

Supporting Information

for

**Synthesis and evaluation of N^α, N^ϵ -diacetyl-L-lysine-inositol conjugates as cancer-selective probes for
metabolic engineering of GPIs and GPI-anchored proteins**

Mohit Jaiswal[†], Sanyong Zhu[†], Wenjie Jiang, and Zhongwu Guo*

Department of Chemistry, University of Florida, 214 Leigh Hall, Gainesville, FL 32611, United States of America

*Corresponding author e-mail: zguo@chem.ufl.edu

Table of Contents

I. Supplementary Flow Cytometry Data -----	Page S2-S5
II. NMR and MS Spectra of Compounds -----	Page S6-S76

I. Supplementary Flow Cytometry Data

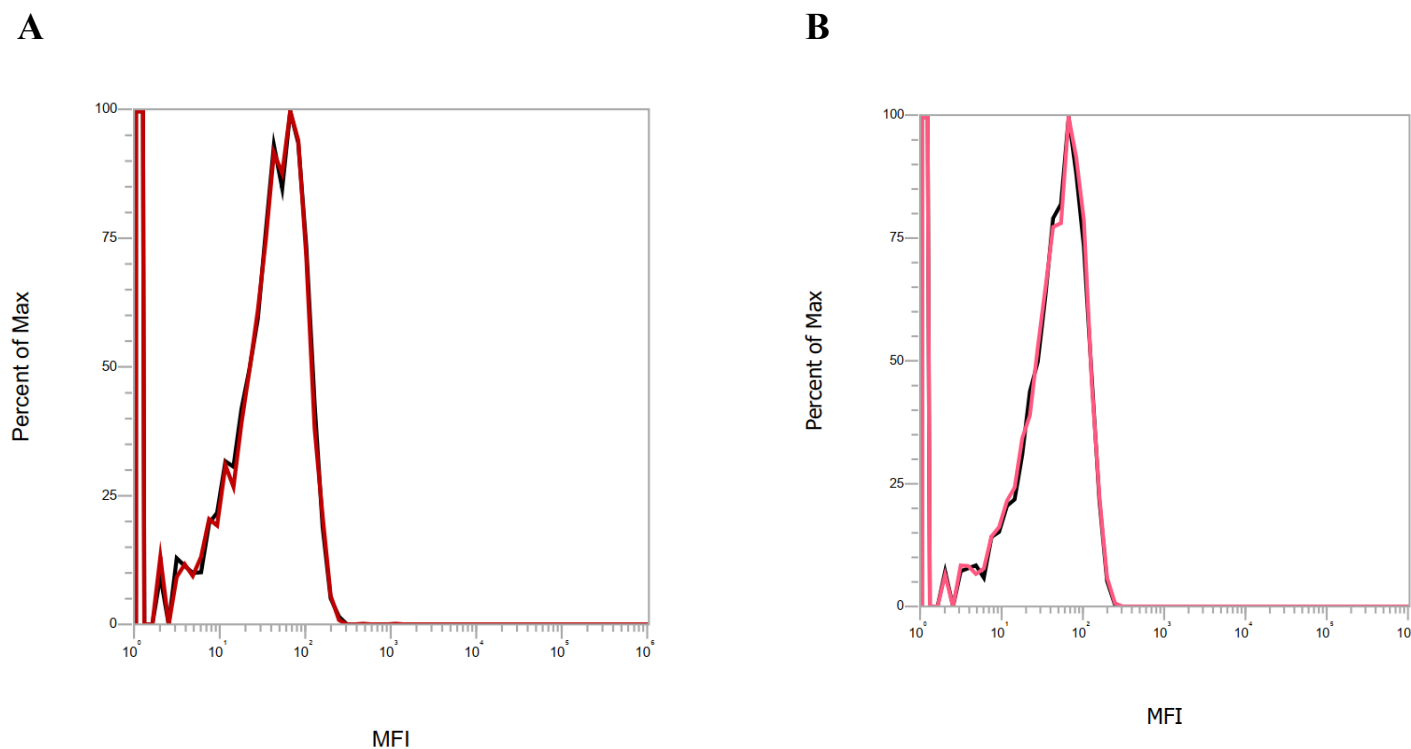


Figure S1: Flow cytometry results of HEK293 cell treated with 200 μ M of **1** (A) or **2** (B) (red) against cell treated with PBS (control group, black) for 48 h, followed by click reaction with DBCO-Cy5 (50 μ M, 37 $^{\circ}$ C, 1 h). More than 10,000 events were recorded during each experiment and three replicate experiments were performed.

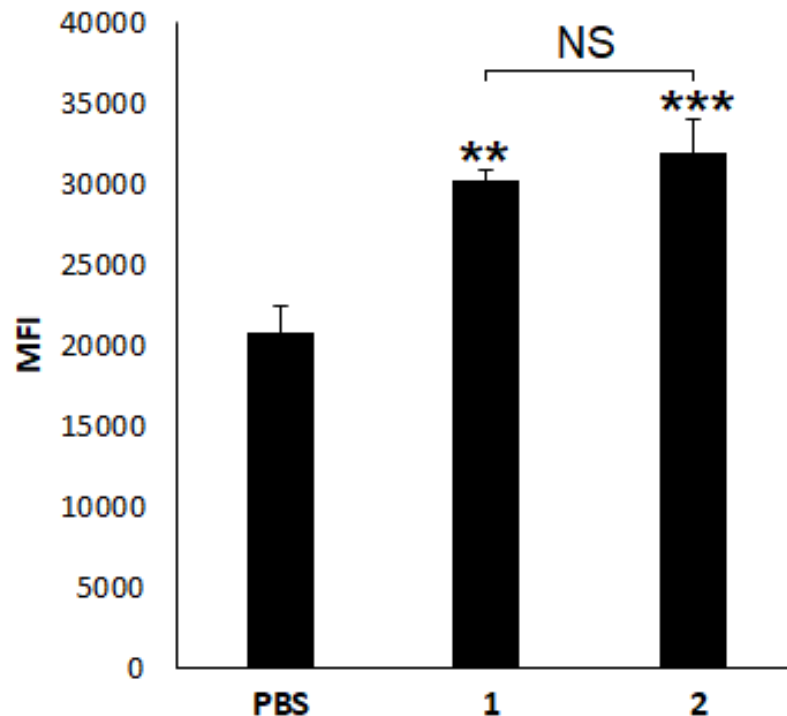


Figure S2: Flow cytometry results of MCF-7 cell treated with PBS (control) or with 200 μ M of **1** or **2** for 48 h and then with DBCO-Cy5 (50 μ M, 37 $^{\circ}$ C, 1 h). Data are presented as mean \pm standard deviation of three parallel experiments and analyzed by two-tailed student's *t* test: **statistically very different ($P \leq 0.01$) from the PBS group; ***statistically extremely different ($P \leq 0.001$) from the PBS group; NS, no significant difference ($P > 0.05$).

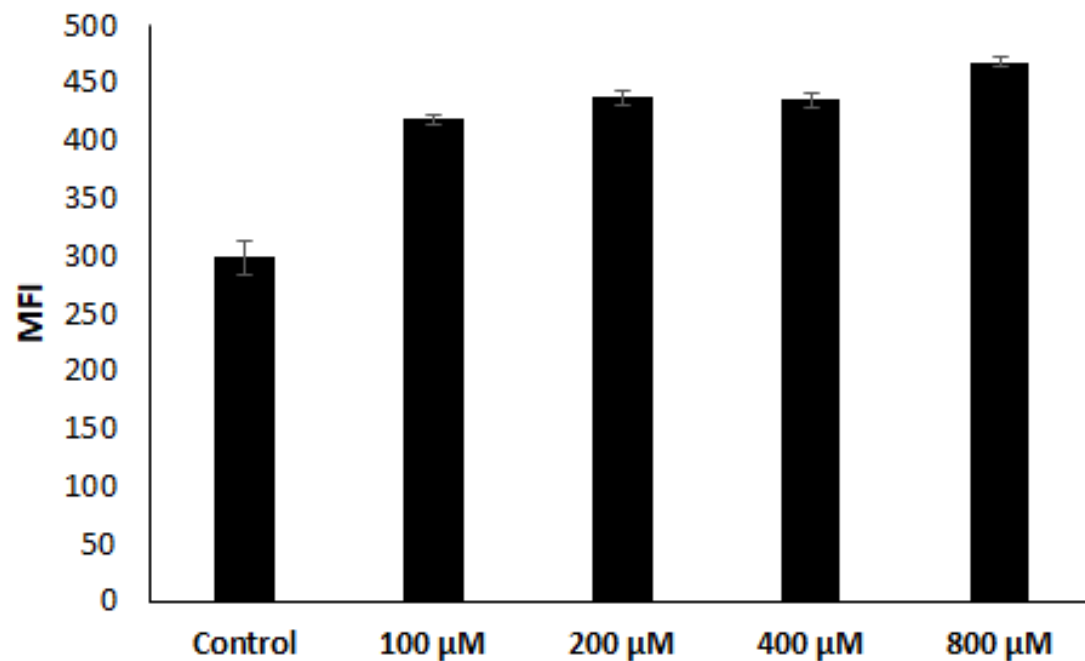


Figure S3: Flow cytometry results of MCF-7 cell treated with PBS (control) or with various concentrations (100-800 μ M) of **1** for 48 h and then with DBCO-Cy5 (50 μ M, 37 $^{\circ}$ C, 1 h). Data are presented as mean \pm standard deviation of three parallel experiments. Difference of the treatment groups with the control (PBS) group was all statistically significant ($P \leq 0.05$).

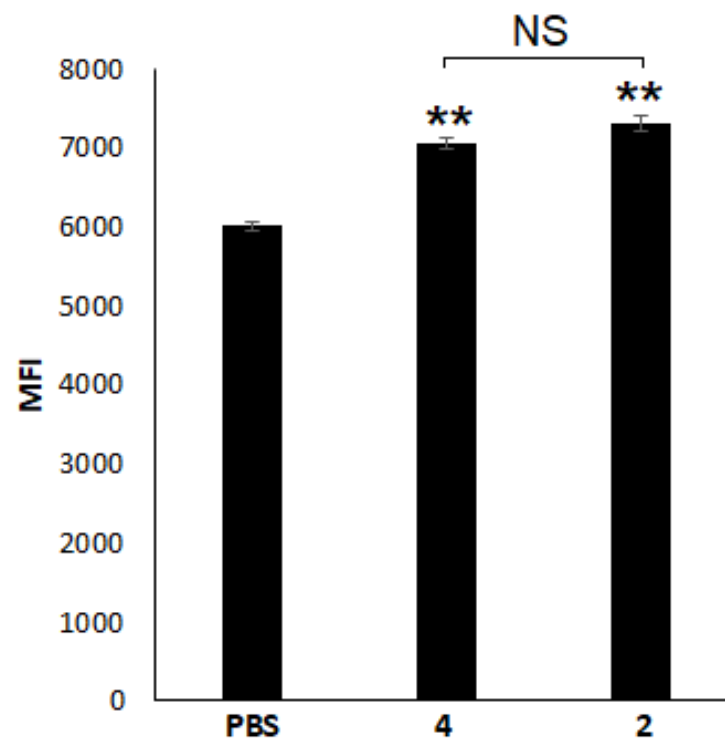
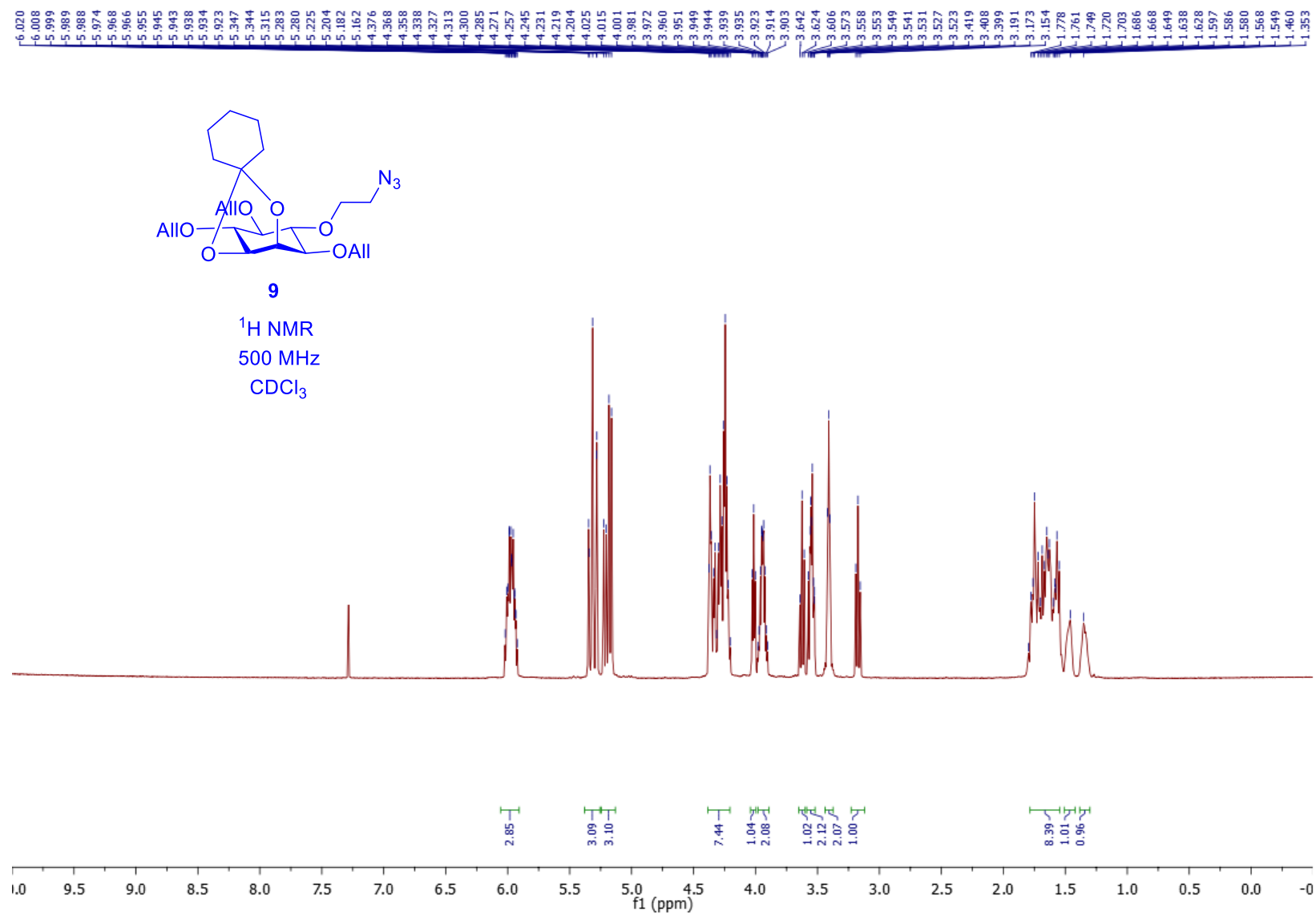
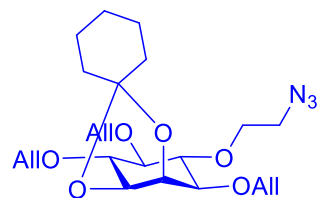


Figure S4: Flow cytometry results of HepG2 cell treated with PBS (control) or with 200 μ M of **2** or **4** for 48 h and then with DBCO-Cy5 (50 μ M, 37 $^{\circ}$ C, 1 h). Data are presented as mean \pm standard deviation of two parallel experiments and analyzed by two-tailed student's *t* test: **statistically very different ($P \leq 0.01$) from the PBS group; NS, no significant difference ($P > 0.05$).

