

Supplementary Information

Facile formation of chiral nanofibers with excellent electrochemical performance via Self-assembly of carbon dots and cysteine molecules

Jiaqi Zhao, Li Li, Fei Li, Huali Liu, Zhen Li, and Yu Wang*

State Key Laboratory of Inorganic Synthesis and Preparative Chemistry, College of Chemistry, Jilin University, Changchun, 130012, China.

* Corresponding authors: wangyu@jlu.edu.cn

Keywords: Chirality; self-assembly; carbon dots; chiroptical response; lithium-oxygen batteries.

Table of Contents

Methods and materials	3
Fig. S1 The TEM and HRTEM images and size distribution image of pristine CDs.	4
Fig. S2 The Raman spectrum of pristine CDs.	5
Fig. S3 The XPS spectrum of pristine CDs.	6
Fig. S4 The FT-IR spectrum of pristine CDs.	7
Fig. S5 The UV-vis spectra of pristine CDs aqueous solution.	8
Fig. S6 The SEM images of L-CDs/cysteine nanofibers at different temperature.	9
Fig. S7 The SEM images of D-CDs/cysteine nanofibers	10
Fig. S8 XPS survey of L-CDs/cysteine nanofibers deconvoluted C _{1s} and S _{2p} spectra	11
Fig. S9 XPS survey of L- cysteine deconvoluted S _{2p} spectra	12
Fig. S10 The CD spectra of the aqueous solution of left-handed CDs@cysteine at different temperatures.	13
Fig. S11 The CD spectra of CDs interacts with other chiral amino acids at different temperatures.	14
Fig. S12 The Emission spectra of the aqueous solution.	15
Fig. S13 UV-vis spectra of CDs and L(D)-CDs/cysteine nanofibers aqueous solution.	16
Fig. S14 The CIE chromaticity diagram.	17
Fig. S15 The Fluorescence decay curves and fitting results of pristine CDs and L-CD/cysteine nanofibers at different temperature.	18
Table S1 The photoluminescence QYs of CDs and L-CDs/cysteine nanofibers.	19
Table S2 The Fluorescence decay value of pristine CDs and L-CD/cysteine nanofibers	20
Table S3 The zeta potentials of CDs and L-CDs/cysteine nanofibers.	21
Supplementary References	22

Methods and Materials

1. Materials

All chemicals were used as received without further purification. Dimethylformamide (DMF), citric acid and urea were purchased from Beijing Chemical Works. L-cysteine (98%), and D-cysteine (98%) were purchased from Shanghai Huishi biochemical Co., Ltd. NaOH were purchased from Aladdin.

2. Synthesis of pristine CDs

The CDs were synthesized via a solvothermal route. Typically, 1 g citric acid and 2 g urea were dissolved in 10 ml DMF. Then the solution was sealed in a Teflon-lined stainless steel autoclave and heated at 180 °C for 24 h under static conditions and cooled to room temperature. Subsequently, the obtained brown solution was mixed with NaOH aqueous solution (50 mg mL⁻¹), and centrifuged. Then precipitate was washed twice with water and dissolved in 10 ml Ultra-pure water.

3. Synthesis of left-handed and right-handed CDs/cysteine

As-synthesized pristine CDs solution (2mL) was mixed with 72.6 mg L-cysteine and D-cysteine respectively in 10 ml Ultra-pure water under gentle stirring for 24h at room temperature ,40°C and 60°C, respectively. Then obtained crude product was centrifuged and washed with water. After that L(D)- CDs/cysteine nanofibers were dispersed in 3.5 ml of water for further characterziations.

4. Characterization techniques

The surface morphologies were performed with a JEOL-7800 F field emission scanning electron microscope at an accelerating voltage of 3 kV and JEOL-6700 F field emission scanning electron microscope at an accelerating voltage of 5 kV. The transmission electron micrograph (TEM) and high-resolution transmission electron micrograph (HRTEM) images were recorded on a FEI Tecnai G2S-Twin with a field emission gun operating at 200 kV. The X-ray photoelectron spectra (XPS) was measured by ESCALab 250 Analytical XPS spectrometer with a monochromatic X-ray source (Al K α , $h = 1486.6$ eV). Fourier transformed infrared (FT-IR) spectra were recorded with IFS-66V/S FT-IR spectrometer.

