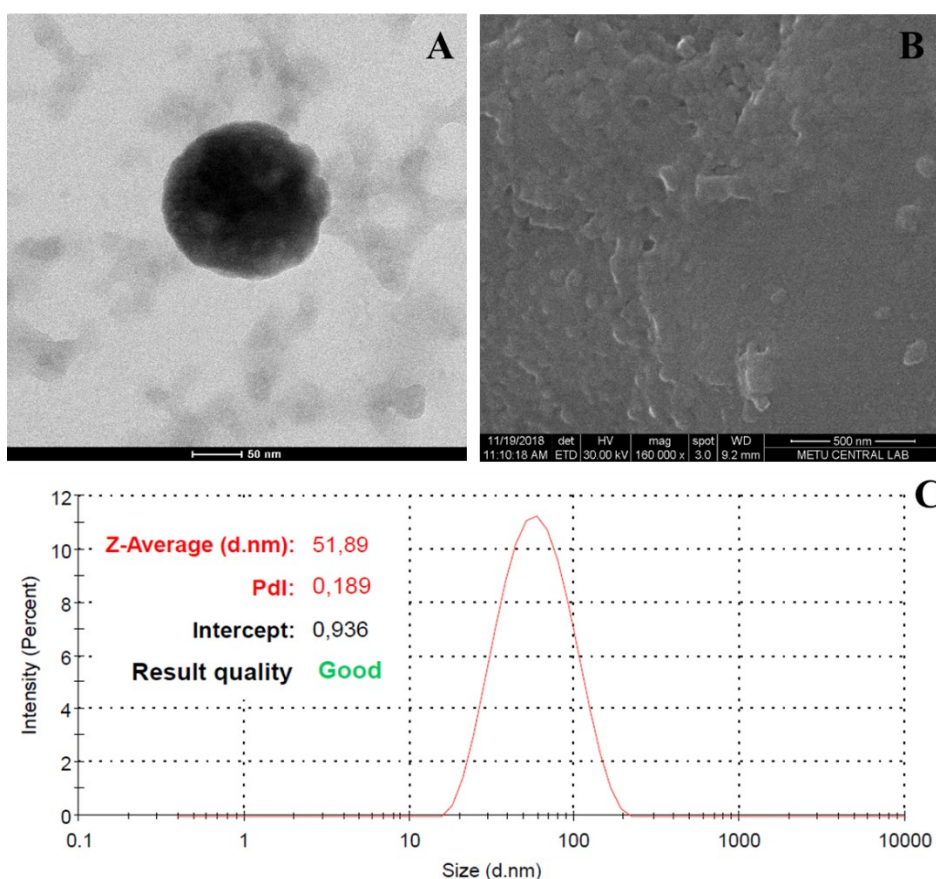


Fig. S1 TEM (A), SEM images (B), and size distribution (C) of non-imprinted nanoparticles.

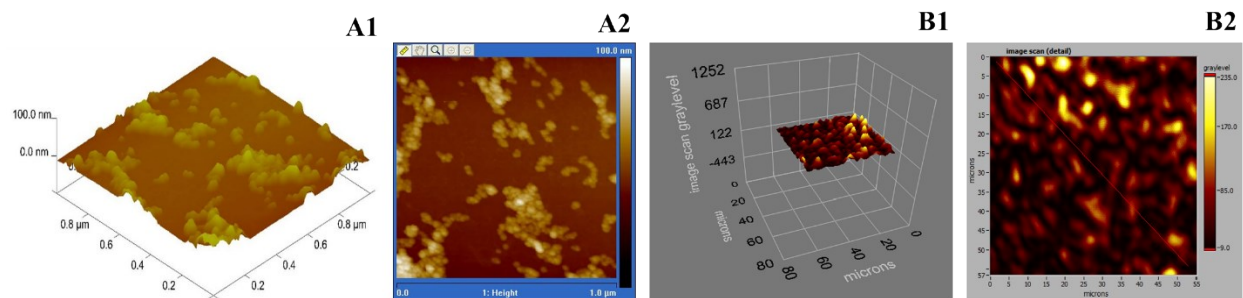


Fig. S2 Three- and two-dimensional AFM (A) and ellipsometry (B) images of non-imprinted nanoparticle-based plasmonic nanosensor.

