Supplementary Information

A light- and heat-driven glycal diazidation approach to nitrogenous carbohydrate derivatives with antiviral activity

Huan He,^{‡a} Ruiyuan Cao,^{‡b} Ruidi Cao,^a Xiao-Yu Liu,^{*a} Wei Li,^b Di Yu,^a Yuexiang Li,^b Miaomiao Liu,^b Yanmei Wu,^a Pingzhou Wu,^a Jin-Song Yang,^a Yunzheng Yan,^b Jingjing Yang,^b Zhi-Bing Zheng,^b Wu Zhong,^{*b} and Yong Qin^{*a}

- Key Laboratory of Drug-Targeting and Drug Delivery System of the Education Ministry and Sichuan Province, Sichuan Engineering Laboratory for Plant-Sourced Drug and Sichuan Research Center for Drug Precision Industrial Technology, West China School of Pharmacy, Sichuan University, Chengdu 610041, China E-mail: xyliu@scu.edu.cn (X.-Y.L.); yongqin@scu.edu.cn (Y.Q.)
- National Engineering Research Center for the Emergence Drugs Beijing Institute
- b. National Engineering Research Center for the Emergence Drugs, Beijing Institute of Pharmacology and Toxicology, Beijing 100850, China E-mail: zhongwu@bmi.ac.cn (W.Z.)

[‡] These authors contributed equally to this work.

Table of Contents

1.	General Information	S3
2.	Optimization Studies	S4
3.	Substrate Preparation	S6
4.	General Procedure and Product Characterization	S13
5.	Product Interconversion and Stereochemistry Determination	S36
6.	Product Diversification	S47
7.	Mechanistic Investigations	S58
8.	Antiviral Activities	S65
9.	References	S67
10.	X-Ray Crystallographic Data	S68
11.	NMR Spectra	S81

1. General Information

All reactions that require anhydrous conditions were performed in flame-dried glassware under argon atmosphere and all reagents were purchased from commercial suppliers. Solvent purification was conducted according to Purification of Laboratory Chemicals 2nd edn (Perrin, D. D., Armarego, W. L. F. and Perrin, D. R., Pergamon Press: Oxford, 1980). The products were purified by flash column chromatography on silica gel (200 - 300 meshes) from the Anhui Liangchen Silicon Material Company (China). Reactions were monitored by thin layer chromatography (TLC, 0.2 mm, HSGF254) supplied by Yantai Chemicals (China). Visualization was accomplished with UV light, exposure to iodine, stained with ethanolic solution of H₂SO₄/EtOH (1:9, v/v) or basic solution of KMnO₄. ¹H NMR and ¹³C NMR spectra were recorded on Varian INOVA-400/54 and Agilent DD2-600/54 spectrometer, in the following solvents (reference peaks include ¹H and ¹³C NMR): CDCl₃ (¹H NMR: 7.26 ppm; ¹³C NMR: 77.00 ppm), CD₃OD (¹H NMR: 3.31 ppm; ¹³C NMR: 49.00 ppm), D₂O (¹H NMR: 4.79 ppm). The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, br = broad, td = triple doublet, dt = double triplet, m = multiplet, and coupling constants (J) were reported in Hertz (Hz). Infrared (IR) spectra were recorded on a Perkin Elmer Spectrum Two FTIR spectrometer. The specific optical rotation was obtained from Rudolph Research Analytical Autopol VI automatic polarimeter. High-resolution mass spectra (HRMS) were recorded on Bruker Apex IV FTMS or Thermo Scientific LTQ Orbitrap XL ESI mass spectrometers. Melting points were obtained with X-4 precision micro melting point apparatus. Kessil H150BLUE LED and Kessil A160WE TUNA BLUE were purchased from http://www.amazon.com.

2. Optimization Studies

Table S1. Effect of oxidant

AcQ OAc AcO O AcO 7a	oxidant (2.0 CO ₂ Me dry MeCN	equiv), TMSN ₃ (I (0.1 M), 8 W blu	3.0 equiv) e LEDs	DAc CO ₂ Me N _{3N3}
Entry ^a	Oxidant	Time	Conversion ^b	Yield ^c
1	O_2^{d}	15 h	trace	0
2	BQ	15 h	0	0
3	$K_2S_2O_8$	15 h	0	0
4	PhIO	15 h	53%	46%
5	BI-OAc ^e	10 h	100%	69%
6	BI-OH ^e	15 h	87%	60%
7	PIDA	15 h	87%	66%
8	PIFA	15 h	77%	63%
9	NaIO ₄	12 h	100%	33%
10	IBX	15 h	33%	25%

^{*a*}Reactions were conducted on 0.075 mmol scale unless otherwise stated. ^{*b*}Conversions were calculated based on the recovered starting glycal **7a** after column chromatography. ^{*c*}Yields were determined according to the isolated material via column chromatography. ^{*d*}The reaction was conducted under a balloon of oxygen; ^{*e*}BI-OAc and BI-OH were prepared according to the literature method.¹

Table S2. Effect of light

AcQ OA AcO AcO 7a	CO ₂ Me dry MeC	quiv), TMSN ₃ (3.0 CN (0.1 M), light	AcO AcO AcO AcO	OAC CO ₂ Me N _{3N3}
Entry ^a	Light	Time	Conversion ^b	Yield ^c
1	26 W CFL	15 h	87%	14%
2	15 W green LEDs	15 h	63%	50%
3	30 W white LEDs	6 h	100%	77%
4	34 W blue LEDs	1.5 h	100%	83%
5	sun light	1.5 h	33%	25%
6	sun light	15 h	67%	41%

"Reactions were conducted on 0.075 mmol scale unless otherwise stated. "Conversions were calculated

based on the recovered starting glycal **7a** after column chromatography. ^{*c*}Yields were determined according to the isolated material via column chromatography.

AcQ OAc AcO O AcO 7a	BI-OAc (2.0 CO ₂ Me solvent (equiv), TMSN ₃ (0.1 M), 34 W blue	3.0 equiv) AcO AcO	OAc CO ₂ Me N _{3N3}
Entry ^a	Solvent	Time	Conversion ^b	Yield ^c
1	MeCN	1.5 h	100%	83%
2	CH_2Cl_2	3 h	100%	36%
3	THF	11 h	<5%	0%
4	DMF	11 h	0%	0%
5	PhMe	11 h	<10%	<5%
6	DMSO	11 h	100%	62%
7	Dioxane	11 h	67%	41%
8	MeOH	11 h	<10%	0%
9	Acetone	3 h	100%	63%

Table S3. Effect of solvent

"Reactions were conducted on 0.075 mmol scale unless otherwise stated. ^bConversions were calculated based on the recovered starting glycal **7a** after column chromatography. ^cYields were determined according to the isolated material via column chromatography.

Table S4. Effect of reagent equivalent

AcQ AcO AcO	OAC OCO2Me Me	BI-OAc , TMSN ₃ CN (0.1 M), 34 W blue LED	AcO OAc AcO AcO	-CO ₂ Me
Entry ^a	Equiv of BI-OAc	Equiv of TMSN ₃	Reaction Time	Yield ^b
1	1	3	20 h	55%
2	2	3	1.5 h	83%
3	3	3	2 h	74%
4	4	3	2 h	65%
5	5	3	2 h	58%
6	2	2	3 h	62%
7	2	4	1.5 h	78%
8	2	5	1.5 h	71%

"Reactions were conducted on 0.075 mmol scale unless otherwise stated. "Yields were determined

according to the isolated material via column chromatography.

A Act AcO	CO ₂ Me BI-OAc (2.0 equiv), TMSN MeCN (0.1 M), 34 W b	I ₃ (3.0 equiv) ► lue LEDs	AcO OAc AcO CO AcO X CO N _{3N3}	_{'2} Me
Entry ^a	Variation from standard conditions	Time	Conversion ^b	Yield ^c
1	none	1.5 h	100%	83%
2	dark	1.5 h	17%	13%
3	dark	15 h	33%	11%
4	no BI-OAc	10 h	0%	0%

Table S5. Control experiments

"Reactions were conducted on 0.075 mmol scale unless otherwise stated. ^bConversions were calculated based on the recovered starting glycal **7a** after column chromatography. ^cYields were determined according to the isolated material via column chromatography.

Table S6. Heating experiments

AcQ OA AcO CO AcO 7a	CO ₂ Me MeC	equiv), TMSN ₃ (3. CN (0.1 M), dark	0 equiv) AcO	DAC O CO ₂ Me N _{3N3}
Entry ^a	Temperature ^b	Time	Conversion ^c	Yield ^d
1	30 °C	15 h	67%	44%
2	40 °C	15 h	83%	65%
3	50 °C	15 h	83%	55%
4	60 °C	15 h	87%	47%

^{*a*}Reactions were conducted on 0.075 mmol scale unless otherwise stated. ^{*b*}The ambient temperature of reactor (heating with oil bath). ^{*c*}Conversions were calculated based on the recovered starting glycal **7a** after column chromatography. ^{*d*}Yields were determined according to the isolated material via column chromatography.

3. Substrate Preparation

Substrates $7a^2$, $7g^3$, and $7m^4$ were prepared according to the literature methods.



Sodium methoxide (492 mg, 9.10 mmol, 0.3 equiv) was added to a solution of compound **7a** (12.2 g, 30.3 mmol, 1.0 equiv) in MeOH (100 mL) at room temperature. After the reaction was stirred for 30 min, it was carefully neutralized with Amberlite IR 120 (H⁺) resin, followed by filtration and concentration. The residue was purified through flash column chromatography (CH₂Cl₂/MeOH = 9:1, v/v) to give **S1** (4.69 g, 66%) as a white amorphous powder. **TLC**: (CH₂Cl₂/MeOH = 9:1, v/v), R_f = 0.13; ¹**H NMR** (400 MHz, CD₃OD) δ 5.84 (t, J = 2.0 Hz, 1H), 4.49 – 4.45 (m, 1H), 4.16 – 4.12 (m, 1H), 3.99 – 3.93 (m, 1H), 3.90 – 3.81 (m, 2H), 3.76 (s, 3H), 3.72 (dd, J = 11.6, 5.2 Hz, 1H); ¹³C **NMR** (100 MHz, CD₃OD) δ 164.3, 144.6, 113.6, 78.6, 70.5, 66.4, 64.3, 64.1, 52.6. [**a**]_D²⁵ = -27.5 (*c* 0.38, CHCl₃/MeOH = 1:3, v/v); **IR** (neat): $v_{max} = 3348$, 2924, 1718, 1648, 1440, 1260, 1154, 1109, 1066, 1021, 863, 844 cm⁻¹; **HRMS (ESI)**: calcd. for C₉H₁₄NaO₇ [M + Na]⁺ *m*/z 257.0637; found *m*/z 257.0637.



Under argon, to a stirred solution of **S1** (4.69 g, 20.0 mmol, 1.0 equiv) in DMF (80 mL) at room temperature were added 2,2-dimethoxypropane (7.41 mL, 60.1 mmol, 3.0 equiv) and catalytic *p*-toluenesulfonic acid monohydrate (382 mg, 2.00 mmol, 0.1 equiv). After the resulting mixture was stirred for 4 h at the same temperature, Et₃N was added to quench the reaction. Concentration of the mixture and purification of the crude product by column chromatography on silica gel (petroleum ether/EtOAc = 9:1 to 1:1, v/v) afforded **7e** (1.01 g, 16%) and **S2** (4.18 g, 76%), respectively. The spectroscopic data of **7e** were in good agreement with those reported in the literature.⁵ Compound **S2**: **TLC**: (petroleum ether/EtOAc = 1:1, v/v), $R_f = 0.20$; ¹H NMR (400 MHz, CDCl₃) δ 5.93 (t, *J* = 2.0 Hz, 1H), 4.55 – 4.43 (m, 2H), 4.23 – 4.14 (m, 2H), 4.06 (dd, *J* = 9.0, 4.8 Hz, 1H), 3.82 – 3.80 (m, 1H), 3.79 (s, 3H), 2.75 – 2.55 (m, 2H), 1.44 (s, 3H), 1.38 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 162.2, 143.6, 112.0, 109.9, 77.8, 73.3, 66.8, 64.8, 63.7, 52.4, 26.8, 25.0; [α]_D²⁵ = -14.6 (*c* 0.23, CHCl₃); **IR** (neat): *v*_{max} = 3451, 2988, 2949, 2934, 1729, 1648, 1372, 1260, 1150, 1077, 1058, 1026, 841, 752 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₂H₁₈NaO₇ [M + Na]⁺ *m/z* 297.0950; found *m/z* 297.0946.



A 100 mL round-bottom flask was charged with compound **S2** (3.68 g, 13.4 mmol, 1.0 equiv) in pyridine (60 mL), diphosgene (3.24 mL, 26.9 mmol, 2.0 equiv) was dropwise added at 0 °C. After being stirred for 30 min at room temperature, the reaction was quenched by water and the mixture was extracted with EtOAc for three times. The combined organic layers were washed with brine and dried over Na₂SO₄, filtered and concentrated. Subjection of the residue to column chromatography on silica gel with petroleum ether/EtOAc (8:1, v/v) yielded **7b** (2.46 g, 61%) as a white amorphous powder. **TLC**: (petroleum ether/EtOAc = 1:1, v/v), R_f = 0.55; ¹**H NMR** (400 MHz, CDCl₃) δ 6.09 (d, *J* = 3.2 Hz, 1H), 5.32 (dd, *J* = 7.6, 3.6 Hz, 1H), 5.07 (d, *J* = 7.6 Hz, 1H), 4.45 – 4.40 (m, 1H), 4.23 – 4.14 (m, 2H), 3.82 (s, 3H), 3.78 (d, *J* = 8.8 Hz, 1H), 1.42 (s, 3H), 1.37 (s, 3H); ¹³C **NMR** (150 MHz, CDCl₃) δ 161.2, 153.2, 147.6, 110.2, 104.8, 75.6, 72.7, 72.2, 68.6, 66.7, 52.8, 26.9, 24.9; $[\alpha]_D^{25} = +1.0$ (*c* 0.40, CHCl₃); **IR** (neat): $v_{max} = 2988$, 2959, 1800, 1738, 1655, 1373, 1257, 1164, 1013, 841, 769 cm⁻¹; **HRMS** (**ESI**): calcd. for C₁₃H₁₆NaO₈ [M + Na]⁺ *m/z* 323.0743; found *m/z* 323.0753.



p-Toluenesulfonic acid monohydrate (951 mg, 5.00 mmol, 3.0 equiv) was added to a solution of compound **7b** (500 mg, 1.67 mmol, 1.0 equiv) in MeOH (17 mL) at 0 °C. The mixture was warmed to room temperature and stirred for 6 h. After quench of the reaction with saturated aqueous NaHCO₃, the aqueous phase was extracted with EtOAc for three times; the combined organic layers were washed with brine, dried over Na₂SO₄ and concentrated in *vacuo*. The crude product, dissolved in CH₂Cl₂ (17 mL) and pyridine (1.0 mL), was used directly in the next step. Benzoyl chloride (0.967 mL, 8.33 mmol, 5.0 equiv) was slowly added to the above solution at 0 °C and then the reaction mixture was stirred at room temperature. After disappearance of the crude product as monitored by TLC, the mixture was diluted with CH₂Cl₂, and washed with 0.5% HCl aqueous solution for three times. Next, the organic phase was washed with saturated aqueous NaHCO₃, dried over Na₂SO₄ and evaporated under reduced pressure to give a residue. Further purification by flash column chromatography (petroleum ether/EtOAc = 3:1, v/v) furnished **7c** (485 mg, 62% over two steps) as white

amorphous powder. **TLC**: (petroleum ether/EtOAc = 1:1, v/v), $R_f = 0.38$; ¹**H NMR** (400 MHz, CDCl₃) δ 8.00 (t, J = 8.4 Hz, 4H), 7.61 – 7.55 (m, 2H), 7.43 (q, J = 7.2 Hz, 4H), 6.17 (d, J = 3.2 Hz, 1H), 5.81 – 5.77 (m, 1H), 5.37 (dd, J = 7.6, 3.6 Hz, 1H), 5.09 (d, J = 8 Hz, 1H), 5.00 (dd, J = 12.8, 2.4 Hz, 1H), 4.78 (dd, J = 12.8, 4.8 Hz, 1H), 4.45 (d, J = 8 Hz, 1H), 3.83 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 165.9, 164.8, 161.0, 152.8, 147.9, 133.7, 133.3, 129.8, 129.6, 129.6, 128.9, 128.6, 128.5, 104.9, 73.4, 71.7, 69.7, 68.6, 62.0, 52.9; $[\alpha]_D^{25} = -67.1$ (*c* 0.17, CHCl₃); **IR** (neat): $v_{max} = 3056$, 2958, 2325, 1814, 1728, 1657, 1264, 1095, 1016, 732, 703 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₄H₂₀NaO₁₀ [M + Na]⁺ *m/z* 491.0954; found *m/z* 491.0952.



To a stirred solution of **S2** (500 mg, 1.82 mmol, 1.0 equiv) in DMF (10 mL) at room temperature under argon were added benzaldehyde dimethyl acetal (1.31 mL, 8.76 mmol, 4.8 equiv) and catalytic *p*-toluenesulfonic acid monohydrate (34.7 mg, 0.182 mmol, 0.1 equiv). After the resulting mixture was stirred overnight at the same temperature, Et₃N was added to quench the reaction. Subsequently, the mixture was concentrated in *vacuo* to afford a residue, which was purified by silical gel column chromatography (petroleum ether/EtOAc = 20:1, v/v) to give **7d** (482 mg, 73%) as colorless oil. **TLC**: (petroleum ether/EtOAc = 8:1, v/v), R_f = 0.50; ¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.46 (m, 2H), 7.37 (t, *J* = 3.2 Hz, 3H), 6.08 (dd, *J* = 3.2, 0.8 Hz, 1H), 5.89 (s, 1H), 5.16 (dd, *J* = 6.4, 3.6 Hz, 1H), 4.62 (d, *J* = 6.4 Hz, 1H), 4.52 – 4.48 (m, 1H), 4.22 – 4.16 (m, 2 H), 3.86 – 3.85 (m, 1H), 3.84 (s, 3H), 1.45 (s, 3H), 1.37 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 162.1, 147.4, 136.9, 129.5, 128.3, 126.7, 126.6, 109.6, 107.7, 103.0, 77.7, 73.9, 71.9, 69.7, 66.6, 52.5, 29.6, 26.8, 25.2; [*a*]_D²⁵ = -3.8 (*c* 0.14, CHCl₃); **IR** (neat): $v_{max} = 2955$, 1736, 1650, 1438, 1373, 1259, 1220, 1100, 1062, 842, 759 cm⁻¹; **HRMS (ESI**): calcd. for C₁₉H₂₂NaO₇[M + Na]⁺ *m/z* 385.1263; found *m/z* 385.1262.



Sodium periodate (817 mg, 3.82 mmol, 1.6 equiv) was added to a solution of compound S4⁵ (654 mg, 2.39 mmol, 1.0 equiv) in a mixture of THF (16 mL) and water (8 mL) at 0 °C. After being stirred for 3 h at room temperature, the reaction was extracted with EtOAc for three

times. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was dissolved in 10 mL MeOH, to which was added sodium borohydride (110 mg, 2.86 mmol, 1.2 equiv) at 0 °C. After 30 min, water was added to quench the reaction and MeOH was removed under reduced pressure. The mixture was subsequently extacted with EtOAc for three times, and the combined organic layers were washed with brine, dried, and concentrated to give the crude product. It was directly dissolved in dry CH₂Cl₂ (6 mL), then pyridine (0.5 mL) and benzoyl chloride (0.831 mL, 7.16 mmol, 3.0 equiv) were added slowly at 0 °C under an argon atmosphere. The reaction mixture was stirred at room temperature until complete consumption of the starting alcohol as monitored by TLC analysis. The mixture was diluted with CH₂Cl₂ and washed with 0.5% aqueous HCl solution for three times. The organic phase was washed with saturated aqueous NaHCO₃, dried over Na₂SO₄, filtered and concentrated in *vacuo*. Purification of the residue via flash column chromatography (petroleum ether/EtOAc = 9:1, v/v) afforded compound 7f (166 mg, 20% over three steps). TLC: (petroleum ether/EtOAc = 4:1, v/v), $R_f = 0.65$; ¹H NMR (400 MHz, CDCl₃) δ 8.08 (d, J = 7.2 Hz, 2H), 7.58 (t, J = 7.2 Hz, 1H), 7.45 (t, J = 8.0 Hz, 2H), 6.05 (d, J = 2.0 Hz, 1H), 4.82 (dd, J = 6.0, 3.2 Hz, 1H), 4.76 - 4.67 (m, 2H), 4.41 (d, J = 6.0)Hz, 1H), 4.36 (t, J = 6.8 Hz, 1H), 3.81 (s, 3H), 1.41 (s, 3H), 1.38 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 166.3, 162.5, 144.0, 133.2, 129.8, 128.4, 111.3, 110.1, 74.1, 71.7, 68.8, 63.8, 52.4, 27.9, 26.5; $[\alpha]_D^{25} = +13.6$ (c 0.12, CHCl₃); **IR** (neat): $v_{max} = 2919$, 1725, 1653, 1451, 1245, 1112, 1028, 712 cm⁻¹; **HRMS (ESI)**: calcd. for $C_{18}H_{20}NaO_7 [M + Na]^+ m/z$ 371.1107; found *m*/*z* 371.1104.



Potassium carbonate (1.46 g, 10.6 mmol, 0.5 equiv) was added to a solution of $7g^3$ (10.0 g, 21.1 mmol, 1.0 equiv) in MeOH (200 mL) at room temperature. After being stirred for 30 min, the mixture was neutralized with Amberlite IR 120 (H⁺) resin, followed by filtration and concentration. The residue, benzoic anhydride (47.8 g, 211 mmol, 10.0 equiv) and 4-dimethylaminopyridine (517 mg, 4.23 mmol, 0.2 equiv) were placed in a 200 mL round bottom flask, followed by addition of dry pyridine (100 mL). After being stirred overnight at 80 °C, the reaction mixture was concentrated in *vacuo*. Flash column chromatography of the residue on silica gel (petroleum ether/EtOAc = 4:1 to 2:1, v/v) gave **7h** (13.1 g, 85%) as a white amorphous powder. **TLC**: (petroleum ether/EtOAc = 1:1, v/v), $R_f = 0.48$; ¹H **NMR**

(400 MHz, CDCl₃) δ 8.10 (d, J = 7.2 Hz 2H), 8.01 – 7.92 (m, 6H), 7.64 – 7.44 (m, 6H), 7.43 – 7.28 (m, 6H), 6.14 (d, J = 3.2 Hz, 1H), 6.11 – 6.06 (m, 1H), 6.05 – 5.90 (m, 2H), 5.85 – 5.75 (m, 1H), 5.16 (dd, J = 12.4, 3.6 Hz, 1H), 4.96 (dd, J = 9.6, 3.2 Hz, 1H), 4.64 (dd, J = 12.0, 6.4 Hz, 1H), 4.39 (q, J = 8.8 Hz, 1H), 3.76 (s, 3H), 1.89 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.2, 170.2, 166.3, 166.1, 165.6, 165.5, 161.7, 145.4, 133.5, 133.3, 133.3, 133.0, 123.0, 129.8, 129.8, 129.7, 129.4, 129.2, 129.1, 128.5, 128.4, 128.4, 128.3, 108.4, 108.3, 76.5, 71.4, 68.9, 68.8, 62.6, 52.4, 47.9, 23.1; $[\alpha]_D^{25}$ = +133.6 (*c* 1.47, CHCl₃); **IR** (neat): v_{max} = 3269, 3065, 2956, 1719, 1248, 1092, 1068, 1025, 708 cm⁻¹; **HRMS (ESI)**: calcd. for C₄₀H₃₅NNaO₁₂ [M + Na]⁺ *m/z* 744.2057; found *m/z* 744.2048.



