Electronic Supplementary Information

C-Glycosylation enabled by N-(glycosyloxy)acetamides

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1. Experimental Procedures

1.1 General methods

Unless otherwise noted, all commercial reagents were used without further purification. Solvents were dried and redistilled commonly before using. Reactions were monitored by thin layer chromatography (TLC) on silica gel-coated plates (60 F₂₅₄). The spots were visualized under UV light (254 nm) and charring with a solution of (NH₄)₆Mo₇O₂₄•4H₂O (19.4 mmol, 24.00 g) and Ce(NH₄)₂(NO₃)₆ (0.90 mmol, 0.50 g) in H₂SO₄ (5%, 500 mL). Silica gel (200 - 300 mesh) was used for flash column chromatography. ¹H and ¹³C NMR spectra were recorded with Bruker AM 400 MHz spectrometer at room temperature. ¹H NMR spectra were reported using tetramethylsilane as the internal standard ($\delta = 0$) in CDCl₃, ¹³C NMR spectra were reported using tetramethylsilane as the internal standard ($\delta = 77.16$) of CDCl₃. Optical rotation values were obtained using Hanon P850 instrument. HRMS were obtained on a Waters Xevo G2 Q-TOF mass spectrometer.

1.2 Optimization of the reaction

Table S1. Solvent screening^{*a*, *b*}

< OI	Bn			~()Bn
BnO BnO	OBn NHAc +	TMS	Cu(OTf) ₂ (4.0 equit Solvent, 30 °C	^{v)} → BnO BnO	-O OBn
1a		2a	4A MS		3a
	Entry	Solvent		Yield (%)	
	1	DCM		10%	
	2	DCE		trace	
	3	Ether		10%	
	4	CH ₃ CN		0	
	5	CH ₃ NO ₂		40%	
	6	CHCl ₃		35%	
	7^c	CHCl ₃		45%	
	8^d	CHCl ₃		25%	
	9 ^c	$CHCl_3: CH_3NO_2=2:1$		37%	
	10^{c}	CHCl ₃ : CH ₃ NO ₂ = 1: 1		75%	
	11^{c}	CHCl ₃ : C	$H_3NO_2 = 1:2$	50%	

^{*a*} Reaction conditions: **1a** (30.0 mg, 0.05 mmol), **2a** (15.9 μL, 0.1 mmol), 4Å MS (100 mg) and solvent (1.0 mL). ^{*b*} Isolated yield. ^{*c*} 50 °C. ^{*d*} 80 °C, microwave (100 w).

Table S2. Additive screening^{*a*, *b*}

BnO BnO 1a	Bn OBn ^{NHAc} +	TMS 2a	Cu(OTf) ₂ (4.0 ec CHCl ₃ / CH ₃ NO ₂ , Additive 4Å MS	quiv) 50 °C BnO	OBn OBn 3a
	Entry	Additi	ve (equiv.)	Yield (%)	
	1	K ₂ CO ₃ (1.0)		35%	
	2	BF ₃	Et ₂ O (0.2)	32%	
	3	TBS	OTf (0.2)	41%	
	4	Cs	F (1.0)	42%	
	5	TBA	AF (1.0)	63%	

^{*a*} Reaction conditions: **1a** (30.0 mg, 0.05 mmol), **2a** (15.9 μL, 0.1 mmol), 4Å MS (100 mg) and solvent (1.0 mL). ^{*b*} Isolated yield.

1.3 Preparation of N-(glycosyloxy) acetamides



Phthalimidyl 2,3,4,6-tetra-O-benzyl-D-glucopyranoside (S2):

To a solution of compound **S1** (1.90 mmol, 1.03 g), *N*-hydroxyphthalimide (2.30 mmol, 0.37 g) and PPh₃ (2.30 mmol, 0.60 g) in dry THF (10 mL) was added diethyl azodicarboxylate (2.30 mmol, 0.5 mL) dropwise at 0 °C under an argon atmosphere. Then the reaction mixture was warmed up to room temperature and stirred overnight. Saturated NaHCO₃ solution was added to quench the reaction. The aqueous layer was extracted with ethyl acetate for three times, then the combined organic layer was washed with saturated NaCl solution, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 4: 1) to give **S2** as a white solid (0.91 g, 70%, α/β = 1: 3). ¹H NMR (400 MHz, CDCl₃) δ 7.89 - 7.80 (m, 7.55H), 7.77 - 7.70 (m, 7.57H), 7.53 (d, *J* = 7.0 Hz, 2.27H), 7.49 (d, *J* = 6.8 Hz, 5.76H), 7.40 - 7.23 (m, 59.29H), 7.21 - 7.13 (m, 8.92H), 5.62 (d, *J* = 3.9 Hz, 1H), 5.18 (d, *J* = 10.6 Hz, 2.79H), 5.14 - 5.07 (m, 3.81H), 5.03 (d, *J* = 10.9 Hz, 1.08H), 4.94 (d, *J* = 11.0 Hz, 2.84H), 4.89 - 4.70 (m, 12.70H), 4.62 - 4.51 (m, 10.41H), 4.40 (d, *J* = 12.0 Hz, 1.07H), 4.13 (t, *J* = 9.5 Hz, 1.07H), 3.90 (dd, *J* = 11.0, 2.5 Hz, 1.10H), 3.84 - 3.61 (m, 17.21H), 3.57 - 3.48 (m, 2.86H). The spectroscopic data coincide with the previous report.¹

Acetamidyl 2,3,4,6-tetra-O-benzyl-D-glucopyranoside (1a):

