1	Tunable nanogels by host-guest interaction with carboxylate
2	pillar[5]arene for controlled encapsulation and release of doxorubicin
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2 Figure S1 ¹H-NMR (D₂O, 300 MHz) spectra of A) H1 (3.5 mg mL⁻¹), B) NG (5 mg mL⁻¹) + 1

 $3 \qquad \text{eq. H1 and C) NG (5 mg mL^{-1}).}$



Figure S2 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) at pH = 5.1 after A) 0 h, B)
24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.



Figure S3 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) + 0.05 eq. H1 at pH = 5.1
after A) 0 h, B) 24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.



Figure S4 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) + 0.1 eq. H1 at pH = 5.1 after
A) 0 h, B) 24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.



Figure S5 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) + 0.2 eq. H1 at pH = 5.1 after
A) 0 h, B) 24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.



Figure S6 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) + 0.5 eq. H1 at pH = 5.1 after
A) 0 h, B) 24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.



Figure S7 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) + 1 eq. H1 at pH = 5.1 after
A) 0 h, B) 24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.



Figure S8 ¹H-NMR (D₂O, 300 MHz) spectra of NG (5 mg mL⁻¹) + 5 eq. H2 at pH = 5.1 after
A) 0 h, B) 24 h, C) 48 h, D) 72 h, E) 96 h, F) 168 h and G) 264 h.





Figure S9 Zeta potential of the respective nanogels at different ratios of H1 to pyridinium unit
of the nanogel; all measurements were performed in phosphate buffer (pH 5.1, 10 mM).









Figure S11 Calibration curve of doxorubicin hydrochloride in phosphate buffer (10 mM, pH = 7.4).



Figure S12 Calibration curve of doxorubicin hydrochloride in acetate buffer (10 mM, pH = 5.1).



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Figure S13 Release of DOX from nanogels with varying amounts of H1 at pH values of 7.4
and 5.1.



5 Figure S14 Release of DOX from nanogels (DLC = 16 wt%) with 1 eq. H1 at pH values of 7.4

6 and 5.1.





Figure S16: Cell viability of L929 mouse fibroblasts after incubation with unloaded nanogels
(500 µg mL⁻¹, DOX loaded nanogels (500 µg polymer and 7.3 µg DOX mL⁻¹) and free DOX
(7.3 µg DOX mL⁻¹) for 4, 24, and 48 h respectively. Data represent mean values ± SD of 6replicates per measured sample (Mann-Whitney test, *p≤0.5;**p≤0.01;***p≤0.001).

The graph shows a clear effect of free DOX on the cell viability. DOX loaded nanogels also
show cytotoxic effect but less than free DOX. This can be due to the difference in the uptake
mechanism of the free DOX and DOX loaded nanogel in the cells.





Figure S17: Cell viability of L929 mouse fibroblasts after incubation with unloaded nanogels
(100 μg mL⁻¹), DOX loaded nanogels (100 μg polymer and 1.4 μg DOX mL⁻¹) and free DOX

4 $(1.4 \ \mu g \ DOX \ mL^{-1})$ for 4, 24, and 48h respectively. Data represent mean values $\pm \ SD$ of 6-fold

5 measured samples (Mann-Whitney test, $p \le 0.5$; $p \le 0.01$; $p \le 0.001$).

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Figure S18: Cell viability of L929 mouse fibroblasts after incubation with unloaded nanogels
(50 μg mL⁻¹), DOX loaded nanogels (50 μg polymer and 0.7 μg DOX mL⁻¹) and free DOX (0.7
μg DOX mL⁻¹) for 4, 24, and 48 h respectively. Data represent mean values ± SD of 6-fold

11 measured samples (Mann-Whitney test, $p \le 0.5$; $p \le 0.01$; $p \le 0.001$).



Figure S19: CLSM images of adherent L929 cells after 24 h incubation at 37 °C with free DOX
at a concentration of 14.7 μg mL⁻¹. Transmitted light (A) and cell nuclei (B), free DOX in the
cells(C) and overlay of all channels (D). The fluorescence signal is correlated with the applied
stain specifically for the nuclei. Overlay of all channels proves (D) an intracellular localization
of the free DOX.