

Supporting Information

O-Cyclopropyl Hydroxylamines: Gram-Scale Synthesis and Utility as Precursors for *N*-Heterocycles

Kaitlyn Lovato^{[a]+}, Urmibhusan Bhakta^{[a]+}, Yi Pin Ng^[b], and László Kurti*^[a]

^[a]Department of Chemistry, Rice University, BioScience Research Collaborative, 6500 Main Street, Room 380, Houston, TX, 77030 (USA)

^[b] Division of Chemistry and Biological Chemistry, School of Physical & Mathematical Sciences, Nanyang Technological University, 50 Nanyang Avenue, Singapore 639798

⁺ These authors contributed equally to this work.

Table of Contents

General Remarks	2
Preparation of <i>N</i>-enoxyphthalimides	2
Preparation of 2-cyclopropoxyisoindoline-1,3-diones	2
Characterization of 2-cyclopropoxyisoindoline-1,3-diones	3
Preparation and Characterization of Ring-Unsubstituted <i>O</i>-cyclopropyl Hydroxylamine Hydrochloride Salt.....	5
Preparation and Characterization of Substituted <i>O</i>-cyclopropyl Hydroxylamine.....	6
Preparation and Characterization of Unsubstituted <i>O</i>-cyclopropyl Hydroxamates.....	6
Preparation and Characterization of Substituted <i>O</i>-cyclopropyl Hydroxamate.....	8
Preparation of <i>N</i>-arylated-<i>O</i>-cyclopropyl Hydroxamates.....	9
Characterization of <i>N</i>-arylated-<i>O</i>-cyclopropyl Hydroxamates.....	9
Preparation of 2-Hydroxy-tetrahydroquinolines via [3,3] Rearrangement of <i>N</i>-arylated-<i>O</i>-cyclopropyl Hydroxamates	14
Characterization of 2-hydroxy-tetrahydroquinolines	15
Preparation of Derivatized 2-hydroxy-tetrahydroquinoline Products.....	21
References	24
NMR Spectra of Reported Compounds	25

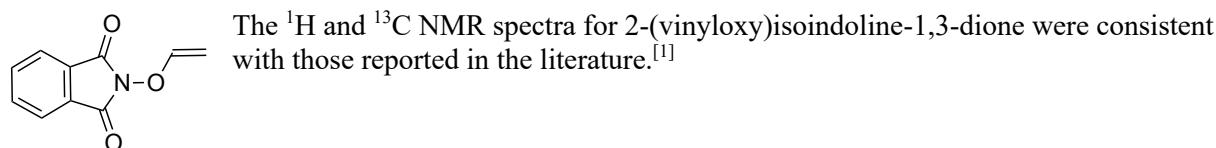
General Remarks

All starting material syntheses were performed in oven-dried 50 mL or 100 mL round-bottomed flasks. Commercially available solvents and reagents were used without further purification. All arylation reactions were carried out in oven-dried 8 mL scintillation vials, while rearrangement reactions were carried out in oven-dried 20 mL scintillation vials. All reactions were monitored by thin-layer chromatography (TLC) with E. Merck silica gel 60 F254 pre-coated plates (0.25 mm). Flash chromatography was carried out using a Biotage Isolera One system with 10g KP-Sil cartridges utilizing ethyl acetate (EA) and hexane (hex) as eluents. ¹H and ¹³C nuclear magnetic resonance (NMR) spectra were obtained using a Bruker DRX-600 NMR spectrometer. Chemical shifts are documented in parts per million (δ , ppm). ¹H NMR spectra are referenced to 7.26 (CDCl₃) and ¹³C NMR spectra are referenced to 77.16 (CDCl₃). The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, p = pentet, br s = broad singlet, dd = doublet of doublets, ddd = doublet of doublet of doublets, dddd = doublet of doublet of doublet of doublets, dt = doublet of triplets, td = triplet of doublets, dtd = doublet of triplet of doublets, tt = triplet of triplets, dp = doublet of pentets, ddt = doublet of doublet of triplets, dtdd = doublet of triplet of doublet of doublets, tddd = triplet of doublet of doublet of doublets, dq = doublet of quartet of triplets, qd = quartet of doublets, dt = doublet of triplet of triplets, qt = quartet of triplets, dq = doublet of quartets, tq = triplet of quartets. High Resolution Mass Spectrometry was performed on an Agilent 1290/6230 LCMS-TOF under electrospray ionization (ESI) conditions in both positive and negative mode. Melting points were recorded on a Mettler Toledo MP50 melting point system.

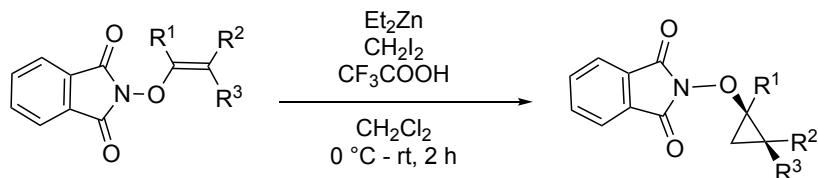
Preparation of *N*-enoxyphthalimides

The preparation of the *N*-enoxyphthalimides was carried out following literature reported protocols. *N*-enoxyphthalimide **15a** was synthesized following protocol A^[1] and the *N*-enoxyphthalimides (**15b-15g**) (i.e those used to synthesize 2-cyclopropoxyisoindolin-1,3diones (**16b-16g**) were synthesized following the copper promoted protocol B^[2]. The ¹H and ¹³C NMR spectra of the obtained *N*-enoxyphthalimides were consistent with those reported in the literature.^[1-3]

2-(vinyloxy)isoindoline-1,3-dione (**15a**)



Preparation of 2-cyclopropoxyisoindoline-1,3-diones

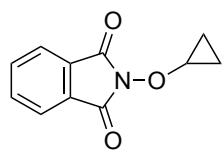


Neat diethylzinc (20.5 mL, 200 mmol, 2.0 equiv) was added to 250 mL of dry DCM under inert atmosphere in a glove box. The diethylzinc solution was removed from the glove box and kept under inert gas. A

solution of trifluoroacetic acid (15.3 mL, 200 mmol, 2.0 equiv) in DCM (125 mL) was slowly added to the diethylzinc solution at 0 °C and stirred until gas evolution ceased. After stirring at 0 °C for about 20 minutes, a solution of diiodomethane (16.13 mL, 200 mmol, 2.0 equiv) in DCM (125 mL) was added. The mixture was stirred for an additional 20 minutes. Upon further stirring, a solution of *N*-enoxyphthalimide **15a** (22.7 g, 120 mmol, 1.0 equiv) in DCM (100 mL) was added. Then the reaction was removed from the ice bath, allowed to warm to rt and stirred for 2 h or until the reaction was complete by TLC analysis. Once the starting material was consumed, the reaction mixture was decanted into a separatory funnel and carefully quenched with 0.1 N HCl (500 mL). The organic layer was separated and washed with saturated NaHCO₃, and brine. The organic layer was dried over Na₂SO₄, filtered and concentrated.^[4] The crude mixture was purified via flash chromatography (30% EA in hexanes) to give 2-cyclopropoxysisoindoline-1,3-dione **16a** as a white solid. The same procedure was used to synthesize **16b–16g** from the corresponding *N*-enoxyphthalimides.

Characterization of 2-cyclopropoxysisoindoline-1,3-diones

2-cyclopropoxysisoindoline-1,3-dione (16a)

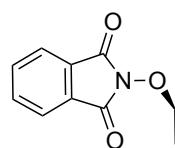


¹H NMR (600 MHz, CDCl₃) δ 7.85 (dd, *J* = 5.4, 3.1 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.40 (tt, *J* = 6.1, 2.7 Hz, 1H), 1.18 – 1.14 (m, 2H), 0.70 – 0.66 (m, 2H).
¹³C NMR (151 MHz, CDCl₃) δ 163.70 (2C), 134.61 (2C), 129.12 (2C), 123.71 (2C), 61.57, 6.90 (2C).

Yield: 2.16 g, 94%. White solid (m.p. 76.0 °C).

HRMS (ESI): m/z calcd for [C₁₁H₉NO₃]⁺ ([M+H]⁺): 204.0655, found 204.0655

2-((1*R*,2*R*)-2-butylcyclopropoxysisoindoline-1,3-dione (16b)

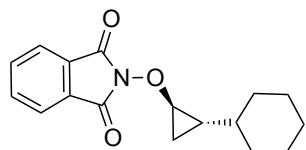


¹H NMR (600 MHz, CDCl₃) δ 7.84 (td, *J* = 5.2, 3.5 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.11 (dt, *J* = 6.5, 2.4 Hz, 1H), 1.50 (dtd, *J* = 12.9, 9.3, 8.3, 6.0 Hz, 1H), 1.32 – 1.23 (m, 5H), 1.17 (dq, *J* = 17.6, 6.9 Hz, 2H), 0.81 (t, *J* = 7.0 Hz, 3H), 0.49 (q, *J* = 6.4 Hz, 1H).
¹³C NMR (151 MHz, CDCl₃) δ 163.68 (2C), 134.57 (2C), 129.10 (2C), 123.66 (2C), 67.20, 30.98, 30.62, 22.34, 20.41, 14.05, 13.67.

Yield: 1.23 g, 95%. Tan solid (m.p. 79.9 °C).

HRMS (ESI): m/z calcd for [C₁₅H₁₇NO₃K]⁺ ([M+K]⁺): 298.0840, found 298.0858

2-((1*R*,2*S*)-2-cyclohexylcyclopropoxy)isoindoline-1,3-dione (16c)



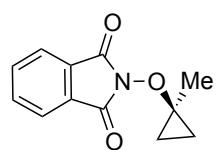
¹H NMR (600 MHz, CDCl₃) δ 7.84 (dd, *J* = 5.4, 3.1 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.16 (dt, *J* = 6.3, 2.5 Hz, 1H), 1.69 – 1.57 (m, 5H), 1.37 (dddd, *J* = 11.3, 9.2, 6.7, 2.3 Hz, 1H), 1.22 (ddd, *J* = 10.6, 6.1, 2.6 Hz, 1H), 1.16 – 1.08 (m, 3H), 1.01 (dddd, *J* = 15.5, 11.6, 7.5, 4.3 Hz, 2H), 0.62 – 0.52 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 163.68 (2C), 134.58 (2C), 129.09 (2C), 123.69 (2C), 66.24, 39.40, 32.49, 31.83, 26.92, 26.41, 26.12, 26.08, 12.47.

Yield: 262.5 mg, 92%. White solid (m.p. 109.8 °C).

HRMS (ESI): m/z calcd for [C₁₇H₂₀NO₃]⁺ ([M+H]⁺): 286.1438, found 286.1441

2-(1-methylcyclopropoxy)isoindoline-1,3-dione (16d)



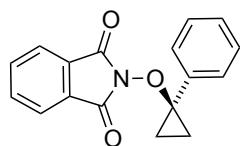
¹H NMR (600 MHz, CDCl₃) δ 8.83 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 1.59 (s, 3H), 1.43 – 1.40 (m, 2H), 0.60 – 0.57 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 164.73 (2C), 134.60 (2C), 129.29 (2C), 123.66 (2C), 67.80, 21.05, 14.13 (2C).

Yield: 206.3 mg, 95%. White solid (m.p. 93.4 °C).

HRMS (ESI): m/z calcd for [C₁₂H₁₂NO₃]⁺ ([M+H]⁺): 218.0812, found 218.0795

2-(1-phenylcyclopropoxy)isoindoline-1,3-dione (16e)

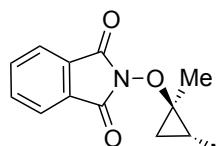


¹H NMR (600 MHz, CDCl₃) δ 7.70 (ddt, *J* = 19.1, 5.5, 3.2 Hz, 4H), 7.57 (dd, *J* = 6.6, 3.0 Hz, 2H), 7.32 (p, *J* = 3.5 Hz, 3H), 1.85 – 1.80 (m, 2H), 1.14 – 1.10 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 163.90 (2C), 137.30, 134.43 (4C), 130.27 (2C), 129.05, 128.24 (2C), 123.52 (2C), 72.57, 13.55 (2C).

Yield: 252.0 mg, 90%. White solid.

2-((1*R*,2*R*)-1,2-dimethylcyclopropoxy)isoindoline-1,3-dione (16f)



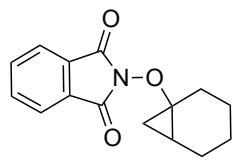
¹H NMR (600 MHz, CDCl₃) δ 7.84 (dd, *J* = 5.4, 3.1 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 1.70 (dp, *J* = 10.5, 6.5 Hz, 1H), 1.55 (s, 3H), 1.55 – 1.52 (m, 1H), 1.04 (d, *J* = 6.4 Hz, 3H), 0.19 (dd, *J* = 7.0, 5.6 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 164.90 (2C), 134.59 (2C), 129.37 (2C), 123.67 (2C), 71.26, 20.72, 19.04, 16.55, 13.93.

Yield: 208.1 mg, 90%. White solid (m.p. 111.0 °C).

HRMS (ESI): m/z calcd for [C₁₃H₁₃NO₃]⁺ ([M+H]⁺): 254.0788, found 254.0774

2-(bicyclo[4.1.0]heptan-1-yloxy)isoindoline-1,3-dione (16g)



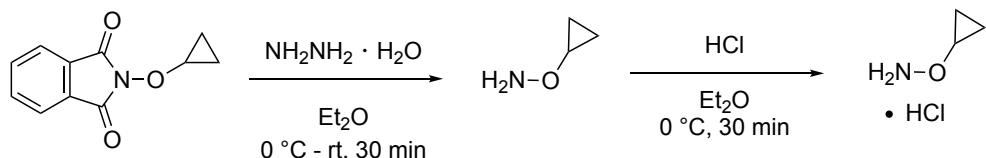
¹H NMR (600 MHz, CDCl₃) δ 7.83 (dd, *J* = 5.4, 3.1 Hz, 2H), 7.74 (dd, *J* = 5.5, 3.0 Hz, 2H), 2.39 (ddd, *J* = 13.4, 9.5, 5.9, 1.6 Hz, 1H), 2.17 (dt, *J* = 13.4, 5.4 Hz, 1H), 2.03 – 1.95 (m, 1H), 1.85 – 1.73 (m, 1H), 1.56 – 1.47 (m, 2H), 1.42 (ddd, *J* = 14.4, 8.1, 5.9, 1.8 Hz, 1H), 1.24 (dtdd, *J* = 25.6, 13.2, 6.5, 2.9 Hz, 2H), 1.08 (tddd, *J* = 12.8, 9.7, 6.5, 3.9 Hz, 1H), 0.46 (t, *J* = 6.2 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 164.91 (2C), 134.57 (2C), 129.31 (2C), 123.64 (2C), 70.70, 28.02, 24.16, 21.60, 21.27, 19.27, 18.46.

Yield: 190.3mg, 74%. Yellow solid (m.p. 75.3 °C).

HRMS (ESI): m/z calcd for [C₁₅H₁₆NO₃]⁺ ([M+H]⁺): 258.1125, found 258.1110

Preparation and Characterization of Ring-unsubstituted *O*-cyclopropyl Hydroxylamine Hydrochloride Salt



Hydrazine hydrate (50-60 %, 7.5 ml, 118 mmol, 2.8 equiv) was added dropwise (over 3 minutes) to a solution of 2-cyclopropoxyisoindoline-1,3-dione **16a** (8.5 g, 42 mmol, 1.0 equiv) in diethyl ether (167 ml, 0.25 M) at 0 °C. The reaction mixture was allowed to warm to rt and stirred for 30 minutes. The turbid mixture turned clear with a white precipitate. The precipitated diazine byproduct was filtered and washed with diethyl ether (60 ml). The combined ether filtrate was re-cooled to 0 °C and 2M HCl in ether (31.5 ml, 63 mmol, 1.5 equiv) was added over 3 minutes. The flask was stirred at 0 °C for an additional 30 minutes. The mixture was filtered, and the white solid was collected and dried to give the desired O-cyclopropyl hydroxylamine hydrochloride salt **17a** as a white solid.

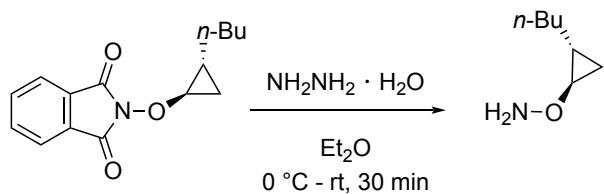
O-cyclopropylhydroxylamine hydrochloride salt (17a)

¹H NMR (600 MHz, DMSO-d6) δ 11.21 (br s, 2H), 4.13 (tt, *J* = 5.8, 2.6 Hz, 1H), 0.88 (q, *J* = 4.6, 3.5 Hz, 2H), 0.67 (qd, *J* = 6.4, 1.4 Hz, 2H).
• HCl **¹³C NMR (151 MHz, DMSO-d6)** δ 57.26, 6.62 (2C).

Yield: 8.55 g, 73%. White solid (m.p. 57.1 °C).

HRMS (CI): m/z calcd for [C₃H₈NO]⁺ ([M]⁺): 74.0600, found 74.0607

Preparation and Characterization of Ring-substituted *O*-cyclopropyl Hydroxylamine



Hydrazine hydrate (50-60 %, 0.60 ml, 9.5 mmol, 2.8 equiv) was added dropwise (over 2 minutes) to a solution of 2-((1*R*,2*R*)-2-butylcyclopropoxy)isoindoline-1,3-dione **16b** (877 mg, 3.38 mmol, 1.0 equiv) in diethyl ether (16.9 ml, 0.2 M) at 0°C . The reaction mixture was removed from the ice bath and allowed to warm to rt. After stirring for 30 minutes, the turbid mixture turned clear with a white precipitate. The precipitated diazine byproduct was filtered and washed with diethyl ether (4.8 ml). The combined filtrate was concentrated and yielded the desired *O*-((1*R*,2*R*)-2-butylcyclopropyl)hydroxylamine product **17b** as a colorless oil.

O-((1*R*,2*R*)-2-butylcyclopropyl)hydroxylamine (**17b**)

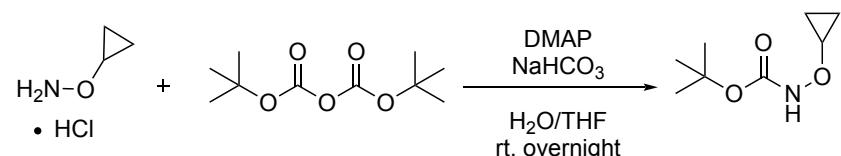
1H NMR (600 MHz, CDCl_3) δ 5.33 (br s, 2H), 3.32 (dt, $J = 5.7, 2.6$ Hz, 1H), 1.40 – 1.29 (m, 4H), 1.17 (dh, $J = 28.7, 7.2$ Hz, 2H), 0.98 (tdt, $J = 16.6, 7.0, 2.3$ Hz, 1H), 0.88 (t, $J = 7.1$ Hz, 3H), 0.76 (ddd, $J = 10.0, 5.7, 2.8$ Hz, 1H), 0.34 (q, $J = 6.0$ Hz, 1H).

13C NMR (151 MHz, CDCl_3) δ 64.42, 31.29, 31.18, 22.53, 19.66, 14.18, 12.90.

Yield: 792 mg, 78%. Colorless oil.

HRMS (CI): m/z calcd for $[\text{C}_7\text{H}_{16}\text{NO}]^+$ ($[\text{M}+\text{H}]^+$): 130.1226, found 130.1234

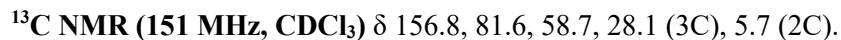
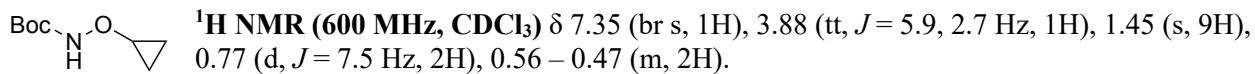
Preparation and Characterization of Unsubstituted *O*-cyclopropyl Hydroxamates



To a suspension of O-cyclopropyl hydroxylamine hydrochloride salt **17a** (3.87 g, 35.3 mmol, 1.0 equiv) in $\text{H}_2\text{O}/\text{THF}$ (3:4, 140 mL, 0.25 M) was slowly added NaHCO_3 (2.52g, 30 mmol, 0.85 equiv) at rt. Once the solution became clear, DMAP (43 mg, 0.353 mmol, 1 mol %) was added. Boc_2O (8.08 g, 37.07 mmol, 1.05 equiv) was dissolved in THF (24 mL) and added dropwise to the solution via syringe. The reaction mixture was left stirring at rt overnight. Upon completion via TLC, the reaction mixture was concentrated and re-dissolved in EA (140 mL). The organic layer was separated and washed with NaHSO_4 , H_2O and brine. The organic layer was then dried over MgSO_4 , filtered and concentrated in vacuo. The crude residue

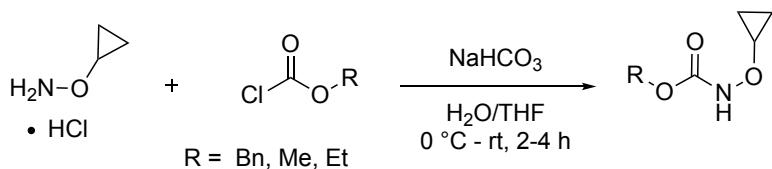
was purified using flash chromatography (30% EA in hexanes) on a Biotage Isolera system to give the corresponding *tert*-butyl cyclopropoxycarbamate **18a** as a colorless oil.

***tert*-butyl cyclopropoxycarbamate (18a)**



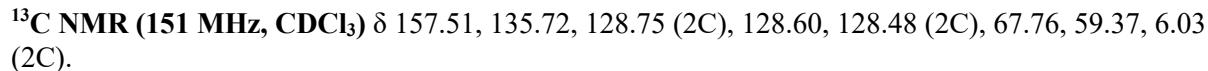
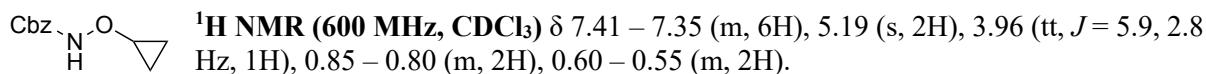
Yield: 4.57 g, 75%. Colorless oil.

HRMS (ESI): m/z calcd for [C₈H₁₅NO₃]⁺ ([M+Na]⁺): 196.0944; found 196.0917



A suspension of O-cyclopropyl hydroxylamine hydrochloride salt **17a** (1.1g, 10 mmol) in H₂O/THF (3:4, 40 mL, 0.25 M) was cooled to 0 °C and stirred for 5 min. Then, NaHCO₃ (2.52g, 30 mmol, 3.0 equiv) followed by the corresponding chloroformate (i.e. benzyl chloroformate, methyl chloroformate, ethyl chloroformate, 15 mmol, 1.5 equiv) were added at 0 °C. The reaction mixture was removed from the ice bath and allowed to warm to rt. Reaction progress was monitored via TLC. Once the reaction was complete, the mixture was diluted with EA (20 mL). The layers were separated, and the organic layer was washed twice with H₂O, followed by NH₄Cl, and brine. The solution was dried with Na₂SO₄, filtered and concentrated in vacuo. The crude residue was purified using flash chromatography (30% EA in hexanes) on a Biotage Isolera system to give the corresponding *N*-protected-*O*-cyclopropyl hydroxamates (**18b-18d**).

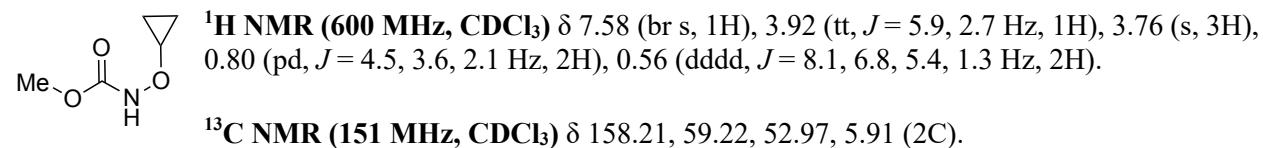
benzyl cyclopropoxycarbamate (18b)



Yield: 497 mg, 78%. White solid (m.p. 56.3 °C).

HRMS (ESI): m/z calcd for [C₁₁H₁₃NO₃Na]⁺ ([M+Na]⁺): 230.0788, found 230.0783

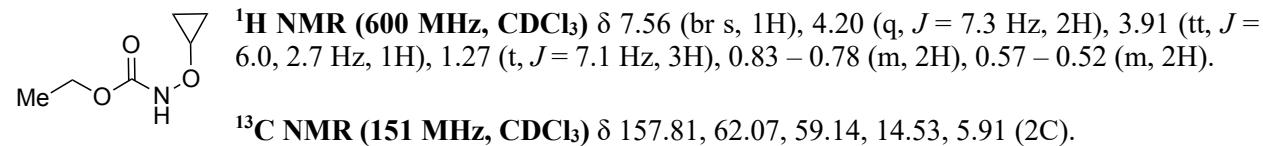
methyl cyclopropoxycarbamate (18c)



Yield: 233.6 mg, 59%. Colorless liquid.

HRMS (ESI): m/z calcd for [C₅H₉NO₃K]⁺ ([M+K]⁺): 170.0214, found 170.0113

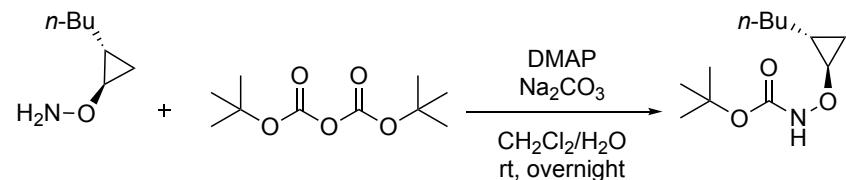
ethyl cyclopropoxycarbamate (18d)



Yield: 275 mg, 52%. Colorless liquid.

HRMS (ESI): m/z calcd for [C₆H₁₁NO₃Na]⁺ ([M+Na]⁺): 168.0631, found 168.0612

Preparation and Characterization of Substituted *O*-cyclopropyl Hydroxamate



To a solution of *O*-((1*R*,2*R*)-2-butylcyclopropyl)hydroxylamine **17b** (370 mg, 2.87 mmol, 1.0 equiv) in CH₂Cl₂/H₂O (4:3, 11.5 mL, 0.25 M) was slowly added NaHCO₃ (253 mg, 3.0 mmol, 1.05 equiv) at rt. Once the solution became clear, DMAP (3.5 mg, 28.7 μmol, 1 mol %) was added. Boc₂O (657 mg, 3.0 mmol, 1.05 equiv) was dissolved in THF (2 mL) and added dropwise to the solution via syringe. The reaction mixture was left stirring at rt overnight. Upon completion via TLC, the reaction mixture was concentrated and re-dissolved in EA (11.5 mL). The organic layer was separated and washed with NaHSO₄, H₂O and brine. The organic layer was then dried over MgSO₄, filtered and concentrated in vacuo. The crude residue was purified using flash chromatography (30% EA in hexanes) on a Biotage Isolera system to give the corresponding *tert*-butyl ((2*R*)-2-butylcyclopropoxy)carbamate **18e** as a colorless oil.

tert-butyl ((2*R*)-2-butylcyclopropoxy)carbamate (18e)

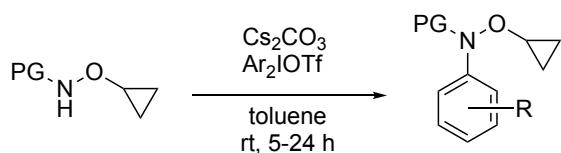
¹H NMR (600 MHz, CDCl₃) δ 7.15 (br s, 1H), 3.62 (dt, *J* = 6.5, 2.1 Hz, 1H), 1.48 (s, 9H), 1.38 – 1.28 (m, 4H), 1.16 (dqt, *J* = 11.7, 5.9, 3.8 Hz, 3H), 0.91 (ddt, *J* = 8.6, 6.0, 2.7 Hz, 1H), 0.87 (t, *J* = 7.0 Hz, 3H), 0.37 (q, *J* = 5.9 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 156.94, 81.82, 64.90, 31.17, 30.94, 28.33 (3C), 22.43, 19.53, 14.15, 12.87.

Yield: 132.0 mg, 72%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₂H₂₃NO₃Na]⁺ ([M+Na]⁺): 252.1570, found 252.1559

Preparation of *N*-arylated-*O*-cyclopropyl Hydroxamates



In an oven-dried 8 mL vial, *O*-cyclopropyl hydroxamate (0.3 mmol, 1.0 equiv) and cesium carbonate (196mg, 0.6 mmol, 2.0 equiv) were suspended in dry toluene (1.5 mL, 0.1 M). The desired diaryliodonium salt (0.45 mmol, 1.5 equiv) was added at room temperature in one portion. The mixture was stirred for 5–24 hours at room temperature, until TLC indicated complete consumption of the *O*-cyclopropyl hydroxamate starting material. Upon reaction completion, the mixture was filtered through celite. The celite was washed four times with ethyl acetate (5 ml each), the filtrate was collected, and the solvent was removed in vacuo. The crude product was purified using flash chromatography (10% EA in hexanes) on a Biotage Isolera system to give the desired *N*-arylated-*O*-cyclopropyl hydroxamate products.

Characterization of *N*-arylated-*O*-cyclopropyl Hydroxamates

tert-butyl cyclopropoxy(phenyl)carbamate (19a)

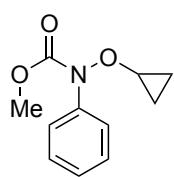
¹H NMR (600 MHz, CDCl₃) δ 7.41 (d, *J* = 7.3 Hz, 2H), 7.33 (d, *J* = 15.8 Hz, 2H), 7.16 (t, *J* = 7.4 Hz, 1H), 4.00 (tt, *J* = 6.1, 2.8 Hz, 1H), 1.53 (s, 9H), 0.87 (dp, *J* = 6.2, 3.2 Hz, 2H), 0.53 (qd, *J* = 6.3, 1.3 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 154.08, 141.53, 128.49 (2C), 125.63, 122.75 (2C), 82.23, 58.04, 28.33 (3C), 5.68 (2C).

Yield: 510 mg, 63%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₄H₁₉NO₃Na]⁺ ([M+Na]⁺): 272.1257, found 272.1268

methyl cyclopropoxy(phenyl)carbamate (19b)



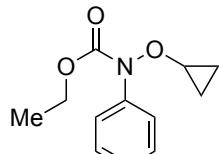
¹H NMR (600 MHz, CDCl₃) δ 7.41 (d, *J* = 7.3 Hz, 2H), 7.36 (t, *J* = 7.9 Hz, 2H), 7.22 (t, *J* = 7.3 Hz, 1H), 4.02 (tt, *J* = 6.0, 2.8 Hz, 1H), 3.82 (s, 3H), 0.85 (dt, *J* = 4.8, 3.2, 2.1 Hz, 2H), 0.56 – 0.52 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 155.68, 141.02, 128.77 (2C), 126.41, 123.31 (2C), 58.38, 53.55, 5.71 (2C).

Yield: 241 mg, 55%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₁H₁₃NO₃+H]⁺ [-H₂O]([M]): 190.0868, found 190.0857

ethyl cyclopropoxy(phenyl)carbamate (19c)



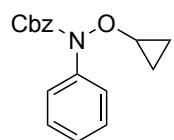
¹H NMR (600 MHz, CDCl₃) δ 7.42 (d, *J* = 8.9 Hz, 2H), 7.35 (t, *J* = 8.0 Hz, 2H), 7.20 (t, *J* = 7.2 Hz, 1H), 4.28 (q, *J* = 7.1 Hz, 2H), 4.02 (dt, *J* = 6.3, 3.3 Hz, 1H), 1.33 (t, *J* = 7.1 Hz, 3H), 0.88 – 0.84 (m, 2H), 0.56 – 0.52 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 155.21, 141.16, 128.73 (2C), 126.16, 123.07 (2C), 62.70, 58.35, 14.65, 5.74 (2C).

Yield: 254 mg, 57%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₂H₁₅NO₃Na]⁺ ([M+Na]⁺): 244.0944, found 244.0926

benzyl cyclopropoxy(phenyl)carbamate (19d)



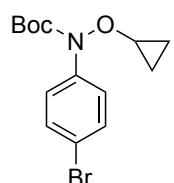
¹H NMR (600 MHz, CDCl₃) δ 7.44 – 7.41 (m, 1H), 7.39 – 7.31 (m, 4H), 7.23 – 7.19 (m, 1H), 5.26 (s, 1H), 4.02 (tt, *J* = 6.0, 2.9 Hz, 1H), 0.86 – 0.82 (m, 1H), 0.52 (ddd, *J* = 5.4, 4.1, 2.7 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 155.03, 140.96, 136.00, 128.75 (3C), 128.66 (2C), 128.35, 128.17 (2C), 126.34, 123.21, 68.11, 58.40, 5.76 (2C).

Yield: 35 mg, 62%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₇H₁₇NO₃Na]⁺ ([M+Na]⁺): 306.1101, found 306.1105

tert-butyl (4-bromophenyl)(cyclopropoxy)carbamate (19e)



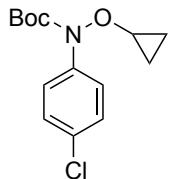
¹H NMR (600 MHz, CDCl₃) δ 7.44 (d, *J* = 8.9 Hz, 2H), 7.29 (d, *J* = 8.9 Hz, 2H), 3.98 (tq, *J* = 6.8, 4.0, 3.4 Hz, 1H), 1.52 (s, 9H), 0.91 – 0.83 (m, 2H), 0.58 – 0.51 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 153.6, 140.6, 131.4, 123.8, 118.5, 82.7, 58.2, 28.2, 5.6.

Yield: 13 mg, 12%. Colorless liquid.

HRMS (ESI): m/z calcd for [C₁₄H₁₈BrNO₃Na]⁺ ([M+Na]⁺): 350.0362; found 350.0381

tert-butyl (4-chlorophenyl)(cyclopropoxy)carbamate (19f)



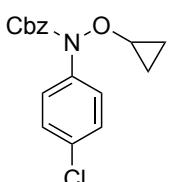
¹H NMR (600 MHz, CDCl₃) δ 7.36 – 7.33 (m, 2H), 7.31 – 7.28 (m, 2H), 4.00 – 3.96 (m, 1H), 1.52 (s, 9H), 0.85 (th, *J* = 2.8, 1.5 Hz, 2H), 0.54 (dq, *J* = 6.1, 1.2 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 153.86, 140.24, 130.87, 128.63 (2C), 123.74 (2C), 82.75, 58.31, 28.36 (3C), 5.76 (2C).

Yield: 18 mg, 21%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₄H₁₈ClNO₃Na]⁺ ([M+Na]⁺): 306.0867, found 306.0856

benzyl (4-chlorophenyl)(cyclopropoxy)carbamate (19g)



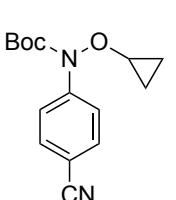
¹H NMR (600 MHz, CDCl₃) δ 7.40 – 7.35 (m, 7H), 7.33 – 7.30 (m, 2H), 5.26 (s, 2H), 4.01 (dt, *J* = 6.2, 3.2 Hz, 1H), 0.83 (tt, *J* = 5.2, 3.1 Hz, 2H), 0.55 – 0.50 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 201.79, 154.80, 139.61, 135.77, 131.56, 128.85 (2C), 128.73 (2C), 128.50, 128.27 (2C), 124.07, 68.36, 58.64, 5.80 (2C).

Yield: 35.2 mg, 38%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₇H₁₆ClNO₃Na]⁺ ([M+Na]⁺): 340.0711, found 340.0697

tert-butyl (4-cyanophenyl)(cyclopropoxy)carbamate (19h)

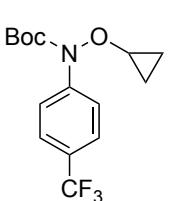


¹H NMR (600 MHz, CDCl₃): δ 7.61 (d, *J* = 8.9 Hz, 2H), 7.57 (d, *J* = 8.9 Hz, 2H), 4.02 (tt, *J* = 6.0, 2.8 Hz, 1H), 1.56 (s, 9H), 0.94 – 0.89 (m, 2H), 0.61 – 0.56 (m, 2H).

¹³C NMR (151 MHz, CDCl₃): δ 152.8, 145.2, 132.5, 120.5, 118.8, 107.5, 83.6, 58.7, 28.2, 5.7.

Yield: 23 mg, 27%. Colorless oil.

tert-butyl cyclopropoxy(4-(trifluoromethyl)phenyl)carbamate (19i)



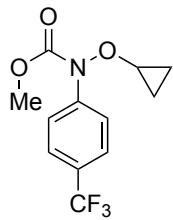
¹H NMR (600 MHz, CDCl₃): δ 7.57 (q, *J* = 8.9 Hz, 4H), 4.02 (s, 1H), 1.55 (s, 9H), 0.90 (s, 2H), 0.57 (q, *J* = 6.0 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃): δ 153.3, 144.5, 126.6 (q, *J* = 32.7 Hz), 125.58 (q, *J* = 3.7 Hz), 124.4 (q, *J* = 271.6 Hz), 120.9, 83.1, 58.5, 28.2, 5.7.

Yield: 39 mg, 41%. Colorless liquid.

HRMS (ESI): m/z calcd for [C₁₅H₁₈F₃NO₃Na]⁺ ([M+Na]⁺): 340.1131; found 340.1150

methyl cyclopropoxy(4-(trifluoromethyl)phenyl)carbamate (19j)



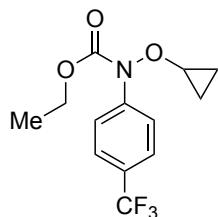
¹H NMR (600 MHz, CDCl₃) δ 7.59 (q, *J* = 9.1 Hz, 4H), 4.05 (tt, *J* = 6.1, 2.8 Hz, 1H), 3.87 (s, 3H), 0.88 (qt, *J* = 3.9, 2.2 Hz, 2H), 0.57 (dq, *J* = 6.2, 1.2 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 155.06, 144.08, 127.34 (q, *J* = 32.8 Hz), 125.91 (2C, q, *J* = 3.8 Hz), 125.04 (q, *J* = 286.9 Hz), 121.31 (2C), 58.99, 53.83, 5.79 (2C).

Yield: 165.6 mg, 60%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₂H₁₃F₃NO₃]⁺ ([M+H]⁺): 276.0842, found 276.0859

ethyl cyclopropoxy(4-(trifluoromethyl)phenyl)carbamate (19k)



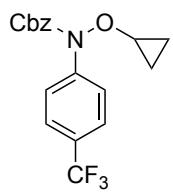
¹H NMR (600 MHz, CDCl₃) δ 7.63 – 7.55 (m, 4H), 4.32 (q, *J* = 7.1 Hz, 2H), 4.05 (tt, *J* = 6.0, 2.8 Hz, 1H), 1.37 (t, *J* = 7.1 Hz, 3H), 0.90 (q, *J* = 5.8, 5.2 Hz, 2H), 0.60 – 0.54 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 154.57, 144.19, 127.15 (q, *J* = 32.7 Hz), 125.87 (2C, q, *J* = 3.7 Hz), 124.18 (q, *J* = 271.8 Hz), 121.14 (2C), 63.16, 58.92, 14.56, 5.82 (2C).

Yield: 39.0 mg, 67%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₃H₁₅F₃NO₃]⁺ ([M+H]⁺): 290.0999, found 290.1002

benzyl cyclopropoxy(4-(trifluoromethyl)phenyl)carbamate (19l)



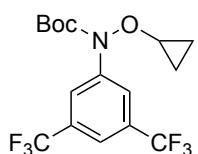
¹H NMR (600 MHz, CDCl₃) δ 7.61 (s, 4H), 7.45 – 7.34 (m, 5H), 5.30 (s, 2H), 4.06 (tt, *J* = 6.1, 2.8 Hz, 1H), 0.92 – 0.86 (m, 2H), 0.56 (h, *J* = 5.2 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 154.33, 143.93, 135.45, 128.68 (2C), 128.53, 128.29 (2C), 127.18 (q, *J* = 32.7 Hz), 125.79 (2C, q, *J* = 3.7 Hz), 124.04 (q, *J* = 271.8 Hz), 121.14 (2C), 68.50, 58.90, 5.74 (2C).

Yield: 69.2 mg, 66%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₈H₁₆F₃NO₃Na]⁺ ([M+Na]⁺): 374.0974, found 374.0985

tert-butyl (3,5-bis(trifluoromethyl)phenyl)(cyclopropoxy)carbamate (19m)



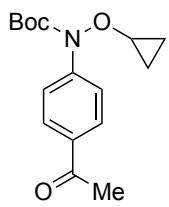
¹H NMR (600 MHz, CDCl₃) δ 7.93 (s, 2H), 7.62 (s, 1H), 4.03 (tt, *J* = 6.0, 2.8 Hz, 1H), 1.56 (s, 9H), 0.99 – 0.88 (m, 2H), 0.61 (q, *J* = 6.7, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 152.8, 143.0, 131.8 (q, *J* = 33.5 Hz), 123.2 (q, *J* = 272.7 Hz), 120.6 (q, *J* = 3.8 Hz), 117.9 (pent, *J* = 3.8 Hz), 83.9, 58.9, 28.1, 5.8.

Yield: 72 mg, 62%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₆H₁₇F₆NO₃Na]⁺ ([M+Na]⁺): 408.1005; found 408.1018

tert-butyl (3-acetylphenyl)(cyclopropoxy)carbamate (19n)



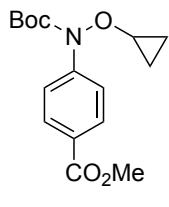
¹H NMR (600 MHz, CDCl₃): δ 7.93 (d, *J* = 8.8 Hz, 2H), 7.53 (d, *J* = 8.8 Hz, 2H), 4.03 (tt, *J* = 6.0, 2.8 Hz, 1H), 2.57 (s, 3H), 1.55 (s, 9H), 0.96 – 0.87 (m, 2H), 0.56 (tt, *J* = 6.7, 3.2 Hz, 2H).

¹³C NMR (151 MHz, CDCl₃): δ 196.9, 152.9, 145.4, 133.2, 128.8, 120.2, 83.0, 58.5, 28.1, 26.4, 5.6.

Yield: 41 mg, 47%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₆H₂₁NO₄Na]⁺ ([M+Na]⁺): 314.1363; found 314.1379

tert-butyl cyclopropoxy(4-(trifluoromethyl)phenyl)carbamate (19o)



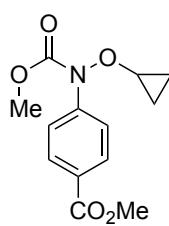
¹H NMR (600 MHz, CDCl₃): δ 8.00 (d, *J* = 8.7 Hz, 2H), 7.51 (d, *J* = 8.7 Hz, 2H), 4.02 (tt, *J* = 5.9, 2.7 Hz, 1H), 3.90 (s, 3H), 1.55 (s, 9H), 0.91 (d, *J* = 7.6 Hz, 2H), 0.55 (q, *J* = 6.1 Hz, 2H);

¹³C NMR (151 MHz, CDCl₃): δ 166.6, 153.1, 145.4, 130.0, 126.1, 120.3, 82.9, 58.5, 52.0, 28.2, 5.6.

Yield: 42 mg, 45%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₆H₂₁NO₅Na]⁺ ([M+Na]⁺): 330.1312; found 330.1333

methyl 4-(cyclopropoxy(methoxycarbonyl)amino)benzoate (19p)



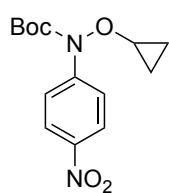
¹H NMR (600 MHz, CDCl₃): δ 8.01 (dd, *J* = 8.8, 2.9 Hz, 2H), 7.52 (dd, *J* = 8.8, 2.8 Hz, 2H), 4.04 (tq, *J* = 6.1, 3.0 Hz, 1H), 3.89 (d, *J* = 3.1 Hz, 3H), 3.85 (d, *J* = 3.0 Hz, 3H), 0.90 – 0.85 (m, 2H), 0.57 – 0.52 (m, 2H).

¹³C NMR (151 MHz, CDCl₃): δ 166.57, 154.86, 144.94, 130.28 (2C), 126.79, 120.60 (2C), 58.94, 53.75, 52.16, 5.74 (2C).

Yield: 46.3 mg, 56%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₃H₁₅NO₅Na]⁺ ([M+Na]⁺): 288.0842; found 288.0838

tert-butyl cyclopropoxy(4-nitrophenyl)carbamate (19q)



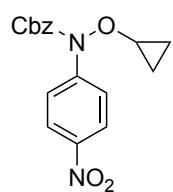
¹H NMR (600 MHz, CDCl₃): δ 8.20 (d, *J* = 9.2 Hz, 2H), 7.63 (d, *J* = 9.2 Hz, 2H), 4.08 – 4.03 (m, 1H), 1.58 (s, 9H), 0.94 (tq, *J* = 5.4, 3.2 Hz, 2H), 0.63 – 0.58 (m, 2H).

¹³C NMR (151 MHz, CDCl₃): δ 152.77, 147.06, 143.95, 124.41 (2C), 120.03 (2C), 83.99, 59.06, 28.32 (3C), 5.92 (2C).

Yield: 195.0 mg, 66%. Yellow solid (m.p. 58.2 °C).

HRMS (ESI): m/z calcd for [C₁₄H₁₈N₂O₅Na]⁺ ([M+Na]⁺): 317.1108; found 317.1080

benzyl cyclopropoxy(4-nitrophenyl)carbamate (19r)



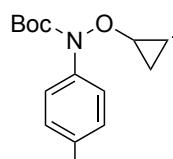
¹H NMR (600 MHz, CDCl₃) δ 8.22 (d, *J* = 9.2 Hz, 2H), 7.66 (d, *J* = 9.2 Hz, 2H), 7.45 – 7.35 (m, 5H), 5.31 (s, 2H), 4.07 (tt, *J* = 6.1, 2.9 Hz, 1H), 0.94 – 0.87 (m, 2H), 0.61 – 0.55 (m, 2H).

¹³C NMR (151 MHz, CDCl₃) δ 153.96, 146.49, 144.32, 135.22, 128.89 (2C), 128.85, 128.55 (2C), 124.53 (2C), 120.21 (2C), 69.02, 59.49, 5.98 (2C).

Yield: 283.4 mg, 86%. Yellow solid (m.p. 86.6 °C).

HRMS (ESI): m/z calcd for [C₁₇H₁₆N₂O₅Na]⁺ ([M+Na]⁺): 351.0951, found 351.0939

tert-butyl ((2*R*)-2-butylcyclopropoxy)(4-(trifluoromethyl)phenyl)carbamate (19s)



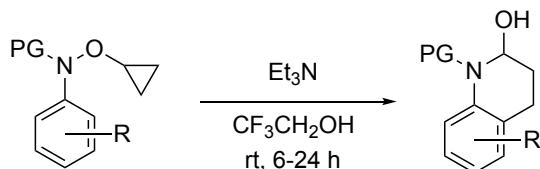
¹H NMR (600 MHz, CDCl₃) δ 7.58 (d, *J* = 8.7 Hz, 2H), 7.54 (d, *J* = 8.7 Hz, 2H), 3.71 (dt, *J* = 6.2, 2.3 Hz, 1H), 1.55 (s, 9H), 1.31 – 1.19 (m, 6H), 1.07 – 0.98 (m, 2H), 0.88 – 0.84 (m, 3H), 0.39 (q, *J* = 6.2 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 153.2, 144.4, 126.6 (q, *J* = 32.7 Hz, 2C), 125.6 (q, *J* = 3.7 Hz), 124.1 (q, *J* = 271.6 Hz), 120.9 (2C), 83.0, 64.4, 31.0, 30.8, 28.2 (3C), 22.2, 19.1, 14.0, 12.7.

Yield: 50 mg, 45%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₉H₂₆F₃NO₃Na]⁺ ([M+Na]⁺): 396.1757; found 396.1736

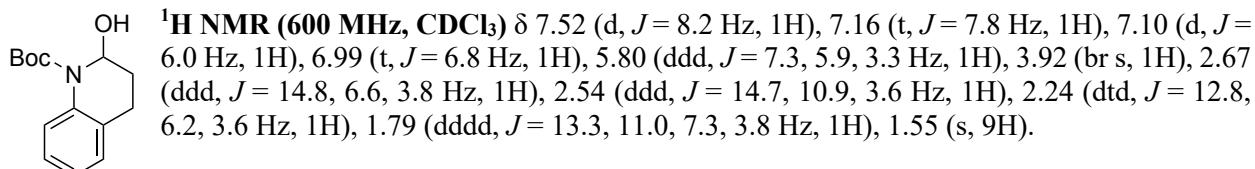
Preparation of 2-Hydroxy-tetrahydroquinolines via [3,3]-Rearrangement of *N*-arylated-O-cyclopropyl Hydroxamates



N-arylated-*O*-cyclopropyl hydroxamate (50 mg, 0.2 mmol, 1.0 equiv) was added to an 8 mL oven-dried vial equipped with a stir bar. The vial was capped, placed under argon atmosphere and trifluoroethanol (2.0 mL, 0.1M) was added. Triethylamine (55.8 µL, 0.4 mmol, 2.0 equiv) was then added to the solution via syringe. The reaction mixture was stirred at rt for 6-24 hours. After complete consumption of the starting material was confirmed via TLC, the reaction mixture was concentrated under reduced pressure. The crude mixture was then purified using flash chromatography (12 % EA in hexanes) on a Biotage Isolera system to give the desired tetrahydroquinoline products.

Characterization of 2-hydroxy-tetrahydroquinolines

tert-butyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20a)

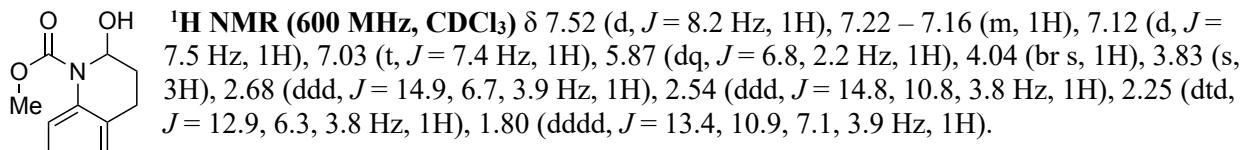


¹³C NMR (151 MHz, CDCl₃) δ 154.79, 136.48, 132.13, 127.34, 126.37, 123.94, 123.65, 82.21, 79.15, 31.16, 28.53 (3C), 24.75.

Yield: 276 mg, 60%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₄H₂₀NO₃]⁺ ([M+H]⁺): 250.1438; found 250.1440

methyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20b)

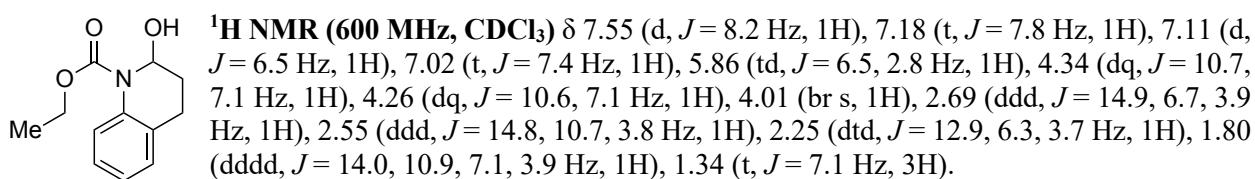


¹³C NMR (151 MHz, CDCl₃) δ 156.12, 135.91, 132.20, 127.39, 126.54, 124.13, 123.84, 79.09, 53.18, 31.08, 24.52.

Yield: 65 mg, 46%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₁H₁₄NO₃]⁺ ([M+H]⁺): 208.0968, found 208.0961

ethyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20c)

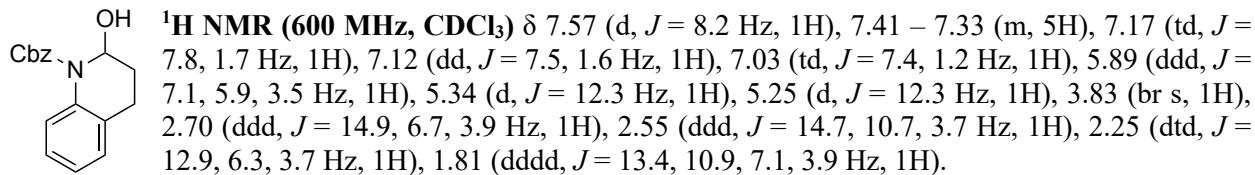


¹³C NMR (151 MHz, CDCl₃) δ 155.69, 136.04, 132.14, 127.39, 126.50, 123.97, 123.85, 79.06, 62.39, 31.05, 24.56, 14.55.

Yield: 25.2 mg, 57%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₂H₁₅NO₃Na]⁺ ([M+Na]⁺): 244.0944, found 244.0924

benzyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20d)

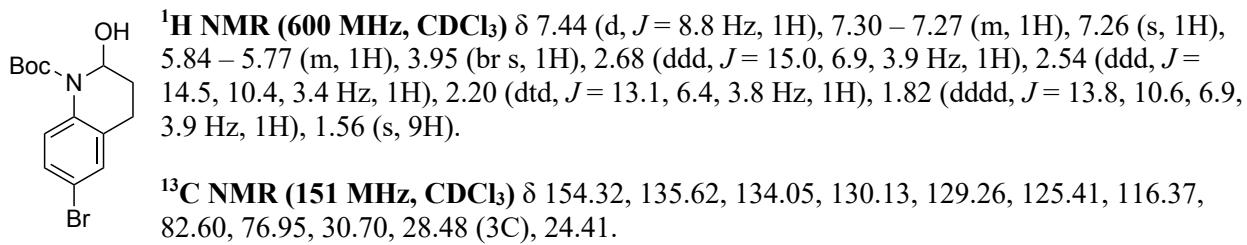


¹³C NMR (151 MHz, CDCl₃) δ 155.50, 135.94, 135.87, 132.19, 128.80 (2C), 128.52, 128.31(2C), 127.45, 126.61, 124.16, 123.91, 79.29, 68.08, 31.06, 24.57.

Yield: 18.6 mg, 53%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₇H₁₇NO₃Na]⁺ ([M+Na]⁺): 306.1101, found 306.1107

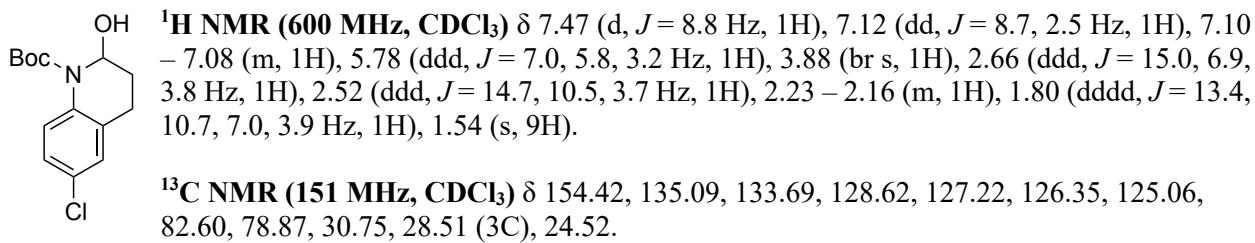
tert-butyl 6-bromo-2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20e)



Yield: 76.8 mg, 52%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₄H₁₉BrNO₃]⁺ ([M+H]⁺): 328.0543; found 328.0534

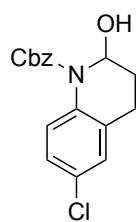
tert-butyl 6-chloro-2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20f)



Yield: 27 mg, 60%. White solid (m.p. 108.4 °C).

HRMS (ESI): m/z calcd for [C₁₄H₁₈ClNO₃Na]⁺ ([M+Na]⁺): 306.0867, found 306.0857

benzyl 6-chloro-2-hydroxy-3,4-dihydroquinoline-1(2H)-carboxylate (20g)



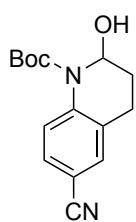
¹H NMR (600 MHz, CDCl₃) δ 7.55 – 7.49 (m, 1H), 7.41 – 7.37 (m, 4H), 7.37 – 7.34 (m, 1H), 7.14 – 7.09 (m, 2H), 5.87 (ddd, *J* = 6.8, 5.7, 3.5 Hz, 1H), 5.32 (d, *J* = 12.2 Hz, 1H), 5.25 (d, *J* = 12.2 Hz, 1H), 3.76 (br s, 1H), 2.69 (ddd, *J* = 15.0, 7.1, 3.9 Hz, 1H), 2.53 (ddd, *J* = 14.7, 10.2, 3.7 Hz, 1H), 2.20 (dddd, *J* = 13.1, 7.0, 5.7, 3.8 Hz, 1H), 1.82 (dddd, *J* = 13.8, 10.5, 6.8, 3.9 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃) 155.16, 135.64, 134.57, 133.69, 129.16, 128.89 (2C), 128.69, 128.39 (2C), 127.37, 126.62, 125.00, 79.02, 68.29, 30.61, 24.34.

Yield: 15.4 mg, 22%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₇H₁₆ClNO₃Na]⁺ ([M+Na]⁺): 340.0711, found 340.0693

tert-butyl 6-cyano-2-hydroxy-3,4-dihydroquinoline-1(2H)-carboxylate (20h)



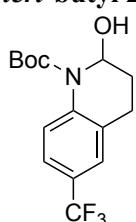
¹H NMR (600 MHz, CDCl₃): δ 7.71 (d, *J* = 8.6 Hz, 1H), 7.44 (dd, *J* = 8.6, 1.8 Hz, 1H), 7.40 (s, 1H), 5.81 (td, *J* = 5.9, 3.5 Hz, 1H), 3.91 (d, *J* = 3.0 Hz, 1H), 2.78 (ddd, *J* = 15.0, 7.9, 3.9 Hz, 1H), 2.58 (ddd, *J* = 14.7, 9.5, 3.7 Hz, 1H), 2.14 (ddt, *J* = 13.2, 8.5, 4.6 Hz, 1H), 1.89 (tdd, *J* = 13.4, 6.4, 3.9 Hz, 1H), 1.56 (s, 9H).

¹³C NMR (151 MHz, CDCl₃): δ 153.8, 140.7, 132.1, 131.2, 130.3, 123.7, 119.0, 106.3, 83.3, 78.6, 29.9, 28.3 (3C), 23.9.

Yield: 54 mg, 66%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₅H₁₉N₂O₃]⁺ ([M+H]⁺): 275.1390; found 275.1379

tert-butyl 2-hydroxy-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2H)-carboxylate (20i)



¹H NMR (600 MHz, CDCl₃): δ 7.68 (d, *J* = 8.5 Hz, 1H), 7.41 (d, *J* = 8.4 Hz, 1H), 7.36 (s, 1H), 5.82 (q, *J* = 6.0 Hz, 1H), 3.92 (s, 1H), 2.77 (ddd, *J* = 14.5, 6.9, 3.6 Hz, 1H), 2.65 – 2.53 (m, 1H), 2.25 – 2.12 (m, 1H), 1.86 (dtt, *J* = 13.5, 6.9, 3.9 Hz, 1H), 1.57 (s, 9H).

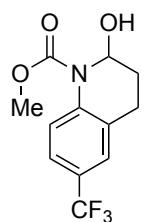
¹³C NMR (151 MHz, CDCl₃): δ 154.2, 139.6, 131.9, 125.2 (q, *J* = 32.6 Hz), 124.4 (q, *J* = 3.6 Hz), 124.3 (q, *J* = 242.3 Hz), 123.5, 123.3 (q, *J* = 3.7 Hz), 82.9, 78.8, 30.3, 28.4 (3C), 24.3.

¹⁹F NMR (471 MHz, CDCl₃) –62.03 (s).

Yield: 74 mg, 78%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₅H₁₈F₃NO₃Na]⁺ ([M+Na]⁺): 340.1131; found 340.1101

methyl 2-hydroxy-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (20j)



¹H NMR (600 MHz, CDCl₃) δ 7.71 (d, *J* = 8.6 Hz, 1H), 7.44 (dd, *J* = 8.6, 2.2 Hz, 1H), 7.38 (s, 1H), 5.88 (t, *J* = 6.0 Hz, 1H), 3.90 (br s, 1H), 3.87 (s, 3H), 2.80 (ddd, *J* = 15.2, 7.7, 4.0 Hz, 1H), 2.60 (ddd, *J* = 14.6, 9.7, 3.8 Hz, 1H), 2.20 (dddd, *J* = 13.3, 7.7, 5.5, 3.9 Hz, 1H), 1.89 (dddd, *J* = 13.6, 10.2, 6.6, 4.0 Hz, 1H).

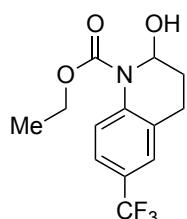
¹³C NMR (151 MHz, CDCl₃) δ 155.74, 139.18, 132.03, 125.83 (q, *J* = 32.6 Hz), 124.67 (q, *J* = 3.9 Hz), 124.26 (q, *J* = 271.6 Hz), 123.75 (q, *J* = 3.7 Hz), 123.58, 78.95, 53.55, 30.33, 24.27.

¹⁹F NMR (471 MHz, CDCl₃) –62.12 (s).

Yield: 25.3 mg, 52%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₂H₁₂F₃NO₃K]⁺ ([M+K]⁺): 314.0401, found 314.0296

ethyl 2-hydroxy-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (20k)



¹H NMR (600 MHz, CDCl₃) δ 7.73 (d, *J* = 8.6 Hz, 1H), 7.44 (dd, *J* = 8.7, 2.2 Hz, 1H), 7.38 (s, 1H), 5.88 (ddd, *J* = 6.5, 5.4, 3.4 Hz, 1H), 4.40 – 4.28 (m, 2H), 3.89 (br s, 1H), 2.80 (ddd, *J* = 15.1, 7.6, 3.9 Hz, 1H), 2.61 (ddd, *J* = 14.6, 9.8, 3.8 Hz, 1H), 2.20 (dddd, *J* = 13.2, 7.6, 5.5, 3.8 Hz, 1H), 1.93 – 1.84 (m, 1H), 1.37 (t, *J* = 7.1 Hz, 3H).

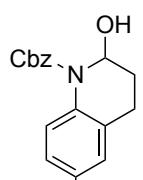
¹³C NMR (151 MHz, CDCl₃) δ 155.30, 139.30, 132.01, 125.69 (q, *J* = 32.6 Hz), 124.64 (q, *J* = 3.8 Hz), 124.30 (q, *J* = 271.6 Hz), 123.57, 123.70 (q, *J* = 3.7 Hz), 78.92, 62.89, 30.33, 24.32, 14.57.

¹⁹F NMR (471 MHz, CDCl₃) –62.09 (s).

Yield: 20.0 mg, 51%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₃H₁₄F₃NO₃Na]⁺ ([M+Na]⁺): 312.0818, found 312.0808

benzyl 2-hydroxy-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (20l)



¹H NMR (600 MHz, CDCl₃) δ 7.74 (d, *J* = 8.6 Hz, 1H), 7.51 – 7.33 (m, 7H), 5.91 (td, *J* = 6.0, 3.4 Hz, 1H), 5.34 (d, *J* = 12.2 Hz, 1H), 5.28 (d, *J* = 12.2 Hz, 1H), 3.85 (br s, 1H), 2.80 (ddd, *J* = 15.2, 7.8, 3.9 Hz, 1H), 2.61 (ddd, *J* = 14.6, 9.7, 3.8 Hz, 1H), 2.19 (dp, *J* = 13.2, 4.3 Hz, 1H), 1.89 (dddd, *J* = 13.6, 10.0, 6.5, 3.9 Hz, 1H).

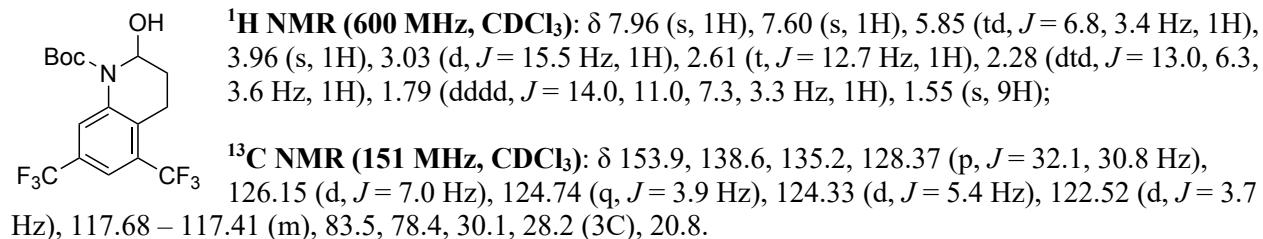
¹³C NMR (151 MHz, CDCl₃) δ 155.06, 139.15, 135.46, 132.00, 128.93 (2C), 128.79, 128.48 (2C), 125.81 (q, *J* = 32.6 Hz), 124.68 (q, *J* = 3.8 Hz), 124.27 (q, *J* = 271.7 Hz), 123.72 (q, *J* = 3.8 Hz), 123.61, 78.95, 68.50, 30.29, 24.22.

¹⁹F NMR (471 MHz, CDCl₃) –62.10 (s).

Yield: 69.2 mg, 65%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₈H₁₆F₃NO₃Na]⁺ ([M+Na]⁺): 374.0974, found 374.0969

tert-butyl 2-hydroxy-5,7-bis(trifluoromethyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (20m)

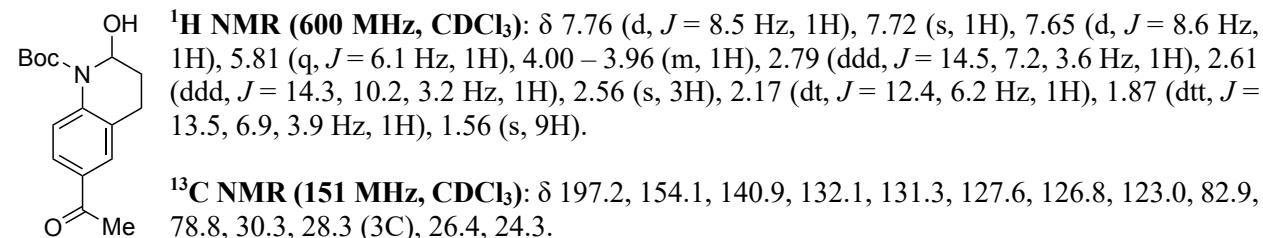


¹⁹F NMR (471 MHz, CDCl₃) –60.39 (s), -62.96 (s).

