Supporting Information

Modulating the Dynamics of Förster Resonance Energy Transfer and Singlet Fission by Variable Molecular Spacers

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General Procedures, Materials, and Methods

Synthesis and characterization

Synthesis and characterization of Pnc₂ derivatives 2a-d. All reagents and solvents were obtained from commercial suppliers and used as received unless otherwise stated. Anhydrous solvents were obtained from a solvent purification system. Purification by column chromatography was carried out on silica gel (SiO₂, 60 Å, 40–63 µm). Thin-layer chromatography (TLC) was carried out using commercially available aluminum sheets precoated with silica gel and a fluorescence indicator with visualization under UV light at 254 or 360 nm. Brine refers to a saturated, aqueous solution of NaCl. ¹H and ¹³C-NMR spectra were recorded on a Varian/Agilent instrument equipped with a cryoprobe at 500 MHz and 126 MHz, respectively. Chemical shift values are quoted in ppm and coupling constants (J) in Hz. ¹H and ¹³C-NMR spectra, respectively) are referenced against the residual solvent peak (CDCl₃ 7.26 ppm, 77.16 ppm; CD₂Cl₂ 5.32 ppm, 53.84 ppm; THF d_8 1.72 ppm, 25.31 ppm). UV-Vis spectroscopic measurements were done on a Cary 400 scan spectrophotometer using a 1 cm path-length quartz cuvette, and the neat solvent was used as baseline; sh = shoulder absorption. IR spectra were recorded on a Thermo Nicolet 8700 FTIR spectrometer and samples were measured as solids or cast films. High-resolution mass spectrometry (HRMS) spectra were recorded on a Bruker 9.4T Apex-Qe FTICR instrument (MALDI), an Agilent Technologies 6220 oaTOF instrument (APPI) or a Kratos MS50G instrument (EI). Differential scanning calorimetry (DSC) measurements were made on a Perkin Elmer Pyris 1 DSC or a Mettler Toledo Polymer DSC. Thermogravimetric analyses (TGA) were made on a Perkin Elmer Pyris 1 TGA or a Mettler Toledo TGA/DSC 1. Thermal analyses were carried out under a flow of nitrogen with a heating rate of 10 °C/min. Thermal decomposition temperature as measured by TGA (as sample weight loss) is reported as T_d in which the temperature listed corresponds to the intersection of the baseline and the tangent lines of the edge of the peak corresponding to the first significant weight loss, typically >5%. Melting points from DSC analysis are reported as the peak maxima, except in cases when the sample decomposed, in which case the onset temperature of the decomposition exothermic peak is reported, as well as the exothermic maxima corresponding to the decomposition. Melting points were measured on a Thomas Hoover capillary melting point apparatus.

Synthesis and characterization of SubPc-Pnc₂ derivatives 1a-d. All reagents and solvents were purchased from commercial suppliers and used without further purification unless otherwise stated. The synthesis and characterization of SubPcCl **9** has been reported elsewhere.¹ Anhydrous solvents were obtained from a solvent purification system. Celite plugs were performed using Celite® 545. Size exclusion chromatography was carried out on Bio-Beads[™] S-X1 Support (200–400 mesh). NMR spectra were

recorded with a BRUKER Avance-300 (300 MHz) and a BRUKER DRX-500 (500 MHz) spectrometer. Chemical shifts (δ) are measured in ppm and coupling constants (*J*) in Hz. ¹H-NMR and ¹³C-NMR spectra, respectively, are referenced against the residual solvent peak (CDCI₃ 7.26 ppm, 77.16 ppm; THF-*d*₈ 1.73 ppm, 67.57 ppm). All the ¹³C-NMR spectra were acquired during 18 hours. High-resolution mass spectrometry (HRMS) spectra were recorded employing Matrix Assisted Laser Desorption/Ionization-Time of Flight (MALDI-TOF) using a BRUKER ULTRAFLEX III instrument equipped with a Nd-YAG laser. A convenient matrix for these measurements is indicated for each compound.

Photophysical Studies

Steady-State Absorption and Emission. A Lambda 2 spectrometer from Perkin Elmer was utilized for all the steady-state UV-Vis absorption spectra at room temperature (rt). A FluoroMax3 emission spectrometer from Horiba was used for the steady-state fluorescence measurements in the visible detection range, again at rt. Fluorescence quantum yields were determined by means of gradient analyses using (1) for the SubPc fluorescence: F_{12} SubPc in toluene (axially substituted with a phenoxy-ligand, $\Phi_F = 0.17)^2$ as well as Rhodamine 6G in ethanol ($\Phi_F = 0.95$)³ and (2) for the Pnc₂ fluorescence: Zinc phthalocyanine in toluene ($\Phi_F = 0.30$)⁴ as reference compounds. All measurements were performed at rt in 10 × 10 mm quartz cuvettes.

Time-Correlated Single Photon Counting (TCSPC). Fluorescence lifetimes were determined by the time-correlated single photon counting technique using a FluoroLog3 emission spectrometer (Horiba) equipped with an R3809U-58 MCP (Hamamatsu) and a SuperK Extreme high-power supercontinuum fiber laser EXB-6 (NKT) exciting at 532 and 632 nm (~150 ps pulse width).

Femtosecond (fsTA) and Nanosecond Transient Absorption (nsTA). Femtosecond (0 to 5500 ps) and nanosecond (1 ns to 400 μ s) transient absorption (fsTA and nsTA) experiments were carried out using transient absorption pump/probe detection systems (HELIOS and EOS, Ultrafast Systems). A 1050 Hz Ti:sapphire output (Clark-MXR, CPA-2101, 775 nm fundamental, ~150 fs pulse width) was applied, from which a fraction is focused onto a sapphire disk (fsTA, 2 mm), generating the white light of the probe beam (~420-800 nm). For nsTA, the white light (~370-900 nm) was generated instead by a photonic crystal fiber supercontinuum laser source (1064 nm fundamental, 2100 kHz, ~1 ns pulse width). Measurements were performed in 2 × 10 mm cuvettes and argon-purged solutions (~10 minutes) at rt. The pump pulses at 530 and 632 nm were generated with a non-collinear optical parametric amplifier (NOPA; Clark-MXR). The

repetition rate of the pump pulses was set to 525 Hz for fsTA (by means of a chopper) and to 1050 Hz for nsTA. Bandpass filters were used to ensure low spectral width of the pump pulses. The spot diameter of the pump beam was always kept larger than that of the probe beam in order to homogeneously excite the complete probed area. The delay of the probe pulses with respect to the pump pulses was reached with a controlled delay stage (fsTA) or electronically (nsTA). A beam splitter was used to split the probe pulses into two (signal and reference beam) before passing the sample. The probe and reference beam were detected independently to correct for fluctuations in the probe beam intensity. The transient spectra were acquired shot-by-shot and averaged at each delay until a reasonable signal-to-noise ratio was reached (~1500 times).

Data Analysis. Data evaluation of the fsTA and nsTA data has been conducted by means of global analysis and fitting of the transient absorption data were performed with the help of the open-source software package GloTarAn.⁵ The latter is a Java-based graphical user interface to the R-package TIMP.^{6,7} The latter was developed for global and target analysis of time-resolved spectroscopy data. The dispersion (chirp of the white-light pulse) of the instrument response function (IRF) was modeled during the fitting procedure.

FRET Rate Constant Determination. For the calculations, the following **Equations S1-S5** were used:

$$R = R_0 \left(\frac{1-E}{E}\right)^{\frac{1}{6}}$$

R = FRET distance; R_0 = critical energy donor-acceptor distance; E = FRET efficiency.

$$E = 1 - \frac{\tau_{DA}}{\tau_D}$$

Equation S2

Equation S1

E = FRET efficiency; τ_{DA} = experimental lifetime (TA) of energy donor-acceptor conjugate; τ_D = experimental fluorescence lifetime of energy donor (TCSPC).

$$R_0 = 0.2108 (\kappa^2 \Phi_F n^{-4} J)^{\frac{1}{6}}$$
 Equation S3

 R_0 = critical energy donor-acceptor distance; κ^2 = orientation factor; ϕ_F = fluorescence quantum yield; n = refractive index of the solvent; J = degree of spectral overlap.

$$J = \int_{0}^{\infty} F_{D}(\lambda)\varepsilon_{A}(\lambda)\lambda^{4}d\lambda$$
 Equation S4

J = degree of spectral overlap; F_D = fluorescence intensity of the donor at wavelength λ . The fluorescence spectra must be normalized to fulfill **Equation S5**; ε_A = extinction coefficient of the acceptor at wavelength λ .

$$\int_{0}^{\infty} F_{D}(\lambda) d\lambda = 1$$

Equation S5

The detailed procedure (including a description of a software-based algorithm to calculate the spectral overlap integral J is described by Hink et al.⁸ For κ^2 we used the values obtained from our computational studies (**Table S3**). The refractive index n was taken as 1.496 for toluene.⁹ We calculated the FRET distances R for conjugates **1a-d**, that is, the distances between SubPc and the Pnc₂ in **1a-d** by means of **Equation S1** through **Equation S5** based on the experimental lifetimes obtained for τ_{DA} (fsTA) and τ_D (TCSPC).¹⁰ See also **Table 2** for a summary of the parameters used for the FRET distance calculation as well as for an overview of the obtained values for R.¹¹

Synthesis and Characterization

Synthesis and characterization of Pnc₂ derivatives 2a-d.