Compound **S2** (0.24 mmol, 0.17 g) was dissolved in methanol (10 mL) and 50% hydrazine hydrate (1.40 mmol, 84 μ L) was added dropwise. After stirring at room temperature for 1 h, the mixture was concentrated in vacuo. Then the residue was dissolved in mixed solvent (CH₂Cl₂/1M K₂CO₃ solution, 1: 1, 16 mL), acetyl chloride (3.70 mmol, 266 μ L) was added to the mixture

slowly at 0 °C. After stirring at room temperature for 2 h, the mixture was extracted with CH₂Cl₂ for three times and the combined organic layer was washed with water, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 2: 1) to give **1a** as yellow oil (0.12 g, 84% yield over 2 steps, $\alpha/\beta = 1$: 2). ¹H NMR (400 MHz,CDCl₃) δ 8.95 (s, 0.88H), 8.42 (s, 0.56H), 7.53 - 7.24 (m, 29.05H), 7.19 - 7.09 (m, 3.52H), 5.12 (s, 0.9H), 4.89 (d, *J* = 11.0 Hz, 2.11H), 4.81 (d, *J* = 10.9 Hz, 2.65H), 4.77 - 4.61 (m, 3.27H), 4.59 - 4.44 (m, 4.89H), 3.80 - 3.65 (m, 6.54H), 3.56 (s, 3.4H), 2.11 (s, 1.63H), 1.78 (s, 3H). The spectroscopic data coincide with the previous report.¹



Phthalimidyl 2-deoxy-3,4,6-tri-O-benzyl-D-glucopyranoside (S4):

The similar procedure for synthesizing **S2** was applied to deliver **S4** as yellow oil (0.72 g, 65%, $\alpha/\beta = 1: 2$). ¹H NMR (400 MHz, CDCl₃) δ 7.78 - 7.71 (m, 6H), 7.68 - 7.63 (m, 6H), 7.29 - 7.12 (m, 45H), 5.48 (d, *J* = 3.8 Hz, 1H), 5.06 (d, *J* = 8.4 Hz, 2H), 4.85 - 4.80 (m, 3H), 4.68 - 4.61 (m, 4H), 4.57 - 4.45 (m, 11H), 4.35 (d, *J* = 12.1 Hz, 1H), 4.06 - 3.96 (m, 1H), 3.84 (dd, *J* = 10.9, 2.4 Hz, 1H), 3.72 - 3.60 (m, 7H), 3.55 (t, *J* = 8.8 Hz, 3H), 3.43 - 3.37(m, 2H), 2.67 - 2.55 (m, 3H), 1.89 - 1.79 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 163.53, 163.28, 138.62, 138.59, 138.54, 138.23, 138.17, 138.15, 134.61, 134.54, 129.07, 129.04, 128.60, 128.55, 128.50, 128.42, 128.36, 128.20, 128.06, 127.95, 127.89, 127.83, 127.77, 127.73, 127.69, 127.65, 127.46, 123.76, 123.63, 103.78, 103.50, 78.78, 77.65, 76.71, 76.37, 75.09, 75.05, 73.65, 73.56, 72.95, 72.19, 71.71, 69.27, 68.39, 34.00, 33.49. HRMS (ESI) calcd for C₃₅H₃₇N₂O₇ [M+NH₄]⁺ 597.2595, found 597.2598.

Acetamidyl 2-deoxy-3,4,6-tri-O-benzyl-D-glucopyranoside (1b):

Glycosyl donor **1b** was obtained according to the similar procedure for the synthesis of **1a** as yellow oil (0.09 g, 80% yield over 2 steps, $\alpha/\beta = 1$: 2). ¹H NMR (400 MHz, CDCl₃) δ 9.07 (s, 0.65H), 8.82 (s, 0.28H), 7.36 - 7.25 (m, 16.14H), 7.22 - 7.16 (m, 2.60H), 5.20 (s, 0.33H), 4.91 - 4.79 (m, 1.91H), 4.67 (t, J = 11.3 Hz, 1.41H), 4.62 - 4.45 (m, 5.11H), 3.94 (s, 0.66H), 3.77 (dd, J = 10.6, 4.6 Hz, 1.01H), 3.72 - 3.64 (m, 2.42H), 3.58 (t, J = 8.5 Hz, 1.11H), 3.52 (s, 1.21H), 2.55 (s, 1H), 2.35 (s, 0.29H), 2.12 (s, 1.07H), 1.85 (s, 1.11H), 1.80 (s, 2.03H). ¹³C NMR (101 MHz, CDCl₃) δ 167.54, 138.30, 138.23, 138.14, 138.00, 137.83, 137.78, 137.70, 128.47, 128.41, 128.39, 128.35, 127.98, 127.91, 127.83, 127.77, 127.74, 127.70, 127.64, 102.47, 101.97, 78.08, 77.73, 74.88, 74.75, 73.58, 73.44, 72.27, 71.75, 71.41, 69.19, 33.32, 33.16, 19.87. HRMS (ESI) calcd for C₂₉H₃₇N₂O₆ [M+NH₄]⁺ 509.2646, found 509.2652.



Phthalimidyl 2-deoxy-3,4,6-tri-O-benzyl-a-D-galactopyranoside (S6):

The similar procedure for synthesizing **S2** was applied to deliver **S6** as yellow oil (0.80 g, 73%, α only). [α]_D²⁵ +116.5 (*c* 1.10, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.81 - 7.77 (m, 2H), 7.75 - 7.70 (m, 2H), 7.37 - 7.25 (m, 15H), 5.59 (d, *J* = 2.5 Hz, 1H), 4.94 (d, *J* = 11.4 Hz, 1H), 4.79 (t, *J* = 6.7 Hz, 1H), 4.68 - 4.61 (m, 3H), 4.54 (d, *J* = 11.9 Hz, 1H), 4.48 (d, *J* = 11.9 Hz, 1H), 4.15 - 4.05 (m, 2H), 3.65 (dd, *J* = 9.2, 7.8 Hz, 1H), 3.54 (dd, *J* = 9.4, 5.8 Hz, 1H), 2.42 - 2.38 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 163.57, 138.75, 138.32, 138.26, 134.39, 129.02, 128.49, 128.28, 128.26, 128.18, 127.65, 127.57, 127.49, 127.35, 123.53, 103.89, 74.52, 73.67, 73.20, 72.60, 71.69, 70.54, 68.68, 28.82. HRMS (ESI) calcd for C₃₅H₃₇N₂O₇ [M+NH₄]⁺ 597.2595, found 597.2603.

