

Supplemental Information

**1,2-*cis*-Selective Glucosylation Enabled
by Halogenated Benzyl Protecting Groups**

Dancan K. Njeri, Claude J. Pertuit, and Justin R. Ragains

**Department of Chemistry
Louisiana State University
232 Choppin Hall
Baton Rouge, LA 70803**

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GENERAL INFORMATION

All reactions were performed under N₂ atmosphere which was achieved by vacuum purge backfill three times. Dried solvents (CH₂Cl₂, CH₃CN, Et₂O, THF and DMF) were used directly from a PureSolv 400-5 solvent purification system. Dry 1,4-dioxane solvent was obtained through distillation from sodium-benzophenone ketyl. Reagents were purchased from commercial sources (Alfa Aesar, Acros Organics, Chem Impex, Matrix scientific (most *p*-substituted benzyl bromides), Sigma Aldrich, TCI). Column chromatography was performed using silica gel (60 Å) purchased from SiliCyle. Preparative TLC was performed on 60 Å silica gel with fluorescent indicator on a glass plate (Agela Technologies). Analytical TLC was performed using 60 Å silica gel with F254 indicator on aluminum sheets (Merck). Compound visualization on TLC was performed using a hand-held UV hand lamp and/or staining with anisaldehyde. ¹H NMR, ¹³C NMR, and ¹⁹F NMR experiments were performed using a Bruker AV-400 or a Bruker AV-500 NMR spectrometer. ¹⁹F NMR spectra were calibrated with CFC₃ (δ = 0.00 ppm) as internal standard. HRMS was performed using an Agilent 6210 electrospray time-of-flight mass spectrometer. Optical rotation values were obtained using a JASCO P-2000 instrument. Deuterated solvents were obtained from Cambridge Isotope Labs.

Determining Anomeric ratios.

For ¹H NMR of anomeric mixtures in both the crude and purified samples, the number of scans used was 16 while relaxation delays were set to 20 seconds. *Alpha:beta* ratios of glycosylation product mixtures were determined using ¹H NMR integration of distinct signals from both the *alpha* and *beta* products. In the case of glycosylation reactions using acceptor **12** bearing a free hydroxyl at C6 and a methyl aglycone at the reducing end, anomeric ratios were determined according to the following set of commands using the GSD algorithm (deconvolution) in MestReNova:

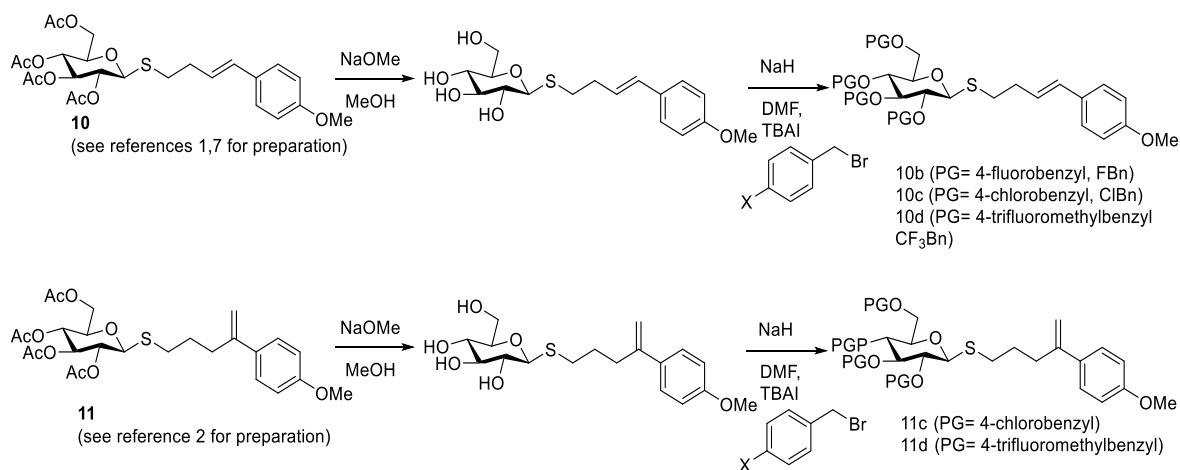
- 1) Phase correction- Processing> Phase correction> Automatic
- 2) Baseline correction- Processing> Baseline> Full auto (Bernstein polynomials)

- 3) Analysis > Peak picking > options = Method - GSD, Refinement level – 5 fitting, Optimized for peaks – average. Then Ok.
- 4) Analysis > Integration > Options = Calculation method – Sum, Source – autodetect, Algorithm- peak picking, Minimum area – 3%. Then Ok.
- 5) Analysis > Peak picking > Automatic
- 6) View > Tables > Peaks.

A GSD table containing all the peaks with their respective height, width and area is then generated. We analyzed reducing-end aglycone methyl signals from both the *alpha* and *beta* products which were located at 3.35-3.36 ppm for *alpha* and 3.32-3.33 ppm for *beta*. Areas of these signals were used to determine anomeric ratios in Tables 1 and 2. Anomeric ratios in Scheme 2 were determined through synthesis and chromatographic purification of *alpha* and *beta* anomers of all products and integration of key signals in the ^1H NMR spectrum of purified material as detailed below.

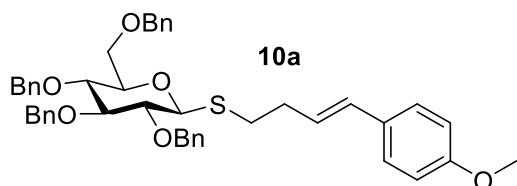
Preparation of MBTG and MPTG Donors

Synthetic scheme detailing synthesis of various MBTG/MPTG donors:



((E)-4-(4-methoxyphenyl)-3-butenyl β-D-1-thio-2,3,4,6-tetra-O-benzyl glucopyranoside)

10a⁷



Compound **10a** was prepared according to a previously published procedure (see reference 7 below).

General Procedure A for preparing *p*-substituted benzyl-protected glycosyl donors.

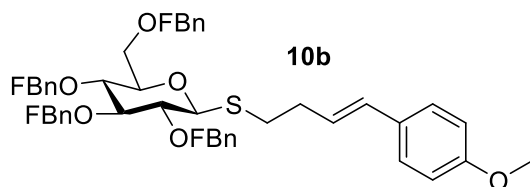
Part 1: To 0.7 g of tetraacetate thioglycoside donor **10**^{1,7} (4-(4-methoxyphenyl)-3-butenyl-1-thio-2,3,4,6-tetraacetyl-β-glucoside) or **11**² (4-(4-methoxyphenyl)-4-pentenyl-1-thio-2,3,4,6-tetraacetyl-β-glucoside, see synthetic scheme above) in a R.B.F was added 10.0 ml of methanol.

A stir bar was added, then a catalytic amount of 5 M sodium methoxide solution was added

dropwise. The reaction was stirred for 10 min after which it was concentrated using a rotary evaporator to give a white powder that was co-evaporated with 2 ml of toluene twice.

Part 2: 50ml of DMF was then added to the aforementioned white powder (tetraol), followed by 0.5 g of tetrabutylammonium iodide (TBAI). The contents of the R.B.F were allowed to stir at 0 °C after which 8-24 equivalents of NaH was added in three portions. Ten minutes later, 8 equivalents of *p*-substituted benzyl bromide was added. The reaction was allowed to stir for 12 hrs while warming to room temperature under a nitrogen atmosphere. The reaction was then placed back into an ice bath quenched at 0°C with the dropwise addition of 10.0 ml of water. Further workup did not commence until gas evolution ceased. 100.0 ml of EtOAc was added. The reaction contents were then transferred to a 500 ml separatory funnel. 200.0 ml of water was then added, and the contents of the separatory funnel were shaken vigorously and then allowed to stand to ensure separation of the aqueous and organic layers. The organic layer was dried using Na₂SO₄ then concentrated to give a crude product which was purified using flash chromatography to give the intended tetrabenzylthioglucoside donor (**10a-d** or **11c-d**).

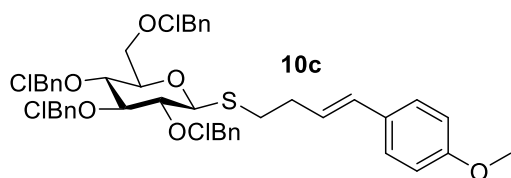
((*E*)-4-(4-methoxyphenyl)-3-butenyl β-D-1-thio-2,3,4,6-tetra-*O*-(4-fluorobenzyl) glucopyranoside) 10b



Prepared following the general procedure A. **Part 1;** used 0.70 g of tetraacetate thioglucoside **10**, 10 ml of methanol, and 0.4 ml of 5 M NaOMe. **Part 2;** 30 ml of DMF, 0.40 g of TBAI, 0.42 g of NaH and 1.33 ml of 4-fluorobenzyl bromide. The reaction crude product was purified by flash column chromatography (15% EtOAc/hexanes) to give 0.683

g of a white solid **10b** (65% yield). **¹H NMR (400 MHz, CDCl₃)** δ 7.35-7.16 (m, 8H), 7.13-7.05 (m, 2H), 7.03-6.91 (m, 8H), 6.83 (d, J = 8.7 Hz, 2H), 6.37 (d, J = 15.9 Hz, 1H), 6.08 (dt, J = 15.6 Hz, 6.9 Hz, 1H), 4.90-4.74 (m, 3H), 4.71 (d, J = 7.4 Hz, 1H), 4.65 (d, J = 10.2 Hz, 1H), 4.59-4.44 (m, 4H), 3.80 (s, 3H), 3.73-3.52 (m, 4H), 3.46-3.37 (m, 2H), 4.94-4.78 (m, 2H), 2.55 (q, J = 7.2 Hz, 2H) **¹³C NMR (100 MHz, CDCl₃)** δ 163.69, 163.61, 163.54, 161.11, 158.97, 134.24, 133.78, 133.70, 130.88, 130.11, 130.08, 130.00, 129.58, 129.53, 129.50, 129.45, 129.30, 129.22, 127.20, 125.98, 115.44-115.10 (m), 113.98, 86.41, 85.29, 81.69, 79.02, 77.82, 74.90, 74.73, 74.21, 72.80, 68.95, 55.31, 33.54, 30.93; **¹⁹F NMR (471 MHz, CDCl₃)** δ -114.74 – -114.88 (m, 2F), -115.07 (m, 1F), -115.22 (m, 1F). **HRMS (m/z)**: Calcd for C₄₅H₄₄F₄O₆SK 827.2432 found 827.2397. **[α]_D²⁵** = +2.3° (c = 1, DCM). **IR (cm⁻¹)** 3038, 2916, 2859, 1604, 1509, 1246, 1221, 1067, 823 (*p*-disubstituted benzene), 767, 501

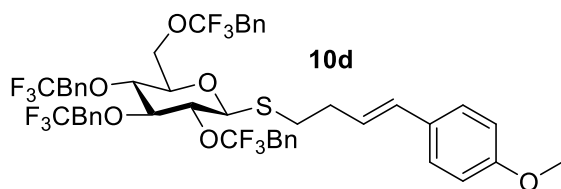
((*E*)-4-(4-methoxyphenyl)-3-butenyl β-D-1-thio-2,3,4,6-tetra-*O*-(4-chlorobenzyl)glucopyranoside) **10c**



Prepared following the general procedure A. **Part 1**; used 0.69 g of tetraacetate thioglucoside **10** (1.3 mmol), 10 ml of methanol, and 0.4 ml of 5 M NaOMe. **Part 2**; 20 ml of DMF, 0.20 g of TBAI, 0.42 g of NaH and 2.16 g of 4-chlorobenzyl bromide. The reaction crude product was purified by flash column chromatography (15% EOAc/hexanes) to give 0.8180 g of a white product **10c** (74% yield). **¹H NMR (400 MHz, CDCl₃)** δ 7.31-7.18 (m, 14H), 7.14 (d, J = 8.3 Hz, 2H), 7.04 (d, J = 8.3 Hz, 2H), 6.83 (d, J = 8.8 Hz, 2H), 6.37 (d, J = 15.8 Hz, 1H), 6.07 (dt, J = 15.8 Hz, 6.9 Hz, 1H), 4.86 (d, J = 10.6 Hz, 1H), 4.82-4.41 (m, 8H), 3.79 (s, 3H), 3.73-3.51 (m, 4H), 3.46-3.36 (m, 2H),

2.91-2.76 (m, 2H), 2.54 (q, $J = 7.2$ Hz, 2H)); ^{13}C NMR (100 MHz, CDCl_3) δ 158.99, 136.84, 136.55, 135.41, 136.33, 133.67, 133.63, 133.45, 130.93, 130.08, 129.48, 129.24, 129.04, 128.98, 128.77, 128.61, 128.57, 127.22, 125.91, 114.01, 86.42, 85.23, 81.75, 78.96, 77.85, 74.75, 74.60, 74.09, 72.71, 68.99, 55.31, 33.48, 30.92; HRMS (m/z): Calcd for $\text{C}_{45}\text{H}_{44}\text{Cl}_4\text{O}_6\text{SK}$ 891.1250 found 891.1225. $[\alpha]_{\text{D}}^{23} = +13.3^\circ$ ($c = 1$, DCM). IR (cm^{-1}) 3029, 2913, 2859, 1602, 1510, 1491, 1246, 1086, 1015, 841, 806 (p -disubstituted benzene), 483.

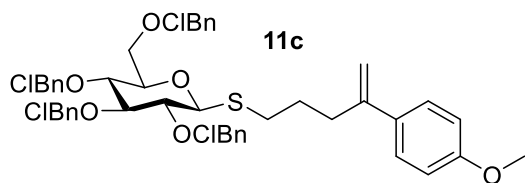
((*E*)-4-(4-methoxyphenyl)-3-butenyl β -D-1-thio-2,3,4,6-tetra-*O*-(4-trifluoromethyl)benzyl glucopyranoside) 10d



Prepared following the general procedure A. **Part 1**; used 0.71 g of tetraacetate thioglucoside **10**, 10.0 ml methanol, and 0.4ml of 5 M NaOMe. **Part 2**; used 20ml of DMF, 0.40g of TBAI, 0.42 g of NaH and 2.59 g of 4-trifluoromethylbenzyl bromide. The reaction crude product was purified by flash column chromatography (20% EOAc/hexanes) to give 1.01g of a white compound **10d** (76% yield). ^1H NMR (400 MHz, CDCl_3) δ 7.59-7.19 (m, 18H), 6.83 (d, $J = 8.7$ Hz, 2H), 6.39 (d, $J = 15.8$ Hz, 1H), 6.09 (dt, $J = 15.8$ Hz, 6.9 Hz, 1H), 4.99 (d, $J = 11.2$ Hz, 1H), 4.89 (d, $J = 12.1$ Hz, 1H), 4.85-4.77 (m, 2H), 4.73-4.56 (m, 4H), 4.52 (d, $J = 9.6$ Hz, 1H), 3.79 (s, 3H), 3.77-3.73 (m, 2H), 3.70-3.63 (m, 2H), 3.54-3.43 (m, 2H), 2.89 (m, 2H), 2.57 (dq, $J = 7.2$ Hz, 1.3 Hz, 2H) ^{13}C NMR (100 MHz, CDCl_3) δ 159.06, 142.28, 142.16, 141.94, 141.79, 131.06, 130.22, 130.14, 130.02, 129.82, 129.71, 127.96, 127.58, 127.39, 127.22, 127.19, 125.76, 125.60-125.16 (m), 122.78, 122.71, 114.00, 86.61, 85.24, 82.00, 78.93, 78.05, 74.67, 74.55, 74.02, 72.73, 69.23,

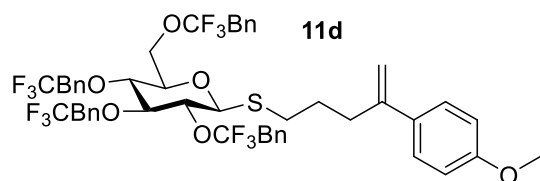
55.25, 55.21, 33.41, 30.98; **¹⁹F NMR (471 MHz, CDCl₃)** δ -63.06 (s, 3F), -63.13 (s, 3F), -63.15 (s, 3F), -63.18 (s, 3F). **HRMS (m/z)**: Calcd for C₄₉H₄₄F₁₂O₆SH 989.2745 found 989.2710. **[α]_D²⁵** = +4.5° (c = 1, DCM). **IR (cm⁻¹)** 2914, 1511, 1324, 1248, 1161, 1120, 1065, 1018, 967, 822 (*p*-disubstituted benzene), 592.

4-(4-methoxyphenyl)pent-4-enyl-2,3,4,6-tetra-O-(4-chlorobenzyl)-1-thio- β -D-glucopyranoside **11c**



Prepared following the general procedure A. **Part 1**; used 0.81 g of tetraacetate thioglucoside **11** 15.0 ml of MeOH, and 0.5 ml of 5 M NaOMe. **Part 2**; used 50 ml of DMF, 0.60 g of TBAI, 0.48 g of NaH, and 2.47 g of 4-chlorobenzyl bromide. The reaction crude product was purified by flash column chromatography (20% EOAc/hexanes) to give 0.80 g of a white compound **11c** (61% yield). **¹H NMR (400 MHz, CDCl₃)** δ 7.34-7.19 (m, 14H), 7.14 (d, *J* = 8.4 Hz, 2H), 7.04 (d, *J* = 8.4 Hz, 2H) 6.83 (d, *J* = 8.8 Hz, 2H), 5.21 (d, *J* = 1.4 Hz, 1H), 4.98 (d, *J* = 1.4 Hz, 1H), 4.85 (d, *J* = 10.6 Hz, 1H), 4.81-4.58 (m, 4H), 4.56-4.41 (m, 3H), 4.35 (d, *J* = 9.8 Hz, 1H), 3.78 (s, 3H), 3.62(d, *J* = 3.2 Hz, 2H), 3.58-3.52 (m, 2H), 3.39-3.31 (m, 2H), 2.81-2.63 (m, 2H), 2.62-2.55 (m, 2H), 1.82 (m, 2H) **¹³C NMR (100 MHz, CDCl₃)** δ 159.12, 146.59, 136.85, 136.59, 136.44, 136.36, 133.67, 133.62, 133.46, 133.44, 133.16, 129.47, 129.05, 128.97, 128.76, 128.61, 128.60, 128.57, 128.55, 127.20, 113.72, 111.41, 86.43, 85.33, 81.70, 78.91, 77.79, 74.74, 74.59, 74.07, 72.67, 68.85, 55.29, 34.25, 30.48, 28.33; **HRMS (m/z)**: Calcd for C₄₆H₄₆Cl₄O₆SH 867.1847 found 867.1778. **[α]_D²⁵** = +14.6° (c = 1, DCM). **IR (cm⁻¹)** 3053, 2912, 1602, 1264, 1086, 1015, 809 (*p*-disubstituted benzene), 734, 704.

4-(4-methoxyphenyl)pent-4-enyl-2,3,4,6-tetra-O-(4-trifluoromethyl)benzyl-1-thio- β -D-glucopyranoside **11d**



Prepared following the general procedure A. **Part 1**; used 1.97 g of tetraacetate thioglucoside **11**, 20.0 ml of methanol, and 0.5 ml of 5 M NaOMe. **Part 2**; 70 ml of DMF, 1.17 g of TBAI, 7.00 g of NaH, and 7.19 g of 4-trifluoromethylbenzyl bromide. The reaction crude product was purified by flash column chromatography (15% EOAc/hexanes) to give 2.82 g of a white compound **11d** (77% yield). **^1H NMR (400 MHz, CDCl_3) δ** 7.59-7.19 (m, 18H), 6.83 (d, J = 8.8 Hz, 2H), 5.22 (d, J = 1.5 Hz, 1H), 5.00-4.95 (m, 2H), 4.90-4.76 (m, 3H), 4.73-4.60 (m, 3H), 4.54 (d, J = 12.7 Hz, 1H), 4.40 (d, J = 9.8 Hz, 1H), 3.77 (s, 3H), 3.72-3.59 (m, 4H), 3.42 (m, 2H), 2.84-2.67 (m, 2H), 2.60 (m, 2H), 1.89-1.78 (m, 2H) **^{13}C NMR (100 MHz, CDCl_3) δ** 159.17, 146.57, 142.29, 142.19, 141.96, 141.83, 133.15, 130.15, 130.03, 129.83, 129.71, 127.96, 127.92, 127.58, 127.37, 127.20, 126.63, 125.60-125.19 (m), 122.84-122.64 (m), 113.71, 111.41, 86.60, 85.42, 81.96, 78.87, 77.98, 74.65, 74.51, 73.98, 72.69, 69.10, 55.23, 34.22, 30.56, 28.38; **^{19}F NMR (471 MHz, CDCl_3) δ** -63.06 (s, 3F), -63.14 (s, 3F), -63.15 (s, 3F), -63.19 (s, 3F). **HRMS (m/z)**: Calcd for $\text{C}_{50}\text{H}_{46}\text{F}_{12}\text{O}_6\text{SH}$ 1003.2902 found 1003.2870. **$[\alpha]_{\text{D}}^{25}$** = +7.9° (c = 1, DCM). **IR (cm^{-1})** 3054, 1325, 1264, 1164, 1165, 822 (p -disubstituted benzene), 734, 705.

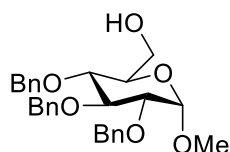
General glycosylation procedure B

To an oven dried 4 ml Wheaton vial with a stir bar, 1 equivalent of a hydroxyl bearing acceptor was added (0.075 mmol), followed by 2 equivalents of tetra-(4-X-benzyl) thioglucoside donor

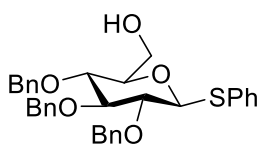
(0.15 mmol X = H,F,Cl or CF₃). Y ml (Y = 1 ml, 2.5 ml or 5 ml) of solvent (DCM or 1,4-dioxane) was added. The vial was capped with a septum then flushed with nitrogen gas. TfOH (10 mol% when using DCM and 40 mol% when using 1,4-dioxane) was added dropwise and magnetic stirring commenced. The reaction was monitored by TLC. Once all of the donor was consumed the reaction was quenched with 0.05 ml of Et₃N. The contents were then transferred to a R.B.F, and 2-3 g of silica gel were added, adsorption onto silica occurred by evaporating solvents under low vacuum. Products were purified by flash chromatography (12%-32% EtOAc/Hexanes). In some cases, a premix of (3:1 DCM/EtOAc) was made (referred to herein as DE). Thus, 12%-48% DE/hexanes solvent system was especially important in separating anomers.

Various acceptors used in the substrate scope study:

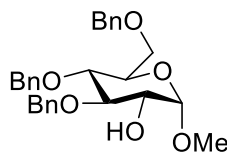
Acceptors utilized herein as well as references for their preparation are provided in the scheme below:



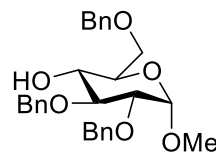
12
(synthesized according to reference 8)



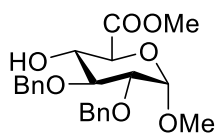
16cc
(synthesized according to reference 11)



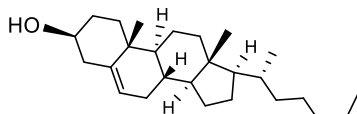
17cc
(synthesized according to reference 9)



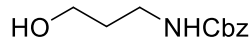
18cc
(synthesized according to reference 8)



19cc
(synthesized according to reference 10)



20cc
(purchased from Sigma-Aldrich)

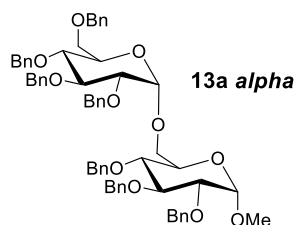


21cc
(synthesized according to reference 12)

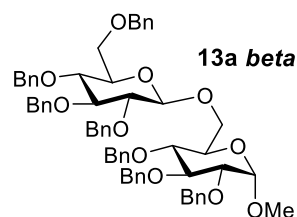
Glycosylation using glycosyl acceptor 12

Several glycosylation reactions were performed using glycosyl acceptor **12**. The general procedure B or slight modification thereof (apparent through study of Table 1 in the manuscript) was used. Reaction conditions, yields, and selectivity achieved can be found in Table 1. Below are characterization details of new compounds. ^1H NMR data for both anomers of compound **13a** (Table 1, entry 1) matched literature values.^{5,6} Tabulated ^1H NMR data for both anomers of **13a** from reference 5 is provided below.

Methyl (2,3,4,6-tetra-O-benzyl-D-glucopyranosyl)-(1→6)-2,3,4-tri-O-benzyl-D-glucopyranoside



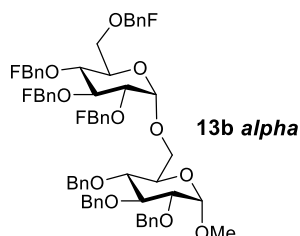
^1H NMR data for *alpha* anomer 13a *alpha*:⁵ (300 MHz, CDCl_3) δ 7.37 – 7.09 (m, 35H), 4.98 – 4.92 (m, 4H), 4.84 – 4.55 (m, 10H), 4.43 (t, $J = 11.9$ Hz, 1H), 4.00 (d, $J = 8.7$ Hz, 1H), 3.94 (d, $J = 9.2$ Hz, 1H), 3.85 – 3.62 (m, 7H), 3.59 – 3.52 (m, 2H), 3.44 (dd, $J = 9.6$, 3.8 Hz, 1H), 3.35 (s, 3H).



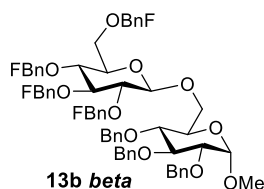
^1H NMR data for *beta* anomer 13a *beta*:⁵ (300 MHz, CDCl_3) δ 7.37 – 7.14 (m, 35H), 4.97 (dd, $J = 11.3$, 2.9 Hz, 1H), 4.90 (d, $J = 11.0$ Hz, 1H), 4.80 (d, $J = 11.0$ Hz, 1H), 4.78 (d, $J = 11.0$ Hz, 2H), 4.73 (d, $J = 3.2$ Hz, 1H), 4.68 (d, $J = 9.2$ Hz, 1H), 4.62 – 4.49 (m,

3H), 4.56 (d, $J = 7.5$ Hz, 2H), 4.35 (d, $J = 7.7$ Hz, 1H), 4.18 (dd, $J = 10.7, 1.9$ Hz, 1H), 3.99 (t, $J = 9.2$ Hz, 1H), 3.85 – 3.80 (m, 1H), 3.74 – 3.38 (m, 9H), 3.32 (s, 3H).

Methyl (2,3,4,6-tetra-O-(4-fluorobenzyl)-D-glucopyranosyl)-(1→6)-2,3,4-tri-O-benzyl-D-glucopyranoside

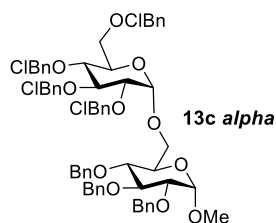


Data for *alpha* anomer 13b *alpha*: ^1H NMR (400 MHz, CDCl_3) δ 7.34-6.88 (m, 31H), 4.99-4.91 (m, 3H), 4.82 (t, $J = 10.8$ Hz, 2H), 4.75-4.51 (m, 9H), 4.37 (t, $J = 11.3$ Hz, 2H), 3.99 (t, $J = 9.2$ Hz, 1H), 3.91-3.82 (m, 2H), 3.78-3.63 (m, 4H), 3.60-3.46 (m, 4H), 3.42 (dd, $J = 9.6$ Hz, 3.6 Hz, 1H), 3.35 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 163.33, 163.29, 163.23, 161.38, 161.33, 161.28, 138.69, 138.40, 138.10, 134.51, 134.49, 134.13, 134.11, 134.04, 134.01, 133.64, 133.62, 129.62, 129.56, 129.50, 129.36, 129.30, 129.20, 129.13, 129.04, 128.45, 128.43, 128.39, 128.23, 128.07, 127.98, 127.93, 127.69, 127.66, 115.34, 115.30, 115.24, 115.17, 115.13, 115.07, 98.05, 97.05, 82.12, 81.45, 80.09, 79.93, 77.68, 77.48, 75.83, 74.95, 74.67, 74.06, 73.39, 72.70, 71.55, 70.36, 70.17, 68.33, 68.99, 65.99, 55.19; ^{19}F NMR (471 MHz, CDCl_3) δ -115.21 (m, 2F), -115.31 (m, 1F), -115.47 (m, 1F). **HRMS (m/z):** Calcd for $\text{C}_{62}\text{H}_{62}\text{F}_4\text{O}_{11}\text{K}$ 1097.3865 found 1097.3890. $[\alpha]_{\text{D}}^{25} = +15.1^\circ$ ($c = 1$, DCM). **IR (cm^{-1})** 3034, 2920, 1603, 1509, 1262, 1222, 1136, 1072, 1049, 1026, 1015, 822 (*p*-disubstituted benzene), 735, 698.

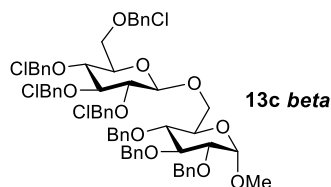


Data for *beta* anomer 13b *beta*: **^1H NMR** (400 MHz, CDCl_3) δ 7.36-6.93 (m, 29H), 6.87-6.84 (t, J = 8.6 Hz, 2H), 4.97 (d, J = 10.8 Hz, 1H), 4.91 (d, J = 11.2 Hz, 1H), 4.81-4.61 (m, 8H), 4.59 (d, J = 3.5 Hz, 1H), 4.56-4.45 (m, 4H), 4.31 (d, J = 7.7 Hz, 1H), 4.16 (d, J = 10.9 Hz, 1H), 4.00 (t, J = 9.2 Hz, 1H), 3.84-3.81 (m, 1H), 3.67-3.62 (m, 3H), 3.57-3.48 (m, 4H), 3.46-3.35 (m, 2H), 3.32 (s, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 163.35, 163.29, 163.27, 163.23, 161.39, 161.33, 161.31, 161.28, 138.71, 138.27, 138.09, 134.25, 134.23, 134.06, 134.04, 133.87, 133.85, 133.82, 129.61, 129.55, 129.52, 129.45, 129.44, 129.37, 129.30, 128.49, 128.42, 128.40, 128.16, 128.03, 129.99, 127.68, 127.63, 115.36, 115.32, 115.30, 115.29, 119.19, 115.15, 115.13, 115.12, 103.79, 98.10, 84.53, 81.94, 81.85, 79.82, 78.02, 77.76, 75.79, 74.94, 74.89, 74.84, 74.16, 74.05, 73.38, 72.73, 69.83, 68.76, 68.70, 55.24; **^{19}F NMR (471 MHz, CDCl_3)** δ -114.98 (m, 1F), -115.27 (m, 2F), -115.39 (m, 1F). **HRMS (m/z):** Calcd for $\text{C}_{62}\text{H}_{62}\text{F}_4\text{O}_{11}\text{K}$ 1097.3865 found 1097.3882. **$[\alpha]_D^{25}$** = +25.1° (c = 1, DCM). **IR (cm^{-1})** 3055, 2918, 2850, 1510, 1260, 1223, 1067, 1013, 793 (*p*-disubstituted benzene), 736, 700, 459.

Methyl (2,3,4,6-tetra-*O*-(4-chlorobenzyl)-D-glucopyranosyl)-(1→6)-2,3,4-tri-*O*-benzyl-D-glucopyranoside



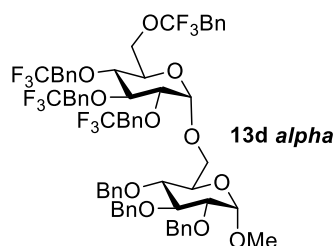
Data for *alpha* anomer 13c *alpha*: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.34-7.13 (m, 29H), 6.99 (d, J = 8.4 Hz, 2H), 4.99-4.91 (m, 3H), 4.80 (dd, J = 11.1 Hz, 6.5 Hz, 2H), 4.73-4.5 (m, 9H), 4.36 (t, J = 11.1 Hz, 2H), 3.98 (t, J = 9.2 Hz, 1H), 3.89-3.82 (m, 3H), 3.77-3.63 (m, 4H), 3.59-3.44 (m, 4H), 3.38 (dd, J = 9.6 Hz, 3.6 Hz, 1H), 3.34 (s, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 138.68, 138.40, 138.10, 137.12, 136.75, 136.71, 136.32, 133.55, 133.44, 133.37, 133.32, 129.16, 129.04, 128.87, 128.66, 128.58, 128.54, 128.47, 128.45, 128.41, 128.10, 127.97, 127.92, 127.71, 127.69, 127.71, 127.69, 127.65, 98.06, 97.01, 82.10, 81.46, 80.10, 79.99, 77.62, 77.51, 75.84, 74.94, 74.52, 73.94, 73.40, 72.63, 71.45, 70.45, 70.36, 70.13, 68.37, 65.96, 55.19; **HRMS (m/z):** Calcd for $\text{C}_{62}\text{H}_{62}\text{Cl}_4\text{O}_{11}\text{K}$ 1161.2683, found 1161.2683. $[\alpha]_{\text{D}}^{25}$ = +22.4° (c = 1, DCM). **IR (cm^{-1})** 2921, 1491, 1359, 1264, 1058, 1014, 806 (*p*-disubstituted benzene), 734, 699.



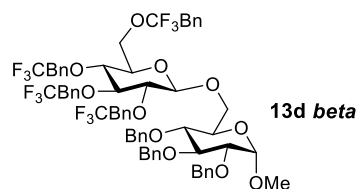
Data for 13c *beta*: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.35-7.02 (m, 31H), 4.96 (d, J = 10.8 Hz, 1H), 4.90 (d, J = 11.4 Hz, 1H), 4.80-4.73 (m, 4H), 4.69-4.44 (m, 9H), 4.30 (d, J = 7.7 Hz, 1H), 4.13-4.09 (m, 1H), 3.99 (t, J = 9.2 Hz, 1H), 3.84-3.80 (m, 1H), 3.67-3.64 (m, 3H), 3.55-3.45 (m, 4H), 3.43-3.35 (m, 2H), 3.32 (s, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 138.69, 138.25, 138.08, 136.84, 136.66, 136.57, 136.47, 133.59, 133.42, 133.40, 133.38, 129.11, 128.95, 128.93, 128.84, 128.57, 128.53, 128.52, 128.49, 128.44, 128.40, 128.15, 128.04, 127.99, 127.69, 127.63, 103.75, 98.11, 84.52, 81.93, 81.84, 79.82, 78.00, 77.78, 75.80, 74.87, 74.70, 74.05, 73.91, 73.39, 72.63, 69.82, 68.77,

68.70, 55.27; **HRMS (m/z)**: Calcd for $C_{62}H_{62}Cl_4O_{11}K$ 1161.2683 found 1161.2689. $[\alpha]_D^{25} = +50.2^\circ$ (c = 1, DCM). **IR (cm⁻¹)** 3030, 2911, 1491, 1357, 1066, 1014, 804 (*p*-disubstituted benzene), 736, 696.

Methyl (2,3,4,6-tetra-O-(4-trifluoromethylbenzyl)-D-glucopyranosyl)-(1→6)-2,3,4-tri-O-benzyl-D-glucopyranoside



Data for *alpha* anomer 13d *alpha*: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.85-7.25 (m, 29H), 7.17 (d, $J = 8.0$ Hz, 2H), 5.05 (d, $J = 3.5$ Hz, 1H), 4.98-4.89 (m, 3H), 4.81-4.65 (m, 6H), 4.62-4.43 (m, 6H), 4.00 (t, $J = 9.1$ Hz, 1H), 3.95-3.86 (m, 2H), 3.80-3.76 (m, 2H), 3.72-3.67 (m, 2H), 3.66-3.50 (m, 4H), 3.39 (dd, $J = 9.6$ Hz, 3.6 Hz, 1H), 3.36 (s, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 142.77, 142.39, 142.33, 142.08, 138.78, 138.59, 138.23, 132.20, 130.29, 130.17, 130.07, 130.01, 129.91, 129.81, 128.66, 128.64, 128.60, 128.32, 128.12, 127.97, 127.91, 127.86, 127.77, 127.72, 127.62, 127.51, 127.29, 125.61, 125.58, 125.54, 125.51, 125.48, 125.46, 125.45, 125.43, 125.42, 125.34, 125.31, 123.18, 114.53, 98.31, 97.13, 82.29, 81.94, 80.47, 80.33, 77.90, 77.77, 76.10, 75.13, 74.68, 74.11, 73.59, 72.85, 71.65, 70.52, 70.31, 68.91, 66.25, 55.40; $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -63.08 (s, 3F), -63.21 (s, 3F), -63.22 (s, 3F), -63.26 (s, 3F). **HRMS (m/z)**: Calcd for $C_{66}H_{62}F_{12}O_{11}K$ 1297.3738 found 1297.3758. $[\alpha]_D^{25} = +11.5^\circ$ (c = 1, DCM). **IR (cm⁻¹)** 3055, 2926, 1325, 1264, 1163, 1123, 1065, 1017, 822 (*p*-disubstituted benzene), 734, 702.

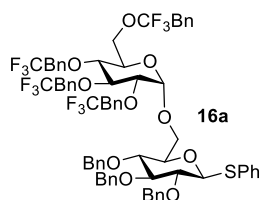


Data for *beta* anomer 13d *beta*: **^1H NMR** (400 MHz, CDCl_3) δ 7.60-7.14 (m, 31H), 5.02 (d, $J = 12.0$ Hz, 1H), 4.96 (d, $J = 10.8$ Hz, 1H), 4.91-4.56 (m, 12H), 4.51 (dd, 12.3 Hz, 6.4 Hz, 2H), 4.36 (d, $J = 7.7$ Hz, 1H), 4.16 (dd, $J = 10.8$ Hz, 2.0 Hz, 1H), 4.00 (t, $J = 9.2$ Hz, 1H), 3.86-3.81 (m, 1H), 3.73-3.66 (m, 3H), 3.62-3.55 (m, 2H), 3.51-3.38 (m, 4H), 3.32 (s, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 142.46, 142.34, 142.25, 142.14, 138.79, 138.38, 138.24, 130.42-129.83 (m), 128.68, 128.62, 128.59, 128.33, 128.24, 128.19, 127.93, 127.90, 127.84, 127.72, 127.67, 127.56, 127.52, 127.47, 127.45, 125.65-125.4 (m), 123.17, 103.92, 98.34, 84.90, 82.19, 82.10, 80.02, 78.22, 78.16, 76.03, 75.06, 74.99, 74.83, 74.18, 74.02, 73.58, 72.83, 69.99, 69.19, 68.99, 55.43; **^{19}F NMR (471 MHz, CDCl_3)** δ -63.11 (s, 3F), -63.15 (s, 3F), -63.26 (s, 3F), -63.28 (s, 3F). **HRMS (m/z):** Calcd for $\text{C}_{66}\text{H}_{62}\text{F}_{12}\text{O}_{11}\text{K}$ 1297.3738 found 1297.3772. **$[\alpha]_{\text{D}}^{25}$** = -0.6° ($c = 1$, DCM). **IR (cm^{-1})** 3063, 2917, 1620, 1324, 1264, 1162, 1121, 1065, 1017, 912, 821 (*p*-disubstituted benzene), 736, 670.

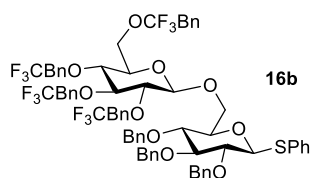
Glycosylation using **16cc** as the acceptor

Following the general procedure B, 5.0 ml of 1,4-dioxane used, 154.9 mg (0.1544 mmol) of donor **11d**, 42.2 mg (0.0778 mmol) of alcohol **16cc** and 5.0 μL (0.057 mmol) of TfOH were used. Purified by flash column chromatography (12-28% DE/hexanes) to give 103.2 mg of a colorless oil **16**, yield 99%, *alpha:beta* ratio 7.2:1

Phenylthio-(2,3,4,6-tetra-O-(4-trifluoromethylbenzyl)-D-glucopyranosyl)-(1→6)-2,3,4-tri-O-benzyl-β-D-glucopyranoside (16)

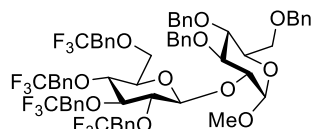


Data for *alpha* anomer 16a: ^1H NMR (500 MHz, CDCl_3) δ 7.57-7.16 (m, 36H), 5.16 (d, J = 3.5 Hz, 1H), 4.98(d, J = 4.2Hz, 1H), 4.94-4.89(m, 2H), 4.86(d, J = 10.9 Hz, 1H), 4.84-4.76(m, 3H), 4.72-4.61(m, 5H), 4.58-4.51(m, 2H), 4.48(d, J = 12.7 Hz, 1H), 3.97(t, J = 9.2 Hz, 1H), 3.93-3.79(m, 3H), 3.74-3.55(m, 6H), 3.47(m, 1H). 3.09(t, J = 8.6Hz, 1H), ^{13}C NMR (125 MHz, CDCl_3) δ 142.63, 142.22, 141.92, 138.31, 138.15, 137.87, 136.87, 134.20, 131.55, 130.31-129.44(m), 129.02, 128.47, 128.44, 128.38, 128.04, 127.85, 127.74, 127.73, 127.65, 127.44, 127.34, 127.22, 127.08, 125.52-125.10(m), 97.09, 88.21, 86.55, 81.80, 81.15, 80.39, 78.77, 77.77, 75.79, 75.41, 74.94, 74.58, 73.90, 72.64, 71.29, 70.10, 68.84, 66.09, ^{19}F NMR (471 MHz, CDCl_3) δ -62.94 (s, 3F), -63.09 (s, 3F), -63.11 (s, 3F), -63.15 (s, 3F). HRMS (m/z): Calcd for $\text{C}_{71}\text{H}_{64}\text{F}_{12}\text{O}_{10}\text{SNa}$ 1359.3921 found 1359.3868. $[\alpha]_{\text{D}}^{25}$ = -49.5 (c = 0.5, DCM). IR (cm^{-1}) 3032, 2922, 2185, 1741, 1620, 1326, 1261, 1162, 1122, 1066, 1018, 821 (p -disubstituted benzene), 745, 698, 643.



Data for *beta* anomer 16b: ^1H NMR (500 MHz, CDCl_3) δ 7.64-7.19 (m, 36H), 5.02 (d, J = 12.3 Hz, 1H), 4.96-4.90 (m, 3H), 4.87-4.54 (m, 11H), 4.45 (d, J = 7.8 Hz, 1H), 4.21 (d, J = 11.0 Hz, 1H), 3.83-3.42(m, 11H) ^{13}C NMR (125 MHz, CDCl_3) δ 142.36, 142.20, 142.09, 142.02, 138.22, 137.91, 134.13, 131.05, 130.26-129.45(m), 128.91, 128.49, 128.46, 128.44, 128.42, 128.15,

80.69, 79.53, 77.93, 77.64, 75.94, 75.05, 74.71, 74.55, 73.88, 73.60, 72.61, 71.53, 70.26, 69.98, 68.37, 54.87; **¹⁹F NMR (471 MHz, CDCl₃)** δ -63.12 (s, 3F), -63.15 (s, 3F), -63.17 (s, 3F), -63.18 (s, 3F). **HRMS (m/z)**: Calcd for C₆₆H₆₂F₁₂O₁₁K 1297.3738 found 1297.3714. **[α]_D²⁵** = +45.2° (c = 1, DCM). **IR (cm⁻¹)** 3032, 2914, 1324, 1162, 1120, 1064, 1018, 851, 822 (*p*-disubstituted benzene), 736, 699.



17b

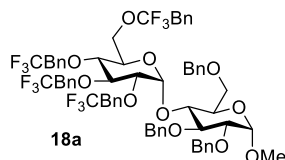
Data for *beta* anomer 17b: **¹H NMR (400 MHz, CDCl₃)** δ 7.58 (d, *J* = 8.0 Hz, 2H), 7.51-7.13 (m, 27H), 7.06 (m, 2H), 5.03 (d, *J* = 12.3 Hz, 1H), 4.95 (d, *J* = 3.5 Hz, 1H), 4.85 (d, *J* = 12.0 Hz, 1H), 4.78-4.44 (m, 13H), 4.02 (t, *J* = 9.8 Hz, 1H), 3.85 (dd, *J* = 9.8 Hz, 3.5 Hz, 1H), 3.82-3.65 (m, 6H), 3.64-3.57 (m, 2H), 3.51 (t, *J* = 8.1 Hz, 1H), 3.44 (m, 1H), 3.40 (s, 3H); **¹³C NMR (125 MHz, CDCl₃)** δ 142.25, 142.11, 141.92, 141.88, 138.39, 138.00, 137.83, 130.12-129.36(m), 128.42, 128.37, 128.26, 127.99, 127.79, 127.77, 127.74, 127.66, 127.56, 127.46, 127.36, 127.30, 127.22, 125.44-125.00(m), 122.99, 104.05, 99.66, 84.78, 81.96, 81.57, 78.93, 78.31, 77.91, 75.08, 75.02, 74.55, 74.53, 73.96, 73.66, 73.60, 72.79, 70.03, 69.11, 68.42, 55.12; **¹⁹F NMR (471 MHz, CDCl₃)** δ -63.13 (s, 3F), -63.17 (s, 3F), -63.22 (s, 3F), -63.24 (s, 3F). **HRMS (m/z)**: Calcd for C₆₆H₆₂F₁₂O₁₁K 1297.3738 found 1297.3718. **[α]_D²⁵** = +27.2° (c = 1, DCM). **IR (cm⁻¹)** 3054, 2927, 1325, 1264, 1165, 1018, 895, 823 (*p*-disubstituted benzene), 734, 703.

Glycosylation using acceptor **18cc**

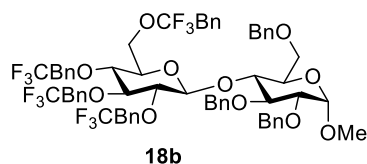
Following the general procedure B, 5.0 ml of 1,4-dioxane, 182.7 mg of thioglucoside donor **11d** (0.1822 mmol), 35.8 mg (0.0771 mmol) of acceptor **18cc** and 5.0 μ L of TfOH (0.057 mmol) were

used. Purified by flash column chromatography (12-36% DE/hexanes) to give 70.7 mg (73%) of a colorless thick oil **18**, *alpha:beta* ratio = 5.0:1.

Methyl (2,3,4,6-tetra-O-(4-trifluoromethylbenzyl)-D-glucopyranosyl)-(1→4)-2,3,6-tri-O-benzyl--D-glucopyranoside (18)



Data for *alpha* anomer 18a : ^1H NMR (400 MHz, CDCl_3) δ 7.51-7.13 (m, 31H), 5.78 (d, J = 3.6 Hz, 1H), 5.10 (d, J = 11.8 Hz, 1H), 4.87 (d, J = 12.0 Hz, 1H), 4.76-4.70 (m, 4H), 4.65 (d, J = 3.7 Hz, 1H), 4.60-4.46 (m, 7H), 4.30 (d, J = 12.8 Hz, 1H), 4.10-4.08 (m, 2H), 3.92-3.86 (m, 3H), 3.74-3.77 (m, 1H), 3.70-3.61 (m, 3H), 3.46-3.42 (m, 2H), 3.40 (s, 3H), 3.39-3.36 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 142.52, 142.16, 141.89, 141.77, 138.85, 138.16, 137.81, 130.31-129.30 (m), 128.49, 128.33, 128.27, 128.18, 128.02, 127.82, 127.50, 127.39, 127.26, 127.24, 127.16, 127.14, 126.39, 125.34-125.12 (m), 122.97, 122.95, 97.73, 96.32, 82.08, 81.95, 80.23, 79.73, 77.78, 74.43, 74.31, 73.97, 73.36, 73.27, 72.68, 72.36, 72.11, 70.77, 69.62, 69.14, 68.48, 55.31; ^{19}F NMR (471 MHz, CDCl_3) δ -63.08 (s, 3F), -63.09 (s, 3F), -63.12 (s, 3F), -63.15 (s, 3F). **HRMS (m/z):** Calcd for $\text{C}_{66}\text{H}_{62}\text{F}_{12}\text{O}_{11}\text{Na}$ 1281.3998 found 1281.3996. $[\alpha]_{\text{D}}^{25}$ = +39.8° (c = 1, DCM). **IR (cm^{-1})** 3059, 2914, 2867, 1620, 1324, 1266, 1161, 1121, 1065, 1017, 850, 822 (*p*-disubstituted benzene), 735, 699, 593.

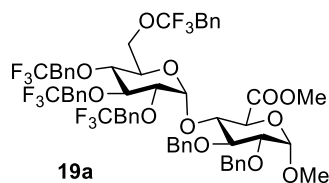


Data for *beta* anomer 18b: ^1H NMR (400 MHz, CDCl_3) δ 7.53-7.17 (m, 31H), 5.05 (d, J = 11.4 Hz, 1H), 4.87-4.58 (m, 12H), 4.45-4.35 (m, 4H), 3.98 (t, J = 9.5 Hz, 1H), 3.85 (t, J = 9.3 Hz, 1H), 3.79 (dd, J = 10.8 Hz, 2.9 Hz, 1H), 3.70 (dd, J = 11.2 Hz, 1.9 Hz, 1H), 3.60-3.38 (m, 6H), 3.36 (s, 3H), 3.31-3.27 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 142.43, 142.35, 142.30, 142.10, 139.42, 138.22, 137.78, 130.05-129.40 (m), 128.53, 128.40, 128.10, 128.04, 128.01, 127.91, 127.88, 127.86, 127.36, 127.32, 127.24, 127.22, 127.15, 127.13, 125.33-125.13 (m), 122.95, 102.19, 98.40, 84.75, 82.79, 80.29, 79.03, 78.25, 76.27, 75.42, 75.00, 74.52, 73.84, 73.82, 73.59, 73.57, 72.54, 69.96, 69.01, 67.85, 55.46; ^{19}F NMR (471 MHz, CDCl_3) δ -62.98 (s, 3F), -63.06 (s, 3F), -63.14 (s, 3F), -63.15 (s, 3F). HRMS (m/z): Calcd for $\text{C}_{66}\text{H}_{62}\text{F}_{12}\text{O}_{11}\text{Na}$ 1281.3998 found 1281.3990. $[\alpha]_{\text{D}}^{25}$ = +25.6° (c = 1, DCM). IR (cm^{-1}) 3056, 2916, 1324, 1265, 1163, 1122, 1065, 1018, 822 (*p*-disubstituted benzene), 736, 701.

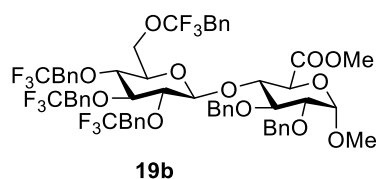
Glucuronate **19cc** as the acceptor

Following the general procedure **B**. 5.0 ml of 1,4-dioxane, 167.7 mg (0.1672 mmol) of thioglucoside donor **11d**, 31.0 mg (0.0770 mmol) of acceptor **19cc**, and 5.0 μL (0.057 mmol) of TfOH were used. Purified by flash column chromatography (12-36% DE/hexanes) to give 65.7 mg of a colorless oil **19** (71%), *alpha:beta* ratio 4.2:1.

Methyl (2,3,4,6-tetra-*O*-(4-trifluoromethylbenzyl)-D-glucopyranosyl)-1 $\rightarrow$ 4-(2,3-di-*O*-benzyl)-D-glucopyranuronic acid methyl ester (19)



Data for *alpha* anomer 19a: ^1H NMR (400 MHz, CDCl_3) δ 7.54 (d, J = 8.0 Hz, 2H), 7.47 (m, 4H), 7.41 (d, J = 8.1 Hz, 4H), 7.27-7.23 (m, 10H), 7.19-7.14 (m, 6H), 5.58 (d, J = 3.6 Hz, 1H), 5.02 (d, J = 11.7 Hz, 1H), 4.89 (d, J = 12.0 Hz, 1H), 4.82-4.45 (m, 11H), 4.25 (d, J = 9.5 Hz, 1H), 4.12 (t, J = 9.1 Hz, 1H), 4.05 (t, J = 9.0 Hz, 1H), 3.89 (t, J = 9.4 Hz, 1H), 3.75-3.70 (m, 1H), 3.69 (s, 3H), 3.66-3.60 (m, 3H), 3.51 (d, J = 9.9 Hz, 1H), 3.46-3.43 (m, 1H), 3.42 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.05, 142.39, 142.17, 141.86, 141.74, 138.65, 137.61, 130.40-129.38 (m), 128.54, 128.31, 128.23, 128.14, 127.85, 127.32, 127.30, 127.00, 126.45, 125.39-125.09 (m), 122.95, 98.50, 97.09, 81.49, 80.98, 79.56, 79.20, 77.50, 75.62, 74.82, 74.47, 73.90, 73.53, 72.79, 72.00, 70.75, 70.29, 68.22, 55.81, 52.54; ^{19}F NMR (471 MHz, CDCl_3) δ -63.15 (s, 6F), -63.20 (s, 3F), -63.22 (s, 3F). HRMS (m/z): Calcd for $\text{C}_{60}\text{H}_{56}\text{F}_{12}\text{O}_{12}\text{K}$ 1235.3217 found 1235.3207. $[\alpha]_D^{25}$ = +37.5° (c = 1, DCM). IR (cm^{-1}) 3057, 2927, 1749, 1324, 1265, 1163, 1122, 1064, 1018, 823 (*p*-disubstituted benzene), 735, 702.



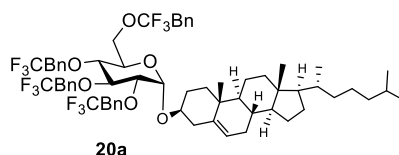
Data for *beta* anomer 19b: ^1H NMR (400 MHz, CDCl_3) δ 7.51-7.21 (m, 26H), 5.03 (d, J = 11.4 Hz, 1H), 4.93 (d, J = 12.1 Hz, 1H), 4.87 (d, J = 12.1 Hz, 1H), 4.83-4.71 (m, 4H), 4.66-4.57 (m, 4H), 4.47-4.38 (m, 3H), 4.17-4.09 (m, 2H), 3.90 (t, J = 8.9 Hz, 1H), 3.74-3.64 (m, 2H), 3.62 (s, 3H), 3.60-3.53 (m, 2H), 3.50 (dd, J = 9.5 Hz, 3.7 Hz, 1H), 3.48-3.43 (m, 1H), 3.41 (s, 3H), 3.39-3.35 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.05, 142.34, 141.97, 139.09, 137.95, 130.39-129.17 (m), 128.47, 128.36, 128.08, 128.00, 127.89, 127.82, 127.75, 127.35, 127.30, 127.26, 127.11, 127.04, 125.42-125.07 (m), 122.98, 102.19, 98.83, 84.73, 82.59, 79.57, 78.63, 78.14, 77.61, 75.51, 75.01, 74.60, 74.01, 73.83, 73.80, 72.50, 69.90, 68.94, 55.92, 52.63; ^{19}F NMR (471 MHz, CDCl_3) δ -63.06 (s, 3F), -63.15 (s, 3F), -63.22 (s, 6F). HRMS (m/z): Calcd for

C₆₀H₅₆F₁₂O₁₂K 1235.3217 found 1235.3200. $[\alpha]_D^{25} = +24.8^\circ$ (c = 1, DCM). IR (cm⁻¹) 3061, 2922, 1747, 1620, 1324, 1265, 1162, 1120, 1065, 1018, 850, 822 (*p*-disubstituted benzene), 736, 700.

Glycosylation using Cholesterol (20cc) as the acceptor

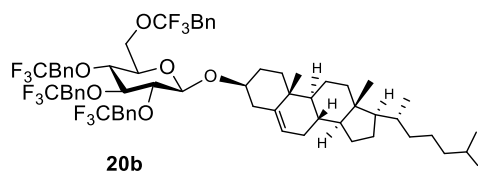
Following the general procedure B. 5.0 mL of 1,4-dioxane, 154.9 mg (0.1544 mmol) of donor **11d**, 29.7 mg (0.0768 mmol) of cholesterol **20cc** and 5.0 μ L (0.057 mmol) of TfOH were used. Purified by flash column chromatography twice since the first purification still had substantial impurity (12-36% DE/hexanes) to give 60.4 mg of a white compound **20** (some product deliberately not collected due to impurity contamination), 67%, *alpha:beta* ratio (after first column) 9.6:1.

cholesteryl 2,3,4,6-tetra-*O*-(4-trifluoromethylbenzyl)-D-glucopyranoside (**20**)



Data for *alpha* anomer 20a: ¹H NMR (500 MHz, CDCl₃) δ 7.56-7.50 (m, 8H), 7.43-7.40 (m, 4H), 7.35 (d, *J* = 8.0 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 2H), 5.29-5.27 (m, 1H), 5.05 (d, *J* = 3.7 Hz, 1H), 5.00 (d, *J* = 12.0 Hz, 1H), 4.81 (t, *J* = 11.9 Hz, 2H), 4.71 (s, 2H), 4.65 (d, *J* = 12.7 Hz, 1H), 4.56 (d, *J* = 11.9 Hz, 1H), 4.50 (d, *J* = 12.7 Hz, 1H), 4.00 (t, *J* = 9.2 Hz, 1H), 3.92 (m, 1H), 3.75 (dd, *J* = 10.7 Hz, 3.8 Hz 1H), 3.69-3.61 (m, 2H), 3.55 (dd, *J* = 9.6 Hz, 3.7 Hz, 1H), 3.46 (m, 1H), 2.44 (t, *J* = 13.3 Hz, 1H), 2.31 (ddd, *J* = 13.2 Hz, 5.0 Hz, 2.0 Hz, 1H), 2.02 (dt, *J* = 16.2 Hz, 3.5 Hz, 1H), 1.99-1.92 (m, 1H), 1.91-1.79 (m, 3H), 1.62-0.96 (m, 20H), 1.01 (s, 3H), 0.92 (d, *J* = 6.6 Hz, 4H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.68 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 142.72, 142.04, 141.91, 140.56, 130.07, 129.82, 127.87, 127.68, 127.32, 125.42-125.04 (m), 122.03, 94.51, 82.10, 80.29, 78.06, 74.57, 74.09, 72.67, 71.92, 70.04, 68.97, 56.75, 56.16, 50.14, 42.33,

40.00, 39.76, 39.53, 37.06, 36.78, 36.20, 35.80, 31.90, 31.88, 28.24, 28.03, 27.72, 24.27, 23.84, 22.83, 22.57, 21.07, 19.38, 18.73, 11.86; **¹⁹F NMR (471 MHz, CDCl₃)** δ -63.07 (s, 3F), -63.09 (s, 3F), -63.10 (s, 3F), -63.17 (s, 3F). **HRMS (m/z):** Calcd for C₆₅H₇₆F₁₂O₆Cl 1215.5144 found 1215.5188. **[α]_D²⁵** = +26.6° (c = 1, DCM). **IR (cm⁻¹)** 2935, 2868, 1620, 1324, 1210, 1109, 1063, 1016, 822 (*p*-disubstituted benzene), 738, 592

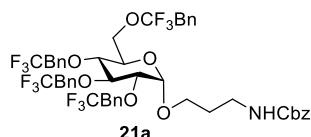


Data for *beta* anomer 20b: **¹H NMR (500 MHz, CDCl₃)** δ 7.56 (d, *J* = 8.0 Hz, 2H), 7.53-7.48 (m, 6H), 7.43 (d, *J* = 8.0 Hz, 2H), 7.39 (d, *J* = 8.0 Hz, 2H), 7.28 (d, *J* = 8.0 Hz, 2H), 7.24 (d, *J* = 8.0 Hz, 2H), 5.33 (d, *J* = 5.4 Hz, 1H), 5.01 (d, *J* = 12.0 Hz, 1H), 4.89 (d, *J* = 12.2 Hz, 1H), 4.80 (d, *J* = 12.0 Hz, 1H), 4.76 (d, *J* = 12.1 Hz, 1H), 4.72 (d, *J* = 12.0 Hz, 1H), 4.67-4.57 (m, 3H), 4.52 (d, *J* = 7.8 Hz, 1H), 3.71 (m, 2H), 3.64-3.55 (m, 3H), 3.47-3.41 (m, 2H), 2.36-2.25 (m, 2H), 2.04-1.95 (m, 3H), 1.88-1.79 (m, 2H), 1.68-0.95 (m, 20H), 1.01 (s, 3H), 0.92 (d, *J* = 6.5 Hz, 4H), 0.87 (d, *J* = 6.7 Hz, 3H), 0.86 (d, *J* = 6.7 Hz, 3H), 0.68 (s, 3H); **¹³C NMR (125 MHz, CDCl₃)** δ 142.39, 142.23, 142.01, 130.08-129.68 (m), 127.88, 127.53, 127.37, 127.30, 125.33, 125.31, 125.28, 125.25, 125.22, 123.01, 122.94, 122.22, 102.08, 84.75, 82.28, 79.77, 78.07, 74.57, 73.97, 73.77, 72.60, 69.15, 56.75, 56.16, 53.42, 50.21, 42.34, 39.75, 39.53, 39.07, 37.29, 36.76, 36.20, 35.80, 31.93, 31.88, 30.01, 29.72, 28.24, 28.03, 24.29, 23.84, 22.83, 22.71, 22.67, 22.57, 21.06, 19.38, 18.73, 14.13, 11.87; **¹⁹F NMR (471 MHz, CDCl₃)** δ -63.04 (s, 3F), -63.08 (s, 3F), -63.13 (s, 3F), -63.16 (s, 3F). **HRMS (m/z):** Calcd for C₆₅H₇₆F₁₂O₆Cl 1215.5144 found 1215.5186. **[α]_D²⁵** = +34.5° (c = 1, DCM). **IR (cm⁻¹)** 2936, 1325, 1264, 1165, 1123, 1065, 1018, 823 (*p*-disubstituted benzene), 735, 704.