Under an argon atmosphere, 2,6-lutidine (2.10 mL, 17.4 mmol, 10.0 equiv) and trifluoromethanesulfonic acid *tert*-butyldimethylsilyl ester (2.79 mL, 12.2 mmol, 7.0 equiv) were successively added to a solution of compound **S5**⁶ (600 mg, 1.74 mmol, 1.0 equiv) in dry CH₂Cl₂ (10 mL). The reaction was stirred overnight, before it was quenched by adding water. The resultant mixture was extracted with EtOAc for three times, the combined organic layers were dried over Na₂SO₄, filtered and concentrated. Subjection of the residue to flash column chromatography (petroleum ether/EtOAc = 5:1, v/v) gave **7i** (902 mg, 90%). **TLC**: (petroleum ether/EtOAc = 3:1, v/v), R_f = 0.30; ¹**H NMR** (400 MHz, CDCl₃) δ 5.97 (d, *J* = 4.4 Hz, 1H), 5.57 (d, *J* = 7.6 Hz, 1H), 4.46 (dd, *J* = 6.8, 2.8 Hz, 1H), 4.37 – 4.27 (m, 2H), 4.22 (t, *J* = 5.6 Hz, 1H), 4.03 – 3.92 (m, 2H), 3.88 (t, *J* = 8.0 Hz, 1H), 3.81 (s, 3H), 1.95 (s, 3H), 1.41 (s, 3H), 1.30 (s, 3H), 0.90 (s, 18H), 0.13 (s, 6H), 0.07 (s, 3H), -0.01 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 169.6, 162.7, 142.8, 110.1, 108.3, 79.4, 75.8, 68.3, 64.0, 63.8, 52.2, 50.3, 26.4, 25.9, 25.7, 25.3, 23.3, 18.2, 17.9, -3.6, -4.6, -4.9, -4.9; **[a]**_D²⁵ = +52.1 (*c* 0.73, CHCl₃); **IR** (neat): $v_{max} = 3271$, 2953, 2930, 2887, 2857, 1738, 1649, 1257, 1063, 834, 777, 756 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₇H₅₁NNaO₈Si₂[M + Na]⁺ *m*/z 596.3051; found *m*/z 596.3083.



Potassium carbonate (68.3 mg, 0.494 mmol, 0.5 equiv) was added to a solution of S6⁷ (520 mg, 0.988 mmol, 1.0 equiv) in MeOH (10 mL). After being stirred for 15 min, the mixture was neutralized with Amberlite IR 120 (H⁺) resin, then filtered and concentrated to give a white product, which was dissolved in 10 mL anhydrous pyridine. Next, to the solution were added benzoic anhydride (2.24 g, 9.88 mmol, 10.0 equiv) and 4-dimethylaminopyridine (24.2 mg, 0.198 mmol, 0.2 equiv). The reaction mixture was allowed to stir at 80 °C for 5 h, before removal of the solvent in vacuo. Flash column chromatography of the residue on silica gel (petroleum ether/EtOAc = 1:1, v/v) yielded 7j (590 mg, 84%) as a white amorphous powder. **TLC**: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.65$; ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, J = 7.2 Hz, 2H), 7.96 – 7.89 (m, 4H), 7.74 (d, J = 8.8 Hz, 1H), 7.60 – 7.29 (m, 9H), 6.35 (d, J= 8.4 Hz, 1H), 6.03 (d, J = 4.8 Hz, 2H), 5.94 - 5.87 (m, 1H), 5.23 (dd, J = 12.4, 3.2 Hz, 1H), 4.97 - 4.88 (m, 1H), 4.71 - 4.60 (m, 2H), 4.45 - 4.35 (m, 1H), 3.72 (s, 3H), 1.91 (s, 3H); ${}^{13}C$ **NMR** (100 MHz, CDCl₃) δ 171.2, 166.2, 166.0, 165.1, 161.7, 157.9 (q, J = 38 Hz), 145.2, 133.7, 133.4, 133.1, 129.9, 129.8, 129.6, 129.5, 129.1, 128.7, 128.6, 128.4, 128.3, 115.5 (g, J = 286 Hz), 108.2, 73.4, 71.7, 69.3, 62.8, 52.5, 48.0, 43.1, 22.8; $[\alpha]_D^{25}$ = +10.6 (*c* 0.97, CHCl₃); **IR** (neat): $v_{\text{max}} = 3260, 3064, 2958, 1721, 1651, 1259, 1163, 1090, 1068, 1025 \text{ cm}^{-1}$; **HRMS** (ESI): calcd. for $C_{35}H_{31}F_{3}N_{2}NaO_{11}[M + Na]^{+} m/z$ 735.1778; found m/z 735.1768.



To a solution of compound **S7**⁴ (2.40 g, 5.26 mmol, 1.0 equiv) in a mixture of THF (20 mL) and H₂O (4 mL) was added triphenylphosphine (1.66 g, 6.32 mmol, 1.2 equiv) at room temperature. The mixture was refluxed for 4 h at 85 °C before being cooled to 0 °C. Triethylamine (1.10 mL, 7.89 mmol, 1.5 equiv) and ditertbutyl dicarbonate (1.81 mL, 7.89 mmol, 1.5 equiv) were added, and the resultant mixture was stirred for 3 h at room temperature. After evaporation of solvent, the residue was purified by silical gel column chromatography (petroleum ether/EtOAc = 1:1, v/v) to give **7k** (2.32 g, 82%) as a white amorphous powder. **TLC**: (petroleum ether/EtOAc = 1:2, v/v), R_f = 0.25; ¹**H NMR** (400 MHz, CDCl₃) δ 6.09 – 5.91 (m, 1H), 5.89 (d, *J* = 2.4 Hz, 1H), 5.47 (dd, *J* = 5.2, 2.0 Hz, 1H), 5.32 – 5.25 (m, 1H), 4.89 – 4.78 (m, 1H), 4.68 (dd, *J* = 12.4, 2.8 Hz, 1H), 4.45 (td, *J* = 9.6, 2.4 Hz, 1H), 4.40 (s, 9H); ¹³**C NMR** (100 MHz, CDCl₃) δ 170.9, 170.6, 170.2, 170.0, 161.7, 156.2, 144.7, 110.8, 80.2, 77.5, 71.3, 67.8, 62.2, 52.4, 50.0, 47.5, 28.2, 23.1, 20.9, 20.7, 20.7; **[a**]_D²⁵ = +37.6

(*c* 1.10, CHCl₃); **IR** (neat): $v_{\text{max}} = 3342$, 2978, 1738, 1663, 1527, 1368, 1247, 1215, 1162, 1091, 1043, 1024, 734 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₃H₃₄N₂NaO₁₂ [M + Na]⁺ m/z 553.2009; found m/z 553.2005.



Triethylamine (112 µL, 0.802 mmol, 1.5 equiv) and benzyl chloroformate (150 µL, 1.07 mmol, 2.0 equiv) were added to a solution of **S8**⁴ (230 mg, 0.535 mmol, 1.0 equiv) in CH₂Cl₂ (2.5 mL) at 0 °C. After being stirred at room temperature for 2 h, the reaction mixture was directly concentrated in *vacuo*. Purification of the crude product by column chromatography with petroleum ether/acetone (1.5:1, v/v) provided **71** (244 mg, 81%) as a white amorphous powder. **TLC**: (petroleum ether/EtOAc = 1:3, v/v), R_f = 0.45; ¹**H NMR** (400 MHz, CDCl₃) δ 7.39 –7.27 (m, 5H), 6.02 (d, *J* = 9.6 Hz, 1H), 5.88 (d, *J* = 2.4 Hz, 1H), 5.50 – 5.45 (m, 1H), 5.32 – 5.20 (m, 2H), 5.04 (q, *J* = 12.4 Hz, 1H), 4.71 (dd, *J* = 12.4, 2.8 Hz, 1H), 4.52 (td, *J* = 9.6, 2.8 Hz, 1H), 4.31 – 4.11 (m, 3H), 3.74 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 2.00 (s, 3H), 1.67 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 171.0, 170.6, 170.4, 170.0, 161.7, 156.8, 144.5, 136.2, 128.5, 128.1, 127.6, 110.6, 77.2, 71.6, 67.8, 67.0, 62.2, 52.4, 50.7, 47.0, 22.9, 20.9, 20.7, 20.6; [**a**]_D²⁵ = +54.0 (*c* 0.90, CHCl₃); **IR** (neat): *v*_{max} = 3333, 3060, 2957, 2917, 1739, 1664, 1539, 1371, 1261, 1220, 1094, 1044 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₆H₃₂N₂NaO₁₂ [M + Na]⁺ *m*/z 587.1853; found *m*/z 587.1848.

4. General Procedure and Product Characterization



Under an argon atmosphere, to an oven-dried flask were successively added substrate (1.0 equiv), BI-OAc (2.0 equiv), dry MeCN (0.1 M) and azidotrimethylsilane (3.0 equiv). The reaction mixture was stirred with irradation of two 34 W blue LED lamps (the average distance between the lamp and the flask is about 11 cm). The ambient temperature of reactor was maintained 25 to 30 °C by a fan. After the reaction was completed (identified by TLC analysis), it was quenched with saturated aqueous KHCO₃, and the mixture was stirred

vigorously at room temperature for 5 min, then extracted with EtOAc for three times. The combined organic extracts were dried over Na_2SO_4 , filtered and concentrated in *vacuo*. The crude residue was purified by silica gel column chromatography using the indicated conditions to afford the corresponding product.

Following the general procedure, the reaction of substrate **7a** (1.01 g, 2.51 mmol, 1.0 equiv), BI-OAc (1.54 g, 5.02 mmol, 2.0 equiv) and azidotrimethylsilane (0.991 mL, 7.53 mmol, 3.0 equiv) proceeded in 1.5 h to afford product **8** as four diastereomers (1.05 g, 86%) after silica gel column chromatography (petroleum ether/acetone = 8:1, v/v). HPLC analysis (Waters e2695 Separations Module; Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50) indicated the ratio of **8a–d** to be 43:30:23:4. Pure products **8a**, **8b**, **8c**, and **8d** were respectively obtained through repeated silica gel column chromatography (petroleum ether/EtOAc = 6:1 or petroleum ether/acetone = 13:1, v/v) combined with reverse-phase C18 HPLC (H₂O/MeOH = 60:40 \rightarrow 40:60 \rightarrow 30:70, v/v).



3H), 2.06 (s, 3H), 2.01 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.6, 170.1, 169.6, 169.5, 165.3, 91.6, 69.8, 69.2, 66.8, 65.4, 61.8, 58.0, 56.2, 54.1, 20.7, 20.6, 20.6; $[\alpha]_D^{25} = +64.5$ (*c* 0.11, CHCl₃); **IR** (neat): $v_{max} = 2927$, 2118, 1752, 1437, 1370, 1226, 1076, 957, 750 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₇H₂₂N₆NaO₁₁ [M + Na]⁺ *m/z* 509.1244; found *m/z* 509.1253; HPLC purity: 93.6% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v; *t_R* = 21.31).

AcO OAc AcO N₃ N₃ CO₂Me **TLC**: (petroleum ether/EtOAc = 2:1, v/v), $R_f = 0.52$; ¹H NMR (400 MHz, CDCl₃) δ 5.48 (d, J = 9.2 Hz, 2H), 5.14 (d, J = 9.2 Hz, 1H), 4.36 (t, J = 8.8 Hz, 2H), 4.22 (dd, J = 12, 2.8 Hz, 1H), 3.92 (s, 3H), 3.76 (d, J = 10 Hz, 1H), 2.14 (s, 3H), 2.07 (s, 3H), 2.04 (s, 3H), 2.01 (s, 3H); ¹³C

NMR (150 MHz, CDCl₃) δ 170.3, 170.1, 169.5, 169.5, 165.7, 91.2, 71.5, 69.4, 67.4, 65.1, 61.8, 61.3, 53.6, 20.7, 20.6, 20.6, 20.6; $[\alpha]_D^{25} = +32.5$ (*c* 0.08, CHCl₃); **IR** (neat): $v_{max} = 2926$, 2118, 1752, 1437, 1372, 1228, 1080, 804, 735 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₇H₂₂N₆NaO₁₁ [M + Na]⁺ *m/z* 509.1244; found *m/z* 509.1253; HPLC purity: 99.8% (Flow rate: 1.0 mL/min;

Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v; *t_R* = 22.76).



TLC: (petroleum ether/EtOAc = 2:1, v/v), $R_f = 0.52$; ¹H NMR (400 MHz, CDCl₃) δ 5.42 (d, J = 2.4 Hz, 2H), 5.34 (d, J = 10 Hz, 1H), 4.55 (dd, J = 12.4, 2.0 Hz, 1H), 4.42 (d, J = 9.6 Hz, 1H), 4.24 (dd, J = 12.8, 2.8 Hz, 1H), 4.07 (t, J = 2.0 Hz, 1H), 3.93 (s, 3H), 2.13 (s, 3H), 2.08 (s,

6H), 1.99 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.5, 170.5, 169.4, 169.2, 164.4, 91.5, $69.4, 67.3, 67.1, 63.3, 61.8, 58.5, 53.7, 20.7, 20.6, 20.6, 20.3; [a]_D^{25} = +116.2 (c 0.32, CHCl_3);$ **IR** (neat): $v_{\text{max}} = 2959$, 2118, 1748, 1438, 1372, 1223, 1053, 924, 732 cm⁻¹; **HRMS** (ESI): calcd. for $C_{17}H_{22}N_6NaO_{11}[M + Na]^+ m/z$ 509.1244; found m/z 509.1231; HPLC purity: 97.7% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = $60:40 \rightarrow$ 50:50, v/v; $t_R = 16.59$); The supplementary crystallographic data of 8c [m.p.: 108 – 110 °C $(hexane/CHCl_3 = 5:1, v/v)$ have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975133). These data be obtained free of can charge via www.ccdc.cam.ac.uk/data request/cif.

TLC: (petroleum ether/EtOAc = 2:1, v/v), $R_f = 0.48$; ¹H NMR (400 MHz, AcQ OAc CDCl₃) δ 5.34 (s, 1H), 5.30 (d, J = 9.6 Hz, 1H), 5.15 (t, J = 2.8 Hz, 1H), AcO-4.46 (d, J = 12.4 Hz, 1H), 4.29 (dd, J = 12.4, 3.2 Hz, 1H), 4.22 (d, J = 3.2ĊO₂Me Hz, 1H), 3.97 (d, *J* = 9.6 Hz, 1H), 3.92 (s, 3H), 2.16 (s, 3H), 2.11 (s, 3H), 2.09 (s, 3H), 1.98 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.5, 170.4, 169.3, 169.3, 165.8, 90.1, 71.9, 67.7, 67.3, 62.9, 61.7, 58.7, 53.9, 20.7, 20.6, 20.5, 20.4; $[\alpha]_D^{25} = +101.8$ (c 0.09, CHCl₃); **IR** (neat): $v_{\text{max}} = 2918$, 2115, 1747, 1436, 1372, 1225, 1045, 923, 733 cm⁻¹; **HRMS** (ESI): calcd. for $C_{17}H_{22}N_6NaO_{11}$ [M + Na]⁺ m/z 509.1244; found m/z 509.1258; HPLC purity: 99.3% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: $H_2O/MeOH = 60:40 \rightarrow$ 50:50, v/v; $t_R = 17.93$); The supplementary crystallographic data of 8d [m.p.: 126 - 128 °C $(hexane/CHCl_3 = 5:1, v/v)$ have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975134). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data request/cif.

HPLC for measuring d.r. value of 8



Run information: column: Agilent ZORBAX 300SB-C8, 5 μ m, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v.

Peak information:	
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Peak	Isomer	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	8c	16.588	15071187	1452881	23.11
2	8d	17.898	2724899	271171	4.18
3	8 a	21.174	27853274	1862561	42.72
4	8 b	22.585	19555577	1234573	29.99

HPLC analysis of 8a under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	6.569	350767	2014	6.40
2	21.315	5132674	388119	93.60

HPLC analysis of 8b under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	14.246	20992	1784	0.15
2	22.766	13601865	914079	99.85

HPLC analysis of 8c under the same condition:



Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	11.107	62605	8406	0.30
2	16.590	20660693	1926998	97.68
3	17.880	333054	34312	1.57
4	21.015	95892	7490	0.45

Peak information:

HPLC analysis of 8d under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	13.866	12363	1339	0.16
2	17.927	7617236	775286	99.26
3	27.881	24075	2627	0.31
4	30.233	20293	2382	0.26

Following the general procedure, the mixture of substrate **7b** (1.20 g, 4.00 mmol, 1.0 equiv), BI-OAc (2.45 g, 8.00 mmol, 2.0 equiv) and azidotrimethylsilane (1.58 mL, 12.0 mmol, 3.0 equiv) was subjected to the diazidation reaction in 1 h to afford two isomers **9a** (537 mg, 35%) and **9b** (845 mg, 53%) after purification by silica gel column chromatography (petroleum ether/EtOAc = 8:1 to 5:1, v/v).



TLC: (petroleum ether/EtOAc = 4:1, v/v), $R_f = 0.50$; ¹H NMR (400 MHz, CDCl₃) δ 4.91 (dd, J = 6.8, 2.0 Hz, 1H), 4.77 (t, J = 6.8 Hz, 1H), 4.37 – 4.32 (m, 1H), 4.21 (d, J = 6.8 Hz, 1H), 4.18 – 4.12 (m, 1H), 4.01 (d, J = 2.0 Hz, 1H), 3.99 – 3.95 (m, 1H), 3.94 (s, 3H), 1.44 (s, 3H), 1.36 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.1, 152.5, 110.2, 90.0, 74.5,

73.3, 72.7, 71.2, 67.0, 60.1, 54.2, 26.8, 24.8; $[\alpha]_D^{25} = +76.7$ (*c* 0.68, CHCl₃); **IR** (neat): $v_{max} = 2988$, 2968, 2121, 1820, 1768, 1079, 846, 759 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₃H₁₆N₆NaO₈ [M + Na]⁺ *m/z* 407.0927; found *m/z* 407.0932; HPLC purity: 99.3% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; $t_R = 1.89$); The supplementary crystallographic data of **9a** [m.p.: 124 – 126 °C (CDCl₃)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975130). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.

TLC: (petroleum ether/EtOAc = 4:1, v/v), $R_f = 0.45$; ¹H NMR (400 MHz, CDCl₃) δ 5.02 (dd, J = 8.0, 5.2 Hz, 1H), 4.96 (dd, J = 8.0, 1.6 Hz, 1H), 4.36 – 4.31 (m, 1H), 4.19 – 4.12 (m, 2H), 3.94 (s, 3H), 3.91 (d, J = 4.8 Hz, 1H), 3.79 (dd, J = 8.4, 1.6 Hz, 1H), 1.41 (s, 3H), 1.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.4, 152.7, 110.2, 89.3, 73.7, 72.8, 72.5, 72.4,

66.7, 61.2, 53.7, 26.9, 24.8; $[\alpha]_D^{25} = +40.0$ (*c* 0.08, CHCl₃); **IR** (neat): $v_{max} = 2988$, 2967, 2111, 1820, 1767, 1078, 846, 759 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₃H₁₆N₆NaO₈ [M + Na]⁺ *m/z* 407.0927; found *m/z* 407.0915; HPLC purity: 99.4% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; $t_R = 1.85$); The supplementary crystallographic data of **9b** [m.p.: 111 – 113 °C (CDCl₃ = 3:1, v/v)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975131). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.

Following the general procedure, the diazidation reaction of **7c** (100 mg, 0.214 mmol, 1.0 equiv) proceeded with BI-OAc (131 mg, 0.427 mmol, 2.0 equiv) and azidotrimethylsilane (84.3 μ L, 0.641 mmol, 3.0 equiv) in 1 h to afford two isomers **10a** (32.0 mg, 27%) and **10b** (63.7 mg, 54%) after purification by column chromatography (petroleum ether/EtOAc = 2:1, v/v).

 $BzO OBz OBz OCO_2Me$

TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.52$; ¹H NMR (400 MHz, CDCl₃) δ 8.03 – 7.98 (m, 4H), 7.58 (q, J = 2.0 Hz, 2H), 7.44 (t, J= 7.6 Hz, 4H), 5.70 – 5.66 (m, 1H), 5.01 – 4.93 (m, 2H), 4.82 – 4.78 (m, 2H), 4.62 (dd, J = 12.4, 3.2 Hz, 1H), 4.28 (d, J = 7.2 Hz, 1H), 3.97 (s,

3H); ¹³C NMR (150 MHz, CDCl₃) δ 165.9, 164.9, 164.6, 152.1, 133.7, 133.3, 129.8, 129.6, 129.5, 128.9, 128.6, 128.5, 90.3, 74.4, 72.7, 69.4, 68.1, 61.7, 59.9, 54.4; **[a]**_D²⁵ = +57.4 (c 0.12, CHCl₃); **IR** (neat): $v_{\text{max}} = 3341$, 2960, 2122, 1827, 1729, 1452, 1265, 1094, 736 cm⁻¹; **HRMS** (ESI): calcd. for C₂₄H₂₀N₆NaO₁₀ [M + Na]⁺ *m/z* 575.1139; found *m/z* 575.1145.

TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.46$; ¹H NMR (400 MHz, CDCl₃) δ 8.00 (t, J = 7.6 Hz, 4H), 7.58 (q, J = 7.6, 2H), 7.48 – 7.43 (m, 4H), 5.68 – 5.65 (m, 1H), 5.05 (t, J = 7.2 Hz, 1H), 4.96 (dd, J = 7.6, 1.6 Hz, 1H), 4.83 – 4.81(m, 2H), 4.50 (dd, J = 9.2, 1.6 Hz, 1H), 3.85 (d, J = 5.6 Hz, 1H), 3.71 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 165.7, 165.2, 164.5, 152.3, 133.7, 133.4, 129.8, 129.6, 129.5, 128.9, 128.6, 128.5, 89.9, 74.4, 72.1, 69.7, 69.6, 62.1, 61.9, 53.7; $[\alpha]_D^{25} = +27.6$ (c 0.12, CHCl₃); IR (neat): $v_{max} = 3341$, 2961, 2120, 1822, 1730, 1452, 1262, 1094, 736 cm⁻¹; HRMS (ESI): calcd. for C₂₄H₂₀N₆NaO₁₀ [M + Na]⁺ *m/z* 575.1139; found *m/z* 575.1135.

