Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2020

Supporting Information

N-Primary-Amine Tetrapeptide-Catalyzed Highly Asymmetric

Michael Addition of Aliphatic Aldehydes to Maleimides†

Zhi-Hong Du,^a Wen-Juan Qin,^a Bao-Xiu Tao,^a Meng Yuan^a and Chao-Shan Da^{*ab} ^aInstitute of Biochemistry and Molecular Biology, School of Life Sciences, Lanzhou University, Lanzhou 730000, China ^bState Key Laboratory of Applied Organic Chemistry, Key Lab of Preclinical Study for New Drugs of Gansu Province, Lanzhou University, Lanzhou 730000, China dachaoshan@lzu.edu.cn

Table of Contents

Supporting Information	1
1. General Experimental Details	1
2. Preparation and characterization of peptide catalysts	2
3. General procedure for the Michael addition reaction.	5
4. Synthesis of product 6	5
5. Synthesis of product 7	6
6. Synthesis of product 8	6
7. Characterization of Michael addition reaction products	7
8. Copies of NMR spectra	42
9. HPLC analytical of the tetrapeptides 1d and 1e	75
10. Images of the HR-MS of the new compounds	76
11. Reference	80

1. General Experimental Details

The reactions were carried out in vials and stirred with a magnetic bar without inert atmosphere unless specified. All commercial reagents were purchased with the analysis purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate (EtOAc) were distilled. Anhydrous DCM and CH₃CN were freshly distilled from CaH₂. THF, Et₂O and toluene were freshly distilled from Sodium/benzophenone before use. Anhydrous methanol and ethanol were distilled from Mg.

The reactions were monitored by TLC (thin layer chromatography) method; column and preparative TLC purifications were carried out using silica gel. Melting points were uncorrected and recorded on XT-5 melting point apparatus.

NMR spectra were acquired on a Bruker 400/600 spectrometer, running at 400/600 MHz and 100/151 MHz for ¹H and ¹³C, respectively. NMR in CDCl₃, D₂O, DMSO-d₆ with TMS as an internal standard, chemical shifts (δ) are reported in ppm relative to residual solvent signals (CDCl₃, 7.26 ppm for ¹H NMR and 77.00 ppm for ¹³C NMR; D₂O, 4.80 ppm for ¹H NMR; DMSO-d₆, 2.50 ppm for ¹H NMR, 40.00 ppm for ¹³C NMR). The following abbreviations are used to describe peak patterns when appropriate: s (singlet), d (doublet), t (triplet), q (quartet), quint (quintet), sept (septuplet), m (multiplet), br (broad). High resolution mass spectra (HR-MS) were measured with ESI-Orbitrap mass spectrometer.

Enantiomeric excess (ee) were decided with chiral HPLC, Waters 1525 Binary HPLC Pump/Waters 2998 Photodiode Array Detector of Lanzhou University State Key Laboratory of Applied Organic Chemistry.

2. Preparation and characterization of peptide catalysts



For the synthesis steps of the tetrapeptide catalyst, please refer to Reference¹

1a: NH₂-Val-DPro-Gly-Leu-OH: white solid, mp 198 – 200 °C; $[\alpha]_D^{25} = + 36.0$ (*c* 0.5, MeOH); ¹H NMR (600 MHz, D₂O) δ 4.53 – 4.51 (m, 1H), 4.30 – 4.25 (m, 2H), 4.05 – 4.02 (m, 1H), 3.92 – 3.83 (m, 3H), 3.77 – 3.73 (m, 1H), 2.39 – 2.34 (m, 2H), 2.12 – 2.05 (m, 3H), 1.73 – 1.63 (m, 4H), 1.13 (d, *J* = 7.2 Hz ,3H), 1.05 (d, *J* = 6.6 Hz ,3H), 0.95 (d, *J* = 6 Hz, 3H), 0.91 (d, *J* = 6 Hz, 3H); ¹³C NMR (151 MHz, D₂O) δ 179.7, 174.3, 170.4, 168.9, 61.1, 57.1, 53.5, 48.1, 42.4, 40.5, 29.2, 28.7, 24.5, 24.3, 22.4, 20.5, 17.9, 15.9; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₁₈H₃₃N₄O₅ 385.2445, found: 385.2438.

1b: **NH₂-Tle-DPro-Gly-Leu-OH:** white solid, mp: 223 – 225 °C, $[\alpha]_D^{25} = + 4.0$ (*c* 0.5, MeOH); ¹H NMR (600 MHz, D₂O) 4.37 – 4.34 (m, 1H), 4.10 (dd, J = 10.8, 3.6 Hz, 1H), 4.05 (s, 1H), 3.85 – 3.77 (m, 2H), 3.72 – 3.63 (m, 2H), 3.21 (d, J = 0.6 Hz, 1H), 2.23 – 2.19 (m, 1H), 1.94 – 1.88 (m, 3H), 1.58 – 1.51 (m, 1H), 1.49 – 1.44 (m, 2H), 0.97 (s, 9H), 7.78 – 7.73 (m, 6H); ¹³C NMR (151 MHz, D₂O) δ 179.8, 174.4, 170.4, 168.2, 61.0, 59.1, 53.7, 49.0, 42.4, 40.6, 33.9, 29.3, 25.3, 24.5, 24.3, 22.4, 20.5; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₁₉H₃₁N₄O₅ 399.2602, found: 399.2599.

1c: NH₂-Chg-DPro-Gly-Leu-OH: white solid, mp 212 – 214 °C; $[\alpha]_D^{25} = +32$ (*c* 0.5, MeOH); ¹H NMR (600 MHz, D₂O) δ 4.51 – 4.48 (m, 1H), 4.30 – 4.23 (m, 2H), 4.07 – 4.01 (m, 1H), 3.93 – 3.72 (m, 3H), 2.41 – 2.32 (m, 1H), 2.11 – 1.96 (m, 3H), 1.83 – 1.58 (m, 7H), 1.35 – 1.12 (m, 5H), 0.96 – 0.90 (m, 6H); ¹³C NMR (151 MHz, D₂O) δ 179.6, 174.3, 170.4, 168.9, 61.1, 56.6, 53.9, 53.4, 48.1, 42.4, 40.5, 38.5, 29.3, 28.5, 27.1, 25.1, 24.5, 24.2, 22.4, 20.5; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₂₁H₃₇N₄O₅ 425.2758, found: 425.2748.