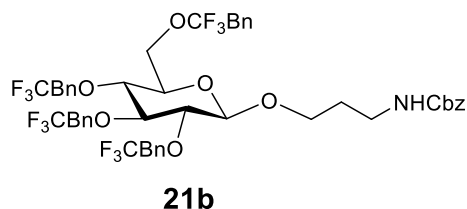
Glycosylation using 21cc as the acceptor

Following the general procedure B, 5.0 ml of 1,4-dioxane used, 155.8 mg (0.1553 mmol) of donor **16b**, 16.6 mg (0.0793 mmol) of alcohol **21cc** and 5.0 μ L (0.057 mmol) of TfOH were used. Purified by flash column chromatography (12-40% EtOAc/hexanes) to give 76.6 mg of a colorless oil **21** yield 96%, *alpha:beta* ratio 5.5:1

(3'-(N-carbobenzyloxy) propyl) 2,3,4,6-tetra-O-(4-trifluoromethylbenzyl)-D-glucopyranoside (21)

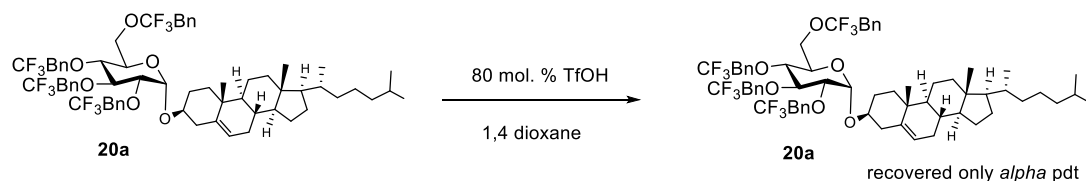


Data for *alpha* anomer 21a: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.59-7.17 (m, 21H), 5.76 (t, J = 5.7 Hz, 1H), 5.04 (s, 2H), 4.86-4.76 (m, 3H), 4.71-4.58 (m, 4H), 4.51 (m, 2H), 3.99-3.86 (m, 2H), 3.80-3.74 (d, J = 9.6 Hz, 1H), 3.70-3.51 (m, 5H), 3.51-3.42 (m, 1H), 3.31-3.21 (m, 1H), 1.85 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 156.50, 142.42, 141.94, 141.79, 136.58, 130.38-128.68 (m), 128.43, 128.05, 128.01, 127.82, 127.74, 127.68, 127.41, 127.25, 125.46-125.08 (m), 122.92, 97.04, 82.15, 80.08, 77.72, 74.63, 74.05, 72.68, 72.26, 70.41, 68.74, 67.69, 66.59, 39.71, 29.19; $^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -63.16 (s, 9F), -63.23 (s, 3F). **HRMS** (m/z): Calcd for $\text{C}_{49}\text{H}_{46}\text{F}_{12}\text{NO}_8$ 1004.3032 found 1004.3061. $[\alpha]_{\text{D}}^{25}$ = +26.6° (c = 1, DCM). **IR** (cm^{-1}) 3054, 2917, 1716, 1516, 1324, 1264, 1163, 1123, 1065, 1018, 823 (*p*-disubstituted benzene), 735, 703.

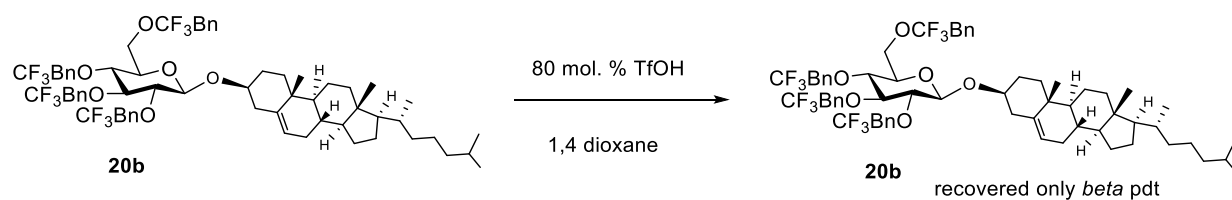


Data for *beta* anomer 21b: **¹H NMR** (400 MHz, CDCl₃) δ 7.56-7.26 (m, 19H), 7.18 (d, *J* = 8.0 Hz, 2H), 5.29 (t, *J* = 5.9 Hz, 1H), 5.06 (d, *J* = 4.7 Hz, 2H), 4.95 (d, *J* = 12.0 Hz, 1H), 4.88 (d, *J* = 12.1 Hz, 1H), 4.76 (m, 2H), 4.69 (d, *J* = 12.0 Hz, 1H), 4.59-4.54 (m, 2H), 4.48 (d, *J* = 12.8 Hz, 1H), 4.40 (d, *J* = 7.7 Hz, 1H), 3.97-3.91 (m, 1H), 3.73-3.64 (m, 3H), 3.63-3.56 (m, 2H), 3.50-3.27 (m, 4H), 1.83 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 156.43, 142.30, 142.23, 141.92, 141.83, 136.57, 130.39-129.62 (m), 128.50, 128.19, 128.14, 127.75, 127.65, 127.35, 127.23, 125.37-125.08 (m), 122.99, 122.96, 122.91, 120.83, 103.45, 84.63, 82.20, 77.96, 74.57, 74.43, 73.98, 73.79, 72.61, 68.89, 67.50, 66.65, 38.12, 29.74; **¹⁹F NMR (471 MHz, CDCl₃)** δ -63.12 (s, 3F), -63.14 (s, 3F), -63.19 (s, 3F), -63.24 (s, 3F). **HRMS** (*m/z*): Calcd for C₄₉H₄₆F₁₂NO₈ 1004.3032 found 1004.3061. **[α]_D²⁵** = +52.4° (*c* = 1, DCM). **IR** (cm⁻¹) 3054, 2917, 1716, 1516, 1324, 1264, 1163, 1123, 1065, 1018, 823 (*p*-disubstituted benzene), 735, 703.

Probing for anomerization.



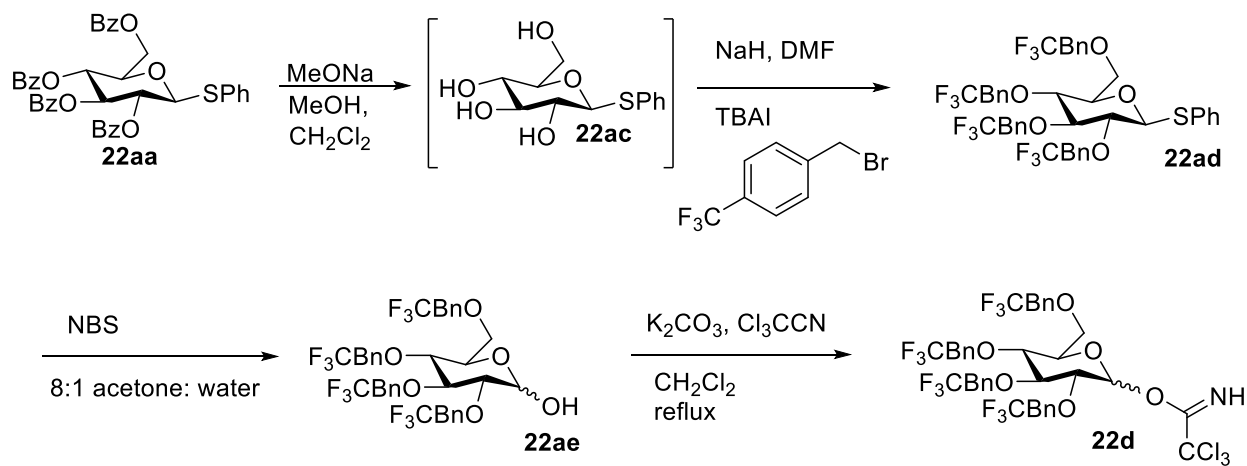
To an oven dried 4ml Wheaton vial with a stir bar, 24.2 mg of **20a** (0.0205 mmol) was added. The vial was capped with a septum then Nitrogen gas was flushed through it. 1.4 mL of 1,4-dioxane was then added under nitrogen atmosphere. Finally, 1.4 μL of TfOH was added and the reaction was allowed to stir at 20 °C for 12 hrs. The reaction mixture was quenched by addition of 0.05 mL of Et₃N then concentrated. ¹H NMR of the concentrated material showed no formation of the *beta* product.



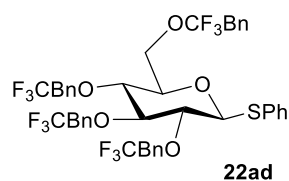
To an oven dried 4ml Wheaton vial with a stir bar, 35.6 mg of **20b** (0.0301 mmol) was added. The vial was capped with a septum then Nitrogen gas was flushed through it. 2.2 mL of 1,4-dioxane was then added under nitrogen atmosphere. Finally, 2.0 μ L of TfOH was added and the reaction was allowed to stir at 20 $^{\circ}$ C for 12 hrs. The reaction mixture was quenched by addition of 0.05 mL of Et₃N then concentrated. NMR of the concentrated material showed no formation of the *alpha* product.

Synthesis of trichloroacetimidate glucosyl donor with *p*-trifluoromethylbenzyl protecting groups (**22d**)

Synthesis plan:



Synthesis of **22ad**.

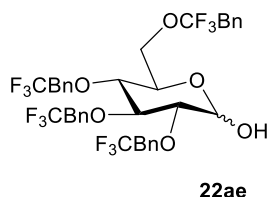


To 2.21g of **22aa** (3.21 mmol) in a R.B.F with a magnetic stir bar, 30.0 ml of methanol was added followed by 25.0 ml of dichloromethane. The R.B.F was capped with a septum. Three cycles of nitrogen purge backfill were performed followed by dropwise addition of 1.0 ml of 5 M NaOMe. After 30 min, TLC showed reaction was complete. 5.45 g of Dowex ® 50WX8 50-100 (H) resin (Alfa Aesar) was added to neutralize the NaOMe. The mixture was filtered through celite and rinsed with 20.0 ml of methanol. Collected solution was concentrated then co-evaporated with 2.0 ml of toluene twice to give an off white powder (**22ac**). To the powder (**22ac**) in R.B.F with a stir bar, 8.06g of *p*-trifluoromethylbenzyl bromide (33.7 mmol) was added followed by 50.0 ml of DMF. 1.02g of TBAI was then added. After all the solids had dissolved, the R.B.F was placed in an ice bath and cooled to 0 °C. Excess NaH (5.0 g) was then added carefully. After 10 min the reaction was allowed to warm to room temperature under N₂ atmosphere and stir for 16hrs. The reaction was quenched at 0 °C by dropwise addition of water until gas evolution ceased. 200.0ml of EtOAc was added and the contents transferred to a separatory funnel. 1000.0 ml of water was then added, the contents were shaken vigorously to get rid of DMF and then allowed to stand to ensure separation of aqueous and organic layer. The organic layer was dried using Na₂SO₄ then concentrated to give a crude product which was purified using flash chromatography (5-15% EtOAc/hexanes) to give 2.19g (2.42 mmol) of **22ad** white solid (70% two steps).

¹H NMR (500 MHz, CDCl₃) δ 7.63-7.51 (m, 10H), 7.49-7.40 (m, 4H), 7.34-7.25 (m, 7H), 5.00(d, *J* = 11.2 Hz, 1H), 4.89 (d, *J* = 12.1 Hz, 1H), 4.86-4.79(m, 2H), 4.77-4.65 (m, 4H), 4.61(d, *J* = 12.6Hz, 1H), 3.86-3.76 (m, 2H), 3.75-3.68 (m, 2H), 3.60-3.51(m, 2H). **¹³C NMR (125 MHz, CDCl₃)** δ 142.15, 142.11, 141.84, 141.80, 133.43, 131.81, 130.16-129.76(m), 129.03, 127.83, 127.75, 127.54, 127.36, 127.20, 125.40, 125.37, 125.34, 125.31, 87.49, 86.70, 81.10, 78.91, 77.88, 74.72,

74.48, 74.01, 72.67, 69.08. **^{19}F NMR (471 MHz, CDCl_3) δ** -63.04 (s, 3F), -63.12 (s, 3F), -63.15 (s, 3F), -63.17 (s, 3F). **HRMS (m/z):** Calcd for $\text{C}_{44}\text{H}_{36}\text{F}_{12}\text{O}_5\text{SNa}$ 927.1990 found 927.1997 **$[\alpha]_{\text{D}}^{25} = +66.3^\circ$** (c = 1, DCM). **IR (cm^{-1})** 2874, 1326, 1264, 1165, 1111, 1066, 1019, 823 (p-disubstituted benzene), 733, 703.

Synthesis of **22ae**.



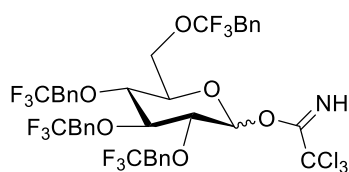
To 2.19g (2.42 mmol) of **22ad** in a R.B.F with a stir bar, 24.0 ml of 8:1 acetone:water was added, followed by addition of 1.72g of NBS (9.66 mmol). The R.B.F was then capped with a septum then three cycles of nitrogen purge backfill were performed. T.L.C showed the reaction was complete after 1hr. The contents were then concentrated followed by addition of 30.0 ml of dichloromethane. The resulting solution was then transferred to a separatory funnel and washed with 100.0 ml of water once. The organic layer was then concentrated and purified by flash chromatography (10-40% EtOAc/hexanes) to give 1.15g (1.42mmol) of **22ae** as a colorless gum (59% yield).

^1H NMR (500 MHz, CDCl_3) δ 7.62-7.29 (m, 21H), 7.26-7.20 (m, 3H), 5.35 (t, $J = 3.0$ Hz, 1H), 5.04 (d, $J = 12.0$ Hz, 1H), 4.99-4.90 (m, 2H), 4.86-4.72 (m, 6H), 4.69-4.64(m, 2H), 4.63-4.58(m, 2H), 4.54(d, $J = 12.8$ Hz, 1H), 4.09(ddd, $J = 10.2, 3.9, 2.0$ Hz, 1H), 4.01(t, $J = 9.2$ Hz, 1H), 3.79-3.54(m, 7H), 3.42(t, $J = 7.7$ Hz, 1H), 3.05-2.96(m, 1H). **^{13}C NMR (125 MHz, CDCl_3) δ** 142.39-141.57(m), 130.42-129.38(m), 127.83, 127.73, 127.72, 127.36, 127.32, 127.26, 127.22, 125.57-125.21(m), 125.13, 122.90, 97.56, 91.03, 84.50, 83.13, 81.67, 80.35, 77.85, 74.64, 74.61, 73.99, 73.69, 72.78, 72.73, 72.32, 70.24, 69.06, 68.84. **HRMS (m/z):** Calcd for $\text{C}_{38}\text{H}_{32}\text{F}_{12}\text{O}_6\text{Na}$

835.1905 found 835.1947 $[\alpha]_D^{25} = +32.6^\circ$ (c = 1, DCM). IR (cm^{-1}) 3421, 2926, 1326, 1161, 1121, 1110, 1066, 1018, 822 (*p*-disubstituted benzene), 637.

Synthesis of 22d

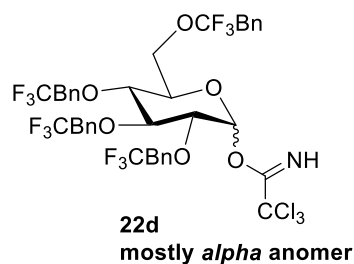
To 1.11 g (1.37 mmol) of **22ae** in R.B.F with stir bar, 25.0 ml of dichloromethane was added followed by 4.5 g of K_2CO_3 (excess). Lastly, trichloroacetonitrile 5.0 ml (excess) was added. The reaction was allowed to stir at room temperature, however, after 5 hrs, little product formation was observed as shown by T.L.C. A reflux condenser was then attached and the reaction was refluxed for two hours. Product was cleanly formed as judged by T.L.C and NMR. The contents were filtered through celite then the filtrate was concentrated and then put under high vacuum to give 1.20g of a white powder **22d** (91%).



22d
mostly *beta* anomer

Data for *beta* anomer: ^1H NMR (500 MHz, CDCl_3) δ 8.75 (s, 1H), 7.56-7.24 (m, 16H), 5.82 (dd, $J = 5.4, 2.2\text{ Hz}$, 1H), 4.99 (d, $J = 11.8\text{ Hz}$, 1H), 4.88 (d, $J = 12.2\text{ Hz}$, 1H), 4.84-4.71 (m, 3H), 4.69-4.61 (m, 2H), 4.58 (d, $J = 12.8\text{ Hz}$, 1H), 3.83-3.63 (m, 6H). ^{13}C NMR (125 MHz, CDCl_3) δ 161.07, 142.10, 141.79, 141.69, 130.31-129.64 (m), 127.84, 127.65, 127.37, 127.19, 125.36, 125.33, 125.30, 98.08, 90.78, 84.51, 80.96, 77.38, 77.21, 75.64, 74.55, 73.97, 73.88, 72.63, 68.37. **HRMS (m/z):** Calcd for $\text{C}_{40}\text{H}_{32}\text{Cl}_3\text{F}_{12}\text{NO}_6\text{Na}$ 978.1001 found 978.1024 $[\alpha]_D^{25} = -15.5^\circ$ (c

= 1, DCM). IR (cm⁻¹) 2923, 1325, 1263, 1121, 1066, 1018, 822(*p*-disubstituted benzene), 797, 736, 703.



Data for *alpha* anomer: ¹H NMR (500 MHz, CDCl₃) δ 8.63 (s, 1H), 7.64-7.22 (m, 16H), 6.60 (d, *J* = 3.4 Hz, 1H), 4.98 (d, *J* = 12.0 Hz, 1H), 4.92-4.74 (m, 3H), 4.74-4.59 (m, 3H), 4.54 (d, *J* = 12.7 Hz, 1H), 4.12-4.00 (m, 2H), 3.87-3.66 (m, 4H). ¹³C NMR (125 MHz, CDCl₃) δ 161.18, 142.31, 141.81, 141.66, 130.51-129.36(m), 127.69, 127.48, 127.46, 127.42, 125.36-125.27(m), 125.12, 122.96, 93.84, 81.45, 79.71, 77.22, 76.97, 74.65, 74.30, 73.04, 72.73, 72.09, 68.36.

HRMS (m/z): Calcd for C₄₀H₃₂Cl₃F₁₂NO₆Na 978.1001 found 978.1003 [*α*]_D²⁵ = +43.5° (*c* = 1, DCM). IR (cm⁻¹) 2921, 1738, 1672, 1325, 1163, 1121, 1066, 1018, 822 (*p*-disubstituted benzene), 795, 737, 643.

General procedure C

Glycosylation using trichloroacetimidate glucosyl donors

0.15 mmol of 2,3,4,6-tetra-O-(4-trifluoromethylbenzyl)-D-glucopyranose trichloroacetimidate donor (**22d**) or 2,3,4,6-tetra-O-benzyl-D-glucopyranose trichloroacetimidate (**22a**) (1 equiv) was put in an oven dried test tube with a stir bar followed by addition of 0.105 mmol of acceptor **12** (0.7 eq). The test tube was capped with a septum and flushed with nitrogen gas for 1 min. 1,4

dioxane (volume is specified below) was then added with nitrogen line still attached. Finally, 13.2 μ L (0.15 mmol) of TfOH (1equiv) was added using a micro-syringe and the reaction allowed to stir at 20 °C for 12hrs. The reaction was quenched by addition of 0.05 ml of Et₃N. Concentration using rotary evaporator was followed by purification using flash column chromatography (12-28% EtOAc/hexanes).

Entry 1 Table 2

Following the general procedure **C**. 2.5 ml of 1,4-dioxane, 102.9 mg (0.150 mmol) of 2,3,4,6-tetra-O-benzyl-D-glucopyranose trichloroacetimidate donor (**22a**), 50.1 mg (0.108 mmol) of acceptor **12**, and 13.2 μ L (0.15 mmol) of TfOH were used. Purified by flash column chromatography (12-28% EtOAc/hexanes) to give 80.7 mg of a colorless oil (76%) which was a mixture of **13a-alpha** and **13a-beta**, *alpha:beta* ratio 5.2:1.

Entry 2 Table 2

Following the general procedure **C**. 2.5 ml of 1,4-dioxane, 144.2 mg (0.1507 mmol) of 2,3,4,6-tetra-O-(4-trifluoromethylbenzyl)-D-glucopyranose trichloroacetimidate donor (**22d**), 49.9 mg (0.1074 mmol) of acceptor **12**, and 13.2 μ L (0.15 mmol) of TfOH were used. Purified by flash column chromatography (12-28% EtOAc/hexanes) to give 111.6 mg of colorless oil (83%) which was a mixture of **13d-alpha** and **13d-beta**, *alpha:beta* ratio 10.0:1.

Entry 3 Table 2

Following the general procedure **C**. 5.0 ml of 1,4-dioxane, 104.8 mg (0.1530 mmol) of 2,3,4,6-tetra-O-benzyl-D-glucopyranose trichloroacetimidate donor (**22a**), 49.7 mg (0.107 mmol) of acceptor **12**, and 13.2 μ L (0.15 mmol) of TfOH were used. Purified by flash column

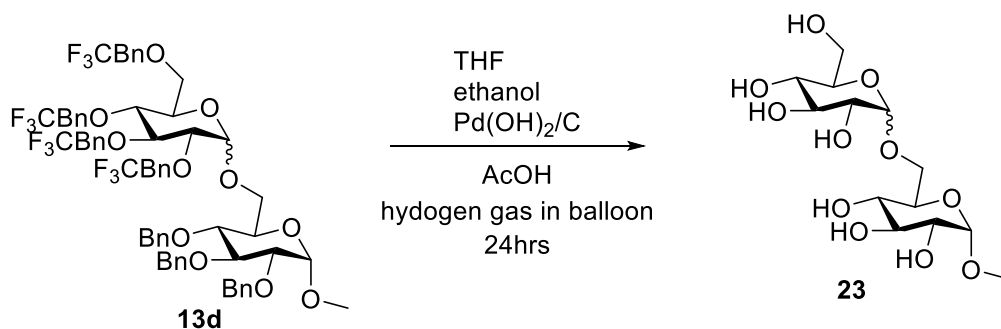
chromatography (12-28% EtOAc/hexanes) to give 99.2 mg of a colorless oil (94%) which was a mixture of **13a-*alpha*** and **13a-*beta***, *alpha:beta* ratio 5.0:1.

Entry 4 Table 2

Following the general procedure **C**. 5.0 ml of 1,4-dioxane, 144.6 mg (0.1511 mmol) of 2,3,4,6-tetra-*O*-(4-trifluoromethylbenzyl)-D-glucopyranose trichloroacetimidate donor (**22d**), 49.9 mg (0.1074 mmol) of acceptor **12**, and 13.2 μ L (0.15 mmol) of TfOH were used. Purified by flash column chromatography (12-28% EtOAc/hexanes) to give 105.7 mg of colorless oil (78%) which was a mixture of **13d-*alpha*** and **13d-*beta***, *alpha:beta* ratio 14.0:1

Debenzylation Procedure

Methyl 6-*O*-(D-Glucopyranosyl)- α -D-glucopyranoside (**23**)



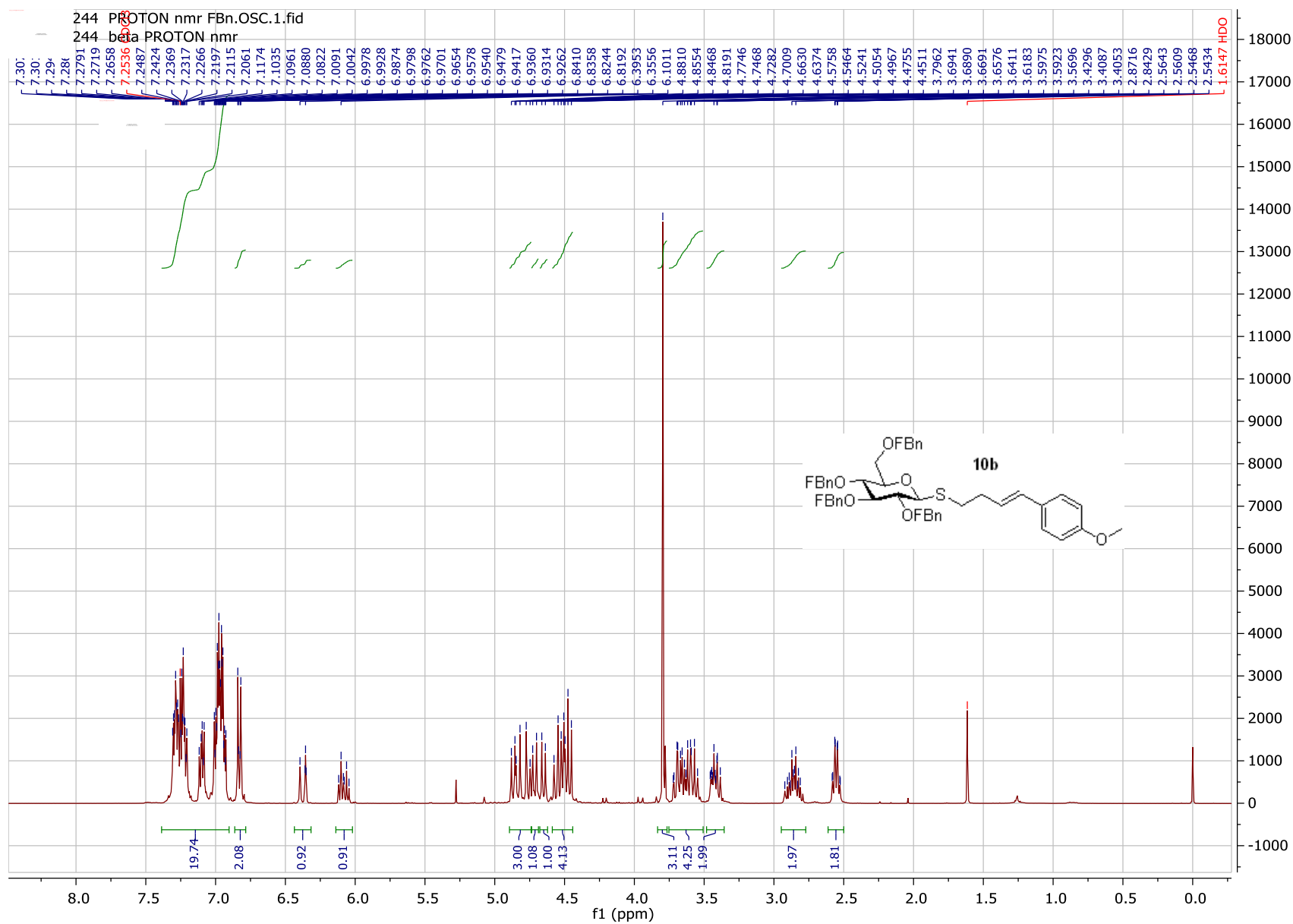
To 74.8 mg of **13d** ($\alpha:\beta$ 8.7:1) (0.0594 mmols) in a vial, 1.5 ml of THF was added followed by 2.2 ml of ethanol then 0.5 ml of water. 156 mg of 20% Pd(OH)₂ on carbon was then added in one portion. Lastly, 0.5 ml of acetic acid was added. The vial was capped with a septum and hydrogen gas in a balloon was connected via a needle. Three cycles of vacuuming and back-filling with hydrogen gas were performed. The reaction was allowed to stir under hydrogen gas atmosphere at 20 °C for 24 hrs. The solid particles were filtered out by passing the reaction mixture through a cotton plug and rinsing with MeOH. The crude Product was purified by flash

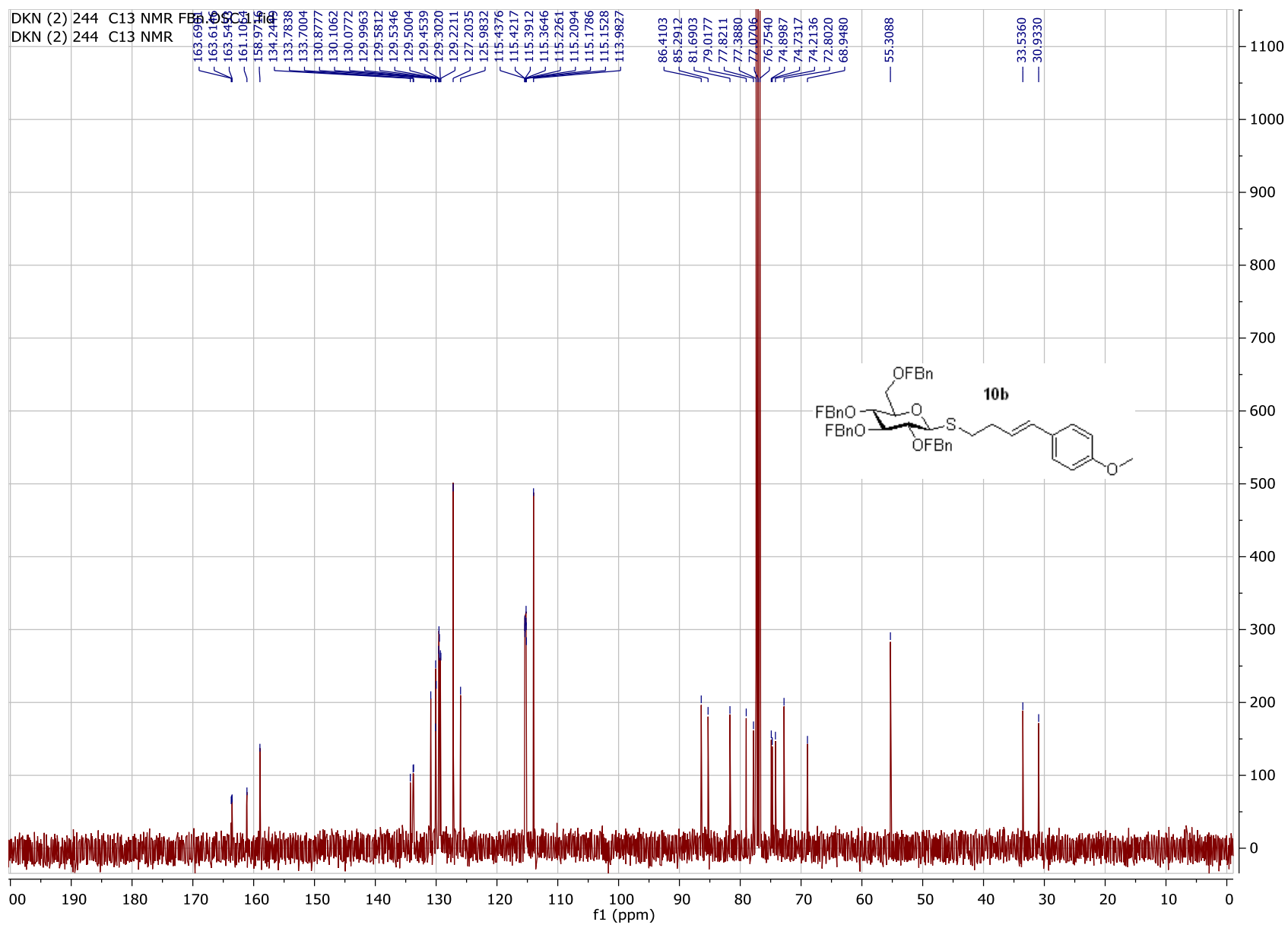
chromatography (10-50% MeOH/CH₂Cl₂) to give 20.0 mg of a white solid (94%). ¹H NMR and ¹³C NMR spectra match those reported in the literature.⁴

References

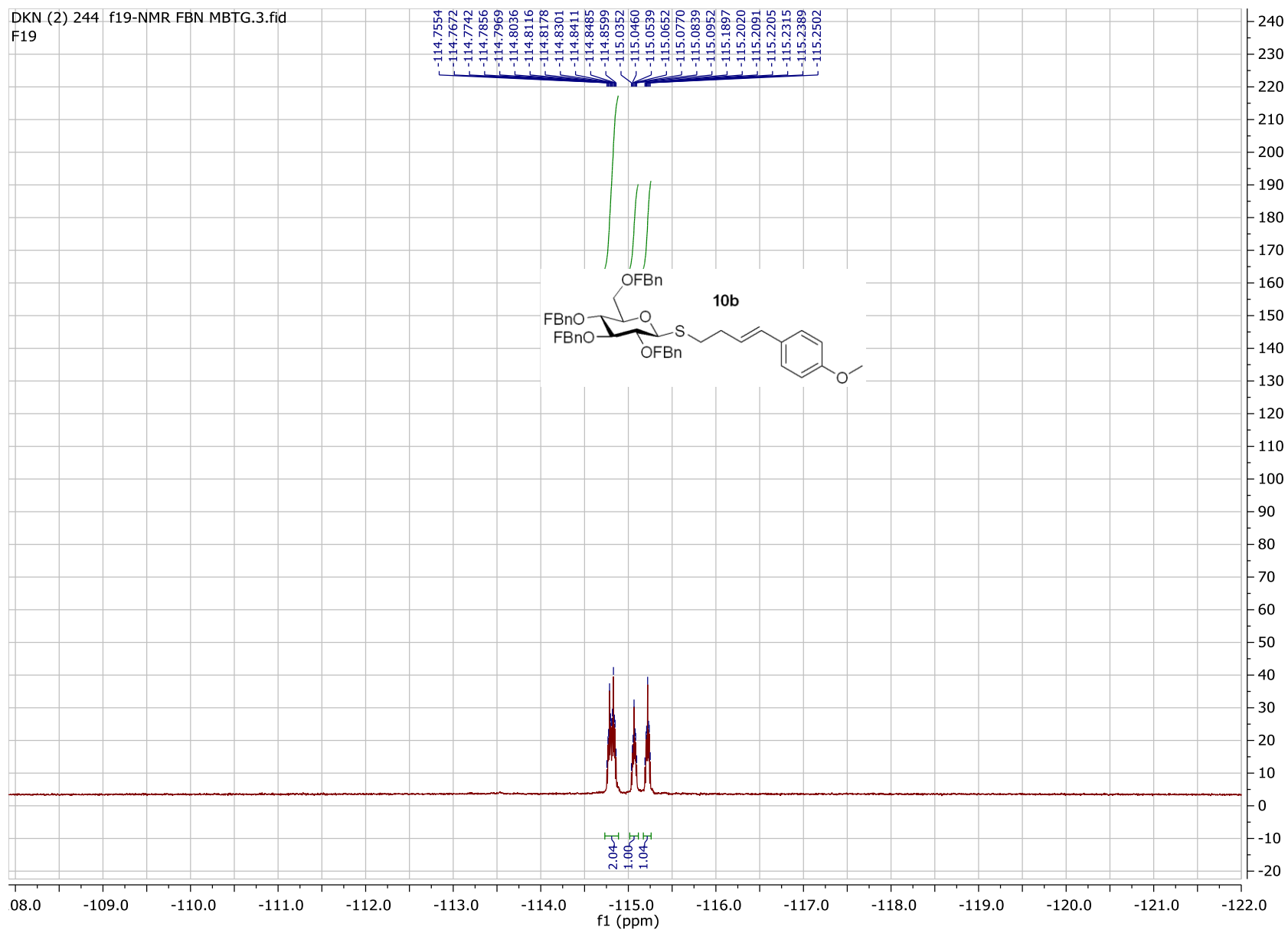
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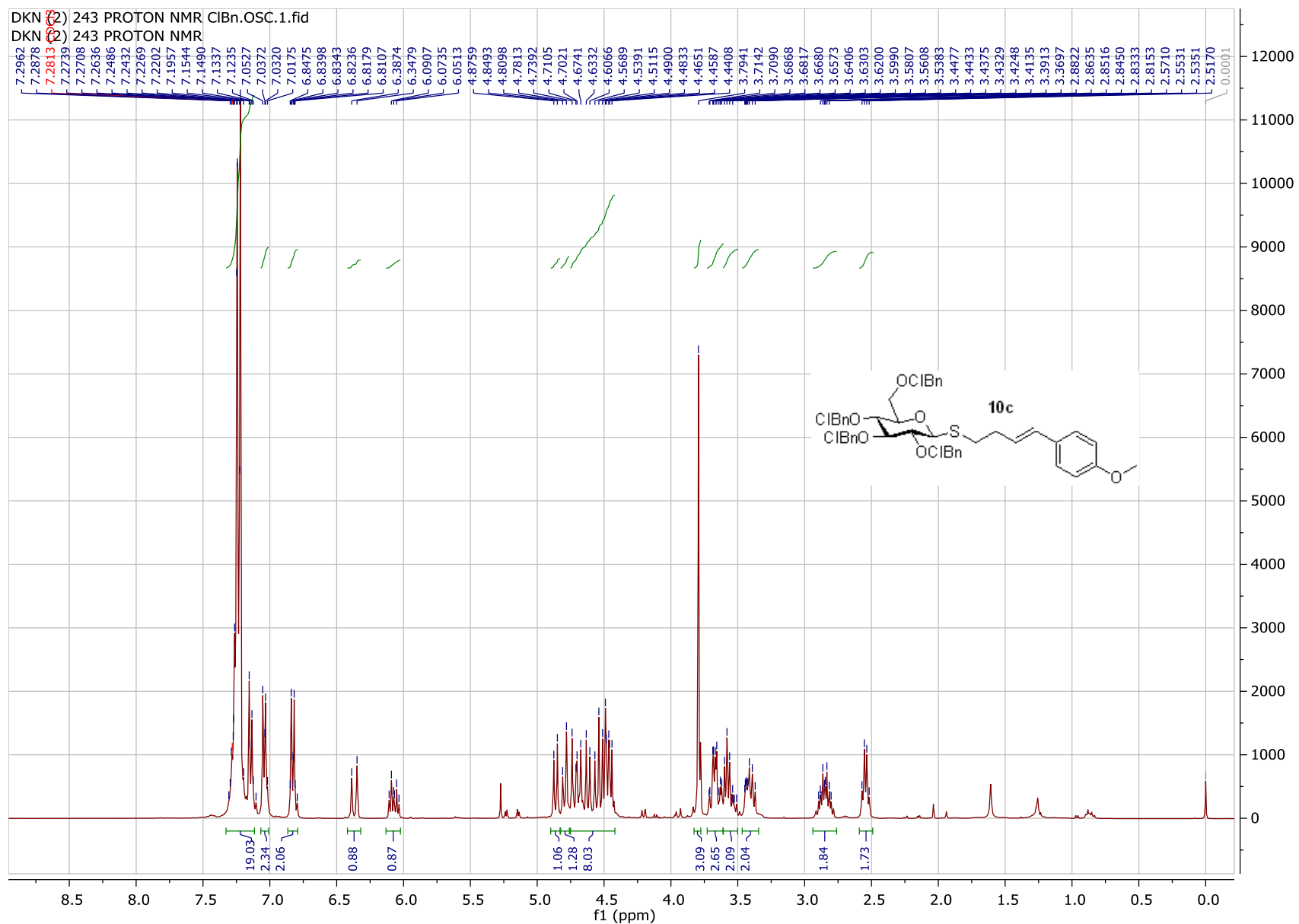
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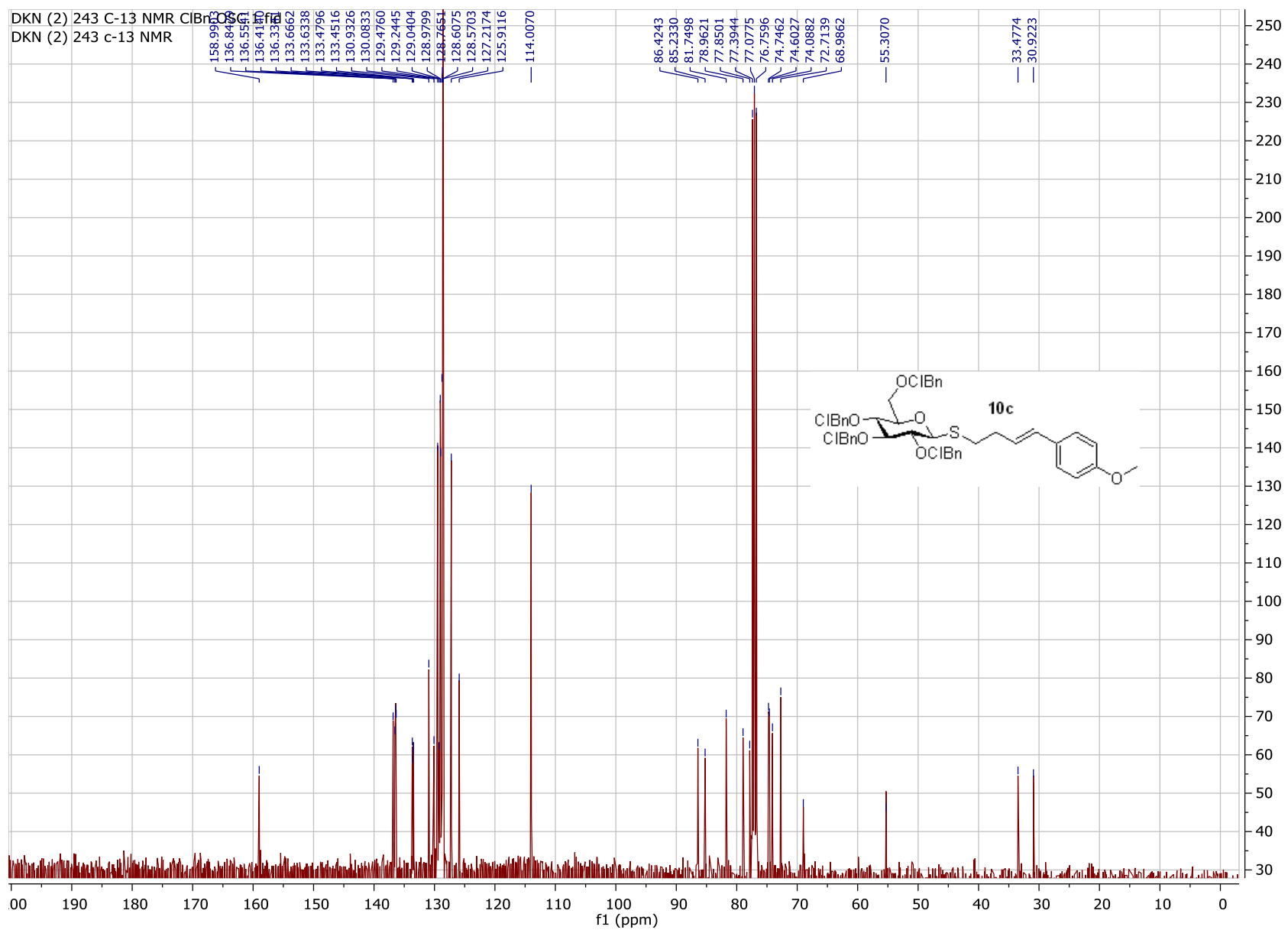


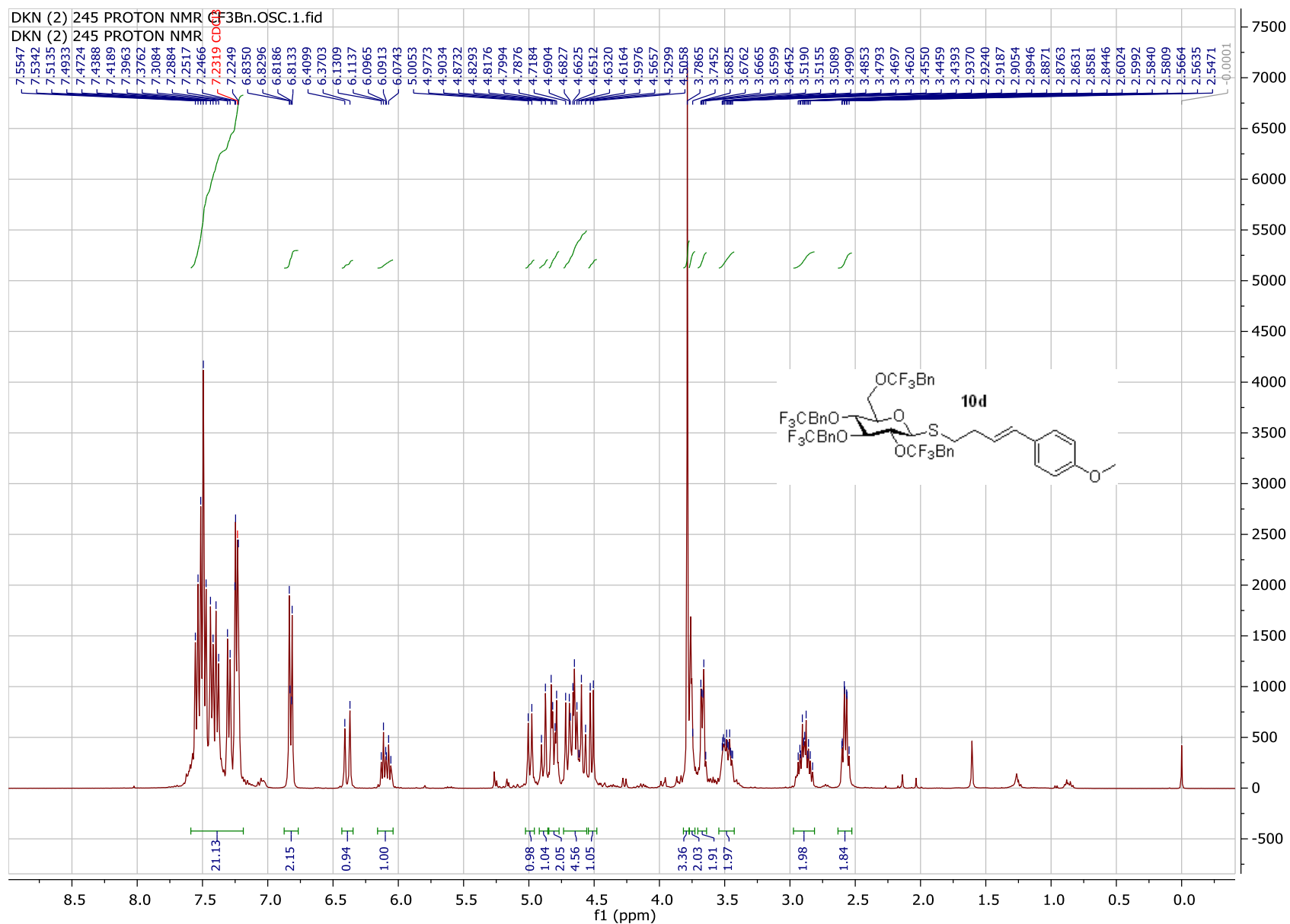


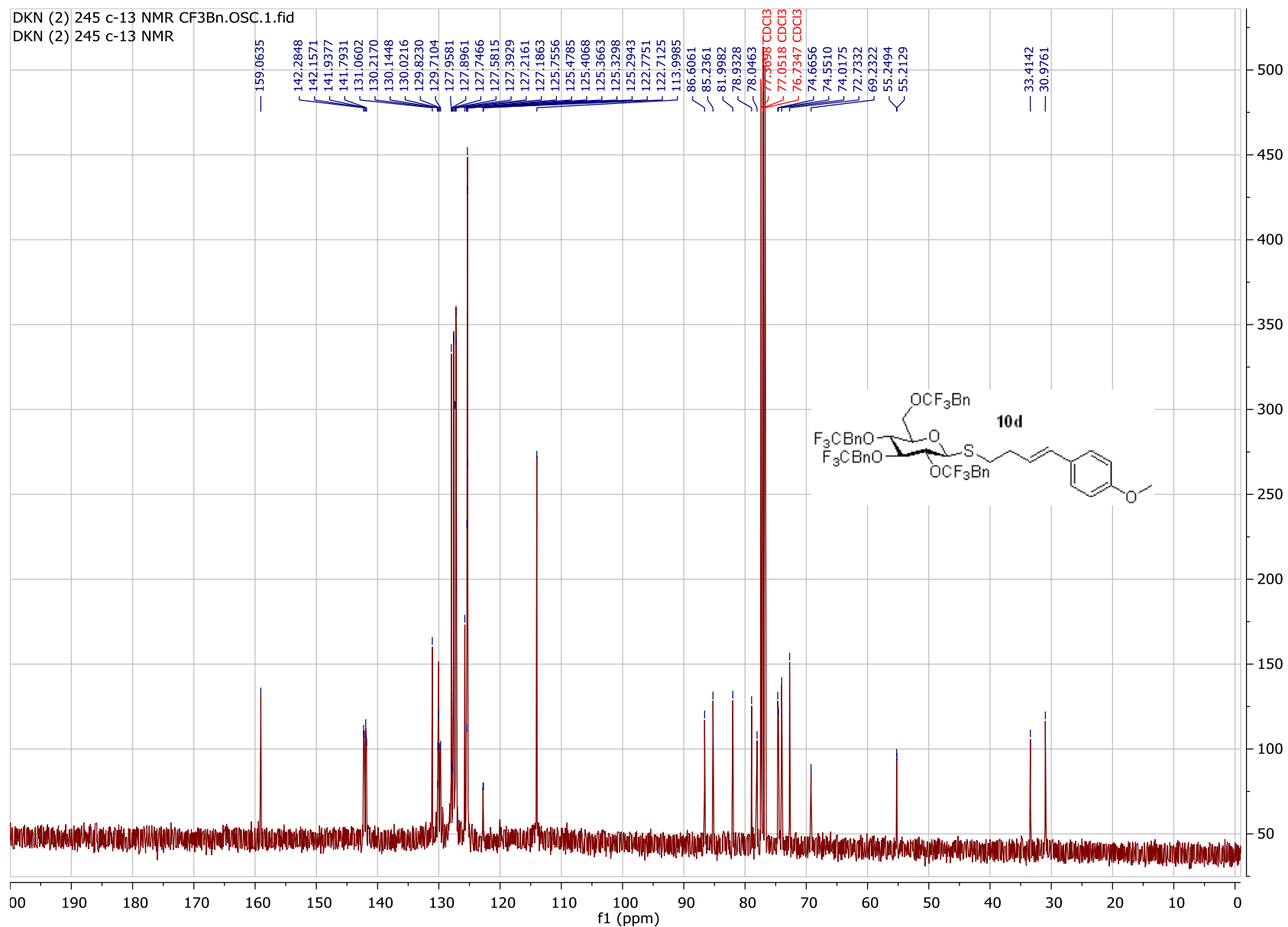
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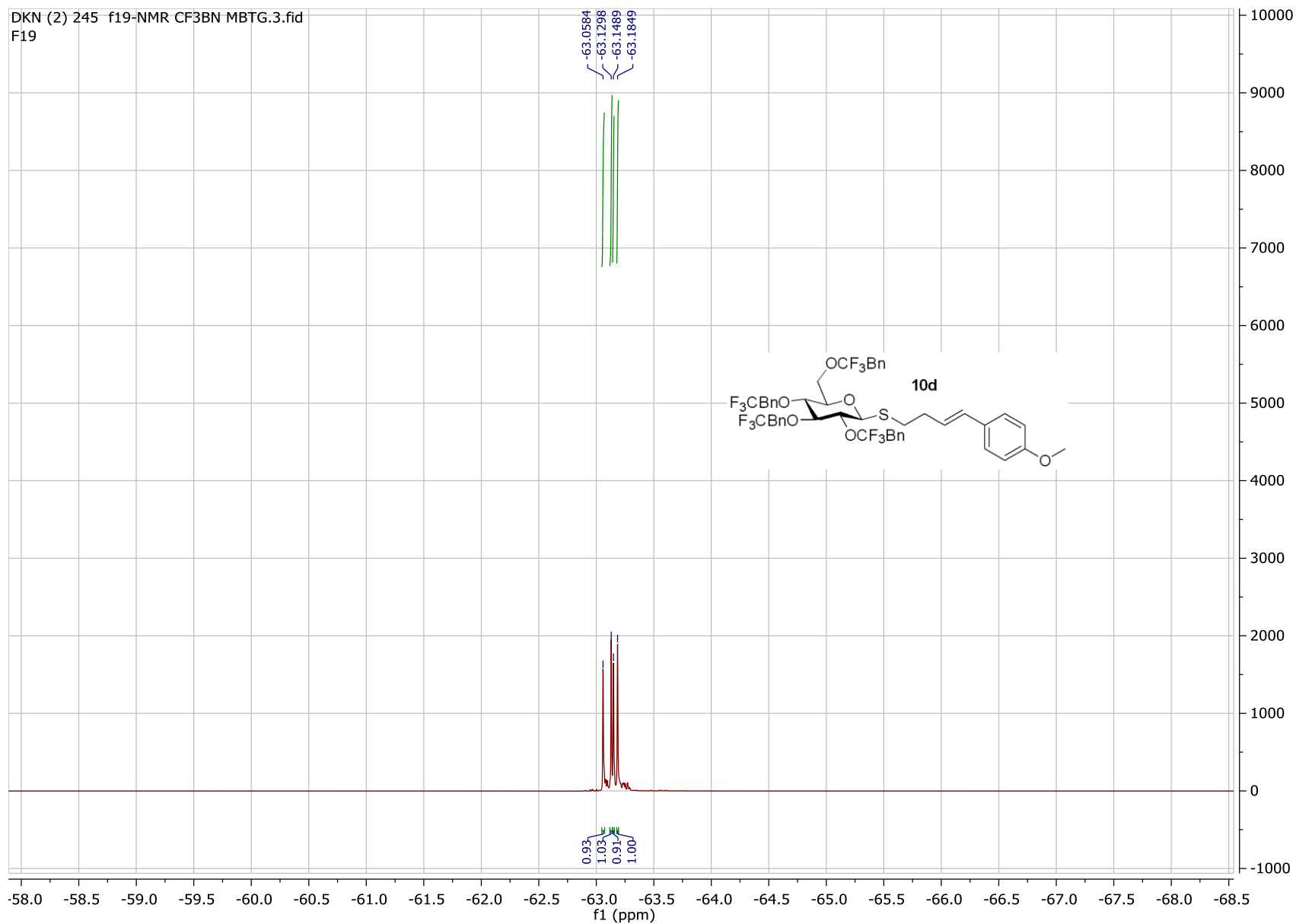


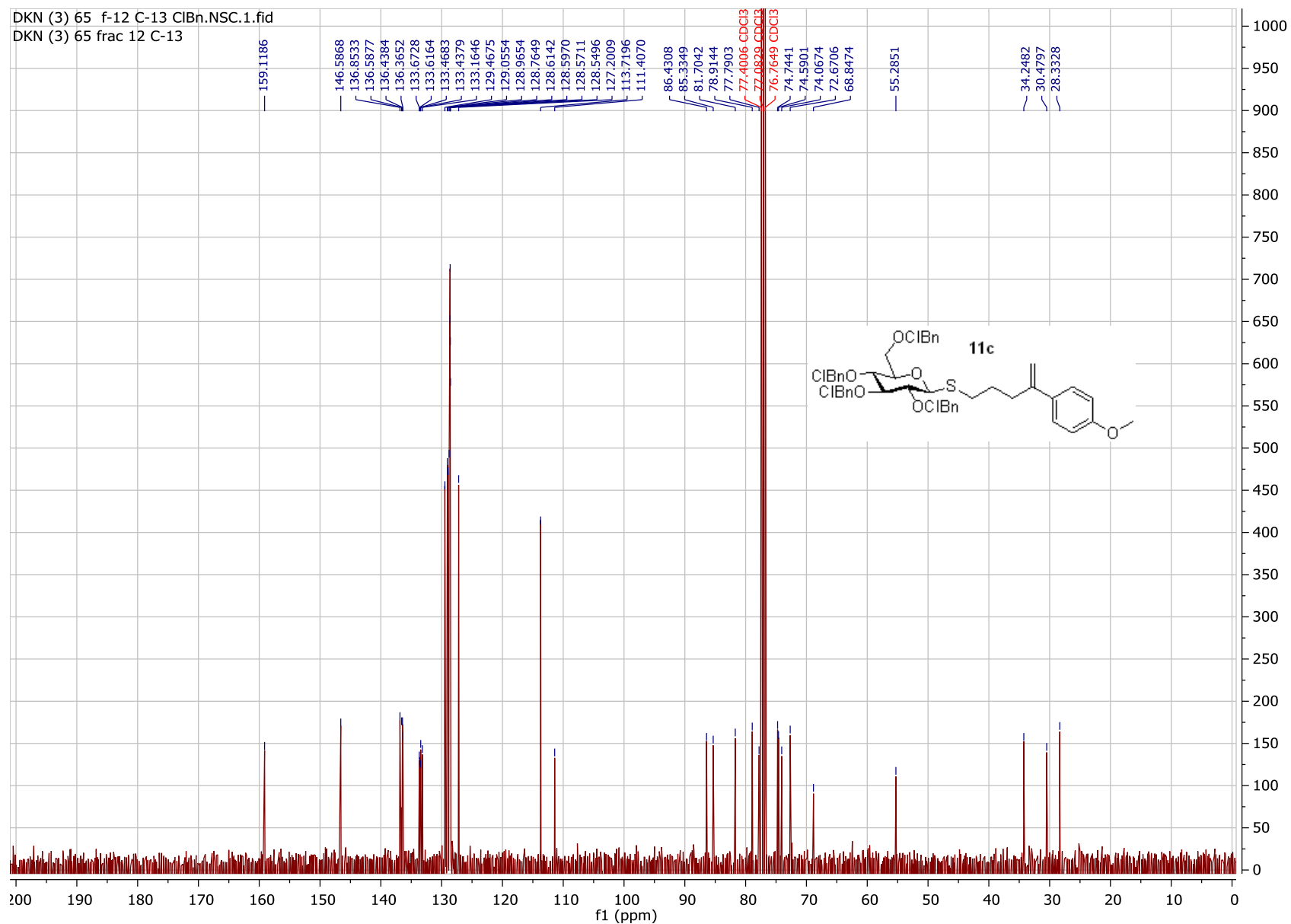


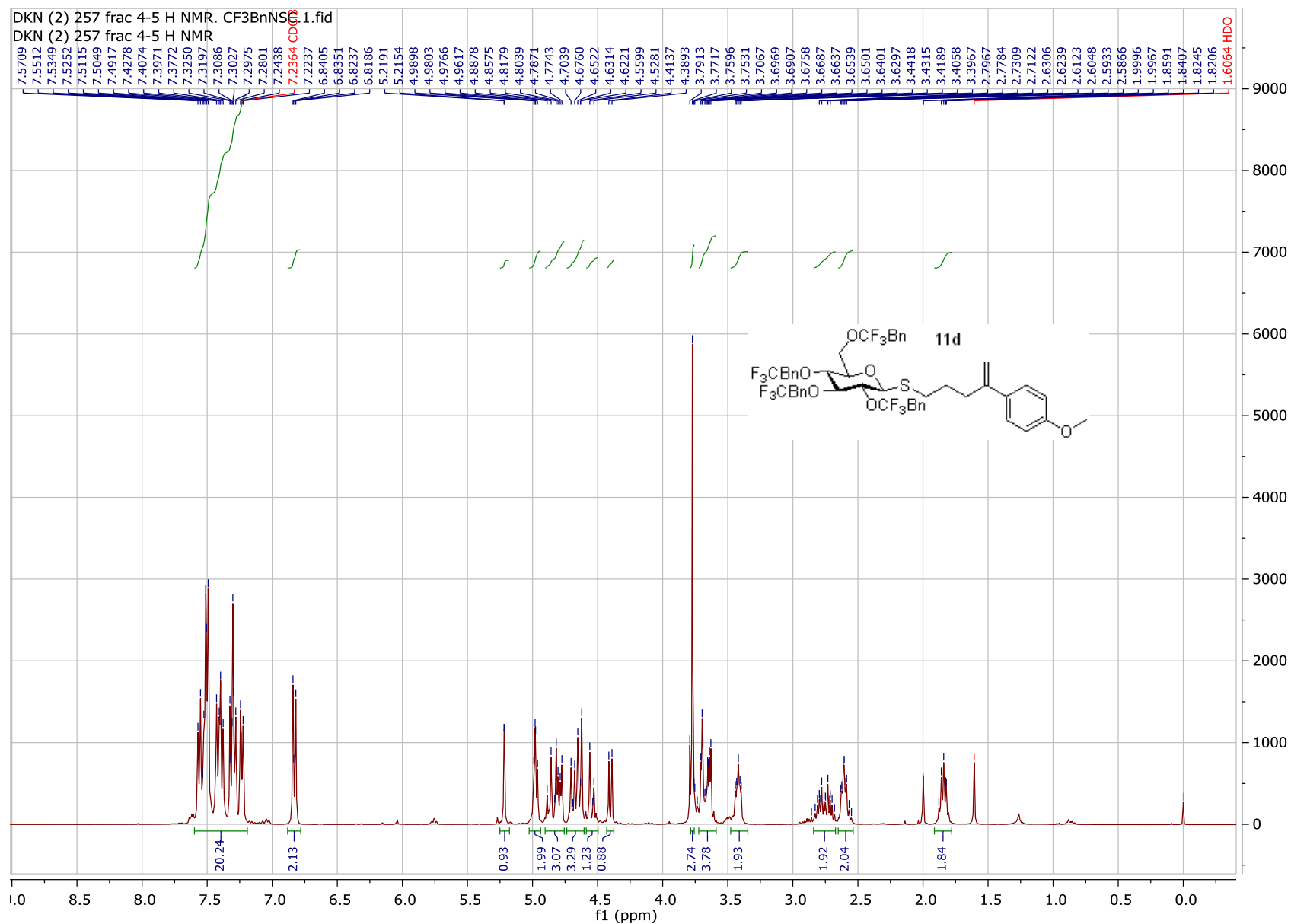




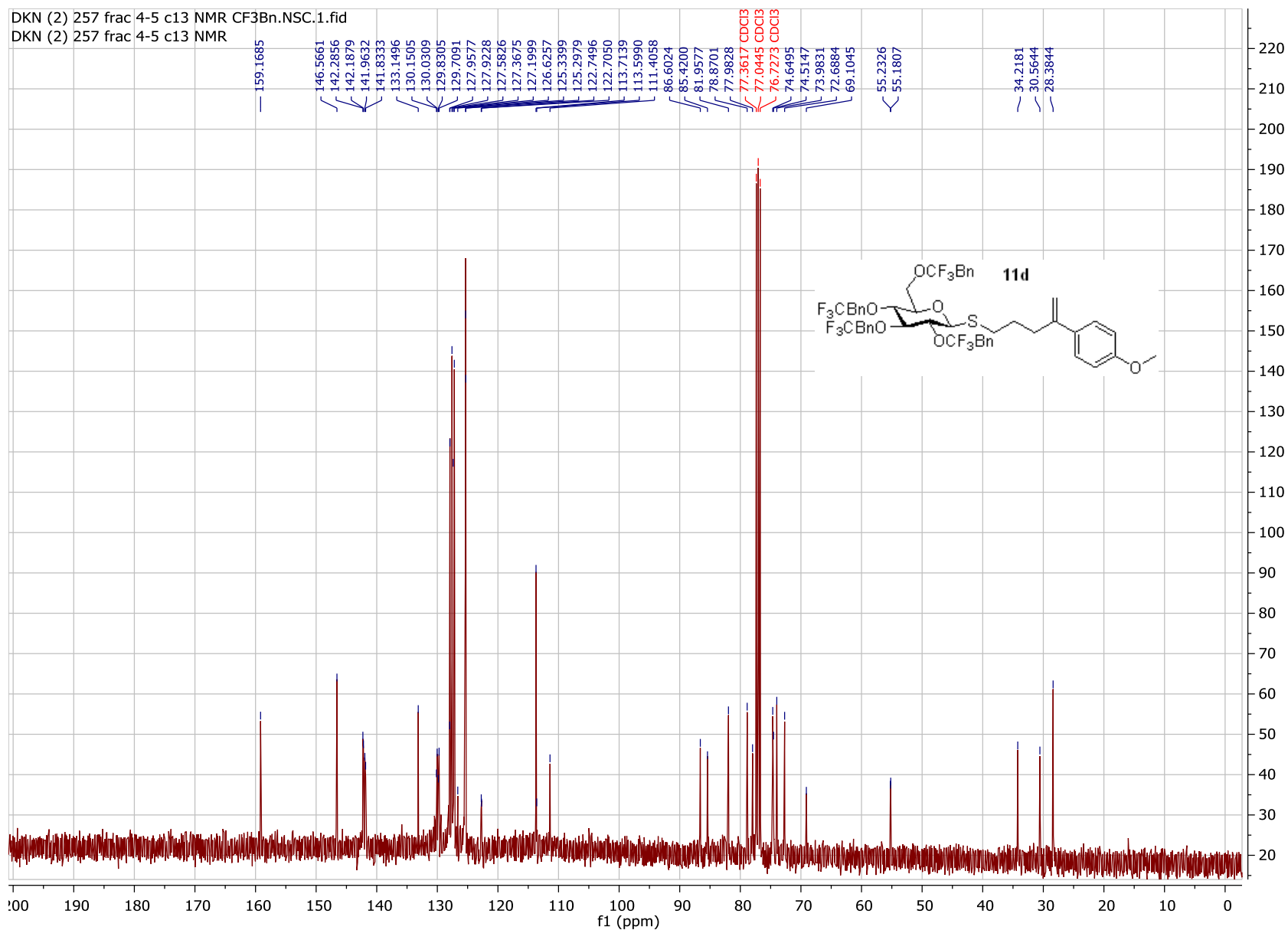
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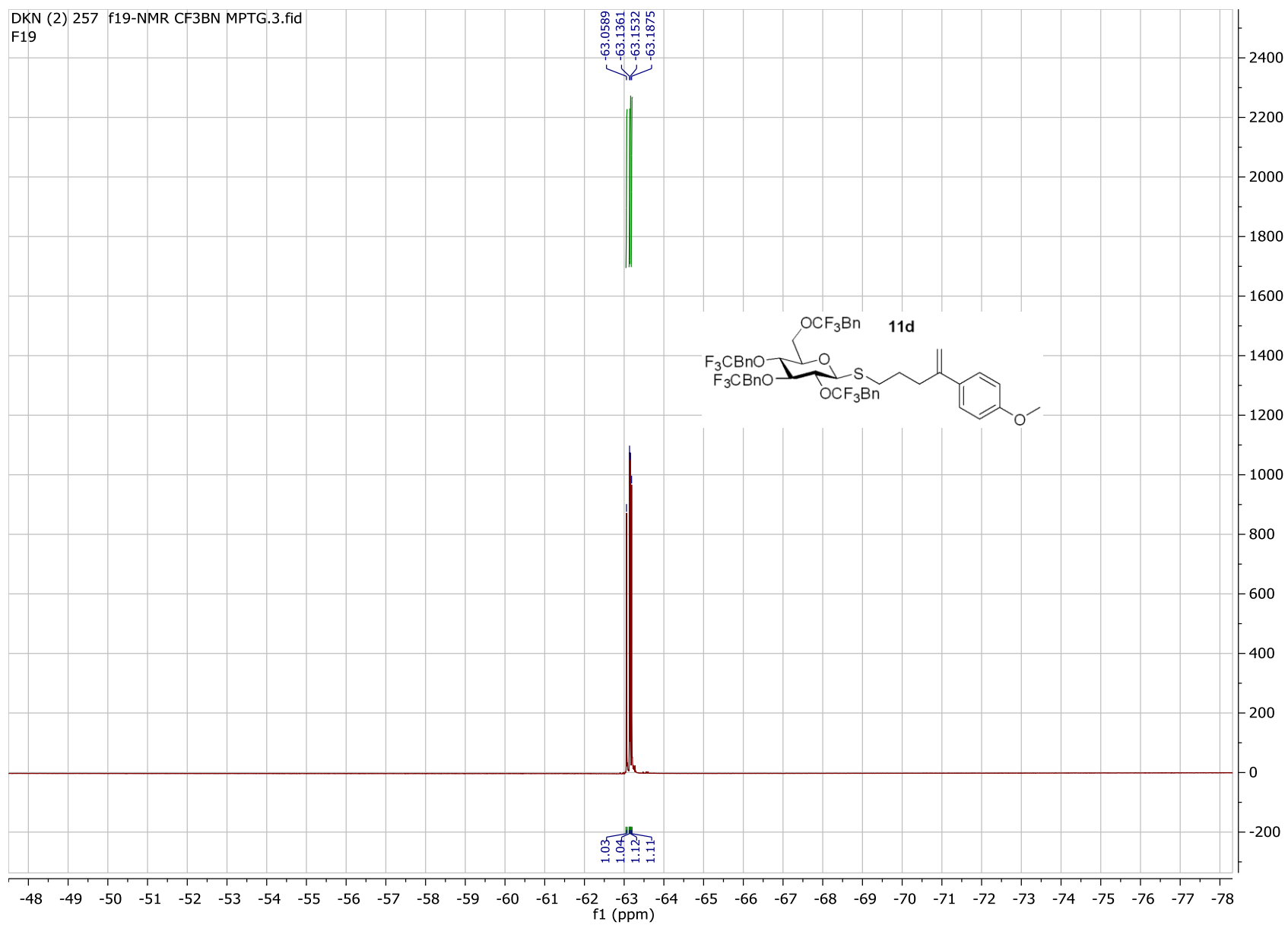