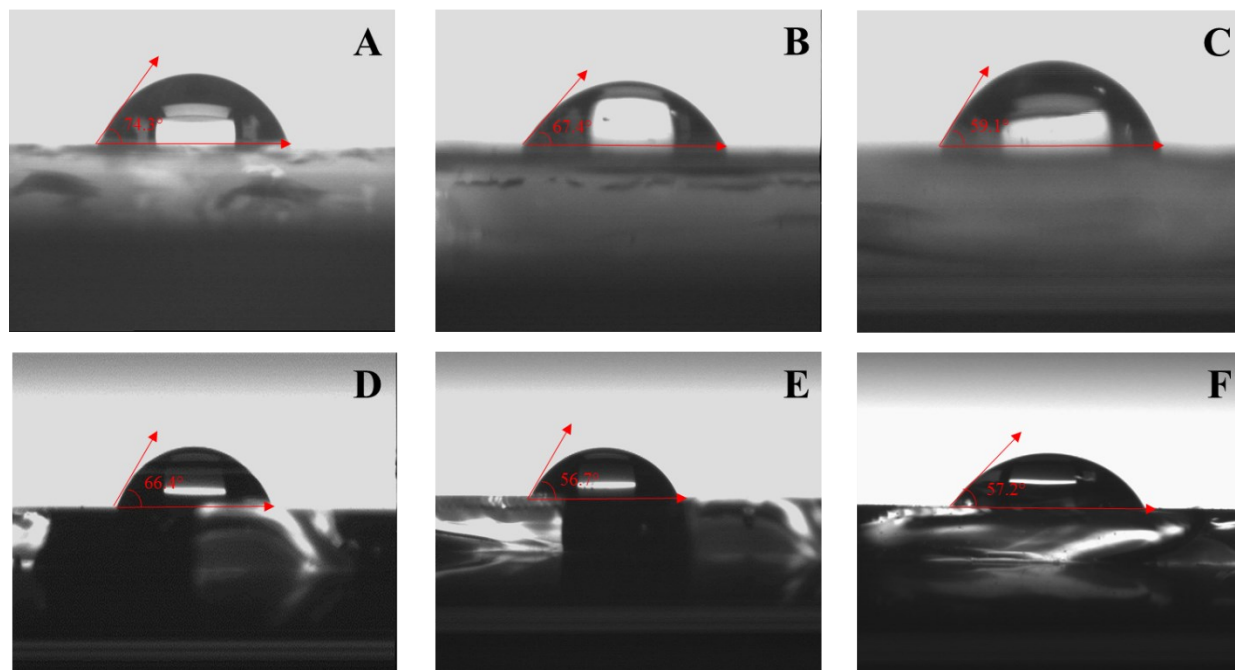


Fig. S3 Contact angle images of nanoparticle- (A: non-modified, B: non-imprinted and C: imprinted) and nanofilm- (D: modified, E: non-imprinted, F: imprinted) based plasmonic nanosensors.

