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SUPPORTING INFORMATION

Synthesis of C11-to-C14 Methyl-shifted all-*trans*-Retinal Analogues and Their Activities On Human Aldo-Keto Reductases

Aurea Rivas,^a Raquel Pequerul^b, Vito Barracco,^{b,c} Marta Domínguez,^a Susana López,^d Rafael Jiménez,^b Xavier Parés,^b Rosana Alvarez,^{a,*} Jaume Farrés^{b,*} and Angel R. de Lera^{a,*}

^a Departamento de Química Orgánica, Facultade de Química, CINBIO and IIS Galicia Sur, Universidade de Vigo, E-36310 Vigo, Spain

^b Department of Biochemistry and Molecular Biology, Faculty of Biosciences, Universitat Autònoma de Barcelona, E-08193 Bellaterra, Barcelona, Spain

^c Department of Biology, Biochemistry Unit, University of Pisa, I-56126 Pisa, Italy

^d Departamento de Química Orgánica, Facultade de Química, Universidade de Santiago de Compostela, E-15782 Santiago, Spain

* To whom correspondence should be addressed: Angel R. de Lera, Tel: +34-986-812316 Fax +34-986-818622, E-mail: qolera@uvigo.es (AdL); Rosana Alvarez, Tel: +34-986-812632 Fax +34-986-818622, E-mail: rar@uvigo.es; Jaume Farrés, Tel: +34-93-5812557 Fax +34-93-5811264, E-mail : jaume.farres@uab.cat

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1. General information

Solvents were dried according to published methods and distilled before use. All other reagents were commercial compounds of the highest purity available. Unless otherwise indicated all reactions were carried out under argon atmosphere, and those not involving aqueous reagents were carried out in oven-dried glassware. Analytical thin layer chromatography (TLC) was performed on aluminum plates with Merck Kieselgel 60F₂₅₄ and visualized by UV irradiation (254 nm) or by staining with an ethanolic solution of phosphomolybdic acid. Flash column chromatography was carried out using Merck Kieselgel 60 (230-400 mesh) C18-SiO₂ (Waters, 55–105 µm 125Å) and CN-SiO₂ (Silicycle 40-63 µm) under pressure. Electron impact (EI) mass spectra were obtained on a Hewlett-Packard HP59970 instrument operating at 70 eV. Alternatively an APEX III FT-ICR MS (Bruker Daltonics), equipped with a 7T actively shielded magnet was used and ions were generated using an Apollo API electrospray ionization (ESI) source, with a voltage between 1800 and 2200 V (to optimize ionization efficiency) applied to the needle, and a counter voltage of 450 V applied to the capillary. For ESI spectra samples were prepared by adding a spray solution of 70:29.9:0.1 (v/v/v) CH₃OH/water/formic acid to a solution of the sample at a v/v ratio of 1 to 5% to give the best signal-to-noise ratio. High Resolution Mass Spectra were taken on a VG Autospec instrument. ¹H NMR spectra were recorded in C₆D₆ and acetone-d₆ at ambient temperature on a Bruker AMX-400 spectrometer at 400 MHz with residual protic solvent as the internal reference.

2. Experimental procedures

Additional observations related to the reactivity of dienylstannanes and dienylsilanes

In our experience with stannyldienals for the synthesis of retinoids, we have observed isomerizations in Stille-Kosugi-Migita cross-coupling reactions. As an example, the non-conjugated iododiene **30** was coupled to δ -stannyldienol, δ -stannyldienal and methyl δ -stannyldienoate (**9a**, **31** and **32**, respectively) to afford the corresponding all-trans-C7,C8-dihydroretinoids (**33-35**) in 31, 83 and 80% yields, respectively. Reaction conditions are also indicated, showing that the δ -stannyldienoate **32** at 80 °C (1h), and the δ -stannyldienal **31** at ambient temperature (25 °C, 30 min). Moreover, despite the mild reaction conditions for the latter, a 11:4 isomer ratio favouring the 13-cis isomer (**36**) was found, thus indicating the incompatibility of the intact polyene structure with the Stille reaction conditions.



As an example of the unsuccessful Hiyama-Denmark cross-coupling of δ silyldienyl esters, the reaction of **26a** with trienyliodide **3** afforded the recovery of the latter and extensive protodesilylation to give the corresponding terminal ethyl (*E*)-3methylpentadienoate **37** as main product.¹



¹ Li, H. X., H.; Zhang, Z.; Xu, Y.; Lu, J.; Gao, L.; Song, Z. Chem. Commun. 2015, 51, 8484.