Following the general procedure, compound **7d** (320 mg, 0.884 mmol, 1.0 equiv), BI-OAc (541 mg, 1.77 mmol, 2.0 equiv), azidotrimethylsilane (0.349 mL, 2.65 mmol, 3.0 equiv) and dry MeCN (8.8 mL) were used. The reaction was completed in 1 h to provide the isomers **11a** (104 mg, 26%) and **11b** (157 mg, 40%) after purification by column chromatography (petroleum ether/EtOAc = 30:1, v/v).



TLC: (petroleum ether/EtOAc = 7:1, v/v), $R_f = 0.45$; ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.36 (m, 5H), 6.24 (s, 1H), 4.51 (dd, J = 8.0, 5.2 Hz, 1H), 4.43 (dd, J = 12.4, 6.0 Hz, 1H), 4.35 (dd, J = 5.2, 2.4 Hz, 1H), 4.13 (dd, J = 8.8, 6.0 Hz, 1H), 4.07 – 4.00 (m, 3H), 3.95 (s, 3H), 1.41 (s, 3H), 1.36 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.9,

138.2, 129.2, 128.5, 126.0, 109.7, 103.4, 90.9, 75.1, 73.7, 72.6, 72.4, 66.9, 59.5, 53.9, 26.8, 25.3; $[\alpha]_D^{25} = +53.7$ (*c* 0.20, CHCl₃); **IR** (neat): $v_{max} = 2988$, 2114, 1756, 1372, 1260, 1225, 1098, 1074, 847, 750 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₉H₂₂N₆NaO₇ [M + Na]⁺ *m/z* 469.1448; found *m/z* 469.1469.



TLC: (petroleum ether/EtOAc = 7:1, v/v), $R_f = 0.35$; ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.38 (m, 5H), 6.25 (s, 1H), 4.77 (t, J = 6.0 Hz, 1H), 4.42 (dd, J = 12.0, 5.2 Hz, 1H), 4.36 (dd, J = 5.6, 1.6 Hz, 1H), 4.14 (d, J = 5.6 Hz, 2H), 3.92 (s, 3H), 3.81 (dd, J = 7.2, 1.6 Hz, 1H), 3.74 (d, J = 6.8 Hz, 1H), 1.40 (s, 3H), 1.36 (s, 3H); ¹³C NMR (100 MHz, CDCl₃)

δ 166.0, 138.1, 129.2, 128.5, 126.1, 109.6, 103.7, 90.1, 75.2, 73.9, 73.8, 72.0, 66.5, 62.6, 53.4, 26.8, 25.2; **[α]**_D²⁵ = +24.2 (*c* 0.16, CHCl₃); **IR** (neat): *v*_{max} = 2988, 2113, 1745, 1372, 1259, 1221, 1097, 1074, 847, 750 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₉H₂₂N₆NaO₇ [M + Na]⁺ *m/z* 469.1448; found *m/z* 469.1463.

Following the general procedure, the diazidation reaction of 7e (200 mg, 0.637 mmol, 1.0

equiv) occurred in the presence of BI-OAc (390 mg, 1.27 mmol, 2.0 equiv) and azidotrimethylsilane (0.251 mL, 1.91 mmol, 3.0 equiv) in 30 min to provide **12a** (103 mg, 41%) and **12b** (41.2 mg, 27%) after purification by column chromatography (petroleum ether/EtOAc = 30:1, v/v).



TLC: (petroleum ether/EtOAc = 5:1, v/v), $R_f = 0.51$; ¹H NMR (400 MHz, CDCl₃) δ 4.40 – 4.28 (m, 3H), 4.11 (dd, J = 9.2, 6.4 Hz, 1H), 4.02 – 3.98 (m, 3H), 3.90 (s, 3H), 1.53 (s, 3H), 1.43 (s, 3H), 1.40 (s, 3H), 1.37 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 166.0, 110.5, 109.7, 90.7, 74.0, 73.5, 72.6, 72.0, 66.9, 61.3, 53.8, 27.7, 26.9, 26.1, 25.3;

 $[\alpha]_D^{25} = +59.7$ (*c* 0.31, CHCl₃); **IR** (neat): $v_{max} = 2930$, 2116, 1757, 1374, 1265, 1074, 810, 733 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₅H₂₂N₆NaO₇ [M + Na]⁺ *m/z* 421.1448; found *m/z* 421.1455; HPLC purity: 91.9% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; $t_R = 2.37$).



TLC: (petroleum ether/EtOAc = 5:1, v/v), $R_f = 0.42$; ¹H NMR (400 MHz, CDCl₃) δ 4.57 (t, J = 6.4 Hz, 1H), 4.38 – 4.32 (m, 2H), 4.15 – 4.08 (m, 2H), 3.90 (s, 3H), 3.75 (dd, J = 7.6, 1.2 Hz, 1H), 3.63 (d, J = 6.4 Hz, 1H), 1.56 (s, 3H), 1.42 (s, 3H), 1.40 (s, 3H), 1.37 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 166.2, 110.4, 109.6, 89.7, 74.3, 73.7, 73.5, 71.5, 66.5,

64.1, 53.4, 27.5, 26.9, 25.7, 25.2; $[\alpha]_D^{25} = +33.6$ (*c* 0.22, CHCl₃); **IR** (neat): $v_{max} = 2929$, 2117, 1746, 1374, 1264, 1081, 807, 732 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₅H₂₂N₆NaO₇ [M + Na]⁺ *m/z* 421.1448; found *m/z* 421.1443.

Following the general procedure, the mixture of **7f** (50.2 mg, 0.144 mmol, 1.0 equiv), BI-OAc (88.3 mg, 0.288 mmol, 2.0 equiv), azidotrimethylsilane (56.9 μ L, 0.433 mmol, 3.0 equiv) and dry MeCN (1.4 mL) was subjected to the diazidation reaction in 1 h to generate two isomers **13a** (27.5 mg, 45%) and **13b** (13.5 mg, 21%) after purification by silica gel column chromatography (petroleum ether/EtOAc = 25:1, v/v).



TLC: (petroleum ether/EtOAc = 9:1, v/v), $R_f = 0.45$; ¹H NMR (400 MHz, CDCl₃) δ 8.04 (d, J = 7.2 Hz, 2H), 7.58 (t, J = 7.6 Hz, 1H), 7.45 (t, J = 7.6 Hz, 2H), 4.65 (t, J = 8.4 Hz, 1H), 4.59 – 4.53 (m, 2H), 4.33 – 4.32 (m, 2H), 4.00 – 3.97 (m, 1H), 3.93 (s, 3H), 1.56 (s, 3H), 1.38 (s,

3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.2, 166.0, 133.3, 129.7, 128.4, 110.8, 90.7, 77.3, 77.0, 76.7, 74.3, 72.4, 70.5, 63.5, 61.4, 53.8, 27.9, 26.1; **[a]**_D²⁵ = +59.5 (*c* 0.09, CHCl₃); **IR** (neat):

 $v_{\text{max}} = 2919, 2116, 1723, 1245, 1081, 713 \text{ cm}^{-1}$; **HRMS (ESI)**: calcd. for C₁₈H₂₀N₆NaO₇ [M + Na]⁺ *m/z* 455.1291; found *m/z* 455.1255.



TLC: (petroleum ether/EtOAc = 9:1, v/v), $R_f = 0.36$; ¹H NMR (400 MHz, CDCl₃) δ 8.04 (d, J = 7.2 Hz, 2H), 7.58 (t, J = 7.2 Hz, 1H), 7.45 (t, J = 7.6 Hz, 2H), 4.66 – 4.55 (m, 3H), 4.32 – 4.28 (m, 2H), 3.85 (s, 3H), 3.59 (d, J = 7.2, 1H), 1.57 (s, 3H), 1.38 (s, 3H); ¹³C NMR (100 MHz,

CDCl₃) δ 166.2, 166.1, 133.2, 129.7, 128.5, 110.6, 90.0, 74.8, 71.9, 71.6, 64.7, 63.4, 53.3, 27.8, 25.9; $[\alpha]_D^{25} = +21.7$ (c 0.05, CHCl₃); **IR** (neat): $v_{max} = 2918$, 2115, 1723, 1271, 1081, 712 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₈H₂₀N₆NaO₇ [M + Na]⁺ m/z 455.1291; found m/z 455.1282.

Following the general procedure, substrate **7g** (300 mg, 0.634 mmol, 1.0 equiv), BI-OAc (388 mg, 1.27 mmol, 2.0 equiv), azidotrimethylsilane (0.250 mL, 1.90 mmol, 3.0 equiv) and dry MeCN (6 mL) were used. The reaction proceeded in 2 h to provide **14** (307 mg, 87%) after column chromatography (petroleum ether/acetone = 3:1, v/v) on silica gel. HPLC analysis (Waters e2695 Separations Module; Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 μ m, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v) indicated that this material was a 5:9:66:20 mixture of four isomers (**14a**, **14b**, **14c**, **14d**). Each pure product was respectively obtained through repeated silica gel column chromatography (petroleum ether/EtOAc = 3.5:1 or petroleum ether/acetone = 5:1, v/v) combined with reverse-phase C18 HPLC (H₂O/MeOH = 50:50 \rightarrow 40:60 \rightarrow 30:70, v/v).



TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.61$; ¹H NMR (400 MHz, CDCl₃) δ 7.19 (d, J = 10.0 Hz, 1H), 5.45 – 4.28 (m, 2H), 5.17 (td, J = 6.0, 2.8 Hz, 1H), 4.46 – 4.32 (m, 2H), 4.24 (q, J = 10.0 Hz, 1H), 4.13 – 4.01 (m, 2H), 3.98 (s, 3H), 2.13 (s, 6H),

2.08 (s,3H), 2.03 (s,3H); ¹³C NMR (150 MHz, CDCl₃) δ 171.4, 170.6, 170.2, 169.5, 164.8, 157.8, 157.5, 91.1, 71.7, 70.8, 70.1, 66.7, 62.4, 61.6, 54.2, 49.3, 20.9, 20.6, 20.5, 20.4; $[\alpha]_D^{25} = -25.3$ (*c* 0.30, CHCl₃); **IR** (neat): $v_{max} = 3254$, 2958, 2925, 2114, 1748, 1246, 1210, 1059, 1030 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₀H₂₇N₇NaO₁₂ [M + Na]⁺ *m/z* 580.1615; found *m/z* 580.1611; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v; *t_R* = 21.26).



TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.59$; ¹H NMR (400 MHz, CDCl₃) δ 6.85 (d, J = 10.0 Hz, 1H), 5.54 (t, J = 10.4 Hz, 1H), 5.35 – 5.28 (m, 1H), 5.25 (dd, J = 8.0, 2.4 Hz, 1H), 4.50 (dd, J = 10.4, 2.0 Hz, 1H), 4.30 – 4.14 (m, 2H), 4.07 (dd, J = 12.4, 5.6 Hz,

1H), 3.93 (s, 3H), 3.62 (d, J = 10.0 Hz, 1H), 2.15 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H), 2.03 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.8, 170.5, 169.7, 169.6, 166.0, 157.7, 116.3, 90.9, 72.7, 70.6, 68.8, 66.7, 65.0, 62.1, 53.8, 49.3, 20.8, 20.6, 20.6, 20.4; $[\alpha]_D^{25} = -17.0$ (*c* 0.20, CHCl₃); **IR** (neat): $v_{max} = 3257$, 2959, 2117, 1753, 1289, 1217 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₀H₂₇N₇NaO₁₂ [M + Na]⁺ *m*/*z* 580.1615; found *m*/*z* 580.1611; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v; *t_R* = 24.67).



TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.59$; ¹H NMR (400 MHz, CDCl₃) δ 7.05 (d, J = 9.2 Hz, 1H), 5.53 (dd, J = 10.0, 3.2 Hz, 1H), 5.37 (dd, J = 5.2, 2.0 Hz, 1H), 5.23 (td, J = 5.2, 2.4 Hz, 1H), 4.65 (dd, J = 12.8, 2.4 Hz, 1H), 4.40 – 4.27 (m, 2H),

4.21 (d, J = 3.2 Hz, 1H), 4.16 (dd, J = 12.8, 6.8 Hz, 1H), 3.94 (s, 3H), 2.17 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H), 2.04 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ 170.9, 170.6, 170.5, 170.2, 164.2, 157.6, 116.3, 114.4, 90.9, 71.7, 69.1, 67.3, 61.9, 61.9, 53.8, 45.8, 20.9, 20.6, 20.5, 20.3; $[\alpha]_D^{25} = -22.3$ (*c* 0.66, CHCl₃); **IR** (neat): $v_{max} = 3275$, 2112, 1742, 1370, 1211, 1031 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₀H₂₇N₇NaO₁₂ [M + Na]⁺ *m*/*z* 580.1615; found *m*/*z* 580.1615; HPLC purity: 98.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v; *t*_{*R*} = 16.72); The supplementary crystallographic data of **14c** [m.p.: 203 – 205 °C (petroleum ether/acetone = 5:1, v/v)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975132). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.



TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.40$; ¹H NMR (400 MHz, CDCl₃) δ 6.72 (d, J = 8.8 Hz, 1H), 5.41 – 5.33 (m, 2H), 5.24 (dd, J = 7.6, 1.6 Hz, 1H), 4.38 – 4.28 (m, 2H), 4.25 – 4.03 (m, 3H), 3.93 (s, 3H), 2.19 (s, 3H), 2.14 (s, 3H), 2.13 (s, 3H), 2.03 (s, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 170.6, 170.5, 170.3, 170.1, 165.4, 157.5, 116.2, 89.6, 72.3, 68.7, 68.7, 67.2, 62.1, 61.8, 54.0, 46.1, 20.9, 20.6, 20.6, 20.3; **[a]**_D²⁵ = +10.4 (*c* 0.77, CHCl₃); **IR** (neat): $v_{\text{max}} = 3251$, 2923, 2133, 2116, 1368, 1252, 1205, 1036 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₀H₂₇N₇NaO₁₂ [M + Na]⁺ *m/z* 580.1615; found *m/z* 580.1612; HPLC purity: 96.8% (Flow

rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = $60:40 \rightarrow 50:50$, v/v; t_R = 19.79); The supplementary crystallographic data of **14d** [m.p.: 197 – 199 °C (hexane/CHCl₃ = 5:1, v/v)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975126). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data request/cif.



HPLC for measuring d.r. value of 14

Run information: column: Agilent ZORBAX 300SB-C8, 5 μ m, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 60:40 \rightarrow 50:50, v/v.

D				
Р	69	Z 1	into	rmation
	cai	x 1		i mation.

Peak	Isomer	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	14c	16.579	7581466	557811	66.10
2	14d	20.215	2306804	137140	20.11
3	14a	21.161	621575	34369	5.42
4	14b	24.925	959431	72483	8.37

HPLC analysis of 14a under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	11.896	14458	5473	0.17
2	21.256	8552093	422624	99.83

HPLC analysis of 14b under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	21.260	14851	1232	0.12
2	24.669	12730552	781236	99.88

HPLC analysis of 14c under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	12.605	37655	4763	1.17
2	16.721	3173270	243176	98.83

HPLC analysis of 14d under the same condition:



Peak information:

Pea	k RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	11.045	813849	116367	1.59
2	11.854	153570	18705	0.30
3	13.147	85940	11640	0.17
4	13.993	54020	6173	0.11

5	14.380	324602	33539	0.63
6	16.734	181810	15181	0.36
7	19.788	49593832	1902242	96.85

Following the general procedure, the diazidation reaction of **7h** (2.03 g, 2.82 mmol, 1.0 equiv) took place with BI-OAc (1.72 g, 5.63 mmol, 2.0 equiv), azidotrimethylsilane (1.11 mL, 8.45 mmol, 3.0 equiv) and dry MeCN (28 mL) in 2 h to give a mixture of three isomers **15a–c** (1.50 g, 66.4%) and pure **15d** (420 mg, 18.6%) after column chromatography (petroleum ether/EtOAc = 5:1 to 2:1, v/v). The ratio of **15a–c** was indicated to be 14:13:73 by HPLC analysis (Waters e2695 Separations Module; Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 μ m, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 50:50 \rightarrow 40:60 \rightarrow 30:70, v/v). Accordingly, the ratio of four isomers (**15a–d**) was 10:10:52:28. Pure **15a–c** was respectively obtained through repeated silica gel column chromatography (petroleum ether/EtOAc = 5:1 or petroleum ether/acetone = 8:1, v/v) combined with reverse-phase C18 HPLC (H₂O/MeOH = 40:60 \rightarrow 30:70, v/v).

$$\begin{array}{c} \text{BzO} \overset{\text{BzO}}{\underset{\text{AcHN}}{\text{DzO}}} \overset{\text{OBz}}{\underset{\text{BzO}}{\text{N}_3}} \overset{\text{N}_3}{\underset{\text{AcHN}}{\text{DzO}_2\text{Me}}} \\ \text{BzO} \overset{\text{OBz}}{\underset{\text{BzO}}{\text{TLC}: (petroleum ether/EtOAc = 1.5:1, v/v), R_f = 0.55; ^{1}\text{H NMR}} \\ (400 \text{ MHz, CDCl}_3) \delta 8.09 - 7.91 (m, 8H), 7.63 - 7.32 (m, 12H), \\ 5.90 (dd, J = 6.4, 1.6 \text{ Hz}, 1H), 5.87 - 5.81 (m, 1H), 5.64 (d, J = 9.6 \text{ Hz}, 1H), 5.55 (t, J = 2.0 \text{ Hz}, 1H), 4.94 (dd, J = 12.4, 3.2 \text{ Hz}, 1H), \\ \end{array}$$

1H), 4.64 (dd, J = 10.8, 2.0 Hz, 1H), 4.50 – 4.35 (m, 2H), 4.21 (d, J = 10.0 Hz, 1H), 3.96 (s, 3H), 1.78 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.0, 166.6, 166.0, 165.6, 165.2, 165.2, 133.8, 133.5, 133.4, 133.1, 130.0, 129.9, 129.9, 129.7, 129.5, 129.4, 129.2, 128.6, 128.6, 128.5, 128.3, 91.6, 72.9, 71.5, 70.6, 68.4, 62.9, 62.8, 54.1, 49.8, 23.0; $[\alpha]_D^{25} = +6.1$ (*c* 0.80, CHCl₃); **IR** (neat): $v_{max} = 2967$, 2132, 2113, 1722, 1451, 1257, 1091, 1067, 1024, 800, 707, 686 cm⁻¹; **HRMS (ESI)**: calcd. for C₄₀H₃₅N₇NaO₁₂ [M + Na]⁺ *m/z* 828.2241; found *m/z* 828.2232; HPLC purity: 92.2% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 40:60 \rightarrow 30:70, v/v; *t_R* = 35.69).

$$\begin{array}{c} \text{BzO} \overset{\text{BzO}}{\longrightarrow} \underset{\text{AcHN}}{\overset{\text{OBz}}{\longrightarrow}} \underset{\text{BzO}}{\overset{\text{OBz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{OBz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{ODz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{N}_3}} \underset{\text{N}_3}{\overset{\text{ODz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{ODz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{ODz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{ODz}}{\longrightarrow}} \underset{\text{N}_3}{\overset{\text{N}_3}} \underset{\text{N}_3} \underset{\text{N}_3}{\overset{\text{N}_3}} \underset{\text{N}_3}{\overset{\text{N}_3}} \underset{\text{N}_3}{\overset{\text{N}_3}} \underset{\text{N}_3} \underset{\text{N}_3}{\overset{\text{N}_3}} \underset{\text{N}_3} \underset{\text{N}$$

(d, J = 10.0 Hz, 1H), 3.48 (s, 3H), 1.80 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 170.0, 166.5, 166.0, 165.6, 165.3, 164.8, 133.7, 133.5, 133.0, 130.0, 129.9, 129.9, 129.7, 129.6, 129.2,

129.2, 128.6, 128.6, 128.5, 128.3, 91.2, 73.8, 71.5, 69.3, 67.9, 65.7, 63.0, 53.3, 49.3, 23.0; $[\alpha]_D^{25} = +29.5$ (*c* 0.9, CHCl₃); **IR** (neat): $v_{max} = 3364$, 2963, 2932, 2114, 1726, 1249, 1093, 1068, 1026, 709 cm⁻¹; **HRMS (ESI)**: calcd. for C₄₀H₃₅N₇NaO₁₂ [M + Na]⁺ *m/z* 828.2241; found 828.2235; HPLC purity: 96.7% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 40:60 \rightarrow 30:70, v/v; *t_R* = 37.20); The supplementary crystallographic data of **15b** [m.p.: 115 – 117 °C (petroleum ether/acetone = 5:1, v/v)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975127). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.



TLC: (petroleum ether/EtOAc = 1.5:1, v/v), $R_f = 0.53$; ¹H NMR (400 MHz, CDCl₃) δ 8.18 – 8.13 (m, 2H), 8.05 – 7.98 (m, 4H), 7.98 – 7.92 (m, 2H), 7.64 – 7.45 (m, 6H), 7.45 – 7.31 (m, 6H), 5.94 – 5.86 (m, 2H), 5.84 (dd, J = 10.4, 3.2 Hz, 1H), 5.69 – 5.94

(m, 1H), 5.11 (dd, J = 12.4, 2.8 Hz, 1H), 4.68 (dd, J = 10.8, 2.0 Hz, 1H), 4.59 (dd, J = 12.4, 6.4 Hz, 1H), 4.49 (q, J = 10.4 Hz, 1H), 4.36 (d, J = 3.2 Hz, 1H), 3.96 (s, 3H), 1.81 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.1, 166.1, 166.0, 165.7, 165.6, 164.6, 133.8, 133.5, 133.3, 133.0, 130.1, 130.0, 129.9, 129.7, 129.7, 129.5, 129.2, 128.6, 128.6, 128.4, 128.4, 128.3, 91.2, 72.78, 71.5, 670.0, 69.0, 63.1, 62.5, 53.7, 45.9, 23.2; **[a]**_D²⁵ = -1.3 (*c* 0.70, CHCl₃); **IR** (neat): $v_{\text{max}} = 3375$, 2970, 2924, 2112, 1723, 1695, 1257, 1092, 1068, 1026, 803, 709 cm⁻¹; **HRMS (ESI)**: calcd. for C₄₀H₃₅N₇NaO₁₂ [M + Na]⁺ *m/z* 828.2241; found *m/z* 828.2232; HPLC purity: 93.7% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 40:60 \rightarrow 30:70, v/v; *t_R* = 34.73).