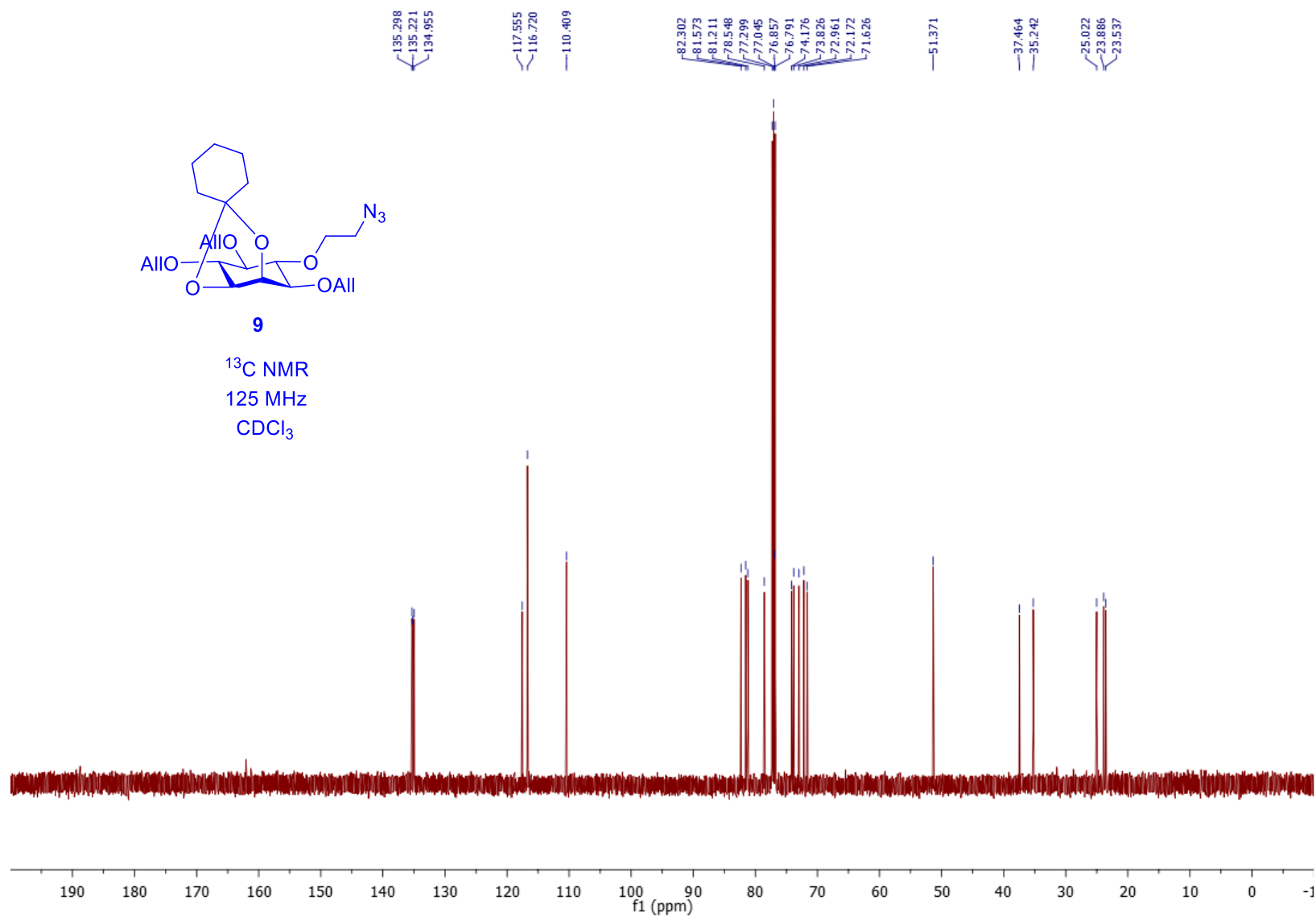
II. NMR and MS Spectra of Compounds

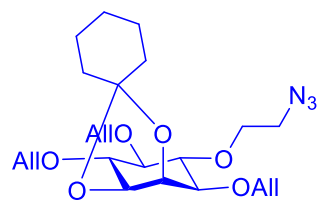




9

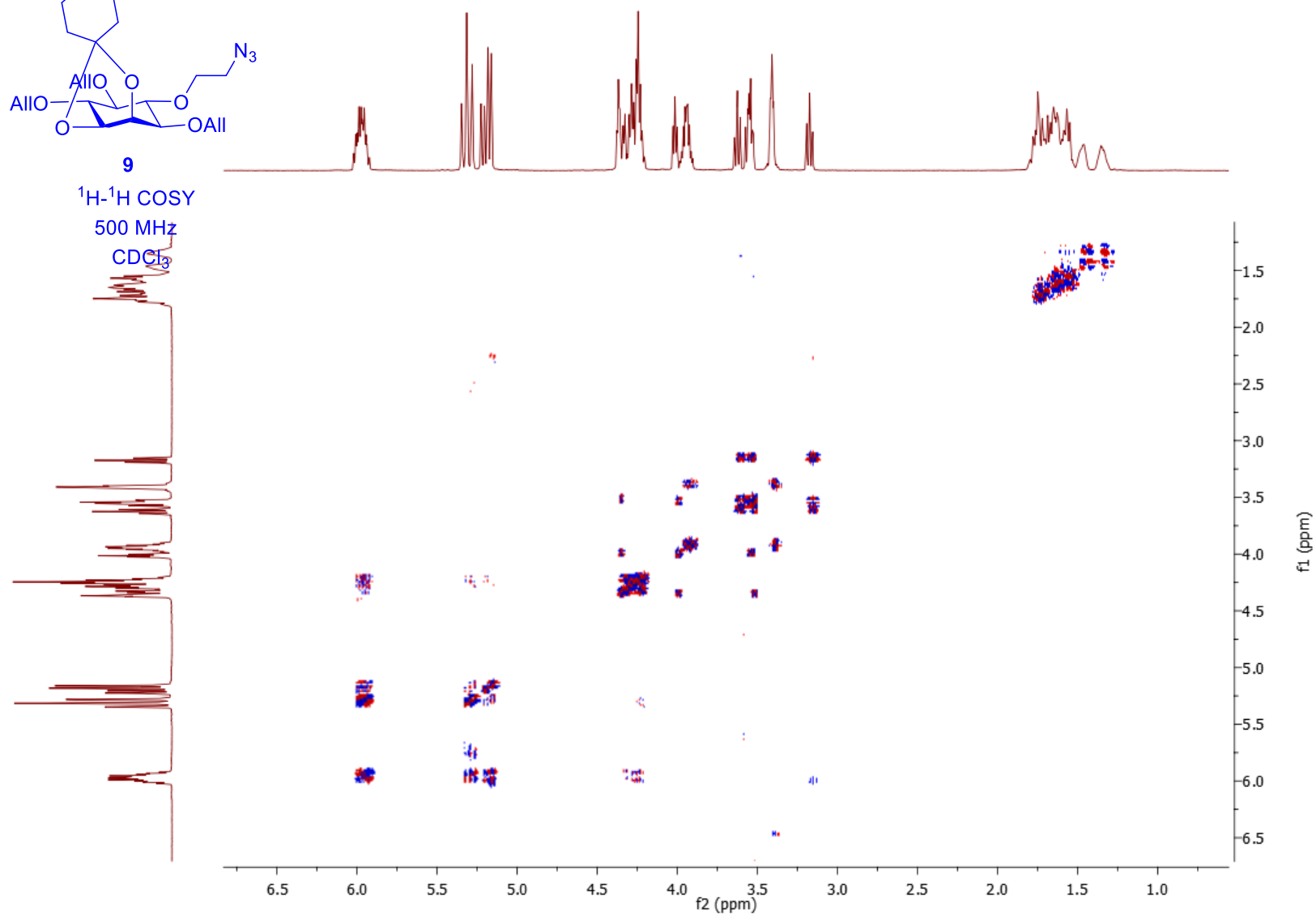
¹³C NMR
125 MHz
CDCl₃

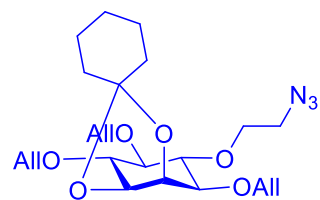




9

^1H - ^1H COSY
500 MHz
 CDCl_3



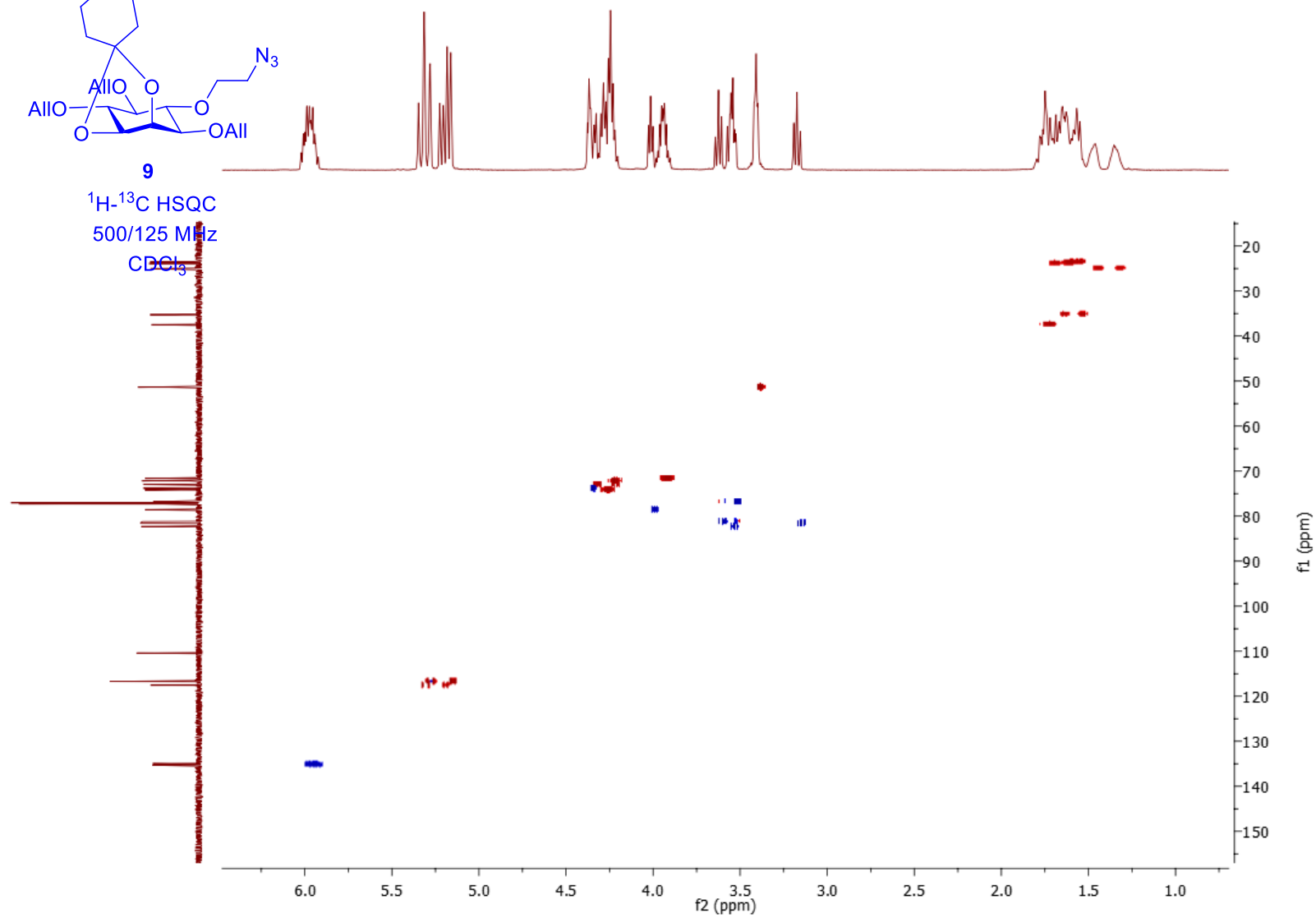


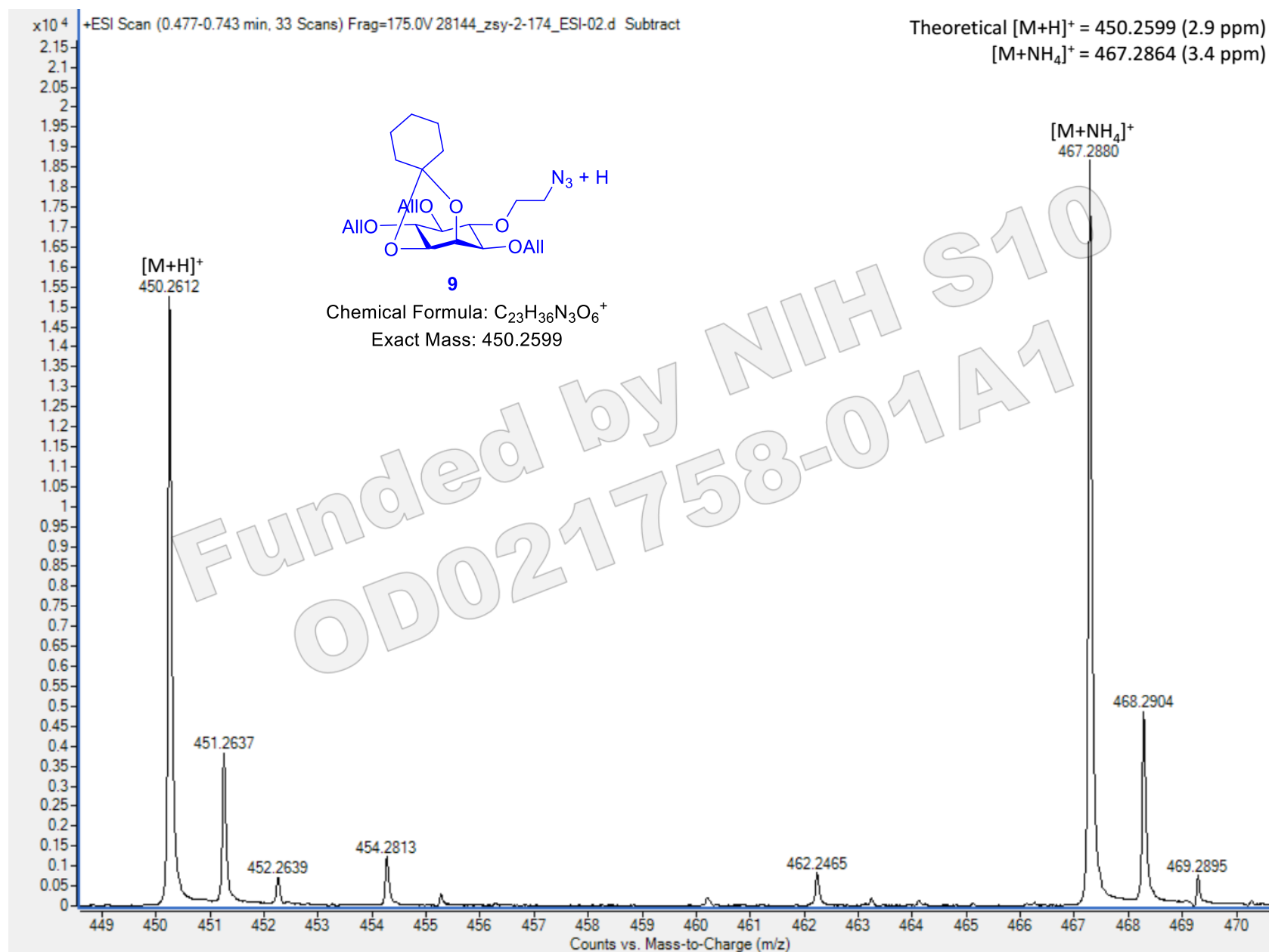
9

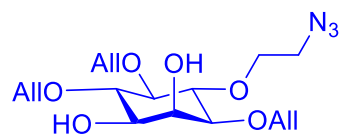
^1H - ^{13}C HSQC

500/125 MHz

CDCl_3

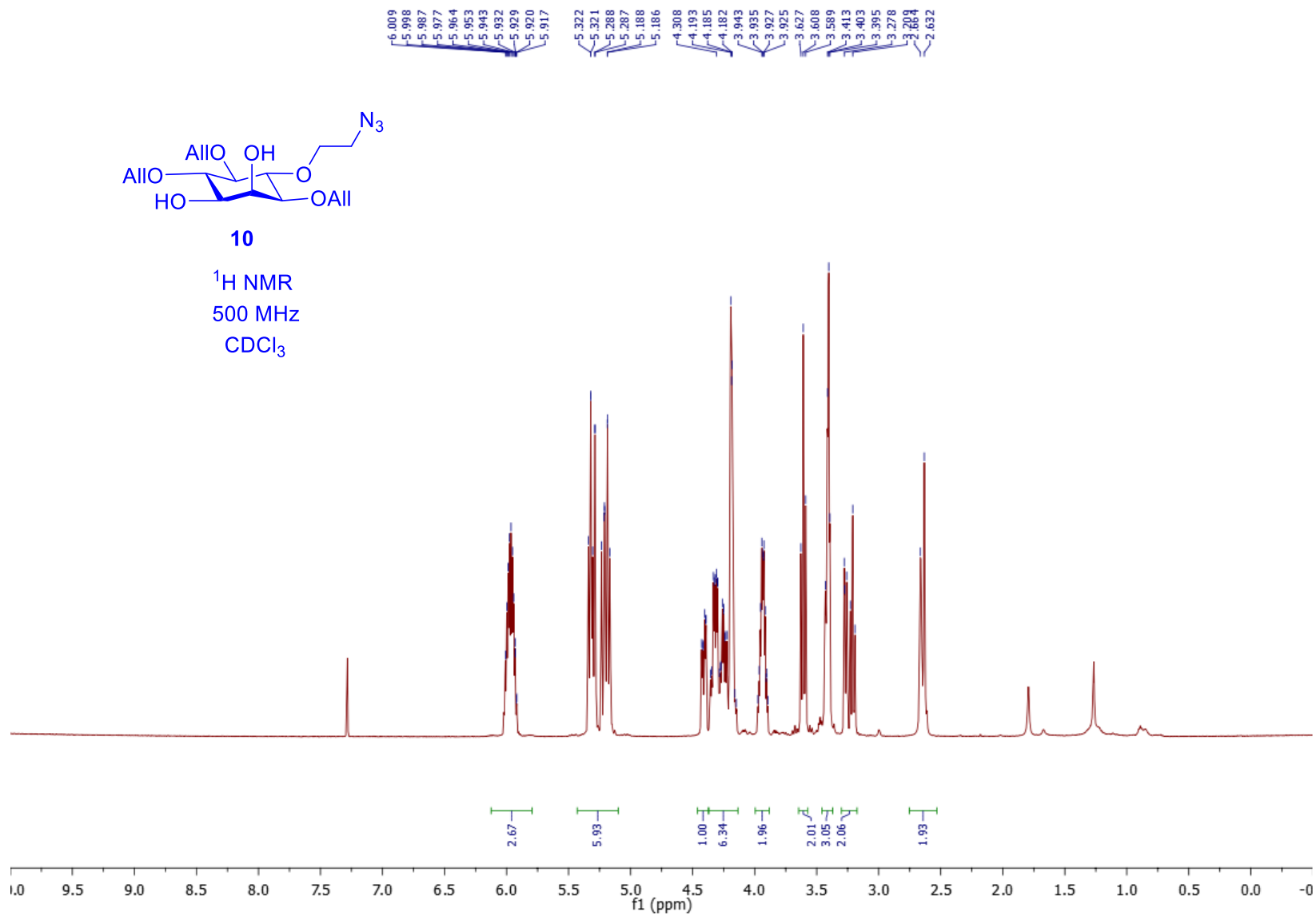


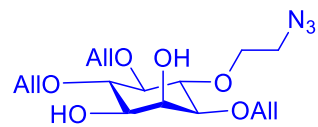




10

^1H NMR
500 MHz
 CDCl_3



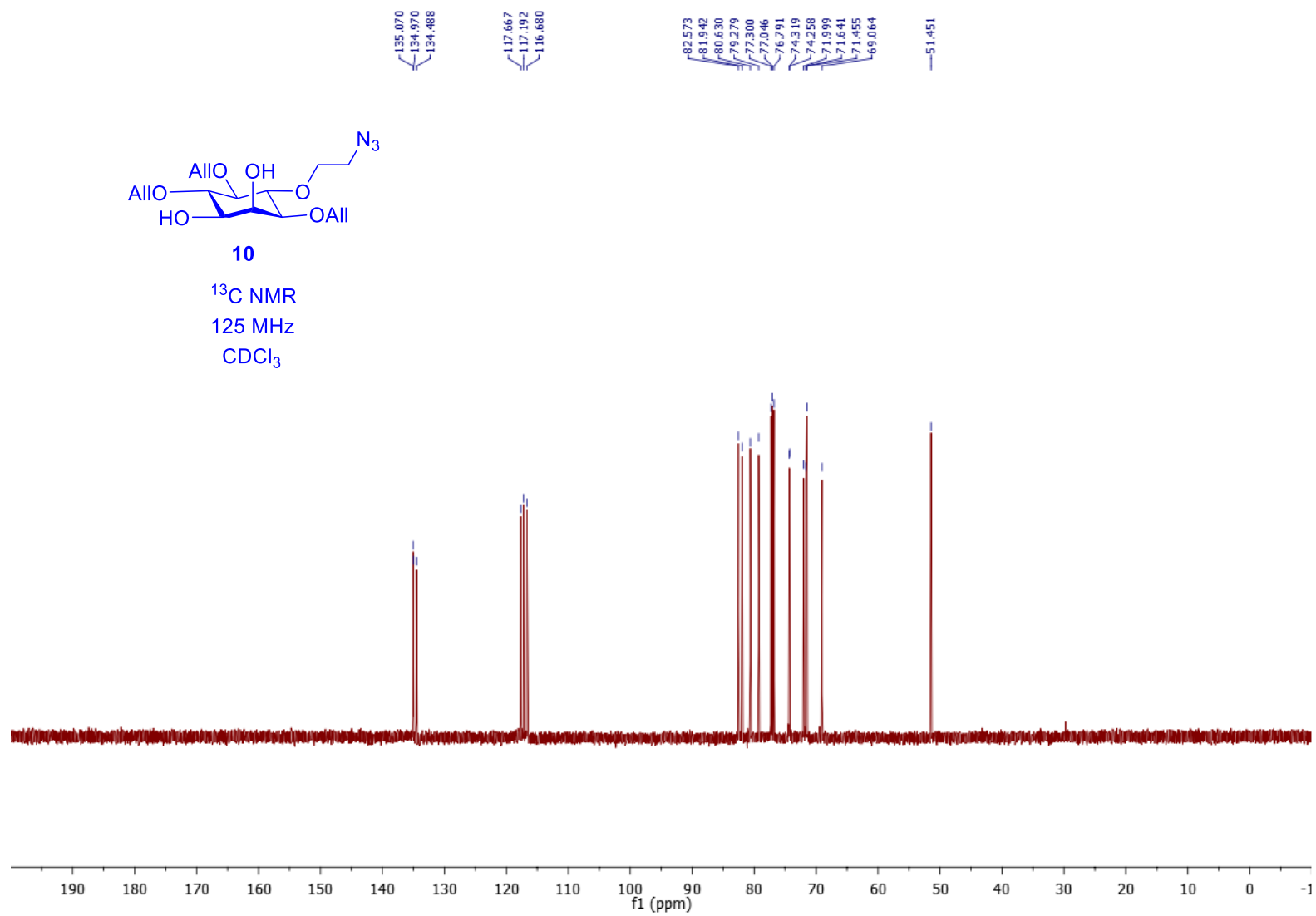


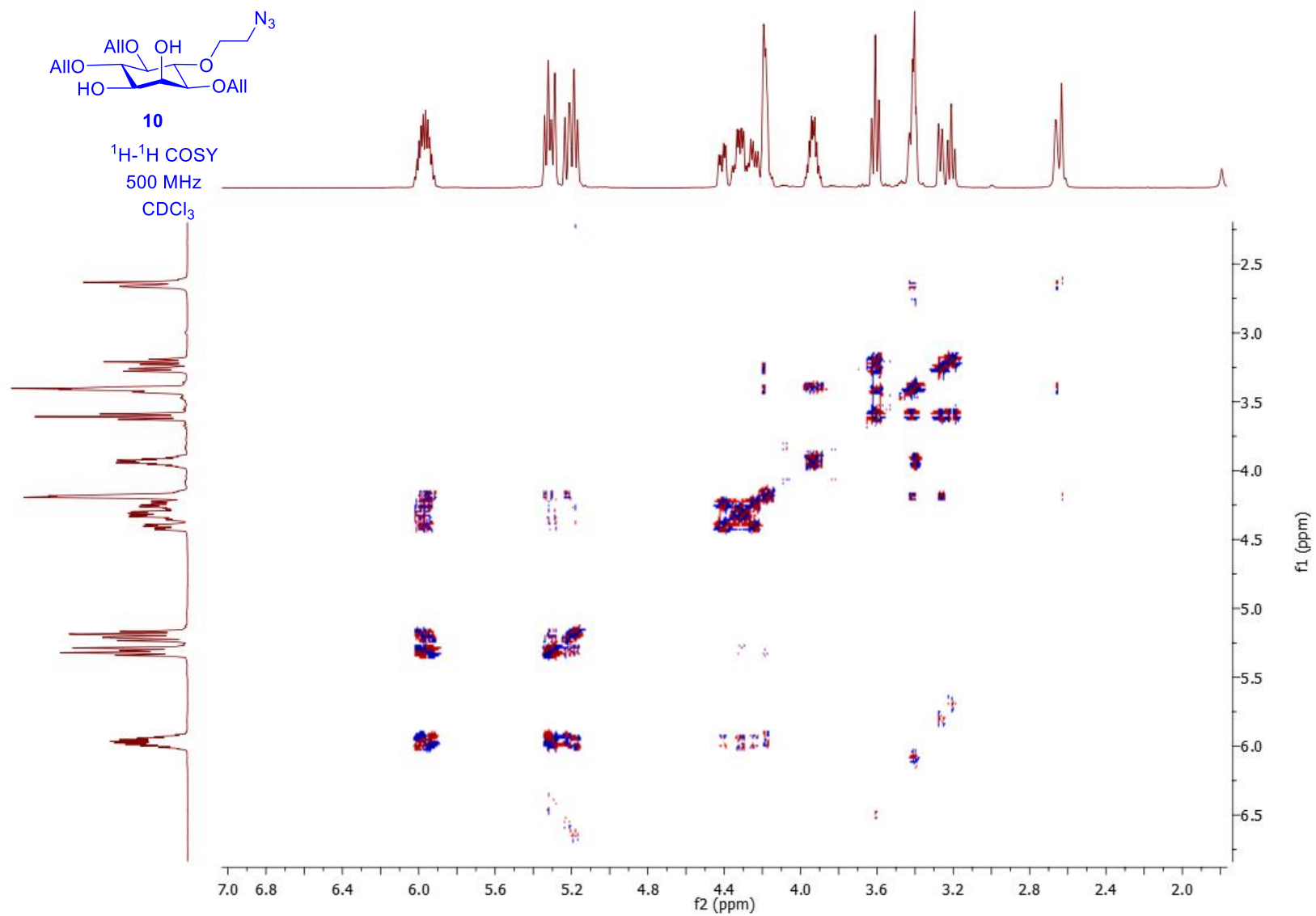
10

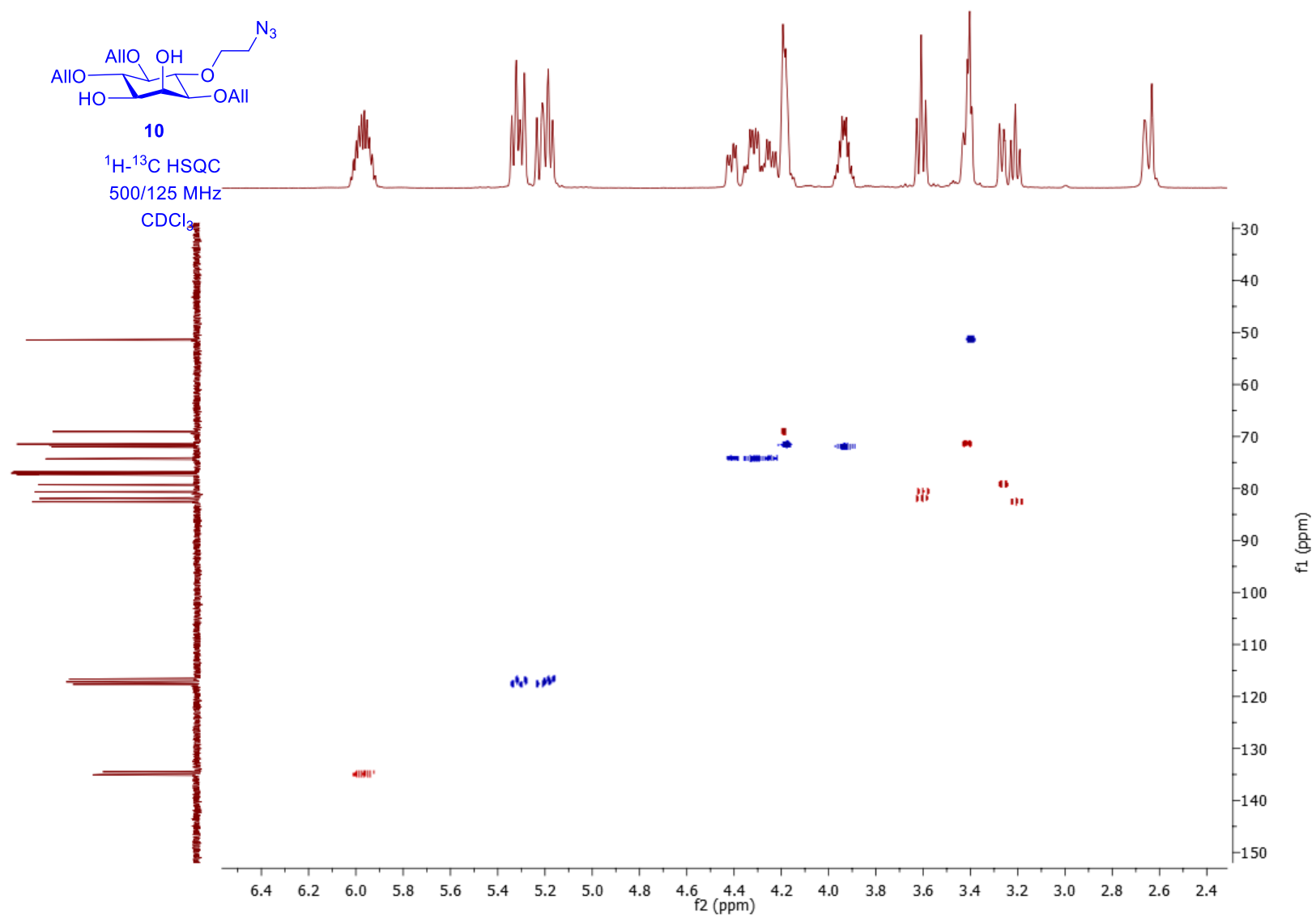
^{13}C NMR

125 MHz

CDCl_3

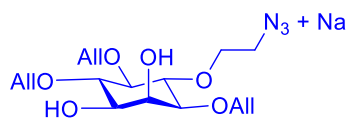






x10⁴ +ESI Scan (0.332-0.664 min, 41 Scans) Frag=175.0V 28120_zsy-2-175_ESI.d Subtract

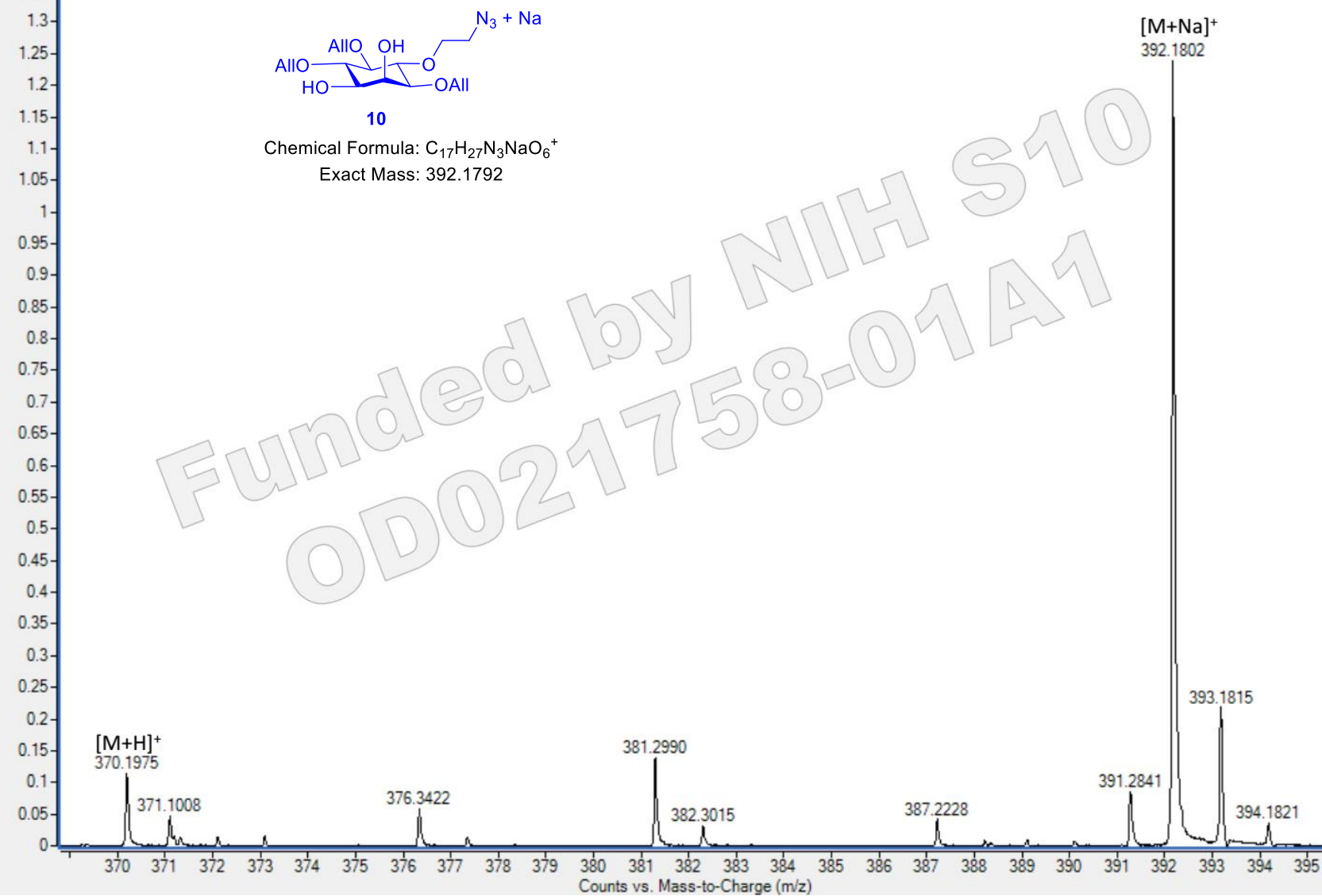
Theoretical [M+H]⁺ = 370.1973 (0.5 ppm)
[M+Na]⁺ = 392.1792 (2.5 ppm)