In contrast, δ -stannyldienyl esters afforded in high yields (50-80%) the corresponding methyl retinoates, as previously reported.²

Synthetic schemes and procedures for the preparation of known dienylstannanes and dienylsilanes

(2E,4E)-5-(Tributylstannyl)-hexa-2,4-dien-1-ol 9b.³





(2E,4E)-2-Methyl-5-(tributylstannyl)-penta-2,4-dien-1-ol 9d.^{5,6}

⁵ Fontán, N.; Vaz, B.; Álvarez, R.; de Lera, A. R. Chem. Commun. 2013, 49, 2694.

² Domínguez, B.; Iglesias, B.; de Lera, A. R. *Tetrahedron* 1999, 55, 15071.

³ Otero, L.; Vaz, B.; Álvarez, R.; de Lera, A. R. Chem. Commun, 2013, 49, 5043.

⁴ Betzer, J. F.; Delaloge, F.; Muller, B.; Pancrazi, A.; Prunet, J. J. Org. Chem. 1997, 62, 7768.

⁶ Evans, D. A.; Gage, J. R.; Leighton, J. L. J. Am. Chem. Soc. 1992, 114, 9434.



(2E,4E)-5-(Tributylstannyl)-penta-2,4-dien-1-ol 9e.⁷



3-(Benzyldimethylsilyl)prop-2-yn-1-ol 20.



To a cooled (10 °C) solution of EtMgBr (29.9 mL, 1 M in THF, 29.94 mmol), a solution of prop-2-yn-1-ol **19** (0.62 g, 11.09 mmol) in THF (11.9 mL) was slowly added. After stirring overnight at 25 °C, the reaction was cooled down to 5 °C, BnMe₂SiCl (5.9 mL, 29.94 mmol) was added and the solution was heated for 2 h at 70 °C. A 1.4 M aqueous solution of H₂SO₄ (38 mL) was added slowly without letting the temperature rise to higher than 40 °C. The mixture was extracted with Et₂O (3x) and the combined organic layers were washed with H₂O (2x) and dried (Na₂SO₄) and the solvent was evaporated. The residue was purified by flash-column chromatography (silica gel, 70:30 hexane/EtOAc) to afford 1.98 g (87%) of a yellow oil, which was identified as 3-(benzyldimethylsilyl)prop-2-yn-1-ol **20**.⁸ **1H-NMR** (400.16 MHz, C₆D₆): δ 7.21-7.13 (m, 3H, ArH), 7.08 – 7.01 (m, 2H, ArH), 3.81 (d, *J* = 6.3 Hz, 2H, 2H₁), 2.12 (s, 2H, SiMe₂CH₂Ph), 0.10 (s, 6H, SiMe₂Bn) ppm. ¹³C-NMR (100.63 MHz, C₆D₆): δ 139.1 (s), 128.7 (d, 2x), 128.6 (d, 2x), 124.9 (d), 106.4 (s), 88.6 (s), 51.2 (t), 26.3 (t), -2.1 (q, 2x) ppm. **IR** (NaCl): v 3500 - 3100 (br, O-H), 2915 (m, C-H), 2854 (m, C-H), 2174 (w, C=C), 1039 (s), 832 (s) cm⁻¹. **MS** (ESI⁺-TOF): *m/z* (%) 205 ([M+H]⁺, 74), 190

⁷ Dias, L. C.; Jr. De Lucca, E. E. J. Org. Chem. 2017, 82, 3019.

⁸ Gallenkamp, D.; Fürstner, A. J. Am. Chem. Soc. 2011, 133, 9232.

(17), 187 (74), 180 (14), 138 (100). **HRMS** (ESI⁺): Calcd. for $C_{12}H_{17}OSi$ ([M+H]⁺), 205.1043; found, 205.1045.

(E)-3-(Benzyldimethylsilyl)-3-methylprop-2-en-1-ol 21a.