5. Optical studies

CD spectra were obtained on a Bio-Logic MOS-450 CD spectrometer. VCD spectra were measured with a ChiralIR-2X spectrometer. The pellets were prepared by adding 1-2 mg of L(D)-CDs/cysteine, CDs and L/D-cysteine powder to 100 mg of KBr. All VCD spectra were collected for 12 h at a resolution of 4 cm⁻¹. Photoluminescence spectra were recorded at room temperature by using FLUOROMAX-4 spectrometer, the time-resolved PL decay was carried out on Edinburgh Instrument FLS920 spectrophotometer. UV-vis absorption (UV) spectra were performed either by a Shimadzu UV-2450 spectrophotometer.

6. Electrochemical test

The coin cells were galvanostatically discharged/charged by Land CT2001 battery testers (Wuhan Land ElectricCo. Ltd., China)

Supplemental figures

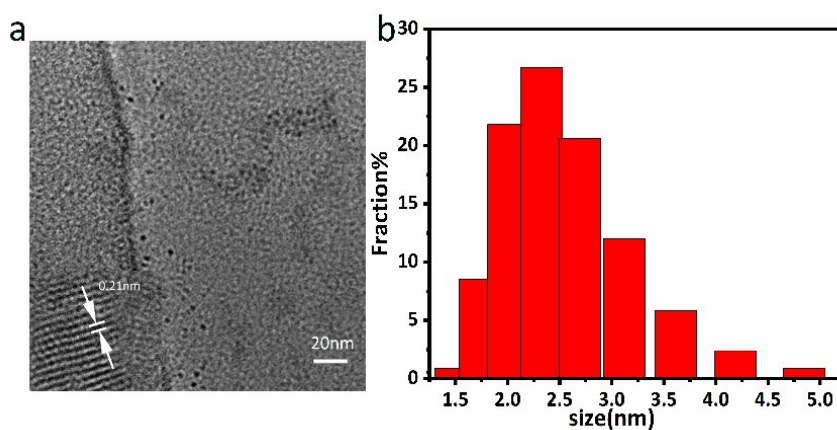


Fig. S1 a) The TEM and HRTEM images and b) size distribution image of pristine CDs. It shows the diameter of pristine CDs with a uniform size distribution around 2 nm. The inset HRTEM image demonstrate the crystallinity of pristine CDs with a lattice parameter of 0.21 nm, which agrees with (100) lattice fringes of graphite. ^[1]

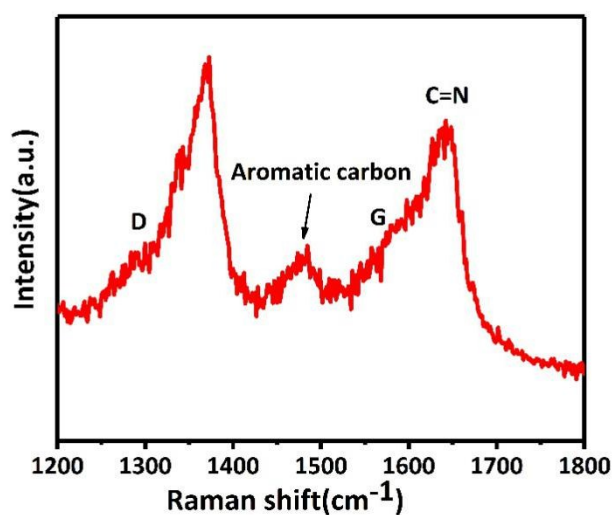


Fig. S2 Raman spectrum of pristine CDs. Common features were observed in the 1200-1800 cm⁻¹ region, where the G band originates from graphitic sp² carbon while D band indicates the existence of disorder structures or defects in the CDs. Except for the D band and G band, strong signals assigned to C=N and aromatic carbon.^[2]