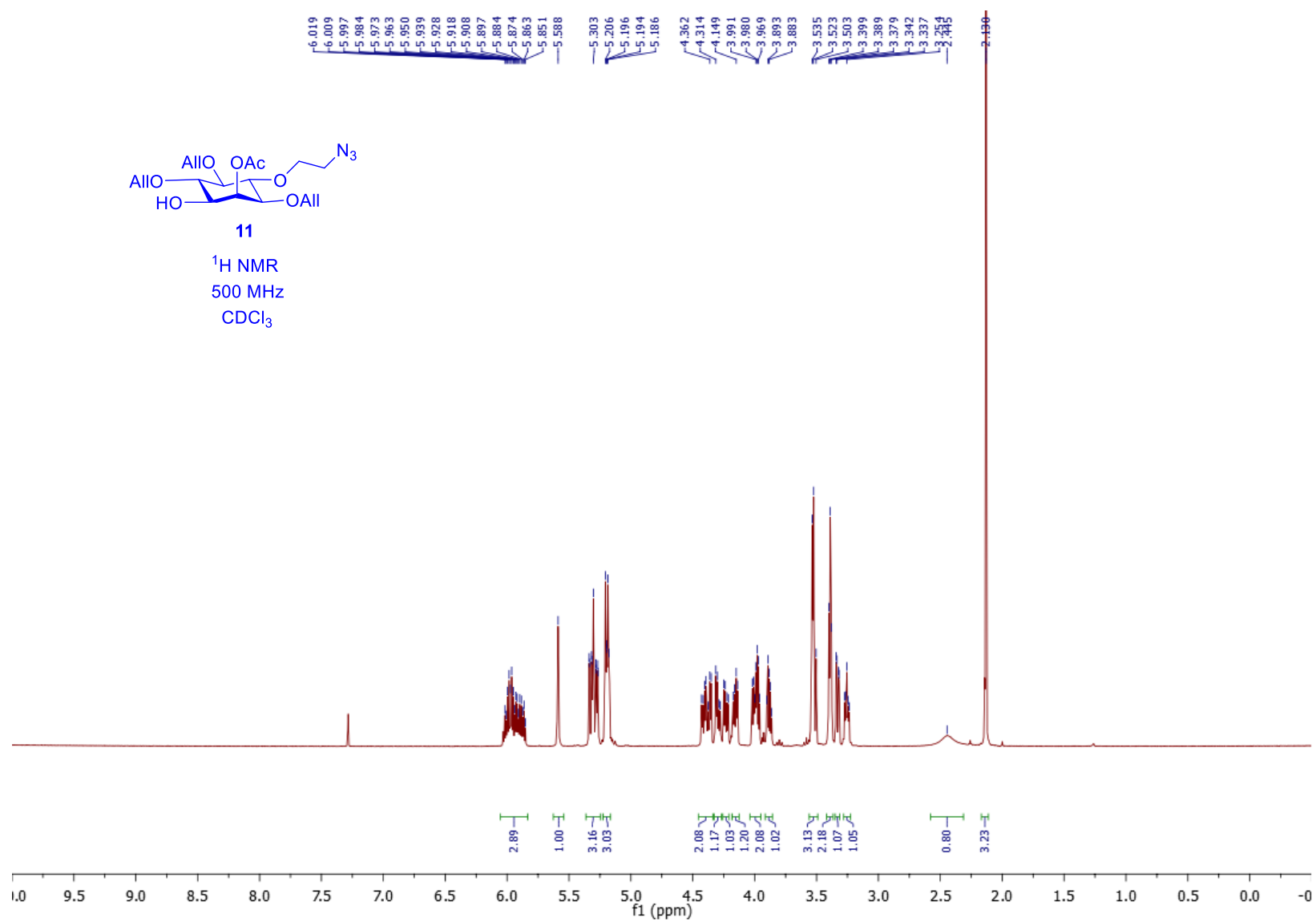


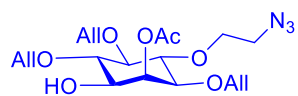
10

Chemical Formula: C₁₇H₂₇N₃NaO₆⁺

Exact Mass: 392.1792





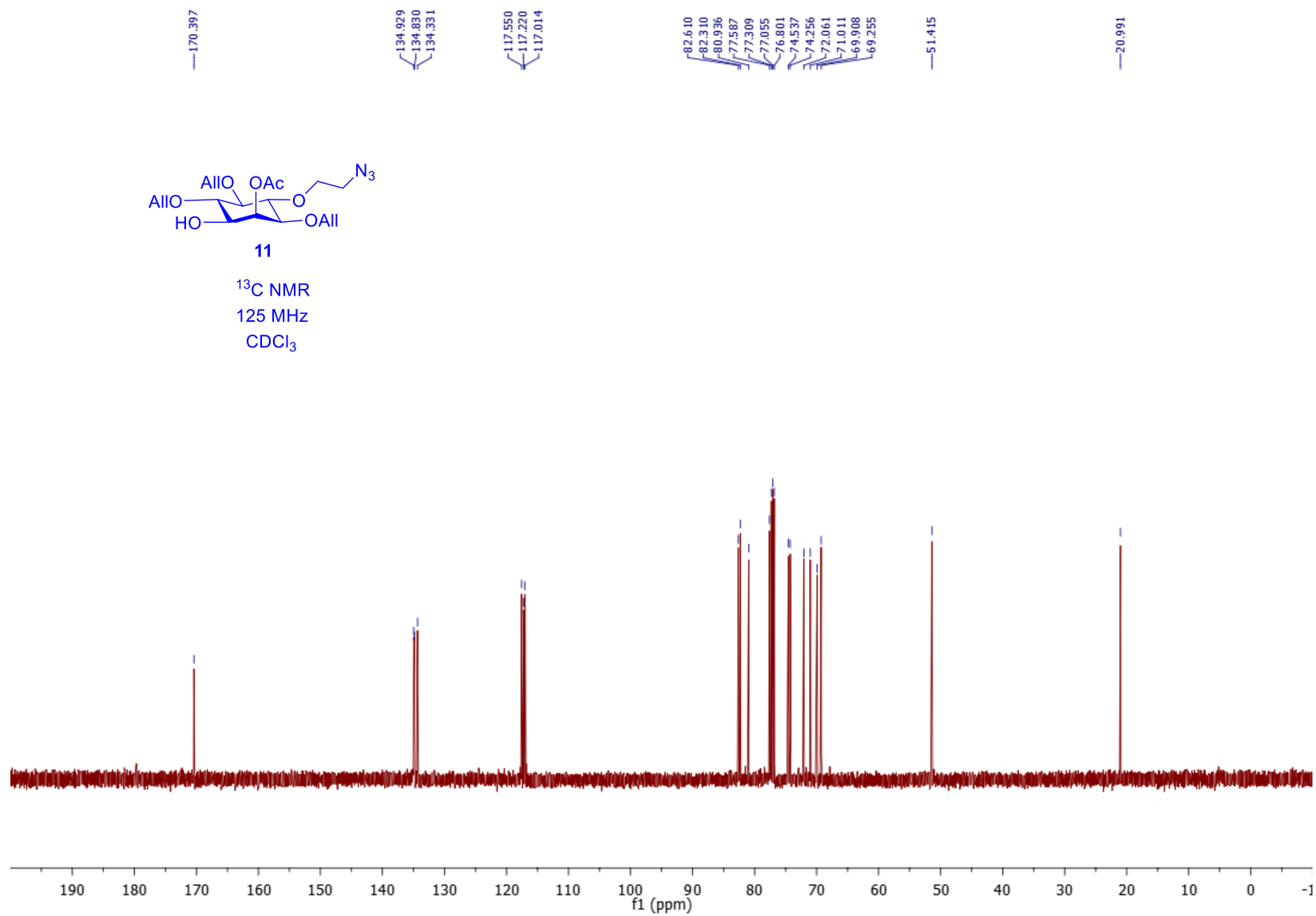


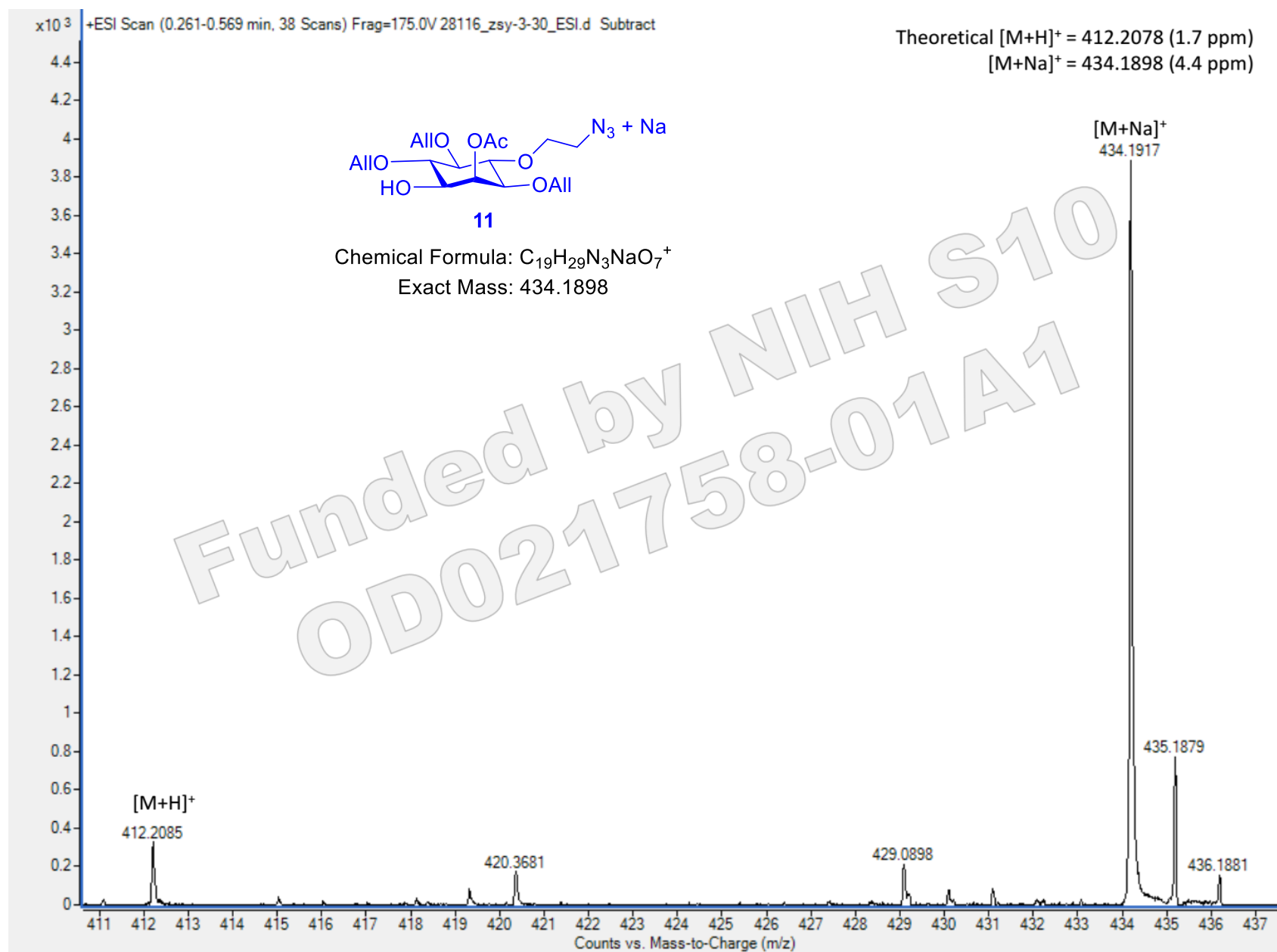
11

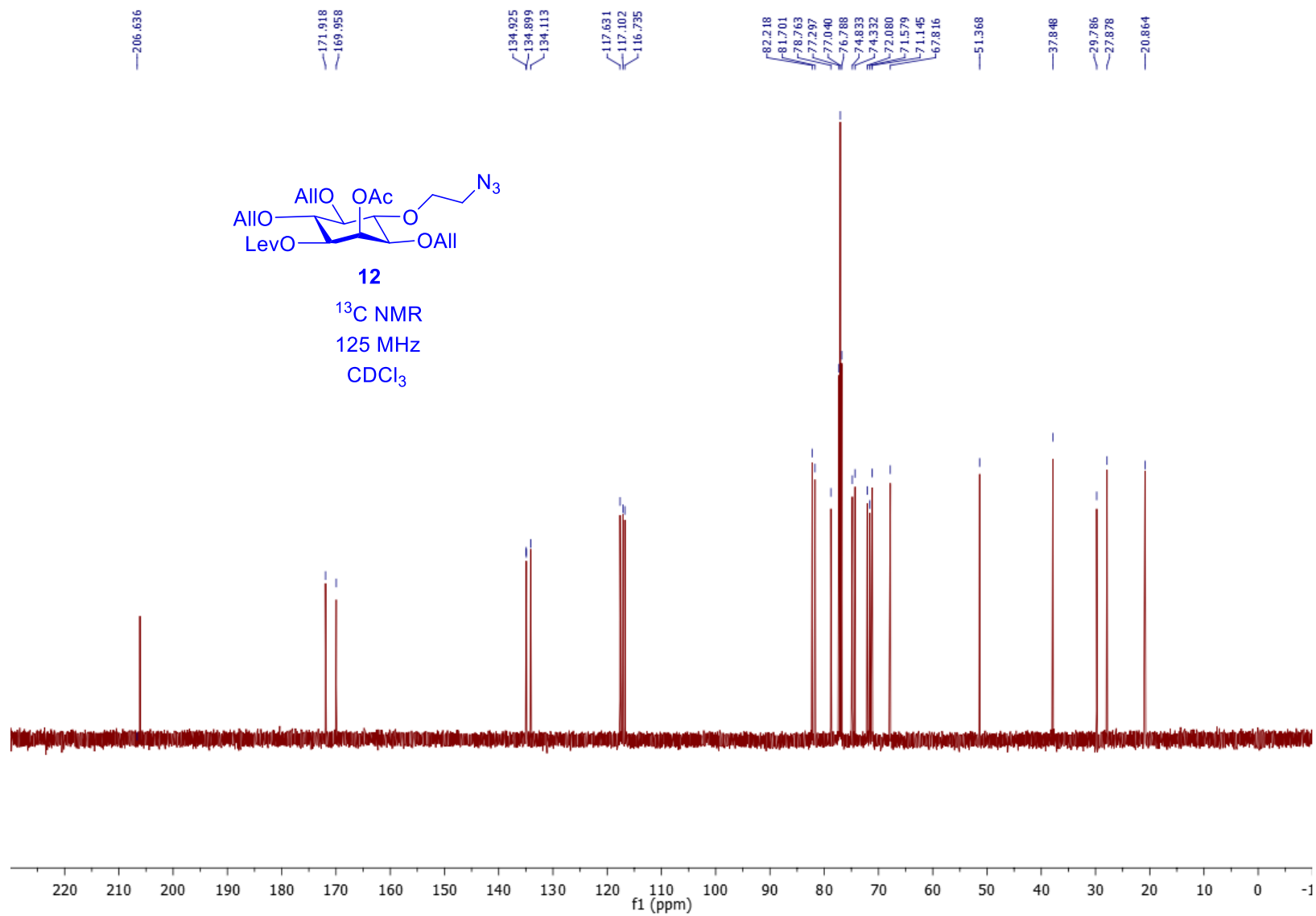
¹³C NMR

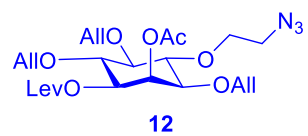
125 MHz

CDCl₃





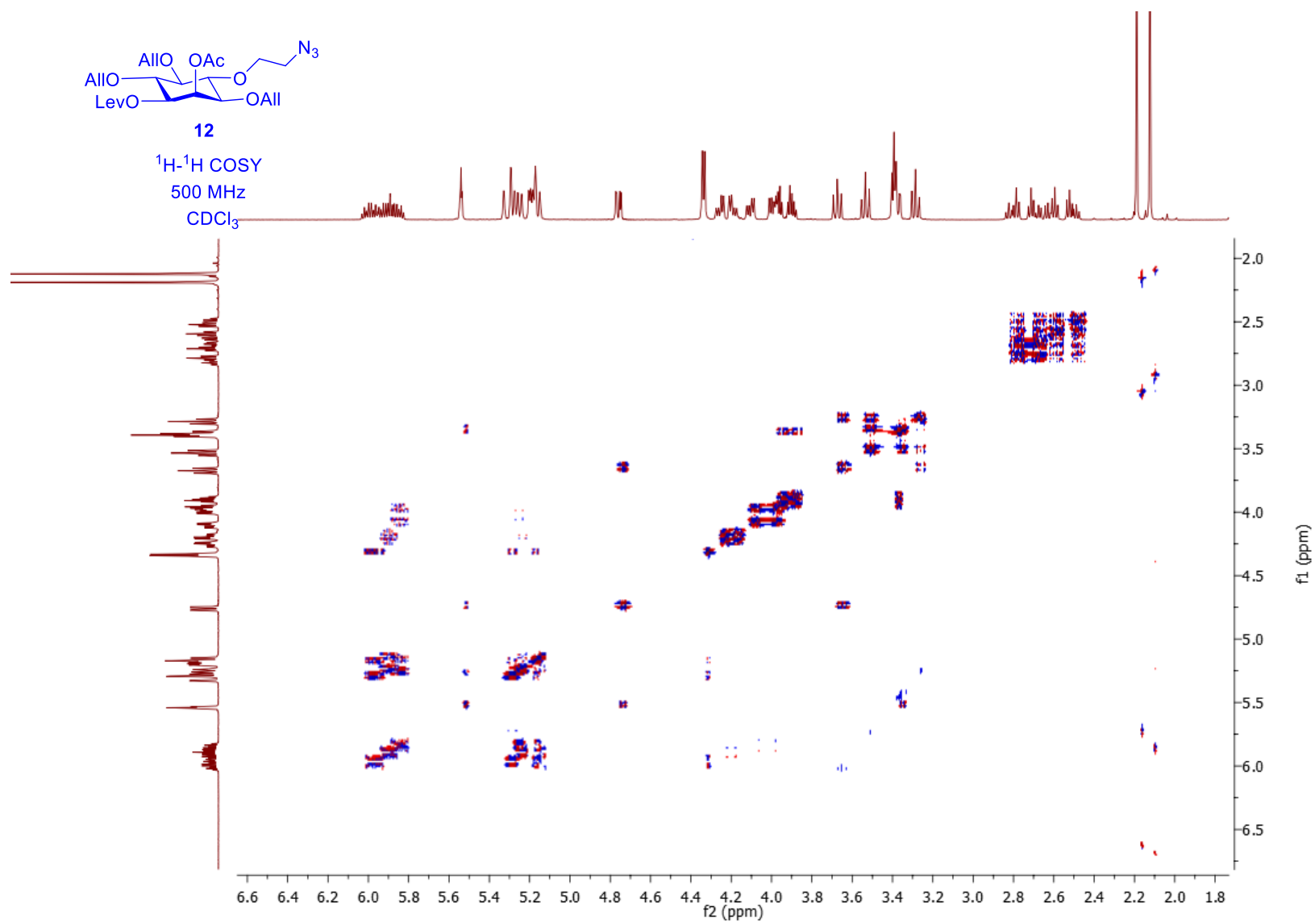


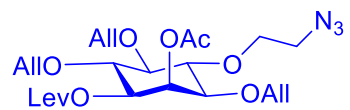


12

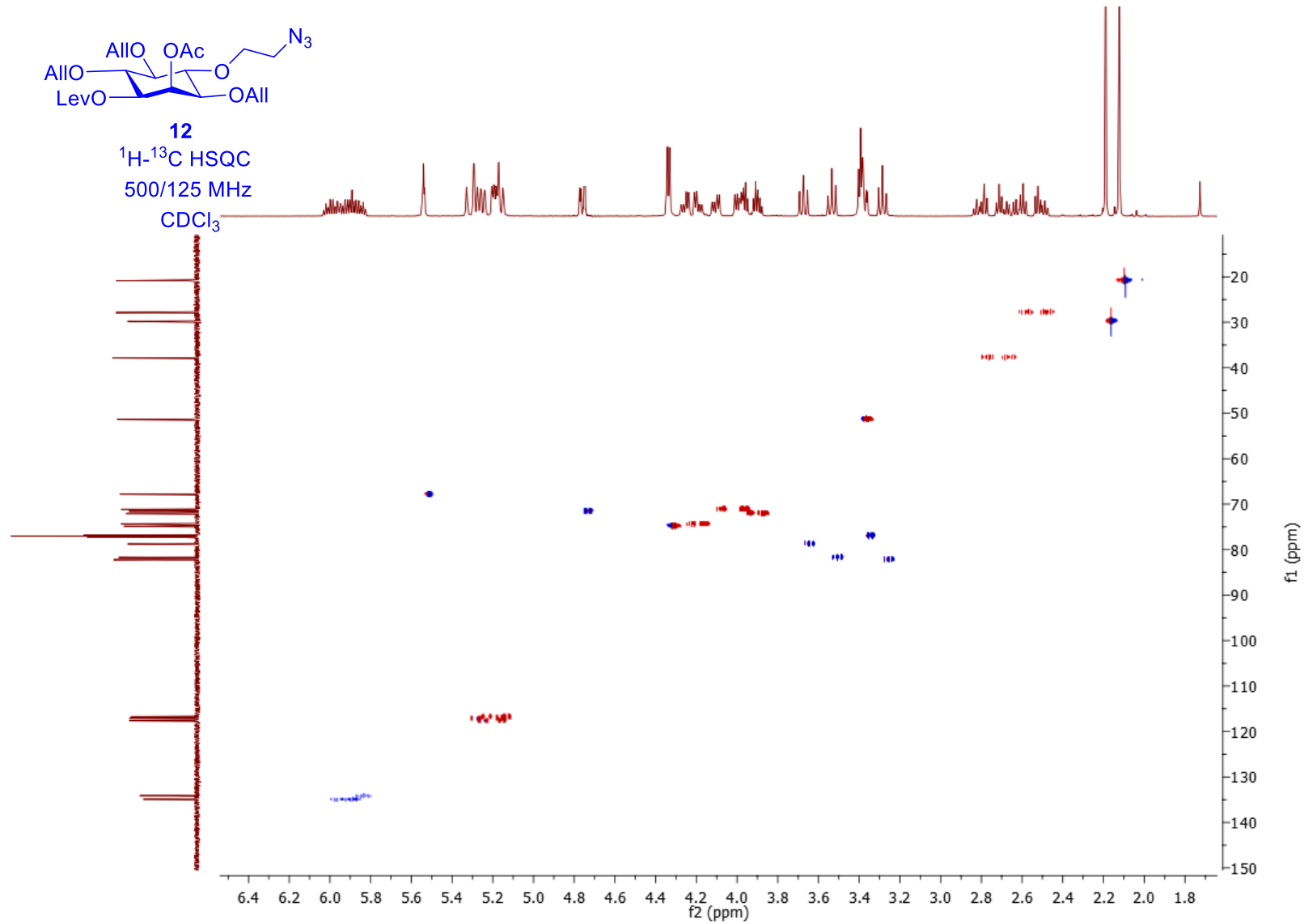
 ^1H - ^1H COSY

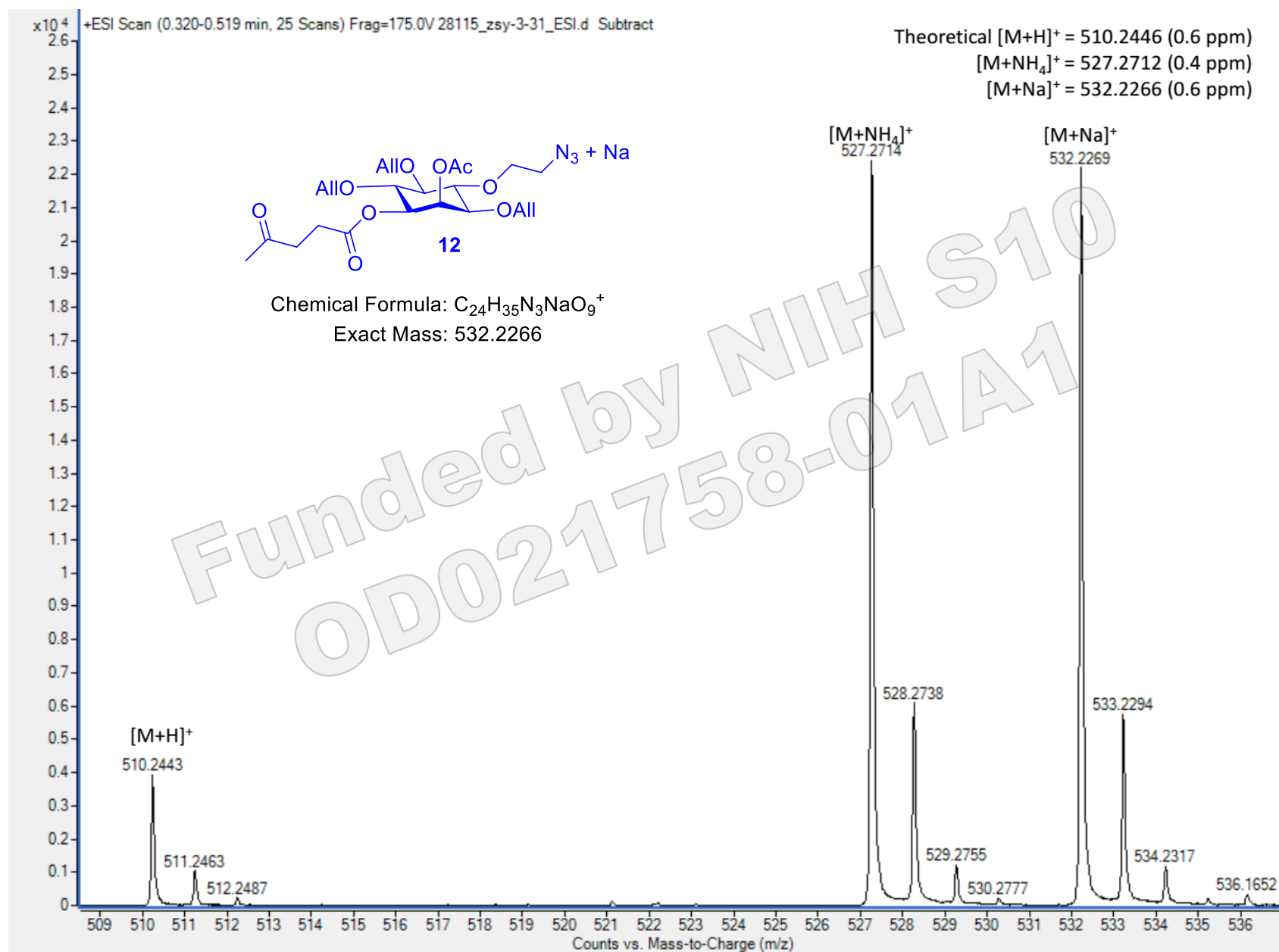
500 MHz

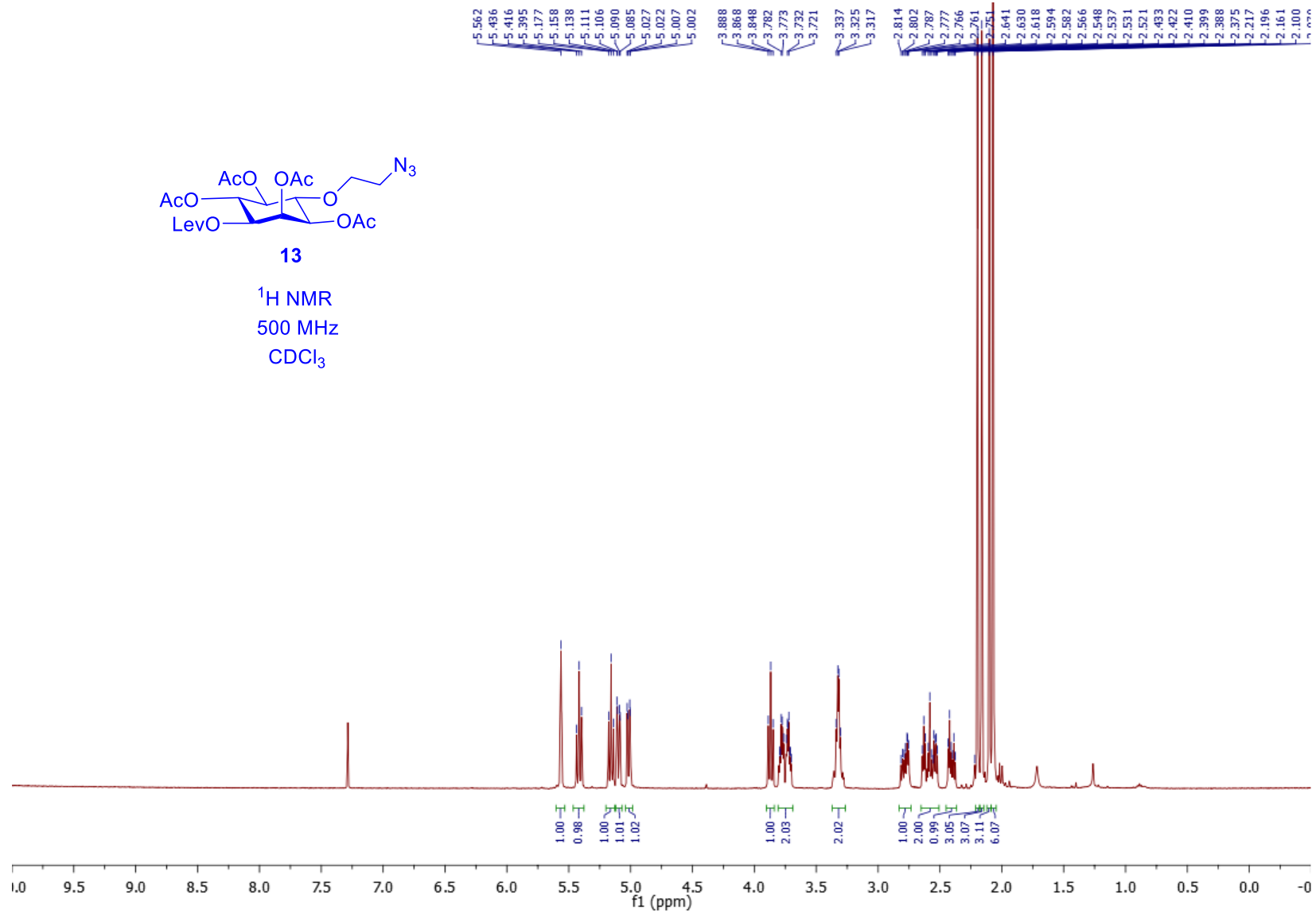
 CDCl_3 

