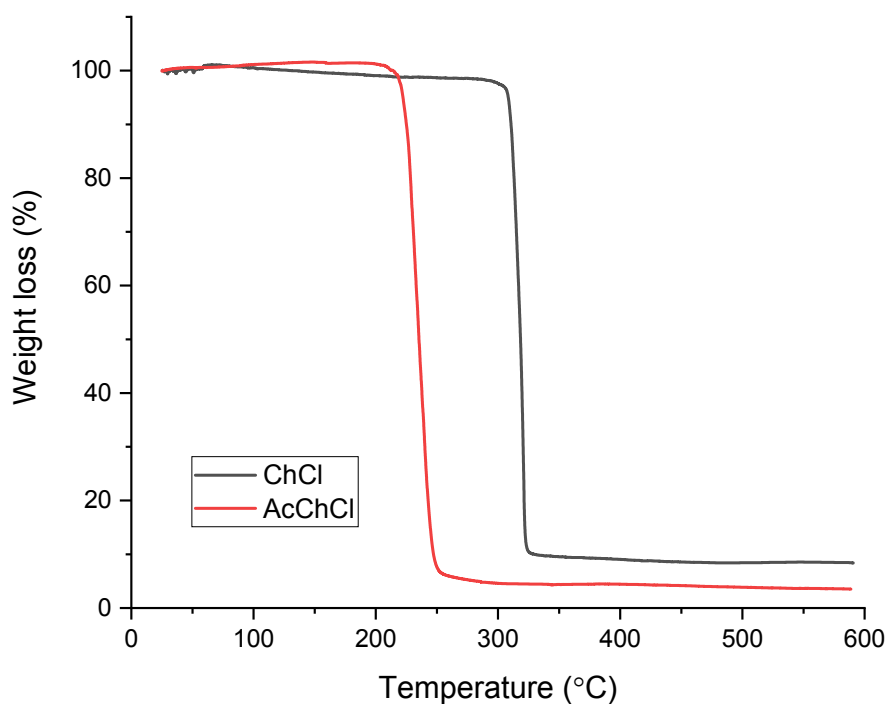


**Electronic Supplementary Information (ESI) for  
Preparation of Thermoresponsive Hydrogels via  
Polymerizable Deep Eutectic Monomer Solvents**

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Thickett\*

**Table S1.** Elemental Analysis Results of Deep Eutectic Monomer Solvents

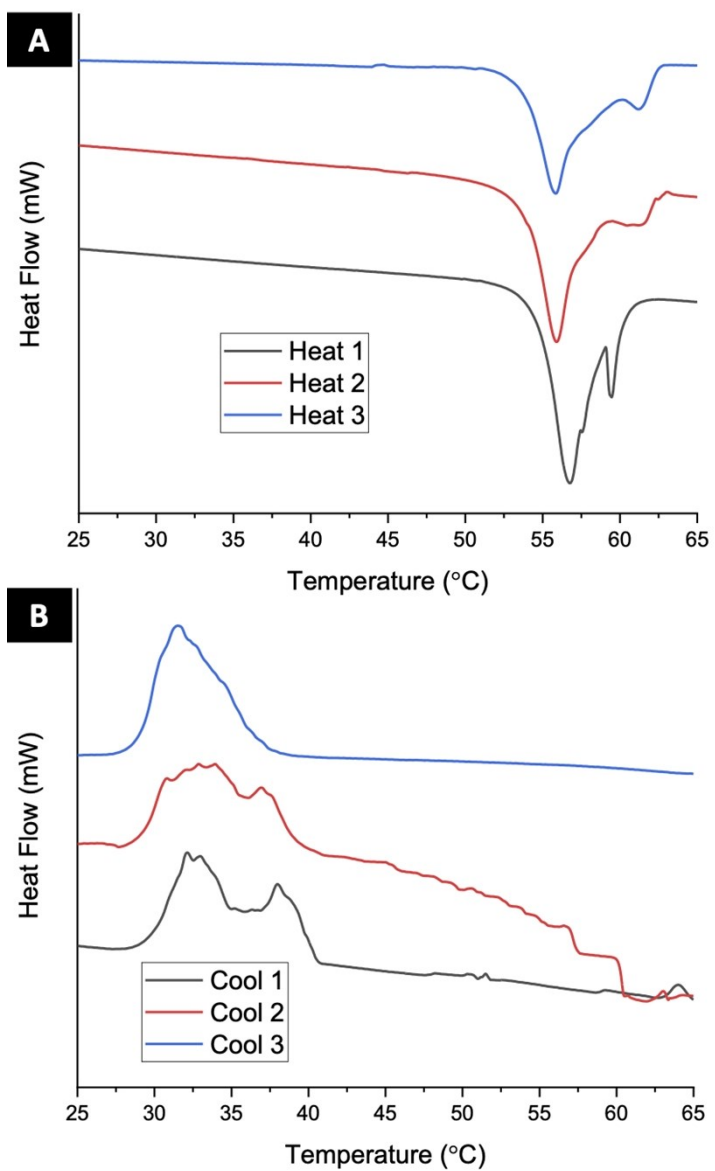
<b>Sample</b>	<b>C %</b>	<b>H %</b>	<b>N %</b>	<b>C/N ratio</b>
NIPAM:ChCl (initial ratio 3:1)	61.52	10.32	12.18	5.05
<i>Predicted values:</i>				
NIPAM:ChCl 3:1	57.65	9.88	11.69	5.03
NIPAM:ChCl 10.5:1	61.54	9.77	12.14	5.07
NIPAM:AcChCl (initial ratio 3:1)	58.84	9.68	11.15	5.27
<i>Predicted values:</i>				
NIPAM:AcChCl 3:1	57.64	9.41	10.76	5.35
NIPAM:AcChCl 4:1	58.72	9.47	11.04	5.31



