

Live-cell Imaging of Lipid Droplets by Solvatochromic Styrylcoumarin Derivatives

Palash Jana^{#‡}, Aravintha Siva^{†‡}, Virupakshi Soppina^{†*} and Sriram Kanvah^{#*}

[#]Department of Chemistry, Indian Institute of Technology Gandhinagar,
Palaj, Gandhinagar 382355, India.

e-mail: sriram@iitgn.ac.in, kanvah@gatech.edu

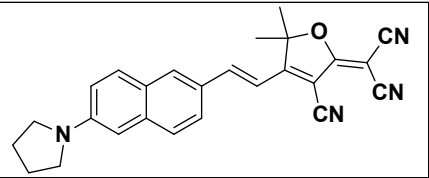
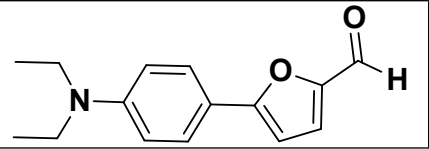
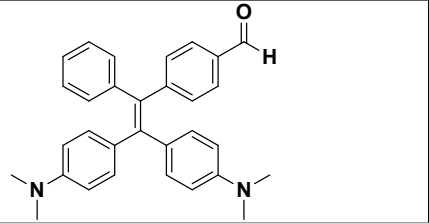
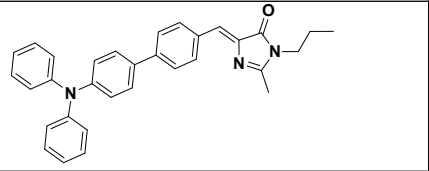
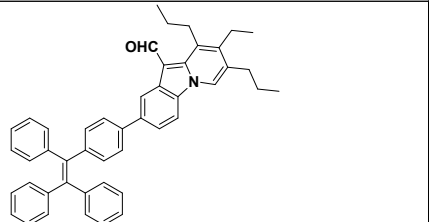
[†]Department of Biological Engineering, Indian Institute of Technology Gandhinagar

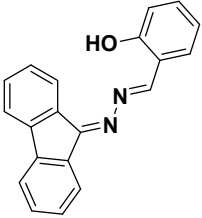
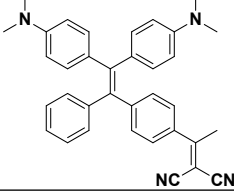
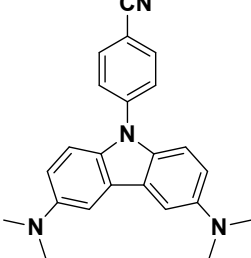
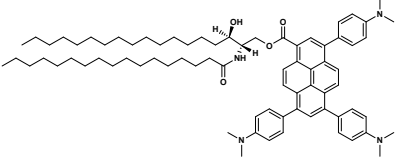
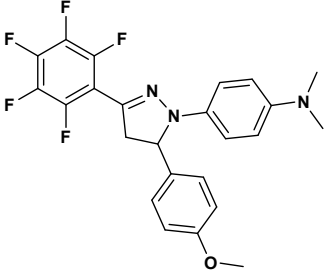
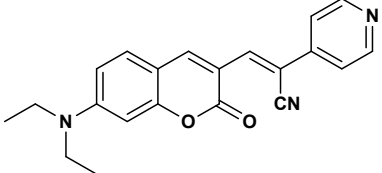
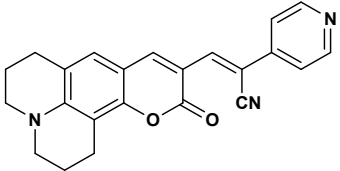
Palaj Gandhinagar 382355

e-mail: vsoppina@iitgn.ac.in, vsoppina@gmail.com

‡ Equal Contribution

Table-S1: Some known structures that were used for imaging lipid droplets

	3 steps + synthesis of starting materials.	λ_{abs} 599 nm, λ_{em} 789 nm (DMSO)	5 μ M	Dyes Pigments 171 (2019) 107718
	1 step,	λ_{abs} .403nm, λ_{em} 520nm	2 μ M	Anal. Chem. 2019, 91, 1928–1935
	2 steps; Coupling Reaction	λ_{abs} . 405nm, λ_{em} . 520nm	10 μ M	J. Mater. Chem. B , 2014, 2 , 2013-2019
	3 steps; Heck Couling and synthesis of starting materials	λ_{abs} . 380nm, λ_{em} 620nm	5 μ M	Chem. Sci. , 2017, 8 , 5440- 5446
	5 steps with Heck reaction	λ_{abs} . 383 nm, λ_{em} 470 nm.	100 μ M	Chem. Asian J. 2017, 12, 2501 – 2509