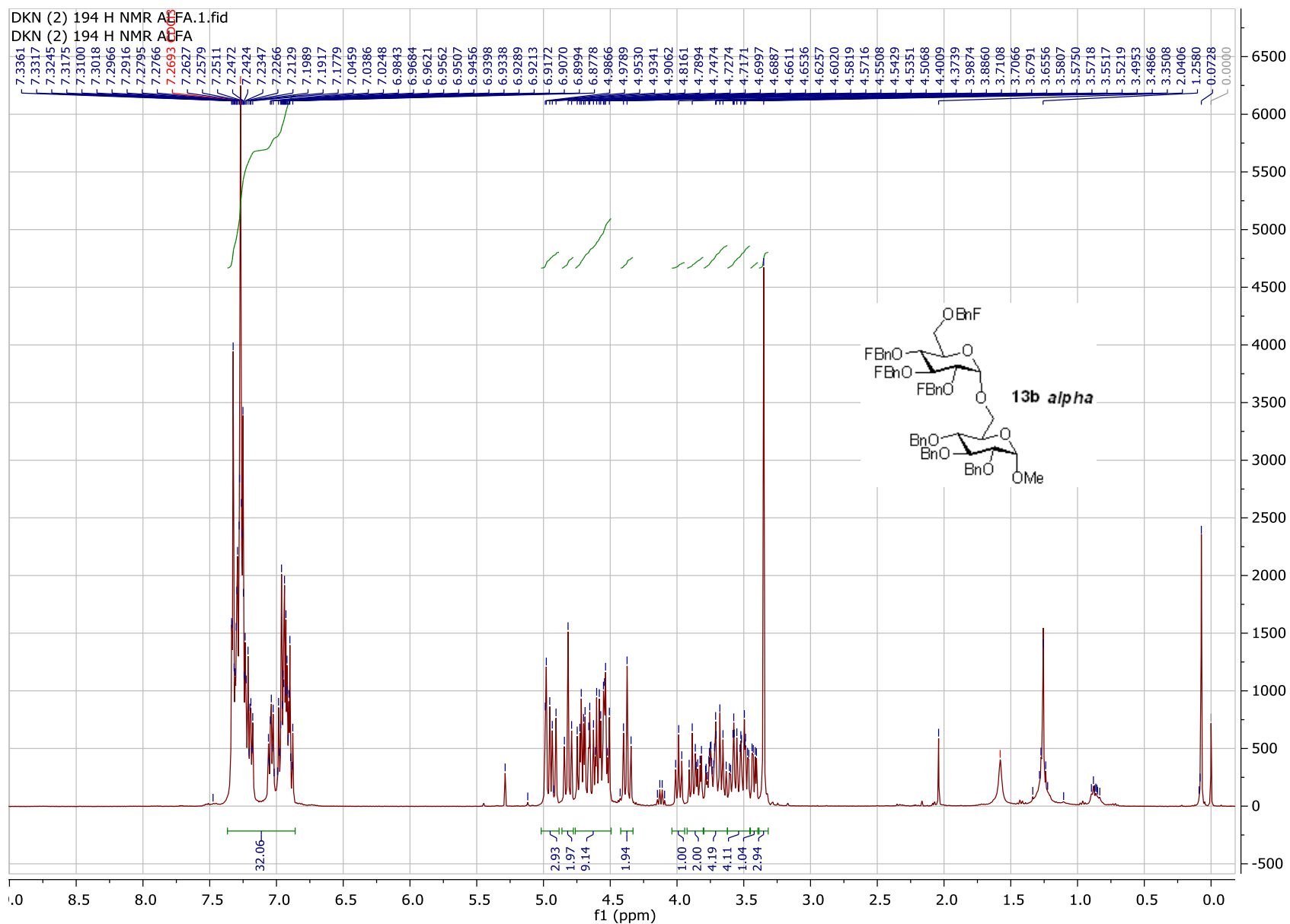


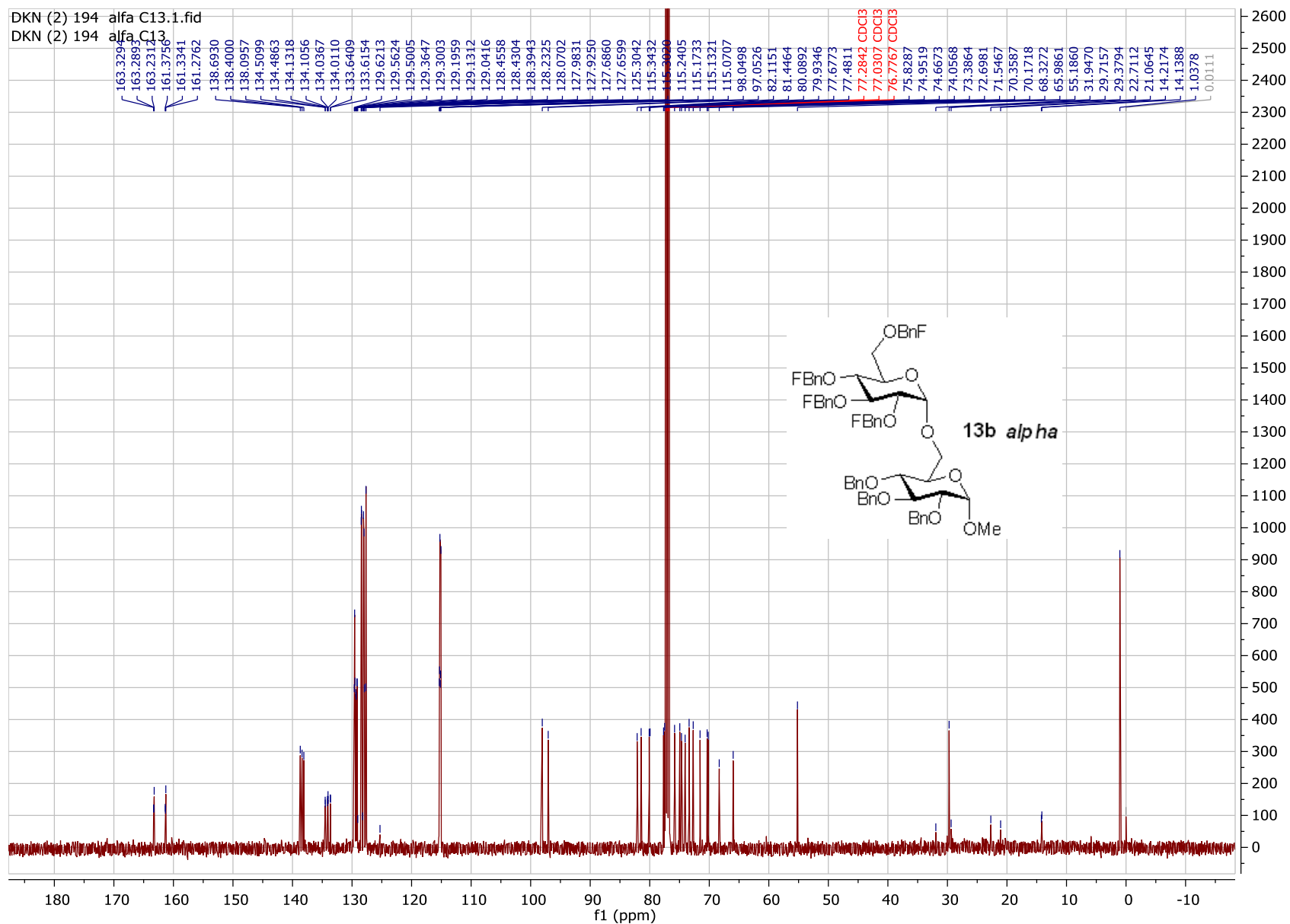
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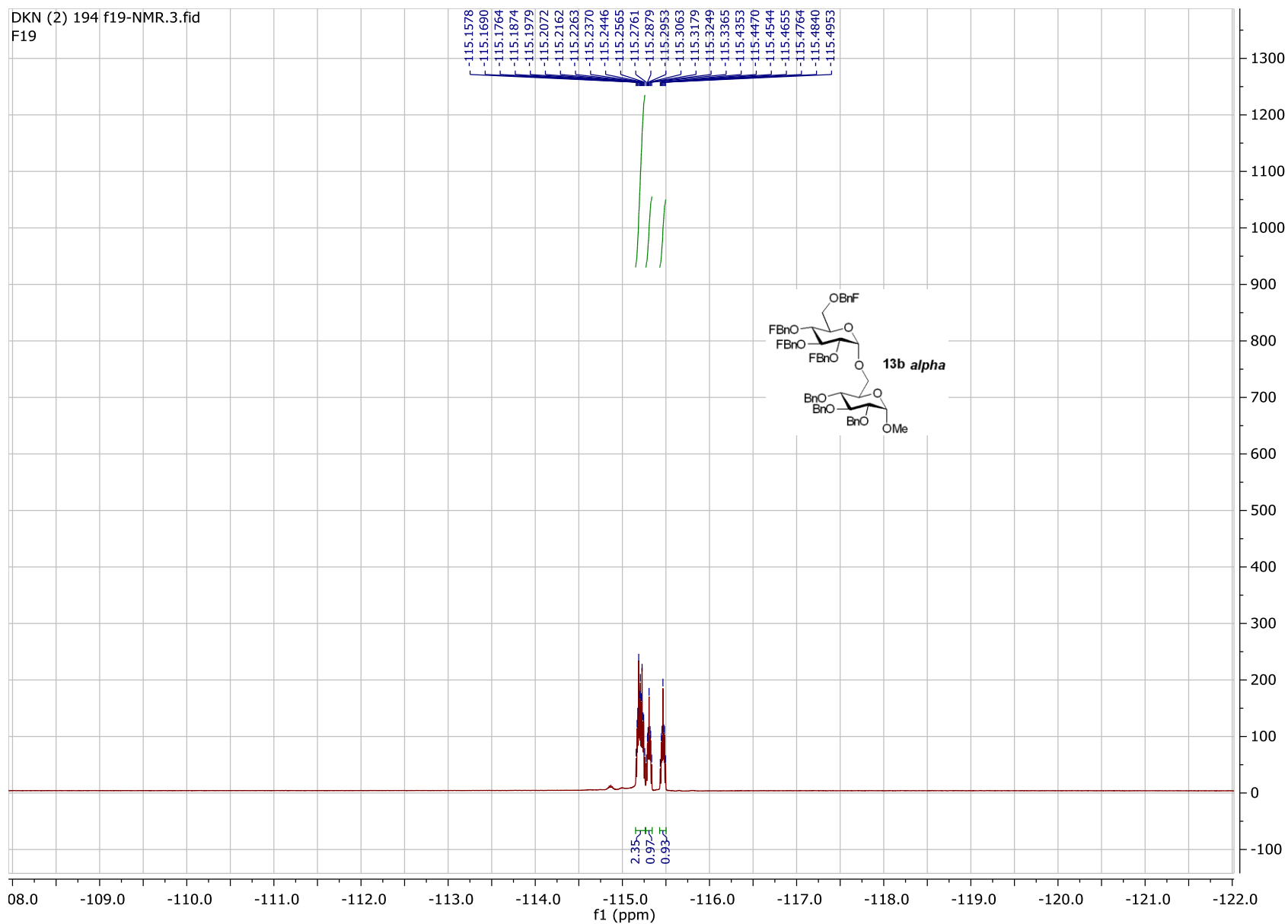
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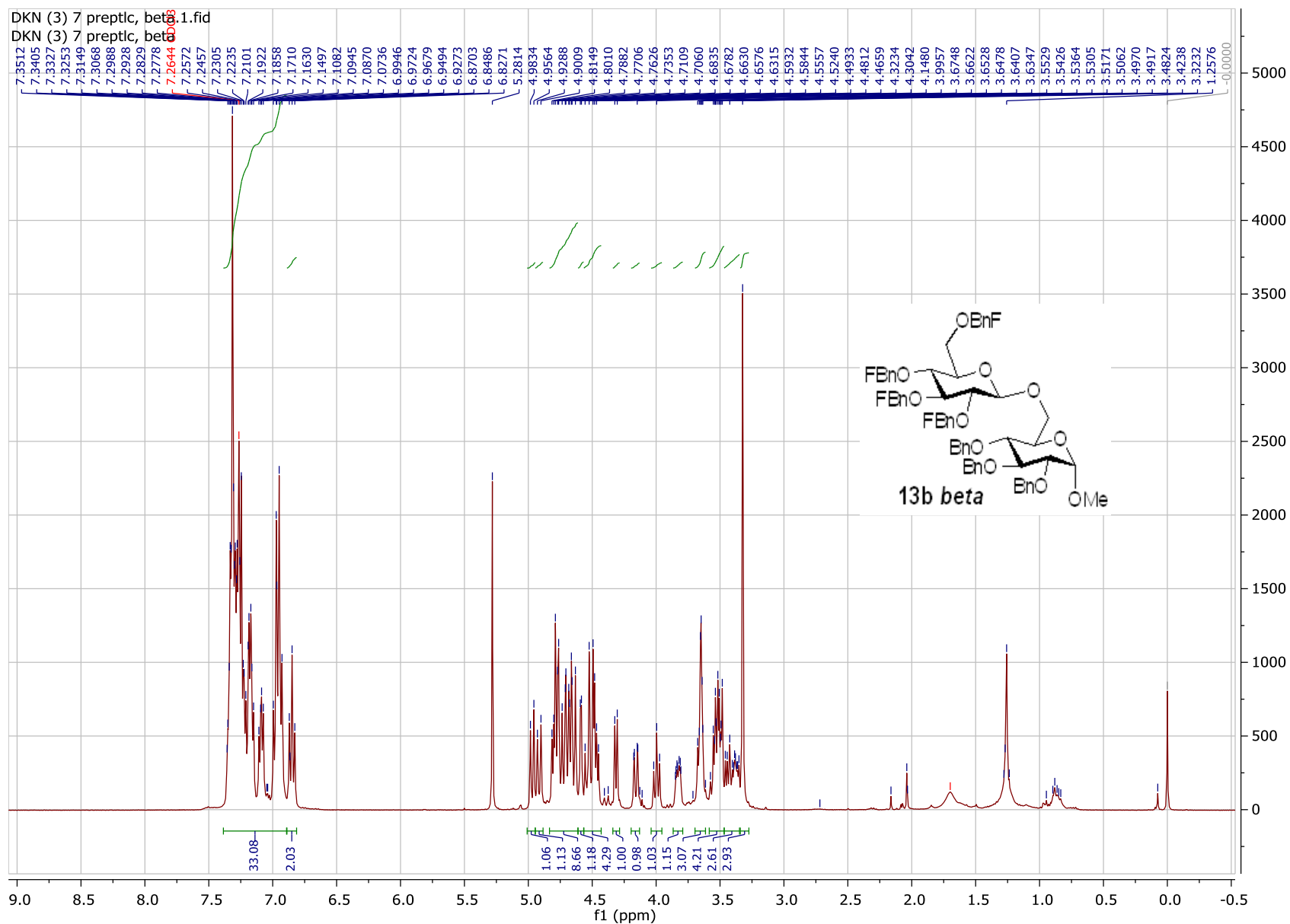


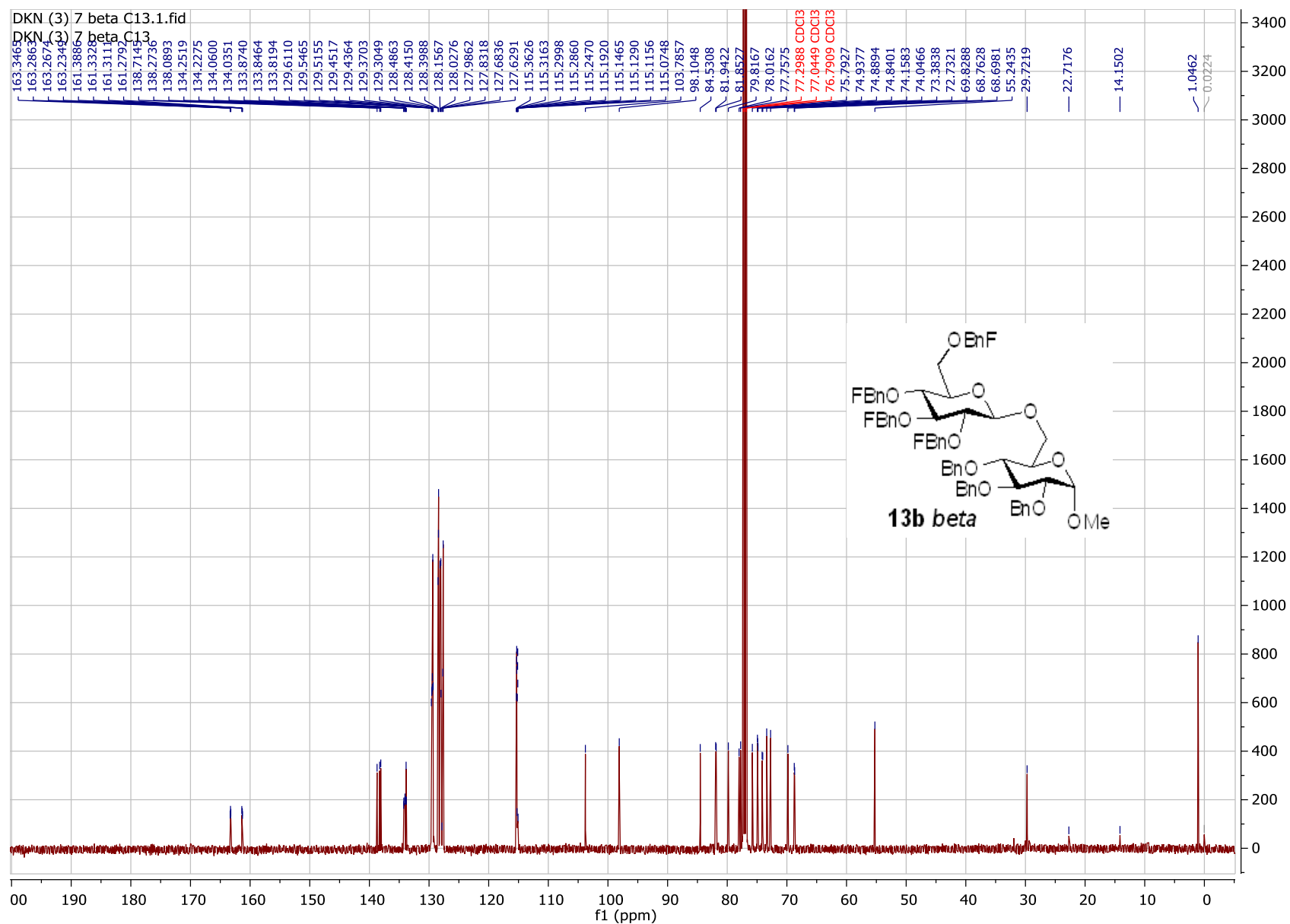




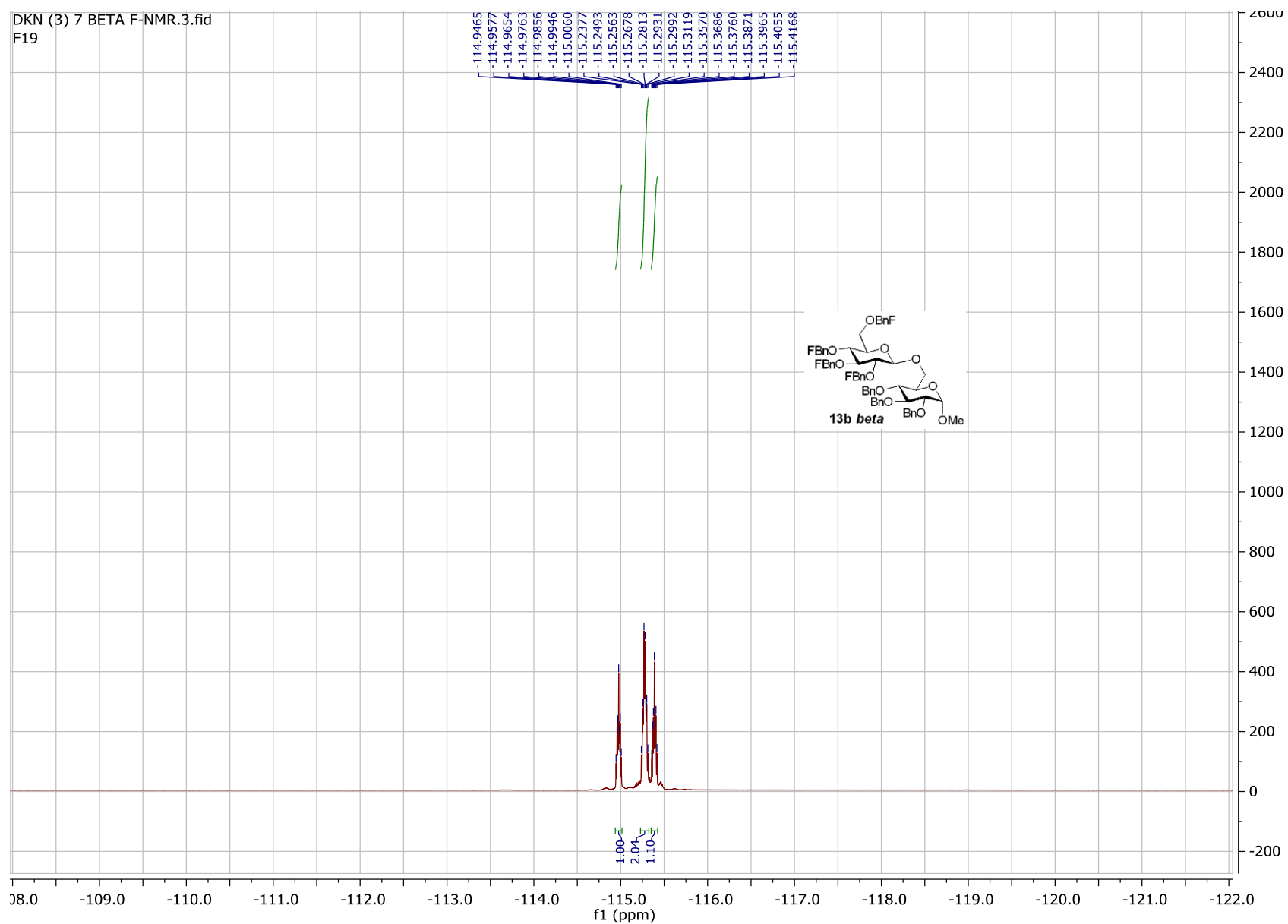
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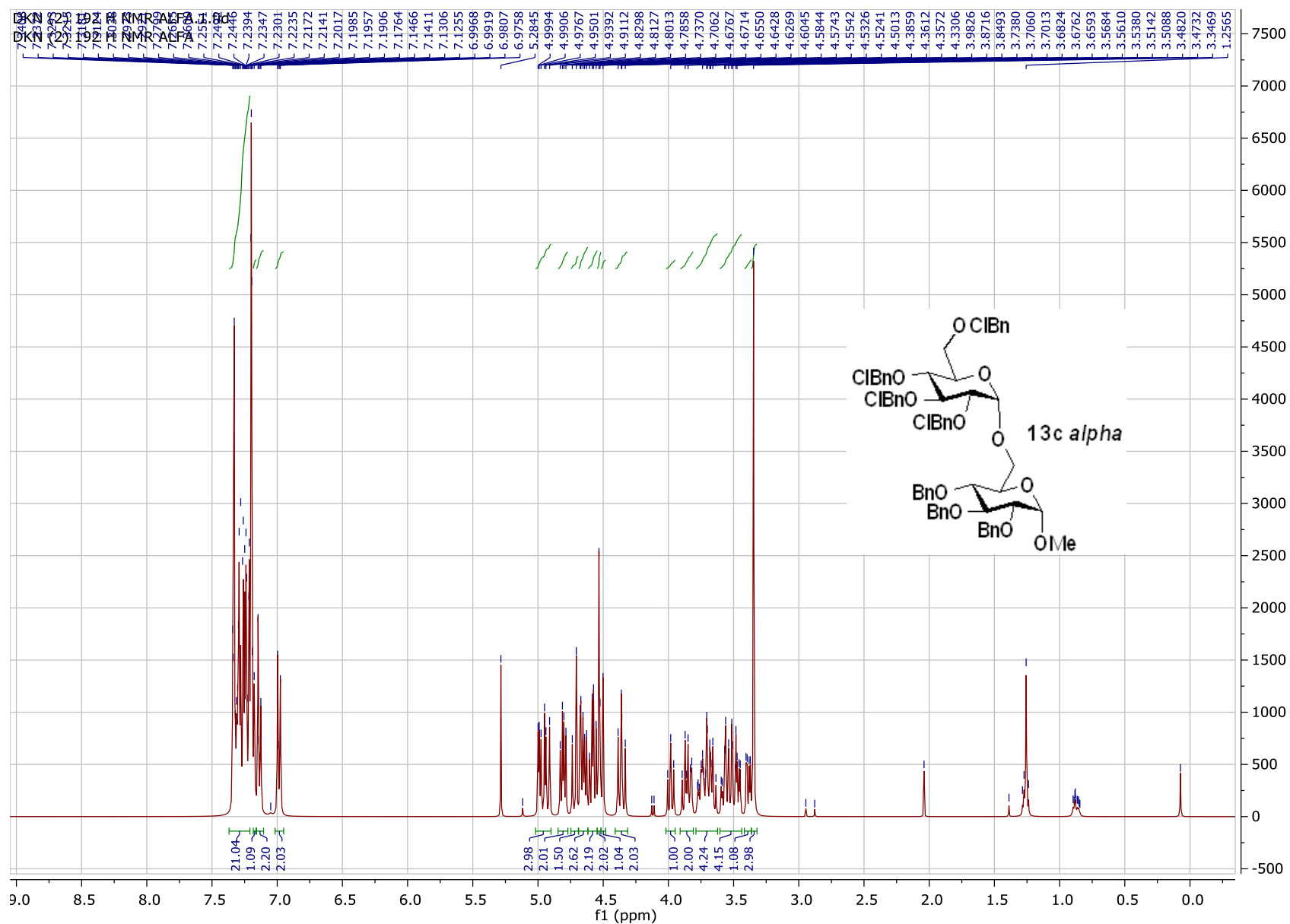


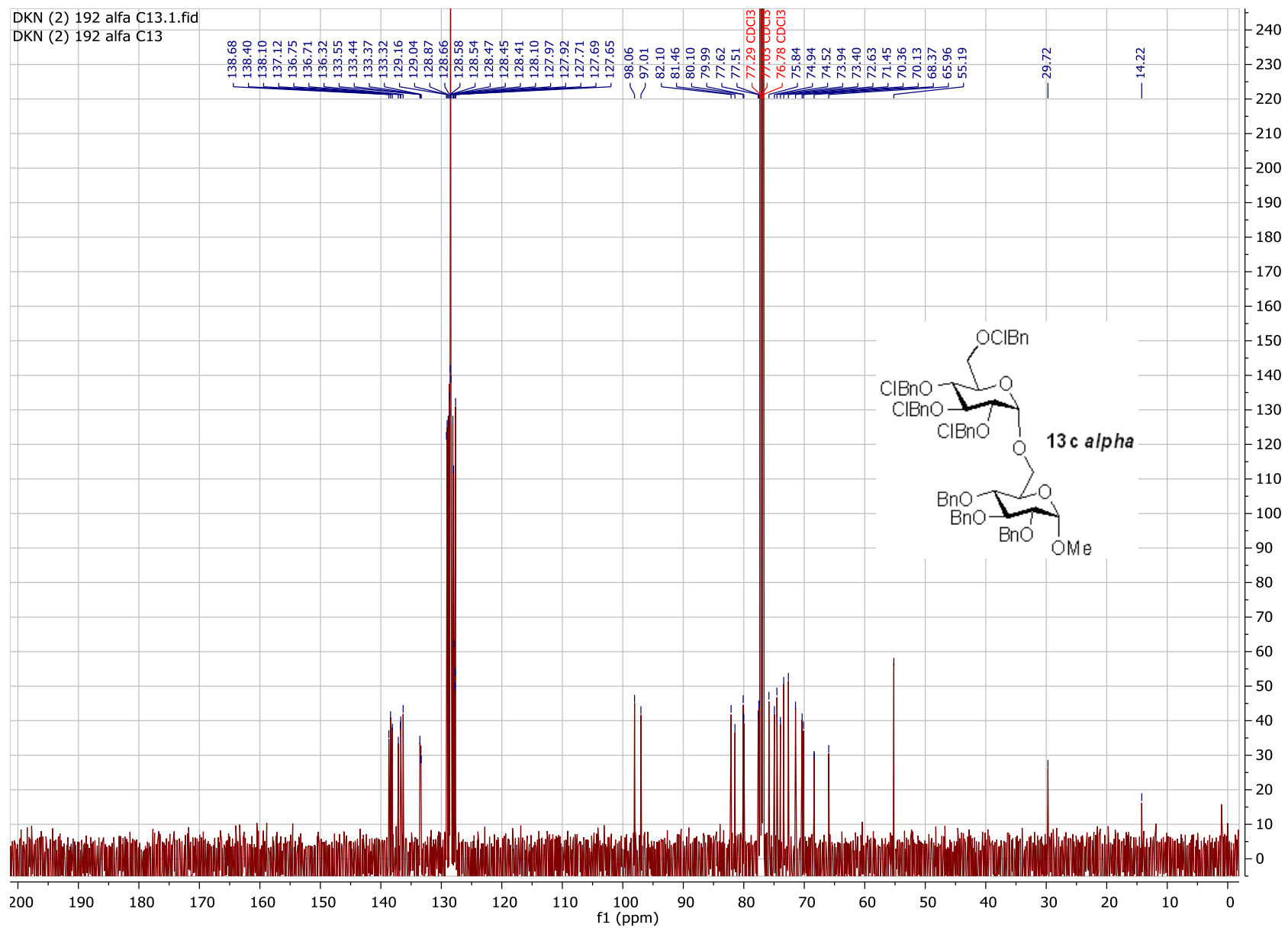


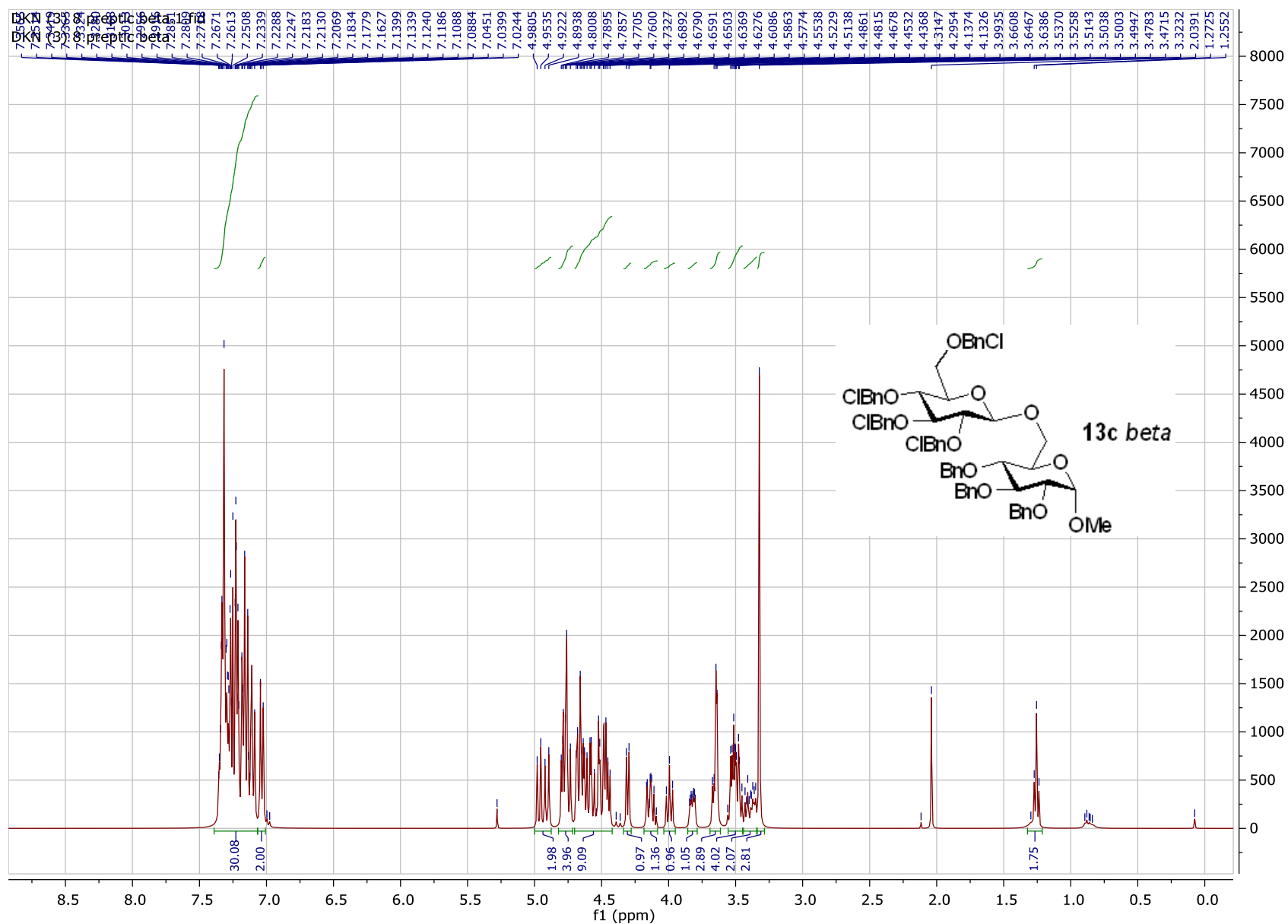


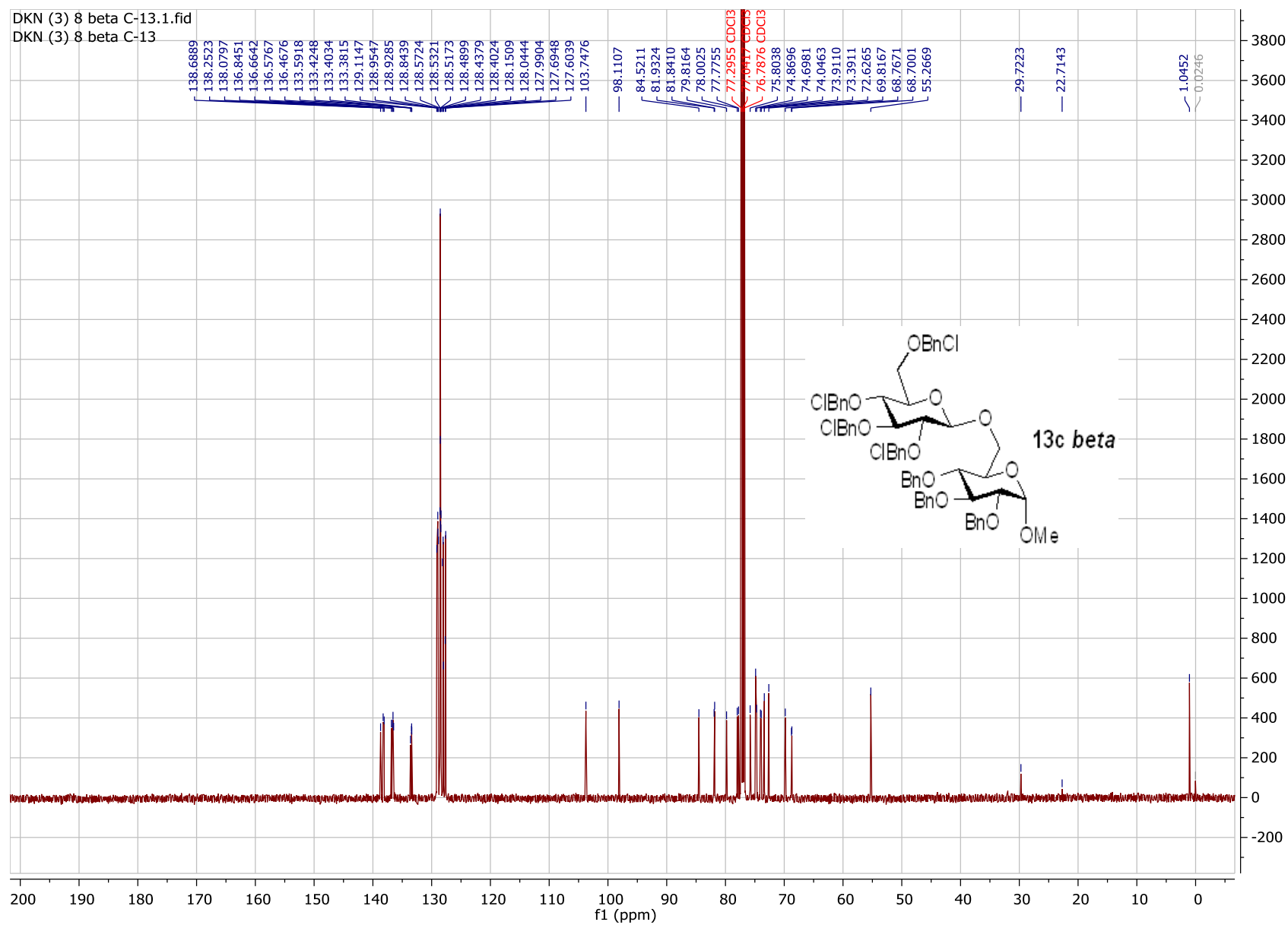
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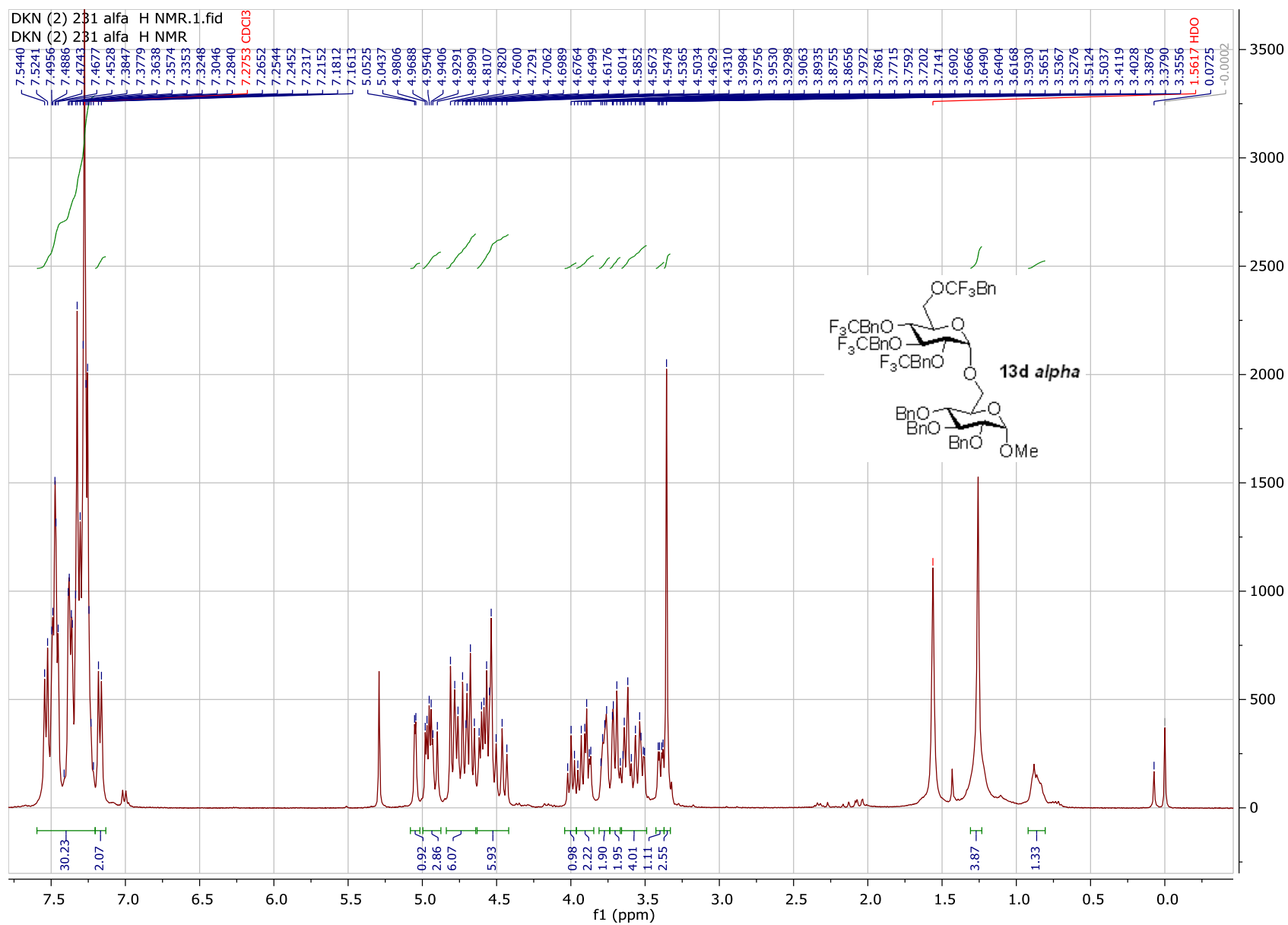


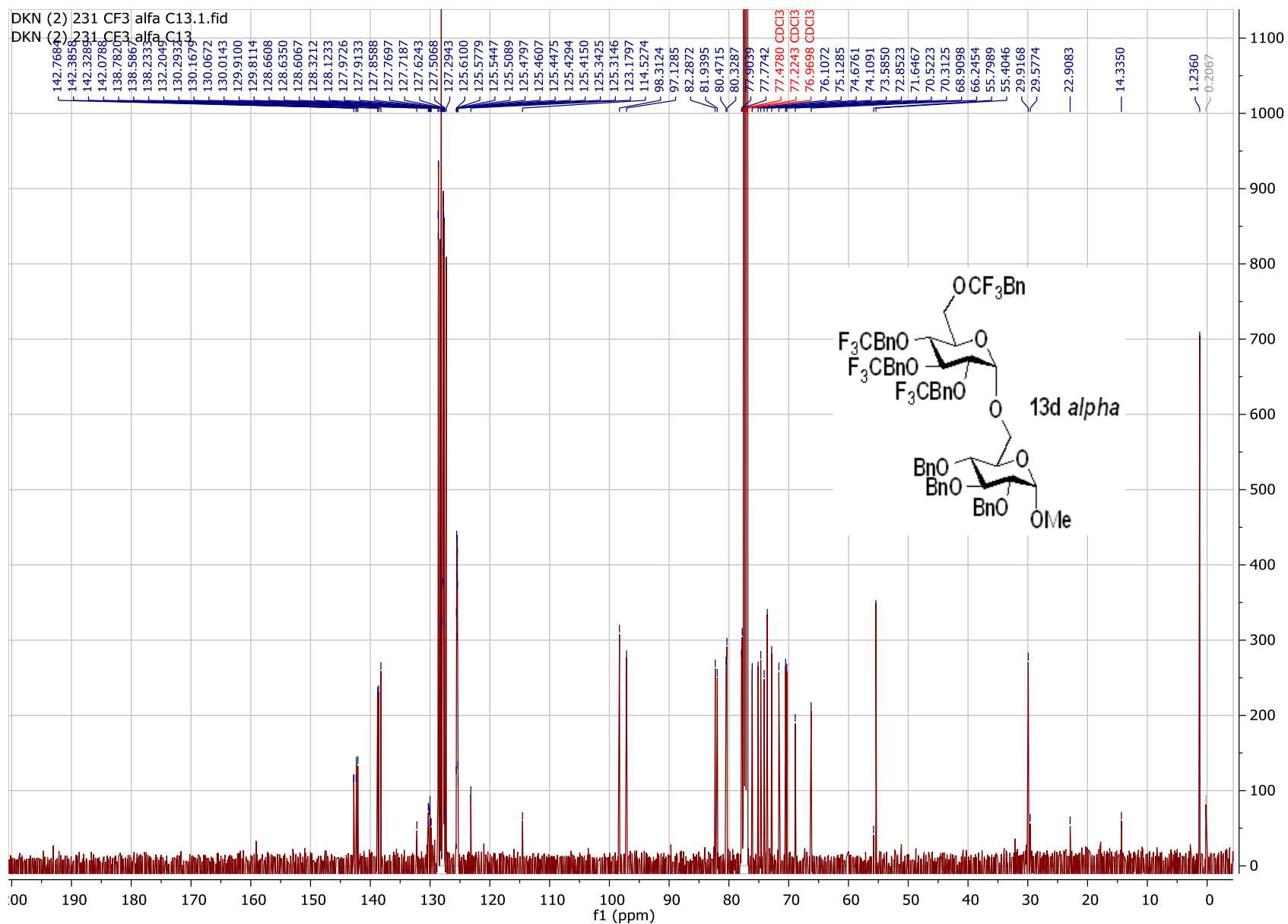




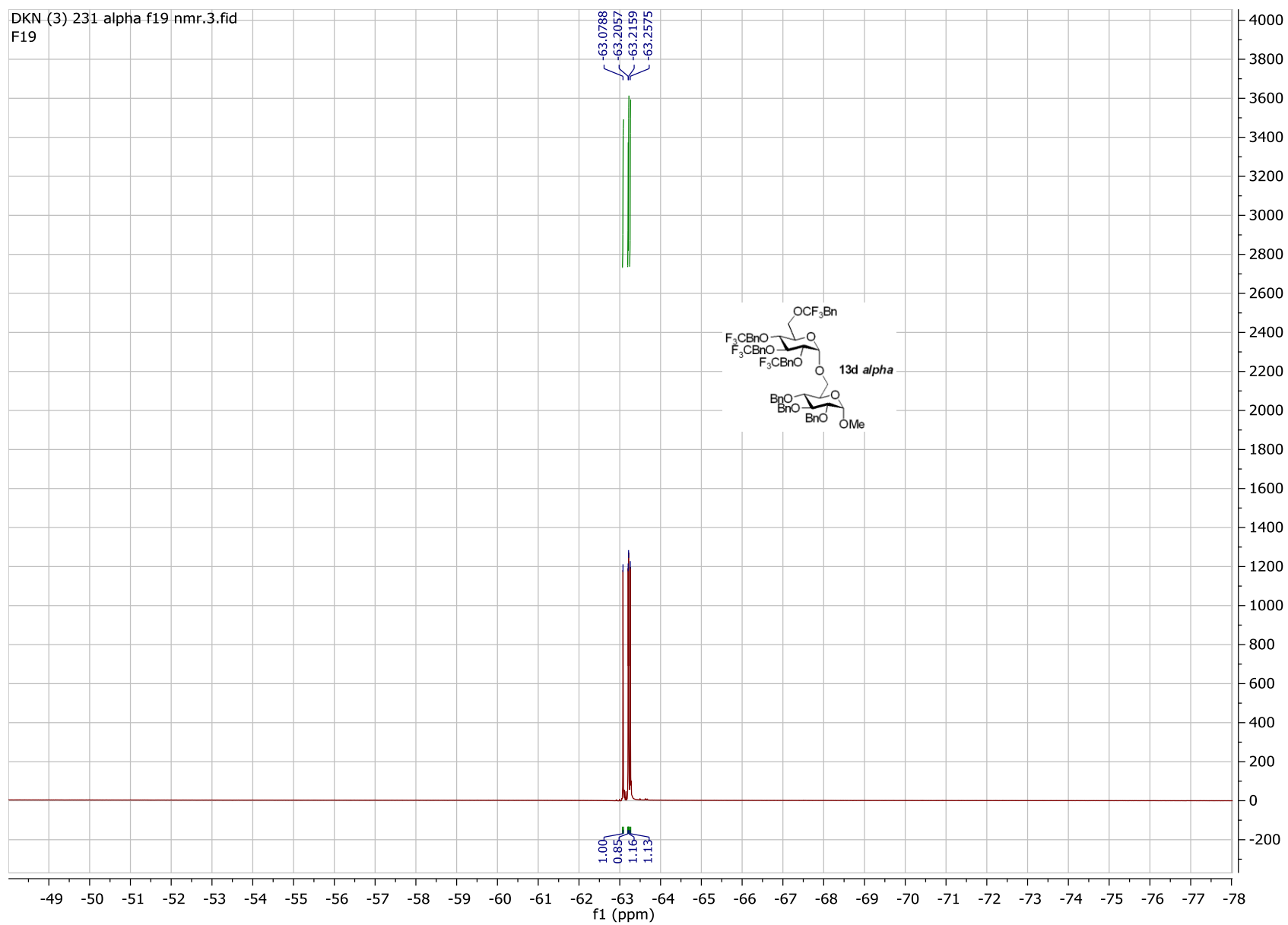


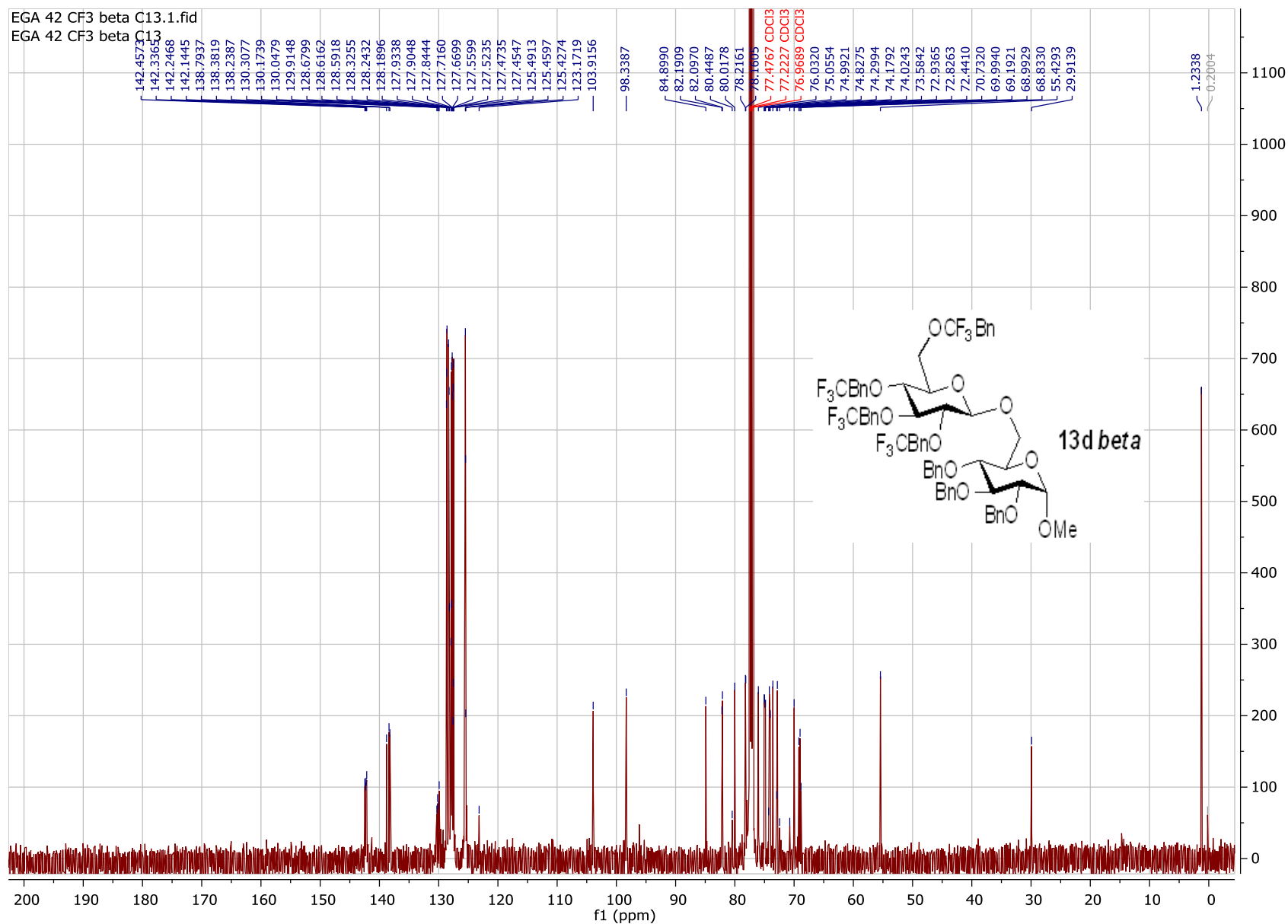


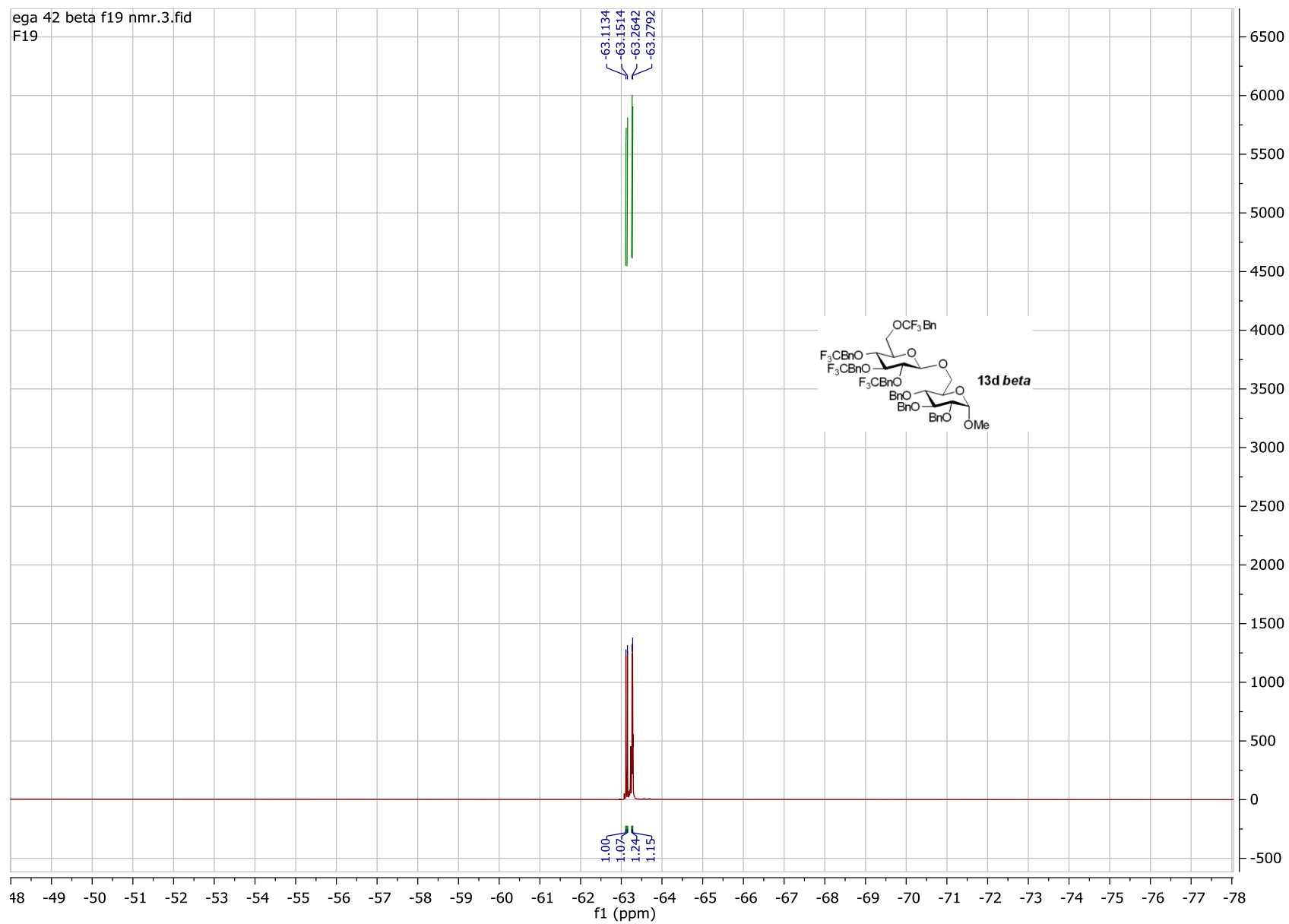




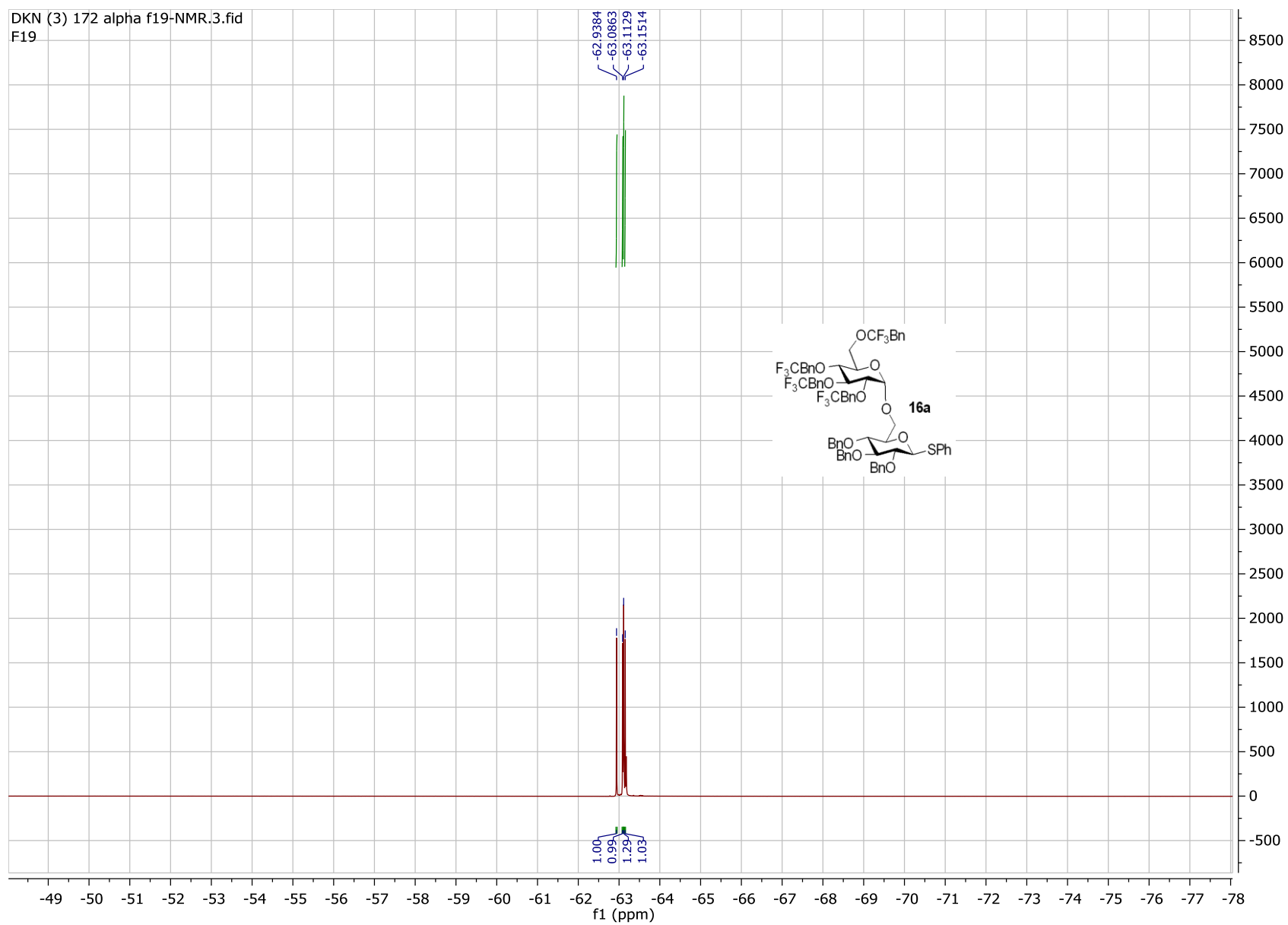
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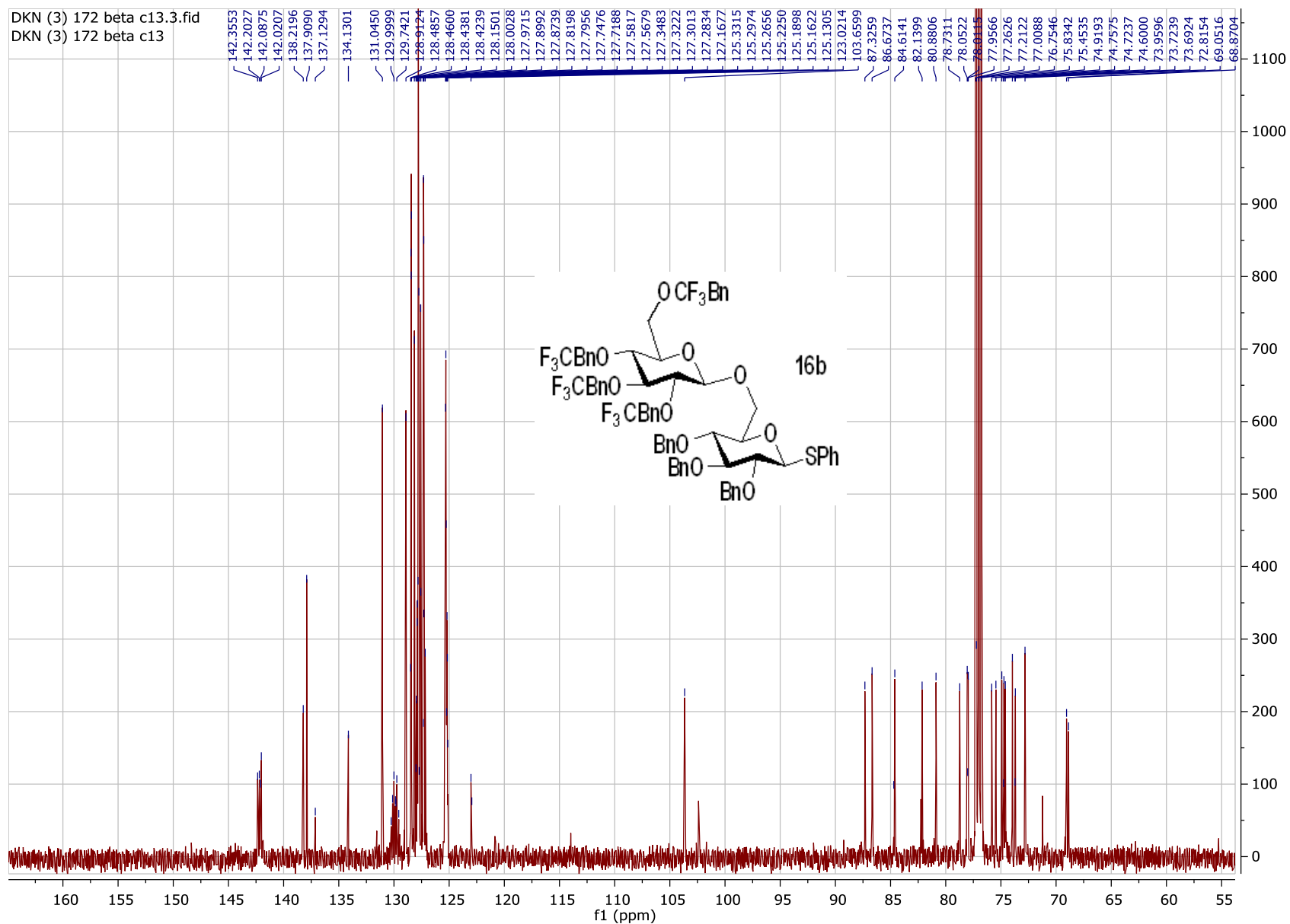