Yield: 60 mg, 52%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₆H₁₈F₆NO₃]⁺ ([M+H]⁺): 386.1185; found 386.1190

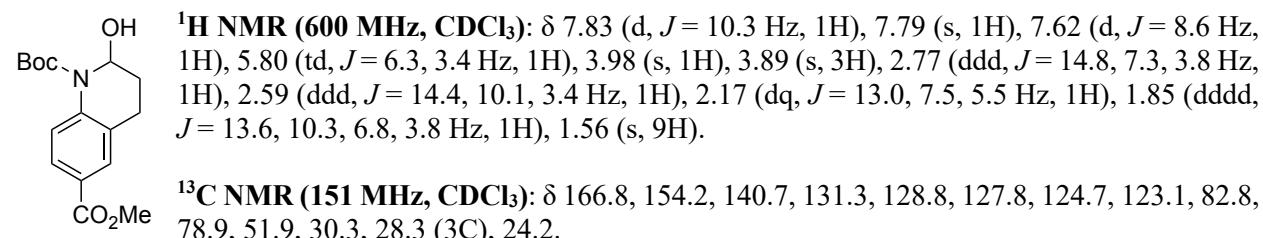
tert-butyl 7-acetyl-2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (20n)



Yield: 58 mg, 67%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₆H₂₂NO₄]⁺ ([M+H]⁺): 292.1543; found 292.1542

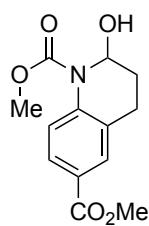
1-(tert-butyl) 6-methyl 2-hydroxy-3,4-dihydroquinoline-1,6(2*H*)-dicarboxylate (20o)



Yield: 63 mg, 68%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₆H₂₂NO₅]⁺ ([M+H]⁺): 308.1492; found 308.1480

dimethyl 2-hydroxy-3,4-dihydroquinoline-1,6(2H)-dicarboxylate (20p)



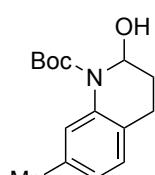
¹H NMR (600 MHz, CDCl₃) δ 7.84 (d, *J* = 8.6 Hz, 1H), 7.80 (s, 1H), 7.66 (d, *J* = 8.6 Hz, 1H), 5.87 (td, *J* = 6.1, 3.0 Hz, 1H), 3.97 (br s, 1H), 3.88 (s, 3H), 3.86 (s, 3H), 2.79 (ddd, *J* = 15.1, 7.7, 4.0 Hz, 1H), 2.59 (ddd, *J* = 14.5, 9.7, 3.8 Hz, 1H), 2.18 (dddd, *J* = 13.3, 7.7, 5.4, 3.8 Hz, 1H), 1.90 – 1.83 (m, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 166.87, 155.79, 140.33, 131.42, 129.08, 128.24, 125.37, 123.11, 79.12, 53.53, 52.14, 30.36, 24.20.

Yield: 35 mg, 45%. Yellow oil.

HRMS (ESI): m/z calcd for [C₁₃H₁₅NO₅Na]⁺ ([M+Na]⁺): 288.0842, found 288.0838

tert-butyl 2-hydroxy-7-methyl-3,4-dihydroquinoline-1(2H)-carboxylate (20q)



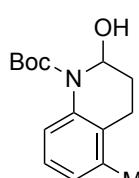
¹H NMR (600 MHz, CDCl₃): δ 7.37 (s, 1H), 6.98 (d, *J* = 7.6 Hz, 1H), 6.81 (d, *J* = 7.6 Hz, 1H), 5.78 (td, *J* = 7.2, 3.2 Hz, 1H), 3.88 (s, 1H), 2.63 (ddd, *J* = 14.7, 6.4, 3.9 Hz, 1H), 2.50 (ddd, *J* = 14.6, 11.0, 3.4 Hz, 1H), 2.31 (s, 3H), 2.22 (dtd, *J* = 12.7, 6.2, 3.7 Hz, 1H), 1.81 – 1.72 (m, 1H), 1.56 (s, 9H).

¹³C NMR (151 MHz, CDCl₃): δ 154.6, 136.1, 135.8, 128.9, 126.9, 124.4, 124.2, 81.9, 78.9, 31.1, 28.4 (3C), 24.1, 21.3.

Yield: 16 mg, 20%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₅H₂₂NO₃]⁺ ([M+H]⁺): 264.1594; found 264.1577

tert-butyl 2-hydroxy-5-methyl-3,4-dihydroquinoline-1(2H)-carboxylate (20r)



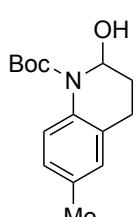
¹H NMR (600 MHz, CDCl₃): δ 7.33 (d, *J* = 8.2 Hz, 1H), 7.06 (t, *J* = 7.9 Hz, 1H), 6.89 (d, *J* = 7.5 Hz, 1H), 5.83 (td, *J* = 7.0, 3.2 Hz, 1H), 3.74 (s, 1H), 2.74 (ddd, *J* = 15.3, 6.6, 4.2 Hz, 1H), 2.40 (ddd, *J* = 14.9, 10.6, 3.8 Hz, 1H), 2.28 (s, 3H), 2.24 (dtd, *J* = 13.0, 6.2, 4.1 Hz, 1H), 1.76 (dddd, *J* = 13.4, 11.0, 7.2, 4.1 Hz, 1H), 1.54 (s, 9H).

¹³C NMR (151 MHz, CDCl₃): δ 154.7, 136.1, 134.5, 130.6, 125.4, 125.3, 122.1, 81.9, 78.5, 30.6, 28.4 (3C), 20.5, 19.6.

Yield: 18 mg, 22%. Colorless oil.

HRMS (ESI): m/z calcd for [C₁₅H₂₁NO₃Na]⁺ ([M+Na]⁺): 286.1414; found 286.1418

tert-butyl 2-hydroxy-6-methyl-3,4-dihydroquinoline-1(2H)-carboxylate (20s)



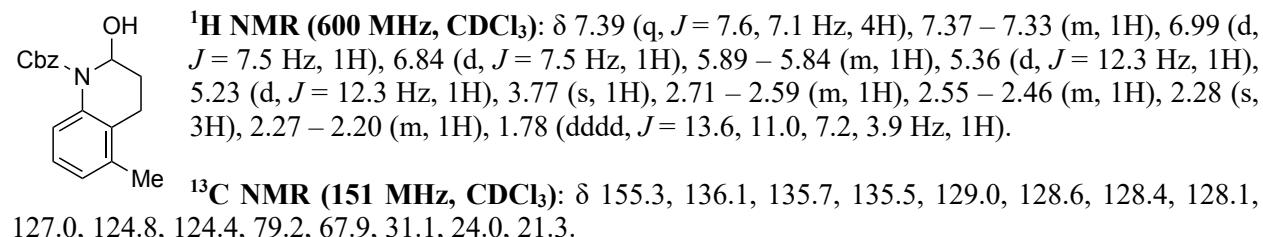
¹H NMR (600 MHz, CDCl₃): δ 7.40 (d, *J* = 8.3 Hz, 1H), 6.97 (d, *J* = 8.3 Hz, 1H), 6.91 (s, 1H), 5.78 (td, *J* = 7.2, 3.3 Hz, 1H), 3.90 (s, 1H), 2.62 (ddd, *J* = 14.7, 6.3, 3.9 Hz, 1H), 2.51 (ddd, *J* = 14.6, 11.1, 3.4 Hz, 1H), 2.29 (s, 3H), 2.26 – 2.18 (m, 1H), 1.77 (dddd, *J* = 13.3, 11.1, 7.3, 3.8 Hz, 1H), 1.55 (s, 9H).

^{13}C NMR (151 MHz, CDCl_3): δ 154.7, 133.8, 133.0, 131.8, 127.8, 126.8, 123.6, 81.9, 79.0, 31.1, 28.4 (3C), 24.6, 20.7.

Yield: 30 mg, 38%. Colorless oil.

HRMS (ESI): m/z calcd for $[\text{C}_{15}\text{H}_{21}\text{NO}_3\text{Na}]^+$ ($[\text{M}+\text{Na}]^+$): 286.1414, found 286.1404

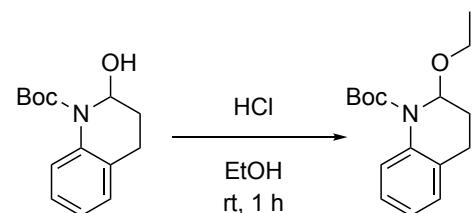
benzyl 2-hydroxy-5-methyl-3,4-dihydroquinoline-1(2*H*)-carboxylate (20t)



Yield: 37 mg, 41%. Colorless oil.

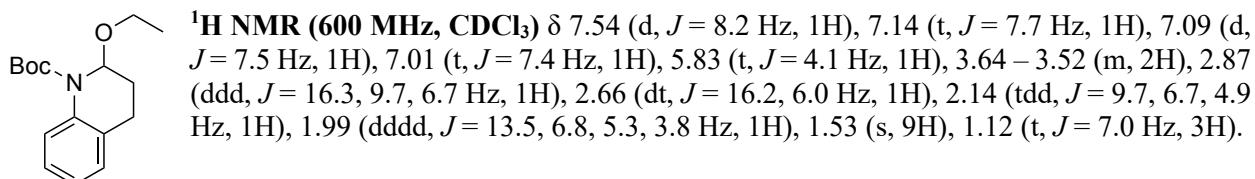
HRMS (ESI): m/z calcd for $[\text{C}_{18}\text{H}_{19}\text{NO}_3\text{Na}]^+$ ($[\text{M}+\text{Na}]^+$): 320.1257; found 320.1243

Preparation of Derivatized 2-hydroxy-tetrahydroquinoline Products



Tert-butyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **20a** (50 mg, 0.2 mmol, 1.0 equiv) was dissolved in EtOH (1 mL, 0.2 M). At room temperature, 2M HCl in Et_2O (200 μL , 0.4 mmol, 2.0 equiv) was added in two equal portions (100 μL each). The reaction was left stirring at rt for 1 h. Upon reaction completion the mixture was quenched with aq. NaHCO_3 (2 mL). The organic layer was separated, washed with brine, dried over MgSO_4 and concentrated. The crude residue was purified using flash chromatography (20% EA in hexanes) on a Biotage Isolera system to give the corresponding *tert*-butyl 2-ethoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **21** as a colorless liquid.

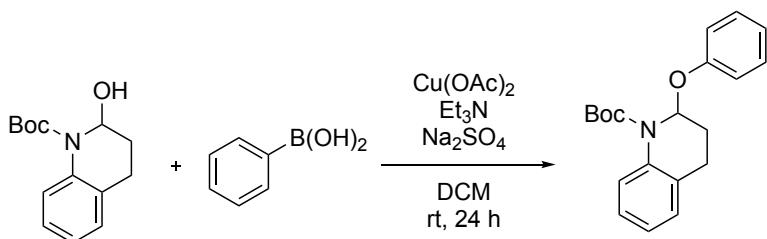
***tert*-butyl 2-ethoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (21)**



¹³C NMR (151 MHz, CDCl₃) δ 153.81, 136.12, 130.28, 128.28, 125.81, 125.06, 123.92, 81.41, 81.26, 62.79, 29.25, 28.48 (3C), 23.37, 15.15.

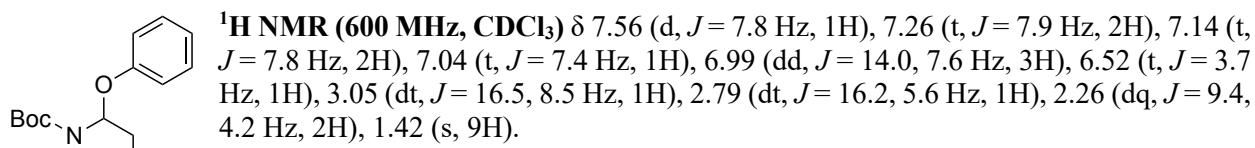
Yield: 33.2 mg, 60%. Colorless liquid.

HRMS (ESI): m/z calcd for [C₁₆H₂₃NO₃Na]⁺ ([M+Na]⁺): 300.1570, found 300.1568



Tert-butyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **20a** (50 mg, 0.2 mmol, 1.0 equiv) was dissolved in DCM (2 mL, 0.1 M). To this solution at rt was added Cu(OAc)₂ (36 mg, 0.2 mmol, 1.0 equiv), Et₃N (101 mg, 1 mmol, 5.0 equiv), Na₂SO₄ (142 mg, 1 mmol, 5.0 equiv) and phenylboronic acid (49 mg, 0.4 mmol, 2.0 equiv), in that order. The mixture was stirred at rt for 24 hours.^[5] Upon reaction completion, the mixture was then passed through a pad of celite and washed three times with DCM (5 mL). The combined organic washes were concentrated, and the crude residue was purified using flash chromatography (20-30% EA in hexanes) on a Biotage Isolera system to give the corresponding *tert*-butyl 2-phenoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **22** as a colorless oil.

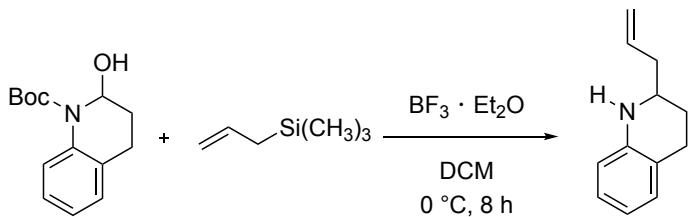
***tert*-butyl 2-phenoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (22)**



¹³C NMR (151 MHz, CDCl₃) δ 156.6, 152.8, 135.7, 129.5, 129.3, 128.2, 125.9, 124.7, 123.9, 122.1, 117.7, 81.6, 81.2, 28.5, 28.1, 23.0.

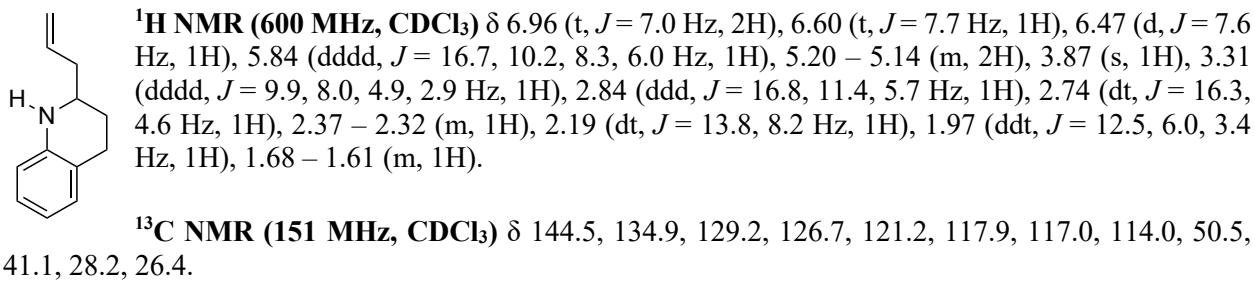
Yield: 98 mg, 39%. Colorless oil.

HRMS (ESI): m/z calcd for [C₂₀H₂₄NO₃]⁺ ([M+H]⁺): 326.1751; found 326.1742



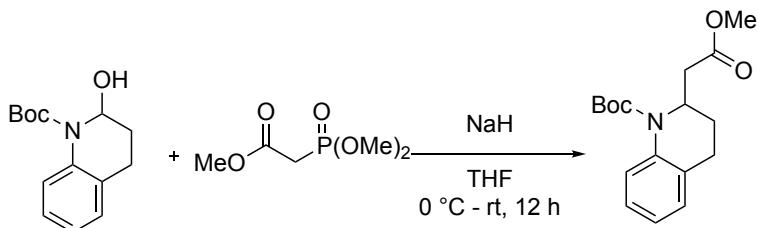
Tert-butyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **20a** (50 mg, 0.2 mmol, 1.0 equiv) was dissolved in DCM (2 mL, 0.1 M). Then allyltrimethylsilane (39 mg, 0.34 mmol, 1.7 equiv) was added. The solution was cooled to 0 °C and $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (48 mg, 0.34 mmol, 1.7 equiv) was added to the solution dropwise. After stirring for 8 hours at 0 °C, the reaction was removed from the ice bath and allowed to warm to rt.^[6] The mixture was concentrated to give the corresponding *tert*-butyl 2-(allyloxy)-3,4-dihydroquinoline-1(2*H*)-carboxylate **23** as a pure colorless oil.

tert-butyl 2-(allyloxy)-3,4-dihydroquinoline-1(2*H*)-carboxylate (**23**)



Yield: 33 mg, 63%. Colorless oil.

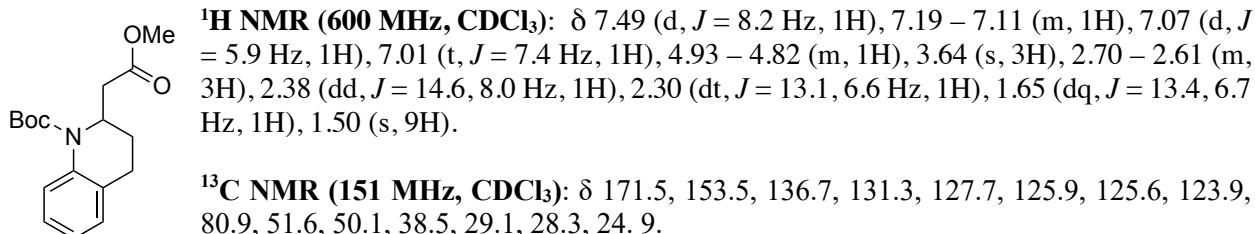
HRMS (ESI): m/z calcd for $[\text{C}_{12}\text{H}_{16}\text{N}]^+$ ($[\text{M}+\text{H}]^+$): 174.1277; found 174.1289



Methyl 2-(dimethoxyphosphoryl)acetate (109 mg, 0.6 mmol, 2.0 equiv) was dissolved in THF (2 mL) and cooled to 0 °C. NaH (0.6 mmol, 2.0 equiv) was slowly added to the solution. Then a *tert*-butyl 2-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **20a** (75 mg, 0.3 mmol, 1.0 equiv) solution in THF (1.0 mL, 0.3 M), was added dropwise. The reaction was allowed to warm to room temperature and stirred overnight. Upon reaction completion, water was added to quench the reaction. The aqueous layer was extracted with DCM (3 x 5 mL). The combined organic layers were dried over anhydrous sodium sulfate and

concentrated.^[7] The crude mixture was purified using flash chromatography (10-20% EA in hexanes) on a Biotage Isolera system to give the corresponding *tert*-butyl 2-(2-oxopropoxy)-3,4-dihydroquinoline-1(2*H*)-carboxylate **24** as a colorless oil.

***tert*-butyl 2-(2-oxopropoxy)-3,4-dihydroquinoline-1(2*H*)-carboxylate (24)**



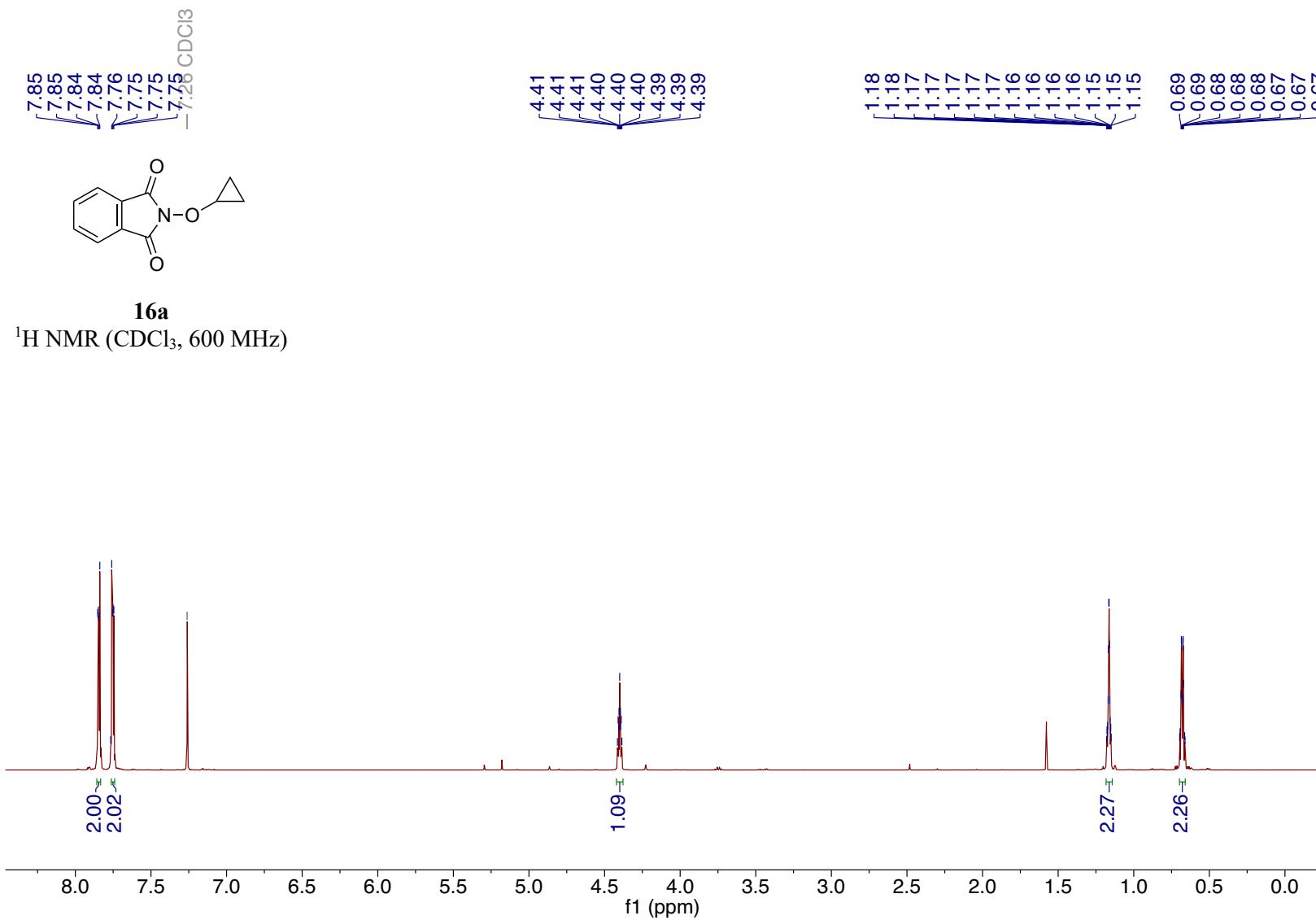
Yield: 63 mg, 69%. Colorless oil.

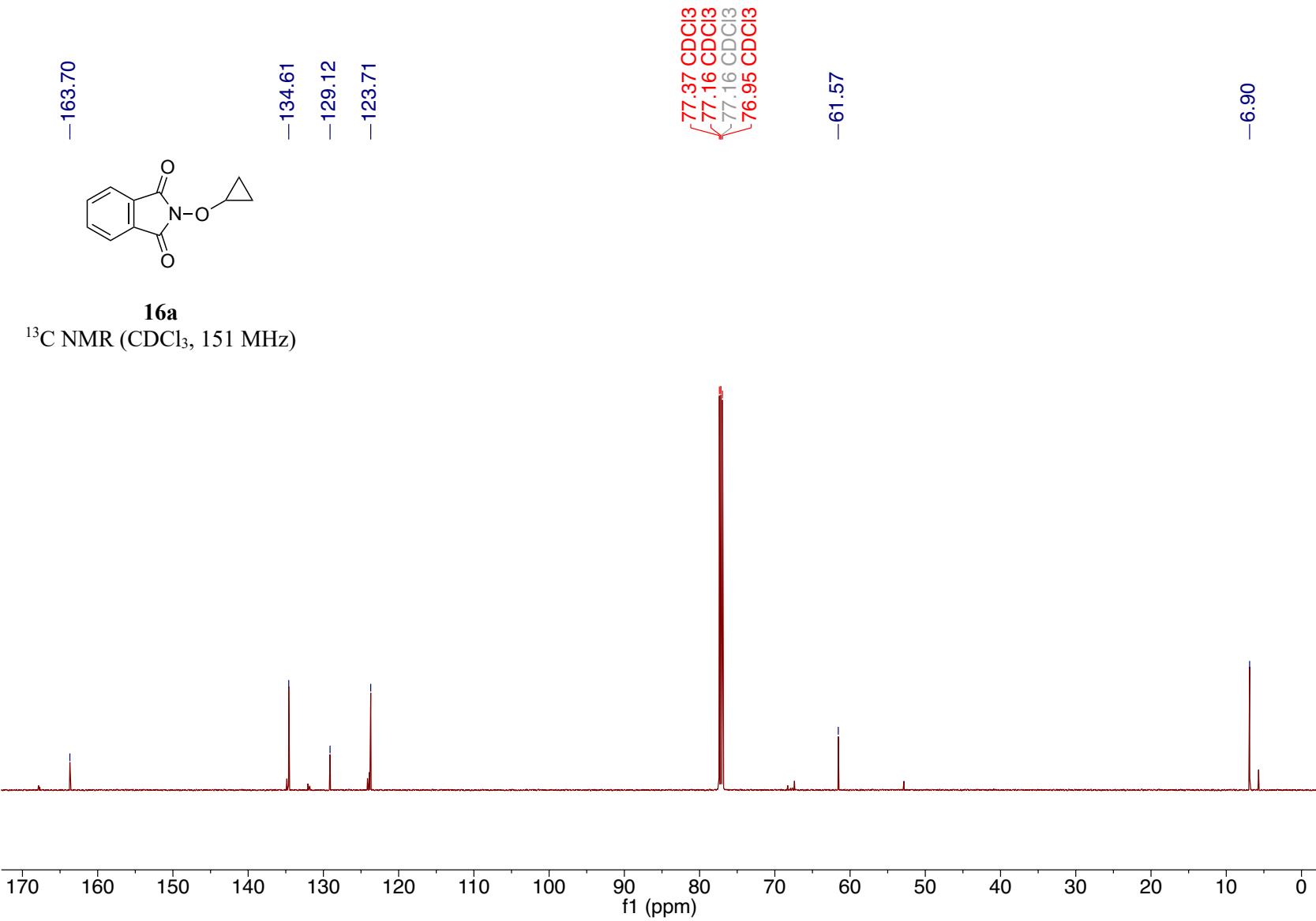
HRMS (ESI-TOF): calc'd for C₁₇H₂₄NO₄ [M+H]⁺ 306.1700; found 306.1704.

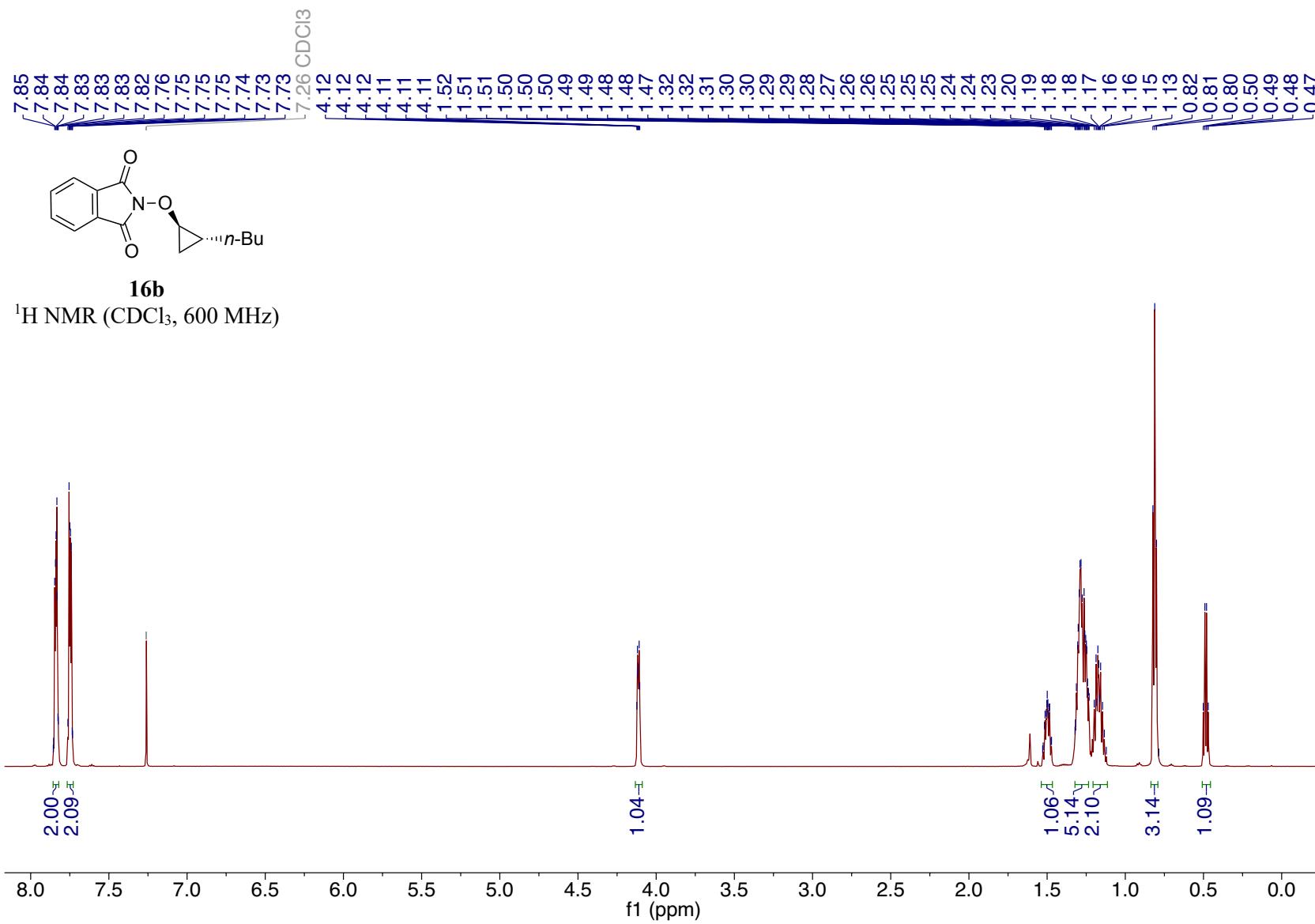
References

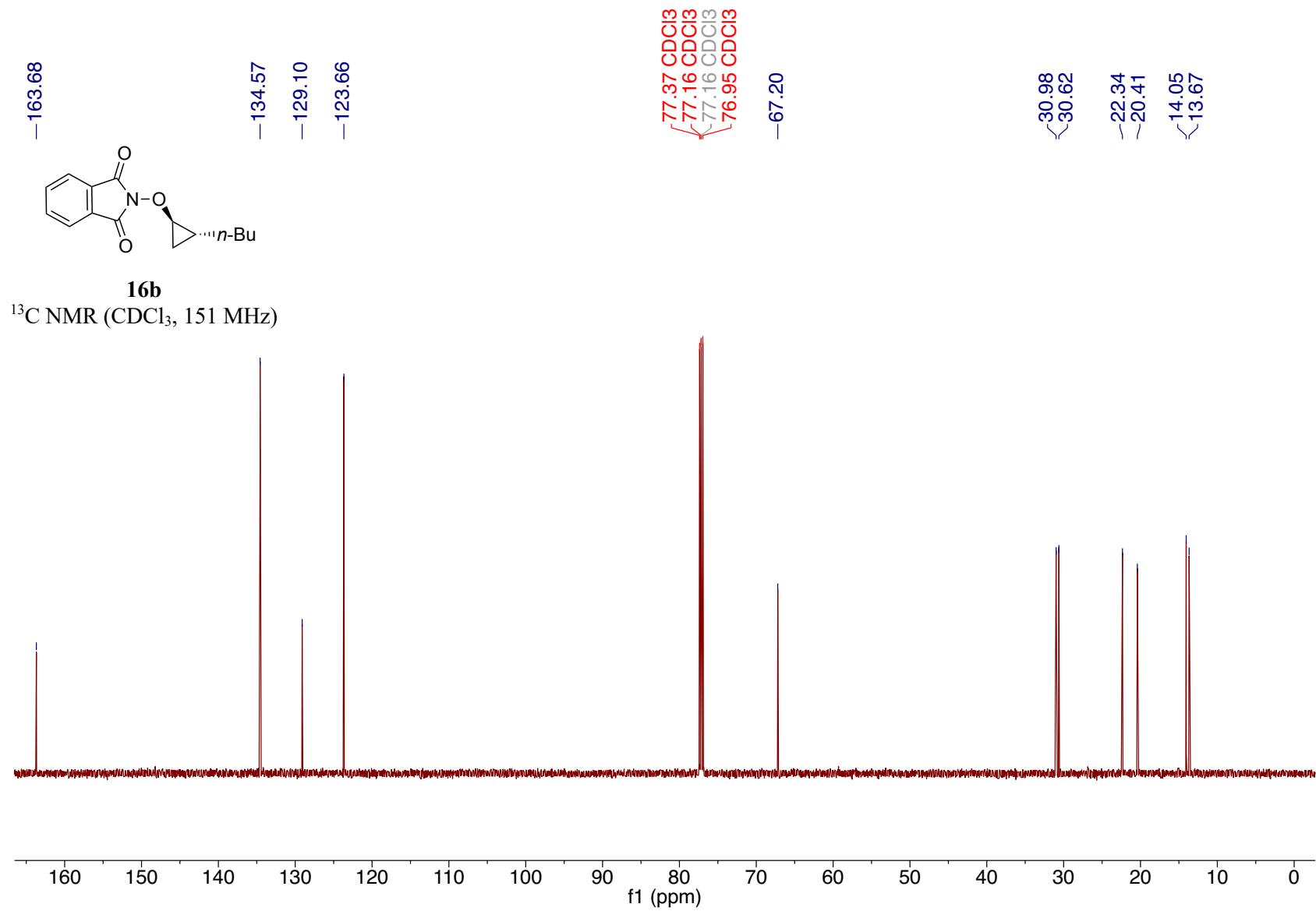
- [1] A. J. Pearce, D. S. Walter, C. S. Frampton, T. Gallagher, *J. Chem. Soc., Perkin Trans. 1* **1998**, 847-852.
- [2] A. S. Patil, D. L. Mo, H. Y. Wang, D. S. Mueller, L. L. Anderson, *Angew. Chem. Int. Ed.* **2012**, 51, 7799-7803.
- [3] L. Steemers, L. Wijsman, J. H. van Maarseveen, *Adv. Synth. Catal.* **2018**, 360, 4241-4245.
- [4] J. C. Lorenz, J. Long, Z. Yang, S. Xue, Y. Xie, Y. Shi, *J. Org. Chem.* **2004**, 69, 327-334.
- [5] D. G. Hall, in *Boronic acids : preparation and applications in organic synthesis, medicine and materials*, Second ed., Wiley-VCH, Weinheim, **2011**.
- [6] H. Sakurai, *Pure & Appl. Chem.* **1982**, 54, 1-22.
- [7] J. D. Scott, M. W. Miller, S. W. Li, S. I. Lin, H. A. Vaccaro, L. W. Hong, D. E. Mullins, M. Guzzi, J. Weinstein, R. A. Hodgson, G. B. Varty, A. W. Stamford, T. Y. Chan, B. A. McKittrick, W. J. Greenlee, T. Priestley, E. M. Parker, *Bioorg. Med. Chem. Lett.* **2009**, 19, 6018-6022.