To a solution of 3-(benzyldimethylsilyl)-prop-2-yn-1-ol **20** (0.2 g, 0.98 mmol) in Et₂O (0.52 mL), a solution of Red-Al[®] (0.35 mL, 1.25 mmol) in Et₂O (1.6 mL) was slowly added for 1h via addition funnel. Then, the mixture was cooled down to -30 °C and a solution of I₂ (0.5 g, 1.96 mmol) in THF (1 mL) was added. After stirring the resulting mixture for 30 min at 0 °C and for 2h at 25 °C, it was poured into a 10% aqueous solution of HCl and extracted with Et₂O (3x). The combined organic layers were washed with a saturated aqueous solution of Na₂S₂O₃ (3x), brine (3x) and dried (Na₂SO₄) and the solvent was removed to afford 0.294 g of a colorless oil which was used immediately without further purification.

To a cooled (-20 °C) suspension of CuI (0.841 g, 4.41 mmol) in Et₂O (1.18 mL), MeLi (5.52 mL, 1.6 M in Et₂O, 0.88 mmol) was added. After stirring for 1h at -20 °C, a solution of the residue obtained above (0.294 g, 0.88 mmol) in Et₂O (0.59 mL) was added and the mixture was stirred for 14 h at -20 °C and for 4.5 h at 0 °C. A saturated aqueous solution of NH₄Cl was added and the mixture was extracted with Et₂O (3x). The combined organic layers were dried (Na₂SO₄) and the solvent was evaporated. The residue was purified by flash-column chromatography (silica gel, 90:10 hexane/EtOAc) to afford 0.201 g (86%) of a colourless oil, which was identified as (E)-3-(benzyldimethylsilyl)-3-methylprop-2-en-1-ol 21a.⁹ ¹H-NMR (400.16 MHz, C₆D₆): δ 7.30 - 7.22 (m, 2H, ArH), 7.18 - 7.08 (m, 1H, ArH), 7.08 - 7.02 (m, 2H, ArH), 5.95 (t, J = 5.5 Hz, 1H, H₂), 4.10 (d, J = 5.5 Hz, 2H, 2H₁), 2.15 (s, 2H, SiMe₂CH₂Ph), 1.60 (s, 3H, C₃-CH₃), 0.12 (s, 6H, SiMe₂Bn) ppm. ¹³C-NMR (100.63 MHz, C₆D₆): δ 140.6 (d, 2x), 140.0 (s), 135.7 (s), 128.3 (d), 128.3 (d), 124.3 (d, 2x), 59.5 (t), 24.9 (t), 14.8 (q), -4.4 (q) ppm. IR (NaCl): v 3500- 3100 (br, O-H), 2955 (m, C-H), 1599 (m), 1492 (m), 831 (s) cm⁻¹. **HRMS** (ESI⁺): Calcd. for $C_{13}H_{21}OSi$ ([M+H]⁺), 221.1356; found, 221.1364.

(*E*)-3-(Benzyldimethylsilyl)-2-methylprop-2-en-1-ol 21b.



⁹ Anderl, F.; Grobl, S.; Wirtz, C.; Fürstner, A. Angew. Chem. Int. Ed. 2018, 57, 10712.

To a solution of 3-(benzyldimethylsilyl)-prop-2-yn-1-ol **20** (0.29 g, 1.46 mmol) in Et₂O (3.8 mL), CuI (0.03 g, 0.15 mmol) and MeMgCl (3.3 mL, 3 M in THF, 9.88 mmol) were sequentially added. After stirring at 40 °C for 21 h, the reaction was cooled down to 0 °C and an aqueous solution of NH₄Cl/NH₄OH (3.13 mL, 9:1 v/v) was slowly added. The mixture was extracted with Et₂O (3x) and dried (Na₂SO₄) and the solvent was evaporated to afford 0.22 g (68%) of a yellow oil, which was identified as (*E*)-3-(benzyldimethylsilyl)-2-methylprop-2-en-1-ol **21b**.¹⁰ **1H-NMR** (400.16 MHz, C₆D₆): δ 7.20 - 7.12 (m, 3H, ArH), 7.09 - 6.95 (m, 2H, ArH), 5.53 (s, 1H, H₃), 3.67 (s, 2H, 2H₁), 2.12 (s, 2H, SiMe₂CH₂Ph), 1.47 (s, 3H, C₂-CH₃), 0.11 (s, 6H, SiMe₂Bn) ppm. ¹³C-**NMR** (100.63 MHz, C₆D₆): δ 155.1 (s), 140.4 (s), 128.6 (d, 2x), 128.5 (d, 2x), 124.5 (d), 118.9 (d), 68.7 (t), 26.9 (t), 18.6 (q), -1.7 (q, 2x) ppm. **UV** (MeOH): λ_{max} 218, 210 nm. **IR** (NaCl): v 3500 - 3100 (br, O-H), 2957 (m, C-H), 2925 (m, C-H), 1460 (m), 961 (m) cm⁻¹. **MS** (ESI⁺-TOF): *m/z* (%) 221 ([M+H]⁺, 64), 204 (18), 203 (100), 190 (51). **HRMS** (ESI⁺): Calcd. for C₁₃H₂₁OSi ([M+H]⁺), 221.1356; found, 221.1360.