Acetamidyl 2-deoxy-3,4,6-tri-*O*-benzyl-α-D-galactopyranoside (1c):

Glycosyl donor **1c** was obtained according to the similar procedure for the synthesis of **1a** as yellow oil (0.10 g, 85% yield over 2 steps, α only). [α]_D²⁵ +139.3 (*c* 0.30, CHCl₃). ¹H NMR (400 MHz,CDCl₃) δ 8.59 (s, 1H), 7.39 - 7.19 (m, 15H), 5.23 (s, 1H), 4.92 (d, *J* = 11.6 Hz, 1H), 4.62 - 4.55 (m, 3H), 4.50 (d, *J* = 11.8 Hz, 1H), 4.41 (d, *J* = 11.8 Hz, 1H), 4.04 (s, 1H), 3.85 (s, 2H), 3.67 - 3.57 (m, 1H), 3.53 - 3.44 (m, 1H), 2.34 - 2.21 (m, 2H), 1.83 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 167.60, 138.54, 138.22, 137.82, 128.48, 128.32, 127.91, 127.89, 127.72, 127.69, 127.35, 102.46, 74.31, 73.93, 73.56, 72.73, 71.69, 70.55, 70.04, 28.82, 19.91. HRMS (ESI) calcd for C₂₉H₃₇N₂O₆ [M+NH₄]⁺ 509.2646, found 509.2649.



Acetamidyl 2,3,5-tri-*O*-benzyl-α-D-arabinofuranoside (1d):

To a solution of compound $S7^2$ (1.90 mmol, 0.80 g), *N*-hydroxyphthalimide (2.30 mmol, 0.37 g) and PPh₃ (2.30 mmol, 0.60 g) in dry THF (10 mL) was added diethyl azodicarboxylate (2.30 mmol, 0.5 mL) dropwise at 0 °C under an argon atmosphere. Then the reaction mixture was warmed up to room temperature and stirred overnight. Saturated NaHCO₃ solution was added to quench the reaction. The aqueous layer was extracted with ethyl acetate for three times, then the combined organic layer was washed with saturated NaCl solution, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 5: 1) to give S8. Compound S8 (1.23 mmol, 0.70 g) was dissolved in methanol (20 mL) and 50% hydrazine hydrate (7.20 mmol, 504 µL) was added dropwise. After stirring at room temperature for 1 h, the mixture was concentrated in vacuo. Then the residue was dissolved in mixed solvent (CH₂Cl₂/1M K₂CO₃ solution, 1: 1, 30 mL), acetyl chloride (22.20 mmol, 1.60 mL) was added to the mixture slowly at 0 °C. After stirring at room temperature for 2 h, the mixture was extracted with CH₂Cl₂ for three times and the combined organic layer was washed with water, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 1: 1) to give 1d as yellow oil (0.44 g, 48% yield over 3 steps, $\alpha/\beta = 1$: 1.5). ¹H NMR (400 MHz, CDCl₃) & 8.53 (s, 0.97H), 8.23 (s, 1.57H), 7.65 - 7.51 (m, 1.07H), 7.48 - 7.30 (m, 36.53H), 7.22 (d, J = 7.1 Hz, 2.03H), 5.40 (s, 1.00H), 5.34 (s, 1.53H), 4.96 (d, J = 9.8 Hz, 1.38H), 4.75 (d, J = 11.7Hz, 2.82H), 4.66 (d, J = 11.1 Hz, 3.33H), 4.62 - 4.41 (m, 12.62H), 4.33 - 4.22 (m, 3.28H), 4.16 (s, 3.25H, 4.11 - 4.03 (m, 2.36H), 3.90 (d, J = 7.5 Hz, 1.07H), 3.71 - 3.55 (m, 4.24H), 3.54 - 3.44 (m,

2.07H), 2.18 - 1.84 (m, 5.83H). ¹³C NMR (101 MHz, CDCl₃) δ 167.55, 167.29, 137.93, 137.81, 137.61, 137.47, 137.15, 130.03, 129.96, 129.91, 129.84, 129.78, 128.60, 128.51, 128.48, 128.43, 128.33, 128.07, 127.96, 127.92, 127.85, 127.78, 110.09, 102.75, 85.62, 83.34, 83.01, 81.46, 80.60, 80.34, 73.54, 73.41, 72.46, 72.34, 72.21, 70.44, 69.80, 19.95, 19.86. HRMS (ESI) calcd for C₂₈H₃₂NO₆ [M+H]⁺ 478.2224, found 478.2230.

1.4 C-Glycosylation procedure

To a solution of glycosyl donor (0.05 mmol), *C*-nucleophile (0.1 mmol), activated 4 Å MS (100 mg), dry CHCl₃ (0.5 mL) and CH₃NO₂ (0.5 mL) was added SnBr₄ (0.075 mmol) at 0 °C under an argon atmosphere. Then the reaction mixture was warmed up to room temperature and stirred for 0.5 h. Then triethylamine (15.0 μ L) was added, the reaction mixture was filtered through a pad of celite and the filtrate was concentrated in vacuo. The residue was purified by column chromatography on silica gel to deliver the desired product.

1.5 Characterization of the *C***-glycosides Compound 3a**



C-Glycoside **3a** was obtained (26.0 mg, 95%, $\alpha/\beta = 5$: 1) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz,CDCl₃) δ 7.37 - 7.22 (m, 21.47H), 7.20 - 7.16 (m, 0.40H), 7.15 - 7.09 (m, 2.07H), 6.01 - 5.87 (m, 0.23H), 5.86 - 5.73 (m, 1.00H), 5.16 - 5.02 (m, 2.39H), 4.96 - 4.85 (m, 1.57H), 4.81 (dd, J = 10.8, 3.2 Hz, 2.16H), 4.71 - 4.59 (m, 3.46H), 4.58 - 4.54 (m, 0.30H), 4.50 - 4.42 (m, 2.03H), 4.17 - 4.07 (m, 1.00H), 3.84 - 3.56 (m, 6.87H), 3.45 - 3.38 (m, 0.20H), 3.36 - 3.30 (m, 0.31H), 2.64 - 2.55 (m, 0.23H), 2.54 - 2.41 (m, 1.97H), 2.37 - 2.27 (m, 0.21H). The spectroscopic data coincide with the previous report.³