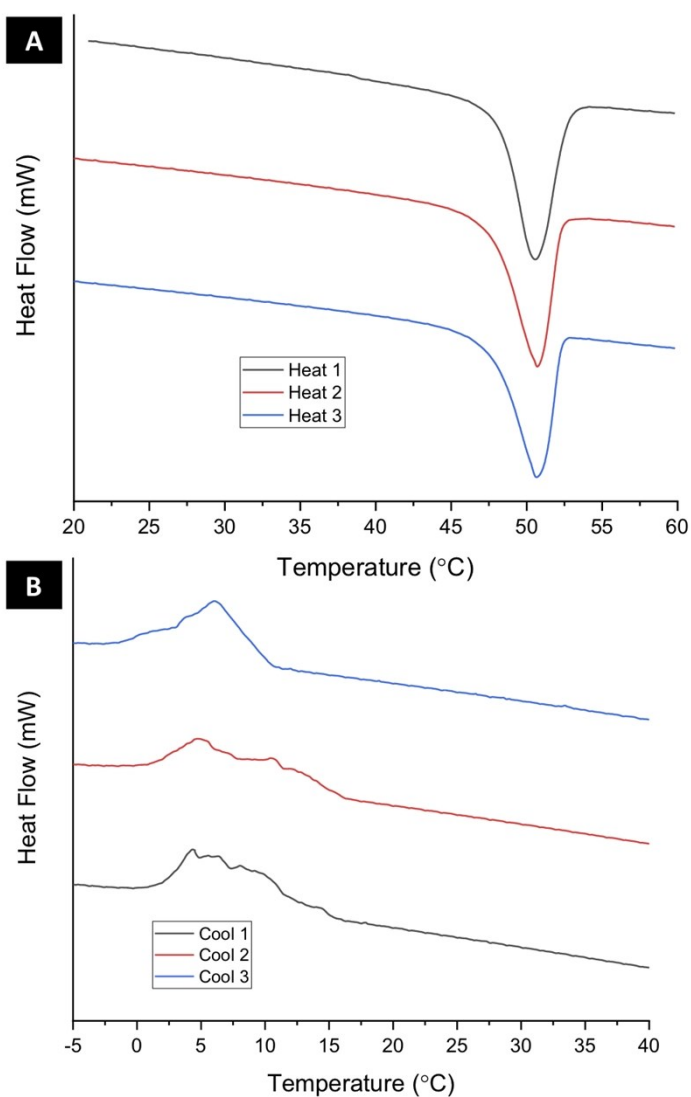
**Figure S1.** Thermogravimetric analysis (TGA) of choline chloride and acetylcholine chloride.

**Table S2.** Onset Temperature and Weight Loss Data from TGA Analysis.

Sample	$T_{\text{onset}}$ (°C)	Total Weight loss (%)
NIPAM	160	98.55
ChCl	307	89.93
NIPAM-ChCl 3:1 eutectic	236	93.91
AcChCl	222	96.06
NIPAM-AcChCl 3:1 eutectic	178	93.11



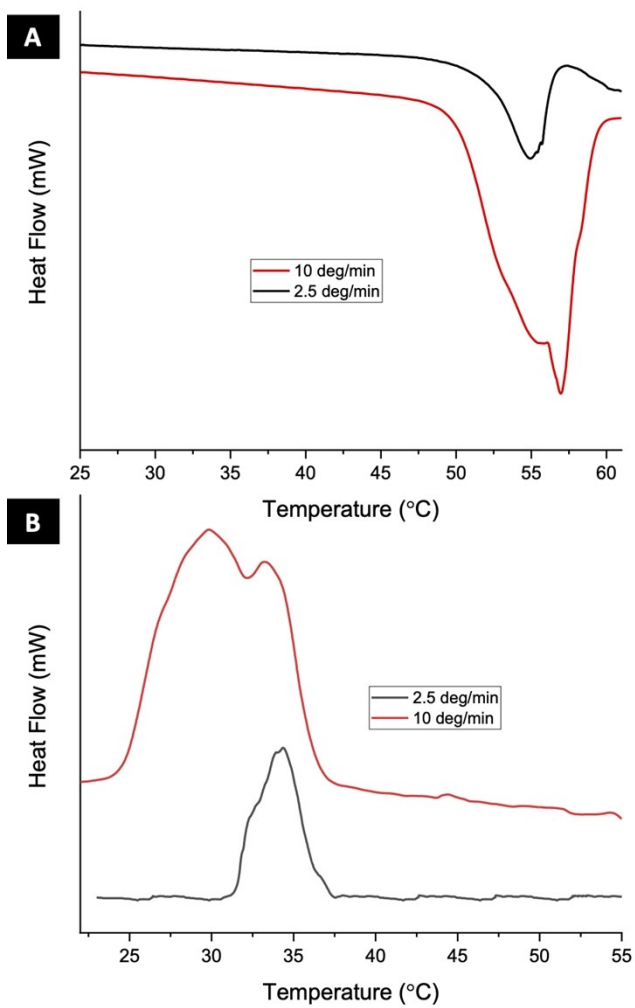
**Figure S2.** DSC analysis of NIPAM:ChCl 3:1 DEM systems and reproducibility of multiple heating (panel A) and cooling (panel B) loops. Heating/cooling rate = 5 °C min<sup>-1</sup>.