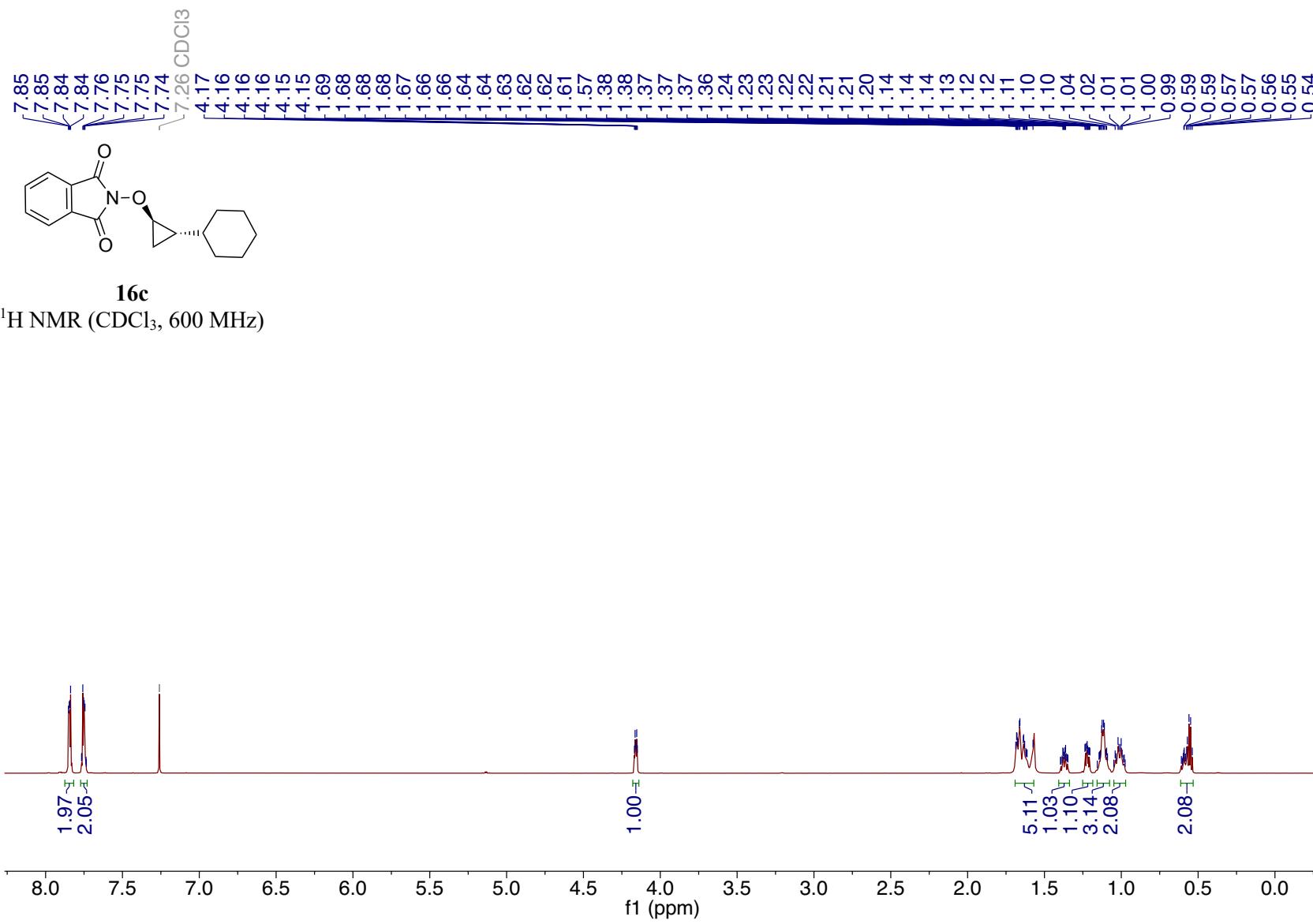
NMR Spectra of Reported Compounds

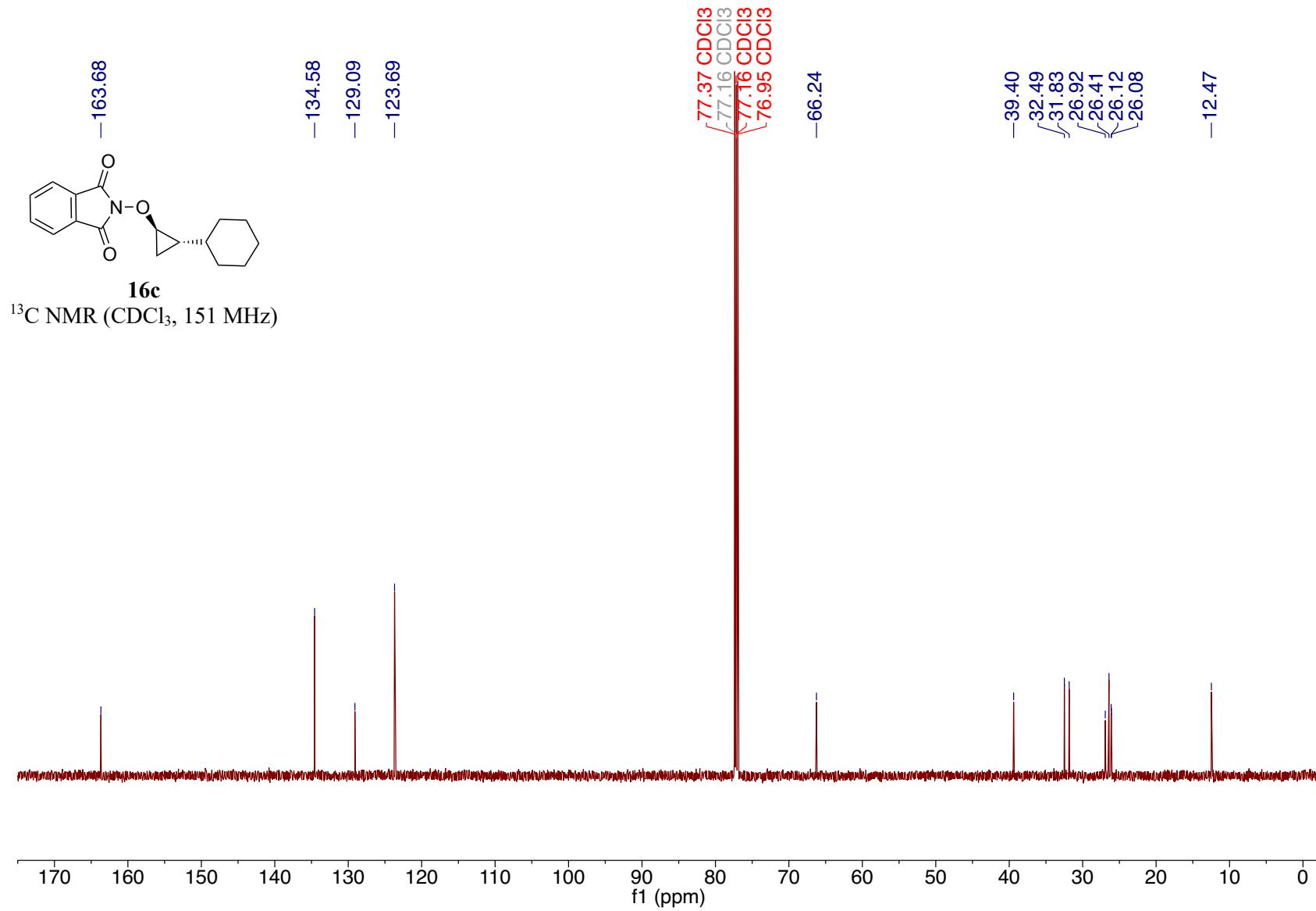


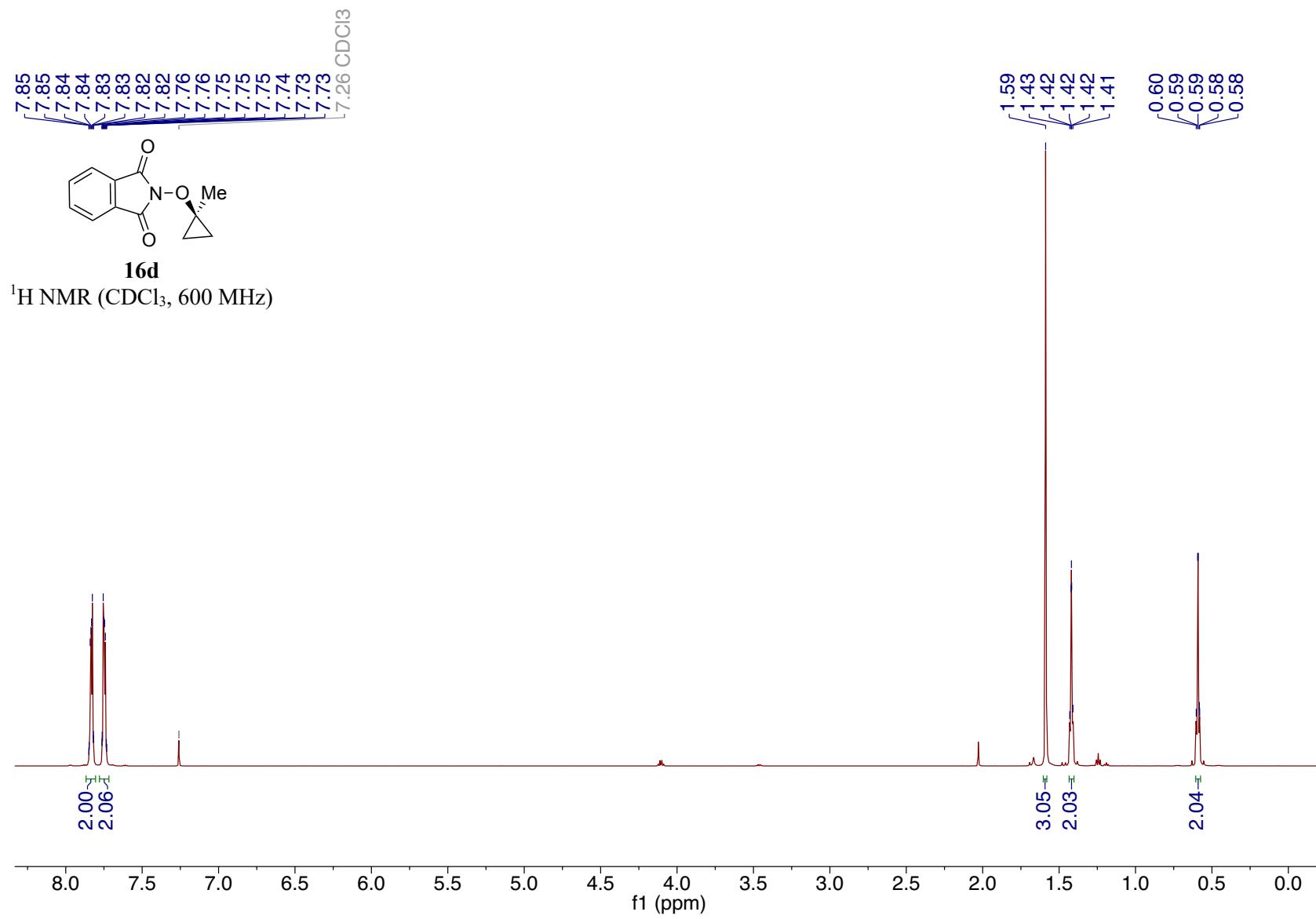


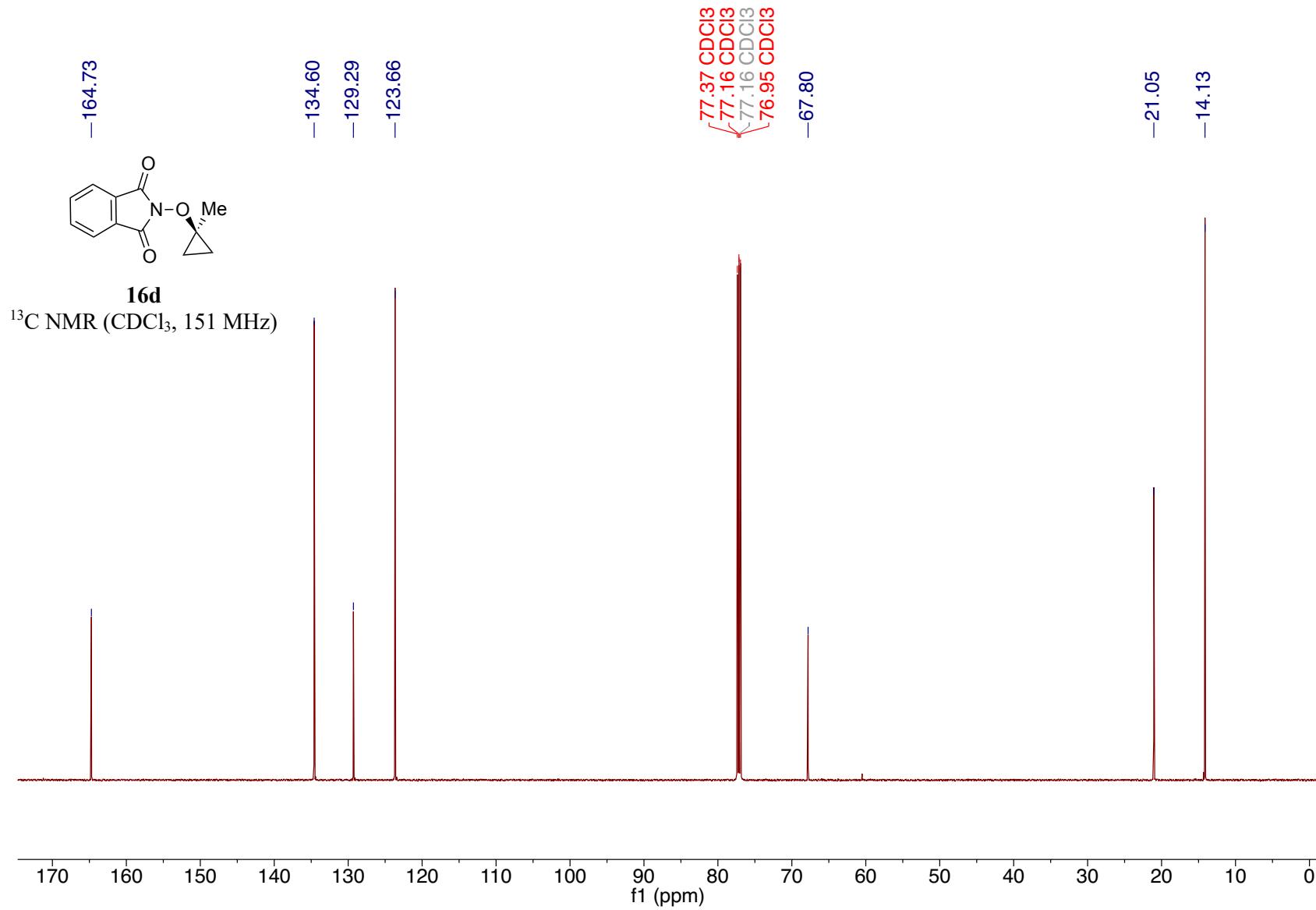


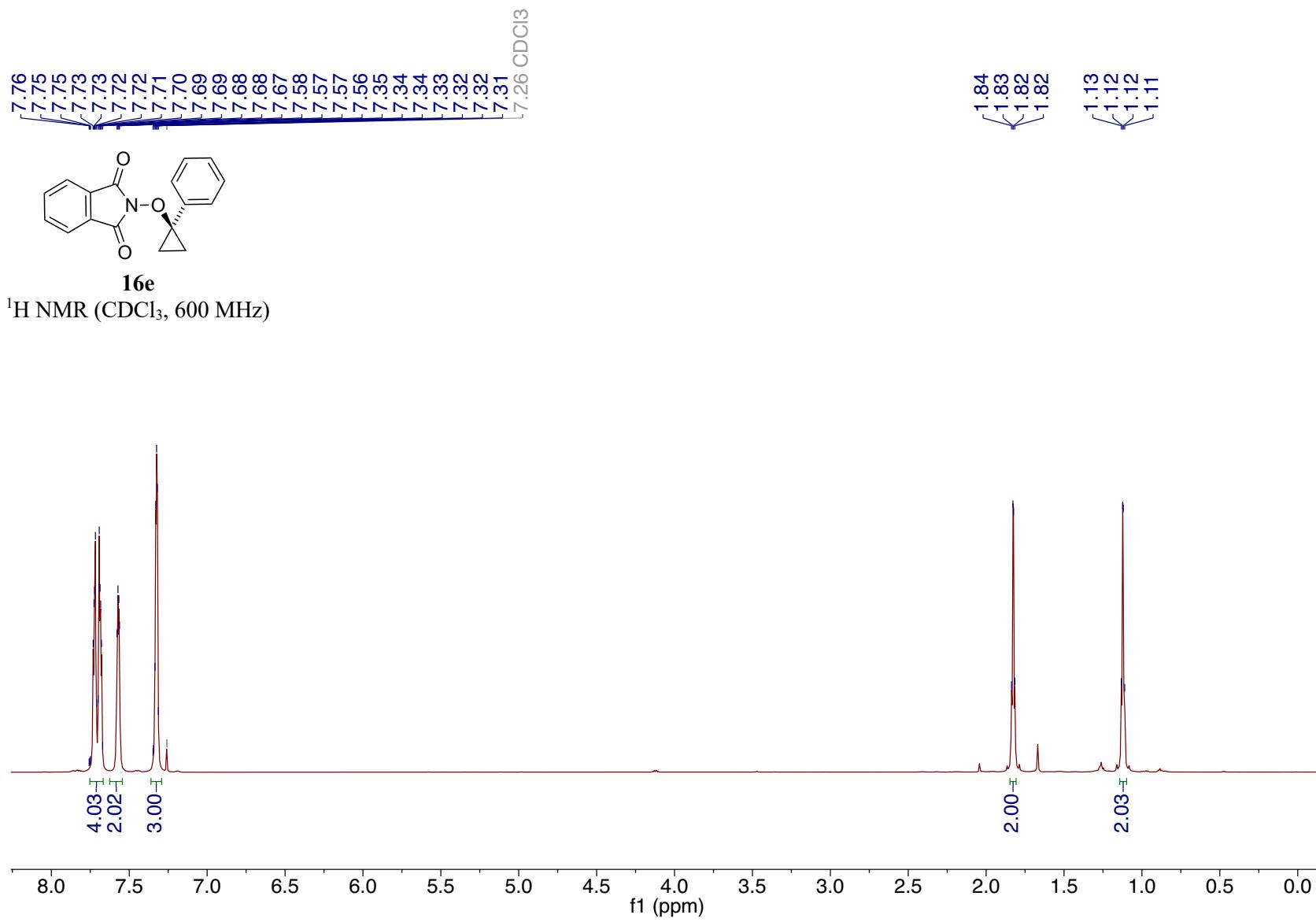


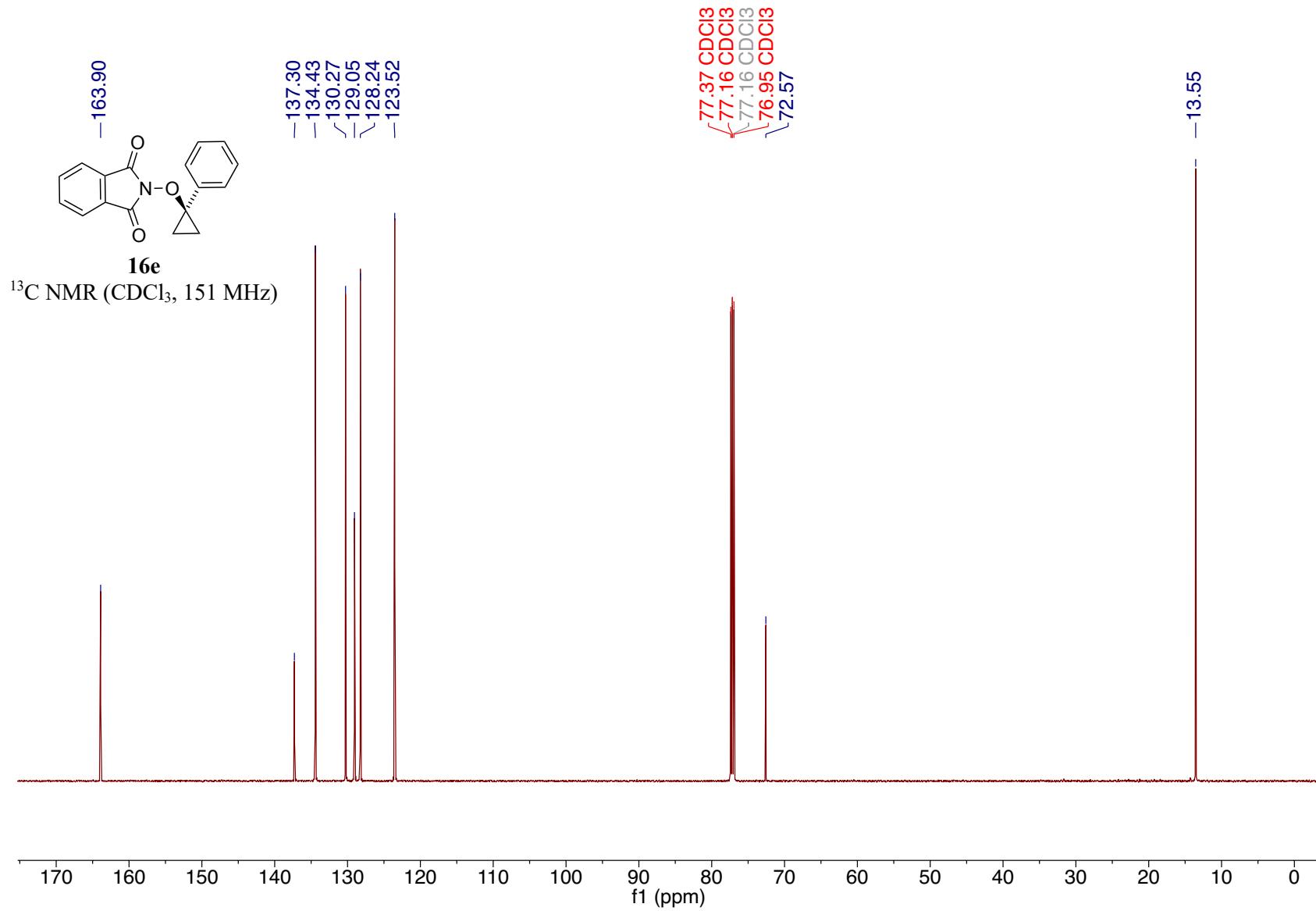


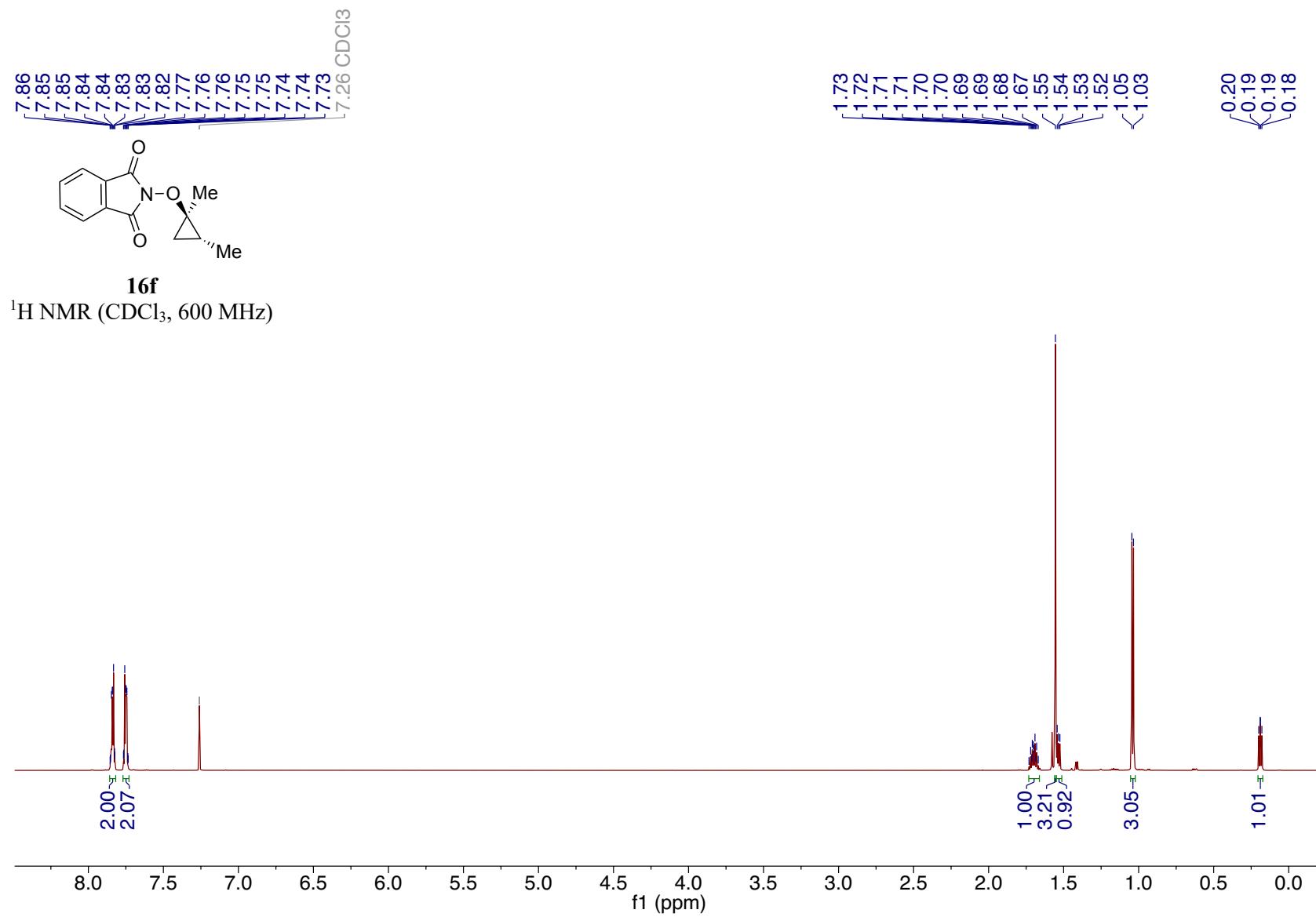


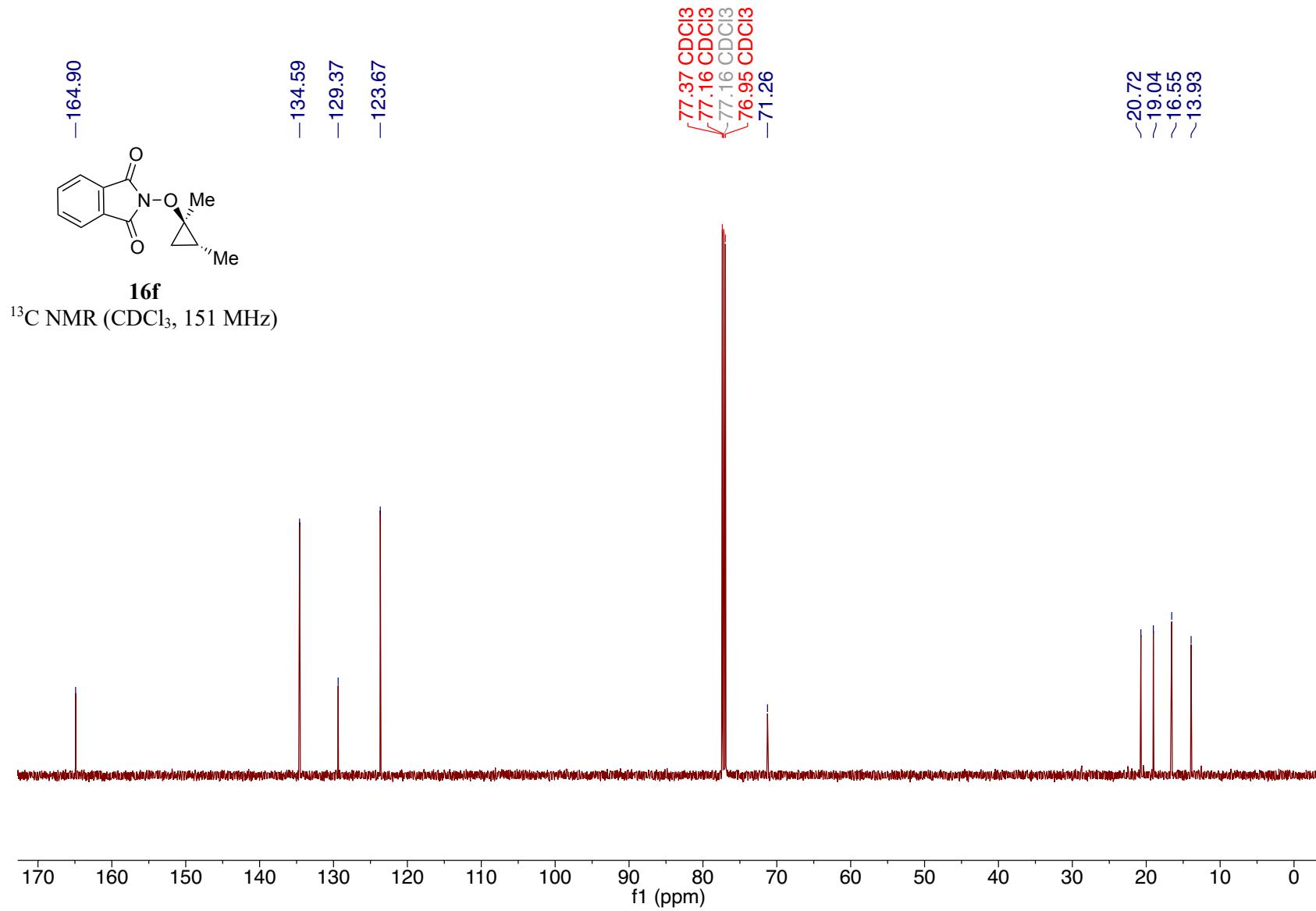


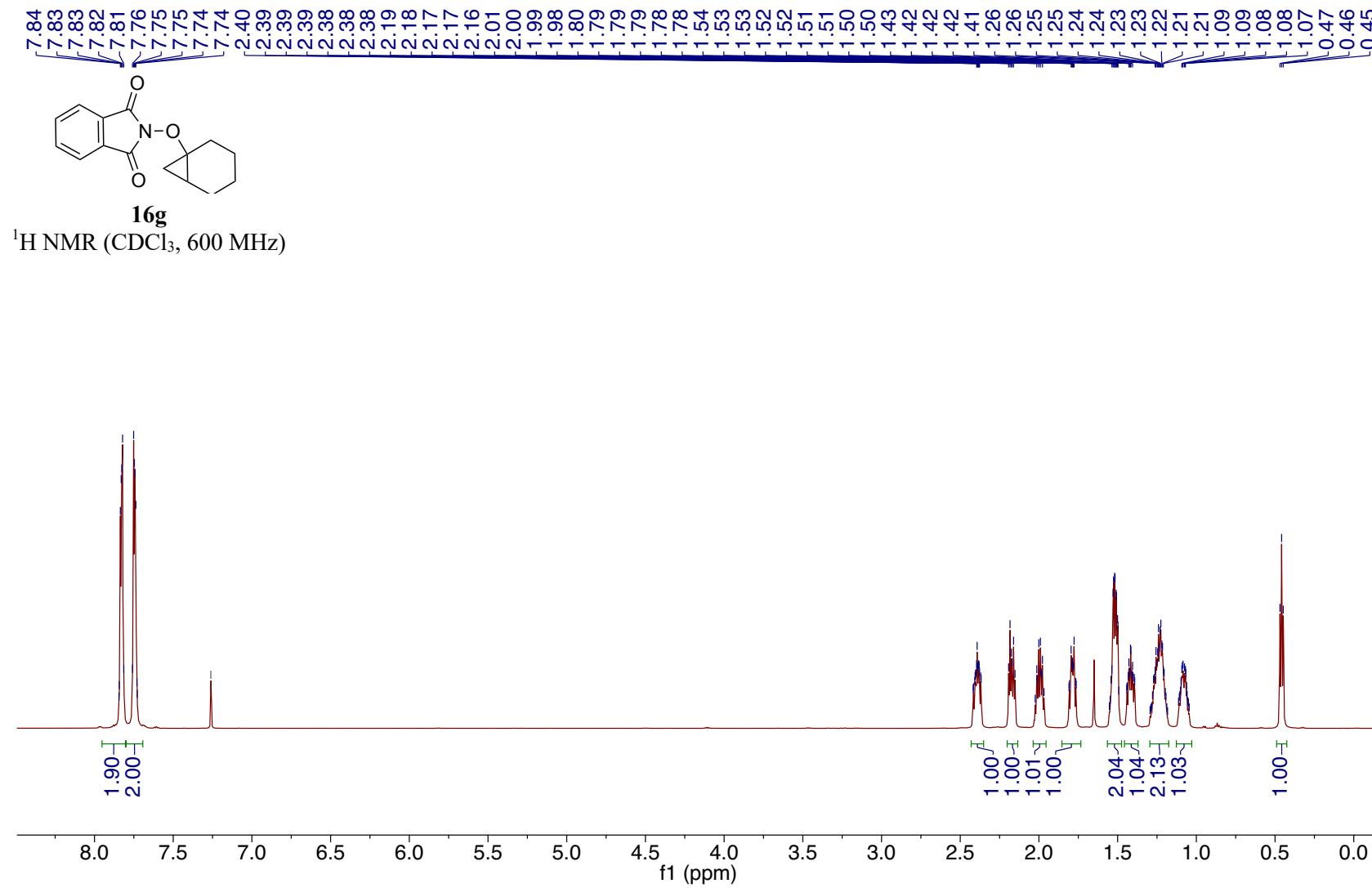


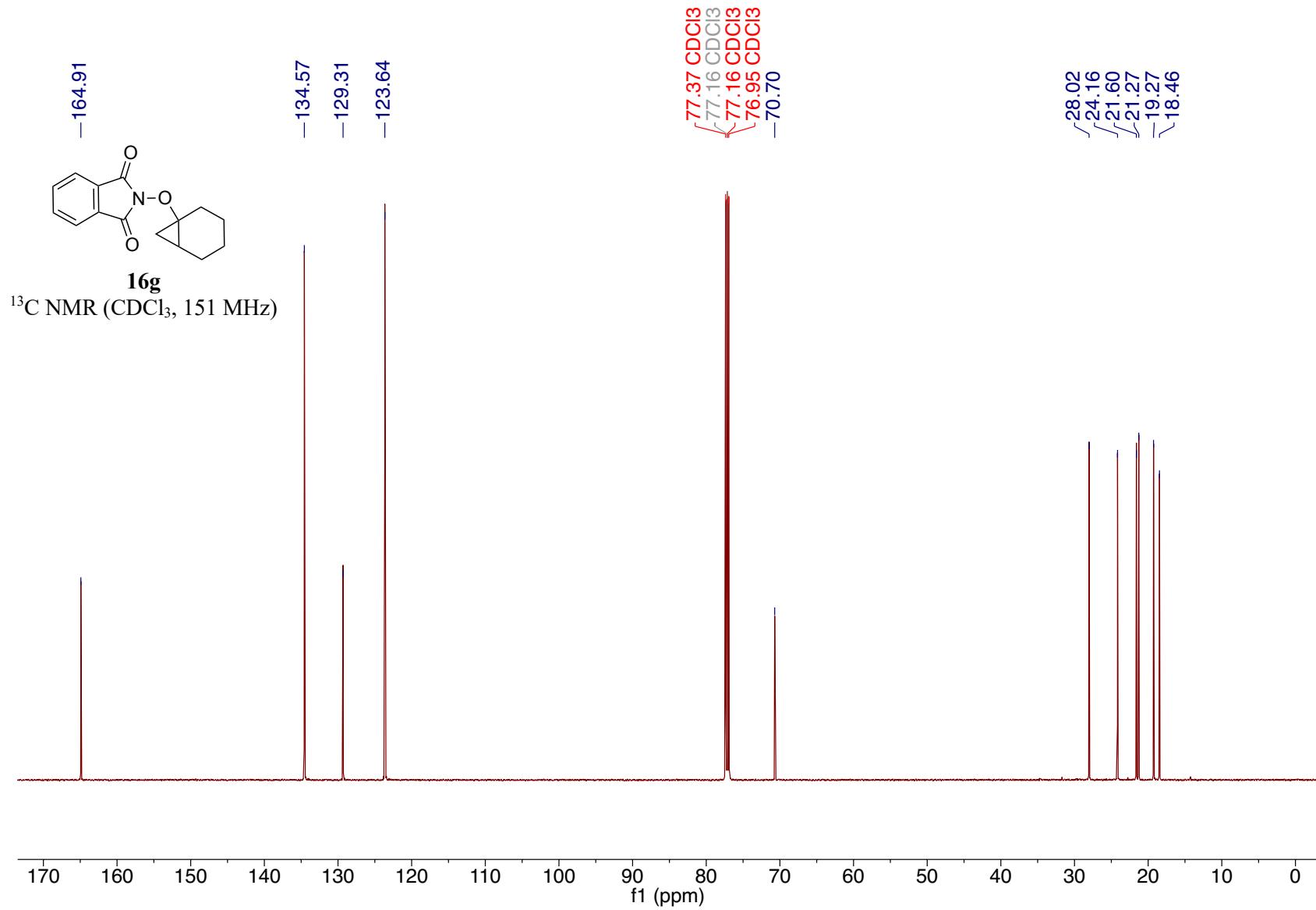


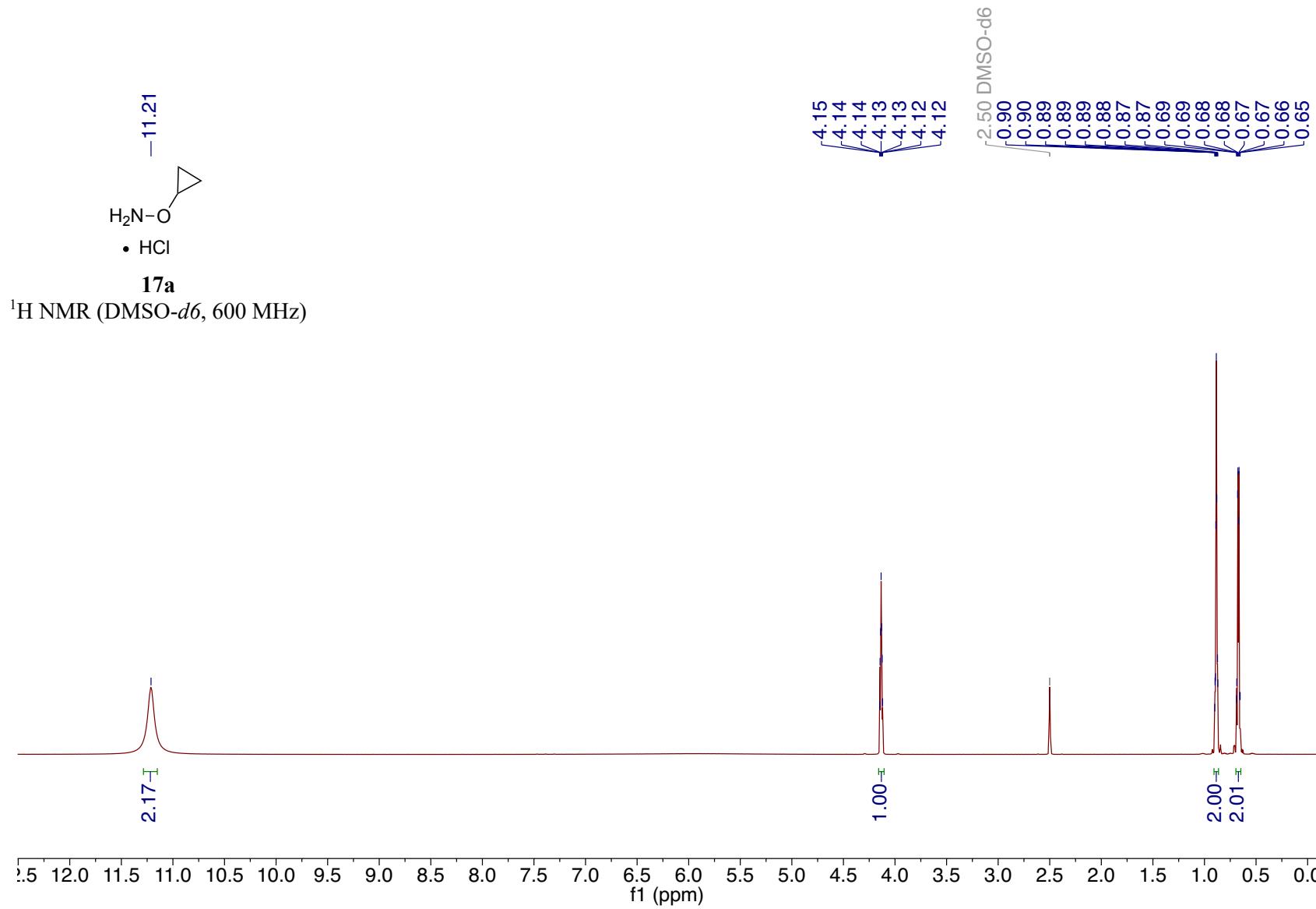


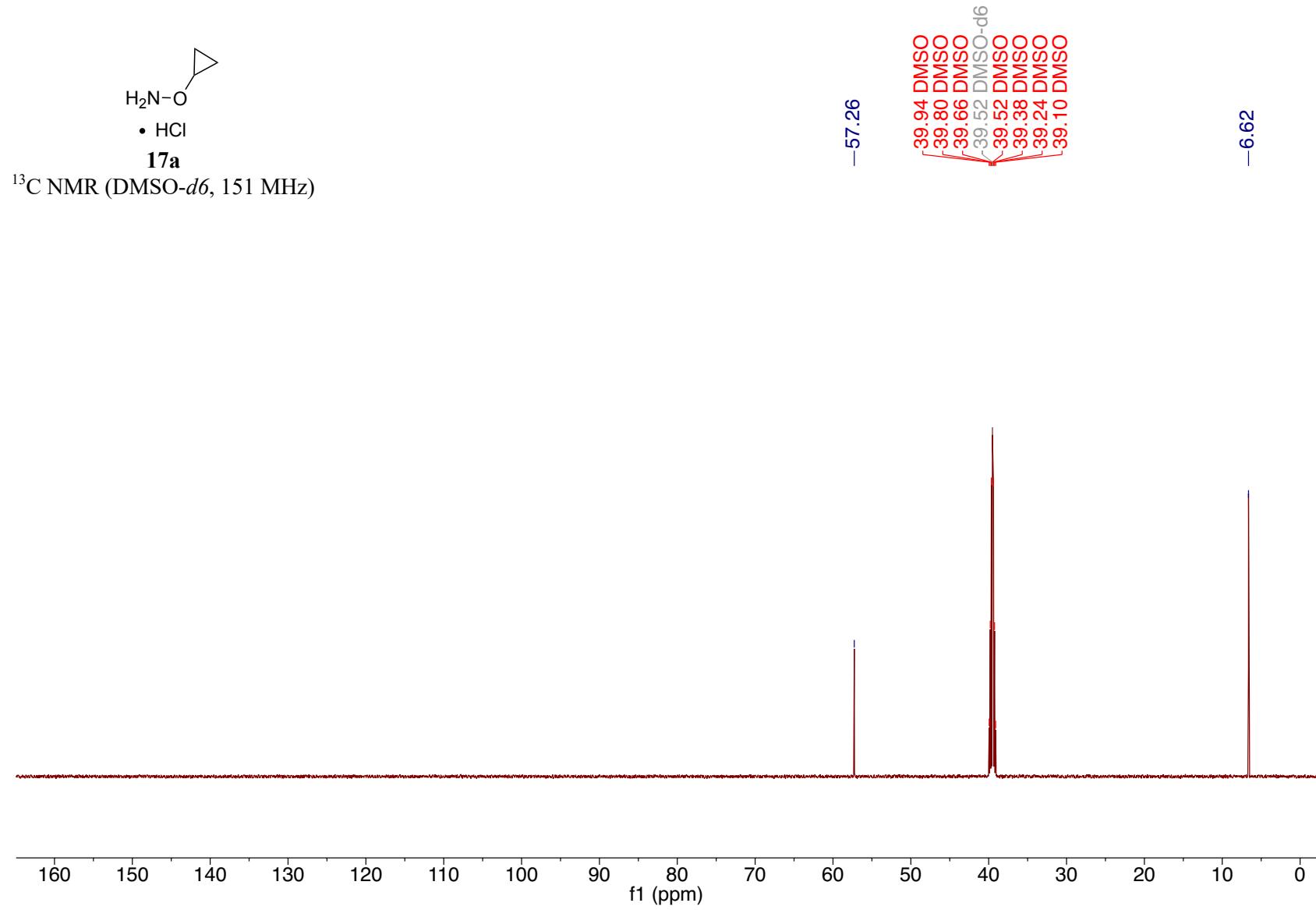


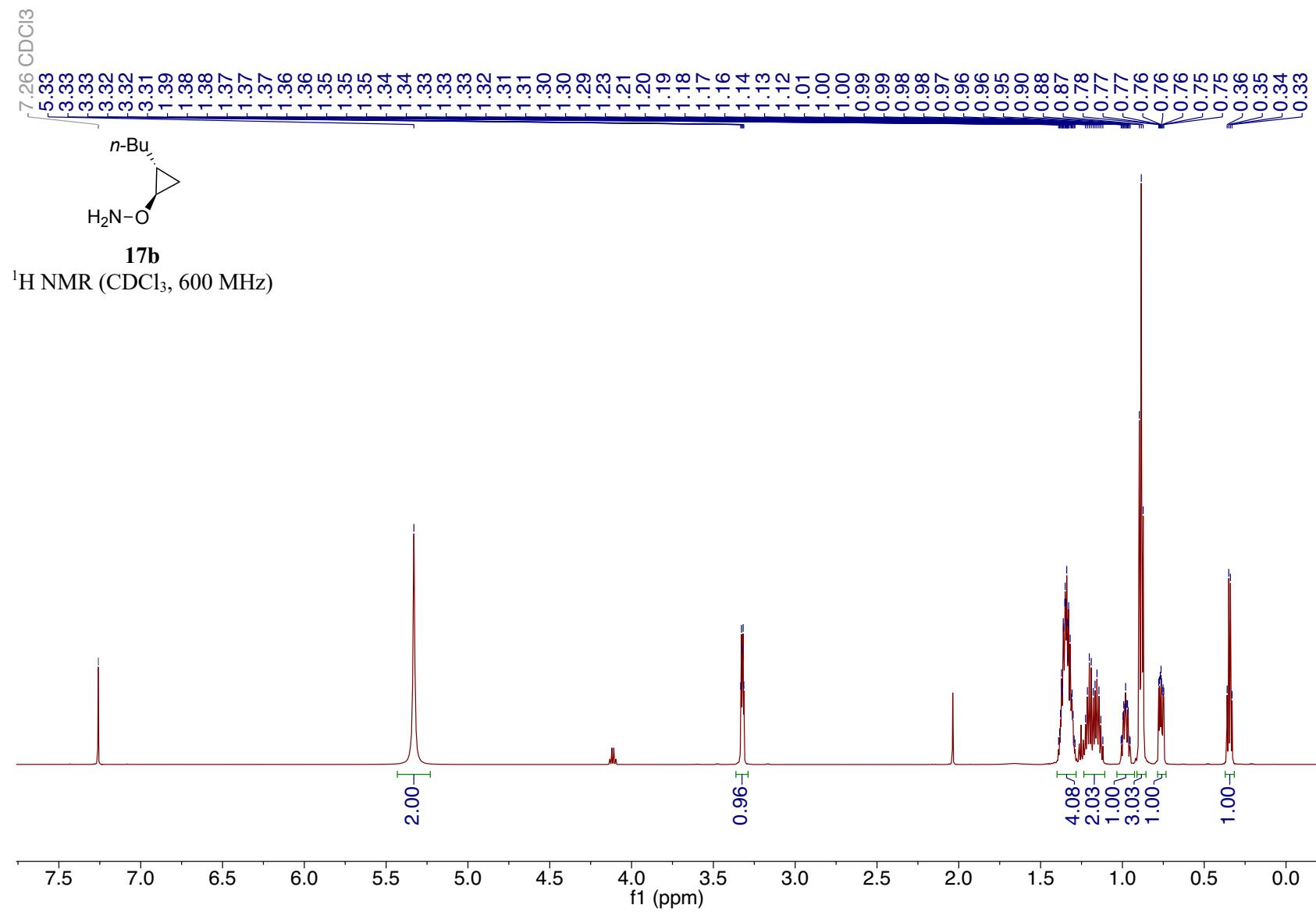


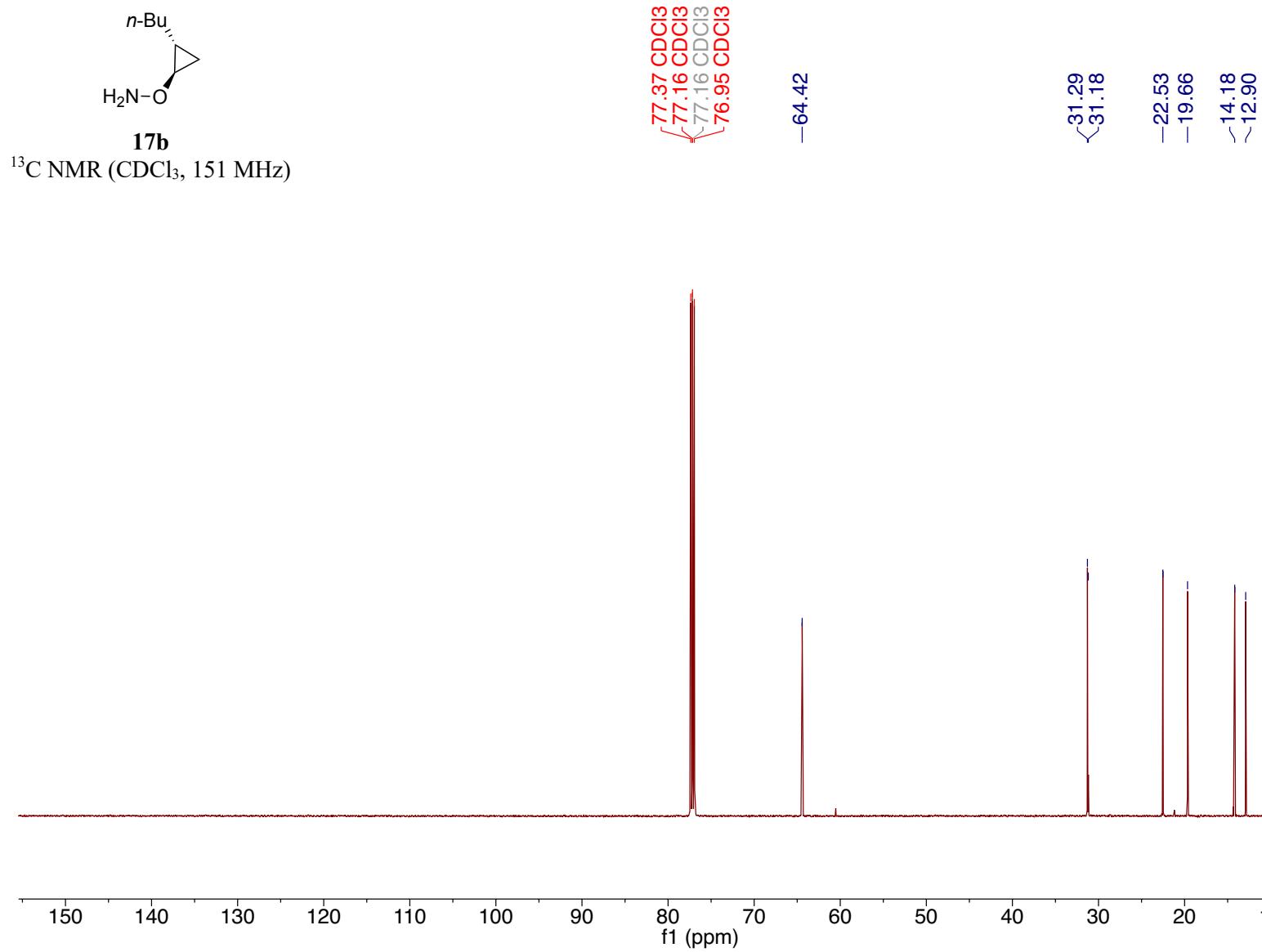


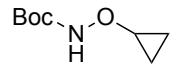






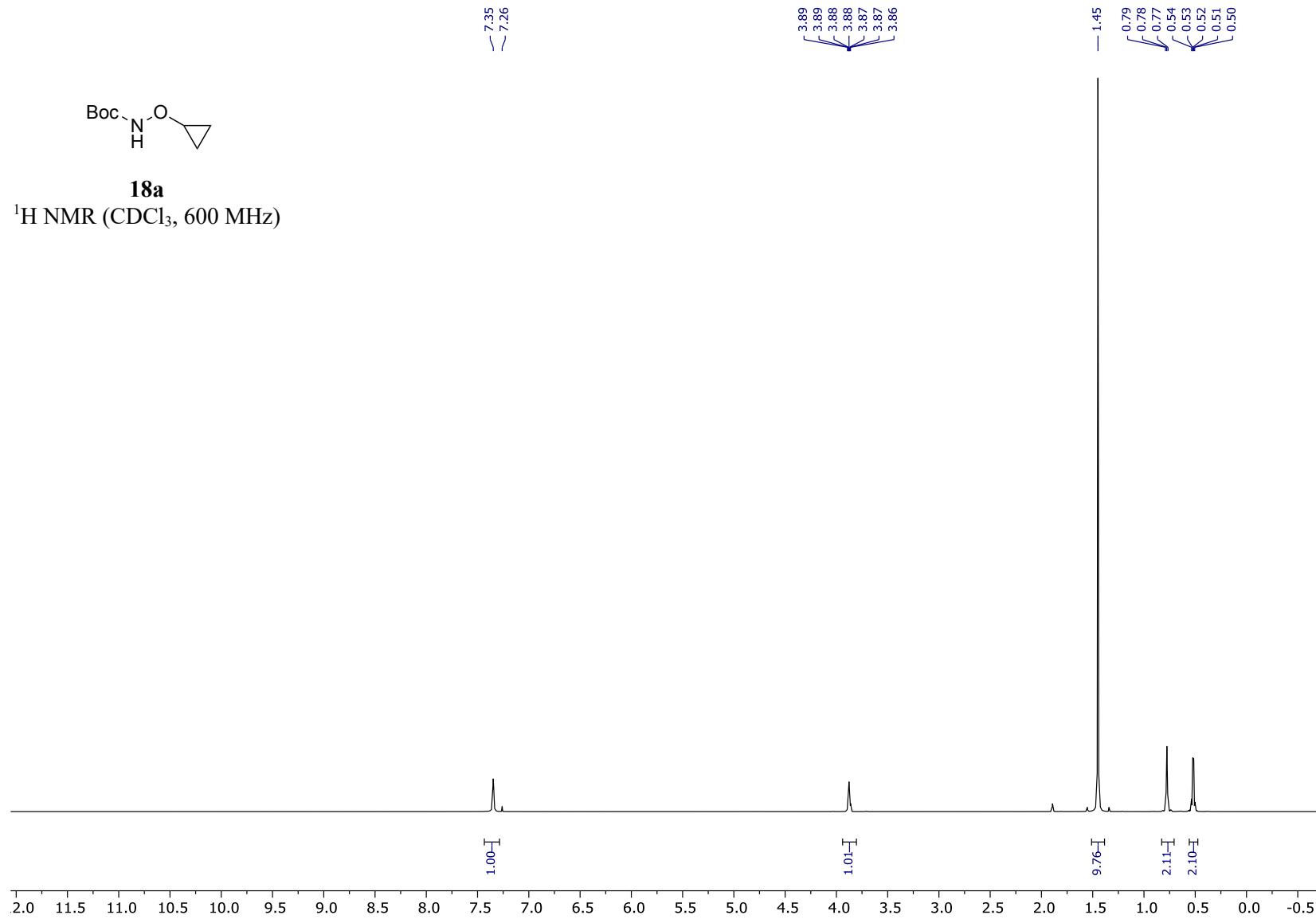


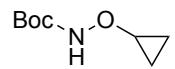




18a

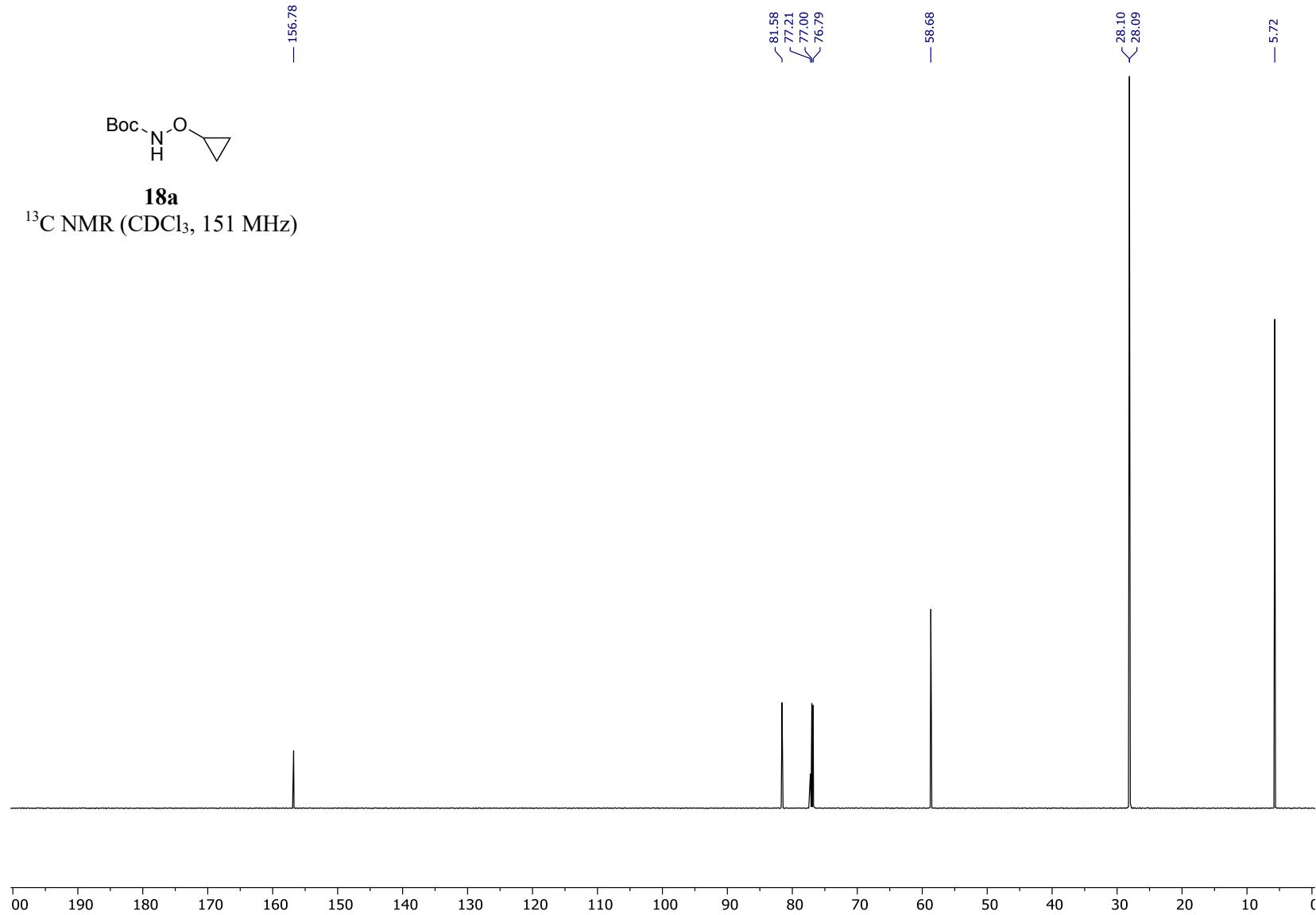
^1H NMR (CDCl_3 , 600 MHz)

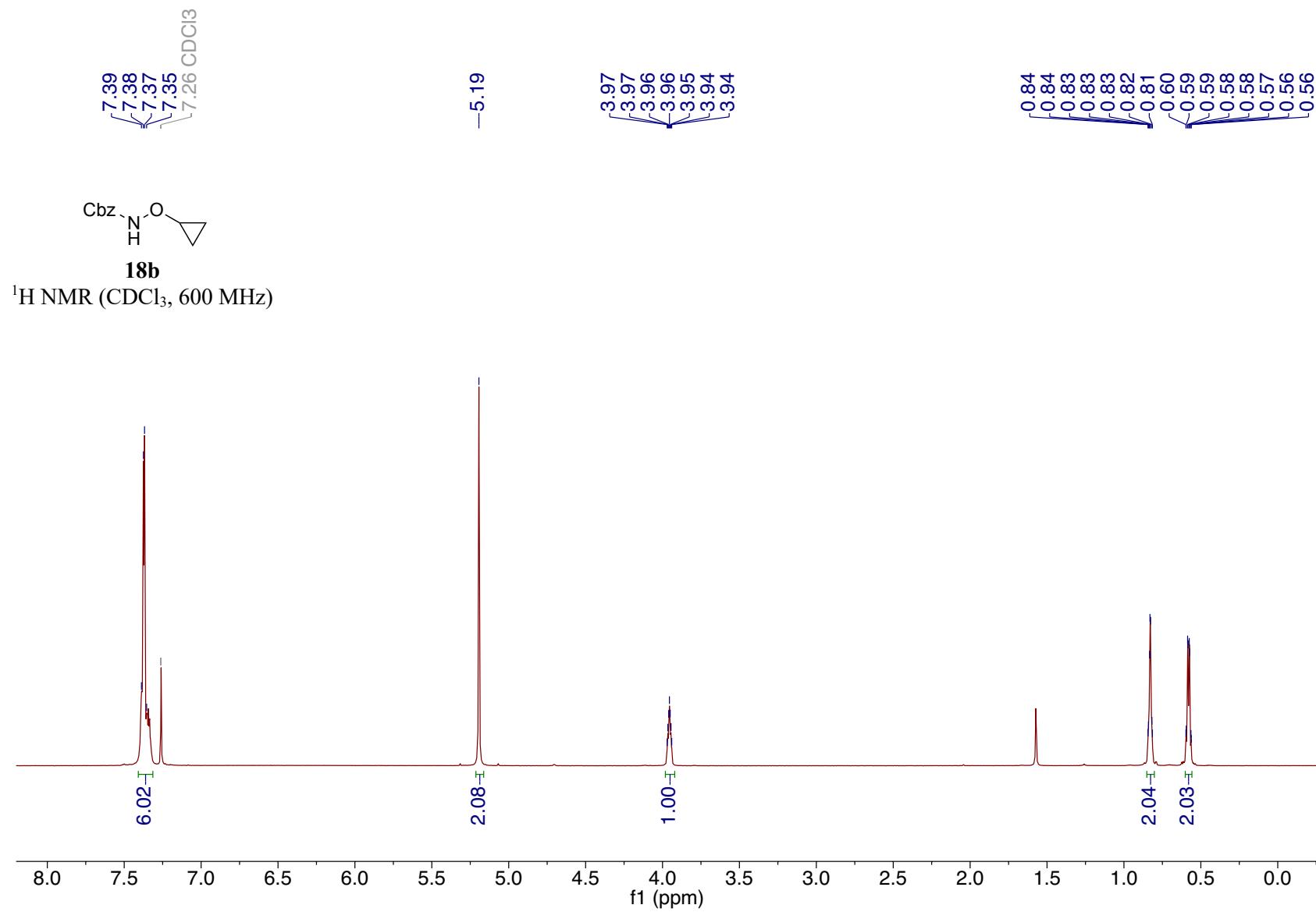


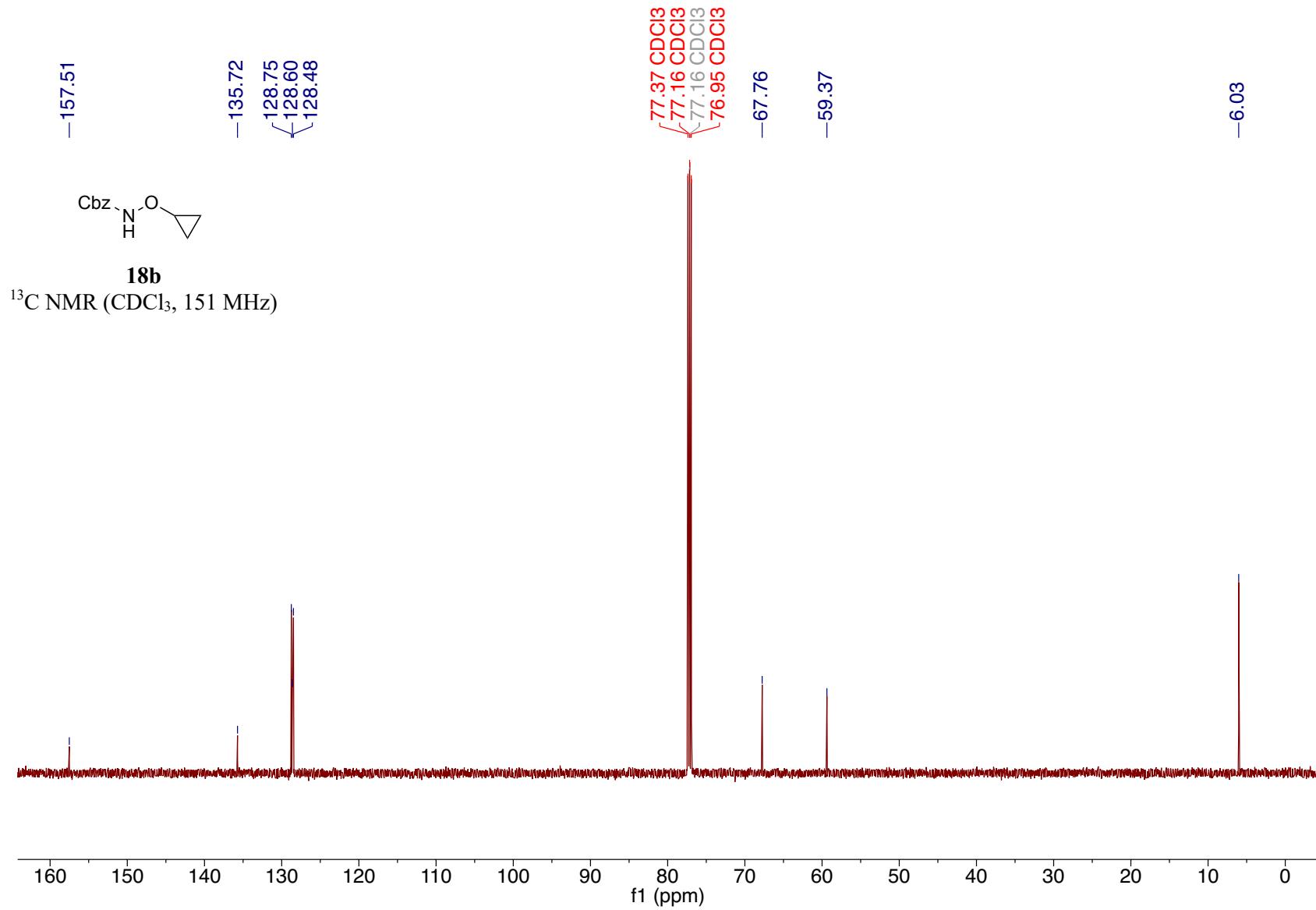


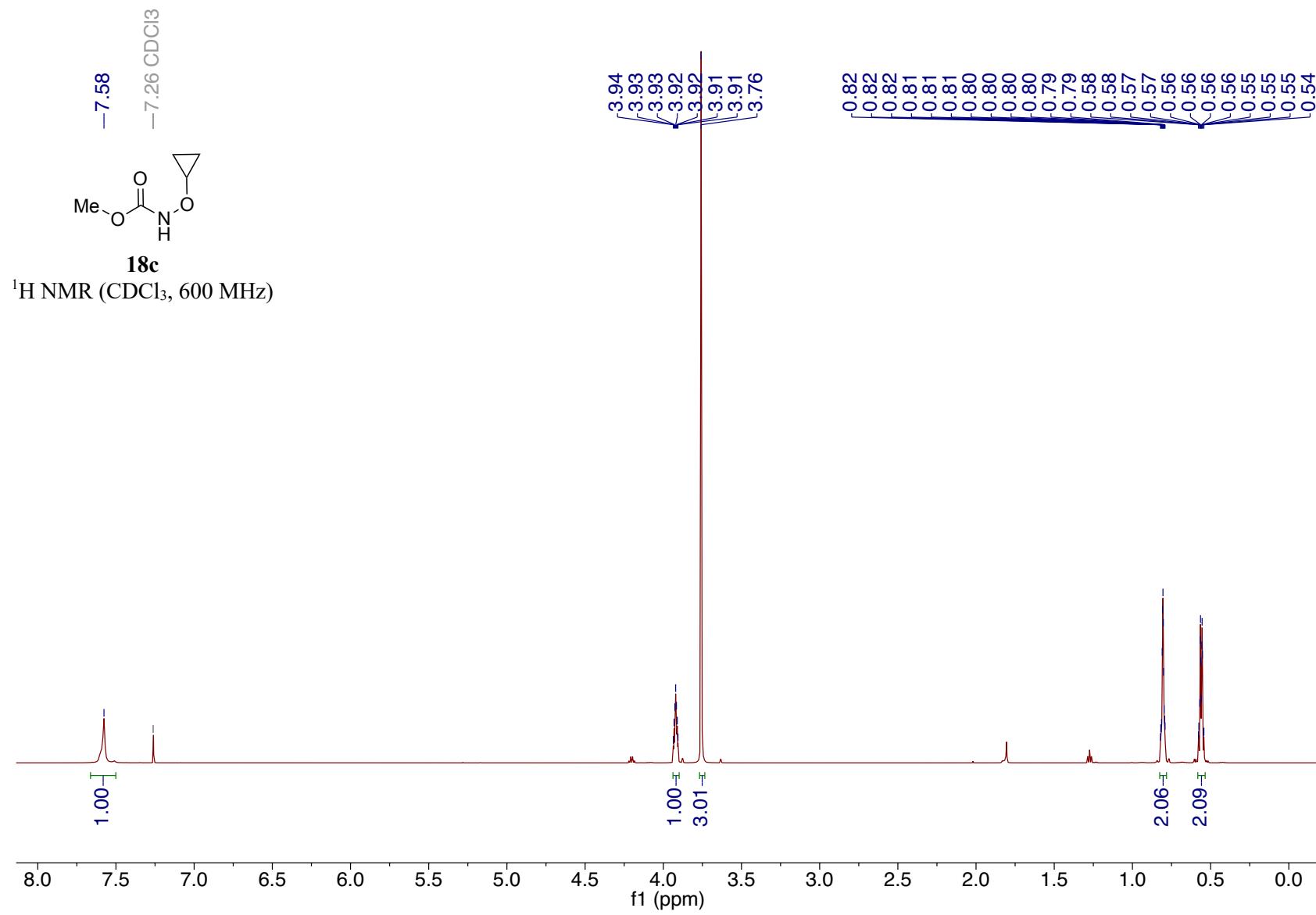
18a

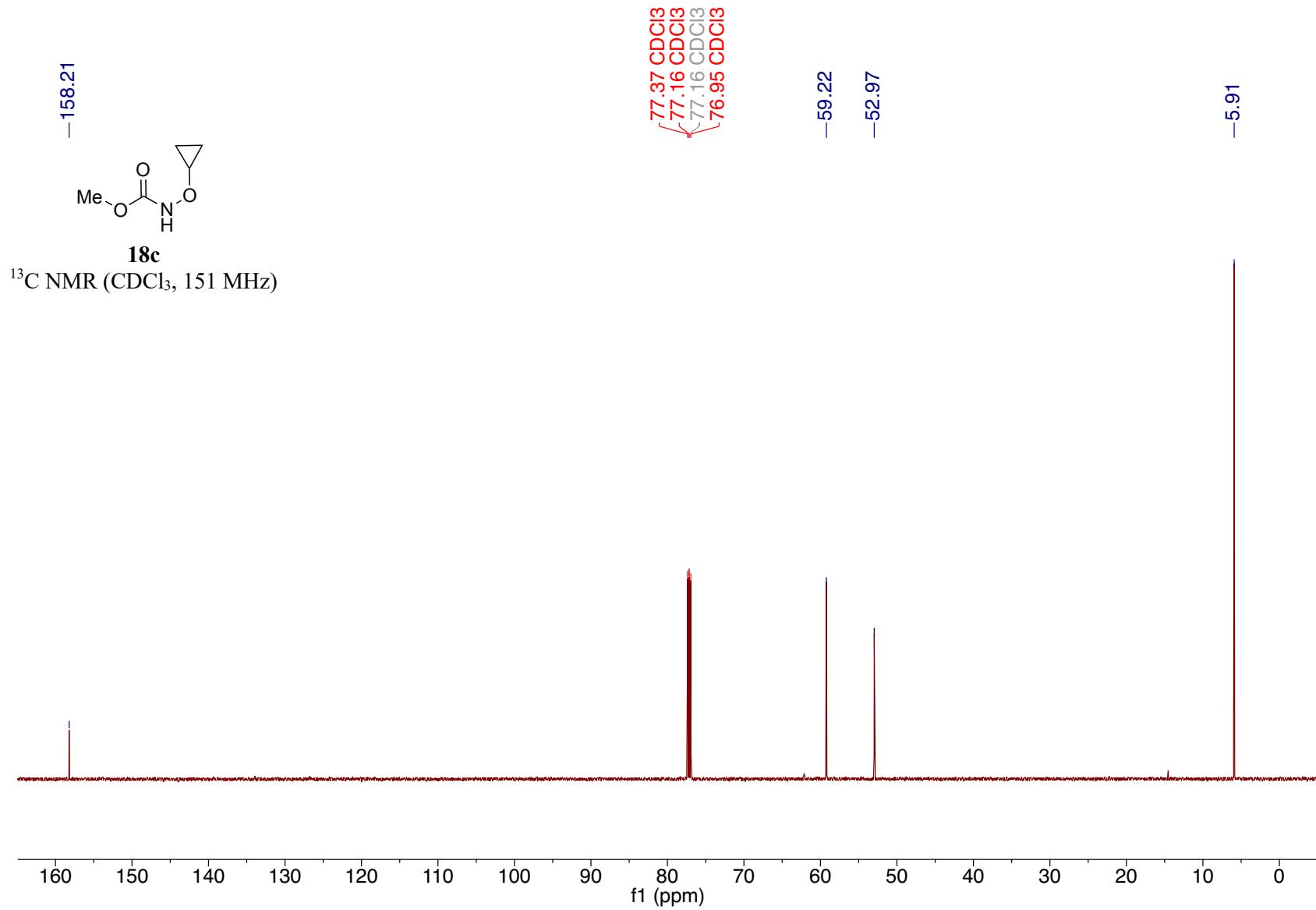
^{13}C NMR (CDCl_3 , 151 MHz)

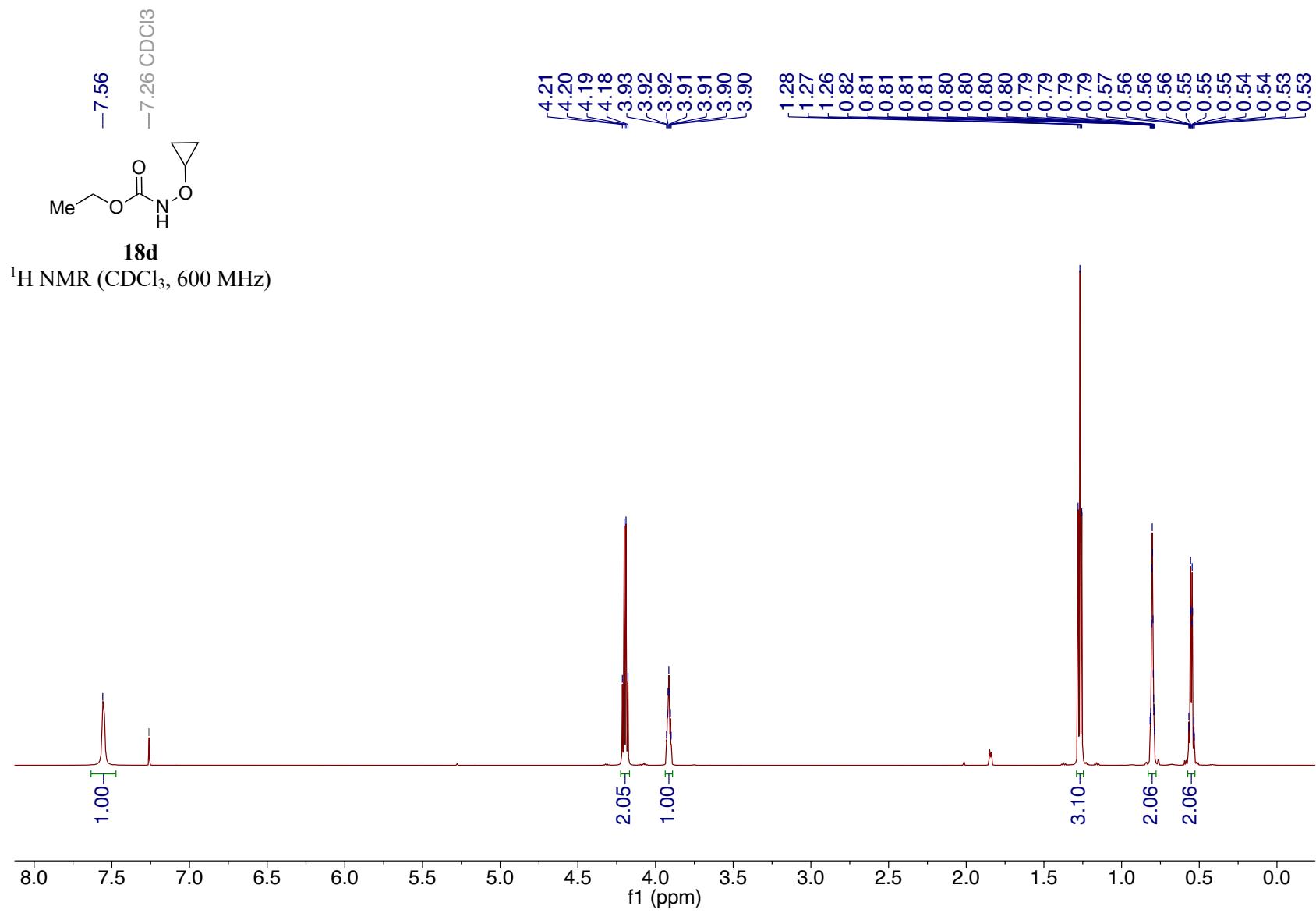


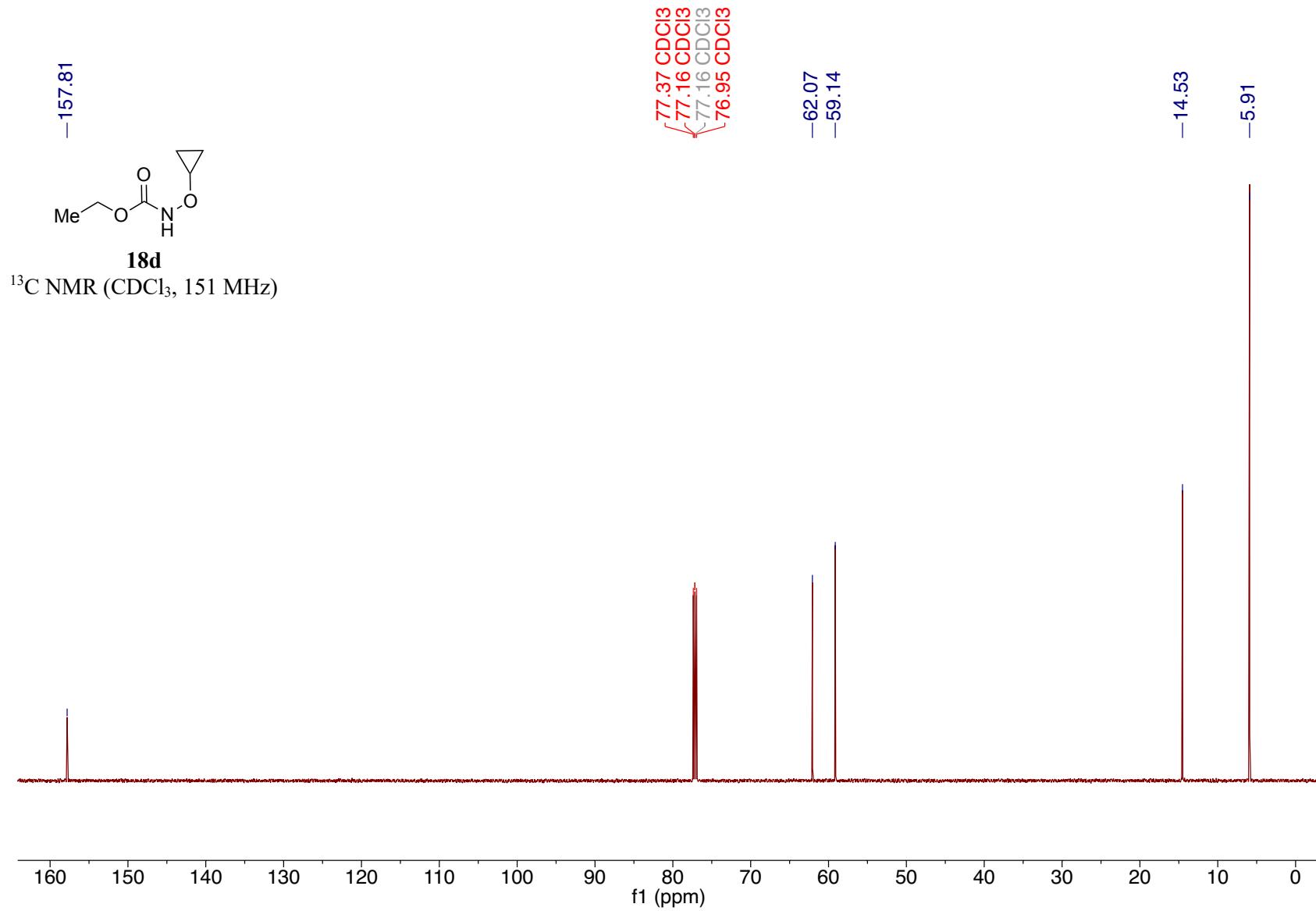


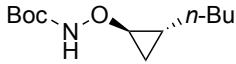




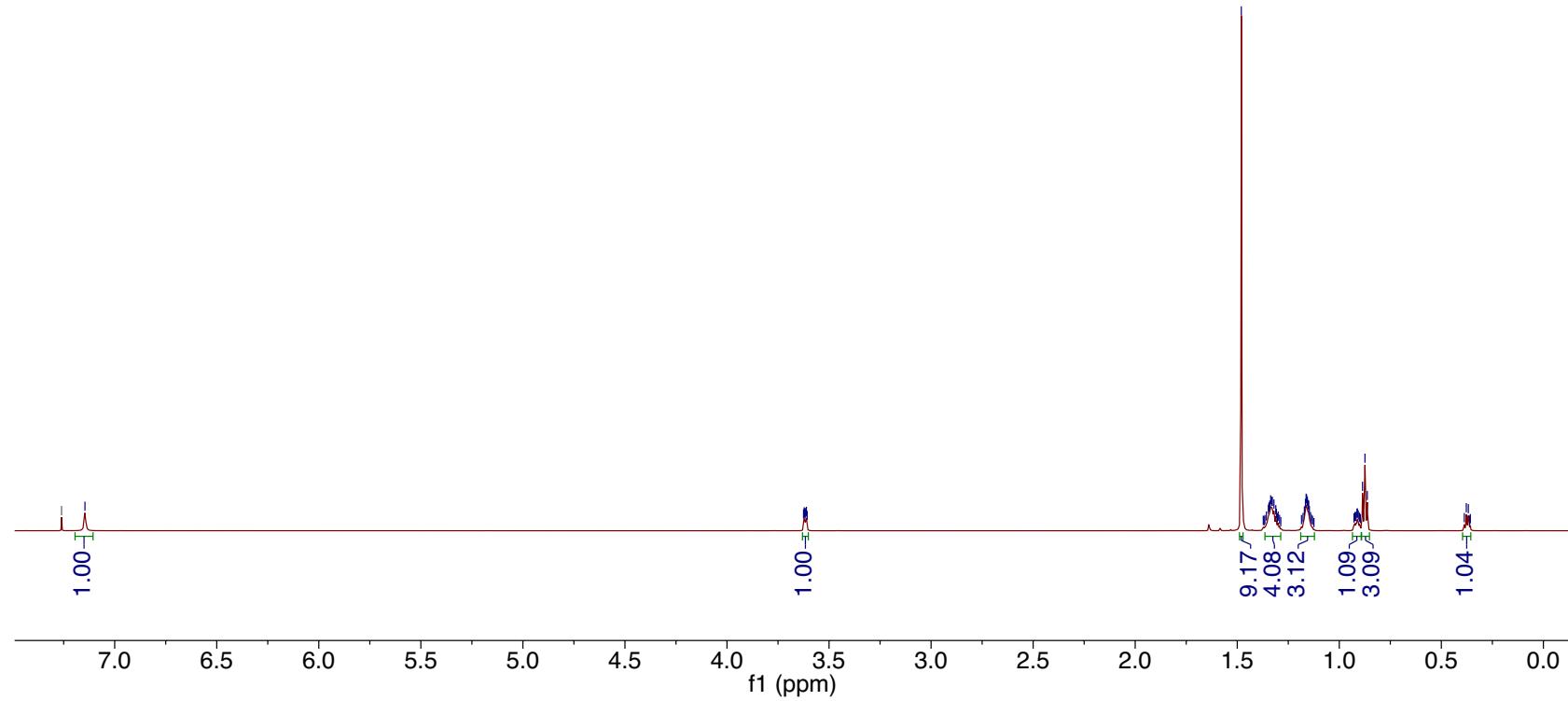


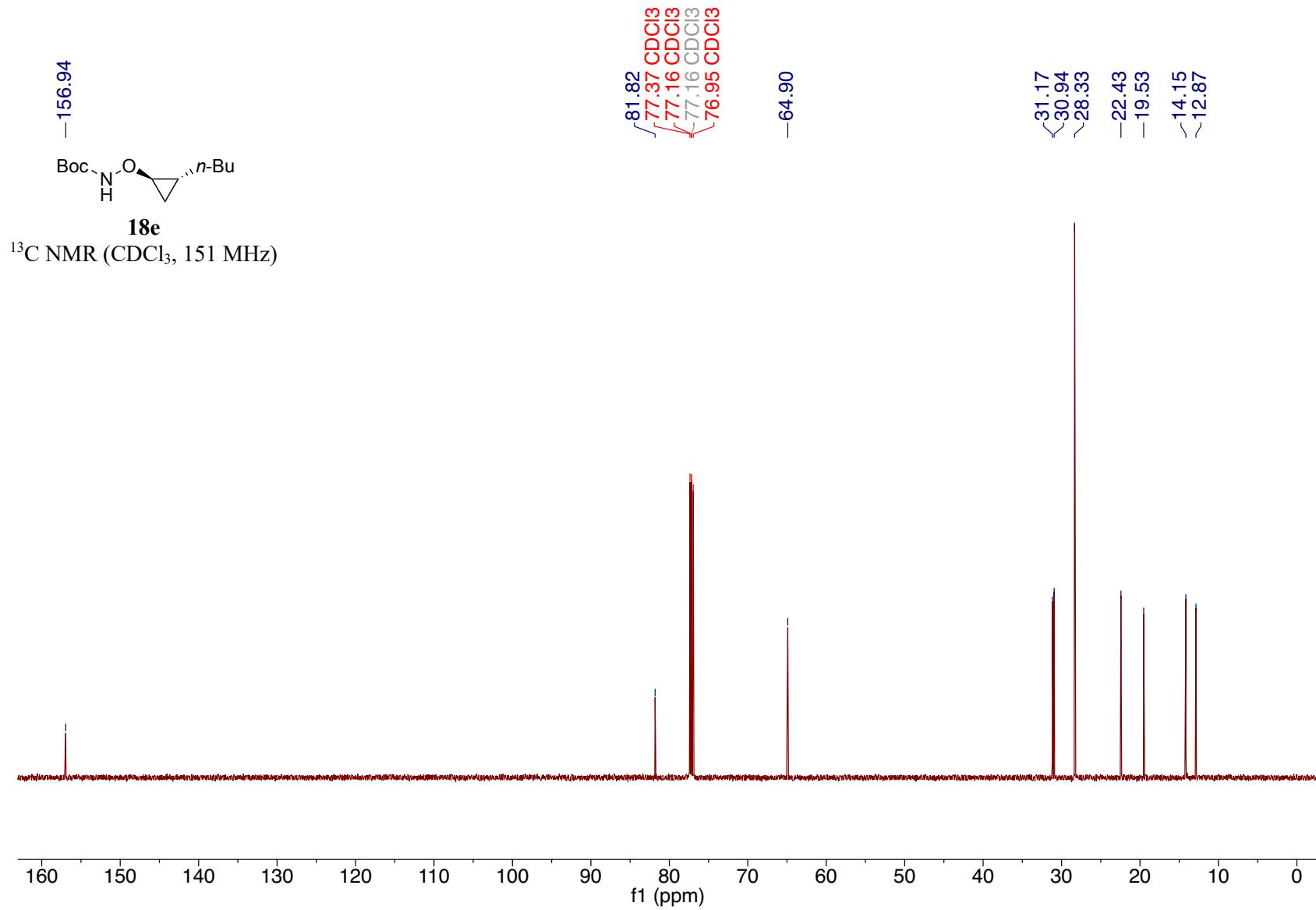


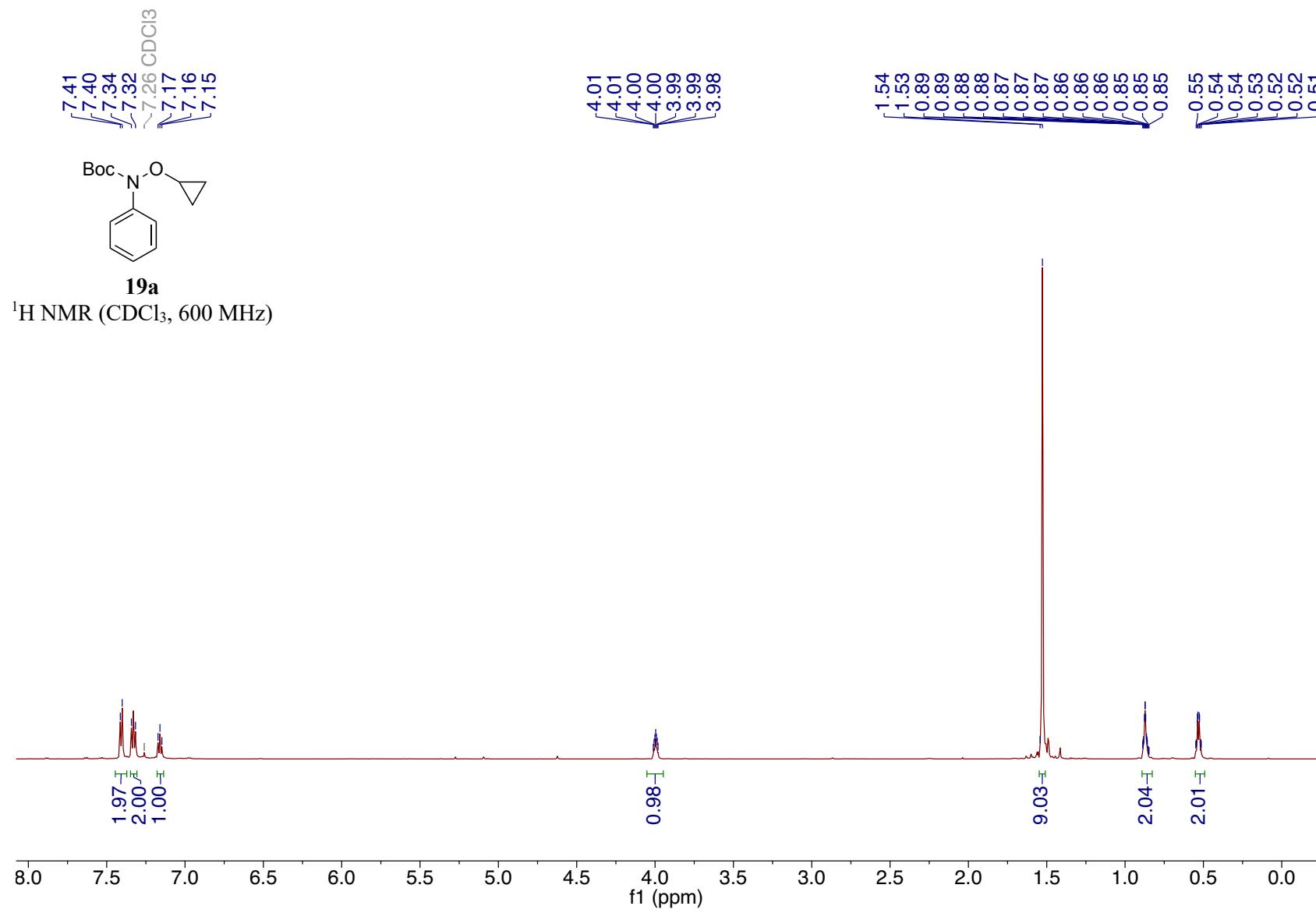


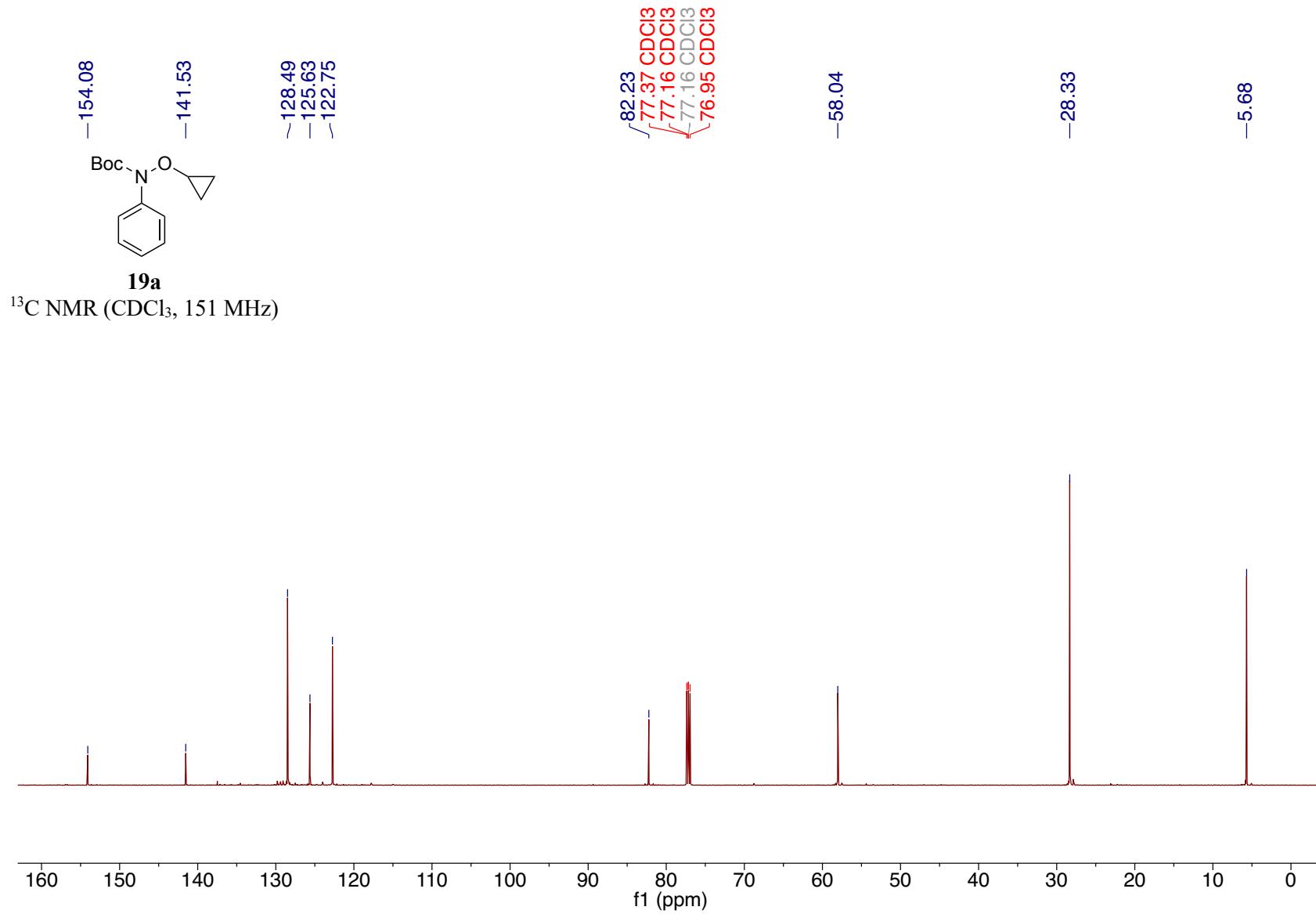


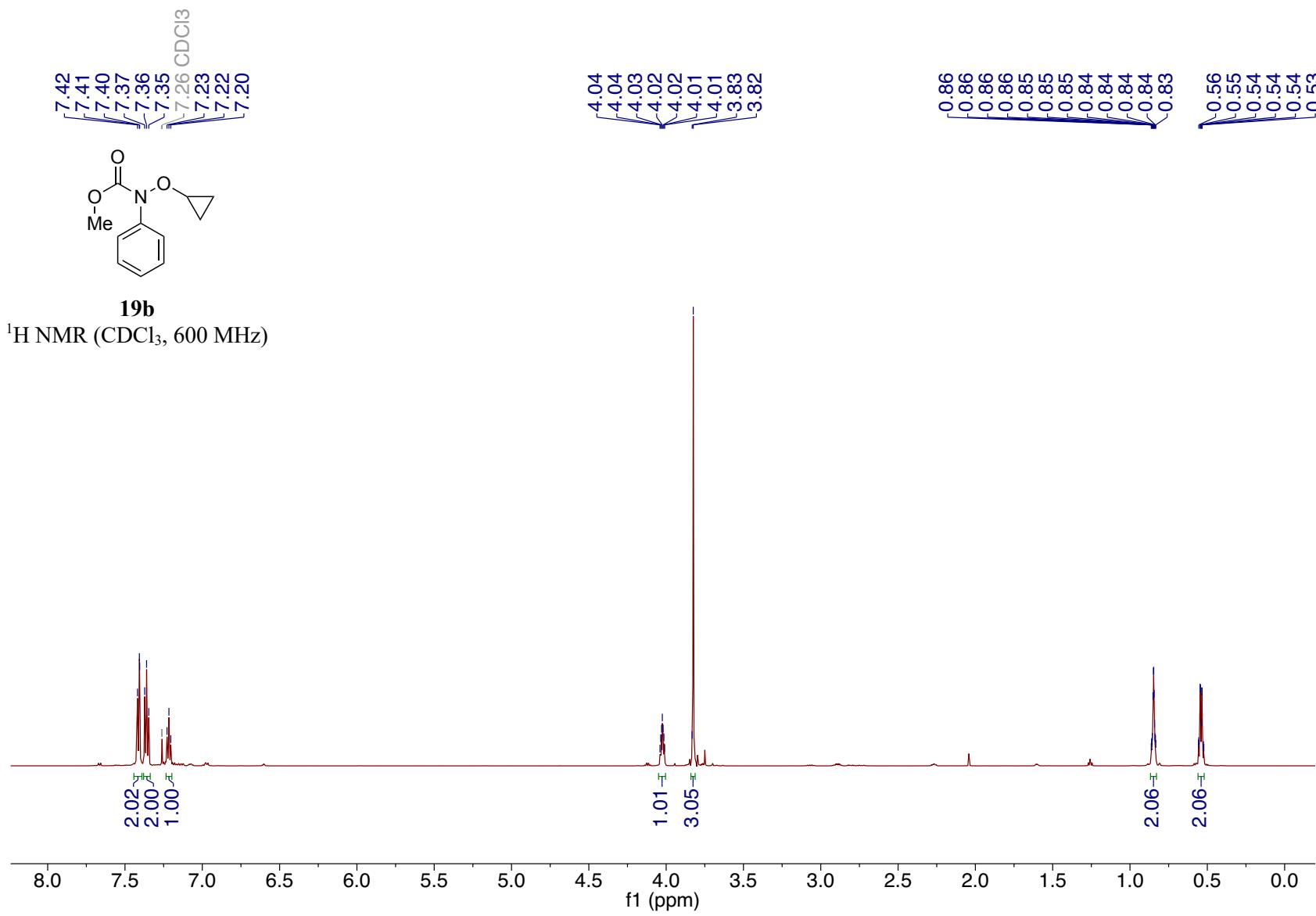
18e
 ^1H NMR (CDCl_3 , 600 MHz)