Compound 3b



C-Glycoside **3b** was obtained (25.8 mg, 83%, $\alpha/\beta = 3$: 1) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 7.39 - 7.21 (m, 28.94H), 7.21 - 7.16 (m, 0.84H), 7.16 - 7.09 (m, 2.23H), 4.96 - 4.87 (m, 2.74H), 4.85 - 4.74 (m, 2.39H), 4.66 - 4.52 (m, 4.06H), 4.51 - 4.43 (m, 2.36H), 3.83 (d, *J* = 8.6 Hz, 0.24H), 3.77 (dd, *J* = 9.0, 6.0 Hz, 1.20H), 3.73 - 3.64 (m, 2.99H), 3.64 - 3.55 (m, 3.37H), 3.40 (d, *J* = 10.1 Hz, 0.31H), 3.34 (t, *J* = 9.1 Hz, 0.30H), 2.89 (dd, *J* = 16.8, 5.4 Hz, 1H), 2.76 (dd, *J* = 16.8, 7.0 Hz, 1H), 2.63 (dd, *J* = 16.5, 9.1 Hz, 0.30H), 2.50 (d, *J* = 14.8 Hz, 0.30H), 1.08 (s, 8.84H), 1.05 (s, 2.77H). ¹³C NMR (101 MHz, CDCl₃) δ 213.16, 212.10, 138.65, 138.61, 138.28, 138.23, 138.19, 138.07, 137.87, 128.46, 128.44, 128.40, 128.34, 128.24, 128.15, 128.07, 127.91, 127.89, 127.82, 127.80, 127.76, 127.70, 127.63, 127.55, 87.41, 82.08, 80.81, 79.30, 78.81, 78.47, 77.79, 75.56, 75.39, 75.30, 75.12, 74.93, 74.80, 73.57,

73.42, 73.11, 72.84, 70.34, 69.02, 68.77, 44.39, 44.18, 38.64, 33.48, 26.14, 26.01. HRMS (ESI) calcd for $C_{40}H_{50}NO_6$ [M+NH₄]⁺ 640.3633, found 640.3639.

Compound 3c



C-Glycoside **3c** was obtained (21.0 mg, 92%, $\alpha/\beta = 13$: 1) as a colorless oil according to the *C*-glycosylation procedure. The α isomer: ¹H NMR (400 MHz, CDCl₃) δ 7.34 - 7.25 (m, 13H), 7.23 - 7.17 (m, 2H), 5.82 - 5.70 (m, 1H), 5.06 - 5.01 (m, 2H), 4.78 (d, *J* = 11.1 Hz, 1H), 4.63 - 4.48 (m, 5H), 4.08 - 4.00 (m, 1H), 3.82 - 3.72 (m, 3H), 3.69 - 3.63 (m, 1H), 3.54 (t, *J* = 7.1 Hz, 1H), 2.49 - 2.40 (m, 1H), 2.26 - 2.18 (m, 1H), 2.03 - 1.96 (m, 1H), 1.81 - 1.71 (m, 1H). The spectroscopic data coincide with the previous report.⁴

Compound 3d



C-Glycoside **3d** was obtained (22.4 mg, 87%, $\alpha/\beta = 1.5$: 1) as a colorless oil according to the *C*-glycosylation procedure. The α isomer: $[\alpha]_D^{25}$ -1.0 (*c* 0.60, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.34 - 7.27 (m, 13H), 7.24 - 7.19 (m, 2H), 4.74 (d, *J* = 11.3 Hz, 1H), 4.62 - 4.49 (m, 6H), 3.79 - 3.69 (m, 4H), 3.51 (t, *J* = 6.4 Hz, 1H), 2.85 (dd, *J* = 17.0, 5.5 Hz, 1H), 2.66 (dd, *J* = 17.0, 7.9 Hz, 1H), 1.97 - 1.91 (m, 1H), 1.88 - 1.81 (m, 1H), 1.10 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 213.74, 138.74, 138.67, 138.40, 128.53, 128.46, 128.10, 127.98, 127.79, 127.71, 81.09, 78.98, 78.41, 75.10, 73.58, 72.13, 71.61, 69.52, 44.37, 42.74, 36.74, 26.26; HRMS (ESI) calcd for C₃₃H₄₄NO₅ [M+NH₄]⁺ 534.3214, found 534.3222. The β isomer: $[\alpha]_D^{25}$ +7.5 (*c* 0.80, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.36 - 7.25 (m, 13H), 7.20 - 7.14 (m, 2H), 4.89 (d, *J* = 10.8 Hz, 1H), 4.69 (d, *J* = 11.6 Hz, 1H), 4.63 - 4.48 (m, 4H), 3.92 - 3.82 (m, 1H), 3.74 - 3.62 (m, 3H), 3.50 (t, *J* = 9.1 Hz, 1H), 3.44 - 3.36 (m, 1H), 2.98 (dd, *J* = 17.2, 5.5 Hz, 1H), 2.59 (dd, *J* = 17.2, 7.2 Hz, 1H), 2.33 - 2.20 (m, 1H), 1.40 - 1.25 (m, 1H), 1.13 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 213.32, 138.46, 138.39, 128.53, 128.45, 128.13, 127.94, 127.85, 127.74, 127.68, 76.12, 75.75, 74.20, 73.86, 73.55, 71.15, 69.21, 66.68, 44.49, 39.90, 32.41, 26.27; HRMS (ESI) calcd for C₃₃H₄₄NO₅ [M+NH₄]⁺ 534.3214, found 534.3216.