3-(Benzyldimethylsilyl)prop-2-en-1-ol 21c.



To a solution of bis(1,5-cyclooctadiene)rhodium(I) tetrafluoroborate (11 mg, 0.03 mmol) and PPh₃ (14 mg, 0.05 mmol) in acetone (17.3 mL), propargylic alcohol **19** (0.3 g, 5.35 mmol) and benzyldimethylsilane (0.80 g, 5.35 mmol) were sequentially added and the reaction mixture was stirred for 15h at 25 °C. The solvent was evaporated and the residue was purified by flash-column chromatography (silica gel, 95:5 to 90:10 hexane/EtOAc) to afford 0.79 g (72%) of a yellow oil, which was identified as 3-(benzyldimethylsilyl)prop-2-en-1-ol **21c**.¹¹ **1H-NMR** (400.13 MHz, C₆D₆): δ 7.16-7.13 (m, 2H, ArH), 7.05 – 6.99 (m, 1H, ArH), 6.99 – 6.95 (m, 2H, ArH), 5.96 (dt, *J* = 18.8, 3.9 Hz, 1H, H₂), 5.82 (dd, *J* = 18.9, 1.7 Hz, 1H, H₃), 3.81 (br s, 2H, 2H₁), 2.04 (s, 2H, SiMe₂CH₂Ph), 0.02 (s, 6H, SiMe₂Bn) ppm. ¹³C-NMR (100.63 MHz, C₆D₆): δ 147.1 (d), 140.1 (s), 128.6 (d, 2x), 128.5 (d, 2x), 126.6 (d), 124.5 (d), 65.1 (t), 26.2 (t), -3.2 (q, 2x) ppm. IR (NaCl): v 3500 - 3100 (br, O-H), 2955 (m, C-H), 1490 (m, C-H), 837 (m) cm⁻¹. MS (ESI⁺-TOF): *m/z* (%) 208 (3), 207 ([M+H]⁺), 10), 190 (13), 189 (100). HRMS (ESI⁺): Calcd. for C₁₂H₁₉OSi ([M+H]⁺), 207.1198; found, 207.1199.

(E)-3-(Benzyldimethylsilyl)-2-methylacrylaldehyde 22b.

¹⁰ Spino, C.; Grobdout, C. J. Am. Chem. Soc. 2003, 125, 12106.

¹¹ Takeuchi, R.; Nitta, S.; Watamabe, D. J. Org. Chem. 1995, 60, 3045.



*General procedure for the oxidation of alcohols with MnO*₂: To a cooled (0 °C) solution of (*E*)-3-(benzyldimethylsilyl)-2-methylprop-2-en-1-ol **21b** (0.22 g, 0.98 mmol) in CH₂Cl₂ (39 mL), MnO₂ (1.53 g, 17.54 mmol) and Na₂CO₃ (1.87 g, 17.64 mmol) were added and the reaction was stirred at 25 °C for 1.5 h. The mixture was filtered through Celite® and washed with CH₂Cl₂, and the solvent was evaporated to afford 0.16 g (73%) of a yellow oil, which was identified as (*E*)-3-(benzyldimethylsilyl)-2-methylacrylaldehyde **22b**.¹² ¹**H-NMR** (400.16 MHz, C₆D₆): δ 9.25 (s, 1H, H₁), 7.12 (t, *J* = 7.6 Hz, 2H, ArH), 7.04 - 6.94 (m, 1H, ArH), 6.85 (d, *J* = 7.2 Hz, 2H, ArH), 6.15 (s, 1H, H₃), 1.94 (s, 2H, SiMe₂CH₂Ph), 1.68 (s, 3H, C₂-CH₃), -0.05 (s, 6H, SiMe₂Bn) ppm. ¹³C-NMR (100.63 MHz, C₆D₆): δ 194.6 (d), 153.6 (s), 149.3 (d), 139.1 (s), 128.7 (d, 2x), 128.4 (d, 2x), 124.9 (d), 25.6 (t), 12.8 (q), -2.8 (q, 2x) ppm. UV (MeOH): λ_{max} 220 nm. IR (NaCl): v 3024 (w, C-H), 2956 (w, C-H), 2797 (w, C-H), 1693 (s, C=O), 845 (s) cm⁻¹. HRMS (ESI⁺): Calcd. for C₁₃H₁₉OSi ([M+H]⁺), 219.1199; found, 219.1200.