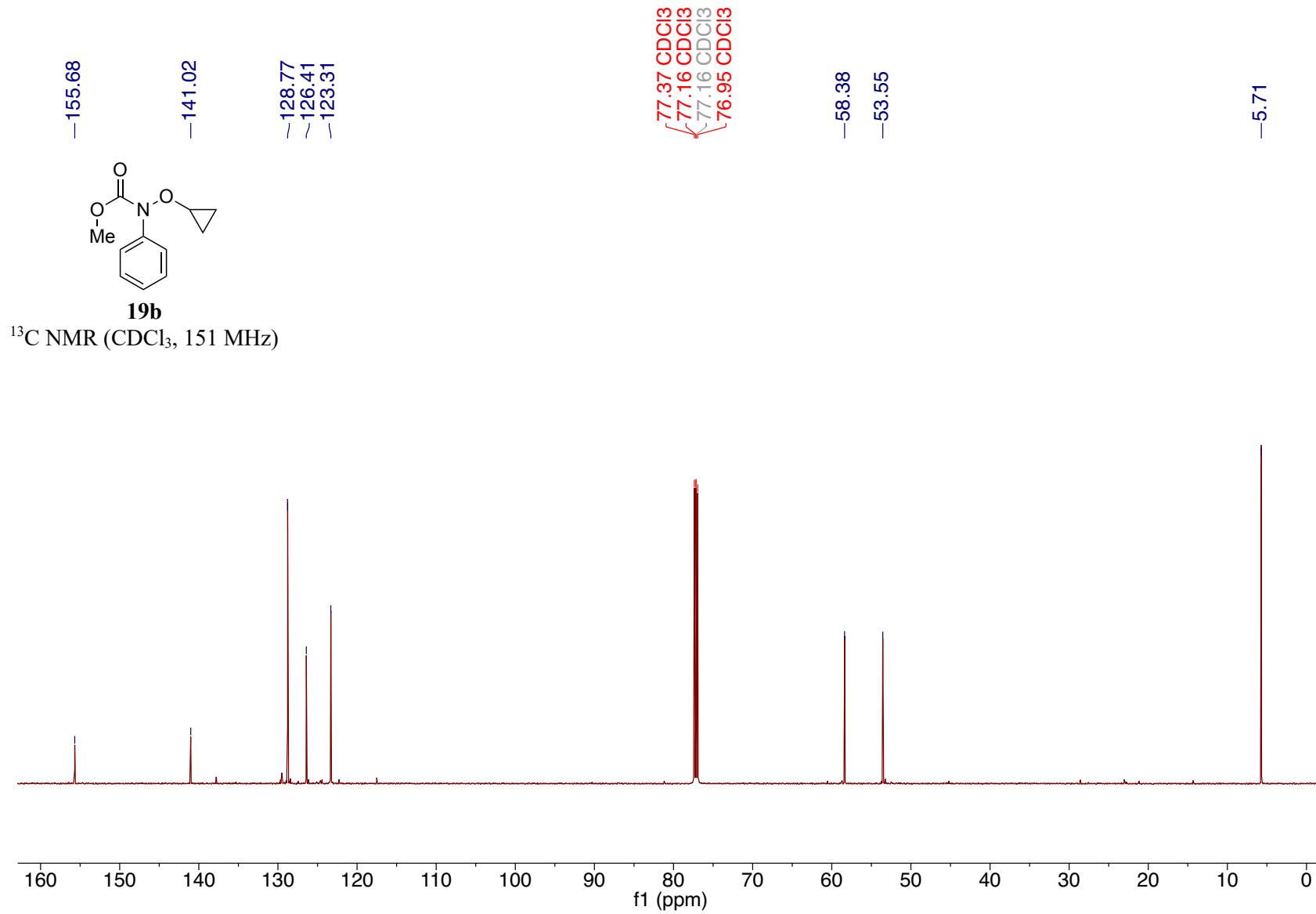


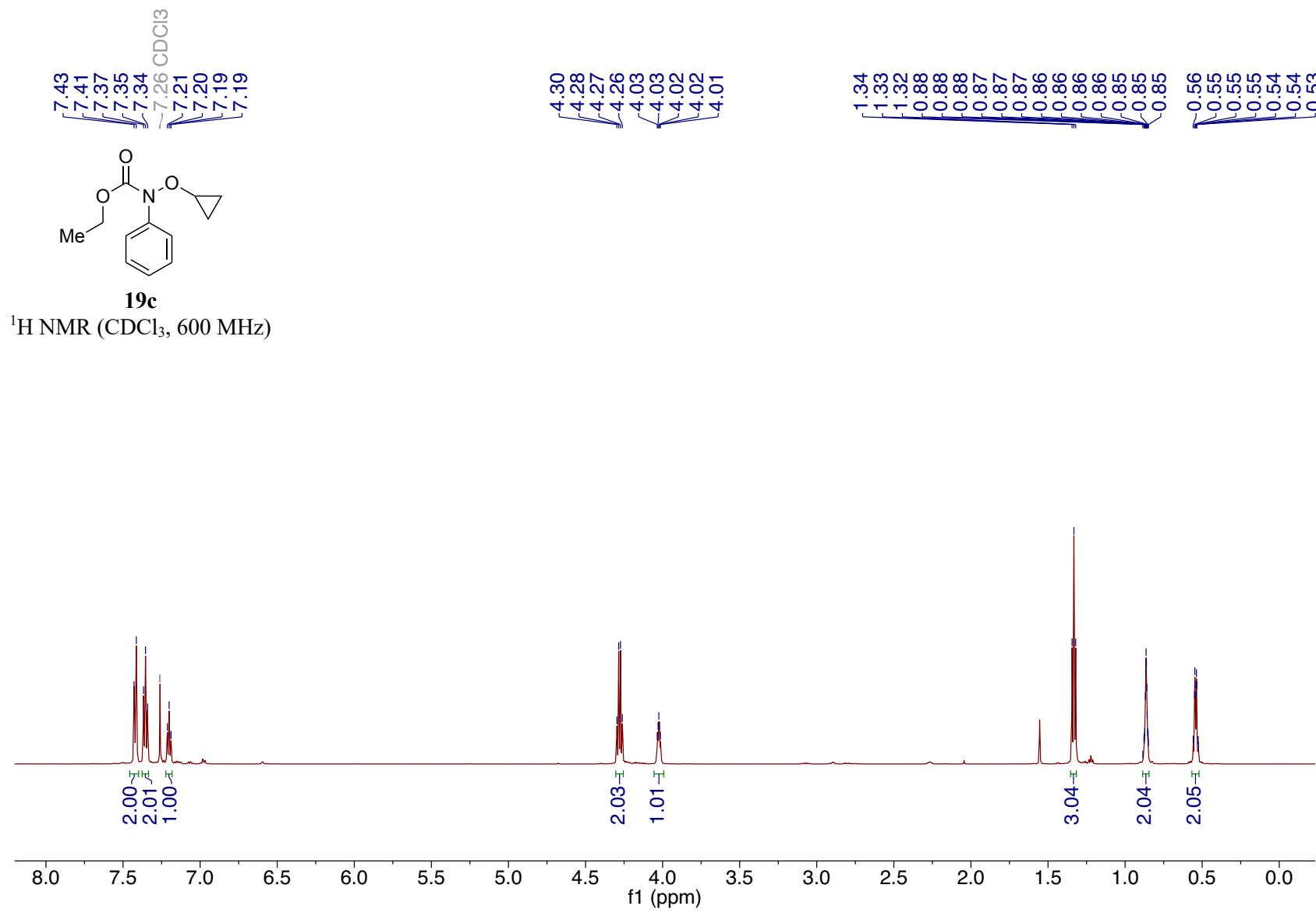


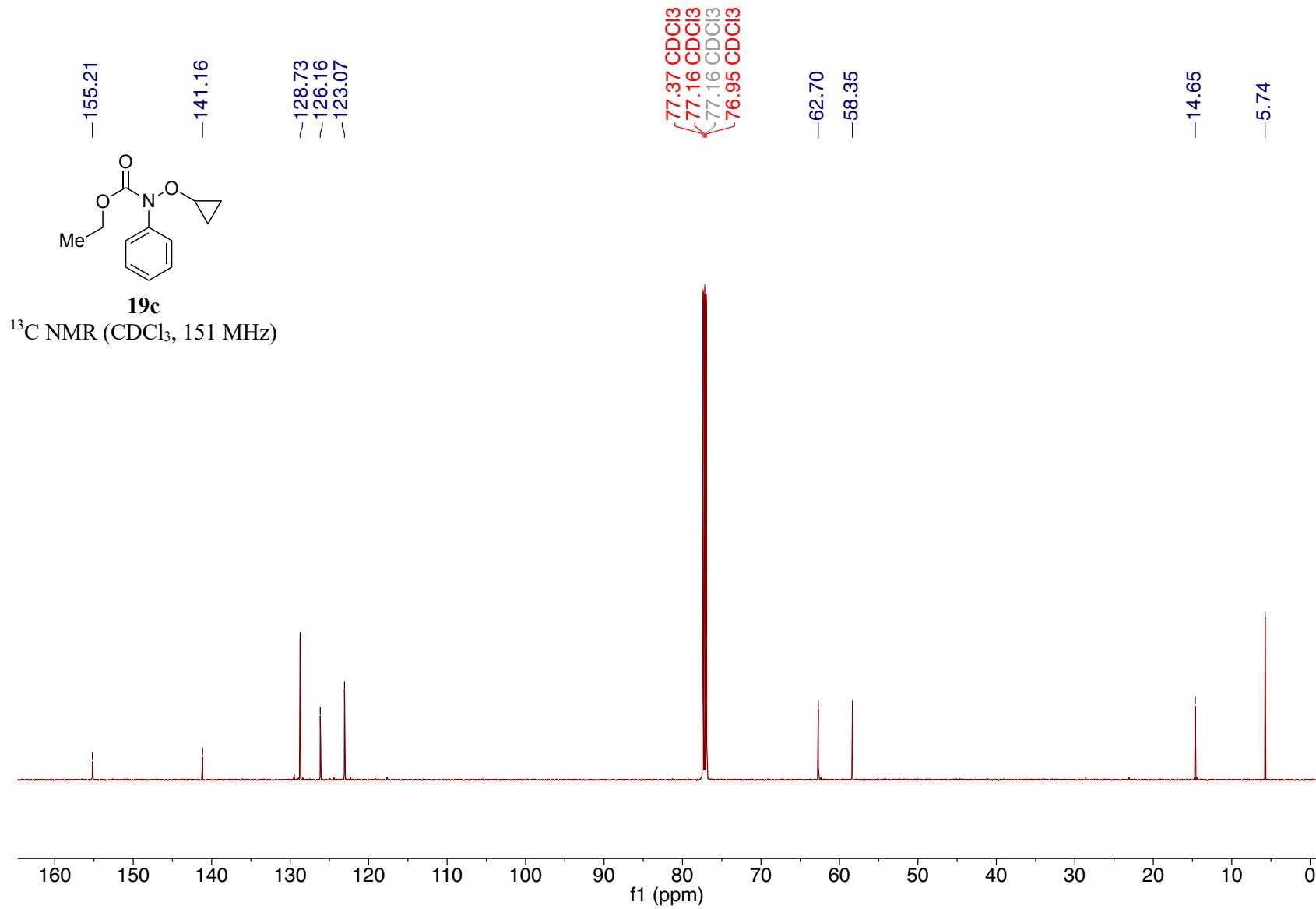


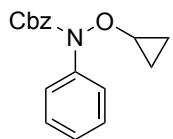
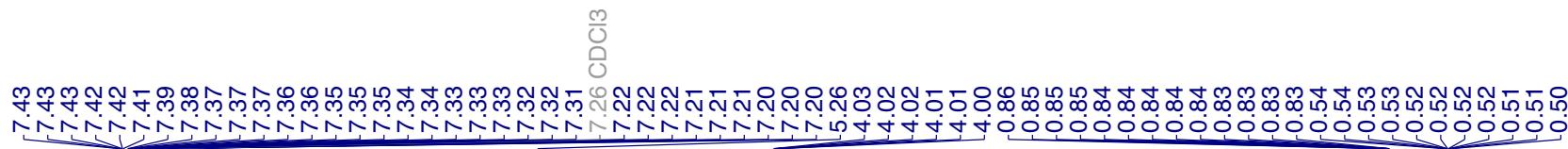






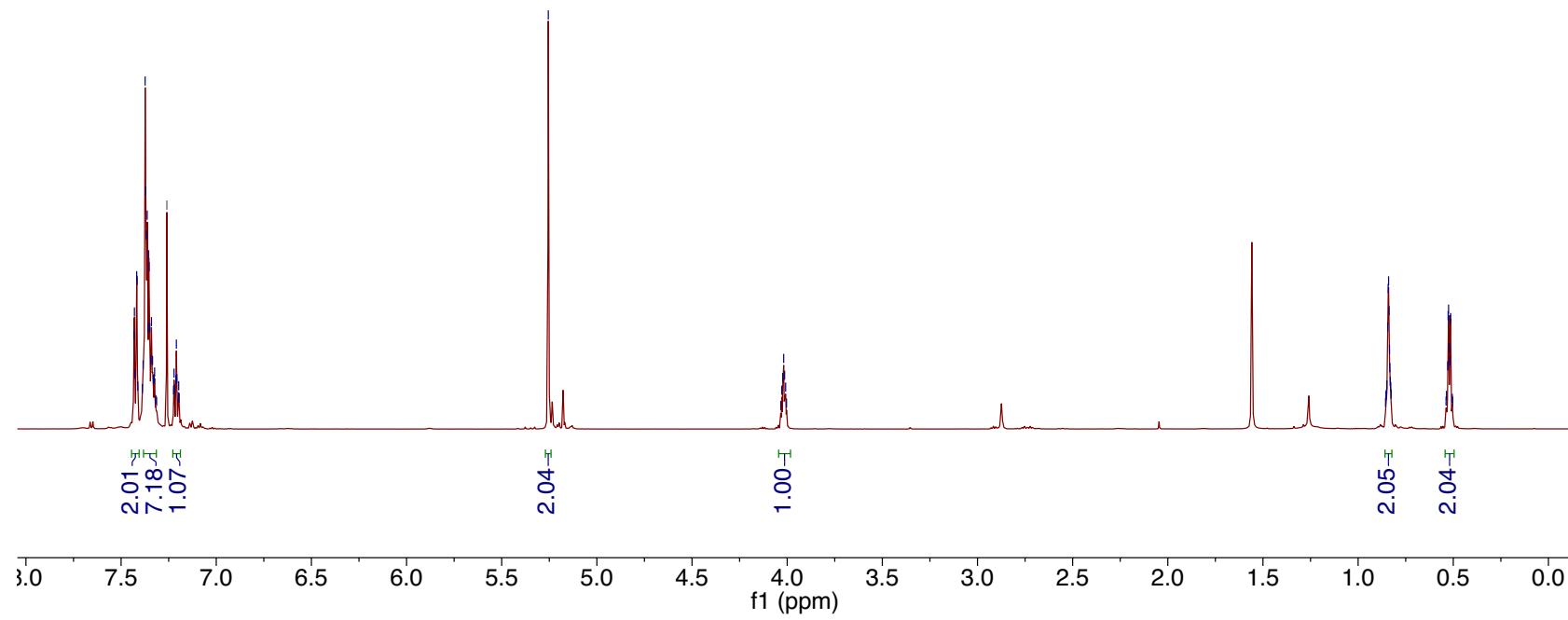


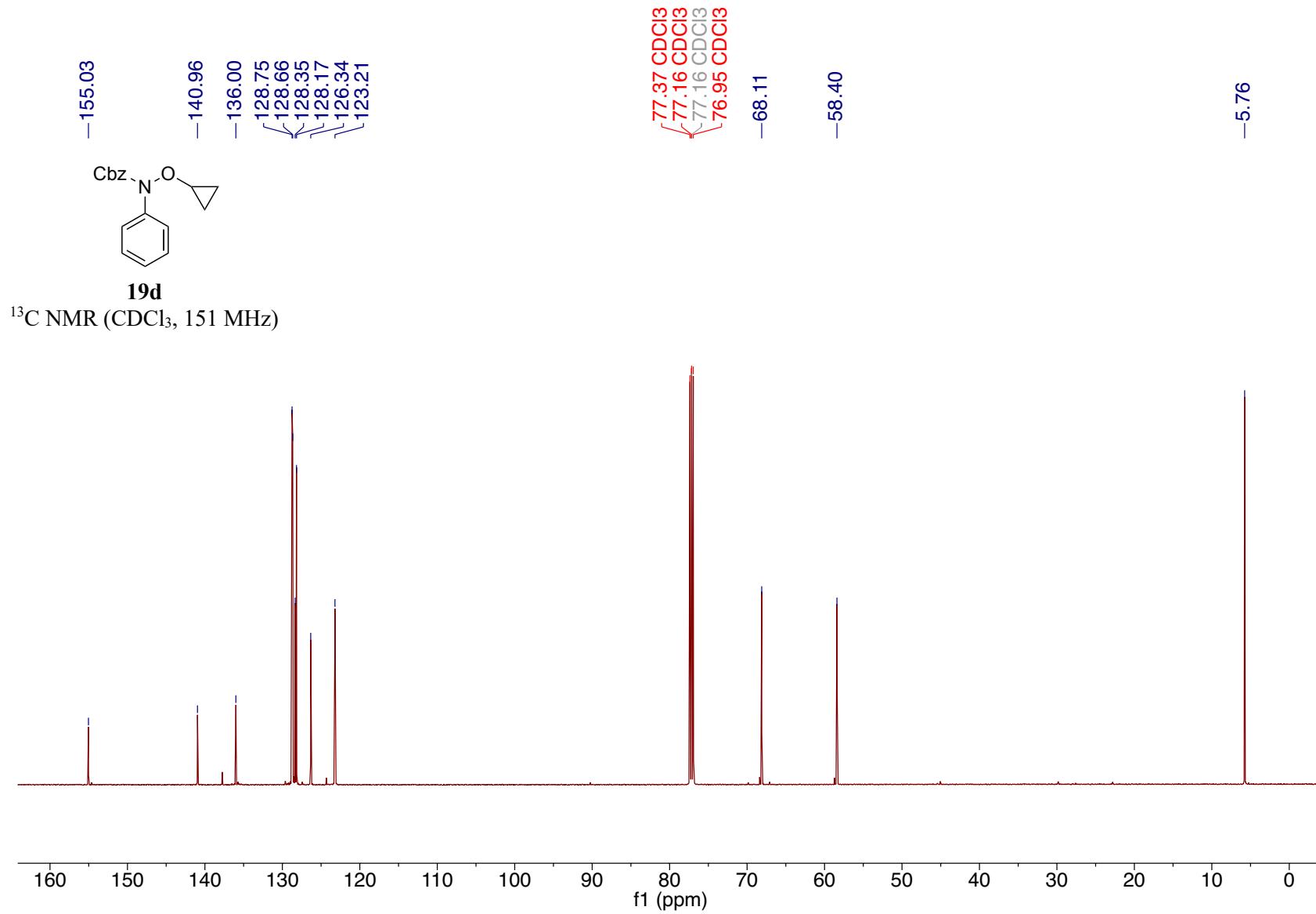


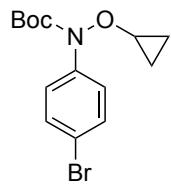


19d

¹H NMR (CDCl₃, 600 MHz)

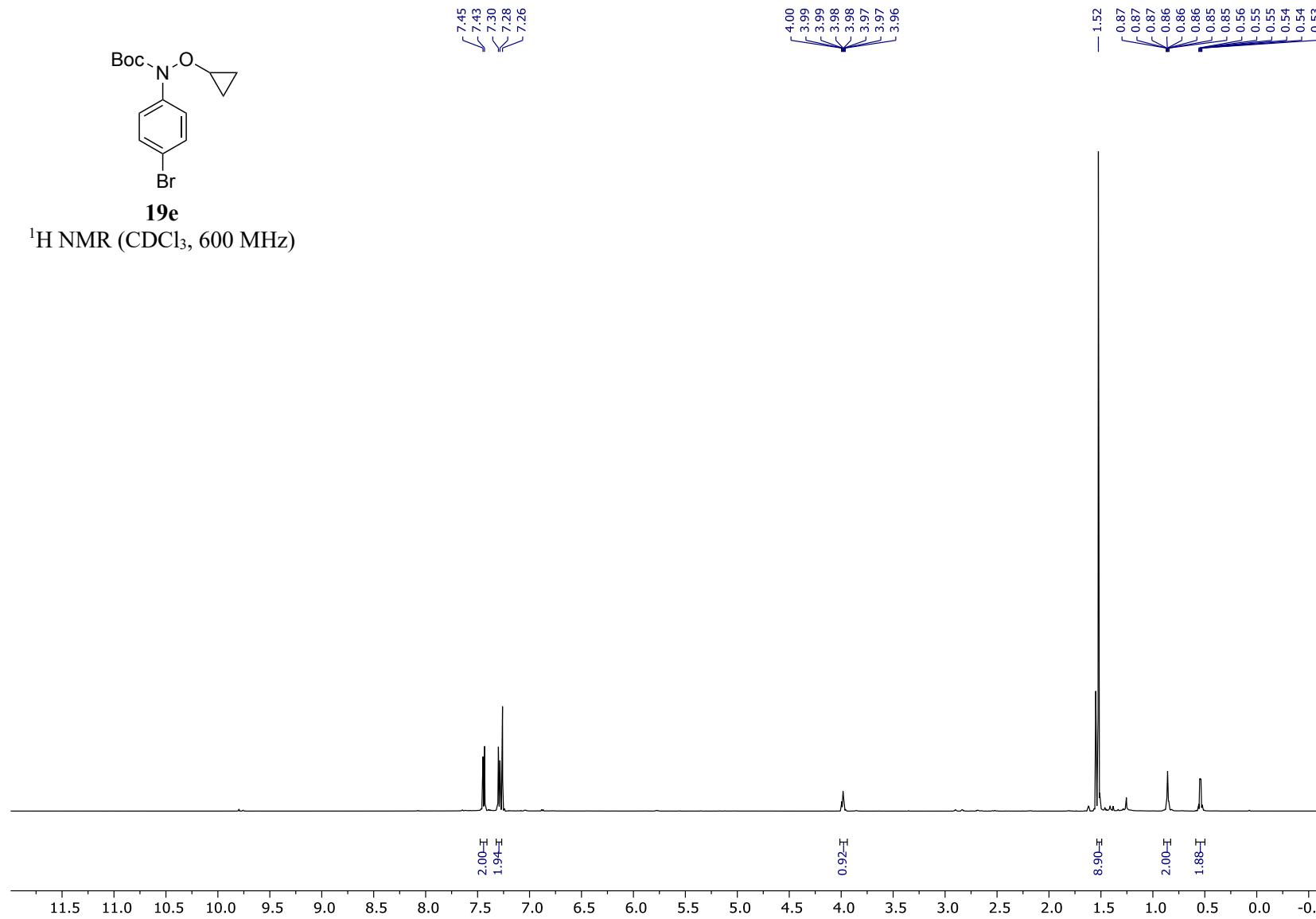


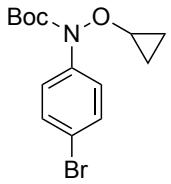




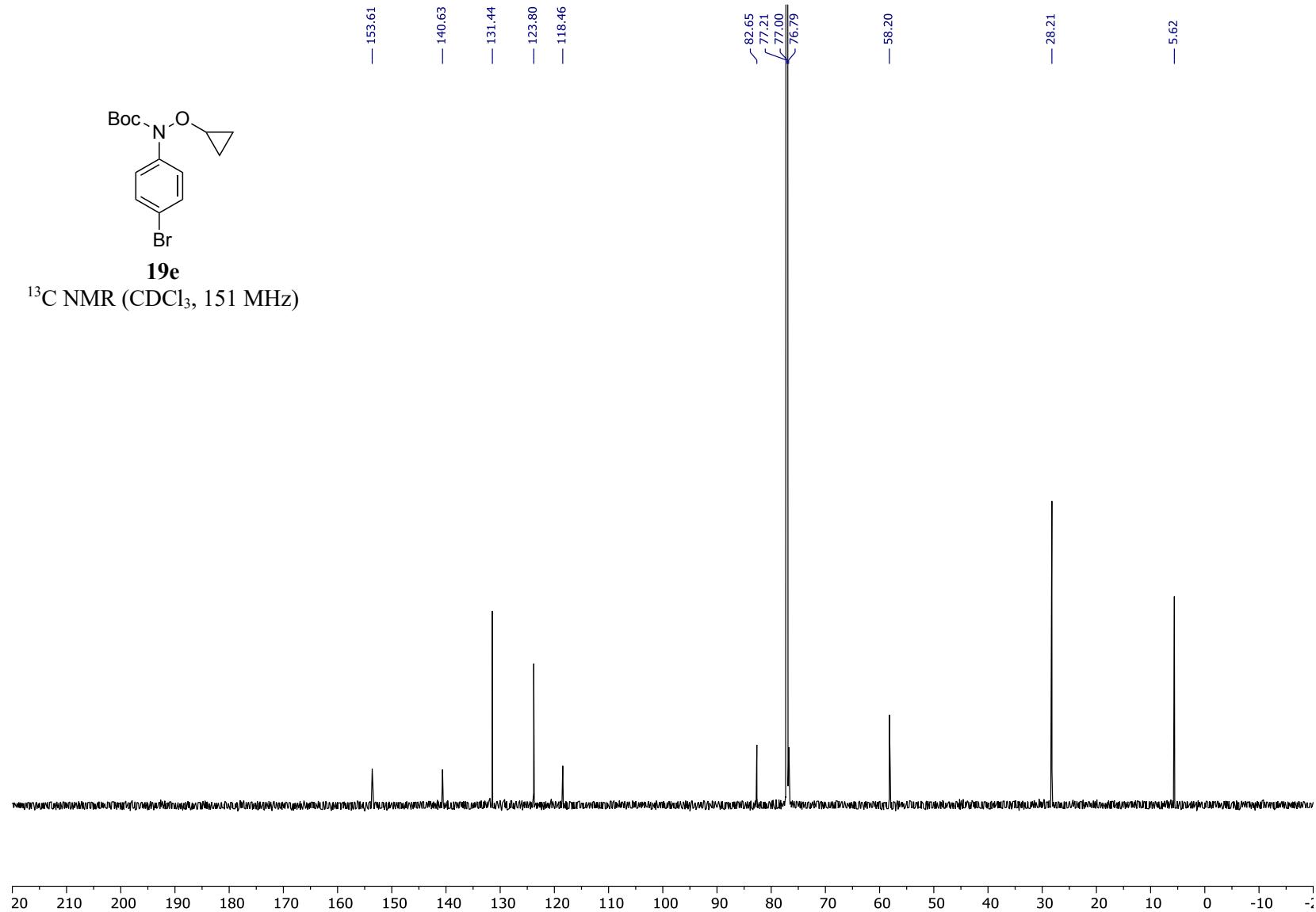
19e

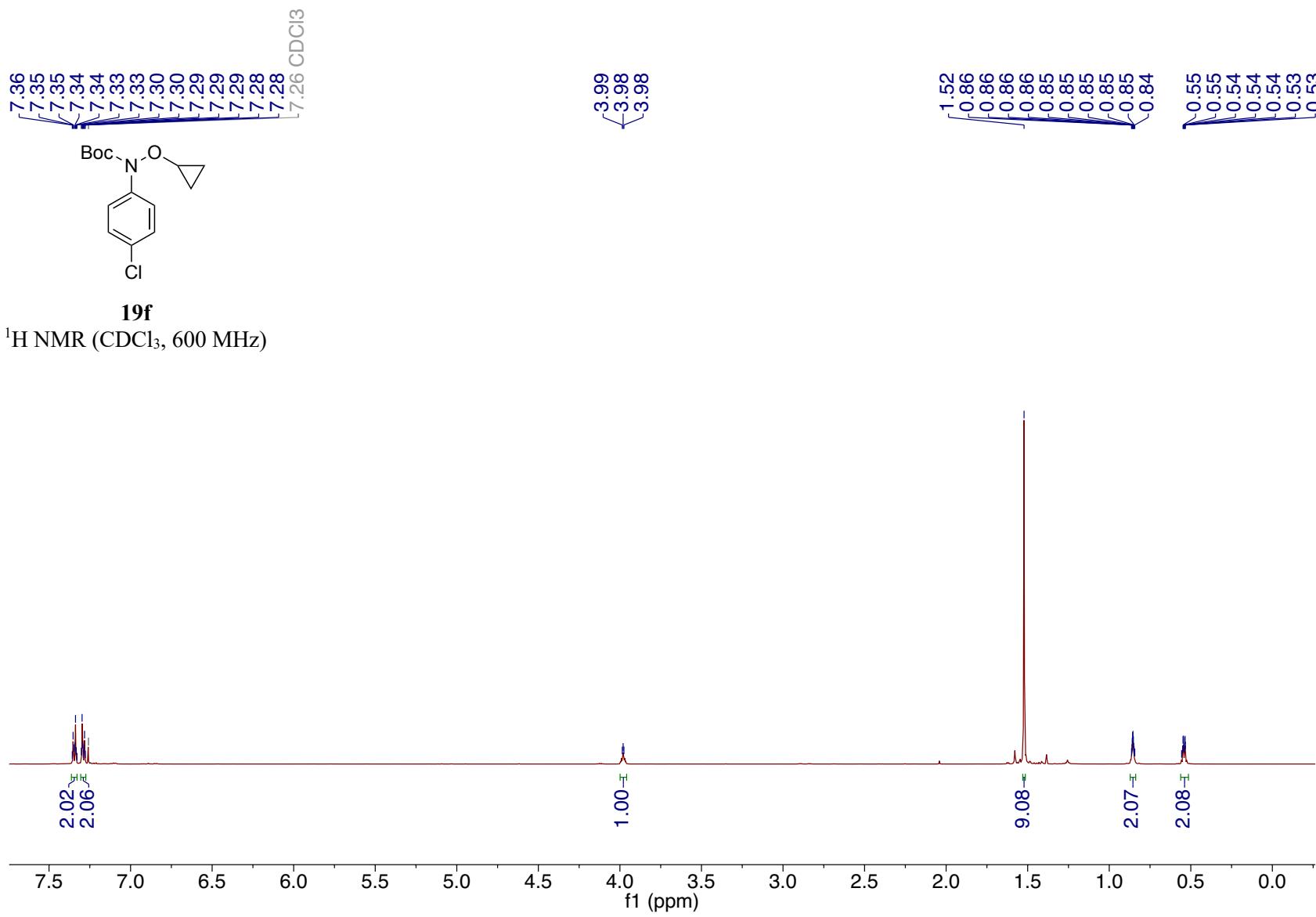
¹H NMR (CDCl₃, 600 MHz)

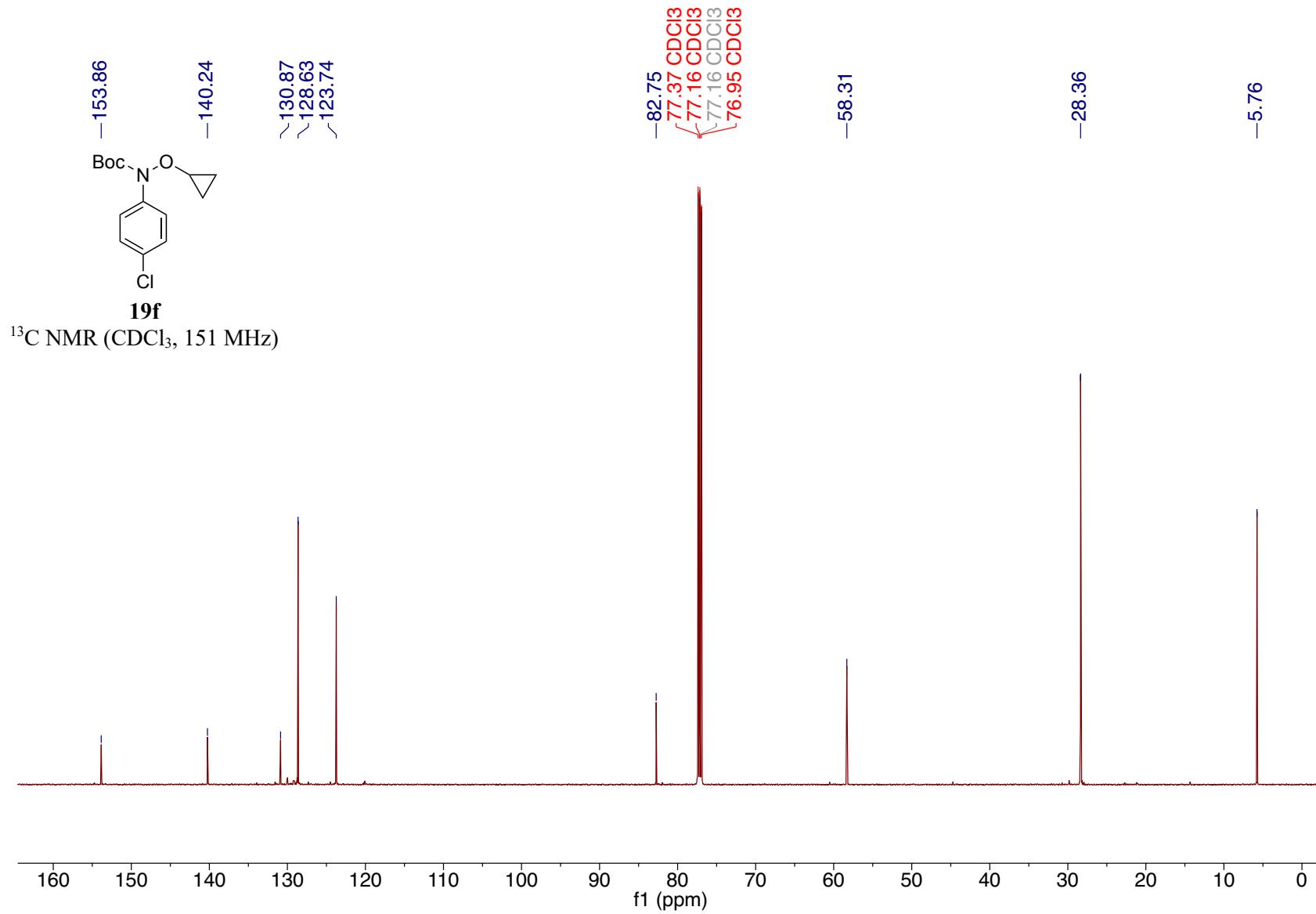


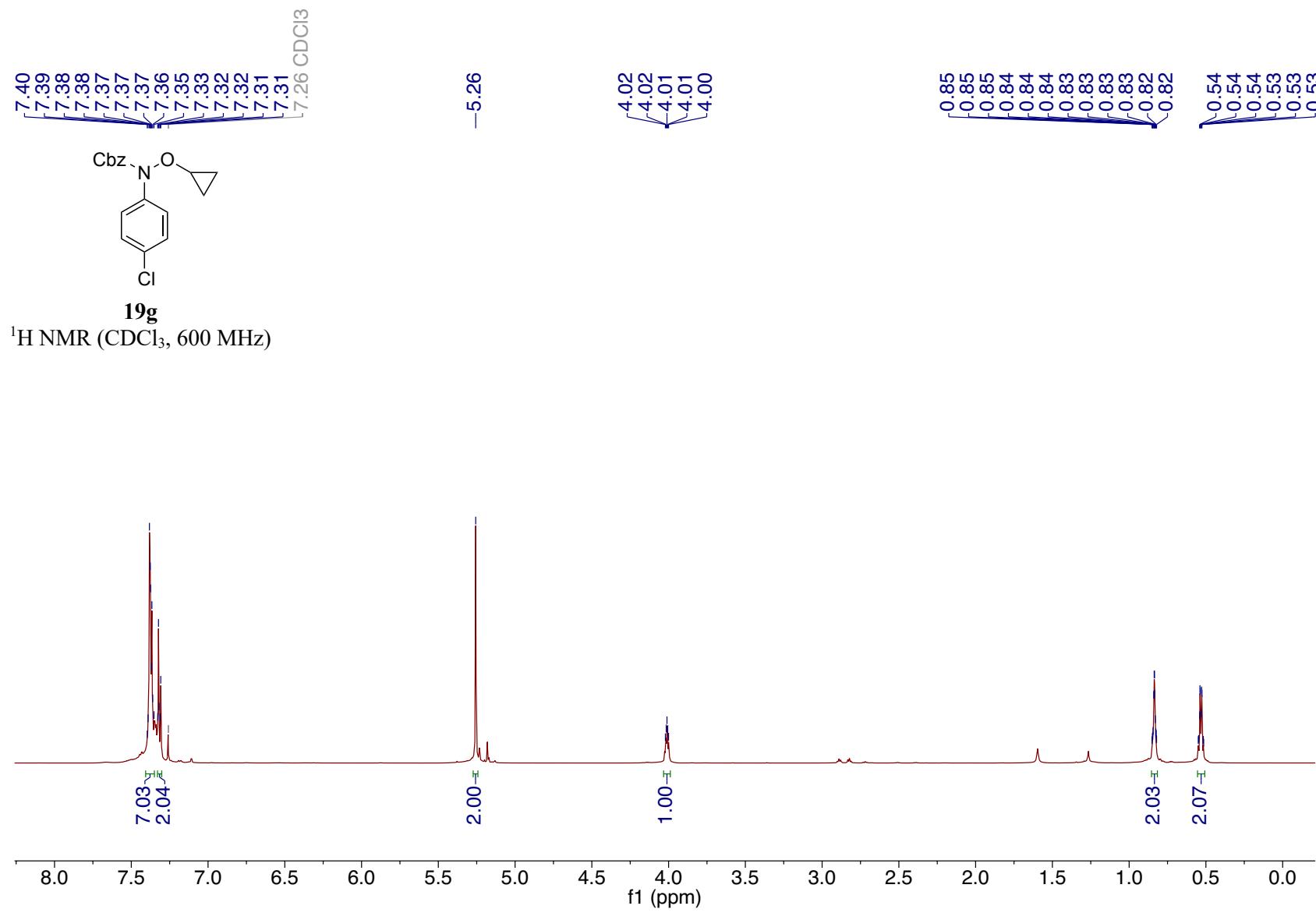


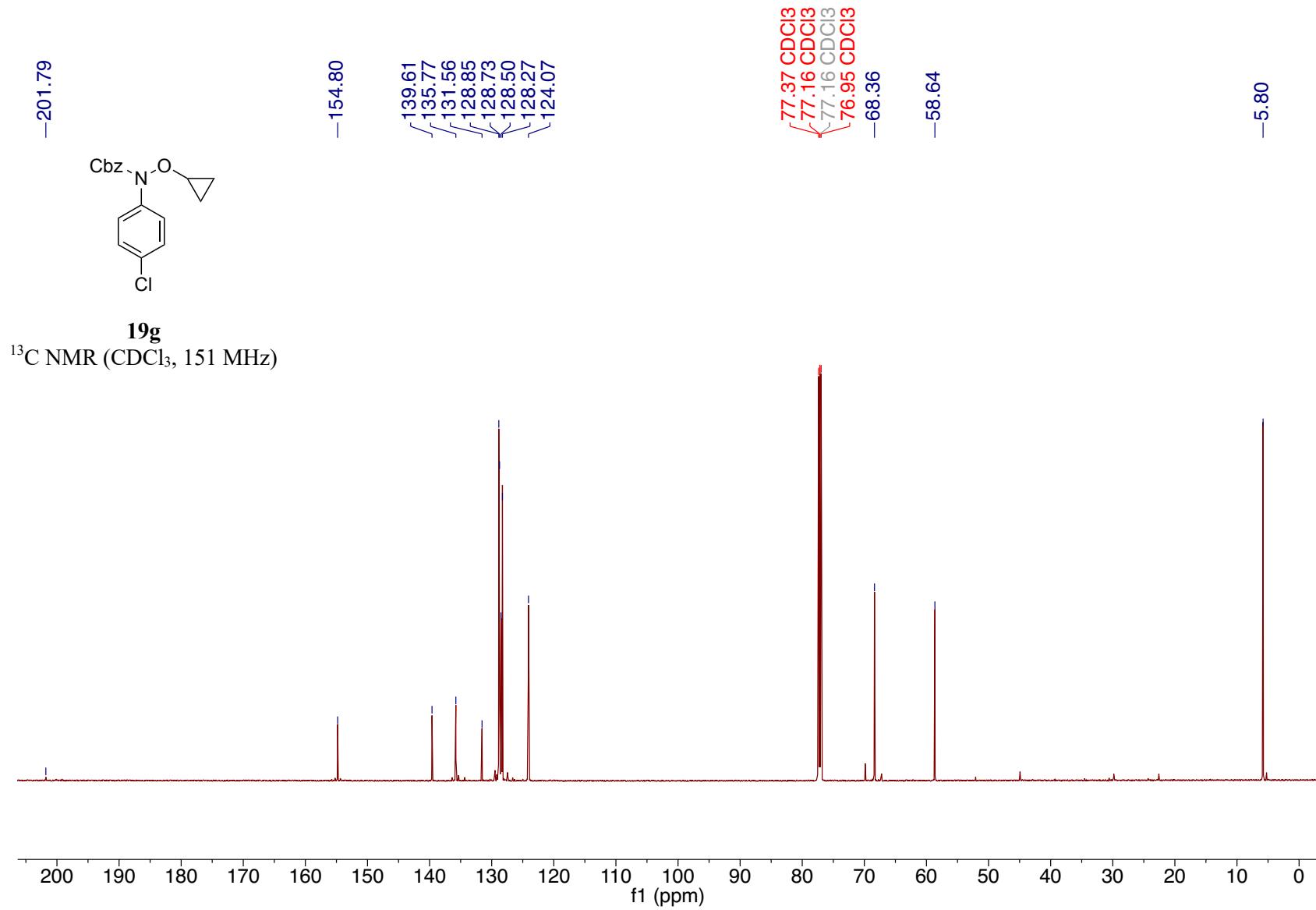
19e
 ^{13}C NMR (CDCl_3 , 151 MHz)

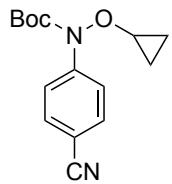






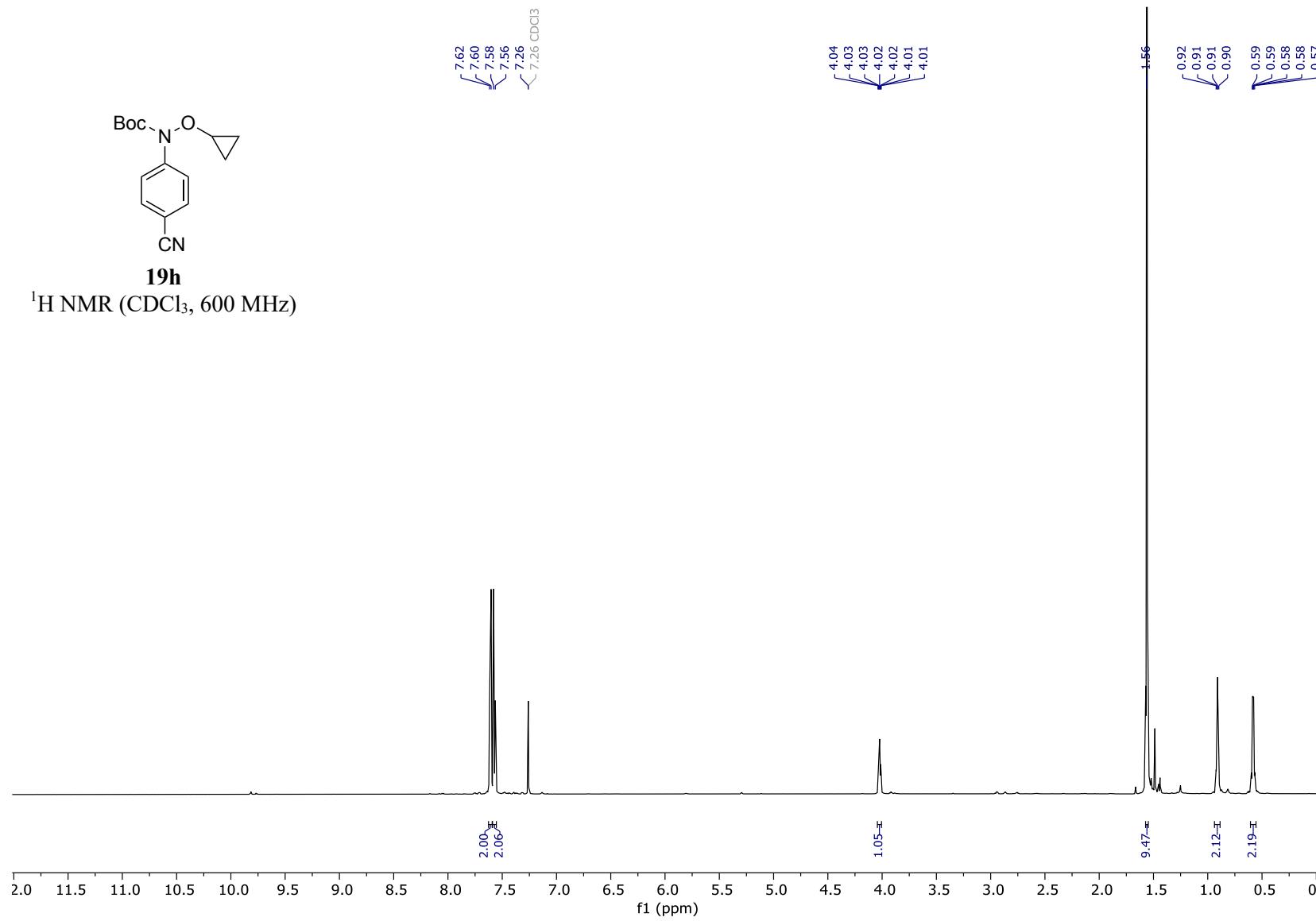


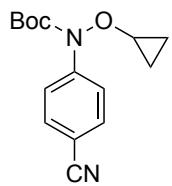




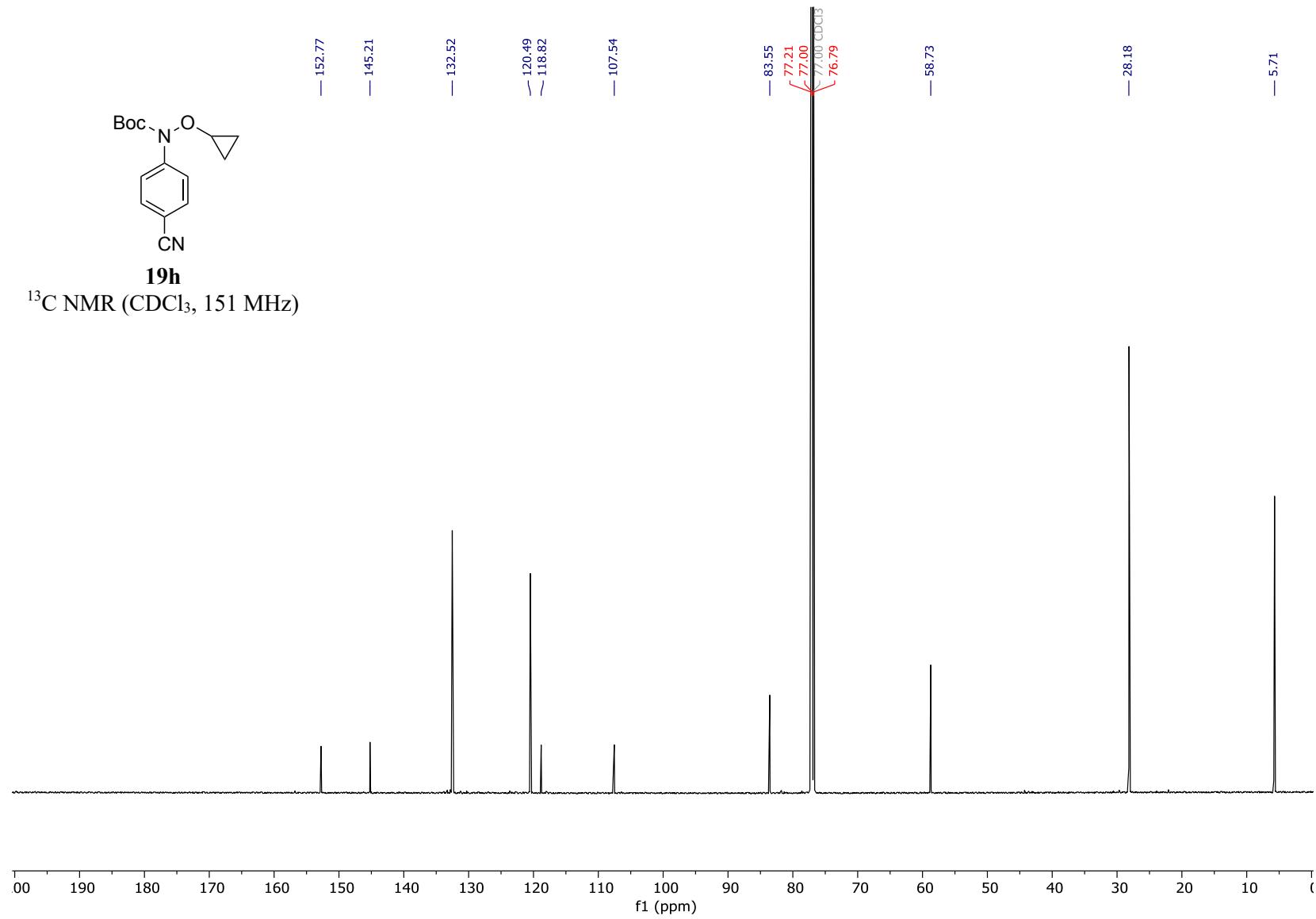
19h

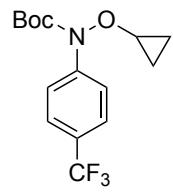
^1H NMR (CDCl_3 , 600 MHz)





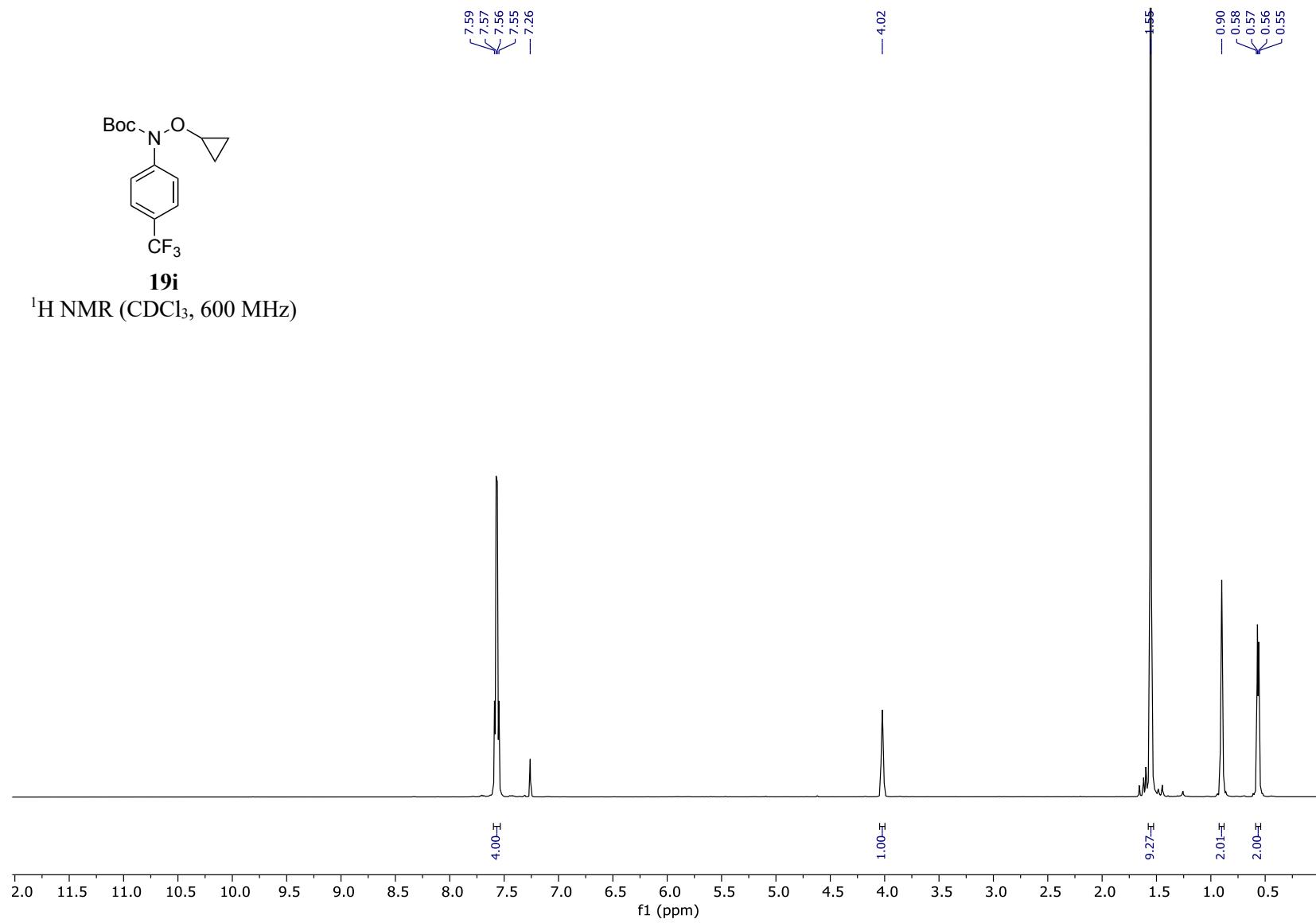
^{13}C NMR (CDCl_3 , 151 MHz)

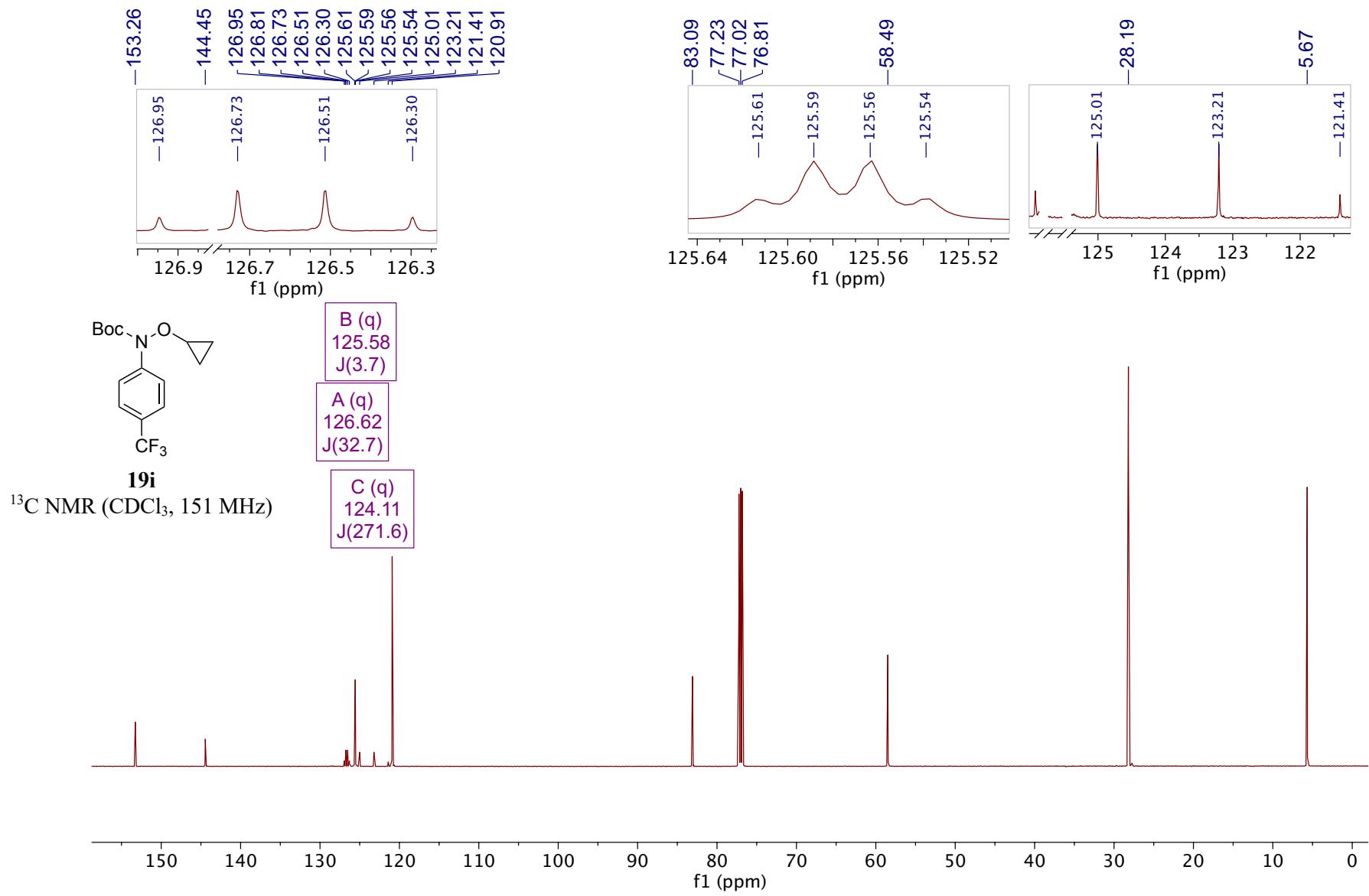


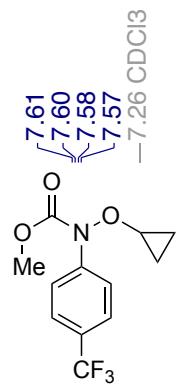


19i

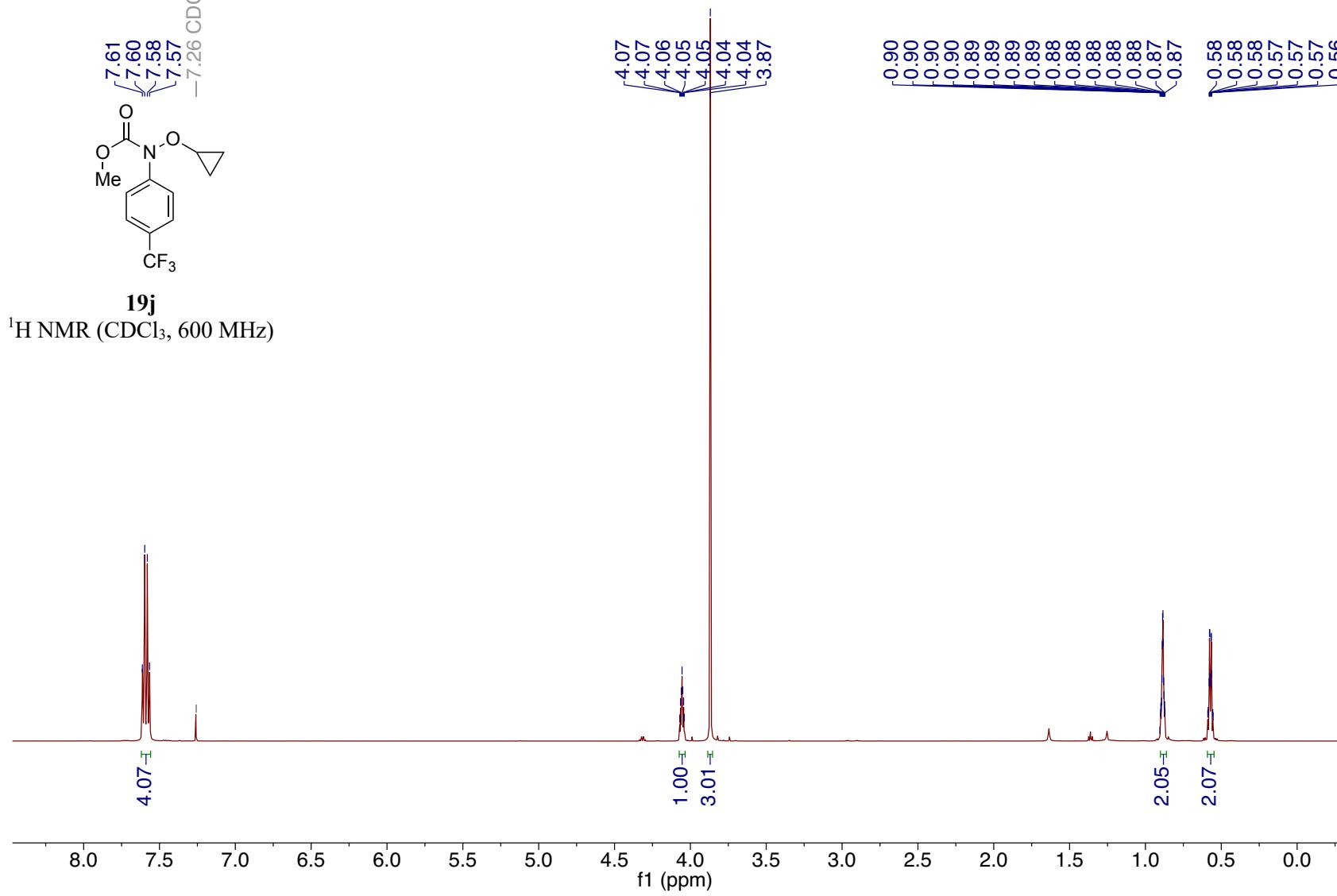
^1H NMR (CDCl_3 , 600 MHz)

