## **Supporting Information**

On the intimate connection between nanoscale adhesion of Yad fimbriae and macroscale attachment of Yad-decorated bacteria to glycosylated, hydrophobic and hydrophilic surfaces

Grégory Francius <sup>1*</sup> , Florian Petit <sup>1</sup> , Eloïse Clément <sup>1</sup> , Yankel Chekli <sup>2,3</sup> , Jean-Marc Ghigo <sup>2</sup> ,
Christophe Beloin <sup>2,#</sup> , Jérôme F.L. Duval <sup>4,#</sup>

# equivalent contribution

\* Corresponding authors:

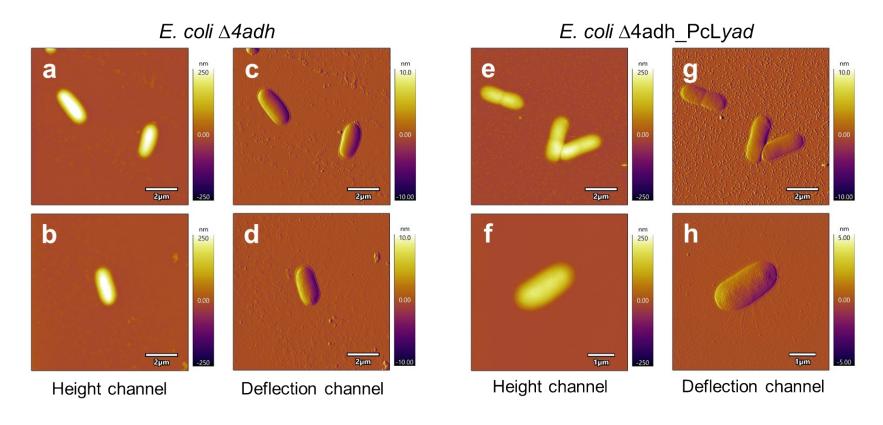
gregory.francius@univ-lorraine.fr

<sup>&</sup>lt;sup>1</sup> Université de Lorraine, LCPME, UMR 7564, Villers-lès-Nancy, F-54600, France.

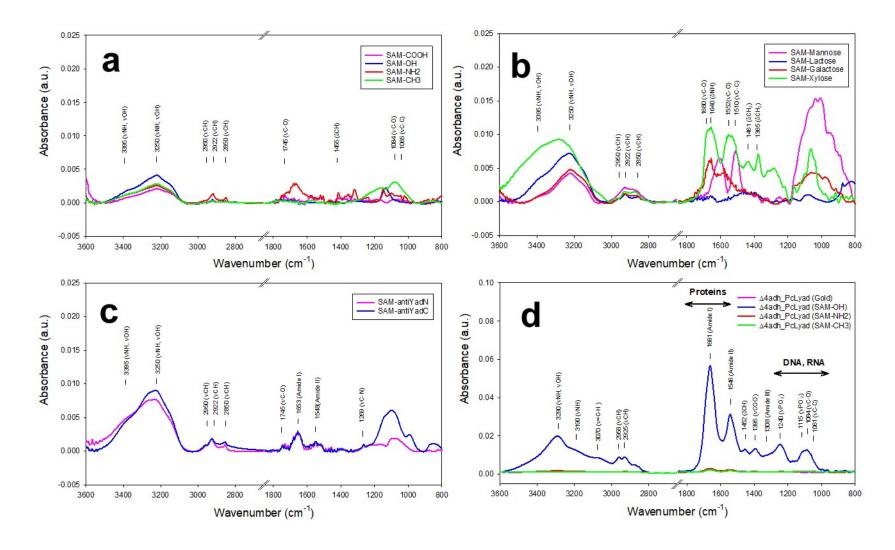
<sup>&</sup>lt;sup>2</sup> Genetics of Biofilms Laboratory, Institut Pasteur, UMR CNRS2001, Paris, 75015, France.

<sup>&</sup>lt;sup>3</sup> Université de Paris, Sorbonne Paris Cité, Paris, France.

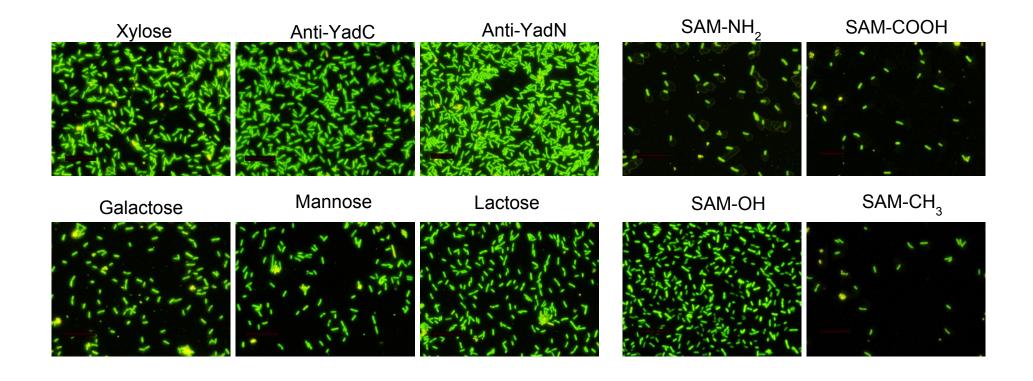
<sup>&</sup>lt;sup>4</sup> Université de Lorraine, LIEC, UMR 7360, Vandœuvre-lès-Nancy, F-54501, France.