TLC: (petroleum ether/EtOAc = 1.5:1, v/v), $R_f = 0.40$; ¹H NMR (400 MHz, CDCl₃) δ 8.19 – 8.13 (m, 2H), 8.07 – 7.98 (m, 4H), 7.97 – 7.92 (m, 2H), 7.67 – 7.32 (m, 12H), 5.97 – 5.90 (m, 1H), 5.86 (dd, J = 7.6, 1.6 Hz, 1H), 5.80 (dd, J = 10.8, 3.2 Hz, 1H), 5.64 – 5.55 (m,

1H), 4.95 (dd, J = 12.4, 3.2 Hz, 1H), 4.64 (dd, J = 10.8, 1.6 Hz, 1H), 4.59 – 4.49 (m, 2H), 4.20 – 4.08 (m, 1H), 3.62 (s, 3H), 1.85 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 170.4, 166.0, 165.9, 165.9, 165.7, 165.3, 133.8, 133.7, 133.3, 133.0, 130.2, 123.0, 129.9, 129.7, 129.6, 129.0, 128.7, 128.6, 128.5, 128.3, 89.5, 73.0, 69.9, 69.4, 68.7, 63.1, 62.8, 53.7, 46.6, 23.3; $[a]_D^{25} = +41.2$ (*c* 0.70, CHCl₃); **IR** (neat): $v_{max} = 3375$, 2963, 2111, 1724, 1255, 1106, 1094, 1069, 1026, 709 cm⁻¹; **HRMS (ESI)**: calcd. for C₄₀H₃₅N₇NaO₁₂ [M + Na]⁺ *m/z* 828.2241; found *m/z* 828.2237; HPLC purity: 96.6% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 40:60 \rightarrow 30:70, v/v; t_R = 36.64).



HPLC for measuring d.r. value of 15a, 15b and 15c

Run information: column: Agilent ZORBAX 300SB-C8, 5 μ m, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 40:60 \rightarrow 30:70, v/v.

n		• •	
Р	eak	into	rmation:
-			

Peak	Isomer	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	15c	34.654	4349581	238456	72.31
2	15a	35.563	856881	47596	14.25
3	15b	37.313	808326	37360	13.44

HPLC analysis of 15a under the same condition:



Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	8.672	34786	4920	0.95
2	14.265	185201	9208	5.06
3	33.539	65218	4490	1.78
4	35.699	3373911	177500	92.21

Peak information:

HPLC analysis of 15b under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	4.820	19324	2916	0.53
2	8.673	22458	3402	0.61
3	27.416	27751	2209	0.75
4	33.439	50962	3482	1.38
5	37.200	3559823	164298	96.73

HPLC analysis of 15c under the same condition:



Peak information:

Peak	RetTime [min]	Area [mAU*s]	Height [mAU]	Area (%)
1	34.735	10484801	500193	93.70
2	35.571	114388	6129	1.02
3	37.077	215188	5575	1.92
4	43.361	293747	11800	2.63
5	44.038	81737	4097	0.73

Following the general procedure, substrate **7i** (201 mg, 0.351 mmol, 1.0 equiv), BI-OAc (215 mg, 0.702 mmol, 2.0 equiv) and azidotrimethylsilane (138 μ L, 1.05 mmol, 3.0 equiv) were used. The crude product was isolated by silica gel column chromatography (petroleum ether/EtOAc = 12:1 to 8:1, v/v) to afford two isomers **16a** (65.3 mg, 28%) and **16b** (131 mg, 57%).

TLC: (petroleum ether/EtOAc = 4:1, v/v), R_f = 0.58; ¹H NMR (400 MHz, CDCl₃) δ 5.26 (d, J = 8.4 Hz, 1H), 4.41 (dd, J = 10.8, 1.2 Hz, 1H), 4.14 (q, J = 6.4 Hz, 1H), 4.07 – 3.86 (m, 3H), 3.92 (s, 3H), 1.29 (s, 3H), 0.89 (s, 18H), 0.17 (s, 3H), 0.13 (s, 6H), 0.06 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 169.6, 166.3, 108.6, 91.4, 76.0, 73.2, 71.5, 70.6, 66.4, 66.3, 53.9, 53.6, 26.6, 25.9, 25.6, 24.8, 23.9, 18.5, 18.0, -3.6, -4.0, -4.6, -4.6; $[\alpha]_D^{25} = -16.0$ (c 0.40, CHCl₃); **IR** (neat): $v_{max} = 2955$, 2930, 2858, 2112, 1757, 1666, 1253, 1120, 1069, 1023, 836, 777, 736 cm⁻¹;

HRMS (ESI): calcd. for C₂₇H₅₁N₇NaO₈Si₂ [M + Na]⁺ m/z 680.3235; found m/z 680.3249; HPLC purity: 98.9% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH $= 10:90, v/v; t_R = 6.46$).



3H), 0.13 (s, 3H), 0.12 (s, 3H), 0.07 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.2, 165.4, 108.1, 91.0, 72.7, 70.6, 67.8, 65.7, 64.9, 53.2, 50.8, 29.7, 26.4, 25.9, 25.6, 24.7, 23.9, 18.5, 17.9, -3.4, -4.1, -4.4, -5.1; $[\alpha]_D^{25} = -0.9$ (*c* 0.41, CHCl₃); **IR** (neat): $v_{max} = 2955$, 2930, 2858, 2111, 1758, 1668, 1256, 1068, 1039, 837, 779, 734 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₇H₅₁N₇NaO₈Si₂ [M + Na]⁺ *m/z* 680.3235; found *m/z* 680. 3235; HPLC purity: 99.3% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; *t_R* = 5.10).

Following the general procedure, the diazidation reaction of **7j** (4.21 g, 5.91 mmol, 1.0 equiv) proceeded with BI-OAc (3.62 g, 11.8 mmol, 2.0 equiv) and azidotrimethylsilane (2.33 mL, 17.7 mmol, 3.0 equiv) in 2 h to give two isomers **17a** (2.68 g, 57%) and **17b** (1.50 g, 32%) after purification by silica gel column chromatography (petroleum ether/EtOAc = 2:1, v/v).



TLC: (petroleum ether/EtOAc = 1:1.5, v/v), $R_f = 0.53$; ¹H NMR (400 MHz, CDCl₃) δ 8.10 - 7.98 (m, 4H), 7.97 - 7.90 (m, 2H), 7.65 - 7.31 (m, 9H), 7.16 - 7.07 (m, 1H), 6.72 (d, *J* = 8.8 Hz, 1H), 5.91 - 5.81 (m, 2H), 5.12 (dd, *J* = 12.8, 2.4 Hz, 1H), 4.93 (d, *J* =

10.8 Hz, 1H), 4.70 – 4.61 (m, 1H), 4.53 (dd, J = 12.4, 5.6 Hz, 1H), 4.35 – 4.25 (m, 1H), 4.02 (d, J = 3.6 Hz, 1H), 3.94 (s, 3H), 2.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.1, 166.1, 166.0, 165.3, 164.4, 157.9 (q, J = 38 Hz), 133.8, 133.6, 133.1, 130.1, 129.9, 129.6, 129.5, 129.2, 128.8, 128.6, 128.6, 128.5, 128.3, 115.4 (q, J = 286 Hz), 91.2, 71.4, 68.9, 68.5, 62.7, 61.1, 53.7, 49.8, 44.6, 23.2; **[a]** $p^{25} = +63.1$ (*c* 1.71, CHCl₃); **IR** (Neat): $v_{max} = 2114$, 1725, 1259, 1088, 1068, 735, 709 cm⁻¹; **HRMS (ESI)**: calcd. for C₃₅H₃₁F₃N₈NaO₁₁ [M + Na]⁺ *m/z* 819.1962; found *m/z* 819.1954; HPLC purity: 98.5% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 100:0 → 5:95, v/v; *t_R* = 18.58); The supplementary crystallographic data of **17a** [m.p.: 194 – 196 °C (petroleum ether/acetone = 5:1, v/v)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975128). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.



TLC: (petroleum ether/EtOAc = 1:1.5, v/v), $R_f = 0.50$; ¹H NMR (400 MHz, CDCl₃) δ 8.09 – 7.98 (m, 4H), 7.94 (d, J = 7.6 Hz, 2H), 7.64 – 7.49 (m, 4H), 7.49 – 7.41 (m, 3H), 7.41 – 7.32 (m, 2H), 6.71 – 6.56 (m, 1H), 5.92 – 5.85 (m, 1H), 5.80 (dd, J = 7.6, 1.6 Hz, 1H),

5.22 (d, J = 11.2 Hz, 1H), 4.92 (dd, J = 12.8, 2.8 Hz, 1H), 4.61 (d, J = 7.2 Hz, 1H), 4.46 (dd, J = 12.4, 4.8 Hz, 1H), 4.33 – 4.20 (m, 1H), 4.15 (q, J = 6.8 Hz, 1H), 3.68 (s, 3H), 1.96 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.4, 166.4, 166.0, 165.6, 165.5, 158.6 (q, J = 38 Hz), 134.0, 133.6, 133.1, 130.1, 129.8, 129.6, 129.4, 129.2, 128.7, 128.6, 128.3, 128.3, 115.5 (q, J = 287Hz), 89.6, 70.1, 69.9, 68.5, 62.5, 59.0, 53.6, 50.6, 46.0, 22.9; $[a]_D^{25} = +53.4$ (*c* 1.00, CHCl₃); IR (neat): $v_{max} = 2116$, 1726, 1263, 1247, 1091, 1069, 734, 708 cm⁻¹; HRMS (ESI): calcd. for C₃₅H₃₁F₃N₈NaO₁₁ [M + Na]⁺ *m/z* 819.1962; found *m/z* 819.1961; HPLC purity: 97.6% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 100:0 \rightarrow 5:95; *t_R* = 19.90).

Following the general procedure, the diazidation of **7k** (2.20 g, 4.15 mmol, 1.0 equiv) proceeded employing BI-OAc (2.54 g, 8.30 mmol, 2.0 equiv) and azidotrimethylsilane (1.64 mL, 12.4 mmol, 3.0 equiv) in 2 h to afford two isomers **18a** (1.64 g, 64%) and **18b** (650 mg, 26%) after purification by repeated column chromatography (petroleum ether/EtOAc = 1.5:1, v/v).



(s, 3H), 2.04 (s, 3H), 1.91 (s, 3H), 1.41 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 170.6, 170.3, 169.9, 164.8, 155.8, 90.7, 80.5, 73.0, 71.0, 67.5, 63.0, 62.1, 53.6, 50.9, 44.5, 28.2, 28.2, 23.0, 21.0, 20.7, 20.6; $[\alpha]_D^{25} = -9.3$ (*c* 1.07, CHCl₃); **IR** (neat): $v_{max} = 3357$, 2962, 2113, 1745, 1368, 1248, 1215, 1043, 734 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₃H₃₄N₈NaO₁₂ [M + Na]⁺ *m/z* 637.2194; found *m/z* 637.2189; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: HRC-SIL SHIM-PACK, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: hexane/2-isopropanol = 80:20, v/v; *t_R* = 3.22).



2.04 (s, 3H), 1.88 (s, 3H), 1.41 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.8, 170.6, 170.0, 169.9, 165.9, 155.8, 90.5, 80.6, 74.8, 68.9, 67.3, 62.9, 62.1, 53.8, 51.3, 45.1, 28.2, 28.2, 23.1, 21.0, 20.7, 20.7; $[\alpha]_D^{25} = +20.8$ (c 0.83, CHCl₃); **IR** (neat): $v_{max} = 3357, 2962, 2112, 1747,$ 1368, 1249, 1214, 1040, 734, 703 cm⁻¹; HRMS (ESI): calcd. for C₂₃H₃₄N₈NaO₁₂ [M + Na]⁺ m/z 637.2194; found m/z 637.2186; HPLC purity: 98.3% (Flow rate: 1.0 mL/min; Column: HRC-SIL SHIM-PACK, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: hexane/2-isopropanol = 80:20, v/v; t_R = 3.64).

Following the general procedure, the diazidation reaction of 71 (512 mg, 0.908 mmol, 1.0 equiv) occurred using BI-OAc (556 mg, 1.82 mmol, 2.0 equiv) and azidotrimethylsilane (358 µL, 2.72 mmol, 3.0 equiv) in 2 h to provide two isomers 19a (387 mg, 66%) and 19b (52 mg, 9%) after purification by repeated column chromatography (petroleum ether/EtOAc = 3:1, v/v).



TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.54$; ¹H NMR $\begin{array}{c} \text{AcO} & \text{AcO$ 12.4 Hz, 1H), 4.22 – 4.05 (m, 5H), 3.95 (s, 3H), 2.12 (s, 3H), 2.08

(s, 3H), 2.04 (s, 3H), 1.75 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 170.6, 170.3, 169.8, 164.7, 156.4, 136.0, 128.6, 128.2, 127.8, 90.7, 72.9, 71.1, 67.4, 67.2, 63.0, 62.1, 53.6, 51.8, 44.4, 22.8, 21.0, 20.7, 20.6; $[\alpha]_D^{25} = +2.0$ (c 1.95, CHCl₃); **IR** (neat): $v_{max} = 3350, 2959, 2113,$ 1743, 1672, 1529, 1370, 1246, 1214, 1040, 734 cm⁻¹; HRMS (ESI): calcd. for $C_{26}H_{32}N_8NaO_{12}$ [M + Na]⁺ m/z 671.2037; found m/z 671.2033; HPLC purity: 97.5% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: $H_2O/MeOH = 10:90$; $t_R = 1.86$).



TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.40$; ¹H NMR (400 MHz, CDCl₃) δ 7.40 - 7.28 (m, 5H), 5.45 - 5.24 (m, 4H), 5.19 -4.95 (m, 2H), 4.32 (dd, J = 12.4, 2.8 Hz, 1H), 4.18 (d, J = 3.2 Hz, 1H), 4.16 – 4.05 (m, 2H), 3.92 (s, 3H), 3.90 – 3.82 (m, 2H), 2.14 (s,

6H), 2.03 (s, 3H), 1.69 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.9, 170.6, 169.9, 169.9, 165.8, 156.4, 135.9, 128.6, 128.3, 127.9, 90.4, 74.6, 68.8, 67.2, 62.7, 62.1, 53.9, 52.1, 44.7, 22.8, 21.0, 20.7, 20.7; $[\alpha]_{D^{25}} = +15.5$ (c 0.37, CHCl₃); **IR** (neat): $v_{max} = 3333$, 2922, 2115, 1738, 1662, 1537, 1370, 1247, 1216, 1093, 1043 cm⁻¹; HRMS (ESI): calcd. for $C_{26}H_{32}N_8NaO_{12}$ [M + Na]⁺ m/z 671.2037; found m/z 671.2032; HPLC purity: 99.1% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; t_R =

1.91).

Following the general procedure, **7m** (180 mg, 0.268 mmol, 1.0 equiv) was subjected to diazidation in the presence of BI-OAc (164 mg, 0.536 mmol, 2.0 equiv) and azidotrimethylsilane (106 μ L, 0.804 mmol, 3.0 equiv) in 2 h to yield two isomers **20a** (142 mg, 70%) and **20b** (48.6 mg, 24%) after purification by silica gel column chromatography (petroleum ether/EtOAc = 2.5:1 to 1.5:1, v/v).



TLC: (petroleum ether/EtOAc = 1:1, v/v), $R_f = 0.53$; ¹H NMR (400 MHz, CDCl₃) δ 11.32 (s, 1H), 8.87 (d, J = 8.4 Hz, 1H), 6.11 (d, J = 9.2 Hz, 1H), 5.34 (dd, J = 5.2, 2.4 Hz, 1H), 5.26 – 5.20 (m, 1H), 4.70 – 4.58 (m, 2H), 4.21 (q, J = 10.0 Hz, 1H), 4.13 – 4,02 (m, 3H), 3.95 (s, 3H), 2.15 (s, 3H), 2.06 (s, 3H), 2.03 (s, 3H),

1.84 (s, 3H), 1.49 (s, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 170.5, 170.4, 170.1, 164.9, 162.9, 156.9, 152.6, 90.6, 84.0, 79.8, 73.8, 71.4, 67.9, 62.7, 62.3, 53.6, 49.8, 45.7, 28.2, 28.2, 27.9, 27.9, 22.9, 21.0, 20.8, 20.7; $[\alpha]_D^{25} = -62.1$ (*c* 0.70, CHCl₃); **IR** (neat): $v_{max} = 3270, 2963, 2932, 2114, 1745, 1729, 1643, 1610, 1368, 1255, 1215, 1143, 1113, 1053, 1028, 803 cm⁻¹;$ **HRMS (ESI)**: calcd. for C₂₉H₄₄N₁₀NaO₁₄ [M + Na]⁺*m/z*779.2936; found*m/z*779.2934; HPLC purity: 91.3% (Flow rate: 1.0 mL/min; Column: CHIRALCEL OD-H, 5 µm, 4.6 × 250 mm; Detector: Agilent G1314F 1260 VWD; Temperature: 25 °C; Mobile phase: hexane/2-isopropanol = 91:9, v/v;*t_R*= 6.23).



TLC: (petroleum ether/EtOAc = 1:1, v/v), $R_f = 0.49$; ¹H NMR (400 MHz, CDCl₃) δ 11.27 (s, 1H), 8.81 (d, J = 8.0 Hz, 1H), 5.95 (d, J = 9.2 Hz, 1H), 5.40 (td, J = 6.8, 2.8 Hz, 1H), 5.28 (dd, J = 6.8, 2.0 Hz, 1H), 4.40 – 4.30 (m, 2H), 4.28 – 4.16 (m, 2H), 4.12 (dd, J = 12.4, 6.0 Hz, 1H), 3.95 (s, 3H), 3.87 (dd, J = 10.4, 2.0 Hz, 1H), 2.17 (s,

3H), 2.14 (s, 3H), 2.04 (s, 3H), 1.83 (s, 3H), 1.50 (s, 9H), 1.47 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 170.5, 170.1, 170.0, 165.8, 162.8, 156.6, 152.4, 90.4, 83.9, 79.6, 75.3, 69.3, 67.6, 62.3, 62.2, 53.7, 50.9, 45.1, 28.2, 28.2, 28.0, 28.0, 23.0, 21.1, 20.8, 20.7; $[\alpha]_D^{25} = -5.8$ (*c* 1.50, CHCl₃); **IR** (neat): *v*_{max} = 3312, 2961, 2931, 2111, 1748, 1728, 1643, 1610, 1368, 1300, 1251, 1144, 1074, 1041, 734 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₉H₄₄N₁₀NaO₁₄ [M + Na]⁺ *m/z* 779.2936; found *m/z* 779.2930; HPLC purity: 93.5% (Flow rate: 1.0 mL/min; Column: CHIRALCEL OD-H, 5 µm, 4.6 × 250 mm; Detector: Agilent G1314F 1260 VWD; Temperature: 25 °C; Mobile phase: hexane/2-isopropanol = 95:5, v/v; *t_R* = 8.67).

5. Product Interconversion and Stereochemistry Determination



Condition a: Sodium methoxide was added to a stirred solution of substrate (1.0 equiv) in MeOH at room temperature. After being stirred overnight, the mixture was neutralized with Amberlite IR 120 (H^+) resin, then filtered and concentrated under *vacuum*. The residue was purified through flash column chromatography.

Condition b: To a stirred solution of substrate (1.0 equiv) in MeOH was added sodium methoxide (5 M in MeOH, 3.0 equiv) at room temperature. When the reaction was completed as indicated by TLC analysis, it was neutralized with Amberlite IR 120 (H+) resin and filtered. The filtrate was evaporated under reduced pressure and dissolved in MeOH, then *p*-toluenesulfonic acid monohydrate (3.0 equiv) was added. After being stirred overnight at room temperature, the reaction was quenched by addition of triethylamine (5.0 equiv). The mixture was concentrated in *vacuo* to afford the crude residue, which was purified through flash column chromatography.
Condition c: *p*-Toluenesulfonic acid monohydrate (3.0 equiv) was added to a stirred solution of substrate (1.0 equiv) in MeOH at room temperature. Triethylamine (5.0 equiv) was added after the reaction was completed (identified by TLC analysis). The mixture was next evaporated under reduced pressure; the resultant crude product was purified by silica gel column chromatography.

Condition d: To a solution of substrate (1.0 equiv) in MeCN at room temperature was added zinc nitrate hexahydrate (3.0 equiv). The resulting mixture was stirred at 50 °C until the reaction was completed as detected by TLC analysis. Then the mixture was diluted with water and extracted with EtOAc for three times. The combined organic extracts were washed with brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo to afford the crude residue. Sodium periodate (1.6 equiv) was added to the above diol (not shown) in THF/H₂O (2:1, v/v) at 0 °C. After being stirred for 3 h at the same temperature, the mixture was diluted with water and extracted with EtOAc for three times. The combined organic extracts were dried, filtered and concentrated in vacuo. The residue was dissolved in MeOH, to which was slowly added sodium borohydride (1.2 equiv) at 0 °C. The reaction was completed in 30 min according to TLC. Thus, water was added to quench the reaction and MeOH was removed under reduced pressure. The mixture was extracted with EtOAc for three times, before the combined organic layers were washed with brine, dried over Na₂SO₄, filtered, and concentrated under vacuum. To a solution of the above crude product in dry CH₂Cl₂ were added pyridine and benzoyl chloride (3.0 equiv) at 0 °C. Then the reaction mixture occurred at room temperature. After removal of solvent in *vacuo*, the residue was purified by silica gel column chromatography.

According to the procedure described for condition a, **8a** or **8b** (20.1 mg, 0.041 mmol, 1.0 equiv), sodium methoxide (1.1 mg, 0.021 mmol, 0.5 equiv) and MeOH (0.5 mL) were used. The mixture was stirred for 20 min at room temperature to give the hydrolyzed products **S9** (9.5 mg, 73%) or **S10** (9.0 mg, 69%), respectively, after purification by silica gel column chromatography (CH₂Cl₂/MeOH = 15:1 to 10:1, v/v). Similarly, **10a** or **10b** (10.0 mg, 0.018 mmol, 1.0 equiv) were converted into the corresponding **S9** (3.7 mg, 64%) or **S10** (3.8 mg, 66%) using sodium methoxide (5 M in MeOH, 10.9 μ L, 0.054 mmol, 3.0 equiv) and MeOH (0.5 mL) overnight.

According to the procedure of condition b, compounds **9a** or **9b** (8.2 mg, 0.021 mmol, 1.0 equiv), sodium methoxide (5 M in MeOH, 12.8 μ L, 0.064 mmol, 3.0 equiv) and *p*-toluenesulfonic acid monohydrate (12.2 mg, 0.064 mmol, 3.0 equiv) were used. As a result, the corresponding products **S9** (2.9 mg, 43%) or **S10** (2.6 mg, 39%) were prepared.

According to the procedure of condition c, 11a or 11b (13.5 mg, 0.030 mmol, 1.0 equiv),

p-toluenesulfonic acid monohydrate (5.8 mg, 0.091 mmol, 3.0 equiv) and MeOH (0.5 mL) were used. The mixture was stirred overnight at room temperature to respectively give **S9** (6.5 mg, 68%) or **S10** (6.4 mg, 67%). Similarly, both **S9** (5.8 mg, 71%) or **S10** (5.4 mg, 66%) were obtained employing **12a** or **12b** (10.2 mg, 0.026 mmol, 1.0 equiv), *p*-toluenesulfonic acid monohydrate (14.6 mg, 0.077 mmol, 3.0 equiv) and MeOH (0.5 mL).