	2 steps	λ_{abs} 380nm, λ_{em} 595nm	7.5 μ M	ACS Appl. Mater. Interfaces 2016, 8, 16, 10193-10200
	3 steps ,	λ_{abs} 450nm, λ_{em} 705nm	10 μ M	Chem. Commun. , 2016, 52 , 5957-5960
	4 steps, 65% yield	λ_{abs} 384nm, λ_{em} 578nm	12 μ M	J. Org. Chem. 2019, 84, 5535–5547
	4 steps and synthesis of starting materials– 8.5% yield	λ_{abs} 405, λ_{em} 540nm	2 μ M	Analyst, 2019, 144, 1608–1621
	3 steps	λ_{abs} 395 nm, λ_{em} 560nm	0.5 μ M	Chem. Sci., 2019, 10, 9009–9016
	3 steps 85% yield	λ_{abs} 485nm, λ_{em} 580nm	0.2 μ M	This work
	3 step 90% yield	λ_{abs} 512nm, λ_{em} 605nm	0.2 μ M	This work

Highlights

1. Easy synthetic steps with high yields
2. Coumarin and pyridine functionality provides greater biocompatibility

3. Nanomolar concentration of probes used for live cell lipid droplet imaging
4. High photostability with red emission

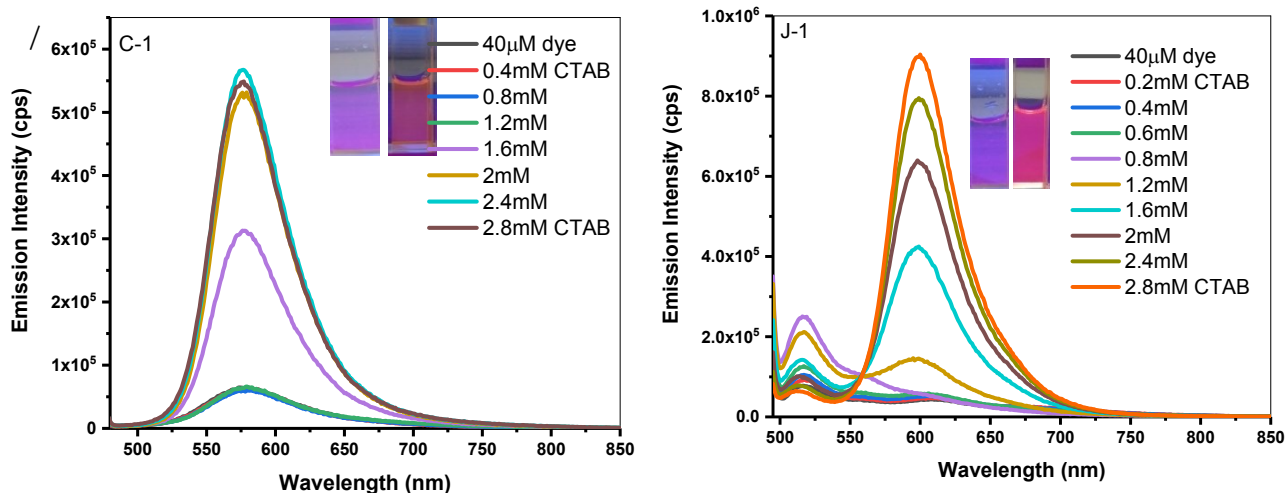


Fig S1: Effect of surfactant (CTAB) concentration on the emission of C-1 and J-1. With increase in concentration, notable emission intensity shifts were noted along with distinct color changes indicating localization preferences.