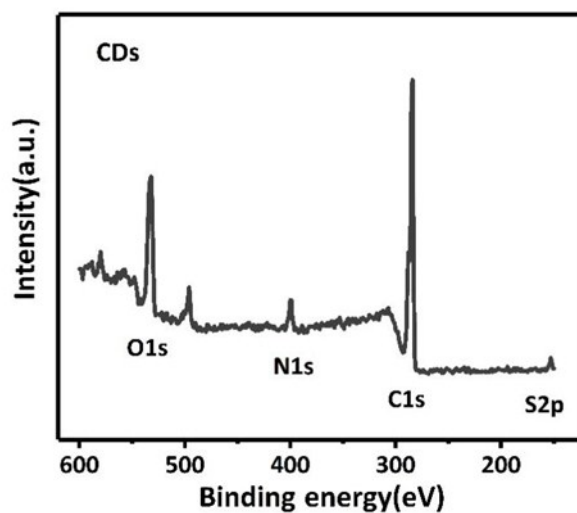


Fig. S3 XPS spectrum of pristine CDs. From the full-scan XPS spectrum of pristine CDs, C, N, O were detected (C1s at 297.2eV, N1s at 411.2eV and O1s at 543.2 eV).

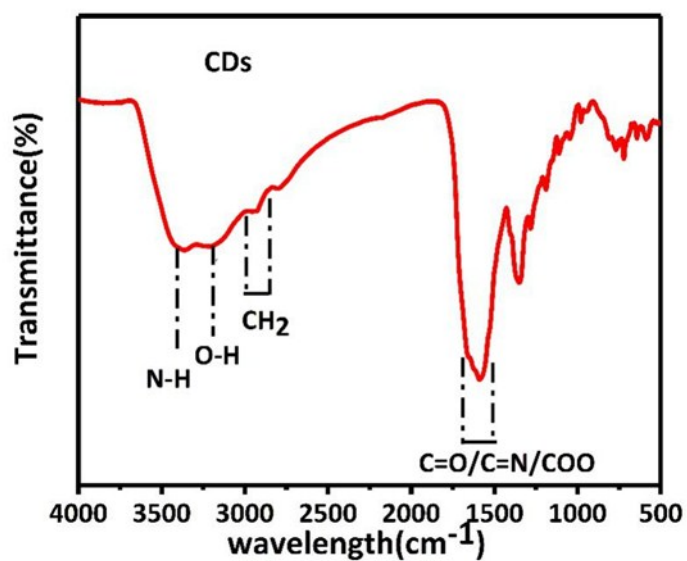


Fig. S4 FT-IR spectrum of pristine CDs. In FT-IR spectra, broad absorption bands at 3050–3450 cm^{-1} are assigned to ν (O-H) and ν (N-H). The strong absorption band from 1600–1700 cm^{-1} assigned to ν_{as} (COO) in carboxylate groups, ν (C=O) in carboxyl groups and C=N in CNH group.^[3-11]