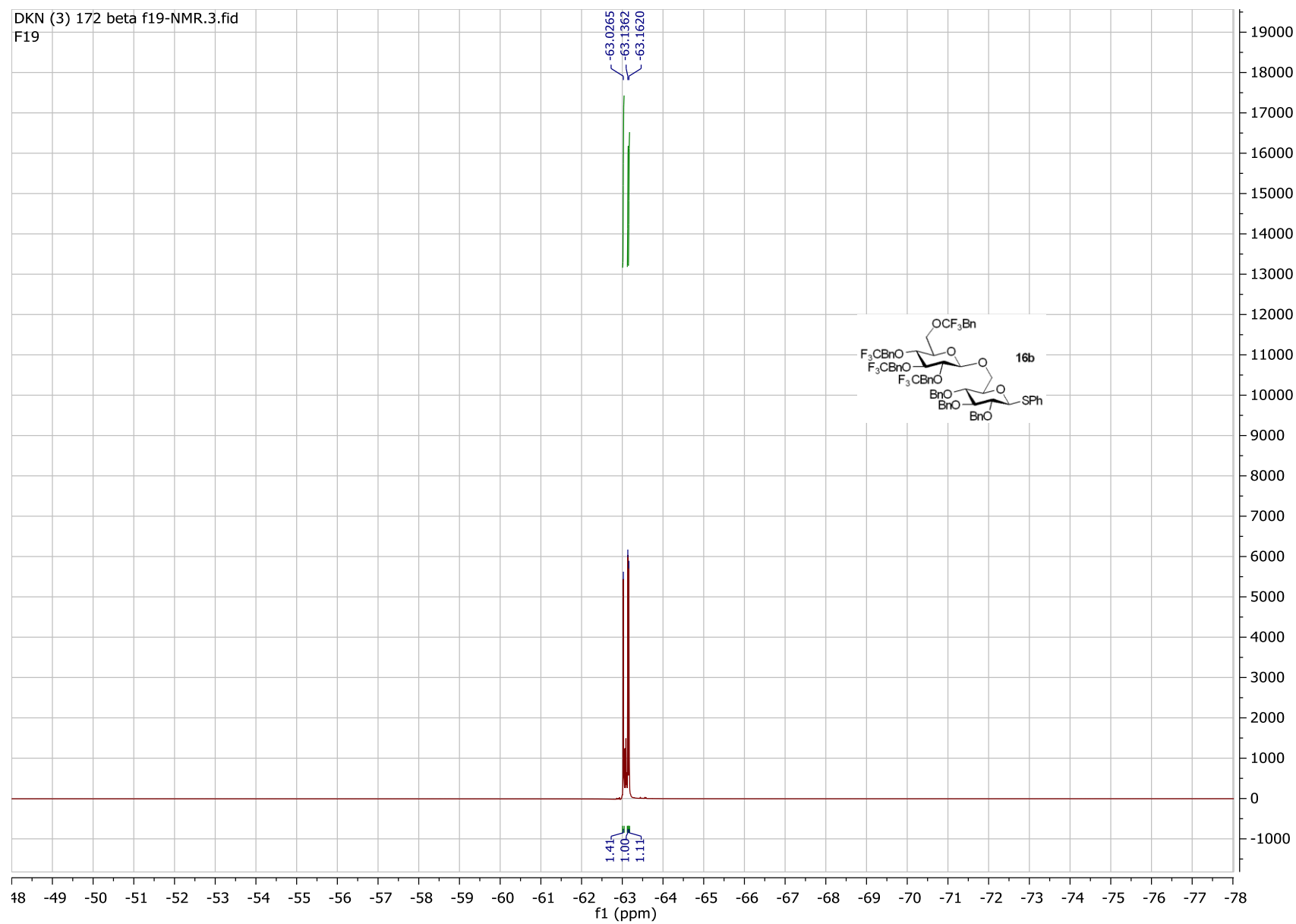


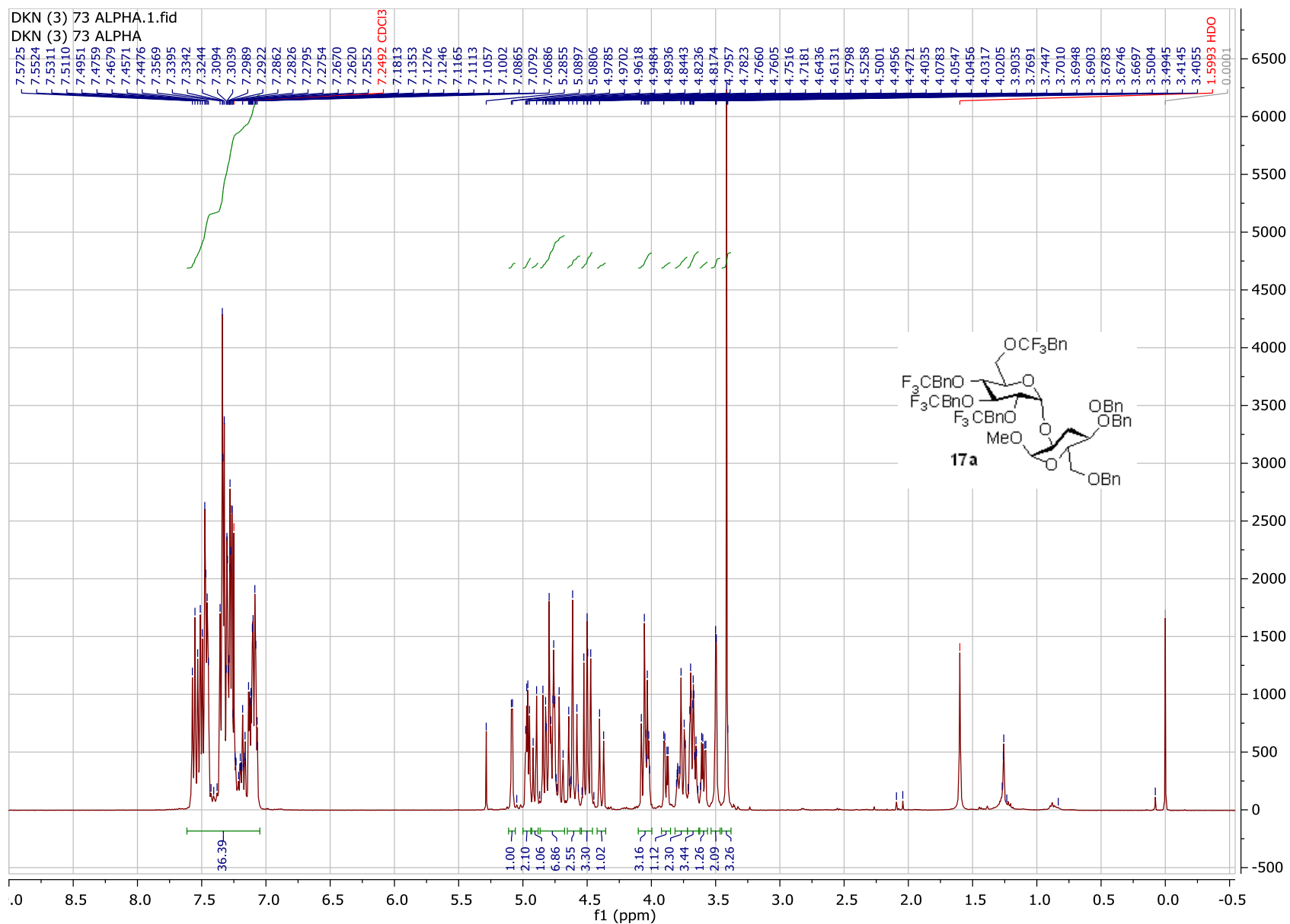
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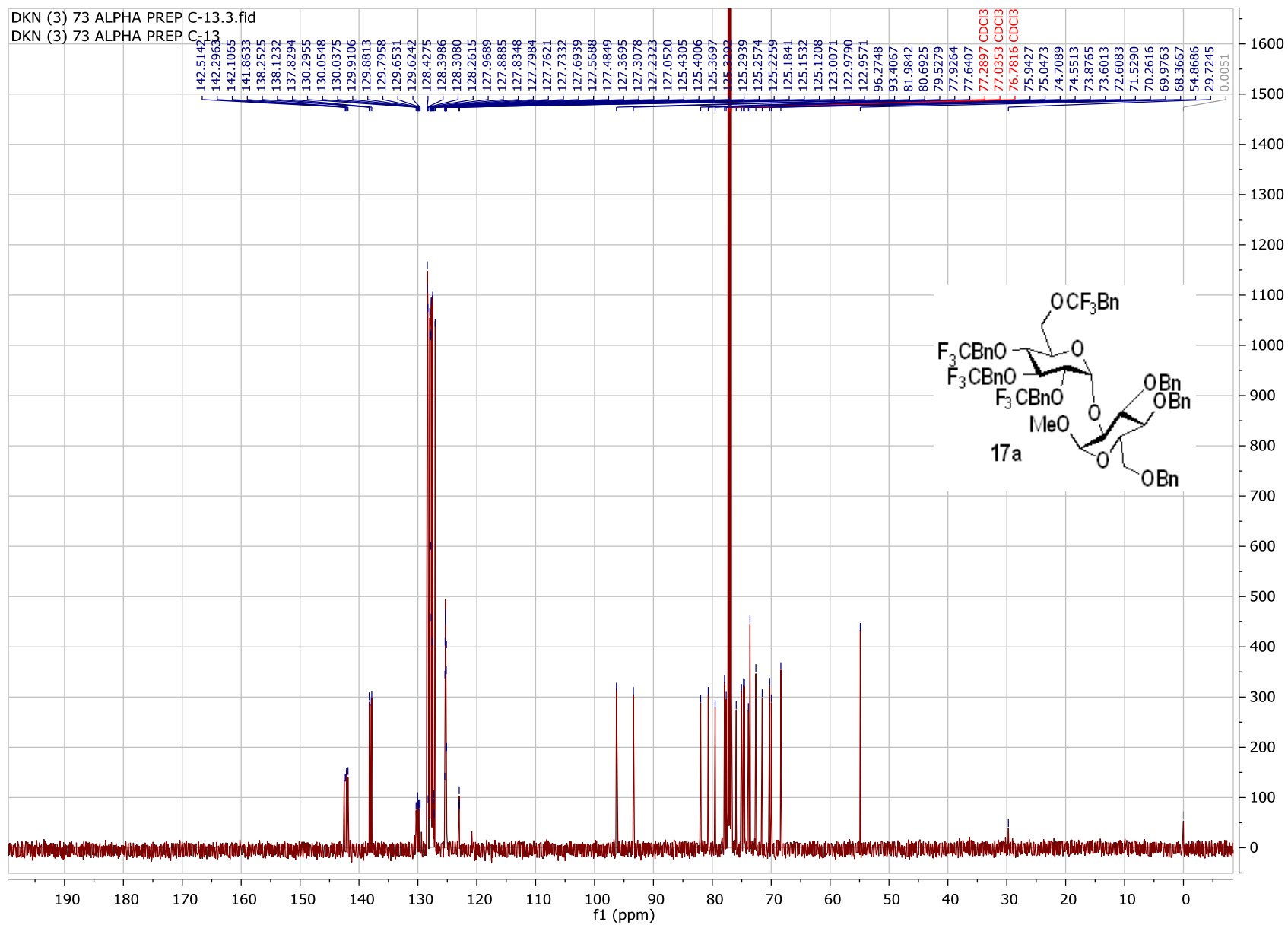


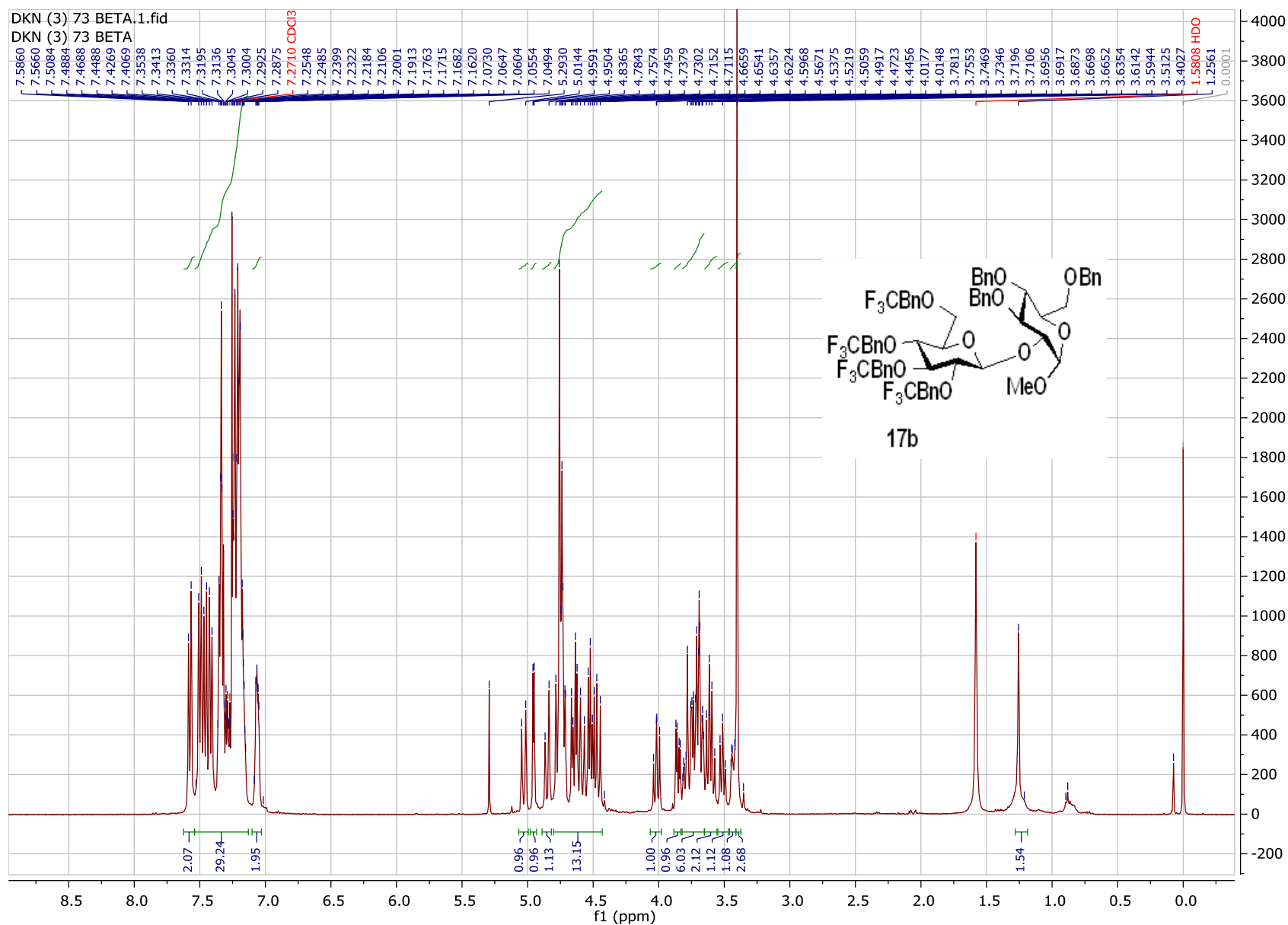


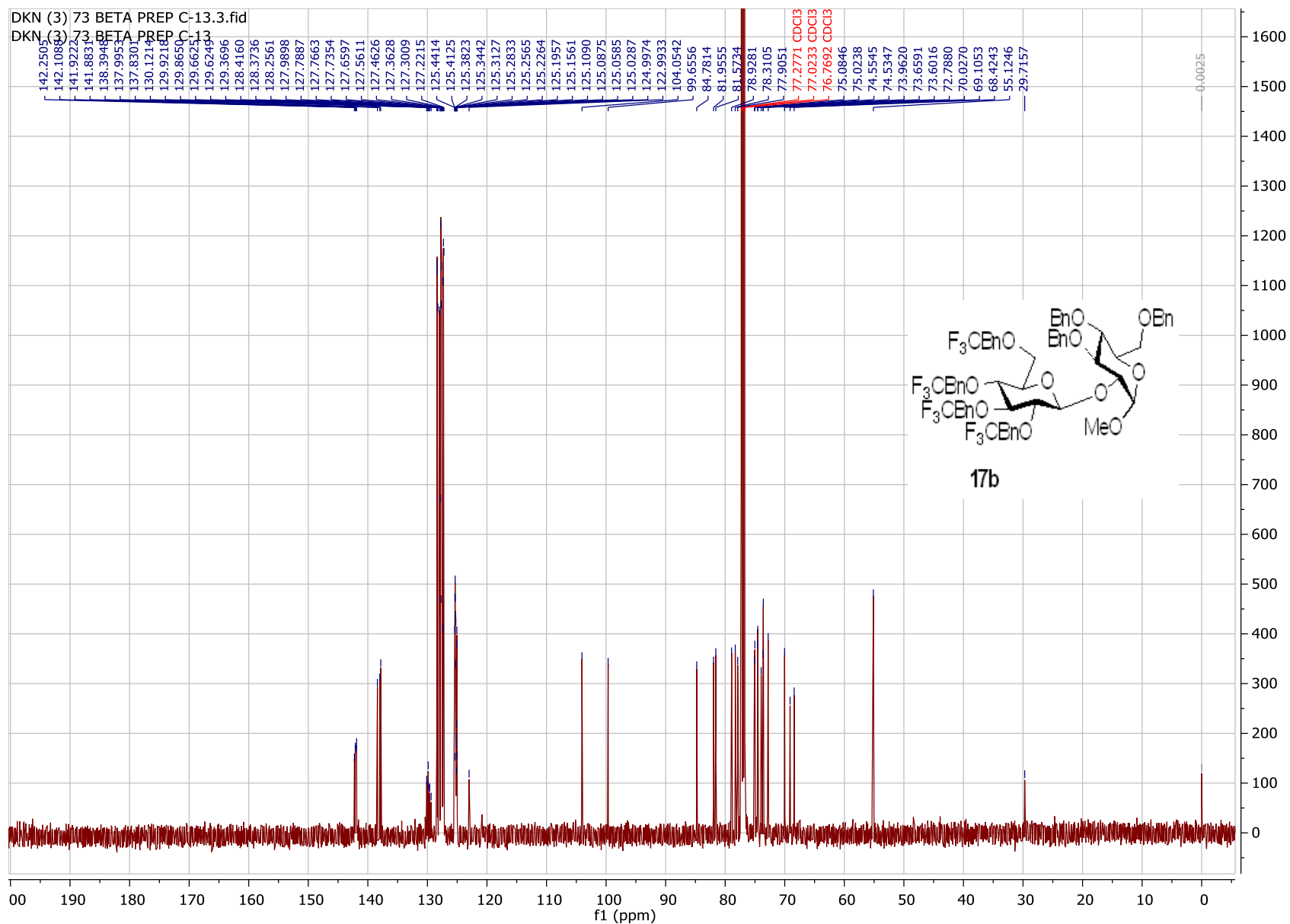
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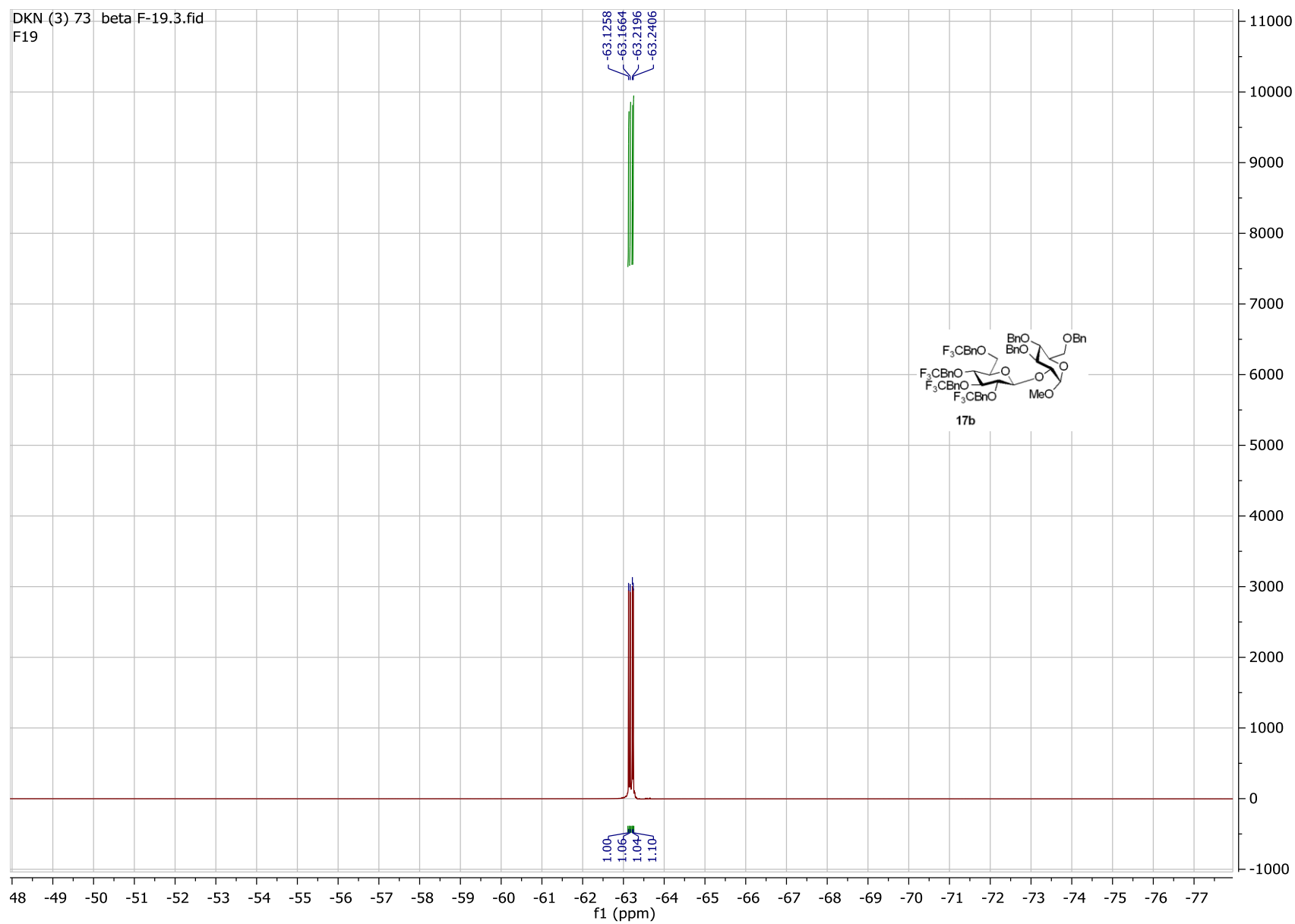


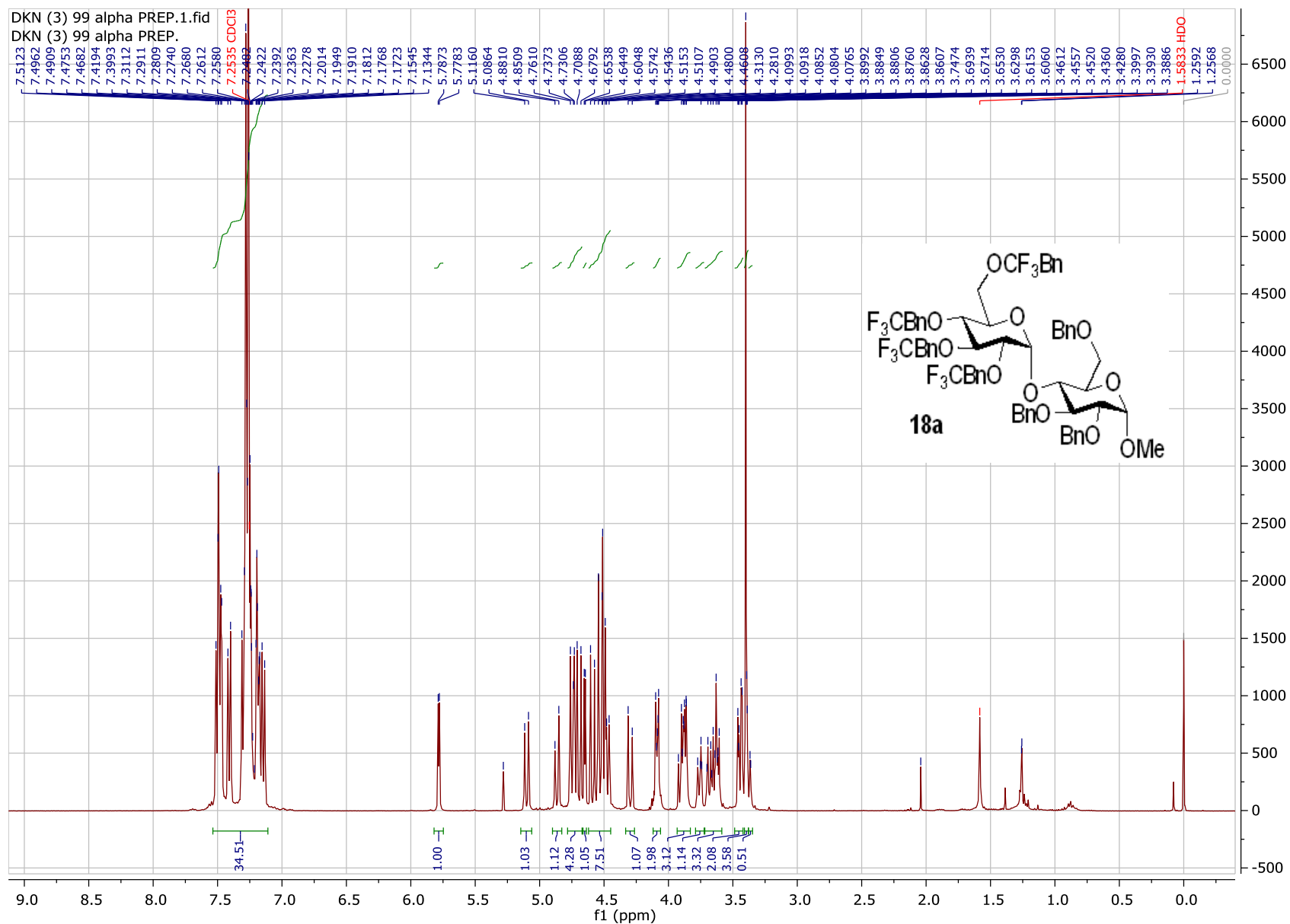


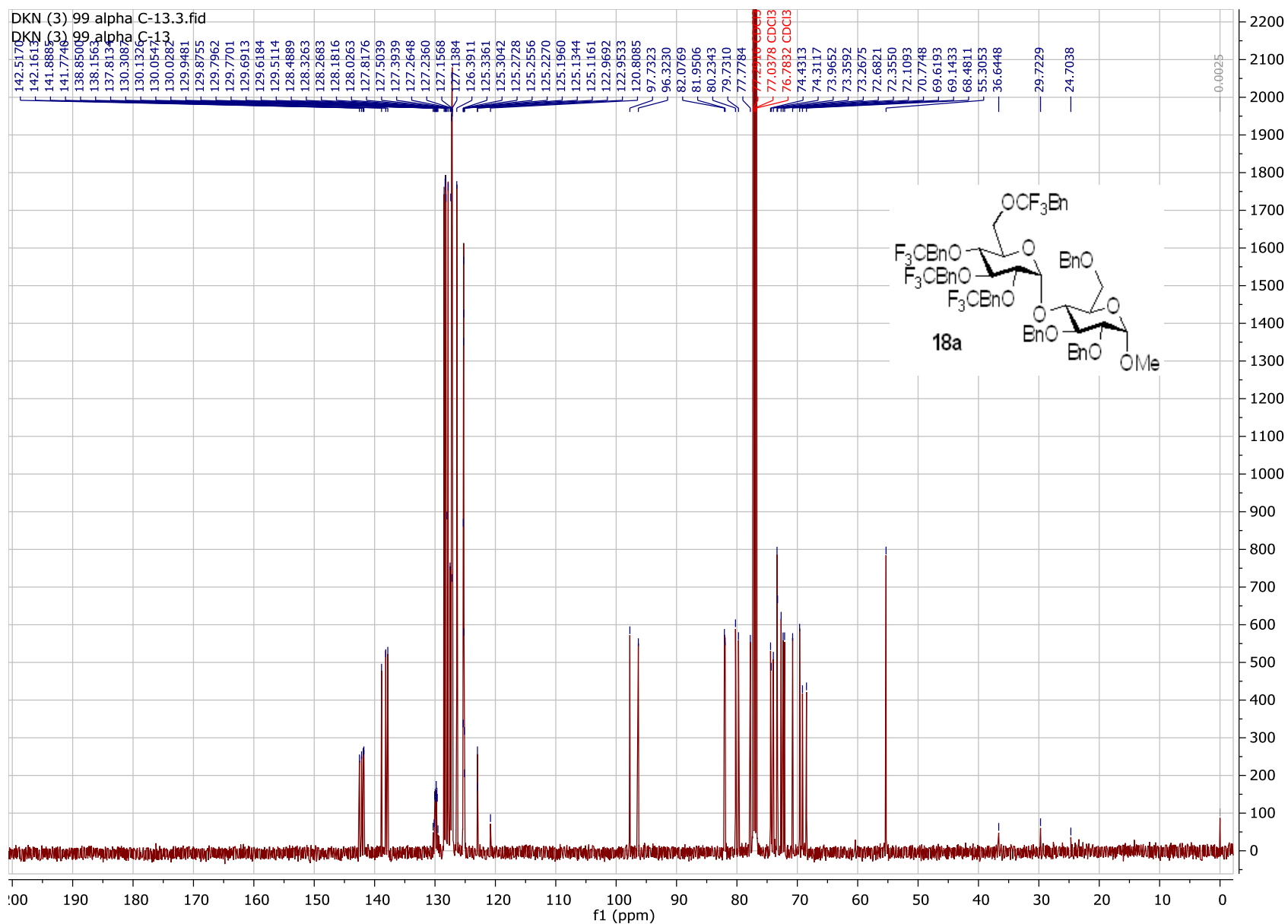




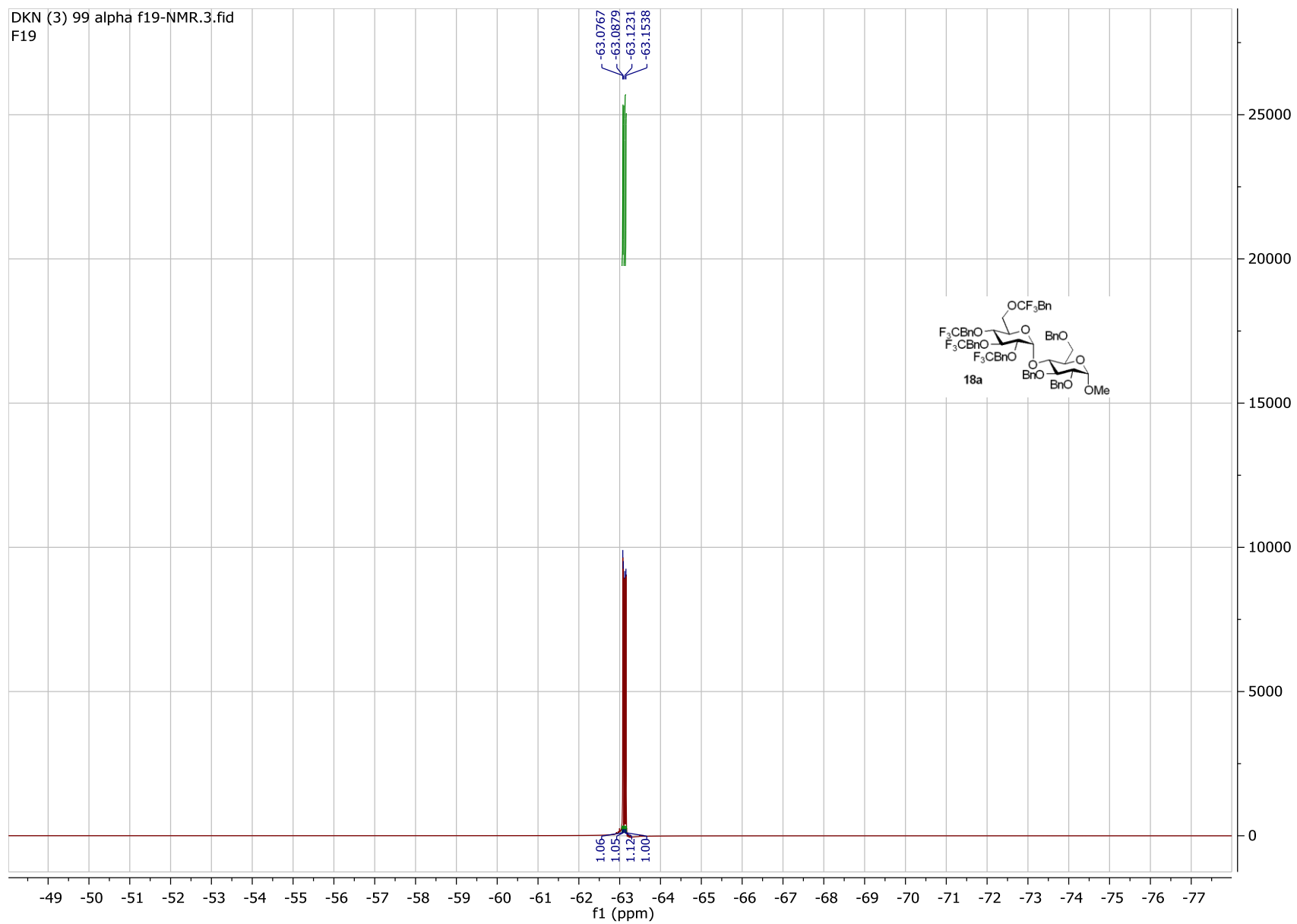


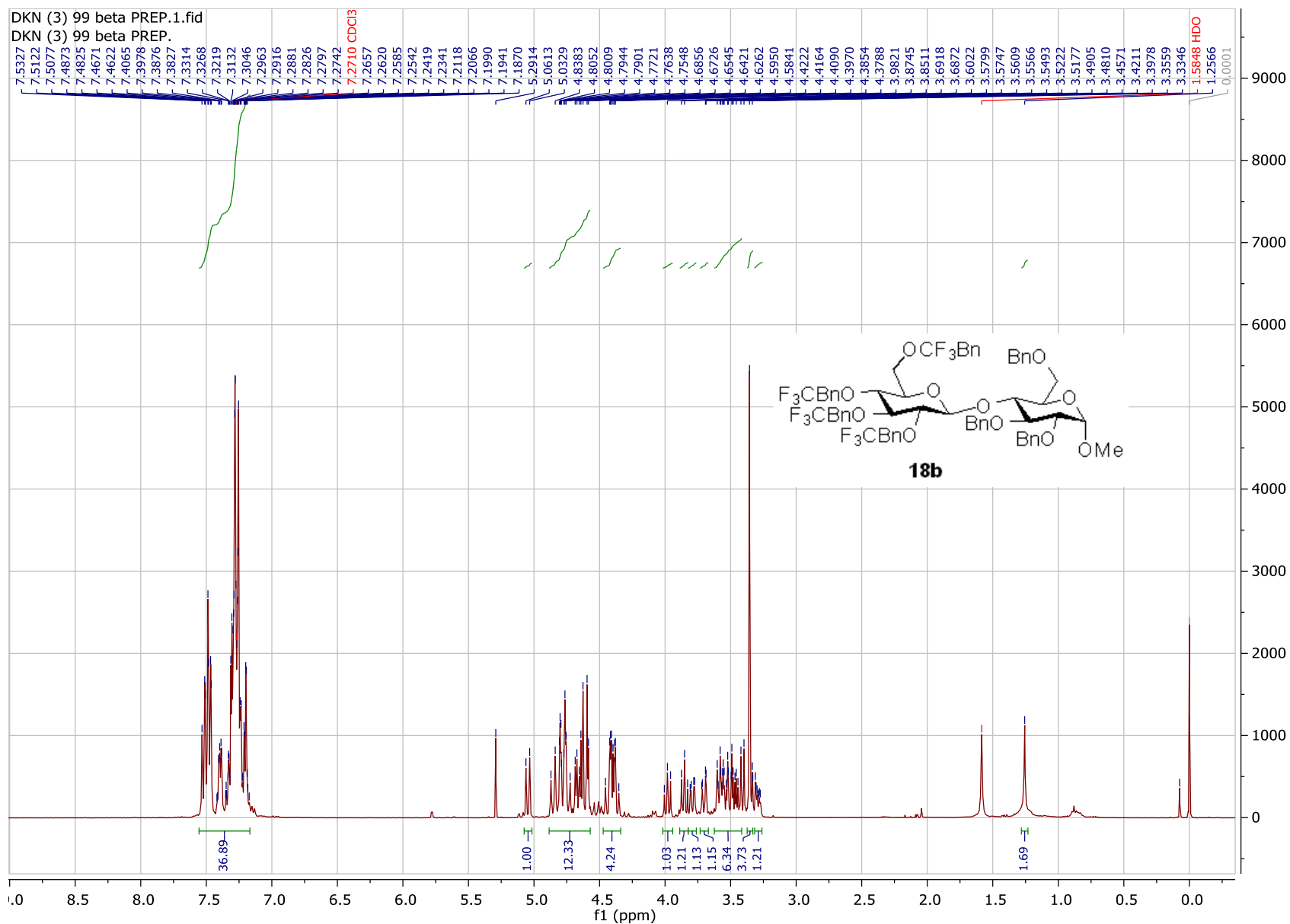


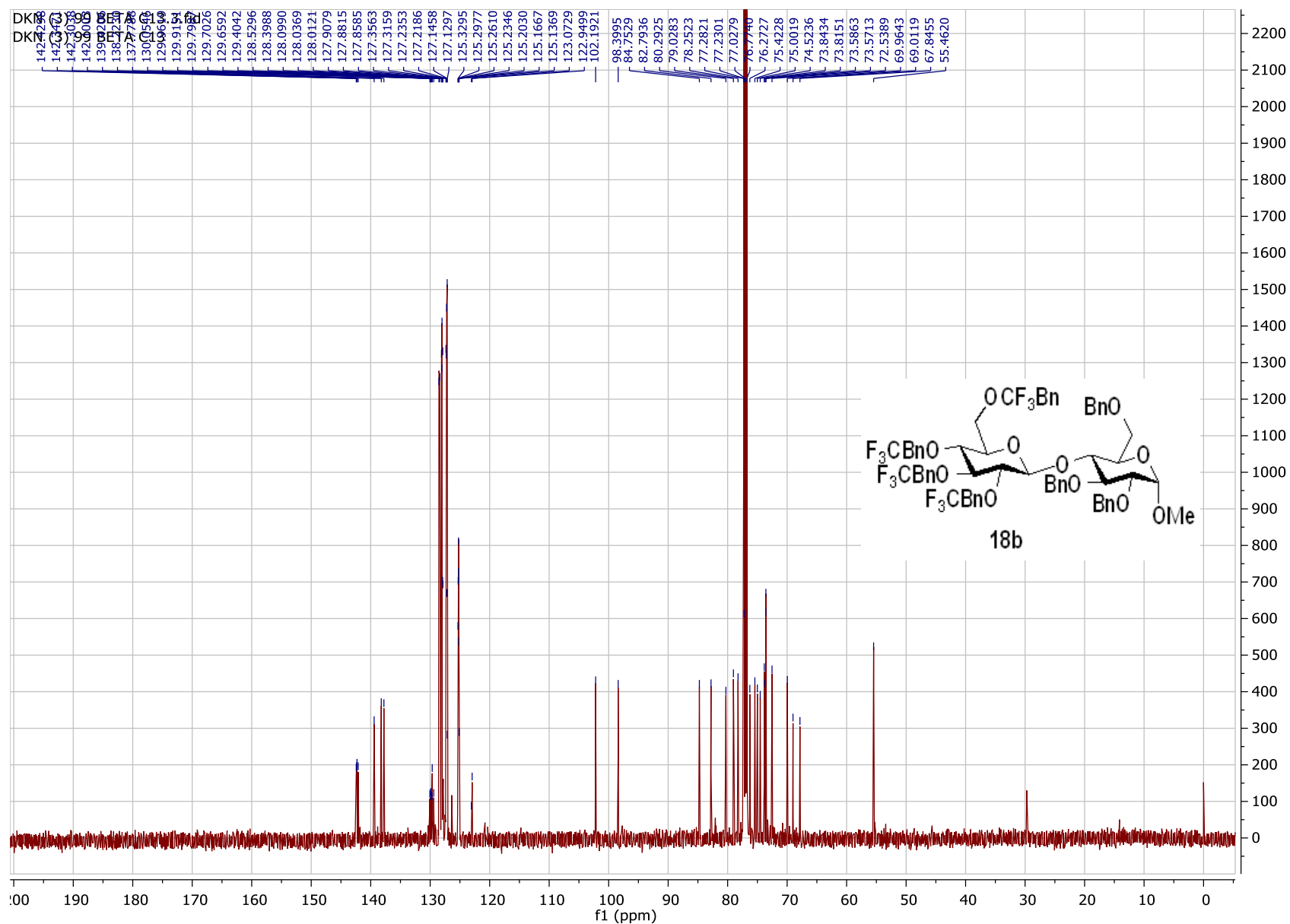




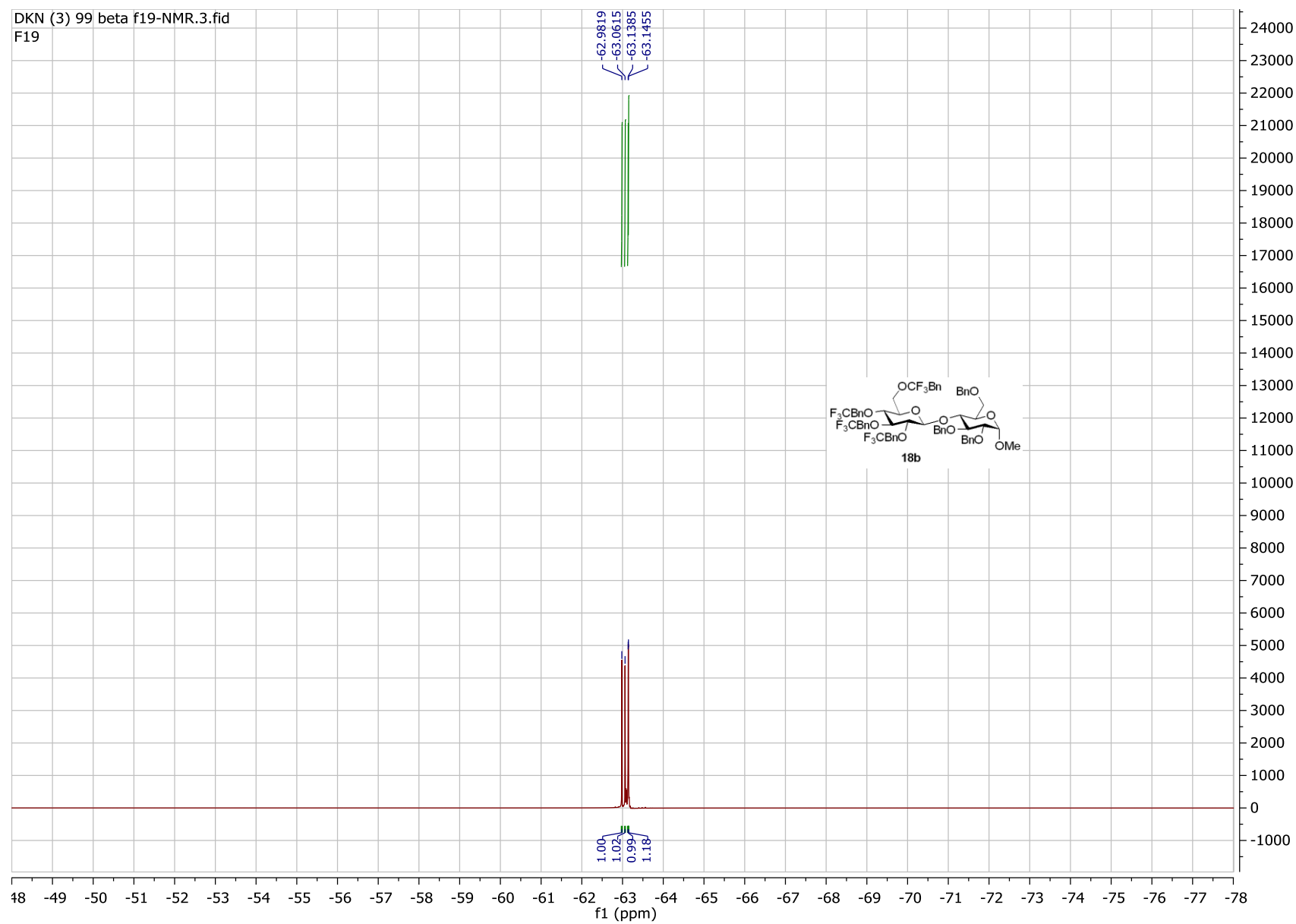
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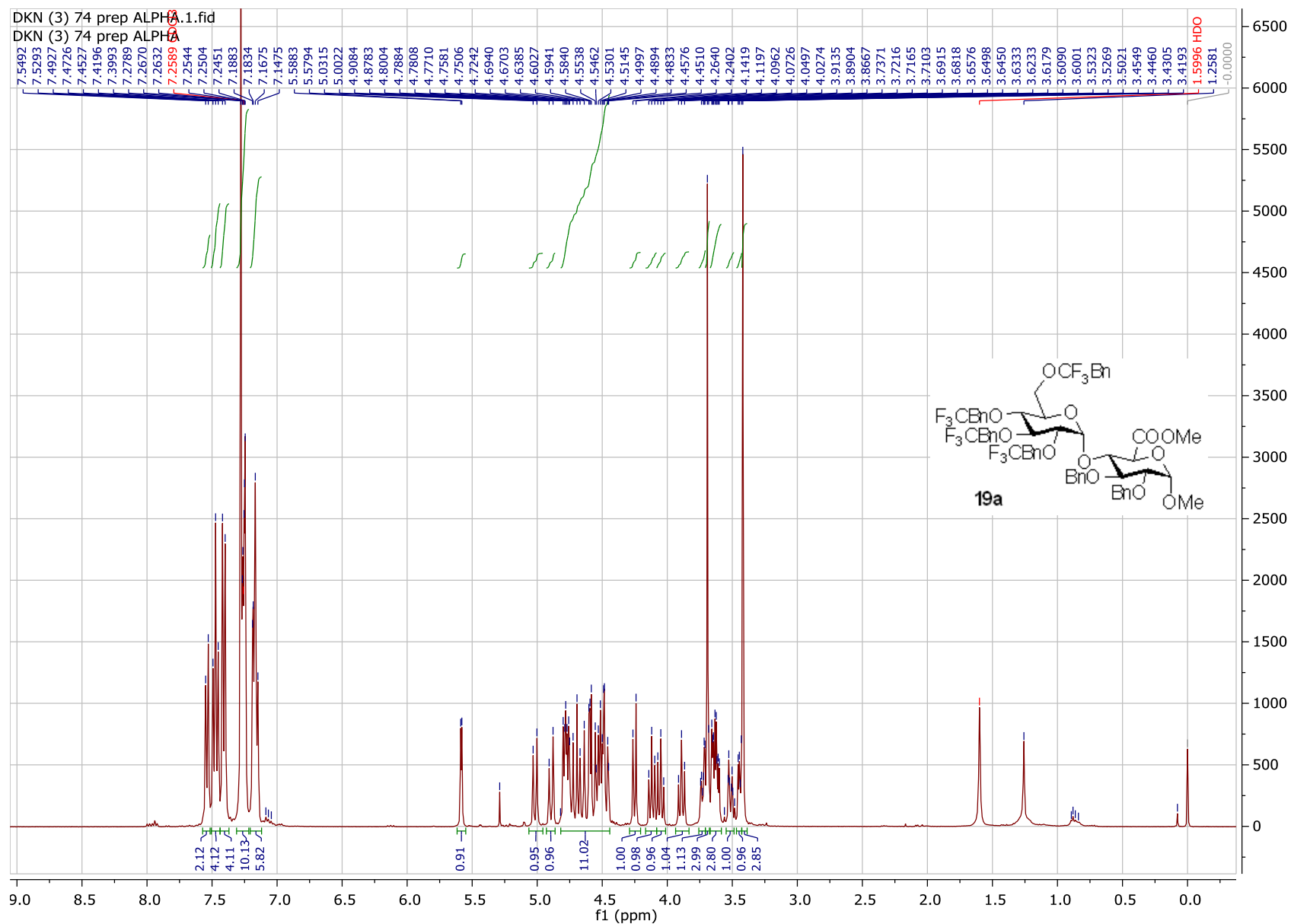




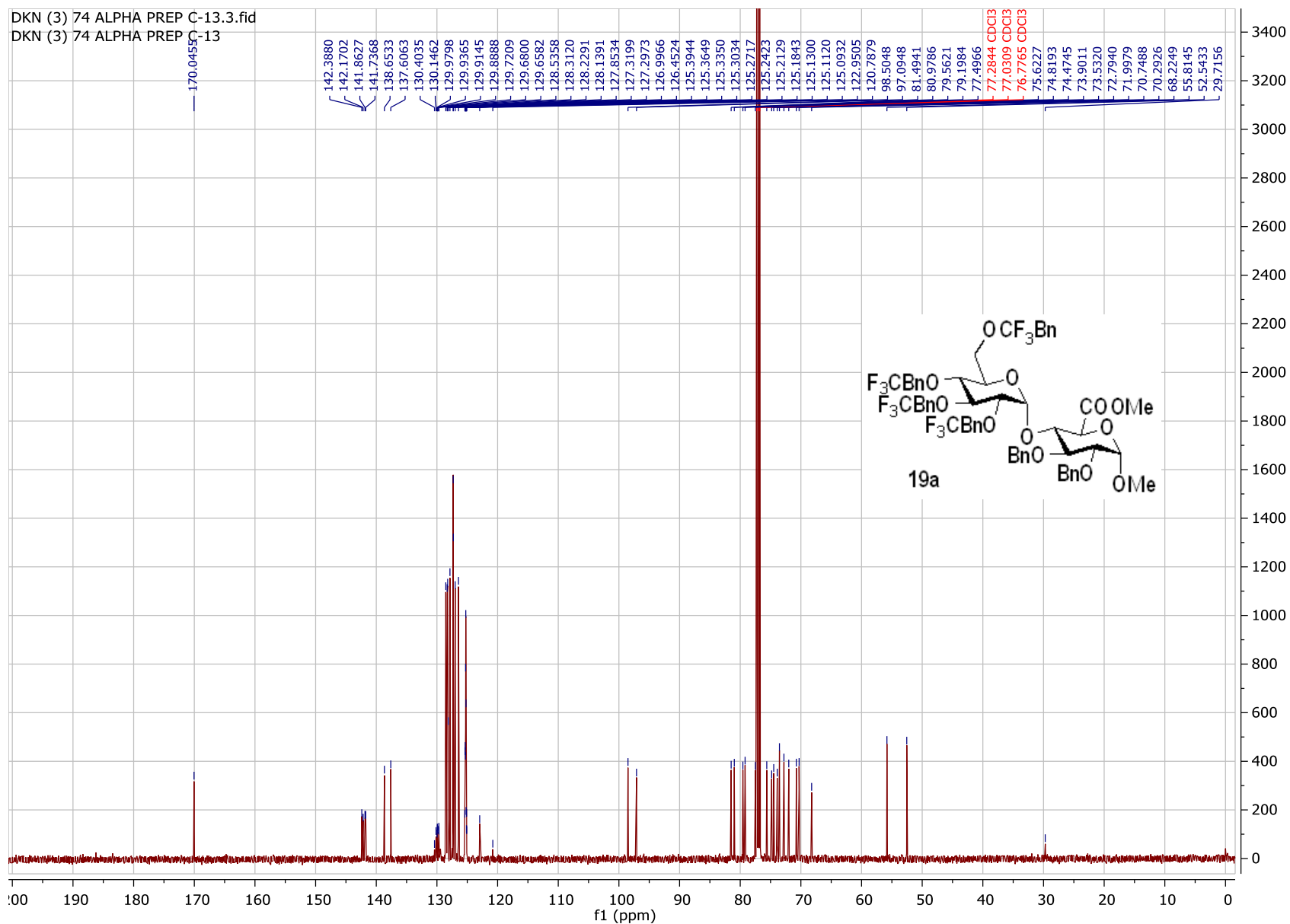


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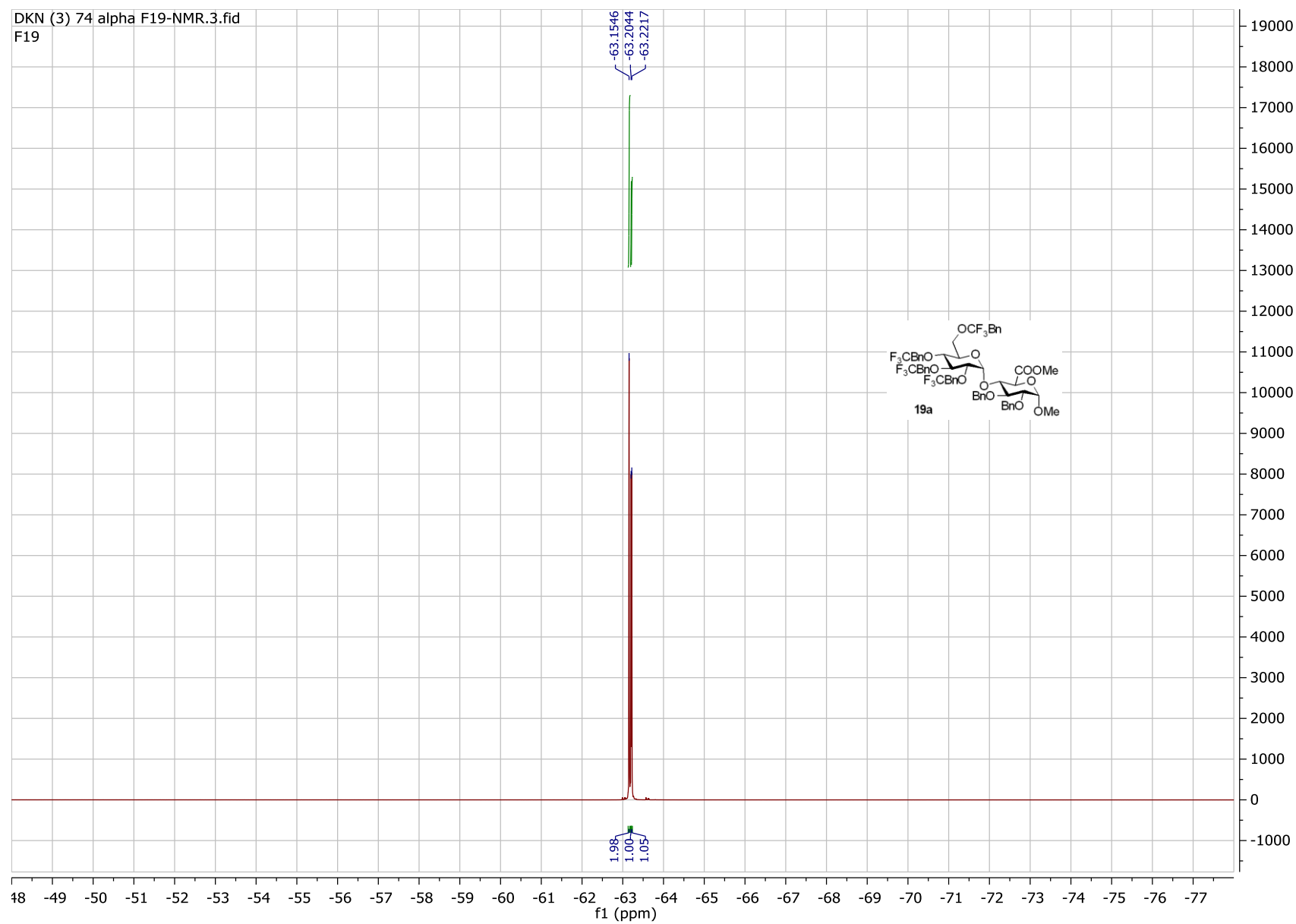