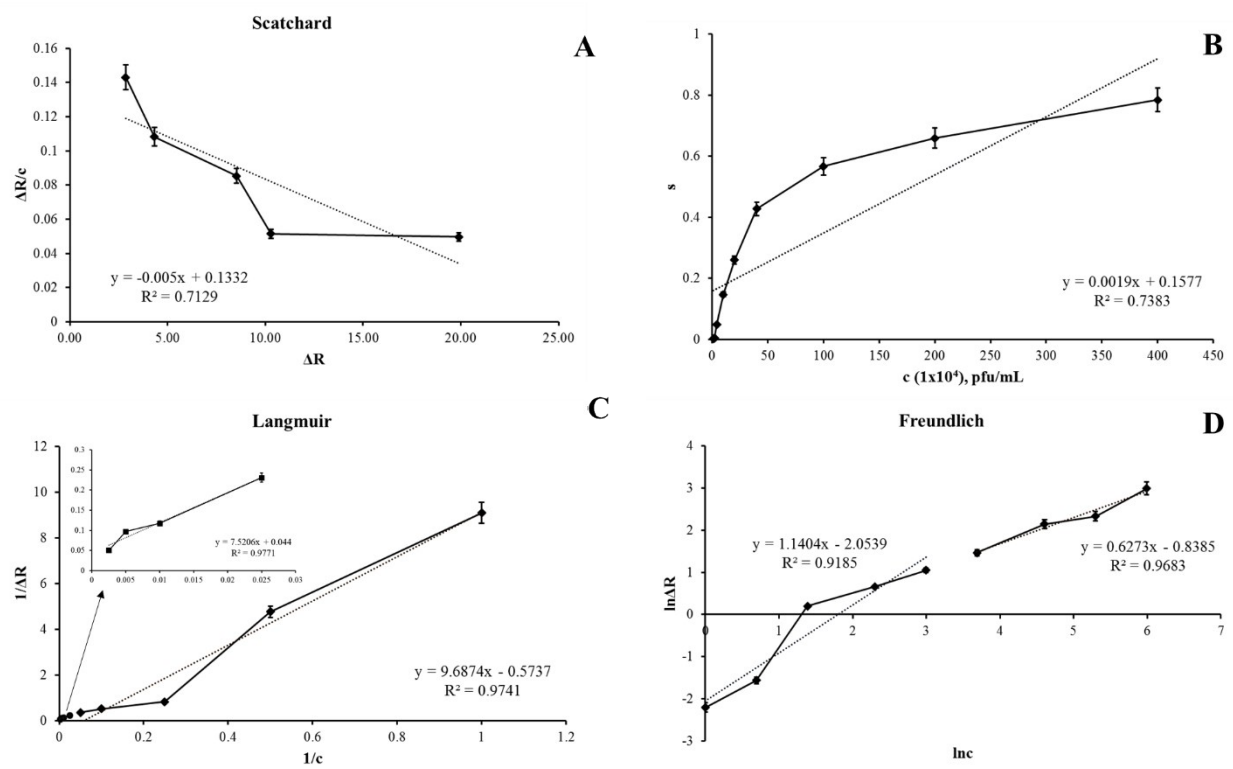


Fig. S4 Adsorption isotherm models for nanoparticle-based plasmonic nanosensor (A: Scatchard; B: Association; C: Langmuir and D: Freundlich).

Table S1. Binding parameters for nanoparticle-based plasmonic nanosensor.

Scatchard		Association	
ΔR_{\max}	26.64	$k_a, \text{pfu/mL}^{-1} \cdot \text{s}^{-1}$	0.002
$K_A, \text{mL/pfu}$	0.005	k_d, s^{-1}	0.158
$K_D, \text{pfu/mL}$	200	$K_A, \text{mL/pfu}$	0.012
R^2	0.71	$K_D, \text{pfu/mL}$	83.0
		R^2	0.74

Table S2. Kinetic parameters for nanoparticle-based plasmonic nanosensor.

Langmuir		Freundlich	
ΔR_{\max}	22.73	ΔR_{\max}	2.31
K_D , pfu/mL	170.9	$1/n$	0.63
K_A , mL/pfu	0.006	R^2	0.92
R^2	0.98		