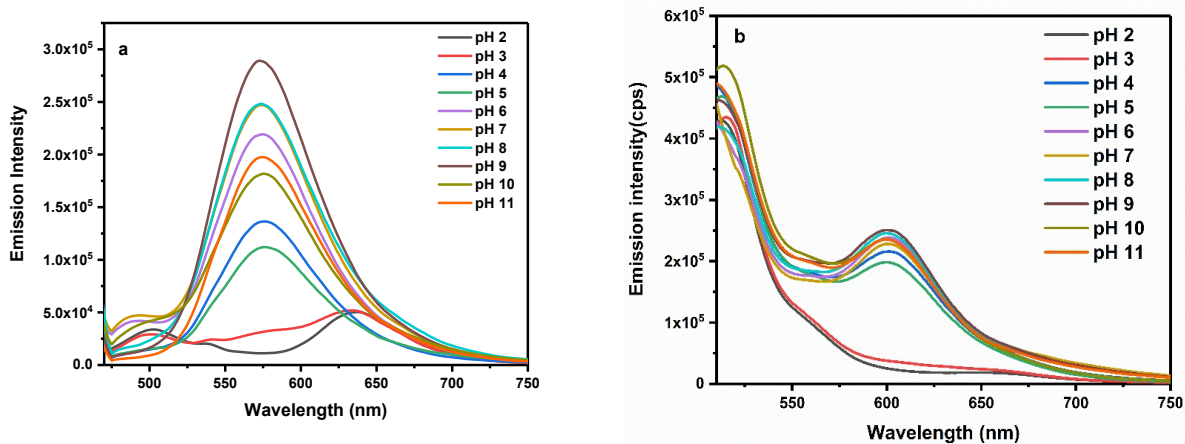


Fig S2: Effect of pH on the emission of C-1 and J-1 (C-1 Ex.wl 450 nm and J-1 Ex Wl 510 nm)

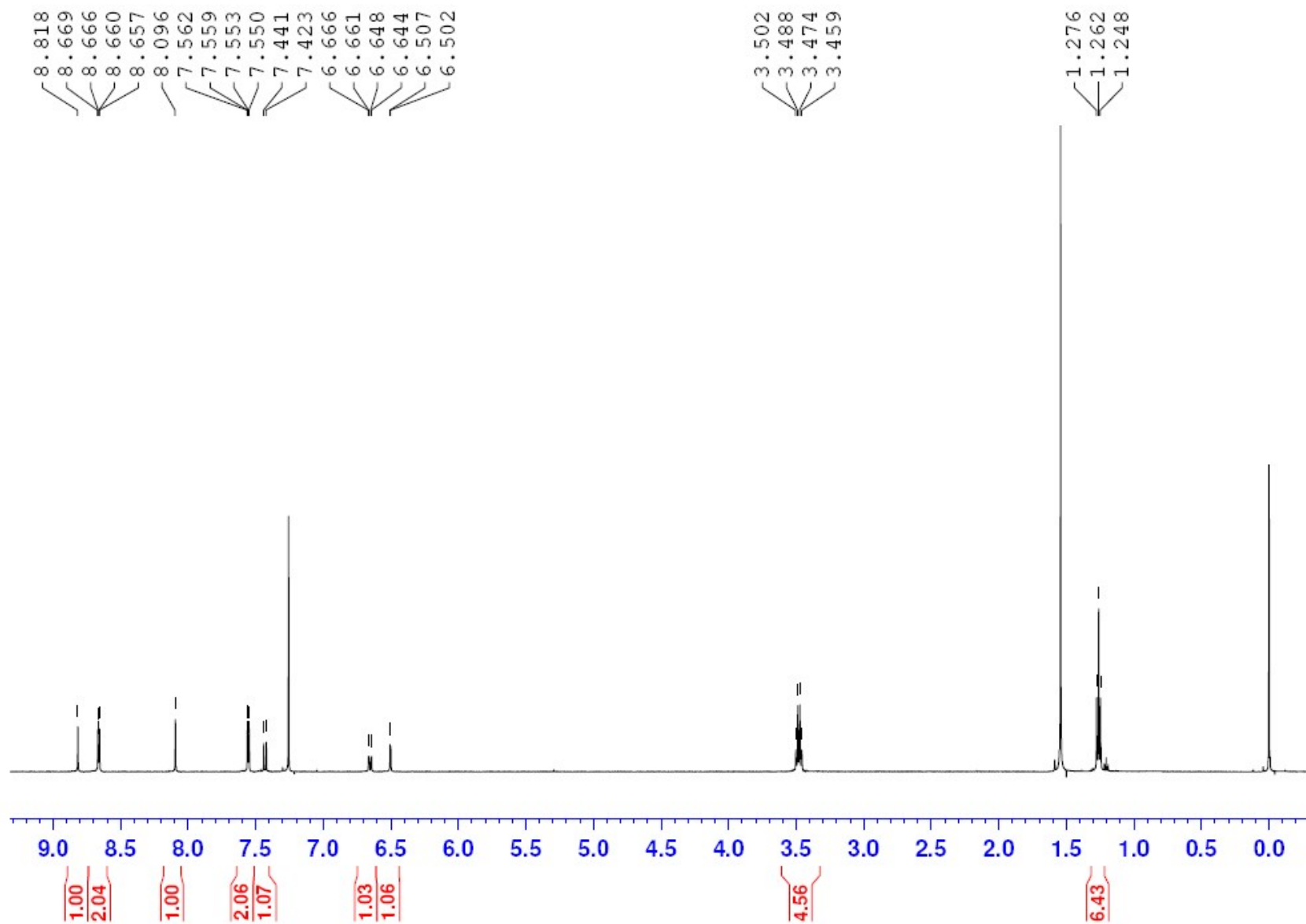
Supporting Information

3-(7-Diethylamino-2-oxo-2*H*-chromen-3-yl)-2-pyridin-4-yl-acrylonitrile (C-1)

