Electronic Supplementary Information

Total Synthesis of Thioamycolamide A via a Biomimetic Route

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28 pages

Table of contentsS2General remarksS2NMR data of natural and synthetic 1 in DMSO-d6S3Cytotoxicity assay of natural 1 and synthetic 1 against HT1080 and HeLa S3S4Synthetic procedures and compound characterizationsS5–S9Spectral dataS10–S28

General remarks

¹H and ¹³C NMR spectra were recorded on Bruker AVANCE DMX 600 NMR (600 MHz for ¹H NMR and 150 MHz for ¹³C NMR), JEOL ECA500 (500 MHz for ¹H NMR and 125 MHz for ¹³C NMR), or JEOL ECZ600 (600 MHz for ¹H NMR and 150 MHz for ¹³C NMR) spectrometer. Chemical shifts are denoted in δ (ppm) relative to residual solvent peaks as internal standard (CDCl₃, ¹H δ 7.25, ¹³C δ 77.2, CD₃OD, ¹H δ 3.31, ¹³C δ 49.0, DMSO-*d*₆, ¹H δ 2.50, ¹³C δ 39.5). ESI-MS and LC-MS experiments were recorded on a Shimadzu LCMS-IT-TOF. Optical rotations were recorded on JASCO P-2200 polarimeter. CD spectra were measured on JASCO J-715 circular dichroism spectrometer with a 1 mm path length cell. High performance liquid chromatography (HPLC) experiments were performed with SHIMADZU HPLC system equipped with LC-20AD intelligent pump. All reactions sensitive to air and/or moisture were used as supplied unless otherwise stated. Analytical thin-layer chromatography (TLC) was performed using E. Merck Silica gel 60 F₂₅₄ pre-coated plates. Silica gel column chromatography was performed using 40-50 µm Silica Gel 60N (Kanto Chemical Co., Inc.).

nos	natural 1		nos	synthetic 1	
pos	δ_C , type	$\delta_{\rm H} (J \text{ in Hz})$	Pop	$\delta_{\rm C}$, type	$\delta_{\rm H} (J \text{ in Hz})$
1	34.9, CH ₂	2.79, dd (13.0, 8.8) 3.10, dd (13.0, 4.3)	1	34.9, CH ₂	2.79, dd (13.1, 8.8) 3.10, dd (13.0, 4.3)
2	50.6, CH	3.39, m	2	50.6, CH	3.39, m
3	61.9, CH ₂	3.21, m 3.51, m	3	61.9, CH ₂	3.22, m 3.51, d (10.1)
4	170.2, C		4	170.3, C	
5	77.7, CH	5.08, t (6.7)	5	77.7, CH	5.08, m
6	36.8, CH ₂	3.56, d (6.7)	6	36.8, CH ₂	3.56, m
7	177.7, C		7	177.7, C	
8	54.3, CH	4.54, m	8	54.3, CH	4.54, m
9	37.6, CH ₂	2.96, dd (13.9, 9.9)	9	37.6, CH ₂	2.95, dd (13.9, 10.0)
		3.03, dd (13.9, 5.0)			3.03, dd (13.9, 5.0)
10	137.4, C		10	137.4, C	
11/15	129.1, CH	7.32, t (7.9)	11/15	129.1, CH	7.32, t (7.8)
12/14	128.3, CH	7.29, t (7.6)	12/14	128.3, CH	7.30, t (7.6)
13	126.6, CH	7.22, t (7.1)	13	126.7, CH	7.22, m
16	171.6, C		16	171.6, C	
17	42.5, CH ₂	2.39, d(8.3)	17	42.5, CH ₂	2.39, m
18	45.7, CH	2.90, m	18	45.8, CH	2.90, m
19	36.8, CH ₂	1.42, m	19	36.9, CH ₂	1.42, m
		1.54, m			1.55, m
20	28.8, CH ₂	1.35, m	20	28.8, CH ₂	1.35, m
		1.45, m			1.45, m
21	21.8, CH ₂	1.28, m	21	21.9, CH ₂	1.28, m
22	14.0, CH ₃	0.87, t (7.3)	22	14.0, CH ₃	0.87, t (7.3)
2-NH		6.77, d (5.7)	2-NH		6.77, d (5.7)
3-ОН		4.82, s	3-OH		4.82, s
8-NH		8.85, d (6.7)	8-NH		8.84, d (6.8)