1d: NH₂-Phg-DPro-Gly-Leu-OH: white solid, mp 210 – 212 °C; $[\alpha]_D^{25} = +72.0$ (*c* 0.5, MeOH); ¹H NMR (600 MHz, D₂O) δ 7.60 – 7.44 (m, 5H), 5.47 (s, 1H), 4.50 (dd, J = 9.0, 3.6 Hz, 1H), 4.31 (dd, J = 10.2, 3.0 Hz, 1H), 4.11 – 4.08 (m, 1H), 3.90 – 3.87 (m, 1H), 3.77 – 3.73 (m, 1H), 3.05 – 3.02 (m, 1H), 2.23 – 2.16 (m, 1H), 2.02 – 1.95 (m, 2H), 1.85 – 1.73 (m, 2H), 1.68 – 1.63 (m, 2H), 0.98 – 0.87 (m, 6H); ¹³C NMR (151 MHz, D₂O) δ 179.9, 174.3, 170.3, 167.4, 130.6, 129.9, 129.6, 129.0, 128.5, 61.5, 56.4, 53.6, 47.5, 42.4, 40.6, 29.0, 24.6, 24.2, 22.5, 20.6; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₂₁H₃₁N₄O₅ 419.2289, found: 419.2295.

ent-1d: NH₂-DPhg-Pro-Gly-DLeu-OH: white solid, mp 210 - 212 °C; $[\alpha]_D^{25} = -72.0$ (*c* 0.5, MeOH).

1e: NH₂-Phe-DPro-Gly-Leu-OH: white solid, mp 224 – 226 °C, $[\alpha]_D^{25} = +60.0$ (*c* 0.5, MeOH); ¹H NMR (600 MHz, D₂O) 7.47 – 7.41 (m, 3H), 7.35 (d, J = 6.6 Hz, 2H), 4.60 (dd, J = 9.0, 6.6 Hz, 1H), 4.37 (dd, J = 9.0, 4.8 Hz, 1H), 4.26 (dd, J = 10.2, 3.6 Hz, 1H), 4.02 – 3.99 (m, 1H), 3.90 – 3.87 (m, 1H), 3.62 – 3.58 (m, 1H), 3.28 – 3.19 (m, 2H), 2.85 – 2.81 (m, 1H), 2.16 – 2.09 (m, 1H), 1.96 – 1.85 (m, 2H), 1.72 – 1.62 (m, 4H), 0.95 – 0.89 (m, 6H); ¹³C NMR (151 MHz, D₂O) δ 179.8, 174.1, 170.3, 168.5, 133.4, 129.4, 129.1, 128.1, 61.0, 53.6, 53.0, 47.7, 42.4, 40.5, 36.3, 29.1, 24.5, 24.0, 22.4, 20.5; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₂₂H₃₃N₄O₅ 433.2445, found: 433.2439.

1f: NH₂-Phe-DPro-Gly-Phe-OH: white solid, mp 203 – 205 °C, $[\alpha]_D^{25} = +$ 74.0 (*c* 0.5, MeOH); ¹H NMR (600 MHz, DMSO-d₆) δ 8.42 – 8.18 (m, 2H), 7.35 – 7.12 (m, 10H), 4.36 –

4.14 (m, 3H), 3.72 - 3.68 (m, 1H), 3.48 - 3.37 (m, 2H), 3.20 - 3.12 (m, 1H), 3.02 - 2.85 (m, 3H), 1.84 - 1.68 (m, 3H), 1.39 - 1.35 (m, 1H), 1.00 (d, J = 6.6 Hz, 1H); ¹³C NMR (151 MHz, DMSO-d₆) δ 175.5, 171.5, 170.2, 168.3, 139.3, 136.0, 129.9, 129.7, 128.9, 128.3, 127.6, 126.3, 61.2, 52.9, 47.1, 42.1, 38.8, 29.4, 24.4, 23.7; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₂₅H₃₁N₄O₅ 467.2288, found: 467.2289.

Boc-Gly-DPro-Gly-Leu-OBn: yellow oil, $[\alpha]_D^{25} = +32.0$ (*c* 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 7.38 – 7.27 (m, 4H), 7.17 (d, *J* = 7.8 Hz, 1H), 7.09 (s, 1H), 5.45 (s, 1H), 5.14 (s, 2H), 4.68 – 4.65(m, 1H), 4.44 (dd, *J* = 7.8, 4.2 Hz, 1H), 4.01 – 3.92 (m, 3H), 3.82 (dd, *J* = 17.4, 3.0 Hz, 1H), 3.62 – 3.59 (m, 1H), 3.46 (dd, *J* = 16.2, 7.2 Hz, 1H), 2.22 – 2.07 (m, 3H), 2.02 – 1.98 (m, 1H), 1.72 – 1.65 (m, 3H), 1.43 (s, 9H), 0.92 (dd, *J* = 9.0, 6.0 Hz, 6H). ¹³C NMR (151 MHz, CDCl₃) δ 173.1, 171.5, 169.4, 168.9, 135.4, 128.6, 128.3, 128.1, 79.8,67.0, 61.0), 50.8, 46.6, 43.1, 41.0, 28.4, 25.1, 24.8, 22.9, 21.8.

Boc-Ala-DPro-Gly-Leu-OBn: yellow oil, $[\alpha]_D^{25} = -7.0$ (*c* 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 7.36 – 7.35 (m, 4H), 7.21 (d, *J* = 7.8 Hz, 1H), 5.37 (d, *J* = 6.0 Hz, 1H), 5.16 (d, *J* = 3.6 Hz, 2H), 4.65 – 4.62 (m 1H), 4.48 (dd, *J* = 7.8, 4.2 Hz, 1H), 4.36 – 4.32 (m, 1H), 4.12 – 4.04 (m, 1H), 3.96 (d, *J* = 12.6 Hz, 1H), 3.72 (dd, *J* = 16.8, 5.4 Hz, 1H), 3.54 (dd, *J* = 17.4, 7.8 Hz, 1H), 2.21 – 2.15 (m, 3H), 2.11 – 2.06 (m, 1H), 2.02 – 1.98 (m, 2H), 1.69 – 1.64 (m, 4H), 1.40 (s, 9H), 1.27 (d, *J* = 7.2 Hz, 3H), 0.90 (dd, *J* = 10.2, 6.0 Hz, 6H). ¹³C NMR (151 MHz, CDCl₃) δ 173.0, 172.7, 171.6, 169.0, 135.7, 128.5, 128.2, 128.0, 80.4, 66.6, 61.2, 50.8, 47.5, 43.3, 41.0, 29.0, 28.3, 24.8, 24.7, 22.9, 21.8.