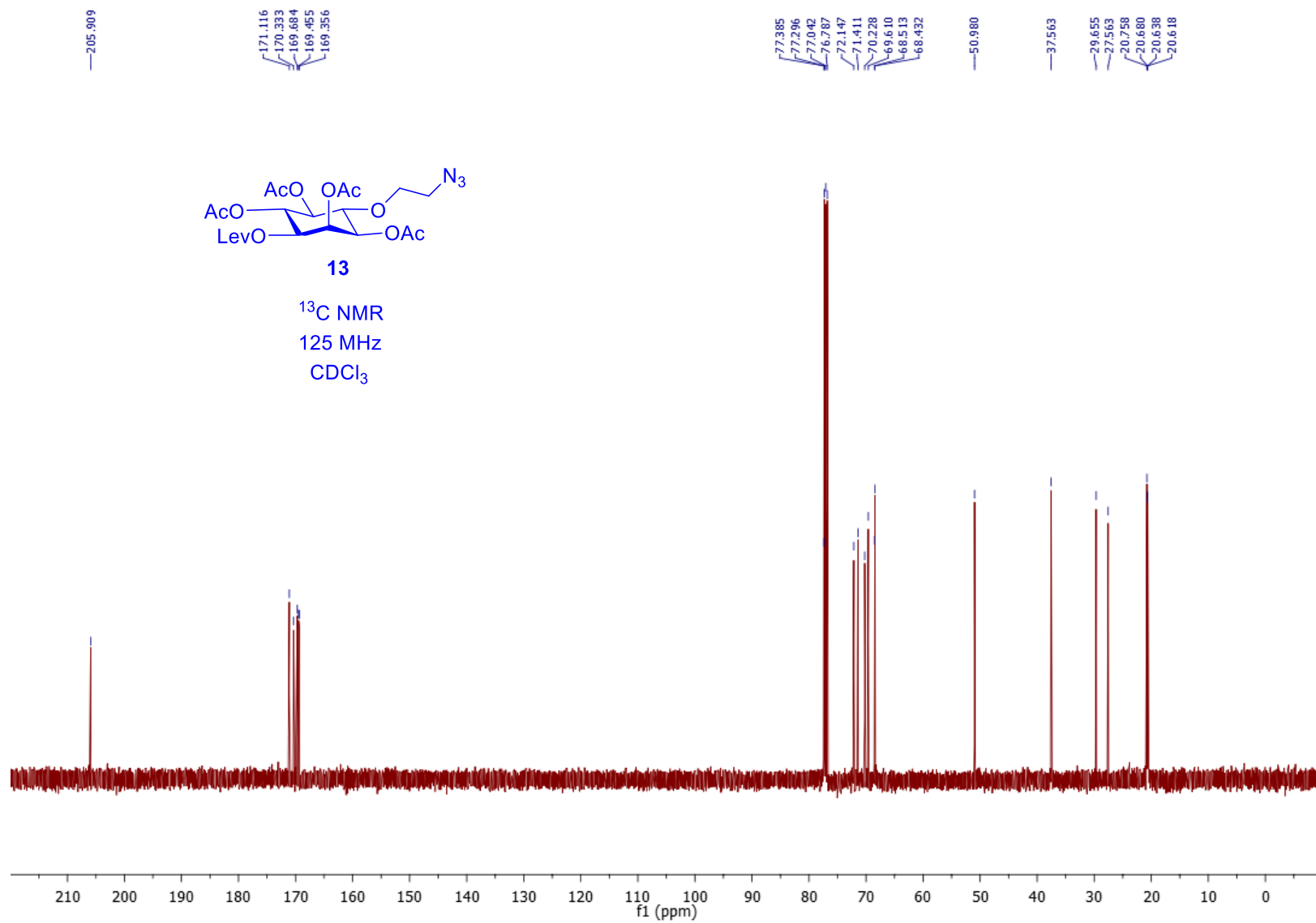


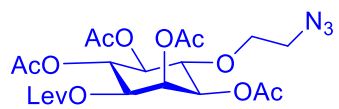
12
¹H-¹³C HSQC
 500/125 MHz
 CDCl₃









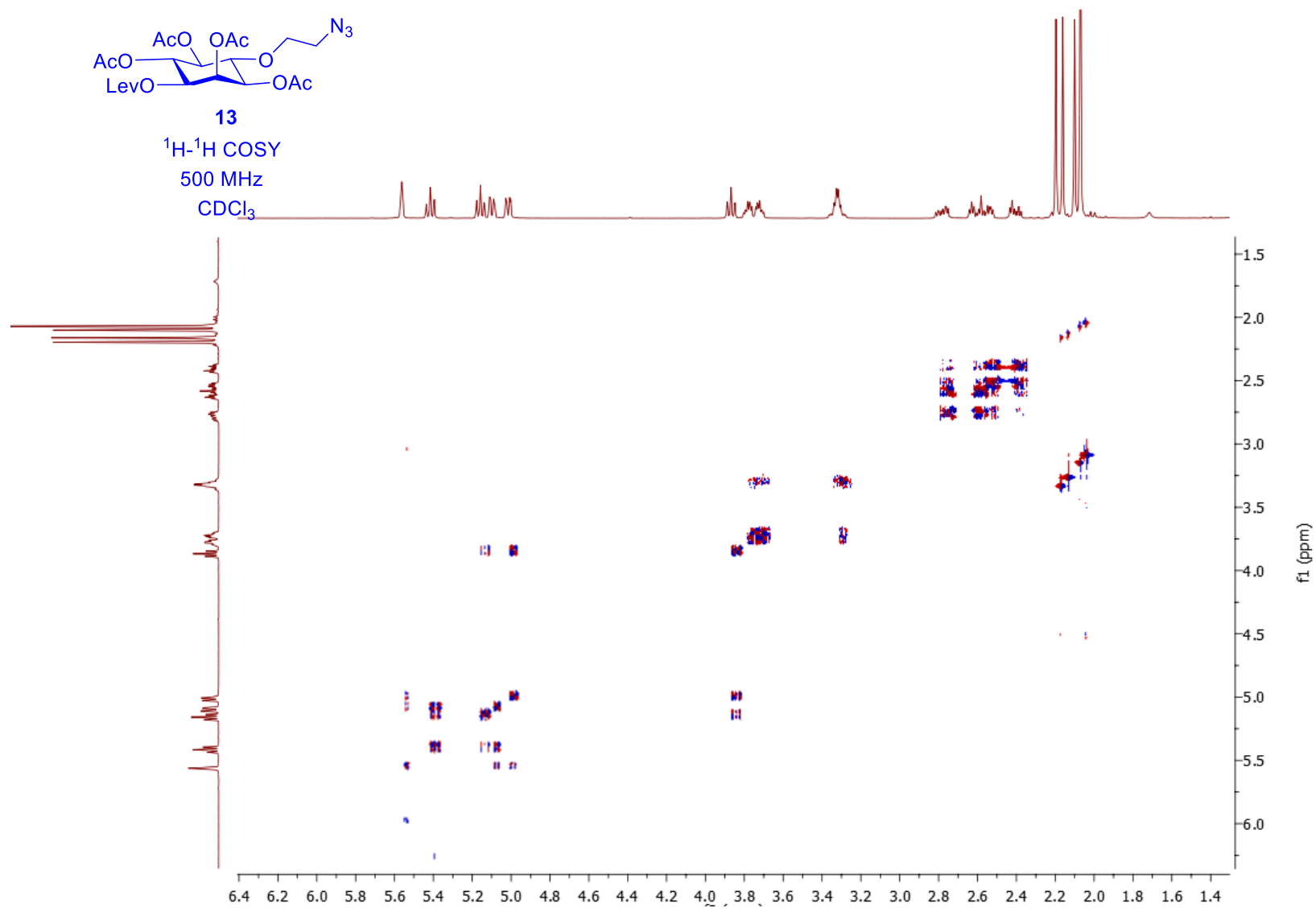


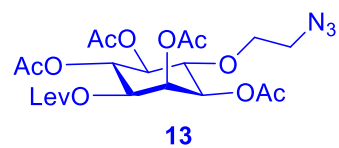
13

^1H - ^1H COSY

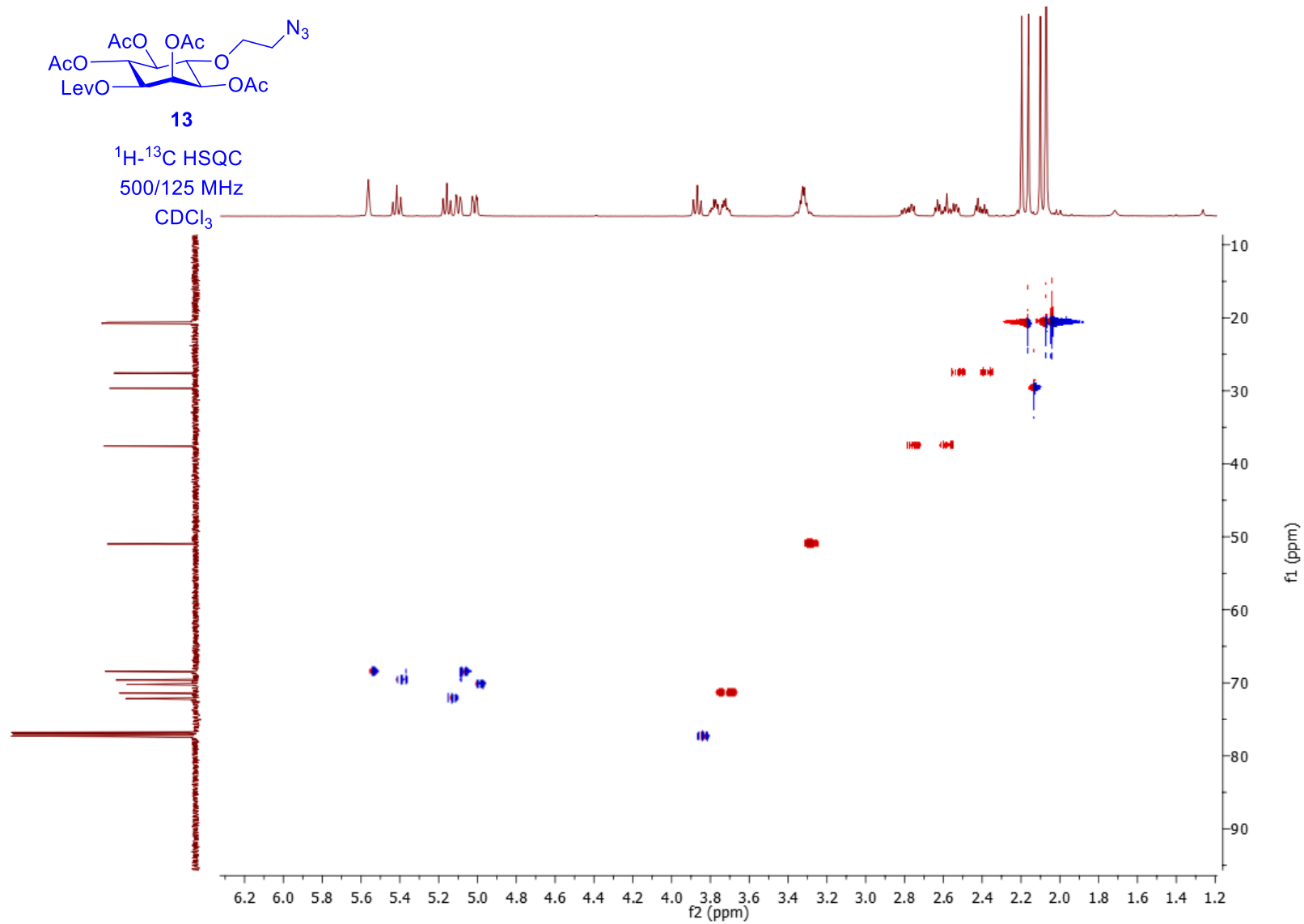
500 MHz

CDCl_3





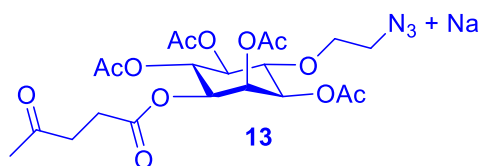
^1H - ^{13}C HSQC
 500/125 MHz
 CDCl_3



x10⁴ +ESI Scan (0.282-0.673 min, 48 Scans) Frag=175.0V 28145_zsy-3-33_ESI.d Subtract

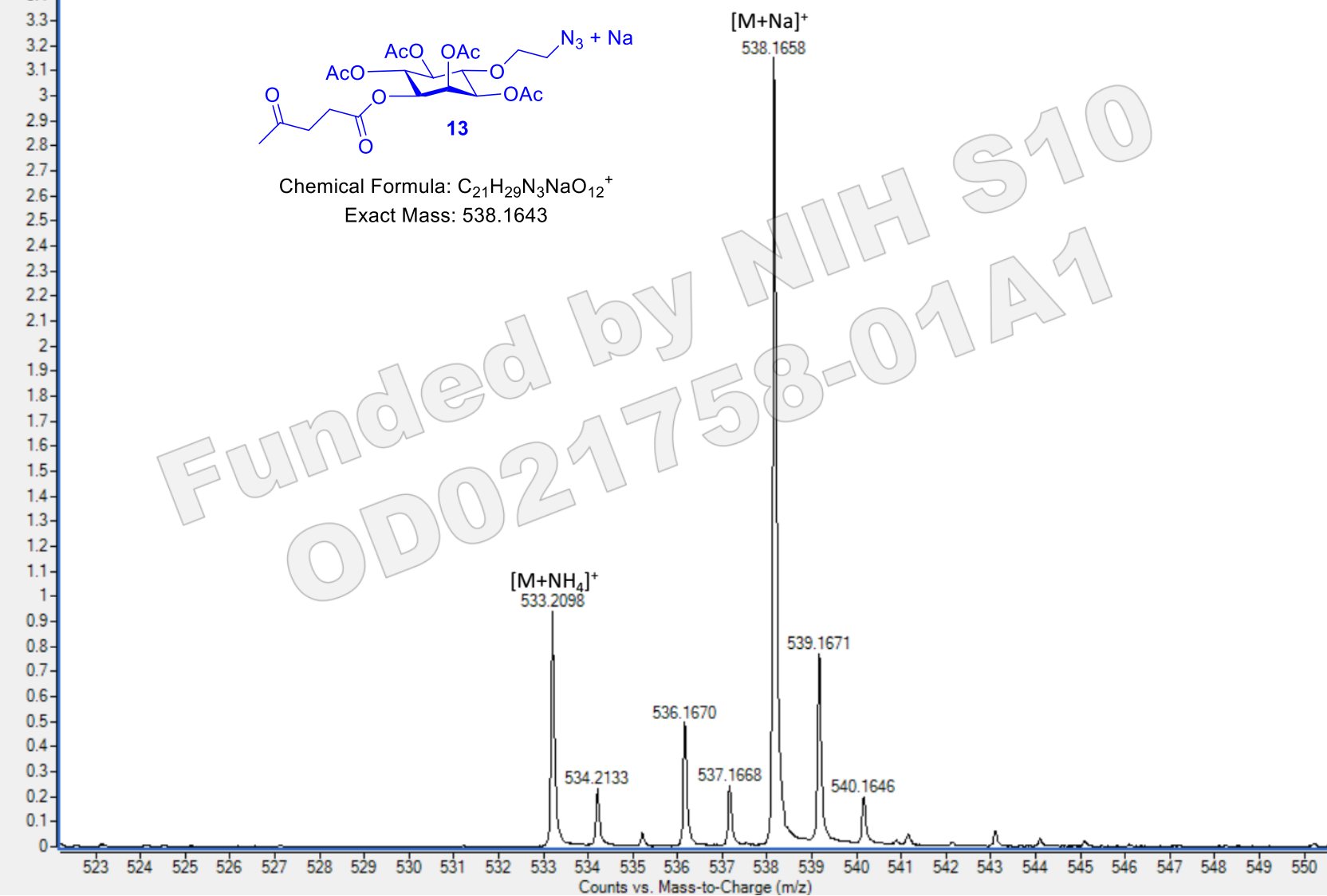
Theoretical $[M+NH_4]^+ = 533.2089$ (1.7 ppm)

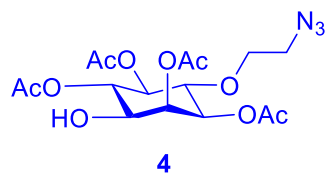
$[M+Na]^+ = 538.1643$ (2.8 ppm)