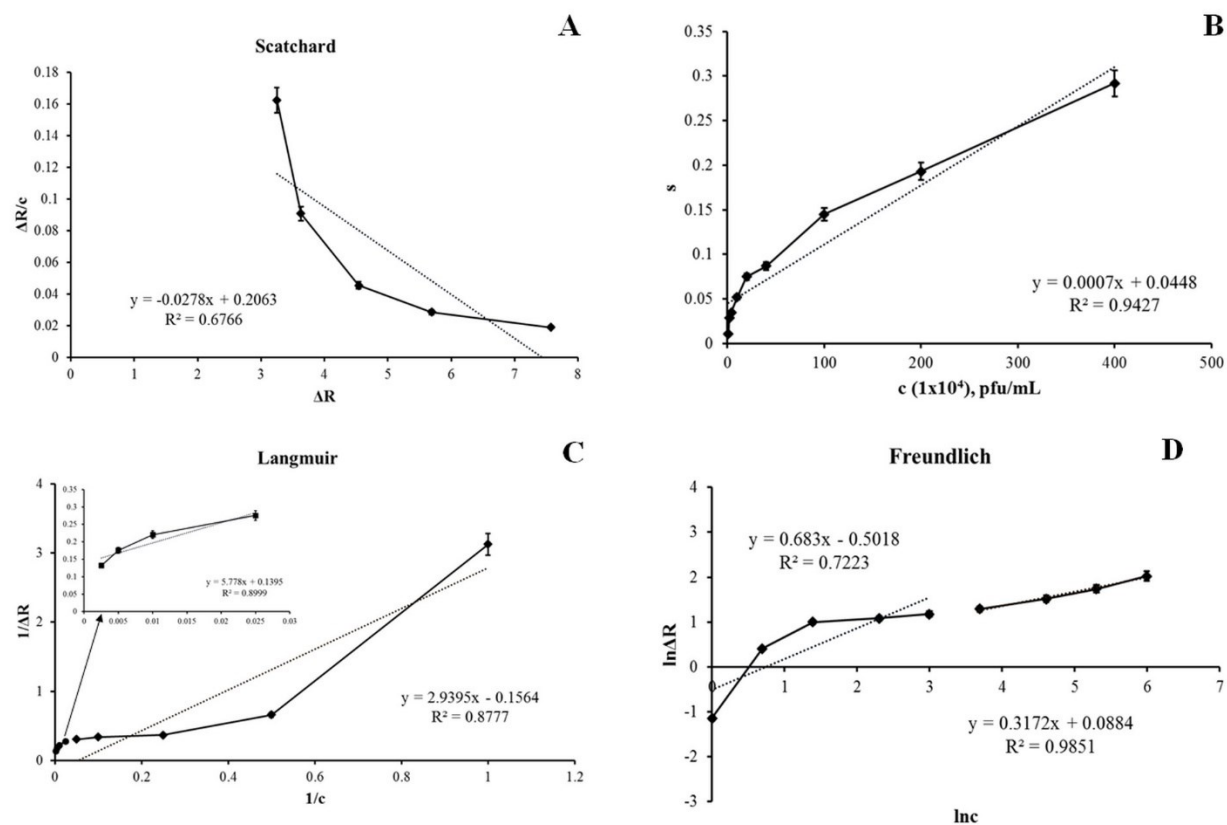


Fig. S5 Adsorption isotherm models for nanofilm-based plasmonic nanosensor (A: Scatchard; B: Association; C: Langmuir and D: Freundlich).

Table S3. Binding parameters for nanofilm-based plasmonic nanosensor.

Scatchard		Association	
ΔR_{\max}	7.42	$k_a, \text{pfu/mL}^{-1} \cdot \text{s}^{-1}$	0.001
$K_A, \text{mL/pfu}$	0.03	k_d, s^{-1}	0.045
$K_D, \text{pfu/mL}$	36.0	$K_A, \text{mL/pfu}$	0.02
R^2	0.68	$K_D, \text{pfu/mL}$	64.0
		R^2	0.94

Table S4. Kinetic parameters for nanoparticle-based plasmonic nanosensor.