Boc-Phg-Gly-Leu-OBn: yellow oil, $[\alpha]_D^{25} = +53.0$ (*c* 1.0, CHCl₃); ¹H NMR (600 MHz,CDCl₃) δ 7.37 – 7.26 (m, 9H), 6.89 (s, 1H), 6.77 (s, 1H), 5.72 (d, *J* = 6.0 Hz, 1H), 5.17 – 5.11(m, 3H), 4.62 – 4.58 (m, 1H), 4.03 (dd, *J* = 16.8, 6.0Hz, 1H), 3.84 (dd, *J* = 16.8, 4.8Hz, 1H), 1.61 –1.54 (m, 3H), 1.41 (s, 9H), 0.86 (t, *J* = 6.0 Hz, 6H). ¹³C NMR (151 MHz, CDCl₃) δ 172.5, 170.8, 168.4, 135.4, 129.1, 128.6, 128.4, 128.2, 127.3, 67.1, 51.0, 43.2, 41.1, 28.3, 24.7, 22.7, 21.8.

1g: NH₂-Gly-DPro-Gly-Leu-OH: white solid, mp 167 – 169 °C, $[\alpha]_D^{25} = + 34.0$ (*c* 1.0, MeOH); ¹H NMR (600 MHz, D₂O) δ 4.32 – 4.26 (m, 2H), 4.12 (dd, J = 10.2, 3.0 Hz, 1H), 3.88 (d, J = 17.4 Hz, 1H), 3.71 (d, J = 17.4 Hz, 1H), 3.68 – 3.65 (m, 1H), 3.54 – 3.50 (m, 1H), 2.22 – 2.19 (m, 1H), 1.96 – 1.88 (m, 2H), 1.54 – 1.46 (m, 3H), 1.38 (d, J = 6.6 Hz, 2H), 1.08 (s, 1H), 0.79 – 0.73 (m, 6H). ¹³C NMR (151 MHz, D₂O) δ 182.4, 177.0, 172.9, 168.9, 63.6, 56.2, 51.2, 49.4, 44.9, 43.1, 31.9, 28.4, 26.9, 25.0, 23.2; HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₅H₂₆N₄NaO₅ 365.1795, Found 365.1807.

1h: NH₂-Ala-DPro-Gly-Leu-OH: white solid, mp 234.3 – 236.2 °C, $[\alpha]_D^{25} = +19.0$ (*c* 1.0, MeOH); ¹H NMR (600 MHz, D₂O) δ 4.32 (dd, *J* = 9.0, 5.4 Hz, 1H), 4.11 (dd, *J* = 10.2, 3.6 Hz, 1H), 3.90 – 3.87 (m, 2H), 3.72 (d, *J* = 16.8 Hz, 1H), 3.53 – 3.50 (m, 1H), 3.48 – 344 (m, 1H), 3.09 (s, 1H), 2.23 – 2.20 (m, 1H), 1.94 – 1.87 (m, 3H), 1.55 – 1.46 (m, 3H), 1.08 (s, 3H), 0.76 (dd, *J* = 24.0, 6.0 Hz, 6H). ¹³C NMR (151 MHz, D₂O) δ 179.6, 174.4, 170.4, 169.8, 61.3, 53.5, 48.3, 47.6, 42.3, 40.5, 29.2, 25.9, 24.3, 22.4, 20.5, 14.6; HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₆H₂₈N₄NaO₅ 379.1952, Found 379.1956.

1i: NH₂-Phg-Gly-Leu-OH: white solid, mp 233 – 235 °C, $[\alpha]_D^{25} = +36.0$ (*c* 0.5, MeOH); ¹H NMR (600 MHz, DMSO-d₆) δ 8.57 (s, 1H), 7.98 (d, *J* = 8.4 Hz, 1H), 7.45 (d, *J* = 7.8 Hz, 2H), 7.34 – 7.26 (m, 3H), 4.61 (s, 1H), 4.22 – 4.19 (m, 1H), 3.78 – 3.66 (m, 2H), 1.59 – 1.56 (m, 1H), 1.51 – 1.41 (m, 2H), 0.84 (dd, *J* = 17.4, 6.6 Hz, 6H). ¹³C NMR (151 MHz, DMSO-d₆) δ 174.4, 171.6, 167.9, 140.0, 128.1, 127.4, 127.1, 57.8, 50.8, 42.0, 40.9, 39.8, 39.7, 39.5, 39.4, 39.3 39.1, 39.0, 24.1, 22.9, 22.0; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₁₆H₂₄N₃O₄ 322.1761, Found 3221764.

3. General procedure for the Michael addition reaction.



In a 5 mL round bottom flask equipped with a magnetic stirrer, the catalyst 1d (5.2 mg, 0.0125 mmol), maleimide (0.5 mmol), aldehyde 3 (1 mmol) and acetonitrile (1.0 mL) were sequentially added and stirred at room temperature and detected by TLC. After the reaction was completed, the mixture was directly purified by column chromatography on silica gel (PE: EA=5:1) to give products.

4. Synthesis of product **6**.²



Aldehyde **4a** (122.5 mg, 0.5 mmol) was dissolved in dry DMF (5 ml), potassium peroxymonosulfate (622 mg, 1.0 mmol) was added and the mixture was stirred at room temperature until the reaction completed. The reaction mixture was extracted with EtOAc ($3 \times 20 \text{ mL}$) and 1 M aq. HCl (20 mL). The combined organic layers were washed with 1 M aq. HCl (50 mL) and brine (50 mL), dried over Na₂SO₄, filtered and all volatiles were removed at reduced pressure. The crude product was purified by column chromatography on silica gel (PE: EA=5:1) to give products **6** as a colorless solid (115 mg, 88% yield).

(*S*)-2-(2,5-dioxo-1-phenylpyrrolidin-3-yl)-2-methylpropanoic acid, ¹H NMR (600 MHz, DMSO-d₆) δ 12.64 (s, 1H), 7.52 – 7.47 (m, 2H), 7.44 – 7.39 (m, 1H), 7.23 – 7.19 (m, 2H), 3.18 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.94 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.70 – 2.60 (m, 1H), 1.37 (s, 3H), 1.22 (s, 3H).

