

Supplementary Information for

**Natural Carbolines Inspired the Discovery of Chiral CarOx Ligands
for Asymmetric Synthesis and Antifungal Leads**

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Content

General Information	3
Synthesis and Structural Elucidation of β^1-CarOx Ligands	4
Synthetic Route to the Chiral β^1 -CarOx Ligands	4
Characterization of the Synthesized Chiral β^1 -CarOx Ligands	6
Synthesis and Structural Elucidation of β^3-CarOx Ligands	12
Synthetic Route to the Chiral β^3 -CarOx Ligands	12
Characterization of the Synthesized Chiral β^3 -CarOx Ligands	15
Enantioselective Michael Addition of Arylboronic Acids to Nitroalkenes:	16
General procedure	16
Optimization of the Reaction Conditions	17
Substrate Scope	18
HPLC traces of the Enantioenriched β -aryl nitroethanes	20
Enantioselective Addition of Arylboronic Acids to β-substituted Cyclic Enones	38
General procedure	38
Concise optimization	38
Substrate Scope	39
HPLC traces of the Enantioenriched β -aryl ketones	40
Synthesis and Application of Chiral Ligands L12 and L13	48
Antifungal Bioassay of CarOx Ligands	51
Compounds Selected for Biotest	51
Initial Screening	52
Precise Antifungal Test	53
References	54
NMR Spectra Traces	55

General Information

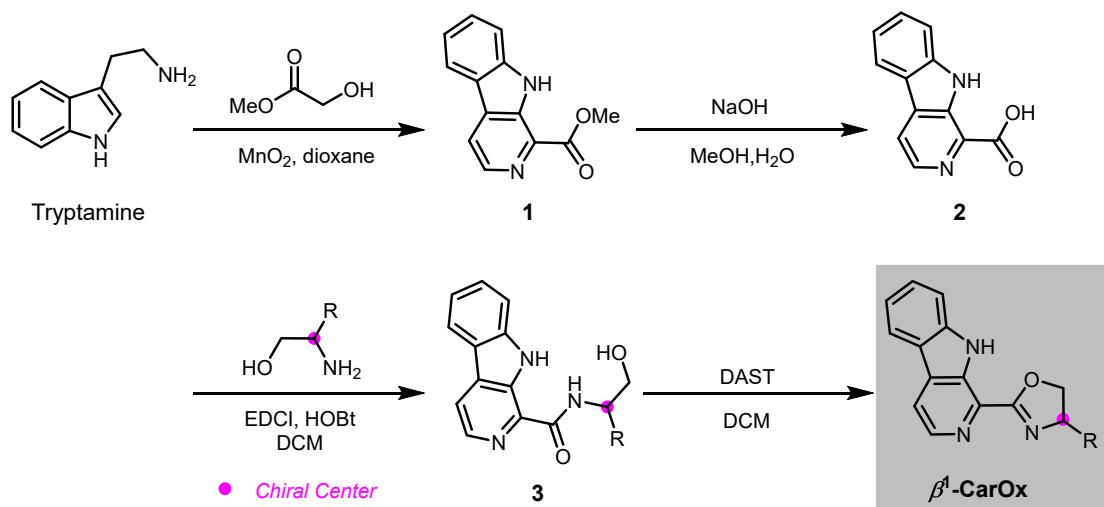
Unless otherwise stated, all solvents and reagents were purchased from commercial sources (Energy or Meryer Chemicals etc.), they were analytically pure and used without further purification. Anhydrous solvents were dried and distilled by standard techniques before use or were purchased from commercial sources (Energy Chemicals etc.).

Silica gel GF₂₅₄ and column chromatography silica gel for isolation (200-300 mesh) were both purchased from Qingdao Broadchem Industrial Co., Ltd. Reaction progress was monitored by thin-layer chromatography (TLC) on silica gel GF₂₅₄ with ultraviolet (UV_{254nm} or UV_{365nm}) detection. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker AV 400 or Bruker AV 500 spectrometers with CDCl₃ as solvent and tetramethylsilane as the internal standard. The chemical shifts (δ) were recorded in parts per million (ppm). Data for ¹H NMR are reported as follows: chemical shift (δ : ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; and m, multiplet), coupling constant (Hz), integration and assignment (H). Data for ¹³C NMR are reported in terms of chemical shift (δ : ppm). Electrospray ionization high-resolution mass spectrometry (ESI-HRMS) data were also obtained with the Waters XEVO G2-XS Q-TOF mass spectrometer.