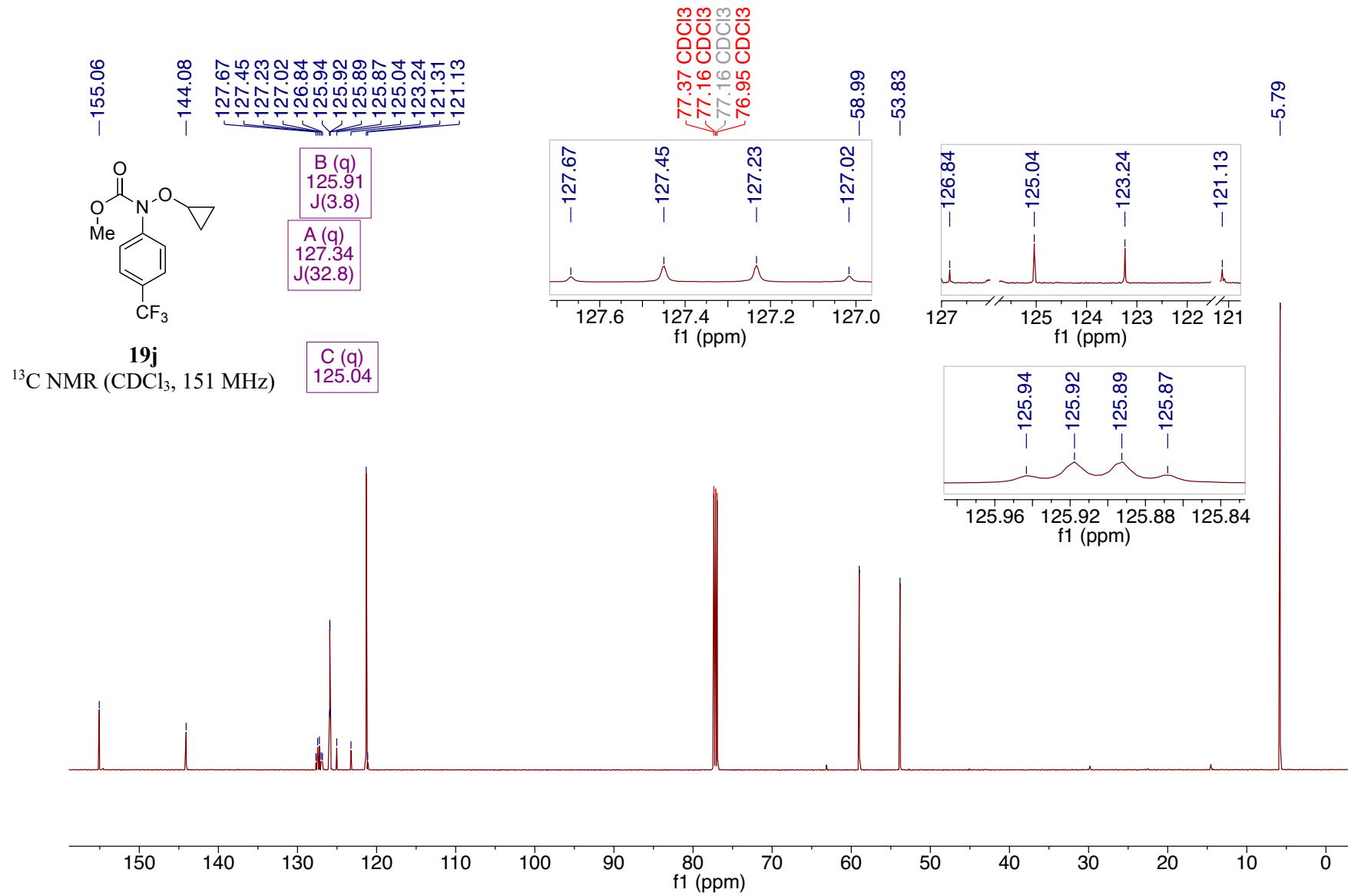




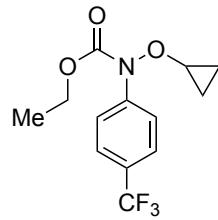


19j
¹H NMR (CDCl₃, 600 MHz)



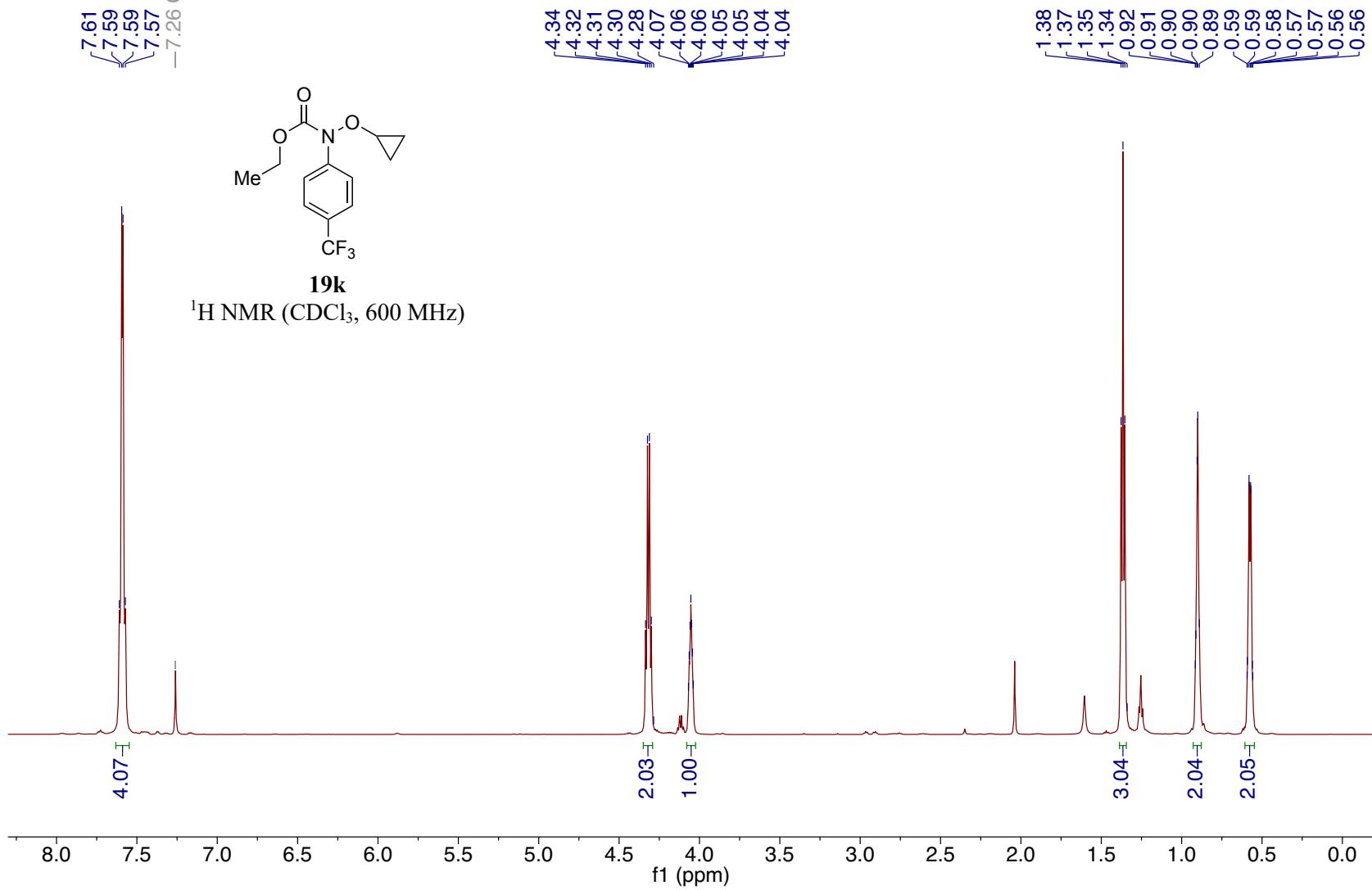


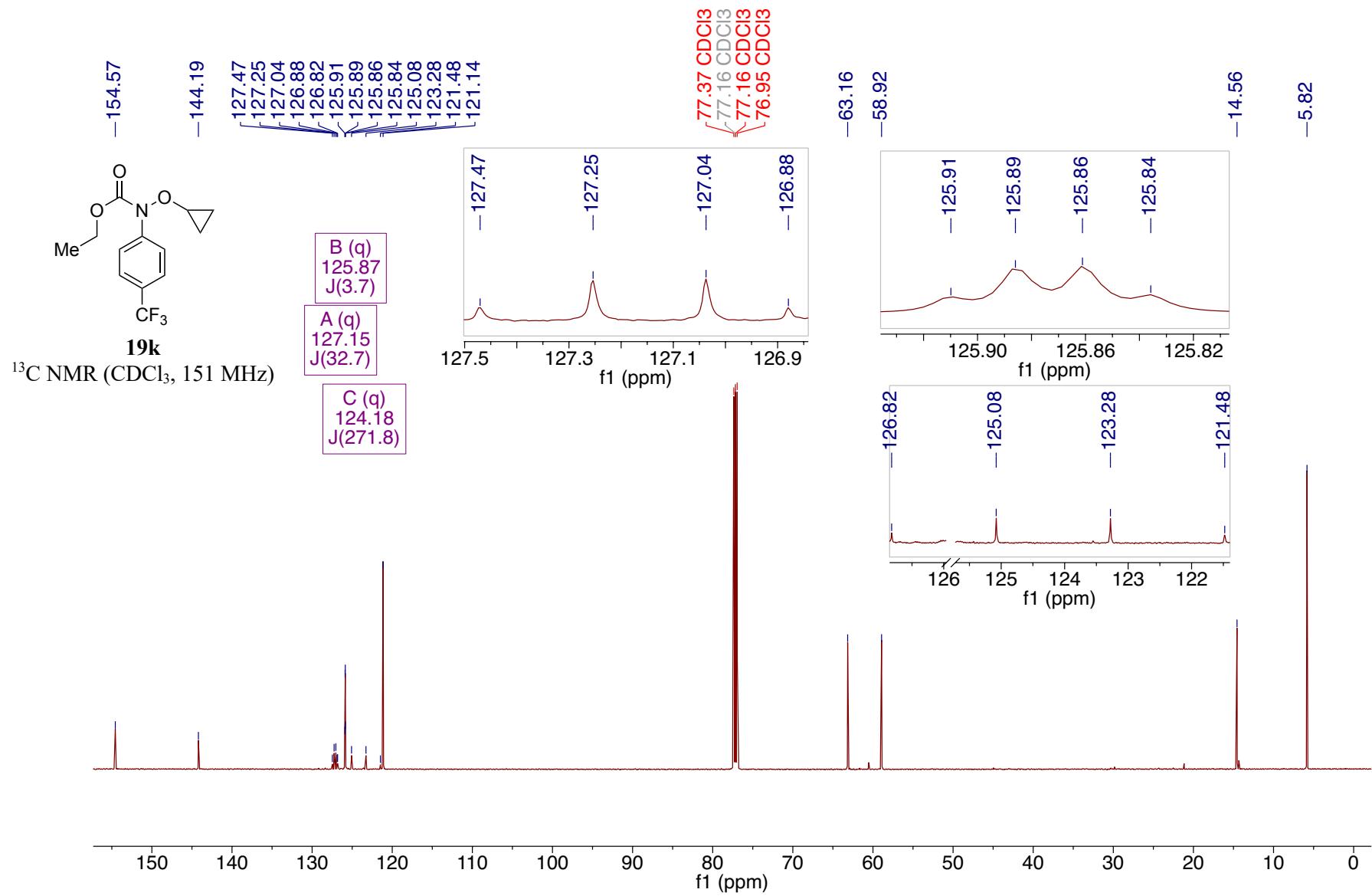
7.61
7.59
7.59
7.57
-7.26 CDCl₃

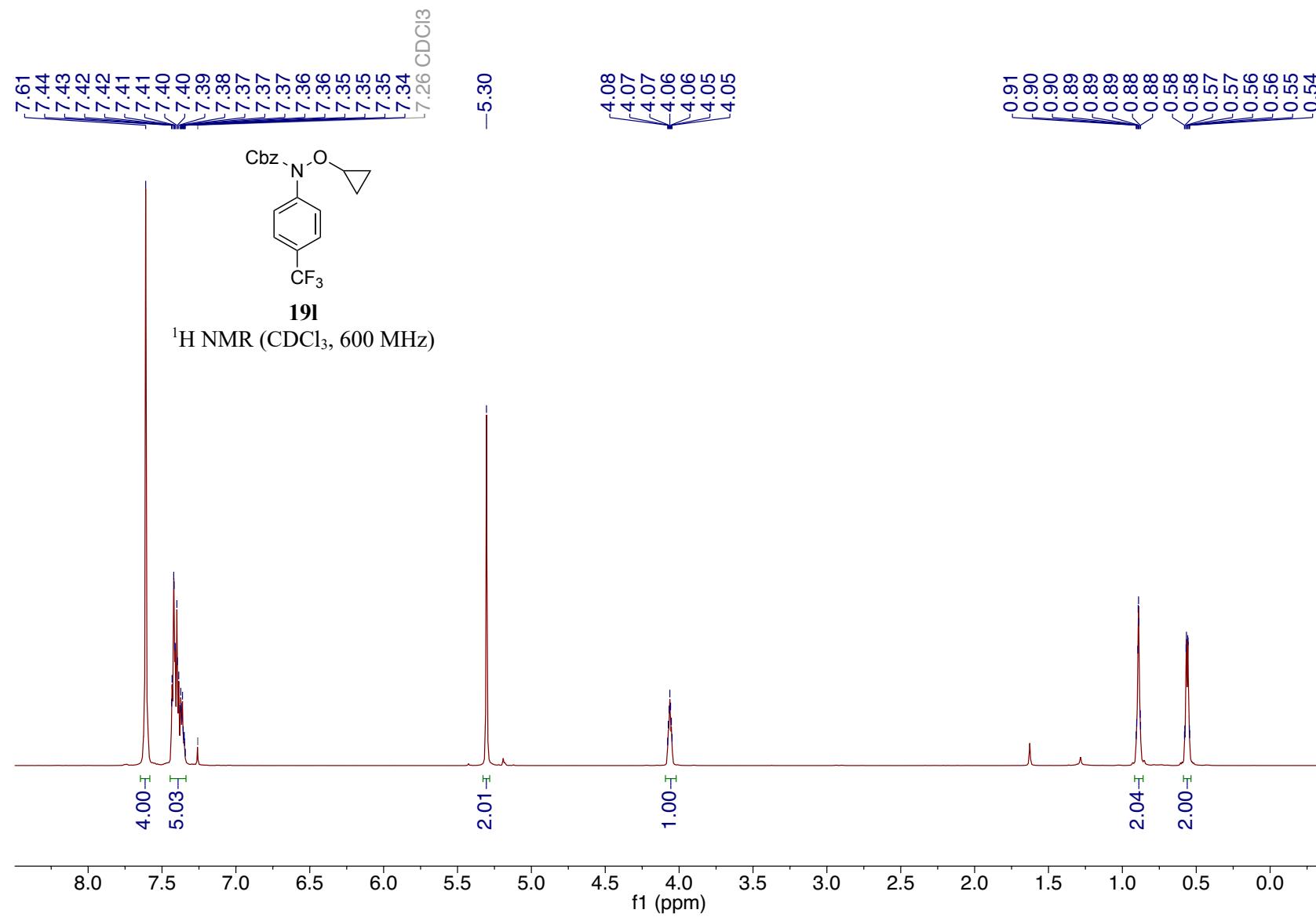


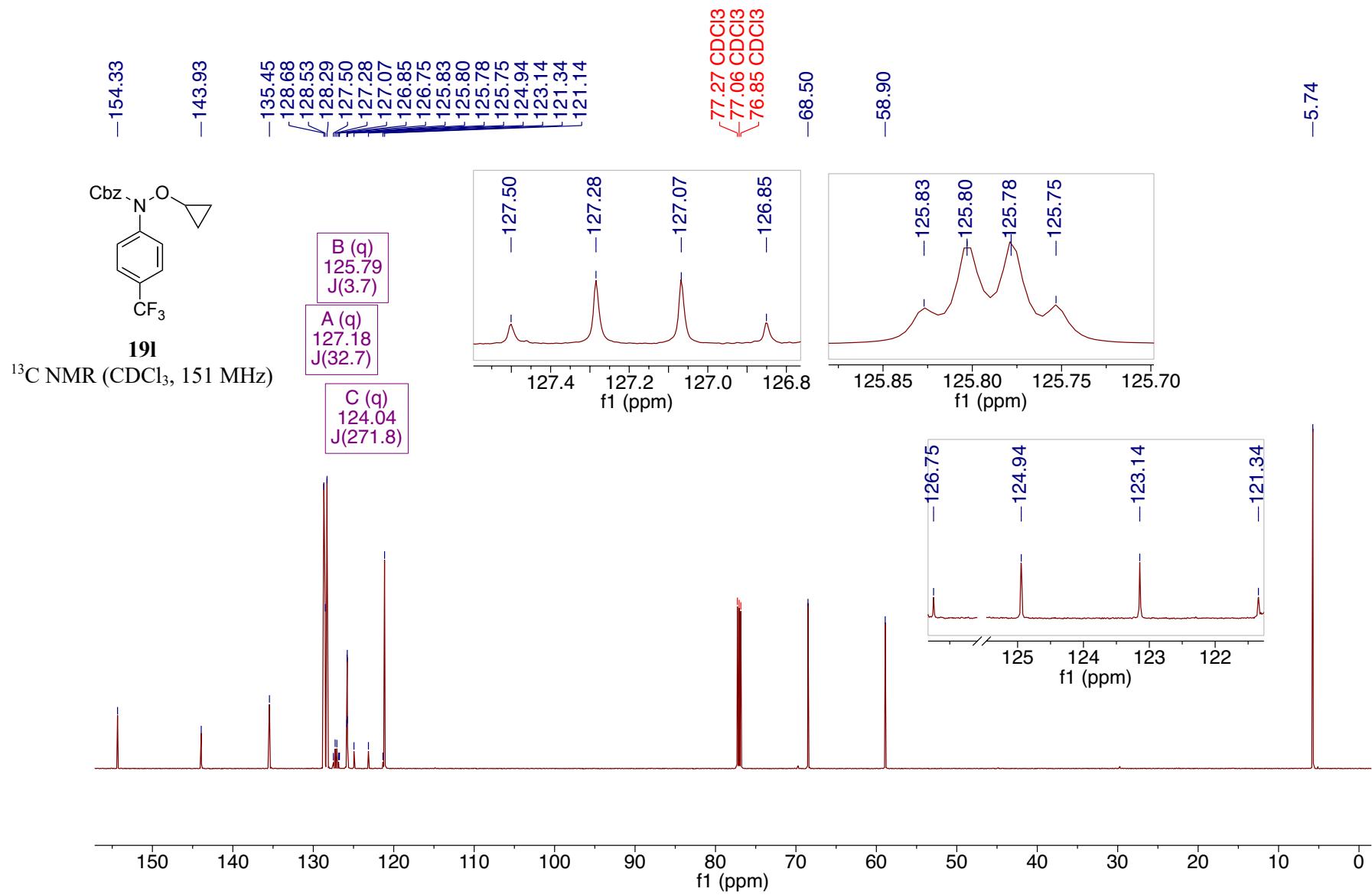
19k

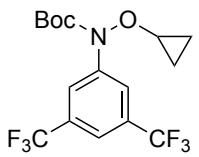
¹H NMR (CDCl₃, 600 MHz)



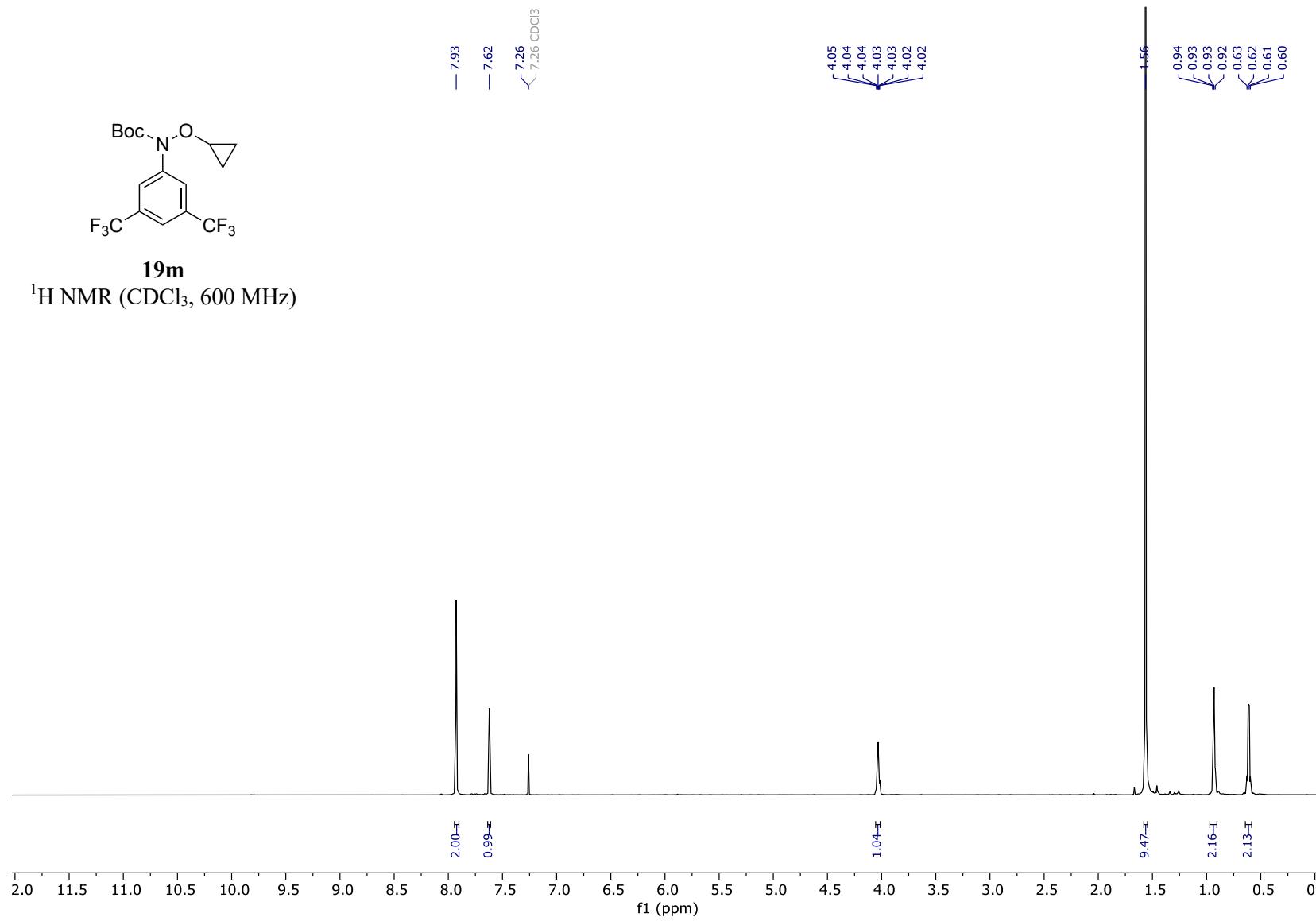


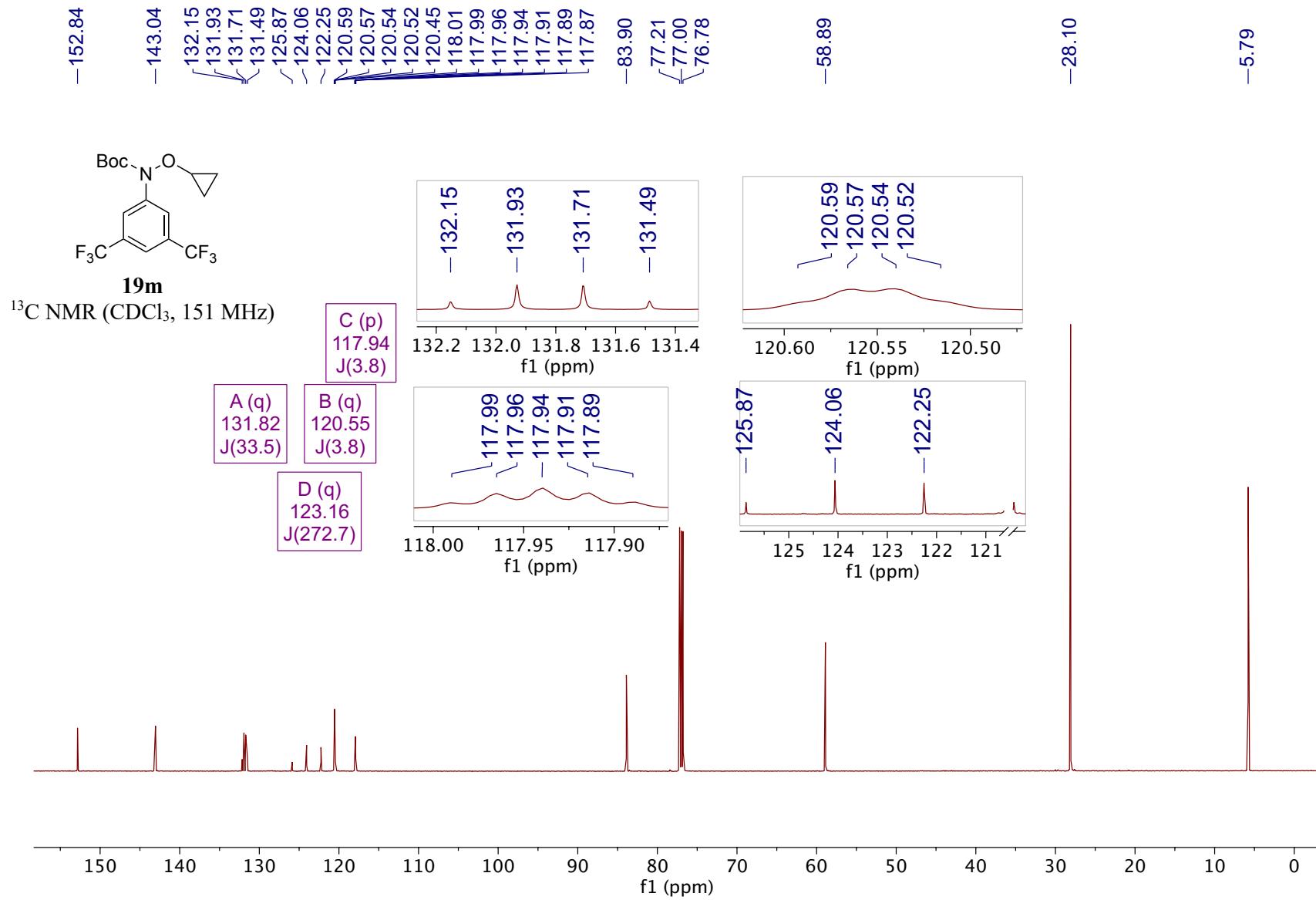


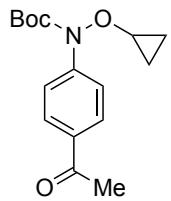




¹H NMR (CDCl₃, 600 MHz)

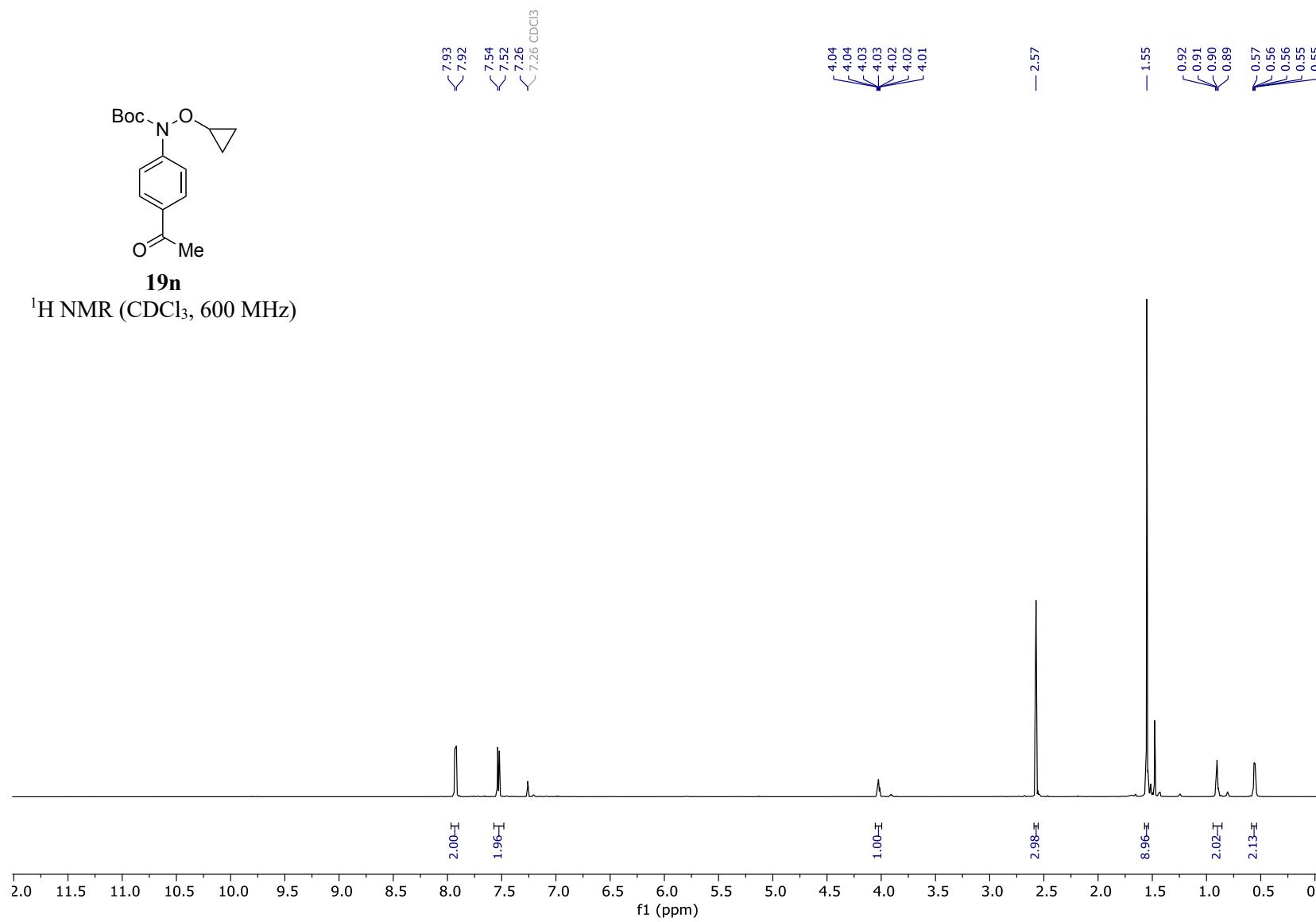


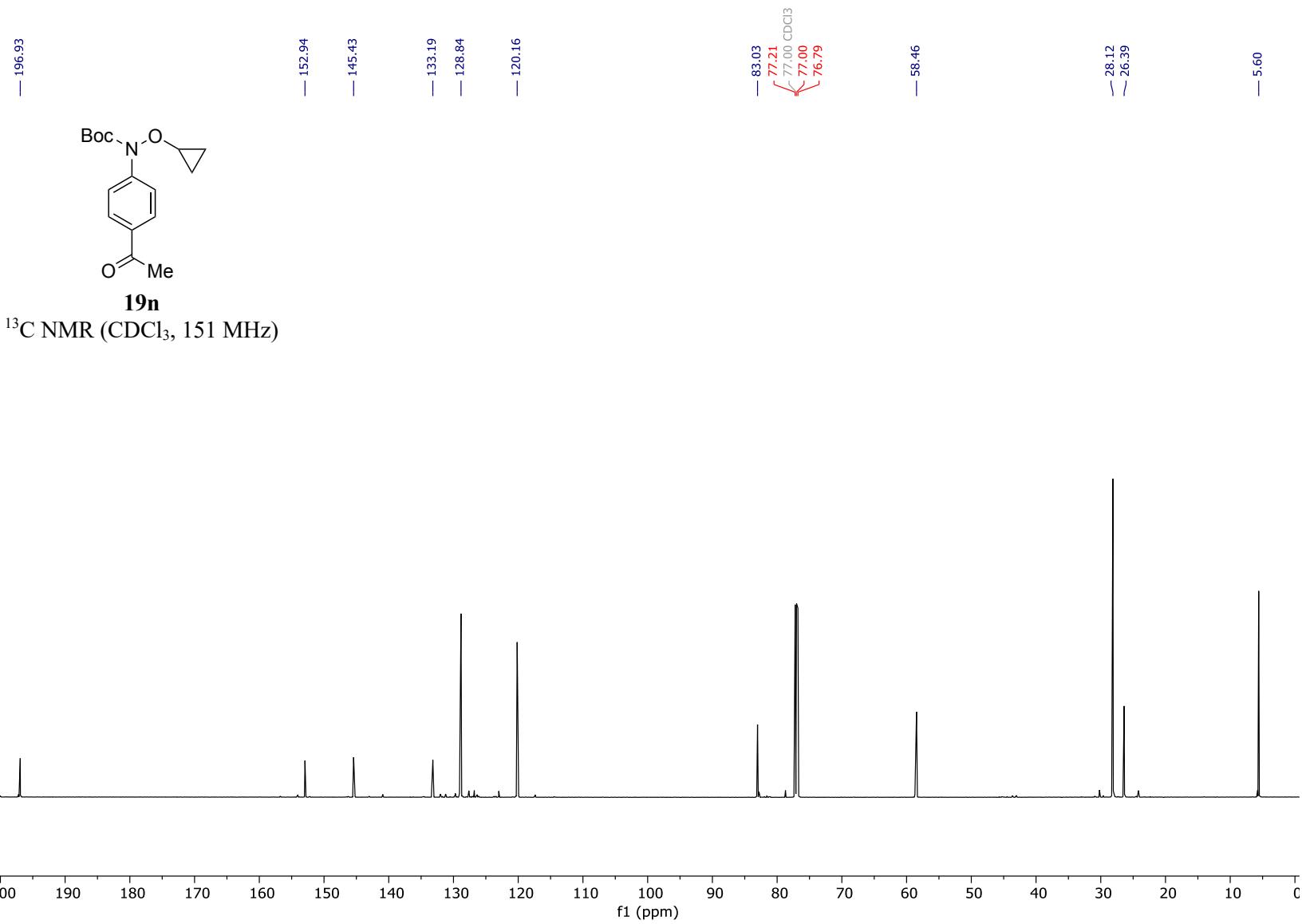


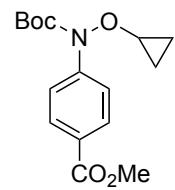


19n

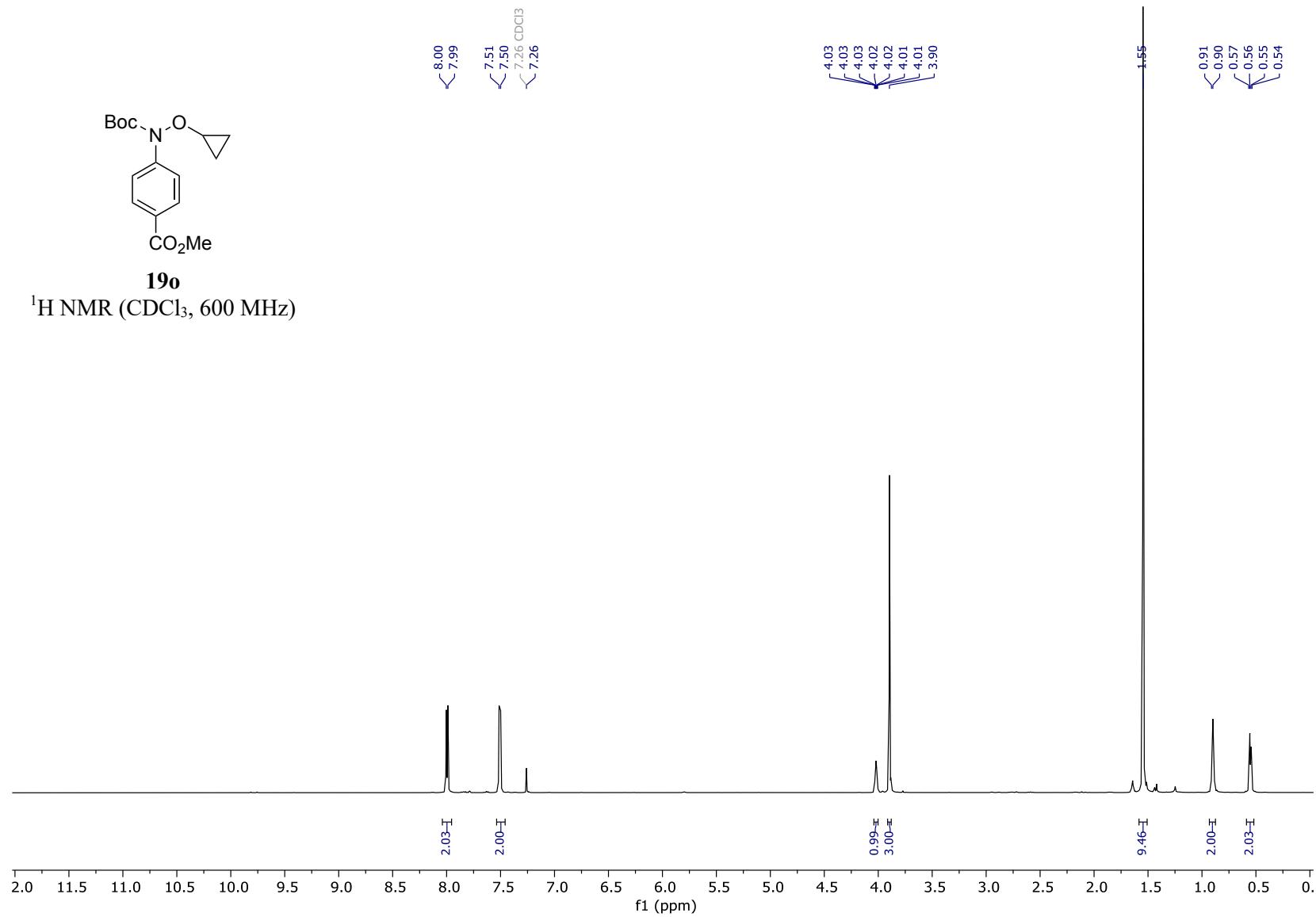
¹H NMR (CDCl₃, 600 MHz)

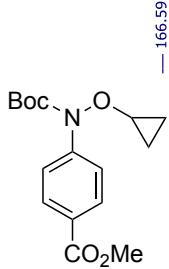






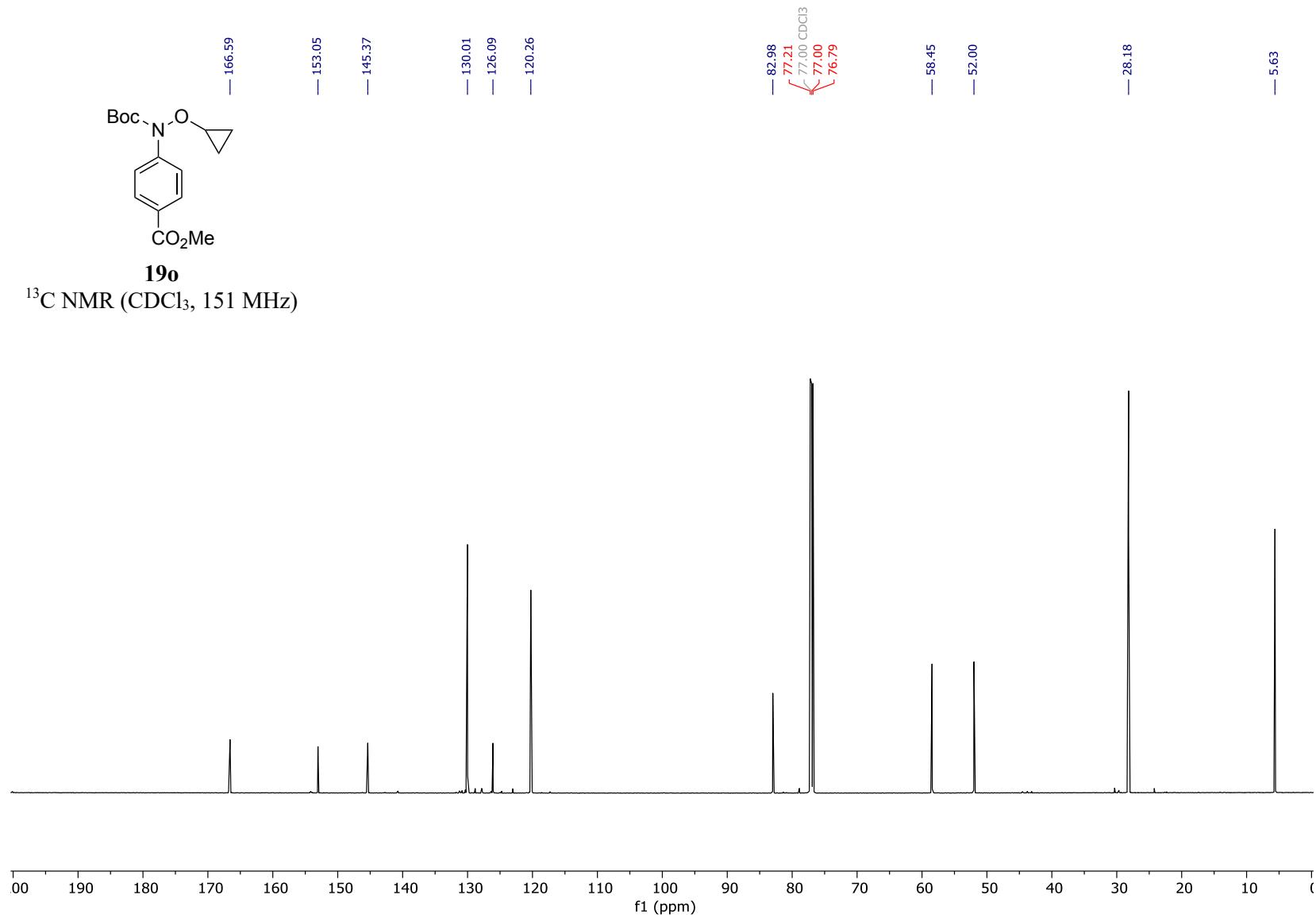
19o
 ^1H NMR (CDCl_3 , 600 MHz)

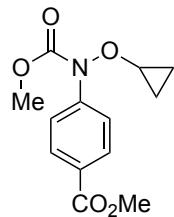
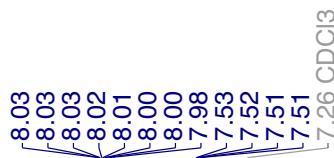




19o

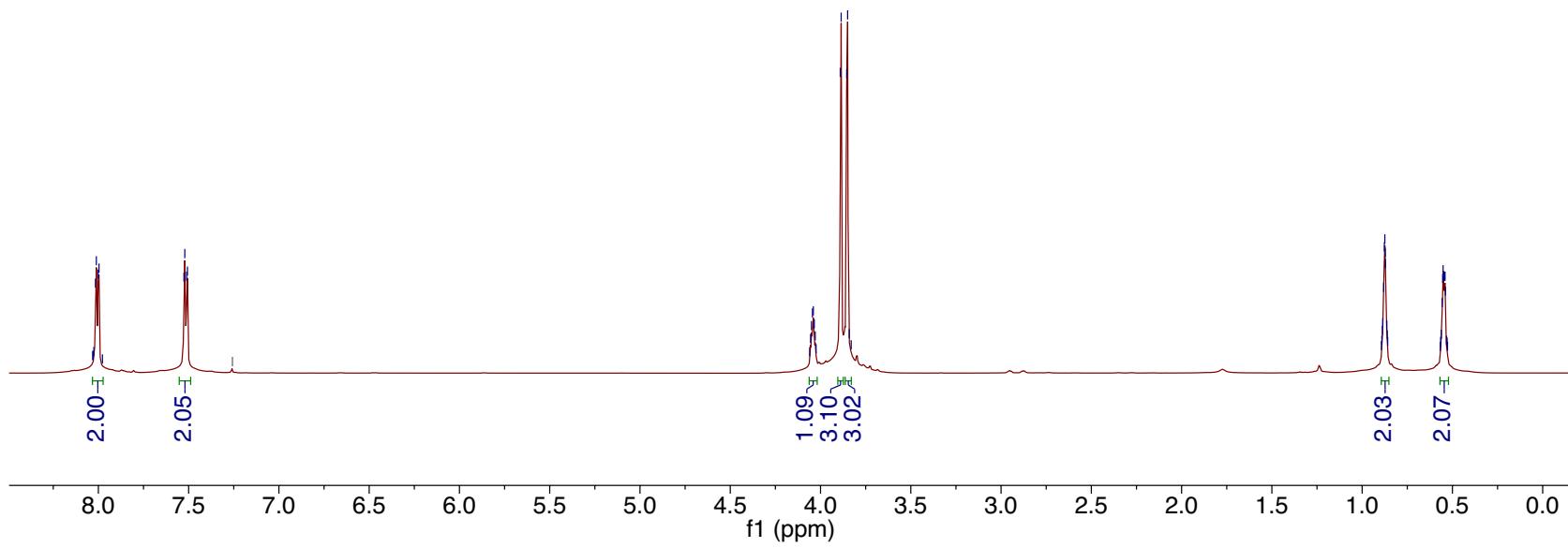
^{13}C NMR (CDCl_3 , 151 MHz)

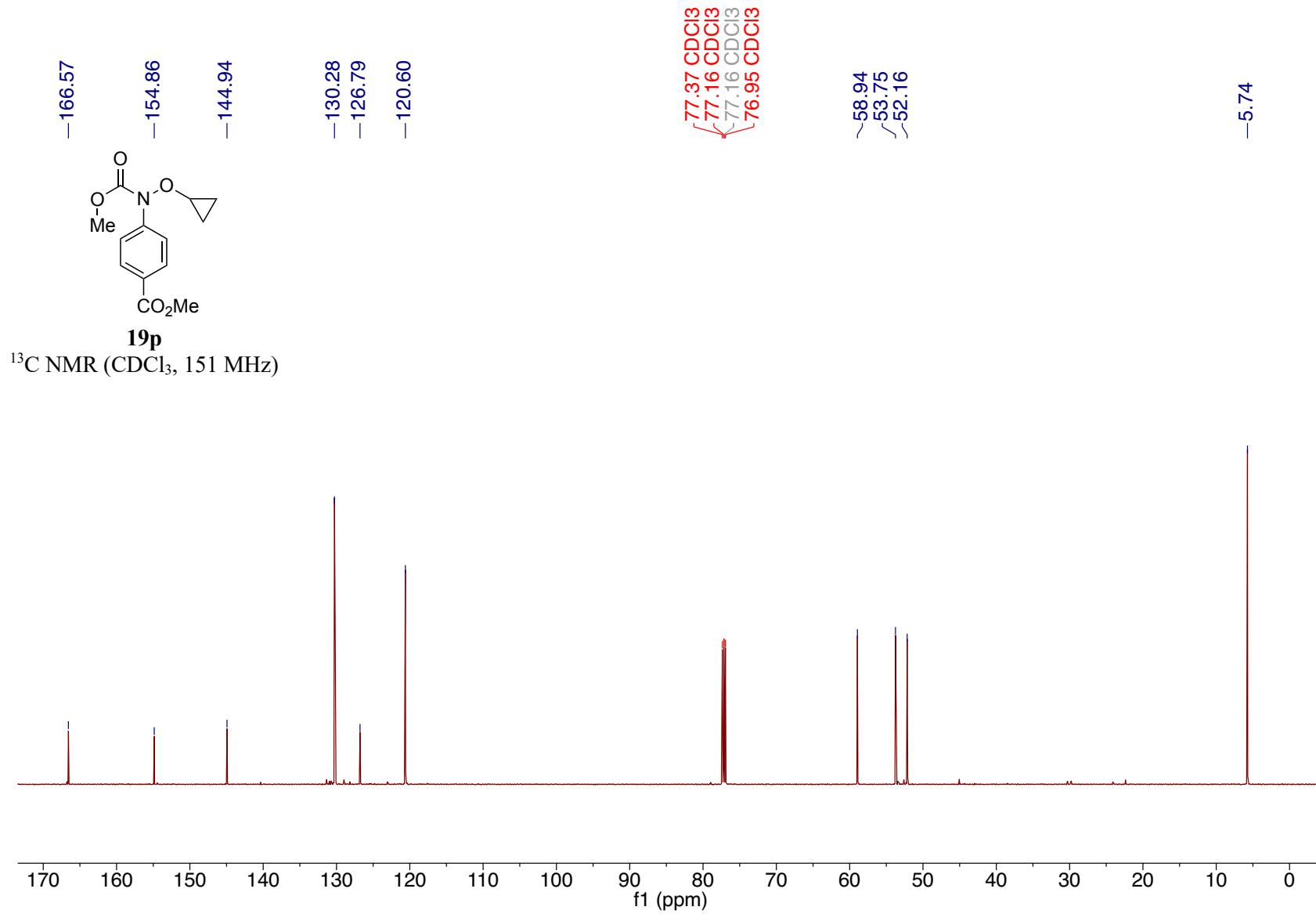


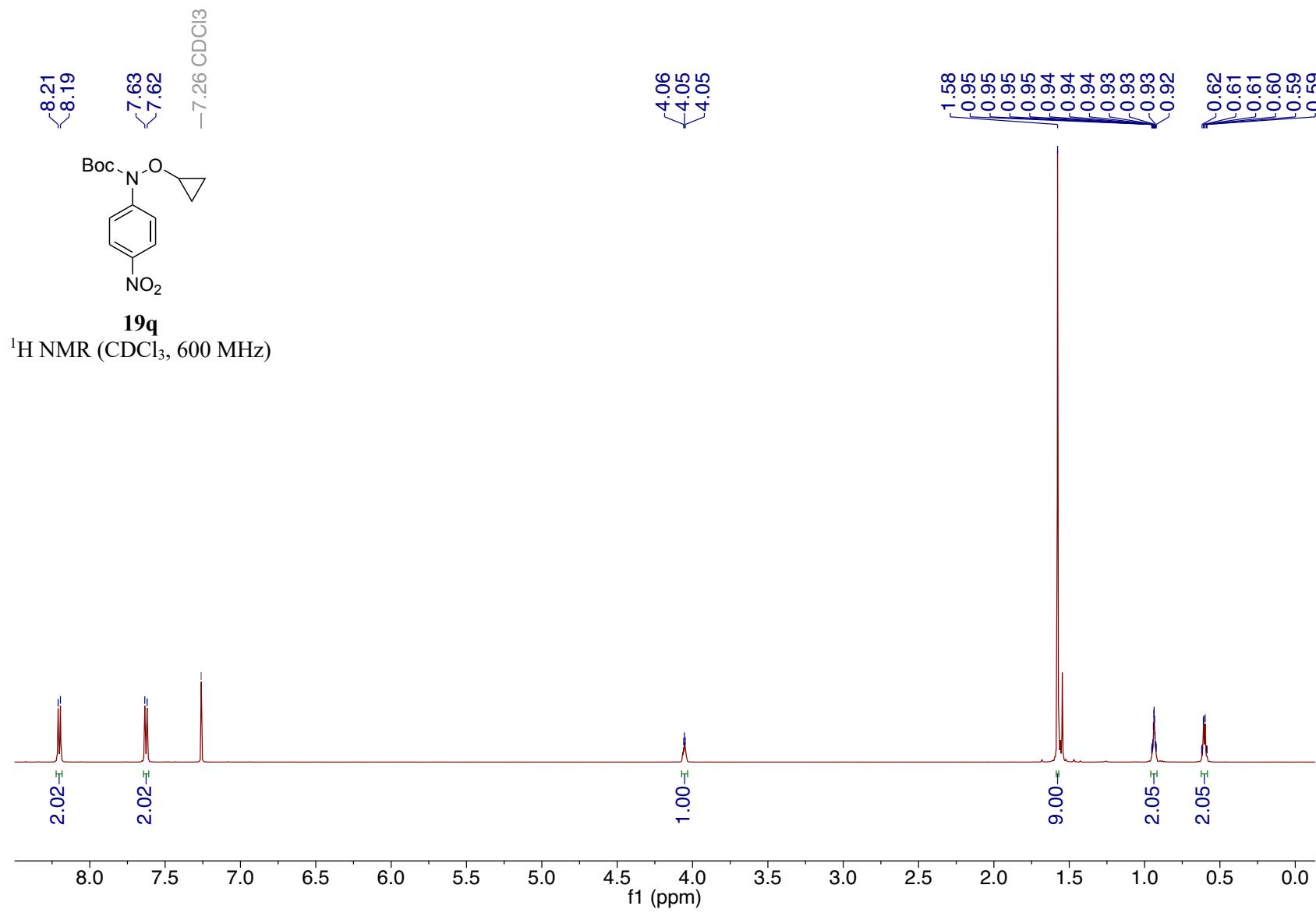


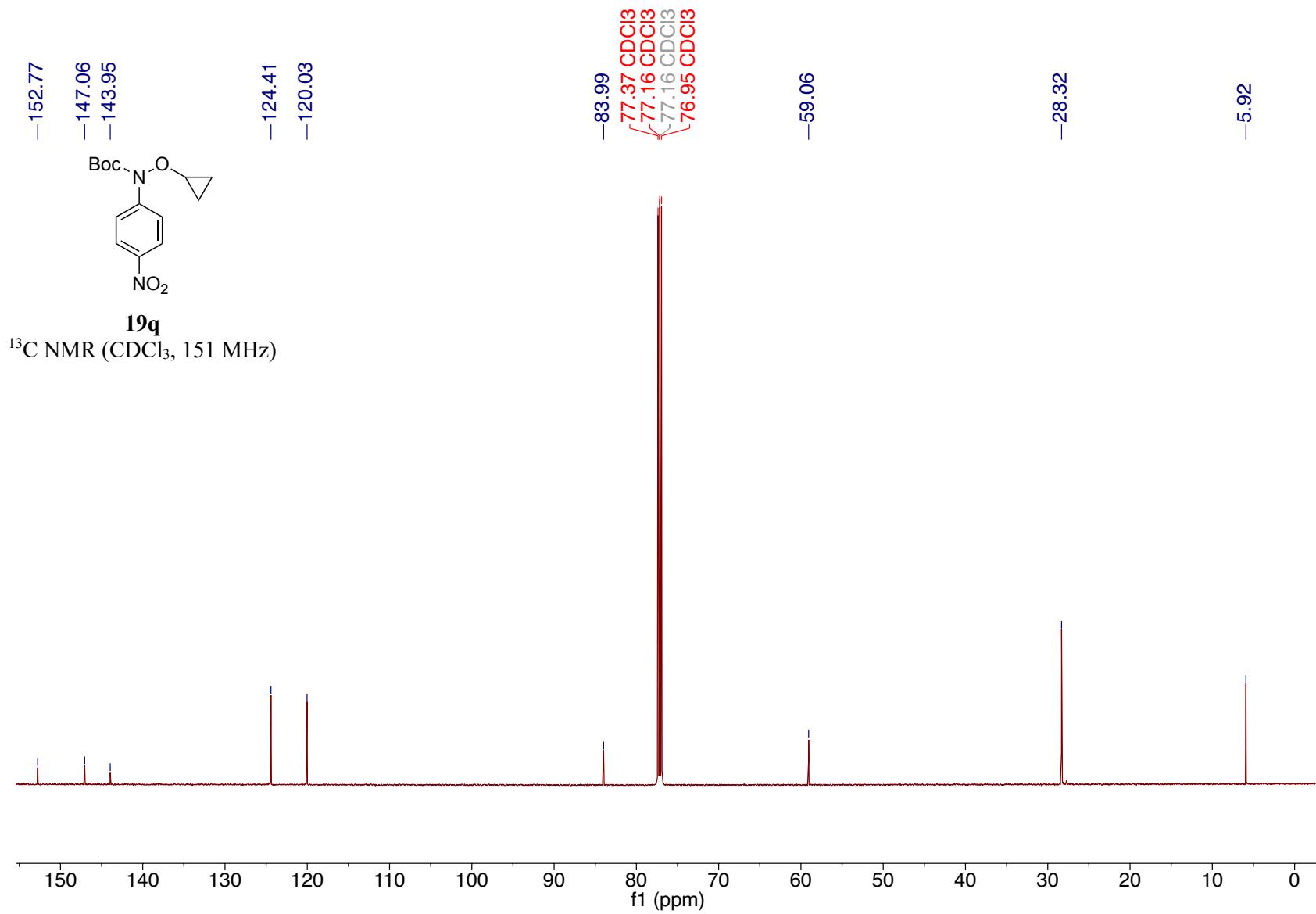
19p

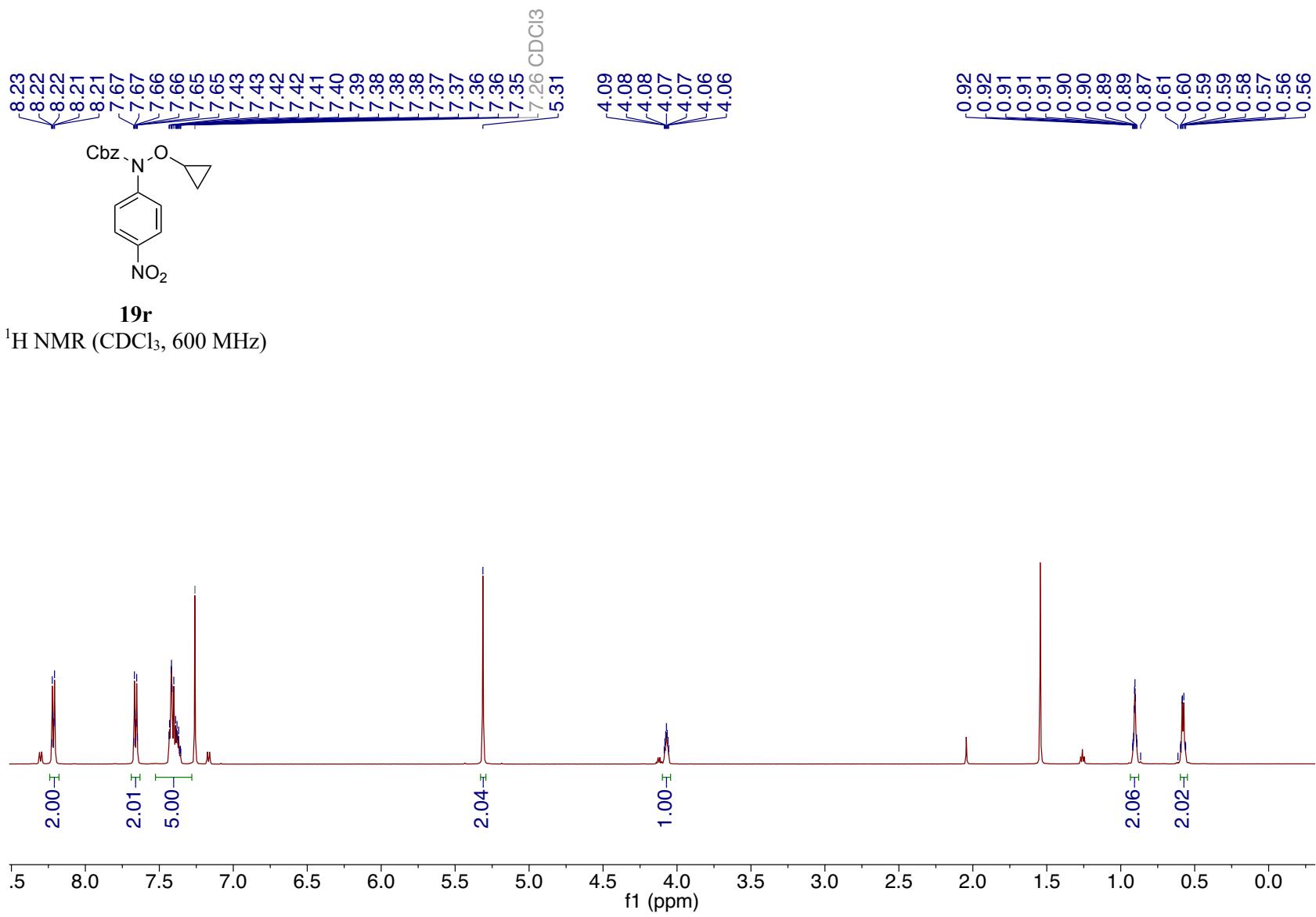
¹H NMR (CDCl₃, 600 MHz)