5. Synthesis of product 7.



Aldehyde **4a** (122.5 mg, 0.5 mmol) was dissolved in dry MeOH (5.0 mL) under an argon atmosphere and the solution was cooled to 0 $^{\circ}$ C. NaBH₄ (95 mg, 2.5 mmol) was added. Then the mixture was allowed to warm up to room temperature and kept stirring until the reaction completed. The reaction mixture was extracted with CH₂Cl₂ (3 × 20 mL), the combined organic layers were dried over Na₂SO₄, filtered and all volatiles were removed at reduced pressure. The crude product was purified by column chromatography on silica gel (PE: EA=5:1) to give lactone **7** as a colorless solid (94 mg, 76% yield).

6. Synthesis of product 8.



Aldehyde **4a** (122.5 mg, 0.5 mmol) was dissolved in dry CH_2Cl_2 (5.0 mL) under an argon atmosphere and the solution was cooled to 0 °C. AcOH (60 µL, 1 mmol), benzylamine (110 µL, 1.0 mmol) and NaBH(OAc)₃ (328 mg, 1.50 mmol) were added, the mixture was allowed to warm up to room temperature and kept stirring for 4 h. DIEA (175 µl, 1.0 mmol) was added and the stirring was continued for 20 h. The mixture was then extracted with CH_2Cl_2 (3 × 20 mL). The combined organic layers were dried over Na₂SO₄, filtered and all volatiles were removed at reduced pressure. The crude product was purified by column chromatography on silica gel (PE: EA=5:1) to give lactam **8** as a colorless solid (104 mg, 62%).

7. Characterization of Michael addition reaction products

(S)-2-(2, 5-dioxo-1-phenylpyrrolidin-3-yl)-2-methylpropanal³



4a: 120 mg, 98% yield, white solid, $[\alpha]_D^{25} = -6.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.51 (s, 1H), 7.48 – 7.44 (m, 2H), 7.40 – 7.36 (m, 1H), 7.29 – 7.25 (m, 2H), 3.14 (dd, *J* = 9.6, 5.6 Hz, 1H), 2.96 (dd, *J* = 18.6, 9.6 Hz, 1H), 2.61 (dd, *J* = 18.4, 5.6 Hz, 1H), 1.32 (s, 3H), 1.28 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 225.1 nm; retention time: 13.8 min (major) and 17.3 min (minor).



(*R*)-2-(2, 5-dioxo-1-phenylpyrrolidin-3-yl)-2-methylpropanal³



*ent-***4a:** 120 mg, 98% yield, white solid, $[\alpha]_D^{25} = +6.0$ (c = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.51 (s, 1H), 7.48-7.44 (m, 2H), 7.40 – 7.36 (m, 1H), 7.29 – 7.25 (m, 2H), 3.14 (dd, J = 9.6, 5.6 Hz, 1H), 2.96 (dd, J = 18.6, 9.6 Hz, 1H), 2.61 (dd, J = 18.4, 5.6 Hz, 1H), 1.32 (s, 3H), 1.28 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 225.1 nm; retention time: 14.9 min (minor) and 17.8 min (major).





4b, 106 mg, 82% yield, white solid, $[\alpha]_D^{25} = -3(c = 1.0, CHCl_3)$; ¹H NMR (600 MHz, CDCl₃) δ 9.51 (s, 1H), 7.35 – 7.30 (m, 2H), 7.30 – 7.27 (m, 1H), 7.12 – 7.01 (m, 1H), 3.20 – 3.15 (m, 1H), 2.98 – 2.94 (m, 1H), 2.70 – 2.65 (m, 1H), 2.21(s, 2H), 2.14 (s, 1H), 1.35 (d, *J* = 5.4 Hz, 3H), 1.29 (d, *J* = 2.8 Hz, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 223.3 nm; retention time: 12.6 min (major) and 22.2 min (minor).





4c, 116 mg, 90% yield, white solid, $[\alpha]_D^{25} = -4$ (c =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.52 (s, 1H), 7.35 (t, *J* = 7.8 Hz, 1H), 7.20 (d, *J* = 6.6 Hz, 1H), 7.09 – 4.04 (m, 2H), 3.14 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.97 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.61 (dd, *J* = 18.0, 3.6 Hz, 1H), 2.38 (s, 3H), 1.32 (s, 3H), 1.28 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 202.7, 176.9, 174.8, 139.3, 131.7, 129.6, 129.0, 127.1, 123.6, 48.5, 45.0, 31.8, 21.3, 20.3, 19.6; HRMS (ESI-Orbitrap) m/z: $[M+H]^+$ Calcd for C₁₅H₁₇NO₃ 260.1281, Found 261.1282.

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 218.0 nm; retention time: 11.6 min (major) and 13.5 min (minor).





4d, 116 mg, 90% yield, white solid, $[\alpha]_D^{25} = -6.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.52 (s, 1H), 7.27 (d, *J* = 7.2 Hz, 2H), 7.14 (d, *J* = 8.4 Hz, 2H), 3.14 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.97 (dd, *J* = 18.4, 9.6 Hz, 1H), 2.61 (dd, *J* = 18.4, 5.6 Hz, 1H), 2.37 (s, 3H), 1.32 (s, 3H), 1.28 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 220.3 nm; retention time: 14.3 min (major) and 17.1 min (minor).



442339

1512

2.04

17.133

2



4e, 93 mg, 68% yield, white solid, $[\alpha]_D^{25} = -4$ (c = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.52 (s, 1H), 7.14 – 7.06 (m, 2H), 6.98 – 6.91 (m, 1H), 3.22 – 3.14 (m, 1H), 3.06 – 2.94 (m, 1H), 2.72 – 7.60 (m, 1H), 2.34 (s, 3H), 2.16 – 2.04(m, 3H), 1.35 (d, J = 2.4 Hz, 3H), 1.30 (d, J = 1.8 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 203.1, 176.8, 174.7, 131.7, 129.1, 128.7, 126.6, 126.4, 51.3, 43.7, 31.1, 27.3, 15.1, 7.9; HRMS (ESI-Orbitrap) m/z: $[M+H]^+$ Calcd for C₁₆H₁₉NO₃ 274.1438, Found 274.1441.

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 212.0 nm; retention time: 12.6 min (major) and 22.0 min (minor).



