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Electronic Supplementary Material

Broadening the reaction scope of unprotected aldoses via their corresponding nitrones: 1,3-dipolar cycloadditions with alkenes.

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¹³C-NMR spectrum of 5b (CD₃OD)





¹³C-NMR spectrum of 5c (CD₃OD)

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¹³C-NMR spectrum (J-MOD) of 7a (CD₃OD)



















¹H-NMR spectrum of 10' (CD₃OD)




























































































Hydrogenation of compounds 10, 10' and 10" for chemical correlation

Procedure : compound **10**, **10'** or **10''** (0.53 mmol) in MeOH (14 mL) and AcOH (1.6 mL) were stirred under H₂ atmosphere for 24 h in the presence of Pearlman's catalyst (Pd(OH)₂/C, 74 mg). Filtration of the solution over a small pad of silica followed by evaporation of the solvents afforded a yellow oil, which was purified by chromatography on silical gel (CHCl₃/MeOH/NH₄OH, 6/4/1; Rf = 0.2) to afford pyrrolidone **13** in quantitative yield.





¹H-NMR spectrum of 2-pyrrolidinones isolated after hydrogenation of 10' (reaction was performed on a fraction which contained the 2 major diastereoisomers)


¹³C-NMR spectrum of 2-pyrrolidinones isolated after hydrogenation of 10' (reaction was performed on a fraction which contained the 2 major diastereoisomers) – extension shows C-4





¹H-NMR spectrum of 2-pyrrolidinones isolated after hydrogenation of 10" (4 diastereoisomers)



13C-NMR spectrum of 2-pyrrolidinones isolated after hydrogenation of 10" (4 diastereoisomers) – extension shows C-4





Enzyme assay: all enzymes were purchased from Sigma Chemical Co. In a typical experiment, the glycosidase (0.013 U/mL) was pre-incubated at 33 °C for 5 min in the presence of the **13** in 50 mM acetate buffer (pH 5.6, except for rice α -glucosidase pH 5.1 and yeast α -glucosidase pH 6.2). The reaction was started by addition of the appropriate substrate (*p*-nitrophenyl glycoside, 1 mM concentration) to a final volume of 250 µl. The reaction was stopped after 10-15 min (depending on the enzyme) by addition of 300 µL of 0.4 M Na₂CO₃. The released *p*-nitrophenolate was quantified spectrophotometrically at 415 nm with a microplate reader (300 µL of the reaction mixture in a well, OD *ca* 0.7 for the control, without inhibitor). All the assays were done in duplicate (less than 10% variability in each case). Absorbance was corrected by subtracting the blank. The blank was measured in an additional experiment, conducted in the absence of enzyme. The control experiment contained no inhibitor.

Percentage of inhibition was calculated as follows:

$$% = \left(1 - \frac{\text{Asample}}{\text{Acontrol}}\right) * 100$$