The agriculturally important plant pathogens were provided by the College of Plant Protection, Nanjing Agricultural University (Nanjing, China). The *in vitro* antifungal activities of the synthesized natural product alangiolbussinine and its analogues were carried out according to the procedures we used previously^[1].

Synthesis and Structural Elucidation of β^1 -CarOx Ligands

Synthetic Route to the Chiral β^1 -CarOx Ligands



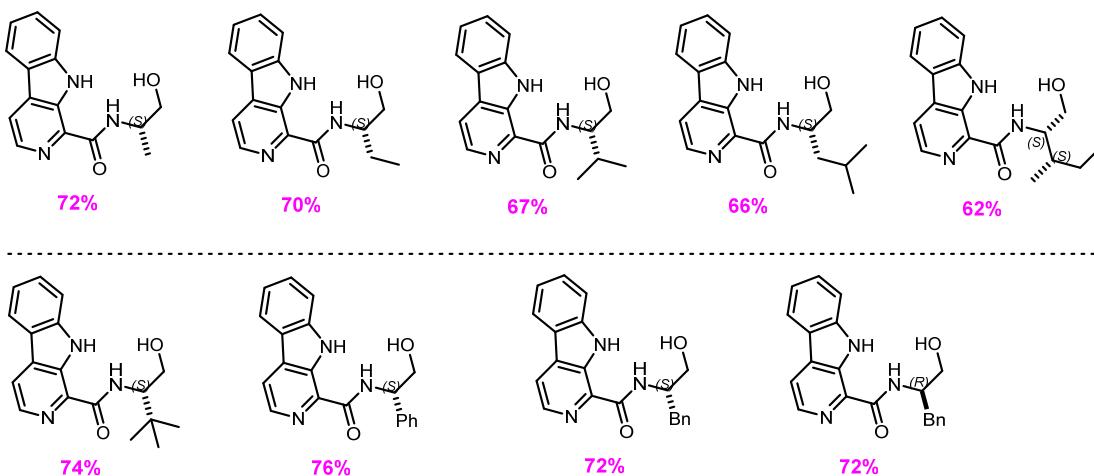
Step 1, Synthesis of the intermediate **1** according to the report by Lang,^[2] To a stirred suspension of tryptamine (3.2 g, 20 mmol) and 3A MS (20 g) was added 1,4-dioxane (50 mL). MnO₂ (17 g) and methyl glycolate (2.7 mL) was added dropwise at 0 °C. The heterogeneous mixture was stirred at room temperature for 3h and then was immersed in a preheated oil bath (110 °C) until the full consumption of the starting material was detected by thin layer chromatography (TLC). The heterogeneous mixture was filtered and rinsed with ethyl acetate. The organic phase concentrated under vacuum. Purification by silica gel column chromatography on silica gel (200-300m) with hexane/EtOAc (2:1, v/v) as the eluent gave the β -carboline-1-carboxylic methyl ester (intermediate **1**) as yellow solid in 42% yield.

Step 2, To a stirred suspension of β -carboline-1-carboxylic methyl ester **1** (2.26 g, 10 mmol) was added methanol (30 mL). Aqueous sodium hydroxide solution (10 mL, 1 M) was added dropwise at 0 °C. The heterogeneous mixture was immersed in a preheated oil bath (50 °C) and was stirred until the full consumption of the starting material was detected by thin layer chromatography (TLC). The mixture was concentrated under vacuum to remove methanol, and was adjusted to pH 6-7 with hydrochloric acid, from which the β -carboline-1-carboxylic acid (intermediate **2**) was precipitated, the heterogeneous mixture was filtered, and the β -carboline-1-carboxylic

acid intermediate **2** was collected and dried as a yellow solid in 89% yield.