<u>NMR data of natural 1 and synthetic 1 in DMSO- d_6 </u>

Table S1. ¹H NMR data (600 MHz) and ¹³C NMR data (150 MHz) of natural 1 and synthetic 1 in DMSO-*d*₆.



Cytotoxicity assay of natural 1 and synthetic 1 against HT1080 and HeLa S3 cells

Cytotoxicity of compounds against HT1080 and HeLa S3 cell lines was evaluated by a WST-8 colorimetric assay (Cell Counting Kit-8, Dojindo). Cells were cultured in 96-well plates (3000 cells/well) for 24 h followed by exposure to natural **1** and synthetic **1** for 72 h, and then the viability was assessed by WST-8. The absorbance was measured at 450 nm using an iMark microplate reader (BIO-RAD). Adriamycin was evaluated as a positive control. IC_{50} values are shown as the mean \pm SD (n = 4).

SampleCytotoxic activity (μ M)HT 1080Hela S3natural 19.60 ± 1.0315.47 ± 3.54synthetic 111.61 ± 1.4414.74 ± 5.16Adriamycin0.30 ± 0.050.37 ± 0.06

 Table S2. Cytotoxic activities (IC₅₀) of natural 1 and synthetic 1.



Fig. S1 Cytotoxicity assay against HT1080 cell.



Fig. S2 Cytotoxicity assay against HeLa-S3 cell.

Synthetic procedures and compound characterizations



To a solution of Boc-L-Phe-OH (**21**) (1.00 g, 3.77 mmol) and H-D-Ser-OMe·HCl (**22**) (645 mg, 4.15 mmol) in DMF (15 mL) were added *N*-methylmorpholine (0.47 mL, 4.17 mmol), Oxyma (589 mg, 4.14 mmol), and EDCI·HCl (795 mg, 4.15 mmol) at 0 °C. After being stirred at room temperature overnight, saturated aqueous NH₄Cl (20 mL) was added to the reaction mixture. The resulting solution was extracted with EtOAc (40 mL × 2). The combined organic layer was washed with saturated aqueous NaHCO₃ (50 mL) and brine (50 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane = 1:1) to afford dipeptide **23** (1.31 g, 95%) as a white foam: $[a]^{20}_{D} = -6.5$ (*c* 0.14, MeOH); ¹H NMR (500 MHz, CDCl₃) δ 7.32 (t, *J* = 7.3 Hz, 2H), 7.25 (m, 3H), 6.72 (d, *J* = 7.3 Hz, 1H), 5.12 (d, *J* = 6.5 Hz, 1H), 4.57 (m, 1H), 4.38 (d, *J* = 6.3 Hz, 1H), 3.78 (m, 1H), 3.74 (s, 3H), 3.06 (d, *J* = 7.2 Hz, 2H), 1.40 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.3, 170.7, 155.8, 136.7, 129.4, 128.9, 127.3, 80.8, 62.7, 56.3, 54.8, 52.9, 38.8, 28.4; HRMS (ESI) calcd for C₁₈H₂₆N₂O₆Na⁺ [M+H]⁺ 389.1683, found 389.1703.