Compound 3e



C-Glycoside **3e** was obtained (20.1 mg, 75%, $\alpha/\beta = 2$: 1) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 8.00 - 7.88 (m, 5.39H), 7.61 - 7.51 (m, 3.02H), 7.51 - 7.40 (m, 6.19H), 7.38 - 7.12 (m, 40.86H), 4.91 (d, *J* = 10.9 Hz, 1H), 4.78 - 4.46 (m, 16.54H), 4.10 - 3.99 (m, 1.09H), 3.89 - 3.87 (m, 1.70H), 3.84 - 3.65 (m, 7.87H), 3.60 - 3.50 (m, 2.67H), 3.51 - 3.42 (m, 2.12H), 3.39 (d, *J* = 5.9 Hz, 0.86H), 3.35 (d, *J* = 5.8 Hz, 1.04H), 3.16 - 3.01 (m, 2.73H), 2.40 (dd, *J* = 12.4, 3.8 Hz, 0.97H), 2.13 - 1.99 (m, 2.19H), 1.98 - 1.84 (m, 2.04H),

1.75 - 1.53 (m, 1.81H), 1.50 - 1.40 (m, 1.47H). ¹³C NMR (101 MHz, CDCl₃) δ 198.00, 197.94, 138.66, 138.41, 137.17, 137.10, 133.37, 128.79, 128.75, 128.52, 128.44, 128.34, 128.30, 128.09, 128.08, 127.93, 127.82, 127.81, 127.75, 127.73, 127.67, 81.04, 79.13, 78.37, 76.00, 75.85, 75.12, 74.14, 73.81, 73.57, 73.51, 72.19, 71.58, 71.26, 69.46, 69.08, 67.08, 42.04, 36.98, 32.61. HRMS (ESI) calcd for C₃₅H₄₀NO₅ [M+NH₄]⁺ 554.2901, found 554.2909.

Compound 3f



C-Glycoside **3f** was obtained (22.2 mg, 97%, α only) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 7.33 - 7.25 (m, 15H), 5.82 - 5.72 (m, 1H), 5.07 - 5.05 (m, 1H), 5.02 (s, 1H), 4.72 (d, *J* = 11.9 Hz, 1H), 4.64 - 4.49 (m, 5H), 4.07 - 4.02 (m, 1H), 4.01 - 3.96 (m, 1H), 3.93 - 3.88 (m, 1H), 3.83 - 3.79 (m, 1H), 3.77 - 3.76 (m, 1H), 3.71 - 3.68 (m, 1H), 2.40 - 2.33 (m, 1H), 2.21 - 2.14 (m, 1H), 2.09 - 2.03 (m, 1H), 1.58 - 1.52 (m, 1H). The spectroscopic data coincide with the previous report.⁵

Compound 3g



C-Glycoside **3g** was obtained (24.5 mg, 95%, α only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ +6.0 (*c* 0.10, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.32 - 7.25 (m, 15H), 4.69 (d, *J* = 11.9 Hz, 1H), 4.66 - 4.54 (m, 4H), 4.53 - 4.45 (m, 2H), 4.05 - 4.01 (m, 1H), 3.96 - 3.91 (m, 1H), 3.79 - 3.77 (m, 1H), 3.75 - 3.71 (m, 2H), 2.84 (dd, *J* = 16.9, 5.4 Hz, 1H), 2.55 (dd, *J* = 16.9, 7.7 Hz, 1H), 2.23 - 2.17 (m, 1H), 1.49 - 1.43 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 213.19, 138.60, 138.57, 138.49, 128.32, 128.30, 127.84, 127.74, 127.56, 127.49, 127.42, 127.37, 74.92, 74.39, 73.35, 72.95, 72.17, 70.90, 67.87, 64.77, 44.30, 40.57, 32.43, 26.17. HRMS (ESI) calcd for C₃₃H₄₄NO₅ [M+NH₄]⁺ 534.3214, found 534.3221.

Compound 3h



C-Glycoside **3h** was obtained (20.4 mg, 92%, $\alpha/\beta = 1$: 1) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 7.26 - 7.18 (m, 15.00H), 5.82 - 5.65 (m, 1.07H), 5.07 - 4.93 (m, 2.09H), 4.53 - 4.40 (m, 5.54H), 4.29 (d, *J* = 11.9 Hz, 0.58H), 4.19 - 4.11 (m, 0.53H), 4.03 - 3.93 (m, 2.05H), 3.85 (d, *J* = 2.6 Hz, 0.57H), 3.80 (dd, *J* = 4.3, 2.8 Hz, 0.52H),

3.74 (d, J = 3.5 Hz, 0.54H), 3.58 - 3.41 (m, 2.04H), 2.45 - 2.39 (m, 1.00H), 2.37 - 2.29 (m, 1.02H).The spectroscopic data coincide with the previous report.⁶

Compound 5a



Glycoside **5a** was obtained (29.4 mg, 91%, $\alpha/\beta = 2$: 1) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 7.39 - 7.24 (m, 30.12H), 7.21 - 7.12 (m, 4.75H), 6.71 - 6.64 (m, 3.06H), 6.62 - 6.56 (m, 1.55H), 5.47 (d, *J* = 3.5 Hz, 1.00H), 5.04 (dd, *J* = 10.9, 7.1 Hz, 1.60H), 4.99 (dd, *J* = 5.4, 1.9 Hz, 0.59H), 4.94 (d, *J* = 11.0 Hz, 0.62H), 4.91 - 4.75 (m, 4.88H), 4.68 (d, *J* = 12.0 Hz, 1.03H), 4.61 - 4.47 (m, 3.86H), 4.41 (d, *J* = 12.0 Hz, 1.03H), 4.19 (t, *J* = 9.2 Hz, 1.00H), 3.91 - 3.85 (m, 1.04H), 3.81 - 3.55 (m, 12.72H). The spectroscopic data coincide with the previous report.⁷

Compound 5b



Glycoside **5b** was obtained (29.7 mg, 92%, α only) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 7.37 - 7.26 (m, 18H), 7.18 - 7.12 (m, 2H), 7.05 - 6.98 (m, 2H), 6.83 - 6.77 (m, 2H), 5.36 (d, *J* = 3.5 Hz, 1H), 5.04 (d, *J* = 10.8 Hz, 1H), 4.87 (dd, *J* = 10.8, 6.7 Hz, 2H), 4.79 (d, *J* = 12.0 Hz, 1H), 4.69 (d, *J* = 12.0 Hz, 1H), 4.58 (d, *J* = 12.0 Hz, 1H), 4.50 (d, *J* = 10.8 Hz, 1H), 4.41 (d, *J* = 12.0 Hz, 1H), 4.18 (t, *J* = 9.3 Hz, 1H), 3.97 - 3.89 (m, 1H), 3.79 - 3.68 (m, 6H), 3.59 (dd, *J* = 10.7, 1.8 Hz, 1H). The spectroscopic data coincide with the previous report.⁸