(S)-2-(1-(4-methoxyphenyl)-2, 5-dioxopyrrolidin-3-yl)-2-methylpropanal³



4f, 135 mg, 98% yield, white solid, $[\alpha]_D^{25} = -5.0$ (c = 1.0, CHCl₃);¹H NMR (600 MHz, CDCl₃) δ 9.52 (s, 1H), 7.21 – 7.15 (m, 2H), 6.99 – 6.95 (m, 2H), 3.82 (s, 3H), 3.13 (dd, J = 9.6, 5.4 Hz, 1H), 2.96 (dd, J = 18.4, 9.6 Hz, 1H), 2.60 (dd, J = 18.6, 5.4 Hz, 1H), 1.32 (s, 3H), 1.28 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 229.8 nm; retention time: 29.5 min (major).





4g, 121 mg, 92 % yield, white solid, $[\alpha]_D^{25} = -7$ (c =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.49 (s, 1H), 7.43 – 7.36 (m, 2H), 7.34 – 7.32 (m, 1H), 7.23 – 7.20 (m, 1H), 3.11 (dd, *J* = 9.6, 5.4Hz, 1H), 2.98 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.62 (dd, *J* = 18.0, 5.4 Hz, 1H), 1.36 (s, 3H), 1.29 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 223.8 nm; retention time: 13.3 min (major) and 18.9 min (minor).





4h, 124 mg, 94% yield, white solid, $[\alpha]_D^{25} = -4$ (c =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 9.48 (s, 1H), 7.41 – 7.35 (m, 2H), 7.33 – 7.31 (m, 1H), 7.22 – 7.18 (m, 1H), 3.10 (dd, J = 9.6, 5.4 Hz, 1H), 2.97 (dd, J = 18.0, 9.6 Hz, 1H), 2.61 (dd, J = 18.0, 5.4 Hz, 1H), 1.35 (s, 3H), 1.28 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 223.8 nm; retention time: 13.3 min (major) and 18.9 min (minor).





4i, 125 mg, 95% yield, white solid, $[\alpha]_D^{25} = -1.0$ (c = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.49 (s, 1H), 7.28 – 7.24 (m, 2H), 7.17 – 7.12 (m, 2H), 3.11 (dd, J = 9.6, 5.4 Hz, 1H), 2.97 (dd, J = 18.6, 9.6 Hz, 1H), 2.61 (dd, J = 18.0, 5.4 Hz, 1H), 1.35 (s, 3H), 1.28 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 218.0 nm; retention time: 13.4 min (major) and 23.3 min (minor).





4j, 124 mg, 89% yield, white solid, $[\alpha]_D^{25} = -7$ (c =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.49 (s, 1H), 7.43 – 7.36 (m, 2H), 7.33 (t, *J* = 1.8 Hz, 1H), 7.23 – 7.19 (m, 1H), 3.12 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.98 (dd, *J* = 18.6, 9.6 Hz, 1H), 2.62 (dd, *J* = 18.6, 6.0 Hz, 1H), 1.37 (s, 3H), 1.30 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 212.2 nm; retention time: 12.2 min (major) and 15.2 min (minor).



(S)-2-(1-(4-chlorophenyl)-2,5-dioxopyrrolidin-3-yl)-2-methylpropanal⁶



4k, 125 mg, 90% yield, white solid, $[\alpha]_D^{25} = -3.0$ (*c* =1.0, CHCl₃);¹H NMR (600 MHz, CDCl₃) δ 9.49 (s, 1H), 7.49 – 7.40 (m, 2H), 7.28 – 7.22 (m, 2H), 3.11 (dd, *J* = 9.6, 6.0 Hz, 1H), 2.97 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.62 (dd, *J* = 18.6, 5.4 Hz, 1H), 1.36 (s, 3H), 1.29 (s, 3H). The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 225.1 nm; retention time: 14.4 min major) and 25.6 min minor).





41, 170 mg,98% yield, white solid, $[\alpha]_D^{25} = -10$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.55 (s, 1H), 7.48 – 7.45 (m, 2H), 3.37 (dd, *J* = 9.6, 6.0 Hz, 1H), 3.12 – 3.06 (m, 1H), 2.72 (dd, *J* = 18.6, 6.0 Hz, 1H), 1.34 (s, 3H), 1.30 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 202.3, 174.8, 172.7, 136.6, 135.1, 134.9, 128.8, 127.1, 48.1, 45.5, 31.9, 19.7, 19.5; HRMS (ESI-Orbitrap) m/z: [M+H]⁺ Calcd for C₁₄H₁₂Cl₃NO₃ 347.9956, Found 347.9958.

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 210.0 nm; retention time: 10.0 min major) and 12.0 in minor).





4m, 153 mg, 95% yield, white solid, $[\alpha]_D^{25} = -3.0$ (*c* =1.0, CHCl₃);¹H NMR (600 MHz, CDCl₃) δ 9.49 (s, 1H), 7.62 - 7.57 (m, 1H), 7.23 - 7.15 (m, 2H), 3.11 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.97 (dd, *J* = 18.6, 9.6 Hz, 1H), 2.62 (dd, *J* = 18.6, 6.0 Hz, 1H), 1.36 (s, 3H), 1.29 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 28.6 nm; retention time: 15.2 in major) and 25.7 min minor).





4n, 116 mg, 80% yield, white solid, $[\alpha]_D^{25} = -6.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.46 (s, 1H), 8.35 – 8.29 (m, 2H), 7.61 – 7.54 (m, 2H), 3.10 (dd, *J* = 9.6, 5.4 Hz, 1H), 3.00 (dd, *J* = 18.6, 9.6 Hz, 1H), 2.65 (dd, *J* = 18.6, 5.4 Hz, 1H), 1.41 (s, 3H), 1.30 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 212.1 nm; retention time: 29.0 min (major) and 35.6 min (minor).





40, 72 mg, 85% yield, white solid, $[\alpha]_D^{25} = +5.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.46 (s, 1H), 8.81 (s, 1H), 3.07 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.83 (dd, *J* = 18.4, 9.6 Hz, 1H), 2.48 (dd, *J* = 18.4, 5.4 Hz, 1H), 1.21 (d, *J* = 6.9 Hz, 6H).

The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 220.3 nm; retention time: 14.0 min minor) and 17.3 min major).



S22



4p, 82.4 mg, 90% yield, white solid, $[\alpha]_D^{25} = +$ 9.0 (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.51 (s, 1H), 3.04 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.99 (s, 3H), 2.82 (q, *J* = 9.0 Hz, 1H), 2.45 (dd, *J* = 18.0, 5.4 Hz, 1H), 1.22 (d, *J* = 6.6 Hz, 6H).