To a solution of **23** (760 mg, 2.07 mmol) in DMF (15 mL) were added TBSCl (469 mg, 3.11 mmol) and imidazole (424 mg, 6.23 mmol) at 0 °C. After being stirred at room temperature for 30 min, saturated aqueous NH₄Cl (20 mL) was added to the reaction mixture. The mixture was extracted with EtOAc (20 mL × 2). The combined organic layer was washed with brine (40 mL × 2), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane = 1:4) to afford **24** (991 mg, 99%) as a white foam: $[\alpha]^{20}{}_{\rm D} = -17.1$ (*c* 0.63, MeOH); ¹H NMR (500 MHz, CDCl₃) δ 7.21 (m, 2H), 7.14 (m, 3H), 6.82 (d, *J* = 7.4 Hz, 1H), 5.23 (d, *J* = 7.7 Hz, 1H), 4.56 (d, *J* = 7.7 Hz, 1H), 4.44 (d, *J* = 7.2 Hz, 1H), 3.93 (d, *J* = 9.7 Hz, 1H), 3.63 (s, 3H), 3.57 (d, *J* = 7.7 Hz, 1H), 3.10 (dd, *J* = 13.8, 6.5 Hz, 1H), 2.96 (dd, *J* = 13.4, 7.2 Hz, 1H), 1.32 (s, 9H), 0.78 (s, 9H), -0.06 (s, 3H), -0.07 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.0, 170.5, 155.3, 136.8, 129.2, 128.4, 126.7, 79.8, 63.2, 55.5, 54.0, 52.2, 38.3, 28.1, 25.6, 18.0, -5.6, -5.9; HRMS (ESI) calcd for C₂₄H₄₀N₂O₆SiNa⁺ [M+H]⁺ 503.2548, found 503.2541.



To a solution of **24** (984 mg, 2.05 mmol) in THF (15 mL) was added Lawesson's reagent (498 mg, 1.23 mmol) at room temperature. After being stirred at 70 °C for 1 h in a sealed tube, the reaction mixture was cooled to room temperature and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane = 1:10) to afford practically pure **25** (1.02 g, Figures S11, S12, and S43), which was used in the next reaction without further purification.

To a solution of the above **25** (1.02 g, ca 2.05 mmol) in CH₂Cl₂ (20 mL) was added 2,6-lutidine (1.40 mL, 12.1 mmol) at room temperature, followed by dropwise addition of TMSOTf (1.46 mL, 8.08 mmol). The mixture was stirred at room temperature for 8 h, then quenched by addition of MeOH (5 mL). The reaction mixture was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (CH₂Cl₂ /MeOH/Et₃N = 200:1:0.5) to afford **26** as a light yellow oil (763 mg, 94% for 2 steps from **24**): $[\alpha]^{20}_{D} = -16.1 (c 0.33, MeOH); {}^{1}_{H} NMR (500 MHz, CDCl₃) <math>\delta$ 7.30 (m, 2H), 7.22 (m, 3H), 5.25 (s, 1H), 4.11 (dd, *J* = 10.3, 2.6 Hz, 1H), 4.06 (dd, *J* = 10.1, 3.7 Hz, 1H), 3.97 (dd, *J* = 10.3, 3.1 Hz, 1H), 3.75 (s, 3H), 3.60 (dd, *J* = 13.8, 3.7 Hz, 1H), 2.60 (dd, *J* = 13.8, 10.1 Hz, 1H), 0.83 (s, 9H), 0.00 (s, 6H); {}^{13}_{C} NMR (125 MHz, CDCl₃) δ 205.6, 169.7, 138.1, 129.2, 128.7, 126.9, 63.7, 62.3, 58.9, 52.4, 43.6, 25.6, 18.0, -5.6, -5.7; HRMS (ESI) calcd for C₁₉H₃₃N₂O₃SSi⁺ [M + H]⁺ 397.1976, found 397.1990.