TLC: (CH₂Cl₂/MeOH = 7:1, v/v), R_f = 0.45; ¹H NMR (400 MHz, CDCl₃/CD₃OD) δ 4.07 (d, J = 2.8 Hz, 1H), 3.97 (d, J = 10.4 Hz 1H), 3.82 (s, 3H), 3.75 – 3.69 (m, 3H), 3.55 – 3.51 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 166.4, 91.7, 73.9, 69.3, 69.0, 67.3, 63.0, 60.6, 53.6;

 $[\alpha]_{D}^{25} = +79.0 \ (c \ 0.07, MeOH); IR \ (neat): v_{max} = 3374, 2927, 2854, 2117, 1752, 1237, 1079, 925, 798 \ cm^{-1}; HRMS \ (ESI): calcd. for C₉H₁₄N₆NaO₇ <math>[M + Na]^+ m/z \ 341.0822;$ found $m/z \ 341.0820.$



0.12, MeOH); **IR** (neat): $v_{\text{max}} = 3389$, 2923, 2852, 2117, 1745, 1245, 1085, 924, 800 cm⁻¹; **HRMS (ESI)**: calcd. for C₉H₁₄N₆NaO₇ [M + Na]⁺ m/z 341.0822; found m/z 341.0818.

According to the procedure described for condition d, **12a** or **12b** (100 mg, 0.251 mmol, 1.0 equiv) were subjected to a sequence of transformation using zinc nitrate hexahydrate (224 mg, 0.754 mmol, 3.0 equiv), sodium periodate (83.5 mg, 0.402 mmol, 1.6 equiv), sodium borohydride (11.0 mg, 0.302 mmol, 1.2 equiv), and benzoyl chloride (106 μ L, 0.754 mmol, 3.0 equiv) in each step. Purification through flash column chromatography (petroleum ether/EtOAc = 25:1, V/V) afforded the corresponding products **13a** (17.4 mg, 16%, 4 steps) or **13b** (21.5 mg, 20%, 4 steps), respectively.



Condition a: A mixture of substrate (1.0 equiv) and potassium carbonate (0.5 equiv) in MeOH was stirred for 20 min at room temperature before being neutralized with Amberlite IR 120 (H^+) resin. The mixture was filtered and evaporated to give the crude product, which was purified through column chromatography.

Condition b: Sodium methoxide (3.0 equiv) was added to a stirring solution of substrate (1.0

equiv) in MeOH at room temperature. After being stirred overnight, the mixture was neutralized with Amberlite IR 120 (H^+) resin, then filtered and concentrated under *vacuum*. Purification by silica gel column chromatography with CH₂Cl₂/MeOH afforded the pure product.

Condition c: To a solution of substrate (1.0 equiv) in dry THF was added tetrabutylammonium fluoride (1 M in THF, 3.0 equiv) at room temperature. After being stirred for 3 h, the reaction was quenched by adding saturated NH₄Cl solution. The mixture was extracted with EtOAc for five times. The combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated *in vacuo*. The residue was dissolved in MeOH, to which was added *p*-toluenesulfonic acid monohydrate (1.0 equiv) at room temperature. Triethylamine was used to quench the reaction after overnight. The solvent was removed in *vacuo* and the crude product was purified via column chromatography to yield the corresponding product.

According to the procedure of condition a, compound **14b** (7.5 mg, 0.013 mmol, 1.0 equiv), potassium carbonate (1.0 mg, 0.007 mmol, 0.5 equiv) and MeOH (0.5 mL) were used for the synthesis of **S11** (3.8 mg, 73%) as a white amorphous powder after purification by silica gel column chromatography (CH₂Cl₂/MeOH = 8:1, v/v).

According to the procedure of condition b, compound **15b** (5.0 mg, 0.006 mmol, 1.0 equiv), sodium methoxide (1.0 mg, 0.019 mmol, 3.0 equiv) and MeOH (1 mL) were used for the synthesis of **S11** (1.3 mg, 52%) after purification by silica gel column chromatography ($CH_2Cl_2/MeOH = 8:1, v/v$).

9.6, 1.6 Hz, 1H), 3.35 (d, J = 1.6 Hz, 1H), 2.01 (s, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 175.0, 167.6, 92.0, 75.5, 71.5, 71.1, 69.8, 69.7, 65.0, 53.6, 52.9, 22.6; $[\alpha]_D^{25} = -59.0$ (*c* 0.29, CHCl₃/MeOH = 5:1, v/v); **IR** (neat): $v_{max} = 3279$, 2924, 2113, 1740, 1639, 1555, 1243, 1058, 1022 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₂H₁₉N₇NaO₈ [M + Na]⁺ *m/z* 412.1193; found *m/z* 412.1192.

According to the procedure described for condition a, compound 14c (20.0 mg, 0.036 mmol, 1.0 equiv), potassium carbonate (2.5 mg, 0.018 mmol, 0.5 equiv) and MeOH (1 mL) were used for the synthesis of 22 (10.6 mg, 76%) as a white amorphous powder after purification

by silica gel column chromatography ($CH_2Cl_2/MeOH = 8:1, v/v$).

According to the procedure described for condition b, compound **15c** (20.0 mg, 0.025 mmol, 1.0 equiv), sodium methoxide (4.0 mg, 0.075 mmol, 3.0 equiv) and MeOH (1 mL) were used for preparation of **22**. After purification by silica gel column chromatography (CH₂Cl₂/MeOH = 8:1, v/v) to give product **22** (6.1 mg, 63%).

According to the procedure described for condition c, compound **16b** (30.0 mg, 0.046 mmol, 1.0 equiv), tetrabutylammonium fluoride (1 M in THF, 137 μ L, 0.137 mmol, 3.0 equiv), 1 mL THF, *p*-toluenesulfonic acid monohydrate (8.7 mg, 0.046 mmol, 1.0 equiv) and MeOH (MeOH) were used for the synthesis of **22** (6.9 mg, 39%) after purification by silica gel column chromatography (CH₂Cl₂/MeOH = 8:1, v/v).



175.0, 167.3, 92.6, 74.4, 71.3, 69.7, 69.5, 66.2, 65.1, 53.8, 48.9, 22.7; $[a]_D^{25} = -51.5$ (*c* 0.60, CHCl₃/MeOH = 5:1, v/v); **IR** (neat): $v_{max} = 3364$, 2955, 2923, 2853, 2112, 1018, 800 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₂H₁₉N₇NaO₈ [M + Na]⁺ *m/z* 412.1193; found *m/z* 412.1189; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 70:30, v/v; *t_R* = 3.19).

According to the procedure described for condition a, compound **14d** (15.2 mg, 0.027 mmol, 1.0 equiv), potassium carbonate (1.9 mg, 0.014 mmol, 0.5 equiv) and MeOH (1 mL) were used for the synthesis of **S12** (7.5 mg, 71%) as a white amorphous powder after purification by silica gel column chromatography (CH₂Cl₂/MeOH = 8:1, v/v).

According to the procedure of condition b, compound **15d** (20.0 mg, 0.025 mmol, 1.0 equiv), sodium methoxide (4.0 mg, 0.075 mmol, 3.0 equiv) and MeOH (1 mL) were used for the preparation of **S12** (6.5 mg, 67%) after purification by silica gel column chromatography ($CH_2Cl_2/MeOH = 8:1, v/v$).



70.3, 70.1, 66.4, 64.9, 54.0, 48.7, 22.7; $[a]_D^{25} = -19.1$ (*c* 0.67, CHCl₃/MeOH = 5:1, v/v); **IR** (neat): $v_{max} = 3266$, 2112, 1744, 1657, 1252, 1034, 589, 568, 552 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₂H₁₉N₇NaO₈ [M + Na]⁺ *m/z* 412.1193; found *m/z* 412.1191; HPLC purity: 99.6% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 70:30, v/v; $t_R = 4.88$).

According to the procedure of condition a, compound **14a** (5.1 mg, 0.009 mmol, 1.0 equiv), potassium carbonate (0.6 mg, 0.005 mmol, 0.5 equiv) and MeOH (0.5 mL) were used. Purification by silica gel column chromatography with $CH_2Cl_2/MeOH$ (8:1, v/v) yielded **S13** (2.4 mg, 67%) as a white amorphous powder.

According to the procedure of condition b, compound **15a** (9.3 mg, 0.012 mmol, 1.0 equiv), sodium methoxide (1.9 mg, 0.035 mmol, 3.0 equiv) and MeOH (1 mL) were used for the synthesis of **S13** (2.3 mg, 52%) after purification by silica gel column chromatography ($CH_2Cl_2/MeOH = 8:1, v/v$).

According to the procedure of condition c, compound **16a** (30.1 mg, 0.046 mmol, 1.0 equiv), tetrabutylammonium fluoride (1 M in THF, 137 μ L, 0.137 mmol, 3.0 equiv), 1 mL THF, *p*-toluenesulfonic acid monohydrate (8.7 mg, 0.046 mmol, 1.0 equiv) and 1 mL MeOH were used. Purification by silica gel column chromatography with CH₂Cl₂/MeOH (8:1, v/v) provided **S13** (6.2 mg, 35%).

HO HO
$$N_3$$

ACHN N_3
S13
HO N_3
ACHN N_3
S13
HILC: (CH₂Cl₂/MeOH = 5:1, v/v), R_f = 0.53; ¹H NMR (400 MHz, CD₃OD) δ 4.14 (dd, J = 10.8, 1.2 Hz, 1H), 4.04 (t, J = 10.0 Hz, 1H), 3.90 (s, 3H), 3.86 (t, J = 10.0 Hz, 1H), 3.78 – 3.71 (m, 2H), 3.70 – 3.58 (m, 2H), 3.54 (dd, J = 9.2, 1.2 Hz, 1H), 2.02 (s, 3H);

¹³**C** NMR (100 MHz, CD₃OD) δ 174.8, 167.8, 93.2, 74.7, 71.2, 69.5, 66.2, 65.1, 54.1, 53.1, 29.5, 22.7; $[\alpha]_D^{25} = -88.3$ (*c* 0.12, CHCl₃/MeOH = 10:1, v/v); **IR** (neat): $v_{max} = 3293$, 2954, 2922, 2852, 2114, 1745, 1640, 1557, 1307, 1235, 1115, 1017, 972 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₂H₁₉N₇NaO₈ [M + Na]⁺ *m/z* 412.1193; found *m/z* 412.1190.



To a stirred solution of compound **17b** (72.0 mg, 0.090 mmol, 1.0 equiv) in MeOH (1 mL) was added sodium methoxide (5 M in MeOH, 54.3 μ L, 0.271 mmol, 3.0 equiv) at room temperature. The mixture was stirred at 30 °C overnight, before being neutralized with Amberlite IR 120 (H⁺) resin. Then the mixture was filtered, and the filtrate was concentrated under reduced pressure. Purification of the crude residue by column chromatography (CH₂Cl₂/MeOH = 15:1 to 10:1, v/v) afforded **24** (22.6 mg, 61%).

 $\begin{array}{c} \text{HO} \\ \text{HO} \\ \text{CF}_{3}\text{COHN} \\ \text{24} \\ \begin{array}{c} \text{N}_{3} \\ \text{24} \end{array} \right) \begin{array}{c} \text{TLC:} (\text{CH}_{2}\text{Cl}_{2}/\text{MeOH} = 7:1, \text{ v/v}), \text{ R}_{f} = 0.50; \ ^{1}\text{H} \text{ NMR} (400 \text{ MHz}, \text{ CD}_{3}\text{OD}) \\ \text{CD}_{3}\text{OD}) \delta 4.68 (\text{dd}, J = 9.6, 1.6 \text{ Hz}, 1\text{H}), 4.23 (\text{dd}, J = 9.6, 1.2 \text{ Hz}, \text{ 1H}), 4.16 (\text{d}, J = 5.6 \text{ Hz}, 1\text{H}), 3.96 (\text{dd}, J = 5.6, 1.6 \text{ Hz}, 1\text{H}), 3.87 - 3.78 (\text{m}, 2\text{H}), 3.66 (\text{dd}, J = 11.6, 5.6 \text{ Hz}, 1\text{H}), 3.42 (\text{dd}, J = 9.2, 1.2 \text{ Hz}, \text{ CD}_{3}\text{OD}) \end{array}$

Hz, 1H); ¹³C NMR (100 MHz, CD₃OD) δ 170.1, 159.0 (q, J = 37.5 Hz), 117.2 (q, J = 285.1), 90.1, 74.8, 70.7, 70.2, 64.9, 64.7, 54.9, 46.4; $[\alpha]_D^{25} = -3.2$ (*c* 0.51, CHCl₃/MeOH = 6:1, v/v); IR (neat): $v_{max} = 3273$, 2926, 2118, 1709, 1213, 1182, 1158, 1072, 1018, 729 cm⁻¹; HRMS (ESI): calcd. for C₁₁H₁₃F₃N₈NaO₆ [M + Na]⁺ *m/z* 433.0808; found *m/z* 433.0802; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 50:50, v/v; *t_R* = 2.10).



 $AcO AcO OAc N_3 AcHN OAc OAc O2M AcHN OAC$

To a stirred solution of compound **18a** (200 mg, 0.326 mmol) in CH_2Cl_2 (4 mL) was added trifluoroacetic acid (0.4 mL) at 0 °C. The mixture was allowed to stir at room temperature for 3 h, before it was concentrated directly in *vacuo*. The crude product

was purified by column chromatography (CH₂Cl₂/MeOH = 10:1 to 8:1, +5‰ NH₃·H₂O, v/v) to give 140 mg (84%) of product **28** as a white amorphous powder. **TLC**: (CH₂Cl₂/MeOH = 8:1, v/v), R_f = 0.48; ¹**H NMR** (400 MHz, CDCl₃ + CD₃OD) δ 5.35 (dd, *J* = 5.4, 2.2 Hz, 1H), 5.22 - 5.14 (m, 1H), 4.61 (dd, *J* = 12.4, 2.4 Hz, 1H), 4.14 - 4.00 (m, 3H), 3.92 (s, 3H), 3.91 - 3.86 (m, 1H), 3.35 (dd, *J* = 10.8, 3.6 Hz, 1H), 2.09 (s, 3H), 2.04 (s, 3H), 2.01 (s, 3H), 1.92 (s, 3H); ¹³C **NMR** (150 MHz, CDCl₃ + MeOD) δ 172.5, 170.8, 170.7, 170.24, 164.9, 90.7, 72.4, 71.2, 67.5, 63.6, 62.0, 53.6, 50.7, 45.84, 22.7, 20.9, 20.6, 20.5; **[a]**_D²⁵ = +15.7 (*c* 0.70, CHCl₃); **IR** (neat): *v*_{max} = 3372, 3059, 2119, 1743, 1669, 1373, 1200, 1136, 1047, 722 cm⁻¹; **HRMS** (**ESI**): calcd. for C₁₈H₂₆N₈NaO₁₀ [M + Na]⁺ *m*/*z* 537.1670; found *m*/*z* 537.1668; HPLC purity: 99.9% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70; *t_R* = 3.43).

Preparation of TfN₃: Sodium azide (13.0 mg, 0.200 mmol, 10.0 equiv) was dissolved in water (1 mL) and cooled in an ice bath and treated with CH₂Cl₂ (1.5 mL). Trifluoromethanesulfonic

anhydride (16.4 μ L, 0.100 mmol, 5.0 equiv) was added slowly at 0 °C to the resulting biphasic mixture. The reaction was stirred at 0 °C for 2 h, the organic layer was separated and the aqueous phase was washed with CH₂Cl₂ as less volume as possible. The combined organic layers were washed with saturated NaHCO₃ solution, dried over anhydrous Na₂SO₄, filtered and used without further purification.



To a stirred solution of compound **28** (10.2 mg, 0.019 mmol, 1.0 equiv) in CH_2Cl_2 (1 mL) was added 4-(dimethylamino)pyridine (7.1 mg, 0.058 mmol, 3.0 equiv). The above freshly prepared trifluoromethanesulfonic azide (TfN₃) solution was added

dropwise at 0 °C. The ice-bath was removed and the mixture was stirred overnight at room temperature. The reaction was quenched with saturated aqueous NaHCO₃ and extracted with CH₂Cl₂, before the combined organic layers were dried and concentrated. Purification of the residue by a flash column chromatography (petroleum ether/EtOAc = 2:1 to 1:1, v/v) furnished **29** (5.0 mg, 47%, 68% brsm) with **28** (3.0 mg) recovered. **TLC**: (petroleum ether/EtOAc = 1:3, v/v), R_f = 0.54; ¹**H NMR** (400 MHz, CDCl₃) δ 5.90 (d, *J* = 8.0 Hz, 1H), 5.38 – 5.24 (m 2H), 4.81 (d, *J* = 10.8 Hz, 1H), 4.57 – 4.48 (m, 2H), 4.20 (dd, *J* = 12.8, 5.2 Hz, 1H), 4.06 (d, *J* = 3.2 Hz, 1H), 3.93 (s, 3H), 3.45 (s, 1H), 2.17 (s, 3H), 2.07 (3, 3H), 2.04 (s, 3H), 2.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 171.1, 170.5, 169.9, 164.6, 90.5, 70.3, 69.9, 67.9, 63.0, 61.8, 57.4, 53.6, 48.2, 23.5, 21.0, 20.8, 20.7; **[a**]_D²⁵ = -2.0 (*c* 0.70, CHCl₃); **IR** (neat): *v*_{max} = 3269, 2960, 2110, 1743, 1370, 1255, 1212, 1048, 1030, 755 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₈H₂₄N₁₀NaO₁₀ [M + Na]⁺ *m*/z 563.1575; found *m*/z 563.1573. The supplementary crystallographic data of **29** [m.p.: 186 – 188 °C (hexane/CHCl₃ = 5:1, v/v)] have been deposited in Cambridge Crystallographic Data Centre (CCDC 1975129). These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.

Triethylamine (5.41 μ L, 0.039 mmol, 2.0 equiv) and benzyl chloroformate (5.48 μ L, 0.039 mmol, 2.0 equiv) were successively added to a solution of **28** (10.0 mg, 0.019 mmol, 1.0 equiv) in dry THF (1 mL) under an argon atmosphere. After being stirred for 3 h at room temperature, the reaction mixture was concentrated in *vacuo*. The crude product was purified by column chromatography (petroleum ether/EtOAc = 1.5:1 to 1:2, v/v) to afford **19a** (10.2 mg, 81%).

To a stirred solution of **28** (10.0 mg, 0.019 mmol, 1.0 equiv) in dry DMF (1 mL) were added *N*,*N'*-bis-tert-butoxycarbonylthiourea (5.9 mg, 0.021 mmol, 1.1 equiv) and triethylamine (54.1 μ L, 0.389 mmol, 20.0 equiv). The mixture was cooled to 0 °C, followed by addition of mercurous chloride (9.0 mg, 0.019 mmol, 1.0 equiv). The reaction was concentrated in *vacuo*

after being stirred for 5 h at the same temperature. The residue was dissolved in EtOAc, which was further subjected to filtration through Celite[®]. After the resultant filtrate was washed with water and brine, the organic layer was dried and evaporated under reduced pressure. Purification of the crude product via column chromatography (petroleum ether/EtOAc = 2:1, v/v) yielded **20a** (9.3 mg, 63%).



According to the procedure for the synthesis of **28**, the diazidation product **18b** (60.2 mg, 0.098 mmol) and trifluoroacetic acid (0.4 mL) were used for the synthesis of **S14**. The crude product was purified by silica gel column chromatography (petroleum ether/acetone =

3:1 to 1:2 , +5‰ NH₃·H₂O, v/v) to give **S14** (39.2 mg, 78%). **TLC**: (CH₂Cl₂/MeOH = 8:1, v/v), R_f = 0.47; ¹**H NMR** (600 MHz, CD₃OD) δ 5.38 – 5.30 (m, 2H), 4.33 (dd, *J* = 12.6, 2.4 Hz, 1H), 4.16 (d, *J* = 2.4 Hz, 1H), 4.11 (dd, *J* = 12.0, 4.8 Hz, 1H), 3.95 – 3.84 (m, 5H), 2.79 (dd, *J* = 10.8, 3.6 Hz, 1H), 2.12 (s, 3H), 2.11 (s, 3H), 2.01 (s, 3H), 1.92 (s, 3H); ¹³C **NMR** (150 MHz, CD₃OD) δ 174.3, 172.4, 171.6, 171.4, 167.5, 91.8, 75.5, 70.0, 68.6, 66.5, 63.2, 54.2, 53.7, 47.6, 22.9, 21.1, 20.8, 20.6; [*a*]_D²⁵ = +32.8 (*c* 0.43, CHCl₃); **IR** (neat): *v*_{max} = 3275, 2959, 2925, 2855, 2109, 1743, 1731, 1368, 1215, 1039, 735 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₈H₂₆N₈NaO₁₀ [M + Na]⁺ *m*/*z* 537.1670; found *m*/*z* 537.1668; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 20:80, v/v; *t_R* = 3.40).

According to the above procedure for synthesis of compound **19a**, triethylamine (4.38 μ L, 0.032 mmol, 2.0 equiv), benzyl chloroformate (4.44 μ L, 0.032 mmol, 2.0 equiv) and compound **S14** (8.1 mg, 0.016 mmol, 1.0 equiv) were used to access **19b** (7.7 mg, 76%) after the crude product was purified by silica gel column chromatography (petroleum ether/EtOAc = 1.5:1 to 1:2).

According to the above procedure for the synthesis of compound **20a**, compound **S14** (10.2 mg, 0.020 mmol, 1.0 equiv), *N*,*N*'-bis-tert-butoxycarbonylthiourea (6.0 mg, 0.022 mmol, 1.1 equiv), triethylamine (55.2 μ L, 0.397 mmol, 20.0 equiv) and mercurous chloride (7.4 mg, 0.020 mmol, 1.0 equiv) were used. The crude product was purified by silica gel column chromatography (petroleum ether/EtOAc = 2:1 to 1:1, v/v) to afford **20b** (9.2 mg, 61%).

