# Supplementary Information

for

# Structure and Activity Relationship Studies of *N*-Heterocyclic Olefins and Thioureas/Ureas Catalytic System: Application in Ring-opening Polymerization of Lactones

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#### 1. General Experimental Procedure

#### **Preparation of NHOs**

The depicted *N*-Heterocyclic Olefins (NHOs) have been synthesized according to literature-known procedures. NHOs was synthesized by deprotonation of the corresponding precursor salt (Scheme S1) using KHMDS and they were stored in a glove box at -25 °C. For full characterization of NHO1, NHO2, NHO3, NHO4 and NHO5 see the cited literature.<sup>1-5</sup>



Scheme S1. Synthetic routes for the NHOs employed in this study.

### **Preparation of TUs/Us**

The depicted thioureas (TUs) and ureas (Us) are prepared by mixing the appropriate amine and isothiocyanate in a solution of MeOH (Scheme S2). The solution was stirred in the room temperature for 30 min. Then MeOH is removed under vacuum and the products were purified by washing with hexanes three times. After vacuum filtration, the filter residue was vacuum drying at 50 °C for 24 h. For full characterization of TU1, TU2, TU3, TU4, TU5, TU6, U1, U2 and U8 see the cited literature.<sup>6-10</sup> U3, U4, U5, U6 and U7 were purchased from commercial suppliers and dried under vacuum at 50 °C for 24 hours.



Scheme S2. Synthetic routes for the TUs/Us.

# 2. pK<sub>a</sub> determination of NHOs

When NHOs and BnOH were mixed together, the methylene group of BnOH shifted downfield. The stronger the basicity of NHOs, the greater the chemical shift of the methylene group of BnOH.



Figure S1. <sup>1</sup>H NMR ( $d_8$ -Toluene, 400 MHz) of NHOs and BnOH (NHOs:BnOH =1:1).



4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.6 3.5 3.4 3.3 3.2 3.1 3.0 2.9 2.8 2.7 2.6 2.5 2.4 2.3 2.2 2.1 2.0 1.9 1.8 1.7 1.6 1.5 1.4 1.3 1.2 1.1 1.0 0.9 0.8 0.7 0.6 0 chemical shift (ppm)

**Figure S2.** <sup>1</sup>H NMR ( $d_3$ -MeCN, 400 MHz) of NHO5 precursor salt, 'Bu-P<sub>1</sub> and combinations (NHO5 precursor salt: 'Bu-P<sub>1</sub> =2:1).

The NMR signal of the "d" methyne group remained unchanged when 'Bu-P<sub>1</sub> ( $pK_a = 26.9$ ) was

used to deprotonate the NHO5 precursor salt, which proves that NHO5 exhibits a higher  $pK_a$  than 'Bu-P<sub>1</sub> ( $pK_a[NHO5] > 26.9$ ).



**Figure S3.** <sup>1</sup>H NMR ( $d_3$ -MeCN, 400 MHz) of NHO5 precursor salt, 'Bu-P<sub>2</sub> and combinations (NHO5 precursor salt: 'Bu-P<sub>2</sub> =2:1).

The signal of the "d" methyne group disappeared when the 'Bu-P<sub>2</sub> ( $pK_a = 33.5$ ) base was used for the deprotonation of NHO5 precursor salt, which proves that 'Bu-P<sub>2</sub> exhibits a higher  $pK_a$  than NHO5 ( $pK_a[NHO5] < 33.5$ ).



**Figure S4.** <sup>1</sup>H NMR ( $d_3$ -MeCN, 400 MHz) of NHO1 precursor salt, DBU and combinations (NHO1 precursor salt: DBU =2:1).

The NMR signal of the "a" methyl group remained unchanged when DBU ( $pK_a = 24.3$ ) was used to deprotonate the NHO1 precursor salt, which proves that NHO1 exhibits a higher  $pK_a$  than DBU ( $pK_a[NHO1] > 24.3$ ).



Figure S5. <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of NHO1, TU5 and combinations (NHO1:TU5=1:1).



Figure S6. <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of NHO2, TU5 and combinations (NHO2:TU5=1:1).



Figure S7.<sup>1</sup>H NMR (*d*<sub>8</sub>-THF, 400 MHz) of NHO3, U8 and combinations (NHO3:U8=1:1).



Figure S8. <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of NHO3 and U6 (NHO3:U6=1:1).



Figure S9. <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of NHO3, U5 and combinations (NHO3:U5=1:1).



Figure S10. <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of NHO3 and U5 (NHO3:U5=1:1).

# 3. <sup>1</sup>H NMR Spectra

For homonuclear decoupled <sup>1</sup>H NMR analysis, acquisition time was measured and fixed to 2.04 s. Samples were obtained in CDCl<sub>3</sub> solutions with the decoupling pulse

based on the methyl region ( $\delta = 1.5$  ppm). In case of the good separation of methine region ( $\delta = 5.00-5.20$  ppm), both Bernouillan statistics (chain-end control mechanism, CEC) and non-Bernouillan statistics (enantiomorphic site control mechanism, ESC) were used to calculate  $P_{\rm m}$  values.<sup>11</sup>

	Probability of ESC (non-Bernouillan)	Probability of CEC (Bernouillan)	
[mmm]	$[P_{\rm m}^2 + (1 - P_{\rm m})^2 + P_{\rm m}^3 + (1 - P_{\rm m})^3]/2$	$P_{\rm m}^2$ +0.5 $P_{\rm m}P_{\rm r}$	
[rmm]	$[P_{\rm m}^2(1-P_{\rm m}) + P_{\rm m}(1-P_{\rm m})^2]/2$	$0.5 P_{\rm m}P_{\rm r}$	
[mmr]	$[P_{\rm m}^2(1-P_{\rm m}) + P_{\rm m}(1-P_{\rm m})^2]/2$	$0.5 P_{\rm m}P_{\rm r}$	
[rmr]	$[P_{\rm m}^2(1-P_{\rm m}) + P_{\rm m}(1-P_{\rm m})^2]/2$	$0.5 P_{\rm r}^2$	
[mrm]	$[P_{\rm m}(1-P_{\rm m})]$	$0.5 (P_{\rm m}^2 + P_{\rm m}P_{\rm r})$	

 $P_{\rm r}$  has meaning of racemic enchainment, and  $P_{\rm m}+P_{\rm r}=1$ .