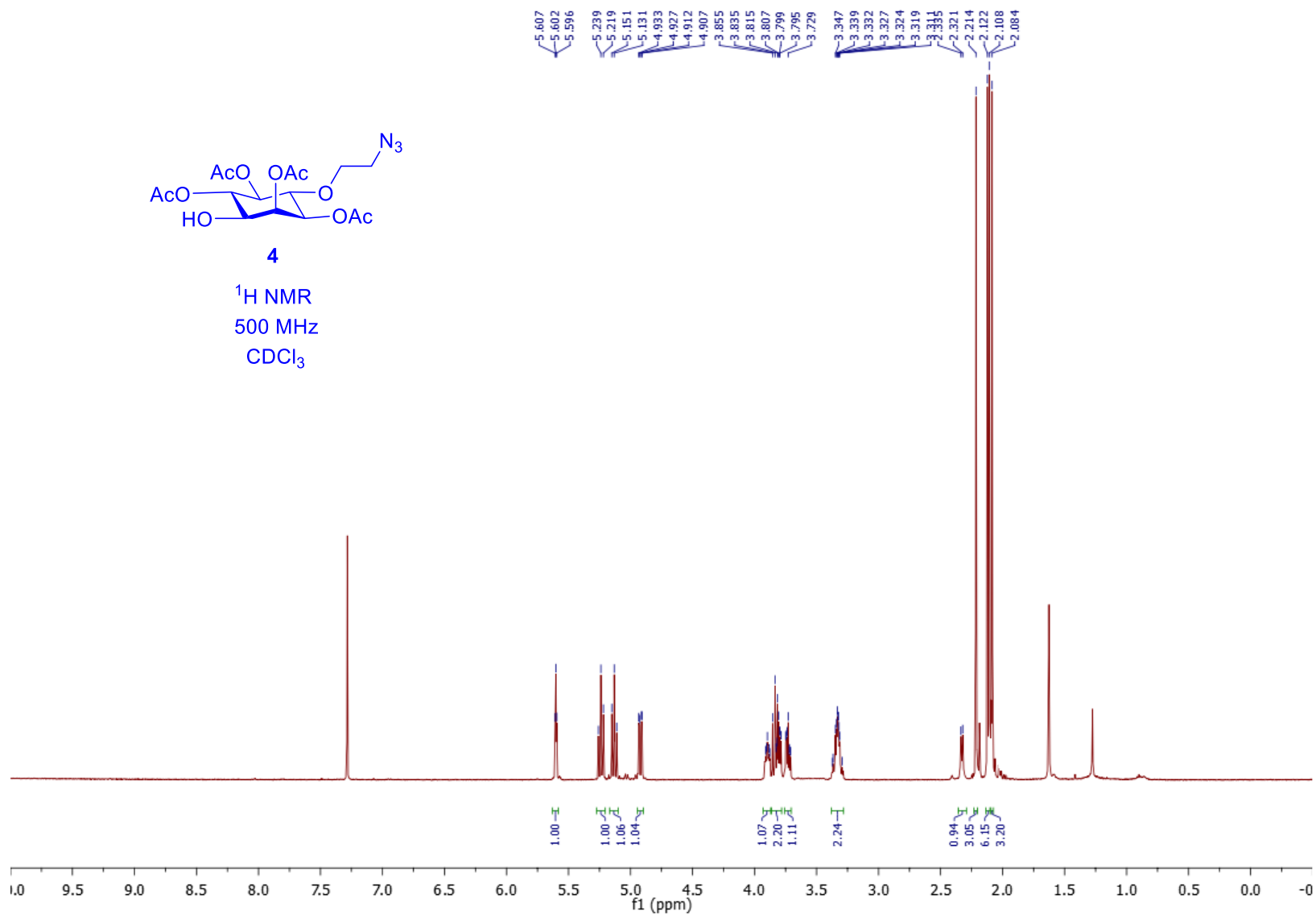
Chemical Formula: C₂₁H₂₉N₃NaO₁₂⁺

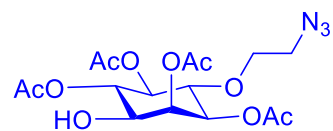
Exact Mass: 538.1643





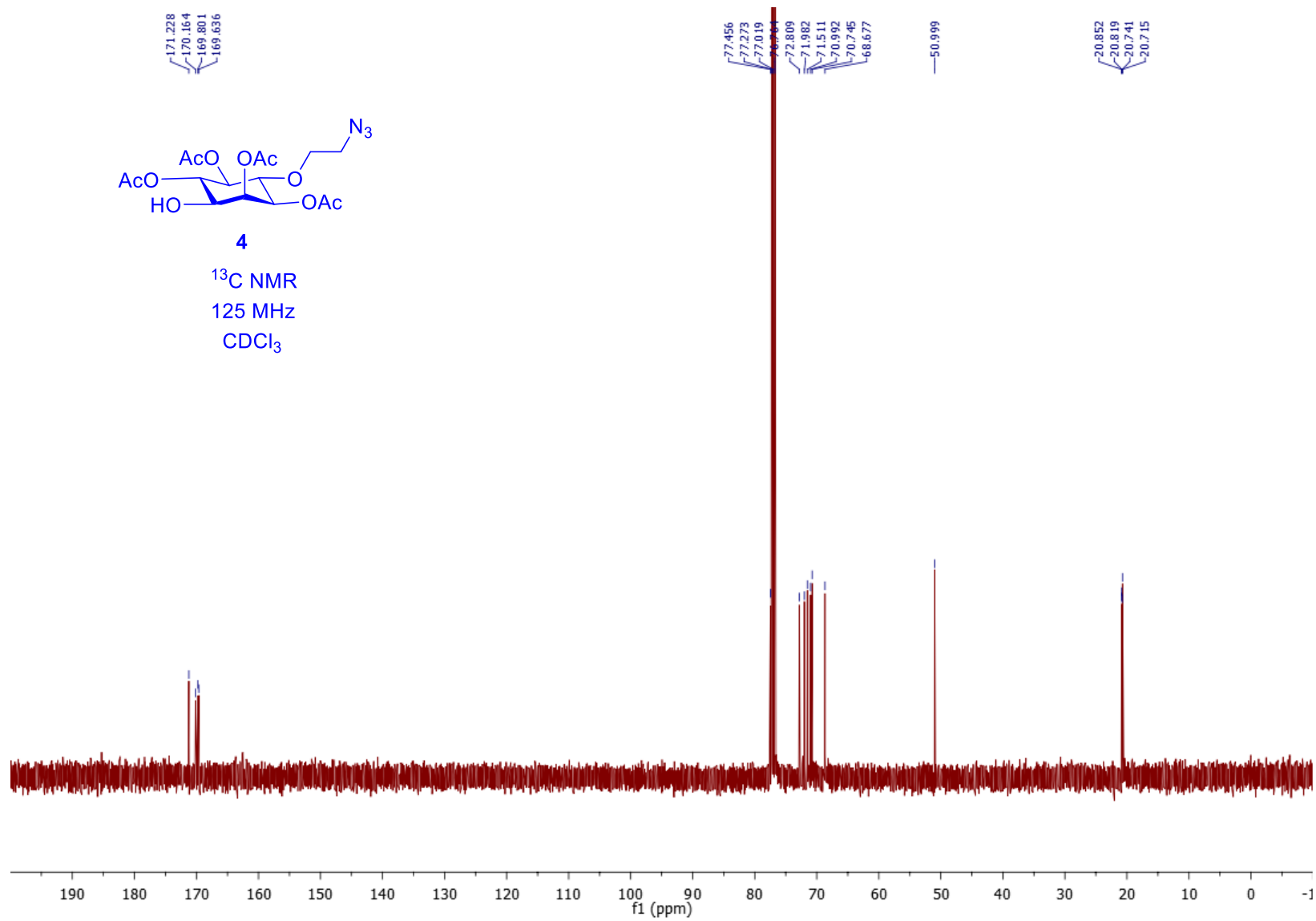
¹H NMR
 500 MHz
 CDCl₃

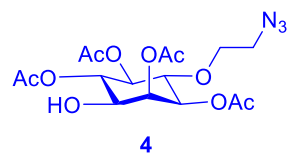




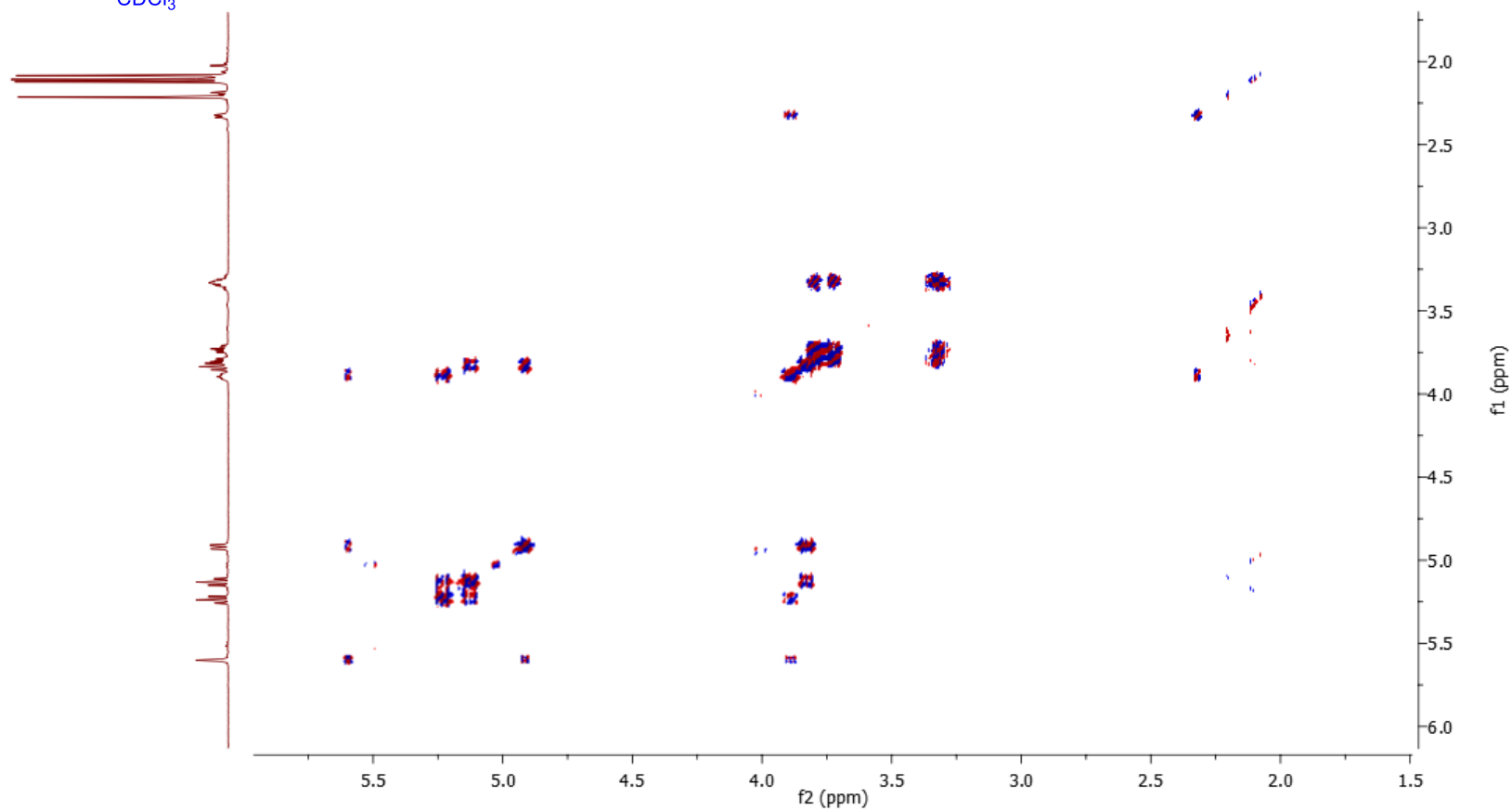
4

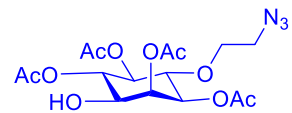
^{13}C NMR
125 MHz
 CDCl_3





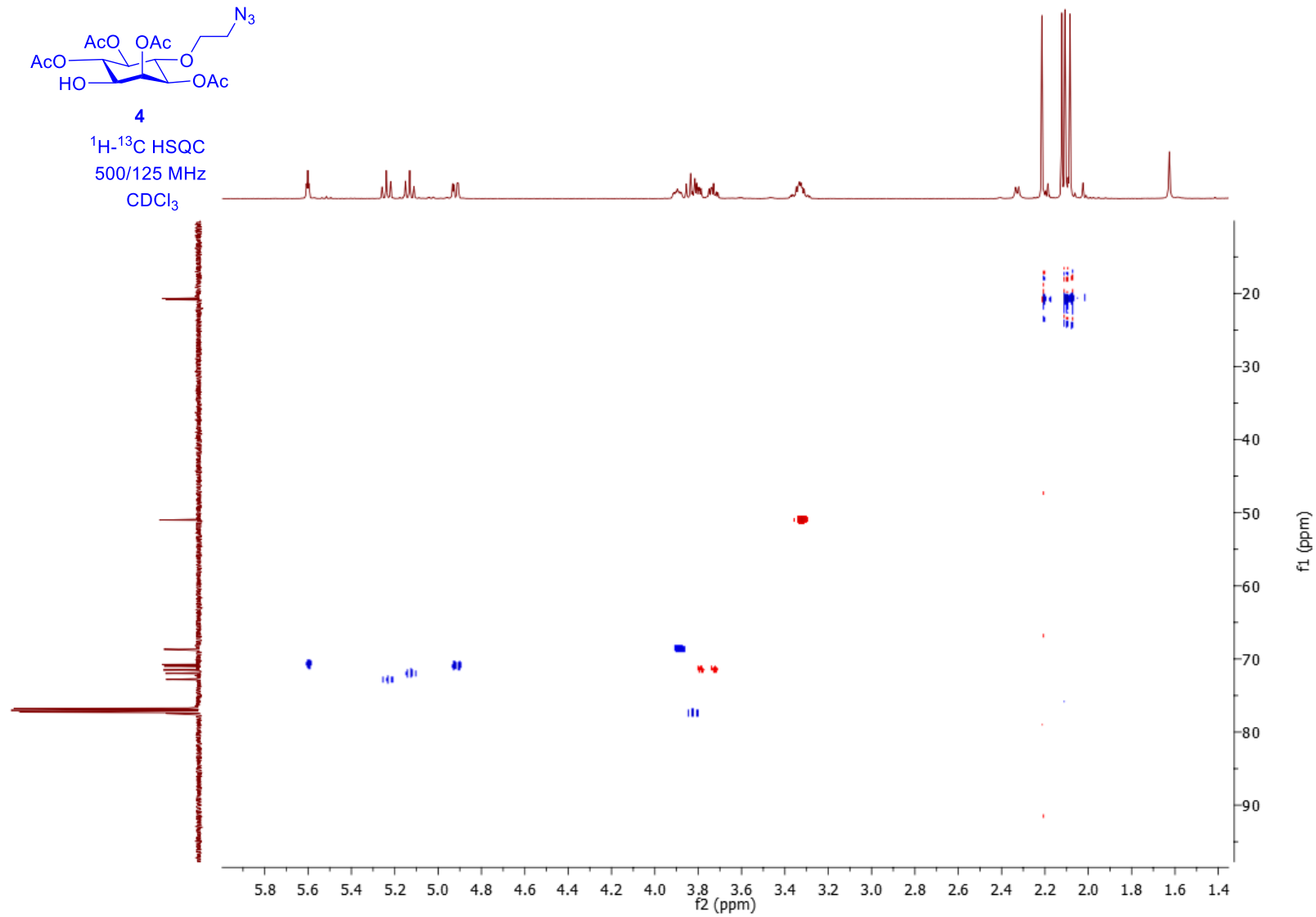
^1H - ^1H COSY
 500 MHz
 CDCl_3





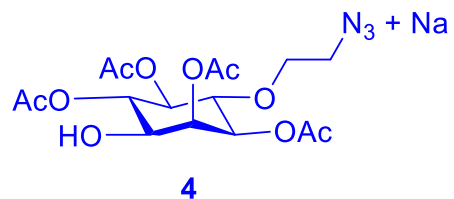
4

^1H - ^{13}C HSQC
500/125 MHz
 CDCl_3

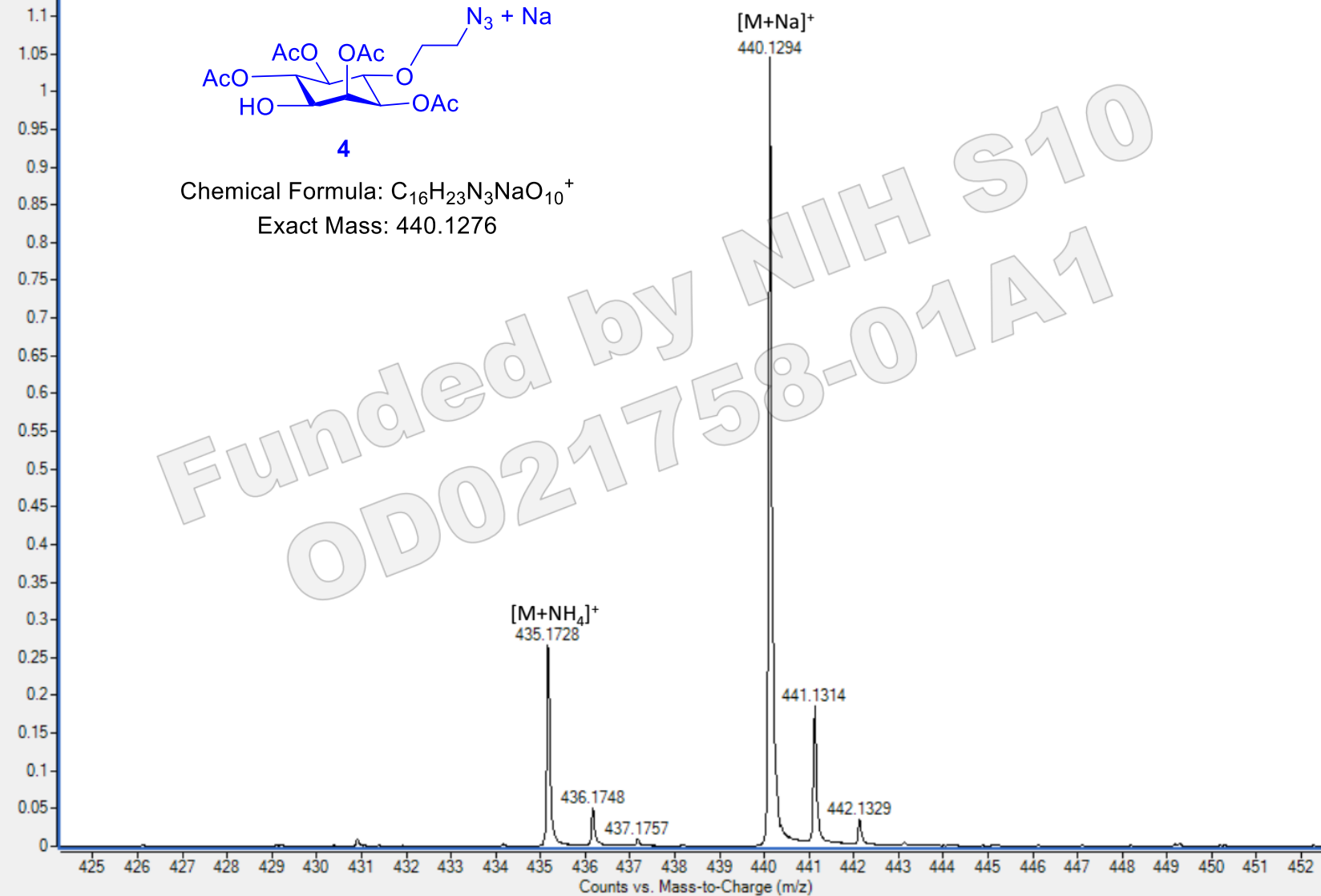


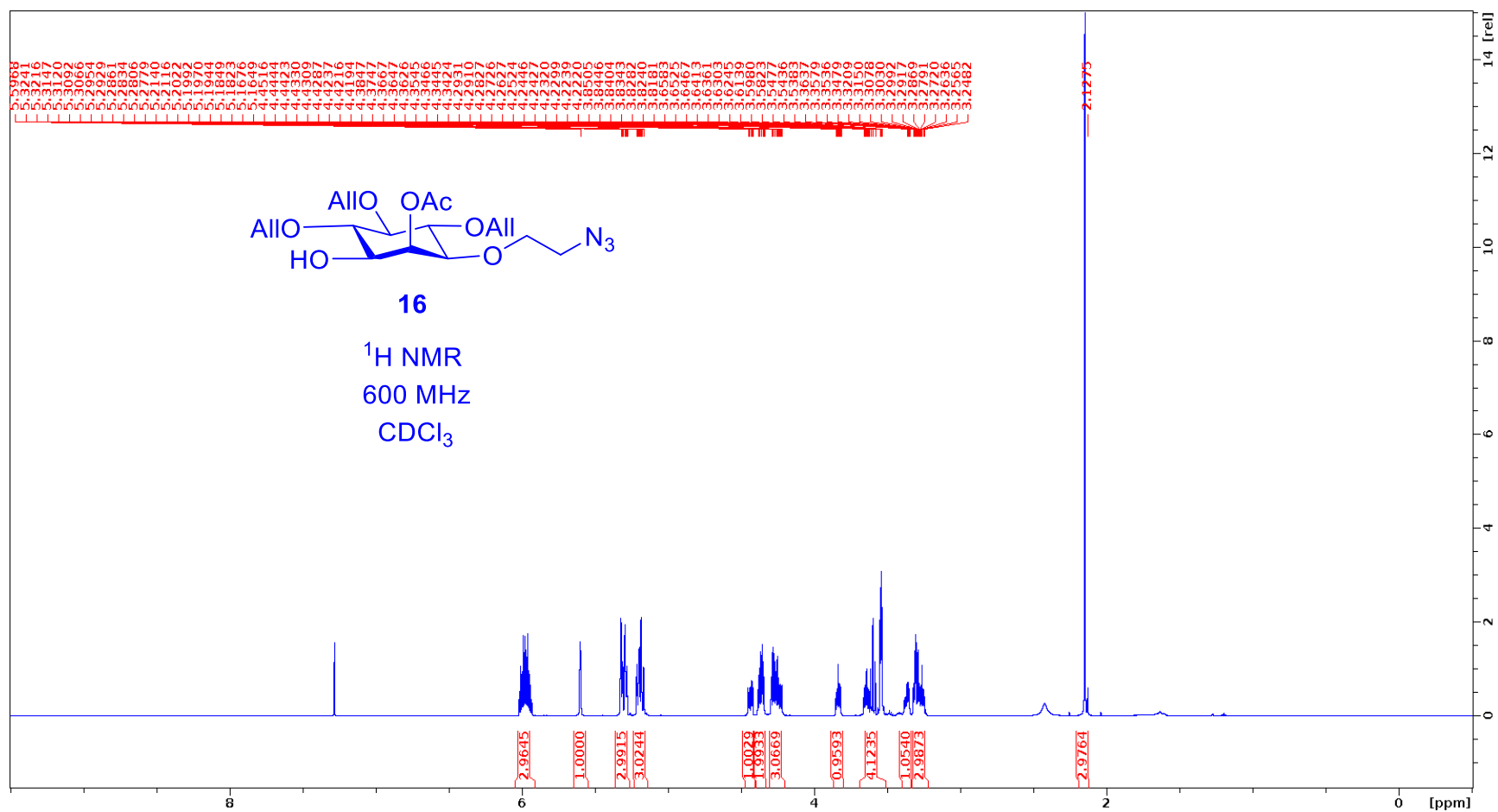
+ESI Scan (0.231-0.505 min, 34 Scans) Frag=175.0V 28155_zsy-2-184_ESI.d Subtract

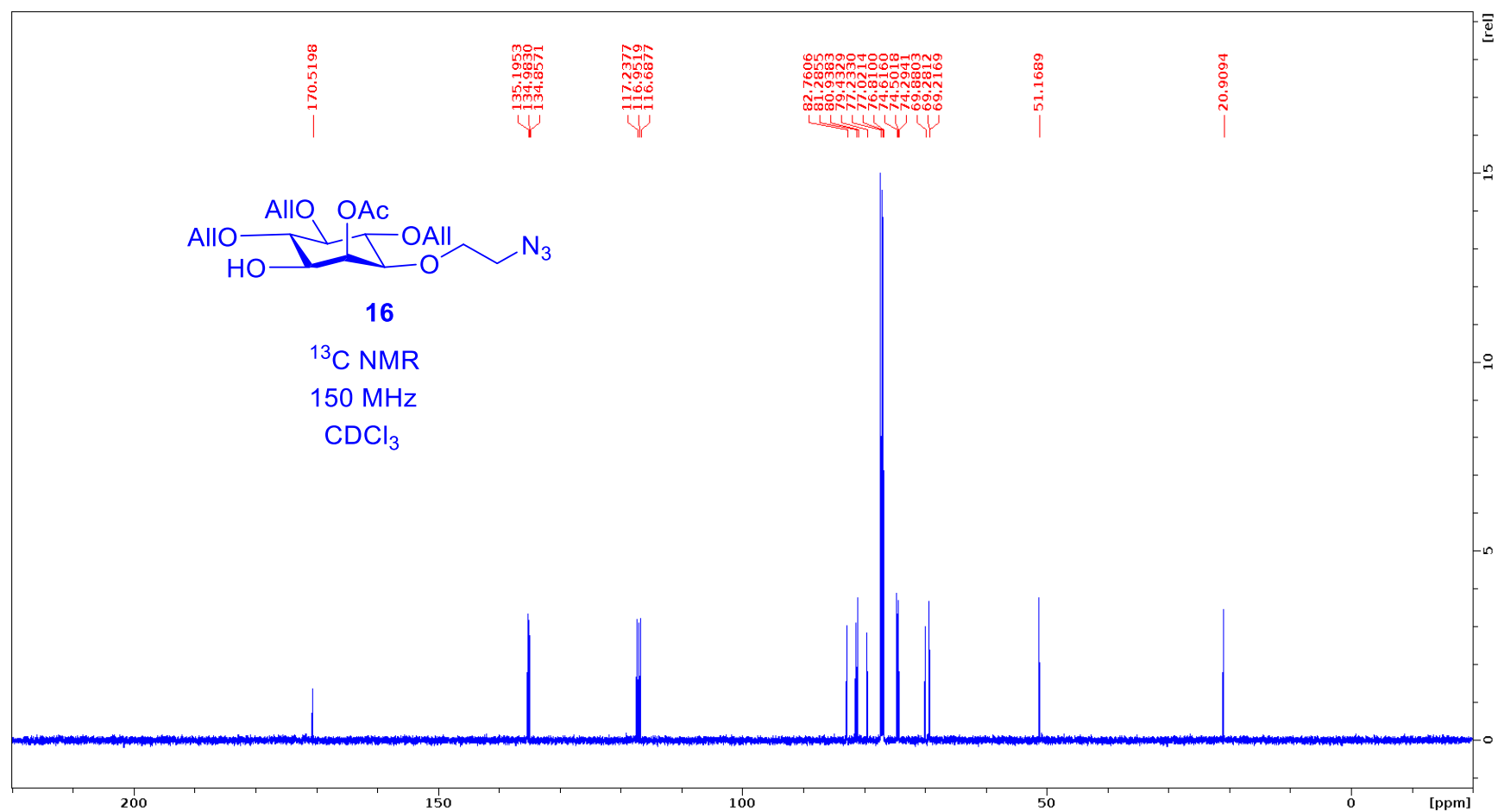
Theoretical $[M+NH_4]^+ = 435.1722$ (1.4 ppm)
 $[M+Na]^+ = 440.1276$ (4.1 ppm)

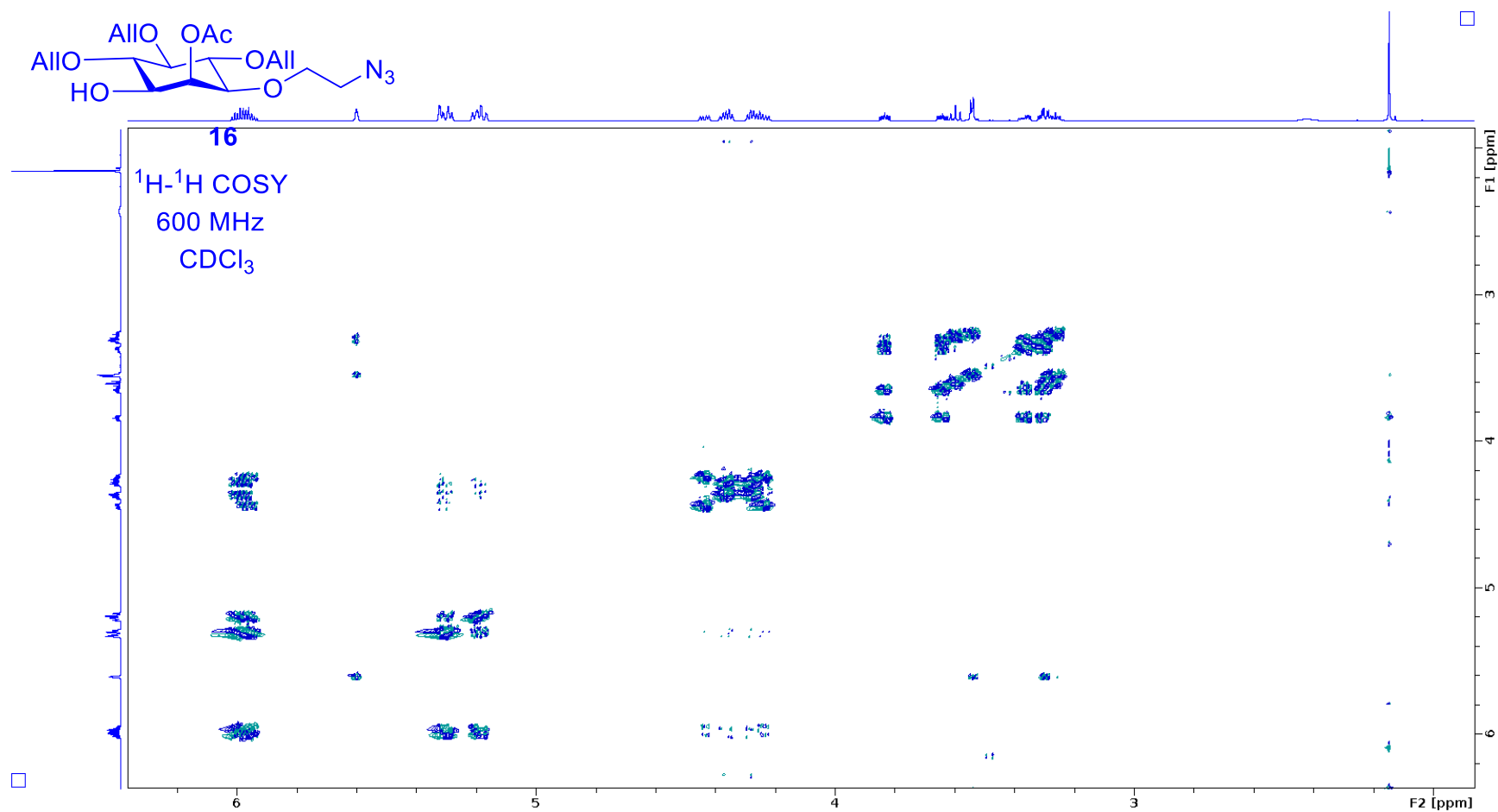


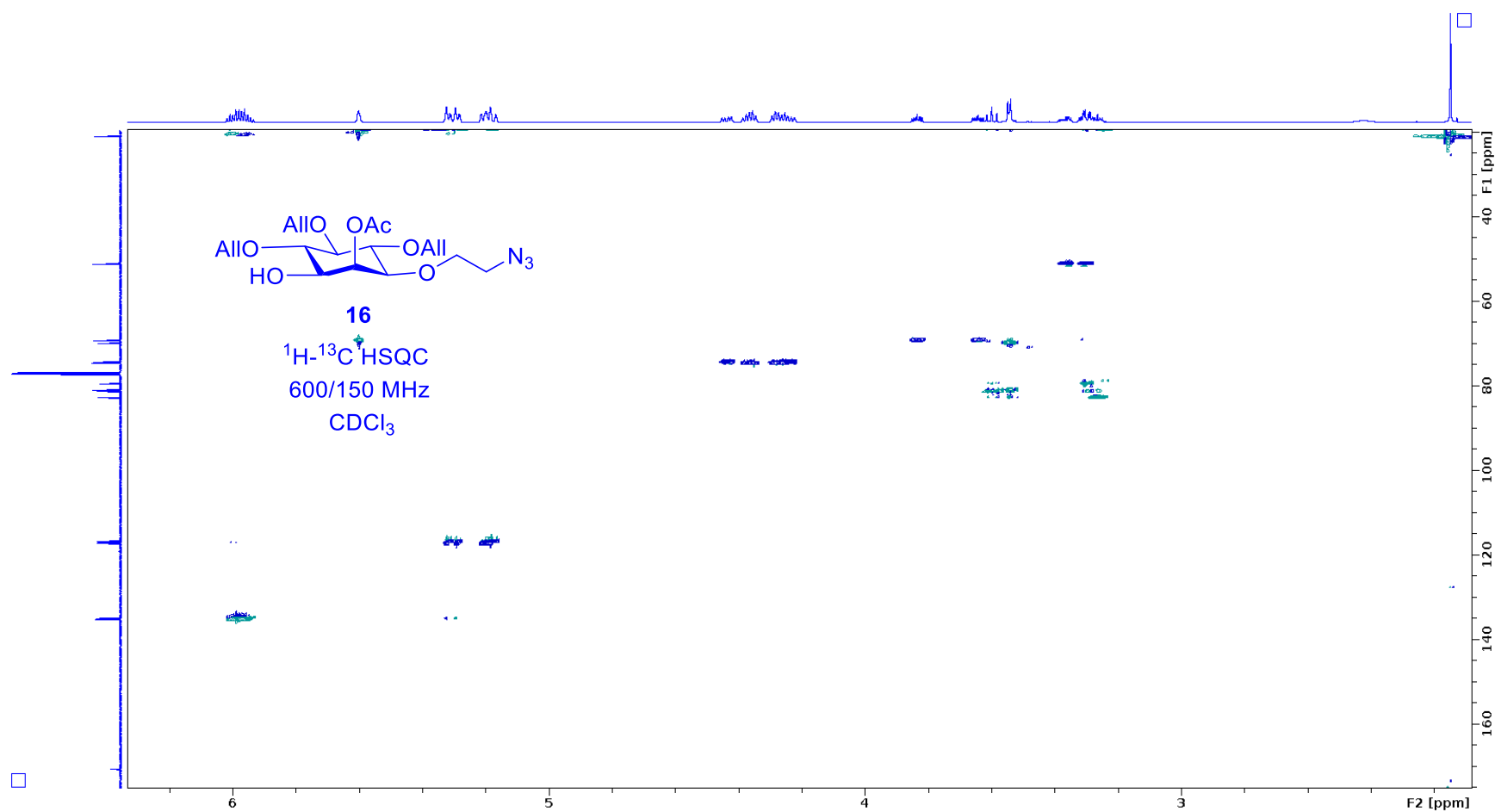
Chemical Formula: $C_{16}H_{23}N_3NaO_{10}^+$
Exact Mass: 440.1276





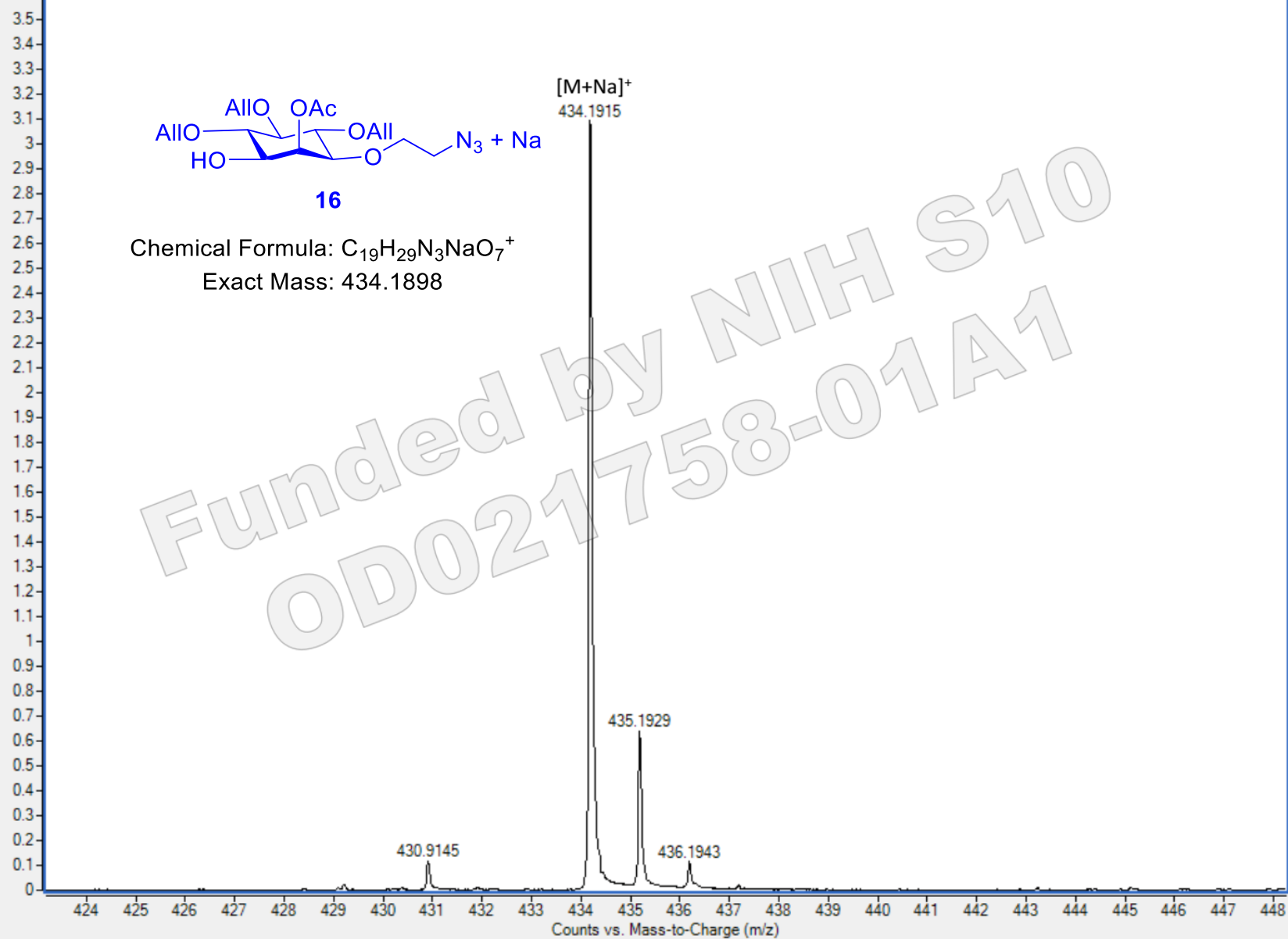
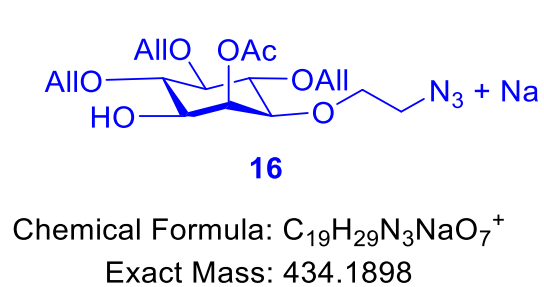