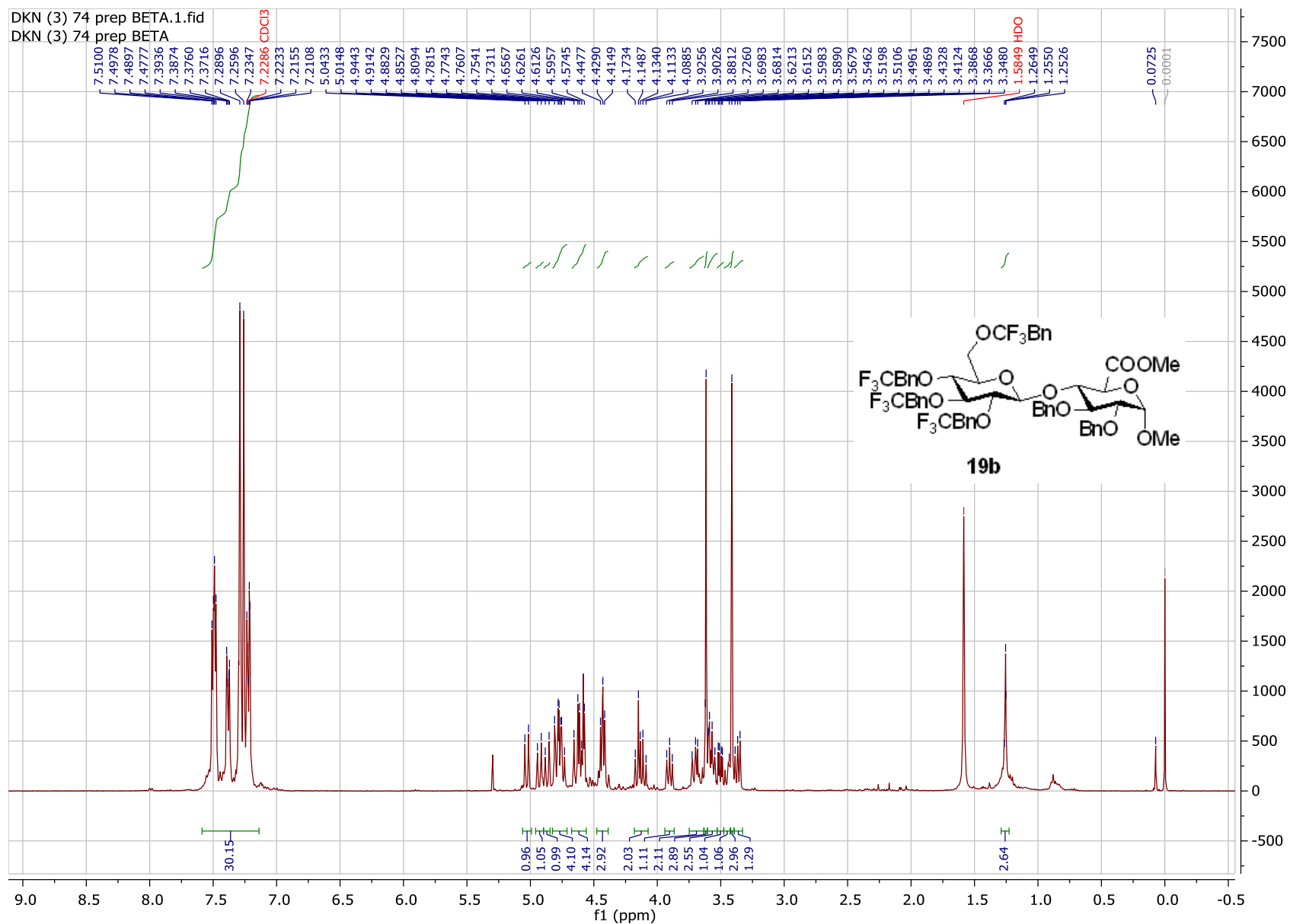


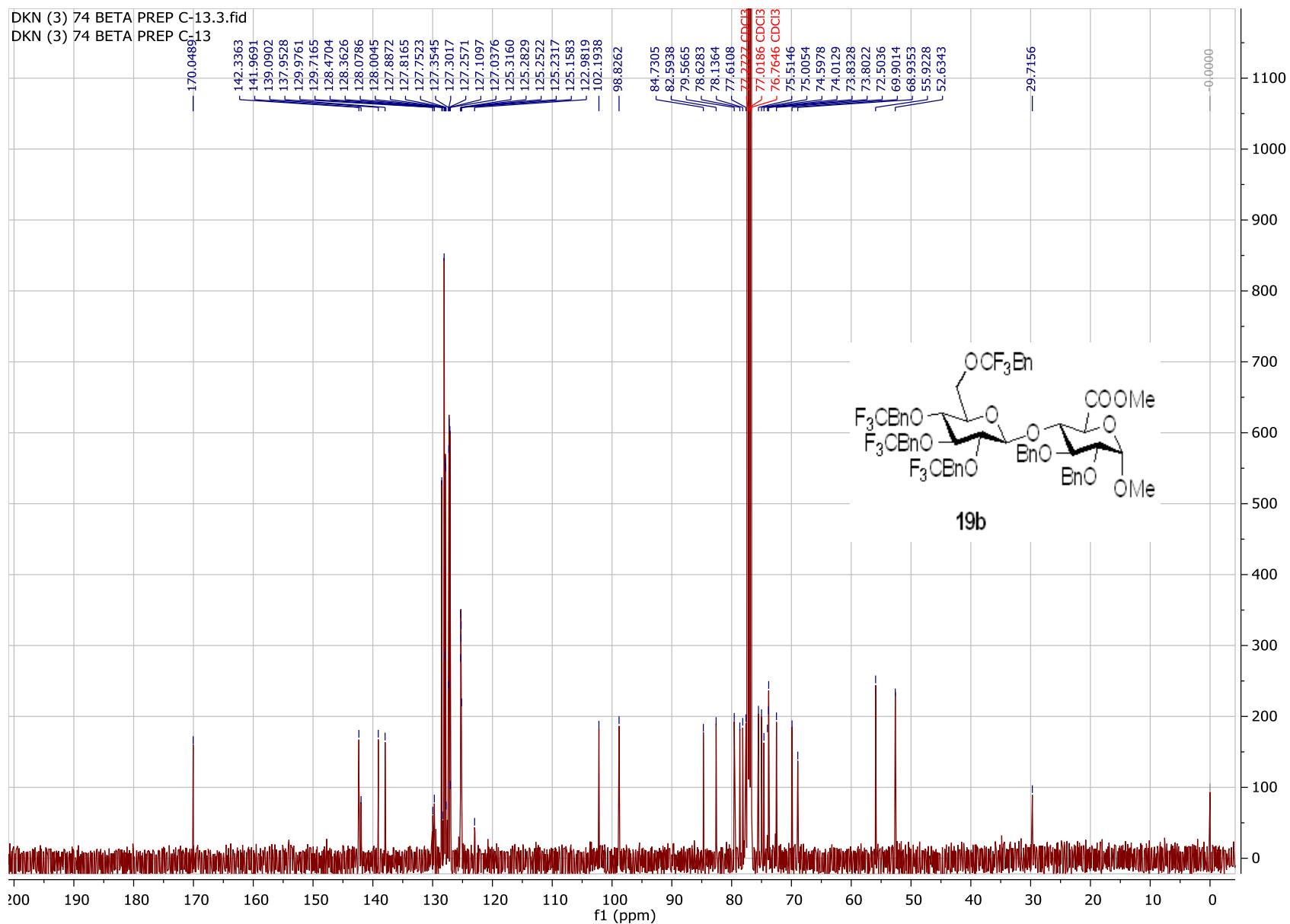
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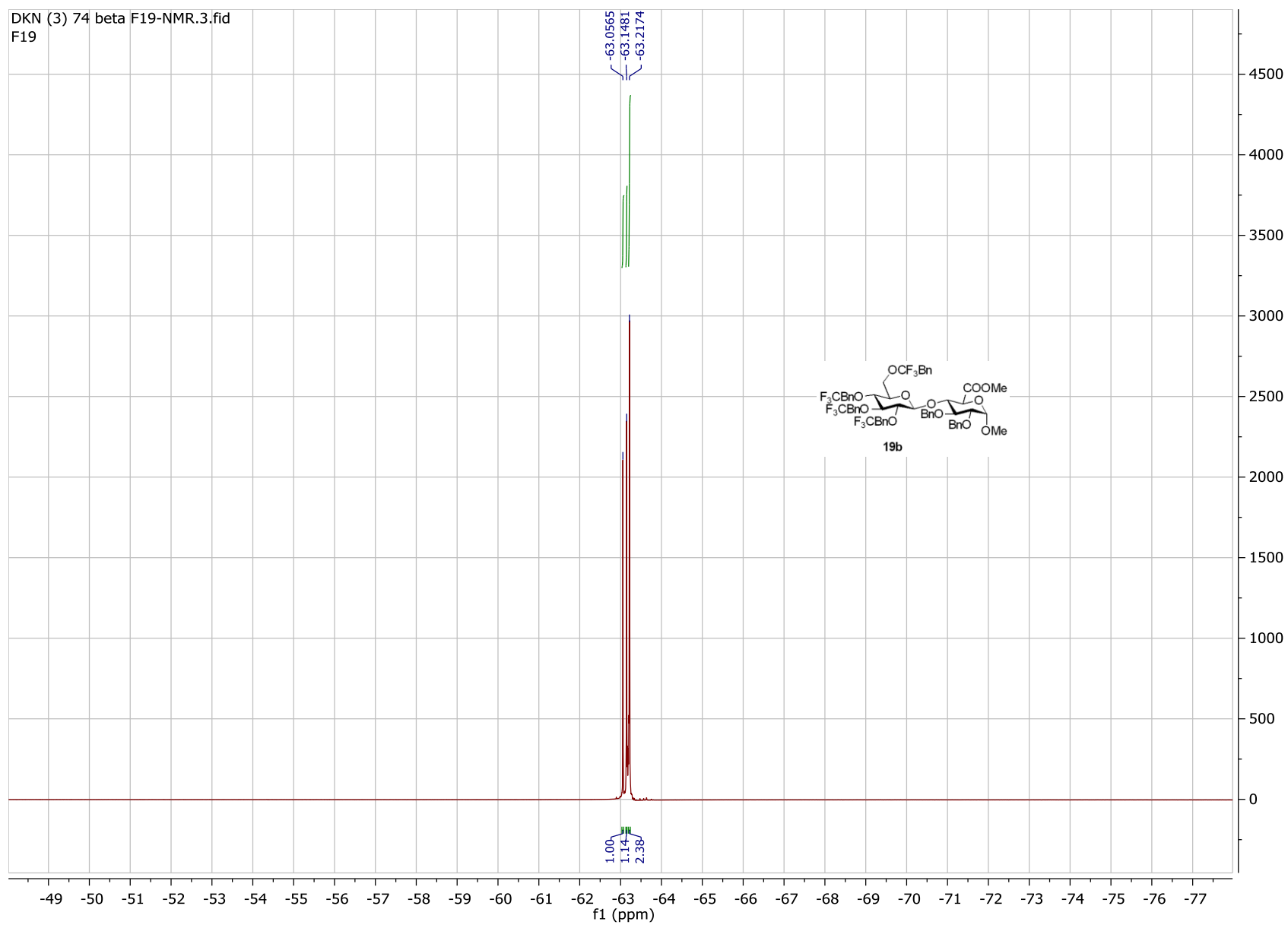
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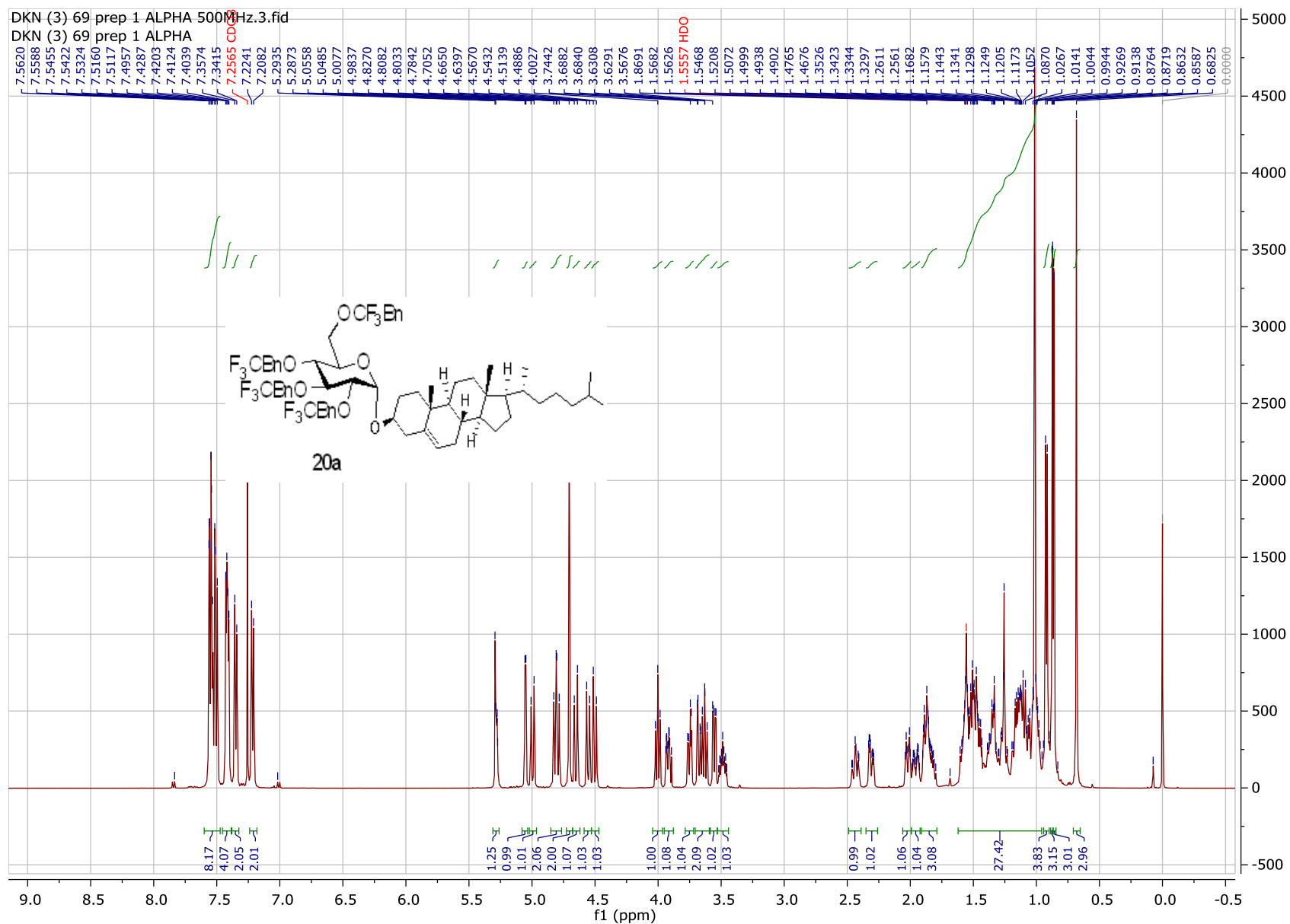




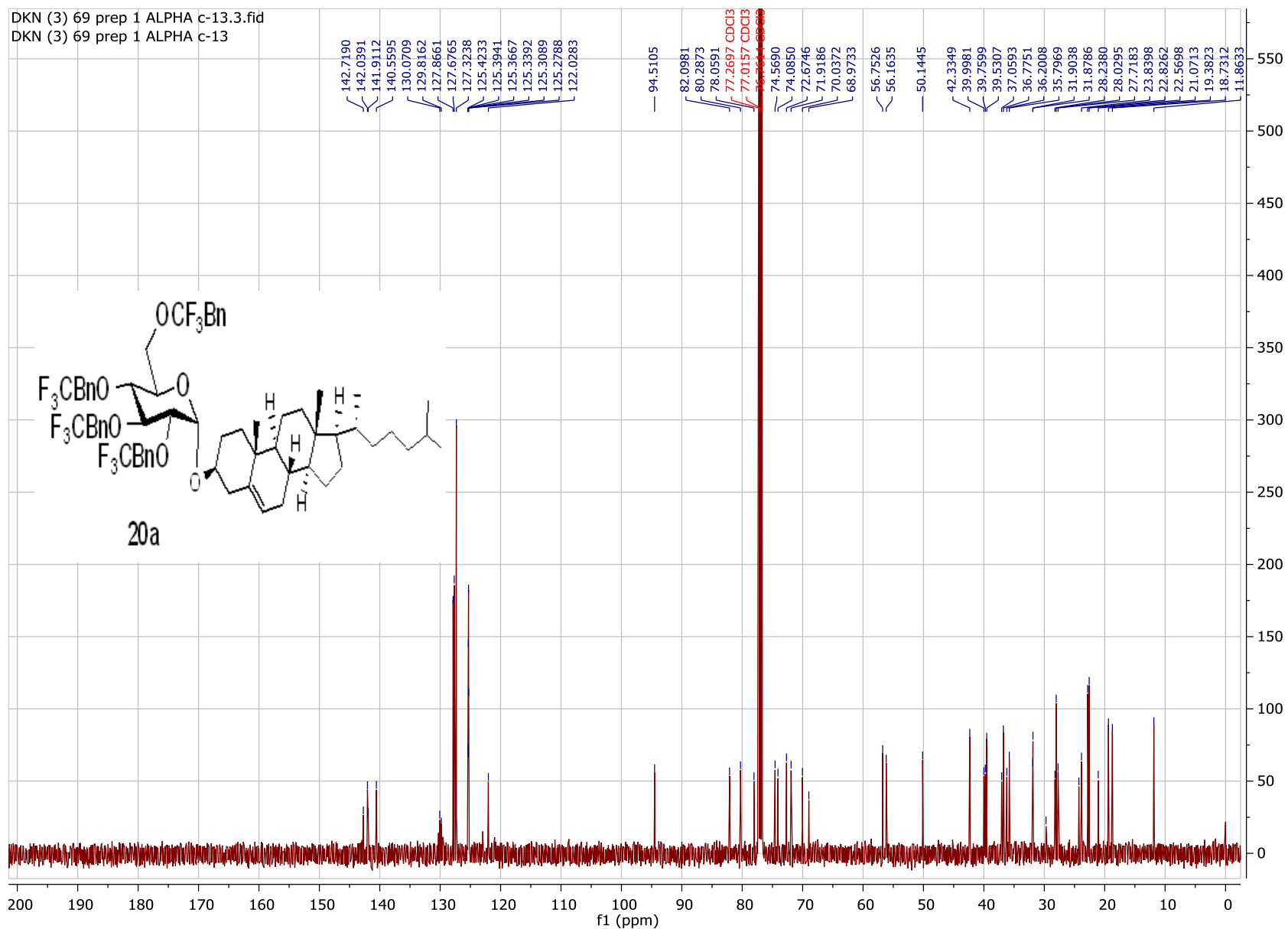


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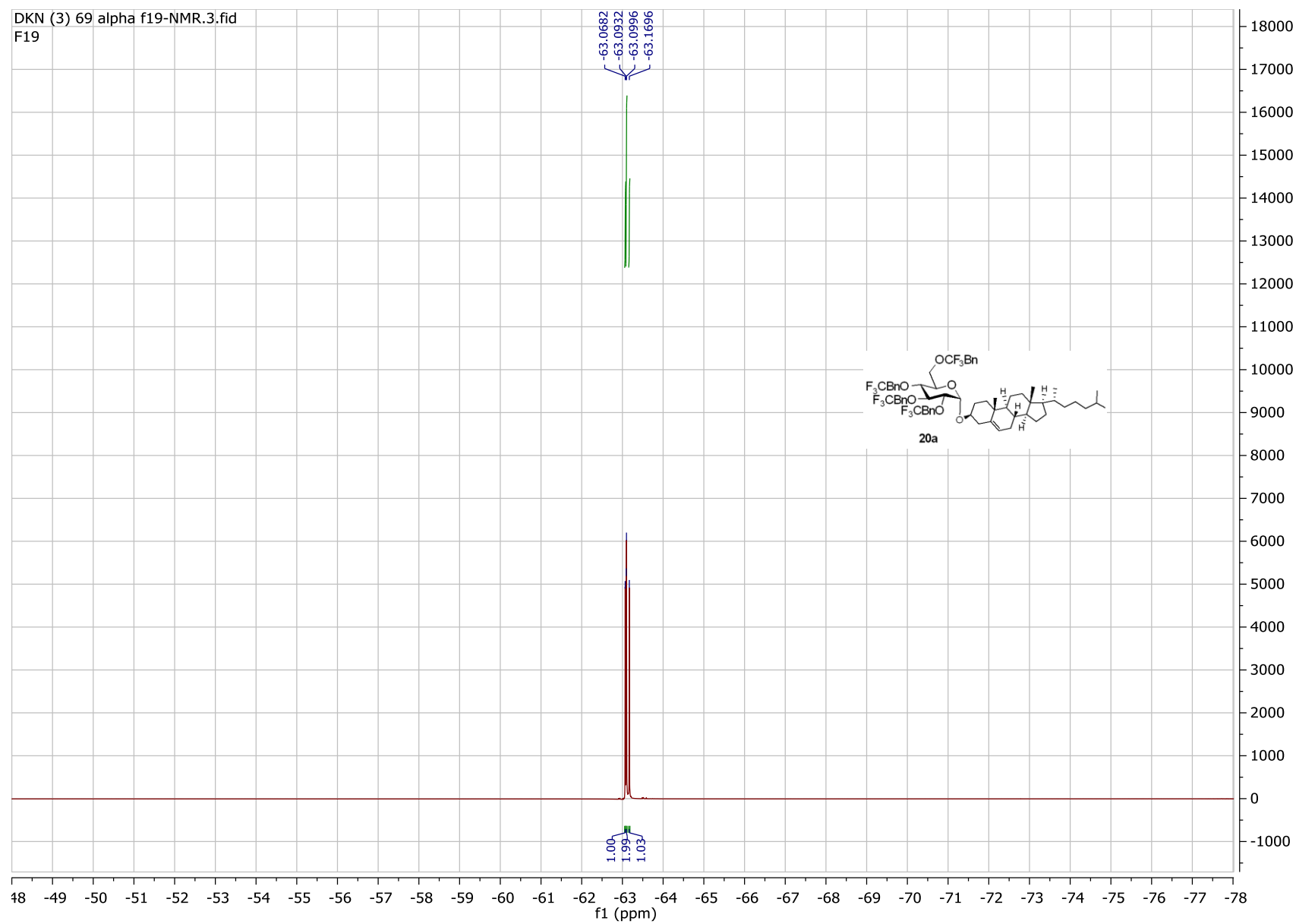


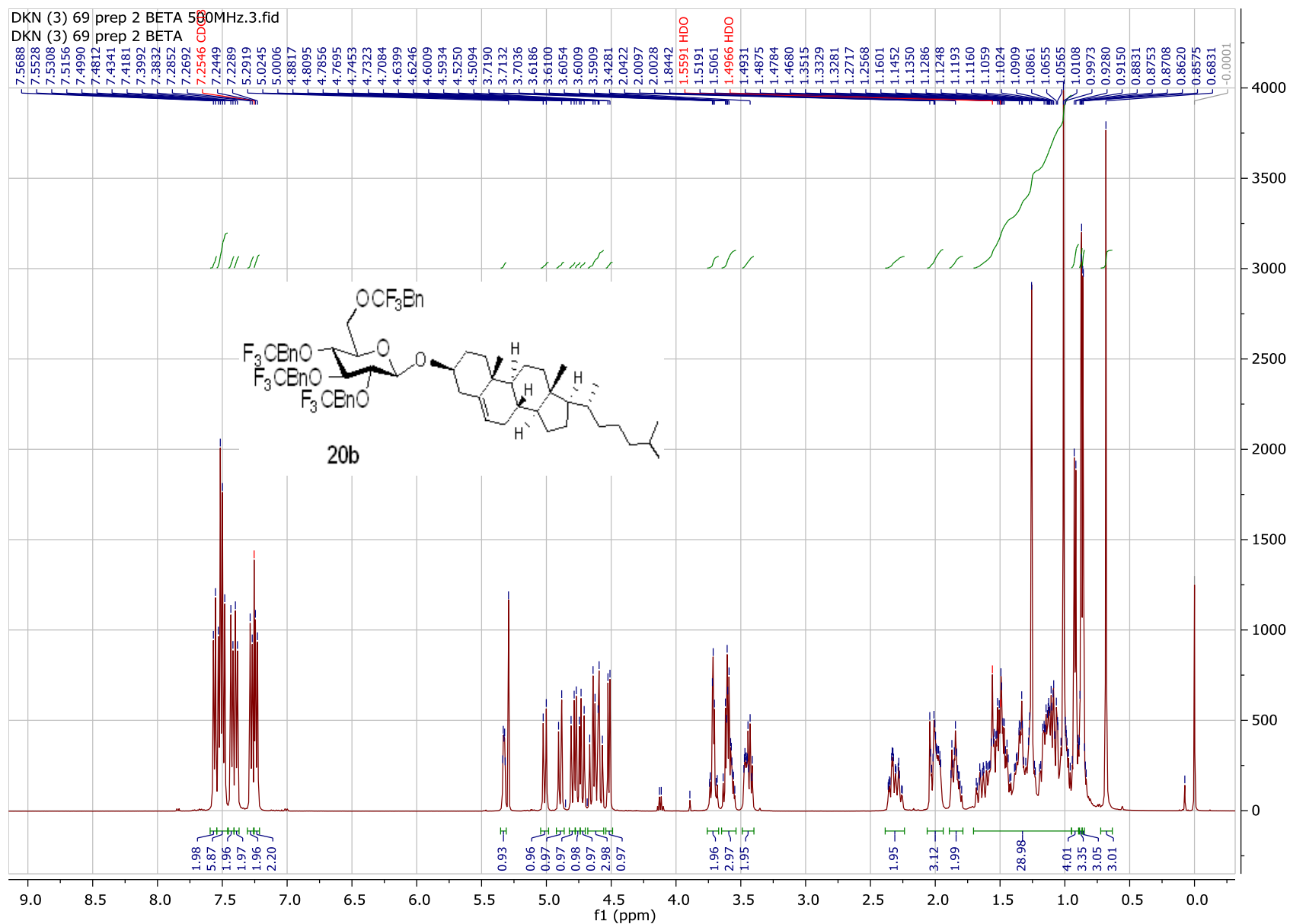


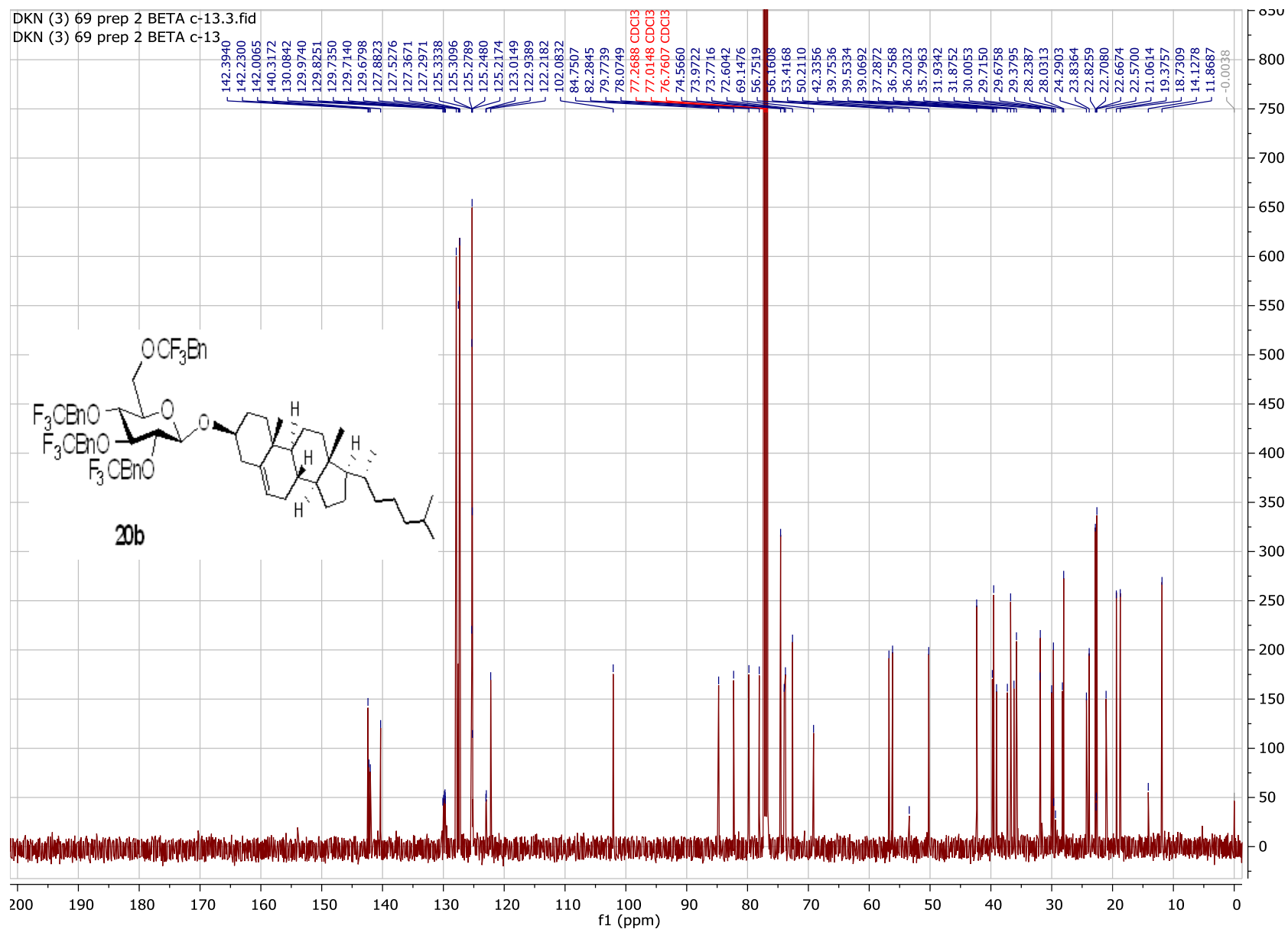
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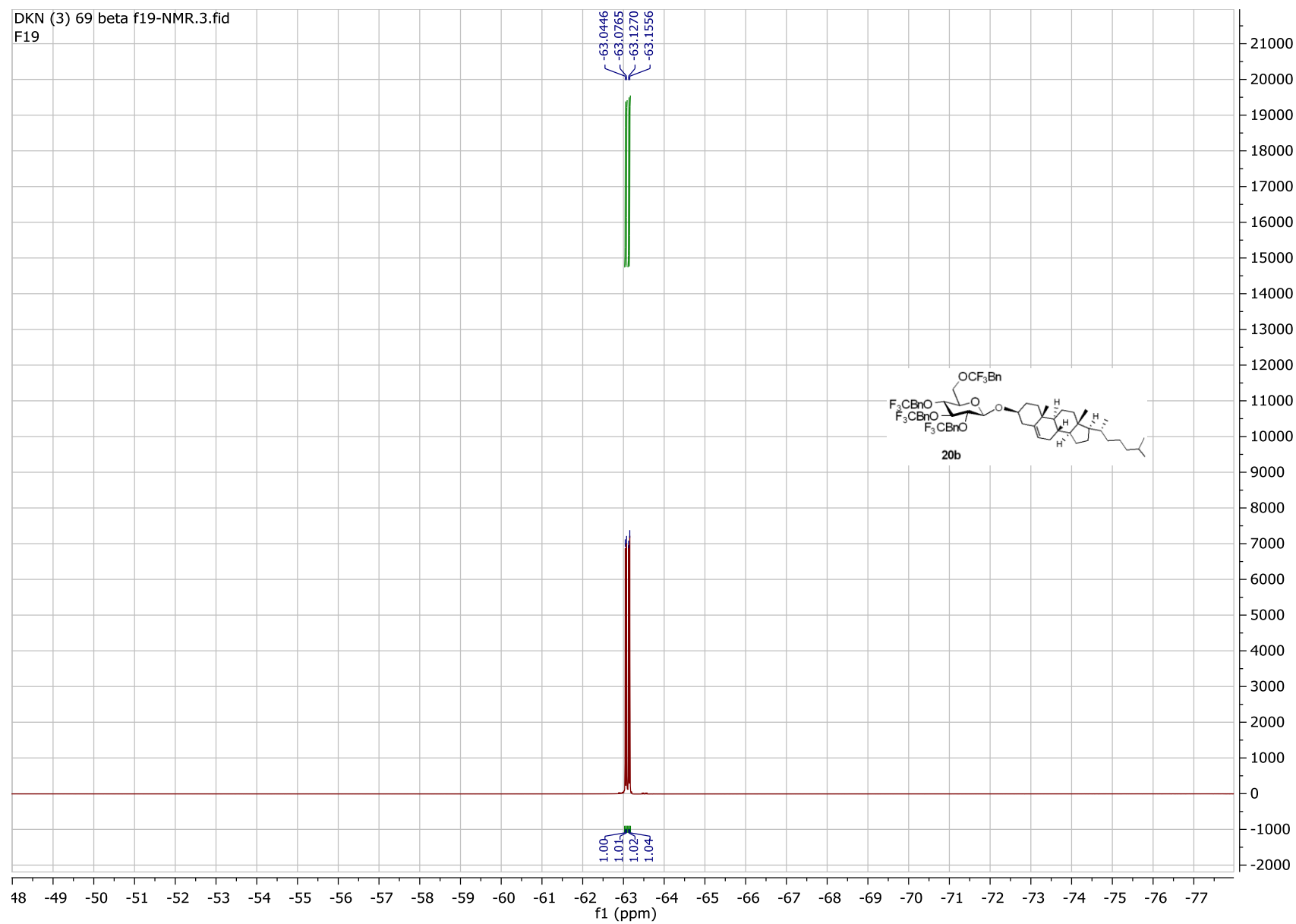
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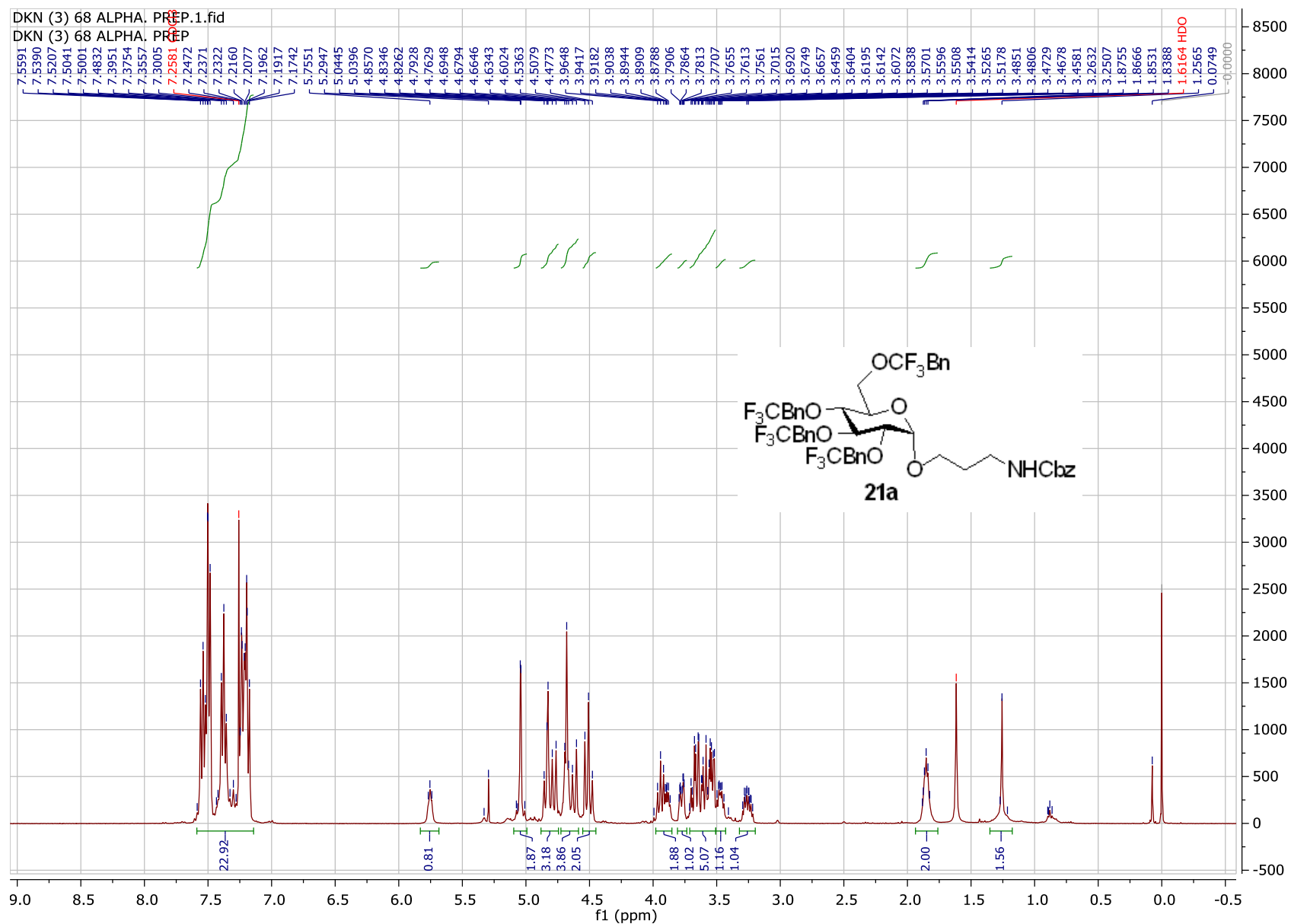




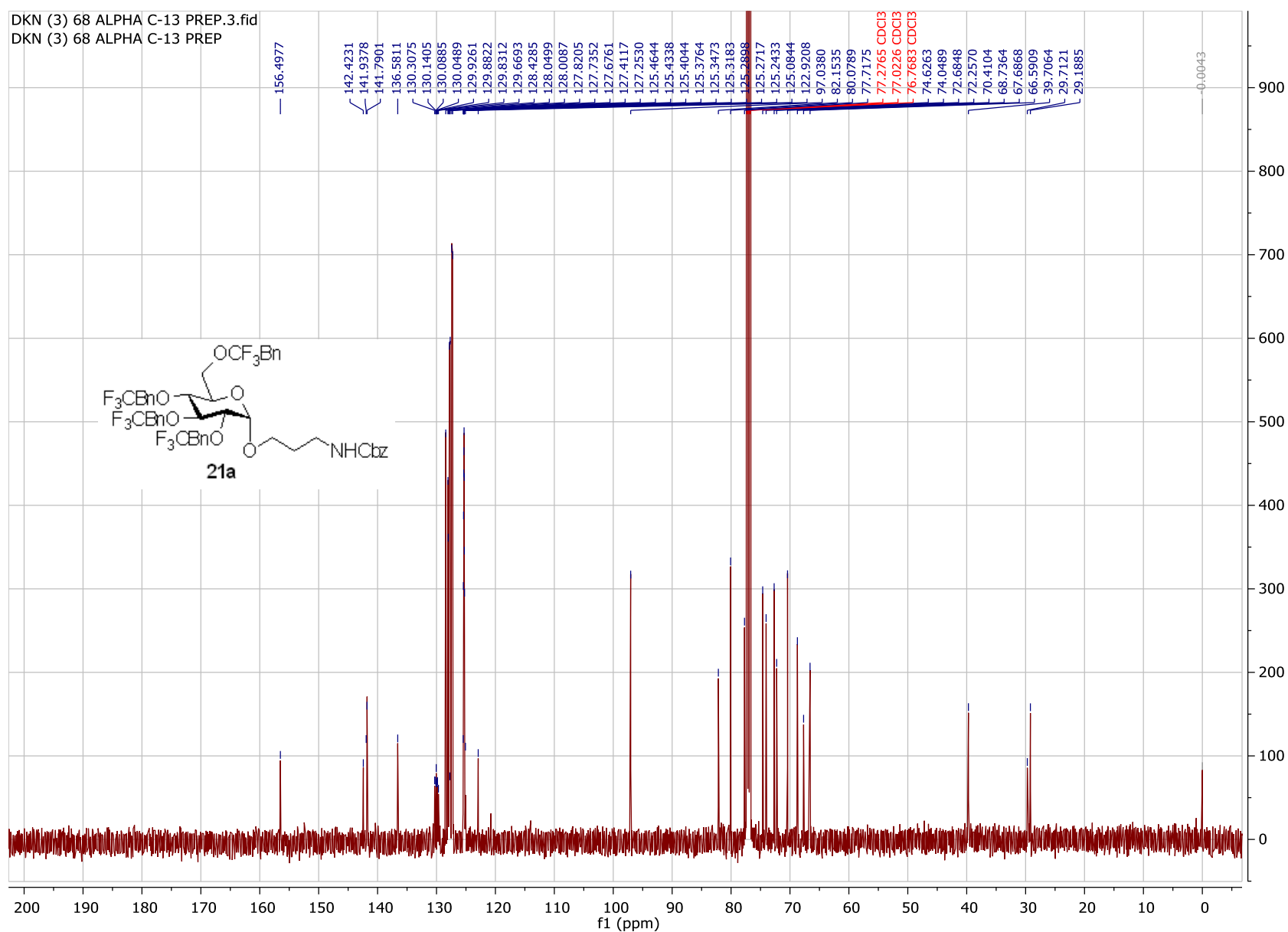


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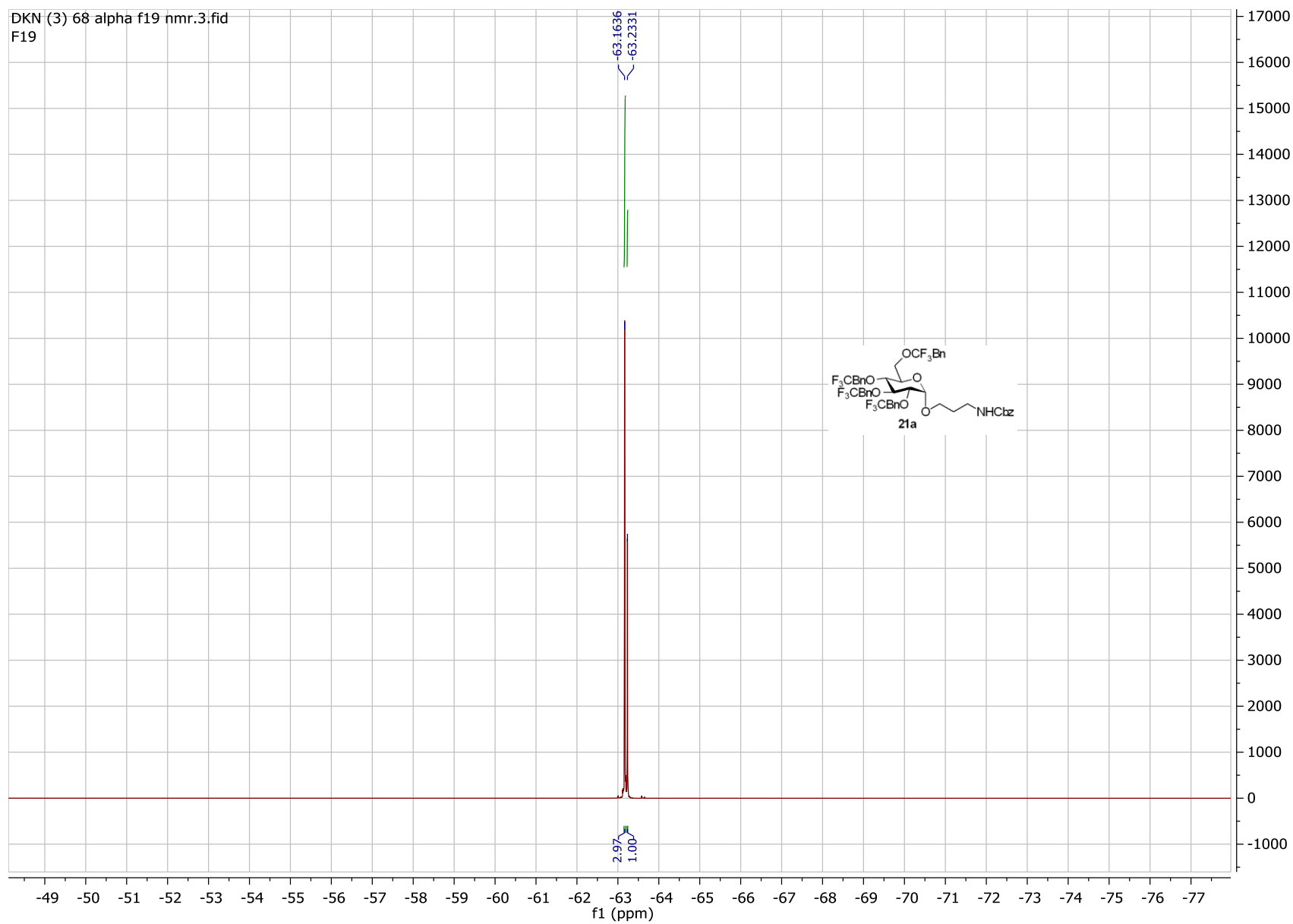


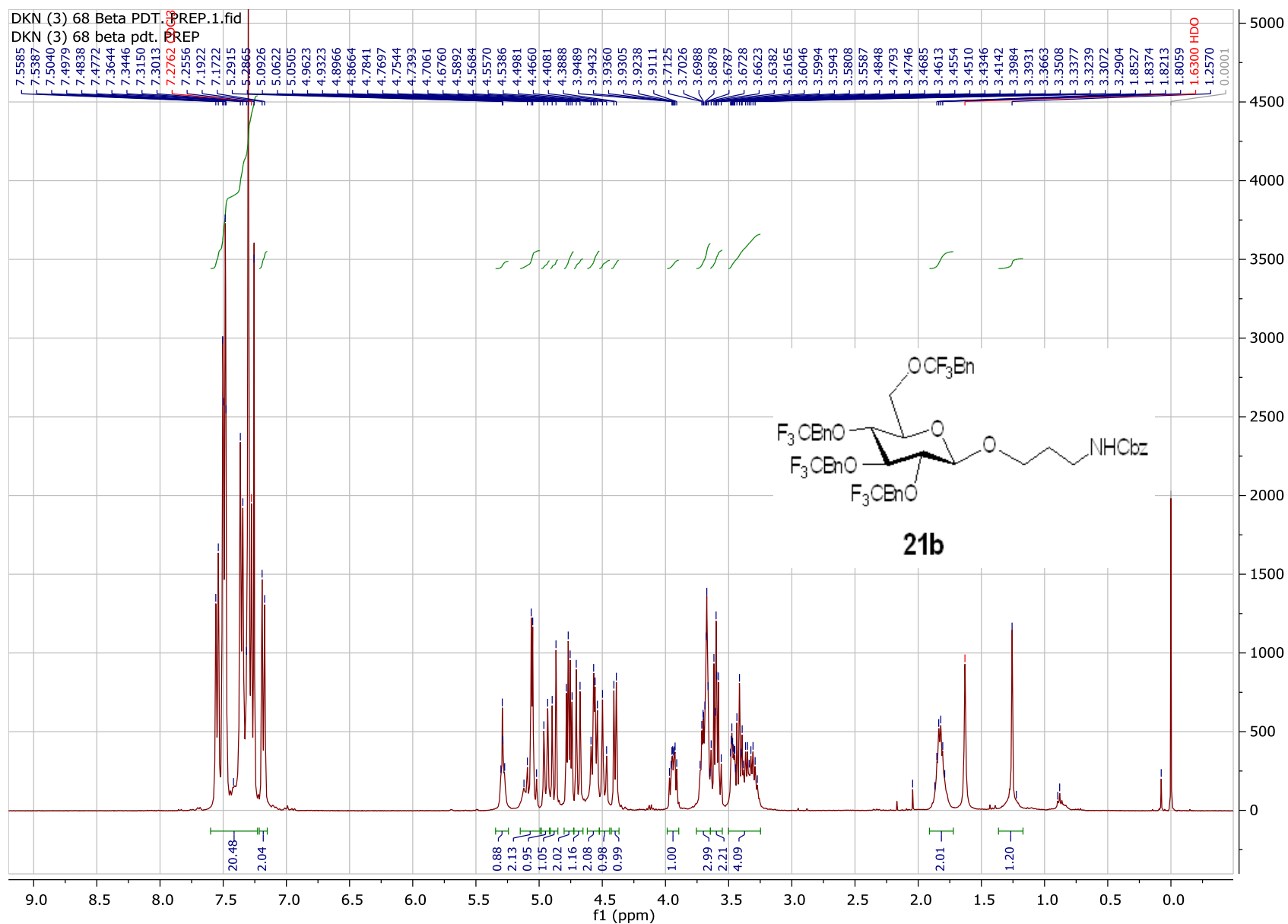


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 DKN (3) 68 ALPHA C-13 PREP

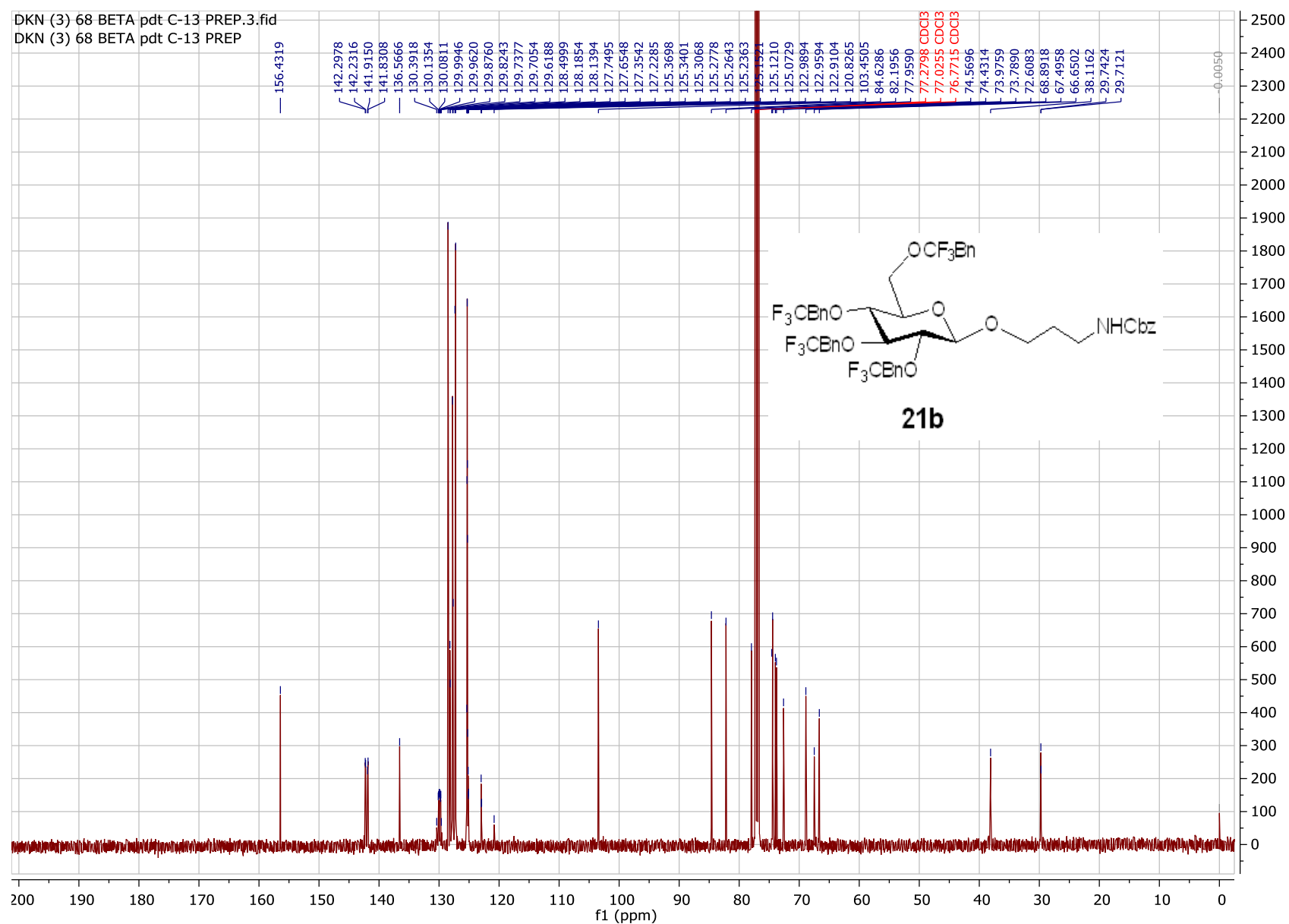


DKN (3) 68 alpha f19 nmr.3.fid
F19

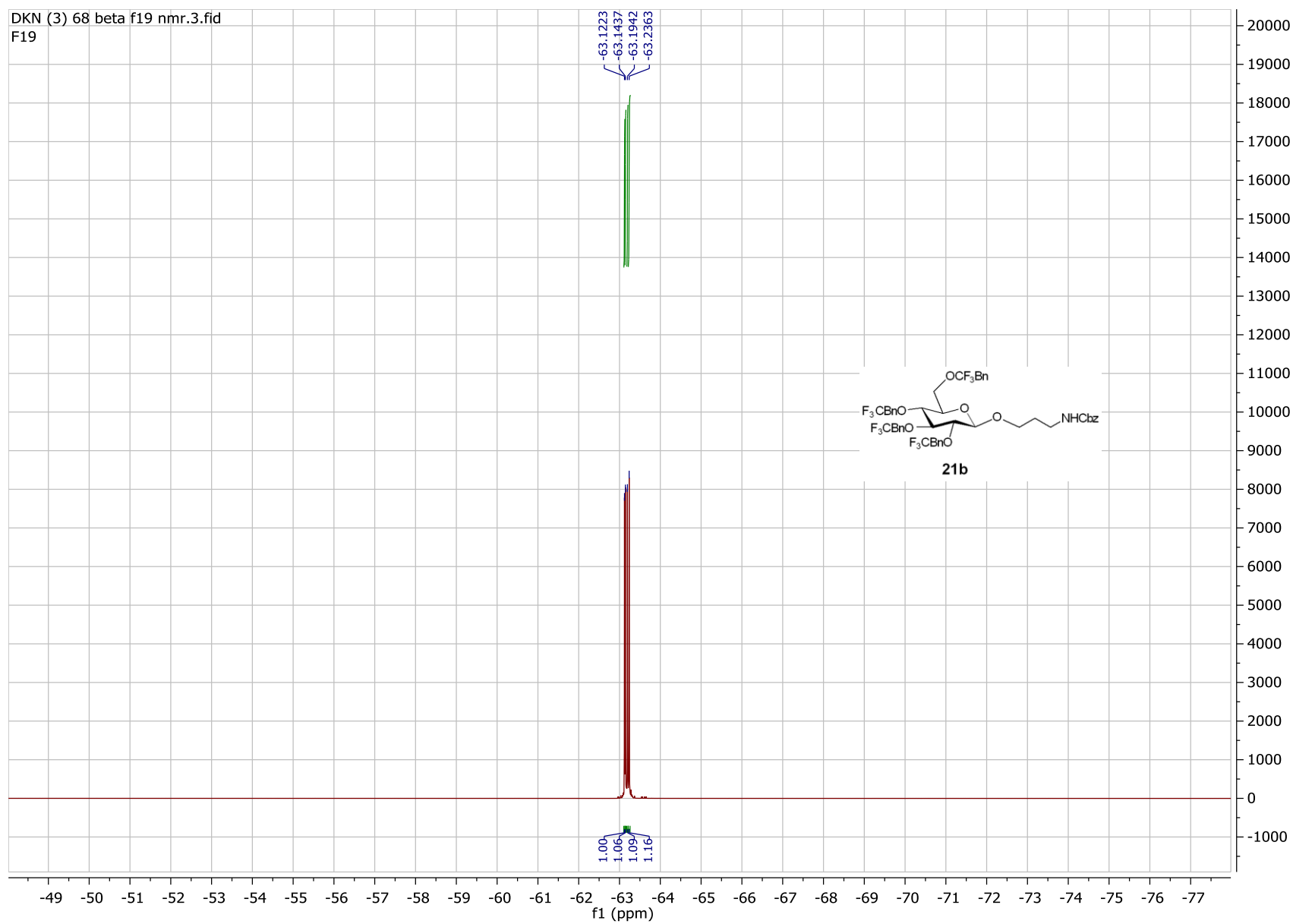




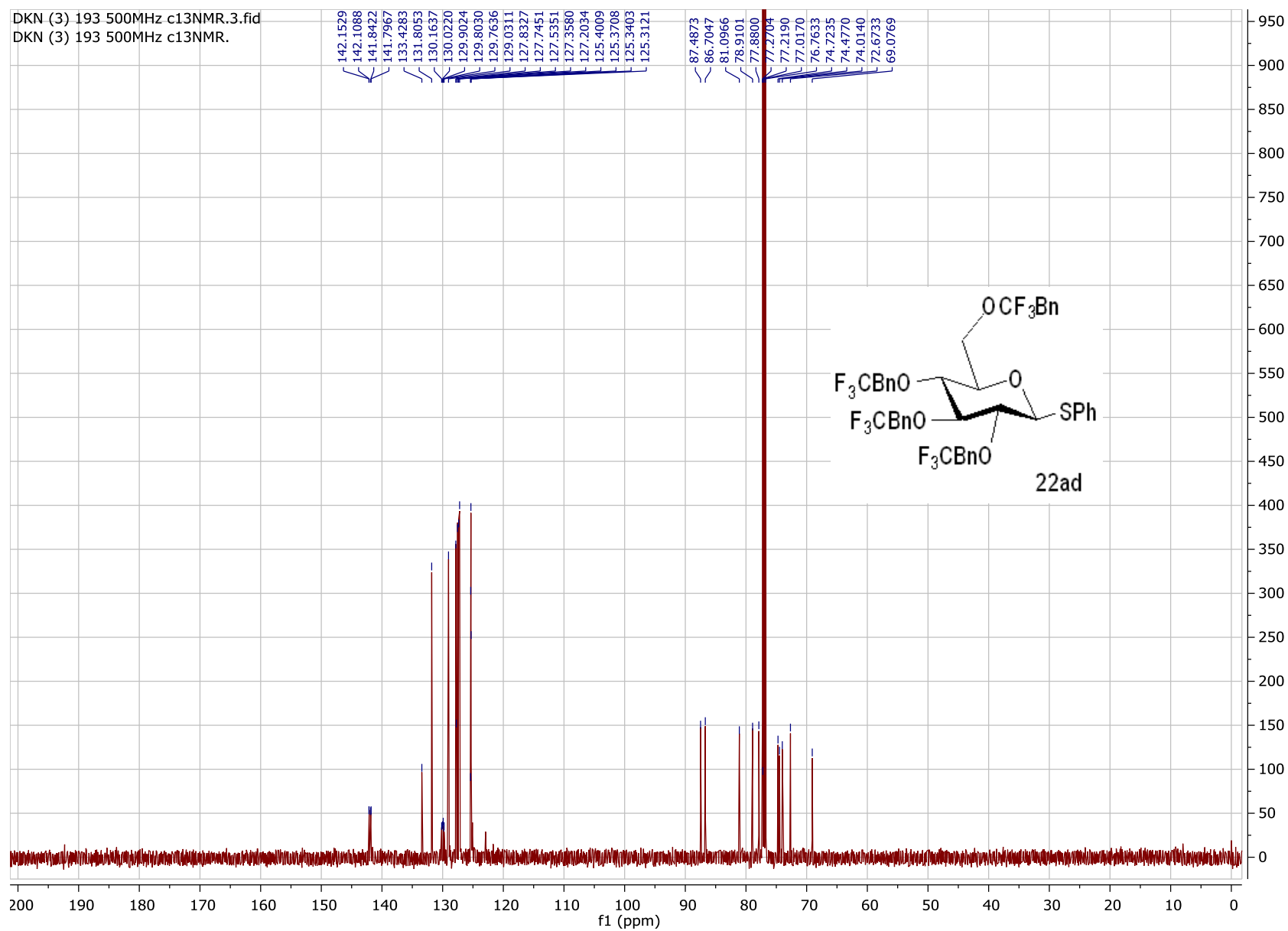
DKN (3) 68 BETA pdt C-13 PREP.3.fid
DKN (3) 68 BETA pdt C-13 PREP