To a solution of **26** (148 mg, 0.370 mmol) and 2-heptenoic acid (7) (52.5 mg, 0.410 mmol) in DMF (3 mL) were added Oxyma (58.3 mg, 0.410 mmol) and EDCI·HCl (78.6 mg, 0.410 mmol) at 0 °C. After being stirred at room temperature overnight, saturated aqueous NH₄Cl (10 mL) was added to the reaction mixture. The mixture was extracted with EtOAc (10 mL × 2). The combined organic layer was washed with saturated aqueous NHHCO₃ (20 mL) and brine (20 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane = 1:5) to afford **27** (171 mg, 90%) as a colorless oil: $[\alpha]^{20}_{D} = -29.5$ (*c* 0.12, MeOH); ¹H NMR (500 MHz, CDCl₃) δ 8.15 (d, *J* = 7.7 Hz, 1H), 7.23 (m, 2H), 7.18 (m, 3H), 6.80 (dt, *J* = 15.1, 6.9 Hz, 1H), 6.64 (d, *J* = 7.9 Hz, 1H), 5.77 (d, *J* = 15.3 Hz, 1H), 5.03 (m, 2H), 3.90 (dd, *J* = 10.2, 2.6 Hz, 1H), 3.67 (s, 3H), 3.43 (dd, *J* = 10.2, 3.4 Hz, 1H), 3.25 (dd, *J* = 13.4, 6.2 Hz, 1H), 3.08 (dd, *J* = 13.4, 8.1 Hz, 1H), 2.14 (td, *J* = 8.1, 1.2 Hz, 2H), 1.39 (m, 2H), 1.29 (m, 2H), 0.87 (t, *J* = 7.3 Hz, 3H), 0.79 (s, 9H), -0.05 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 203.0, 169.3, 165.3, 145.8, 136.7, 129.3, 128.6, 127.0, 123.2, 62.2, 60.7, 59.4, 52.5,

42.1, 31.8, 30.3, 25.6, 22.3, 18.1, 13.9, -5.5, -5.8; HRMS (ESI) calcd for $C_{26}H_{43}N_2O_4SSi^+$ [M + H]⁺ 507.2707, found 507.2724.



To a solution of **27** (169 mg, 0.33 mmol) in THF (3 mL) were added AcOH (22.6 μ L, 0.4 mmol) and TBAF (1 M in THF, 0.4 mL, 0.4 mmol) at 0 °C. After being stirred at room temperature for 3 h, the reaction mixture was diluted with EtOAc (10 mL), and then washed with saturated aqueous NH₄Cl (10 mL) and brine (10 mL). The combined organic layer was dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane = 1:2) to afford **28** (122 mg, 93%) as a white foam: $[\alpha]^{20}_{D} = -0.9 (c 0.30, MeOH)$; ¹H NMR (400 MHz, CDCl₃) δ 8.95 (d, J = 7.3 Hz, 1H), 7.20 (m, 5H), 6.80 (d, J = 8.3 Hz, 1H), 6.73 (dt, J = 15.3, 6.9 Hz, 1H), 5.77 (dt, J = 15.3, 1.3 Hz, 1H), 5.18 (dt, J = 8.4, 6.7 Hz, 1H), 5.00 (m, 1H), 3.82 (dd, J = 11.8, 3.2 Hz, 1H), 3.66 (s, 3H), 3.56 (dd, J = 11.9, 3.1 Hz, 1H), 3.21 (dd, J = 13.1, 6.4 Hz, 1H), 3.06 (dd, J = 13.1, 8.6 Hz, 1H), 2.12 (m, 2H), 1.37 (m, 2H), 1.28 (m, 2H), 0.87 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 203.4, 169.7, 165.8, 146.4, 136.6, 129.6, 128.5, 127.0, 123.0, 61.5, 60.4, 60.1, 52.8, 42.5, 31.8, 30.2, 22.3, 13.9; HRMS (ESI) calcd for C₂₀H₂₈N₂O₄SNa⁺ [M + Na]⁺415.1662, found 415.1661.



To a solution of **28** (115 mg, 0.29 mmol) in CH₂Cl₂ (3 mL) was added DAST (100 μ L, 0.80 mmol) at -78 °C. After being stirred at -78 °C for 1 h, the reaction mixture was poured to ice-water (10 mL) and extracted with EtOAc (10 mL × 2). The combined organic layer was washed with brine (10 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure to afford practically pure **29** (107 mg, Figures S19, S20, and S47) as a white powder, which was used in the next reaction without further purification to avoid the epimerization.