(E)-tert-Butyldimethyl-((3-methylpent-2-en-4-yn-1-yl)oxy)silane 23.



To a solution of imidazole (1.13 g, 16.645 mmol) in DMF (9 mL), a solution of (*E*)-3-methylpent-2-en-4-yn-1-ol **17** (1.00 g, 10.403 mmol) in DMF (9 mL) and then a solution of TBDMSCl (2.04 g, 13.53 mmol) in DMF (9 mL) were sequentially added. After stirring the resulting mixture overnight, water was slowly added and the reaction mixture was extracted with Et₂O (3x). The combined organic layers were dried and the solvent was removed. The residue was purified by flash-column chromatography (silica gel, 97:3 hexane/EtOAc) to afford 1.9 g (91%) of a yellow oil, which was identified as (*E*)-tert-butyldimethyl((3-methylpent-2-en-4-yn-1-yl)oxy)silane **23**.¹³ **1H-NMR** (400.16 MHz, CDCl₃): δ 6.01 (t, J = 6.0 Hz, 1H, H₂), 4.25 (d, J = 6.8 Hz, 2H, 2H₁), 2.79 (s, 1H, H₅), 1.80 (s, 3H, C₃-CH₃), 0.90 (s, 9H, Si(Me₂)/Bu), 0.07 (s, 6H, Si(Me₂)/Bu) ppm. ¹³C-**NMR** (101.63 MHz, CDCl₃): δ 138.7 (d), 117.8 (s), 86.2 (s), 74.7 (d), 60.0 (t), 26.0 (q, 3x), 18.5 (s), 17.5 (q), -5.1 (q, 2x) ppm. **IR (NaCl):** v 2933 (s, C-H), 2890 (s, C-H), 2859 (s, C-H), 1563 (m), 1219 (s), 772 (s) cm⁻¹. **MS** (ESI⁺-TOF): *m/z* (%) 211 (2, [M+H]⁺), 190 (3), 156 (100), 145 (60). **HRMS (ESI⁺):** Calcd. for C₁₂H₂₃OSi ([M+H]⁺), 211.1513; found, 211.1526.

¹² Zhang, Y.; Panek, J. S. Z. Org. Lett. 2007, 9, 3142.

¹³ Betzer, J. F.; Francette, F.; Muller, B.; Pancrazi, A.; Prunet, J. J. Org. Chem. 1997, 62, 7768.

(E)-tert-Butyldimethyl((3-methylhex-2-en-4-yn-1-yl)oxy)silane 24.



To a cooled (-30 °C) solution of (E)-tert-butyldimethyl((3-methylpent-2-en-4yn-1-yl)oxyl)silane 23 (1.48 g, 7.04 mmol) in THF (70.3 mL), n-BuLi (3.12 mL, 2.48 M in hexanes, 7.74 mmol) was added and the mixture was stirred at this temperature for 30 min. MeI (2.19 mL, 35.17 mmol) was then added and the temperature was allowed to rise to 25 °C for 45 min. A saturated aqueous solution of NH₄Cl was added and the mixture was extracted with Et₂O (3x). The combined organic layers were dried and the solvent was removed. After purification by flash-column chromatography (silica gel, 95:5 hexane/EtOAc), 1.18 g (71%) of a yellow oil, which was identified as (E)-tertbutyldimethyl((3-methylhex-2-en-4-yn-1-yl)oxy)silane 24, were obtained.¹¹ ¹H-NMR $(400.16 \text{ MHz}, \text{CDCl}_3)$: δ 5.82 (t, $J = 6.7 \text{ Hz}, 1\text{H}, \text{H}_2$), 4.22 (d, $J = 6.4 \text{ Hz}, 2\text{H}, 2\text{H}_1$), 1.93 (s, 3H, C₅-CH₃), 1.76 (s, 3H, C₃-CH₃), 0.89 (s, 9H, Si(Me₂)/Bu), 0.06 (s, 6H, Si(Me₂)/Bu) ppm. ¹³C-NMR (101.63 MHz, CDCl₃): δ 135.5 (d), 119.3 (s), 83.3 (s), 82.3 (s), 60.1 (t), 26.1 (q, 3x), 18.5 (s), 18.0 (q), 4.3 (q), -5.0 (q, 2x) ppm. IR (NaCl): v 2955 (m, C-H), 2360 (w, C=C), 1706 (s), 1271 (m), 1208 (m), 837 (s) cm⁻¹. MS (ESI⁺-TOF): *m/z* (%) 225 ([M+H]⁺, 5), 215 (3), 203 (3), 185 (7), 174 (3), 156 (100). **HRMS** (ESI⁺): Calcd. for C₁₃H₂₄OSi ([M+H]⁺), 225.1669; found 225.1663.