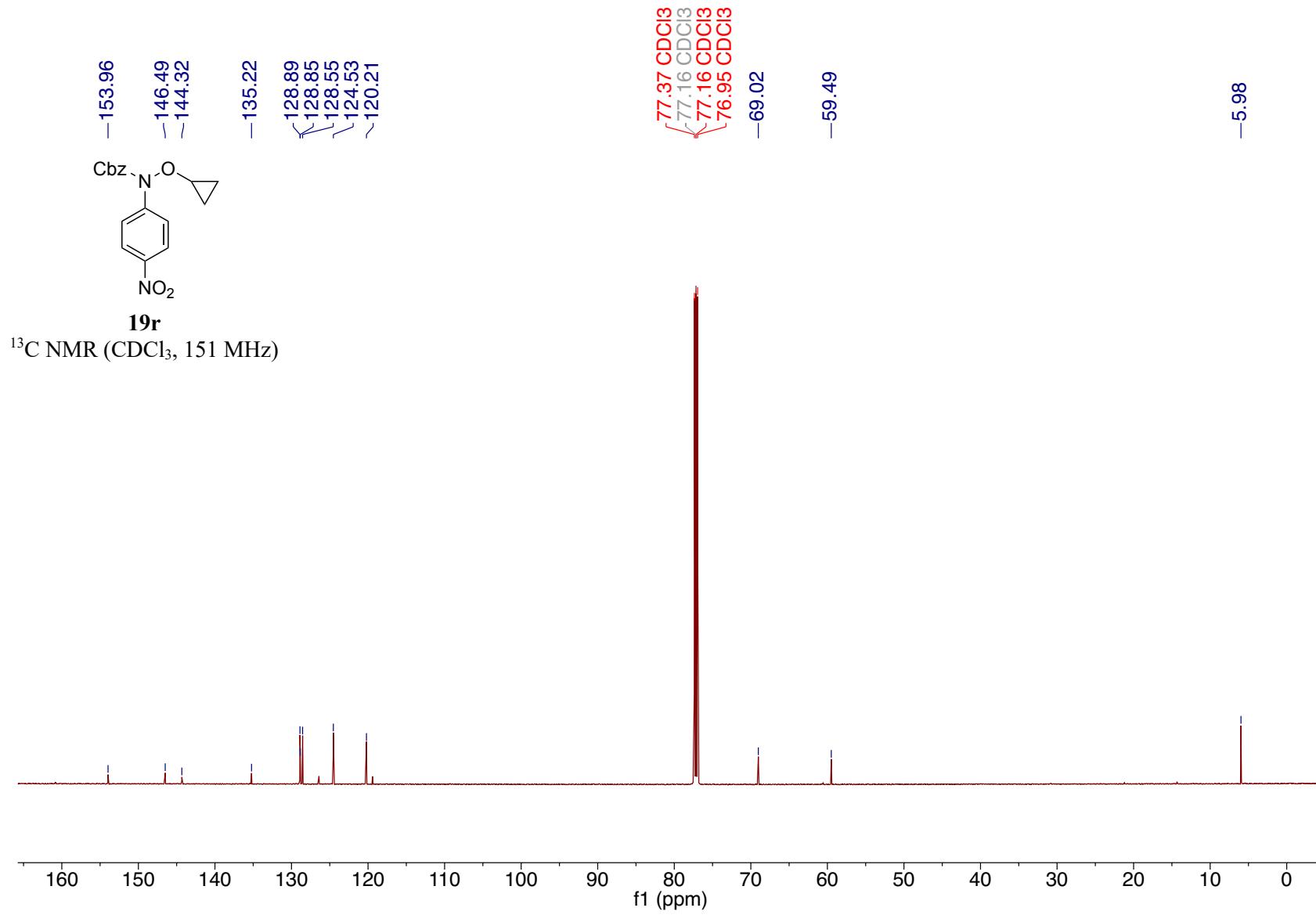


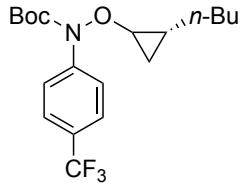




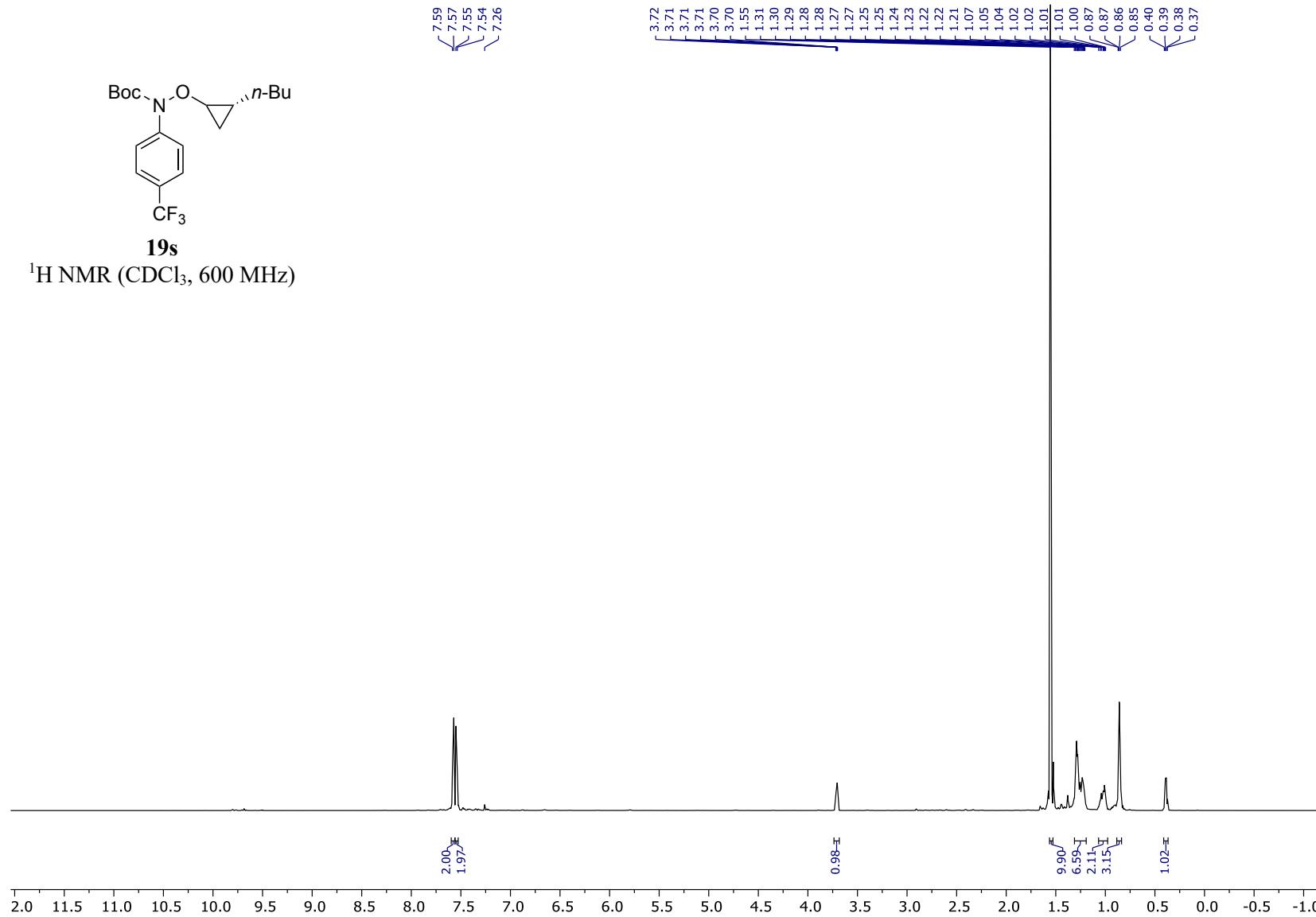


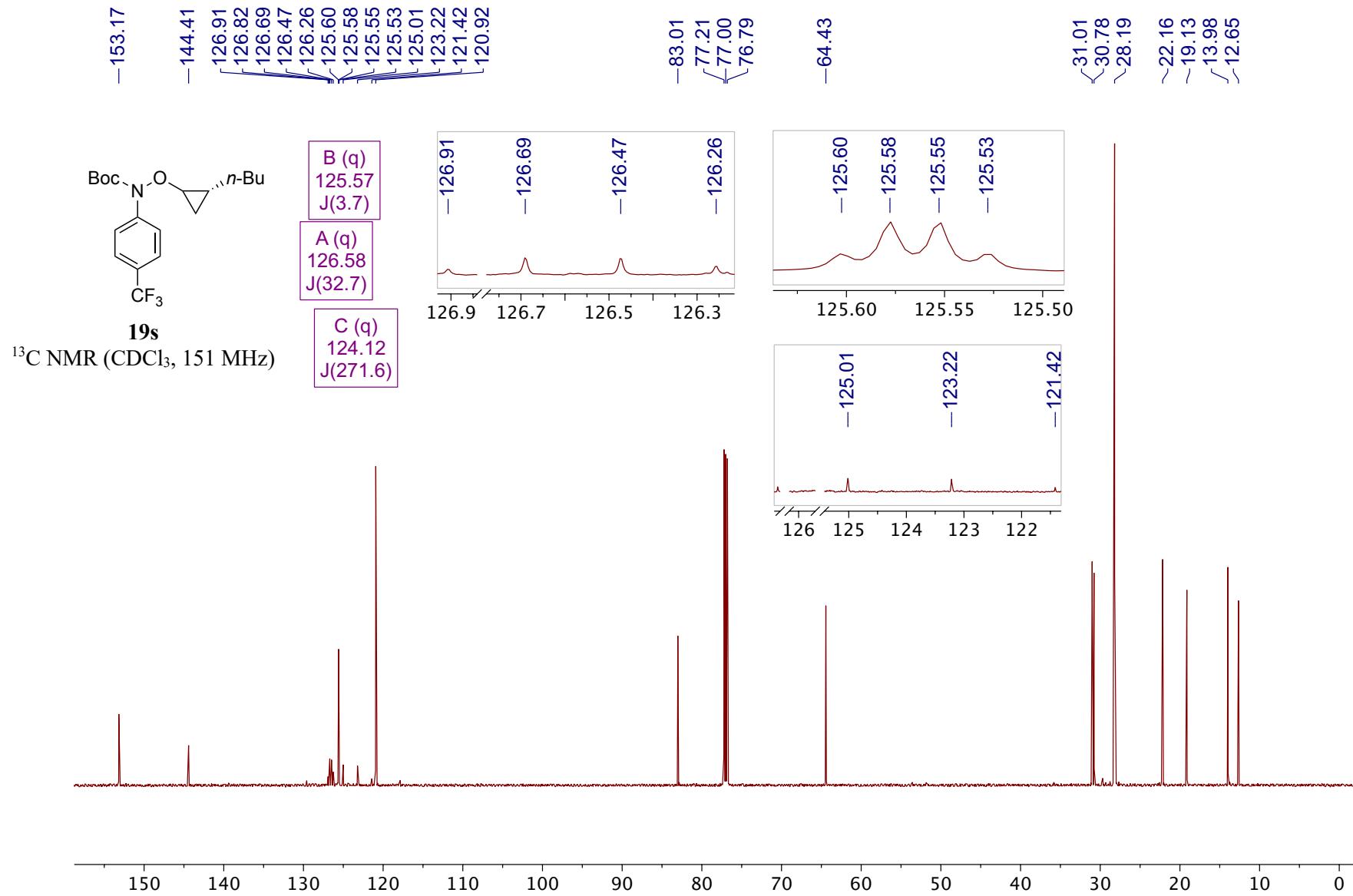


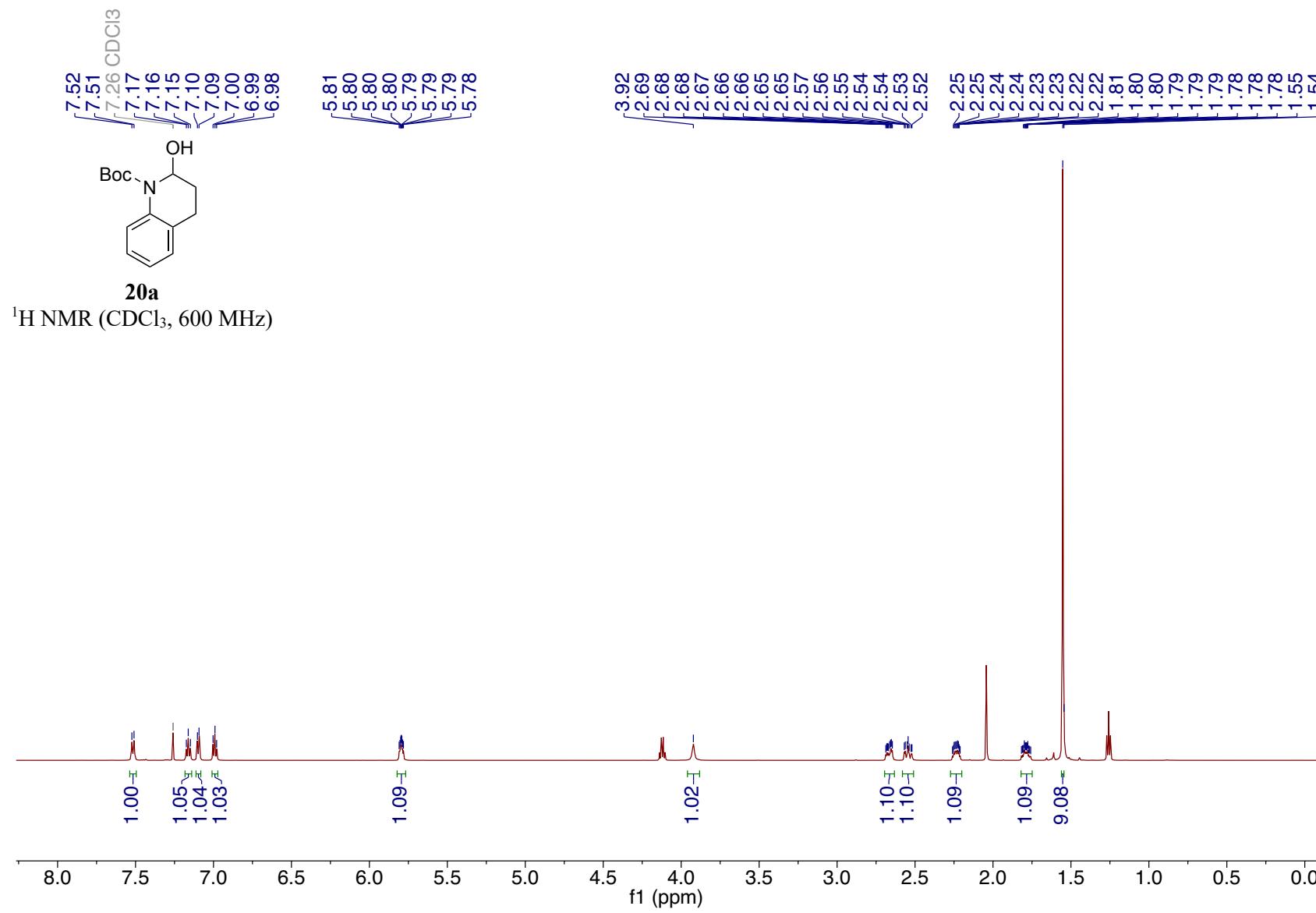


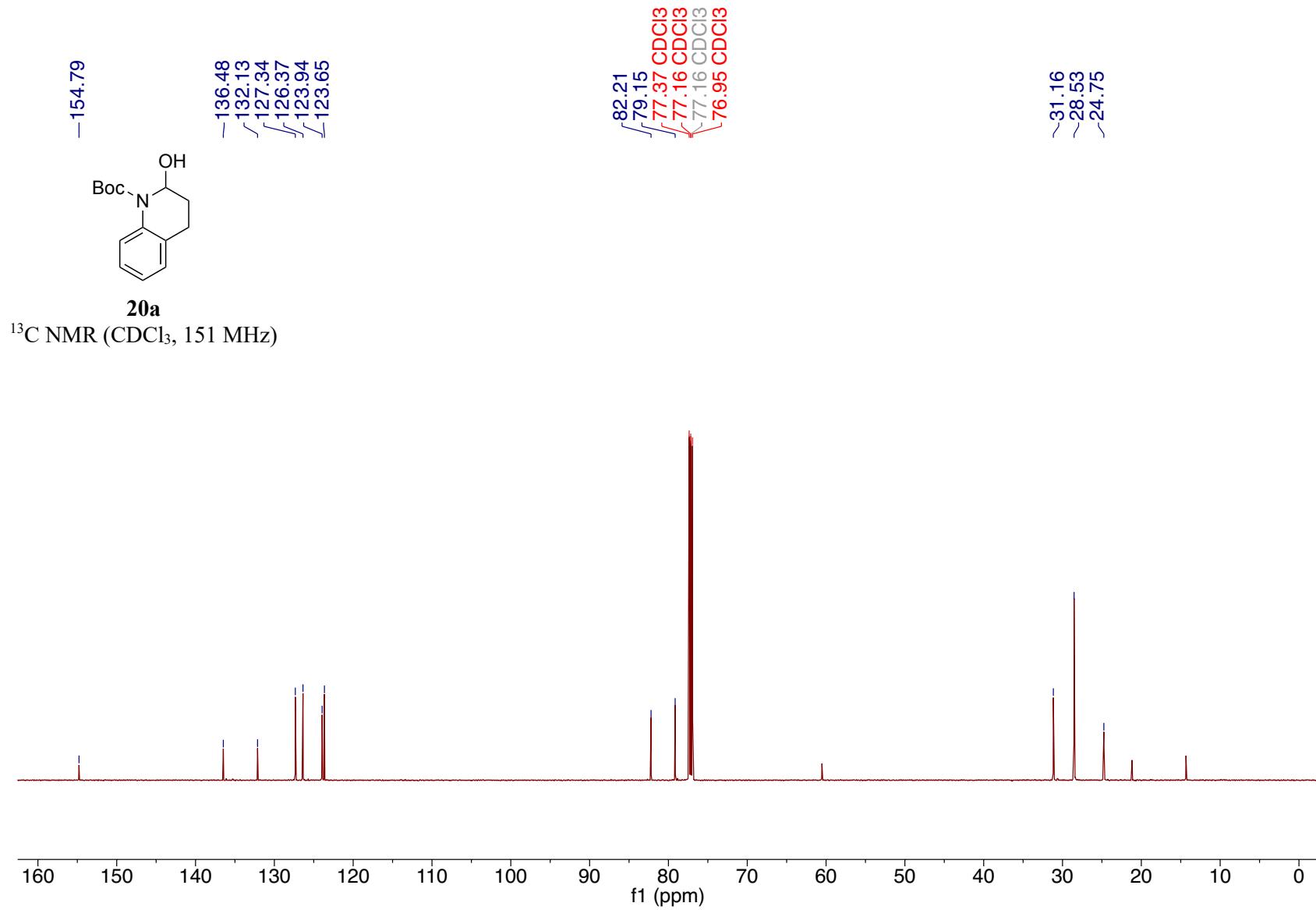


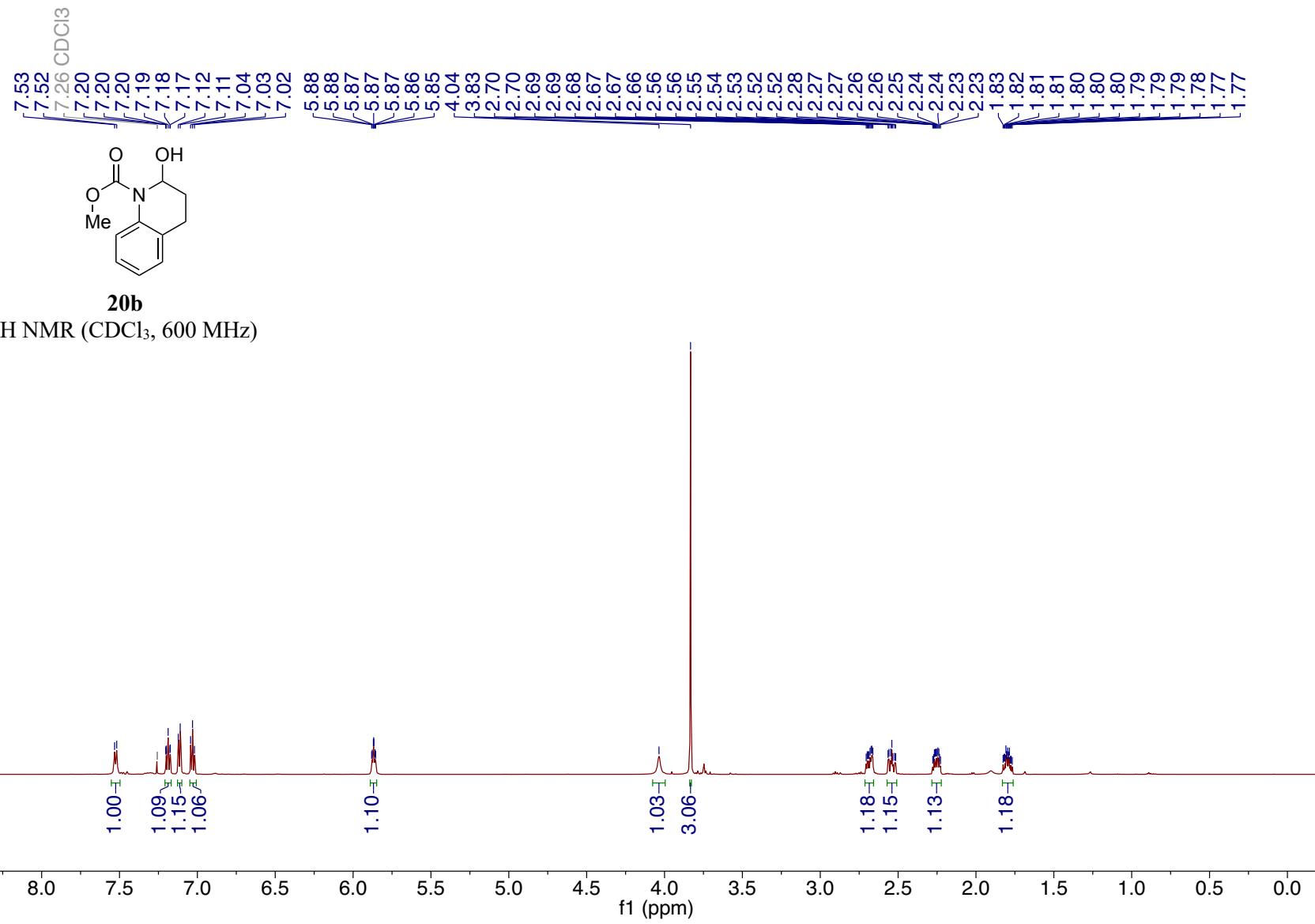
¹H NMR (CDCl₃, 600 MHz)

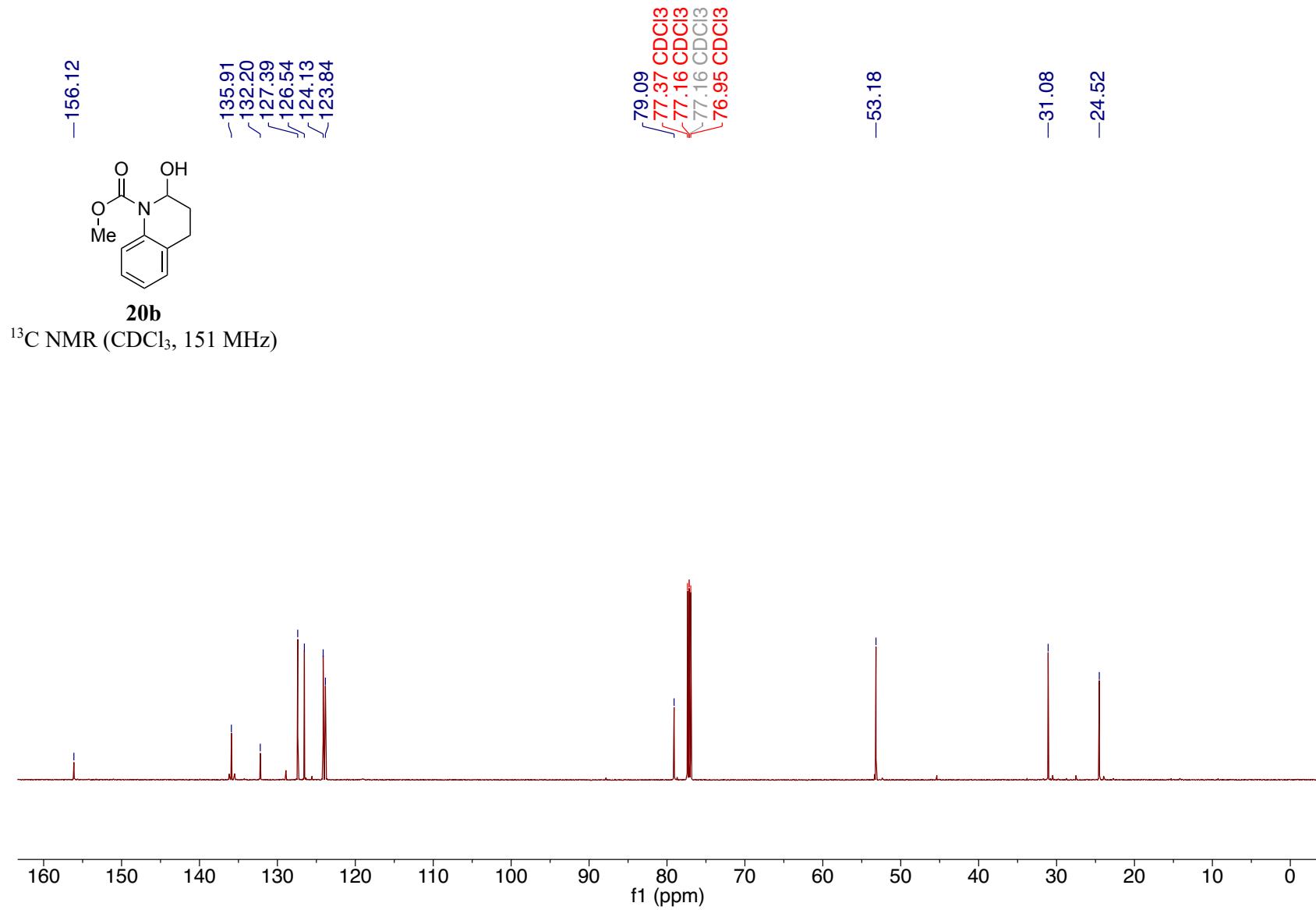


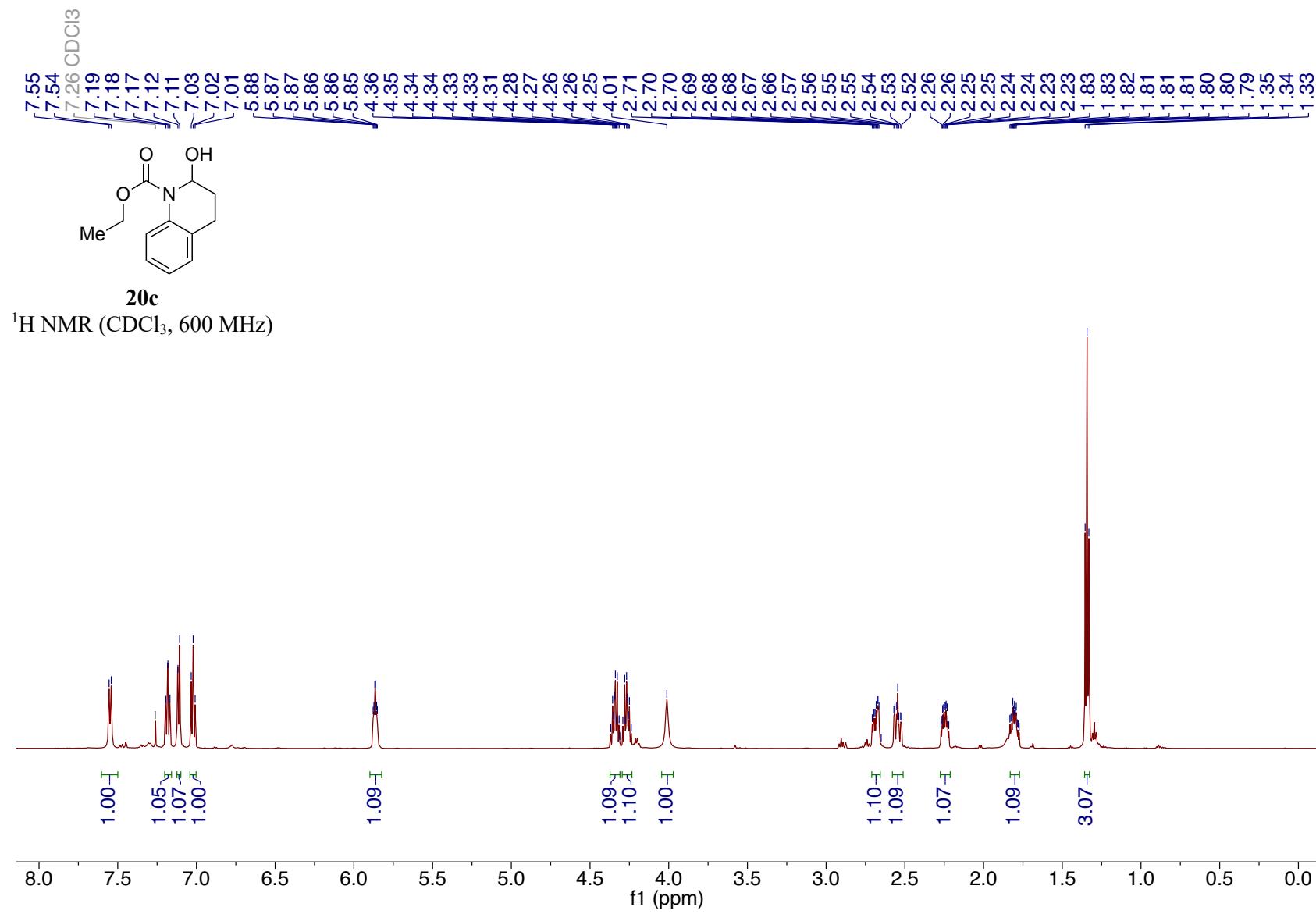


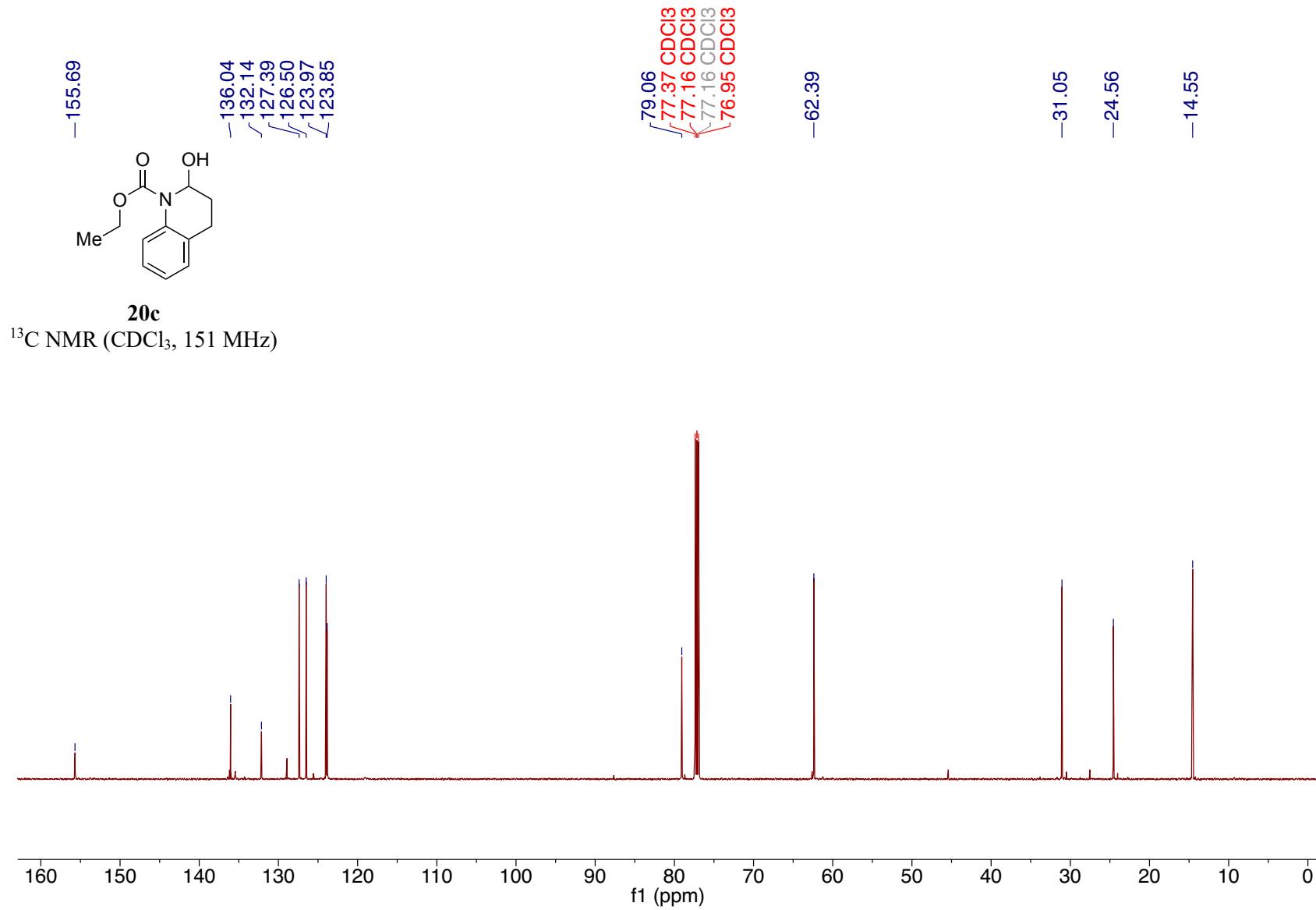


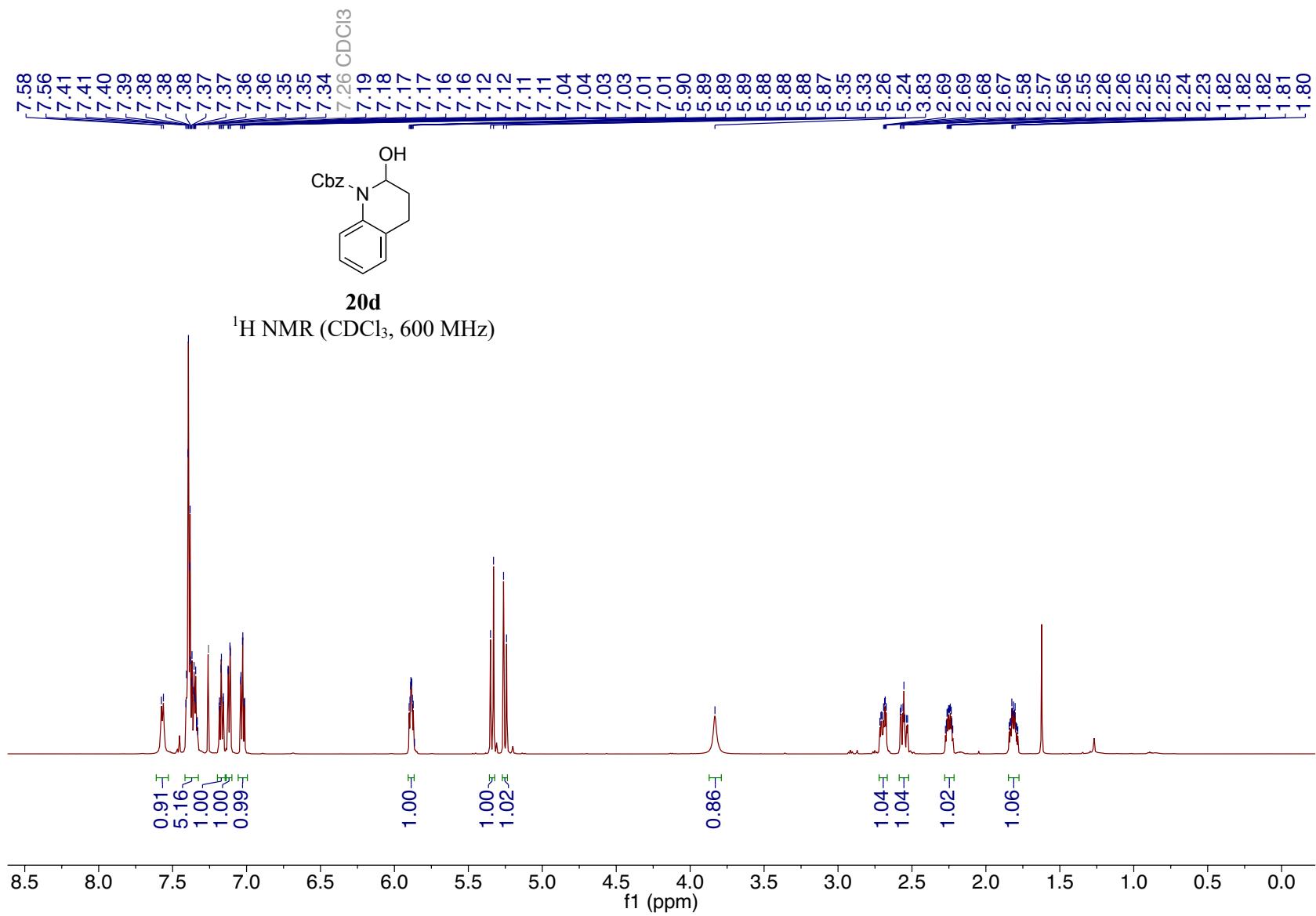


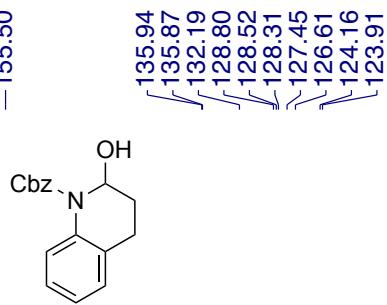






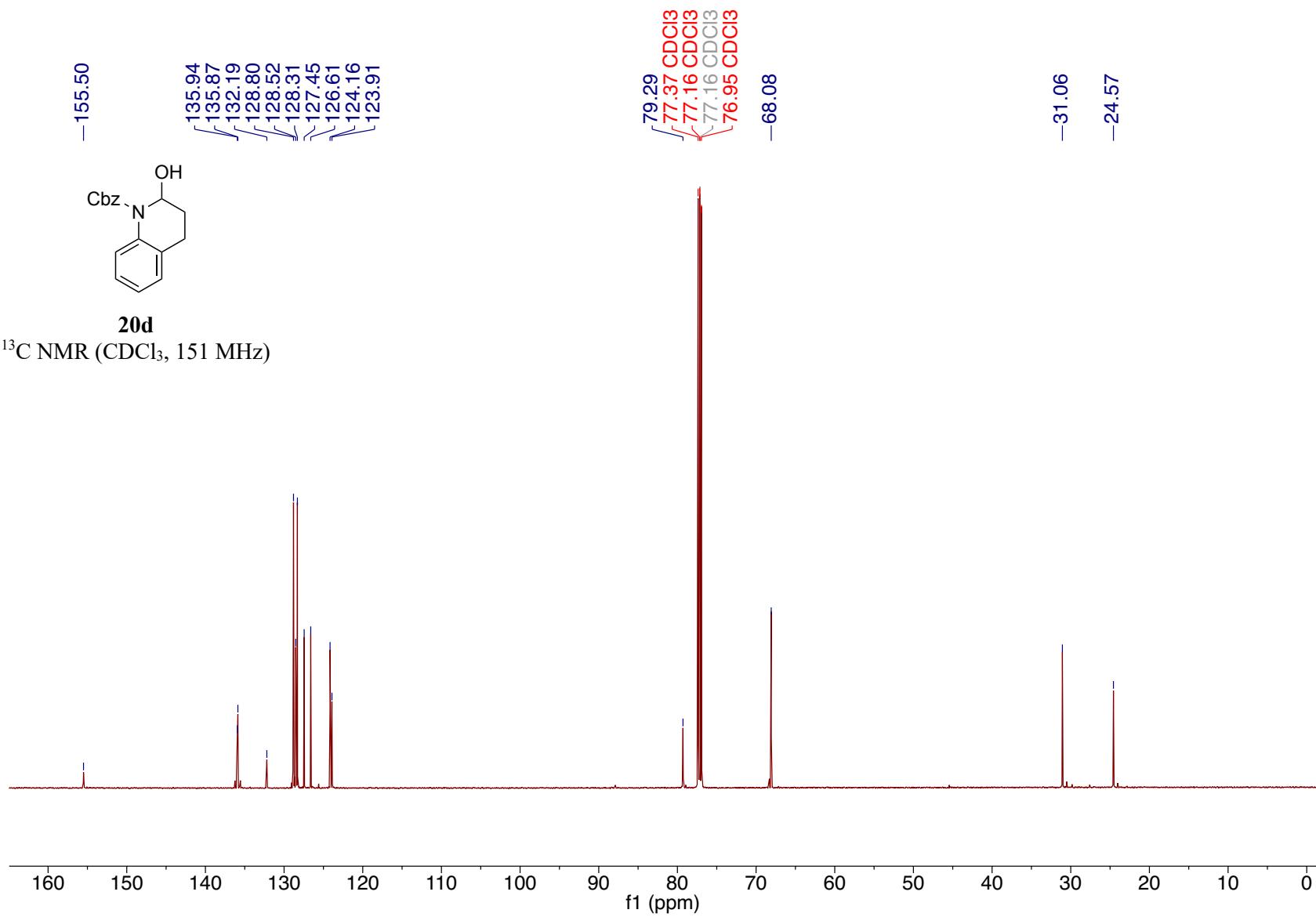


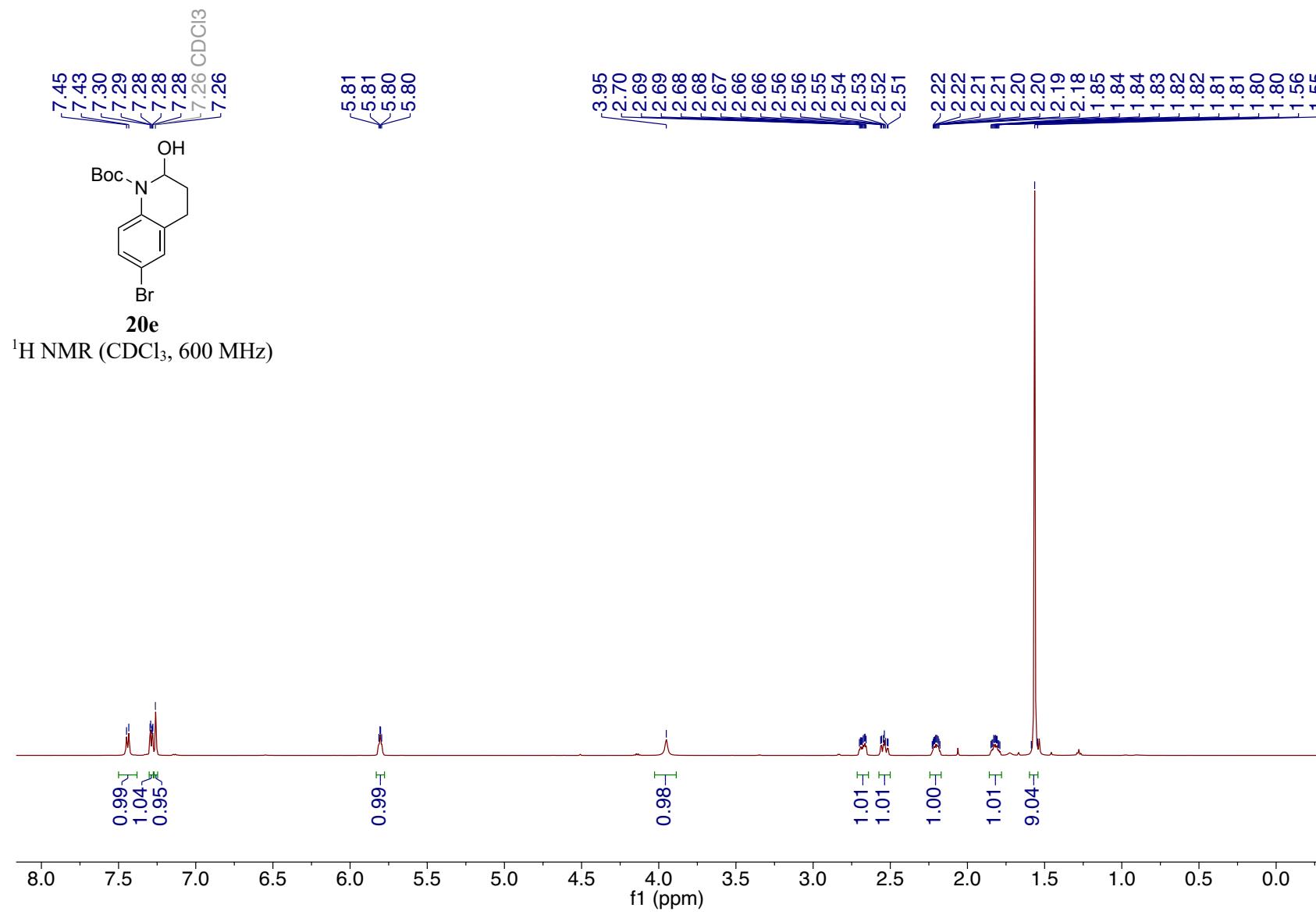


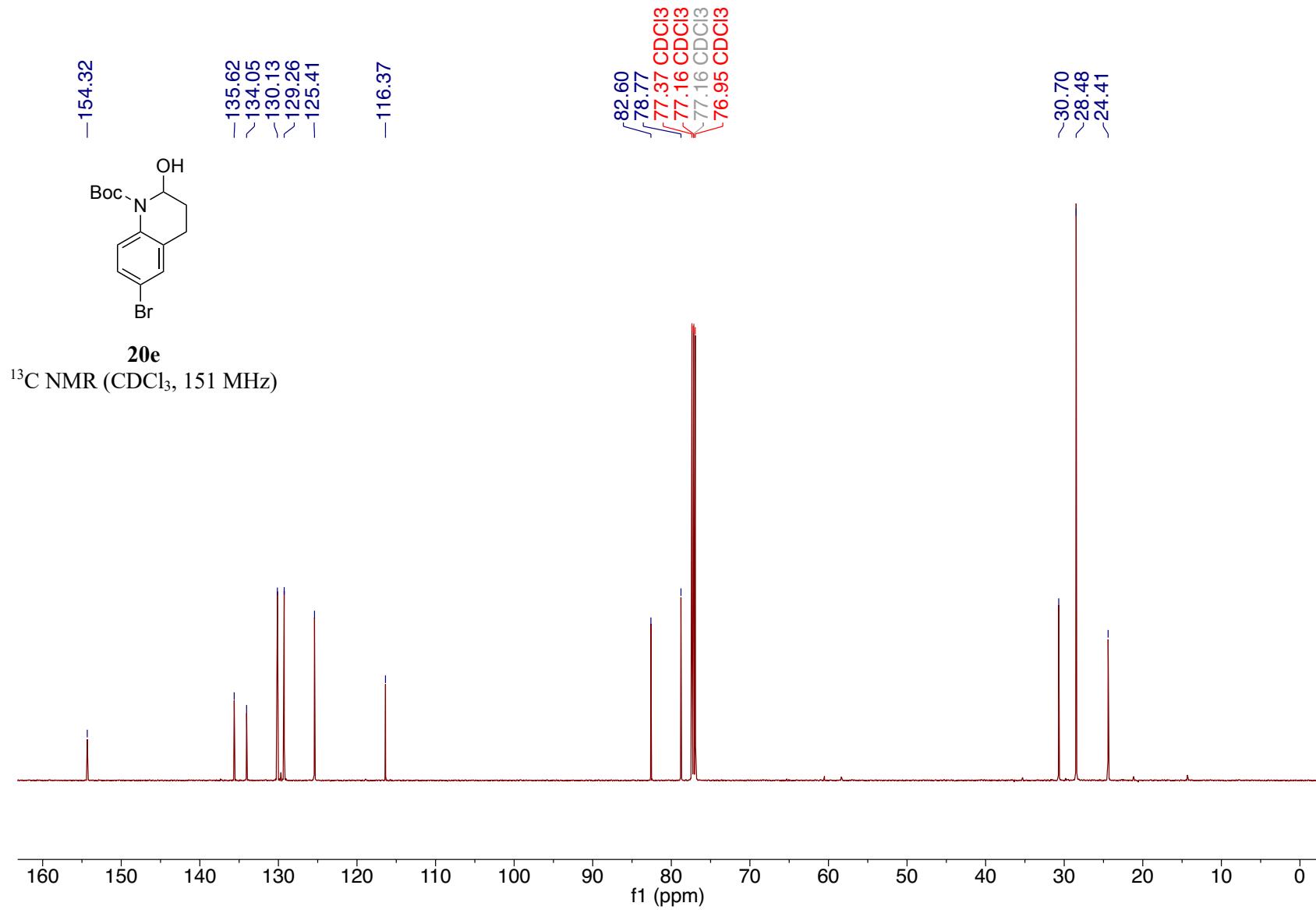


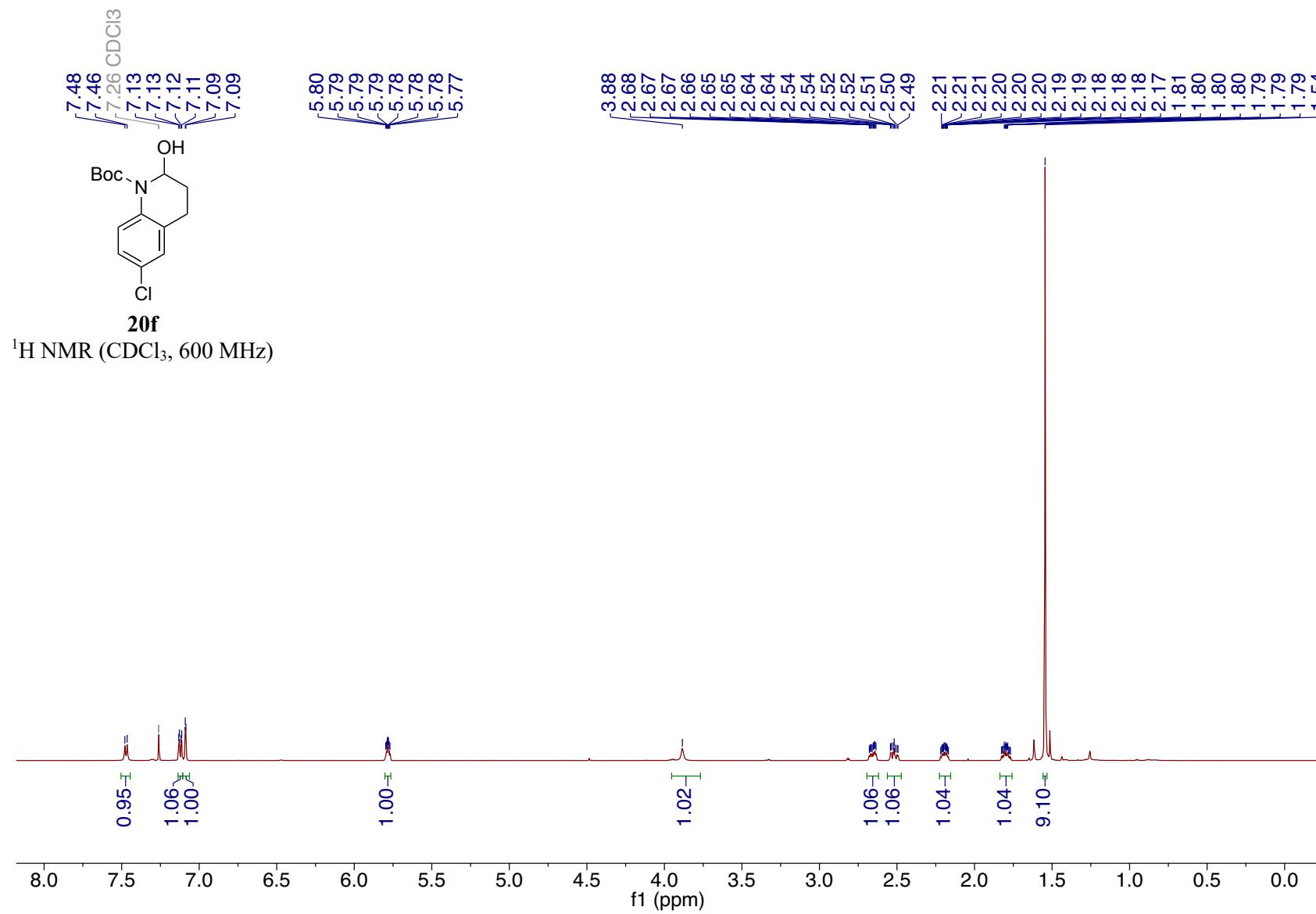
20d

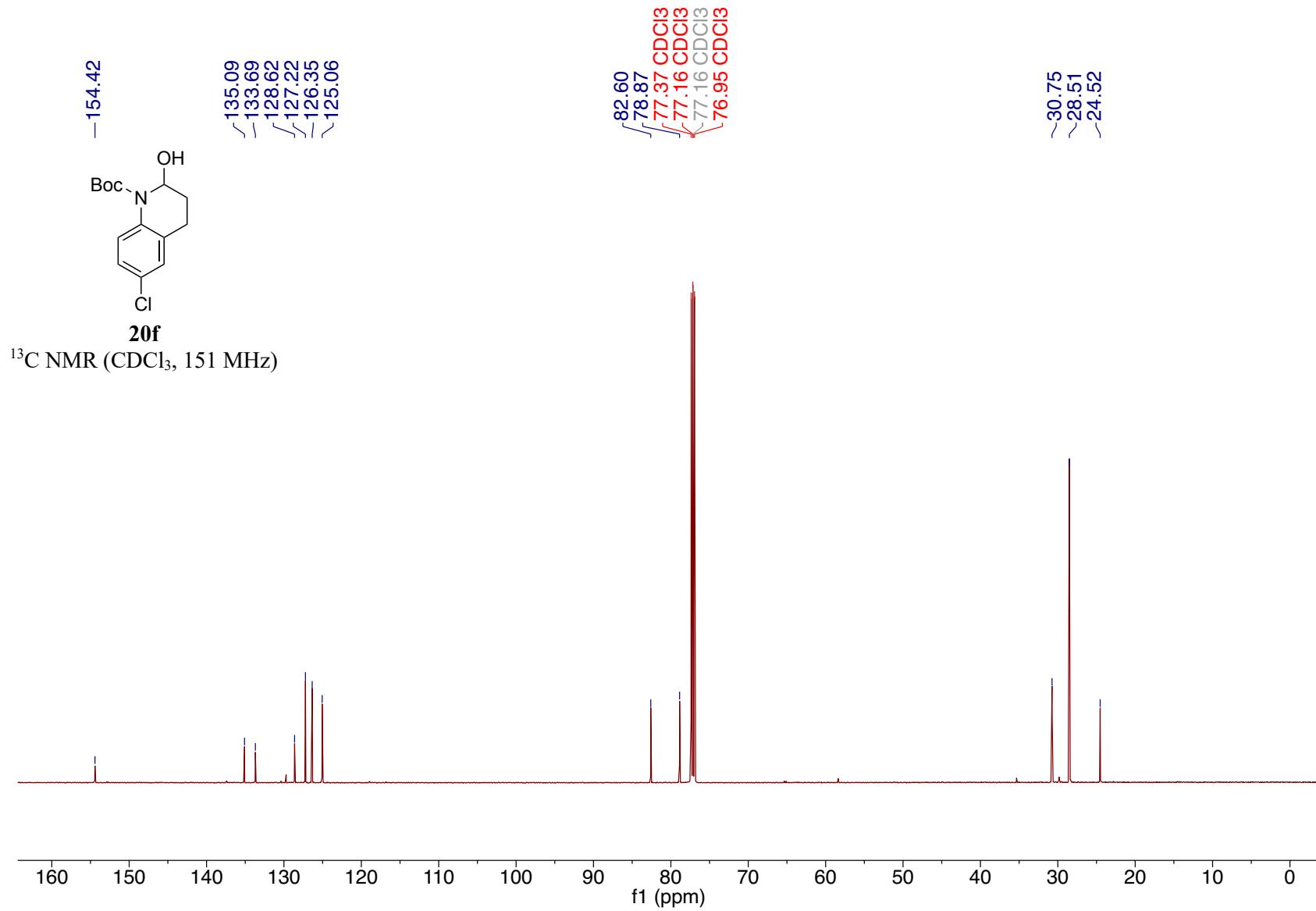
¹³C NMR (CDCl₃, 151 MHz)

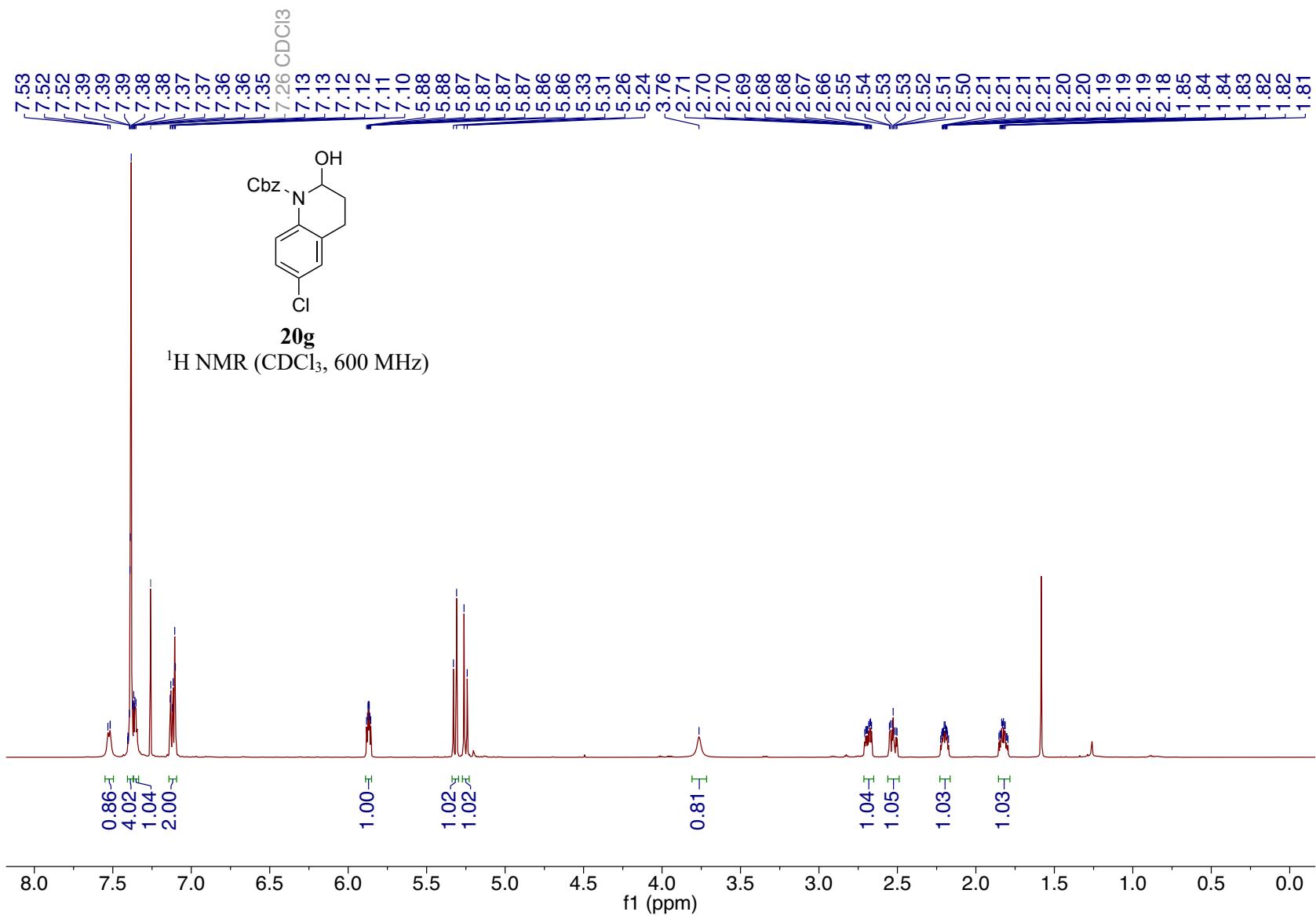


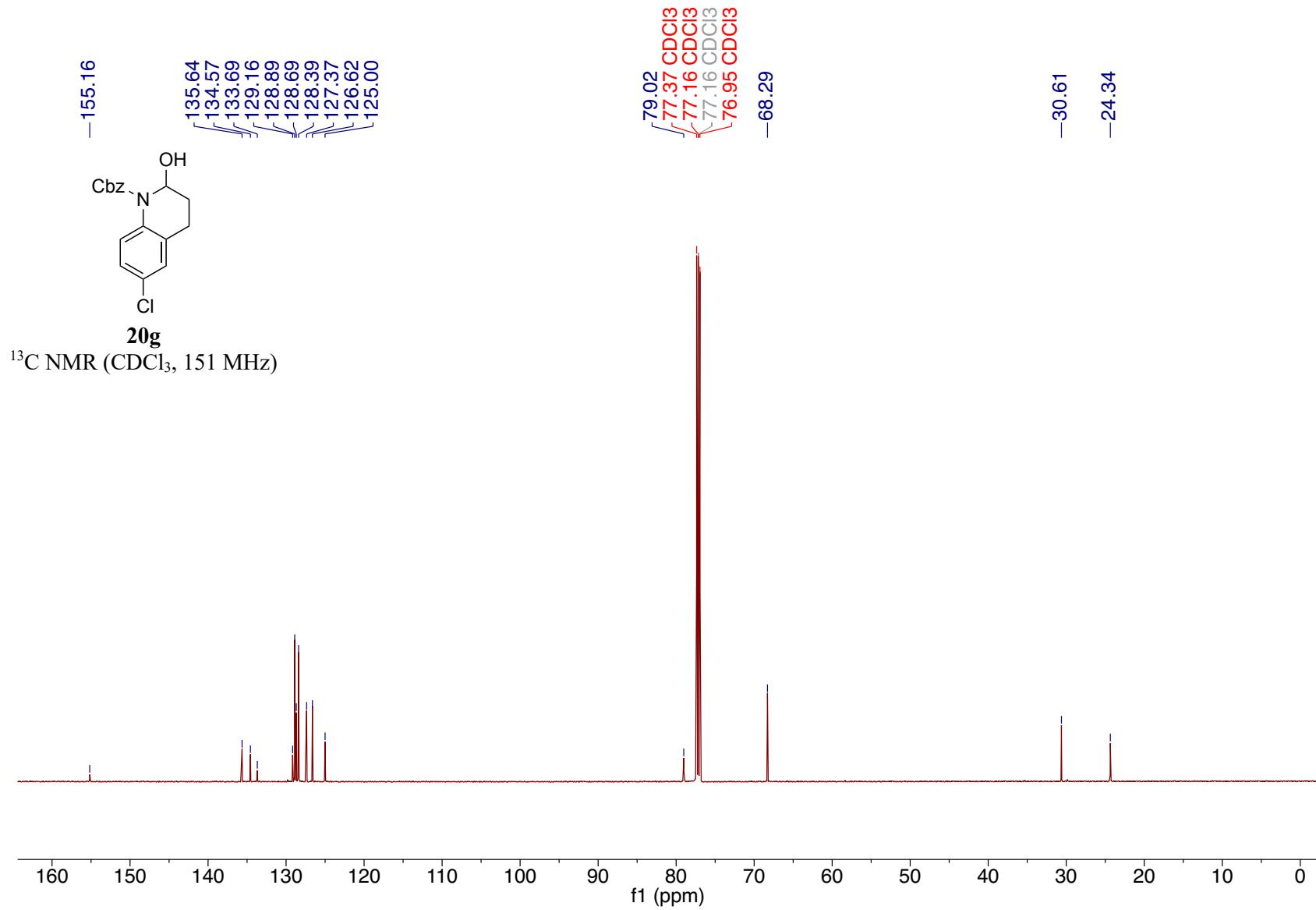


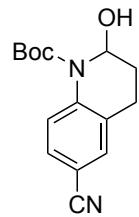






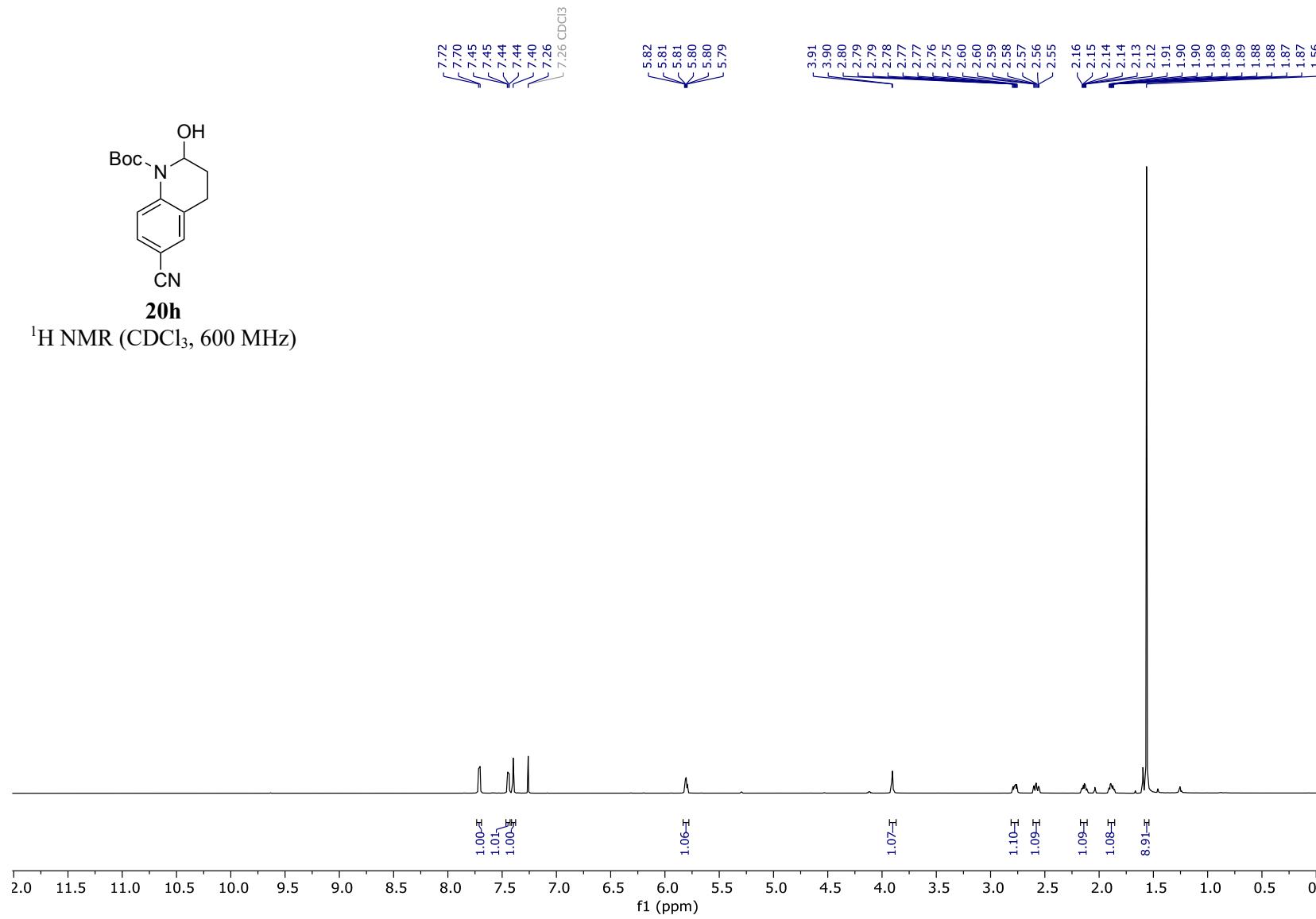


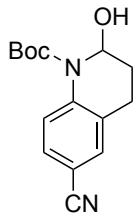




20h

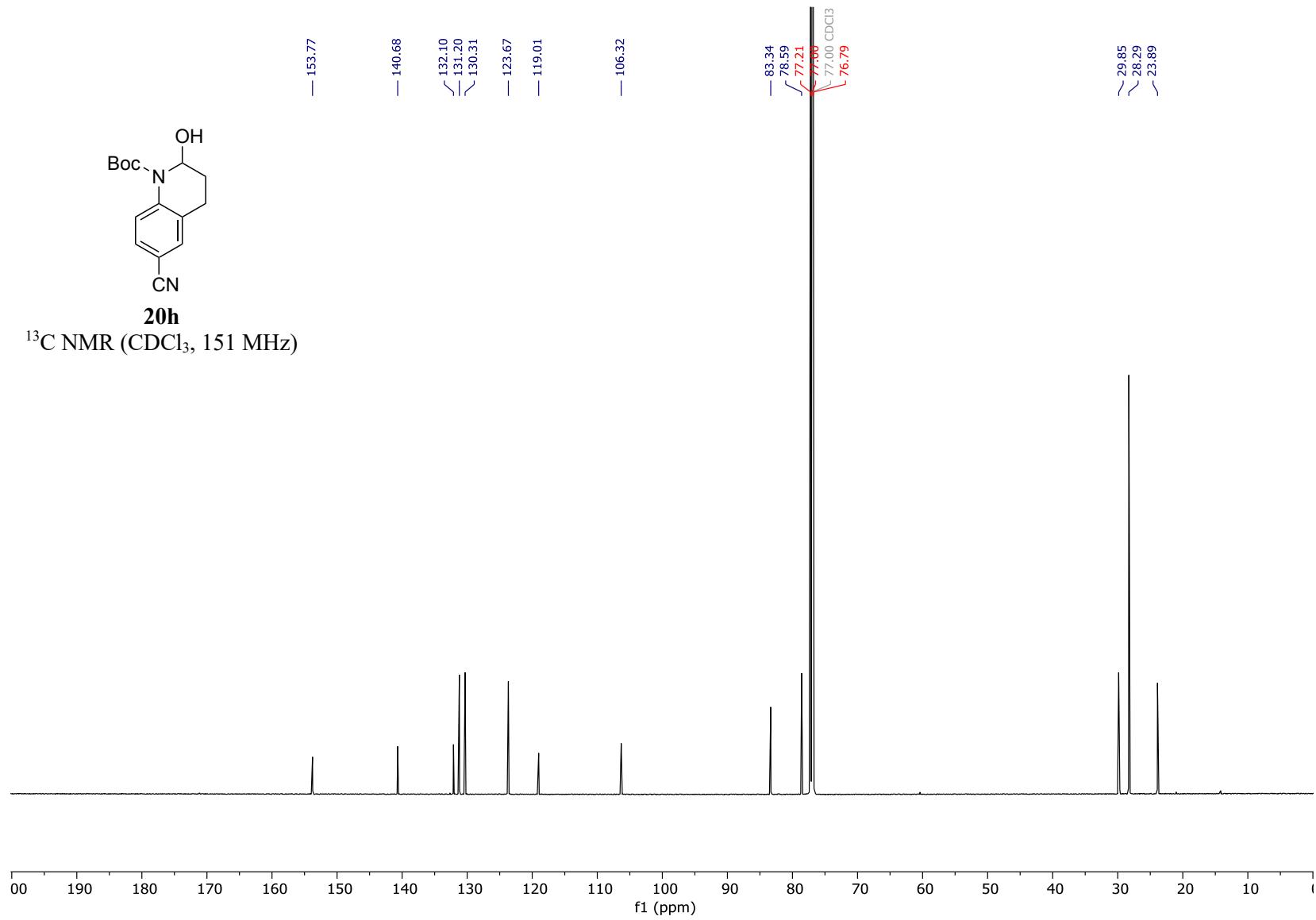
¹H NMR (CDCl₃, 600 MHz)