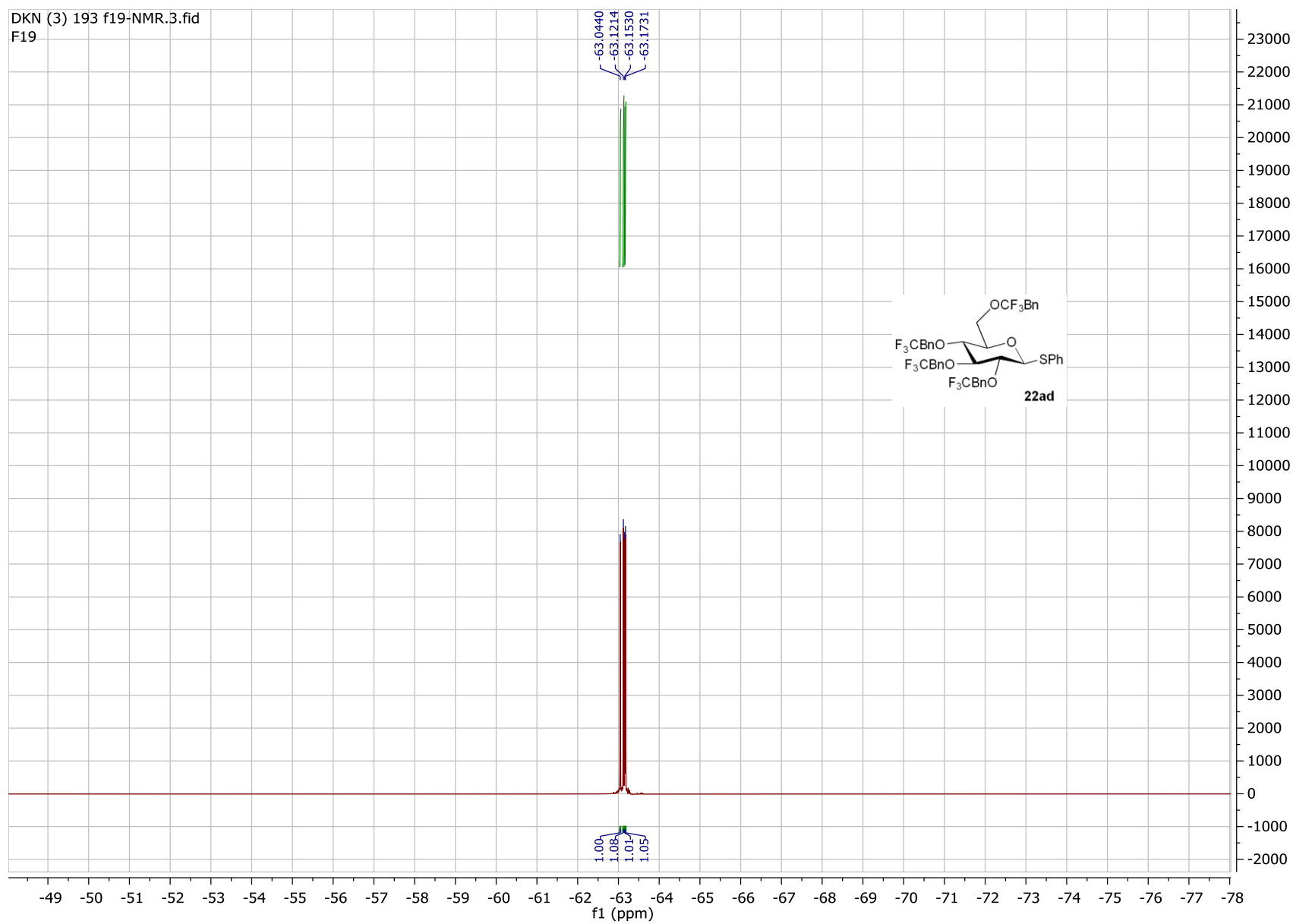
DKN (3) 68 beta f19 nmr.3.fid
F19

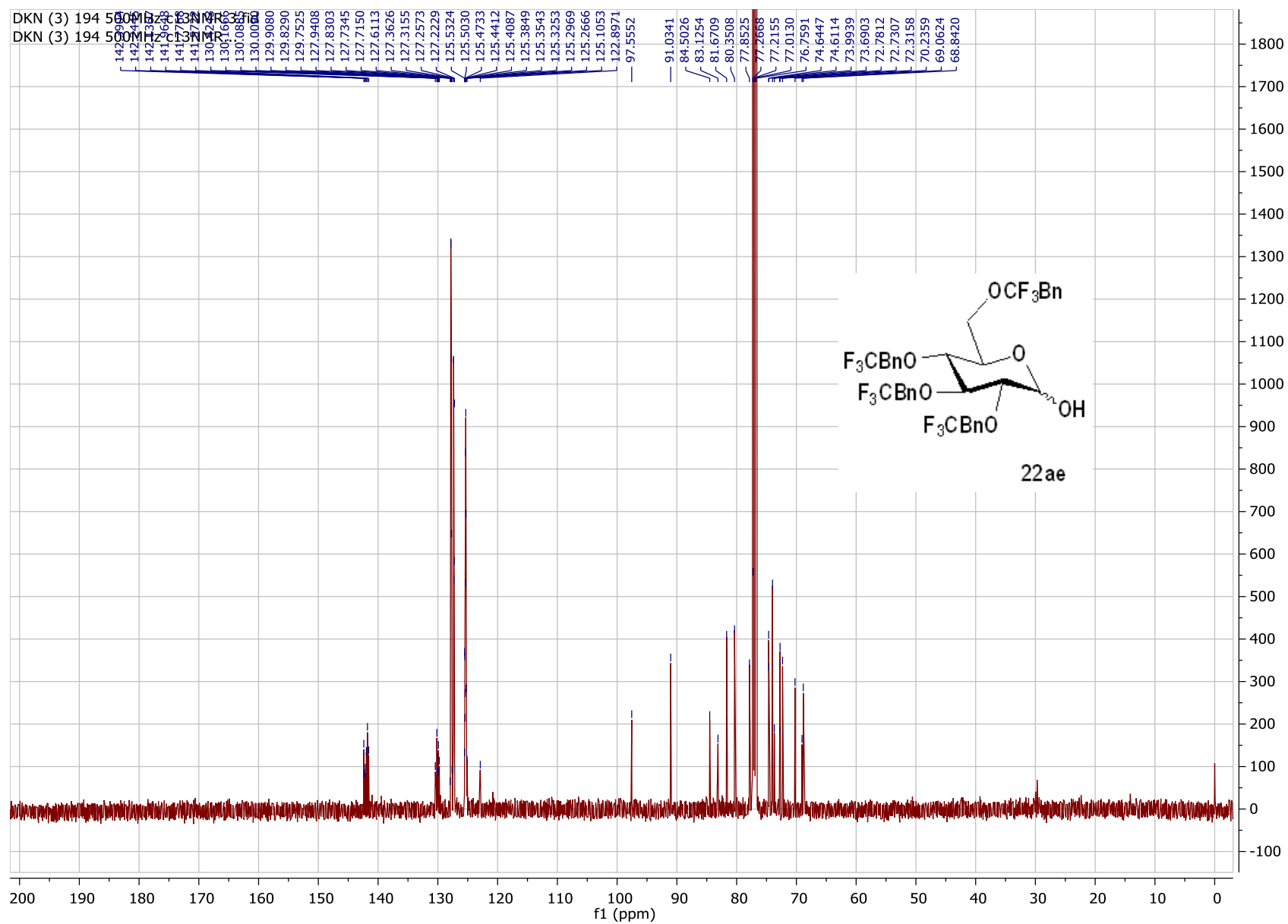


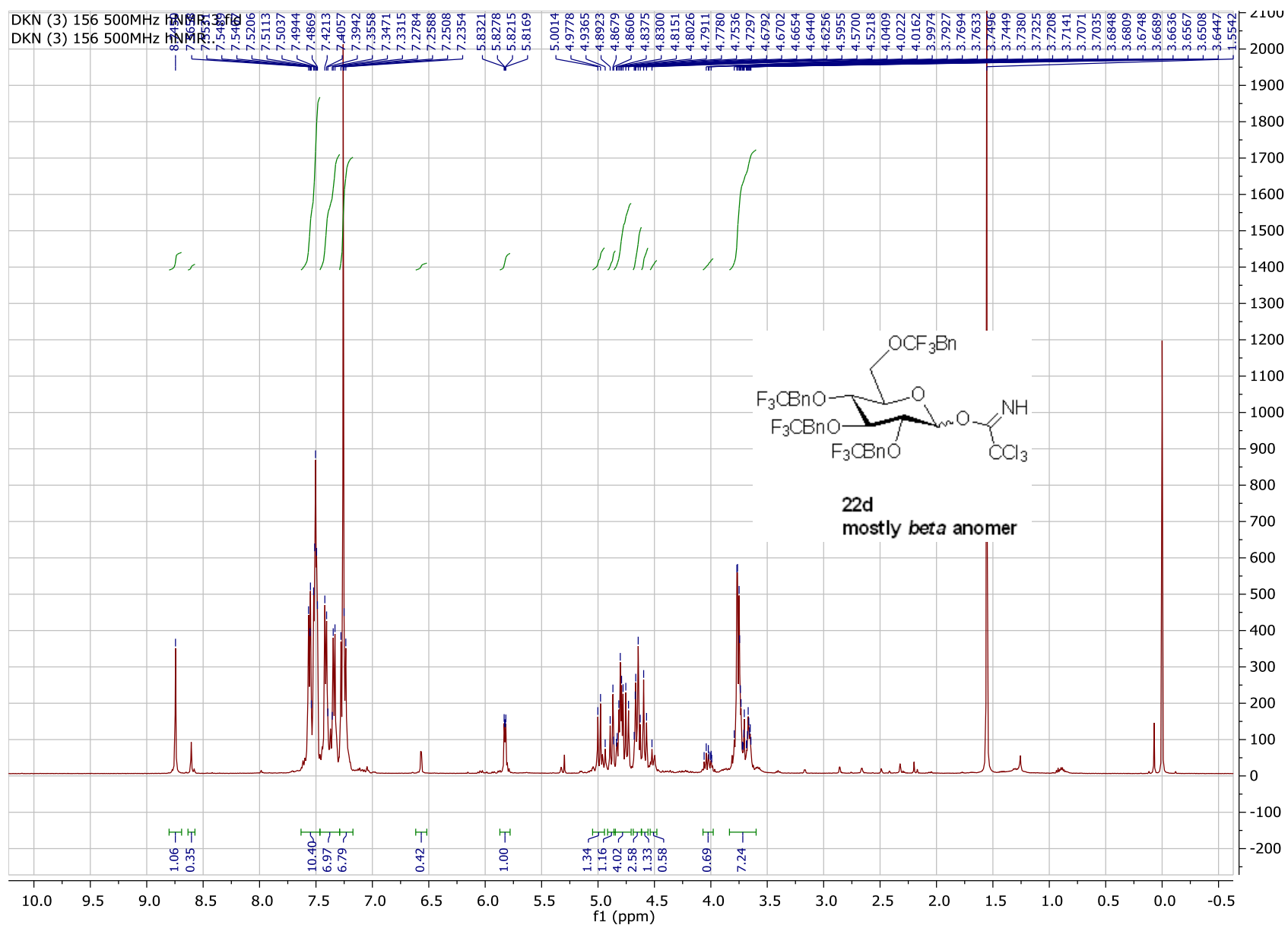
DKN (3) 193 500MHz c13NMR.3.fid
DKN (3) 193 500MHz c13NMR.

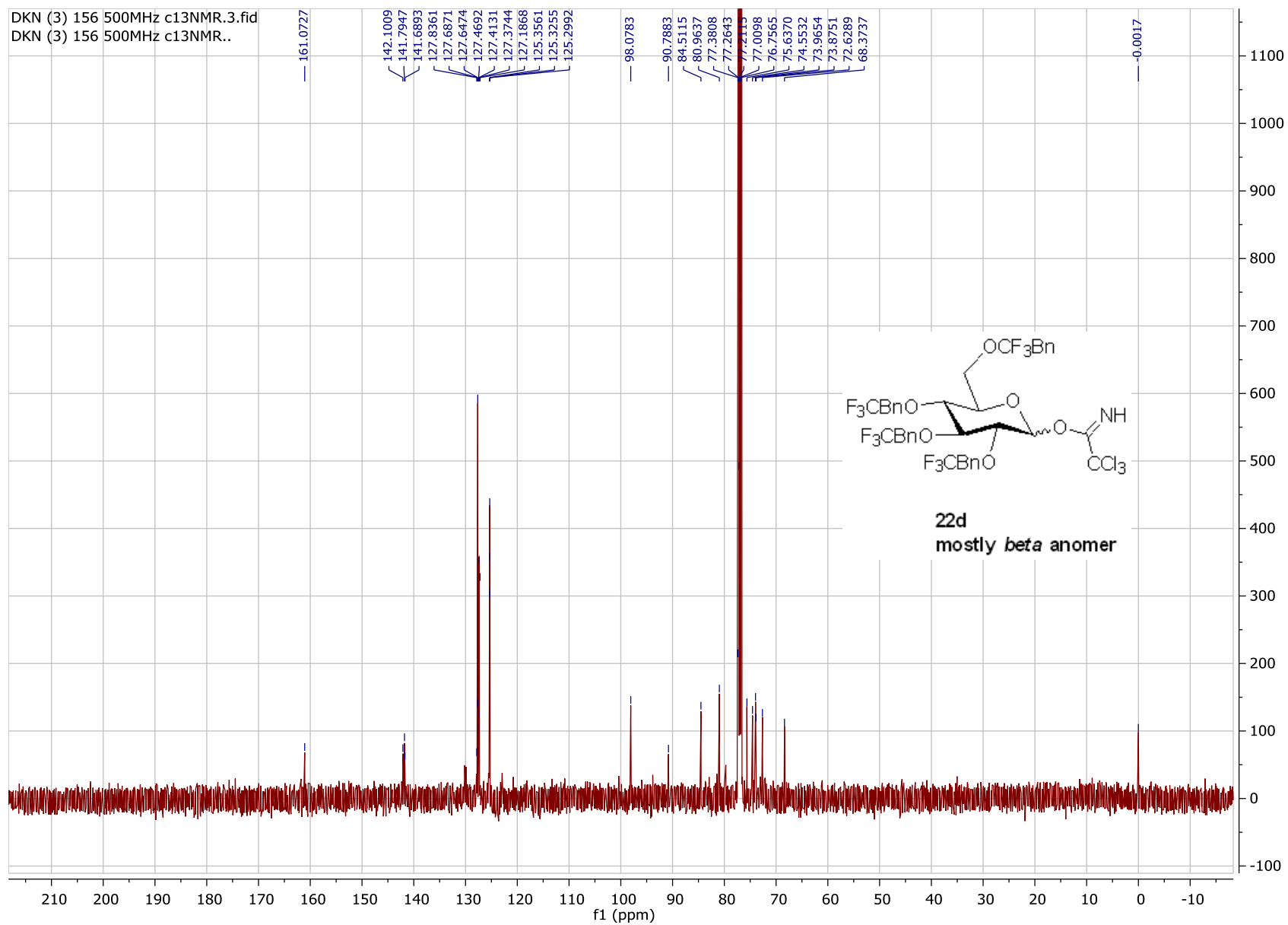


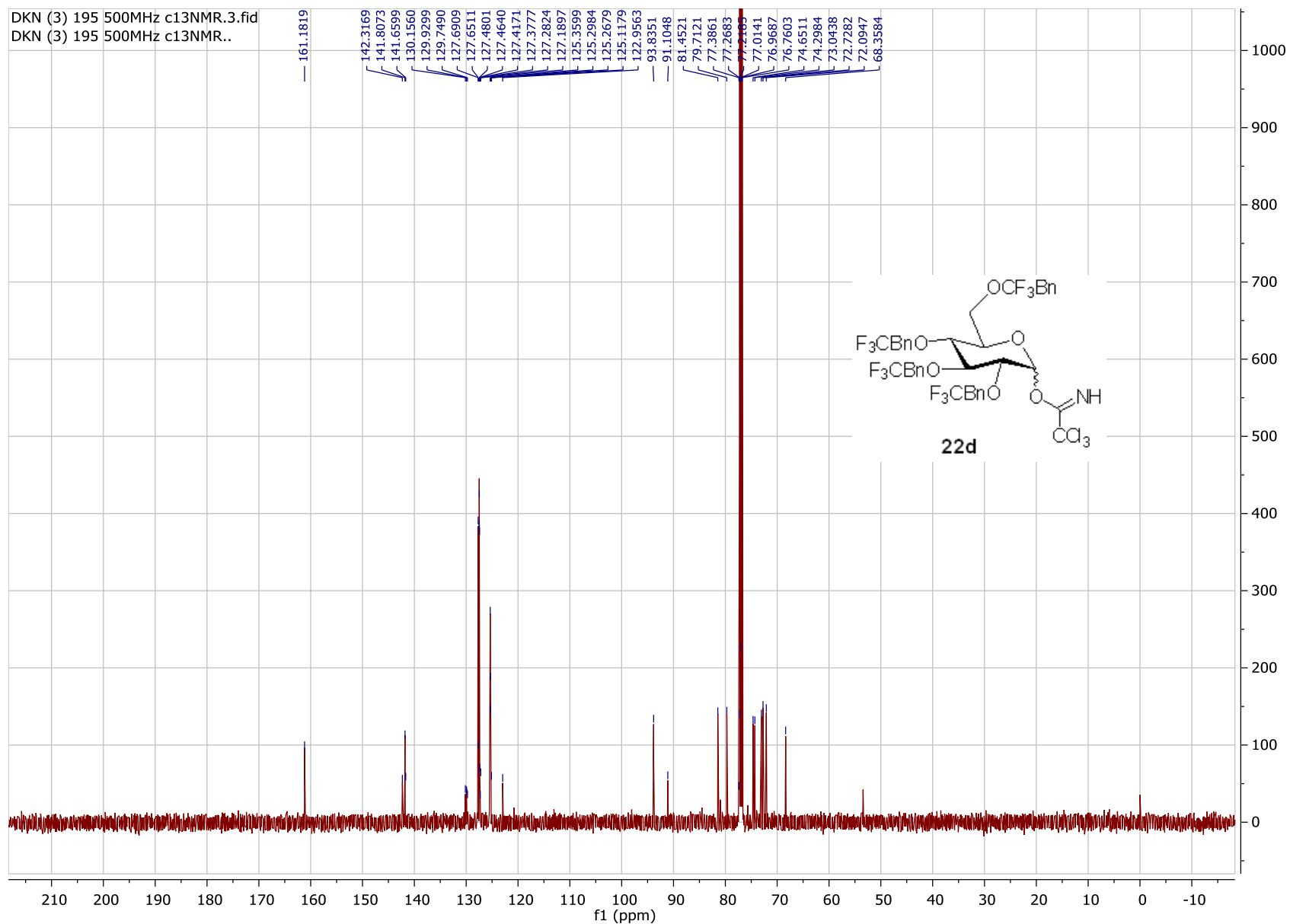
DKN (3) 193 f19-NMR.3.fid
F19



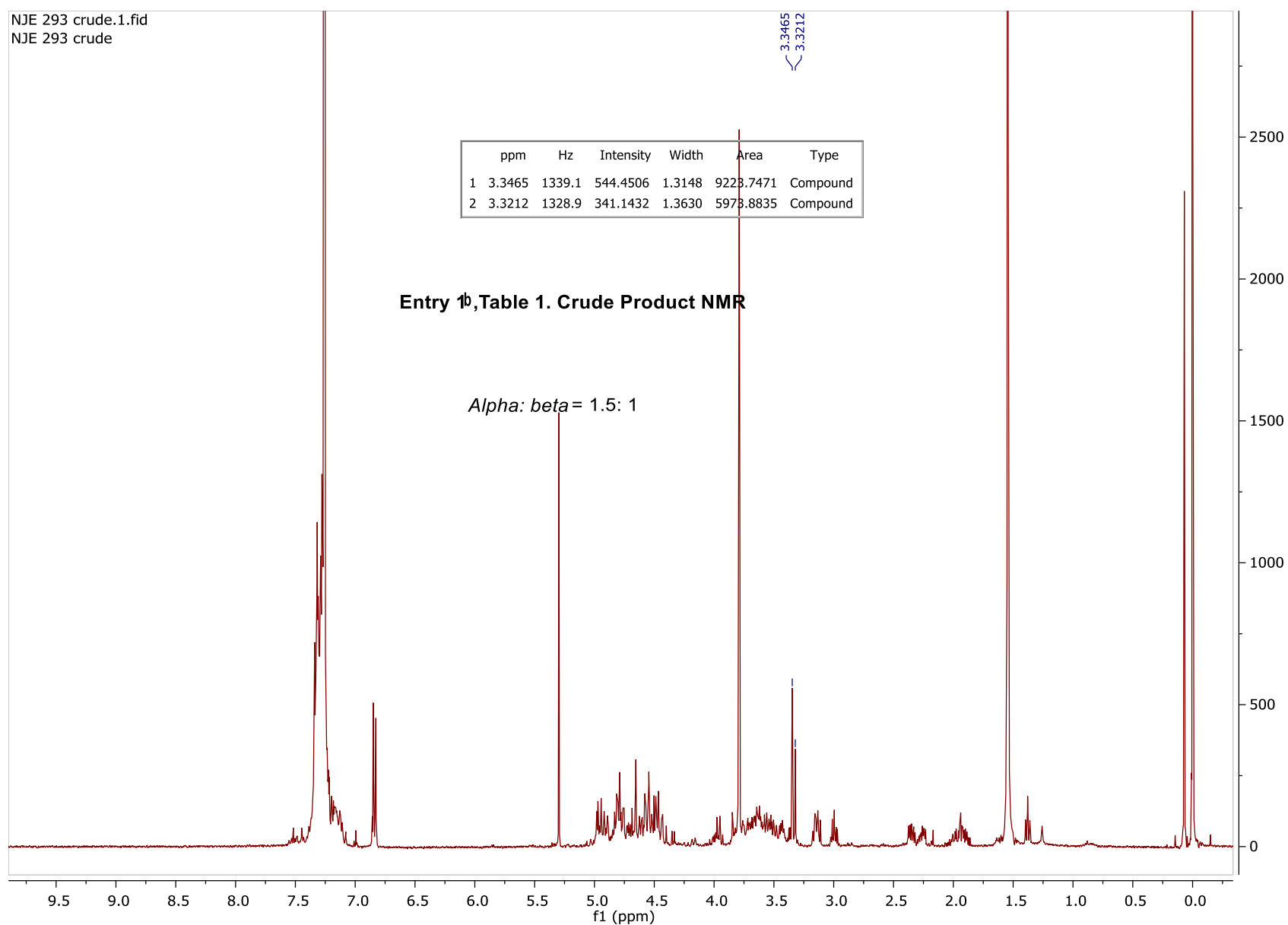


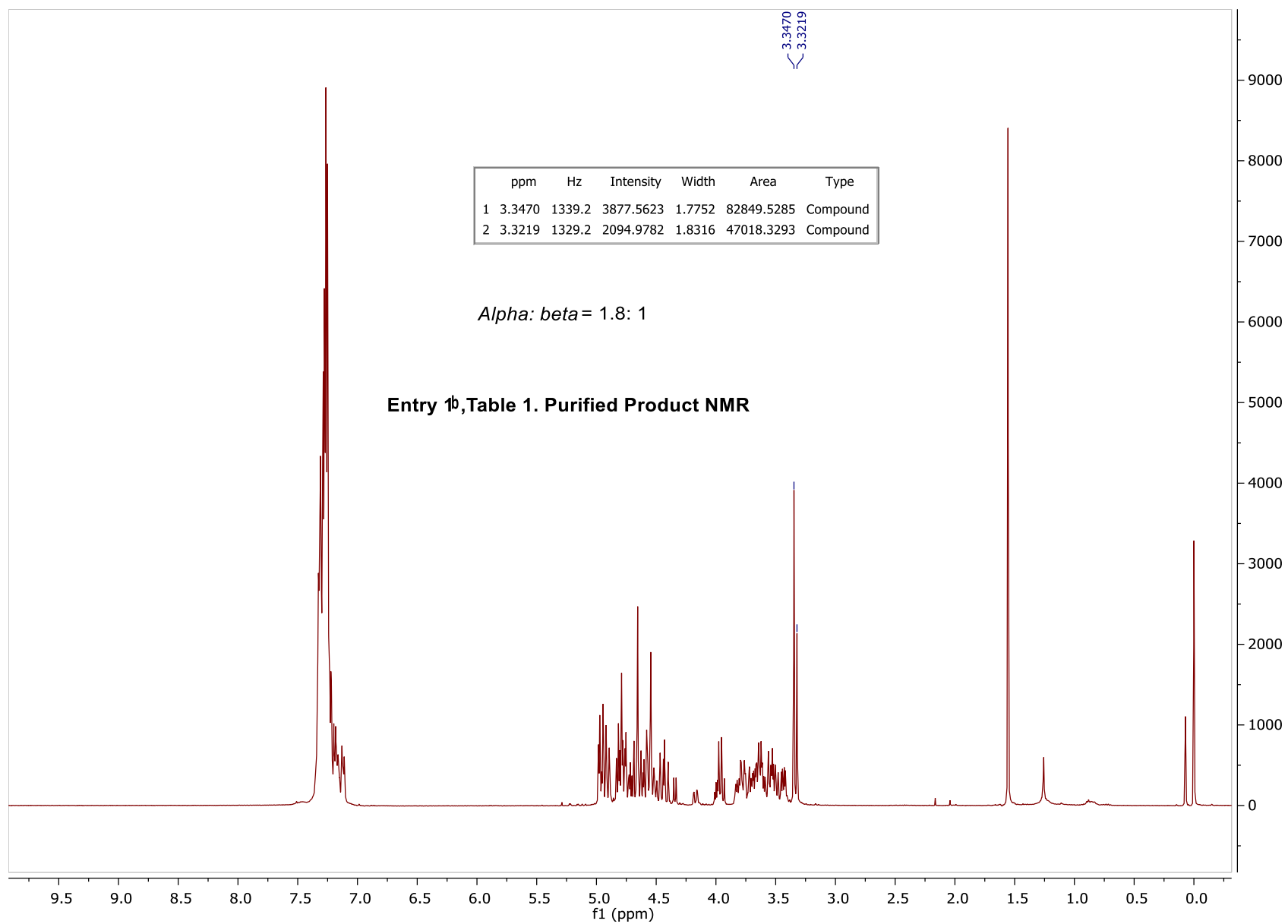




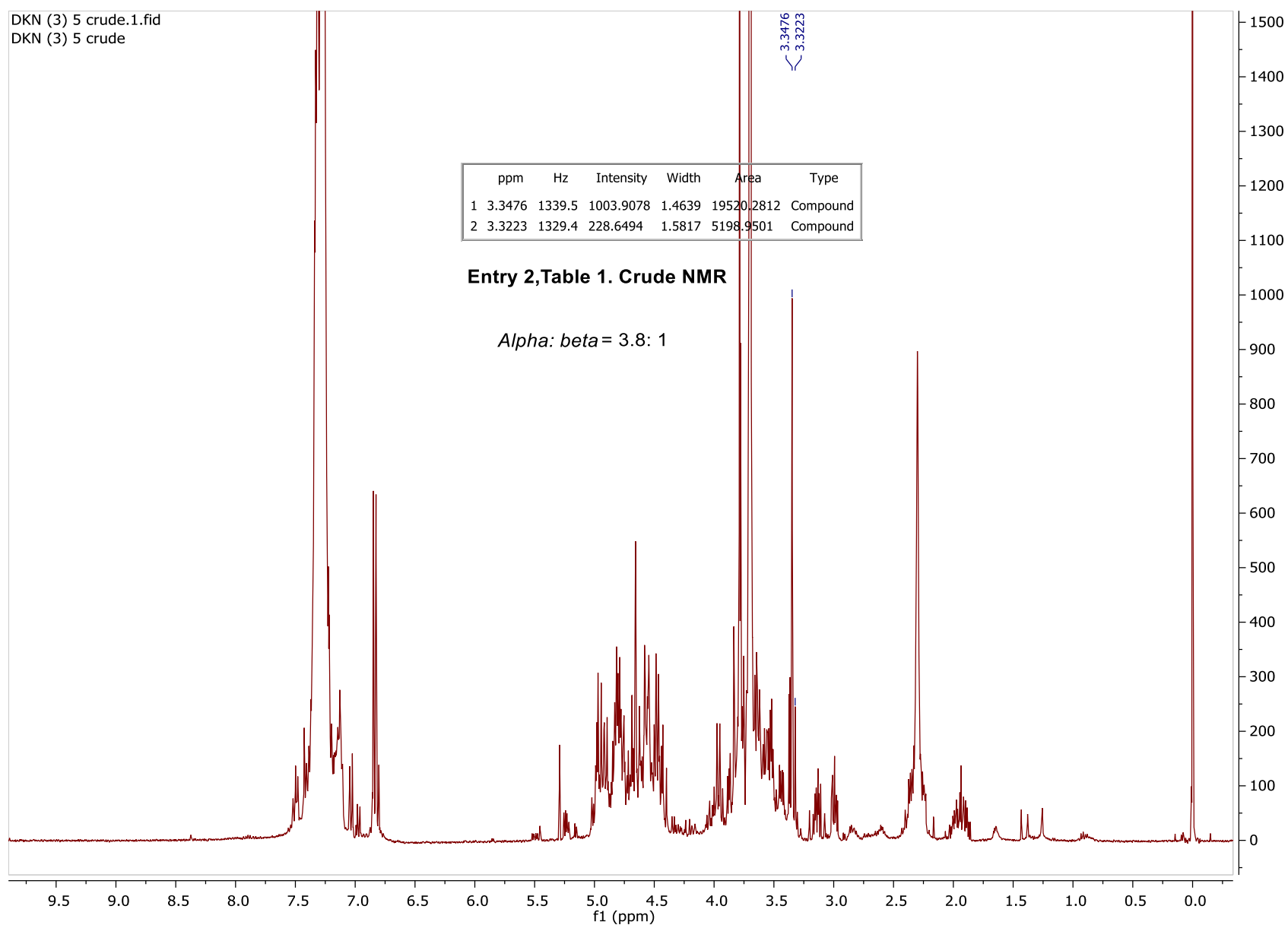


NJE 293 crude.1.fid
NJE 293 crude

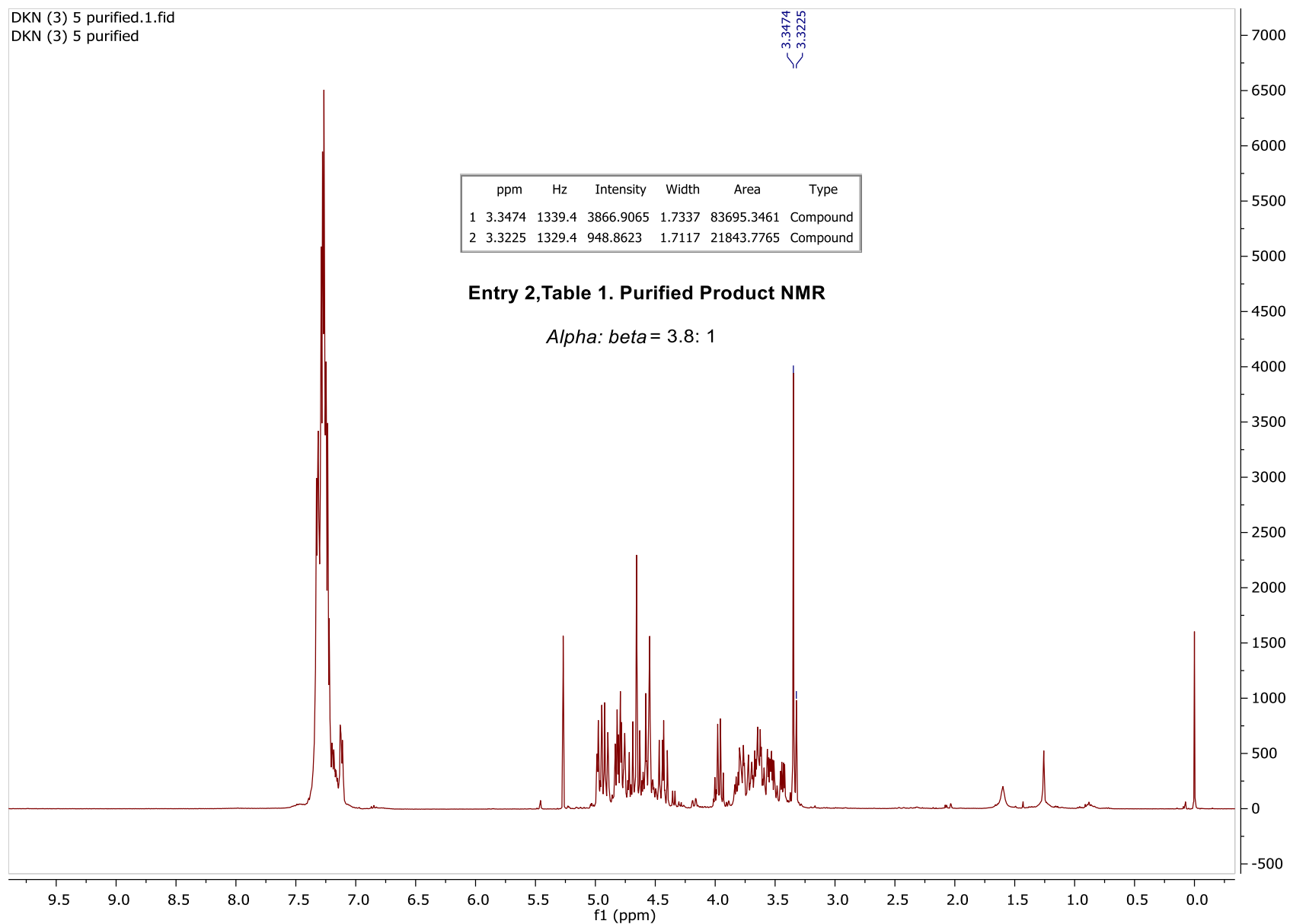




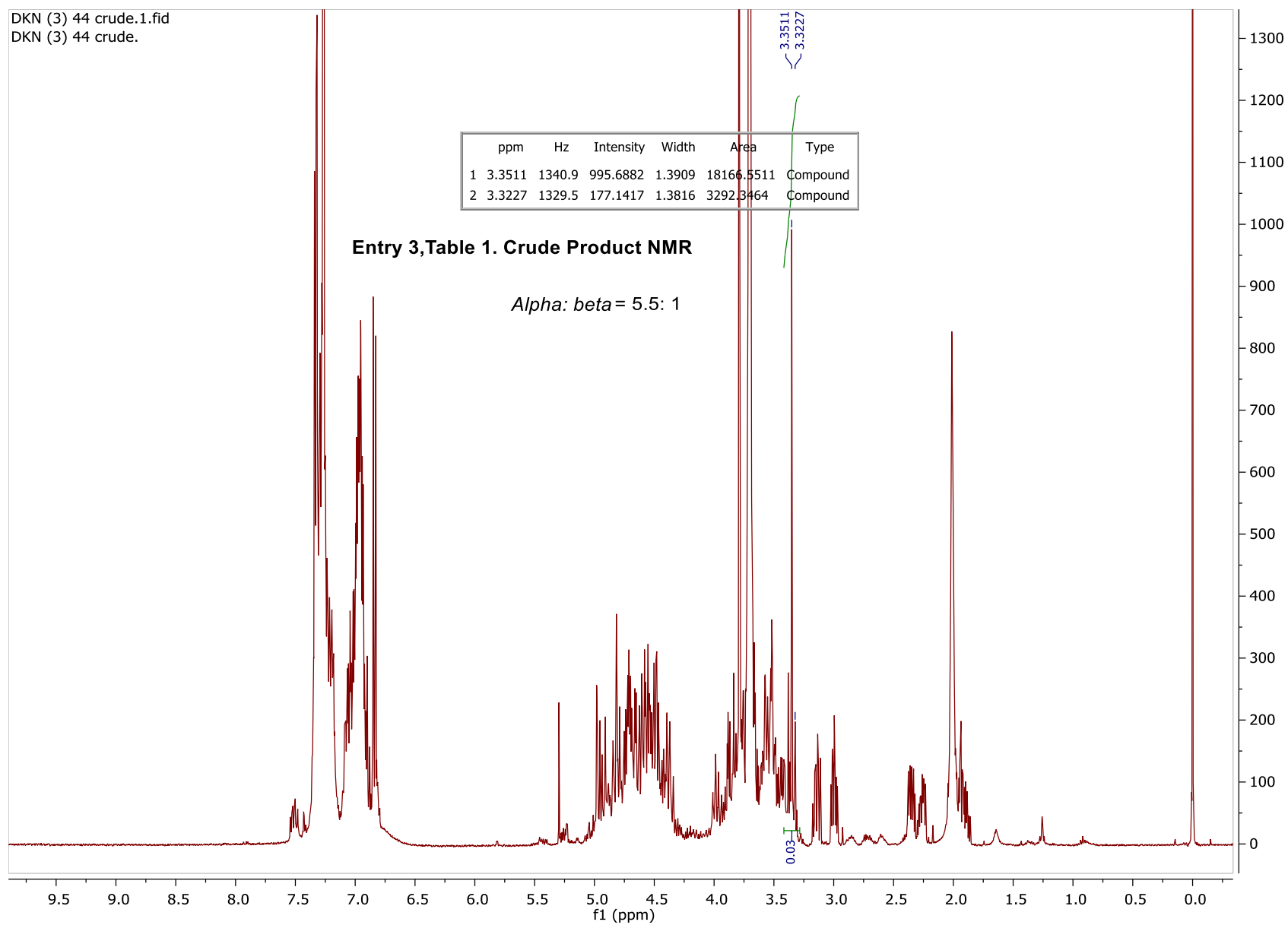
DKN (3) 5 crude.1.fid
DKN (3) 5 crude



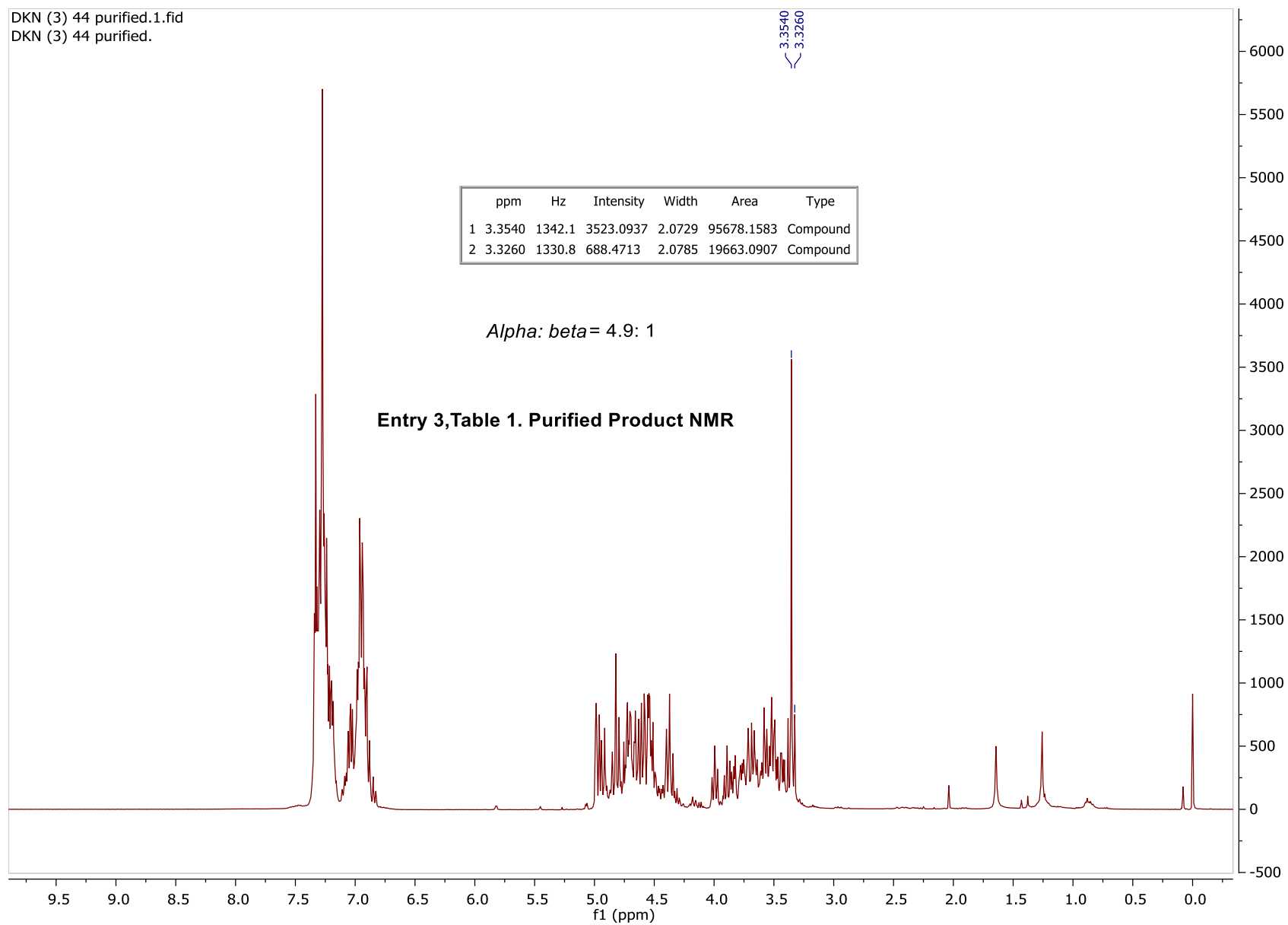
DKN (3) 5 purified.1.fid
DKN (3) 5 purified



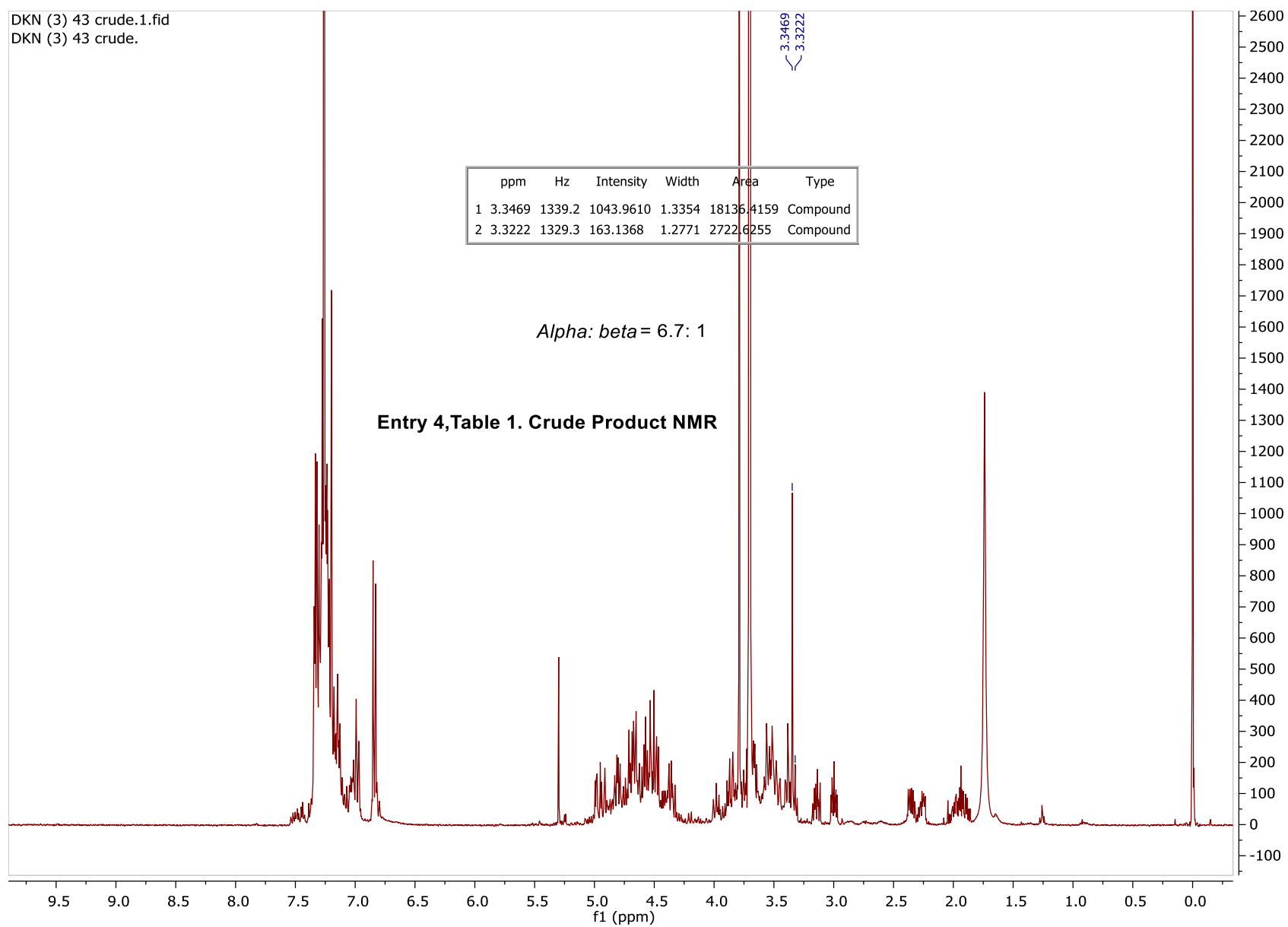
DKN (3) 44 crude.1.fid
DKN (3) 44 crude.



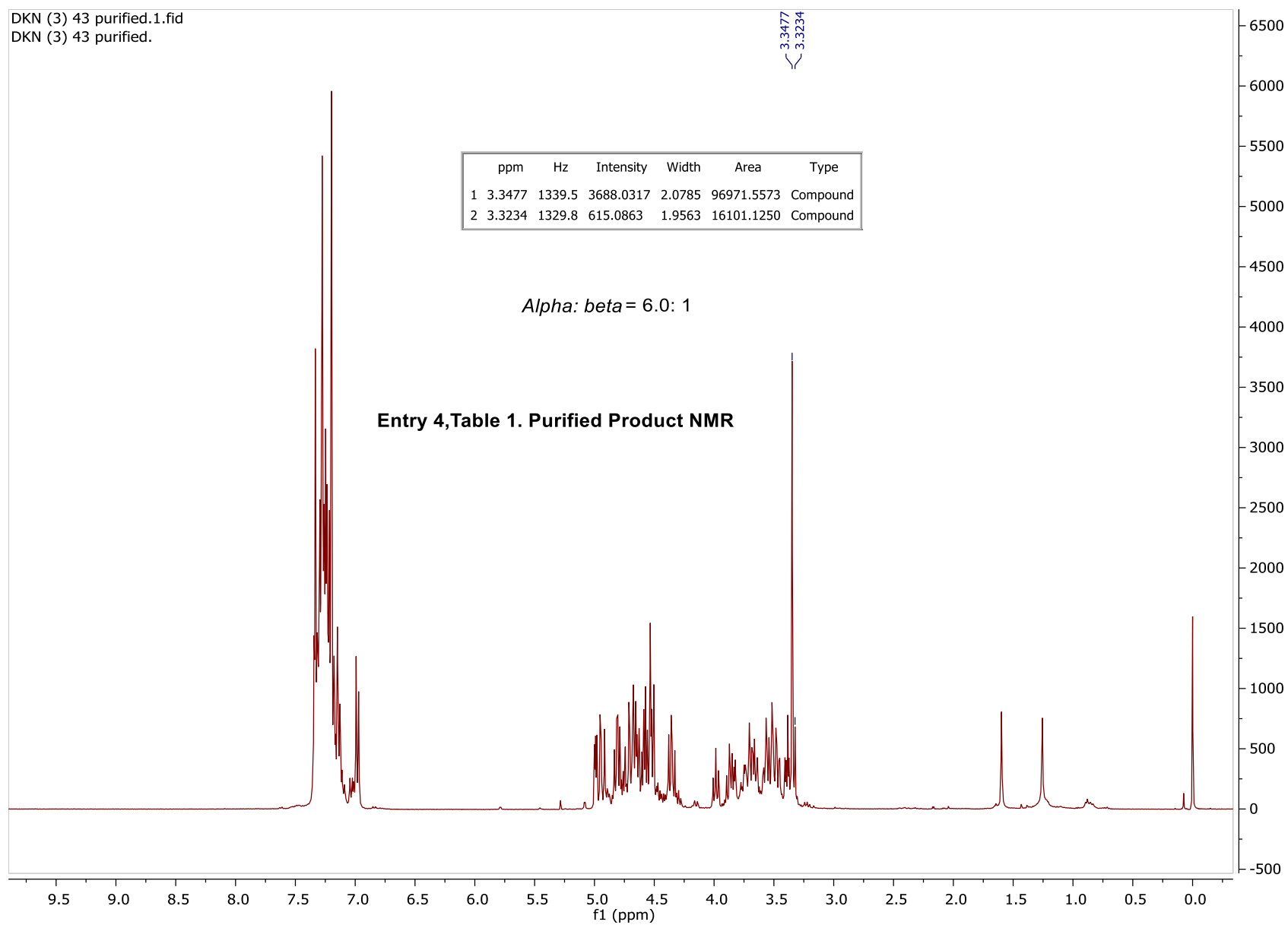
DKN (3) 44 purified.1.fid
DKN (3) 44 purified.



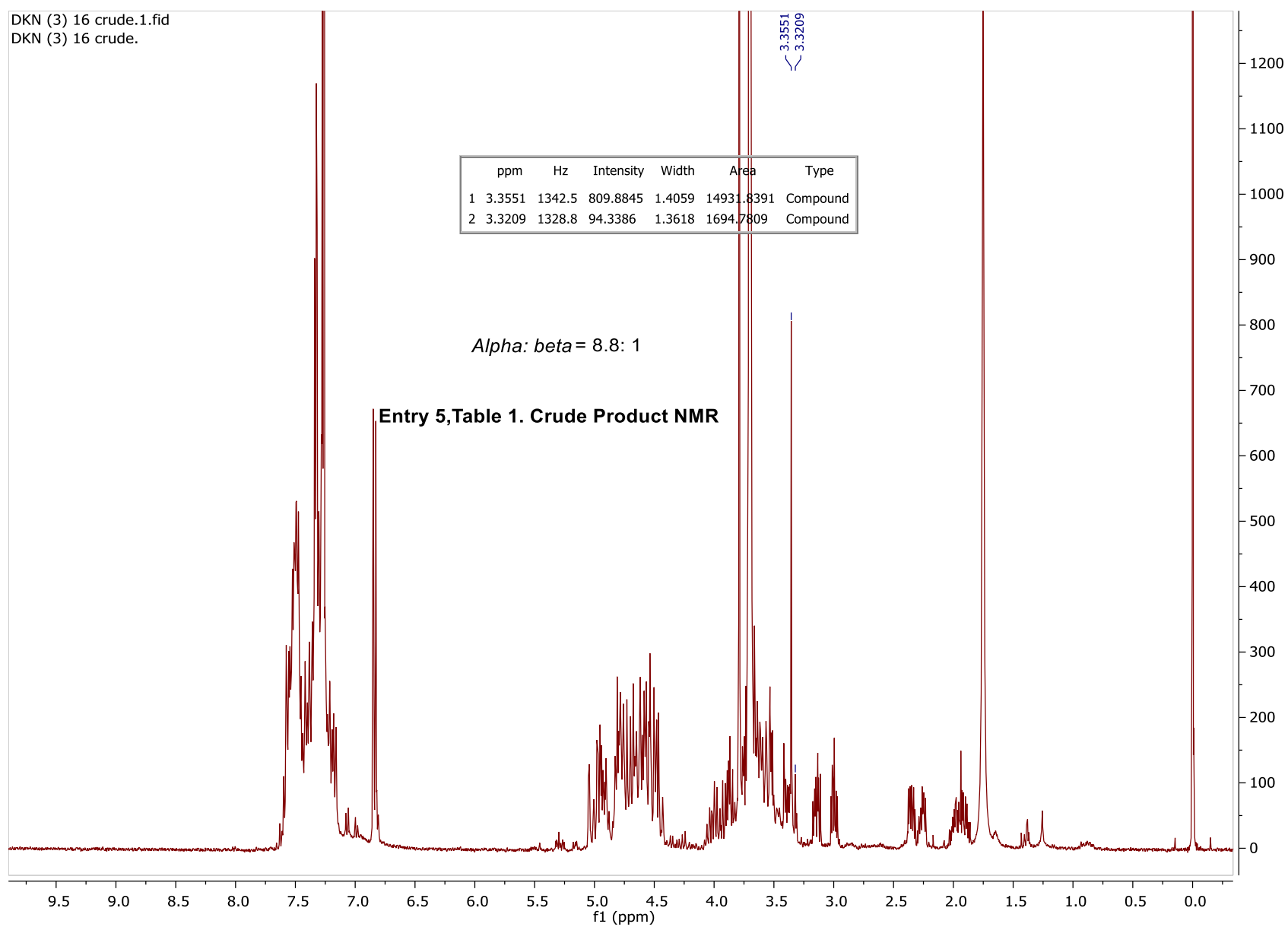
DKN (3) 43 crude.1.fid
DKN (3) 43 crude.



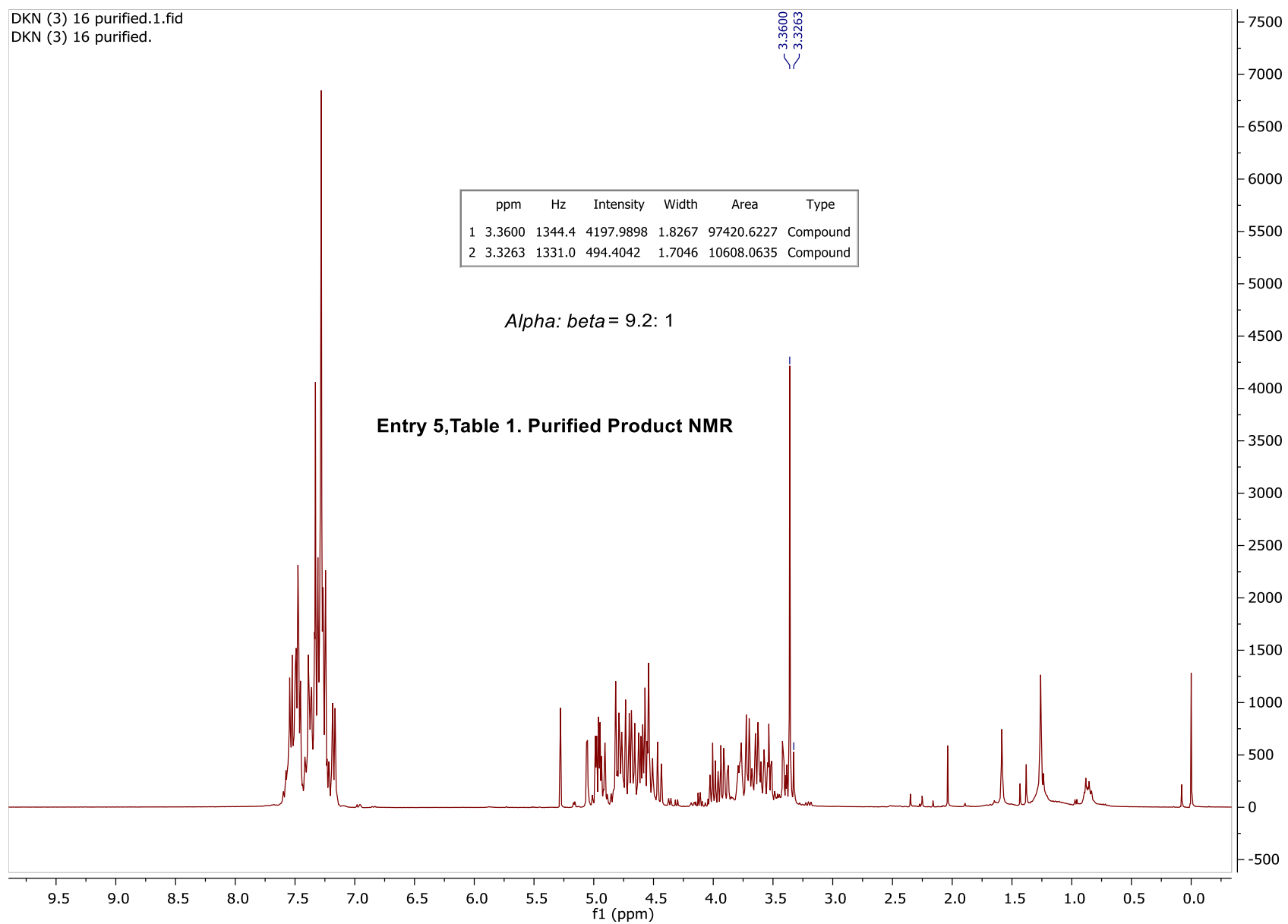
DKN (3) 43 purified.1.fid
DKN (3) 43 purified.



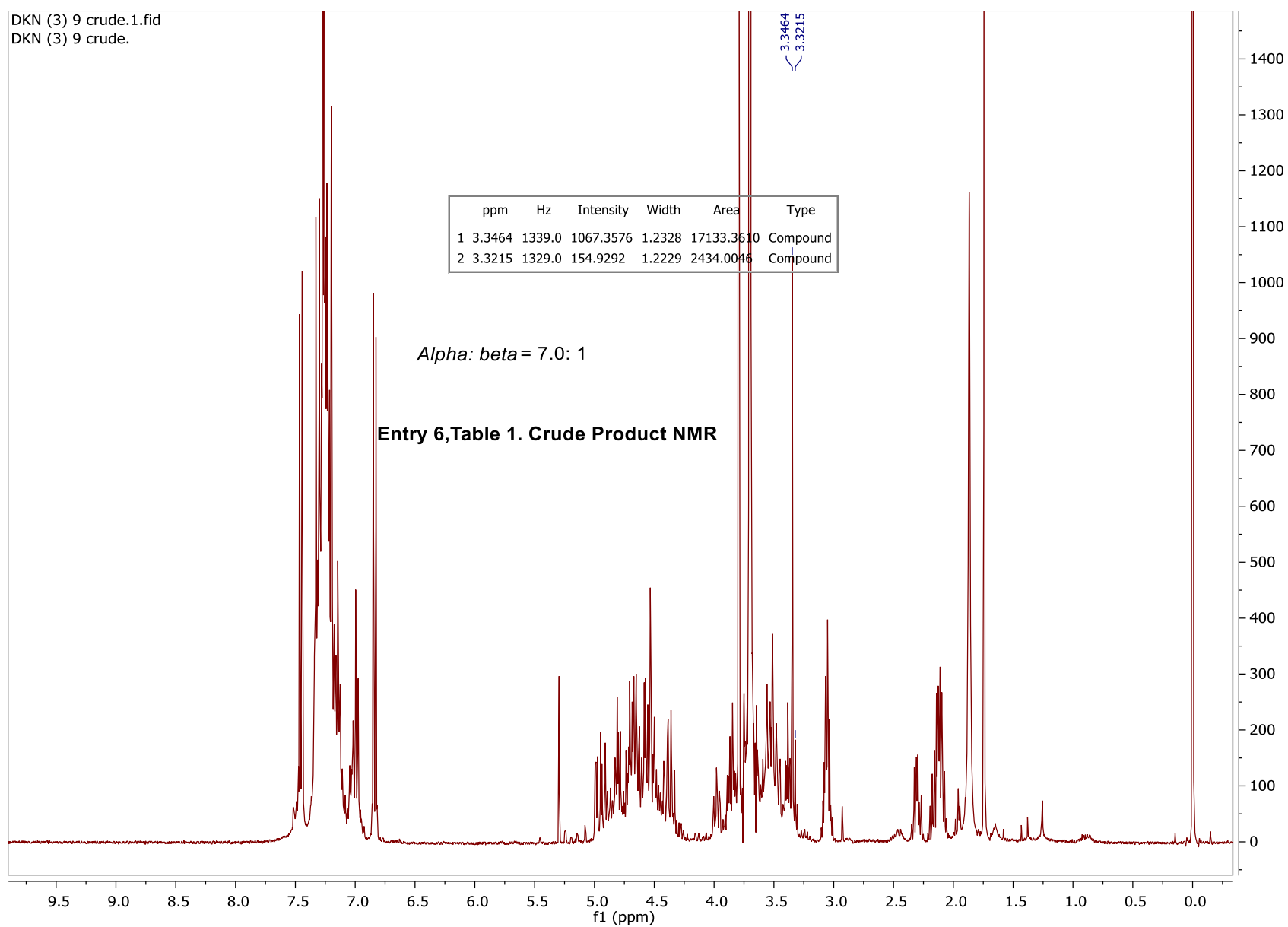
DKN (3) 16 crude.1.fid
DKN (3) 16 crude.