Langmuir		Freundlich	
ΔR_{\max}	7.17	ΔR_{\max}	1.09
$K_D, \text{pfu/mL}$	41.4	$1/n$	0.32
$K_A, \text{mL/pfu}$	0.03	R^2	0.72
R^2	0.90		

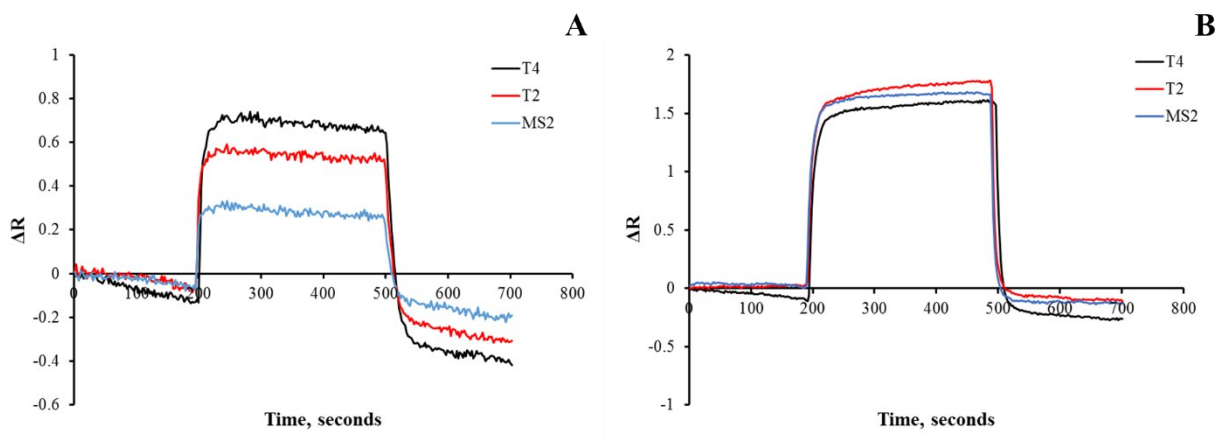


Fig. S6 Selectivity studies of non-imprinted nanoparticle- (A) and nanofilm- (B) based plasmonic nanosensors.

Table S5. Selectivity and relative selectivity of nanoparticle- (A) and nanofilm- (B) based plasmonic nanosensors.

A	Imprinted	k	Non-imprinted	k	k'
T4	19.89		0.68		
T2	3.82	5.21	0.53	1.28	4.07
MS2	3.61	5.51	0.26	2.62	2.11
B					
T4	7.57		1.61		
T2	2.51	3.02	1.77	0.91	3.32
MS2	1.88	4.03	1.67	0.96	4.21