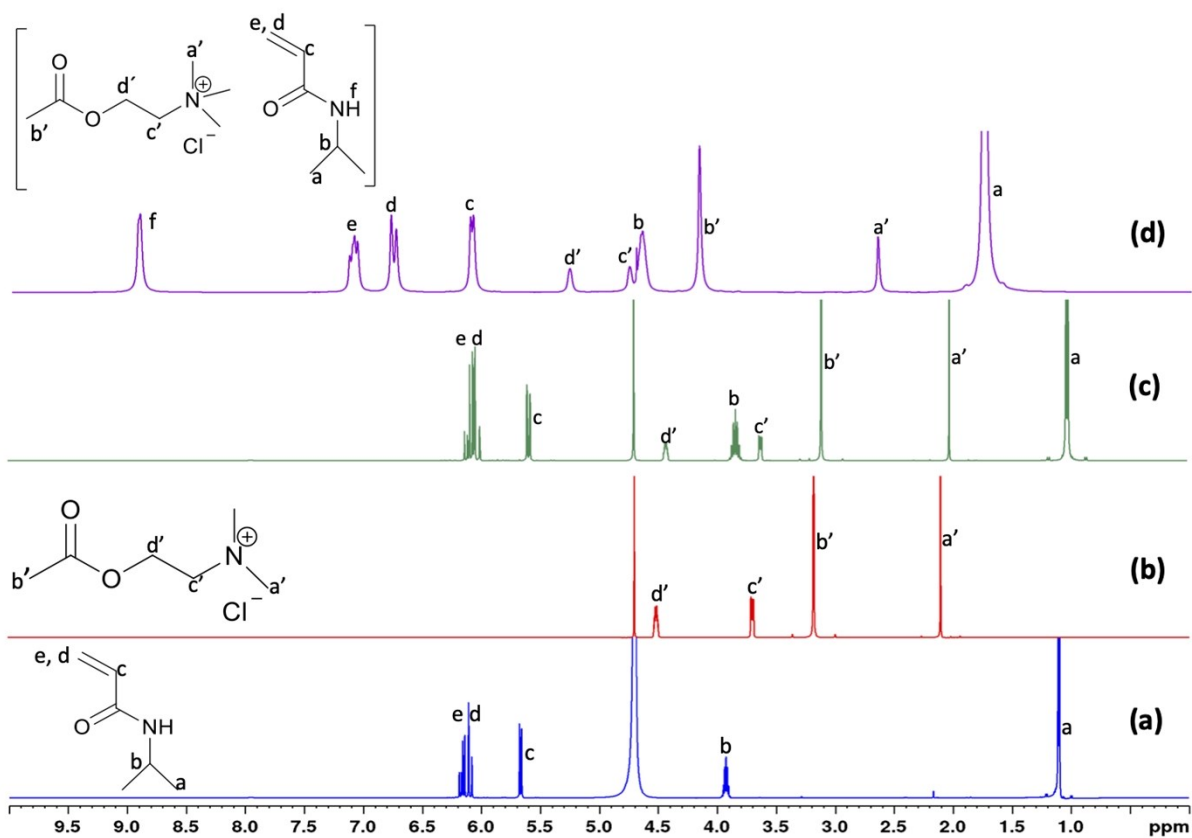
**Figure S3.** DSC analysis of NIPAM:AcChCl 3:1 DEM systems and reproducibility of multiple heating (panel A) and cooling (panel B) loops. Heating/cooling rate = 5 °C min<sup>-1</sup>.