Compounds $\mathbf{8}$, ¹² $\mathbf{7}$, ¹³ $\mathbf{6b}$, ¹⁴ and $\mathbf{5a}^{15}$ were synthesized as reported.



Scheme S1. Synthesis of aryl precursors 5a-d.dibromide





Compound 6c. To a N₂-purged solution of 1,3,5-tribromobenzene (11.41 g, 36.25) mmol), 4-hydroxylphenylboronic acid (2.00 g, 14.5 mmol) and K₂CO₃ (3.01 g, 21.8 mmol) in toluene (120 mL) and EtOH (40 mL) was added Pd(PPh₃)₄ (168 mg, 0.145 mmol). The reaction mixture was heated to reflux and stirred for 12 h. After cooling to room temperature, the reaction mixture was poured into sat. aq. NH₄Cl (150 mL), the phases were separated, and the aqueous phase extracted with CH₂Cl₂ (2 x 50 mL). The combined organic phases were washed with H₂O (200 mL) and brine (150 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 4:1) afforded 6c (3.40 g, 71%) as an off-white crystalline solid. $R_f = 0.20$ (EtOAc/CH₂Cl₂ 1:19). Mp = 150–152 °C. IR (cast film CH₂Cl₂): 3315 (br s), 3106 (w), 3067 (w), 3026 (w), 1882 (w), 1756 (w), 1692 (w), 1610 (m), 1595 (m), 1584 (s), 1544 (m), 1520 (s) cm⁻¹. ¹**H-NMR** (500 MHz, CDCl₃) δ 7.60 (d, J = 1.6 Hz, 2H), 7.59 (t, J = 1.6 Hz, 1H), 7.42 (d, J = 8.6 Hz, 2H), 6.91 (d, J = 8.6 Hz, 2H), 4.79 (s, 1H). ¹³C-NMR (126 MHz, CDCl₃) δ 156.1, 144.5, 132.1, 131.3, 128.7, 128.6, 123.4, 116.1. EI MS m/z 327.9 (M⁺, 100), 168.1 ([M - ⁷⁹Br⁸¹Br]⁺, 49). EI HRMS m/z calcd for C₁₂H₈O⁷⁹Br⁸¹Br (M⁺) 327.8922, found 327.8928.



Compound 6d. To a solution of **7**¹³ (1.00 g, 2.94 mmol) in CH₂Cl₂ (15 mL) and ethanol (15 mL) at rt was added NaBH₄ (111 mg, 2.94 mmol) in several portions. After stirring for 2 h, the reaction mixture was poured into sat. aq. NH₄Cl (100 mL) and extracted twice with CH₂Cl₂ (100 and 50 mL). The combined organic phases were washed with H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The remaining solid was suspended in hexane and collected by vacuum filtration, washed with hexanes (3 x 5 mL), and dried *in vacuo* to afford **6d** (851 mg, 84%) as a white solid. *R*_f = 0.13 (CH₂Cl₂). **Mp** = 128–130 °C. **IR** (solid): 3257 (br, s), 3064 (m), 3027 (m), 2922 (m), 2889 (m), 2869 (m), 1909 (w), 1770 (w), 1586 (s), 1545 (s), 1516 (m) cm⁻¹. ¹H-NMR (500 MHz, CDCl₃) δ 7.65 (d, *J* = 1.7 Hz, 2H), 7.64 (t, *J* = 1.7 Hz, 1H), 7.53 (d, *J* = 8.3 Hz, 2H), 7.45 (d, *J* = 8.3 Hz, 2H), 4.76 (d, *J* = 5.7 Hz, 2H), 1.69 (t, *J* = 5.7 Hz, 1H). ¹³C-NMR (126 MHz, CDCl₃) δ 144.6, 141.4, 137.8, 132.8, 129.1, 127.7, 127.4, 123.5, 65.0. **APPI HRMS** *m/z* calcd for C₁₃H₁₀⁷⁹Br₂O⁺ (M⁺) 339.9093, found 341.9077.



Compound 5b. A solution of **6b**¹⁴ (2.00 g, 7.52 mmol) in acetic anhydride (15 mL) was heated to 80 °C. After stirring for 3 h, the reaction mixture was cooled to rt, poured onto ice (ca. 150 mL), and neutralized by addition of K₂CO₃ until the effervescences ceased. The aqueous suspension was extracted twice with CH₂Cl₂ (100 and 50 mL), and the combined organic phases were washed with H₂O (150 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 4:1) afforded **5b** (2.07 g, 89%) as a colorless oil. **R**_f = 0.84 (CH₂Cl₂/hexanes 7:3). **IR** (neat): 3073 (w), 2955 (w), 2895 (w), 1745 (s), 1590 (m), 1559 (s) cm⁻¹. ¹H-NMR (500 MHz, CDCl₃) δ 7.62 (t, *J* = 1.7 Hz, 1H), 7.43 (d, *J* = 1.7 Hz, 2H), 5.03 (s, 2H), 2.13 (s, 3H). ¹³C-NMR (126 MHz, CDCl₃) δ 170.6, 140.0, 134.0, 129.8, 123.2, 64.6, 21.0. **EI MS** *m*/*z* 307.9 (M⁺, 29), 265.9 ([M – C₂H₂O]⁺, 100), 248.9 ([M – AcO]⁺, 19). **EI HRMS** *m*/*z* calcd for C₉H₈O₂⁷⁹Br⁸¹Br (M⁺) 307.8871, found 307.8871.



Compound 5c. A solution of **6c** (3.18 g, 9.70 mmol) in acetic anhydride (15 mL) was heated to 80 °C. After stirring for 3 h, the reaction mixture was cooled to rt, poured onto ice (ca. 150 mL), and neutralized by addition of K₂CO₃ until the effervescences ceased. The aqueous suspension was extracted twice with CH₂Cl₂ (150 and 50 mL), and the combined organic phases were washed with H₂O (150 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 3:2) afforded **5c** (3.27 g, 91%) as a white solid. **R**_f = 0.84 (CH₂Cl₂/hexanes 7:3). **Mp** = 98 °C. **IR** (cast film CH₂Cl₂): 3070 (w), 1760 (s), 1606 (w), 1584 (s), 1547 (s), 1511 (s) cm⁻¹. ¹**H-NMR** (500 MHz, CDCl₃) δ 7.64 (t, *J* = 1.7 Hz, 1H), 7.62 (d, *J* = 1.7 Hz, 2H), 7.53 (d, *J* = 8.7 Hz, 2H), 7.18 (d, *J* = 8.7 Hz, 2H), 2.33 (s, 3H). ¹³**C-NMR** (126 MHz, CDCl₃) δ 169.5, 151.1, 144.0, 136.2, 132.9, 129.1, 128.4, 123.5, 122.4, 21.3. **EI MS** *m*/*z* 369.9 (M⁺, 7), 327.9 ([M – C₂H₂O]⁺, 100). **EI HRMS** *m*/*z* calcd for C₁₄H₁₀O₂⁷⁹Br⁸¹Br (M⁺) 369.9029, found 369.9027.



Compound 5d. A solution of **6d** (820 mg, 2.40 mmol) in acetic anhydride (10 mL) was heated to 80 °C. After stirring for 4 h, the reaction mixture was cooled to rt, poured onto ice (ca. 150 mL), and neutralized by addition of K₂CO₃ until the effervescences ceased. The aqueous suspension was extracted twice with CH₂Cl₂ (100 and 50 mL), and the combined organic phases were washed with H₂O (150 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 4:1) afforded **5d** (885 mg, 96%) as a white solid. *R*_f = 0.51 (CH₂Cl₂/hexanes 4:1). **Mp** = 52 °C. **IR** (solid): 3088 (m), 3070 (m), 3033 (w), 2963 (m), 2874 (w), 1743 (s), 1613 (m), 1585 (s), 1551 (m), 1522 (m) cm⁻¹. ¹**H-NMR** (500 MHz, CDCl₃) δ 7.64 (s, 3H), 7.52 (d, *J* = 8.2 Hz, 2H), 7.44 (d, *J* = 8.2 Hz, 2H), 5.15 (s, 2H), 2.13 (s, 3H). ¹³**C-NMR** (126 MHz, CDCl₃) δ 171.0, 144.4, 138.4, 136.5, 132.9, 129.1, 129.0, 127.5, 123.5, 65.9, 21.2. **EI MS** *m*/*z* 383.9 (M⁺, 52), 341.9 ([M – C₂H₂O]⁺, 100), 324.9 ([M – AcO]⁺, 64). **EI HRMS** *m*/*z* calcd for C₁₅H₁₂O₂⁷⁹Br⁸¹Br (M⁺) 383.9184, found 383.9184.