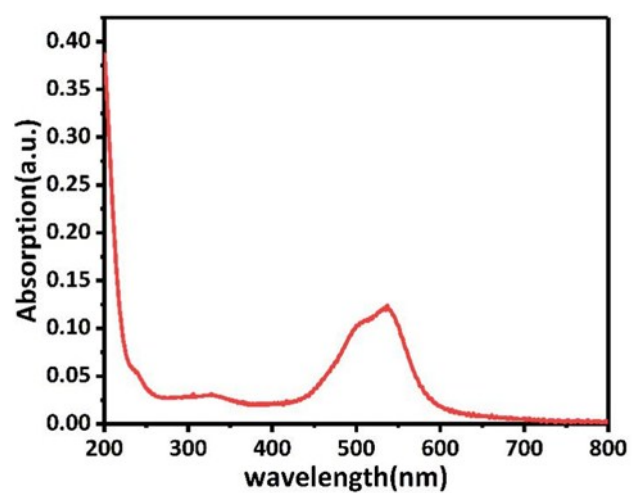


Fig. S5 UV-vis spectra of pristine CDs aqueous solution. The absorption at 260 nm and 350 nm are the intrinsic absorption peak of carbon cores which are assigned to the $\pi \rightarrow \pi^*$ transition and the $n \rightarrow \pi^*$ transitions, respectively. The obvious visible absorption band peaked at 542 nm, indicating large sized conjugated sp^2 -domain in the particles. ^[12]

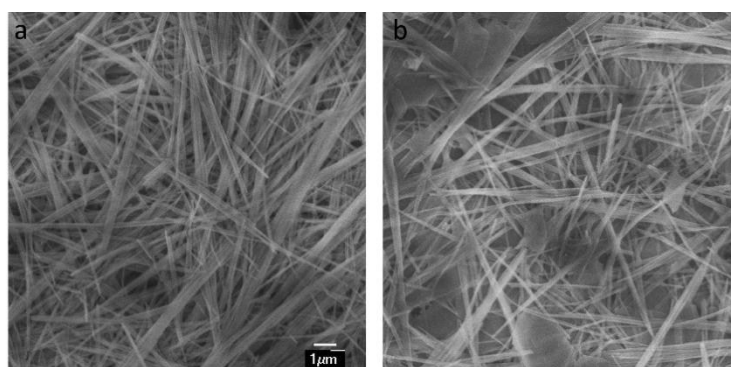


Fig. S6 SEM images of L-CDs/cysteine nanofibers at a) room temperature and b) 60°C

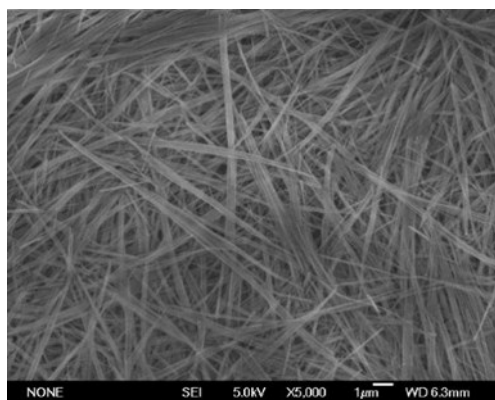


Fig. S7 SEM images of D-CDs/cysteine nanofibers at 40°C

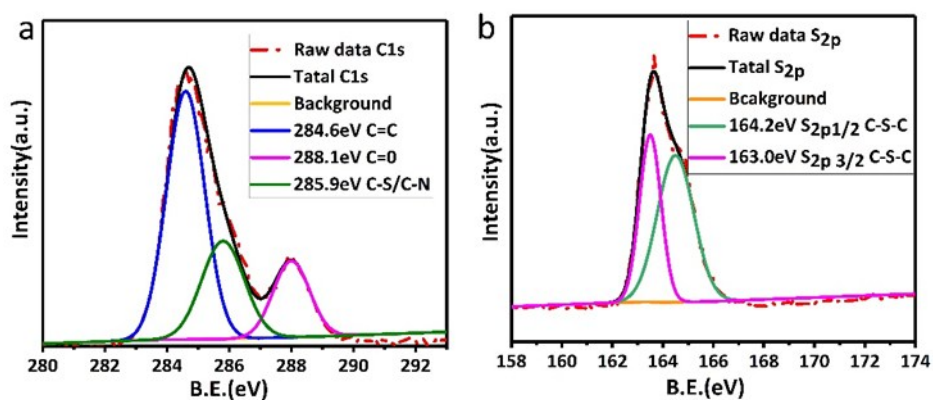


Fig. S8 XPS survey of L-CDs/cysteine nanofibers a) deconvoluted C_{1s} spectra and b) deconvoluted S_{2p} spectra