The ee was determined by HPLC analysis. CHIRALPAK AS-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 212.1 nm; retention time: 16.6 min minor) and 18.4 min major).





4q, 79 mg, 80% yield, white solid, $[\alpha]_D^{25} = +7$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.52 (s, 1H), 3.56 (q, *J* = 7.2 Hz, 2H), 3.02 (dd, *J* = 9.0, 5.4 Hz, 1H), 2.83 – 2.77 (m, 1H), 2.43 (dd, *J* = 18.0, 4.8 Hz, 1H), 1.21 (d, *J* = 6.6 Hz, 6H), 1.17 (t, *J* = 7.2 Hz, 3H). The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 210 nm; retention time: 12.1 min major) and 14.7 min minor).





4r, 119 mg, 92% yield, white solid, $[\alpha]_D^{25} = +14.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.48 (s, 1H), 7.37 – 7.34 (m, 2H), 7.31 – 7.28 (m, 3H), 4.65 (q, *J* = 13.8 Hz, 2H), 3.03 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.81 (q, *J* = 9.0 Hz, 1H), 2.45 (dd, *J* = 18.0, 5.4 Hz, 1H), 1.16 (d, *J* = 2.4 Hz, 6H).

The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 218.0 nm; retention time: 8.7 min major) and 18.8 min minor).



(S)-2-(1-cyclohexyl-2,5-dioxopyrrolidin-3-yl)-2-methylpropanal²



4s, 119 mg, 95% yield, white solid, $[\alpha]_D^{25} = +14(c = 1.0, \text{CHCl}_3)$; ¹H NMR (600 MHz, CDCl₃) δ 9.51 (s, 1H), 3.98 – 3.91 (m, 1H), 2.97 (dd, *J* = 9.6, 5.4 Hz, 1H), 2.74 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.37 (dd, *J* = 18.0, 4.8 Hz, 1H), 2.16 – 2.08 (m, 2H), 1.85 – 1.74 (m, 2H), 1.66 – 1.53 (m, 3H), 1.32 – 1.23 (m, 2H), 1.23 – 1.12 (m, 7H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=90:10; flow rate 1.0 mL/min; 290.2 nm; retention time: 10.7 min (minor) and 11.9 min (major).





(S)-2-(1-(tert-butyl)-2,5-dioxopyrrolidin-3-yl)-2-methylpropanal⁸



4t, 82 mg, 73% yield, white solid, $[\alpha]_D^{25} = +10$ (c = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.56 (s, 1H), 2.97 (dd, J = 10.2, 6.0 Hz, 1H), 2.69 (dd, J = 18.0, 9.6 Hz, 1H), 2.36 (dd, J = 18.0, 5.4 Hz, 1H), 1.56 (s, 9H), 1.14 (s, 6H).

The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 210.0 nm; retention time: 6.7 min major) and 8.1 min minor).





(S)-2-(2,5-dioxo-1-phenylpyrrolidin-3-yl)-2-ethylbutanal²



5a, 129 mg, 95% yield, white solid, $[\alpha]_D^{25} = +10.0$ (c =1.0, CHCl₃); 1H NMR (600 MHz, CDCl₃) δ 9.63 (s, 1H), 7.49 – 7.44 (m, 2H), 7.41 – 7.37 (m, 1H), 7.29 – 7.25 (m, 2H), 3.25 (dd, *J* = 9.6, 6.0 Hz, 1H), 2.97 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.69 (dd, *J* = 18.0, 5.4 Hz, 1H), 2.02 – 1.83 (m, 3H), 1.78 – 1.70 (m, 1H), 1.03 – 0.97 (m, 6H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 218.0 nm; retention time: 13.0 min (major) and 19.9 min (minor).



(S)-1-(2,5-dioxo-1-phenylpyrrolidin-3-yl)cyclopentanecarbaldehyde²



5b, 129 mg, 95% yield, white solid, $[\alpha]_D^{25} = +40.0 \ (c = +1.0, \text{CHCl}_3)$; ¹H NMR (600 MHz, CDCl₃) δ 9.39 (s, 1H), 7.49 – 7.46 (m, 2H), 7.40 – 7.37 (m, 1H), 7.34 – 7.29 (m, 2H), 3.05 – 2.94 (m, 2H), 2.58 (dd, J = 18.0, 5.4 Hz, 1H), 2.37 – 2.27 (m, 1H), 2.14 – 2.03 (m, 2H), 1.88 – 1.69 (m, 5H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 226.2 nm; retention time: 14.7 min (major) and 20.6 min (minor).



(S)-1-(2,5-dioxo-1-phenylpyrrolidin-3-yl)cyclohexanecarbaldehyde²



5c, 136 mg, 96% yield, white solid, $[\alpha]_D^{25} = +5.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.55 (s, 1H), 7.47 (t, *J* = 7.8 Hz, 2H), 7.40 – 7.38 (m, 1H), 7.31 – 7.27 (m, 2H), 3.22 (dd, *J* = 9.0, 6.0 Hz, 1H), 2.87 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.68 (dd, *J* = 18.2, 5.9 Hz, 1H), 1.99 – 1.85 (m, 3H), 1.67 – 1.49 (m, 6H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 220.3 nm; retention time: 8.8 min (major) and 12.9 min (minor).



(R)-2-((S)-2,5-dioxo-1-phenylpyrrolidin-3-yl)-2-methylbutanal⁹



5d, 136 mg, 96% yield, white solid, $[\alpha]_D^{25} = -14.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.62 (s, 1H), 7.48 – 7.44 (m, 2H), 7.41 – 7.39 (m, 1H), 7.29 – 7.25 (m, 2H), 3.35 (dd, *J* = 9.6, 60 Hz, 1H), 2.95 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.66 (dd, *J* = 18.0, 6.0 Hz, 1H), 1.80 – 1.70 (m, 2H), 1.19 (s, 3H), 0.95 (t, *J* = 7.8 Hz, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 220.3 nm; retention time: major diastereomer: 12.0 min (major), 16.5 min (minor); minor diastereomer: 14.9min (major), 20.6 min (minor).