6. Product Diversification



BZO ACHN BZO NH₂ S15 A mixture of compound **15c** (140 mg, 0.174 mmol), 10% Pd/C (14.0 mg), and MeOH (10 mL) was stirred under a balloon of hydrogen at ambient pressure and room temperature for 4 h. The reaction mixture was then filtered through a short plug of Celite[®]

and the filtrate was concentrated in *vacuo*. Purification of the residue by silica gel column chromatography (petroleum ether/EtOAc = 2:1, v/v) afforded **S15** (110 mg, 81%). **TLC**: (petroleum ether/EtOAc = 1:1, v/v), R_f = 0.50; ¹**H NMR** (400 MHz, CDCl₃) δ 8.21 – 8.14 (m, 2H), 8.03 – 7.97 (m, 4H), 7.96 – 7.90 (m, 2H), 7.64 – 7.45 (m, 6H), 7.45 – 7.30 (m, 6H), 5.98 (dd, *J* = 10.8, 3.6 Hz, 1H), 5.95 – 5.80 (m, 1H), 5.84 (dd, *J* = 5.6, 2.0 Hz, 1H), 5.58 (d, *J* = 9.6 Hz, 1H), 5.19 (dd, *J* = 12.4, 2.8 Hz, 1H), 4.81 (dd, *J* = 10.4, 2.0 Hz, 1H), 4.56 (dd, *J* = 12.0, 7.6 Hz, 1H), 4.44 (q, *J* = 10.0 Hz, 1H), 4.32 (d, *J* = 3.6 Hz, 1H), 3.88 (s, 3H), 2.13 (s, 2H), 1.81 (s, 3H); ¹³C **NMR** (100 MHz, CDCl₃) δ 170.3, 170.1, 166.4, 166.2, 165.9, 133.7, 133.4, 132.9, 130.2, 130.0, 129.8, 129.8, 129.6, 129.4, 129.4, 128.6, 128.5, 128.5, 128.5, 128.3, 86.3, 72.0, 71.2, 70.3, 69.5, 63.8, 63.7, 53.0, 46.5, 23.3; [**a**] \mathbf{p}^{25} = +4.4 (*c* 0.50, CHCl₃); **IR** (neat): *v*_{max} = 3378, 2928, 2855, 2111, 1718, 1261, 1107, 1094, 1069, 1026, 708 cm⁻¹; **HRMS (ESI)**: calcd. for C₄₀H₃₇N₅NaO₁₂[M + Na]⁺ *m/z* 802.2336; found *m/z* 802.2325; HPLC purity: 97.3% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2489 UV/Vis; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 100:0 → 5:95, v/v; *t_R* = 18.64).



A mixture of **15c** (50.3 mg, 0.062 mmol, 1.0 equiv), lithium iodide (83.1 mg, 0.621 mmol, 10.0 equiv), and pyridine (1 mL) was stirred overnight at 90 °C under an argon atmosphere. After being cooled to room temperature, the reaction was concentrated under

reduced pressure. Subjection of the crude residue to purification via column chromatography (CH₂Cl₂/MeOH = 20:1, v/v) afforded **21** (42.5 mg, 86%). **TLC**: (CH₂Cl₂/MeOH = 8:1, v/v), R_f = 0.45; ¹**H NMR** (400 MHz, CD₃OD) δ 8.17 (d, *J* = 7.2 Hz, 2H), 8.05 (d, *J* = 7.2 Hz, 2H), 8.00 (d, *J* = 7.2 Hz, 2H), 7.94 (d, *J* = 7.2 Hz, 2H), 7.69 – 7.34 (m, 12H), 6.05 – 5.93 (m, 2H),

5.54 (dd, J = 10.4, 3.6 Hz, 1H), 5.08 (dd, J = 12.4, 2.4 Hz, 1H), 4.68 – 4.47 (m, 4H), 4.44 (d, J = 3.6 Hz, 1H), 1.80 (s, 3H); ¹³**C** NMR (100 MHz, CD₃OD) δ 173.3, 167.6, 167.1, 167.0, 166.8, 150.0, 134.7, 134.6, 134.6, 134.3, 131.3, 131.0, 131.0, 131.0, 130.9, 130.8, 130.7, 130.3, 129.7, 129.6, 129.6, 129.5, 73.3, 73.1, 71.6, 69.6, 64.9, 63.6, 46.0, 30.7, 22.7; $[\alpha]_D^{25} = +21.4$ (*c* 0.44, CHCl₃ : MeOH = 4:1, v/v); **IR** (neat): $v_{max} = 3398$, 2119, 1725, 1662, 1261, 1090, 1069, 1025, 707, 686 cm⁻¹; **HRMS (ESI)**: calcd.for C₃₉H₃₃N₇NaO₁₂ [M + Na]⁺ *m/z* 814.2085; found *m/z* 814.2055; HPLC purity: 99.1% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; *t_R* = 2.28).



To a stirred solution of **15c** (20.0 mg, 0.025 mmol, 1.0 equiv) in MeOH (1 mL) was added NaOH aq. (1 M in H₂O, 50.0 μ L, 0.050 mmol, 2.0 equiv) at ambient temperature. The resultant mixture was allowed to stir for 5 h, then neutralized with Amberlite IR 120 (H⁺)

resin and filtered. The filtrate was evaporated under reduced pressure to afford the crude residue, which was purified through reverse phase silica gel column chromatography (MeOH/H₂O = 1:9, v/v) to give acid **23** (8.4 mg, 81%). ¹**H NMR** (400 MHz, CD₃OD) δ 4.27 – 4.13 (m, 2H), 4.11 – 4.01 (m, 2H), 3.88 – 3.70 (m, 2H), 3.66 (dd, *J* = 10.8, 4.8 Hz, 1H), 3.57 (d, *J* = 8.0 Hz, 1H), 2.01 (s, 3H); ¹³**C NMR** (100 MHz, CD₃OD) δ 174.8, 169.7, 93.7, 74.1, 72.1, 69.9, 66.6, 64.8, 22.8; $[\alpha]_D^{25} = -82.0$ (*c* 0.60, MeOH); **IR** (neat): $v_{max} = 3277$, 2946, 2110, 1631, 1558, 1376, 1070, 1019, 679 cm⁻¹; **HRMS** (**ESI**): calcd. for C₁₁H₁₇N₇NaO₈ [M + Na]⁺ *m/z* 398.1036; found *m/z* 398.1031; HPLC purity: 97.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 100:0, v/v; *t_R* = 1.74).



According to the procedure for the preparation of **23**, compound **15d** (20.3 mg, 0.025 mmol, 1.0 equiv), NaOH aq. (1 M in H₂O, 75.6 μ L, 0.076 mmol, 3.0 equiv) and MeOH (1 mL) were used for the synthesis of **S16**. The crude product was purified through reverse phase silica gel column chromatography (MeOH/H₂O = 1:9 to 1:4, v/v) to give **S16** (7.4 mg, 78%). ¹H **NMR** (400 MHz, CD₃OD) δ 4.33 (d, *J* = 1.6 Hz, 1H), 4.03 (d, *J* = 6.0 Hz, 2H), 3.95 – 3.82 (m, 2H), 3.67 (dd, *J* = 11.2, 5.2 Hz, 1H), 3.59 – 3.59 (m, 2H), 2.02 (s, 3H); ¹³C **NMR** (150 MHz, CD₃OD) δ 175.5, 171.6, 92.7, 76.1, 72.6, 71.0, 69.9, 67.6, 64.0, 48.9, 22.7; **[a]**_D²⁵ = +2.1 (*c*

0.57, MeOH); **IR** (neat): $v_{\text{max}} = 3273$, 2948, 2840, 2109, 1624, 1563, 1375, 1247, 1023 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₁H₁₇N₇NaO₈ [M + Na]⁺ *m*/*z* 398.1036; found *m*/*z* 398.1020; HPLC purity: 97.2% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 90:10, v/v; $t_R = 1.68$).



condition a: 4-pentynyl alcohol, CuSO₄· 5H₂O sodium ascorbate, THF/H₂O, RT

OBz _{NHA}d

25

CE₂COHN

condition b (for **32**): NaOH, MeOH, RT condition c (for **S17**): Lil, pyridine, 90 °C condition d (for **31**): Pd/C, H₂, MeOH, RT

4-Pentynyl alcohol (38.6 μ L, 0.414 mmol, 2.2 equiv), CuSO₄·5H₂O (18.8 mg, 0.075 mmol, 0.4 equiv) and sodium ascorbate (149 mg, 0.754 mmol, 4.0 equiv) were added sequentially to a stirred solution of **17a** (150 mg, 0.188 mmol, 1.0 equiv) in THF (7.5 mL) and H₂O (7.5 mL) at room temperature. The mixture was allowed to stir at the same temperature for 2 h

and diluted with H₂O. Filtration through a pad of Celite[®] was conducted to remove insoluble salts, and the filtrate was extracted with EtOAc. Next, the combined organic layers were dried over Na₂SO₄, filtered and concentrated. Subjection of the residue to flash column chromatography (CH₂Cl₂/MeOH = 15:1, v/v) on silica gel afforded compound **25** (151 mg, 83%). **TLC**: (CH₂Cl₂/MeOH = 8:1, v/v), R_f = 0.54; ¹**H NMR** (400 MHz, CD₃OD) δ 8.25 – 8.18 (m, 2H), 8.02 (s, 1H), 7.93 – 7.87 (m, 4H), 7.85 (s, 1H), 7.74 – 7.53 (m, 5H), 7.46 (t, *J* = 8.0 Hz, 2H), 7.39 (t, *J* = 8.0 Hz, 2H), 6.95 (d, *J* = 8.0 Hz, 1H), 5.95 (dd, *J* = 8.0, 1.6 Hz, 1H), 5.78 – 5.71 (m, 1H), 5.17 – 5.06 (m, 2H), 5.02 (dd, *J* = 10.8, 6.8 Hz, 1H), 4.63 (dd, *J* = 10.8, 1.6 Hz, 1H), 4.60 (s, 1H), 4.39 (dd, *J* = 12.8, 4.8 Hz, 1H), 3.62 (s, 3H), 3.57 – 3.50 (m, 4H), 2.74 (t, *J* = 7.2 Hz, 2H), 2.49 (t, *J* = 7.6 Hz, 2H), 1.88 – 1.70 (m, 4H), 1.82 (s, 3H); ¹³C **NMR** (100 MHz, CD₃OD) δ 173.7, 167.5, 166.7, 166.6, 165.4, 159.4 (q, *J* = 37 Hz), 150.8, 148.3, 135.1, 134.9, 134.5, 131.3, 130.8, 130.7, 130.7, 130.4, 130.2, 129.9, 129.8, 129.7, 129.6, 129.5, 126.2, 123.0, 117.2 (q, *J* = 285 Hz), 92.6, 71.3, 70.8, 69.8, 63.1, 62.0, 61.8, 58.5, 54.8, 49.9, 47.2, 33.1, 32.5, 22.8, 22.5, 22.4; [**a**]_D²⁵ = +36.4 (*c* 0.52, CHCl₃); **IR** (neat): *v*_{max} = 3339, 2936, 1721, 1260, 1106, 1068, 708 cm⁻¹; **HRMS (ESI)**: calcd. for C4₅H₄₇F₃N₈NaO₁₃ [M +

Na]⁺ m/z 987.3112; found m/z 987.3072; HPLC purity: 99.9% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; t_R = 1.90).



According to the procedure for the preparation of 23, compound 17a (80.1 mg, 0.101 mmol, 1.0 equiv), NaOH aq. (1 M in H₂O, 0.302 mL, 0.302 mmol, 3.0 equiv) and 2 mL MeOH were employed for synthesis of 32 (32.1 mg, 68%) after the crude

product was purified by silica gel column chromatography (CH₂Cl₂/MeOH = 5:1 to 2:1, v/v). **TLC**: (CH₂Cl₂/MeOH = 1:1, v/v), R_f = 0.50; ¹**H NMR** (400 MHz, CD₃OD) δ 4.68 (t, *J* = 3.6 Hz, 1H), 4.59 (dd, *J* = 11.2, 4.0 Hz, 1H), 4.48 (d, *J* = 11.2 Hz, 1H), 4.02 (d, *J* = 2.8 Hz, 1H), 3.92 – 3.83 (m, 1H), 3.83 – 3.68 (m, 2H), 3.62 (d, *J* = 8.0 Hz, 1H), 2.07 (s, 3H); ¹³**C NMR** (100 MHz, CD₃OD) δ 173.5, 170.6, 158.9 (q, *J* = 37 Hz), 117.2 (q, *J* = 285 Hz), 93.5, 72.3, 69.4, 69.0, 64.4, 63.1, 49.9, 44.8, 22.9; $[\alpha]_D^{25} = -154.4$ (*c* 1.00, MeOH); **IR** (neat): $v_{max} = 3259$, 2933, 2113, 1714, 1635, 1380, 1214, 1186, 1153 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₃H₁₇F₃N₈NaO₈ [M + Na]⁺ *m/z* 493.1019; found *m/z* 493.1018; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; *t_R* = 1.69).



According to the procedure for synthesis of **21**, a mixture of compound **17a** (100 mg, 0.126 mmol, 1.0 equiv), lithium iodide (168 mg, 1.26 mmol, 10.0 equiv) and pyridine (2 mL) was stirred overnight at 90 °C. Purification of crude product by flash column

chromatography (CH₂Cl₂/MeOH = 20:1 to 15:1, v/v) yielded **S17** (74.7 mg, 76%). **TLC**: (CH₂Cl₂/MeOH = 6:1, v/v), R_f = 0.61; ¹**H NMR** (400 MHz, CD₃OD) δ 8.13 (d, *J* = 7.6 Hz, 2H), 8.02 (d, *J* = 7.2 Hz, 2H), 7.93 (d, *J* = 7.2 Hz, 2H), 7.70 – 7.34 (m, 9H), 6.10 – 6.02 (m, 1H), 5.98 (dd, *J* = 8.0, 2.0 Hz, 1H), 5.16 (dd, *J* = 12.4, 2.0 Hz, 1H), 4.70 (dd, *J* = 11.2, 1.6 Hz, 1H), 4.65 – 4.55 (m, 3H), 4.49 (dd, *J* = 12.4, 4.8 Hz, 1H), 4.39 (dd, *J* = 11.2, 4.4 Hz, 1H), 4.00 (d, *J* = 3.2 Hz, 1H), 2.01 (s, 3H); ¹³**C NMR** (150 MHz, CD₃OD) δ 173.4, 170.3, 167.6, 166.9, 166.7, 158.9 (q, *J* = 37.5 Hz), 134.8, 134.6, 134.4, 131.2, 131.0, 130.9, 130.7, 130.5, 130.5, 129.6, 129.6, 129.5, 129.4, 117.1 (q, *J* = 285 Hz), 93.7, 71.5, 69.9, 68.9, 63.5, 63.3, 50.3, 44.9, 22.9; [**a**]_D²⁵ = +45.0 (*c* 0.50, CHCl₃); **IR** (neat): *v*_{max} = 2117, 1718, 1654, 1452, 1259, 1177, 1091, 1068, 709 cm⁻¹; **HRMS (ESI)**: calcd. for C₃₄H₂₉F₃N₈NaO₁₁ [M + Na]⁺ *m/z* 805.1806; found *m/z* 805.1803; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; *t_R* = 2.22).



According to the procedure for synthesis of **S15**, compound **17a** (100 mg, 0.126 mmol) was subjected to hydrogenation with 10% Pd/C (10.2 mg) in MeOH (5 mL) under a balloon of hydrogen. Amine **31** (58.9 mg, 63%) was obtained after purification by silica

gel column chromatography (petroleum ether/EtOAc = 1.5:1, v/v). TLC: (petroleum ether/EtOAc = 1:2, v/v), $R_f = 0.56$; ¹H NMR (400 MHz, CDCl₃) δ 8.15 – 8.07 (m, 2H), 8.06 – 7.99 (m, 2H), 7.98 – 7.89 (m, 2H), 7.72 (d, J = 9.2 Hz, 1H), 7.64 – 7.32 (m, 9H), 6.76 (d, J = 9.2 Hz, 1H), 5.91 – 5.85 (m, 1H), 5.75 (dd, J = 4.8, 1.6 Hz, 1H), 5.31 (dd, J = 12.4, 2.4 Hz, 1H), 4.78 – 4.69 (m, 2H), 4.46 (dd, J = 12.4, 7.2 Hz, 1H), 4.43 – 4.35 (m, 1H), 3.89 (s, 3H), 3.84 (d, J = 3.2 Hz, 1H), 2.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 170.0, 166.4, 166.4, 165.3, 157.3 (q, J = 38 Hz), 133.6, 133.5, 133.1, 130.1, 129.9, 129.6, 129.6, 129.2, 129.0, 128.6, 128.4, 115.4 (q, J = 286 Hz), 86.0, 72.4, 69.2, 67.6, 63.8, 62.7, 53.2, 49.8, 44.5, 23.5; $[\mathbf{a}]_{D}^{25} = +124.8$ (*c* 0.40, CHCl₃); **IR** (neat): $v_{max} = 3377$, 3287, 2115, 1718, 1259, 1250, 1089, 1068, 1025, 708 cm⁻¹; **HRMS (ESI)**: calcd. for C₃₅H₃₃F₃N₆NaO₁₁ [M + Na]⁺ *m*/*z* 793.2057; found *m*/*z* 793.2049; HPLC purity: 92.9% (Flow rate: 1.0 mL/min; Column: CHIRALCEL OD-H, 5 µm, 4.6 × 250 mm; Detector: Agilent G1314F 1260 VWD; Temperature: 25 °C; Mobile phase: hexane/2-isopropanol (0.1% Et₂NH) = 25:75, v/v; *t_R* = 4.90).

$$\begin{array}{c|c} BZO & CO_2Me \\ \hline CF_3COHN & OBZ & NHAC| \\ \hline 17b & N_3 \end{array} \begin{array}{c} Lil, pyridine \\ 90 \ ^\circ C, \ 83\% \end{array} \begin{array}{c} BZO & OBZ & CO_2H \\ \hline CF_3COHN & OBZ & NHAC| \\ \hline CF_3COHN & OBZ & NHAC| \\ \hline CF_3COHN & OBZ & N_3 \end{array}$$

Following the procedure for synthesis of **21**, a mixture of **17b** (52.3 mg, 0.066 mmol, 1.0 equiv), lithium iodide (87.9 mg, 0.657 mmol, 10.0 equiv) and pyridine (1 mL) was stirred overnight at 90 °C. The residue was purified by column chromatography (CH₂Cl₂/MeOH = 20:1 to 10:1, v/v) to afford acid **30** (42.6 mg, 83%). **TLC**: (CH₂Cl₂/MeOH = 7:1, v/v), R_f = 0.58; ¹**H NMR** (400 MHz, CD₃OD) δ 8.11 (dd, *J* = 8.0, 1.2 Hz, 2H), 7.96 (dd, *J* = 8.0, 1.2 Hz, 2H), 7.90 (dd, *J* = 8.4, 1.6 Hz, 2H); 7.68 – 7.30 (m, 9H), 5.98 – 5.85 (m, 2H), 5.10 (dd, *J* = 12.4, 3.2 Hz, 1H), 4.78 (dd, *J* = 10.4, 2.0 Hz, 1H), 4.70 (dd, *J* = 12.4, 6.4 Hz, 1H), 4.61 – 4.50 (m, 1H), 4.44 (t, *J* = 5.2 Hz, 1H), 4.21 (d, *J* = 4.8 Hz, 1H), 1.95 (s, 3H); ¹³C **NMR** (100 MHz, CD₃OD) δ 173.8, 172.6, 167.6, 167.0, 166.7, 159.0 (q, *J* = 38 Hz), 134.7, 134.4, 134.3, 131.1, 131.0, 130.9, 130.8, 130.6, 130.5, 129.7, 129.5, 129.4, 117.1 (q, *J* = 285 Hz), 93.6, 72.6, 70.9, 63.8, 62.6, 59.5, 50.2, 46.0, 22.9; **[a**]_{D²⁵} = +82.8 (*c* 0.40, CHCl₃); **IR** (neat): *v*_{max} = 3276, 3083, 2889, 2114, 1722, 1647, 1262, 1177, 1093, 1069, 707 cm⁻¹; **HRMS (ESI)**: calcd. for C₃₄H₂₉F₃N₈NaO₁₁ [M + Na]⁺ *m/z* 805.1806; found *m/z* 805.1811; HPLC purity: 99.8% (Flow

rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 μ m, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; t_R = 1.89).



Following the procedure for synthesis of **S15**, **18a** (80.1 mg, 0.130 mmol), 10% Pd/C (8.0 mg) and MeOH (2 mL) were used under a balloon of hydrogen to give **26** (47.5 mg, 62%) and **27** (9.0 mg, 12%), which were purified by silica gel column chromatography (petroleum ether/acetone = 3:1 to 1:1, v/v).



(m, 3H), 3.84 (s, 3H), 2.13 (s, 3H), 2.07 (s, 3H) , 2.03 (s, 3H), 1.92 (s, 3H), 1.42 (s, 9H); ¹³C **NMR** (150 MHz, CDCl₃) δ 171.0, 170.8, 170.6, 170.3, 156.0, 85.8, 80.4, 71.7, 70.5, 68.2, 64.6, 62.9, 53.0, 51.6, 45.4, 28.2, 23.2, 21.0, 20.8, 20.8; $[\alpha]_D^{25} = +29.8$ (*c* 1.20, CHCl₃); **IR** (neat): $v_{\text{max}} = 3363$, 2979, 2112, 1738, 1368, 1247, 1216, 1161, 1041, 735 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₃H₃₆N₆NaO₁₂ [M + Na]⁺ *m*/*z* 611.2289; found *m*/*z* 611.2287; HPLC purity: 99.9% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; *t_R* = 2.70).



TLC: (petroleum ether/acetone = 1:1, v/v), $R_f = 0.32$; ¹H NMR (400 MHz, CD₃OD) δ 5.42 – 5.27 (m, 2H), 4.66 (dd, J = 12.4, 2.4Hz, 1H), 4.59 (s, 1H), 4.47 (dd, J = 10.4, 2.0 Hz, 1H), 4.18 – 4.07 (m, 2H), 3.96 – 3.85 (m, 1H), 3.77 (s, 3H), 3.15 (d, J = 3.6 Hz,

1H), 2.09 (s, 3H), 2.05 (s, 3H), 2.01 (s, 3H), 1.87 (s, 3H), 1.42 (s, 9H); ¹³C NMR (100 MHz, CD₃OD) δ 173.8, 173.2, 172.5, 171.9, 171.8, 157.9, 89.5, 80.5, 72.4, 70.8, 70.1, 63.9, 55.8, 52.9, 52.8, 46.4, 28.6, 22.7, 20.9, 20.8, 20.7; $[\alpha]_D^{25} = +0.78$ (*c* 0.26, CHCl₃); **IR** (neat): $v_{max} =$

3332, 2998, 2925, 2854, 1739, 1690, 1368, 1218, 1164, 1040, 734 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₃H₃₉N₄O₁₂ [M + H]⁺ m/z 563.2564; found m/z 563.2557; HPLC purity: 98.4% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; t_R = 2.51).