Compound 4a. To a N₂-purged solution of **5a**¹⁵ (294 mg, 1.00 mmol) and **8**¹² (1.20 g, 2.20 mmol) in THF (50 mL) and Et₃N (20 mL) was added Pd(PPh₃)₄ (173 mg, 0.150 mmol) and Cul (19 mg, 0.10 mmol). The reaction mixture was heated to reflux and stirred for 18 h. After cooling to room temperature, the reaction mixture was poured into sat. aq. NH₄Cl (50 mL), H₂O (50 mL) was added, and the phases were separated. The aqueous phase was extracted with CH₂Cl₂ (50 mL) and the combined organic phases were washed with H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 85:15) afforded **4a** (856 mg, 70%) as a white solid. **R**_f = 0.20 (CH₂Cl₂/hexanes 85:15). **Mp**: decomposition, 218 °C (turns into dark green liquid accompanied by evolution of gas). **IR** (solid): 3055 (w), 2942 (s), 2891 (m), 2865 (s),

2817 (w), 2164 (w), 1772 (s), 1598 (m), 1578 (m) cm⁻¹. ¹H-NMR (500 MHz, CD₂Cl₂) δ 8.68 (s, 4H), 8.41 (s, 4H), 8.01–7.96 (m, 4H), 7.96–7.90 (m, 4H), 7.61–7.52 (m, 8H), 7.20 (t, *J* = 1.4 Hz, 1H), 6.96 (d, *J* = 1.4 Hz, 2H), 3.08 (s, 6H), 2.99 (s, 6H), 2.15 (s, 3H), 1.40–1.13 (m, 42H). ¹³C-NMR (126 MHz, CD₂Cl₂) δ 169.2, 150.8, 134.5, 134.0, 133.9, 133.4, 132.3, 128.8, 128.72, 128.68, 127.5, 127.4, 127.3, 125.3, 124.6, 106.1, 93.3, 92.4, 84.8, 76.5, 74.1, 52.7, 52.3, 21.2, 19.2, 12.0. MALDI HRMS (DCTB): *m/z* calcd for C₈₂H₈₄O₆Si₂⁺ (M⁺) 1220.5801, found 1220.5794.



Compound 4b. To a N₂-purged solution of **5b** (262 mg, 0.850 mmol) and $\mathbf{8}^{12}$ (1.00 g, 1.84 mmol) in THF (40 mL) and Et₃N (15 mL) was added Pd(PPh₃)₄ (145 mg, 0.125 mmol) and Cul (16 mg, 0.083 mmol). The reaction mixture was heated to reflux and stirred for 10 h. After cooling to room temperature, the reaction mixture was poured into sat. aq. NH₄Cl (50 mL), H₂O (50 mL) was added and the phases were separated. The aqueous phase was extracted with CH_2CI_2 (50 mL) and the combined organic phases were washed with H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated in vacuo. Purification by flash column chromatography (SiO₂, CH₂Cl₂) afforded **4b** (565 mg, 70%) as an off-white solid. $R_{f} = 0.25$ (CH₂Cl₂). **Mp** = 190–194 °C. IR (cast film CH₂Cl₂): 3055 (m), 2942 (s), 2891 (m), 2865 (s), 2817 (w), 2167 (w), 1745 (s), 1593 (m) cm⁻¹. ¹**H-NMR** (500 MHz, CD₂Cl₂) δ 8.69 (s, 4H), 8.41 (s, 4H), 8.01–7.96 (m, 4H), 7.96–7.91 (m, 4H), 7.62–7.51 (m, 8H), 7.28 (s, 1H), 7.22 (s, 2H), 4.87 (s, 2H), 3.07 (s, 6H), 3.00 (s, 6H), 1.97 (s, 3H), 1.44–1.05 (m, 42H). ¹³C-NMR (126 MHz, CD₂Cl₂) δ 170.9, 137.3, 134.6, 134.4, 134.0, 133.9, 133.4, 131.3, 128.8, 128.7, 128.7, 127.5, 127.4, 127.3, 123.7, 106.1, 92.9, 92.4, 85.3, 76.5, 74.1, 65.4, 52.6, 52.3, 21.2, 19.2, 12.0. MALDI HRMS (DCTB) m/z calcd for C₈₃H₈₆O₆Si₂ (M⁺) 1234.5957, found 1234.5943.



Compound 4c. To a N₂-purged solution of **5c** (340 mg, 0.918 mmol) and $\mathbf{8}^{12}$ (1.10 g, 2.02 mmol) in THF (50 mL) and Et₃N (20 mL) was added Pd(PPh₃)₄ (159 mg, 0.138 mmol) and Cul (17 mg, 0.092 mmol). The reaction mixture was heated to reflux and stirred for 12 h. After cooling to room temperature, the reaction mixture was poured into sat. aq. NH₄Cl (50 mL), H₂O (50 mL) was added, and the phases were separated. The aqueous phase was extracted with CH₂Cl₂ (50 mL), and the combined organic phases were washed with H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated in vacuo. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 9:1) afforded 4c (474 mg, 40%) as a pale green solid. $R_f = 0.35$ (CH₂Cl₂/hexanes 9:1). Mp: decomposition, 230 °C (color change to green). IR (cast film CH₂Cl₂): 3054 (m), 2945 (s), 2893 (m), 2867 (s), 2818 (w), 2162 (w), 1771 (s), 1595 (m), 1504 (m) cm⁻¹. ¹**H-NMR** (500 MHz, CD₂Cl₂) δ 8.69 (s, 4H), 8.44 (s, 4H), 8.02–7.97 (m, 4H), 7.97–7.92 (m, 4H), 7.60–7.53 (m, 8H), 7.46 (d, J = 1.5 Hz, 2H), 7.44 (d, J = 8.6 Hz, 2H), 7.32 (t, J = 1.5 Hz, 1H), 7.07 (d, J = 8.6 Hz, 2H), 3.09 (s, 6H), 3.01 (s, 6H), 2.25 (s, 3H), 1.41–1.06 (m, 42H). ¹³C-NMR (126 MHz, CD₂Cl₂) δ 169.8, 151.3, 141.1, 137.5, 134.7, 134.1, 133.9, 133.7, 133.4, 130.5, 128.8, 128.72, 128.68, 128.5, 127.5, 127.4, 127.3, 124.0, 122.6, 106.2, 92.7, 92.3, 85.6, 76.5, 74.2, 52.6, 52.3, 21.5, 19.2, 12.0. **MALDI HRMS** (DCTB): *m*/*z* calcd for C₈₈H₈₈O₆Si₂⁺ (M⁺) 1296.6114, found 1296.6106.



Compound 4d. To a N₂-purged solution of **5d** (352 mg, 0.918 mmol) and 8^{12} (1.10 g, 2.02 mmol) in THF (50 mL) and Et₃N (20 mL) was added Pd(PPh₃)₄ (159 mg, 0.138 mmol) and Cul (17 mg, 0.092 mmol). The reaction mixture was heated to reflux and stirred for 12 h. After cooling to room temperature, the reaction mixture was poured into sat. aq. NH₄Cl (50 mL), H₂O (50 mL) was added and the phases were separated. The aqueous phase was extracted with CH₂Cl₂ (50 mL) and the combined organic phases were washed with H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated in vacuo. Purification by flash column chromatography (SiO₂, CH₂Cl₂/hexanes 9:1) afforded **4d** (567 mg, 47%) as a pale green solid. $R_f = 0.19$ (CH₂Cl₂/hexanes 9:1). Mp = decomposition, 230 °C (color change to green), 245 °C (melts into dark green liquid). IR (cast film CH₂Cl₂): 3055 (m), 2943 (s), 2892 (m), 2865 (s), 2816 (w), 2167 (w), 1741 (s), 1589 (m) cm⁻¹. ¹**H-NMR** (500 MHz, CD₂Cl₂) δ 8.69 (s, 4H), 8.44 (s, 4H), 8.02–7.97 (m, 4H), 7.97–7.92 (m, 4H), 7.60–7.53 (m, 8H), 7.47 (d, J = 1.4 Hz, 2H), 7.44 (d, J = 8.2 Hz, 2H), 7.34 (d, J = 8.2 Hz, 2H), 7.32 (t, J = 1.4 Hz, 1H), 5.06 (s, 2H), 3.09 (s, 6H), 3.01 (s, 6H), 2.06 (s, 3H), 1.38–1.11 (m, 42H). ¹³C-NMR (126 MHz, CD₂Cl₂) δ 171.1, 141.4, 139.6, 136.6, 134.7, 134.1, 133.9, 133.7, 133.4, 130.5, 129.1, 128.8, 128.73, 128.67, 127.6, 127.5, 127.4, 127.3, 124.0, 106.2, 92.7, 92.4, 85.6, 76.5, 74.2, 66.2, 52.6, 52.3, 21.3, 19.2, 12.0. MALDI HRMS (DCTB) m/z calcd for C₈₉H₉₀O₆Si₂ (M⁺) 1310.6271, found 1310.6265.



Compound 3a. To a solution of **4a** (829 mg, 0.679 mmol) in THF (30 mL) and H_2O (15 mL) was added LiOH·H₂O (142 mg, 3.39 mmol) and the reaction mixture was stirred at room temperature for 17 h. The clear solution was poured into sat. ag. NH₄CI (50 mL) and extracted with CH₂Cl₂ (100 mL). The organic phase was washed with sat. NaHCO₃ (2 x 50 mL), H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated in vacuo. The remaining solid was suspended in hexanes (50 mL), collected on a glass frit and washed with additional hexanes (3 x 10 mL) to obtain 3a (744 mg, 93%) as a white solid. $R_f = 0.40$ (EtOAc/CH₂Cl₂ 1:49). Mp: decomposition, 160 °C (color change to grey). IR (solid): 3374 (w, broad), 3056 (w), 2943 (s), 2892 (m), 2865 (s), 2824 (w), 2166 (w), 1600 (m), 1584 (s) cm⁻¹. ¹H-NMR (500 MHz, CD₂Cl₂) δ 8.67 (s, 4H), 8.45 (s, 4H), 8.04–7.96 (m, 4H), 7.96–7.89 (m, 4H), 7.60–7.51 (m, 8H), 6.77-6.73 (m, 1H), 6.55 (s, 2H), 5.41 (s, 1H), 3.08 (s, 6H), 2.98 (s, 6H), 1.45-1.12 (m, 42H). ¹³C-NMR (126 MHz, CD₂Cl₂) δ 155.7, 134.4, 133.9, 133.5, 133.1, 128.6, 128.50, 128.47, 127.3, 127.2, 127.1, 124.1, 118.7, 105.9, 92.2, 92.0, 85.4, 76.4, 73.9, 52.4, 52.1, 18.9, 11.8 (one signal coincident or not observed). MALDI HRMS (DCTB): m/z calcd for C₈₀H₈₂O₅Si₂⁺ (M⁺) 1178.5695, found 1178.5697.