Compound 5c



Glycoside **5c** was obtained (29.7 mg, 88%, $\alpha/\beta = 2$: 1) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 7.40 - 7.25 (m, 27.06H), 7.20 - 7.17 (m, 0.85H), 7.16 - 7.12 (m, 1.96H), 6.74 (d, J = 8.7 Hz, 1.46H), 6.68 (d, J = 2.3 Hz, 1.43H), 6.66 - 6.61 (m, 1.38H), 5.38 (d, J = 3.5 Hz, 1.00H), 5.39 - 5.36 (m, 1.45H), 4.95 (d, J = 10.9 Hz, 0.49H), 4.91 - 4.78 (m, 4.79H), 4.68 (d, J = 12.0 Hz, 1.08H), 4.60 - 4.49 (m, 3.18H), 4.42 (d, J = 12.0 Hz, 1.02H), 4.18 (t, J = 9.3 Hz, 1.02H), 3.94 - 3.90 (m, 1.05H), 3.83 (s, 4.43H), 3.80 (s, 3.36H), 3.77 - 3.56 (m, 8.04H). ¹³C NMR (101 MHz, CDCl₃) δ 151.94, 151.16, 149.59, 144.92, 144.54, 138.81,

138.51, 138.35, 138.19, 138.11, 138.02, 137.81, 128.50, 128.44, 128.41, 128.40, 128.36, 128.17, 128.07, 127.95, 127.89, 127.83, 127.78, 127.74, 127.69, 127.65, 127.63, 111.68, 111.60, 108.18, 107.29, 102.90, 102.87, 102.54, 96.18, 84.76, 82.17, 82.06, 79.83, 77.85, 77.54, 75.83, 75.79, 75.16, 75.06, 73.54, 73.45, 73.36, 70.74, 69.05, 68.41, 56.30, 55.89, 55.80. HRMS (ESI) calcd for $C_{42}H_{48}NO_8$ [M+NH₄]⁺ 694.3374, found 694.3373.

Compound 5d



C-Glycoside **5d** was obtained (19.2 mg, 71%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ +0.8 (*c* 1.00, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.13 (s, 1H), 7.38 - 7.24 (m, 13H), 7.22 - 7.20 (m, 2H), 6.86 (d, *J* = 8.5 Hz, 1H), 6.49 (d, *J* = 2.5 Hz, 1H), 6.39 (dd, *J* = 8.4, 2.6 Hz, 1H), 4.92 (d, *J* = 10.9 Hz, 1H), 4.72 (d, *J* = 11.6 Hz, 1H), 4.65 - 4.47 (m, 5H), 3.80 - 3.75 (m, 4H), 3.72 - 3.64 (m, 3H), 3.61 - 3.55 (m, 1H), 2.41 - 2.36 (m, 1H), 1.93 (dd, *J* = 13.0, 11.7 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 160.65, 156.98, 138.34, 138.28, 137.93, 128.43, 128.40, 128.07, 127.79, 127.76, 127.69, 127.63, 126.99, 117.39, 105.84, 102.91, 80.48, 78.58, 77.96, 77.66, 75.21, 73.42, 71.45, 68.59, 55.29, 36.93. HRMS (ESI) calcd for C₃₄H₃₆NaO₆ [M+Na]⁺ 563.2404, found 563.2408.

Compound 5e



C-Glycoside **5e** was obtained (22.8 mg, 80%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ +34.7 (*c* 0.04, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.60 (s, 1H), 7.40 - 7.24 (m, 13H), 7.23 - 7.19 (m, 2H), 6.11 (d, *J* = 2.2 Hz, 1H), 6.00 (d, *J* = 2.2 Hz, 1H), 5.02 (dd, *J* = 11.7, 2.0 Hz, 1H), 4.93 (d, *J* = 10.9 Hz, 1H), 4.70 (d, *J* = 11.6 Hz, 1H), 4.64 - 4.53 (m, 3H), 4.46 (d, *J* = 12.1 Hz, 1H), 3.79 - 3.73 (m, 9H), 3.67 (dd, *J* = 10.2, 1.9 Hz, 1H), 3.56 - 3.49 (m, 1H), 2.38 - 2.27 (m, 1H), 1.90 - 1.75 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 160.78, 157.64, 156.86, 138.61, 138.55, 138.07, 128.42, 128.39, 128.36, 128.05, 127.79, 127.66, 127.59, 127.55, 106.17, 94.83, 90.74, 80.55, 78.77, 75.19, 73.43, 73.36, 71.24, 68.35, 55.54, 55.30, 36.19. HRMS (ESI) calcd for C₃₅H₃₈NaO₇ [M+Na]⁺ 593.2510, found 593.2520.

Compound 5f



C-Glycoside **5f** was obtained (24.6 mg, 82%, β only) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz,CDCl₃) δ 8.31 (s, 1H), 7.36 - 7.25 (m, 13H), 7.24

- 7.19 (m, 2H), 6.27 (s, 1H), 4.96 - 4.91 (m, 2H), 4.70 (d, J = 11.6 Hz, 1H), 4.63 (d, J = 11.6 Hz, 1H), 4.61 - 4.52 (m, 2H), 4.46 (d, J = 12.1 Hz, 1H), 3.87 (s, 3H), 3.81 (s, 3H), 3.80 - 3.74 (m, 6H), 3.67 (dd, J = 10.2, 2.2 Hz, 1H), 3.57 - 3.51 (m, 1H), 2.33 - 2.23 (m, 1H), 1.94 - 1.82 (m, 1H). The spectroscopic data coincide with the previous report.⁹

Compound 5g



C-Glycoside **5g** was obtained (20.1 mg, 72%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ -5.0 (*c* 0.40, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.86 (s, 1H), 8.35 - 8.28 (m, 1H), 7.78 - 7.71 (m, 1H), 7.49 -7.45 (m, 2H), 7.41 - 7.27 (m, 14H), 7.25 - 7.19 (m, 2H), 7.03 (d, *J* = 8.4 Hz, 1H), 4.95 (d, *J* = 10.9 Hz, 1H), 4.80 (dd, *J* = 11.9, 2.2 Hz, 1H), 4.73 (d, *J* = 11.6 Hz, 1H), 4.67 - 4.52 (m, 4H), 3.87 - 3.74 (m, 4H), 3.68 - 3.62 (m, 1H), 2.49 - 2.44 (m, 1H), 2.08 - 1.96 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 151.32, 138.40, 138.30, 137.97, 134.01, 128.45, 128.41, 128.11, 127.90, 127.78, 127.73, 127.68, 127.62, 127.17, 126.34, 125.77, 125.15, 124.15, 122.44, 119.11, 117.44, 80.51, 79.09, 78.76, 77.56, 75.26, 73.45, 71.41, 68.51, 37.22. HRMS (ESI) calcd for C₃₇H₄₀NO₅ [M+NH₄]⁺ 578.2901, found 578.2906.