**Table S3.** DSC Analysis of NIPAM:ChCl 3:1 DEM systems (heating/cooling rate = 5 °C min<sup>-1</sup>).

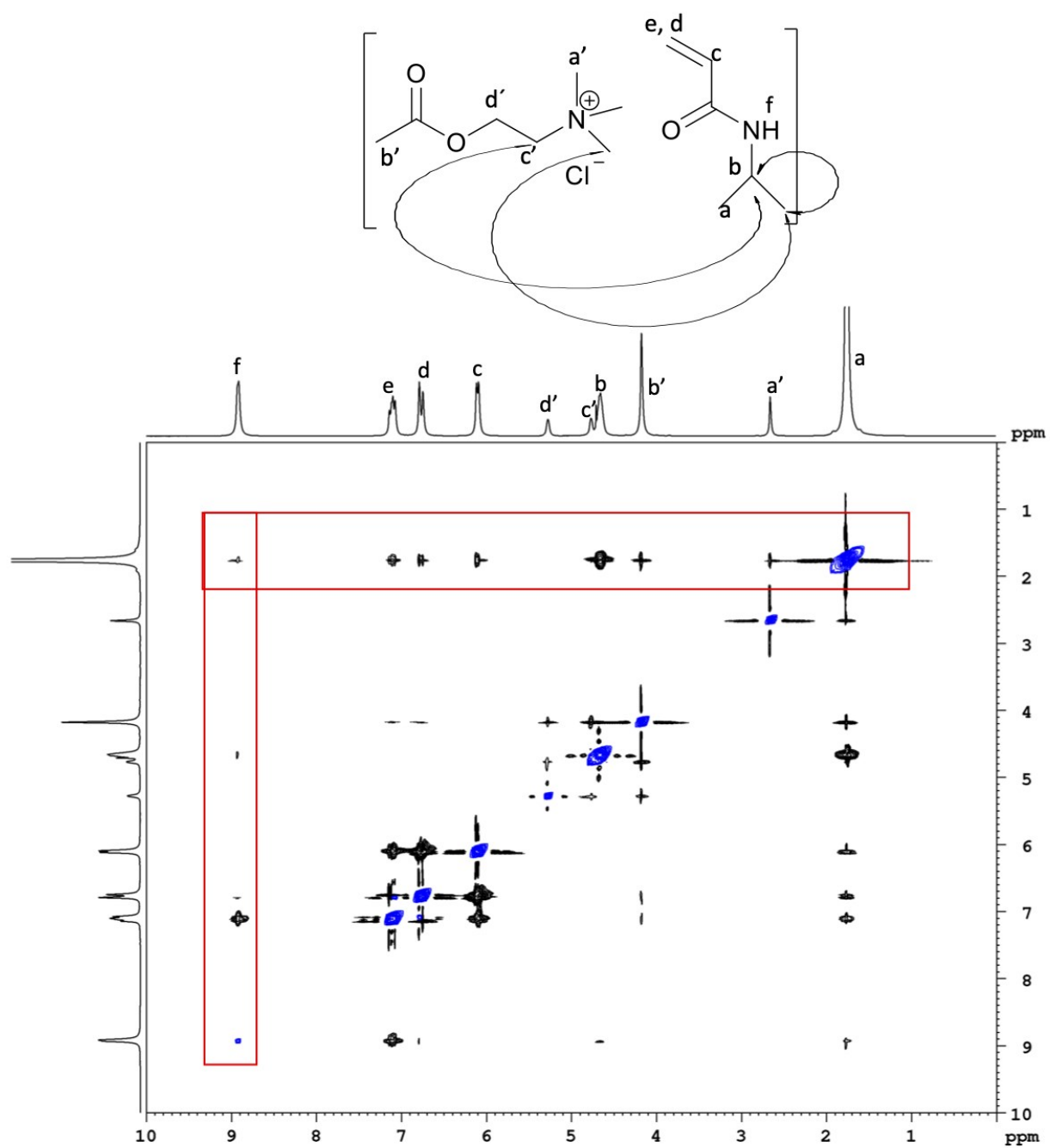
Cycle	$T_{\text{onset}}(^{\circ}\text{C})$	Mean	$T_{\text{max}}(^{\circ}\text{C})$	Mean	$\Delta H$ (J/g)
NIPAM:ChCl 3:1					
Heat 1	54.2	53.9 ± 0.3	56.5	55.9 ± 0.6	69.2 ± 3.4
Heat 2	53.8		55.7		
Heat 3	53.6		55.6		
Cool 1	40.4	39.1 ± 1.6	32.2	32.6 ± 1.4	- 67.1 ± 2.5
Cool 2	39.3		34.0		
Cool 3	37.5		31.6		
NIPAM:AcChCl 3:1					
Heat 1	48.2	48.0 ± 0.2	50.6	50.7 ± 0.1	79.1 ± 7.6
Heat 2	47.8		50.7		
Heat 3	48.1		50.8		
Cool 1	15.2	15.1 ± 1.2	4.4	5.1 ± 1.1	- 50.7 ± 3.0
Cool 2	16.3		4.9		
Cool 3	13.8		6.1		



**Figure S4.** DSC analysis of NIPAM:ChCl 3:1 DEM systems as a function of DSC heating rate.

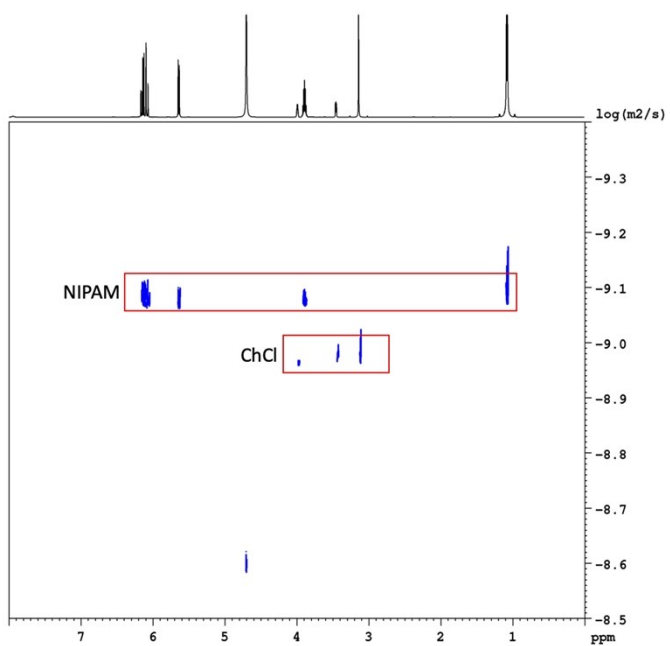
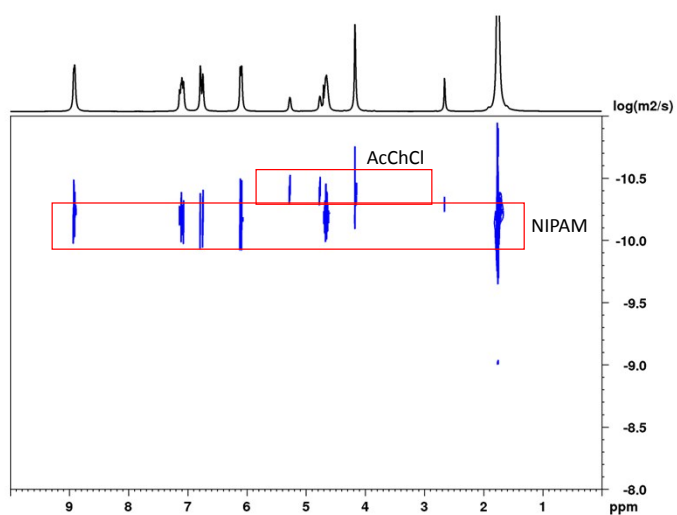
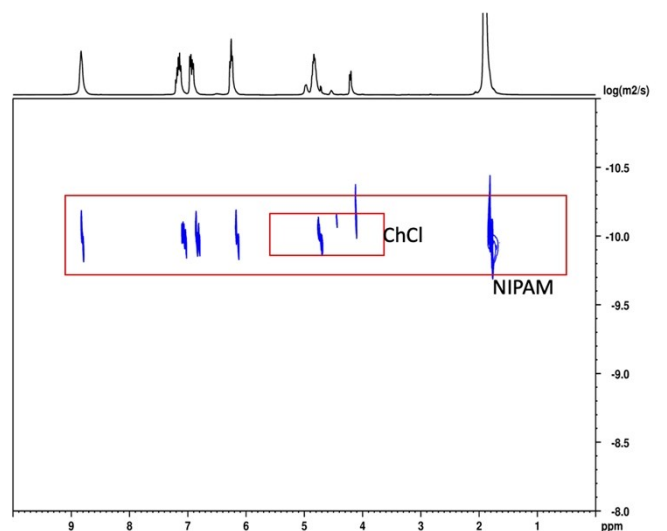