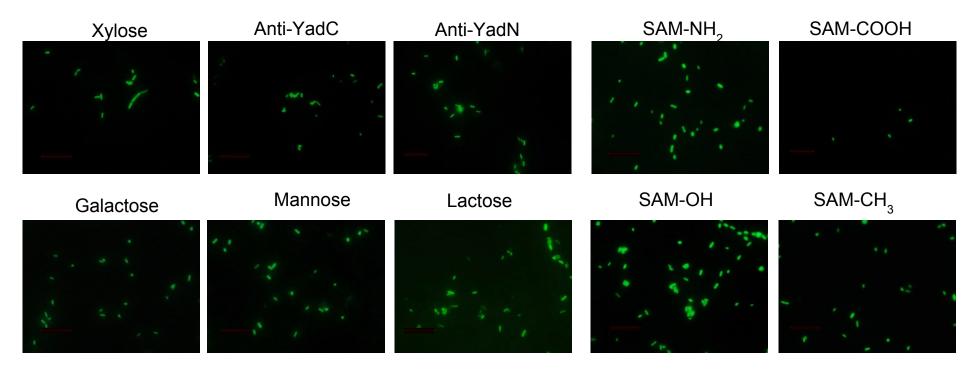
**Figure S1.** Images of *E. coli*  $\Delta 4adh$  and  $\Delta 4adh$ \_PcLyad recorded by AFM operated in contact mode under air conditions at 23°C.



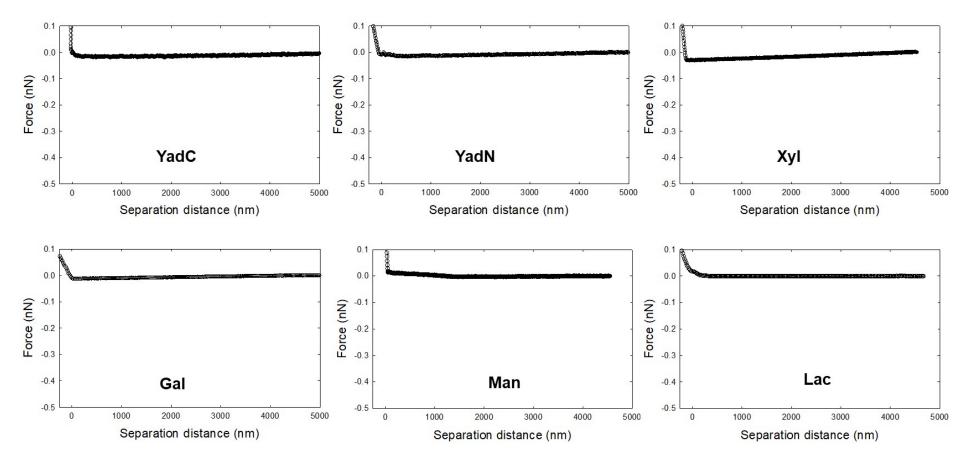
**Figure S2.** Infrared reflection absorption spectroscopy (IRRAS) spectra measured for the various model self-assembled monolayers (SAM) of interest in this work (indicated), and assignments of the bands.