A mixture of **28** (15.0 mg, 0.029 mmol) and 10% Pd/C (1.5 mg) in MeOH (1 mL) was stirred under a balloon of hydrogen at room temperature for 3 h. The reaction mixture was then filtered through a short pad of Celite[®] and the filtrate was concentrated in

vacuo. Purification of the residue by column chromatography (CH₂Cl₂/MeOH = 10:1. v/v) afforded **S18** (10.2 mg, 70%). **TLC**: (CH₂Cl₂/MeOH = 6:1, v/v), R_f = 0.50; ¹**H NMR** (600 MHz, CDCl₃ + CD₃OD) δ 5.28 – 5.07 (m, 2H), 4.69 (d, *J* = 8.4 Hz, 1H), 4.13 (d, *J* = 7.2 Hz, 1H), 3.99 – 3.94 (m, 1H), 3.82 (d, *J* = 2.4 Hz, 1H), 3.76 – 3.65 (m, 4H), 3.22 (s, 1H), 3.02 (dd, *J* = 7.2, 2.4 Hz, 1H), 1.99 (s, 3H), 1.94 (s, 3H), 1.91 (s, 3H), 1.80 (s, 3H); ¹³**C NMR** (150 MHz, CDCl₃ + CD₃OD) δ 172.3, 171.1, 170.9, 170.7, 170.6, 85.9, 71.8, 69.7, 68.4, 66.2, 62.8, 52.5, 51.2, 47.7, 22.3, 20.5, 20.4, 20.3; $[\alpha]_D^{25} = +30.9$ (*c* 0.33, CHCl₃); **IR** (neat): *v*_{max} = 3360, 2957, 2925, 2856, 2114, 1731, 1372, 1224, 1045 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₈H₂₈N₆NaO₁₀ [M + Na]⁺ *m/z* 511.1765; found *m/z* 511.1764; HPLC purity: 94.2% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 10:90, v/v; *t_R* = 5.73).



A mixture of compound **18a** (50.0 mg, 0.081 mmol, 1.0 equiv) and potassium carbonate (5.6 mg, 0.041 mmol, 0.5 equiv) in MeOH (1 mL) was stirred for 30 min at room temperature before being neutralized with Amberlite IR 120 (H^+) resin. The mixture was

filtered and evaporated to give the crude product, which was purified through column chromatography (CH₂Cl₂/MeOH = 10:1, v/v) to deliver **S19** (30.2 mg, 76%). **TLC**: (CH₂Cl₂/MeOH = 6:1, v/v), R_f = 0.62; ¹**H NMR** (400 MHz, CDCl₃) δ 6.73 (d, *J* = 6.0 Hz, 1H), 5.13 (d, *J* = 10.0 Hz, 1H), 4.69 (d, *J* = 5.2 Hz, 1H), 4.32 – 4.20 (m, 1H), 4.13 – 3.79 (m, 10H), 3.54 (dd, *J* = 9.2, 4.0 Hz, 1H), 2.00 (s, 3H), 1.46 (s, 9H); ¹³**C NMR** (150 MHz, CDCl₃) δ 173.3, 165.3, 156.8, 90.8, 81.3, 73.9, 69.6, 69.5, 64.8, 63.1, 53.6, 48.9, 48.2, 28.3, 22.9; [a]_D²⁵ = -54.3 (*c* 0.61, CHCl₃); **IR** (neat): *v*_{max} = 3311, 2959, 2927, 2112, 1691, 1646, 1527, 1250, 1163, 1037, 1020 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₇H₂₈N₈NaO₉ [M + Na]⁺ *m/z* 511.1877; found *m/z* 511.1874; HPLC purity: 99.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 40:60, v/v; *t_R* = 5.08).



To a stirred solution of **18a** (20.0 mg, 0.032 mmol, 1.0 equiv) in MeOH (1 mL) was added NaOH aq. (1 M in H₂O, 65.1 μ L, 0.065 mmol, 2.0 equiv) at ambient temperature. The resultant mixture was allowed to stir for 40 min, then neutralized with Amberlite IR 120

(H⁺) resin and filtered. The filtrate was evaporated under reduced pressure to afford the compound **S20** (10.2 mg, 65%), which was pure enough for further characterization. ¹H NMR (400 MHz, CD₃OD) δ 4.25 – 4.07 (m, 4H), 3.83 – 3.76 (m, 2H), 3.67 (dd, *J* = 12.4, 6.0 Hz, 1H), 3.46 (d, *J* = 8.8 Hz, 1H), 1.97 (s, 3H), 1.47 (s, 9H); ¹³C NMR (150 MHz, CD₃OD) δ 174.2, 170.9, 157.7, 93.9, 80.7, 74.0, 71.7, 70.0, 65.8, 65.0, 52.7, 46.2, 28.7, 22.8; $[\alpha]_D^{25} = -52.1$ (*c* 1.07, CHCl₃); **IR** (neat): $v_{max} = 3278$, 2956, 2924, 2854, 2113, 1727, 1639, 1534, 1074 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₆H₂₆N₈NaO₉ [M + Na]⁺ *m/z* 497.1720; found *m/z* 497.1722; HPLC purity: 98.0% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 85:15, v/v; *t_R* = 3.49).



A mixture of **18a** (10.1 mg, 0.016 mmol, 1.0 equiv), NaOH aq. (1 M in H₂O, 49.3 μ L, 0.049 mmol, 3.0 equiv) and 1 mL MeOH was stirred for 30 min at room temperature before it was neutralized with Amberlite IR 120 (H⁺) resin. After filtration and evaporation, the

residue was dissolved in CH₂Cl₂ (1 mL) and trifluoroacetic acid (0.2 mL) was added. The mixture was stirred for 2 h at room temperature and concentrated in *vacuo* to afford a residue. Purification by reverse phase silical gel column chromatography (MeCN/H₂O = 1:9, v/v) yielded product **S21** (3.8 mg, 62%). ¹**H NMR** (600 MHz, D₂O) δ 4.37 – 4.29 (m, 2H), 4.22 (d, J = 10.2 Hz, 1H), 3.97 (dd, J = 10.8, 3.0 Hz, 1H), 3.93 – 3.88 (m, 1H), 3.86 (dd, J = 12.0, 3.0 Hz, 1H), 3.67 (dd, J = 12.0, 6.0 Hz, 1H), 3.57 (d, J = 9.6 Hz, 1H), 2.06 (s, 3H); ¹³**C NMR** (100 MHz, D₂O) δ 175.0, 169.6, 91.9, 71.6, 69.9, 68.0, 63.3, 61.2, 50.8, 43.8, 22.3; [**a**]_D²⁵ = -23.6 (*c* 0.60, MeOH); **IR** (neat): $v_{max} = 3320$, 2950, 2839, 2359, 2342, 1685, 1661, 1637, 1013 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₁H₁₈N₈NaO₇ [M + Na]⁺ *m*/z 397.1196; found *m*/z 397.1199; HPLC purity: 91.8% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 90:10, v/v; *t_R* = 1.49).





Following the procedure described for synthesis of **S20**, a mixture of **18b** (20.3 mg, 0.033 mmol, 1.0 equiv), NaOH aq. (1 M in H₂O, 99.2 μ L, 0.099 mmol, 3.0 equiv) and MeOH (1 mL) was allowed to stir for 40 min at room temperature, neutralized with Amberlite IR 120 (H⁺)

resin, and filtered. The filtrate was evaporated under reduced pressure to afford product **S22** (11.4 mg, 73%), which was pure enough for further characterization. ¹**H NMR** (400 MHz, CD₃OD) δ 4.60 (s, 2H), 4.36 (s, 1H), 4.21 – 4.08 (m, 1H), 4.03 (t, *J* = 10.4 Hz, 1H), 3.93 – 3.76 (m, 2H), 3.69 – 3.54 (m, 2H), 1.96 (s, 3H), 1.46 (s, 8H); ¹³**C NMR** (100 MHz, CD₃OD) δ 175.1, 157.9, 80.9, 76.8, 72.8, 69.9, 65.7, 64.1, 53.5, 46.8, 28.7, 22.7; [α]_D²⁵ = -1.6 (*c* 0.37, MeOH); **IR** (neat): v_{max} = 3288, 2927, 2109, 1632, 1534, 1369, 1250, 1161, 1037 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₆H₂₆N₈NaO₉ [M + Na]⁺ *m/z* 497.1720; found *m/z* 497.1741; HPLC purity: 97.1% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 80:20, v/v; *t_R* = 4.44).



According to the procedure for synthesis of **S21**, compound **18b** (31.2 mg, 0.051 mmol) was used to prepare **S23** (11.2 mg, 59%). ¹H NMR (400 MHz, CD₃OD) δ 4.49 (d, J = 3.2 Hz, 1H), 4.28 (t, J = 10.8 Hz, 1H), 3.91 – 3.72 (m, 4H), 3.71 – 3.57 (m, 2H), 2.03 (s, 3H); ¹³C NMR

(100 MHz, CD₃OD) δ 175.2, 170.3, 76.1, 72.8, 69.5, 64.3, 62.5, 53.4, 45.3, 23.0; $[\alpha]_D^{25} = +9.5$ (*c* 0.77, MeOH); **IR** (neat): $v_{max} = 3270$, 2930, 2121, 1632, 1553, 1375, 1305, 1248, 1037 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₁H₁₈N₈NaO₇ [M + Na]⁺ *m/z* 397.1196; found *m/z* 397.1183; HPLC purity: 98.1% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX 300SB-C8, 5 µm, 4.6 × 150 mm; Detector: Waters 2424 ELSD; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 100:0, v/v; *t_R* = 2.18).





To a solution of **20a** (68.0 mg, 0.090 mmol, 1.0 equiv) in CH₂Cl₂ (2 mL) was added trifluoroacetic acid (0.4 mL) at room temperature. The resultant mixture was allowed to stir overnight and then concentrated under *vacuum*. The residue was purified through flash column chromatography (CH₂Cl₂/MeOH = 20:1,

v/v) to yield **33** (42.5 mg, 85%). TLC: (CH₂Cl₂/MeOH = 8:1, v/v), $R_f = 0.55$; ¹H NMR (400

MHz, CD₃OD) δ 5.44 (dd, J = 7.2, 2.0 Hz, 1H), 5.31 – 5.25 (m, 1H), 4.52 (dd, J = 12.0, 2.4 Hz, 1H), 4.32 (d, J = 3.2 Hz, 1H), 4.25 (d, J = 10.4 Hz, 1H), 4.21 – 4.14 (m, 1H), 4.11 (dd, J = 12.8, 5.6 Hz, 2H), 3.97 (s, 3H), 2.08 (s, 3H), 2.06 (s, 3H), 2.02 (s, 3H), 1.91 (s, 3H); ¹³C **NMR** (150 MHz, CD₃OD) δ 173.9, 172.4, 171.5, 171.2, 166.2, 159.1, 92.2, 73.0, 71.0, 68.5, 64.4, 62.9, 54.2, 53.4, 46.3, 22.7, 21.0, 20.6; $[\alpha]_D^{25} = -7.8$ (*c* 1.00, CHCl₃); **IR** (neat): $v_{max} = 3338$, 2118, 1745,1667, 1372, 1203, 1134, 1046, 734 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₉H₂₉N₁₀O₁₀ [M + H]⁺ *m*/*z* 557.2068; found *m*/*z* 557.2086; HPLC purity: 99.3% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; *t_R* = 1.32).



A mixture of compound **33** (8.3 mg, 0.015 mmol, 1.0 equiv), NaOH aq. (1 M in H₂O, 44.8 μ L, 0.045 mmol, 3.0 equiv) and MeOH (1 mL) was allowed to stir for 1 h at room temperature, then neutralized with Amberlite IR 120 (H⁺) resin and filtered. After removal of solvent from the filtrate, the residue was purified by

reverse phase silical gel column chromatography (MeOH/H₂O = 1:9, v/v) to give **34** (3.4 mg, 54%). ¹**H NMR** (600 MHz, CD₃OD/D₂O) δ 4.30 – 4.04 (m, 4H), 3.92 – 3.79 (m, 2H), 3.65 (dd, *J* = 12.0, 6.0 Hz, 1H), 3.55 (d, *J* = 9.6 Hz, 1H), 2.00 (s, 3H); ¹³**C NMR** (150 MHz, CD₃OD) δ 173.9, 158.9, 73.3, 71.7, 69.9, 65.5, 64.9, 53.9, 47.3, 45.4, 22.8; $[\boldsymbol{\alpha}]_D^{25} = -63.8$ (*c* 0.13, MeOH); **IR** (neat): $v_{max} = 3195$, 2923, 2854, 2113, 1633, 1612, 1374, 1064, 1038, 1019, 610 cm⁻¹; **HRMS** (**ESI**): calcd. for C₁₂H₂₁N₁₀O₇ [M + H]⁺ *m/z* 417.1595; found *m/z* 417.1581; HPLC purity: 97.4% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 80:20, v/v; *t_R* = 1.51).





According to the procedure described for the synthesis of **33**, a mixture of **20b** (58.2 mg, 0.077 mmol) and trifluoroacetic acid (0.2 mL) in CH₂Cl₂ (1 mL) was stirred for 8 h. The residue was purified by silica gel column chromatography (CH₂Cl₂/MeOH = 20:1, v/v) to afford **S24** (37.2 mg, 87%). **TLC**: (CH₂Cl₂/MeOH = 8:1, v/v), R_f

= 0.50; ¹**H NMR** (400 MHz, CDCl₃) δ 7.95 (brs, 2H), 7.01 (m, 3H), 5.47 – 5.26 (m, 2H), 4.28 (dd, J = 12.8, 2.8 Hz, 1H), 4.23 – 4.05 (m, 4H), 3.91 (s, 3H), 3.90 – 3.85 (m, 1H), 2.12 (s, 3H),

2.07 (s, 3H), 2.04 (s, 3H), 1.97 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.0, 170.8, 167.0, 169.7, 165.8, 157.7, 90.5, 73.6, 68.2, 67.2, 61.7, 53.9, 51.8, 45.4, 29.7, 23.0, 20.8, 20.7, 20.4; $[\alpha]_D^{25} = +20.7$ (*c* 0.58, MeOH); **IR** (neat): $v_{max} = 3339$, 2962, 2116, 1746, 1667, 1610, 1372, 1203, 1179, 1135, 1043, 721 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₉H₂₉N₁₀O₁₀ [M + H]⁺ *m/z* 557.2068; found *m/z* 557.2080; HPLC purity: 97.6% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 30:70, v/v; *t_R* = 1.35).



Following the procedure for the preparation of **34**, **S24** (27.3 mg, 0.049 mmol, 1.0 equiv), NaOH aq. (1 M in H₂O, 147 μ L, 0.147 mmol, 3.0 equiv) and MeOH (1 mL) were used for the synthesis of **S25** (14.2 mg, 70%) as a white amorphous powder after purification by reverse phase silica gel column chromatography (MeOH/H₂O = 1:9, v/v). ¹H

NMR (400 MHz, CD₃OD) δ 4.54 (dd, J = 10.4, 3.6 Hz, 1H), 4.30 (d, J = 3.6 Hz, 1H), 4.21 (t, J = 10.4 Hz, 1H), 3.91 – 3.78 (m, 2H), 3.75 – 3.63 (m, 3H), 1.97 (s, 3H); ¹³C NMR (100 MHz, CD₃OD) δ 174.6, 171.0, 158.7, 93.3, 76.3, 73.5, 69.6, 65.2, 64.5, 54.2, 47.5, 22.7; $[\alpha]_D^{25} = -9.9$ (*c* 0.43, MeOH); **IR** (neat): $v_{max} = 3332$, 2944, 2834, 2113, 1634, 1374, 1020 cm⁻¹; **HRMS (ESI)**: calcd. for C₁₂H₂₀N₁₀NaO₇ [M + Na]⁺ *m*/*z* 439.1414; found *m*/*z* 439.1487; HPLC purity: 95.3% (Flow rate: 1.0 mL/min; Column: Agilent ZORBAX SB-C18, 5 µm, 4.6 × 150 mm; Detector: Alltech ELSD 2000ES; Temperature: 25 °C; Mobile phase: H₂O/MeOH = 90:10, v/v; *t_R* = 3.71).

7. Mechanistic Investigations

7.1 Radical Trapping Experiment with TEMPO



To a solution of **7b** (40.0 mg, 0.133 mmol, 1.0 equiv), BI-OAc (81.6 mg, 0.267 mmol, 2.0 equiv) and TEMPO (62.5 mg, 0.400 mmol, 3.0 equiv) in dry MeCN (1 mL) was added TMSN₃ (52.6 µL, 0.400 mmol, 3.0 equiv) under an argon atmosphere. The mixture was stirred for 3 h under irradiation of 34 W blue LEDs. Saturated aqueous KHCO₃ was added and the mixture was stirred vigorously at room temperature for 5 min. After extraction of the resulting mixture with EtOAc for three times, the organic layers were combined, dried, and concentrated. Purification of the crude residue by silica gel column chromatography (petroleum ether/acetone = 20:1, v/v) afforded S26 (30.0 mg, 45%, 86% brsm) as a white amorphous powder, along with recovery of 7b (19.1 mg). Note that S26 was a mixture of two isomers S26a and S26b. S26a: TLC: (petroleum ether/EtOAc = 5:1, v/v), $R_f = 0.38$; ¹H NMR $(400 \text{ MHz, CDCl}_3) \delta 4.99 \text{ (s, 2 H), } 4.51 \text{ (t, } J = 1.2 \text{ Hz, 1H), } 4.33 \text{ (m, 1H), } 4.20 - 4.13 \text{ (m, 2H), }$ 3.82 (s, 3H), 3.79 (d, J = 9.2 Hz, 1H), 1.54 - 1.23 (m, 12H), 1.21 (s, 3H), 1.17 (s, 3H), 1.14 (s, 3H), 1.12 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.4, 153.0, 110.0, 103.6, 73.1, 72.5, 71.4, 71.0, 67.1, 62.1, 61.5, 59.8, 51.8, 41.2, 40.6, 33.9, 33.4, 26.9, 25.0, 21.7, 21.6, 16.8; $[\alpha]_D^{25} =$ +52.5 (c 0.19, CHCl₃); **IR** (neat): $v_{\text{max}} = 2943$, 2936, 2123, 1817, 1258, 1179, 1075 cm⁻¹; **HRMS (ESI)**: calcd. for C₂₂H₃₄N₄NaO₉ $[M + Na]^+ m/z$ 521.2223; found m/z 521.2217. S26b: **TLC**: (petroleum ether/EtOAc = 5:1, v/v), $R_f = 0.32$; ¹H NMR (400 MHz, CDCl₃) δ 5.16 (d, J = 4.0 Hz, 1H), 4.95 (dd, J = 8.4, 4.0 Hz, 1H), 4.88 (dd, J = 8.4, 1.6 Hz, 1H), 4.34 – 4.27 (m, 1H), 4.26 - 4.16 (m, 2H), 4.06 (dd, J = 8.8, 4.8 Hz, 1H), 3.76 (s, 3H), 1.62 - 1.34 (m, 12H), 1.26 (s, 3H), 1.24 (s, 3H), 1.20 (s, 3H), 1.18 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.9, 152.7, 109.8, 100.8, 72.6, 72.6, 72.3, 71.5, 67.7, 62.1, 61.1, 59.8, 52.4, 41.4, 40.7, 33.7, 33.7, 26.8, 24.9, 21.2, 20.8, 16.6; $[\alpha]_D^{25} = +45.5$ (c 0.09, CHCl₃); **IR** (neat): $v_{\text{max}} = 2923$, 2853, 2119, 1821, 1752, 1370, 1069, 1031 cm⁻¹; HRMS (ESI): calcd. for $C_{22}H_{34}N_4NaO_9 [M + Na]^+$ *m*/*z* 521.2223; found *m*/*z* 521.2216.

7.2 Isolation of 2-Iodobenzoic Acid



To a solution of **7a** (30.1 mg, 0.075 mmol, 1.0 equiv) and BI-OAc (45.8 mg, 0.15 mmol, 2.0 equiv) in 0.75 mL dry MeCN was added TMSN₃ (29.5 μ L, 0.22 mmol, 3.0 equiv) under an argon atmosphere. The reaction mixture was stirred under irradiation of 34 W blue LEDs for 2 h. Then saturated NaHCO₃ solution was added to quench the reaction. The mixture was stirred for additional 5 min before being acidified with 1M HCl. EtOAc was added to dilute the reaction. After separation of the two layers, the aqueous phase was extracted with EtOAc for three times. The combined organic phase was washed with brine, dried over Na₂SO₄, and evaporated to dryness. 2-Iodobenzoic acid was isolated from the residue through preparative TLC (petroleum ether/EtOAc = 2:1, v/v), which could be confirmed by NMR and MS, as well as by comparison with an authentic sample.

7.3 UV/Vis Absorption

No visible color change was observed when mixing the reaction components between each other (Figure S1).



Figure S1. Appearance of the reaction components and their corresponding mixtures

UV/Vis absorption spectra between substrate **7a** (Kdo-G, 0.04 mmol) and azide donor TMSN₃ (0.04 mmol) or oxidant BI-OAc (0.04 mmol) in CHCl₃ (2 mL) were recorded in 1 cm path quartz cuvettes using a Quawell scientific Q6000+ micro volume spectrophotometer. As shown in Figure S2, no bathochromic shift was observed.



Figure S2. UV/Vis absorption spectra between Kdo-G (7a) and TMSN3 or BI-OAc

UV/Vis absorption spectra between oxidant BI-OAc (0.04 mmol) and azide donor TMSN₃ (0.04 mmol) in CHCl₃ (2 mL) were recorded in 1 cm path quartz cuvettes using a Quawell scientific Q6000+ micro volume spectrophotometer. As shown in Figure S3, no bathochromic shift was observed.



Figure S3. UV/Vis absorption spectra between TMSN₃ and BI-OAc

UV/Vis absorption spectra between substrate **7a** (Kdo-G, 0.04 mmol) and oxidant BI-N₃ (0.04 mmol) in 2 mL CHCl₃ were recorded in 1 cm path quartz cuvettes using a Quawell scientific Q6000+ micro volume spectrophotometer. As shown in Figure S4, no bathochromic shift was observed.



Figure S4. UV/Vis absorption spectra between Kdo-G (7a) and BI-N₃

UV/Vis absorption spectra between BI-N₃ (0.04 mmol) and azide donor TMSN₃ (0.04 mmol) in 2 mL CHCl₃ were recorded in 1 cm path quartz cuvettes using a Quawell scientific Q6000+ micro volume spectrophotometer. As shown in Figure S5, no bathochromic shift was observed.



Figure S5. UV/Vis absorption spectra between TMSN₃ and BI-N₃

7.4 NMR Titration Experiments

¹H NMR experiments of solutions containing different ratios of oxidant (BI-OAc) and azide donor (TMSN₃) (covering the ratio 0%, 10%, 20%, 40%, 60%, 80% and 100% TMSN₃, from 1 to 7 in Figure S6) were performed, with constant concentration of the two components (0.01 M in CDCl₃). No resonance signal shift was observed in BI-OAc upon addition of TMSN₃.





Figure S6. ¹H NMR titration between oxidant (BI-OAc) and TMSN₃

¹H NMR experiments of solutions containing different ratios of BI-N₃ and azide donor TMSN₃ (covering the ratio 0%, 10%, 20%, 40%, 60%, 80% and 100% TMSN₃, from 1 to 7 in Figure S7) were performed, with constant concentration of the two components (0.02 M in CDCl₃). No resonance signal shift was observed in BI-N₃ upon addition of TMSN₃.