**Figure S5a.**  $^1\text{H}$  NMR spectra a) NIPAM; b) AcChCl; c) a 3:1 mixture of NIPAM and AcChCl (all in  $\text{D}_2\text{O}$ ); d) NIPAM:AcChCl DEM obtained by using a small capillary of  $\text{D}_2\text{O}$  placed in the sample.

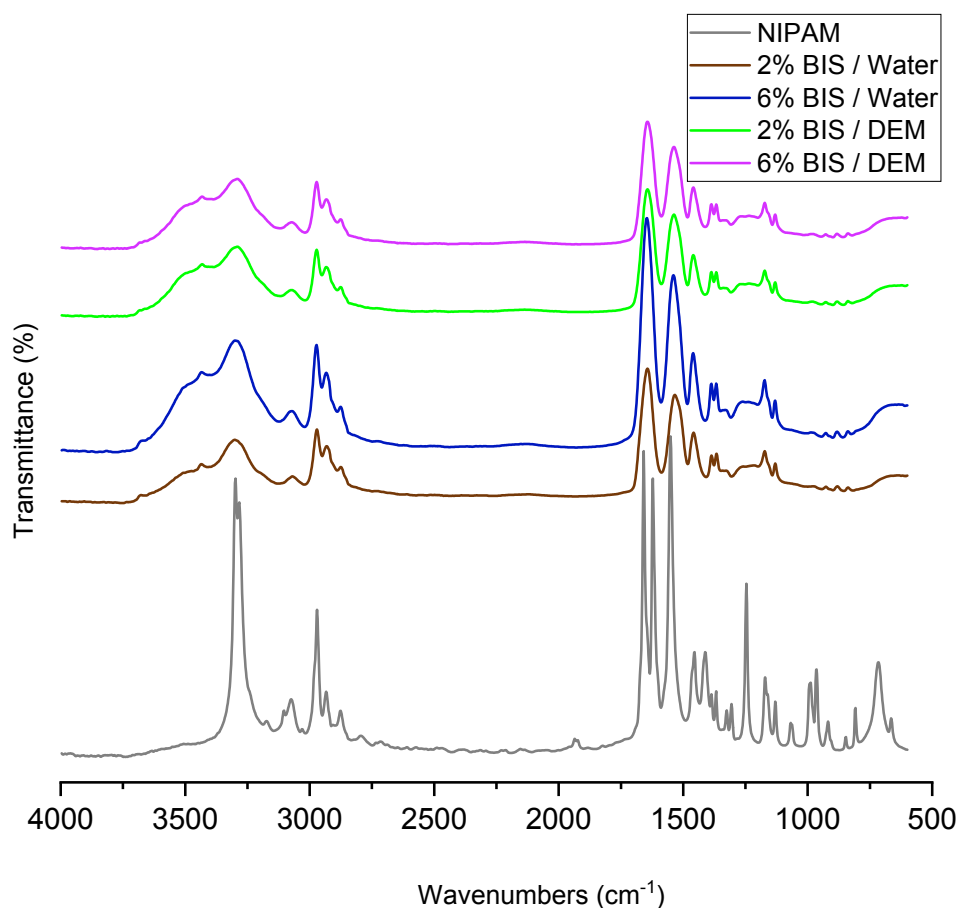


**Figure S5b.**  $^1\text{H}$ - $^1\text{H}$  ROESY spectrum of NIPAM:AcChCl DEM.





**Figure S6.** <sup>1</sup>H DOSY NMR spectra of (top) NIPAM:ChCl DEM; (middle) NIPAM:AcChCl DEM; (bottom) NIPAM and ChCl dissolved in D<sub>2</sub>O.