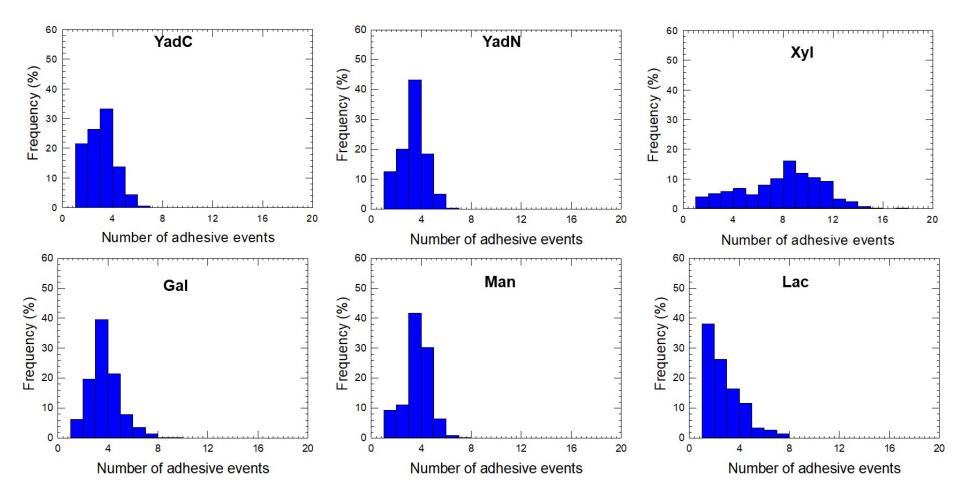
**Figure S3.** Representative epifluorescence images (filter cube U-MWIB3, Olympus, excitation filter: BP 460-495 nm, emission filter: LP 510 nm) of *E. coli*  $\Delta_4 adh$ \_PcLyad bacteria incubated 2 hrs onto the different SAM-coated gold substrates covered by anti-YadC, anti-YadN antibodies, xylose (Xyl), galactose (Gal), lactose (Lac), mannose (Man) (indicated) and onto the SAM-NH2, SAM-COOH, SAM-OH and SAM-CH3 reference substrates. *E. coli*  $\Delta_4 adh$ \_PcLyad bacterial cells constitutively expressing GFP were stained with PI to verify membrane integrity. Green and red fluorescence emissions were simultaneously acquired and damaged bacteria appears in yellow. Bars: 20 μm (white).



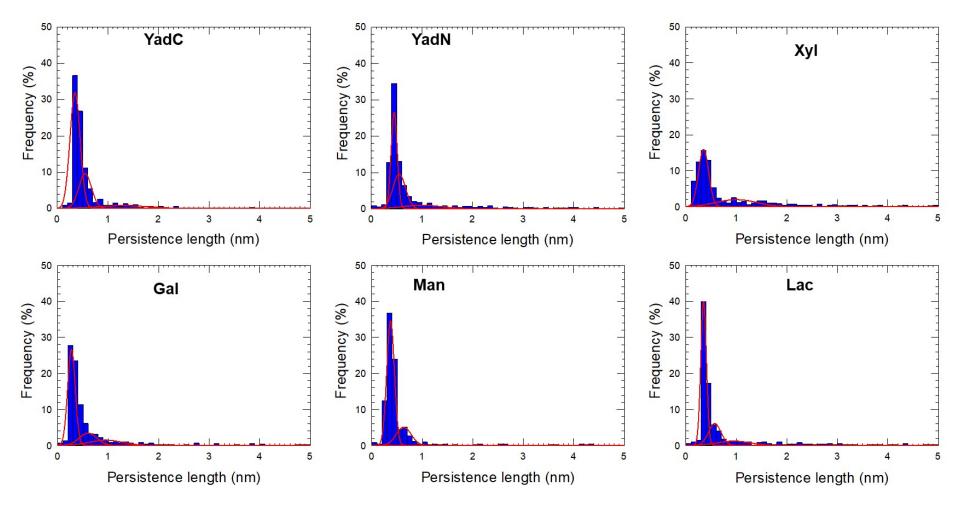
**Figure S4.** Representative epifluorescence images (filter cube U-MWIB3, Olympus, excitation filter: BP 460-495 nm, emission filter: LP 510 nm) of *E. coli*  $\Delta_4 adh$  bacteria incubated 2 hrs onto the different SAM-coated gold substrates covered by YadC, YadN antibodies, xylose (Xyl), galactose (Gal), lactose (Lac), mannose (Man) (indicated) and onto the SAM-NH2, SAM-COOH, SAM-OH and SAM-CH3 reference substrates. *E. coli*  $\Delta_4 adh$  bacterial cells constitutively expressing GFP were stained with PI to verify membrane integrity. Green and red fluorescence emissions were simultaneously acquired and damaged bacteria appears in yellow. Bars: 20 μm (white).



**Figure S5.** Illustrative force vs. separation distance profiles (black circles) measured by SMFS upon withdrawal of several AFM-probes (functionalized with YadC, YadN antibodies, xylose (Xyl), galactose (Gal), lactose (Lac) and mannose (Man)) from E.  $coli \Delta_4 adh$  cell wall. This figure clearly evidences the absence of adhesion and accordingly there is no modelling of the data reported here.



**Figure S6.** Statistic distribution of the number of adhesive events estimated from the SMFS force-distance curves measured for *E. coli*  $\Delta_4 adh$ \_PcL*yad* interacting with AFM-probes functionalized with YadC, YadN antibodies, xylose (Xyl), galactose (Gal), lactose (Lac) and mannose (Man) (indicated).



**Figure S7.** Statistic distribution of the persistence length of Yad fimbriae estimated from the SMFS force-distance curves measured for *E. coli*  $\Delta_4 adh$ \_PcLyad interacting with AFM-probes functionalized with YadC, YadN antibodies, xylose (Xyl), galactose (Gal), lactose (Lac) and mannose (Man) (indicated). Red lines correspond to decomposition of the distribution into fundamental Gaussian components.