Compound 3b. To a solution of **4b** (550 mg, 0.449 mmol) in THF (15 mL) and H_2O (7.5 mL) was added LiOH·H₂O (113 mg, 2.69 mmol) and the reaction mixture was stirred at room temperature for 18 h. The clear solution was poured into sat. ag. NH₄CI (50 mL) and extracted with CH₂Cl₂ (100 mL). The organic phase was washed with sat. aq. NaHCO₃ (2 x 50 mL), H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by flash column chromatography (SiO₂, EtOAc/CH₂Cl₂ 1:49) afforded **3b** (481 mg, 90%) as a white solid. $R_f = 0.42$ (EtOAc/CH₂Cl₂ 1:49). Mp = decomposition, 160 °C (color change to yellow), 180 °C (melt), 210 °C (turns into dark green liquid accompanied by evolution of gas). IR (cast film CH₂Cl₂): 3468 (br, w), 3055 (m), 2943 (s), 2891 (m), 2865 (s), 2818 (w), 2226 (w), 2167 (w), 1738 (w), 1591 (m) cm⁻¹. ¹H-NMR (500 MHz, CD₂Cl₂) δ 8.68 (s, 4H), 8.42 (s, 4H), 8.00–7.96 (m, 4H), 7.96–7.92 (m, 4H), 7.60–7.52 (m, 8H), 7.27–7.24 (m, 1H), 7.24– 7.22 (m, 2H), 4.48 (d, J = 5.8 Hz, 2H), 3.08 (s, 6H), 3.00 (s, 6H), 1.66 (t, J = 5.8 Hz, 1H), 1.33–1.18 (m, 42H). ¹³C-NMR (126 MHz, CD₂Cl₂) δ 142.3, 134.7, 134.0, 133.9, 133.7, 133.4, 130.1, 128.8, 128.72, 128.66, 127.44, 127.38, 127.3, 123.6, 106.2, 92.5, 92.3, 85.7, 76.5, 74.2, 64.5, 52.6, 52.3, 19.2, 12.0. MALDI HRMS (DCTB) m/z calcd for C₈₁H₈₄O₅Si₂ (M⁺) 1192.5852, found 1192.5863.



Compound 3c. To a solution of **4c** (462 mg, 0.356 mmol) in THF (15 mL) and H₂O (7.5 mL) was added LiOH·H₂O (90 mg, 2.1 mmol) and the reaction mixture was stirred at room temperature for 20 h. The clear solution was poured into sat. aq. NH₄Cl (50 mL)

and extracted with CH₂Cl₂ (100 mL). The organic phase was washed with sat. aq. NaHCO₃ (2 x 50 mL), H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by flash column chromatography (SiO₂, CH₂Cl₂) afforded **3c** (400 mg, 89%) as a white solid. $R_f = 0.29$ (CH₂Cl₂). **Mp** = decomposition, 160 °C (color change to dark green). **IR** (cast film CH₂Cl₂): 3372 (br, m), 3055 (m), 2942 (s), 2892 (m), 2865 (m), 2827 (w), 2229 (w), 2166 (w), 1612 (m), 1586 (m), 1516 (m) cm⁻¹. ¹**H-NMR** (500 MHz, CD₂Cl₂) δ 8.69 (s, 4H), 8.44 (s, 4H), 8.01–7.96 (m, 4H), 7.96–7.92 (m, 4H), 7.59–7.53 (m, 8H), 7.38 (d, *J* = 1.5 Hz, 2H), 7.29–7.24 (m, 3H), 6.75 (d, *J* = 8.7 Hz, 2H), 5.12 (s, 1H), 3.08 (s, 6H), 3.01 (s, 6H), 1.44–1.11 (m, 42H). ¹³**C-NMR** (126 MHz, CD₂Cl₂) δ 156.4, 141.5, 134.7, 134.1, 133.9, 133.4, 133.0, 132.3, 130.1, 128.8, 128.73, 128.72, 128.67, 127.5, 127.4, 127.3, 123.8, 116.2, 106.2, 92.42, 92.35, 85.8, 76.5, 74.2, 52.6, 52.3, 19.2, 12.0. **MALDI HRMS** (DCTB): *m/z* calcd for C₈₆H₈₆O₅Si₂⁺ (M⁺) 1254.6008, found 1254.6015.



Compound 3d. To a solution of **4d** (552 mg, 0.421 mmol) in THF (15 mL) and H₂O (7.5 mL) was added LiOH·H₂O (106 mg, 2.52 mmol) and the reaction mixture was stirred at room temperature for 20 h. The clear solution was poured into sat. aq. NH₄Cl (50 mL) and extracted with CH₂Cl₂ (100 mL). The organic phase was washed with sat. aq. NaHCO₃ (2 x 50 mL), H₂O (100 mL) and brine (100 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by flash column chromatography (SiO₂, EtOAc/CH₂Cl₂ 1:99) afforded **3d** (430 mg, 81%) as a white solid. **R**_f = 0.26 (EtOAc/CH₂Cl₂ 1:99). **Mp** = decomposition, 230 °C (color change to pale green); 250 °C (melts into dark green liquid with evolution of gas). **IR** (cast film CH₂Cl₂): 3499 (br, w), 3055 (m), 2943 (s), 2891 (m), 2865 (s), 2815 (w), 2163 (w), 1591 (m) cm⁻¹. ¹**H-NMR** (500 MHz, CD₂Cl₂) δ 8.69 (s, 4H), 8.44 (s, 4H), 8.01–7.96 (m, 4H), 7.96–7.92 (m, 4H), 7.60–7.52 (m, 8H), 7.48 (d, *J* = 1.5 Hz, 2H), 7.43 (d, *J* = 8.4 Hz, 2H), 7.34 (d, *J* = 8.4 Hz, 2H), 7.31 (t, *J* = 1.5 Hz, 1H), 4.65 (d, *J* = 6.0 Hz, 2H), 3.08 (s, 6H), 3.01 (s, 6H), 1.73 (t, *J* = 6.0 Hz, 1H), 1.39–1.13 (m, 42H). ¹³**C-NMR** (126 MHz, CD₂Cl₂) δ 141.64, 141.58, 138.9, 134.7, 134.1, 133.9, 133.6, 133.4, 130.5, 128.8, 128.72, 128.67, 127.8, 127.54,

127.45, 127.4, 127.3, 124.0, 106.2, 92.6, 92.4, 85.7, 76.5, 74.2, 65.2, 52.6, 52.3, 19.2, 12.0. **MALDI HRMS** (DCTB) m/z calcd for $C_{87}H_{88}O_5Si_2$ (M⁺) 1268.6165, found 1268.6156.



Compound 2a. A solution of 3a (704 mg, 0.60 mmol) in THF (60 mL) was purged with N₂ for 2 min. To this solution was added SnCl₂·2H₂O (539 mg, 2.39 mmol) and 10% H_2SO_4 (0.65 mL) in that order, and the reaction mixture was further purged with N_2 for 2 min. To limit light exposure, the reaction flask was covered in aluminum foil. After stirring for 5 h, the reaction mixture was poured into H₂O/MeOH (1:1, 300 mL), and the precipitate was collected on a glass frit, washed with H₂O/MeOH (1:1, 100 mL) and MeOH (50 mL). The remaining solid was dissolved in THF (20 mL) and stirred for 30 minutes before being poured into hexanes (100 mL). The precipitate was collected on a glass frit and washed with hexanes (100 mL). The solid was dissolved in THF (20 mL) and filtered through a glass frit. The filtrate was concentrated in vacuo, and the dry solid was suspended in MeOH, collected on a glass frit and washed with additional MeOH (100 mL) to obtain 2a (612 mg, 97%) as a green-blue solid. $R_f = 0.58$ (CH₂Cl₂/hexanes 4:1). Mp: decomposition, 200 °C (color change to pale-green). UV-Vis (THF): λ_{max} 310, 348, 366, 414, 439, 519, 558, 602, 656 nm. IR (solid): 3672-3113 (w), 3559 (w), 3049 (m), 2942 (s), 2890 (m), 2864 (s), 2127 (m), 1754 (w), 1704 (w), 1596 (m), 1581 (s) cm⁻¹. ¹**H-NMR** (500 MHz, CD₂Cl₂) δ 9.16 (s, 4H), 9.14 (s, 4H), 8.04–7.90 (m, 9H), 7.49–7.35 (m, 10H), 5.51 (s, 1H), 1.48–1.36 (m, 42H). ¹³**C-NMR** (126 MHz, CD₂Cl₂) δ 156.5, 132.80, 132.79, 130.9, 130.6, 129.2, 129.1, 128.2, 126.9, 126.8, 126.7, 126.2, 126.0, 119.6, 119.2, 117.9, 108.2, 105.1, 104.1, 89.3, 19.4, 12.3. MALDI HRMS (DCTB): m/z calcd for C₇₆H₇₀O₁Si₂⁺ (M⁺) 1054.4960, found 1054.4963. **DSC**: decomposition, 121 °C (onset), 174 °C (peak). **TGA**: $T_d \approx 470$ °C.