(*R*)-2-((*S*)-2, 5-dioxo-1-phenylpyrrolidin-3-yl)-2-methylpentanal²



5e, 136 mg, 96% yield, white solid, $[\alpha]_D^{25} = -5.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.62 (s, 1H), 7.46 (t, *J* = 7.8 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 1H), 7.29 – 7.25 (m, 2H), 3.34 (dd, *J* = 9.6, 60. Hz, 1H), 2.94 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.66 (dd, *J* = 18.0, 6.0 Hz, 1H), 1.70 – 1.57 (m, 2H), 1.45 – 1.36 (m, 1H), 1.31 – 1.22 (m, 1H), 1.19 (s, 3H), 0.95 (t, *J* = 7.2 Hz, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 220.3 nm; retention time: major diastereomer: 26.4 min (major), 30.4 min (minor); minor diastereomer: 17.1 min (major), 18.9 min (minor).



(*R*)-2-((*S*)-2,5-dioxo-1-phenylpyrrolidin-3-yl)propanal³



5f, 101 mg, 88% yield, white solid, $[\alpha]_D^{25} = -20.0$ (*c* =1.0, CHCl₃);¹H NMR (600 MHz, CDCl₃) δ 9.63 (s, 1H), 7.48 (t, *J* = 7.8 Hz, 2H), 7.40 (t, *J* = 7.2 Hz, 1H), 7.34 - 7.28 (m, 2H), 3.31-3.26 (m, 1H), 3.12 - 3.06 (m, 1H), 2.93 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.60 (dd, *J* = 18.0, 5.6 Hz, 1H), 1.40 (d, *J* = 7.8 Hz, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 0.5 mL/min; 218.0 nm; retention time: major diastereomer: 20.8 min (major), 18.7 min (minor); minor diastereomer: 26.4 min (major), 30.64 min (minor).



(*R*)-2-((*S*)-2, 5-dioxo-1-phenylpyrrolidin-3-yl) pentanal¹⁰



5g, 124 mg, 96% yield, white solid, $[\alpha]_D^{25} = -15.0$ (*c* =1.0, CHCl₃);¹H NMR (600 MHz, CDCl₃) δ 9.76 (s, 1H), 7.50 - 7.46 (m, 2H), 7.42 - 7.38 (m, 1H), 7.33 - 7.28 (m, 2H), 3.37 - 3.32 (m, 1H), 3.04 - 3.01 (m, 1H), 2.59 (d, *J* = 5.4 Hz, 1H), 2.56 (d, *J* = 6.0 Hz, 1H), 1.94 - 1.85 (m, 1H), 1.69 - 1.60 (m, 1H), 1.58 - 1.52 (m, 2H), 1.01 (t, *J* = 7.2 Hz, 3H).

The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=90:10; flow rate 0.5 mL/min; 210.0 nm; retention time: major diastereomer: 26.2 min (major); minor diastereomer: 31.9 min (major)





5h, 109 mg, 84% yield, white solid, $[\alpha]_D^{25} = -16.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.78 (s, 1H), 7.50 – 7.45 (m, 2H), 7.41 – 7.36 (m, 1H), 7.31 – 7.28 (m, 2H), 3.17 (dd, *J* = 7.2, 3.7 Hz, 1H), 3.11 – 2.99 (m, 1H), 2.87 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.73 (dd, *J* = 18.0, 6.0 Hz, 1H), 2.36 – 2.28 (m, 1H), 1.26 (d, *J* = 6.6 Hz, 3H), 1.10 (d, *J* = 6.6 Hz, 3H). The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=80:20; flow rate 0.5 mL/min; 218.0 nm; retention time: major diastereomer: 27.3 min (major); minor diastereomer: 34.9 min (major).



S35
(R)-2-((S)-2, 5-dioxo-1-phenylpyrrolidin-3-yl)decanal³



5i, 99 mg, 60% yield, white solid, $[\alpha]_D^{25} = -16(c = 1.0, CHCl_3)$; ¹H NMR (600 MHz, CDCl₃) δ 9.64 (s, 1H), 7.51 – 7.45 (m, 2H), 7.42 – 7.38 (m, 1H), 7.36 – 7.27 (m, 2H), 3.25 - 3.23 (m 1H), 3.10 – 3.07 (m, 1H), 2.87 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.63 - 2.59 (m, 1H), 1.52 – 1.51 (m, 3H), 1.33 – 1.26 (m, 11H), 1.37-1.35 (m, 1H), 1.33 – 1.22 (m, 9H), 0.91 – 0.89 (m, 3H).

The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=90:10; flow rate 0.5 mL/min; 218.0 nm; retention time: major diastereomer: 29.2 min (major), 52.9 min (minor); minor diastereomer: 32.7 min (minor), 34.6 min (major).



(R)-2-((S)-2,5-dioxo-1-phenylpyrrolidin-3-yl)-3-phenylpropanal¹¹



5j, 115 mg, 75% yield, white solid, $[\alpha]_D^{25} = +25$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.72 (s, 1H), 7.49 – 7.44 (m, 2H), 7.41 – 7.33 (m, 3H), 7.33 – 7.23 (m, 5H), 3.65 – 3.59 (m, 1H), 3.40 (dd, *J* = 13.8, 5.4 Hz, 1H), 2.97 – 2.85 (m, 1H), 2.85 – 2.73(m, 2H), 2.65 – 2.58 (m, 1H).

The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=85:15; flow rate 1.0 mL/min; 218.0 nm;



(*R*)-2-((*S*)-1-(4-fluorophenyl)-2,5-dioxopyrrolidin-3-yl) hexanal



5k, 108 mg, 74% yield, white solid, $[\alpha]_D^{25} = -33(c = 1.0, CHCl_3)$;¹H NMR (600 MHz, CDCl₃) δ 9.62 (s, 1H), 7.34 – 7.28 (m, 2H), 7.19 – 7.13 (m, 2H), 3.29 – 3.20 (m, 1H), 3.10 – 3.02 (m, 1H), 2.86 (dd, J = 18.0, 9.6 Hz, 1H), 2.60 (dd, J = 18.0, 6.0 Hz, 1H), 1.54 – 1.49 (m, 6H), 0.97 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 201.4, 177.3, 174.7, 134.5, 130.3, 129.4, 127.7, 52.9, 38.5, 32.6, 29.8, 26.1, 22.6,

13.8; HRMS (ESI-Orbitrap) m/z: $[M+H]^+$ Calcd for $C_{16}H_{18}FNO_3$ 292.1343, Found 292.1342. The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 218.0 nm; retention time: minor diastereomer: 18.9 min (minor), 25.0 min (major); major diastereomer: 20.8 min (minor), 30.9 min (major).