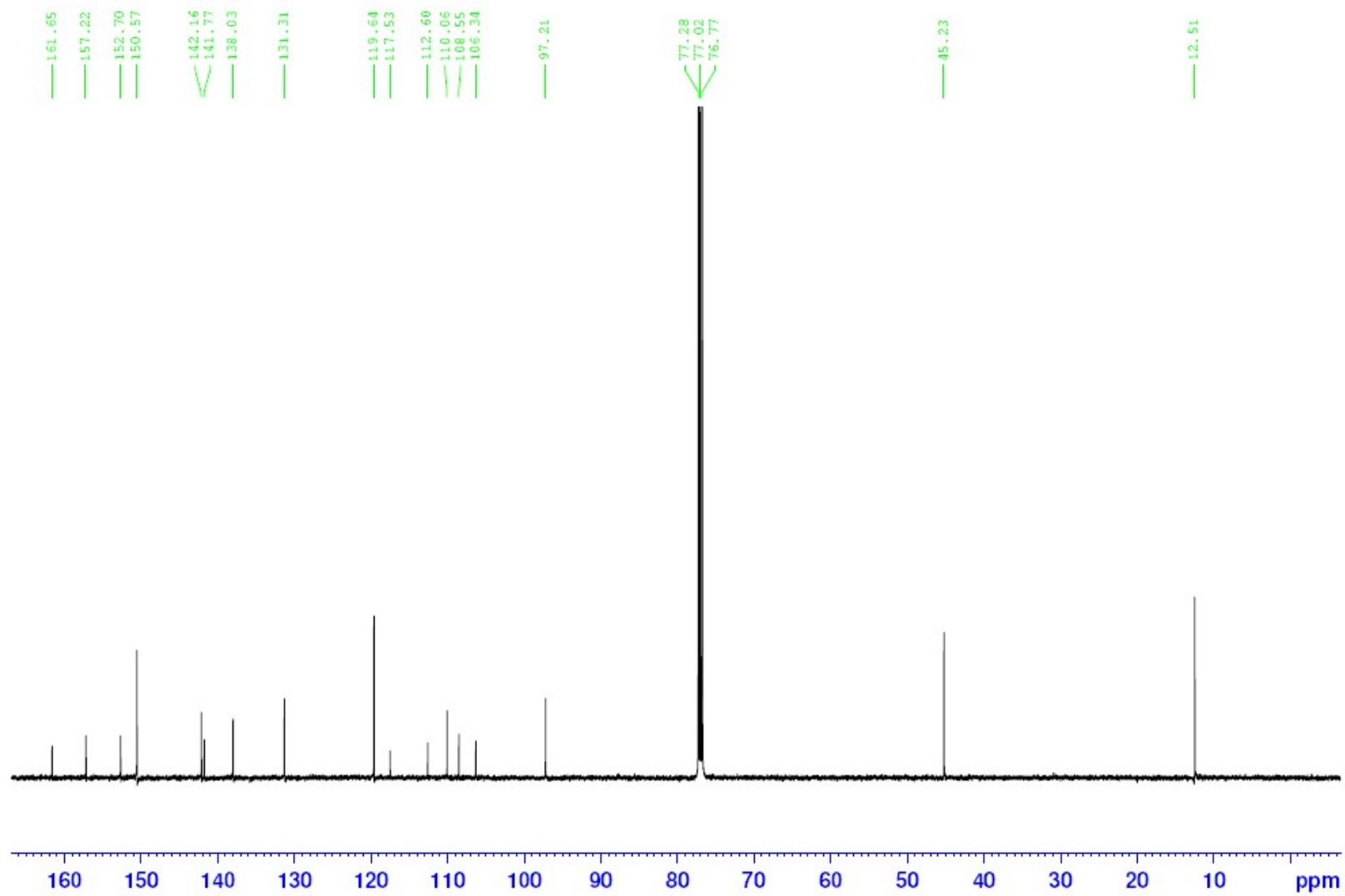
Yield 85 %, (110 mg) ¹H-NMR (500 MHz, CDCl₃) δ (ppm): 8.81 (s, 1H), 8.66–8.65 (d, 2H), 8.09 (s, 1H), 7.56–7.55 (d, 2H), 7.44–7.42 (d, 1H), 6.66–6.64 (d, 1H), 6.50 (s, 1H), 3.50–3.45 (q, 4H), 1.27–1.24 (t, 6H). ¹³C-NMR (125 MHz, DMSO-*d*₆): δ (ppm) 160.69, 157.59, 153.36, 148.30, 143.71, 132.27, 120.81, 117.33, 112, 111, 108.37, 97.13, 55.38, 45.04, 31.16, 22.65, 12.88 HRMS (ESI-Q-TOF): C₂₁H₂₃N₂O₂ [M + H]⁺: cal. *m/z* 346.1585, found, *m/z* 346.1593 (error 2.3ppm)