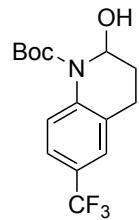




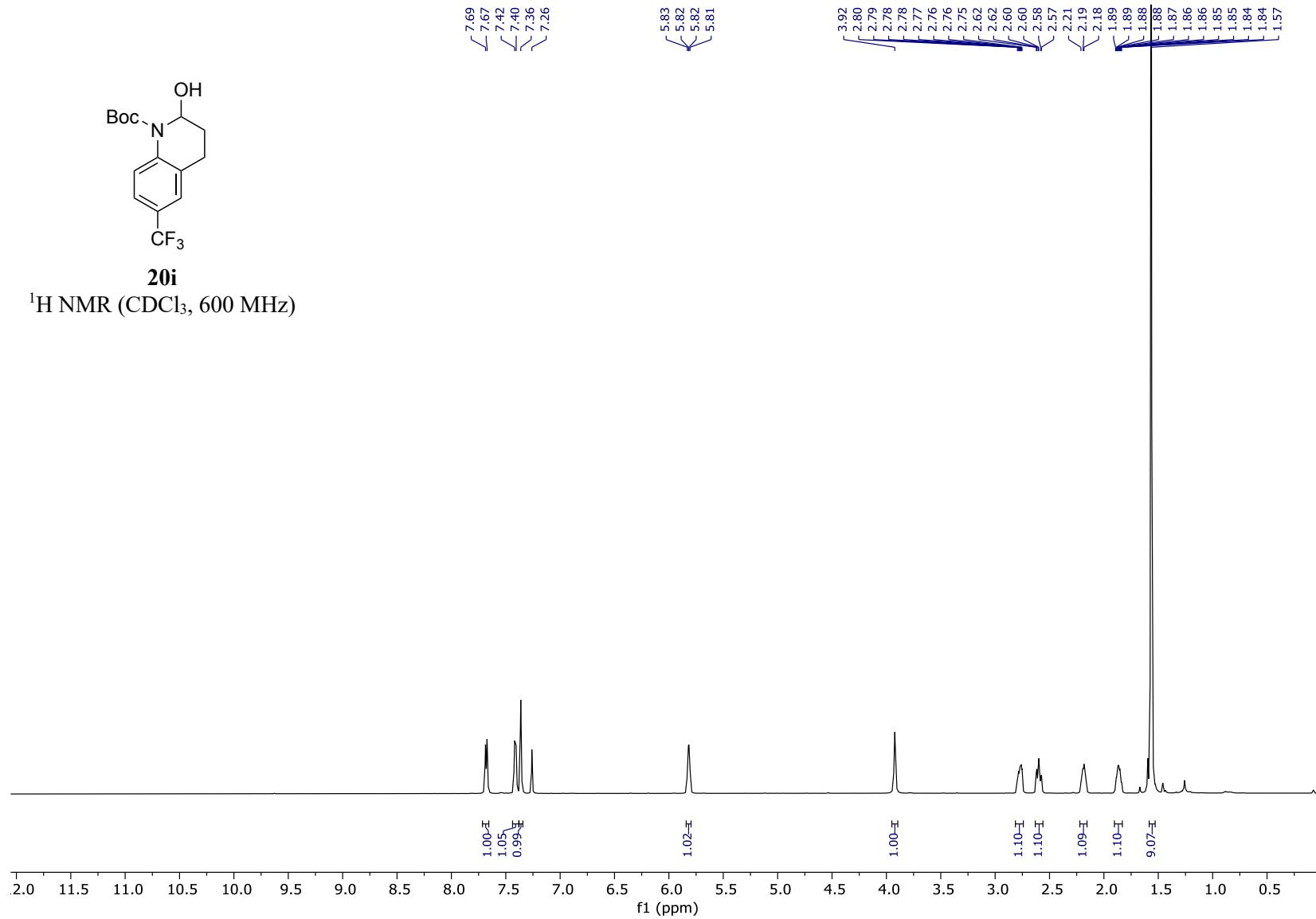
20h

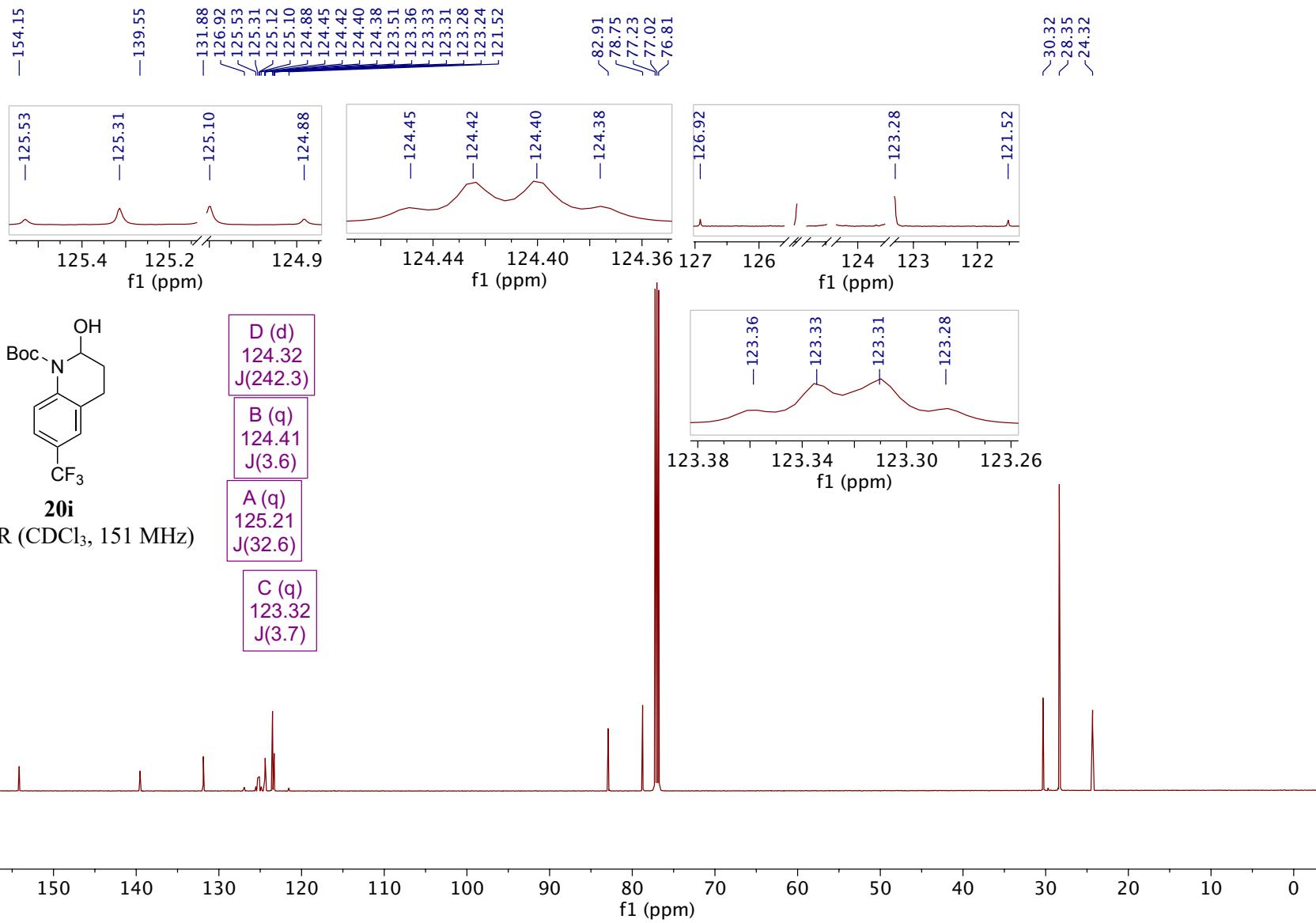
^{13}C NMR (CDCl_3 , 151 MHz)

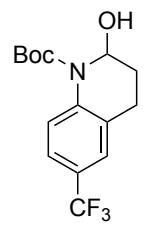




¹H NMR (CDCl₃, 600 MHz)

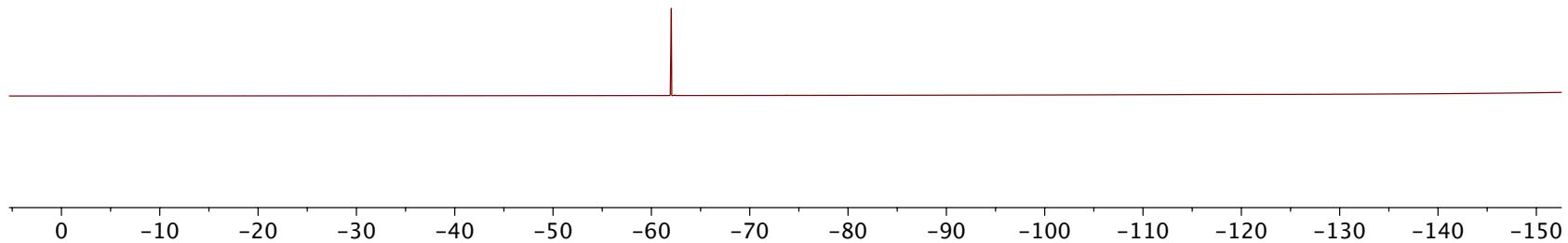


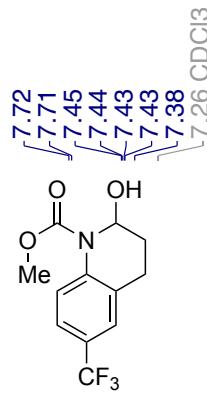




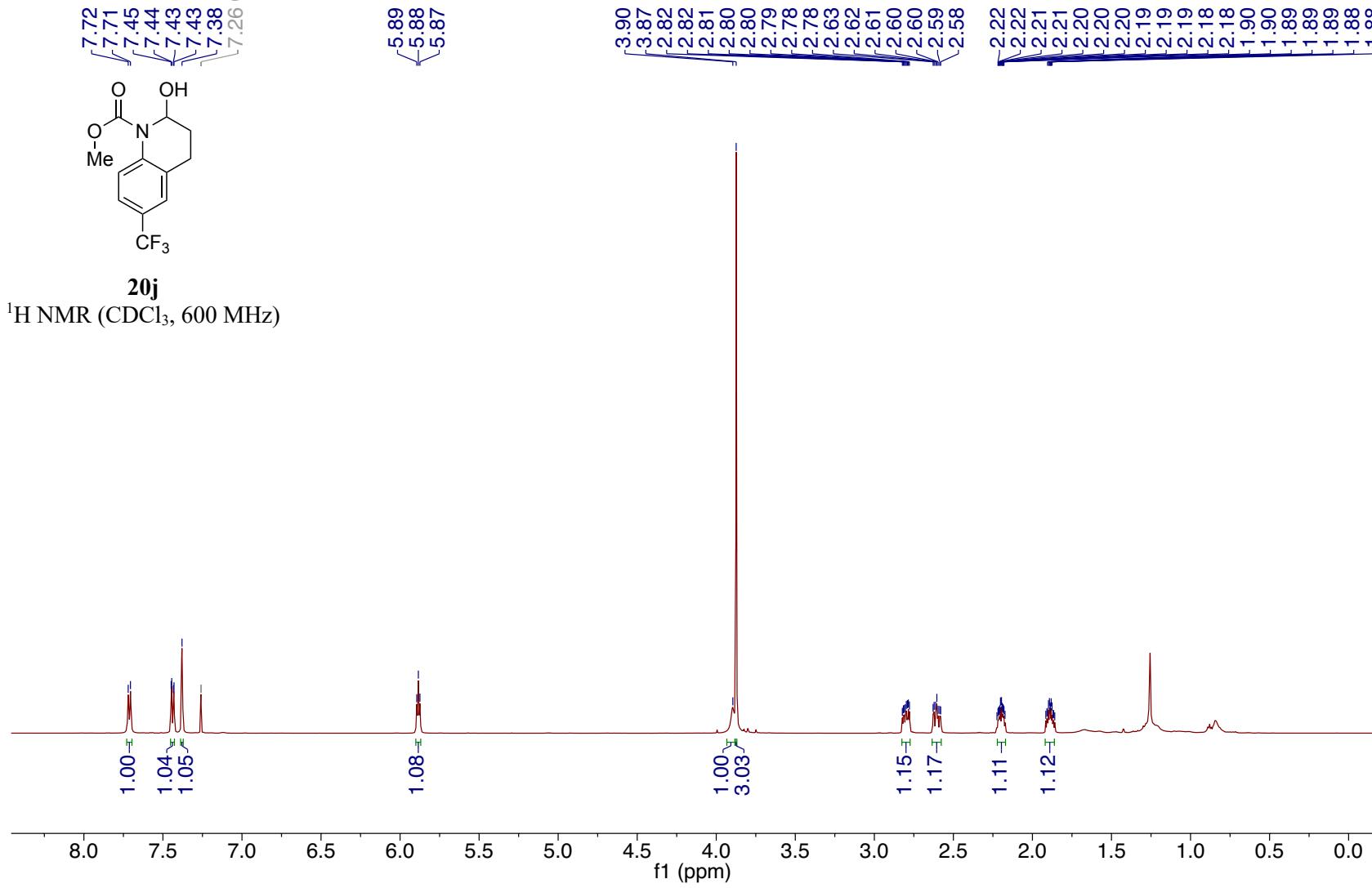
20i

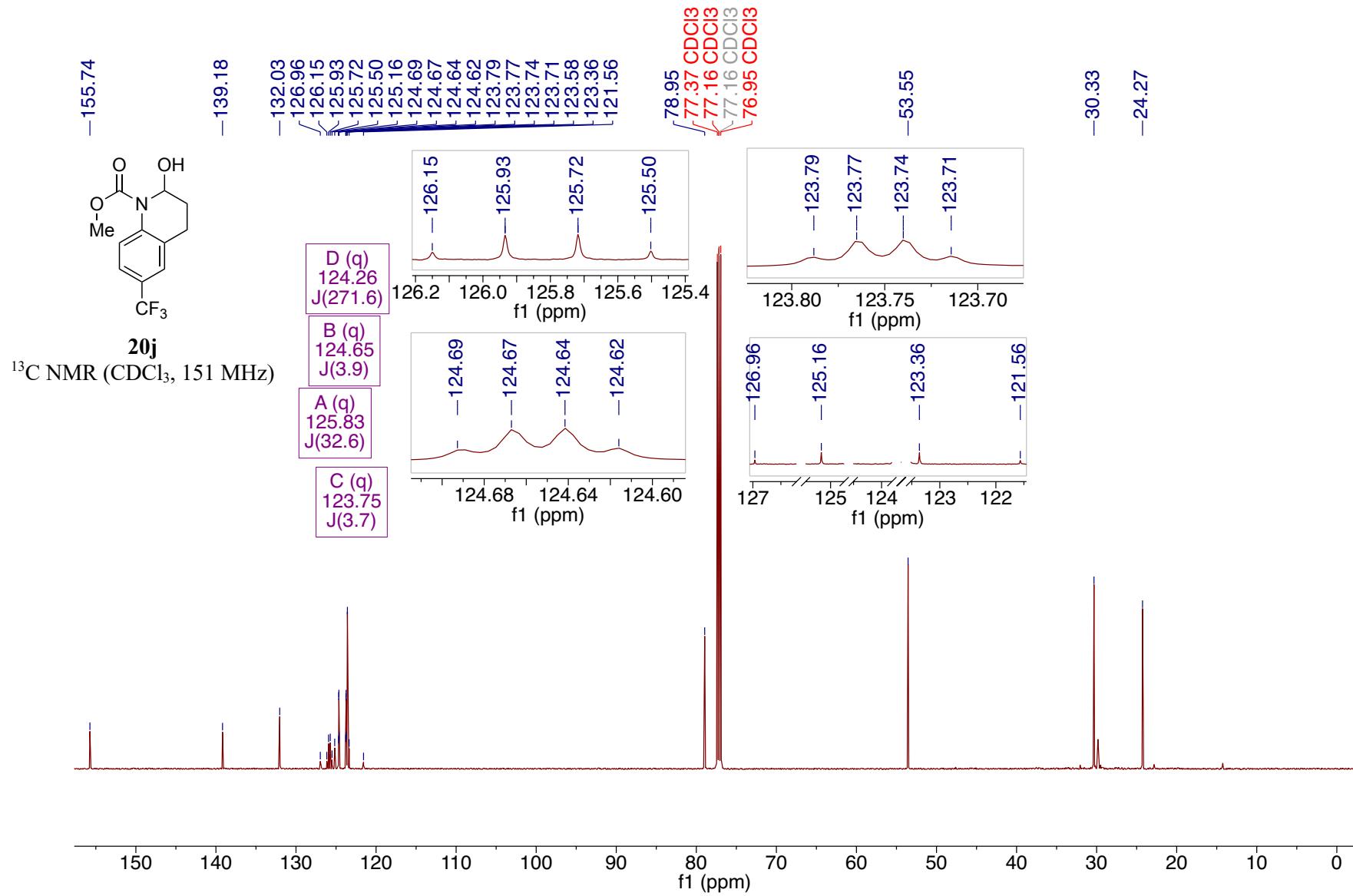
¹⁹F NMR (CDCl₃, 471 MHz)

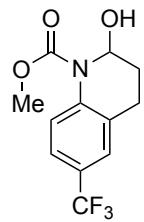




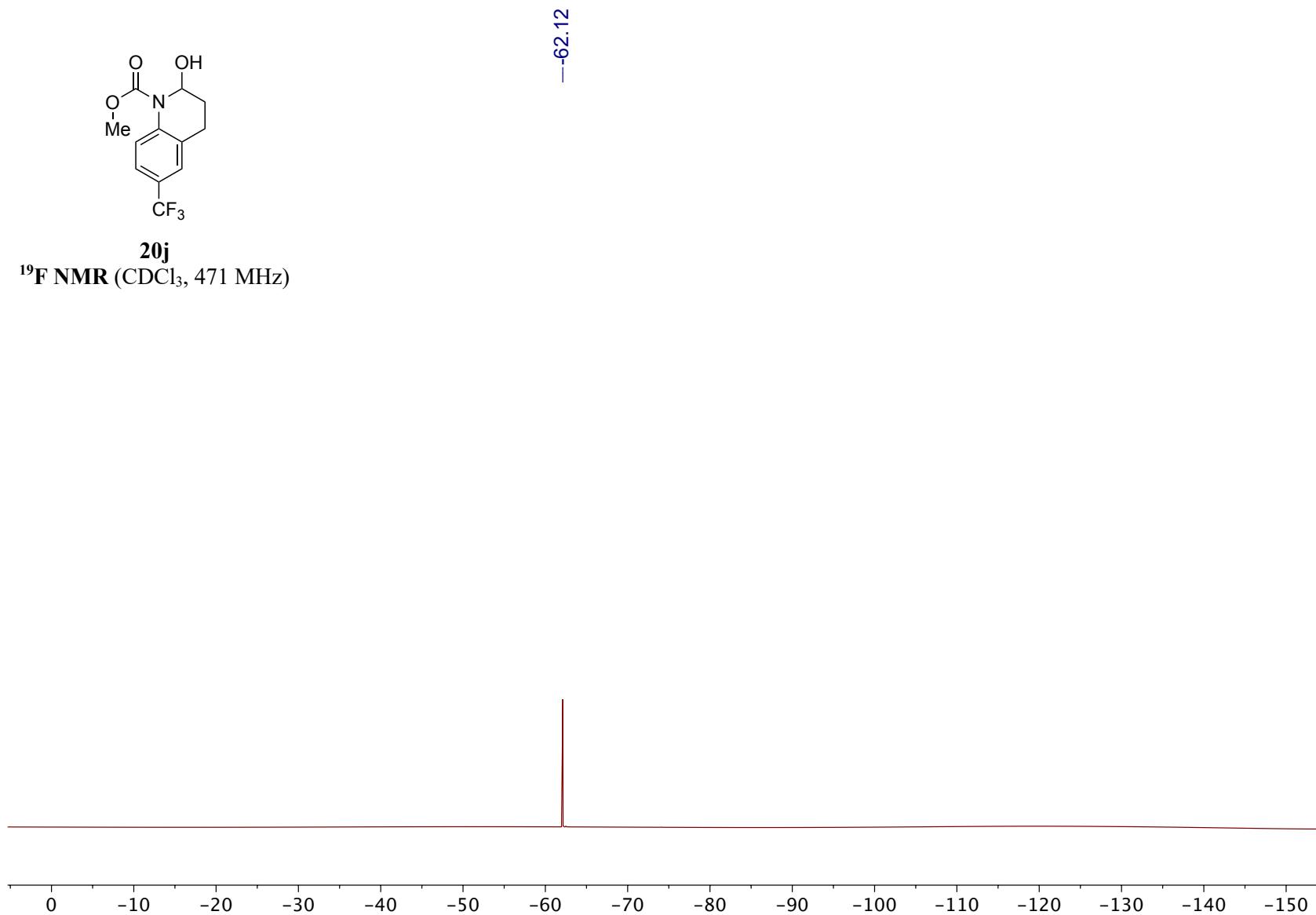
20j
¹H NMR (CDCl₃, 600 MHz)

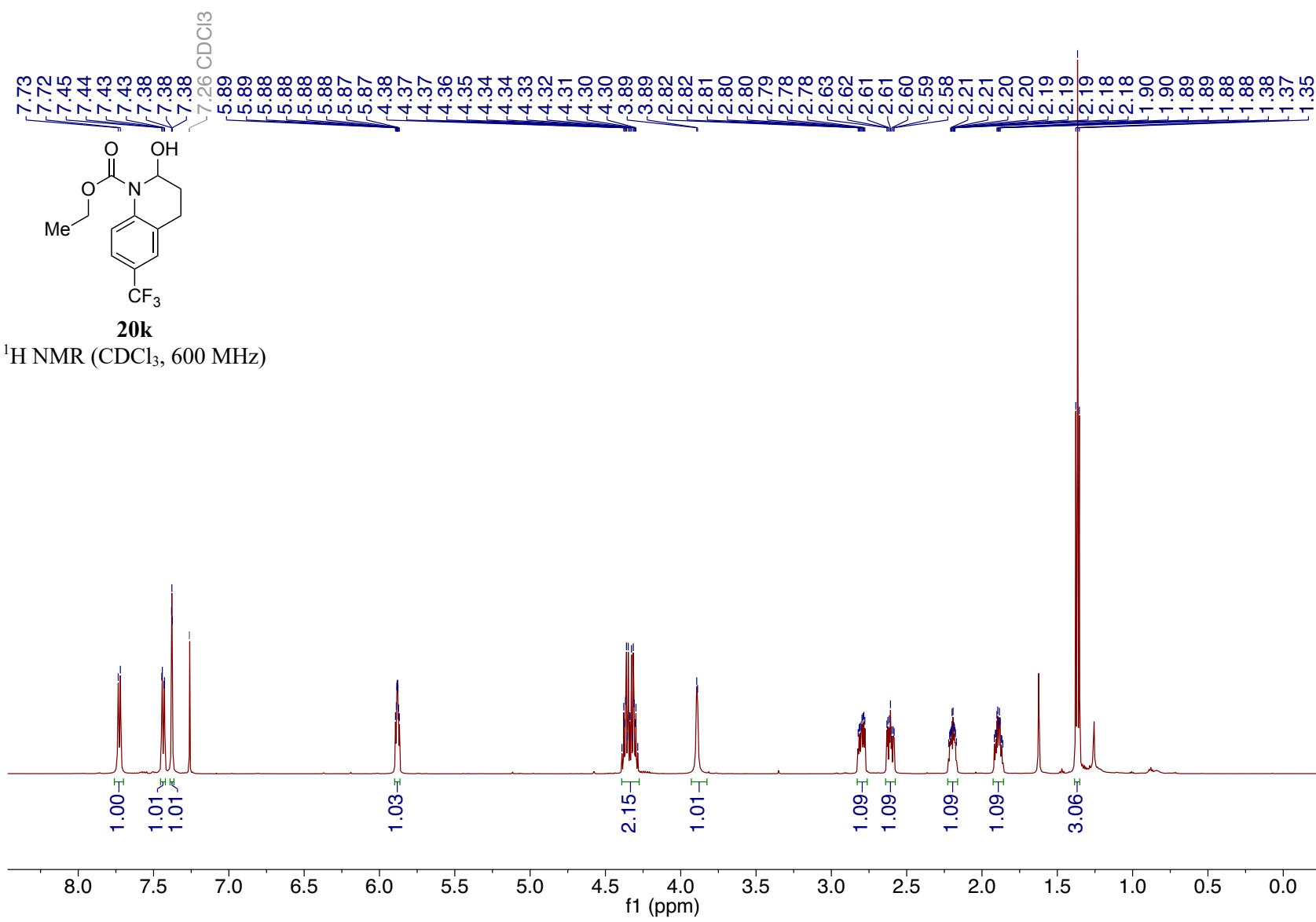


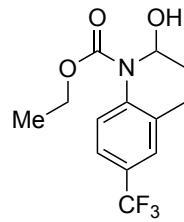




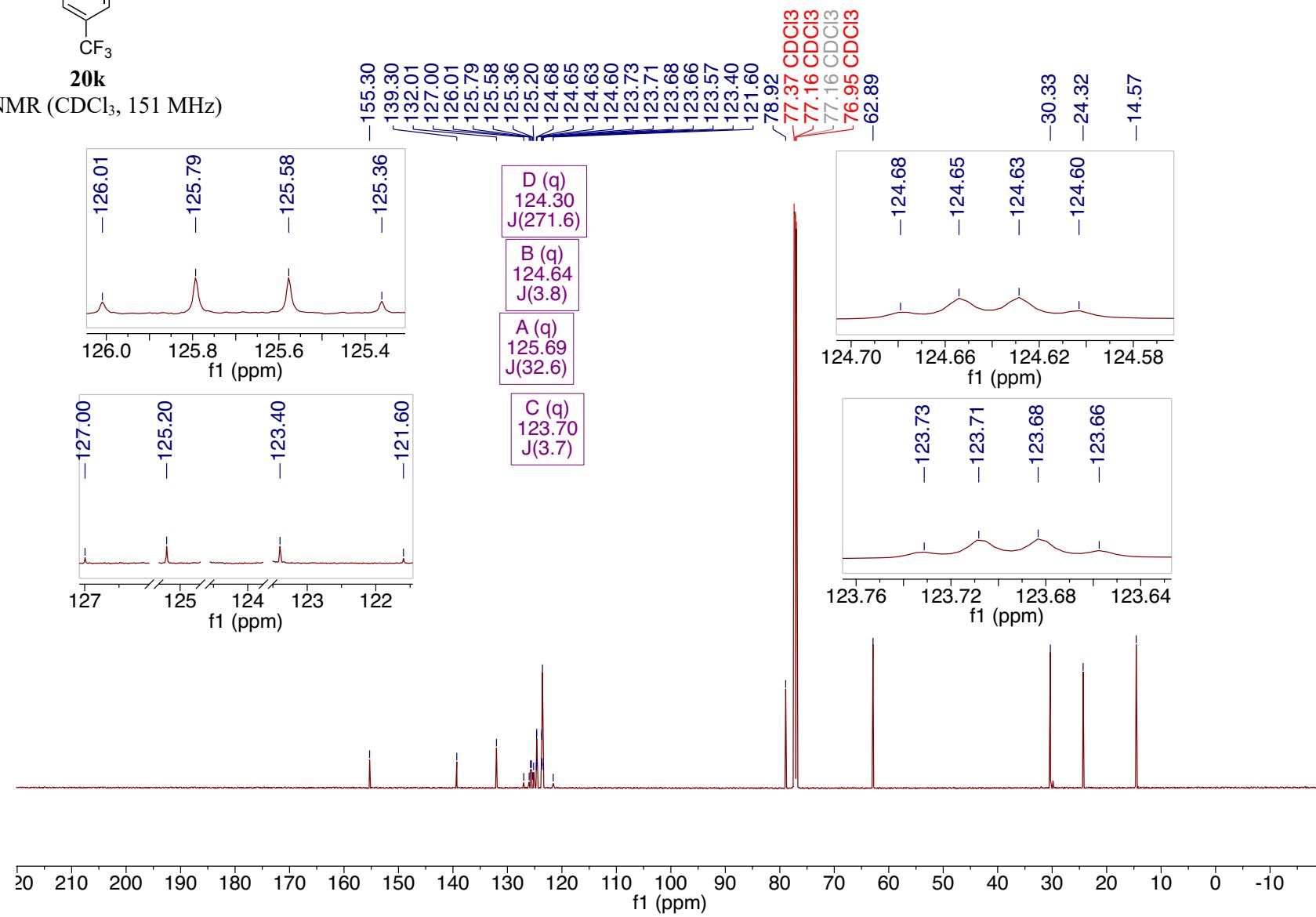
20j
¹⁹F NMR (CDCl₃, 471 MHz)

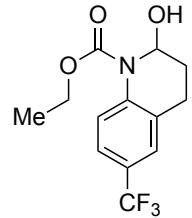




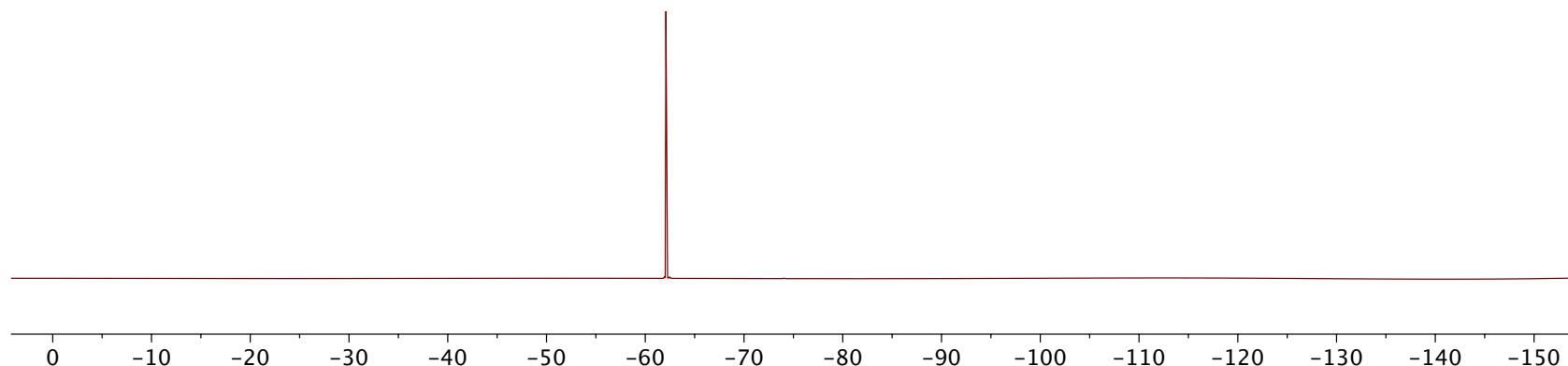


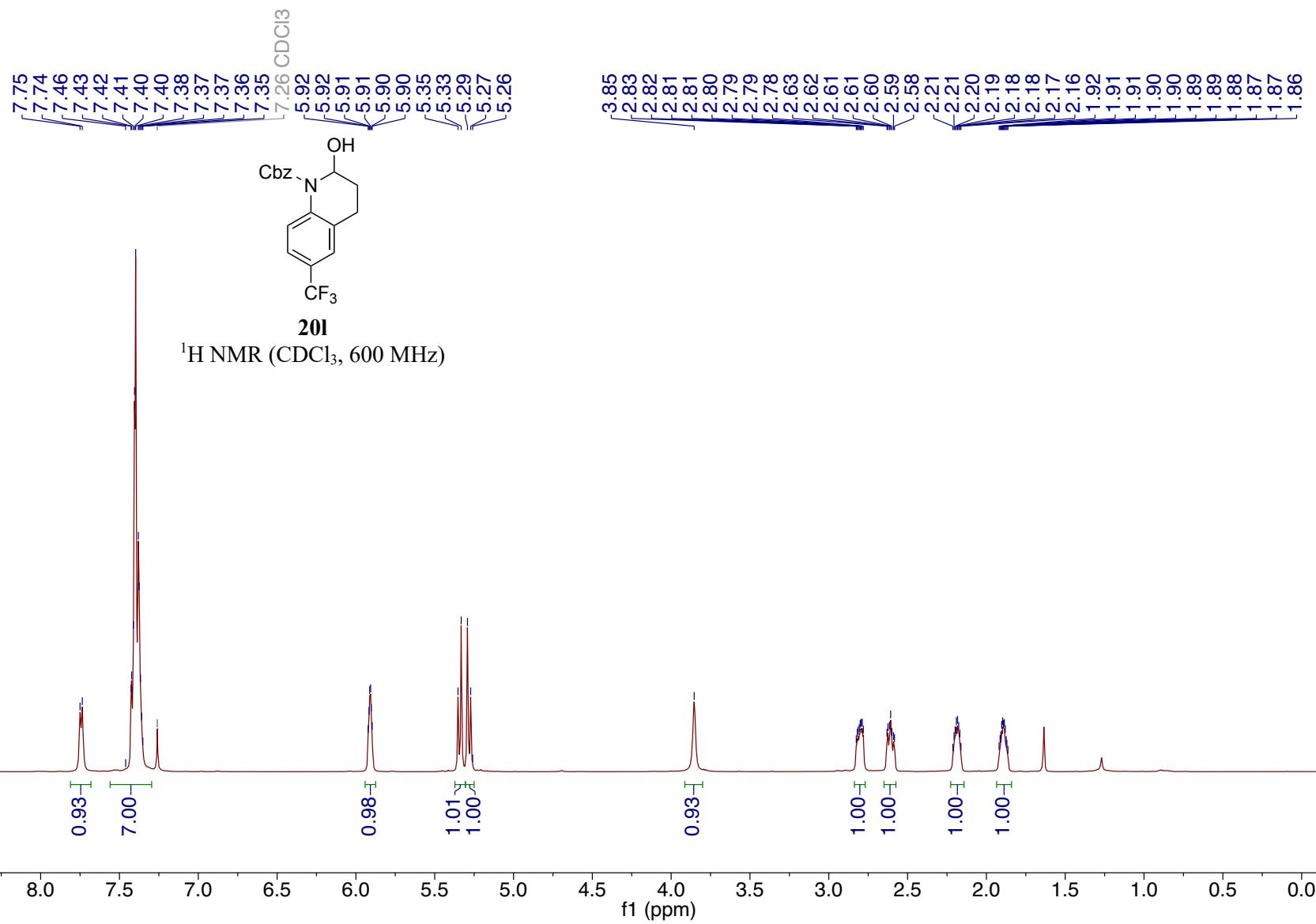
¹³C NMR (CDCl_3 , 151 MHz)

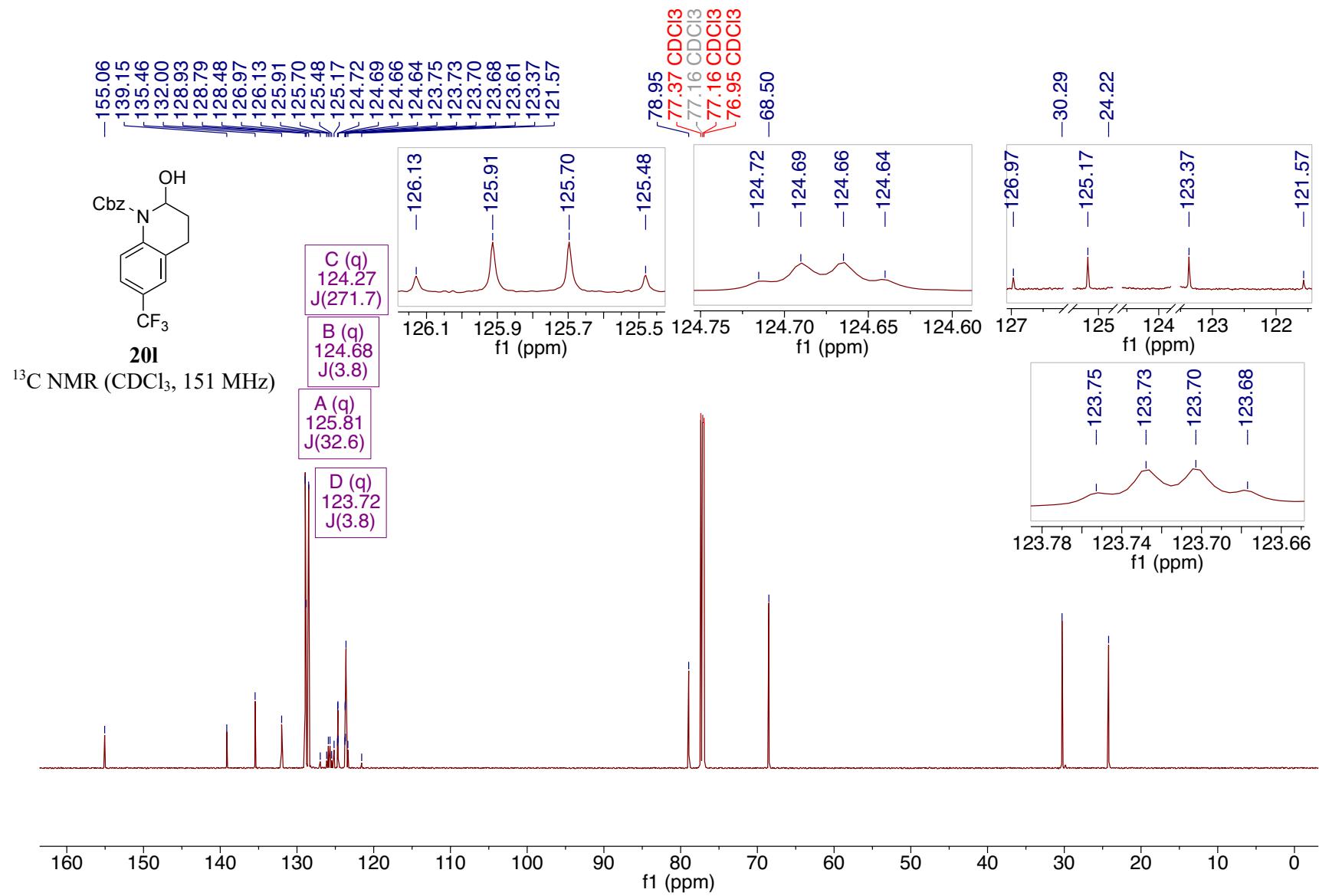


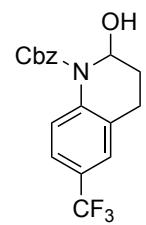


20k
¹⁹F NMR (CDCl₃, 471 MHz)

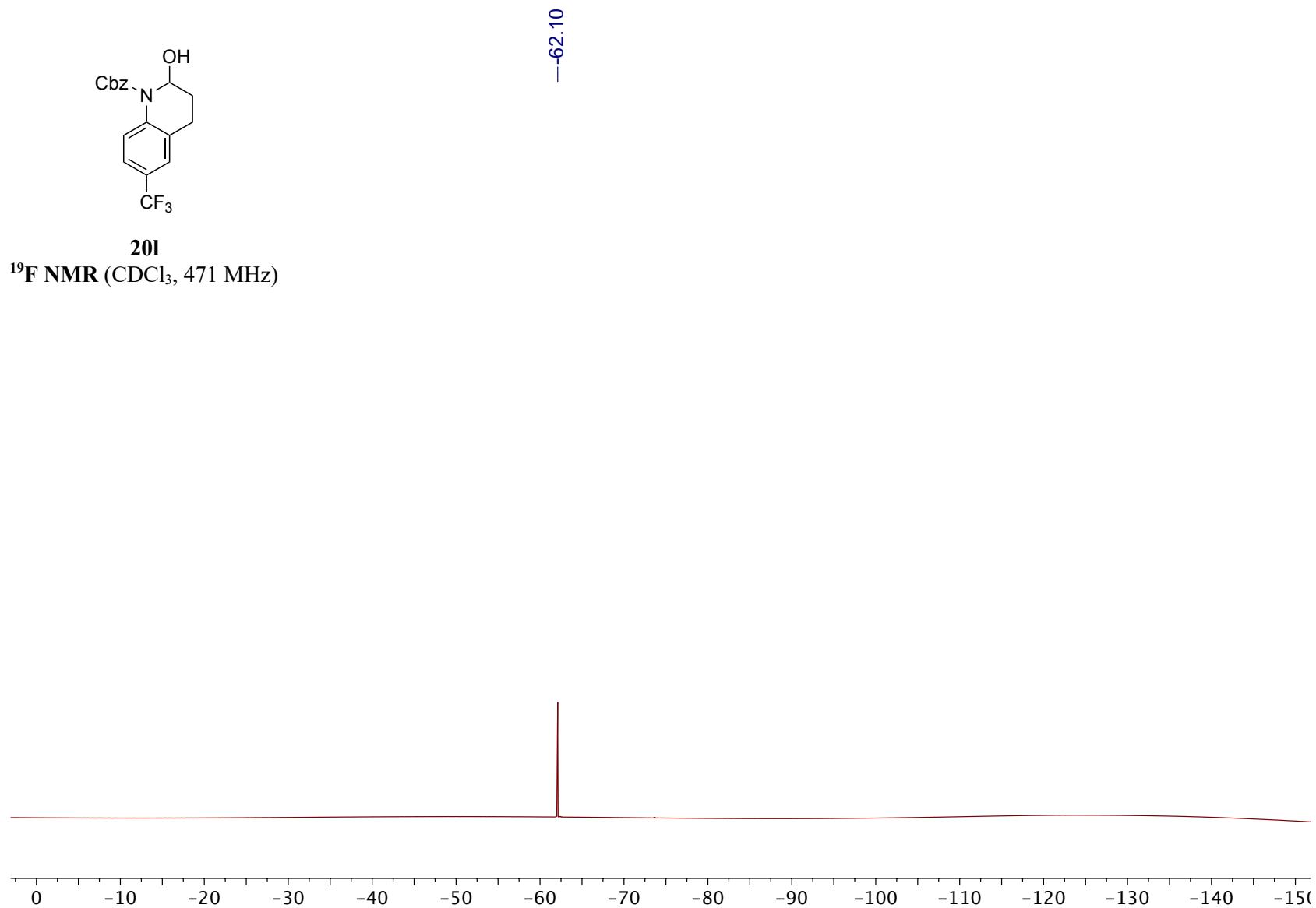


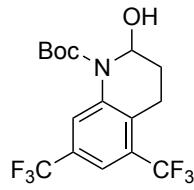






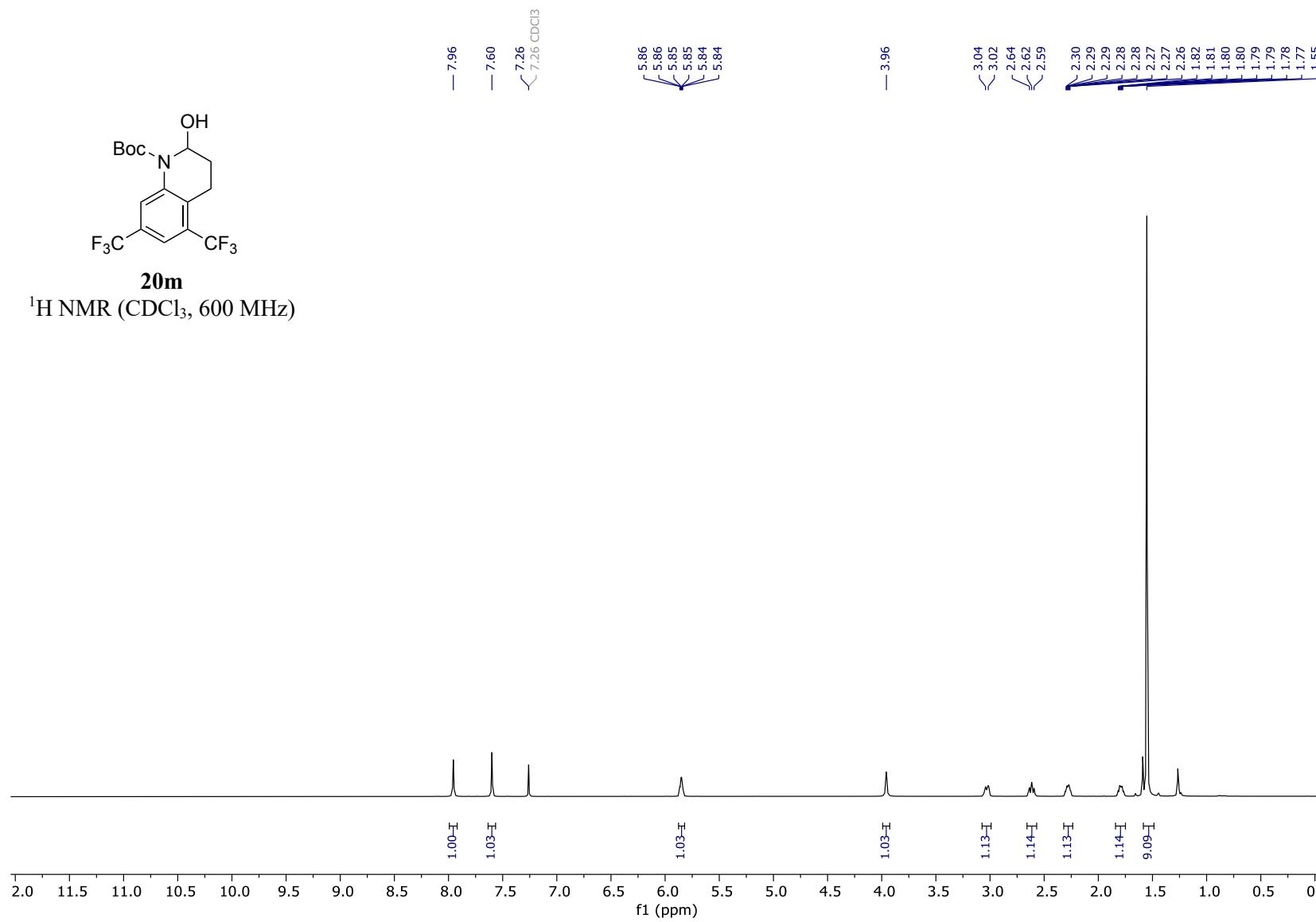
20l
¹⁹F NMR (CDCl₃, 471 MHz)

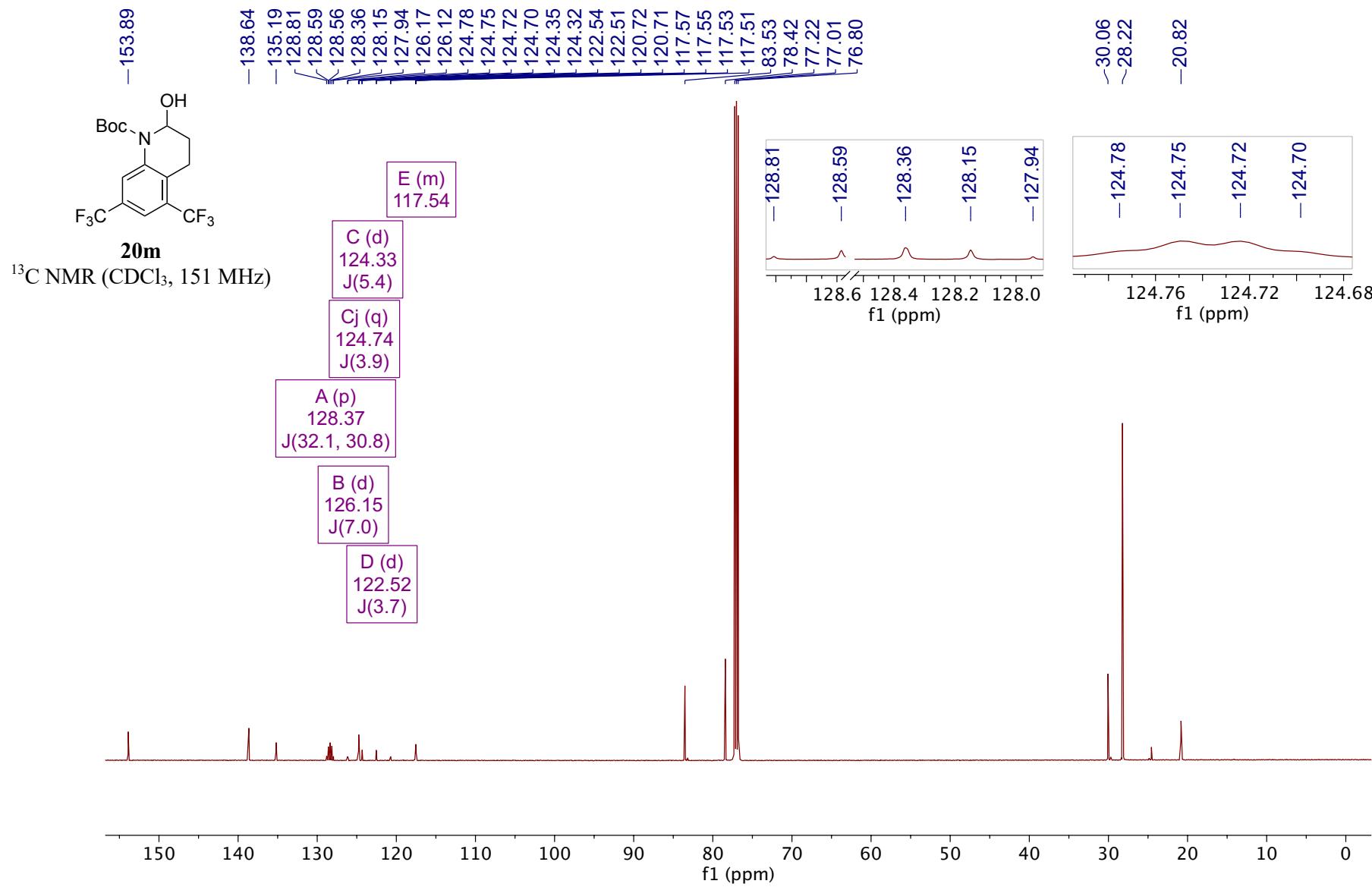


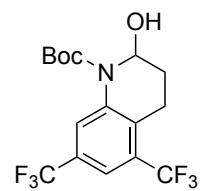


20m

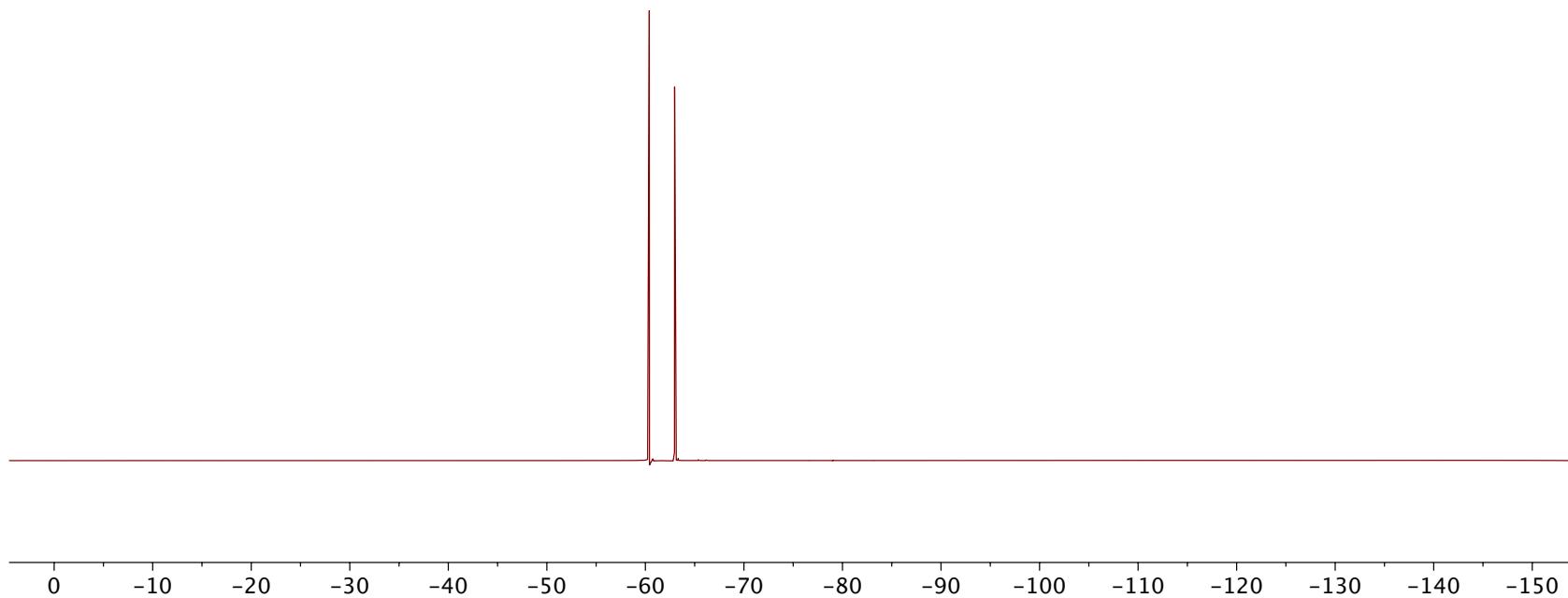
¹H NMR (CDCl₃, 600 MHz)

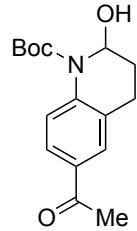




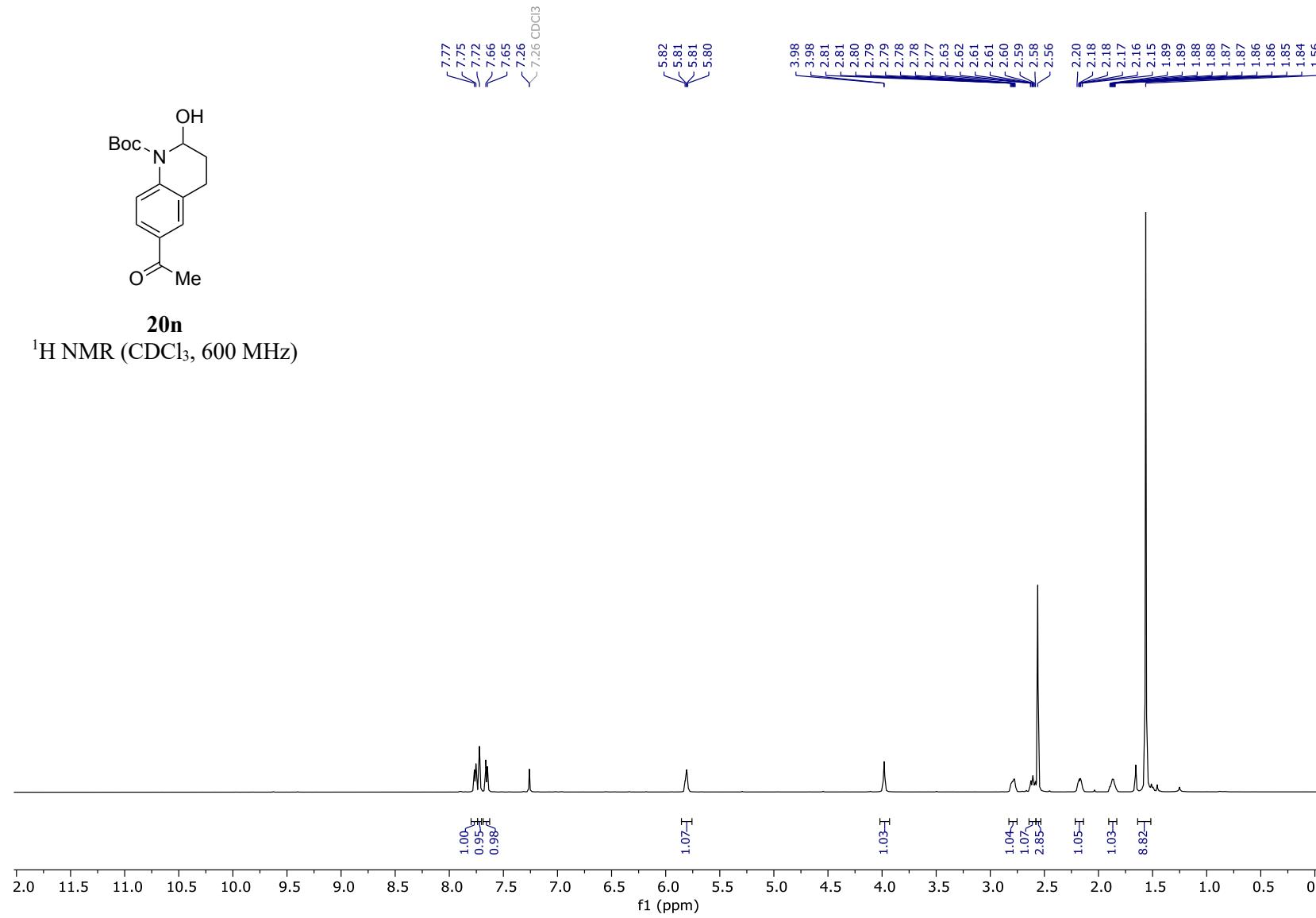


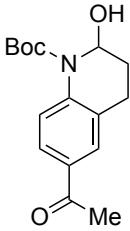
20m
¹⁹F NMR (CDCl₃, 471 MHz)





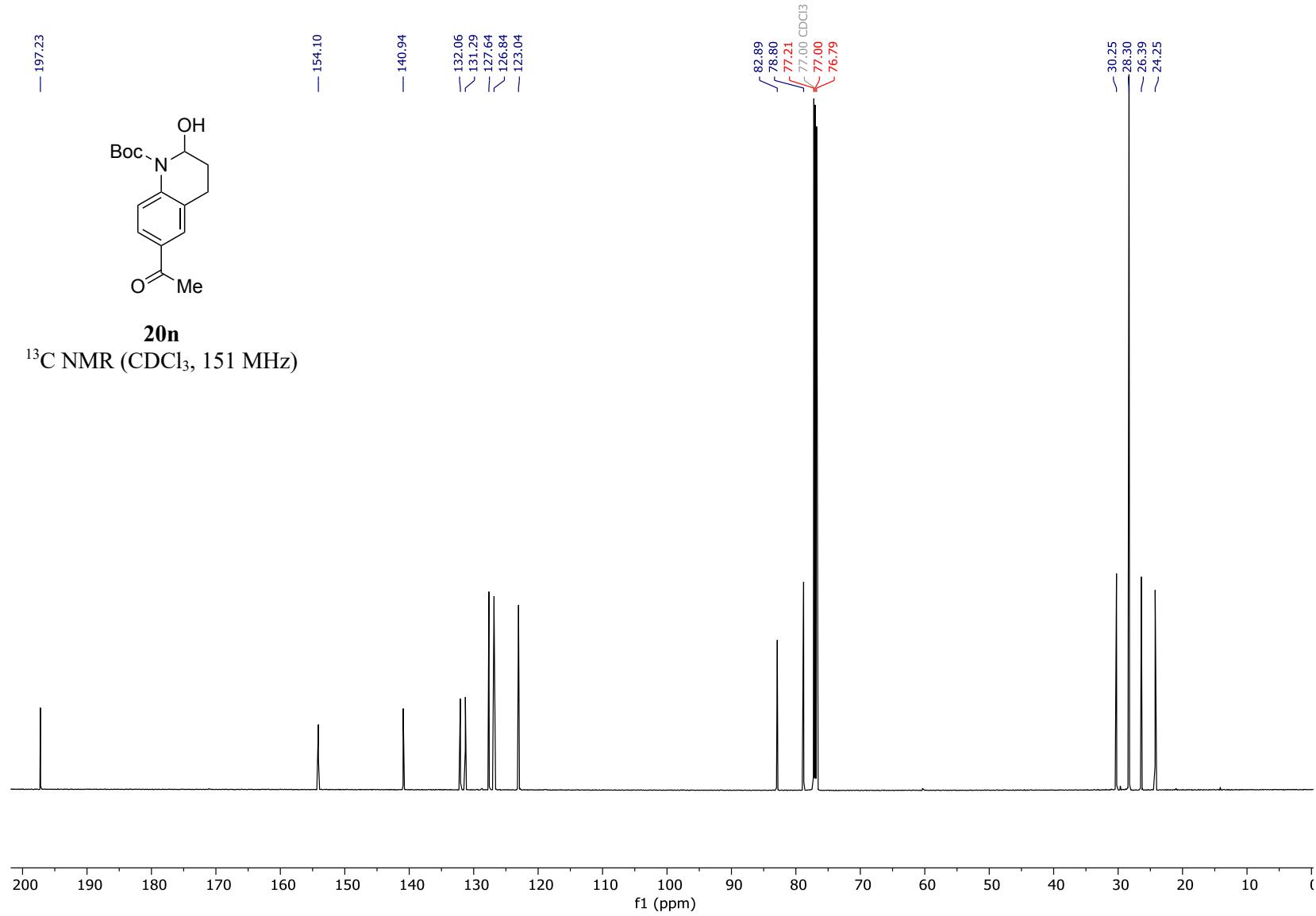
^1H NMR (CDCl_3 , 600 MHz)

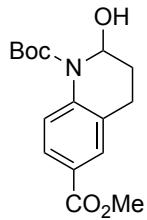




20n

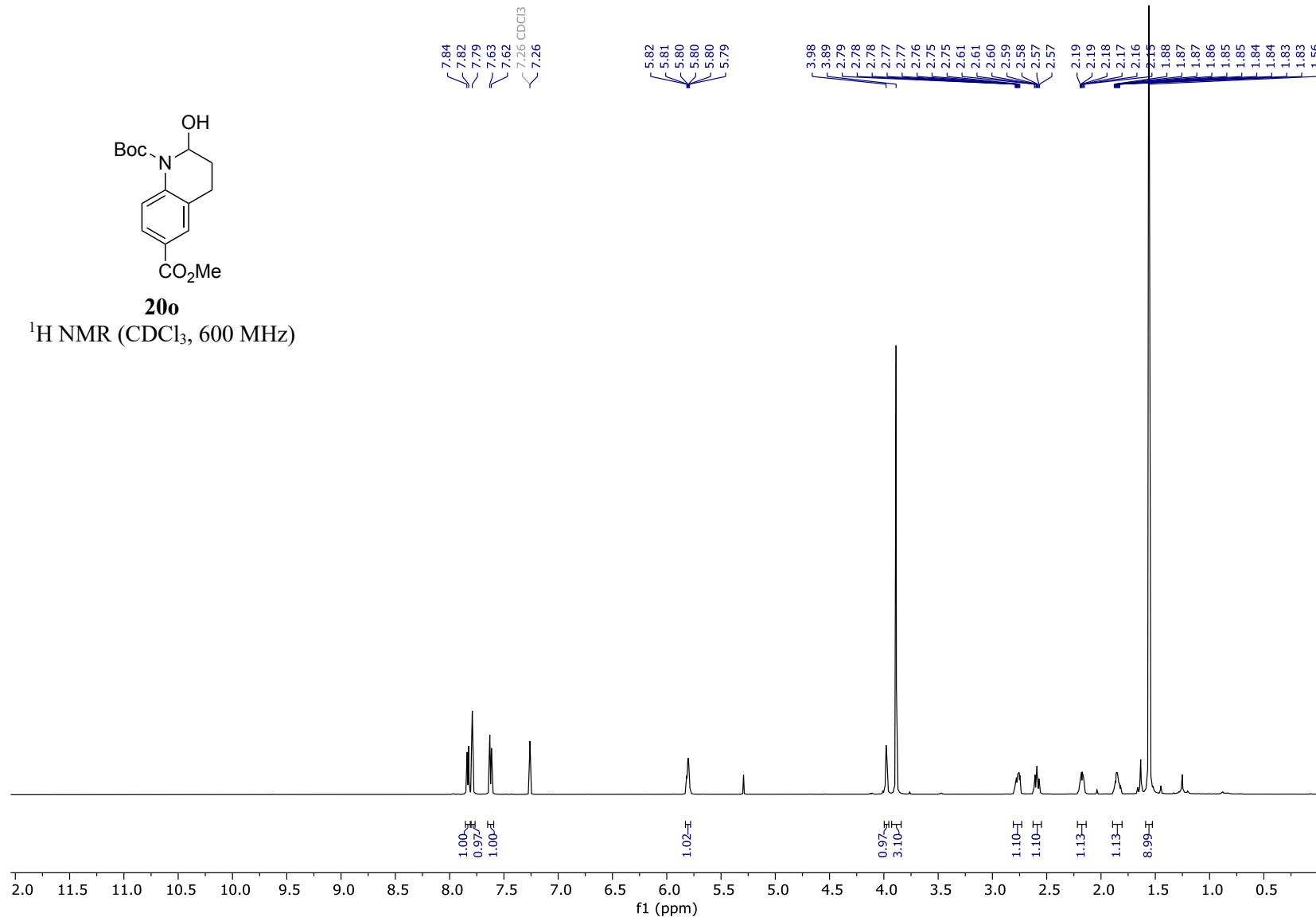
^{13}C NMR (CDCl_3 , 151 MHz)

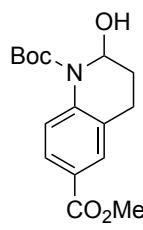




200

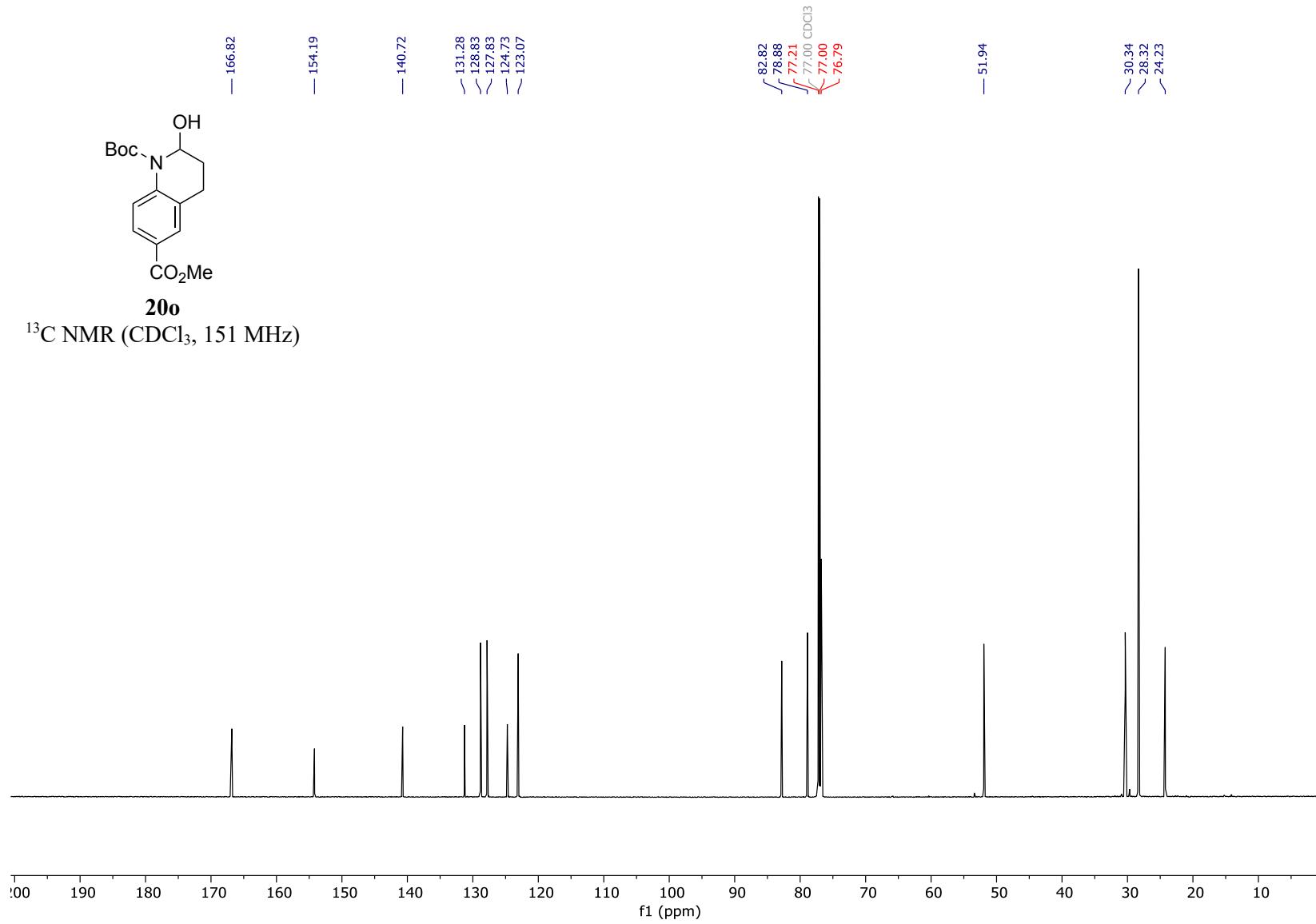
¹H NMR (CDCl₃, 600 MHz)