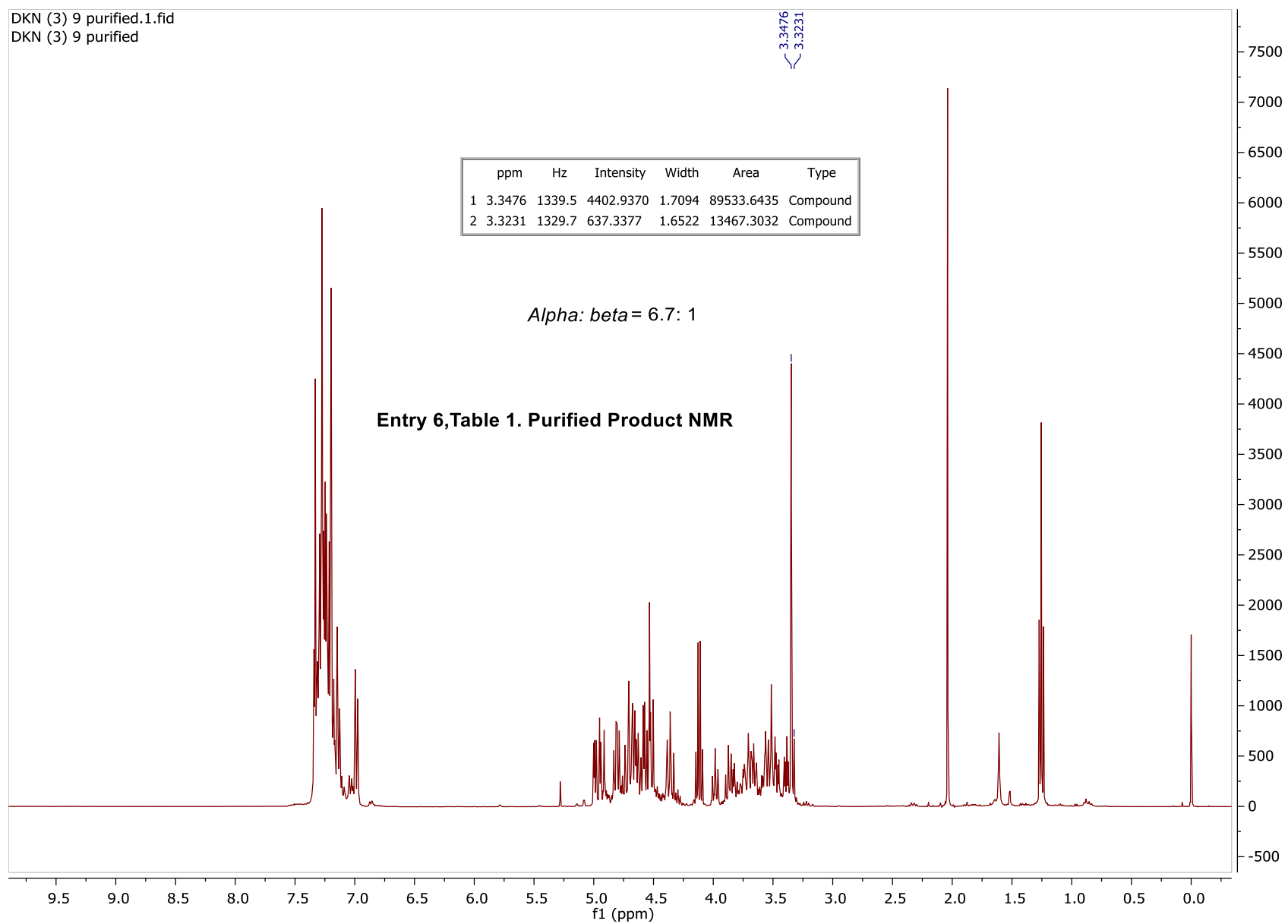
DKN (3) 16 purified.1.fid
DKN (3) 16 purified.



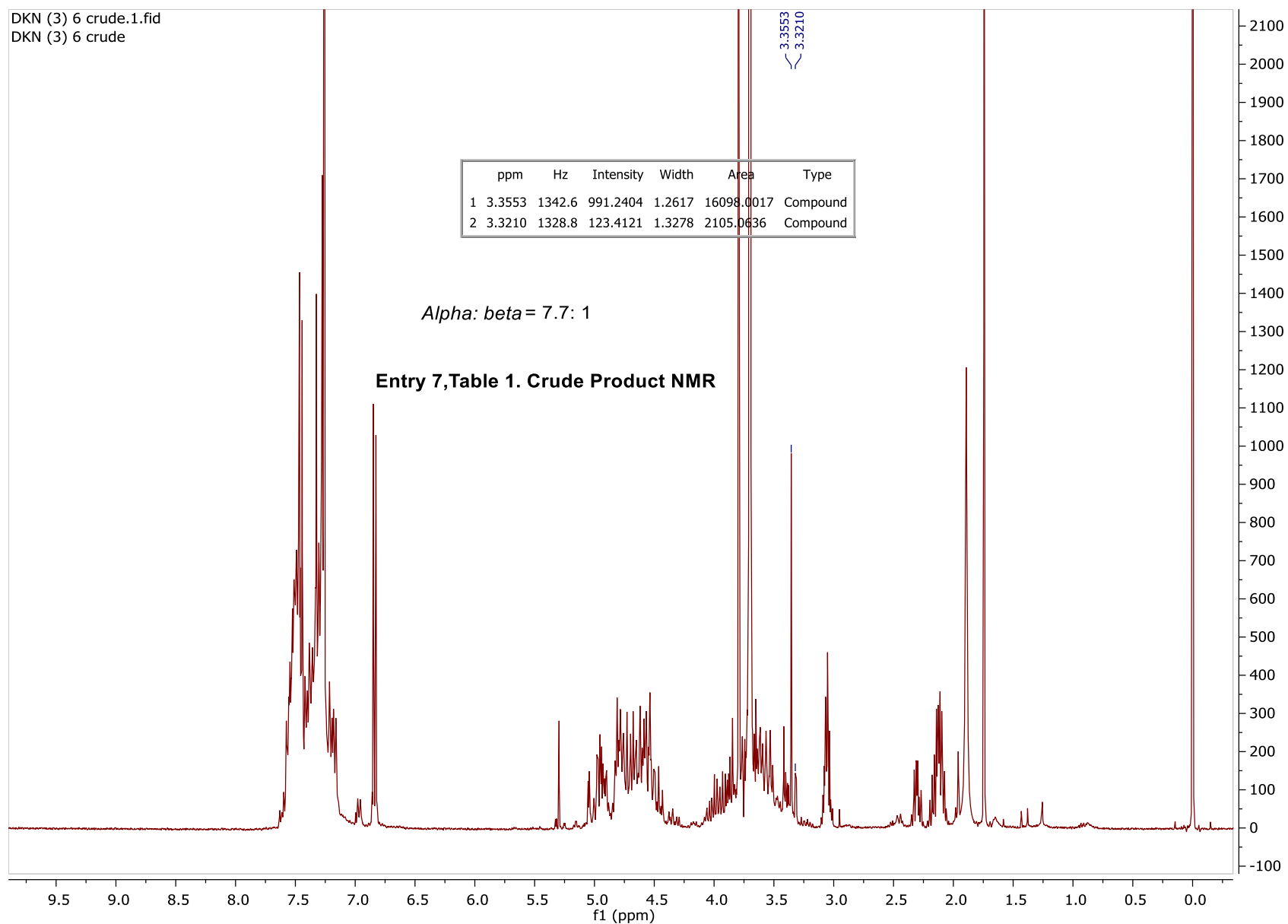
DKN (3) 9 crude.1.fid
DKN (3) 9 crude.



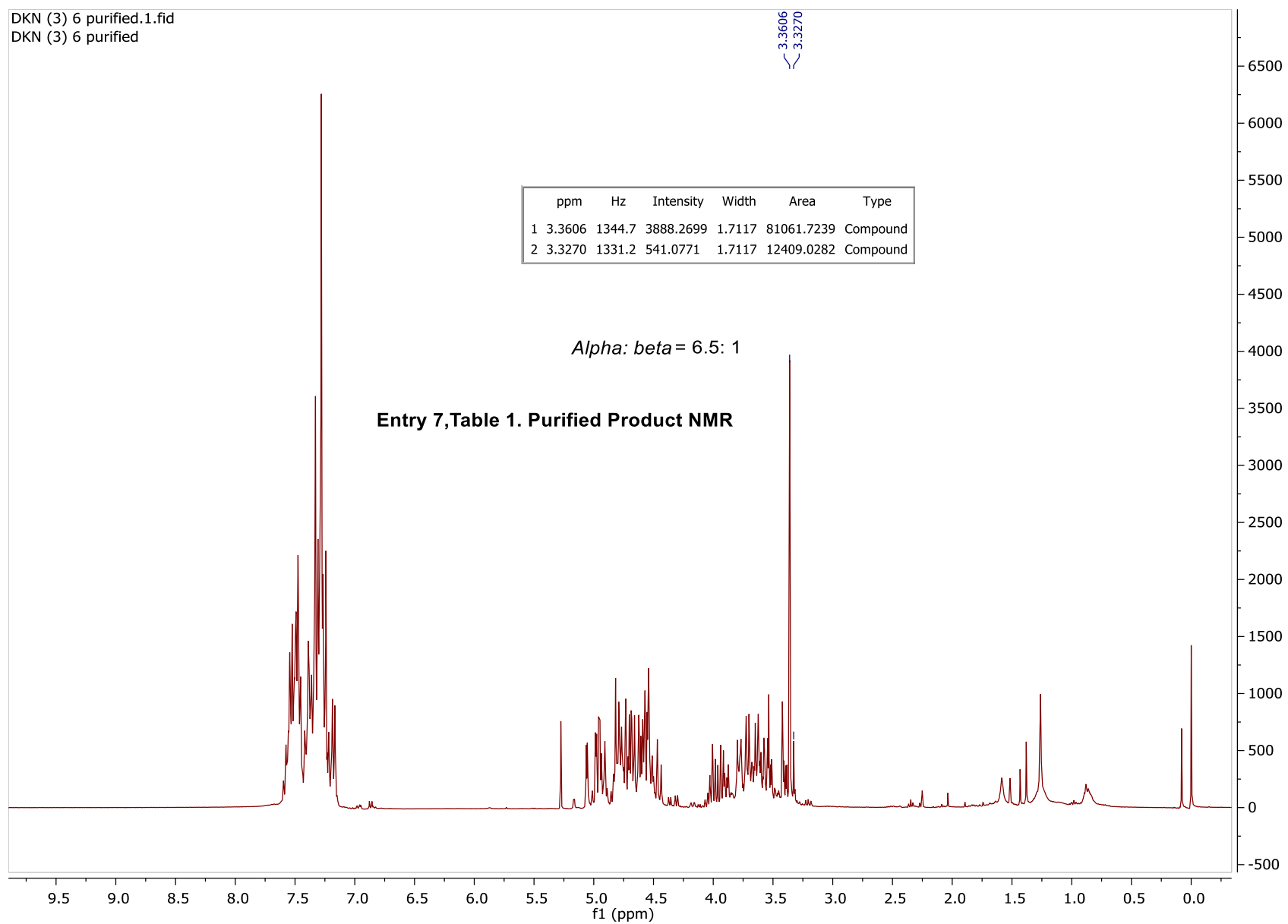
DKN (3) 9 purified.1.fid
DKN (3) 9 purified



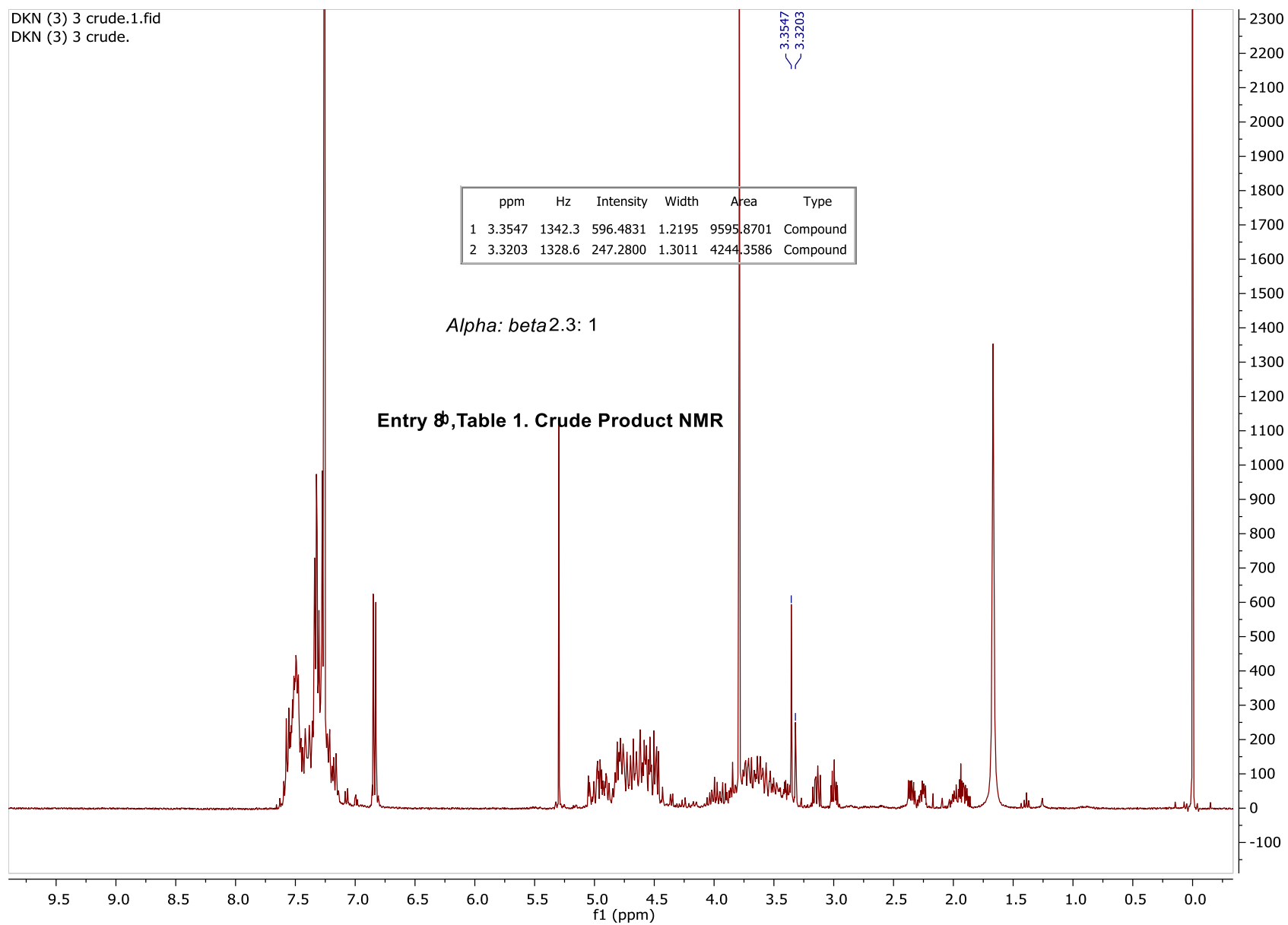
DKN (3) 6 crude.1.fid
DKN (3) 6 crude



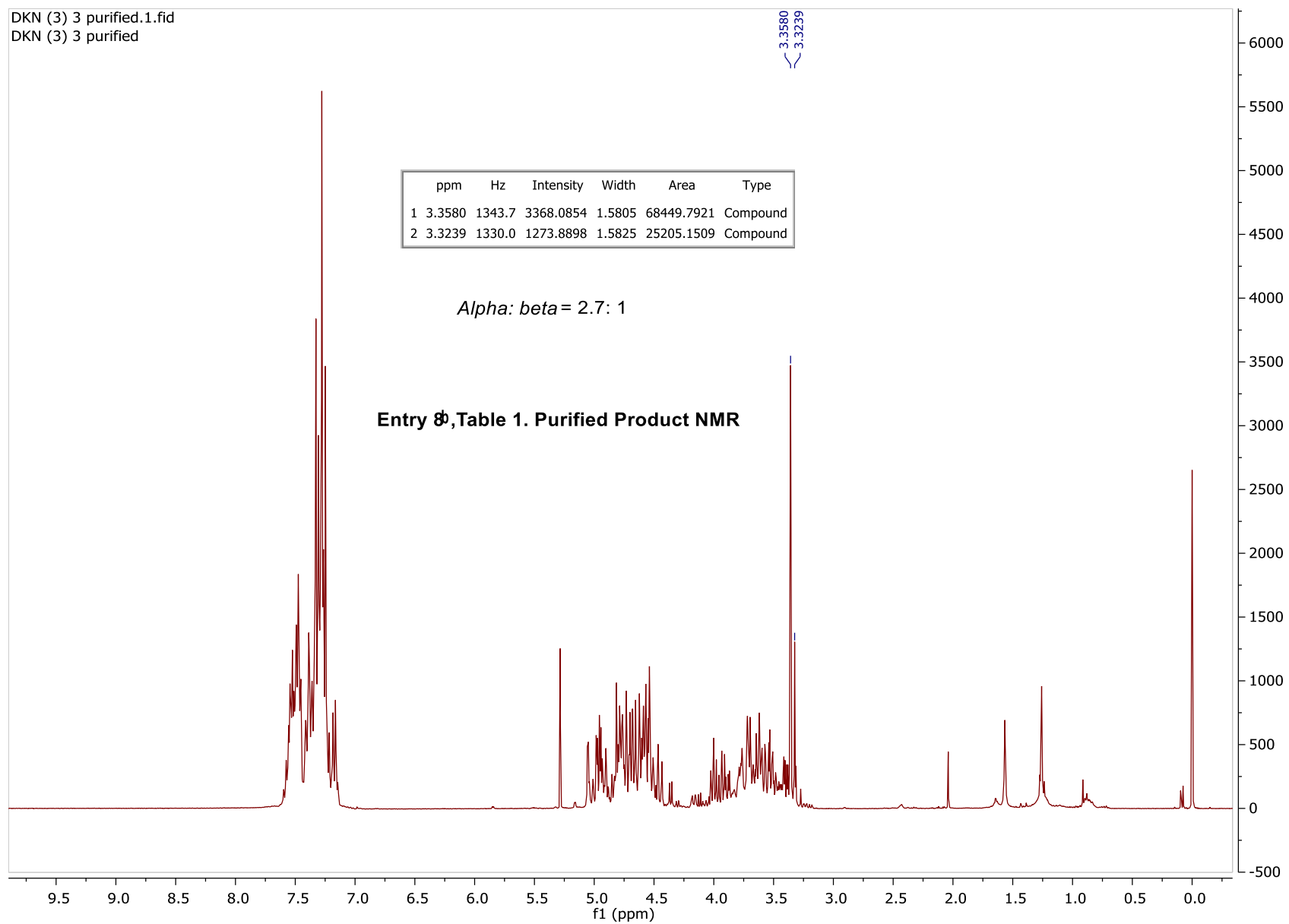
DKN (3) 6 purified.1.fid
DKN (3) 6 purified



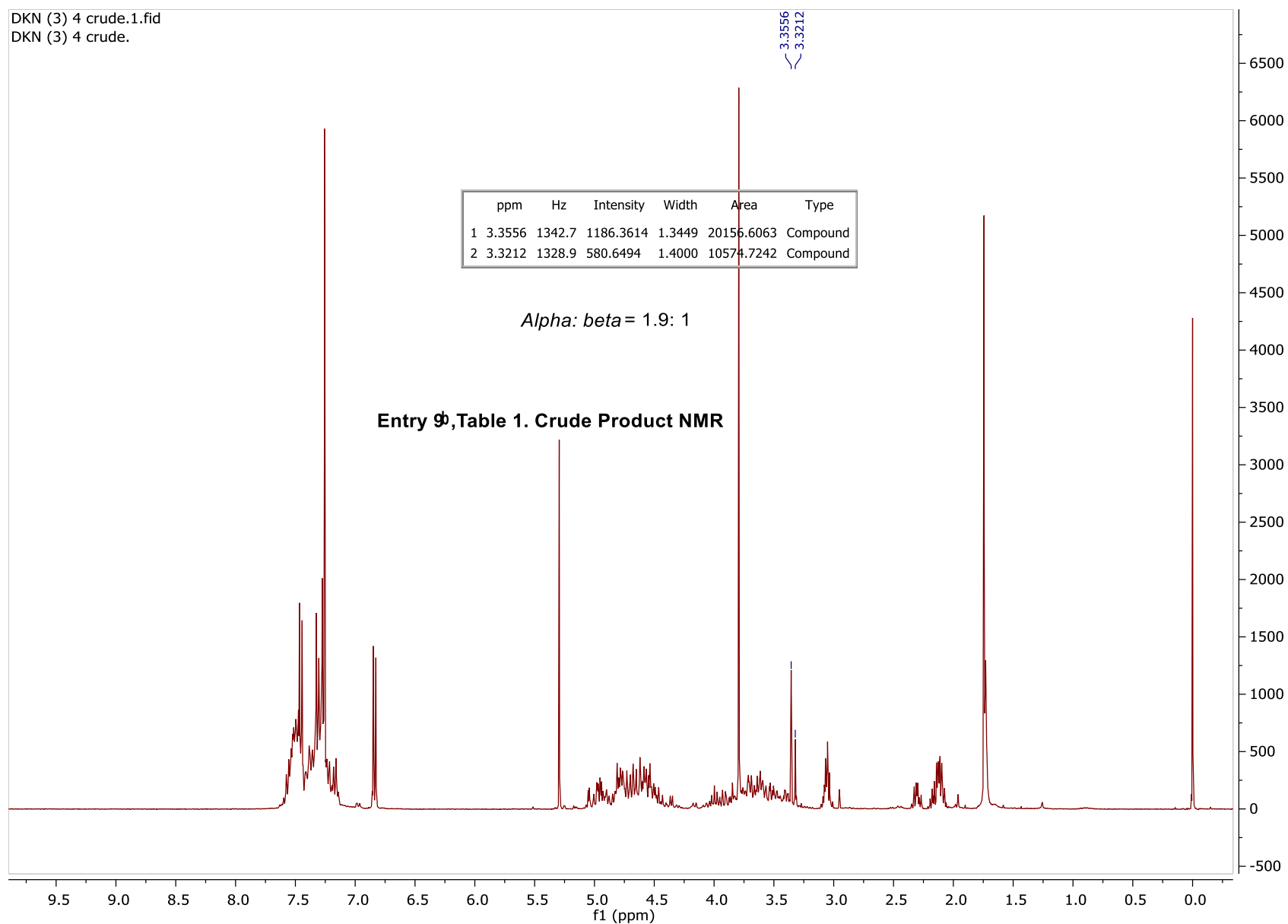
DKN (3) 3 crude.1.fid
DKN (3) 3 crude.



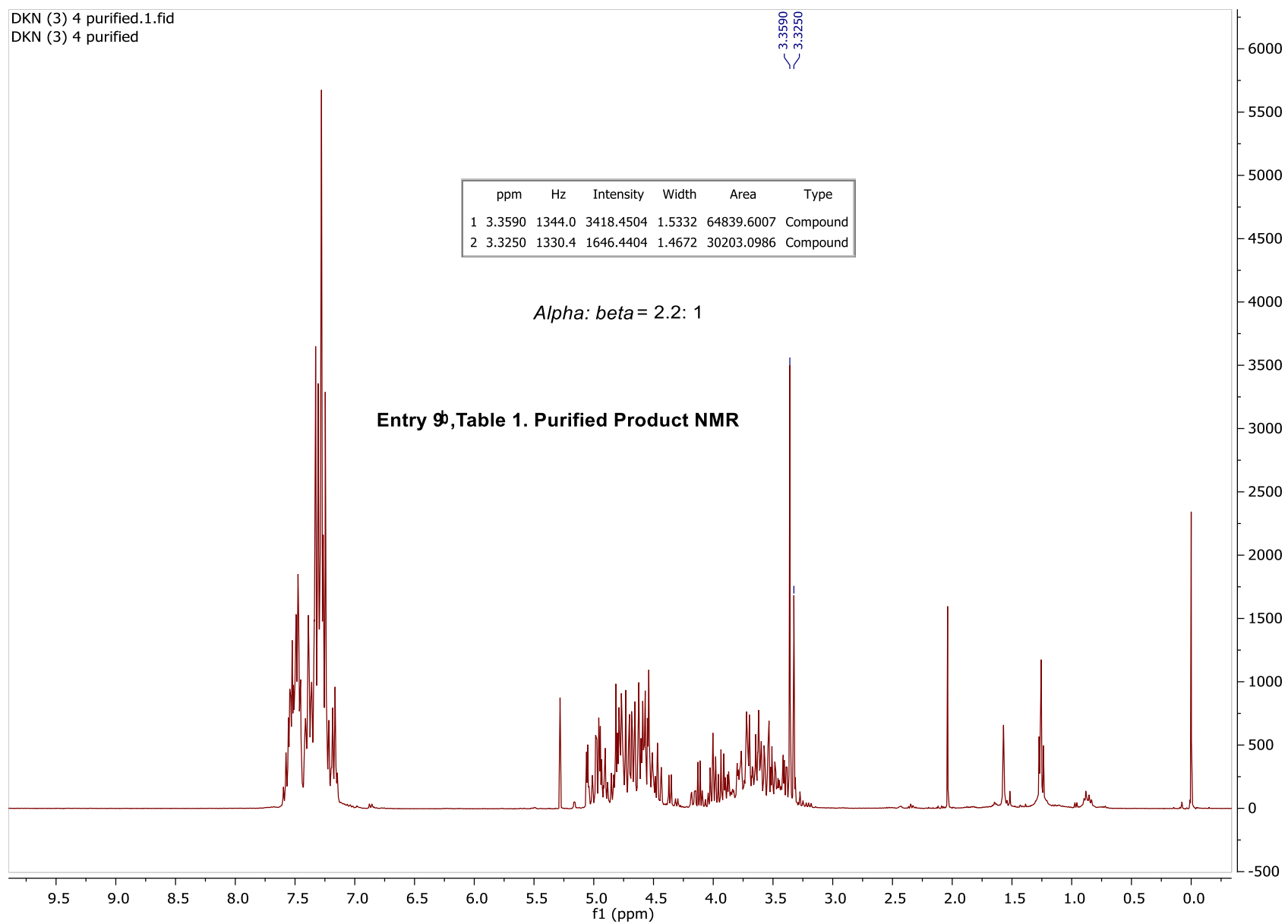
DKN (3) 3 purified.1.fid
DKN (3) 3 purified



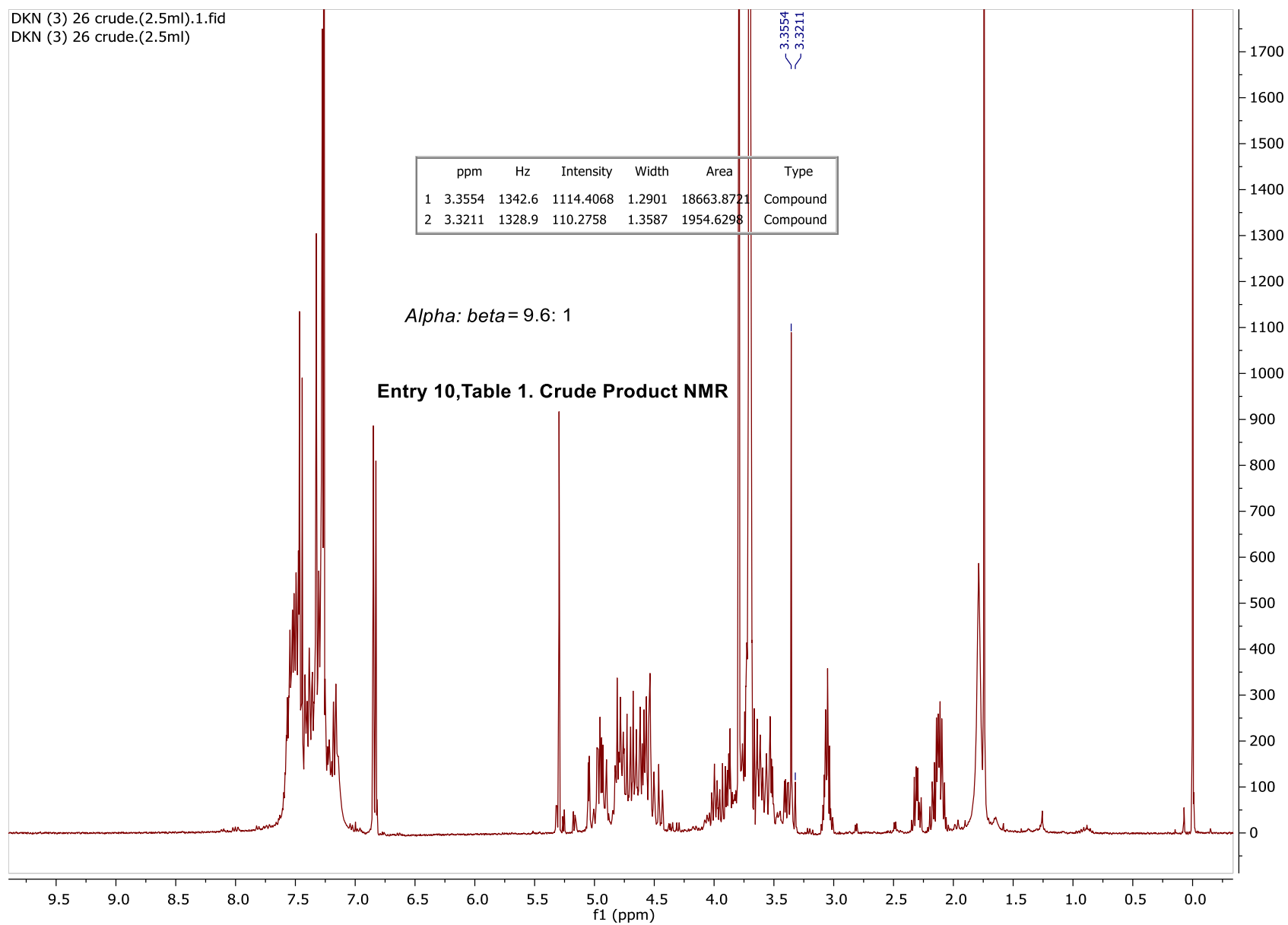
DKN (3) 4 crude.1.fid
DKN (3) 4 crude.



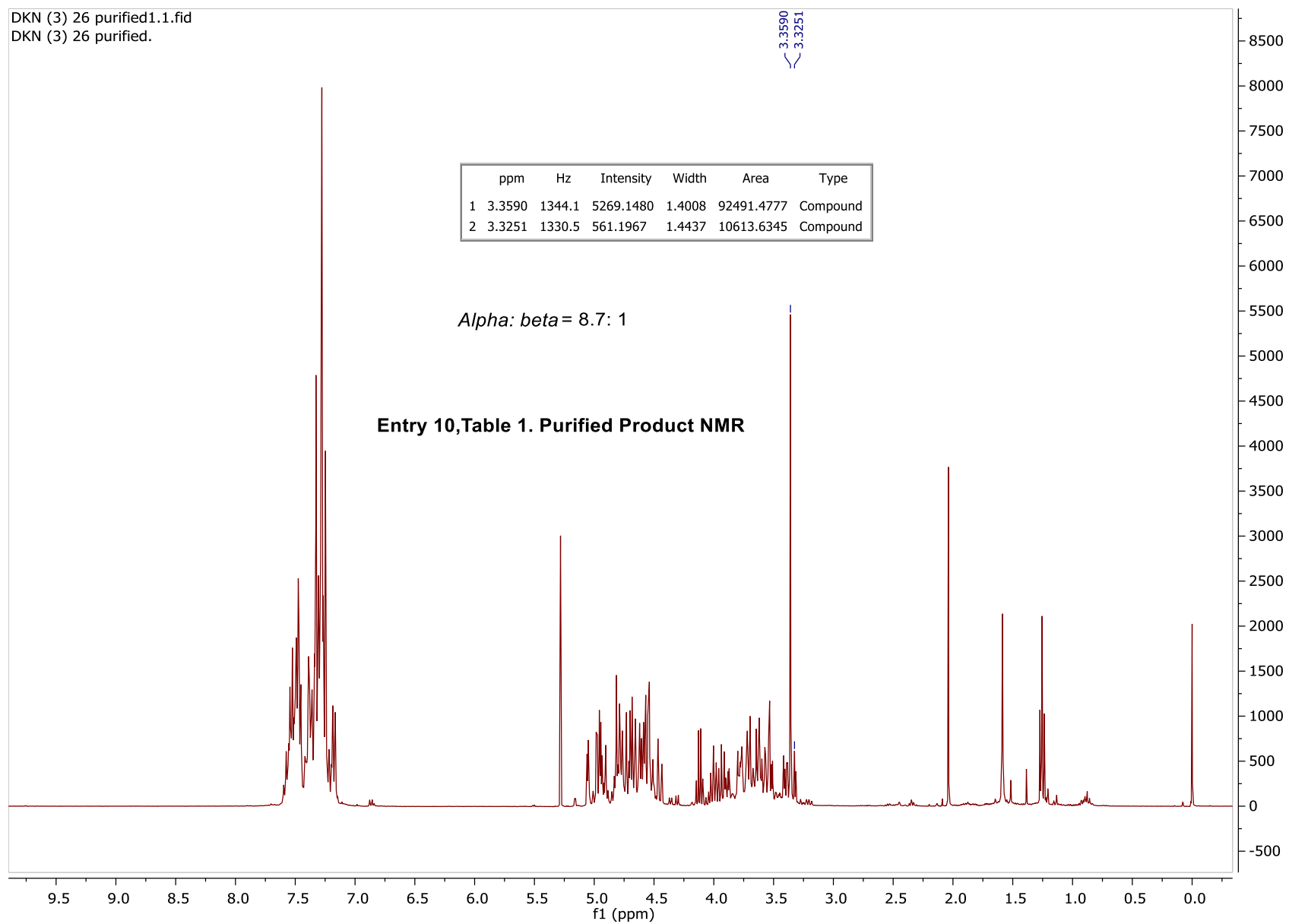
DKN (3) 4 purified.1.fid
DKN (3) 4 purified

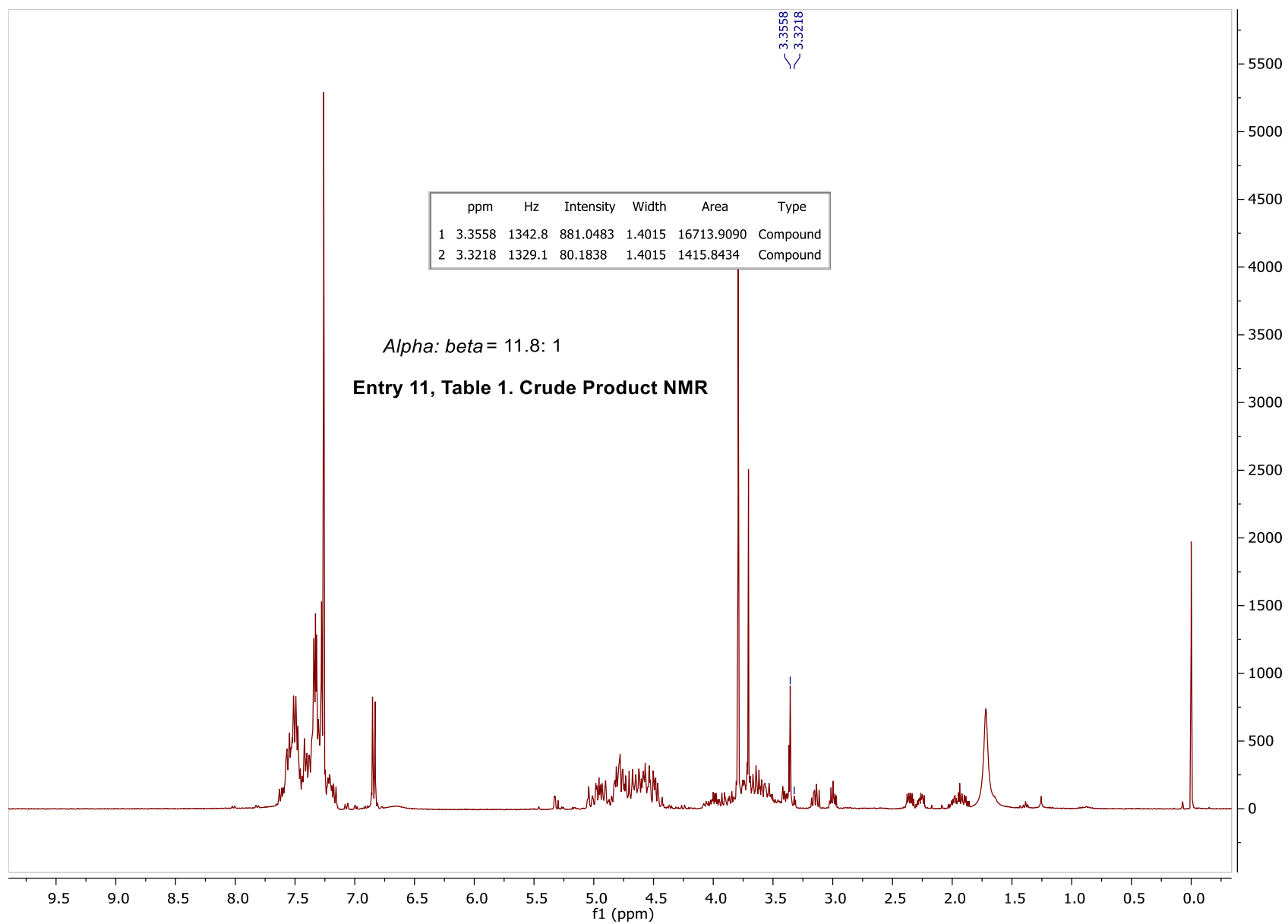


DKN (3) 26 crude.(2.5ml).1.fid
DKN (3) 26 crude.(2.5ml)

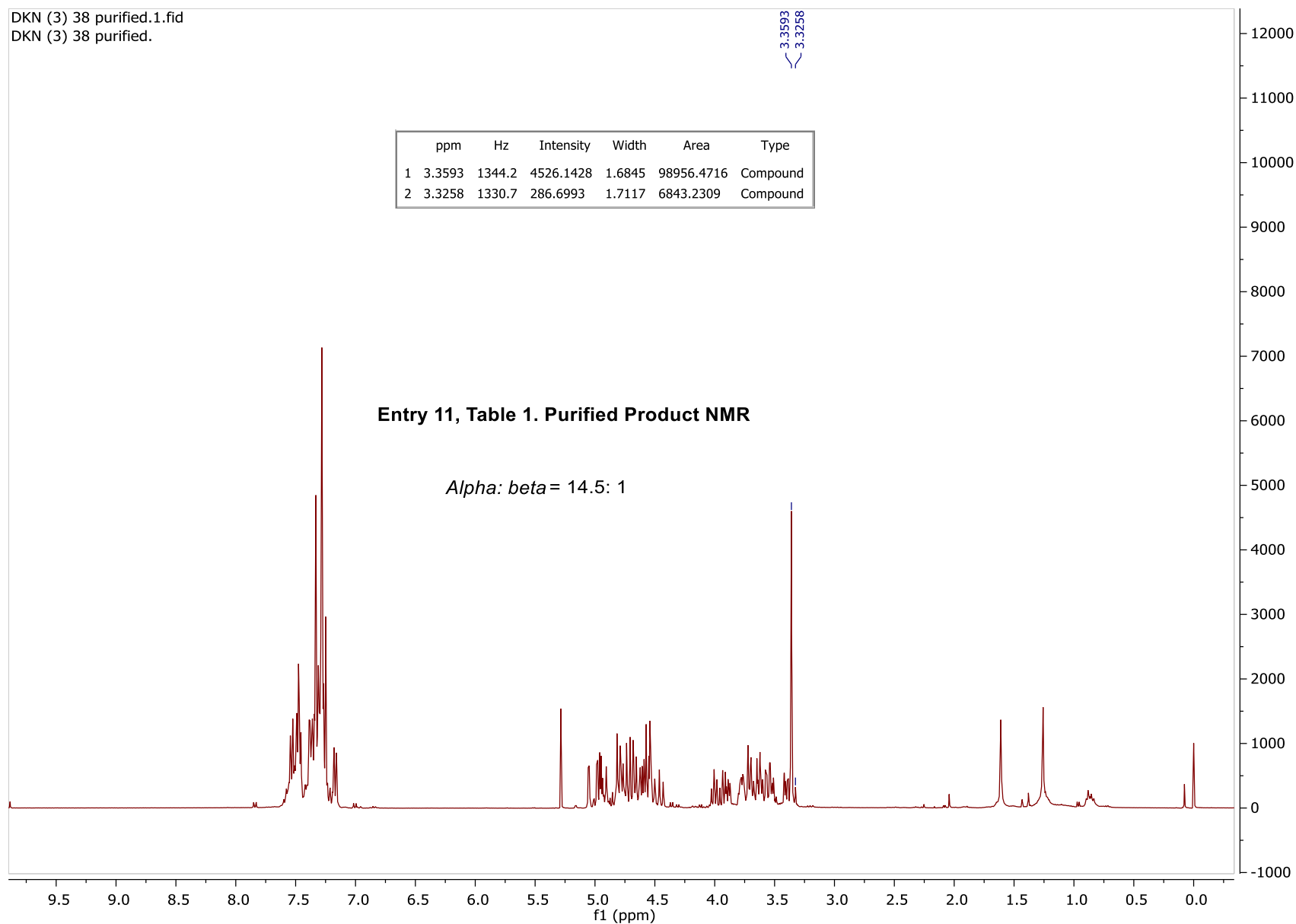


DKN (3) 26 purified1.1.fid
DKN (3) 26 purified.

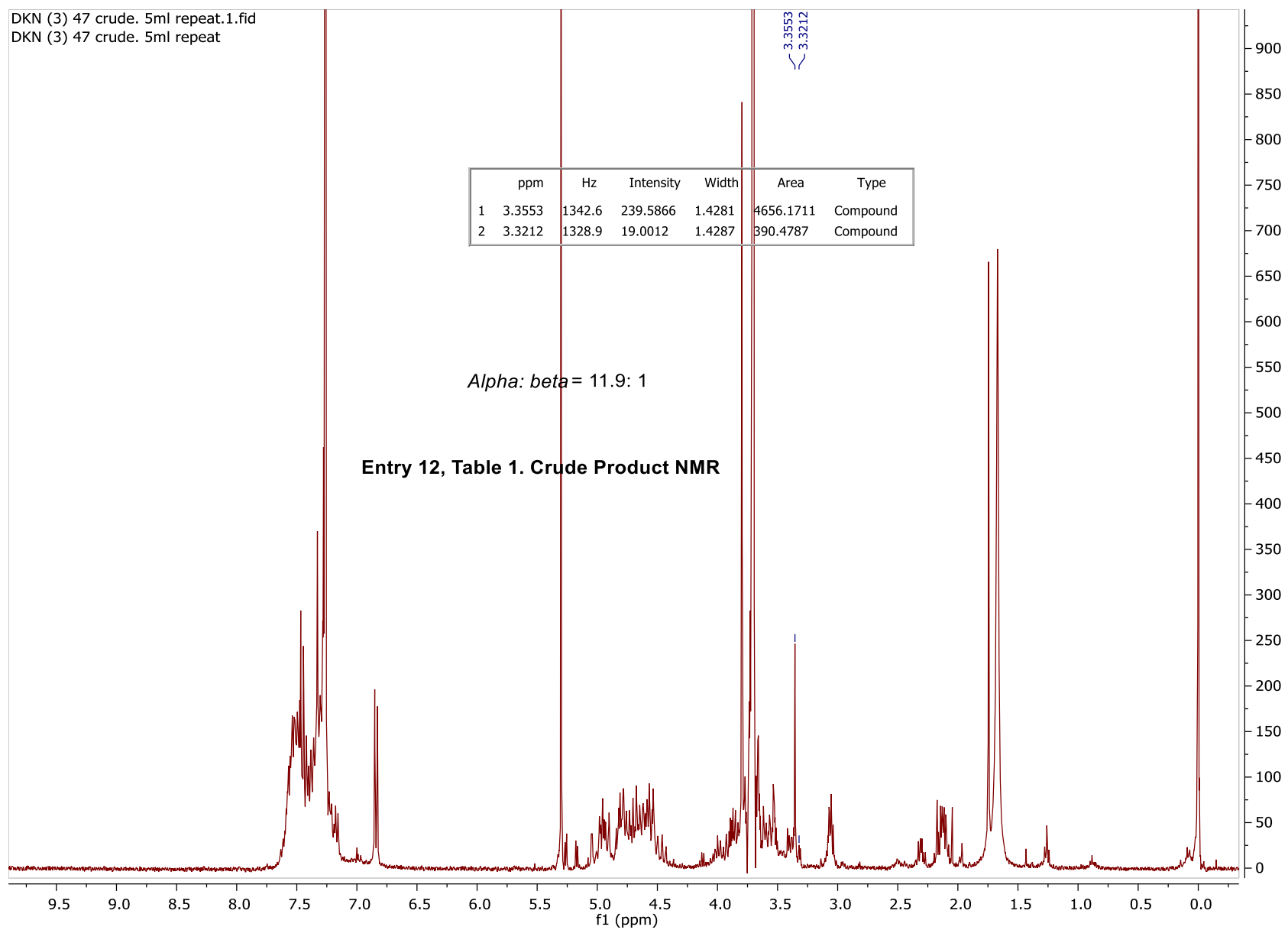




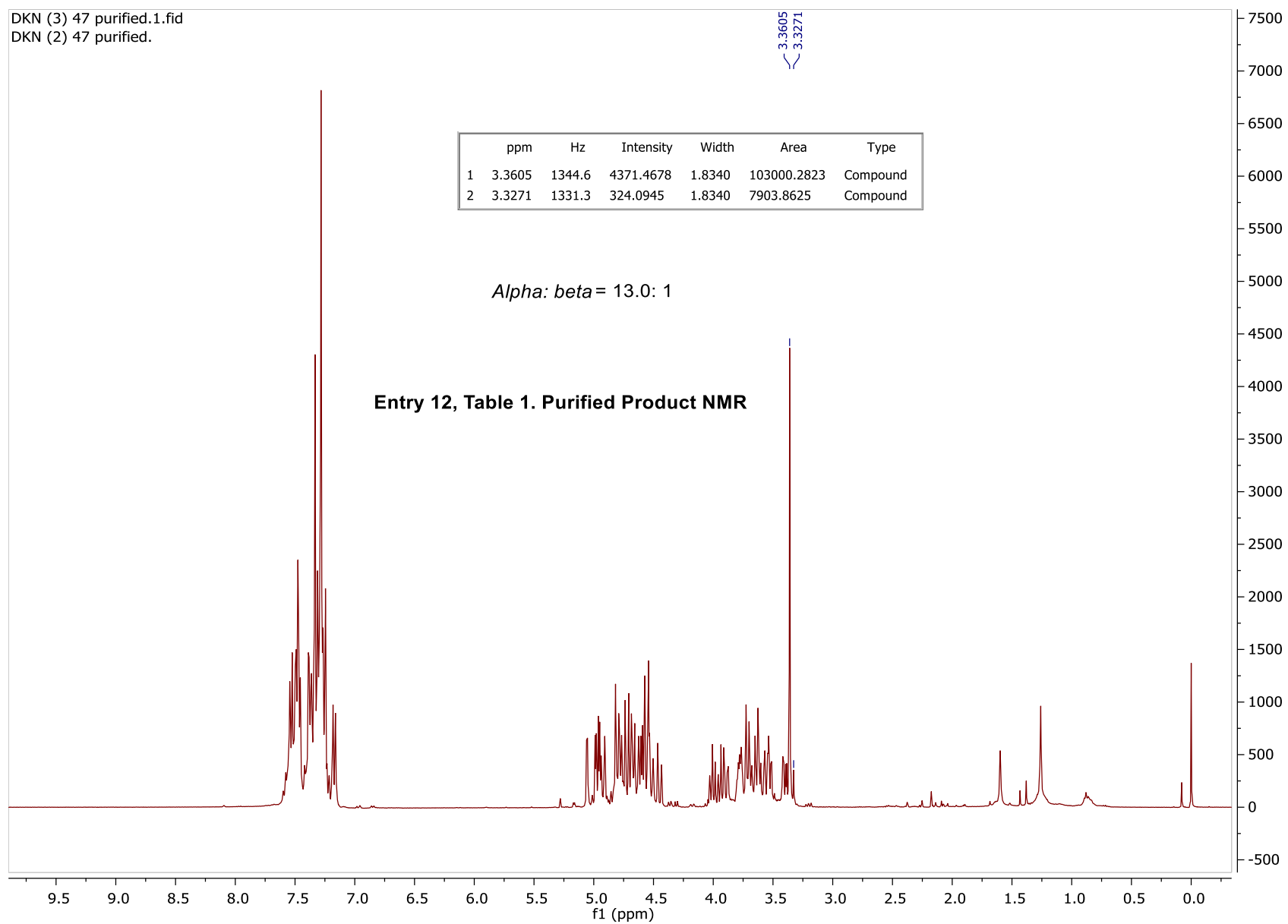
DKN (3) 38 purified.1.fid
DKN (3) 38 purified.



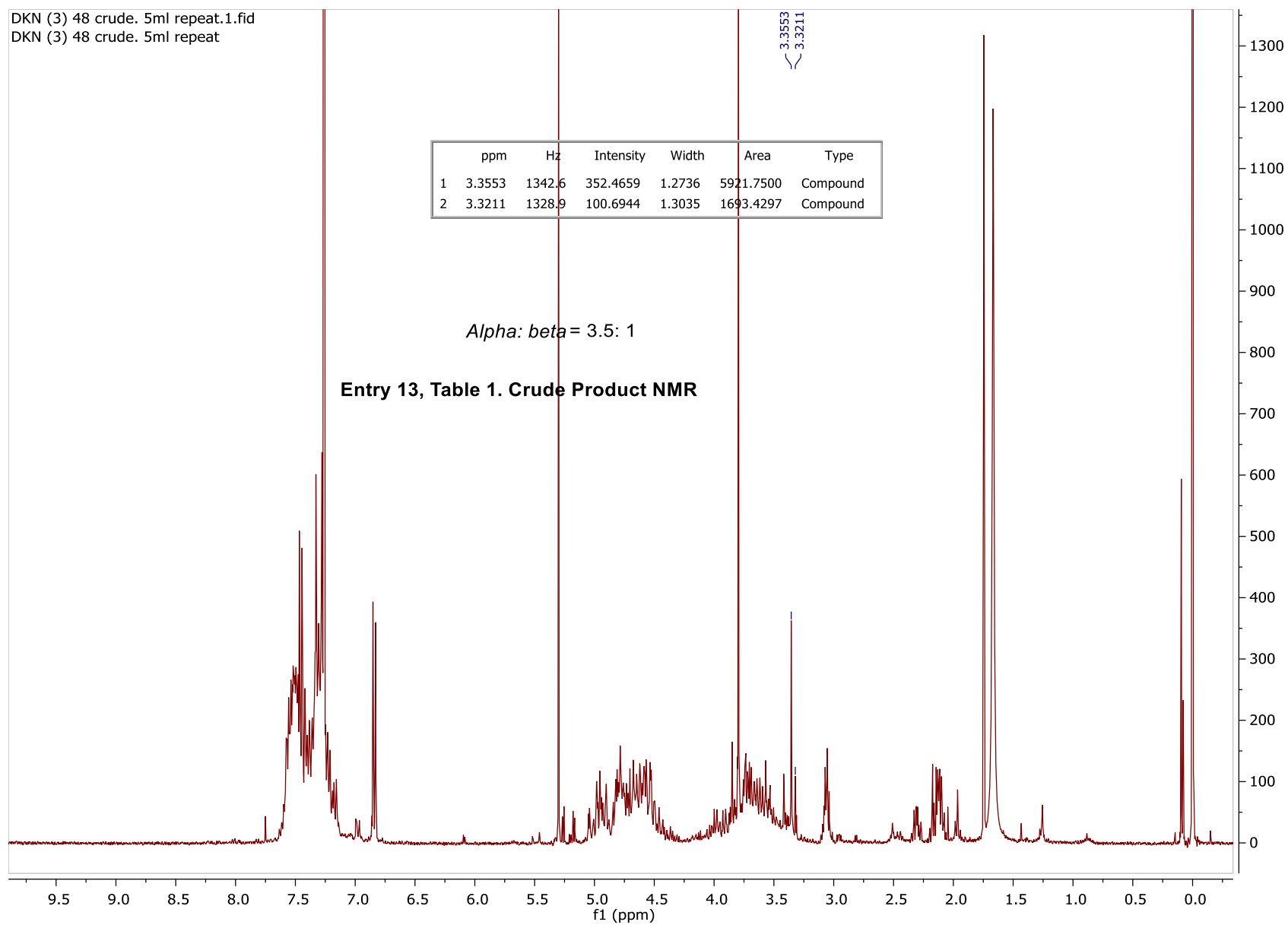
DKN (3) 47 crude. 5ml repeat.1.fid
DKN (3) 47 crude. 5ml repeat



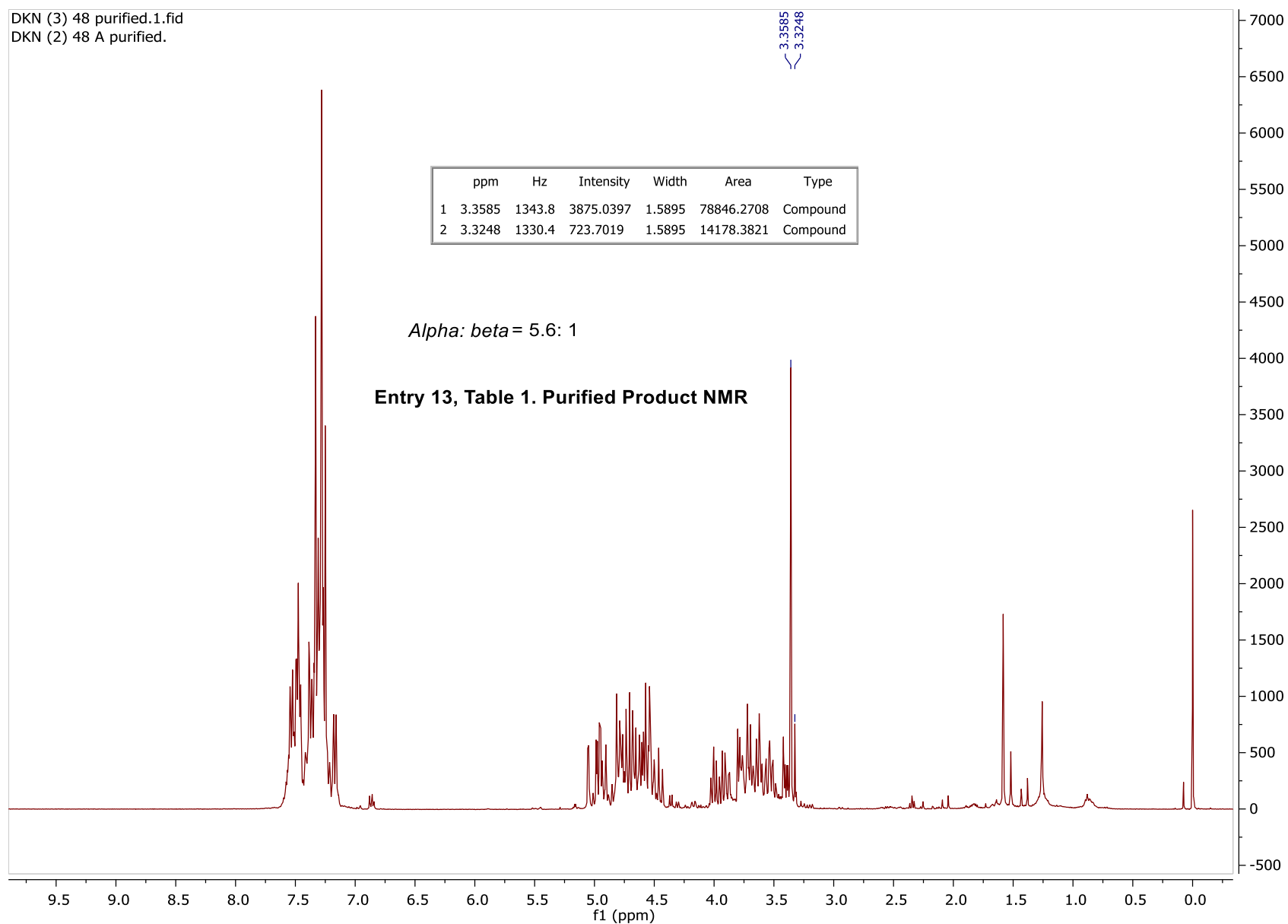
DKN (3) 47 purified.1.fid
DKN (2) 47 purified.

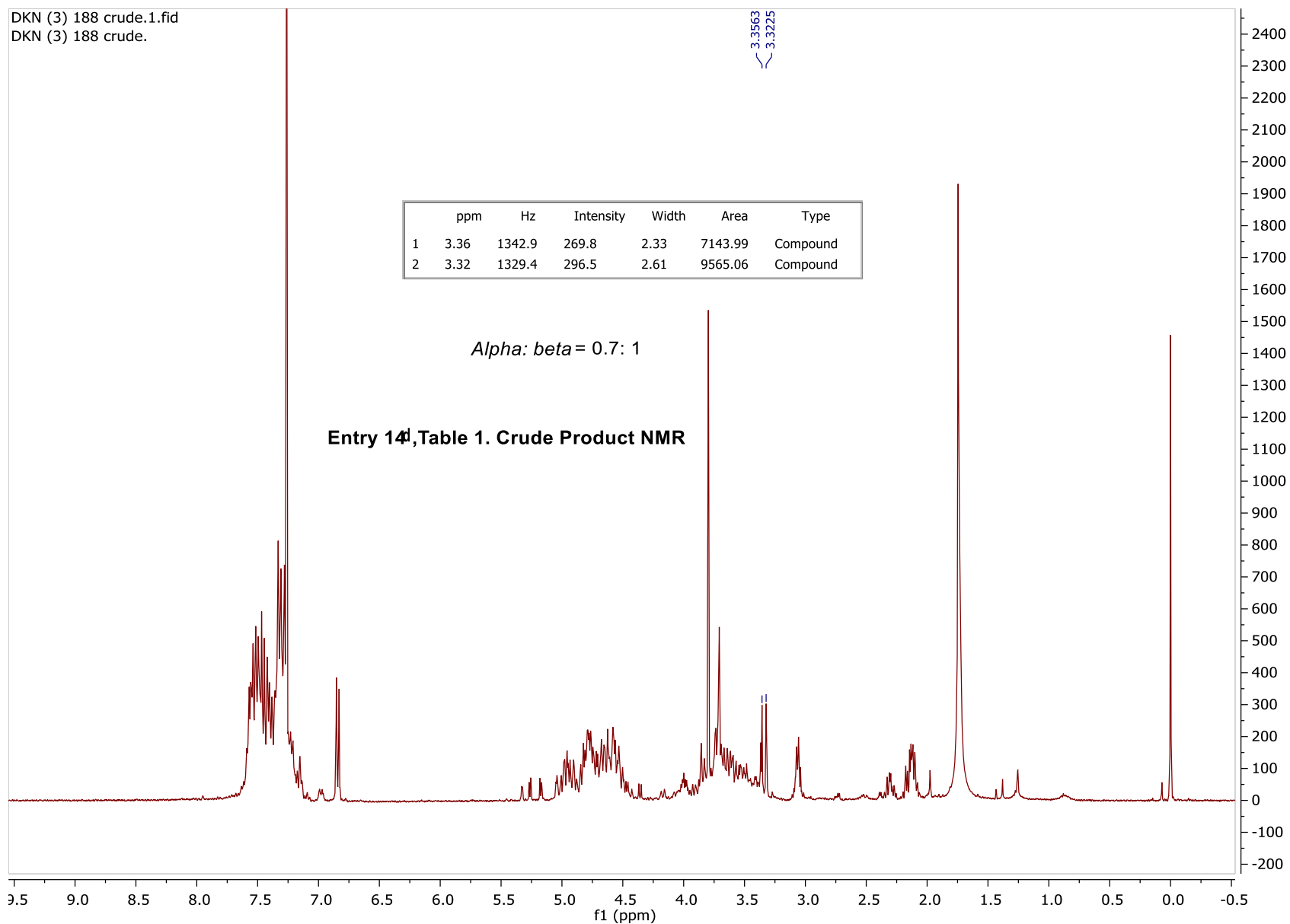


DKN (3) 48 crude. 5ml repeat.1.fid
DKN (3) 48 crude. 5ml repeat

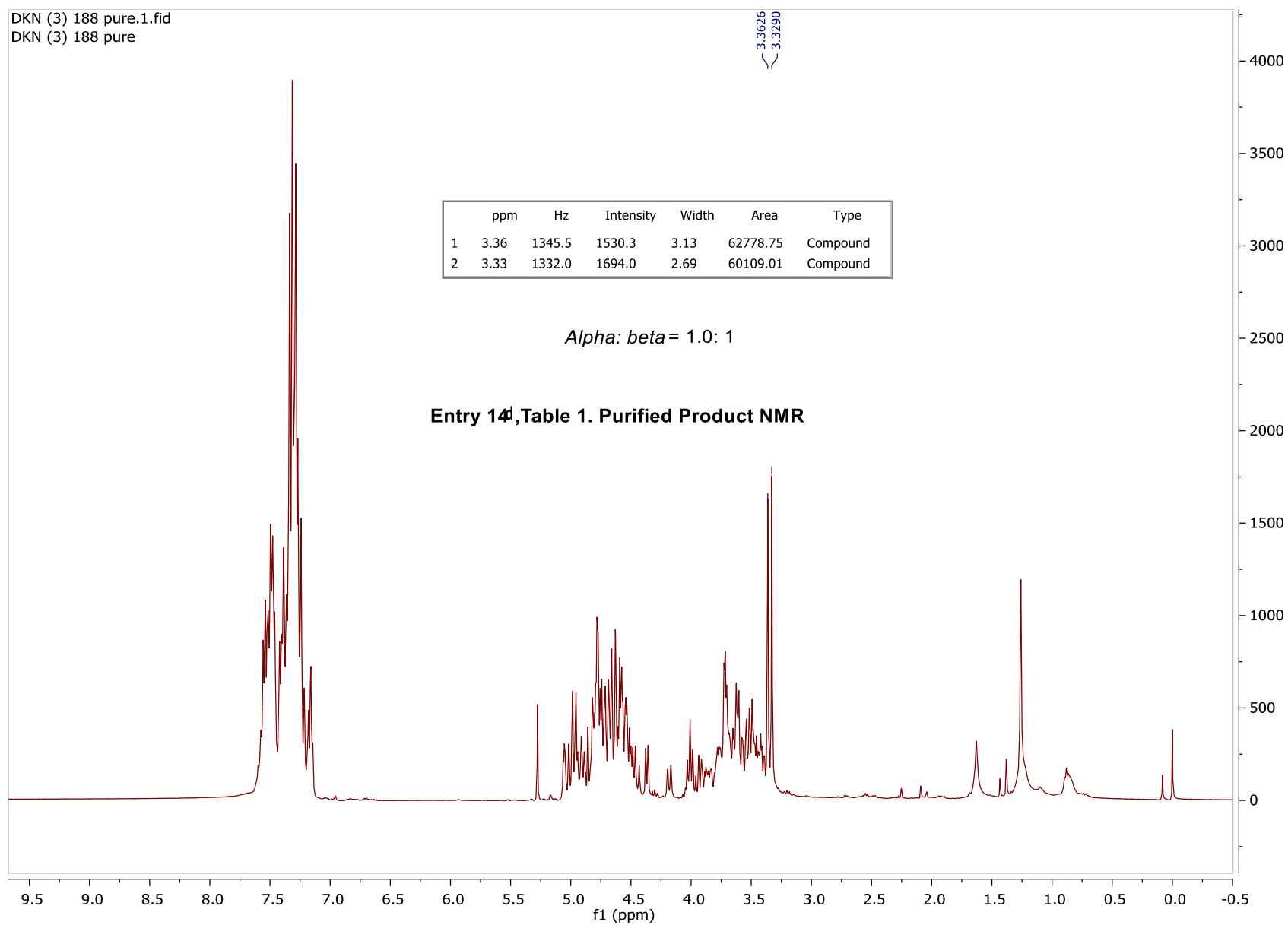


DKN (3) 48 purified.1.fid
DKN (2) 48 A purified.