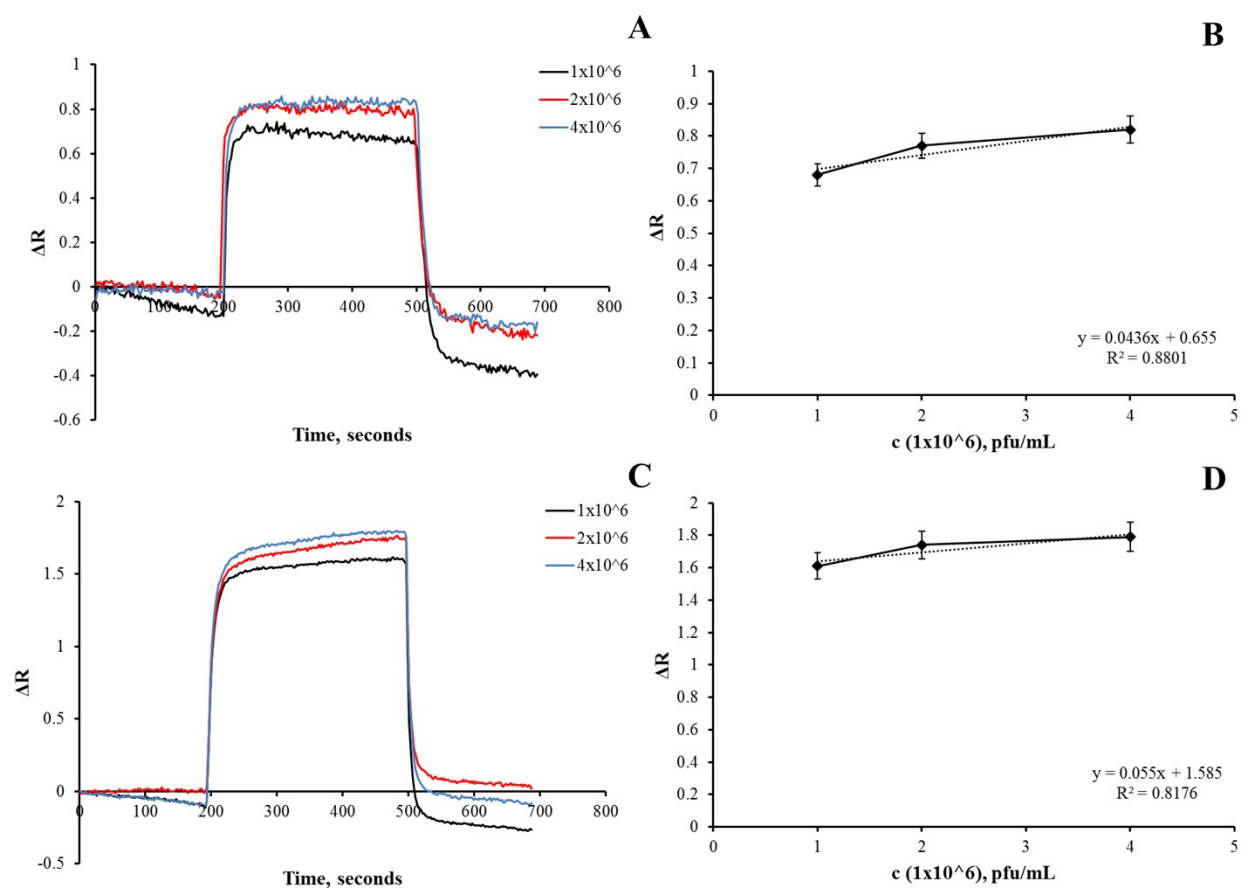


Fig. S7 Real-time bacteriophage detection and the corresponding relationship between the change of reflectivity and bacteriophage concentration for non-imprinted nanoparticle- (A, B) and nanofilm- (C, D) based plasmonic nanosensors.

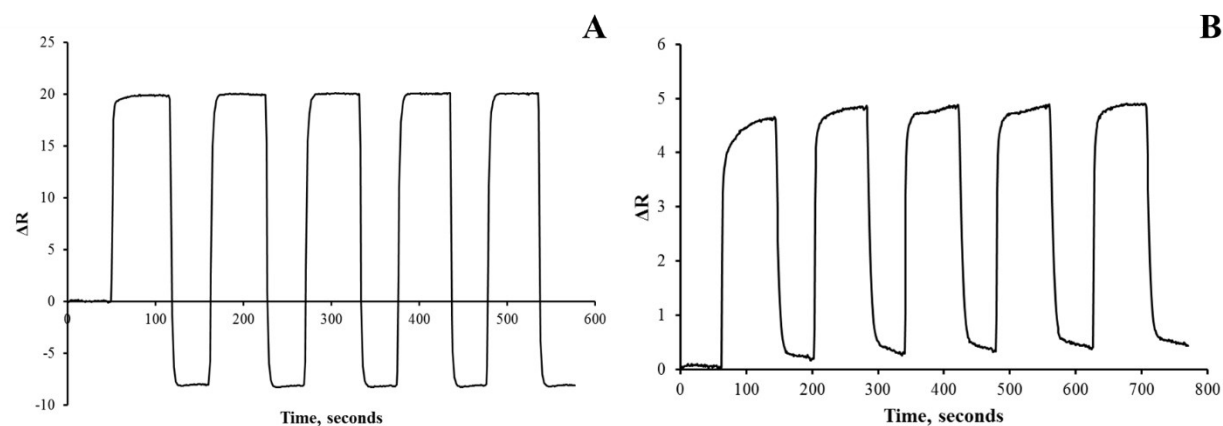


Fig. S8 Reusability studies of nanoparticle- (A) and nanofilm- (B) based plasmonic nanosensors.