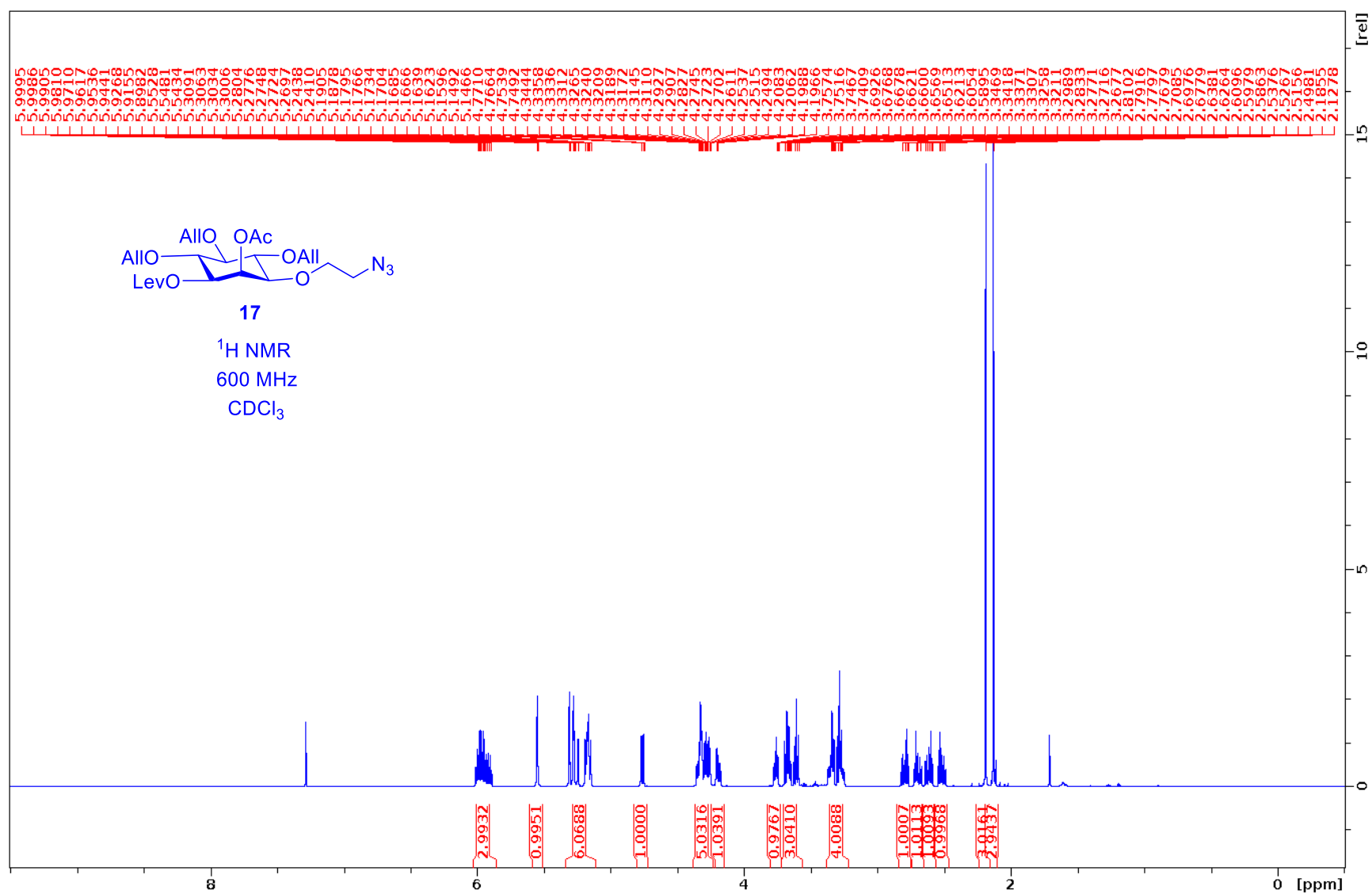


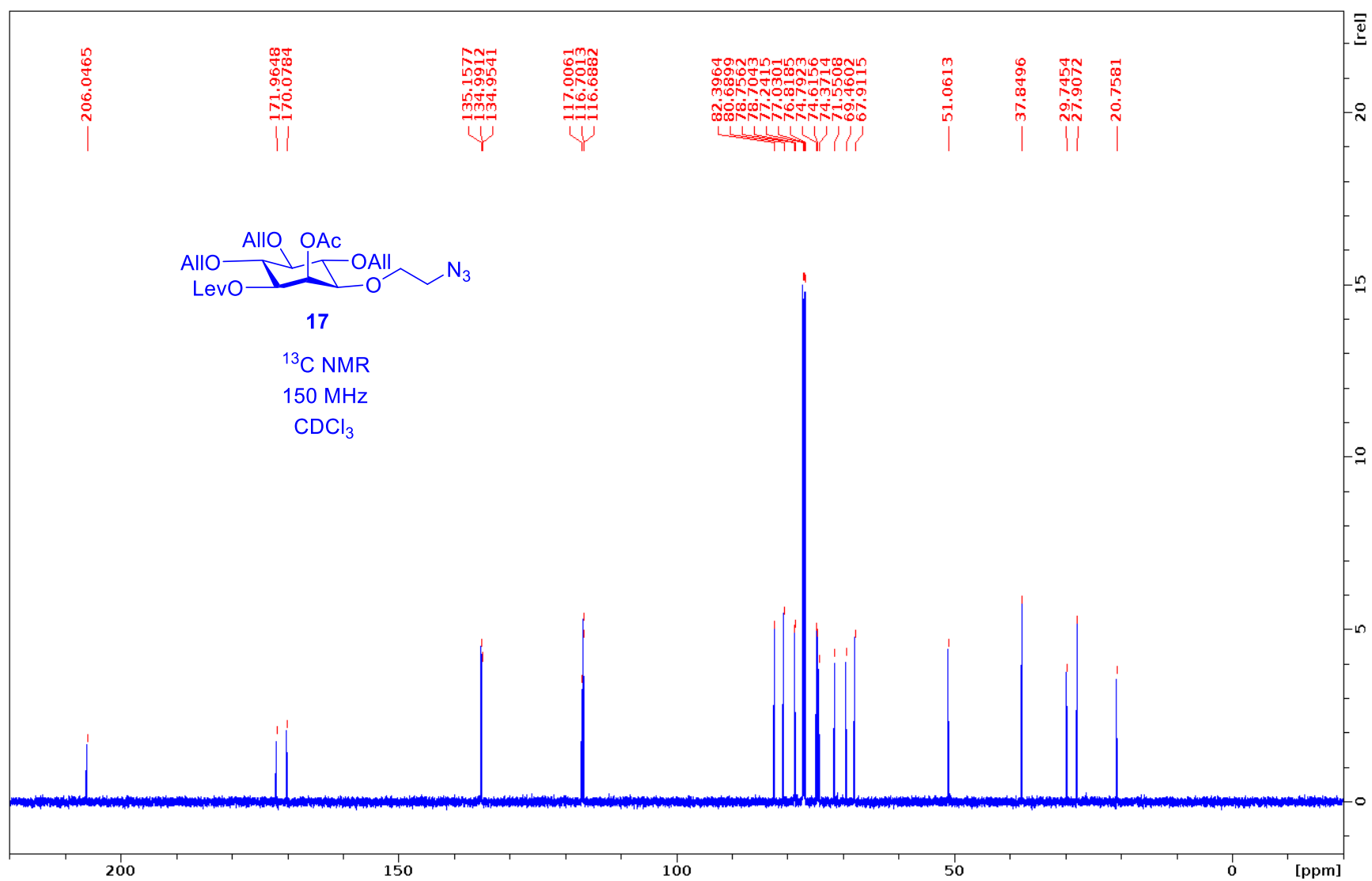


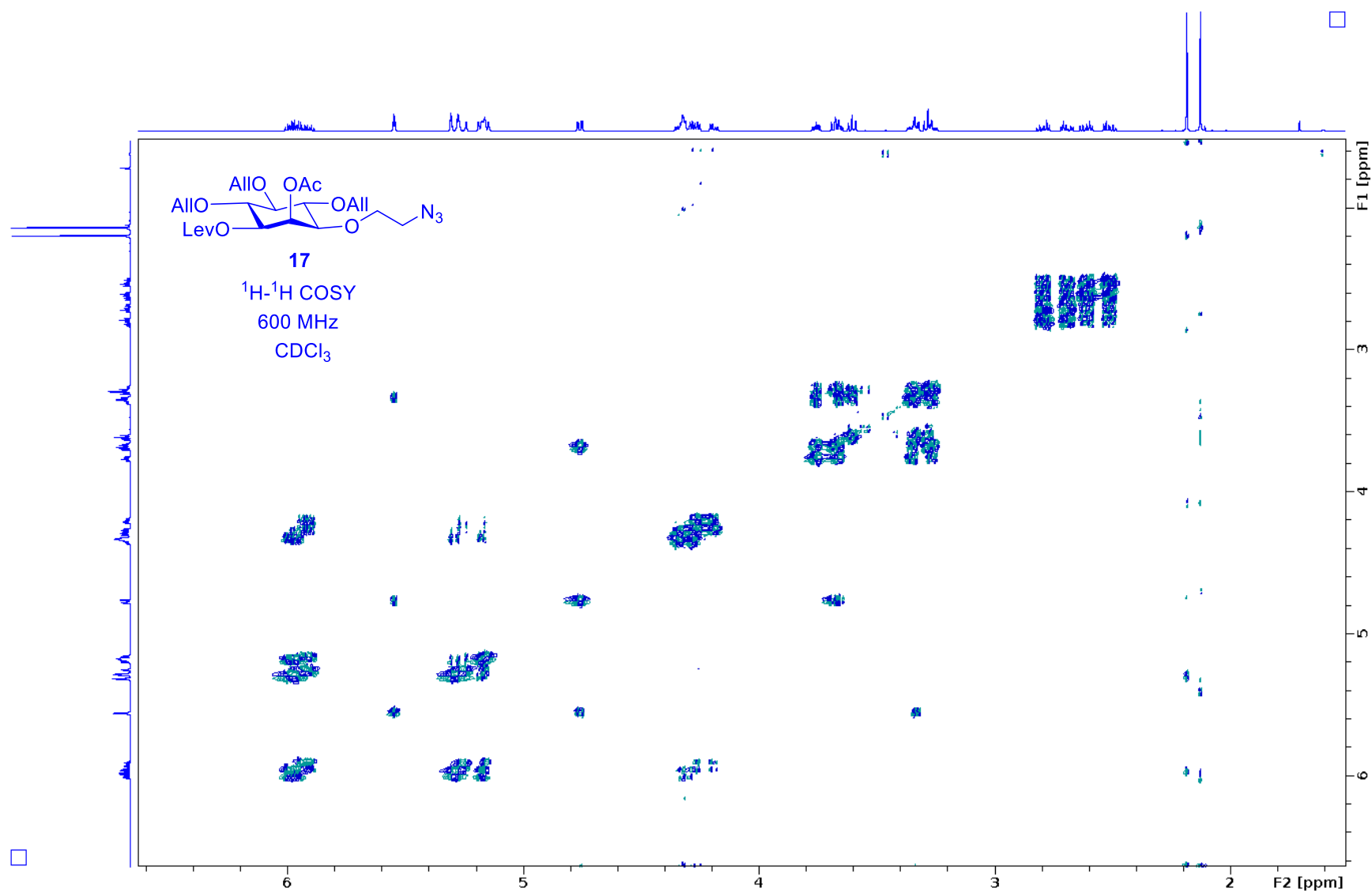
x10⁴ +ESI Scan (0.256-0.613 min, 44 Scans) Frag=175.0V 28151_zsy-3-45_ESI.d Subtract

Theoretical [M+Na]⁺ = 434.1898 (3.9 ppm)



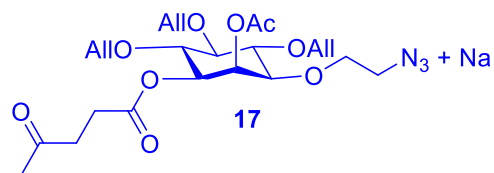




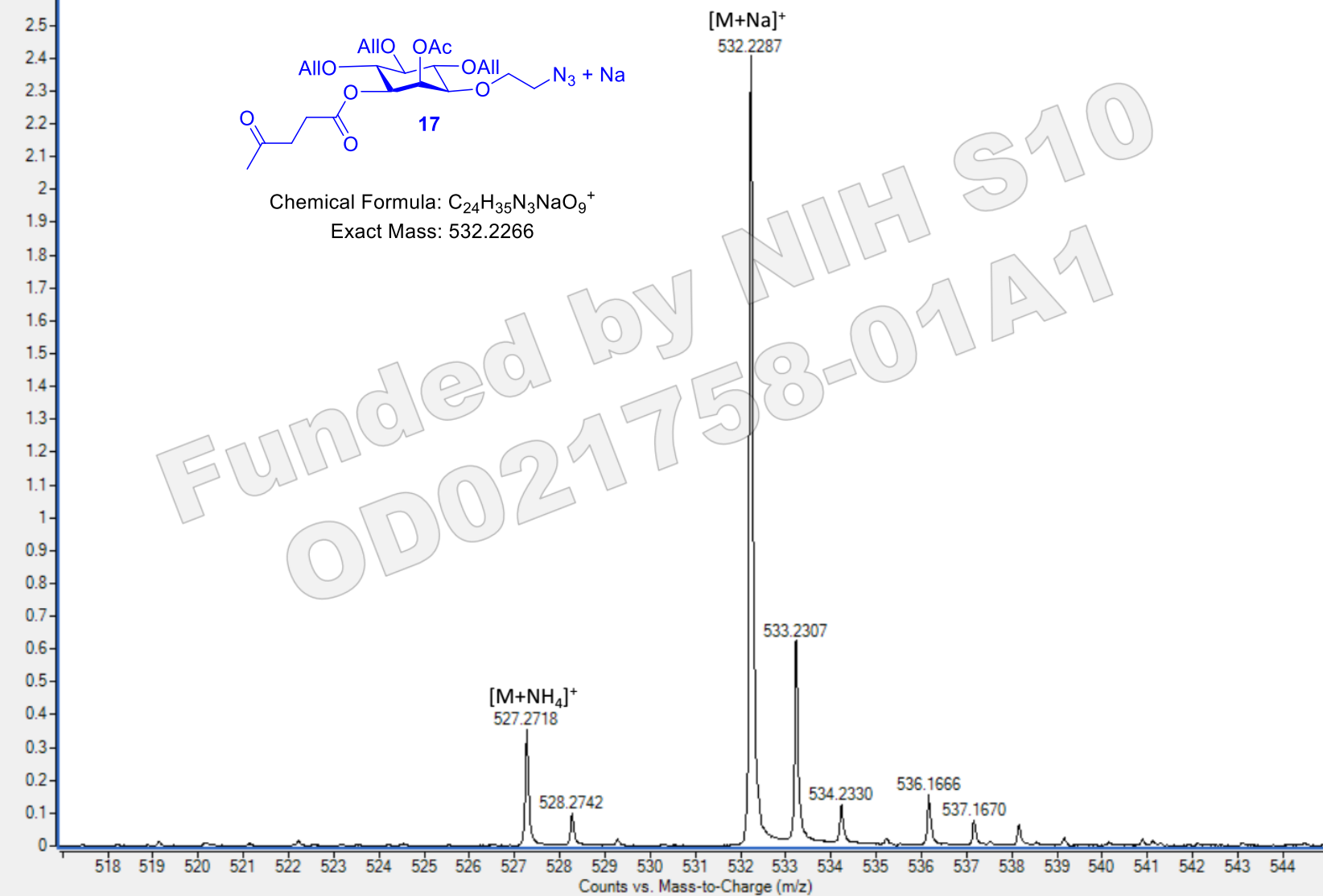


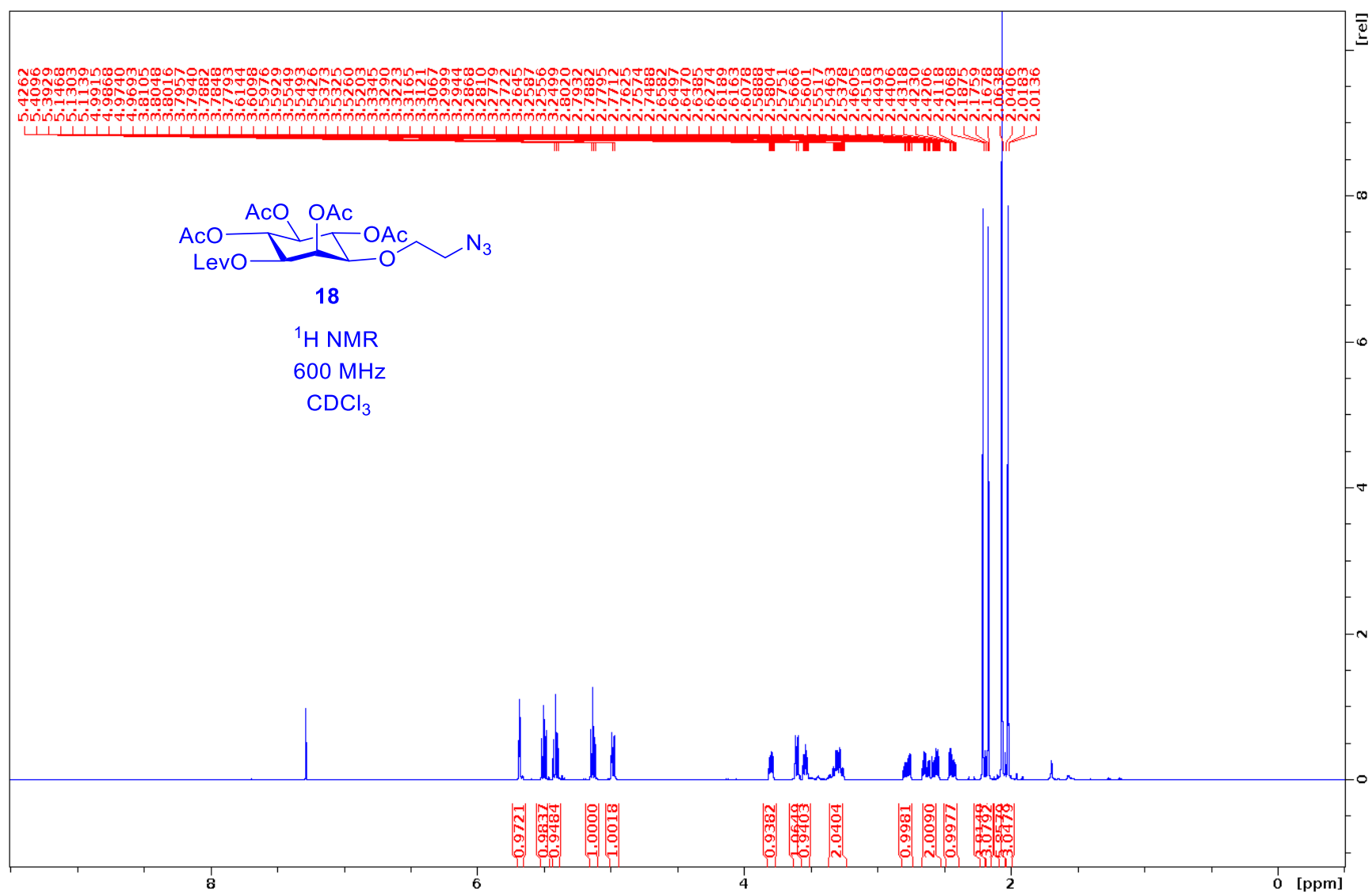
x10⁴ +ESI Scan (0.420-0.736 min, 39 Scans) Frag=175.0V 28156_zsy-3-48_ESI.d Subtract

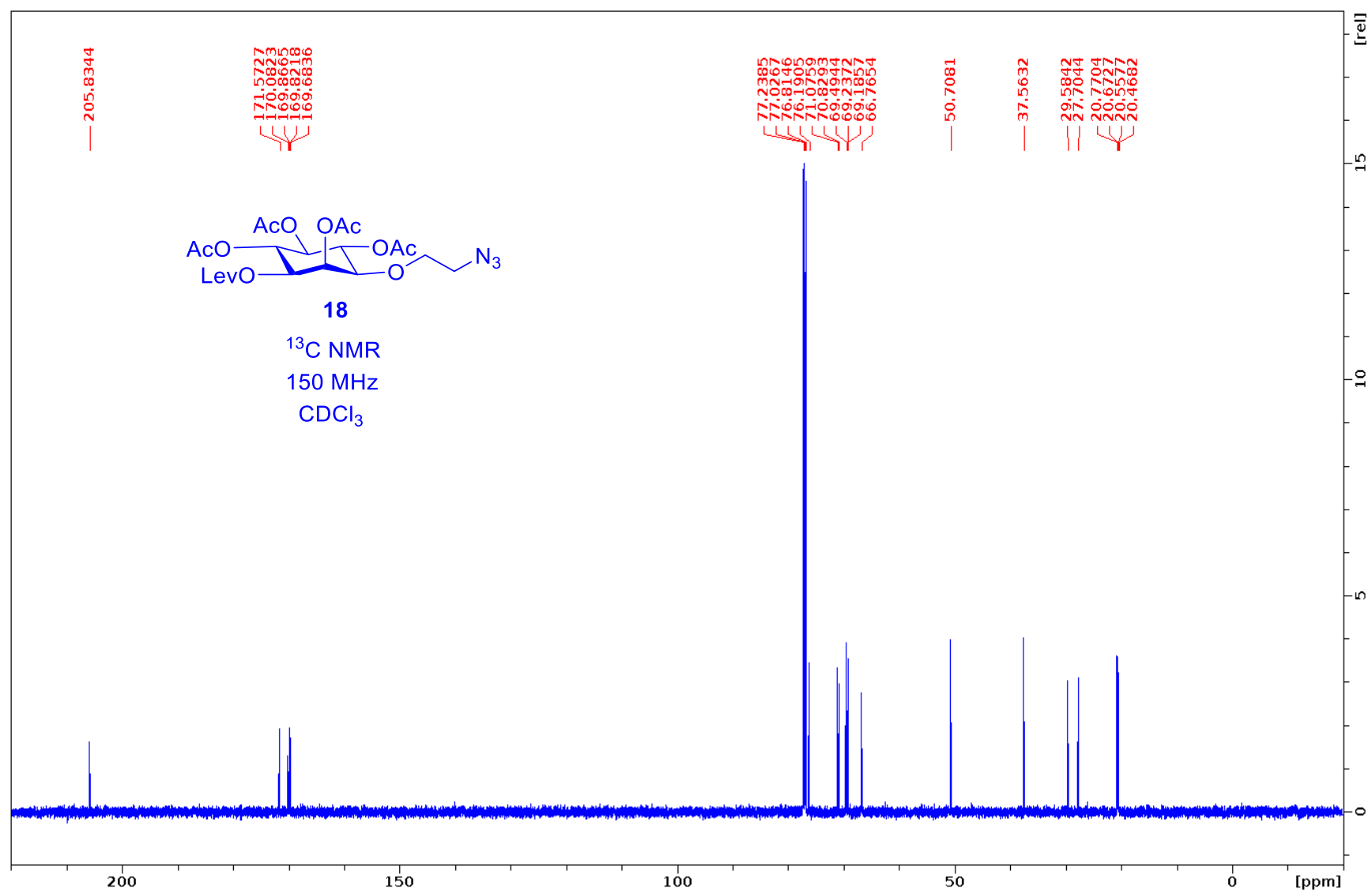
Theoretical $[M+NH_4]^+ = 527.2712$ (1.1 ppm)
 $[M+Na]^+ = 532.2266$ (3.9 ppm)

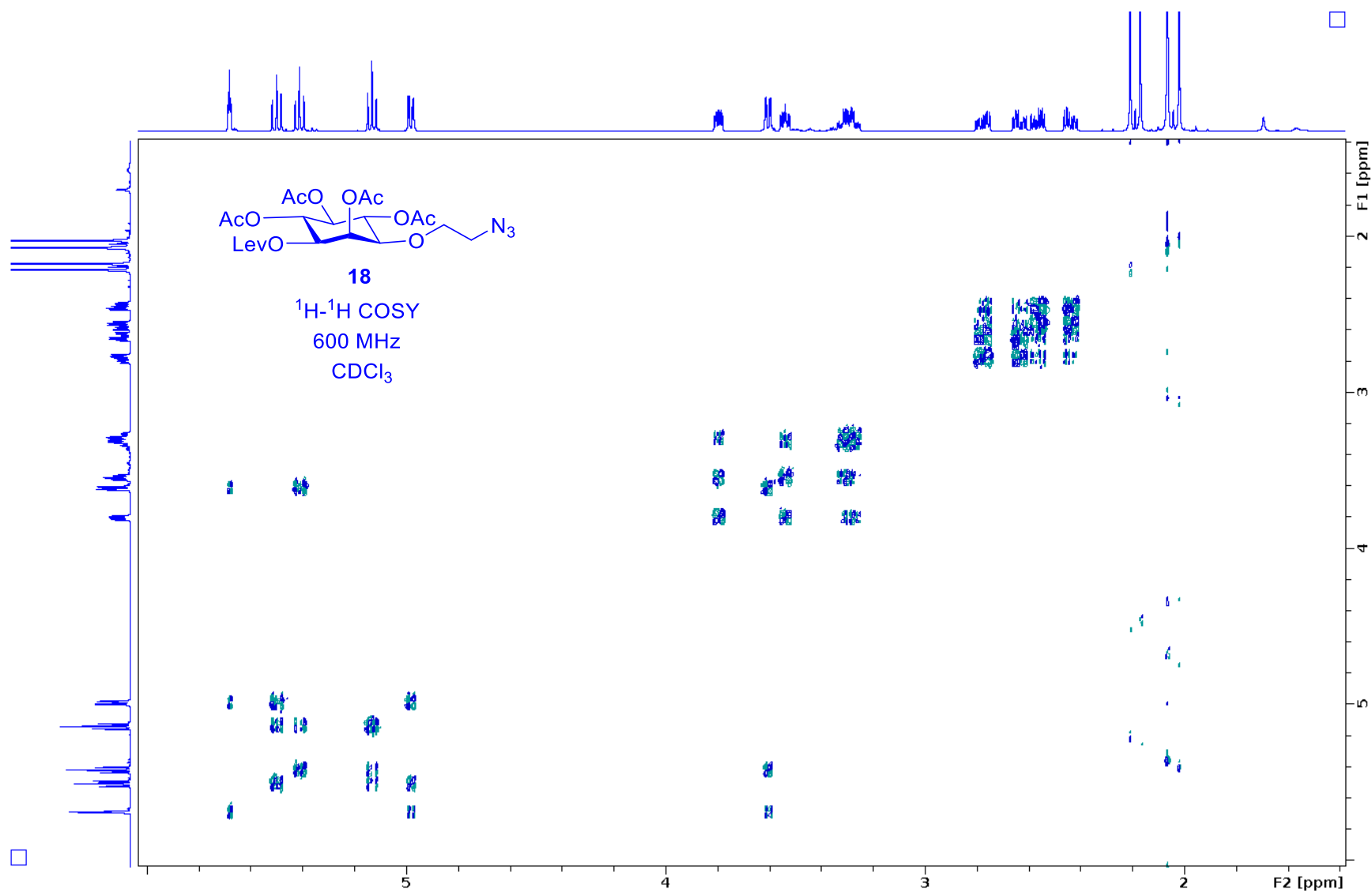


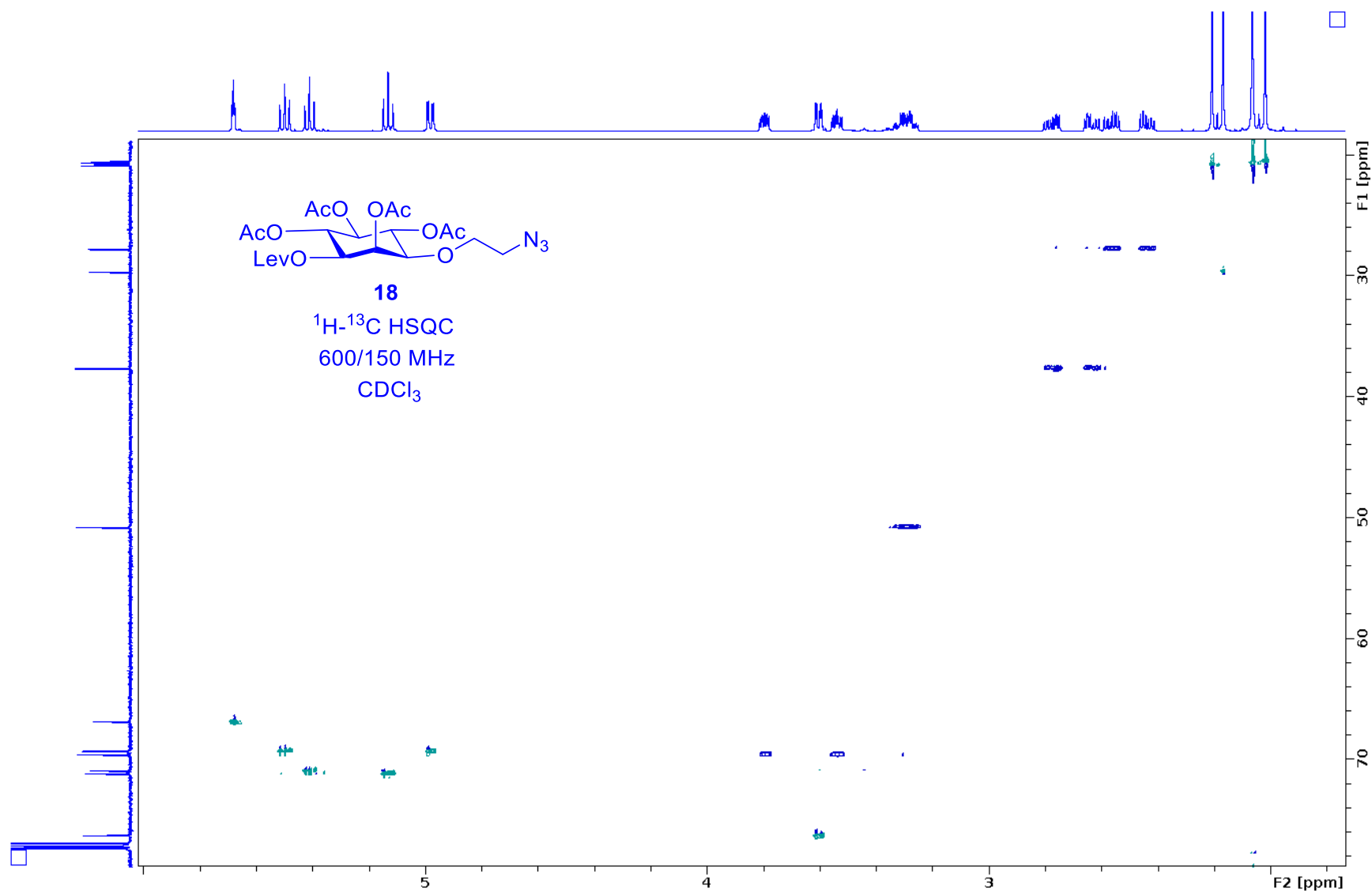
Chemical Formula: C₂₄H₃₅N₃NaO₉⁺
Exact Mass: 532.2266