Step 3, general procedure of Steglich Condensation: To a dried Schlenk flask charged with the synthesized β -carboline-1-carboxylic acid, compound **2**, (1 mmol, 212 mg) and the specific chiral amino alcohol (1 mmol), was added anhydrous dichloromethane (5 mL) for dissolution. Hydroxybenzotriazole (HOEt) (175 mg, 1.3 mmol) and *N*-(3-(dimethyl amino)propyl)-*N'*-ethylcarbodiimide hydrochloride (EDCI-HCl) (0.25 g, 1.3 mmol) were then added while the reaction flask was in an ice bath. The mixture was allowed to gradually warm to room temperature, and it was stirred overnight until full consumption of the carboxylic acid detected by thin layer chromatography (TLC). The mixture was quenched by the addition of a saturated aqueous solution of NaHCO₃ (20 mL) and separated. The water phase was extracted with dichloromethane (10 mL \times 3), and the combined organic phase was sequentially washed with water (10 mL \times 2) and saturated aqueous NaCl (10 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum. Purification by silica gel column chromatography on silica gel (200-300m) with hexane/EtOAc (2:1, v/v) as the eluent gave the amide intermediate **3**.

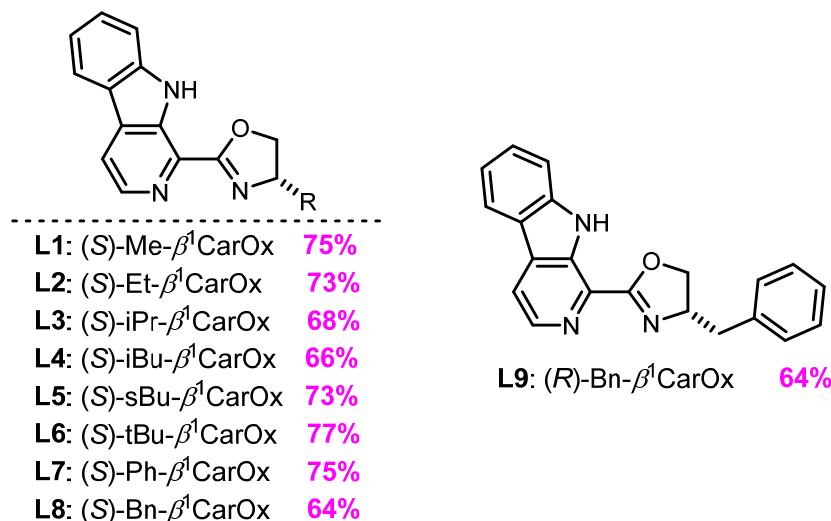
Yields for the intermediate **3** are listed below,



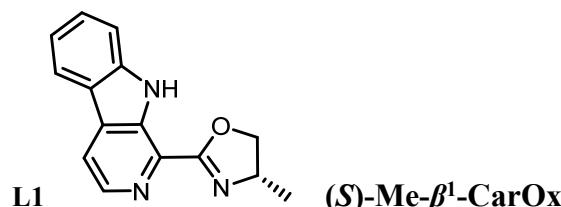
Step 4, General procedure for the DAST mediated cyclization to produce β^L -CarOx Ligand: To a Schlenk tube charged the amide intermediate **3** (1.0 mmol) was added anhydrous DCM (5.0 mL) under N₂ atmosphere. Diethylaminosulfur trifluoride (DAST) (160mg, 1mmol) was added dropwise at -78 °C. The reaction mixture was

stirred at -78 °C until the full consumption of the starting material was detected by TLC. The mixture was quenched by the addition of a saturated aqueous solution of NaHCO₃ (10 mL) and separated, The water phase was extracted with dichloromethane (10 mL × 3), and the combined organic phase was sequentially washed with water (10 mL × 2) and saturated aqueous NaCl (10 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum. Purification by silica gel column chromatography on silica gel (200-300m) with hexane/EtOAc (2:1, v/v) as the eluent gave the chiral ligand β^1 -CarOx.