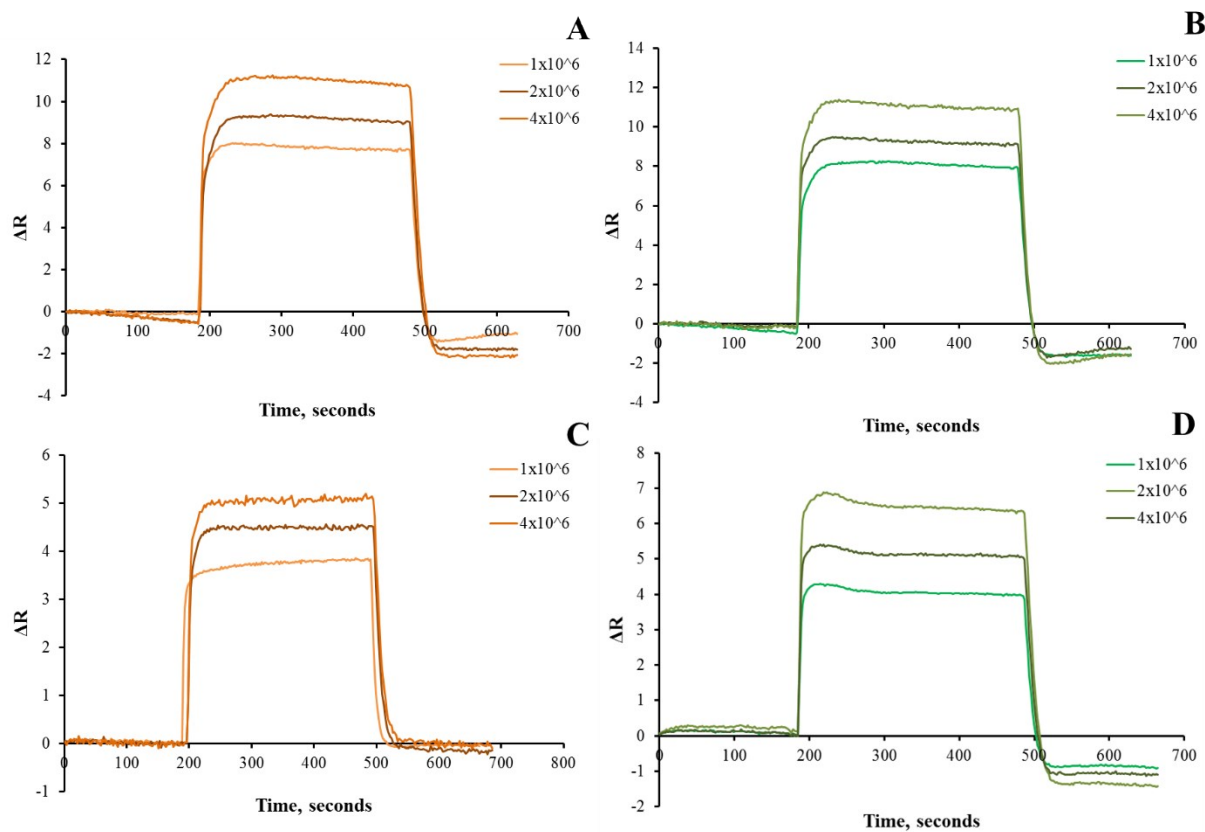


Fig. S9 Water (tap and sea) sample analysis of nanoparticle- (A, B) and nanofilm- (C, D) based plasmonic nanosensors in different bacteriophage concentration.