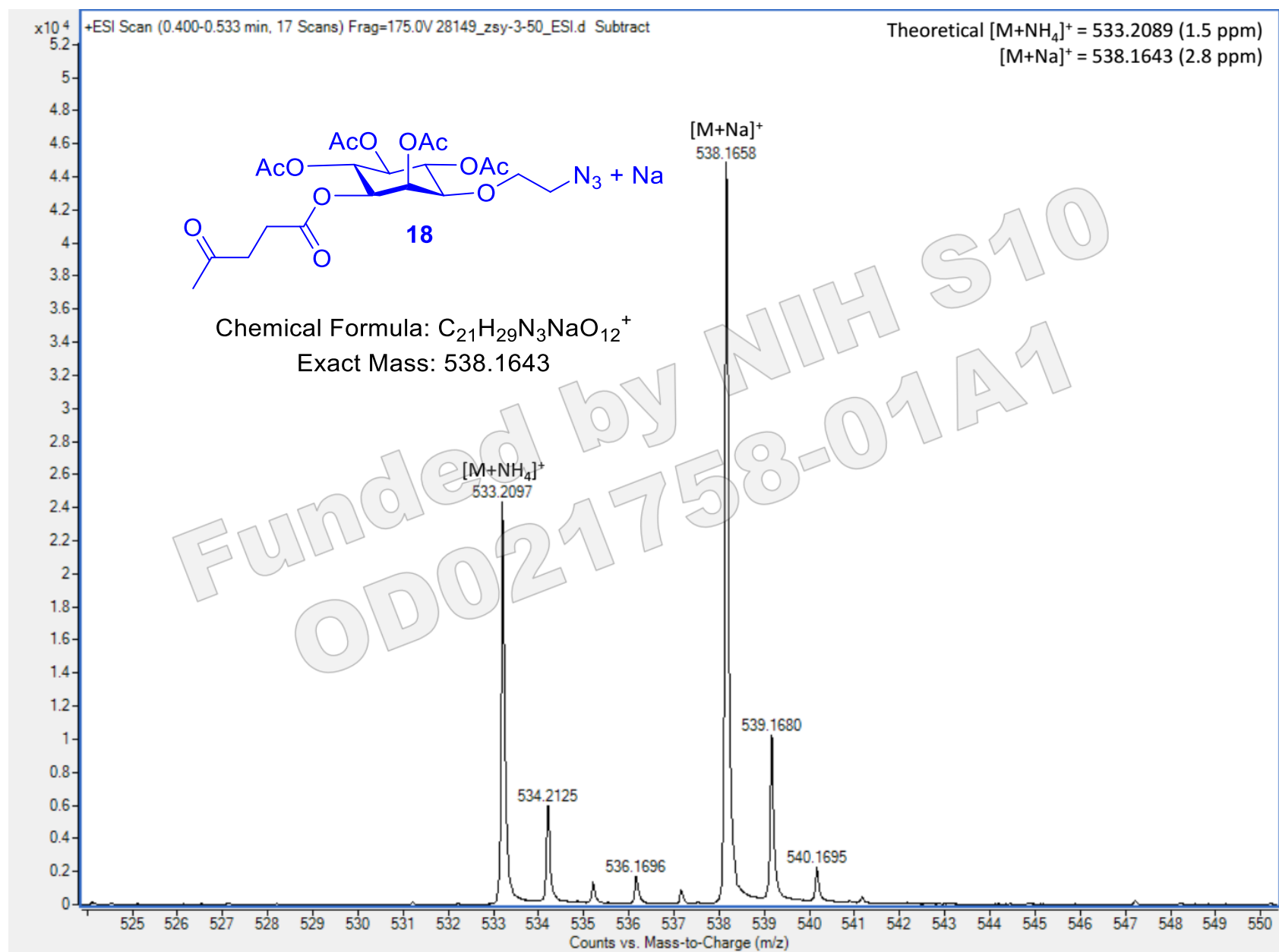


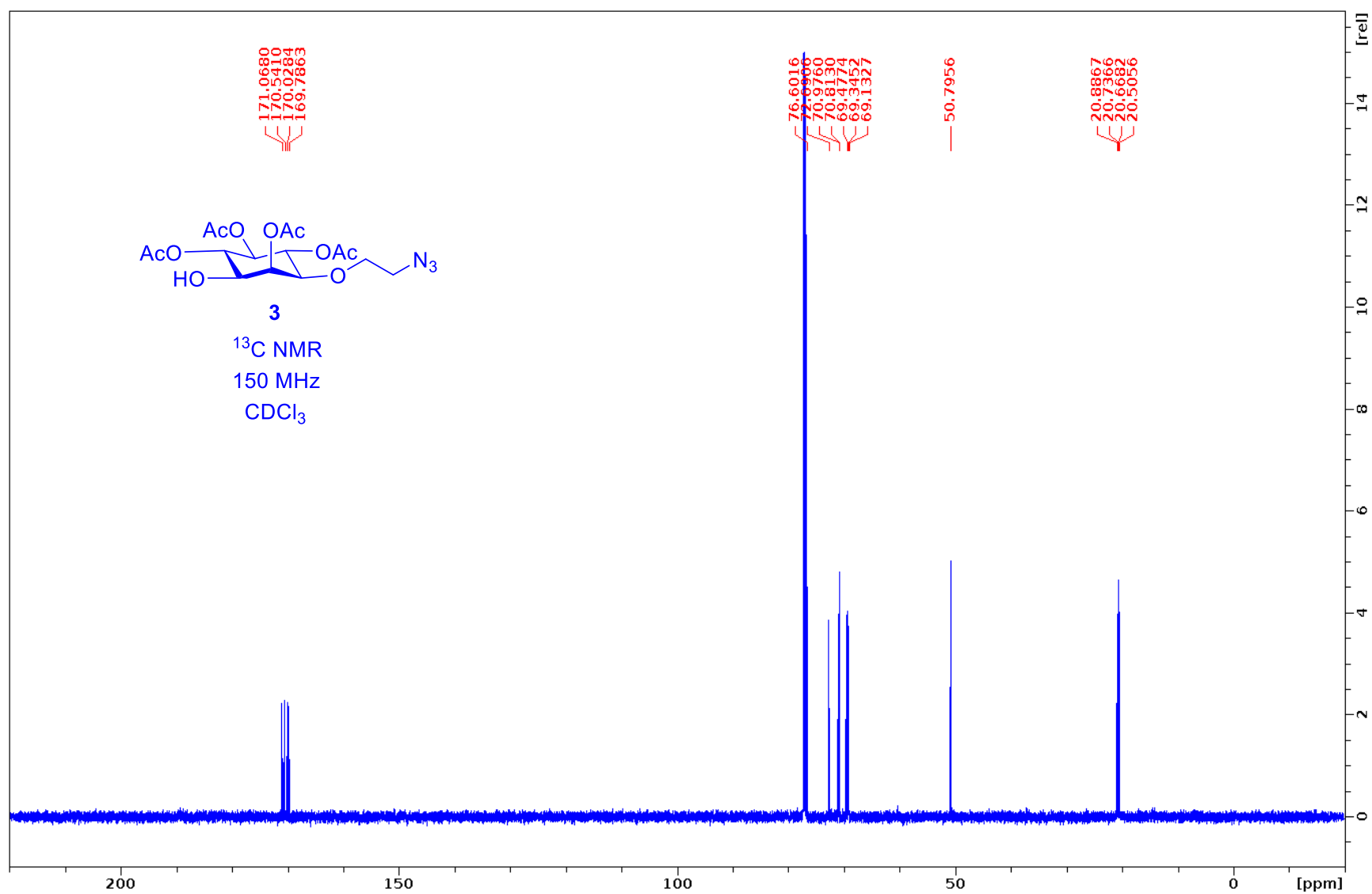


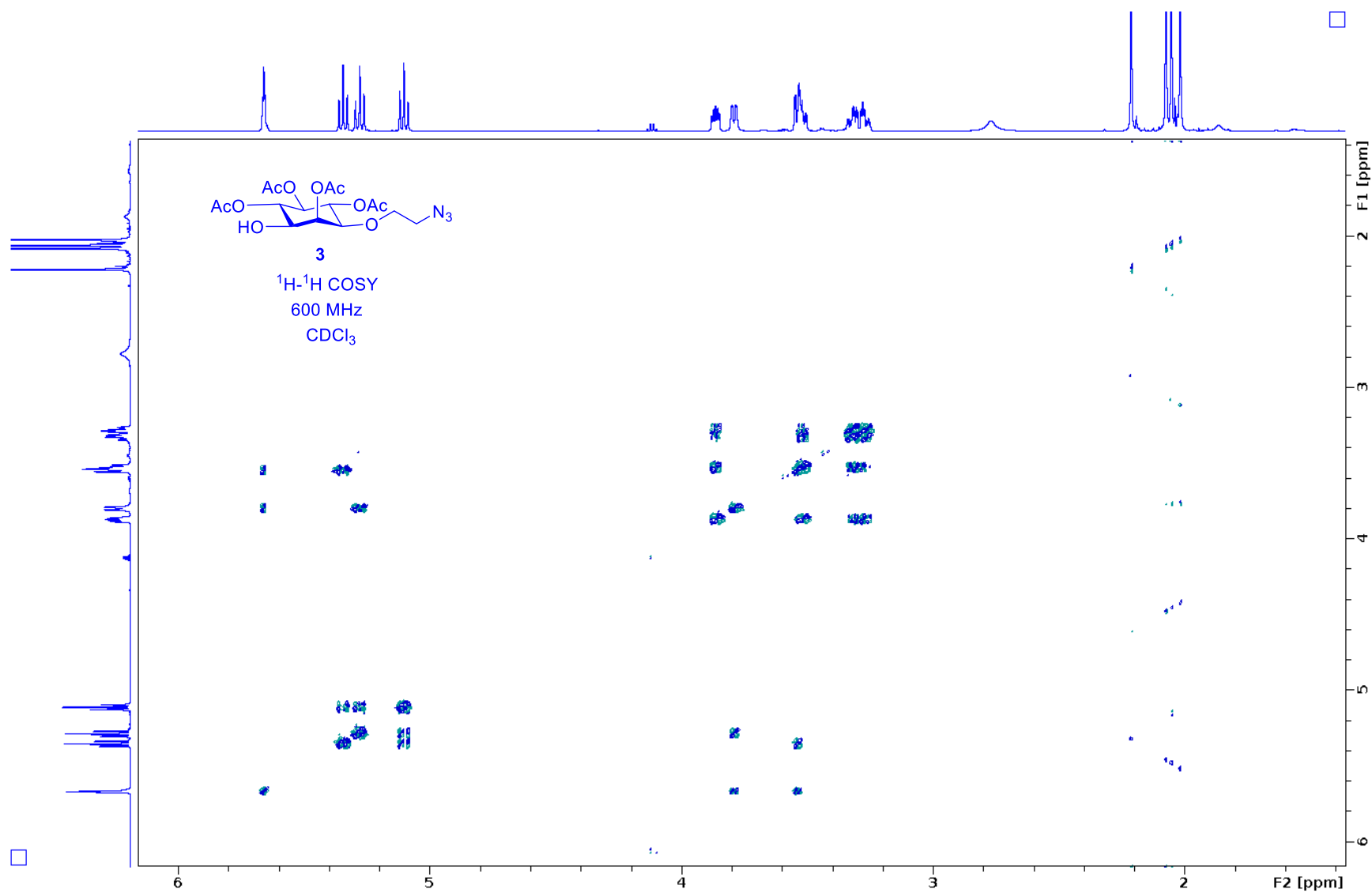


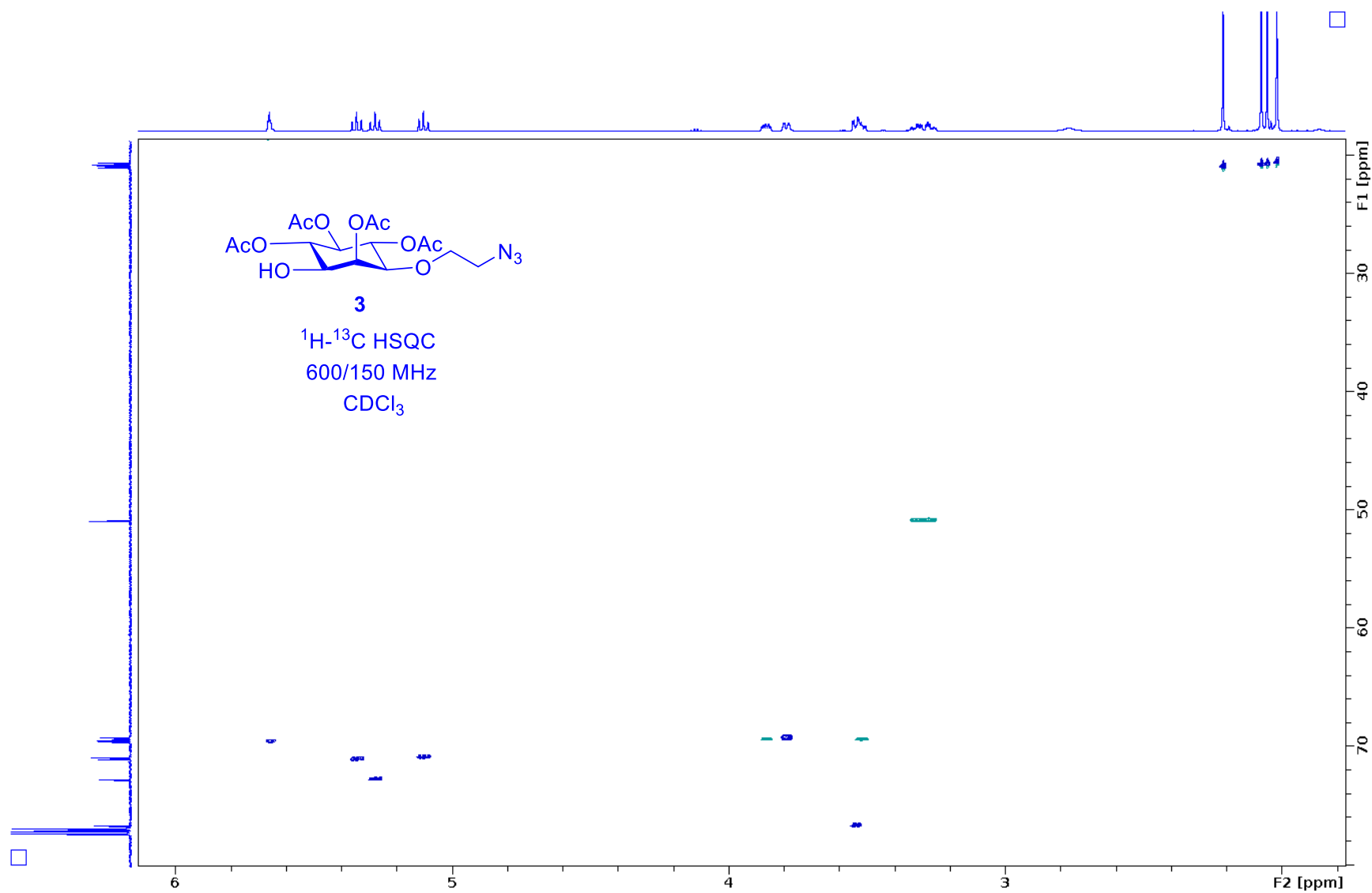








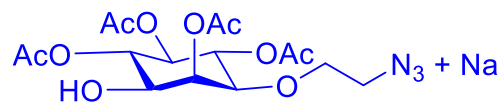




x10⁴ +ESI Scan (0.242-0.691 min, 55 Scans) Frag=175.0V 28146_zsy-3-51_ESI.d Subtract

Theoretical [M+NH₄]⁺ = 435.1722 (0.9 ppm)

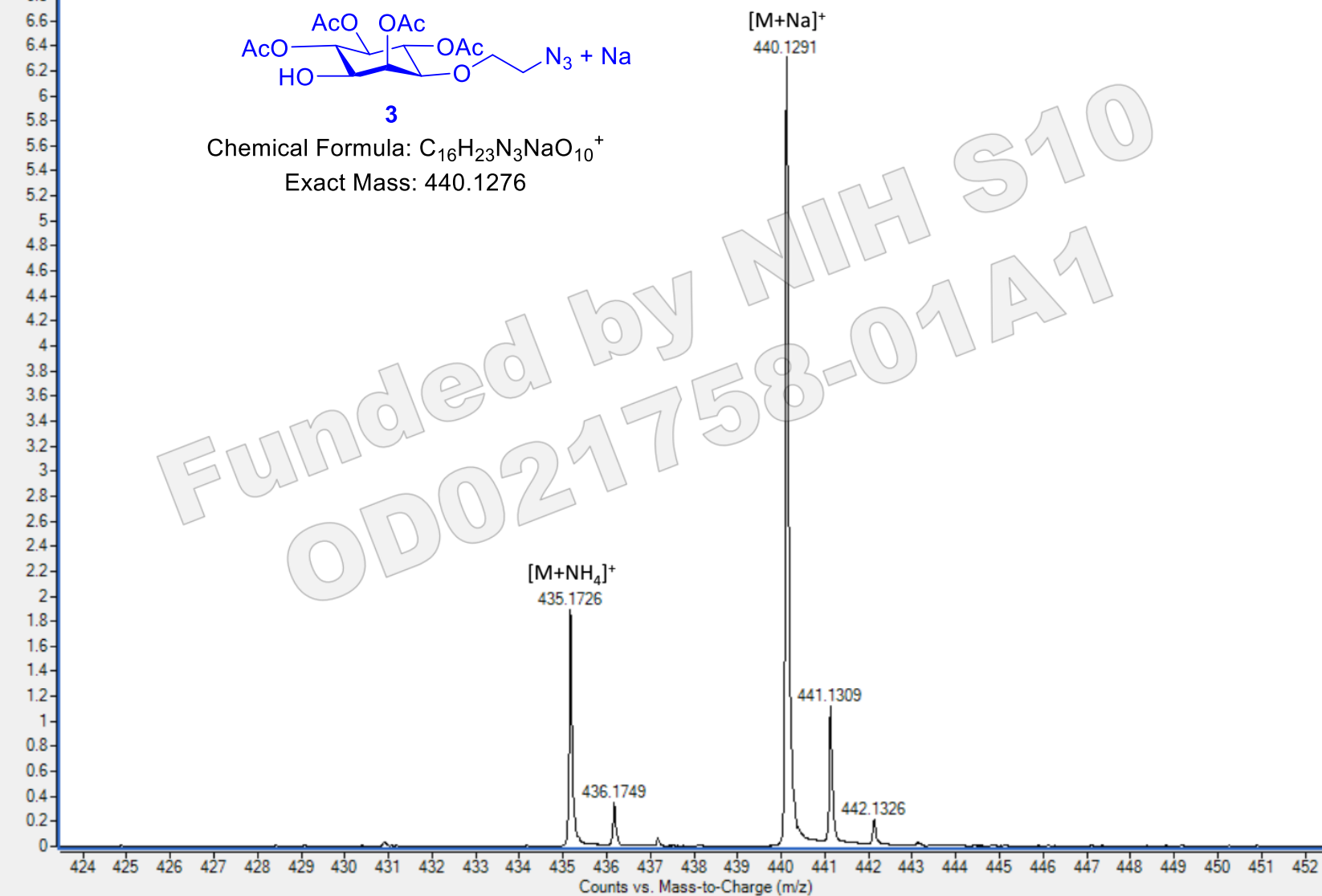
[M+Na]⁺ = 440.1276 (3.4 ppm)