(R)-2-((S)-1-(4-bromophenyl)-2, 5-dioxopyrrolidin-3-yl)hexanal



4

51, 125 mg, 71% yield, white solid, $[\alpha]_D^{25} = -80.0(c = 1.0, CHCl_3)$; ¹H NMR (600 MHz, CDCl₃) δ 9.74 (s, 1H), 7.63 – 7.57 (m, 2H), 7.23 – 7.18 (m, 2H), 3.33 – 3.28(m, 1H), 3.05 – 2.95 (m, 2H), 2.56 (dd, *J* = 18.6, 6.0 Hz, 1H), 1.96 – 1.89 (m, 1H), 1.73 – 1.65 (m, 1H), 1.53 – 1.46 (m, 2H), 1.45 – 1.36(m, 2H), 0.94 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 201.4, 177.3, 174.7, 132.3, 130.9,

128.0, 52.9, 38.5, 32.6, 29.8, 26.1, 22.6, 13.8; HRMS (ESI-Orbitrap) m/z: $[M+H]^+$ Calcd for $C_{16}H_{18}BrNO_3$ 352.0543, Found 352.0545.

The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 218.0 nm; retention time: minor diastereomer: 24.2 min (minor),31.9 min (major); major diastereomer: 26.7 min (minor), 37.3 min (major).



104708543

922576

65.45

37.254

(S)-2-(4,4-dimethyl-2-oxotetrahydrofuran-3-yl)-N-phenylacetamide²



7, 94 mg, 76% yield, white solid, $[\alpha]_D^{25} = +9.0$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 8.64 (s, 1H), 7.53 (d, *J* = 7.8 Hz, 2H), 7.32 – 7.26 (m, 2H), 7.09 (t, *J* = 7.2 Hz, 1H), 4.01 (d, *J* = 9.0 Hz, 1H), 3.97 (d, *J* = 9.0 Hz, 1H), 2.92 (dd, *J* = 8.4, 4.2 Hz, 1H), 2.68 (dd, *J* = 15.0, 8.4 Hz, 1H), 2.38 (dd, *J* = 15.0, 4.2 Hz, 1H), 1.19 (s, 3H), 1.01 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 220.0 nm; retention time: 10.7 min (minor) and 17.8 min(major).





8, 104 mg, 62% yield, white solid, $[\alpha]_D^{25} = -40$ (*c* =1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 10.43 (s, 1H), 7.62 (d, *J* = 8.4 Hz, 2H), 7.35 – 7.20 (m, 7H), 7.05 (t, *J* = 7.2 Hz, 1H), 4.58 (d, *J* = 15.0 Hz, 1H), 4.37 (d, *J* = 15.0 Hz, 1H), 3.09 (d, *J* = 9.6 Hz, 1H), 2.85 (d, *J* = 9.6 Hz, 1H), 2.78 – 2.66 (m, 2H), 2.32 – 2.29 (m, 1H), 1.13 (s, 3H), 0.87 (s, 3H).

The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=75:25; flow rate 1.0 mL/min; 218.0 nm; retention time: 6.6 min (minor) and 8.8 min(major).



8. Copies of NMR spectra



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -j f1 (ppm)





-1.79.58 ~ 174.31 ~ 170.43 168.86

-61.08 55.12 55.15

1c: NH₂-Chg-DPro-Gly-Leu-OH ¹³C NMR, 151 MHz, D₂O



























-61.26 -63.46 -53.46 -53.48 47.56 40.49 -53.87 -53.85 -53.

NH2

1h: NH₂-Ala-DPro-Gly-Leu-OH ¹³C NMR, 151 MHz, D₂O


















































9. HPLC analytical of the tetrapeptides 1d and 1e

Tetrapeptides **1d** and **1e** have passed HPLC purity analysis, and used C18 reversed phase column gradient elution, acetonitrile/ (0.1% TFA in water) = 10: 90 to 90: 10, 1mL/min. λ_{max} = 220nm.



No	name	Retention	Area(µV*s)	Height (μV)	% Area
1	N.A.	9.967	699.50797	82.804	5.034
2	N.A.	13.380	45.30635	6.571	0.326
3	N.A.	13.930	12.72706	2.292	0.092
4	N.A.	14.200	13138.91413	1224.438	94.549



No	name	Retention	Area(µV*s)	Height (μV)	% Area
1	N.A.	10.502	111.03011	15.070	3.000
2	N.A.	13.038	3590.04890	418.667	97.000



10. Images of the HR-MS of the new compounds

S76







S77













11. Reference

- 1. Z-H. Du, B-X. Tao, M. Yuan, W-J. Qin, Y-Li. Xu, P. Wang and C-S. Da, *Org. Lett.*, 2020, **22**, 4444-4450.
- 2. C. G. Kokotos, Org. Lett., 2013, 15, 2406-2409.
- 3. F. Xue, L. Liu, S. Zhang, W. Duan and W. Wang, *Chem. Eur. J.* 2010, **16**, 7979 7982.
- 4. W. Yang, K. Z. Jiang, X. Lu, H. M. Yang, L. Li, Y. Lu and L. W. Xu, *Chem. Asian J.* 2013, **8**, 1182-1190.
- 5. J.-F. Bai, L. Peng, L.-l. Wang, L.-X. Wang and X.-Y. Xu, *Tetrahedron*, 2010, **66**, 8928-8932.
- 6. A. Avila, R. Chinchilla, E. Gómez-Bengoa and C. Nájera, *Eur. J. Org. Chem.*, 2013, 23, 5085-5092.
- 7. F. Yu, Z. Jin, H. Huang, T. Ye, X. Liang and J. Ye, Org. Biomol. chem., 2010, 8, 4767-4674.
- 8. V. Kozma, F. Fül öp and G. Szőllősic, Adv. Synth. Catal., 2020, 362, 2444–2458.
- 9. Z.-w. Ma, X.-f. Liu, J.-t. Liu, Z.-j. Liu and J.-c. Tao, *Tetrahedron Lett.*, 2017, **58**, 4487-4490.
- 10. Z.-w. Ma, Y.-x. Liu, P.-l. Li, H. Ren, Y. Zhu and J.-c. Tao, *Tetrahedron: Asymmetry*, 2011, **22**, 1740-1748.
- 11. G. L. Zhao, Y. Xu, H. Sunden, L. Eriksson, M. Sayah and A. Cordova, *Chem. Commun.*, 2007, 734-735.