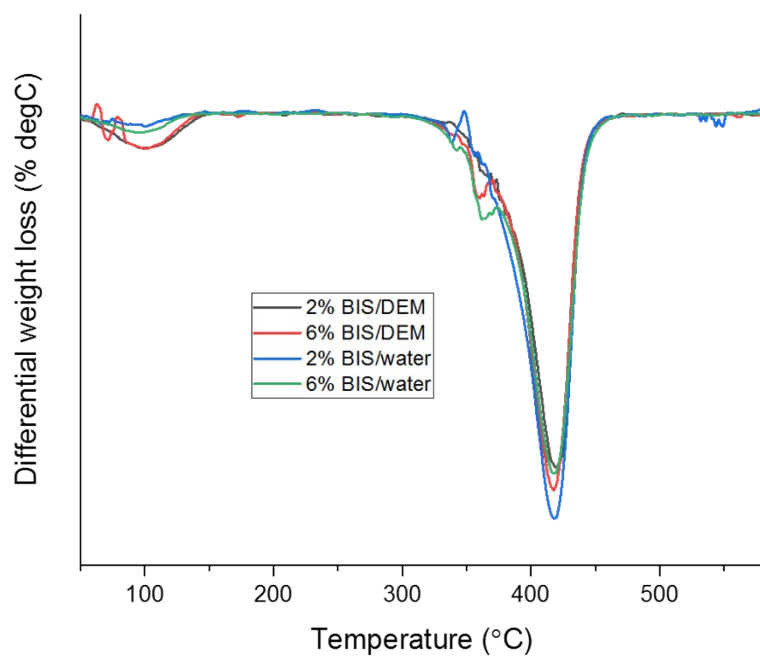


**Figure S7.** ATR-FTIR spectra of poly(NIPAM-co-BIS) hydrogels prepared in both DEMs and water.

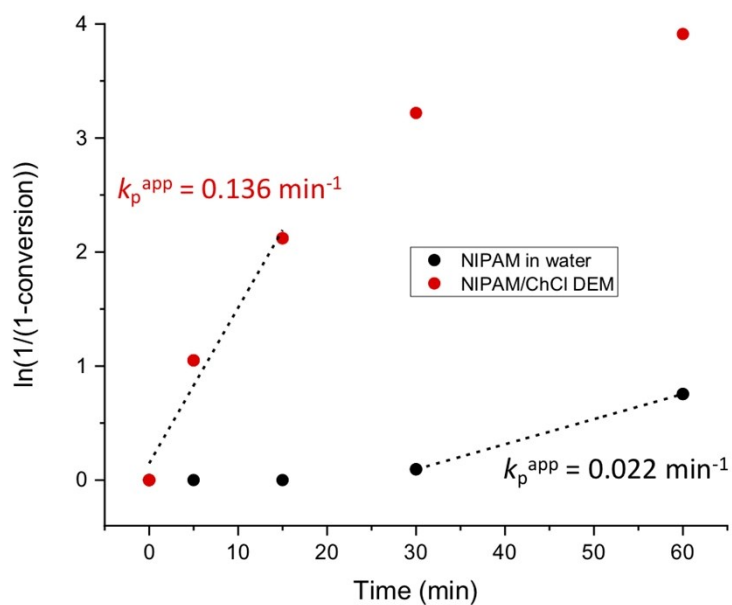
The bands appearing at  $2850\text{ cm}^{-1}$  and  $2900\text{ cm}^{-1}$  are assigned to C–H stretching vibrations. The amide group N–H bending peak is also observed at  $1550\text{ cm}^{-1}$ . The broad band at  $3240\text{ cm}^{-1}$  is consistent with O–H and N–H stretching vibrations of carboxyl and amide groups. Additionally, the broad intense band in the range  $800\text{--}1300\text{ cm}^{-1}$  can be assigned to C–O stretching and C–H bending vibrations.

**Table S4.** TGA Analysis of PNIPAM-BIS Hydrogels.

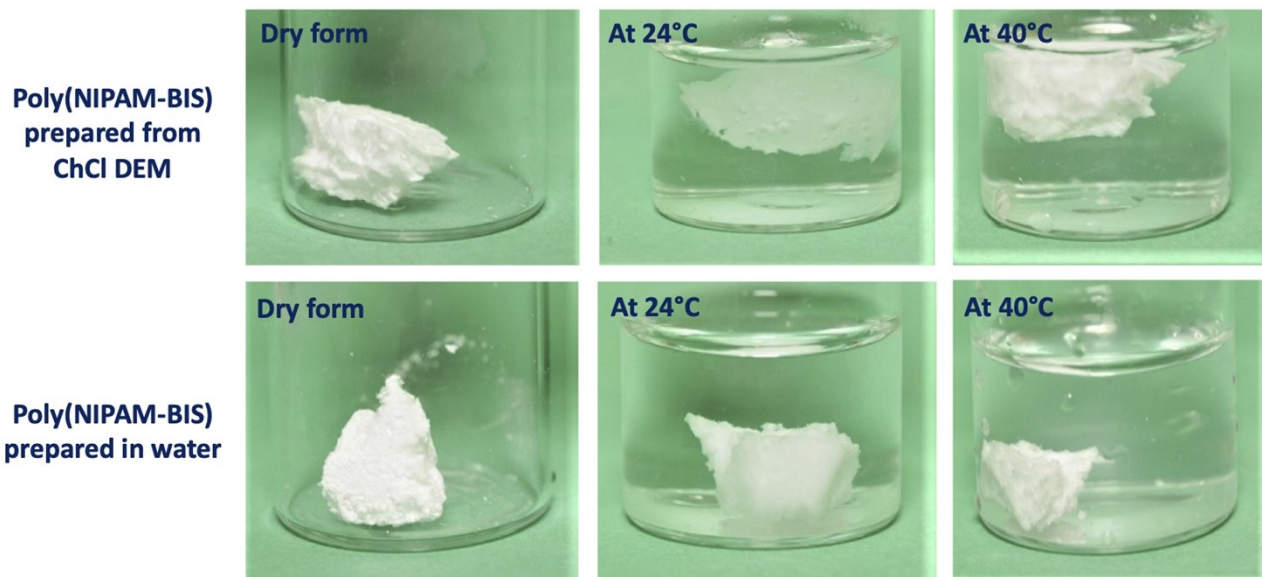
Sample	$T_{\text{onset}}$ (°C)	Weight loss $T < 200$ °C (%)	Weight loss $200 < T < 600$ °C (%)
BIS 2% / DEM	346.9	10.48	80.68
BIS 6 % / DEM	346.3	7.87	86.87
BIS 2 % / water	362.2	2.45	92.73
BIS 6 % / water	345.3	4.88	91.72