Compound 2b. A solution of 3b (471 mg, 0.395 mmol) in THF (35 mL) was purged with N₂ for 2 min. To this solution was added SnCl₂·2H₂O (356 mg, 1.58 mmol) and 10% ag. H_2SO_4 (0.40 mL) in that order, and the reaction mixture was further purged with N_2 for 2 min. To limit light exposure, the reaction flask was covered in aluminum foil. After stirring for 5 h, the reaction mixture was poured into MeOH (600 mL) and the precipitate was collected on a glass frit, washed with MeOH (150 mL) and dried in vacuo to obtain 2b (371 mg, 88%) as a dark blue solid. $R_f = 0.58$ (CH₂Cl₂/hexanes 4:1). Mp = no visible change below 250 °C. **UV-Vis** (THF): λ_{max} (ε) 310 (496 000), 350 (37 500), 368 (51 500), 415 (5 480), 440 (8 010), 512 (sh, 2 810), 557 (10 100), 602 (27 700), 657 (54 200) nm. IR (cast film CH₂Cl₂): 3332 (br, w), 3049 (w), 2942 (s), 2891 (m), 2865 (s), 2126 (m), 1587 (m) cm⁻¹. ¹**H-NMR** (500 MHz, THF-*d*₈) δ 9.42 (s, 4H), 9.31 (s, 4H), 8.46 (s, 1H), 8.21-8.14 (m, 4H), 8.12-8.08 (m, 2H), 8.01-7.95 (m, 4H), 7.49-7.40 (m, 8H), 4.91 (d, J = 5.8 Hz, 2H), 4.67 (t, J = 5.8 Hz, 1H), 1.60–1.22 (m, 42H). ¹³C-NMR (126 MHz, THF- d_8) δ 145. 6, 134.1, 133.6, 133.5, 131.5, 131.1, 131.0, 129.6, 129.3, 127.3, 127.2, 127.0, 126.9, 125.0, 119.2, 118.9, 107.9, 105.9, 105.3, 89.1, 64.3, 19.5, 12.7. MALDI HRMS (DCTB) *m*/*z* calcd for C₇₇H₇₂OSi₂ (M⁺) 1068.5116, found 1068.5110. DSC: decomposition, 141 °C (onset), 183 °C (peak); 403 °C (onset), 460 °C (peak). **TGA**: $T_d \approx$ 485 °C. Crystals suitable for X-ray crystallography were grown by vapor diffusion from MeOH/THF.



Compound 2c. A solution of 3c (390 mg, 0.311 mmol) in THF (30 mL) was purged with N_2 for 2 min. To this solution was added SnCl₂·2H₂O (280 mg, 1.24 mmol) and 10% H_2SO_4 (0.35 mL) in that order, and the reaction mixture was further purged with N_2 for 2 min. To limit light exposure, the reaction flask was covered in aluminum foil. After stirring for 6 h, the reaction mixture was poured into MeOH (450 mL) and the precipitate was collected on a glass frit, washed with MeOH (150 mL), and dried in vacuo to obtain 2c (268 mg, 76%) as a dark blue solid. $R_f = 0.45$ (CH₂Cl₂/hexanes 4:1). Mp: no visible change below 250 °C. **UV-Vis** (THF): λ_{max} 310, 351, 369, 416, 440, 515 (sh), 558, 603, 657 nm. IR (cast film CH₂Cl₂): 3553 (w), 3368 (br, w), 3048 (m), 2942 (s), 2890 (m), 2865 (s), 2126 (m), 1612 (m), 1582 (m), 1515 (m) cm⁻¹. ¹H-NMR (500 MHz, THF-d₈) δ 9.46 (s, 4H), 9.33 (s, 4H), 8.54 (s, 1H), 8.50 (t, J = 1.3 Hz, 1H), 8.30 (d, J = 1.3 Hz, 2H), 8.22–8.16 (m, 4H), 8.01–7.96 (m, 4H), 7.83 (d, J = 8.5 Hz, 2H), 7.48–7.40 (m, 8H), 7.01 (d, J = 8.5 Hz, 2H), 1.56–1.31 (m, 42H). ¹³C-NMR (126 MHz, THF- d_8) δ 159.4, 143.6, 133.59, 133.55, 133.5, 131.5, 131.4, 131.2, 131.0, 129.7, 129.4, 129.3, 127.3, 127.2, 127.02, 126.98, 125.6, 119.2, 118.9, 116.8, 108.0, 105.9, 105.3, 89.2, 19.5, 12.7. **MALDI HRMS** (DCTB): m/z calcd for $C_{82}H_{74}OSi_2^+$ (M⁺) 1130.5273, found 1130.5260. DSC: decomposition, 89 °C (onset), 148 °C (peak); 396 °C (onset), 446 °C (peak). TGA: *T_d* ≈ 490 °C.



Compound 2d. A solution of **3d** (420 mg, 0.331 mmol) in THF (30 mL) was purged with N_2 for 2 min. To this solution was added SnCl₂·2H₂O (299 mg, 1.32 mmol) and 10% aq. H_2SO_4 (0.35 mL) in that order, and the reaction mixture was further purged with N_2 for 2 min. To limit light exposure, the reaction flask was covered in aluminum foil. After stirring for 6 h, the reaction mixture was poured into MeOH (450 mL) and the precipitate was collected on a glass frit, washed with MeOH (150 mL) and dried in vacuo to obtain 2d (341 mg, 90%) as a dark blue solid. $R_f = 0.25$ (CH₂Cl₂/hexanes 4:1). Mp = no visible change below 250 °C. **UV-Vis** (THF): λ_{max} (ϵ) 310 (582 300), 351 (40 400), 371 (57 100), 416 (51 700), 440 (8 400), 510 (sh, 3 110), 557 (11 600), 602 (32 100), 357 (61 800) nm. IR (cast film CH₂Cl₂): 3556 (w), 3443 (w), 3048 (w), 2942 (s), 2890 (m), 2865 (s), 2123 (m), 1586 (m), 1516 (w) cm⁻¹. ¹**H-NMR** (500 MHz, THF- d_8) δ 9.45 (s, 4H), 9.32 (s, 4H), 8.57–8.53 (m, 1H), 8.38 (s, 2H), 8.22–8.13 (m, 4H), 8.04–7.93 (m, 6H), 7.63 (d, J = 7.9 Hz, 2H), 7.52–7.36 (m, 8H), 4.76 (d, J = 5.8 Hz, 2H), 4.32 (t, J = 5.8 Hz, 1H), 1.66–1.20 (m, 42H). ¹³**C-NMR** (126 MHz, THF- d_8) δ 144.3, 143.5, 138.8, 134.2, 133.6, 133.5, 131.47, 131.46, 131.2, 129.6, 129.3, 128.0, 127.9, 127.3, 127.2, 127.0, 126.9, 125.7, 119.3, 118.8, 108.0, 105.9, 105.1, 89.4, 64.6, 19.5, 12.7. MALDI HRMS (DCTB) m/z calcd for C₈₃H₇₆OSi₂ (M⁺) 1144.5429, found 1144.5418. **DSC**: decomposition, 147 °C (onset), 206 °C (peak); 236 °C (onset), 268 °C (peak); 388 °C (onset), 411 °C (peak); 413 °C (onset), 460 °C (peak). **TGA**: *T_d* ≈ 485 °C.

Table S1. Crystallographic Experimental Details of 2b

A. Crystal Data formula $C_{81}H_{80}O_2Si_2$ formula weight 1141.63 crystal dimensions (mm) 0.27 × 0.06 × 0.05 crystal system triclinic space group P-1 (No. 2) unit cell parameters^a a (Å) 8.9356(3) *b* (Å) 17.9940(7) c (Å) 22.2531(9) α (deg) 69.3269(18) β (deg) 80.898(2) y (deg) 85.2585(19) V (Å³) 3304.1(2) Ζ 2 ρ calcd (g cm-3) 1.147 μ (mm⁻¹) 0.840 B. Data Collection and Refinement Conditions diffractometer Bruker D8/APEX II CCD^b Cu Ka (1.54178) (microfocus source) radiation (λ [Å]) temperature (°C) -100 scan type ω and ϕ scans (1.0°) (5 s exposures) data collection 2θ limit (deg) 140.74 total data collected 22808 ($-10 \le h \le 10, -21 \le k \le 21, -27 \le l \le 26$) independent reflections 12150 (*R*int = 0.0291) number of observed reflections (*NO*) 9884 $[F_0^2 \ge 2\sigma (F_0^2)]$ structure solution method intrinsic phasing (SHELXT-2014^c) refinement method full-matrix least-squares on F^2 (SHELXL-2016^d) absorption correction method Gaussian integration (face-indexed)



range of transmission factors1.0000-0.8139data/restraints/parameters 12150 / 0 / 770goodness-of-fit (S)^e [all data]1.047final R indices^f

 $R_1 [F_o^2 \ge 2\sigma (F_o^2)] = 0.0575$

wR2 [all data]0.1620

largest difference peak and hole 1.116 and –0.677 e Å $^{-3}$

^a Obtained from least-squares refinement of 9830 reflections with $5.26^{\circ} < 2\theta < 140.52^{\circ}$.