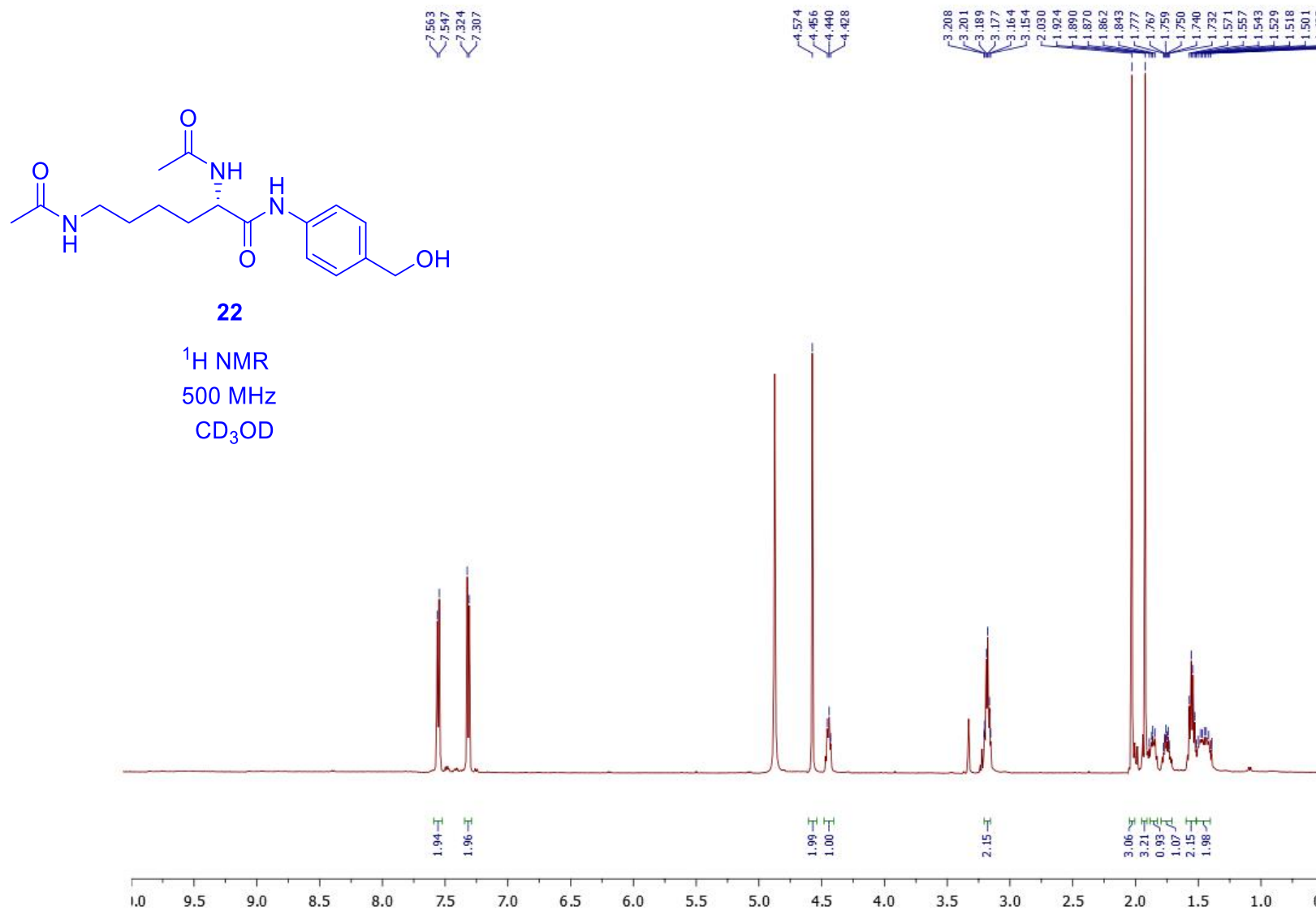


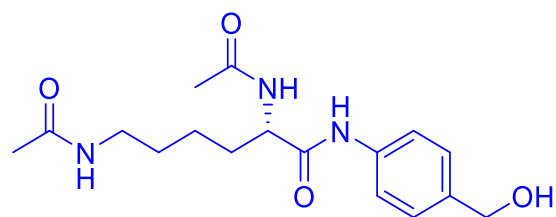
3

Chemical Formula: C₁₆H₂₃N₃NaO₁₀⁺

Exact Mass: 440.1276

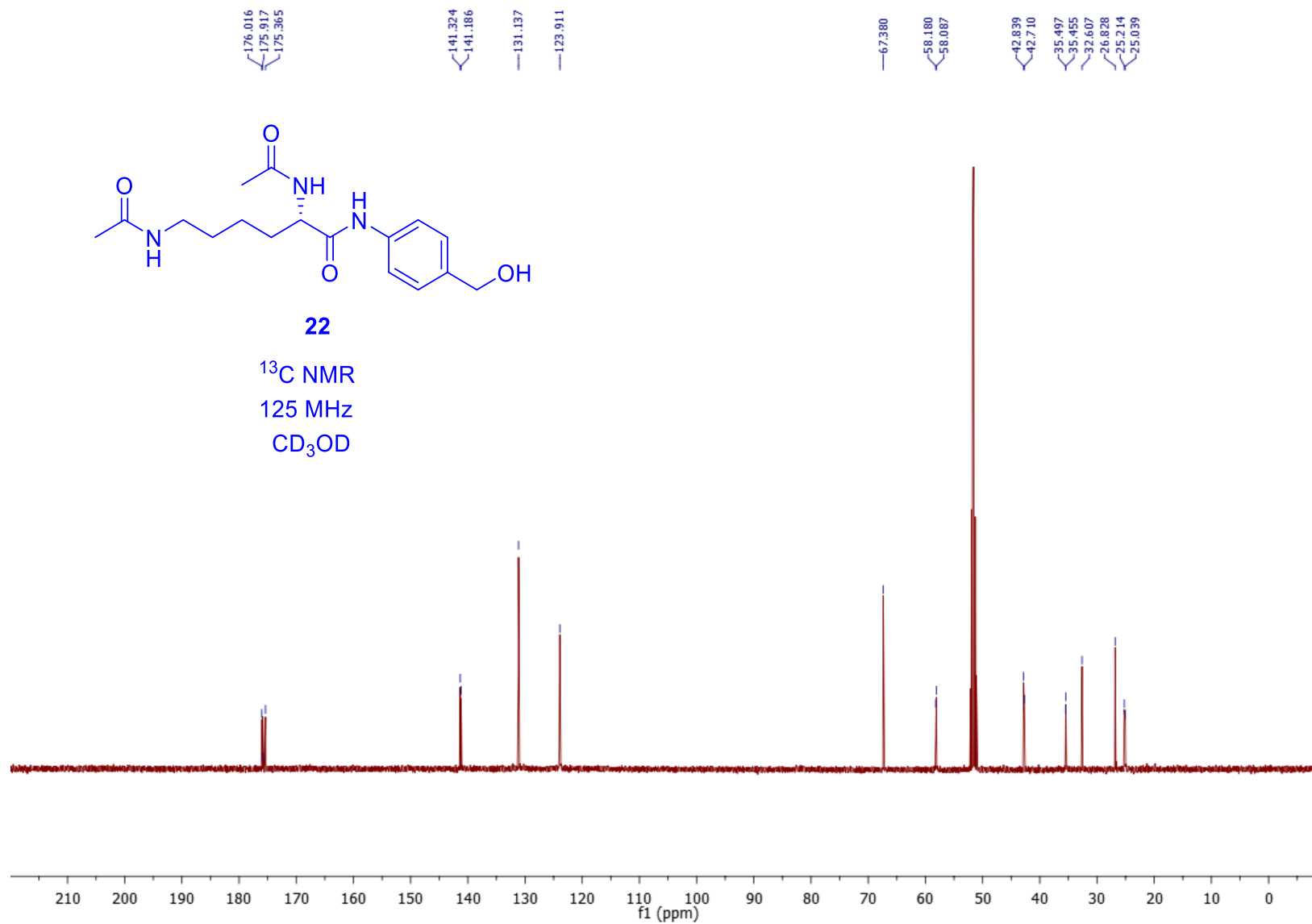


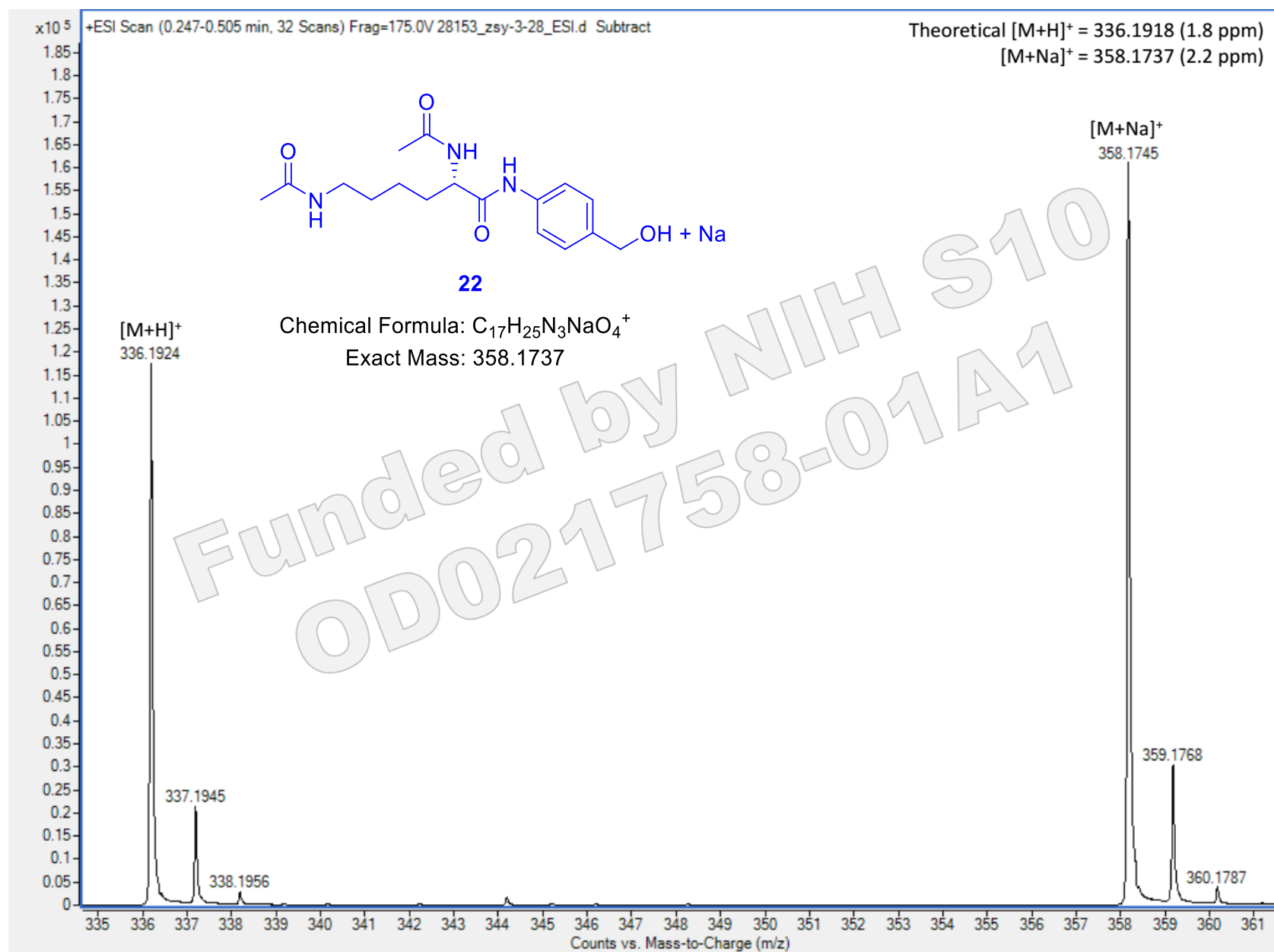


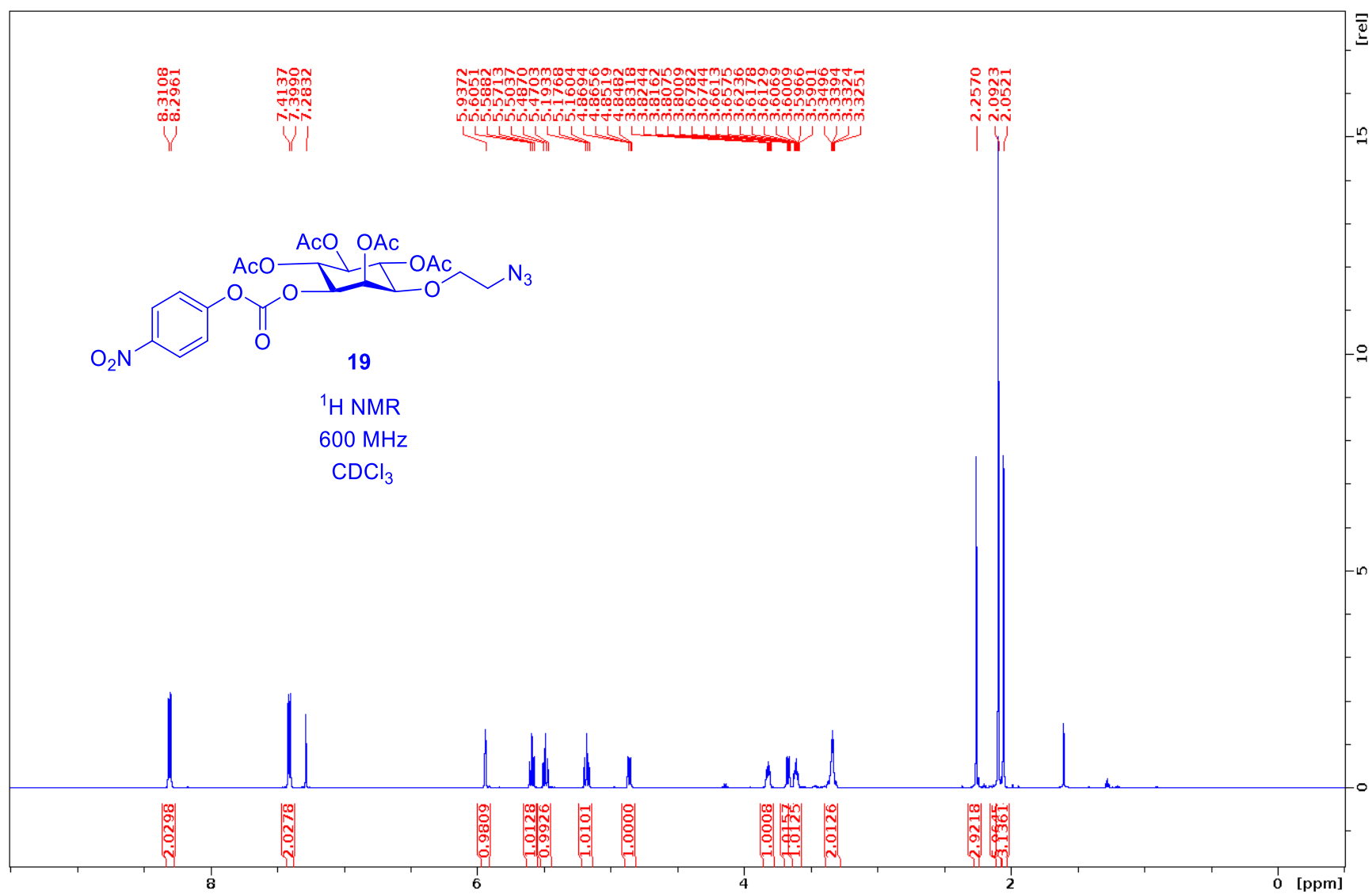


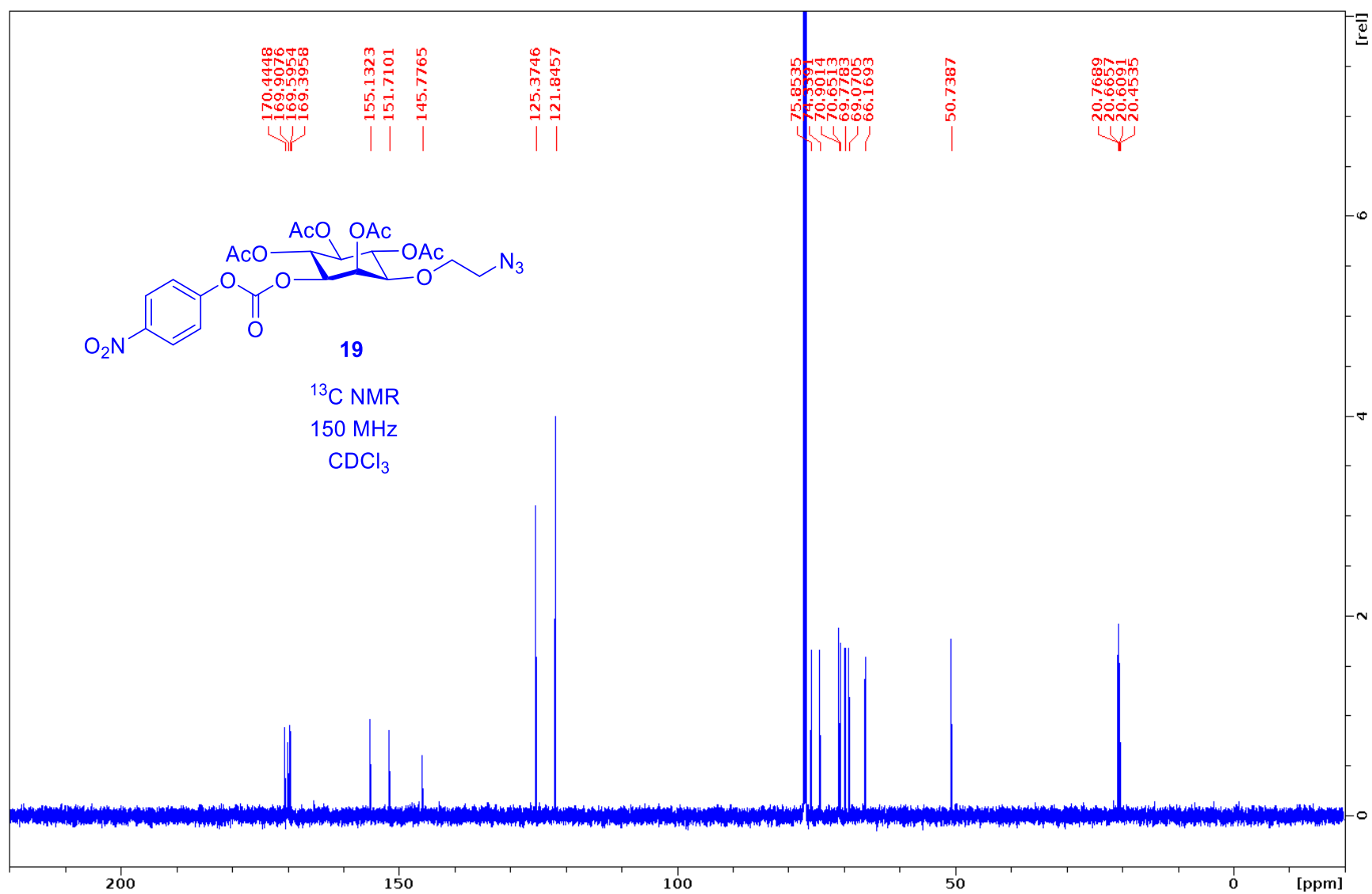
22

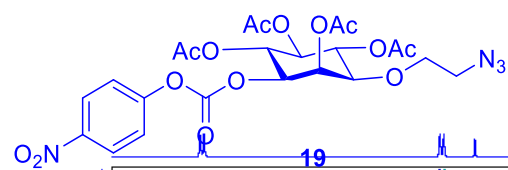
^{13}C NMR
125 MHz
 CD_3OD



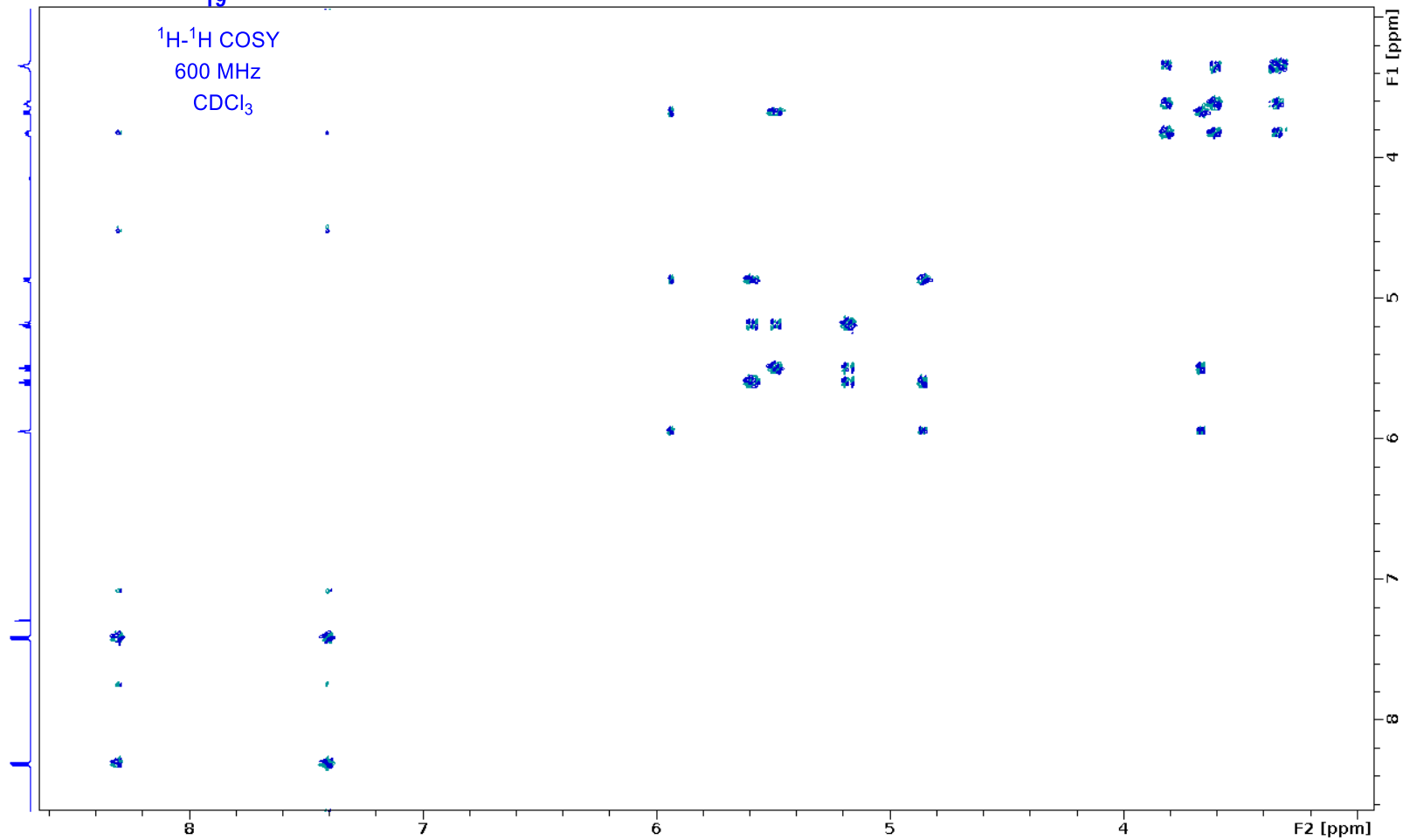


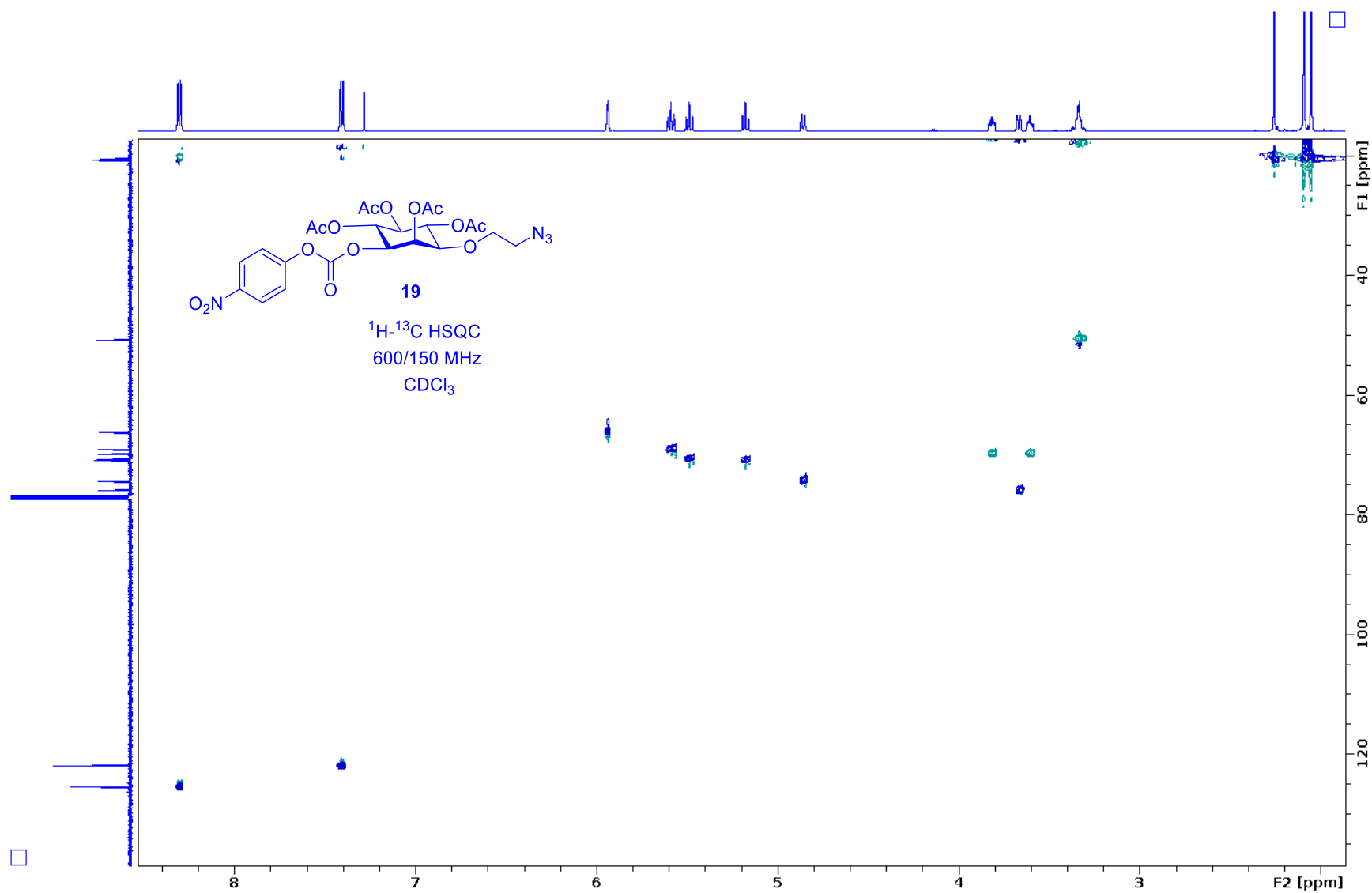


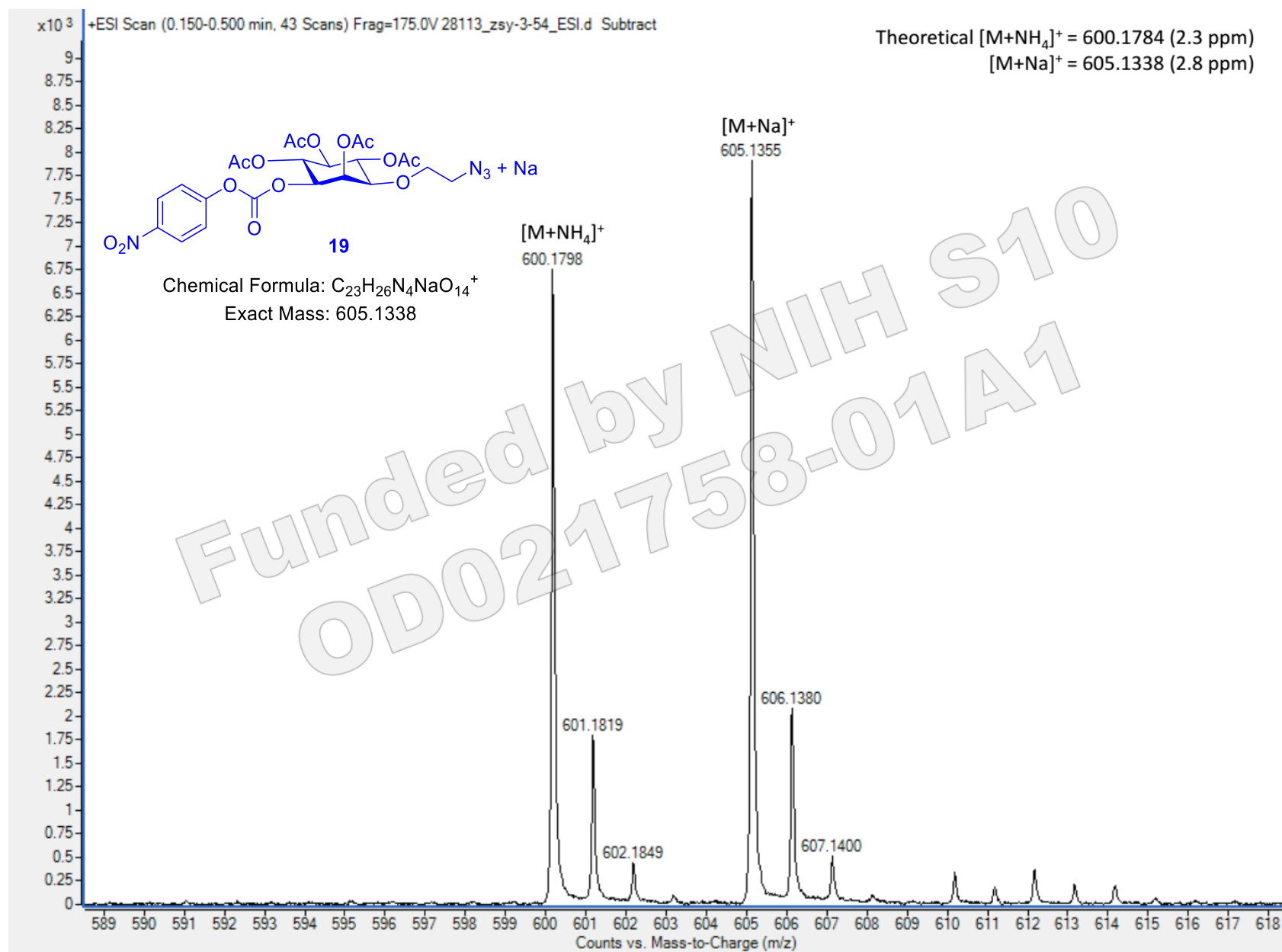


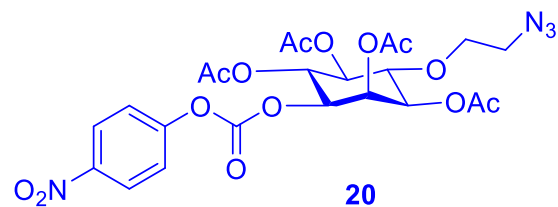


¹H-¹H COSY
600 MHz
CDCl₃

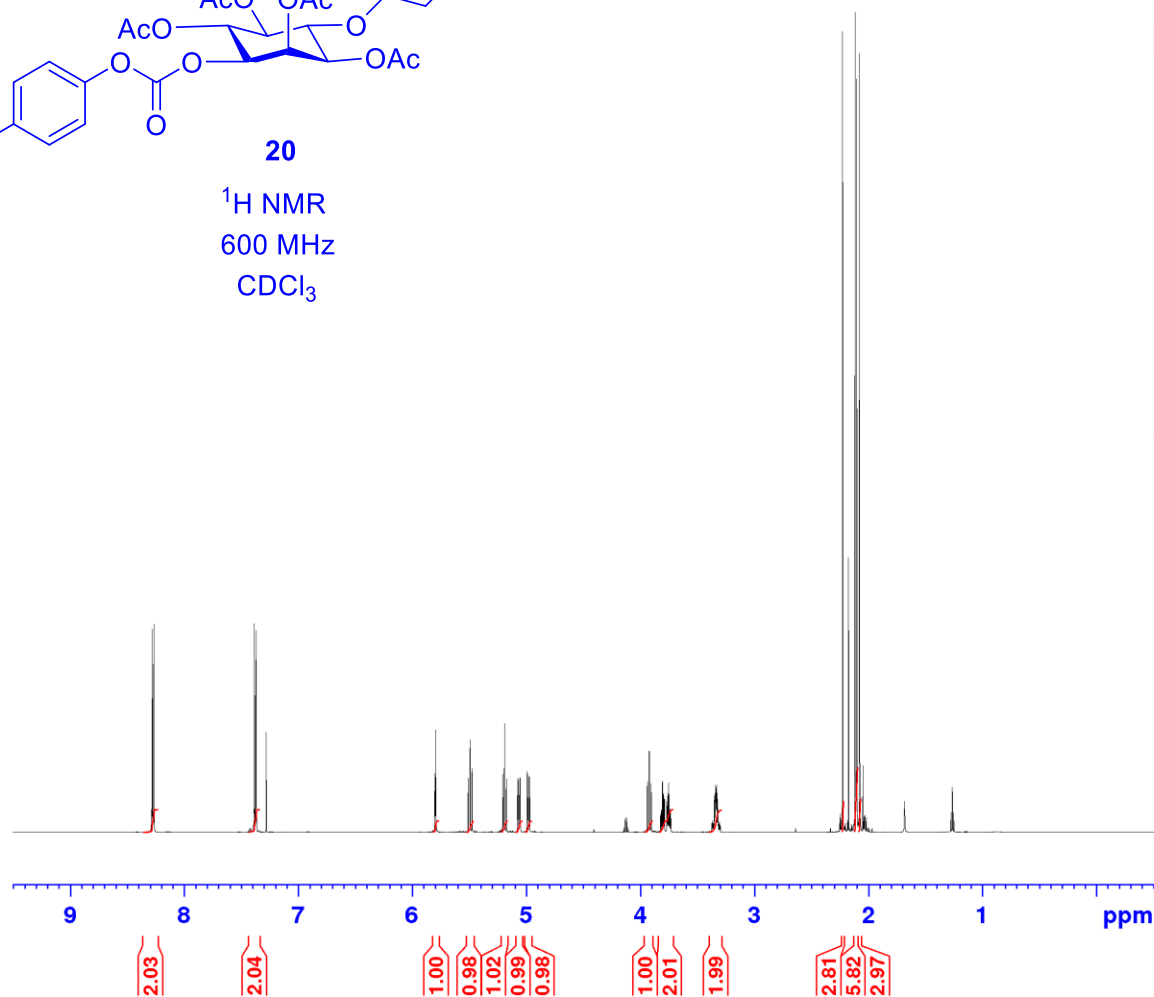








20
¹H NMR
 600 MHz
 CDCl₃



Current Data Parameters
 NAME zsy-3-37
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180413
 Time 16.09 h
 INSTRUM spect
 PROBHD Z847801_0073 (
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 5.4525952 sec
 RG 114
 DW 83.200 usec
 DE 6.50 usec
 TE 302.0 K
 D1 1.00000000 sec
 TD0 1
 SFO1 600.1627007 MHz
 NUC1 1H
 P1 14.00 usec
 PLW1 15.77099991 W

F2 - Processing parameters
 SI 65536
 SF 600.1600000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