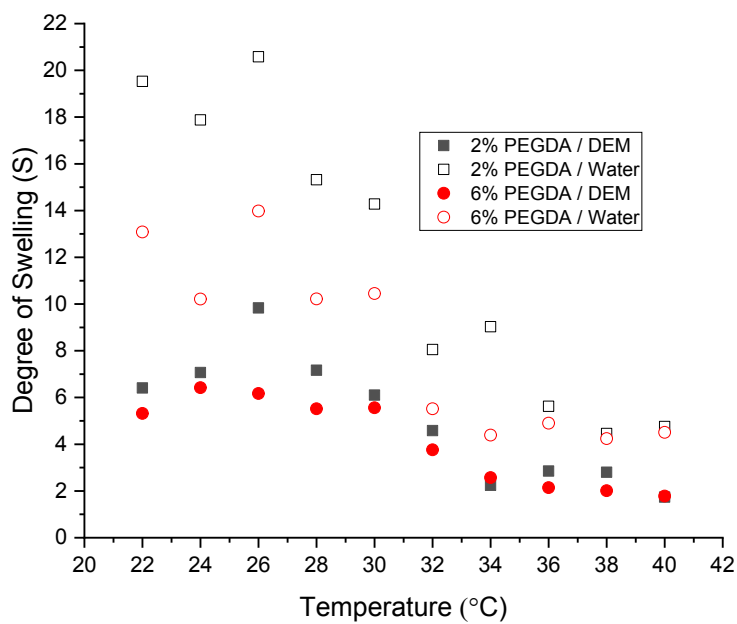
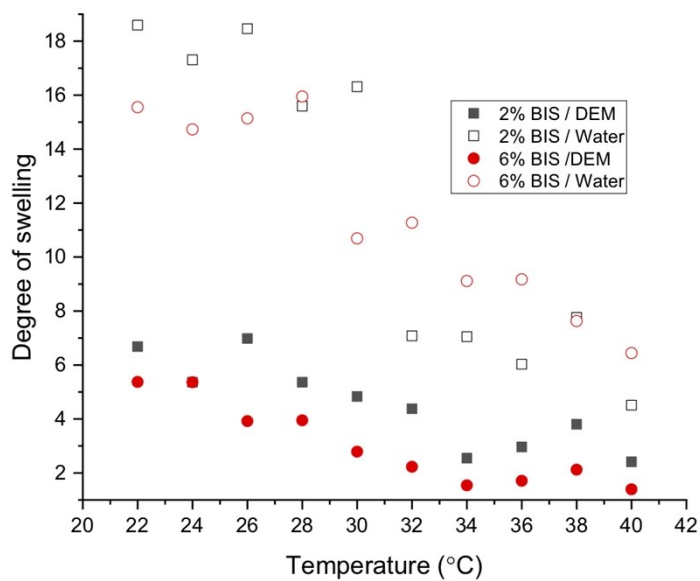
**Figure S8.** Differential TGA Analysis of PNIPAM-BIS Hydrogels.



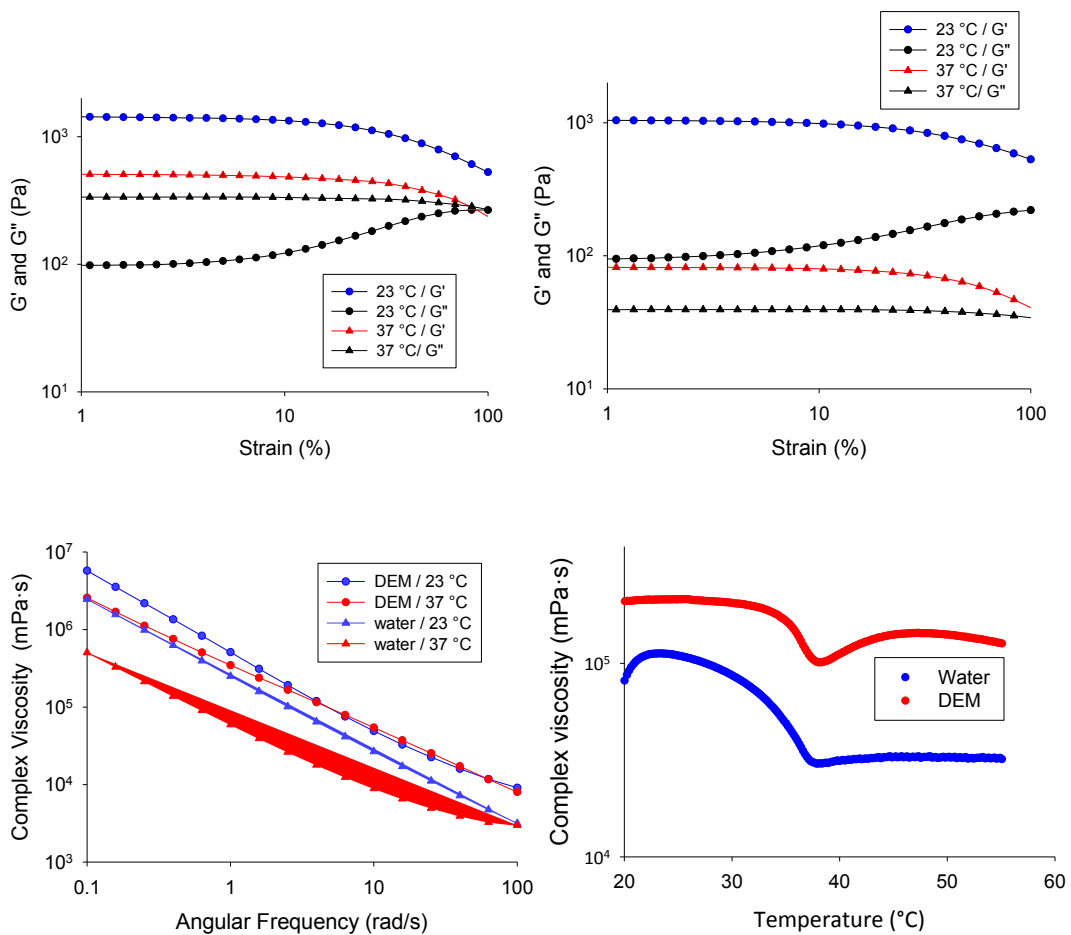
**Figure S9.** Plot of  $\ln(1/(1-\text{conversion}))$  versus time for the polymerization of NIPAM in both a NIPAM:ChCl DEM system as well as in water.