^{*b*} Programs for diffractometer operation, data collection, data reduction and absorption correction were supplied by Bruker.

^c Sheldrick, G. M. Acta Crystallogr. 2015, A71, 3–8. (SHELXT-2014)

^d Sheldrick, G. M. Acta Crystallogr. 2015, C71, 3–8. (SHELXL-2017)

^e S = $[\Box w(F_o^2 - F_c^2)^2/(n - p)]^{1/2}$ (*n* = number of data; *p* = number of parameters varied; *w*

= $[\Box^2 (F_0^2) + 1.6513P]^{-1}$ where $P = [Max(F_0^2, 0) + 2F_c^2]/3)$.

 ${}^{f}R1 = \Box ||F_{o}| - |F_{c}|| / \Box |F_{o}|; wR2 = [\Box w(F_{o}^{2} - F_{c}^{2})^{2} / \Box w(F_{o}^{4})]^{1/2}.$

Figure S1. ¹H-NMR spectrum (500 MHz) of **6c** recorded in $CDCI_3$. Residual solvent signal denoted by asterisk (*).



Figure S2. ¹³C-NMR spectrum (126 MHz) of **6c** recorded in CDCl₃. Residual solvent signals denoted by asterisk (*).



Figure S3. ¹H-NMR spectrum (500 MHz) of **6d** recorded in $CDCI_3$. Residual solvent signal denoted by asterisk (*).



Figure S4. ¹³C-NMR spectrum (126 MHz) of **6d** recorded in CDCl₃. Residual solvent signals denoted by asterisk (*).



Figure S5. ¹H-NMR spectrum (500 MHz) of **5b** recorded in $CDCI_3$. Residual solvent signal denoted by asterisk (*).



Figure S6. ¹³C-NMR spectrum (126 MHz) of **5b** recorded in CDCl₃. Residual solvent signals denoted by asterisk (*).



Figure S7. ¹H-NMR spectrum (500 MHz) of **5c** recorded in $CDCI_3$. Residual solvent signal denoted by asterisk (*).



Figure S8. ¹³C-NMR spectrum (126 MHz) of **5c** recorded in $CDCI_3$. Residual solvent signals denoted by asterisk (*).



Figure S9. ¹H-NMR spectrum (500 MHz) of 5d recorded in $CDCI_3$. Residual solvent signal denoted by asterisk (*).



Figure S10. ¹³C-NMR spectrum (126 MHz) of 5d recorded in $CDCI_3$. Residual solvent signals denoted by asterisk (*).



Figure S11. ¹H-NMR spectrum (500 MHz) of **4a** recorded in CD₂Cl₂. Residual solvent signal denoted by asterisk (*).



Figure S12. ¹³C-NMR spectrum (126 MHz) of **4a** recorded in CD_2CI_2 . Residual solvent signals denoted by asterisk (*).



Figure S13. ¹H-NMR spectrum (500 MHz) of **4b** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S14. ¹³C-NMR spectrum (126 MHz) of **4b** recorded in CD_2Cl_2 . Residual solvent signals denoted by asterisk (*).



Figure S15. ¹H-NMR spectrum (500 MHz) of **4c** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).


Figure S16. ¹³C-NMR spectrum (126 MHz) of **4c** recorded in CD_2CI_2 . Residual solvent signals denoted by asterisk (*).



Figure S17. ¹H-NMR spectrum (500 MHz) of **4d** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S18. ¹³C-NMR spectrum (126 MHz) of **4d** recorded in CD_2Cl_2 . Residual solvent signals denoted by asterisk (*).



Figure S19. ¹H-NMR spectrum (500 MHz) of **3a** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S20. ¹³C-NMR spectrum (126 MHz) of **3a** recorded in CD_2CI_2 . Residual solvent signals denoted by asterisk (*).



Figure S21. ¹H-NMR spectrum (500 MHz) of **3b** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S22. ¹³C-NMR spectrum (126 MHz) of **3b** recorded in CD_2Cl_2 . Residual solvent signals denoted by asterisk (*).



Figure S23. ¹H-NMR spectrum (500 MHz) of **3c** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S24. ¹³C-NMR spectrum (126 MHz) of **3c** recorded in CD_2CI_2 . Residual solvent signals denoted by asterisk (*).



Figure S25. ¹H-NMR spectrum (500 MHz) of **3d** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S26. ¹³C-NMR spectrum (126 MHz) of **3d** recorded in CD_2Cl_2 . Residual solvent signals denoted by asterisk (*).



Figure S27. ¹H-NMR spectrum (500 MHz) of **2a** recorded in CD_2CI_2 . Residual solvent signal denoted by asterisk (*).



Figure S28. ¹³C-NMR spectrum (126 MHz) of **2a** recorded in CD_2CI_2 . Residual solvent signals denoted by asterisk (*).



Figure S29. ¹H-NMR spectrum (500 MHz) of **2b** recorded in THF- d_8 . Residual solvent signal denoted by asterisk (*).



Figure S30. ¹³C-NMR spectrum (126 MHz) of **2b** recorded in THF- d_8 . Residual solvent signals denoted by asterisk (*).



Figure S31. ¹H-NMR spectrum (500 MHz) of **2c** recorded in THF- d_8 . Residual solvent signal denoted by asterisk (*).



Figure S32. ¹³C-NMR spectrum (126 MHz) of **2c** recorded in THF- d_8 . Residual solvent signals denoted by asterisk (*).



Figure S33. ¹H-NMR spectrum (500 MHz) of **2d** recorded in THF- d_8 . Residual solvent signal denoted by asterisk (*).



Figure S34. ¹³C-NMR spectrum (126 MHz) of **2d** recorded in THF- d_8 . Residual solvent signal denoted by asterisk (*).



Synthesis and characterization of SubPc-Pnc₂ derivatives 1a-d.

General method for the synthesis of SubPc-Pnc₂ derivatives **1a-d**: In a 10 mL roundbottomed flask equipped with a magnetic stirrer and a rubber seal, SubPcCl **9** (0.025 mmol) and silver trifluoromethanesulfonate (0.031 mmol, 1.25 equiv) were placed. Anhydrous toluene (1 mL) was added and the mixture was stirred under argon atmosphere at room temperature until the disappearance of the SubPcCl starting material, which was monitored by TLC. At this point Pnc₂ **2a-d** (0.031 mmol, 1.25 equiv) and *N*,*N*-diisopropylethylamine (0.031 mmol, 1.25 equiv) were added, and the reaction mixture was stirred at 50 °C. To limit light exposure, the reaction flask was covered in aluminum foil. The reaction mixture was then passed through a short Celite plug. The solvent was removed by evaporation under reduced pressure and the product was directly purified by size exclusion chromatography using CHCl₃ as eluent. The solvent was evaporated under reduced pressure and the product was then recrystallized from a CH₂Cl₂/hexane mixture. The reaction time and the reaction yields are indicated below for each compound.



SubPc-Pnc₂ 1a. Reaction time: 5 h. SubPc-Pnc₂ **1a** was afforded as a purple solid in 19% yield. ¹H-NMR (500 MHz, CDCl₃): δ (ppm) = 9.31 (s, 4H), 9.26 (s, 4H), 8.94–8.91 (AA'XX' system, 6H), 8.27–8.25 (m, 4H), 8.00–7.98 (m, 4H), 7.90–7.86 (AA'XX' system, 6H), 7.76 (t, *J* = 1.5 Hz, 1H), 7.45–7.43 (m, 8H), 6.00 (d, *J* = 1.5 Hz, 2H), 1.40–1.39 (m, 42H); ¹³C-NMR (125.7 MHz, CDCl₃): δ (ppm) = 151.92, 132.53, 132.49, 131.28, 131.27, 130.77, 130.42, 130.24, 129.11, 128.83, 127.85, 126.60, 126.24, 126.12, 124.73, 122.54, 122.16, 118.75, 117.85, 107.50, 104.93, 103.60, 88.49, 19.18, 11.88 (one signal coincident or not observed); ¹¹B-NMR (160.4 MHz, CDCl₃): δ (ppm) = -14.51; HRMS (MALDI-TOF): Calculated for C₁₀₀H₈₁BN₆OSi₂: 1449.6136; Found: 1449.6167; UV/Vis (toluene): λ_{max} (log ε) = 661 (4.83), 606 (4.54), 565 (4.99), 549 (sh), 525 (4.48), 442 (3.98), 417 (3.78), 374 (4.69), 353 (4.66) nm.