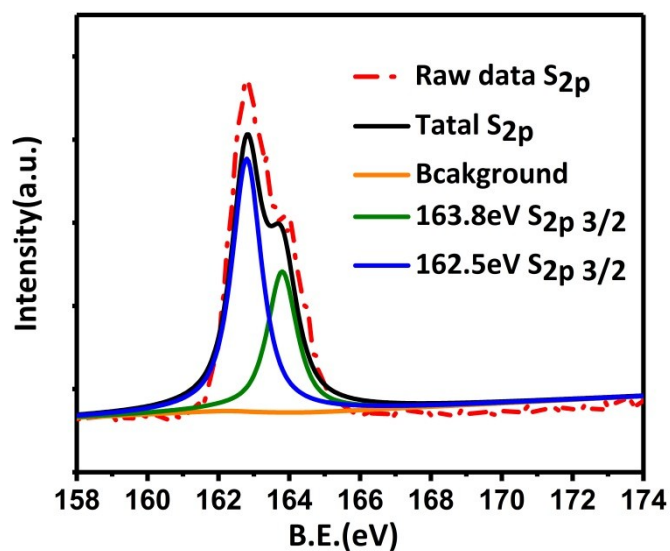


Fig. S9 XPS survey of L- cysteine deconvoluted S_{2p} spectra. The peak deconvolution of the L-cysteine shows the presence of two S chemical states: the first one giving rise to an $S_{2p_{3/2}}$ signal at 162.8 eV and the second one producing a $S_{2p_{3/2}}$ peak at 163.9 eV, which is similar to the previous report.^[13]

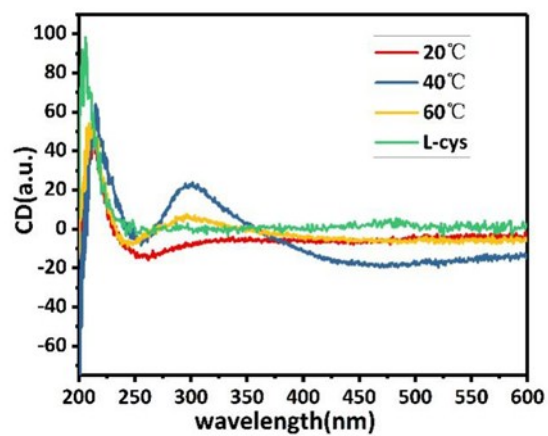


Fig. S10 CD spectra of the aqueous solution of left-handed CDs@cysteine at different temperatures.

