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**Supporting Information** Antibacterial plasticizers based on bio-2 based engineering elastomers for medical 3 **PVC:** synthesis, characterization and 4 properties 5 Yao Xie, <sup>*a,b,c*</sup>, Bingran Yu<sup>*a,b,c*</sup>, Yaocheng Zhang<sup>*a,b,c*</sup>, Yue Wang<sup>*a,b,c*</sup>, Pengfei Li<sup>*a,b,c*</sup>, 6 Qinan Zhang <sup>a,b,c</sup>, Shun Duan <sup>*a,b,c*</sup>, Xuejia Ding <sup>*a,b,c*</sup> \* and Fu-Jian Xu <sup>*a,b,c*</sup> \* 7 8 <sup>a</sup>State Key Laboratory of Chemical Resource Engineering, Beijing University of 9 Chemical Technology, Beijing, 100029, China 10 <sup>b</sup>Key Laboratory of Biomedical Materials of Natural Macromolecules (Beijing 11 University of Chemical Technology), Ministry of Education, Beijing, 100029, China 12 <sup>c</sup>Beijing Laboratory of Biomedical Materials, Beijing University of Chemical 13 Technology, Beijing, 100029, China 14 15 \*To whom correspondence should be addressed 16 Email: dingxj@mail.buct.edu.cn (X. J. Ding), xufj@mail.buct.edu.cn (F. J. Xu) 17 18 **EXPERIMENTAL SECTION** 19 Morphology of Platelet Adhesion and Activation of PVC samples 20 PVC samples were cut into squares with a side length of 5 mm. Fresh rat blood 21 was centrifuged at 1500 rpm for 15 min and the top layer of platelet rich plasma (PRP) 22 was collected. Afterward, each PVC sample was immersed in 200 µL of PRP in the 96-23 well plate and then incubated for 2 h at 37 °C. Then the samples were carefully rinsed 24 with normal saline in order to remove nonfirmly adsorbed platelets. After fixed with 25 2.5 % glutaraldehyde solution for 24 h, the platelets adsorbed on the surfaces were 26 dehydrated with increasing concentrations of ethanol (25 % and 50 % for 2 min, 75 %, 27 87.5 %, 100%, 100 % and 100% for 3 min, respectively). After natural drying, the 28 obtained samples were observed with SEM. 29 Cellular Toxicity Evaluation of PVC samples. 30 L929 cells were seeded into a 96-well plate (100  $\mu$ L, 1 × 10<sup>5</sup> cfu/mL). After the 31 PVC extraction solutions were obtained, each cell culture solution in 96 well plate 32

33 was replaced by 100 µL of extraction solution. The positive and blank controls were 1 the minimum essential medium (MEM) cell culture solution with 10% DMSO and

2 MEM cell culture solution, respectively. After incubation at 37 °C for 24 h, the

3 extraction solution, cell culture solution and DMSO solution were replaced by 50 μL

4 MTT solution, respectively. After another culturing for 2 h, MTT solution was

5 replaced by 100  $\mu$ L of isopropanol and cells and isopropanol were mixed evenly. The

6 corresponding absorbance at 570 nm was recorded.

7 In Vivo Anti-Infection Assay.

The mice were anesthetized with isoflurane and the hair on the back of the mice 8 was shaved off. Two 0.8 -1.0 cm incisions were cut on both sides of the back, which 9 were parallel to the spine of the mice. S. aureus suspension (1  $\mu$ L, 1×10<sup>9</sup> cfu/mL) was 10 inoculated on each PVC sample and then PVC sample with 1×10<sup>6</sup> CFU S. aureus was 11 implanted subcutaneously into each incision of the mice (Figure S9). Two incisions of 12 13 the same mouse were implanted with the same PVC samples. The incision was closed with 4-0 suture. After 1 day the mice were sacrificed and the samples were removed for 14 characterization. The infectious tissues were weighed, collected in normal saline (10 15 mg/mL) and homogenized. After the dilution for 500 times, 50 µL of the diluted tissue 16 17 homogenates were plated on LB-agar and incubated at 37 °C for 24 h to get a single colony. The PVC samples were placed in normal saline solution, and stained with 18 live/dead Bac Light Bacterial Viability Kit and the bacteria were imaged by CLSM. The 19 blood of mice was collected from eyes for blood test. Cells on the surface of PVC 20 samples were dehydrated by ethanol of different concentrations (25 %, 50 %, 75 %, 21 87.5 %, 100 %). The subcutaneous tissues of mice were excised and fixed with 4% 22 paraformaldehyde solution, embedded in paraffin, cut into slices with a thickness of 5 23 µm and stained with hematoxylin-eosin (H&E) according to the standard protocols. 24 25 Finally, the obtained sections were observed under an optical microscope.