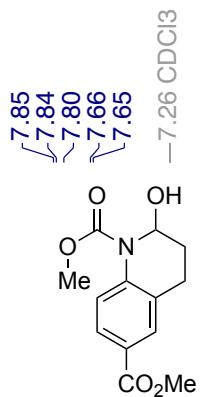




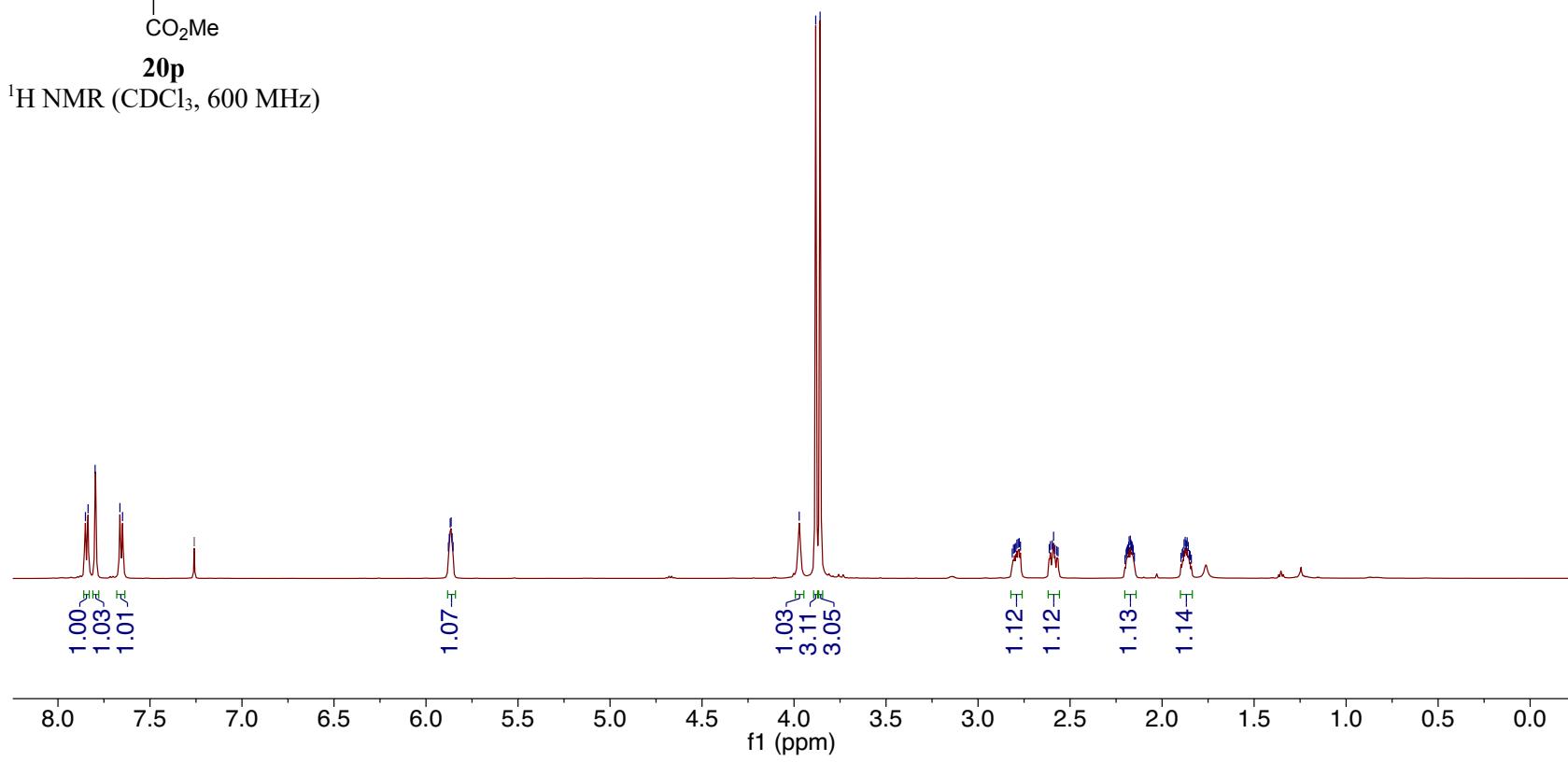
20o

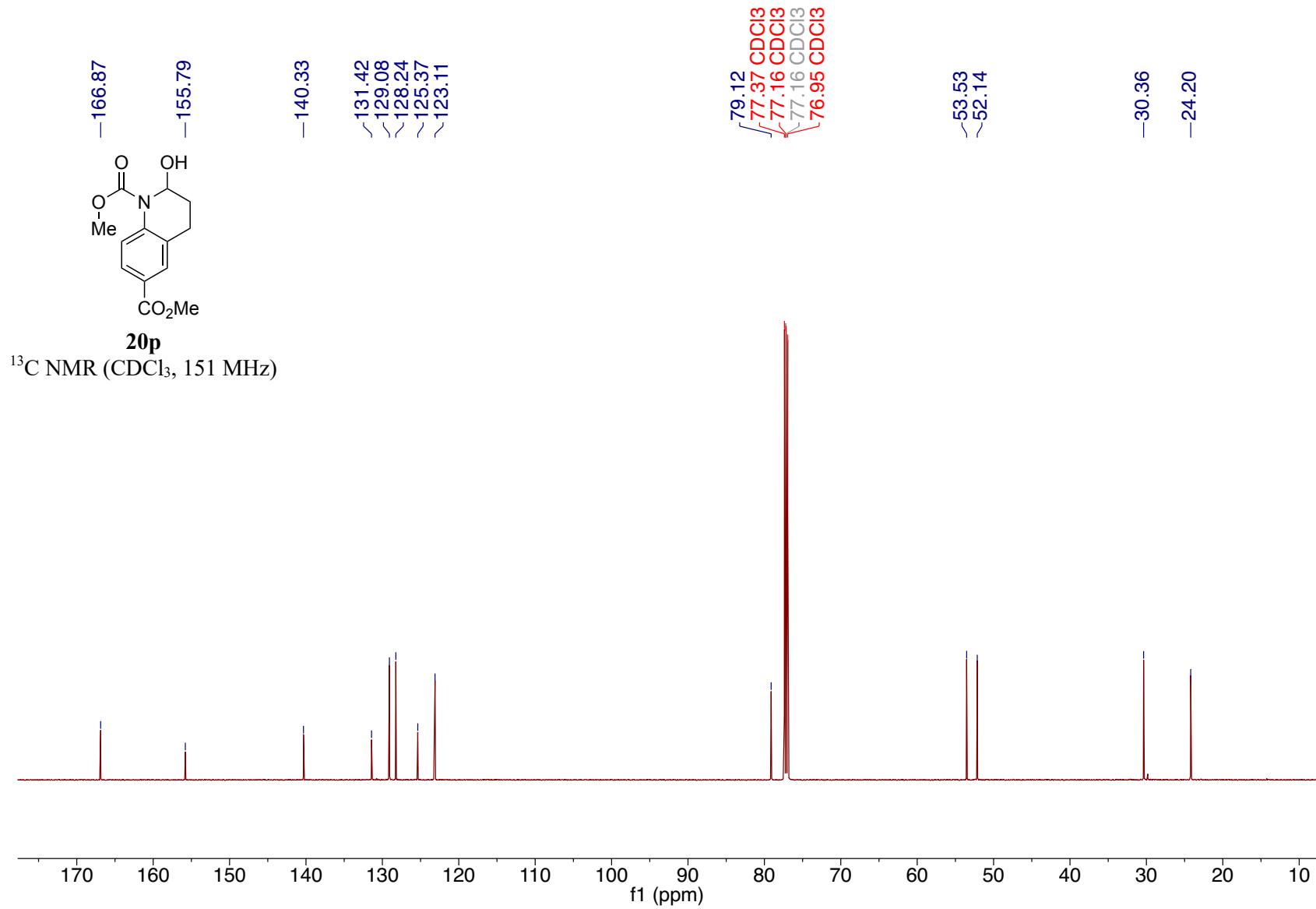
^{13}C NMR (CDCl_3 , 151 MHz)

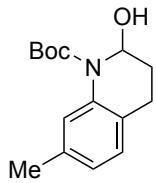




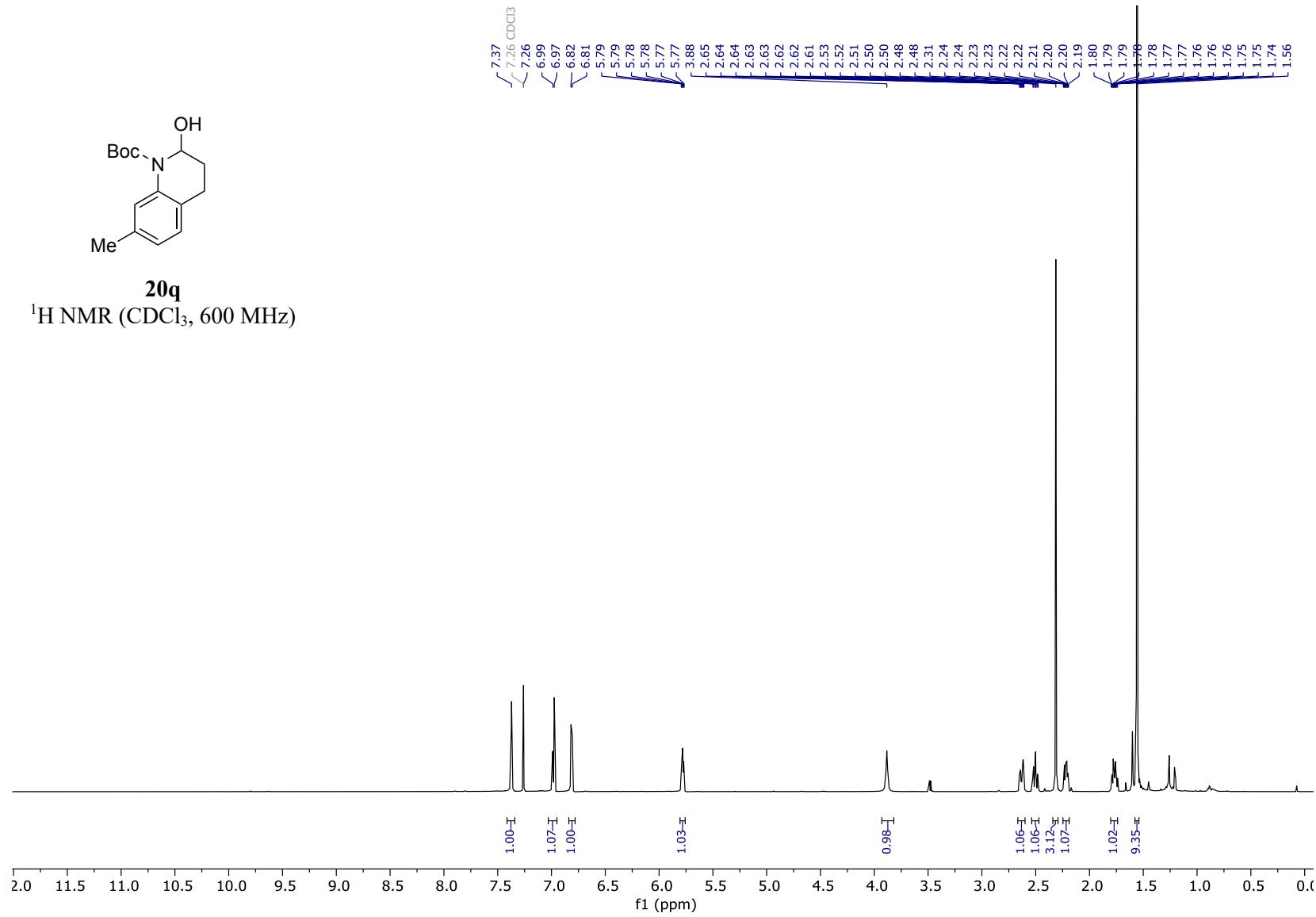
20p

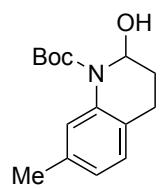




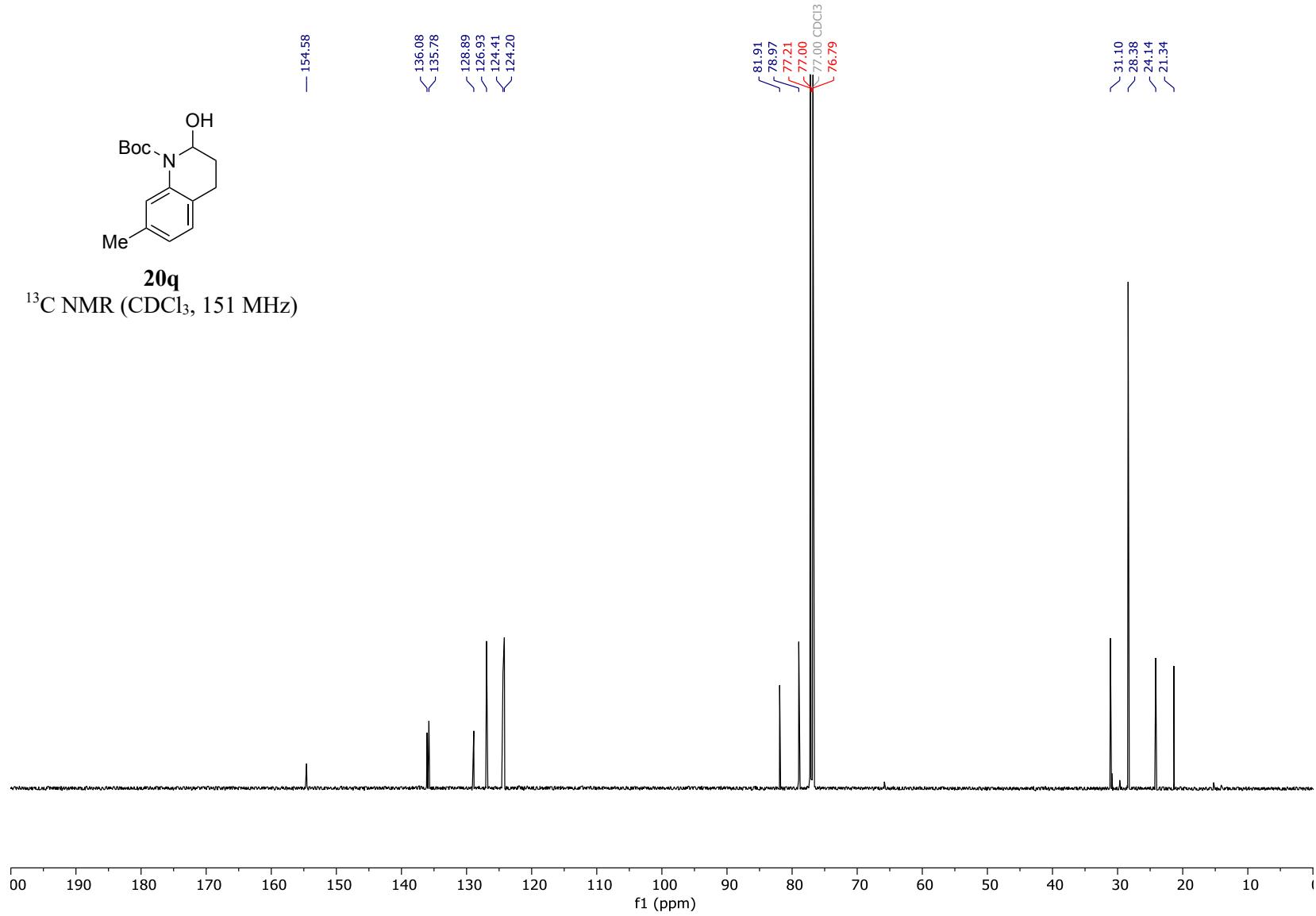


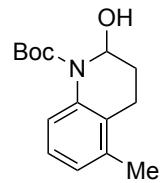
20q
 ^1H NMR (CDCl_3 , 600 MHz)





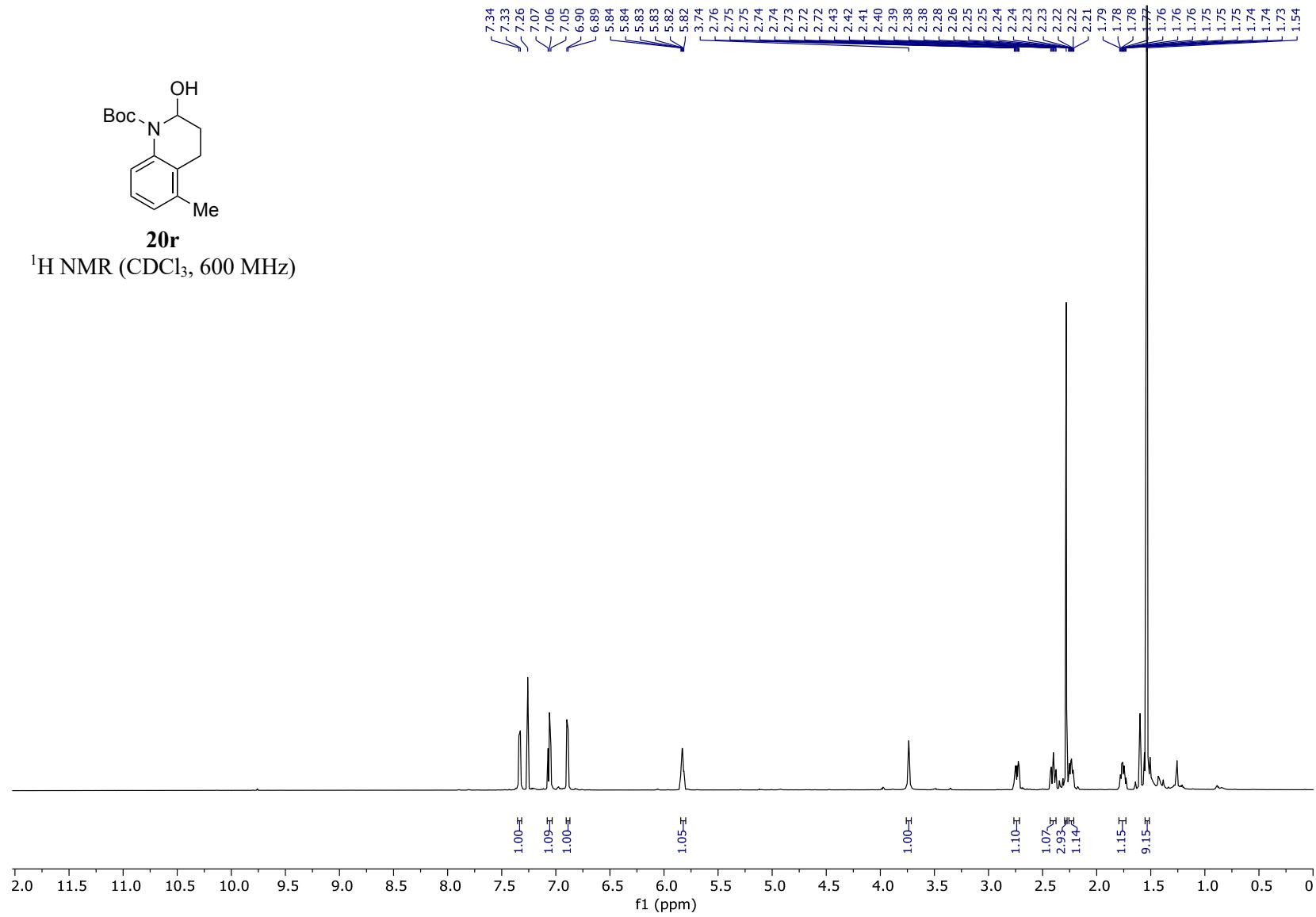
20q
 ^{13}C NMR (CDCl_3 , 151 MHz)

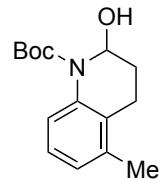




20r

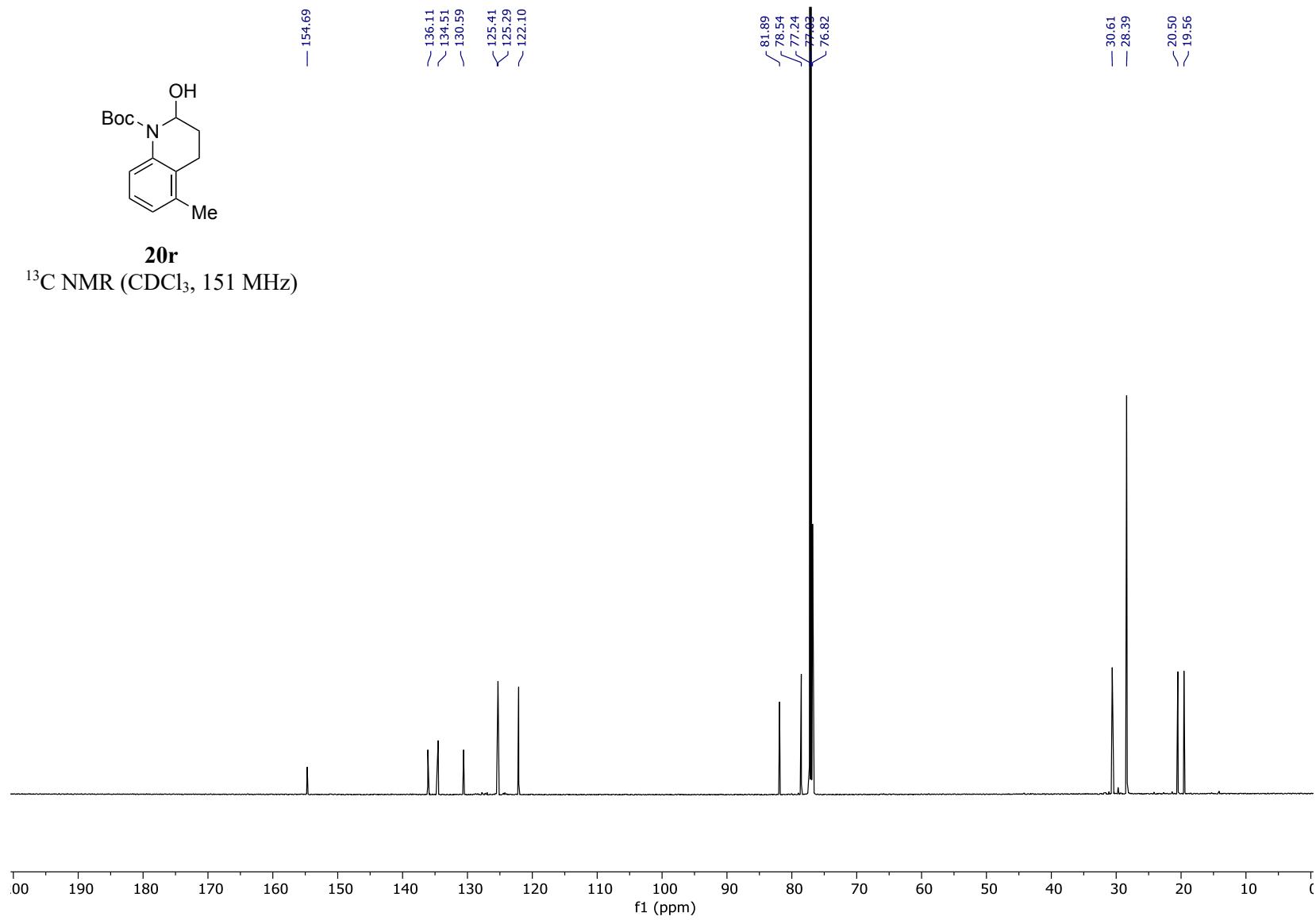
¹H NMR (CDCl₃, 600 MHz)

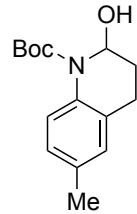




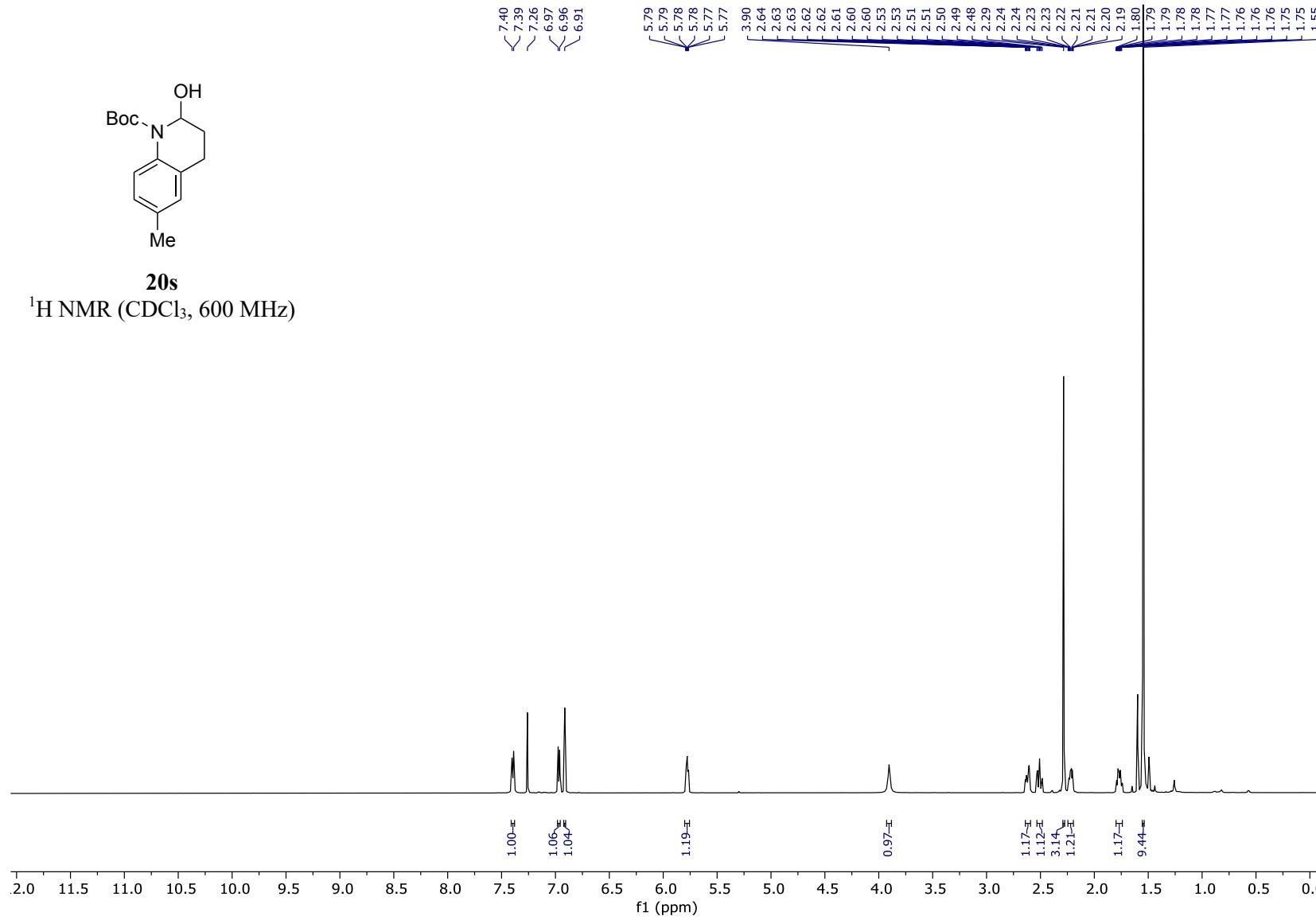
— 154.69

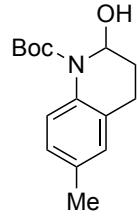
20r
 ^{13}C NMR (CDCl_3 , 151 MHz)





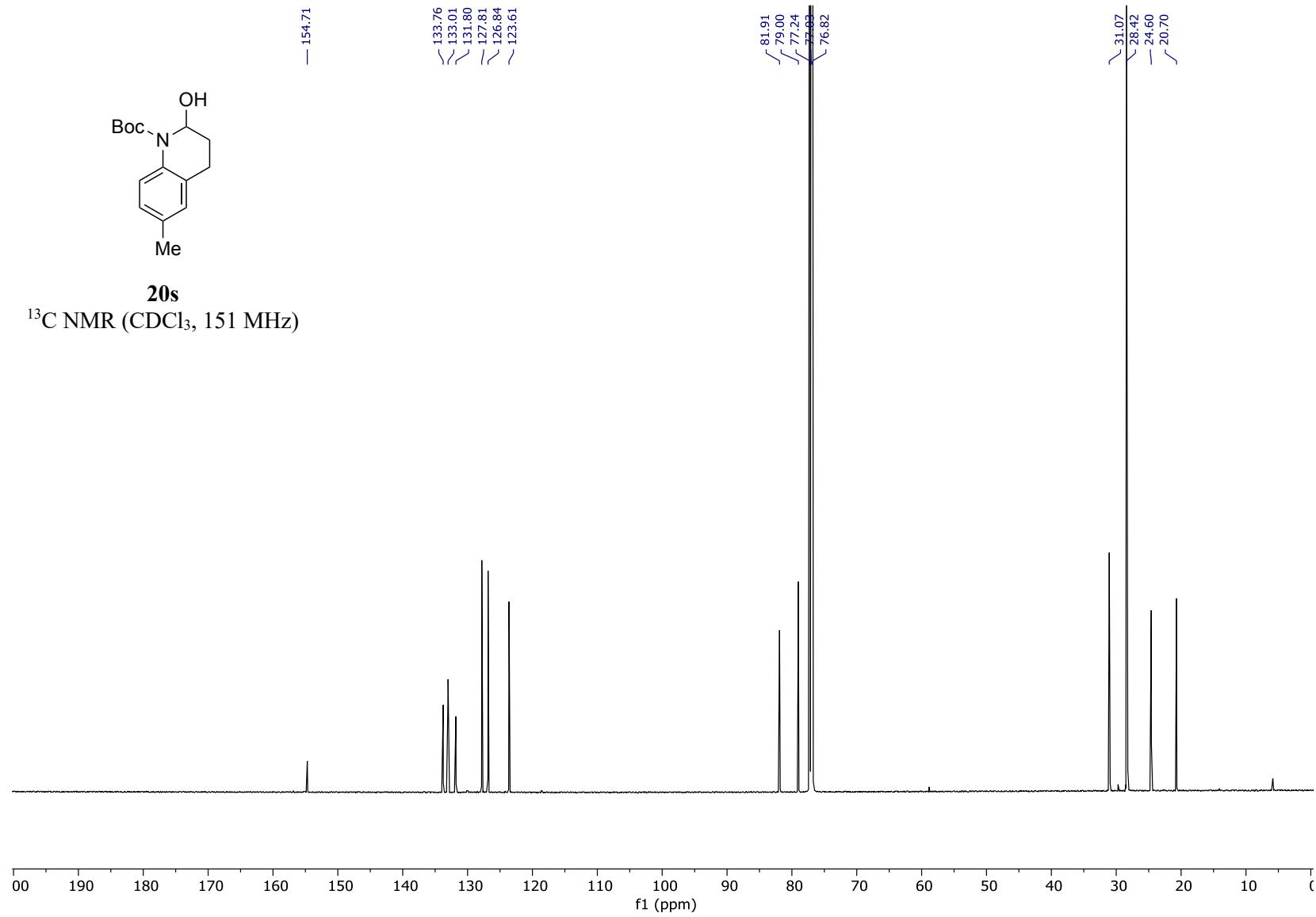
¹H NMR (CDCl₃, 600 MHz)

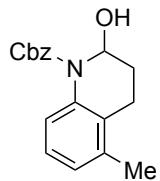




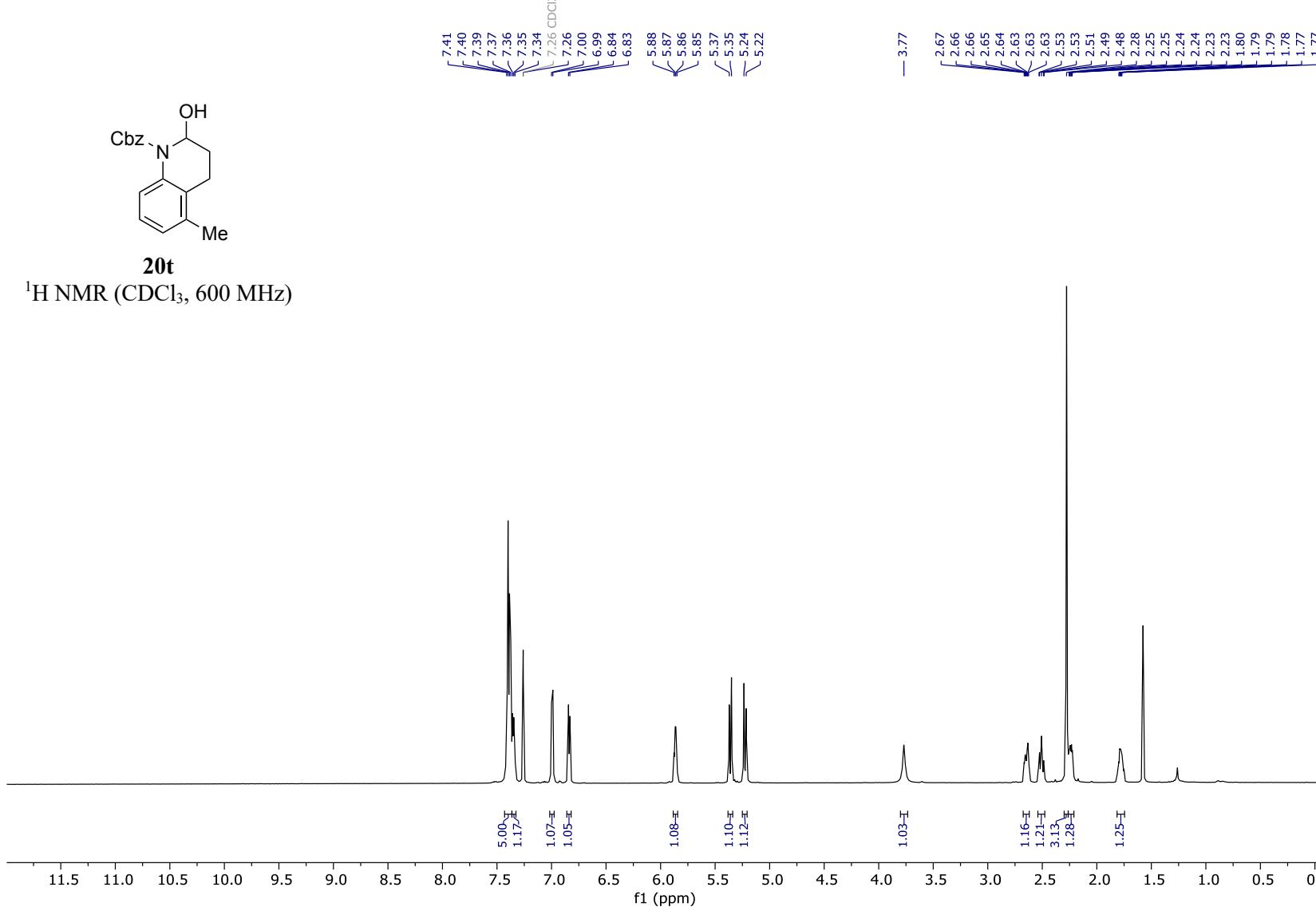
— 154.71

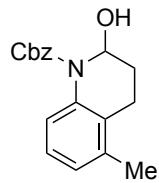
20s
 ^{13}C NMR (CDCl_3 , 151 MHz)





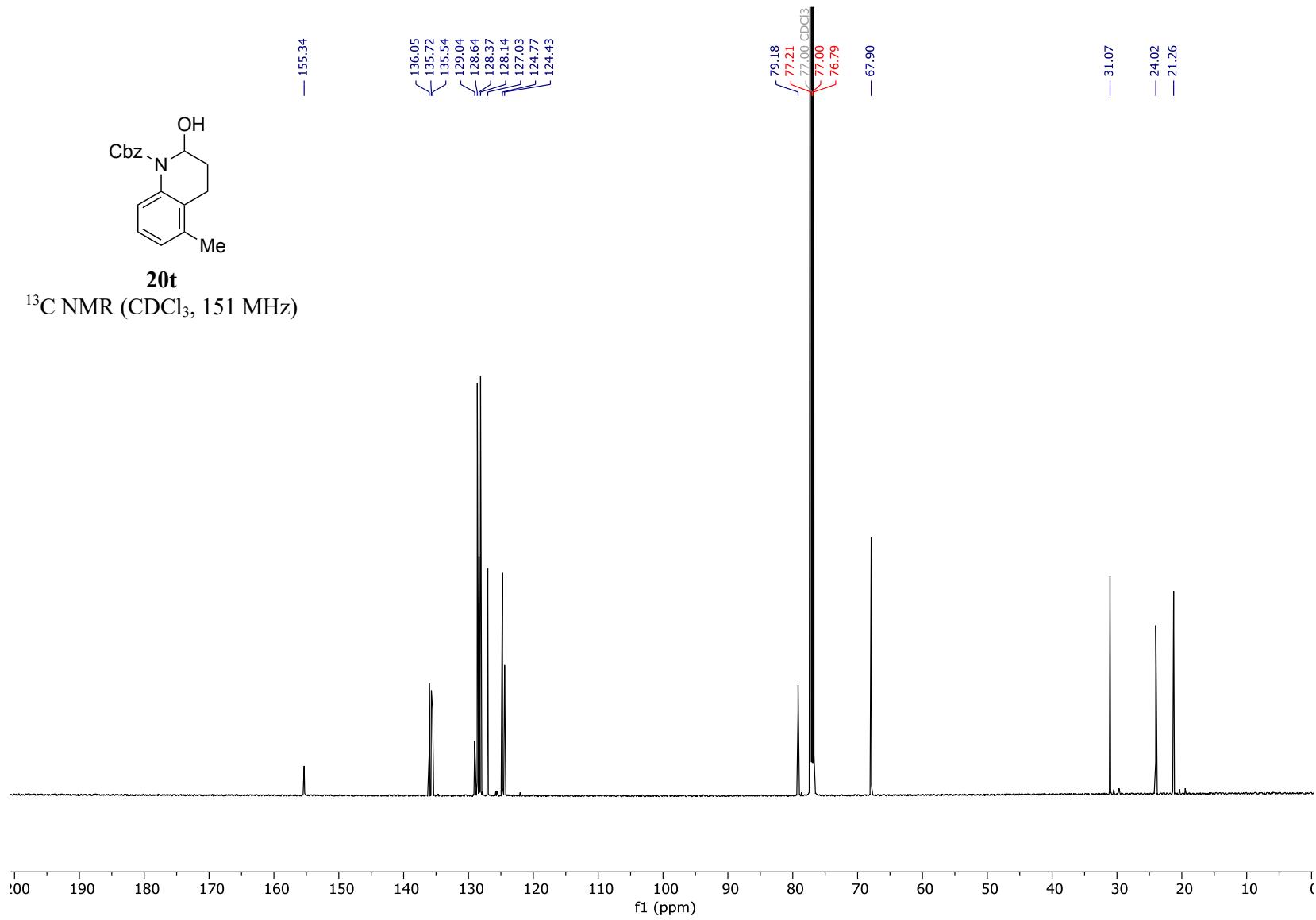
¹H NMR (CDCl₃, 600 MHz)

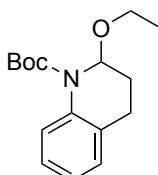




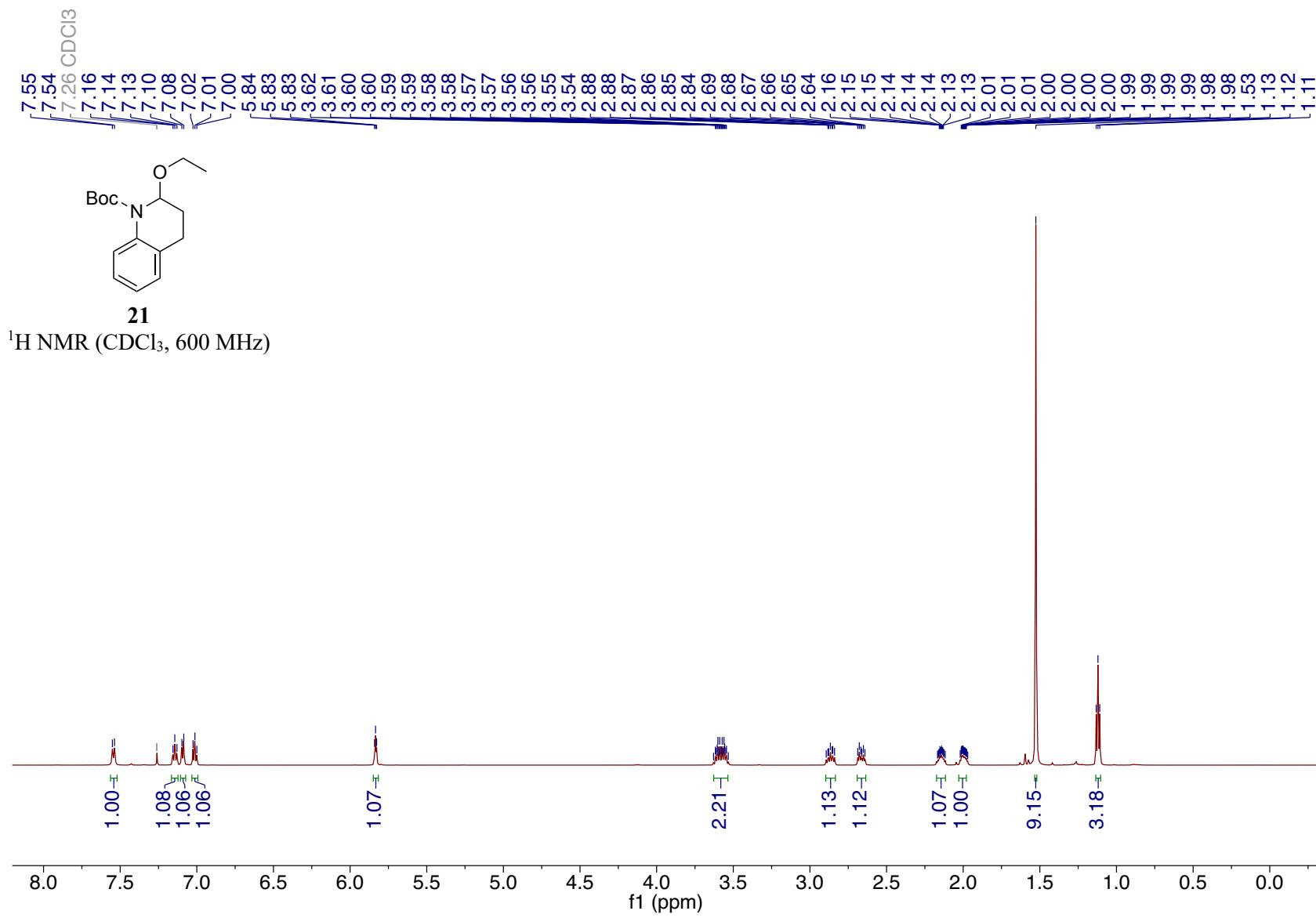
20t

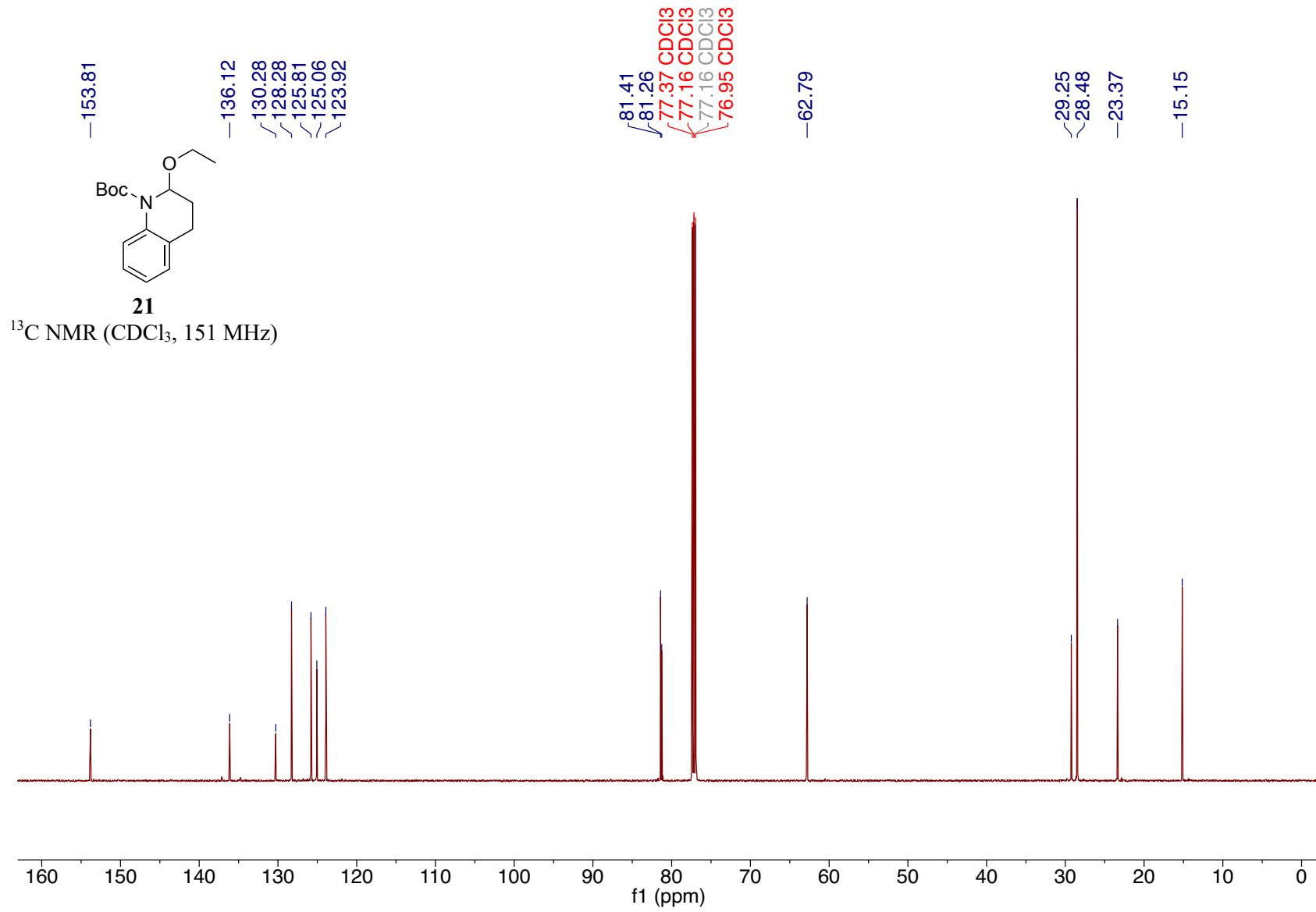
^{13}C NMR (CDCl_3 , 151 MHz)

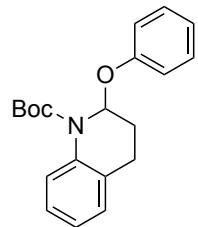




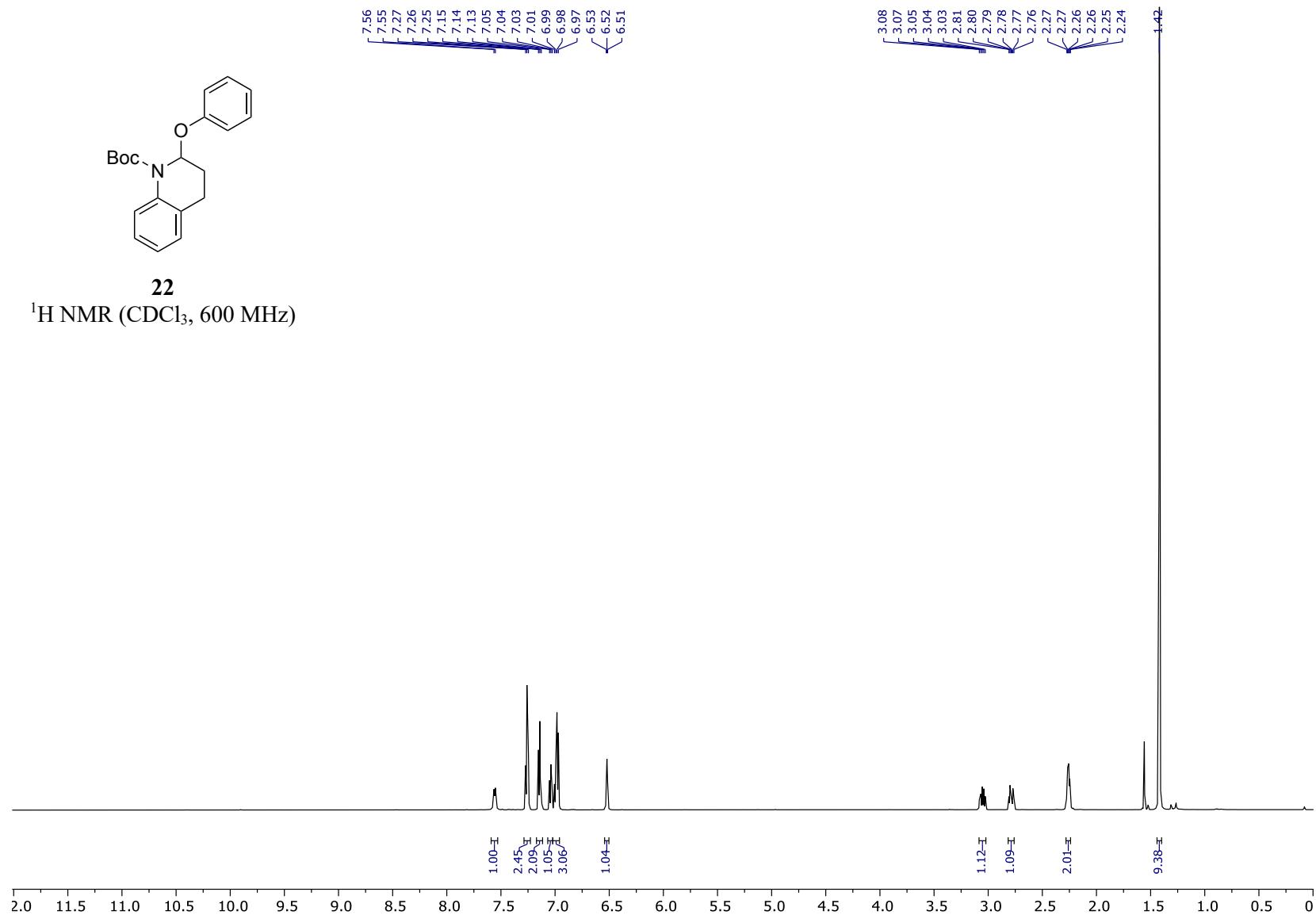
21
 ^1H NMR (CDCl_3 , 600 MHz)

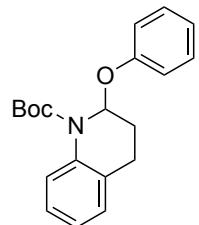






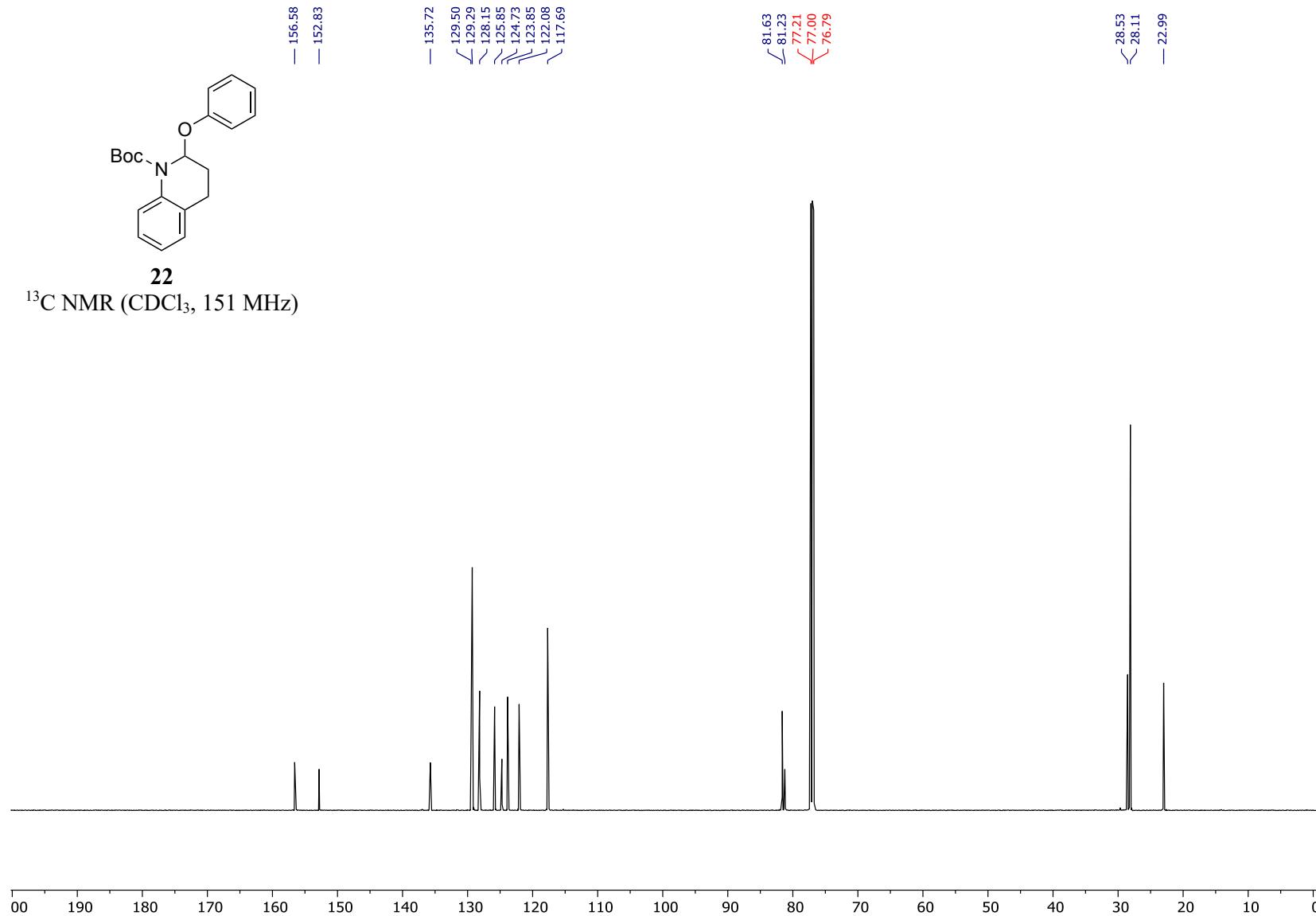
^1H NMR (CDCl_3 , 600 MHz)



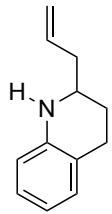


22

^{13}C NMR (CDCl_3 , 151 MHz)

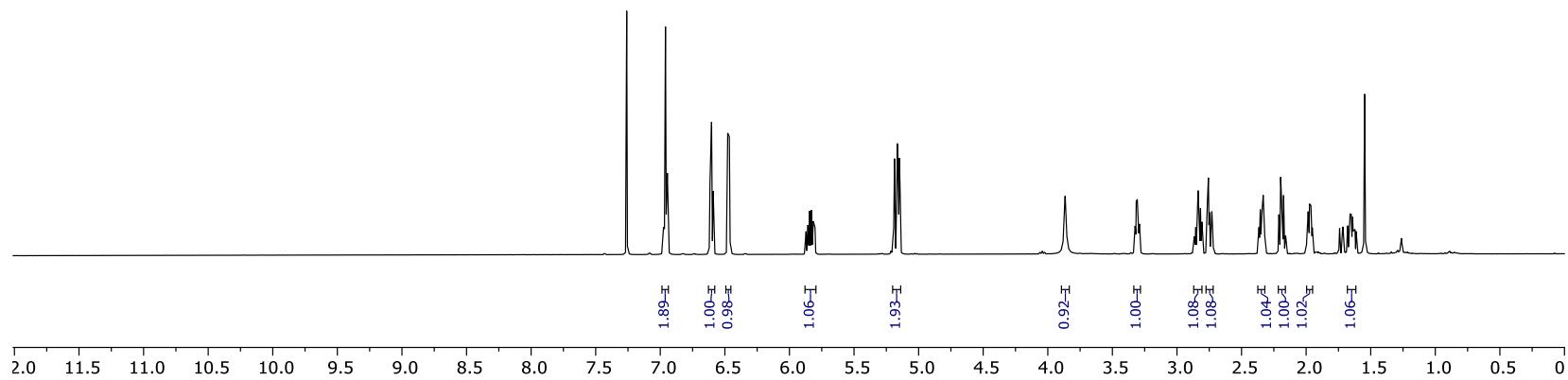


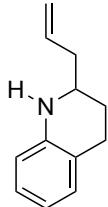
7.26
6.97
6.96
6.95
6.62
6.60
6.59
6.48
6.47
5.86
5.85
5.84
5.84
5.83
5.83
5.83
5.82
5.82
5.81
5.80
5.19
5.19
5.17
5.16
5.16
5.16
5.15
5.15
3.87
3.32
3.32
3.32
3.31
3.31
3.31
3.30
3.30
3.30
3.29
3.29
2.85
2.84
2.84
2.83
2.82
2.81
2.76
2.76
2.75
2.74
2.73
2.72
2.36
2.36
2.34
2.34
2.33
2.33
2.22
2.21
2.20
2.19
2.18
2.17
2.16
1.99
1.99
1.98
1.98
1.97
1.97
1.96
1.96
1.66
1.66
1.66
1.65
1.65
1.65
1.65
1.64
1.64
1.64
1.63
1.63
1.63



23

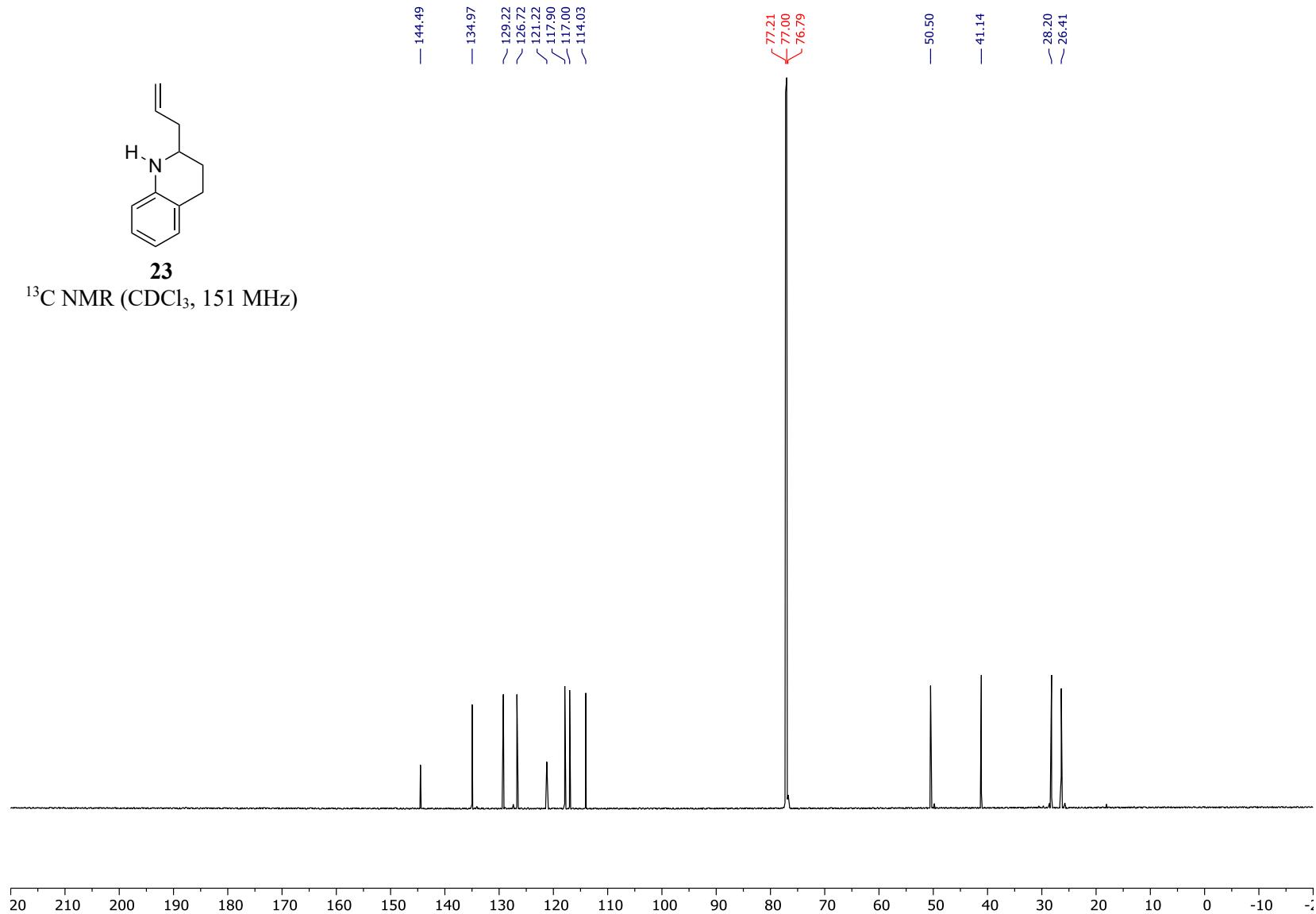
¹H NMR (CDCl₃, 600 MHz)

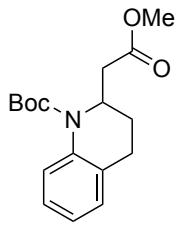




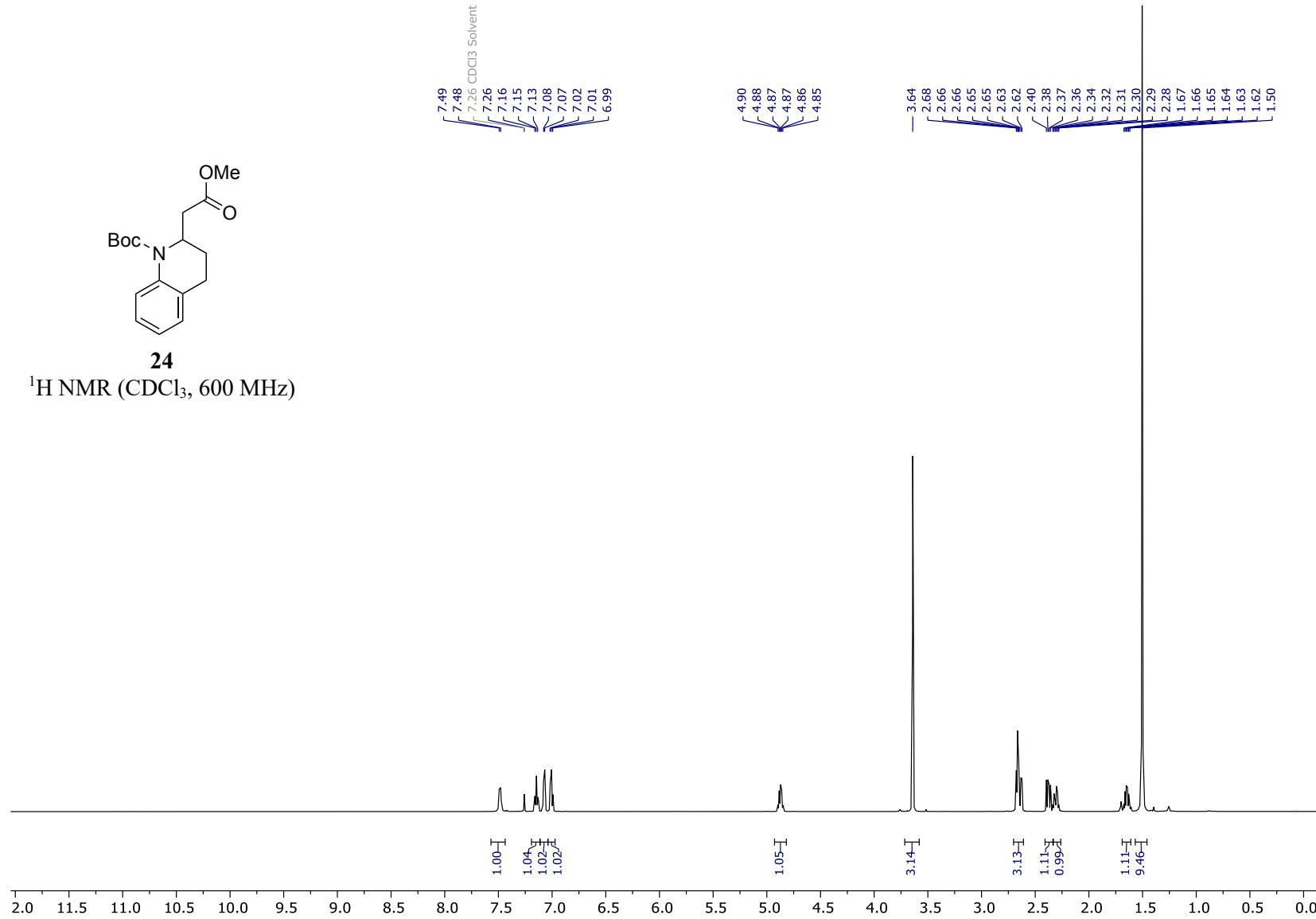
23

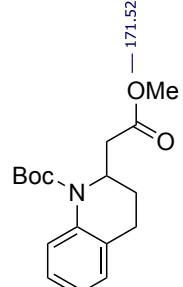
¹³C NMR (CDCl₃, 151 MHz)





^1H NMR (CDCl_3 , 600 MHz)





24

^{13}C NMR (CDCl_3 , 151 MHz)