(E)-Methylhex-2-en-4-yn-1-ol 25.



To a cooled (0 °C) solution of (*E*)-*tert*-butyldimethyl((3-methylhex-2-en-4-yn-1yl)oxyl)silane **24** (1.1 g, 4.01 mmol) in THF (80.5 mL), TBAF (7.4 mL, 1 M in THF, 7.4 mmol) was added and the resulting mixture was stirred for 0.5 h at 25 °C. An aqueous solution of NaHCO₃ was added and the resulting mixture was extracted with Et₂O (3x). The combined organic layers were washed with brine (3x), dried (Na₂SO₄) and the solvent was evaporated. After purification by column chromatography (silica gel, 90:10 hexane/EtOAc), 0.505 g (94%) of a yellow oil, which was identified as (*E*)methylhex-2-en-4-yn-1-ol **25**, were isolated.¹¹ **H-NMR** (400.16 MHz, CDCl₃): δ 5.90 (t, *J* = 7.5 Hz, 1H, H₂), 4.20 (m, 2H, 2H₁), 1.94 (s, 3H, C₃-CH₃), 1.80 (s, C₅-CH₃) ppm. **¹³C-NMR** (101.63 MHz, CDCl₃): δ 134.1 (d), 121.3 (s), 84.1 (s), 82.1 (s), 59.0 (t), 17.8 (q), 4.2 (q) ppm. **IR** (NaCl): v 3362 (br, O-H), 2913 (m, C-H), 1633 (m, C=H), 1435 (m), 1011 (m), 665 (m) cm⁻¹. **HRMS** (ESI⁺): Calcd. for C₇H₁₀NaO ([M+Na]⁺), 133.0623; found, 133.0621.

(2E,4E)-5-(Benzyldimethylsilyl)-3-methylpenta-2,4-dien-1-ol 10a.



To a degassed solution of Pt(DVDS) (0.92 mL, 2% in xylene, 0.04 mmol) and ⁴Bu₃P (0.04 mL, 1M in toluene, 0.04 mmol) in THF (25 mL), HSiMe₂Bn (3.3 mL, 20.6 mmol) was added. After stirring for 30 min at 25 °C, a solution of (E)-3-methylpent-2en-4-yn-1-ol 17 (0.79 g, 8.26 mmol) in THF (25 mL) was added and the reaction mixture was stirred at 25 °C for 1.5 h. The solvent was evaporated and the residue was purified by flash-column chromatography (silica gel, 80:20 hexane/EtOAc) to afford 1.40 g (69%) of a yellow oil, which was identified as (2E, 4E)-5-(benzyldimethylsilyl)-3-methylpenta-2,4-dien-1-ol 10a.¹⁴ ¹H-NMR (400.16 MHz, C₆D₆): δ 7.20 - 7.14 (m, 2H, ArH), 7.08 - 6.94 (m, 3H, ArH), 6.60 (d, J = 19.0 Hz, 1H, H₄), 5.82 (d, J = 19.0 Hz, 1H, H₅), 5.58 (t, J = 6.5 Hz, 1H, H₂), 3.97 (d, J = 6.5 Hz, 2H, 2H₁), 2.09 (s, 2H) SiMe₂CH₂Ph), 1.57 (s, 3H, C₃-CH₃), 0.08 (s, 6H, SiMe₂Bn) ppm. ¹³C-NMR (100.63 MHz, C₆D₆): δ 149.5 (d), 140.1 (s), 136.5 (s), 133.3 (d), 128.6 (d, 2x), 128.5 (d, 2x), 125.7 (d), 124.5 (d), 59.3 (t), 26.4 (t), 12.1 (q), -3.1 (q, 2x) ppm. UV (MeOH): λ_{max} 243 nm. IR (NaCl): v 3500- 3100 (br, O-H), 3023 (m, C-H), 2954 (m, C-H), 1578 (m), 1492 (m), 1246 (m), 1152 (m) cm⁻¹. **HRMS** (ESI⁺): Calcd. for $C_{15}H_{23}OSi$ ([M+H]⁺), 247.1513; found, 247.1512.