To a solution of the above **29** (27.5 mg, ca 73 μ mol) in 1,2-dichloroethane (1 mL) was added trimethyltin hydroxide (40 mg, 0.22 mmol) at room temperature. The reaction mixture was heated at 80 °C for 1 h, and then concentrated under reduced pressure. The residue was dissolved in EtOAc (10 mL), washed with 0.01 M KHSO₄ aqueous solution (5 mL × 5) and brine (10 mL), dried over MgSO₄, filtered and concentrated to afford practically pure **30** (26.4 mg, Figures S21, S22, and S30) as a white foam, which was used in the next reaction without further purification to avoid the epimerization.



To a solution of **31** (2.98 g, 6.8 mmol) in THF (40 mL) were added *N*-methylmorpholine (1.64 mL, 14.9 mmol) and isobutyl chloroformate (1.95 mL, 14.8 mmol) at -10 °C. After being stirred for 15 min, NaBH₄ (1022 mg, 27.0 mmol) was added to the solution. After being stirred for 15 min at -10 °C, saturated aqueous NH₄Cl (40 mL) was added to the reaction mixture. The mixture was extracted with EtOAc (50 mL × 2). The combined organic layer was washed with brine (80 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/hexane = 2:1) to afford **32** (2.27 g, 81%) as a white powder: $[\alpha]^{20}_{D}$ = 85.1 (*c* 0.77, MeOH); ¹H NMR (500 MHz, CD₃OD) δ 6.55 (d, *J* = 8.3 Hz, 1H), 3.85 (m, 1H), 3.62 (dd, *J* = 11.1, 4.9 Hz, 1H), 3.57 (dd, *J* = 11.1, 5.5 Hz, 1H), 2.97 (dd, *J* = 13.7, 5.7 Hz, 1H), 2.82 (dd, *J* = 13.7, 7.8 Hz, 1H), 1.45 (s, 9H); ¹³C NMR (125 MHz, CD₃OD) δ 158.0, 80.2, 63.9, 53.4, 41.5, 28.8; HRMS (ESI) calcd for C₁₆H₃₂N₂O₆S₂Na⁺ [M + Na]⁺ 435.1594, found 435.1587.



To **32** (283 mg, 0.69 mmol) was added 2 M HCl in ethanol (3 mL) at room temperature. After being stirred for 1 h, the reaction mixture was concentrated to afford **33** (196 mg, 100%) as a white foam, which was used in the next reaction without further purification: $[\alpha]_{D}^{20} = -100$ (*c* 0.29, MeOH); ¹H NMR (500 MHz, CD₃OD) δ 3.88 (dd, *J* = 11.8, 3.7 Hz, 1H), 3.76 (dd, *J* = 11.8, 5.4 Hz, 1H), 3.60 (m, 1H), 3.12 (dd, *J* = 14.4, 6.6 Hz, 1H), 3.05 (dd, *J* = 14.4, 7.1 Hz, 1H); ¹³C NMR (125 MHz, CD₃OD) δ 61.2, 53.3, 37.0; HRMS (ESI) calcd for C₆H₁₇N₂O₂S₂⁺ [M + H]⁺ 213.0726, found 213.0730.



To a solution of 33 (8.6 mg, 30.2 µmol) and 30 (24 mg, 67 µmol) in DMF (0.5 mL) were added *i*-Pr₂NEt (12 µL, 67