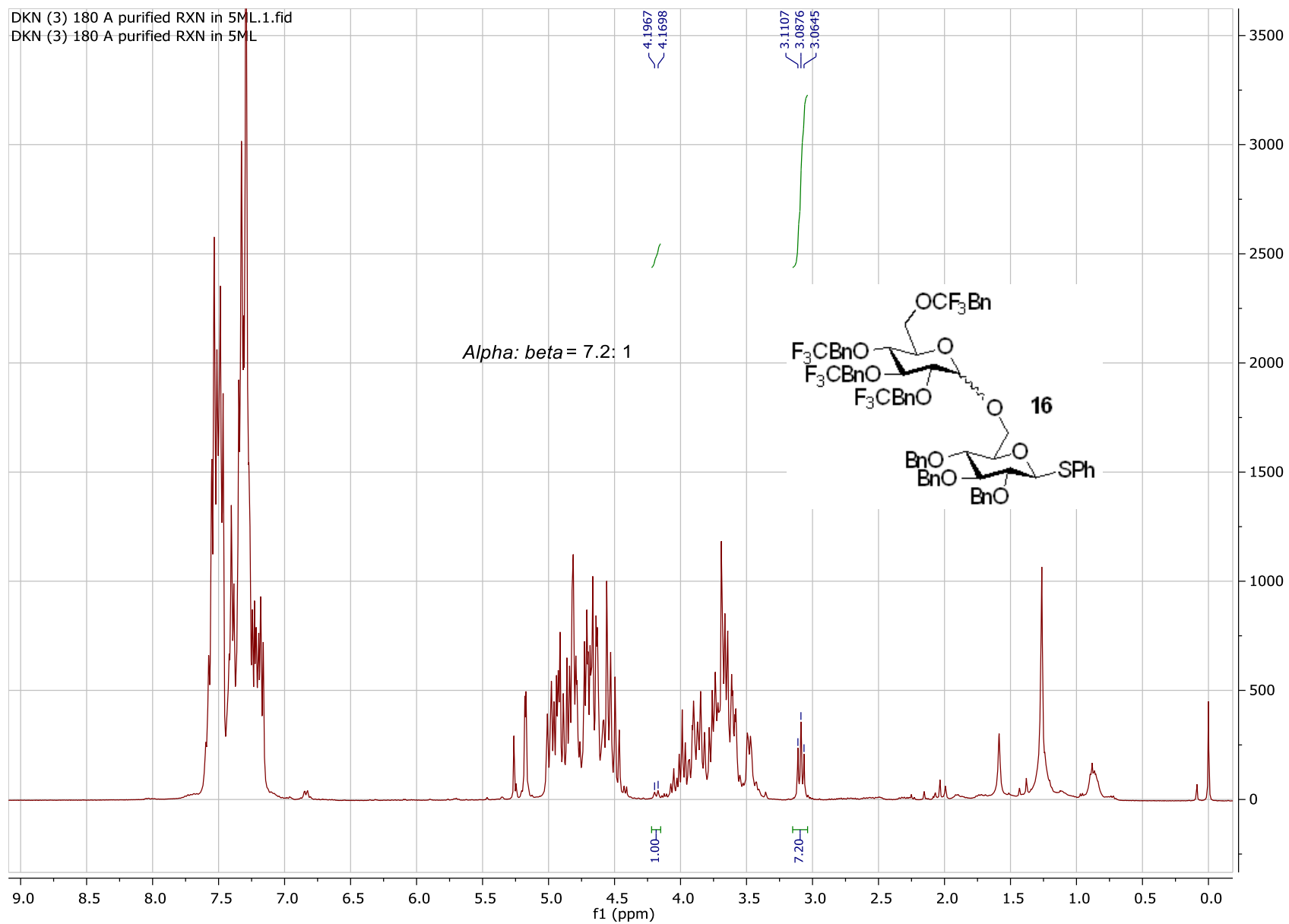




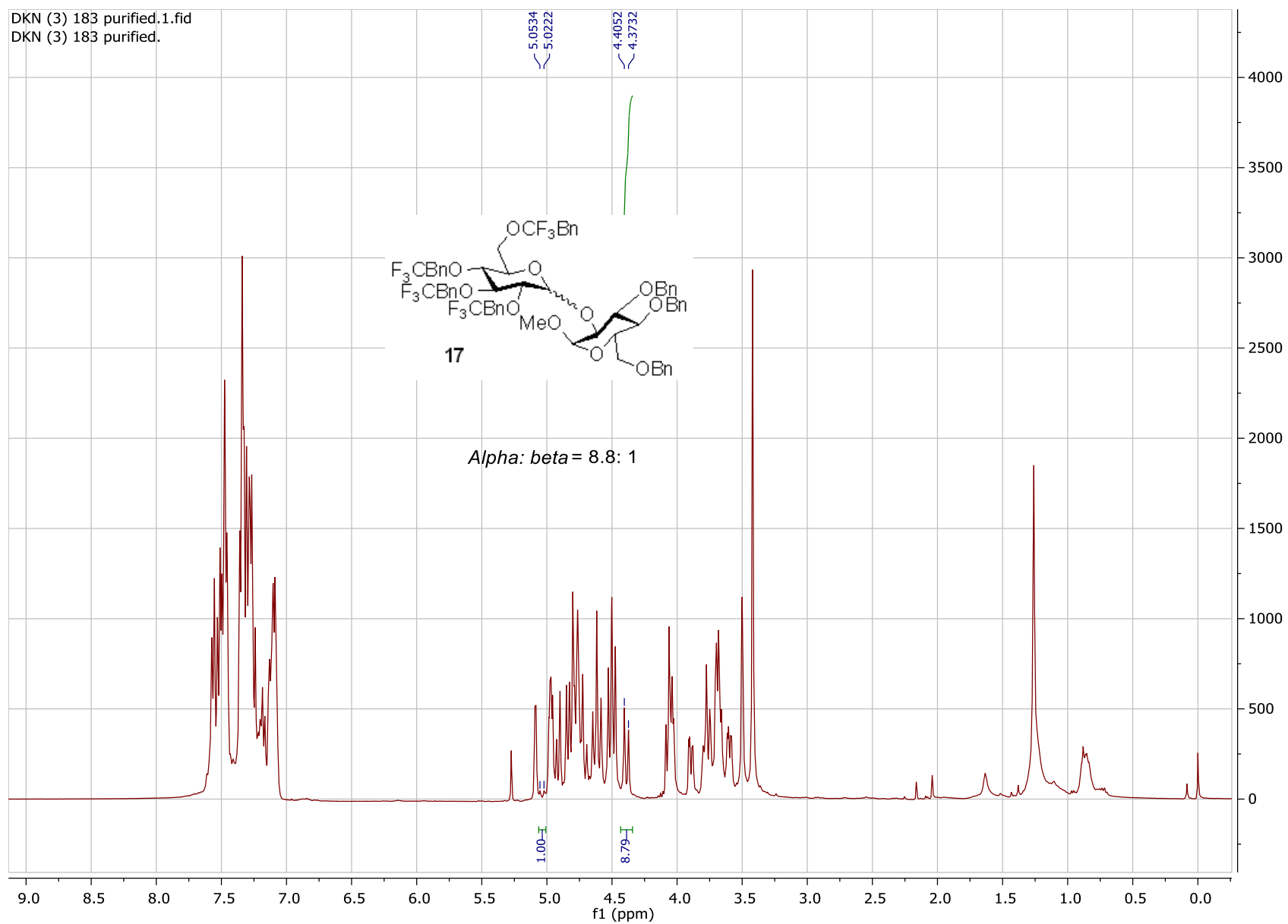
DKN (3) 188 pure.1.fid
DKN (3) 188 pure



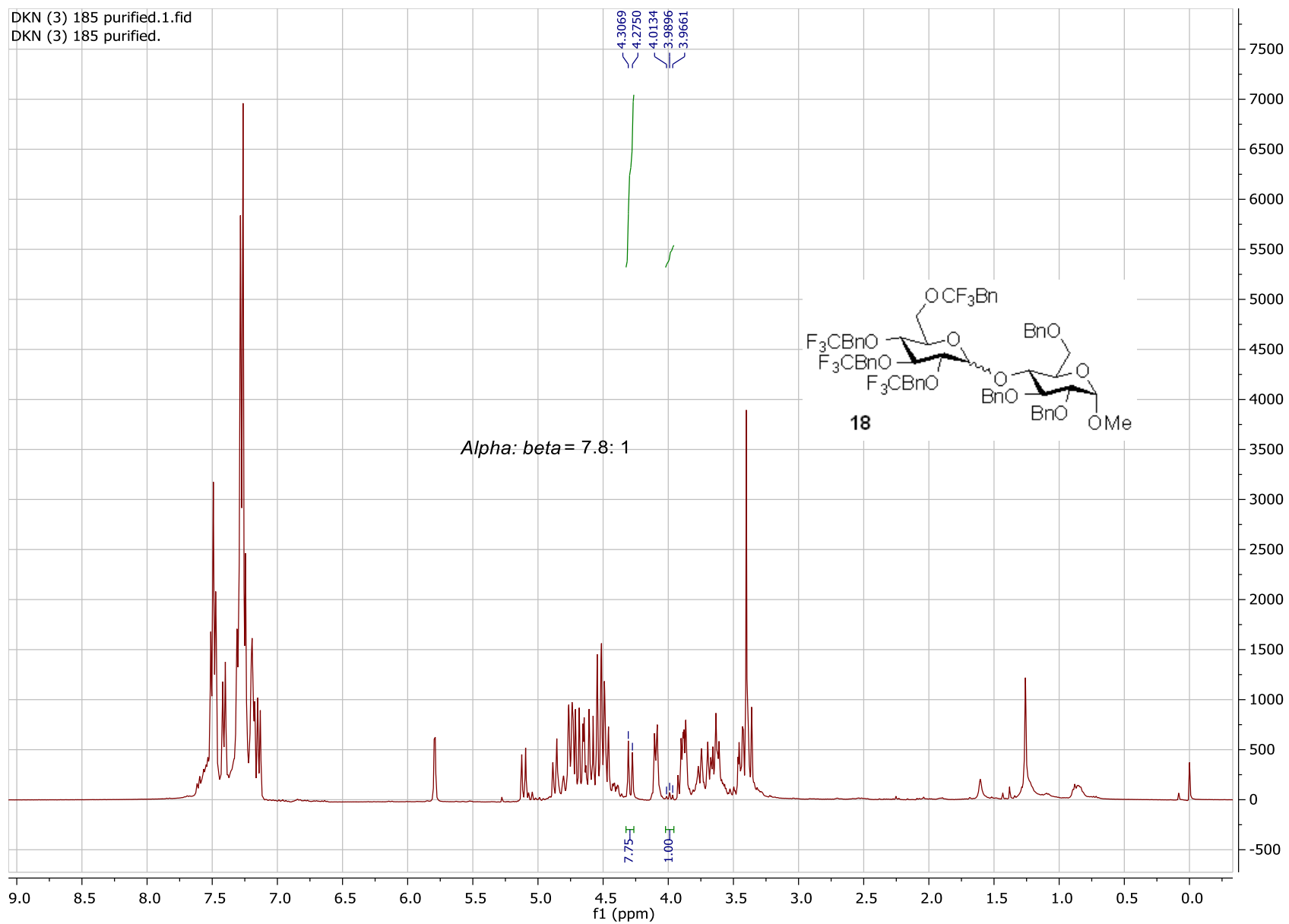
DKN (3) 180 A purified RXN in 5ML.1.fid
 DKN (3) 180 A purified RXN in 5ML



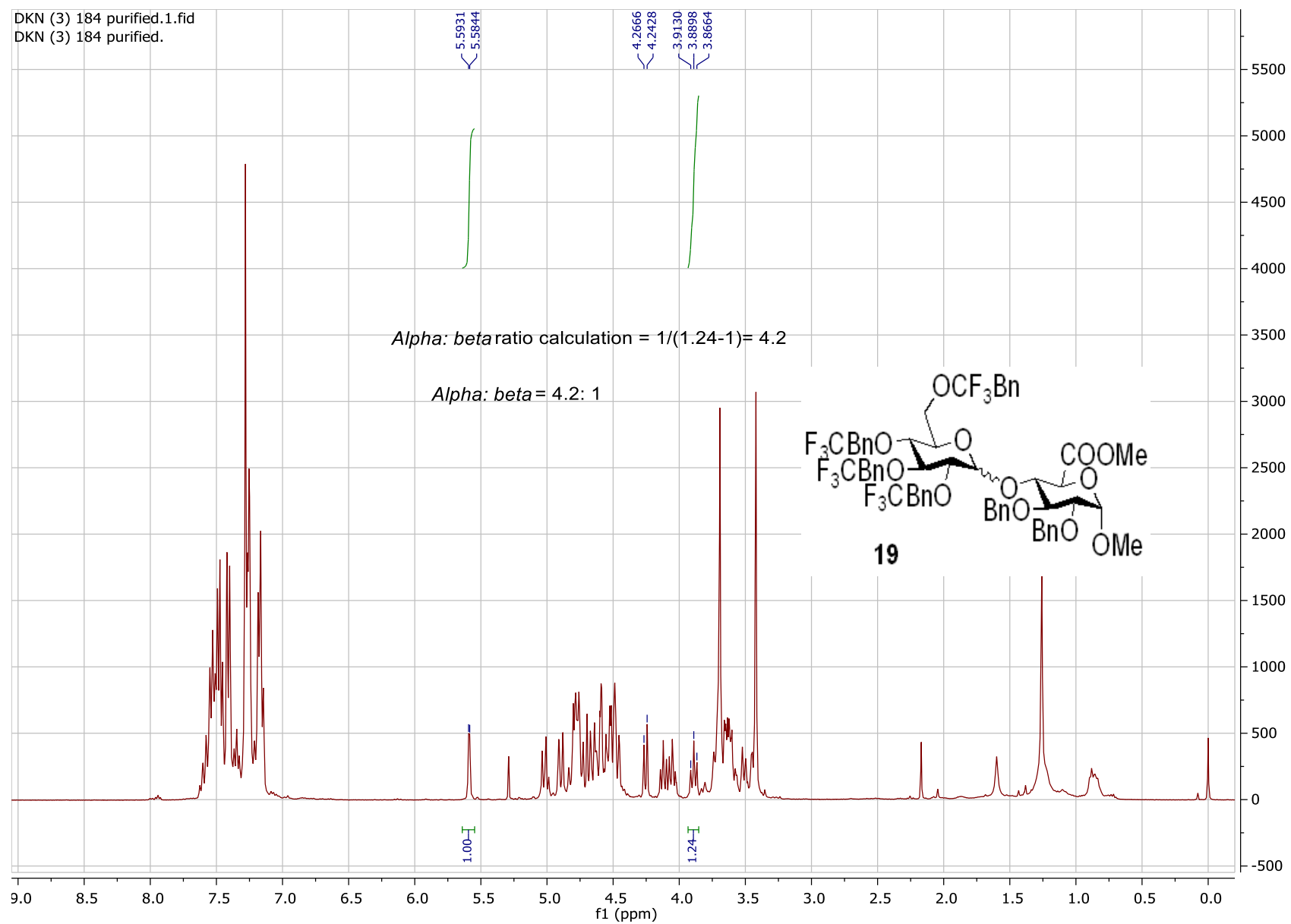
DKN (3) 183 purified.1.fid
DKN (3) 183 purified.



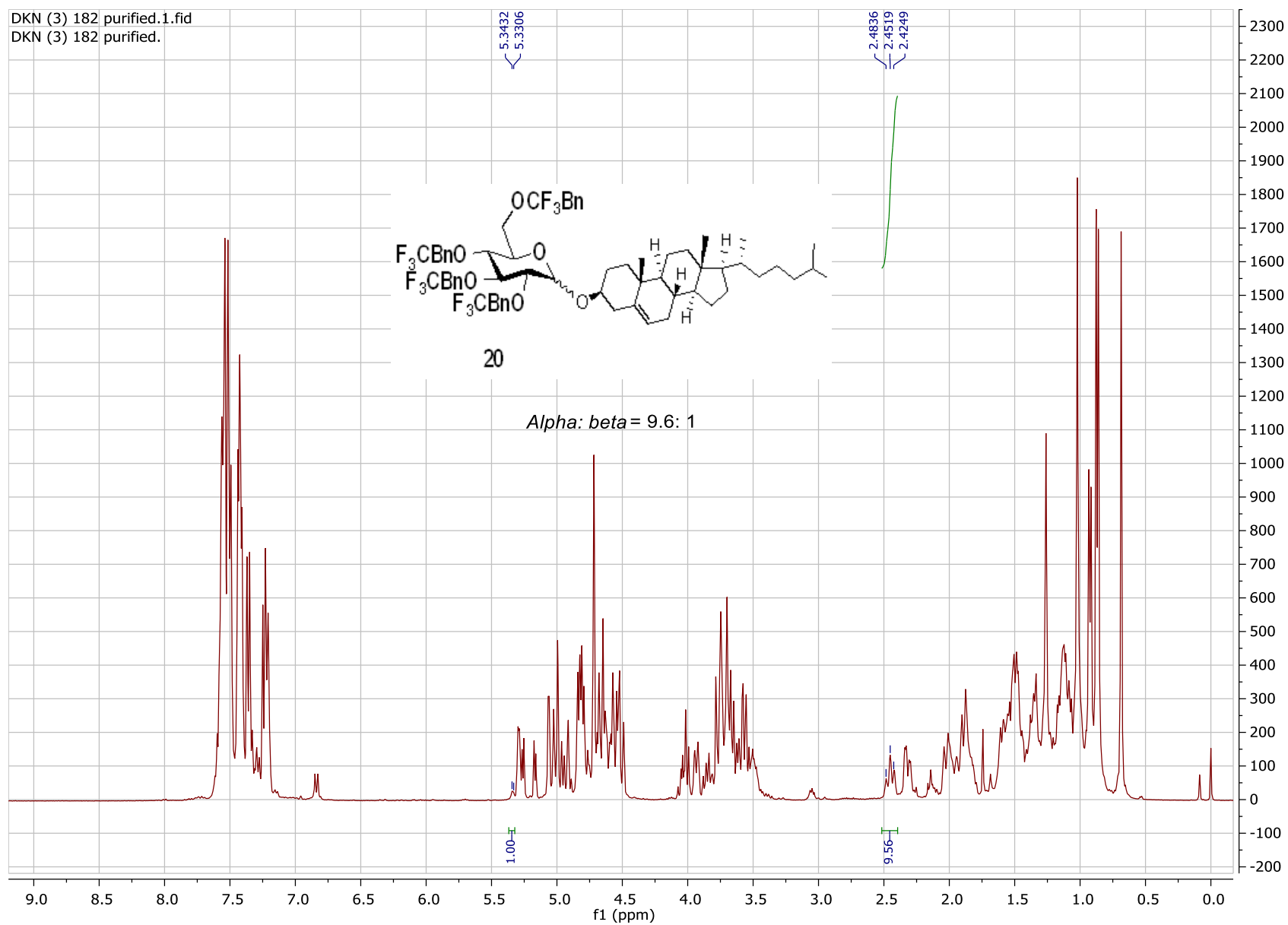
DKN (3) 185 purified.1.fid
DKN (3) 185 purified.



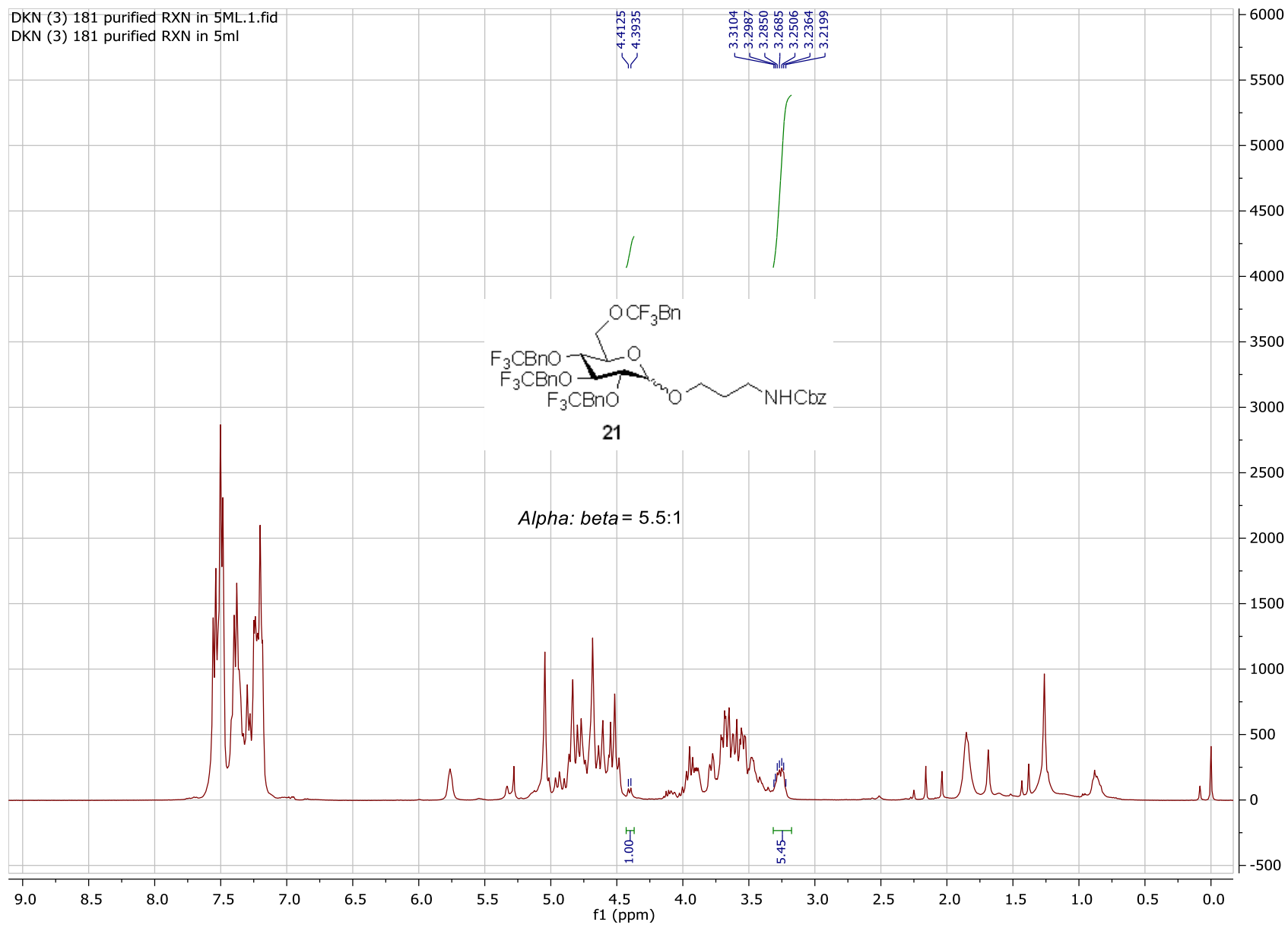
DKN (3) 184 purified.1.fid
DKN (3) 184 purified.



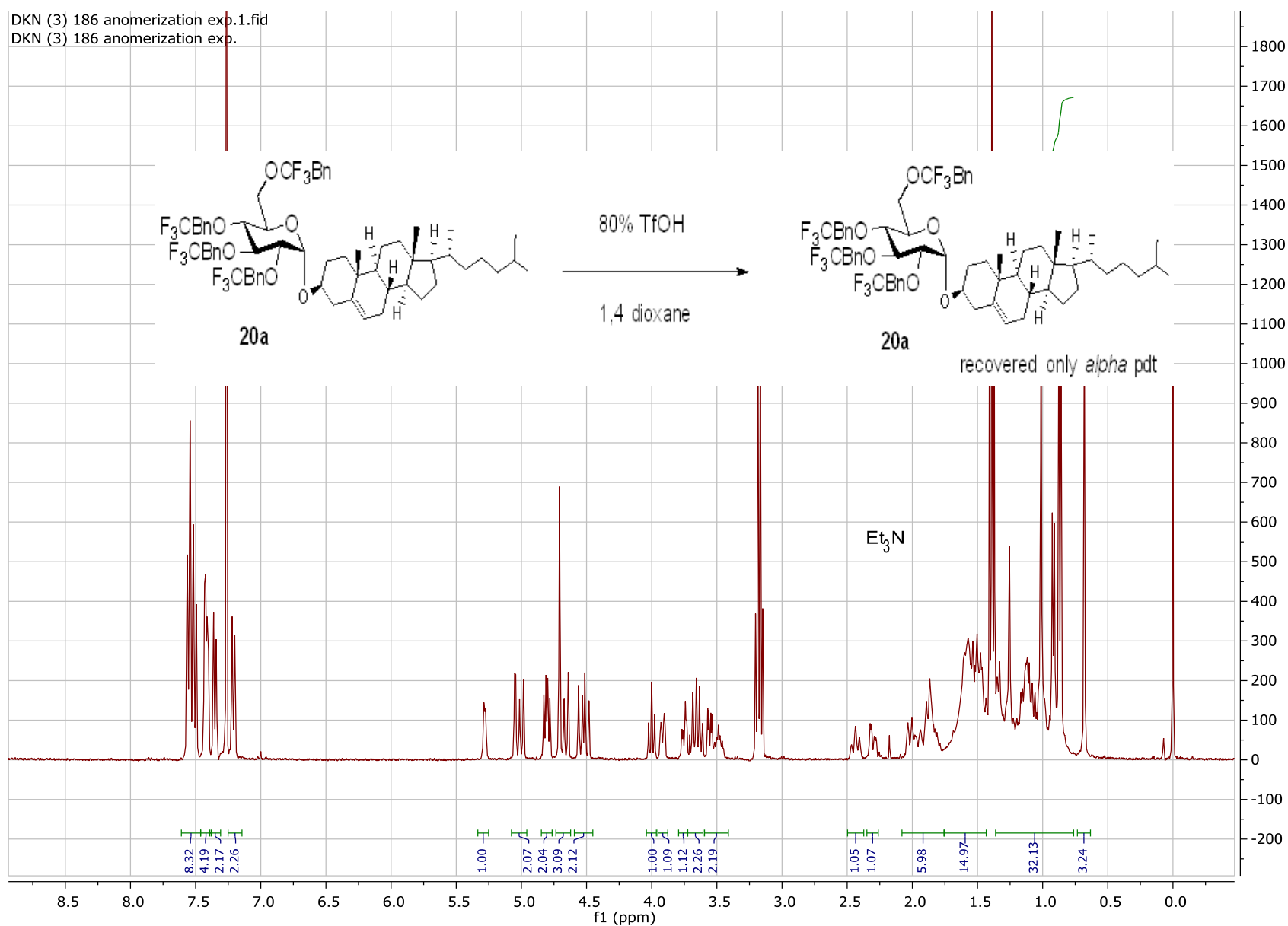
DKN (3) 182 purified.1.fid
DKN (3) 182 purified.



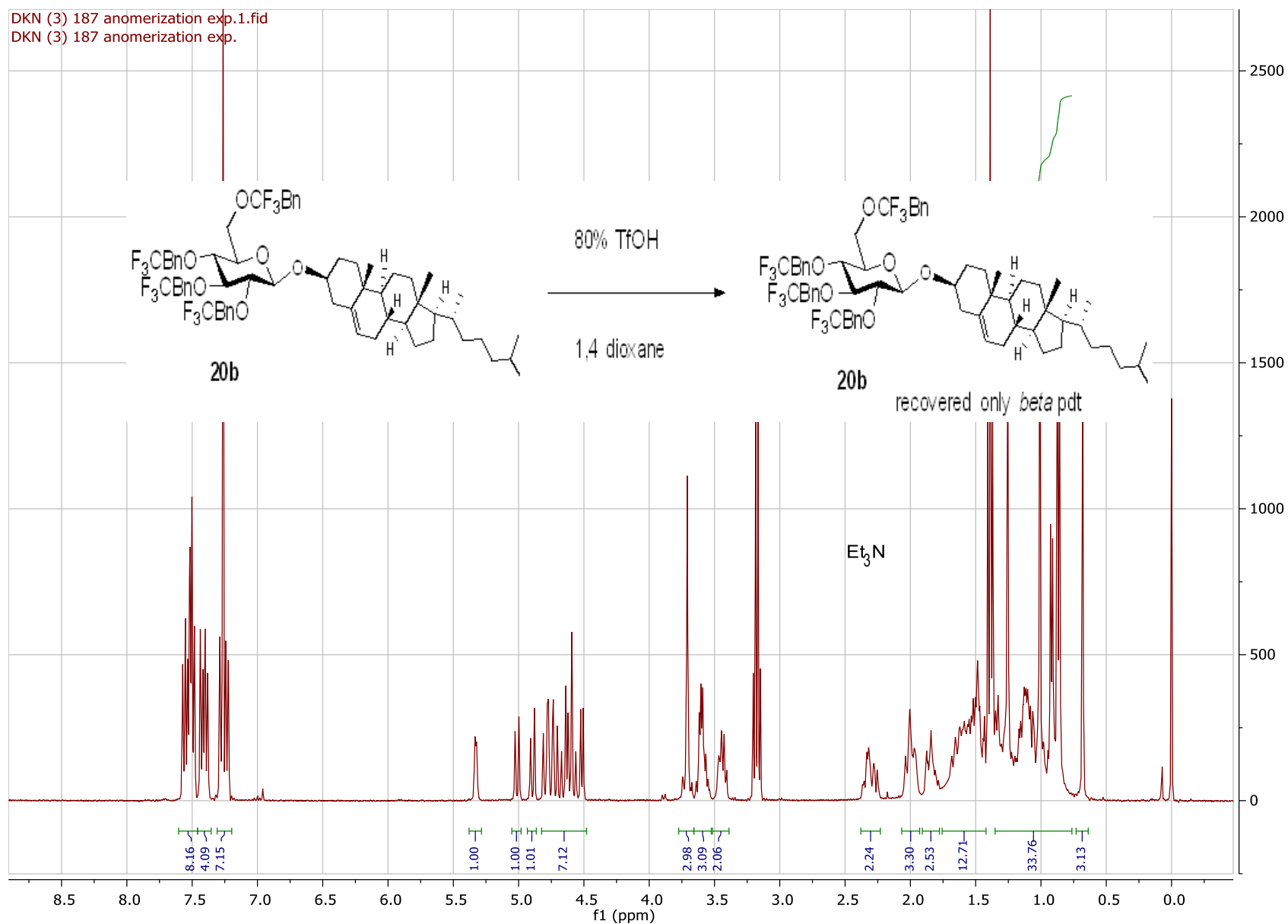
DKN (3) 181 purified RXN in 5ML.1.fid
DKN (3) 181 purified RXN in 5ml

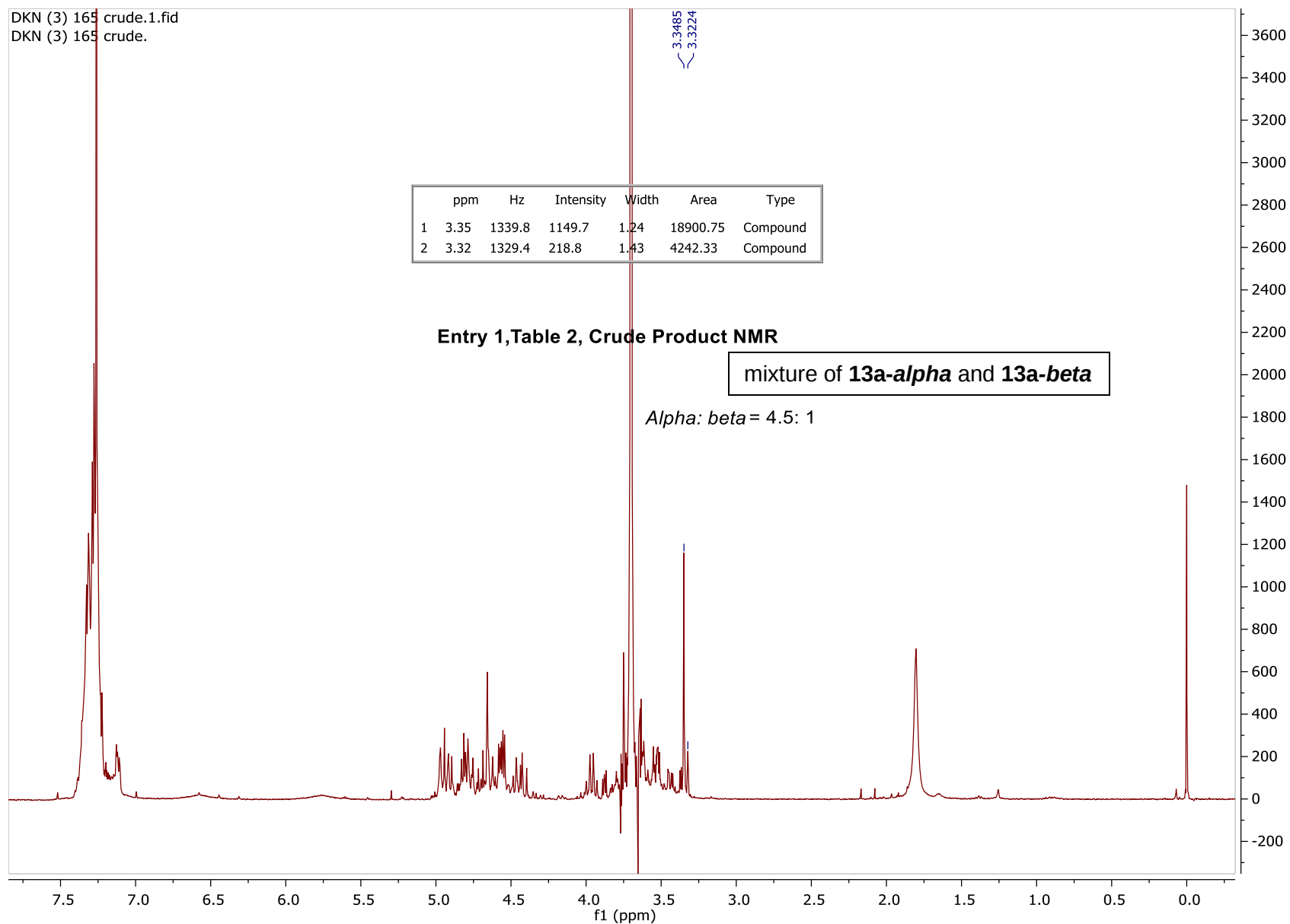


DKN (3) 186 anomerization exp.1.fid
 DKN (3) 186 anomerization exp.

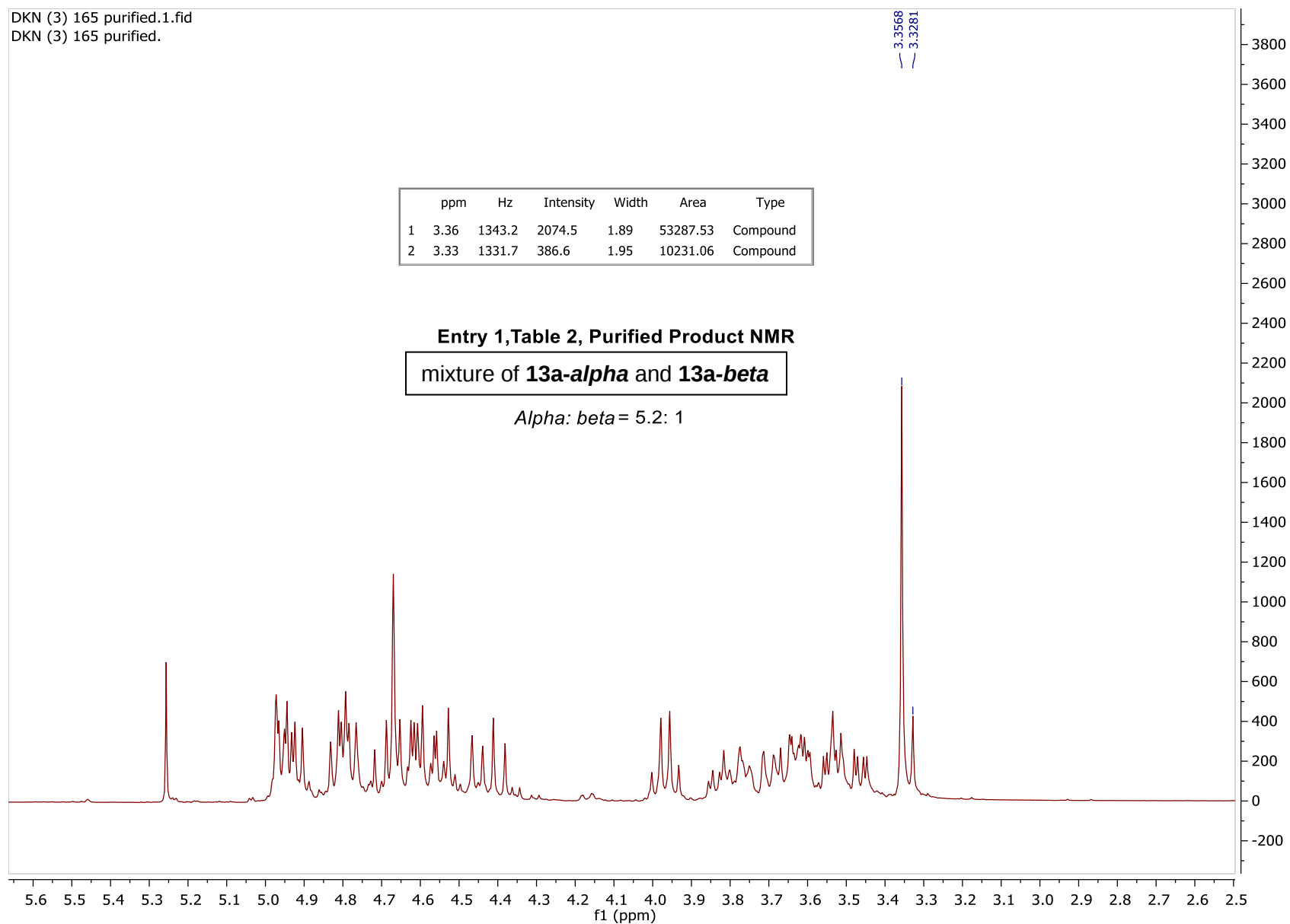


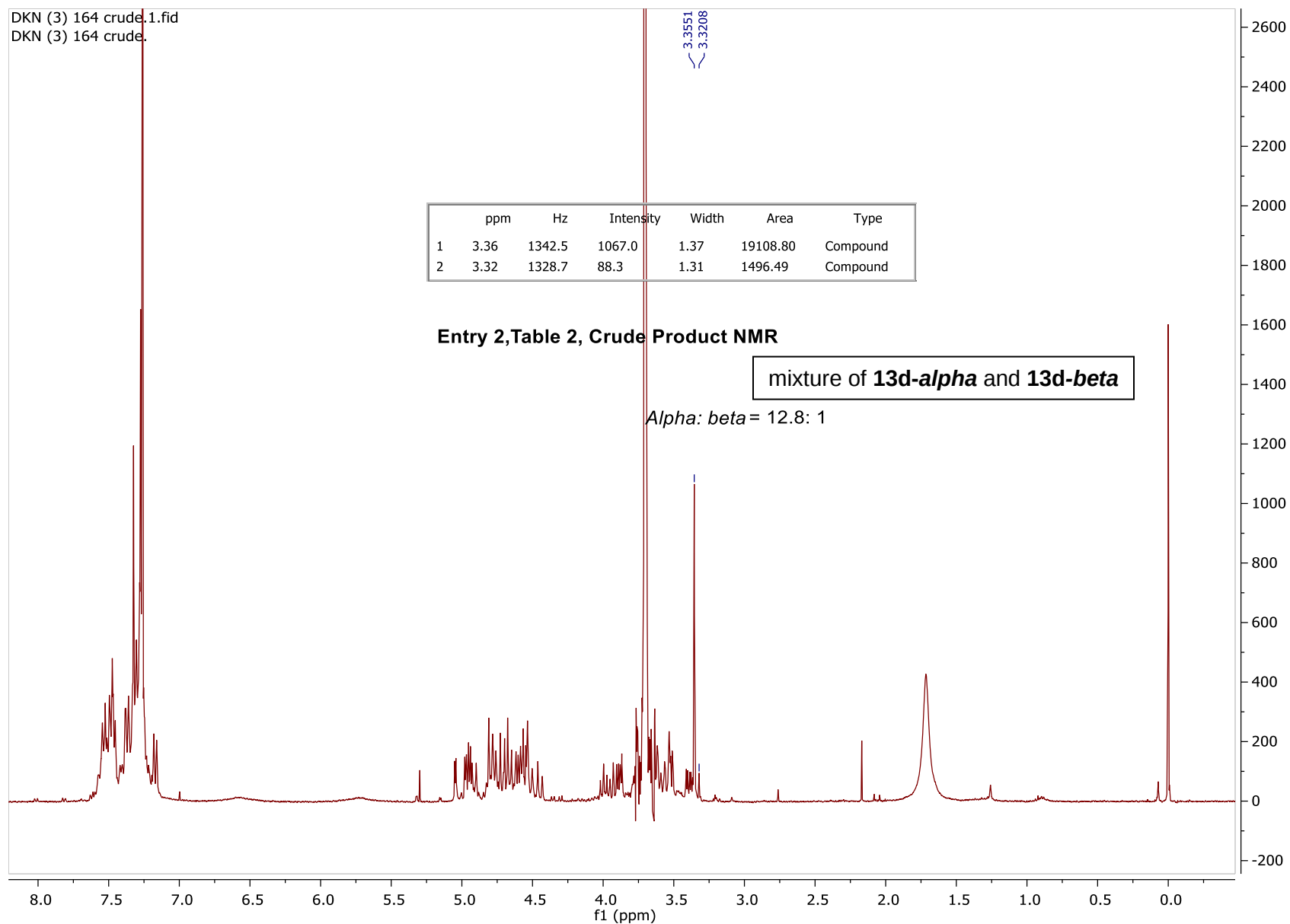
DKN (3) 187 anomerization exp.1.fid
DKN (3) 187 anomerization exp.





DKN (3) 165 purified.1.fid
DKN (3) 165 purified.





DKN (3) 164 purified.1.fid
DKN (3) 164 purified.

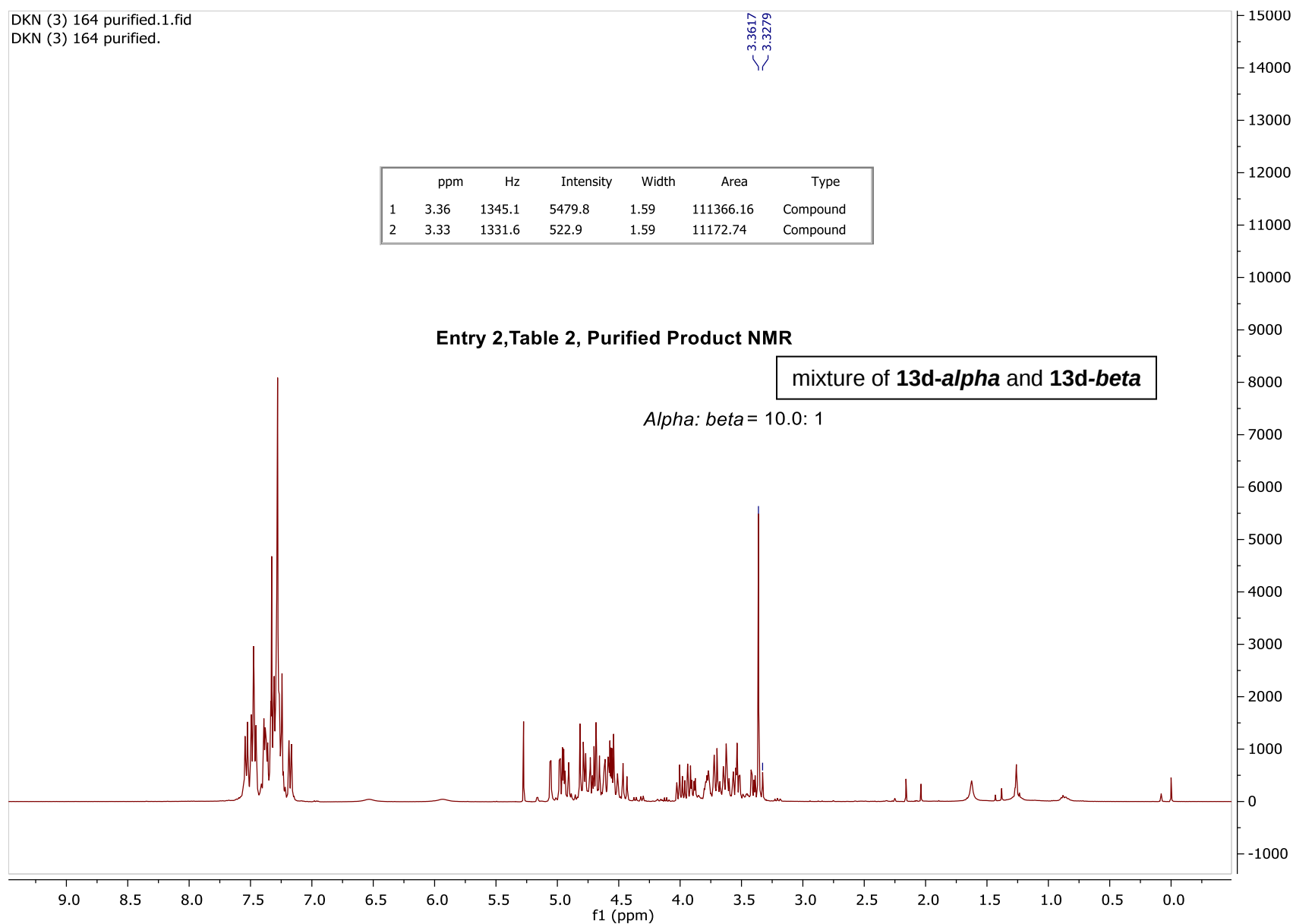
3.3617
3.3279

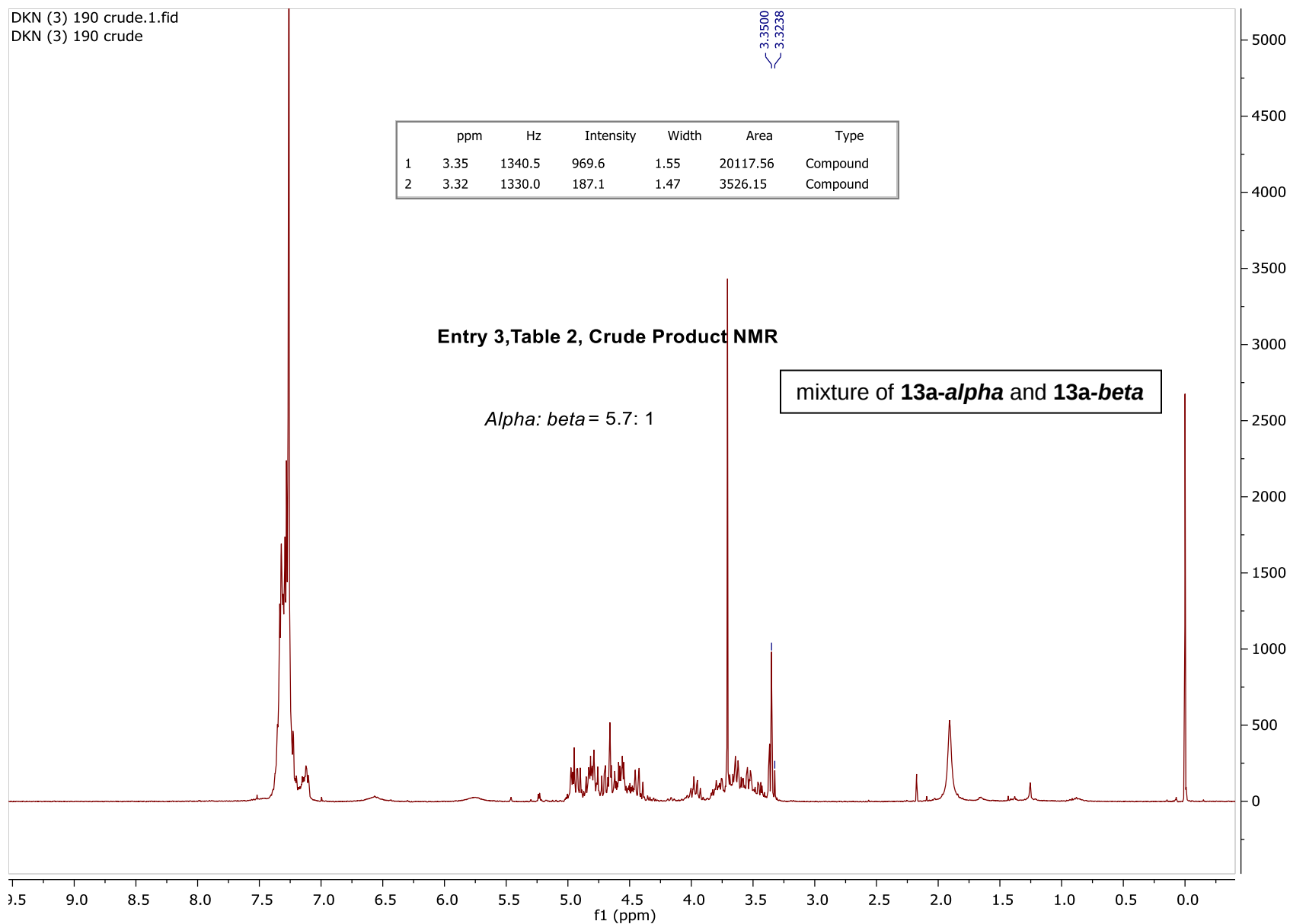
	ppm	Hz	Intensity	Width	Area	Type
1	3.36	1345.1	5479.8	1.59	111366.16	Compound
2	3.33	1331.6	522.9	1.59	11172.74	Compound

Entry 2, Table 2, Purified Product NMR

mixture of **13d-*alpha*** and **13d-*beta***

Alpha: beta = 10.0: 1





DKN (3) 190 purif 500.3.fid
DKN (3) 190 purif 500

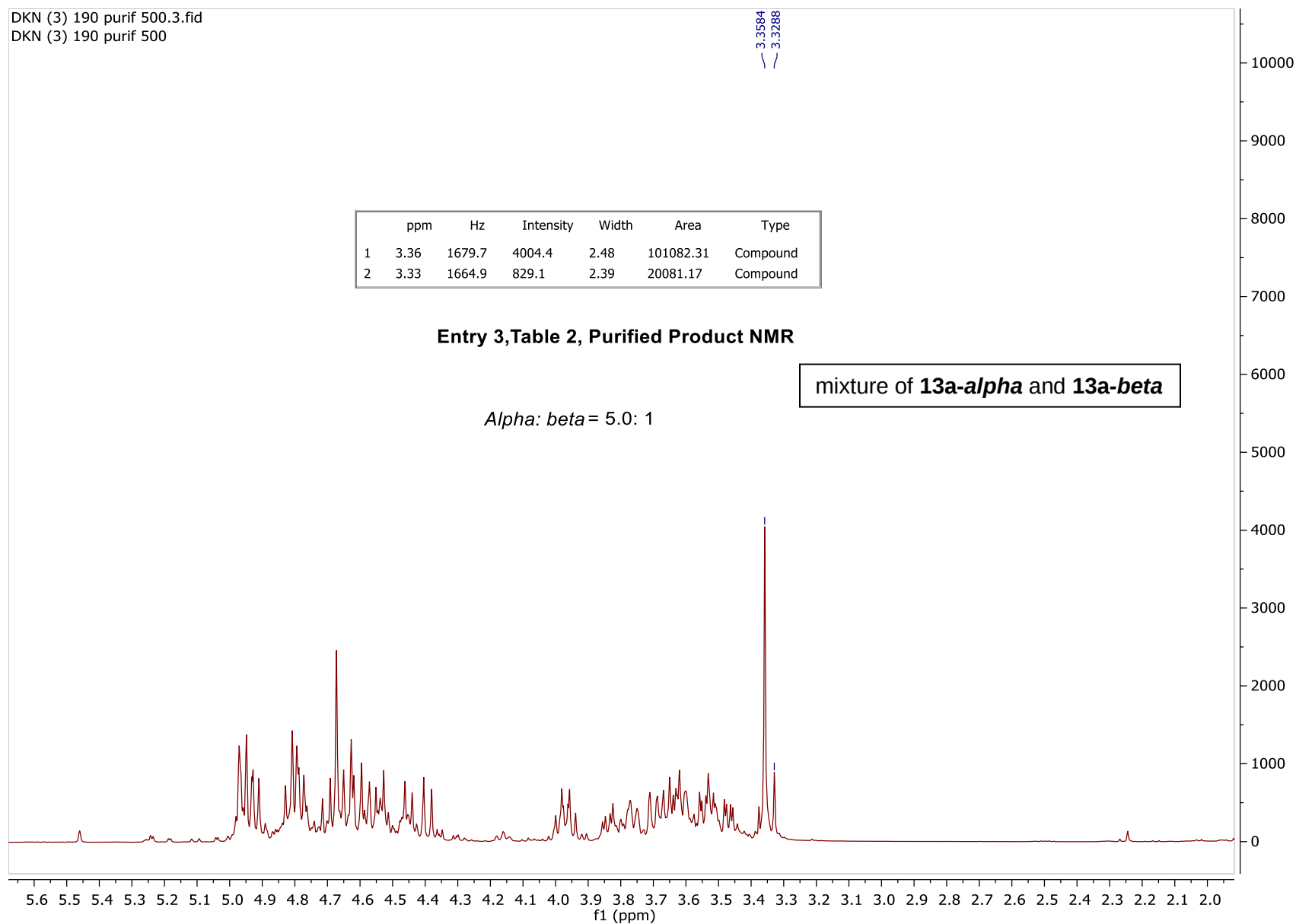
3.3584
3.3288

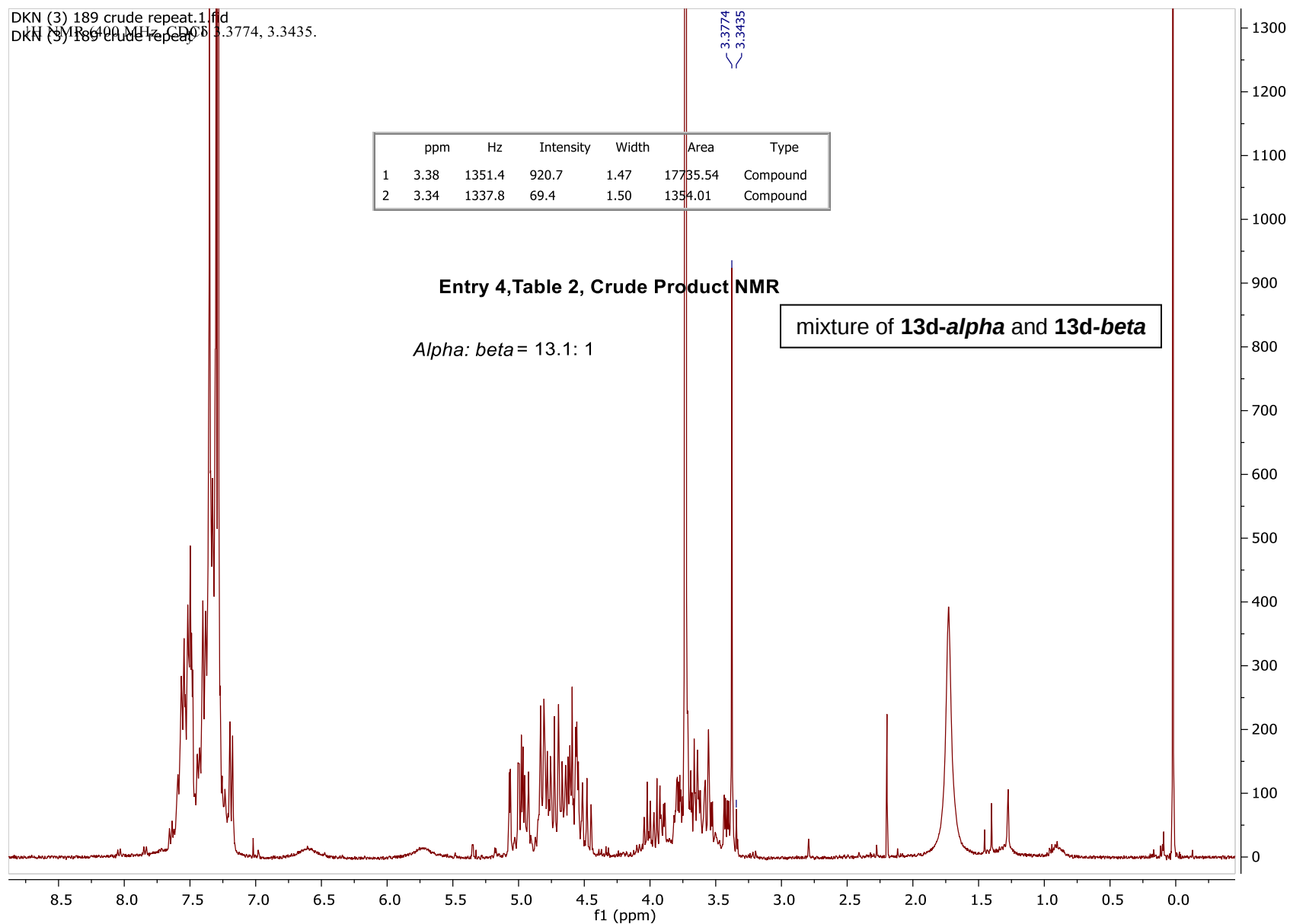
	ppm	Hz	Intensity	Width	Area	Type
1	3.36	1679.7	4004.4	2.48	101082.31	Compound
2	3.33	1664.9	829.1	2.39	20081.17	Compound

Entry 3, Table 2, Purified Product NMR

Alpha: beta = 5.0: 1

mixture of **13a-alpha** and **13a-beta**





DKN (3) 189 purif 500.3.fid
DKN (3) 189 purif 500