¹⁴ Bergueiro, J.; Montenegro, J.; F.; Saá, C.; López, S. Chem. Eur. J. 2012, 18, 14100.

All-trans-retinol 1a.



Spectroscopic data for all-*trans*-retinol **1a**.¹³ **¹H-NMR** (400.16 MHz, (CD₃)₂CO): δ 6.66 (dd, J = 15.2, 11.3 Hz, 1H, H₁₁), 6.34 (d, J = 15.3 Hz, 1H, H₁₂), 6.27 – 6.08 (m, 3H, H₇ + H₈ + H₁₀), 5.70 (t, J = 6.5 Hz, 1H, H₁₄), 4.25 (d, J = 6.5 Hz, 2H, 2H₁₅), 2.04 (t, J = 6.6 Hz, 2H, 2H₄), 1.98 (s, 3H, C₁₃-CH₃), 1.84 (s, 3H, C₉-CH₃), 1.72 (s, 3H, C₅-CH₃), 1.68 – 1.59 (m, 2H, 2H₃), 1.53 – 1.44 (m, 2H, 2H₂), 1.05 (s, 6H, C₁-(CH₃)₂) ppm. ¹³C-NMR (101.63 MHz, (CD₃)₂CO): δ 139.0 (d), 138.8 (s), 138.2 (d), 136.1 (s), 135.7 (s), 133.8 (d), 131.7 (d), 129.6 (s), 127.0 (d), 125.2 (d), 59.5 (t), 40.5 (t), 35.0 (s), 33.7 (t), 29.4 (q, 2x), 22.1 (q), 20.1 (t), 12.8 (q) ppm. UV (MeOH): λ_{max} (ϵ) 378 (14 800) nm. IR (NaCl): v 3500 - 3100 (br, O-H), 2925 (m, C-H), 286.2304; found, 286.2305.

3. Expression and purification of AKR1B enzymes

AKR1B1 and AKR1B10 enzymes were expressed and purified from pET-15b expression vector. The pET-15b constructs allow the expression of proteins with an N-terminal fused histidine tag under the control of phage T7 RNA polymerase promoter and lac operon. For protein expression, transformed E. coli BL21(DE3) pLysS strains were grown in 1L of 2xYT medium in the presence of 33 μ g/mL ampicillin and 50 μ g/mL chloramphenicol, and cells were incubated at 37°C until O.D.595 = 0.6 was reached. Protein expression was then induced by the addition of 1 mM IPTG and cells were grown overnight at 22°C. AKR1B enzymes were purified as described previously for AKR1B10. ¹⁵

4. HPLC enzymatic activity assay

¹⁵ Gallego, O.; Belyaeva, Olga V.; Porté, S.; Ruiz, F. X.; Stetsenko, Anton V.; Shabrova, Elena V.; Kostereva, Natalia V.; Farrés, J.; Parés, X.; Kedishvili, Natalia Y. *Biochem. J.* **2006**, *399*, 101.