Compound 5h



C-Glycoside **5h** was obtained (25.2 mg, 90%, β only) as a colorless oil according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 9.06 (s, 1H), 7.76 (d, *J* = 8.0 Hz, 1H), 7.69 (d, *J* = 8.9 Hz, 1H), 7.60 (d, *J* = 8.5 Hz, 1H), 7.47 - 7.42 (m, 1H), 7.39 - 7.25 (m, 14H), 7.24 - 7.19 (m, 2H), 7.15 (d, *J* = 8.9 Hz, 1H), 5.47 (dd, *J* = 11.9, 2.1 Hz, 1H), 4.96 (d, *J* = 10.9 Hz, 1H), 4.70 (d, *J* = 11.6 Hz, 1H), 4.66 - 4.55 (m, 3H), 4.49 (d, *J* = 12.1 Hz, 1H), 3.92 - 3.86 (m, 2H), 3.83 (dd, *J* = 10.2, 2.6 Hz, 1H), 3.73 (dd, *J* = 10.2, 2.2 Hz, 1H), 3.67 - 3.62 (m, 1H), 2.53 - 2.45 (m, 1H), 2.05 - 1.95 (m, 1H). The spectroscopic data coincide with the previous report.¹⁰

Compound 5i



C-Glycoside **5i** was obtained (26.8 mg, 91%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ +103.8 (*c* 0.13, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 9.01 (s, 1H), 7.67 (d, *J* = 8.9 Hz, 1H), 7.61 (d, *J* = 8.8 Hz, 1H), 7.40 - 7.26 (m, 13H), 7.25 - 7.21 (m, 2H), 6.99 (dd, *J* = 8.9, 2.6 Hz, 2H), 6.88 (d, *J* = 2.1 Hz, 1H), 5.35 (dd, *J* = 11.9, 1.9 Hz, 1H), 4.97 (d, *J* = 10.9 Hz, 1H), 4.72 - 4.56 (m, 4H), 4.49 (d, *J* = 12.1 Hz, 1H), 3.93 - 3.85 (m, 5H), 3.83 (dd, *J* =

10.2, 2.6 Hz, 1H), 3.73 (dd, J = 10.2, 2.2 Hz, 1H), 3.69 - 3.61 (m, 1H), 2.52 - 2.43 (m, 1H), 2.05 - 1.95 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 158.39, 154.66, 138.42, 138.26, 137.94, 131.91, 130.50, 129.48, 128.46, 128.44, 128.40, 128.05, 127.86, 127.74, 127.70, 124.07, 117.79, 114.18, 113.87, 101.07, 80.13, 79.05, 75.69, 75.25, 73.45, 71.28, 68.07, 55.34, 35.69. HRMS (ESI) calcd for C₃₈H₄₂NO₆ [M+NH₄]⁺ 608.3007, found 608.3013.

Compound 5j



C-Glycoside **5j** was obtained (24.0 mg, 78%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ -18.0 (*c* 0.10, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.78 (s, 1H), 7.43 - 7.26 (m, 20H), 6.87 (d, *J* = 8.4 Hz, 1H), 6.54 (d, *J* = 2.5 Hz, 1H), 6.43 (dd, *J* = 8.4, 2.5 Hz, 1H), 5.05 - 4.97 (m, 3H), 4.69 - 4.57 (m, 3H), 4.54 (dd, *J* = 11.8, 2.7 Hz, 1H), 4.47 (d, *J* = 11.9 Hz, 1H), 4.40 (d, *J* = 11.8 Hz, 1H), 3.94 (s, 1H), 3.73 - 3.67 (m, 1H), 3.66 - 3.51 (m, 3H), 2.53 - 2.41 (m, 1H), 2.11 - 2.02 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 159.92, 157.02, 138.72, 138.27, 137.79, 137.04, 128.54, 128.48, 128.43, 128.27, 127.92, 127.88, 127.80, 127.66, 127.48, 127.45, 127.40, 127.29, 118.44, 106.36, 103.74, 78.55, 77.90, 77.72, 74.06, 73.57, 72.18, 70.12, 69.96, 69.24, 32.28. HRMS (ESI) calcd for C₄₀H₄₄NO₆ [M+NH₄]⁺ 634.3163, found 634.3176.

Compound 5k



C-Glycoside **5k** was obtained (24.3 mg, 81%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ +20.4 (*c* 0.46, CHCl₃). ¹H NMR (400 MHz,CDCl₃) δ 7.85 (s, 1H), 7.34 - 7.25 (m, 15H), 6.25 (s, 1H), 5.04 (d, *J* = 11.8 Hz, 1H), 4.95 (dd, *J* = 11.9, 3.1 Hz, 1H), 4.66 - 4.60 (m, 3H), 4.43 (dd, *J* = 11.9 Hz, 2H), 3.97 (s, 1H), 3.87 (s, 3H), 3.79 (s, 3H), 3.77 (s, 3H), 3.75 - 3.69 (m, 1H), 3.68 - 3.63 (m, 1H), 3.62 - 3.57 (m, 2H), 2.49 - 2.40 (m, 1H), 2.01 - 1.96 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 153.56, 152.34, 149.89, 138.88, 138.37, 137.81, 134.85, 128.42, 128.25, 127.89, 127.79, 127.57, 127.36, 127.25, 111.23, 97.13, 77.92, 77.73, 74.02, 73.53, 73.08, 72.32, 70.12, 68.98, 61.37, 60.94, 55.82, 32.17. HRMS (ESI) calcd for C₃₆H₄₄NO₈ [M+NH₄]+ 618.3061, found 618.3068.