3-(10-Oxo-2,3,5,6-tetrahydro-1*H*,4*H*,10*H*-11-oxa-3*a*-aza-benzo[*de*]anthracen-9-yl)-2-pyridin-4-yl-acrylonitrile (J-1)

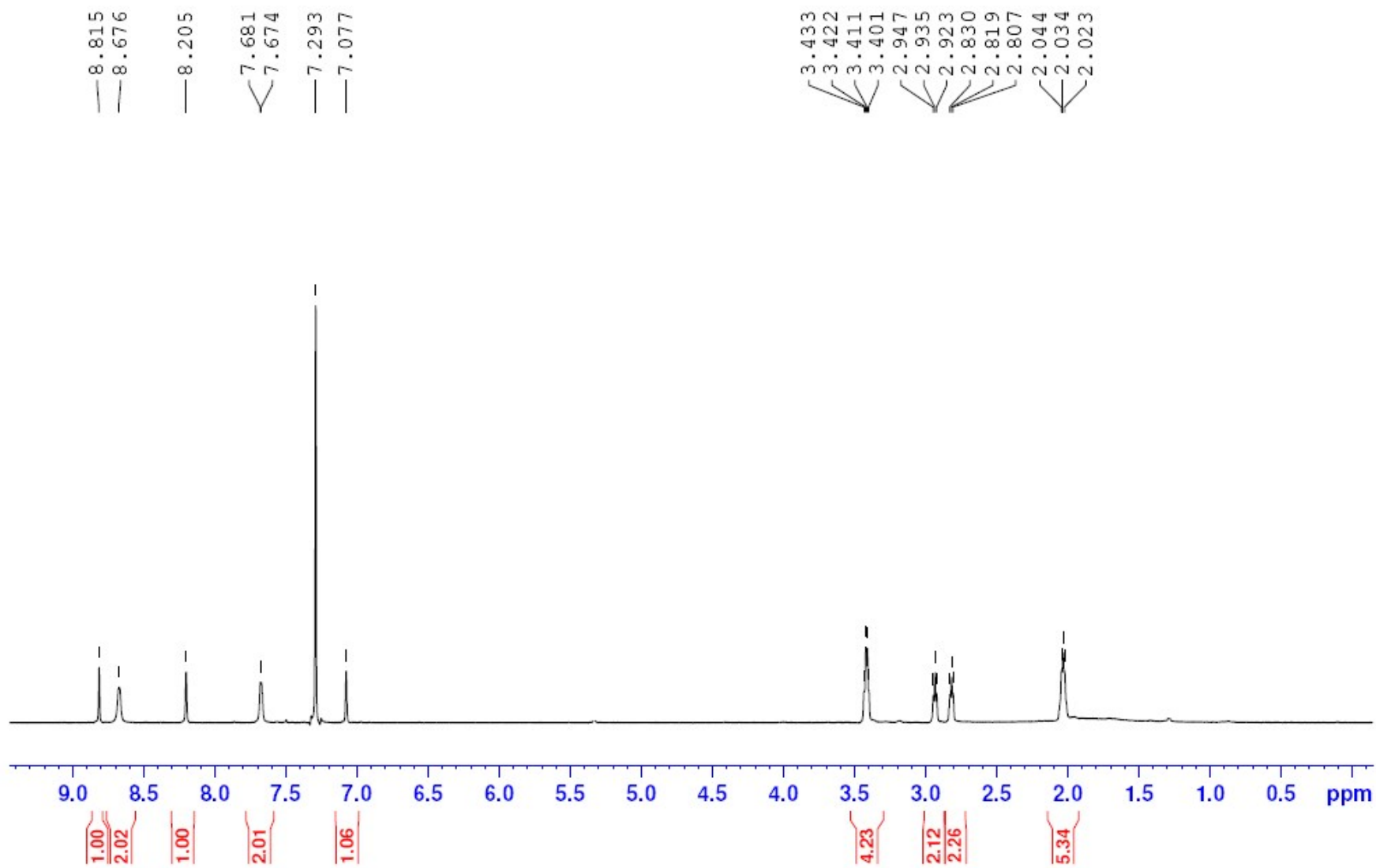
Yield 90%. ¹H-NMR (500 MHz, CDCl₃) δ (ppm): 8.81 (s, 1H), 8.67–8.66 (d, 2H), 8.2 (s, 1H), 7.68–7.67 (d, 2H), 7.68–7.67 (d, 2H), 7.0 (s, 1H), 3.43–3.40 (t, 4H), 2.94–2.92 (m, 2H), 2.83–2.80 (m, 2H), 2.04–2.02 (t, 4H) ¹³C-NMR (125 MHz, CDCl₃): δ (ppm) 161.92, 152.24, 150.14, 148.54, 142.17, 138.53, 127.40, 119.78, 119.61, 117.76, 111.23, 108.54, 106.31, 104.90, 96.14, 50.42, 27.38, 21.16, 20.19, 19.99. HRMS (ESI-Q-TOF): C₂₁H₂₃N₂O₂ [M+ H]⁺: cal. *m/z* 370.1598, found, *m/z* 370.1601 (error 0.8pm)