Activity assays with the C11 to C14-methyl shifted retinal analogues were carried out using an HPLC-based methodology.¹⁷ Briefly, the stock solutions of substrates prepared in ethanol were solubilized using glass tubes by a 10-min sonication at molar ratio 1:1 with fatty acid-free bovine serum albumin in 100 mM potassium chloride, pH 7.4.¹⁷ The actual concentration of solubilized compound was determined based on the corresponding molar absorption coefficient determined in Figure 1: ε_{383} = 14 500 M⁻¹·cm⁻¹ for **2b**, $\varepsilon_{395} = 22$ 400 M⁻¹·cm⁻¹ for **2c**, $\varepsilon_{390} = 15$ 500 M⁻¹·cm⁻¹ for **2d** and $\varepsilon_{390} = 14\ 200\ M^{-1} \cdot cm^{-1}$ for **2f**. The enzymatic reaction was carried out for 15 min at 37 °C in a final volume of 0.5 mL. To measure the steady-state enzymatic activity, the concentration of enzyme was kept from 25- to 100-fold lower than that of the substrate for all the enzymatic assays. The reactions were stopped by the addition of 1 mL of cold methanol and, after two rounds of extraction with hexane and the subsequent evaporation of the organic phase under a N₂ stream, C11 to C14-methyl shifted vitamin A analogues were analysed by HPLC. Briefly, these compounds were dissolved in 200 µL of hexane and injected on a Nova Pak Silica column (4 µm, 3.9 x 150 mm, Waters) as previously described.¹⁶ Quantification of C11 to C14-methyl shifted vitamin A analogues (aldehyde substrates 2b, 2c, 2d and 2f, and alcohol products 1b, 1c, 1d and 1f) was performed by interpolation of HPLC peak areas into a calibration curve of known compound concentrations. The specific activity of AKR1B1 and AKR1B10 enzymes was obtained from the product appearance while the substrate conversion was controlled to assure the optimal enzymatic conditions regarding to initial linear rate (V_0) . The kinetic constants were calculated using the non-linear regression program GraFit 5.0 (Erithacus software) and expressed as the mean \pm standard error from three independent determinations.



¹⁶ Domínguez, M.; Pequerul, R.; Alvarez, R.; Giménez-Dejoz, J.; Birta, E.; Porté, S.; Rühl, R.; Parés, X.; Farrés, J.; de Lera, A. R. *Tetrahedron* **2018**, *74*, 2567.

Figure 1. Linear regression plots to determine the molar extinction coefficients for compounds **2b**, **2c**, **2d** and **2f**. The compound concentrations tested ranged $5 - 40 \mu$ M.

5. NMR Spectra

¹H-NMR (400.16 MHz, C₆D₆) of 9c



7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 (ppm)

¹³C-NMR (100.63 MHz, C₆D₆) of 9c



¹H-NMR (400.16 MHz, C₆D₆) of 16c



¹³C-NMR (100.63 MHz, C₆D₆) of 16c





¹³C-NMR (100.63 MHz, C₆D₆) of 20











13 C-NMR (100.63 MHz, C₆D₆) of 21b

















¹H-NMR (400.16 MHz, $CDCl_3$) of 23



¹³C-NMR (100.63 MHz, CDCl₃) of 23









7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 f1 (ppm)



¹³C-NMR (100.63 MHz, CDCl₃) of 25

34

¹H-NMR (400.16 MHz, C₆D₆), spectrum of 28a.




¹H-NMR (400.16 MHz, C₆D₆) of 26b





¹H-NMR (400.16 MHz, C₆D₆) of 26c





¹H-NMR (400.16 MHz, C_6D_6) of 26d





¹³C-NMR (100.63 MHz, C₆D₆) of 26d









¹³C-NMR (100.63 MHz, C₆D₆) of 26e



¹H-NMR (400.16 MHz, C₆D₆) of 27

¹³C-NMR (100.63 MHz, C₆D₆) of 27





¹³C-NMR (100.63 MHz, C₆D₆) of 10a







¹H-NMR (400.16 MHz, C₆D₆) of 10c



¹³C-NMR (100.63 MHz, C₆D₆), spectrum of 10c.



¹H-NMR (400.16 MHz, C_6D_6) of 10d





¹³C-NMR (100.63 MHz, C₆D₆) of 10d















¹³C-NMR (100.63 MHz, C₆D₆) of 10f



¹H-NMR (400.16 MHz, (CD₃)₂CO) of 1a



7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 0.6 ppm









7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 0.6 f1 (ppm)









¹³C-NMR (100.63 MHz, C₆D₆) of 2b





6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 0.6 ppm

¹³C-NMR (100.63 MHz, C₆D₆) of 1c



¹H-NMR (400.16 MHz, C₆D₆) of 2c



¹H-NMR (400.16 MHz, C₆D₆) of 1d



7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 0.6 f1 (ppm)

¹³C-NMR (100.63 MHz, C₆D₆) of 1d




¹H-NMR (400.16 MHz, C₆D₆) of 1e











¹³C-NMR (100.63 MHz, C₆D₆) of 2e





7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0. f1 (ppm)











¹³C-NMR (100.63 MHz, C₆D₆) of 2f