Figure S11. Homonuclear decoupled <sup>1</sup>H NMR (Table 4, entry 1).



Figure S12. Homonuclear decoupled <sup>1</sup>H NMR (Table 4, entry 3).



Figure S13. Homonuclear decoupled <sup>1</sup>H NMR (Table 4, entry 4).



Figure S14. Homonuclear decoupled <sup>1</sup>H NMR (Table 4, entry 6).



Figure S15. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) spectra of the obtained PVL.



Figure S16. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) spectra of the obtained PCL.



Figure S17. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) spectra of the obtained PLA.

4. GPC curves of various polymer



Figure S18. GPC curves of the obtained PVL (Table 1).



Figure S19. GPC curves of the obtained PCL (Table 2).



Figure S20. GPC curves of the obtained PLA at room temperature (Table 3).



Figure S21. GPC curves of the obtained PLA at low temperature (Table 4).



Figure S22. GPC curves of the obtained PVL (Table 5, entries 2-5).



Figure S23. GPC curves of the obtained PVL (Table 5, entries 7-10).

# 5. Experimental data

TUs/Us	k <sub>obs</sub> (min <sup>-1</sup> )
TU1	$7.1*10^{-6} \pm 2.1*10^{-8}$
TU2	$7.2*10^{-6} \pm 3.2*10^{-8}$
TU3	$0.00072 \pm 2.4*10^{-5}$
TU4	$0.0016 \pm 6.4*10^{-5}$
TU5	$0.0034 \pm 9.5*10^{-5}$
U1	$0.148 \pm 0.0031$
U2	$0.064 \pm 0.0018$
U3	$0.138 \pm 0.0013$
TU6	$0.322 \pm 0.013$
U4	$0.537 \pm 0.019$
U5	$0.0055 \pm 2.3*10^{-4}$
U6	$0.0014 \pm 6.1*10^{-5}$
U7	$0.0013 \pm 6.5*10^{-5}$
U8	$0.00082 \pm 3.3^{*}10^{-5}$

Table S1.  $k_{obs}$  for  $\delta\text{-VL}$  ROP with NHO3 and different TUs (Us)  $^a$ 

 $\label{eq:aReaction conditions: [NHO3]_0: [TU/U]_0: [BnOH]_0: [VL]_0 = 1:2:2:800 \mbox{ in toluene at room temperature, } [VL]_0 = 3.4 \mbox{ mol/L}.$ 

Table S2. ROPs of  $\delta$ -VL with various ratios of monomer/initiator <sup>a</sup>

Entry	NHO3/U1/BnOH/δ-VL	Time	Conv. <sup>b</sup>	$M_{ m n,cal}$ (g/mol)	$M_{\rm n,exp}({ m g/mol})^{ m c}$	а
1	1/2/2/100	30 s	95 %	4800	5400	1.14
2	1/2/2/200	5 min	98 %	9800	12200	1.15
3	1/2/2/400	10 min	94 %	18800	19500	1.16
4	1/2/2/800	30 min	96 %	38400	34300	1.22

<sup>a</sup> Polymerization conditions: room temperature, toluene as solvent, the reactions were carried in sealed tube. <sup>b</sup> Determined by <sup>1</sup>H NMR in CDCl<sub>3</sub> using integrals of the characteristic signals. <sup>c</sup> Number-average molar mass ( $M_n$ ) and dispersity values were determined by GPC in THF at 25 °C using polystyrene standards for calibration, and corrected using the factor 0.57 for PVL.

Time	Conv. <sup>b</sup>	M <sub>n,cal</sub> (g/mol)	$M_{\rm n,exp}({ m g/mol})^{ m c}$	а
30 min	8 %	4600	4200	1.12
2 h	26 %	14800	13300	1.10
4 h	51 %	29100	22300	1.15
6 h	68 %	38800	31000	1.18
8 h	80 %	45700	38100	1.21
10 h	98 %	55900	46100	1.46

Table S3. The results of monomer conversion and molecular weight versus time <sup>a</sup>

<sup>a</sup> Unless otherwise specified, reaction conditions are [NHO3]<sub>0</sub>:[U1]<sub>0</sub>:[BnOH]<sub>0</sub>:[ $\epsilon$ -CL]<sub>0</sub> = 1:2:2:1000 in toluene at room temperature. <sup>b</sup> Determined by <sup>1</sup>H NMR in CDCl<sub>3</sub> using integrals of the characteristic signals. <sup>c</sup> Number-average molar mass ( $M_n$ ) and dispersity values were determined by GPC in THF at 25 °C using polystyrene standards for calibration, and corrected using the factor 0.56 for PCL.

### 6. MALDI-TOF MS spectrum



**Figure S24.** MALDI-TOF MS spectrum of the obtained PCL sample (NHO3/U1/BnOH/ $\epsilon$ -CL= 1:2:2:200).

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