Compound 51



C-Glycoside **51** was obtained (23.8 mg, 85%, β only) as a colorless oil according to the *C*-glycosylation procedure. [α]_D²⁵ +56.9 (*c* 0.16, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.56 (s, 1H), 7.75 (d, *J* = 7.4 Hz, 1H), 7.71 - 7.67 (m, 2H), 7.46 - 7.41 (m, 1H), 7.38 - 7.25 (m, 16H), 7.14 (d, *J* = 8.9 Hz, 1H), 5.45 (dd, *J* = 12.0, 2.9 Hz, 1H), 5.07 (d, *J* = 11.9 Hz, 1H), 4.69 (d, *J* = 11.9 Hz, 1H), 4.64 (d, *J* = 12.1 Hz, 1H), 4.59 (d, *J* = 12.0 Hz, 1H), 4.47 (d, *J* = 11.9 Hz, 1H), 4.41 (d, *J* = 11.9 Hz, 1H), 4.04 (s, 1H), 3.84 - 3.79 (m, 1H), 3.77 (d, *J* = 6.4 Hz, 1H), 3.68 - 3.61 (m, 2H), 2.62 - 2.50 (m, 1H), 2.19 - 2.14 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 154.23, 138.82, 138.25, 137.78, 130.80, 129.72, 128.90, 128.58, 128.48, 128.45, 128.32, 127.93, 127.84, 127.69, 127.66, 127.46, 127.29, 126.58, 122.73, 120.62, 119.90, 116.12, 78.27, 77.90, 75.02, 74.13, 73.61, 72.36, 70.13, 69.08, 31.36. HRMS (ESI) calcd for C₃₇H₄₀NO₅ [M+NH₄]⁺ 578.2901, found 578.2909.

Compound 5m



C-Glycoside **5m** was obtained (25.9 mg, 88%, β only) as a colorless oil according to the *C*-glycosylation procedure. $[\alpha]_D^{25}$ +46.4 (*c* 0.11, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 8.35 (s, 1H), 7.60 (dd, *J* = 12.7, 9.1 Hz, 2H), 7.39 - 7.25 (m, 15H), 7.14 - 7.08 (m, 3H), 5.40 (dd, *J* = 12.0, 2.7 Hz, 1H), 5.07 (d, *J* = 11.9 Hz, 1H), 4.69 (d, *J* = 11.9 Hz, 1H), 4.66 - 4.55 (m, 2H), 4.50 - 4.37 (m, 2H), 4.04 (s, 1H), 3.89 (s, 3H), 3.83 - 3.78 (m, 1H), 3.76 (d, *J* = 6.3 Hz, 1H), 3.64 (d, *J* = 6.3 Hz, 2H), 2.63 - 2.50 (m, 1H), 2.17 - 2.09 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 155.31, 152.50, 138.82, 138.24, 137.77, 129.51, 128.47, 128.45, 128.40, 128.30, 127.93, 127.83, 127.68, 127.65, 127.44, 127.28, 125.99, 122.20, 120.31, 118.95, 116.49, 107.22, 78.23, 77.91, 75.11, 74.11, 73.60, 72.33, 70.13, 69.07, 55.31, 31.42. HRMS (ESI) calcd for C₃₈H₄₂NO₆ [M+NH₄]⁺ 608.3007, found 608.3018.

Compound 5n



C-Glycoside **5n** was obtained (19.4 mg, 70%, β only) as a yellow solid according to the *C*-glycosylation procedure. ¹H NMR (400 MHz, CDCl₃) δ 8.97 (s, 1H), 7.39 - 7.29 (m, 10H), 7.28 - 7.25 (m, 3H), 7.05 (dd, J = 6.6, 2.9 Hz, 2H), 6.13 (d, J = 2.3 Hz, 1H), 6.01 (d, J = 2.4 Hz, 1H), 5.51 (d, J = 3.5 Hz, 1H), 4.61 (d, J = 5.6 Hz, 2H), 4.49 (s, 2H), 4.22 (d, J = 1.6 Hz, 2H), 4.20 - 4.15 (m, 1H), 4.09 (d, J = 3.5 Hz, 1H), 4.00 (d, J = 3.4 Hz, 1H), 3.80 (s, 3H), 3.74 - 3.64 (m, 5H). The spectroscopic data coincide with the previous report.¹¹

3. References

1 M. Liu, B.-H. Li, D.-C. Xiong and X.-S. Ye, J. Org. Chem., 2018, 83, 8292.

- 2 W. C. Wei and C. C. Chang, Eur. J. Org. Chem., 2017, 3033.
- 3 G. J. McGarvey, C. A. LeClair and B. A. Schmidtmann, Org. Lett., 2008, 10, 4727.
- 4 L. G. Xie, J. Rogers, I. Anastasiou, J. A. Leitch and D. J. Dixon, Org. Lett., 2019, 21, 6663.
- 5 X. P. Chen, Q. L. Wang and B. Yu, Chem. Commun., 2016, 52, 12183.
- 6 B. Doboszewski, Nucleosides, Nucleotides and Nucleic acid, 2009, 28, 875.

7 S. Yajima, T. Saitoh, K. Kawa, K. Nakamura, H. Nagase, Y. Einaga and S. Nishiyama, *Tetrahedron*, 2016, **72**, 8428.

- 8 M. Koshiba, N. Suzuki, R. Arihara, T. Tsuda, H. Nambu, S. Nakamura and S. Hashimoto, *Chem. Asian J.*, 2008, **3**, 1664.
- 9 C. Yamada, K. Sasaki, S. Matsumura and K. Toshima, Tetrahedron Lett., 2007, 48, 4223.
- 10 P. Mitra, B. Behera, T. K. Maiti and D. Mal, J. Org. Chem., 2013, 78, 9748.

11 M. Cornia, G. Casiraghi and L. Zetta, Tetrahedron, 1990, 46, 3071.

4. Copies of NMR Spectra



¹H NMR of compound S4























































¹³C NMR of compound **5**c



¹³C NMR of compound **5d**





















¹³C NMR of compound **5**j







¹³C NMR of compound **5**l







¹H NMR of compound **5n**