**Figure S10.** Photographs of freeze-dried hydrogels prepared by different methods.



**Figure S11.** Degree of swelling (S) versus temperature for polyNIPAM-based gels prepared from ChCl DEMs (filled symbols) or water (open symbols). The crosslinker concentration was either 2 % (squares) or 6 % (circles) relative to NIPAM. Top panel: BIS as crosslinker; Bottom panel: PEGDA as crosslinker.



**Figure S12.** Rheological analysis of poly(NIPAM-co-BIS) gels swollen with water. (Top left): amplitude sweep of gels prepared from ChCl DEM. (Top right): amplitude sweep of gels prepared from water. (Bottom left): Frequency sweep comparison of the two gels. (Bottom right): Complex viscosity of the two gels as a function of temperature.

### Flory-Rehner Analysis:

Flory-Rehner theory was used to determine the average molecular weight between crosslinks ( $M_c$ ) based on the equilibrium degree of swelling ( $S_{eq}$ ) of the hydrogels prepared in this work. The Flory-Rehner equation for an affine network model can be stated as:

$$\ln(1-\phi) + \phi + \chi\phi^2 = \frac{d_p V_s}{M_c} \left[ \left( \frac{\phi}{2\phi_0} \right) - \left( \frac{\phi}{\phi_0} \right)^{1/3} \right]$$

Where  $\phi$  = polymer volume fraction in the gel,  $\chi$  the Flory-Huggins parameter between the polymer and solvent,  $d_p$  the density of polymer,  $V_s$  the molar volume of solvent,  $M_c$  the average molecular weight between crosslinks, and  $\phi_0$  the polymer volume fraction of the gel in a reference state (see below discussion).

The following temperature and concentration dependence of  $\chi$  was used:<sup>1</sup>

$$\chi = \frac{1}{2} - A \left( 1 - \frac{\theta}{T} \right) + C\phi + D\phi^2$$

Where  $\theta$  is the theta temperature for NIPAM (303.6 K), and  $A$ ,  $C$  and  $D$  are coefficients. Values of  $A = -2$ ,  $C = 0.32$ ,  $D = 0.24$  were used based on recently published values,<sup>1</sup> however we acknowledge there are numerous functional forms (and values of these coefficients) reported in the literature.

The volume fraction of polymer  $\phi$  was calculated from equilibrium swelling values via the following relation:

$$\phi = \frac{1}{1 + \frac{d_p}{d_s} S_{eq}}$$

Where  $d_p$  and  $d_s$  are the densities of polymer and solvent respectively. The values of  $d_p$  and  $d_s$  used in this work for polyNIPAM and water were 1.1 and 1.0 g mL<sup>-1</sup> respectively.

The volume fraction of polymer for each gel at different temperatures studied, in addition to the temperature and concentration-dependent value of the Flory-Huggins parameter, were used to determine the value of  $M_c$  that provided the best fit to experimental data. The value of  $M_c$  was then used to calculate the effective cross-linking density  $q_{eff}$ , which is defined as  $M_0/M_c$  where  $M_0$  is the molar mass of a NIPAM repeat unit (113 Da).

There is significant debate regarding the value and interpretation of  $\phi_0$ ,<sup>2</sup> numerous authors define this quantity as the polymer volume fraction during gel preparation, however Lopez and Richtering<sup>1</sup> show that this often gives poor fit to data and  $\phi_0$  is typically much smaller (2 – 6 % of this value). In this work,  $\phi_0$  was allowed to be a free parameter in the fit to experimental data, and the following values were obtained:

Gel	$\phi_0$
Water	0.020
ChCl DEM	0.056
AcChCl DEM	0.073



For the gel prepared in water, a value of  $\phi_0 = 0.020$  is comparable to the work of Lopez and Richtering,<sup>1</sup> whereas the DEM-based gels present higher  $\phi_0$  values, indicative of their different method of preparation.

**Table S5.** Swelling properties of poly(NIPAM-co-PEGDA) gels when immersed in water.

Method of gel preparation	Temp (°C)	$S_{eq}^a$	$M_c$ (Da) <sup>b</sup>	$q_{eff}^c$
ChCl DEM	22	$6.29 \pm 0.17$	$4165 \pm 190$	0.0273
	40	$1.64 \pm 0.05$		
Water	22	$21.69 \pm 0.53$	$23620 \pm 980$	$4.82 \times 10^{-3}$
	40	$3.65 \pm 0.10$		

<sup>1</sup> Lopez, C.G. and Richtering, W. *Soft Matter*, **2017**, 13, 8271

<sup>2</sup> Quesada-Pérez, M. *et al*, *Soft Matter*, **2011**, 7, 10536.