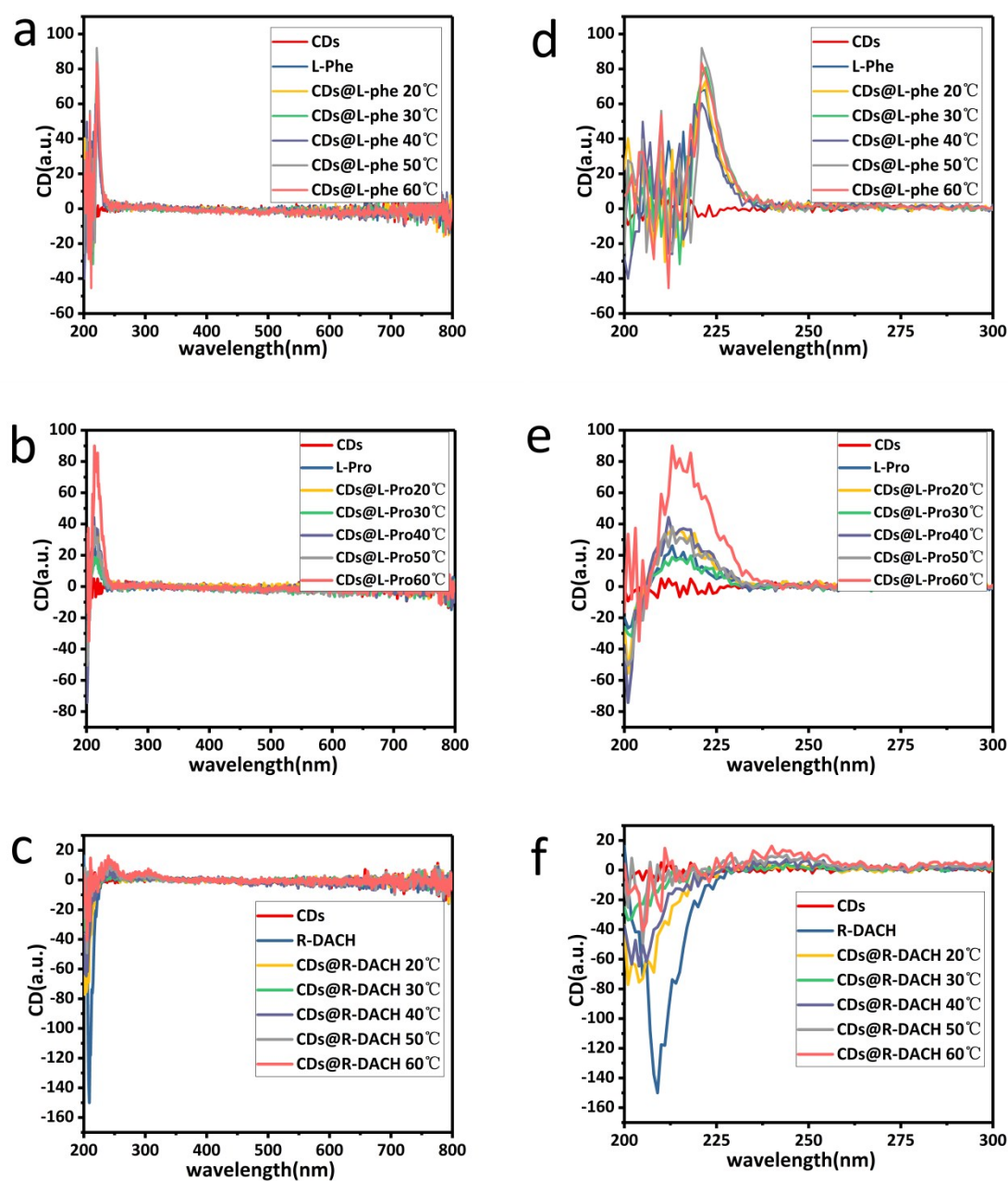


Fig. S11 CD spectra of a) CDs@ L-phenylalanine (L-phe), b) CDs@ L-proline (L-pro) and c) CDs@ (R,R)- 1,2-diaminocyclohexane (R-DACH) d-f) the CD image of the sample a-c)X-axis reduction at different temperatures, which shows that CDs and other chiral amino acids cannot produce similar chiral optical properties.

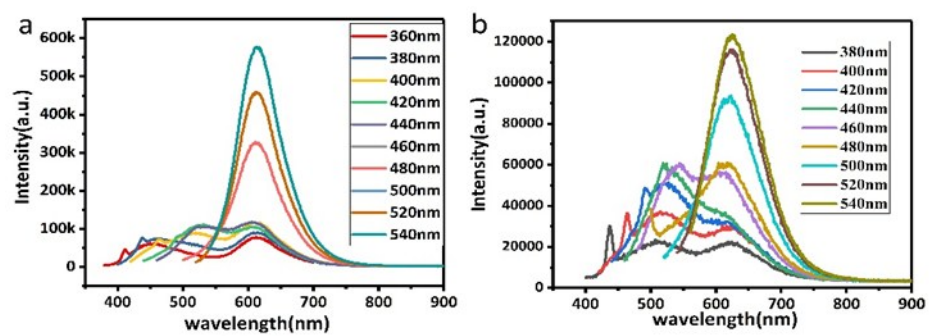


Fig. S12 Emission spectra of the aqueous solution of a) pristine CDs after it has been present in aqueous solution for a while. b) left-handed CDs/cysteine nanofibers excited at different wavelengths.