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27 Statistical Analysis.

Each experiment was repeated at least three times, where data are shown as means ± standard deviation.



**Figure S1.** Quantitative <sup>13</sup>C NMR spectra of BPE-2, BPED-2 and QBED-2.



Figure S2. (a) Analysis of quantitative <sup>13</sup>C NMR spectrum of QBED-1, (b) analysis of
 quantitative <sup>13</sup>C NMR spectrum of QBED-2.

wt.% = 
$$\left(\frac{m_{\text{CDCl}_3}/M_{\text{CDCl}_3}\right)/(A_b/A_a) \times M_{\text{QDED}}}{m_{\text{QBED}}} \times 100\%$$

- $m_{1 \text{ CDCl}_3} = 0.8674 \text{ g}$
- $m_{QBED-1} = 0.1092 \text{ g}$
- $m_{2 \text{ CDCl}_3}=0.8792 \text{ g}$
- 8 m<sub>QBED-2</sub>=0.1083 g
- $M_{CDCl_3} = 120.38 \text{ g/mol}$
- $M_{QDED} = 281.28 \text{ g/mol g}$
- $A_{b1}/A_{a1} = 124.7$
- $A_{b2}/A_{a2} = 150.98$



**Figure S3.** Images of QBED-1 and QBED-2.



**Figure S4.** Images of PVC samples.

-	Log reduction									
	PVC/QBED-1-n				PVC/QBED-2-n					
Immersing time in PBS (Days)	10	20	30	40	10	20	30	40		
0	> 3	> 3	> 3	> 3	> 3	> 3	> 3	> 3		
1	2.0	> 3	> 3	> 3	> 3	> 3	> 3	> 3		
3	2.5	> 3	> 3	> 3	> 3	> 3	> 3	> 3		
5	1.1	1.8	2.0	1.9	> 2	1.6	1.6	2.0		
7	0.2	1.4	1.2	1.3	0.8	1.3	1.4	1.5		

## **Table S1**. Log reduction of PVC samples against *S. aureus*



1 Figure S5. Antibacterial effect against *E.coli* of PVC samples after immersions in PBS

for 0, 1, 3, 5, 7 days, respectively, *in vitro*, (a) figures of LB agar plates and (b)
antibacterial ratio of PVC samples against *E.coli*.

Immersing time in PBS (Days)	Log reduction					
	PVC/QE	BED-1-n	PVC/QBED-2-n			
	30	40	30	40		
0	> 3	> 3	> 3	> 3		
1	1.4	> 3	2.3	3.1		
3	0.6	2.6	2.8	3.2		
5	0.3	0.4	2.5	2.9		
7	0.1	0.3	0.1	0.1		

## **Table S2**. Log reduction of PVC samples against *E.coli*



- 1 Figure S6. Photographs of the zones of inhibition of PVC samples against *S.aureus*
- 2 after immersions in PBS for 0, 1, 3, 5, 7 days, respectively.



PVC/QBED-2-30

45 mm

PVC/QBED-2-40

- 1 Figure S7. Photographs of the zones of inhibition of the control group, PVC/QBED-1-
- 2 30, PVC/QBED-1-40, PVC/QBED-2-30 and PVC/QBED-1-40 against E. coli after
- 3 immersions in PBS for 0, 1, 3, 5, 7 days, respectively.



- 1 Figure S8. Morphology of adherent platelets on the surfaces of PVC samples observed
- 2 by SEM.



**Figure S9.** Overview of the animal model with PVC samples infection.