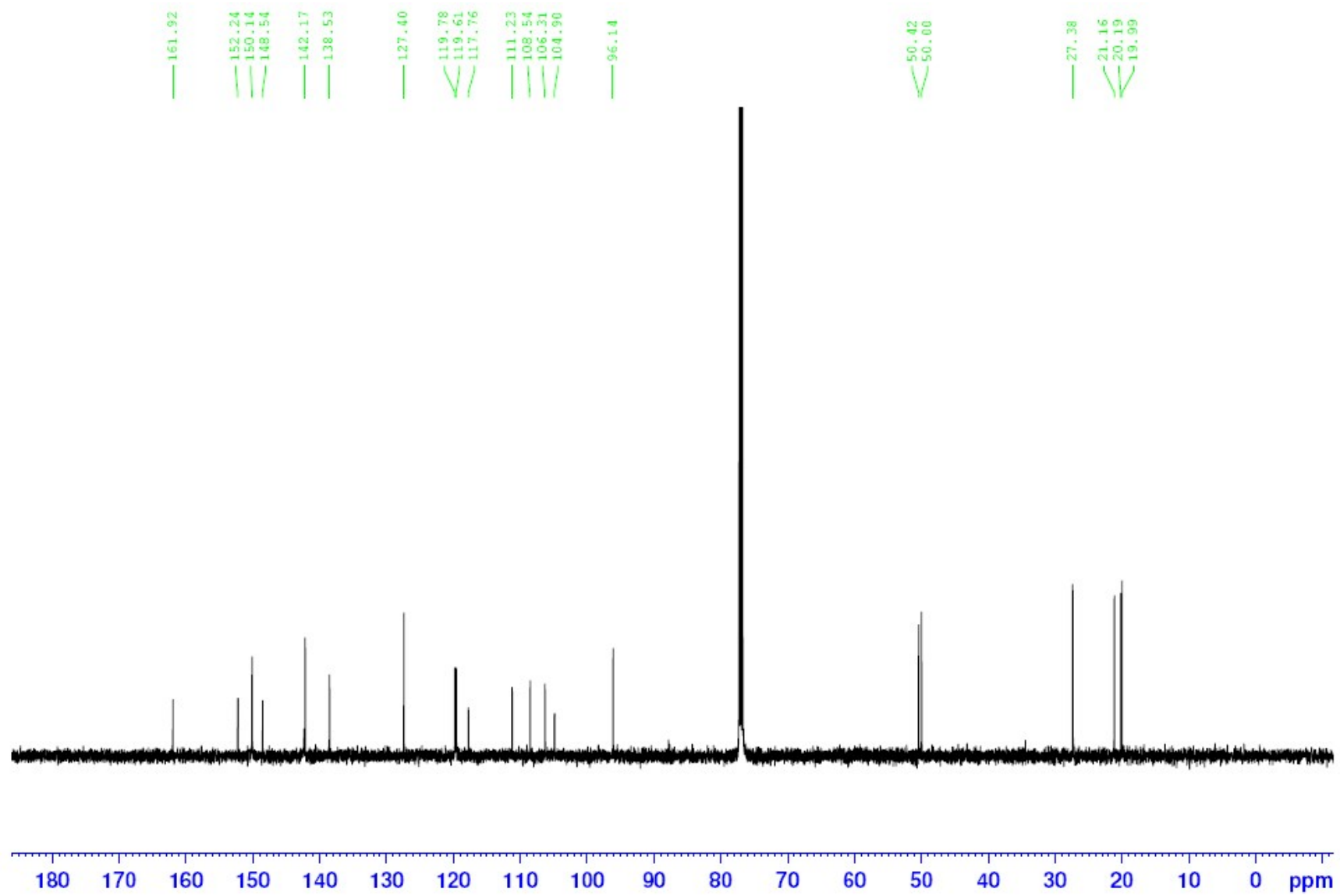
¹H-NMR spectrum of compound C-1



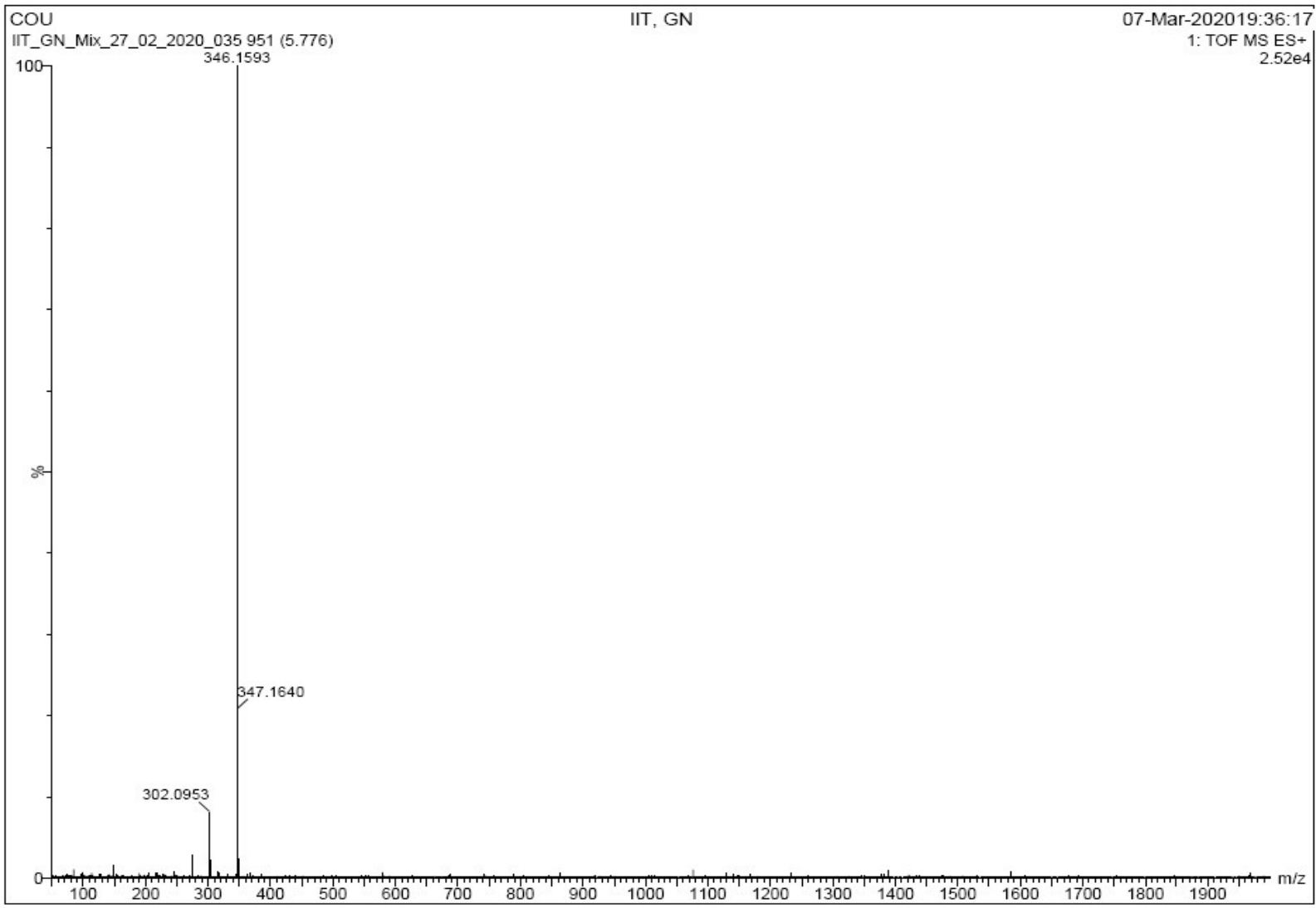