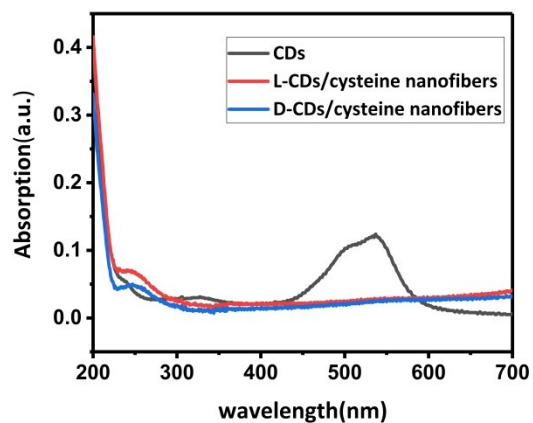


Fig. S13 UV-vis spectra of CDs and L(D)-CDs/cysteine nanofibers aqueous solution.

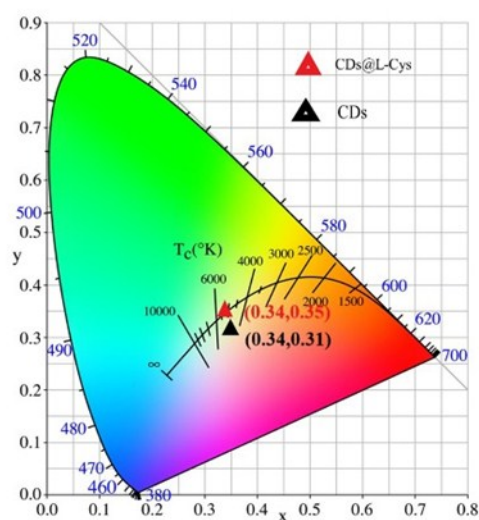


Fig. S14 The CIE chromaticity diagram showing the coordinates of the samples presented in Fig S11. a) and b). Under excitation at 380 nm, L-CDs/cysteine nanofibers exhibits a white-light emission with CIE chromaticity coordinates of (0.34, 0.35) and a correlated color temperature (CCT) of 5300 K, and thus the emission of our L-CDs/cysteine nanofibers are called warm white-light. This warm white-light-emission is ideal applications for indoor light sources.

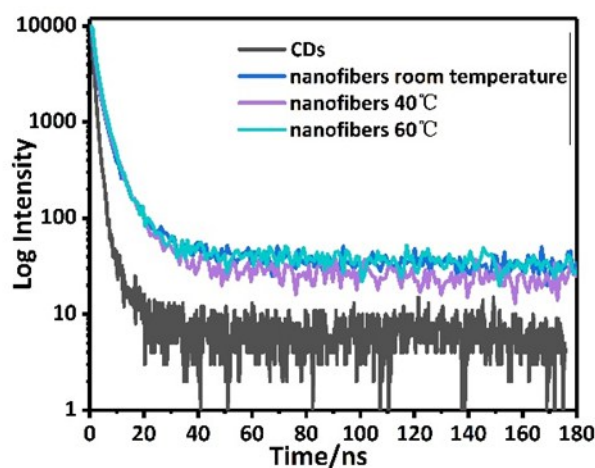


Fig. S15 Fluorescence decay curves and fitting results of pristine CDs and L-CDs/cysteine nanofibers at different temperature recorded at emission wavelengths of 460nm in water. The PL decay of CDs and L-CDs/cysteine nanofibers curves can be well fitted into a multiexponential function with two and three lifetimes respectively. The proportion of each lifetime is shown in Table S1. The average lifetime pristine CDs and CDs@L-cysteine at room temperature, 40, 60 ° C calculated using the following Equation (1)

$$\tau_{av} = (\sum \alpha_i \tau_i^2) / (\sum \alpha_i \tau_i) \quad (1)$$

is 2.189ns, 337.3ns, 359.7ns and 294.1ns respectively. The multiple lifetimes of the phosphorescence imply various emission species, which may be a wide range of chemical environments on the surface of carbon dots for aromatic carbonyls.