μmol), Oxyma (9.5 mg, 67 μmol), and EDCI·HCl (13 mg, 67 μmol) at 0 °C. After being stirred at 0 °C for 2 h and at room temperature for 2 h, saturated aqueous NH₄Cl (5 mL) was added to the reaction mixture. The resulting solution was extracted with EtOAc (5 mL × 2). The combined organic layer was washed with saturated aqueous NaHCO₃ (10 mL) and brine (10 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (CHCl₃/MeOH = 50:1) to afford **34** (25.6 mg, 94%) as a white foam: $[\alpha]^{20}{}_{\rm D}$ = -86.3 (*c* 0.50, MeOH); ¹H NMR (500 MHz, CD₃OD) δ 7.26 (m, 4H), 7.20 (m, 1H), 6.75 (dt, *J* = 15.3, 7.0 Hz, 1H), 5.91 (dt, *J* = 15.4, 1.4 Hz, 1H), 5.09 (t, *J* = 8.9 Hz, 1H), 4.95 (dd, *J* = 9.3, 5.6 Hz, 1H), 4.22 (m, 1H), 3.68 (dd, *J* = 11.4, 4.8 Hz, 2H), 3.59 (m, 2H), 3.52 (dd, *J* = 11.2, 8.2 Hz, 1H), 3.31 (m, 1H), 3.06 (dt, *J* = 13.9, 7.7 Hz, 1H), 2.92 (dd, *J* = 13.8, 8.1 Hz, 1H), 2.18 (m, 2H), 1.43 (m, 2H), 1.34 (m, 2H), 0.92 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (125 MHz, CD₃OD) δ 178.6, 173.1, 168.7, 147.0, 138.5, 130.3, 129.5, 127.8, 124.0, 80.0, 63.4, 55.2, 52.3, 40.6, 39.7, 36.6, 32.8, 23.3, 14.2; HRMS (ESI) calcd for C₄₄H₆₁N₆O₆S₄⁺ [M + H]⁺ 897.3530, found 897.3527.



To a solution of **34** (23.0 mg, 25.6 µmol) in MeOH/aqueous buffer (pH 9, Na₂CO₃/NaHCO₃) (10 mL/10 mL) was added TCEP·HCl (22 mg, 76.8 µmol) at room temperature. The reaction mixture was stirred for 30 min and then diluted with pH 9 Na₂CO₃/NaHCO₃ buffer (20 mL). The reaction mixture was allowed to room temperature for an additional 2 days. The mixture was extracted with EtOAc (50 mL × 2). The combined organic layer was washed with brine (80 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by reversed-phase HPLC [column: PEGASIL ODS SP100 20 × 250 mm; eluent: MeOH/H₂O = 70/30, 8.0 mL/min; detection: UV 210 nm] to afford 1 (15.4 mg, t_R 31.0 min, 67%) as a white foam: $[a]^{20}_{D} = -70.4$ (*c* 0.39, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 8.84 (d, *J* = 6.8 Hz, 1H), 7.31 (m, 4H), 7.22 (t, *J* = 7.1 Hz, 1H), 6.77 (d, *J* = 5.7 Hz, 1H), 5.08 (dd, *J* = 7.1, 6.2 Hz, 1H), 4.82 (s, 1H), 4.54 (td, *J* = 9.3, 5.6 Hz, 1H), 3.56 (d, *J* = 6.8 Hz, 2H), 3.51 (d, *J* = 10.1 Hz, 1H), 3.39 (m, 1H), 3.21 (t, *J* = 7.1 Hz, 1H), 3.10 (dd, *J* = 13.0, 4.3 Hz, 1H), 3.03 (dd, *J* = 13.9, 5.1 Hz, 1H), 2.95 (dd, *J* = 13.9, 9.9 Hz, 1H), 2.90 (m, 1H), 2.79 (dd, *J* = 13.1, 8.8 Hz, 1H), 2.39 (m, 2H), 1.55 (m, 1H), 1.41 (m, 3H), 1.27 (m, 2H), 0.87 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (150 MHz, DMSO-*d*₆) δ 177.7, 171.6, 170.3, 137.4, 129.1, 128.3, 126.7, 77.7, 61.9, 54.3, 50.6, 45.8, 42.5, 37.6, 36.9, 36.8, 34.9, 28.8, 14.0; HRMS (ESI) calcd for C₂₂H₃₂N₃O₃O₃S⁺ [M + H]⁺ 450.1880, found 450.1892.

entry	solvent	yield ^b
1	0.1 M PBS buffer, pH 8/MeOH ^a (= 4:1)	5%
2	MeOH, 5% TEA	<1%
3	0.1 M NaHCO ₃ solution, pH 8.3/MeOH (= 4:1)	45%
4	0.1 M Na ₂ CO ₃ /NaHCO ₃ buffer, pH 9/MeOH (= 4:1)	71% (67%) ^c
5	0.1 M Na ₂ CO ₃ /NaHCO ₃ buffer, pH 10/MeOH (= 4:1)	48%

 Table S3. Optimization of the cyclization condition.