^{13}C -NMR spectrum of compound C-1



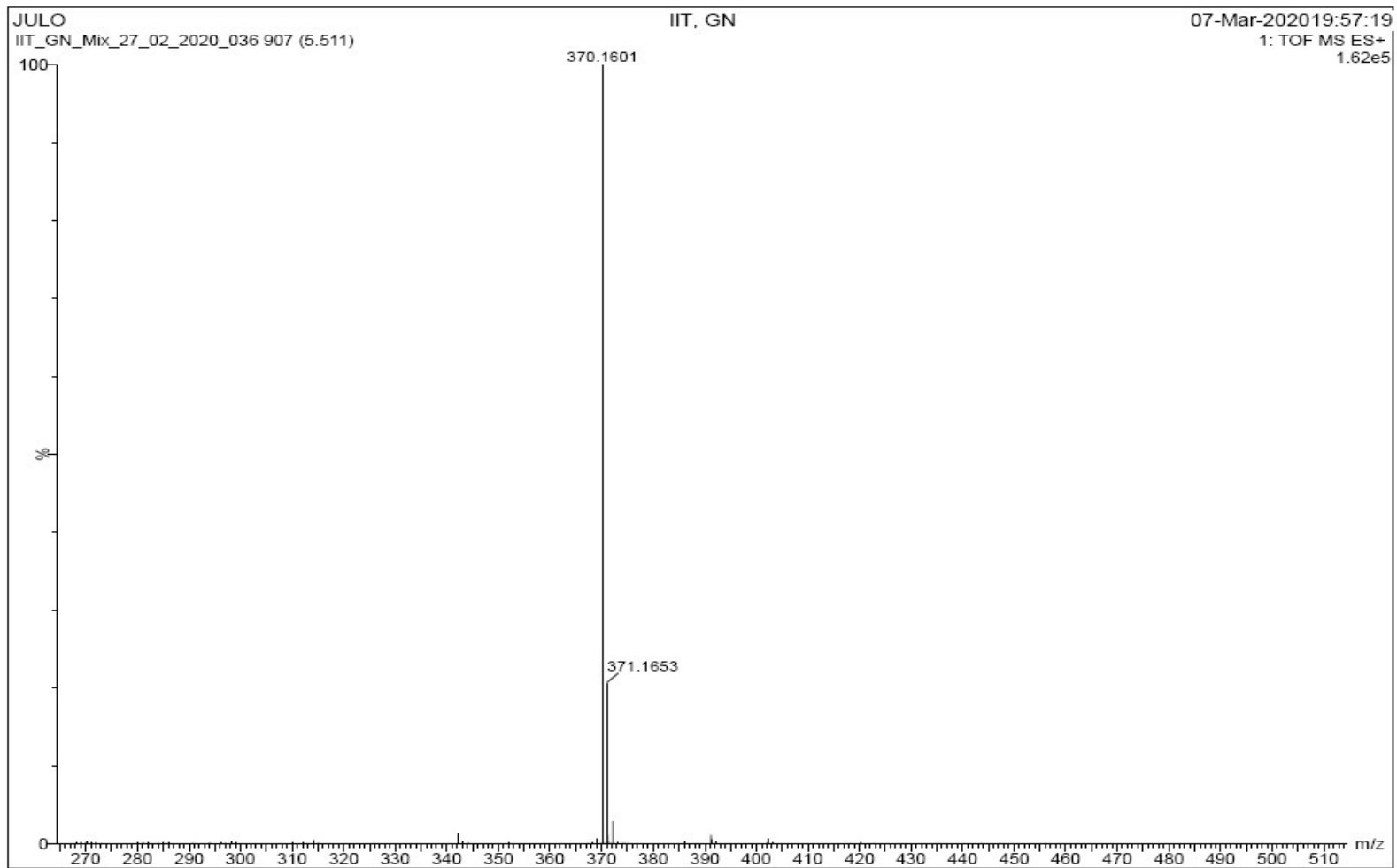
¹H-NMR spectrum of compound J-1



^{13}C -NMR spectrum of compound J-1



HRMS spectrum of compound C-1



HRMS spectrum of compound J-1