Table S1. The photoluminescence QYs of CDs and L-CDs/cysteine nanofibers at different temperature aqueous solution under excitation of 420 nm.

Sample	CDs	20 °C	40 °C	60 °C
QYs	6.19%	1.33%	1.23%	0.04%

Table S2. Fluorescence decay value of pristine CDs and CDs@ L-cysteine nanofibers

Sample	τ_1	A ₁ %	τ_2	A ₂ %	τ_3	A ₃ %	τ_{av}
CDs	1.178ns	93.9	5.515ns	6.10			2.189ns
CDs@L-cysteine at 20 °C	1.414 ns	45.69	5.704ns	30.68	347.0ns	23.63	337.3ns
CDs@L-cysteine at 40 °C	1.320 ns	45.92	5.431ns	30.68	373.1ns	23.63	359.7ns
CDs@L-cysteine at 60 °C	1.461 ns	43.89	4.916ns	36.53	305.9ns	19.58	294.1ns

Table S3. The zeta potentials of CDs and CDs@L-cysteine nanofibers at different temperature aqueous solution.

Sample	CDs	room temperature	40°C	60°C
Zeta potential	-11.8mV	9.77mV	-5.97mV	-3.33mV

Supplementary References

- [1] H. F. Liu, Z. H. Li, Y. Q. Sun, X. Geng, Y. L. Hu, H.M. Meng, J. Ge and L. B. Qu, *Scientific Reports*, 2018, **8**, 1086.
- [2] M.J. Matthews, M.A. Pimenta, G. Dresselhaus, M.S. Dresselhaus, M. Endo, *Physical Review B*, 1999, **59**, R6585-R6588.
- [3] C. J. Reckmeier, J. Schneider, A. S. Sussha and A. L. Rogach, *Opt. Express*, 2016, **24**, A312;
- [4] D. Chen, W. Wu, Y. Yuan, Y. Zhou, Z. Wan and P. Huang, *J. Mater. Chem. C*, 2016, **4**, 9027
- [5] Y. Chen, M. Zheng, Y. Xiao, H. Dong, H. Zhang, J. Zhuang, H. Hu, B. Lei and Y. Liu, *Adv. Mater*, 2016, **28**, 312;
- [6] S. Zhu, J. Shao, Y. Song, X. Zhao, J. Du, L. Wang, H. Wang, K. Zhang, J. Zhang and B. Yang, *Nanoscale*, 2015, **7**, 7927.
- [7] Y. Wang, S. Kalytchuk, L. Wang, O. Zhovtiuk, K. Cepe, R. Zboril and A. L. Rogach, *Chem. Commun*, 2015, **51**, 2950.
- [8] S. Sun, L. Zhang, K. Jiang, A. Wu and H. Lin, *Chem. Mater*, 2016, **28**, 8659.
- [9] H. Li, X. He, Z. Kang, H. Huang, Y. Liu, J. Liu, S. Lian, C. A. Tsang, X. Yang and S. T. Lee, *Angew. Chem. Int. Ed*, 2010, **49**, 4430.
- [10] C. J. Reckmeier, J. Schneider, A. S. Sussha and A. L. Rogach, *Opt. Express*, 2016, **24**, A312.
- [11] D. Chen, W. Wu, Y. Yuan, Y. Zhou, Z. Wan and P. Huang, *J. Mater. Chem. C*, 2016, **4**, 9027.
- [12] C. J. Reckmeier, J. Schneider, A. S. Sussha and A. L. Rogach, *Opt. Express*, 2016, **24**, A312.
- [13] O. Cavalleri, L. Oliveri, A. Dacca, R. Parodi and R. Rolandi, *Applied Surface Science*, 2001, **175**, 357