SubPc-Pnc₂ 1b. Reaction time: 13 h. SubPc-Pnc₂ **1b** was afforded as a purple solid in 26% yield. ¹H-NMR (500 MHz, THF-*d*₈): δ (ppm) = 9.43 (s, 4H), 9.38 (s, 4H), 8.88– 8.85 (AA'XX' system, 6H), 8.32–8.30 (m, 4H), 8.12 (s, 1H), 8.02–8.00 (m, 4H), 7.90– 7.87 (AA'XX' system, 6H), 7.50–7.46 (m, 8H), 7.17 (s, 2H), 2.94 (s, 2H), 1.44–1.43 (m, 42H); ¹³C-NMR (125.7 MHz, THF-*d*₈): δ (ppm) = The low solubility of this product prevented the registration of a clear ¹³C-NMR spectrum; ¹¹B-NMR (160.4 MHz, THF*d*₈): δ (ppm) = -14.22; HRMS (MALDI-TOF): Calculated for C₁₀₁H₈₃BN₆OSi₂: 1463.6292; Found: 1463.6302; UV/Vis (toluene): λ_{max} (log ε) = 660 (4.83), 605 (4.57), 564 (5.02), 547 (sh), 524 (4.50), 441 (3.99), 417 (3.73), 373 (4.77), 353 (4.68) nm.



SubPc-Pnc₂ 1c. Reaction time: 5 h. SubPc-Pnc₂ **1c** was afforded as a purple solid in 16% yield. ¹H-NMR (500 MHz, THF- d_8): δ (ppm) = 9.47 (s, 4H), 9.37 (s, 4H), 8.89–8.86 (AA'XX' system, 6H), 8.45 (s, 1H), 8.22–8.20 (m, 4H), 8.15 (d, J = 1.5 Hz, 2H), 8.01–7.99 (m, 4H), 7.97–7.94 (AA'XX' system, 6H), 7.47–7.43 (m, 8H), 7.41 (d, J = 8.5 Hz, 2H), 5.61 (d, J = 8.5 Hz, 2H), 1.43–1.42 (m, 42H); ¹³C-NMR (125.7 MHz,

THF- d_8): The low solubility of this product prevented the registration of a clear ¹³C-NMR spectrum; ¹¹B-NMR (160.4 MHz, THF- d_8): δ (ppm) = -14.49; HRMS (MALDI-TOF): Calculated for C₁₀₆H₈₅BN₆OSi₂: 1525.6449; Found: 1525.6489; UV/Vis (toluene): λ_{max} (log ϵ) = 660 (4.77), 604 (4.52), 563 (5.02), 548 (sh), 524 (4.49), 441 (3.98), 417 (3.76), 373 (4.71), 353 (4.66) nm.



SubPc-Pnc₂ 1d. Reaction time: 13 h. H₁₂SubPcOPnc₂ **1d** was afforded as a purple solid in 18% yield. ¹H-NMR (300 MHz, CDCl₃): δ (ppm) = 9.17 (s, 8H), 8.89–8.84 (AA'XX' system, 6H), 8.27 (s, 1H), 7.96–7.88 (m, 14H), 7.84 (d, *J* = 1.5 Hz, 2H), 7.42–7.30 (m, 10H), 6.67 (d, *J* = 8.1 Hz, 2H), 2.83 (s, 1H), 1.39–1.38 (m, 42H); ¹³C-NMR (125.7 MHz, CDCl₃): δ (ppm) = 151.54, 142.28, 139.60, 138.19, 133.18, 132.37, 131.17, 130.61, 130.55, 130.33, 129.91, 128.82, 128.78, 126.91, 126.87, 126.60, 126.25, 126.15, 125.90, 124.72, 122.27, 118.80 117.59, 108.11, 107.40, 104.86, 103.78, 89.06, 61.70, 19.19, 11.88; ¹¹B-NMR (160.4 MHz, CDCl₃): δ (ppm) = -14.51; HRMS (MALDI-TOF): Calculated for C₁₀₇H₈₇BN₆OSi₂: 1539.6606; Found: 1539.6541; UV/Vis (toluene): λ_{max} (log ϵ) = 660 (4.78), 604 (4.52), 562 (5.02), 546 (sh), 523 (4.48), 441 (3.95), 417 (3.71), 374 (4.73), 353 (4.65) nm.

Figure S35. ¹H-NMR spectrum (500 MHz, $CDCl_3$) of SubPc-Pnc₂ **1a**. *Inset*: Expansion of the region between 9.43 ppm and 7.33 ppm.



Figure S36. ¹³C-NMR spectrum (125.7 MHz, $CDCl_3$) of SubPc-Pnc₂ **1a**. *Inset*: Expansion of the region between 134 ppm and 116 ppm.



Figure S37. ¹¹B-NMR spectrum (160.4 MHz, CDCl₃) of SubPc-Pnc₂ 1a.







Figure S39. ¹H-NMR spectrum (500 MHz, THF- d_8) of SubPc-Pnc₂ **1b**. *Inset*: Expansion of the region between 9.55 ppm and 6.52 ppm.



Figure S40. ¹¹B-NMR spectrum (160.4 MHz, THF-*d*₈) of SubPc-Pnc₂ 1b.



Figure S41. H-H COSY NMR spectrum (500 MHz, THF-*d*₈) of SubPc-Pnc₂ 1b.



Figure S42. ¹H-NMR spectrum (500 MHz, THF- d_8) of SubPc-Pnc₂ **1c**. *Inset*: Expansion of the region between 9.58 ppm and 7.28 ppm.



Figure S43. ¹¹B-NMR spectrum (160.4 MHz, THF-*d*₈) of SubPc-Pnc₂ **1c**.



Figure S44. H-H COSY NMR spectrum (500 MHz, THF-*d*₈) of SubPc-Pnc₂ 1c.



bpm

Figure S45. ¹H-NMR spectrum (300 MHz, CDCl₃) of SubPc-Pnc₂ **1d**. *Inset*: Expansion of the region between 9.31 ppm and 6.52 ppm.



Figure S46. ¹³C-NMR spectrum (125.7 MHz, $CDCl_3$) of SubPc-Pnc₂ **1d**. *Inset*: Expansion of the region between 144 ppm and 115 ppm.



Figure S47. ¹¹B-NMR spectrum (160.4 MHz, CDCl₃) of SubPc-Pnc₂ 1d.



Figure S48. H-H COSY NMR spectrum (500 MHz, CDCl₃) of SubPc-Pnc₂ 1d.



mdd


Figure S49. MALDI-TOF mass spectrum (in DCTB matrix) of SubPc-Pnc₂ 1a.

Figure S50. *Upper part* – HRMS (MALDI-TOF) spectrum (in DCTB + PPG1000 + 2000 matrix) of SubPc-Pnc₂ **1a**. *Lower part* – Calculated isotopic pattern of SubPc-Pnc₂ **1a**.





Figure S51. MALDI-TOF mass spectrum (in DCTB matrix) of SubPc-Pnc₂ 1b.

Figure S52. *Upper part* – HRMS (MALDI-TOF) spectrum (in DCTB + PPG1000 + 2000 matrix) of SubPc-Pnc₂ **1b**. *Lower part* – Calculated isotopic pattern of SubPc-Pnc₂ **1b**.





Figure S53. MALDI-TOF mass spectrum (in DCTB matrix) of SubPc-Pnc₂ 1c.

Figure S54. *Upper part* – HRMS (MALDI-TOF) spectrum (in DCTB + PPG1000 + 2000 matrix) of SubPc-Pnc₂ **1c**. *Lower part* – Calculated isotopic pattern of SubPc-Pnc₂ **1c**.





Figure S55. MALDI-TOF mass spectrum (in DCTB matrix) of SubPc-Pnc₂ 1d.

Figure S56. *Upper part* – HRMS (MALDI-TOF) spectrum (in DCTB + PPG1000 + 2000 matrix) of SubPc-Pnc₂ **1d**. *Lower part* – Calculated isotopic pattern of SubPc-Pnc₂ **1d**.



Photophysics

Figure S57. Steady-state absorption spectra of SubPcCl **9**, Pnc_2 **2a**, and SubPc-Pnc₂ **1a** in toluene (rt) – see figure legends for details.



Figure S58. Steady-state absorption spectra of SubPcCl **9**, Pnc_2 **2b**, and SubPc-Pnc₂ **1b** in toluene (rt) – see figure legends for details.



Figure S59. Steady-state absorption spectra of SubPcCl **9**, Pnc_2 **2d**, and SubPc-Pnc₂ **1d** in toluene (rt) – see figure legends for details.



Figure S60. Steady-state fluorescence spectra obtained upon 500 nm photoexcitation of SubPcCl **9**, Pnc_2 **2a**, and SubPc-Pnc₂ **1a** in toluene (rt) – see figure legends for details.



Figure S61. Steady-state fluorescence spectra obtained upon 500 nm photoexcitation of SubPcCl **9**, Pnc_2 **2b**, and SubPc-Pnc₂ **1b** in toluene (rt) – see figure legends for detail.



Figure S62. Steady-state fluorescence spectra obtained upon 500 nm photoexcitation of SubPcCl **9**, Pnc_2 **2d**, and SubPc-Pnc₂ **1d** in toluene (rt) – see figure legends for details.



	λ _{abs} / nm		λ _{em} / nm		Φ _F / %	T _F
	SubPc	Pnc ₂	SubPc	Pnc ₂	SubPc / Pnc ₂	SubPc ^[a, d]
1a	565	662	571	665	0.05 / 1.06	<50 ps ^[b]
2a	-	659	-	664	- / 1.47	<100 ps ^[b, c]
1b	564	660	571	665	0.03 / 0.90	<50 ps ^[b]
2b	-	659	-	664	- / 0.75	<100 ps ^[b, c]
1c	563	660	570	664	0.12 / 1.19	<50 ps ^[b]
2c	-	660	-	664	- / 1.03	<100 ps ^[b, c]
1d	562	660	569	665	0.09 / 0.66	<50 ps ^[b]
2d	-	660	-	665	- / 0.80	<100 ps ^[c]
9	565	-	570	-	28.81 / -	3.35 ns

Table S2. Summary of photophysical properties of Pnc₂s **2a-d**, SubPc-Pnc₂ conjugates **1a-d**, and SubPc **9** in toluene.