Figure S7. ¹H NMR titration between BI-N₃ and TMSN₃

8. Antiviral Activities

Materials and Methods

Cells and Viruses. BHK and H1 Hela cells were obtained from National Infrastructure of Cell Line Resource, China. Cells were cultured in Dulbecco's modified Eagle's medium (DMEM, Gibco) supplemented with 10% fetal bovine serum (FBS, Gibco) and 100 U/ml penicillin/streptomycin at 37 °C in 5% CO₂. ZIKV (SMGC-1 strain) was produced by infecting Vero cells one day post-seeding at a multiplicity of infection (MOI) of 0.001. The virus titers were determined by 50% tissue culture infectious dose (TCID₅₀) assay. Human rhinovirus 1059 strain belongs to Enterovirus Rhinovirus B and genotype 14 (ATCC, VR-284), and the virus stocks were propagated in H1 Hela cells according standard protocol.

In Vitro Cytopathic Effect (CPE) Assay. BHK or H1 Hela cells were used for antiviral assays of ZIKV or HRV, respectively. Briefly, 5×10^3 cells were seeded in 96-well plates. At 24 hours post infection (hpi), cells were infected with 100 TCID₅₀ of virus in the presence of gradient dilutions of indicated drugs and cultured for 8 days for ZIKV, and 3 days for HRV. Then, the protection of CPE was measured with CellTiter-Glo® Luminescent Cell Viability Assay (Promega, USA) kit according to the manufacturer's instructions. Values for the vitro efficacy were calculated using OriginPro 8.0 Software.

Cytotoxicity Assay. The cytotoxicity was evaluated using a CellTiter-Glo® Luminescent Cell

Viability Assay (Promega, USA) kit following the manufacturer's instructions. Briefly, 5×10^3 cells were seeded in 96-well plates and incubated with gradient dilutions of drugs for the 8 days or 3 days for BHK and H1 Hela cells, respectively. Values for the cytotoxicity were calculated using the OriginPro 8.0 Software.

J	anti-ZIKV (BHK)			anti-HRV (H1 Hela)		
compa.	IC ₅₀ (µM) ^a	$\mathrm{CC}_{50}(\mu\mathrm{M})^b$	SI^c	IC ₅₀ (µM) ^a	$\mathrm{CC}_{50}(\mu\mathrm{M})^b$	SI^c
NITD008 ^d	0.87	>20	>22.99			
Rupintrivir ^d				0.08	>10	>125
8 a	>200			95.24	>200	>2.10
8b	166.11			59.81		
8c	>200			178.09	>200	>1.12
8d	>200			171.99	>200	>1.16
9a	>200			178.95	>200	>1.12
9b	>200			>200		
12a	>200			71.1	199.79	2.81
14a	>200			81.04	>200	>2.47
14b	>200			68.77		
14c	>200			>200		
14d	>200			>200		
15 a	31.05	>200	>6.44	>200		
15b	8.67	>200	>23.07	>200		
15c	9.24	>200	>21.65	>200		
15d	9.20	>200	>21.74	>200		
16a	1.41	7.68	5.45	4.58	9.25	2.02
16b	1.90	>200	>105.26	18.89	49.11	2.60
17a	5.50	21.89	3.98	0.93	6.68	7.18
17b	7.30	20.29	2.78	2.05	5.74	2.80
18 a	>200			>200		
18b	>200			193.78	>200	>1.03
19a	>200			>200		
19b	>200			59.38	>200	>3.37
20a	69.63	128.12	1.84	74.56	>200	>2.68
20b	165.03	>200	>1.21	104.97	>200	>1.91
21	44.95	>200	>4.45	19.16	200.03	10.44
22	177.50	>200	>1.13	>200		
23	140.65	>200	>1.42	67.39	>200	>2.97
24	108.41	>200	>1.84	>200		

Table S7. Antiviral Activity

25	15.02	193.31	12.87	>200		
26	>200			83.06	>200	>2.41
27	69.32			>200		
28	>200			>200		
30	22.73	29.32	1.29	1.87	64.70	34.60
31	25.05	67.89	2.71	5.39	41.02	7.61
32	>200			113.99	>200	>1.75
33	105.56	>200	>1.89	>200		
34	>200			70.49	>200	>2.84
S12	192.98	>200	>1.04	>200		
S14	>200			>200		
S15	9.64	>200	>20.75	>200		
S16	>200			>200		
S17	74.64	>200	>2.68	12.53	>200	>15.96
S18	>200			181.77	>200	>1.10
S19	>200			>200		
S20	199.68	>200	>1.00	98.73	>200	>2.03
S21	185.68	>200	>1.08	82.09	>200	>2.44
S22	156.62	>200	>1.28	185.29	>200	>1.08
S23	154.46	>200	>1.29	18.89	200.05	10.59
S24	>200			176.55	>200	>1.13
S25	>200			>200		

^{*a*}IC₅₀: the concentration of test compounds that reduced the activity by 50% of the untreated (control) cell cultures. Each value was calculated from duplicate assays. ^{*b*}CC₅₀: the concentration of test compounds that reduced cell viability to 50% of the untreated (control) cell cultures. Each value was calculated from duplicate assays. ^{*c*}SI: selectivity index, the ratio of CC₅₀ to IC₅₀ (CC₅₀/IC₅₀). ^{*d*}NITD008⁸ and rupintrivir⁹ were used as positive controls, respectively.

9. References

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10.X-Ray Crystallographic Data

X-ray Crystallographic Data for Compound 8c: CCDC 1975133 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S8. Crystal data and structure refinement for 190419_s1_hh_yd0417a.

Identification code	190419_s1_hh_yd0417a
Empirical formula	$C_{17}H_{22}N_6O_{11}$
Formula weight	486.40
Temperature/K	293.15
Crystal system	monoclinic
Space group	P21
a/Å	9.2129(4)
b/Å	40.7685(17)
c/Å	12.8563(5)
α/°	90
β/°	90.320(3)

$\gamma/^{\circ}$	90
Volume/Å ³	4828.7(3)
Z	8
$\rho_{calc}g/cm^3$	1.338
µ/mm ⁻¹	0.113
F(000)	2032.0
Crystal size/mm ³	$0.35\times0.3\times0.25$
Radiation	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/°	5.81 to 52.744
Index ranges	$-11 \le h \le 11, -50 \le k \le 49, -15 \le l \le 16$
Reflections collected	35580
Independent reflections	$16488 [R_{int} = 0.0264, R_{sigma} = 0.0466]$
Data/restraints/parameters	16488/1/1253
Goodness-of-fit on F ²	1.015
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0499, wR_2 = 0.1143$
Final R indexes [all data]	$R_1 = 0.0741, wR_2 = 0.1318$
Largest diff. peak/hole / e Å ⁻³	0.28/-0.24
Flack parameter	0.1(4)

X-ray Crystallographic Data for Compound 8d: CCDC 1975134 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S9. Crystal data and structure refinement for 190419_s1_hh_yd0417b.Identification code190419_s2_hh_yd0417bEmpirical formulaC17H22N6O11

Formula weight	486.40
Temperature/K	293.15
Crystal system	monoclinic
Space group	P21
a/Å	7.7778(5)
b/Å	13.2995(10)
c/Å	11.4128(6)
$\alpha/^{\circ}$	90
β/°	91.458(5)
$\gamma^{\prime \circ}$	90
Volume/Å ³	1180.18(13)
Ζ	2
$\rho_{calc}g/cm^3$	1.369
µ/mm ⁻¹	0.116
F(000)	508.0
Crystal size/mm ³	$0.35 \times 0.3 \times 0.25$
Radiation	MoKa ($\lambda = 0.71073$)
2Θ range for data collection/°	6.07 to 52.738
Index ranges	$-9 \le h \le 9, -16 \le k \le 9, -14 \le l \le 12$
Reflections collected	5262
Independent reflections	3233 [$R_{int} = 0.0190, R_{sigma} = 0.0362$]
Data/restraints/parameters	3233/1/312
Goodness-of-fit on F ²	1.047
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0410, wR_2 = 0.0881$
Final R indexes [all data]	$R_1 = 0.0617, wR_2 = 0.0994$
Largest diff. peak/hole / e Å ⁻³	0.19/-0.18
Flack parameter	0.3(7)

X-ray Crystallographic Data for Compound 9a: CCDC 1975130 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S10. Crystal data and structure refinance	nement for 190226_s3_yd_1.
Identification code	190226_s3_yd_1
Empirical formula	$C_{14}H_{16}Cl_3N_6O_8$
Formula weight	502.68
Temperature/K	293.15
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	8.6304(6)
b/Å	10.8741(9)
c/Å	22.9325(14)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	2152.2(3)
Z	4
$\rho_{calc}g/cm^3$	1.551
µ/mm ⁻¹	0.480
F(000)	1028.0
Crystal size/mm ³	$0.35\times0.3\times0.25$
Radiation	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/°	6.026 to 52.728
Index ranges	$\text{-10} \le h \le 10, \text{-8} \le k \le 13, \text{-28} \le l \le 28$
Reflections collected	9826
Independent reflections	4407 [$R_{int} = 0.0216$, $R_{sigma} = 0.0399$]
Data/restraints/parameters	4407/0/293
Goodness-of-fit on F ²	1.030

Final R indexes [I>= 2σ (I)]	$R_1 = 0.0594, wR_2 = 0.1420$
Final R indexes [all data]	$R_1 = 0.0806, wR_2 = 0.1562$
Largest diff. peak/hole / e Å ⁻³	0.31/-0.43
Flack parameter	0.04(3)

X-ray Crystallographic Data for Compound 9b: CCDC 1975131 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S11. Crystal data and structure refinement for 190226_s4_yd_2.			
Identification code	190226_s4_yd_2		
Empirical formula	$C_{13}H_{16}N_6O_8$		
Formula weight	384.32		
Temperature/K	293.15		
Crystal system	orthorhombic		
Space group	$P2_{1}2_{1}2_{1}$		
a/Å	7.2784(7)		
b/Å	9.2019(10)		
c/Å	26.166(3)		
α/°	90		
β/°	90		
γ/°	90		
Volume/Å ³	1752.4(3)		
Ζ	4		
$\rho_{calc}g/cm^3$	1.457		
μ/mm^{-1}	0.123		
F(000)	800.0		
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Crystal size/mm ³	$0.35 \times 0.3 \times 0.25$		
Radiation	MoKa ($\lambda = 0.71073$)		
2Θ range for data collection/°	6.228 to 52.732		
Index ranges	$-9 \le h \le 8, -11 \le k \le 10, -23 \le l \le 32$		
Reflections collected	6084		
Independent reflections	$3278 [R_{int} = 0.0229, R_{sigma} = 0.0467]$		
Data/restraints/parameters	3278/0/251		
Goodness-of-fit on F ²	1.018		
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0461, wR_2 = 0.0845$		
Final R indexes [all data]	$R_1 = 0.0687, wR_2 = 0.0950$		
Largest diff. peak/hole / e Å ⁻³	0.14/-0.14		
Flack parameter	0.2(10)		

X-ray Crystallographic Data for Compound 14c: CCDC 1975132 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



 Table S12. Crystal data and structure refinement for 190321_s2_hh.

Identification code	190321_s2_hh
Empirical formula	$C_{20}H_{27}N_7O_{12}$
Formula weight	557.48
Temperature/K	293.15
Crystal system	monoclinic
Space group	P21
a/Å	9.6273(12)
b/Å	8.8714(10)
c/Å	16.549(2)
$\alpha/^{\circ}$	90
β/°	101.469(11)
$\gamma/^{\circ}$	90
Volume/Å ³	1385.2(3)
Z	2
$\rho_{calc}g/cm^3$	1.337
μ/mm ⁻¹	0.112
F(000)	584.0
Crystal size/mm ³	$0.35\times0.3\times0.25$
Radiation	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/°	5.938 to 52.74
Index ranges	$-11 \le h \le 12, -11 \le k \le 7, -20 \le l \le 13$
Reflections collected	5951
Independent reflections	4170 [$R_{int} = 0.0233$, $R_{sigma} = 0.0662$]
Data/restraints/parameters	4170/5/381
Goodness-of-fit on F ²	0.965
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0484, wR_2 = 0.0959$
Final R indexes [all data]	$R_1 = 0.1013$, $wR_2 = 0.1171$
Largest diff. peak/hole / e Å ⁻³	0.23/-0.16
Flack parameter	0.8(10)

X-ray Crystallographic Data for Compound 14d: CCDC 1975126 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S13. Crystal data and structure	refinement for qy-hh181126e.
Identification code	qy-hh181126e
Empirical formula	$C_{20}H_{27}N_7O_{12}$
Formula weight	557.48
Temperature/K	291.6(2)
Crystal system	orthorhombic
Space group	P212121
a/Å	9.29096(14)
b/Å	14.5006(2)
c/Å	20.3184(3)
$\alpha/^{\circ}$	90
β/°	90
γ/°	90
Volume/Å ³	2737.38(7)
Z	4
$\rho_{calc}g/cm^3$	1.353
μ/mm^{-1}	0.975
F(000)	1168.0
Crystal size/mm ³	0.7 imes 0.4 imes 0.3
Radiation	$CuK\alpha \ (\lambda = 1.54184)$
2Θ range for data collection/°	7.49 to 145.916
Index ranges	$-11 \le h \le 7, -17 \le k \le 16, -23 \le l \le 24$
Reflections collected	15433
Independent reflections	5368 [$R_{int} = 0.0320, R_{sigma} = 0.0264$]
Data/restraints/parameters	5368/0/358
Goodness-of-fit on F ²	1.038
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0540, wR_2 = 0.1430$

Final R indexes [all data]	$R_1 = 0.0574, wR_2 = 0.1492$
Largest diff. peak/hole / e Å ⁻³	0.24/-0.32
Flack parameter	-0.10(10)

X-ray Crystallographic Data for Compound 15b: CCDC 1975127 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S14. Crystal data and structure refinement for 190510_s5_hh_0405b.		
Identification code	190510_s5_hh_0405b	
Empirical formula	$C_{40}H_{35}N_7O_{12}$	
Formula weight	805.75	
Temperature/K	293.15	
Crystal system	trigonal	
Space group	P3 ₁ 21	
a/Å	19.7672(8)	
b/Å	19.7672(8)	
c/Å	23.6968(12)	
α/°	90	
β/°	90	
γ/°	120	
Volume/Å ³	8018.8(8)	
Ζ	6	
$\rho_{calc}g/cm^3$	1.001	

μ/mm ⁻¹	0.076
F(000)	2520.0
Crystal size/mm ³	$0.35 \times 0.3 \times 0.25$
Radiation	MoKa ($\lambda = 0.71073$)
2Θ range for data collection/°	5.872 to 52.74
Index ranges	-24 \leq h \leq 20, -24 \leq k \leq 23, -29 \leq l \leq 29
Reflections collected	34678
Independent reflections	10925 [$R_{int} = 0.0321$, $R_{sigma} = 0.0489$]
Data/restraints/parameters	10925/0/536
Goodness-of-fit on F ²	1.035
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0819, wR_2 = 0.2096$
Final R indexes [all data]	$R_1 = 0.1515, wR_2 = 0.2662$
Largest diff. peak/hole / e Å ⁻³	0.47/-0.30
Flack parameter	0.8(4)

X-ray Crystallographic Data for Compound 17a: CCDC 1975128 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S15. Crystal data and structure refinement for 190419_s2_hh_hh190410a.

Identification code	190419_s2_hh_hh190410a
Empirical formula	$C_{37}H_{39}F_3N_8O_{13}$
Formula weight	860.76
Temperature/K	293.15
Crystal system	monoclinic

Space group	P21
a/Å	12.9906(4)
b/Å	11.1438(4)
c/Å	14.8377(7)
$\alpha/^{\circ}$	90
β/°	103.257(4)
$\gamma/^{\circ}$	90
Volume/Å ³	2090.74(15)
Z	2
$\rho_{calc}g/cm^3$	1.367
µ/mm ⁻¹	0.113
F(000)	896.0
Crystal size/mm ³	$0.35 \times 0.3 \times 0.25$
Radiation	MoKa ($\lambda = 0.71073$)
2Θ range for data collection/°	5.99 to 52.738
Index ranges	$-15 \le h \le 16, -13 \le k \le 13, -18 \le l \le 12$
Reflections collected	9421
Independent reflections	7012 [$R_{int} = 0.0172$, $R_{sigma} = 0.0448$]
Data/restraints/parameters	7012/1/569
Goodness-of-fit on F ²	1.031
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0533, wR_2 = 0.1204$
Final R indexes [all data]	$R_1 = 0.0802, wR_2 = 0.1378$
Largest diff. peak/hole / e Å-3	0.26/-0.24
Flack parameter	-0.5(5)

X-ray Crystallographic Data for Compound 29: CCDC 1975129 contains the supplementary crystallographic data for this structure. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Table S16 . Crystal data and structure refinement for 190226_s5_yd_hh181130b.		
Identification code	190226_s5_yd_hh181130b	
Empirical formula	$C_{18}H_{24}N_{10}O_{10}\\$	
Formula weight	540.47	
Temperature/K	293.15	
Crystal system	orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
a/Å	8.6075(7)	
b/Å	10.7485(9)	
c/Å	33.736(3)	
$\alpha/^{\circ}$	90	
β/°	90	
$\gamma^{/\circ}$	90	
Volume/Å ³	3121.1(5)	
Z	4	
$\rho_{calc}g/cm^3$	1.150	
μ/mm^{-1}	0.095	
F(000)	1128.0	
Crystal size/mm ³	0.35 imes 0.3 imes 0.25	
Radiation	MoKa ($\lambda = 0.71073$)	
2Θ range for data collection/°	5.96 to 52.74	
Index ranges	-10 \leq h \leq 10, -9 \leq k \leq 13, -24 \leq l \leq 42	
Reflections collected	9470	
Independent reflections	5968 [$R_{int} = 0.0270$, $R_{sigma} = 0.0541$]	
Data/restraints/parameters	5968/0/342	
Goodness-of-fit on F ²	0.966	

Final R indexes [I>= 2σ (I)]	$R_1 = 0.0717, wR_2 = 0.1976$
Final R indexes [all data]	$R_1 = 0.0974, wR_2 = 0.2188$
Largest diff. peak/hole / e Å ⁻³	0.29/-0.28
Flack parameter	0.7(10)

11. NMR Spectra



¹³C NMR spectrum of compound **S1**



 ^{13}C NMR spectrum of compound S2



¹³C NMR spectrum of compound **7b**









 ^{13}C NMR spectrum of compound 7c



¹³C NMR spectrum of compound **7d**



 ^{13}C NMR spectrum of compound 7f





 $^{13}\mathrm{C}$ NMR spectrum of compound 7h



¹³C NMR spectrum of compound 7i



¹³C NMR spectrum of compound **7**j



 ^{13}C NMR spectrum of compound 7k



 $^{13}\mathrm{C}$ NMR spectrum of compound 71



¹³C NMR spectrum of compound 8a



 $^{13}\mathrm{C}$ NMR spectrum of compound $\mathbf{8b}$



¹³C NMR spectrum of compound 8c



 $^{13}\mathrm{C}$ NMR spectrum of compound 8d



¹³C NMR spectrum of compound **9a**



 ^{13}C NMR spectrum of compound $\mathbf{9b}$



¹³C NMR spectrum of compound **10a**

$\begin{array}{c} 8.0.24\\ 8.0.24\\ 7.621\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.762\\ 7.7260\\ 7.7$



¹H NMR spectrum of compound **10b**



¹³C NMR spectrum of compound **10b**



¹³C NMR spectrum of compound **11a**



¹³C NMR spectrum of compound **11b**



¹³C NMR spectrum of compound **12a**



¹³C NMR spectrum of compound **12b**



¹³C NMR spectrum of compound **13a**



¹³C NMR spectrum of compound **13b**



¹³C NMR spectrum of compound 14a



¹³C NMR spectrum of compound **14b**



 ^{13}C NMR spectrum of compound 14c


¹³C NMR spectrum of compound **14d**













¹³C NMR spectrum of compound **15a**



¹³C NMR spectrum of compound **15b**



 $^{13}\mathrm{C}$ NMR spectrum of compound 15c



¹³C NMR spectrum of compound **15d**



¹³C NMR spectrum of compound **16a**



¹³C NMR spectrum of compound **16b**



¹³C NMR spectrum of compound **17a**











¹³C NMR spectrum of compound **17b**



¹³C NMR spectrum of compound **18a**



¹³C NMR spectrum of compound **18b**



¹³C NMR spectrum of compound **19a**



¹³C NMR spectrum of compound **19b**



¹³C NMR spectrum of compound **20a**



¹³C NMR spectrum of compound **20b**



¹³C NMR spectrum of compound **S9**



¹³C NMR spectrum of compound S10



¹³C NMR spectrum of compound S11



100 90 f1 (ppm)

 ^{13}C NMR spectrum of compound 22





¹³C NMR spectrum of compound S12



¹³C NMR spectrum of compound S13



 ^{13}C NMR spectrum of compound $\mathbf{24}$







HSQC spectrum of compound 24



HMBC spectrum of compound 24



NOE spectrum of compound 24



¹³C NMR spectrum of compound **28**



¹³C NMR spectrum of compound **29**



¹³C NMR spectrum of compound S14



HSQC spectrum of compound S14



HMBC spectrum of compound S14













 1 H NMR spectrum of compound **23**



¹H NMR spectrum of compound **S16**









 ^{1}H NMR spectrum of compound **32**



F 58.0 -4.0 F 90'E ₩ 60.1 4.5 fl (ppm) 2.5 1.82 0.0 8.5 8.0 7.0 6.0 5.5 5.0 3.5 3.0 2.5 1.5 1.0 0.5 6.5



9.0










¹H NMR spectrum of compound **26**

$$= 156.0404$$

$$= 156.0404$$

$$= 156.0404$$

$$= 156.0404$$

$$= 156.0404$$

$$= 85.8404$$

$$= 85.8404$$

$$= 85.8404$$

$$= 85.8404$$

$$= 85.8404$$

$$= 77.0001$$

$$= 77.0001$$

$$= 77.0001$$

$$= 77.0001$$

$$= 77.3403$$

$$= 23.2778$$

$$= 23.278$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$

$$= 23.3404$$



¹³C NMR spectrum of compound **26**

















¹³C NMR spectrum of compound S19



¹H NMR spectrum of compound **S20**



¹³C NMR spectrum of compound S20





HO HO OH N_3 CO_2H ACHN H_2N N_3 S21



¹³C NMR spectrum of compound S21



 $^1\mathrm{H}$ NMR spectrum of compound S22



¹³C NMR spectrum of compound S22



¹H NMR spectrum of compound **S23**







¹³C NMR spectrum of compound **33**



¹H NMR spectrum of compound **34**







¹³C NMR spectrum of compound S24

















¹³C NMR spectrum of compound **S26b**