(a) MeOH was added to dissolve **34**. (b) Estimated by HPLC. (c) Isolated yield.



Fig. S3 Comparison of the ¹H NMR spectra of natural 1 and synthetic 1. 600 MHz in DMSO- d_6



Fig. S4 HPLC charts of natural 1 and synthetic 1. (column: PEGASIL ODS SP100 4.6×250 mm; eluent: MeOH/H₂O = 70/30, 1.0 mL/min; detection: UV 210 nm)



Fig. S5 Comparison of the CD spectra of natural 1 and synthetic 1 in MeOH.



Fig. S6 ¹H NMR spectrum (500 MHz) of 23 in CDCl₃.



Fig. S7 ¹³C NMR spectrum (125 MHz) of 23 in CDCl₃.



Fig. S8 ¹H NMR spectrum (500 MHz) of 24 in CDCl₃.



Fig. S9¹³C NMR spectrum (125 MHz) of 24 in CDCl₃.



Fig. S10 ¹H NMR spectrum (500 MHz) of 25 in CDCl₃.



Fig. S11 13 C NMR spectrum (125 MHz) of 25 in CDCl₃.



Fig. S12 ¹H NMR spectrum (500 MHz) of 26 in CDCl₃.



Fig. S13 13 C NMR spectrum (125 MHz) of 26 in CDCl₃.



Fig. S14 ¹H NMR spectrum (500 MHz) of 27 in CDCl₃.



Fig. S15¹³C NMR spectrum (125 MHz) of 27 in CDCl₃.



Fig. S16¹H NMR spectrum (500 MHz) of 28 in CDCl₃.



Fig. S17¹³C NMR spectrum (125 MHz) of 28 in CDCl₃.



Fig. S18 ¹H NMR spectrum (500 MHz) of 29 in CDCl₃.



Fig. S19¹³C NMR spectrum (125 MHz) of 29 in CDCl₃.



Fig. S20 1 H NMR spectrum (500 MHz) of 30 in CD₃OD.



Fig. S21 13 C NMR spectrum (125 MHz) of 30 in CD₃OD.



Fig. S22 ¹H NMR spectrum (500 MHz) of 7 in CDCl₃.



Fig. S23 ¹³C NMR spectrum (125 MHz) of 7 in CDCl₃.



Fig. S24 1 H NMR spectrum (500 MHz) of 31 in CD₃OD.



Fig. S25 13 C NMR spectrum (125 MHz) of 31 in CD₃OD.



Fig. S26 ¹H NMR spectrum (500 MHz) of 32 in CD₃OD.



Fig. S27 13 C NMR spectrum (125 MHz) of 32 in CD₃OD.



Fig. S28 ¹H NMR spectrum (500 MHz) of 33 in CD₃OD.



Fig. S29 13 C NMR spectrum (125 MHz) of 33 in CD₃OD.



Fig. S30 1 H NMR spectrum (500 MHz) of 34 in CD₃OD.



Fig. S31 13 C NMR spectrum (125 MHz) of 34 in CD₃OD.



Fig. S32 ¹H NMR spectrum (600 MHz) of synthetic 1 in DMSO- d_6 .



Fig. S33 13 C NMR spectrum (150 MHz) of synthetic 1 in DMSO- d_6 .



Fig. S34 HRMS for compound 23.











Fig. S37 HRMS for compound 26.











Fig. S40 HRMS for compound 29.



Fig. S41 HRMS for compound 30.







Fig. S43 HRMS for compound 32.



Fig. S44 HRMS for compound 33.



Fig. S45 HRMS for compound 34.



Fig. S46 HRMS for synthetic 1.



Fig. S47 HRMS for natural 1.