[a] values refer to the SubPc centered fluorescence if not stated otherwise. [b] lifetime is below the resolution limit of our time-correlated single photon counting (TCSPC) setup. [c] value refers to the Pnc₂ centered fluorescence. [d] An additional, long-lived component with lifetimes similar to those of the corresponding reference compounds points to minor contributions from free, highly fluorescent SubPcs and Pnc monomer.

Figure S63. (A) Differential absorption spectra (visible) obtained upon fsTA experiments (632 nm) of Pnc₂ **2b** in toluene with several time delays between 0 and 5500 ps at room temperature. Single wavelength kinetics at 450 (black), 505 (red), 601 (blue), and 663 nm (green) monitoring the singlet excited state (${}^{1}(S_{1}S_{0})$): 17.13 ps), intermediate state (${}^{CT}(S_{1}S_{0})$): 88.83 ps), and triplet excited state (${}^{1}(T_{1}T_{1})$): 2.50 ns) dynamics as well as fits to the data (red lines) are shown in the *inset*. (B) Evolution associated spectra of the transient absorption data of Pnc₂ **2b** shown in (A). The red spectrum illustrates the initially formed Pnc₂ ${}^{1}(S_{1}S_{0})$ state, while the blue and green spectra are those of the intermediate Pnc₂ ${}^{CT}(S_{1}S_{0})$ state and the multiexcitonic Pnc₂ ${}^{1}(T_{1}T_{1})$ state, respectively. The *inset* depicts the relative populations of ${}^{1}(S_{1}S_{0})$, ${}^{CT}(S_{1}S_{0})$, and ${}^{1}(T_{1}T_{1})$.



Figure S64. (A) Differential absorption spectra (visible) obtained upon fsTA experiments (632 nm) of Pnc₂ 2d in toluene with several time delays between 0 and 5500 ps at room temperature. Single wavelength kinetics at 450 (black), 505 (red), 603 (blue), and 663 nm (green) monitoring the singlet excited state (${}^{1}(S_{1}S_{0})$): 16.62 ps), intermediate state (${}^{CT}(S_{1}S_{0})$): 104.92 ps), and triplet excited state (${}^{1}(T_{1}T_{1})$): 2.68 ns) dynamics as well as fits to the data (red lines) are shown in the *inset*. (B) Evolution associated spectra of the transient absorption data of Pnc₂ 2d shown in (A). The red spectrum illustrates the initially formed Pnc₂ ${}^{1}(S_{1}S_{0})$ state, while the blue and green spectra are those of the intermediate Pnc₂ ${}^{CT}(S_{1}S_{0})$ state and the multiexcitonic Pnc₂ ${}^{1}(T_{1}T_{1})$ state, respectively. The *inset* depicts the relative populations of ${}^{1}(S_{1}S_{0})$, ${}^{CT}(S_{1}S_{0})$, and ${}^{1}(T_{1}T_{1})$.



Figure S65. (A) Differential absorption spectra (visible) obtained upon fsTA experiments (530 nm) of SubPc-Pnc₂ **1b** in toluene with several time delays between 0 and 5500 ps at room temperature. Single wavelength kinetics at 450 (black), 506 (red), 564 (blue), and 663 nm (green) monitoring the singlet excited state (${}^{1}(S_{1})$: 0.70 ps), singlet excited state (${}^{1}(S_{1}S_{0})$: 7.28 ps), intermediate state (${}^{CT}(S_{1}S_{0})$: 77.38 ps), and triplet excited state (${}^{1}(T_{1}T_{1})$: 2.51 ns) dynamics as well as fits to the data (red lines) are shown in the *inset*. (B) Evolution associated spectra of the transient absorption data of **1b** shown in (A). The black spectrum illustrates the initially formed SubPc ${}^{1}(S_{1}S_{0})$ state, the intermediate Pnc₂ ${}^{CT}(S_{1}S_{0})$ state, respectively. The *inset* depicts the relative populations of ${}^{1}(S_{1})$, ${}^{1}(S_{1}S_{0})$, ${}^{CT}(S_{1}S0)$, and ${}^{1}(T_{1}T_{1})$.



Figure S66. (A) Differential absorption spectra (visible) obtained upon fsTA experiments (530 nm) of SubPc-Pnc₂ **1d** in toluene with several time delays between 0 and 5500 ps at room temperature. Single wavelength kinetics at 450 (black), 505 (red), 562 (blue), and 663 nm (green) monitoring the singlet excited state (${}^{1}(S_{1})$: 1.28 ps), singlet excited state (${}^{1}(S_{1}S_{0})$: 14.74 ps), intermediate state (${}^{CT}(S_{1}S_{0})$: 104.09 ps), and triplet excited state (${}^{1}(T_{1}T_{1})$: 2.88 ns) dynamics as well as fits to the data (red lines) are shown in the *inset*. (B) Evolution associated spectra of the transient absorption data of **1d** shown in (A). The black spectrum illustrates the initially formed SubPc ${}^{1}(S_{1}S_{0})$ state, the intermediate Pnc₂ ${}^{CT}(S_{1}S_{0})$ state, respectively. The *inset* depicts the relative populations of ${}^{1}(S_{1})$, ${}^{1}(S_{1}S_{0})$, ${}^{CT}(S_{1}S0)$, and ${}^{1}(T_{1}T_{1})$.



Figure S67. (A) Differential absorption spectra (visible) obtained upon nsTA experiments (530 nm) of SubPcCl **9** in toluene with several time delays between 0 and 400 µs at room temperature. Single wavelength kinetics at 451 (black), 519 (red), 569 (blue), and 611 nm (green) monitoring the singlet excited state ($^{1}(S_1)$: 3.55 ns) and triplet excited state ($^{1}(T_1T_1)$: 51.64 µs) dynamics as well as fits to the data (red lines) are shown in the *inset*. (B) Evolution associated spectra of the transient absorption data of **9** shown in (A). The black spectrum illustrates the initially formed SubPc $^{1}(S_1)$ state, while the orange spectrum is that of the SubPc $^{3}(T_1)$ state. The *inset* depicts the relative populations of $^{1}(S_1)$ and $^{3}(T_1)$.



Computational Details

Geometries were optimized at the B3LYP¹⁶/6-31G(d)¹⁷ level of density-functional theory with Grimme's D3 dispersion correction.¹⁸ Two alternative conformations were investigated for **1a-c** and three for **1d**. Additional geometry optimizations using a polarizable continuum model for solvation in toluene¹⁹ gave little change compared to the gas-phase calculations. The first two conformations differ in the orientation of the pentacenes to each other and the third conformation for **1d** used a stretched conformation for the linker.

Absorption spectra were calculated using the AM1 semiempirical Hamiltonian²⁰ using a singles-only configuration interaction (CIS) with 32 occupied and 32 virtual orbitals in the active space. Initial tests with larger active spaces and an SCRF continuum solvent treatment²¹ for toluene showed no significant changes in the calculated spectra. FRET orientation factors κ^2 were calculated according to Loura²² using the AM1-CIS transition dipoles for the relevant excited singlets from the ground state.

Figure S68. Optimized geometries of **1a** with the respective energies in kcal mol⁻¹ in the gas phase and toluene (in parentheses).



Figure S69. Optimized geometries of **1b** with the respective energies in kcal mol⁻¹ in the gas phase and toluene (in parentheses).



Figure S70. Optimized geometries of **1c** with the respective energies in kcal mol⁻¹ in the gas phase and toluene (in parentheses).



Figure S71. Optimized geometries of **1d** with the respective energies in kcal mol⁻¹ in the gas phase and toluene (in parentheses).



Table S3. Calculated vertical absorption energies, inter-chromophore distances and angles between transition dipoles for compound **1a-d**. The results are shown for the most stable conformation in every case except **1d**, for which the extended conformation is also shown.

Compound	Excitation energy (eV)			R ^a (Å)	Angle ^b (°)	κ ² /ℝ ⁶
	SubPc	Pentacene	Δ		, g .e ()	
	2.080		I	I		I
1 A	2.093					
		2.378	0.285	2.776	35	0.001268
	2.087					
1B	2.102					
		2.369	0.267	2.661	62	0.002277
	2.099					
1C	2.102					
		2.393	0.291	6.492	62	1.08E-05
1D	2.108					
extended	2.109					
extended		2.395	0.286	12.635	8	2.36E-07
	2.091					
1D	2.107					
		2.393	0.286	2.725	82	0.002937



Figure S72. Correlation between the experimental FRET quenching and calculated κ^2/R^6 for 1a-d.

Author Contributions

R.R.T., T.T., and D.M.G. designed the research. J.Z. performed the photophysical studies. G.L. performed the synthesis of **1a-d** and **9**, while H.G. and L.C. synthesized **2a-d** and the corresponding precursors. J.Z., I.P. and D.M.G. analyzed the photophysical data. T.C. performed the computational studies. All authors contributed to the writing and editing of the paper.

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