Yields for the chiral ligands β^1 -CarOx are listed below,



Characterization of the Synthesized Chiral β^1 -CarOx Ligands



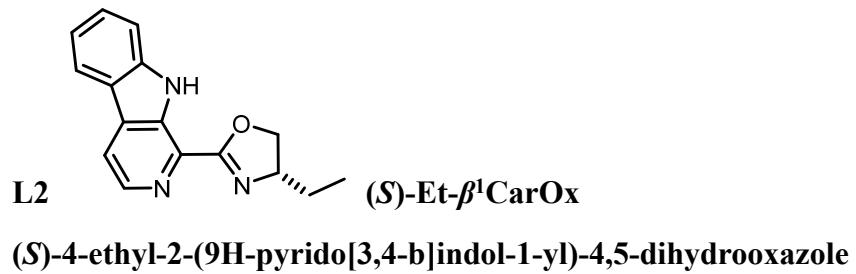
(S)-4-methyl-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

White solid, ¹H-NMR (400 MHz, CDCl₃) δ 1.47 (d, *J* = 6.44 Hz, 3H, CH₃), 4.12 (dd, *J*₁ = 7.60 Hz, *J*₂ = 7.52 Hz, 1H, OCH₂CH), 4.58 (m, 1H, OCH₂CH), 4.66 (dd, *J*₁ = 9.36 Hz, *J*₂ = 7.84 Hz, 1H, OCH₂CH), 7.31 (m, 1H, *H* in Benzene Ring), 7.57-7.60 (m, 2H,

*H in Benzene Ring), 8.04 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*), 8.13 (dd, $J_1 = 7.88$ Hz, $J_2 = 0.96$ Hz, 1H, *H in Benzene Ring*), 8.54 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*), 10.4 (s, br, 1H, *NH*).*

^{13}C NMR (100 MHz, CDCl_3) δ 21.8, 62.3, 73.9, 111.9, 116.9, 120.3, 121.3, 121.9, 128.9, 129.0, 129.9, 135.8, 138.7, 140.6, 163.1.

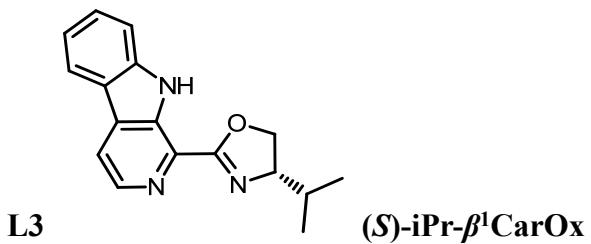
HRESI-MS: calcd for $\text{C}_{15}\text{H}_{14}\text{N}_3\text{O} [\text{M} + \text{H}]^+$: 252.1137, found: 252.1140.



White solid, ^1H -NMR (400 MHz, CDCl_3) δ 1.10 (t, $J = 7.36$ Hz, 3H, CH_2CH_3), 1.75 (m, 1H, CH_2CH_3), 1.87 (m, 1H, CH_2CH_3), 4.21 (dd, $J_1 = 7.96$ Hz, $J_2 = 8.00$ Hz, 1H, OCH_2CH), 4.46 (m, 1H, OCH_2CH), 4.64 (dd, $J_1 = 9.64$ Hz, $J_2 = 8.00$ Hz, 1H, OCH_2CH), 7.31 (m, 1H, *H in Benzene Ring*), 7.56-7.61 (m, 2H, *H in Benzene Ring*), 8.05 (d, $J = 5.10$ Hz, 1H, *H in Pyridine Ring*), 8.16 (dd, $J_1 = 7.92$ Hz, $J_2 = 1.04$ Hz, 1H, *H in Benzene Ring*), 8.54 (d, $J = 5.10$ Hz, 1H, *H in Pyridine Ring*), 10.4 (s, br, 1H, *NH*).

^{13}C NMR (100 MHz, CDCl_3) δ 10.3, 28.9, 68.3, 72.1, 112.0, 117.0, 120.3, 121.3, 121.9, 128.9, 129.1, 129.9, 135.9, 138.7, 140.6, 163.1.

HRESI-MS: calcd for $\text{C}_{16}\text{H}_{16}\text{N}_3\text{O} [\text{M} + \text{H}]^+$: 266.1293, found: 266.1576.

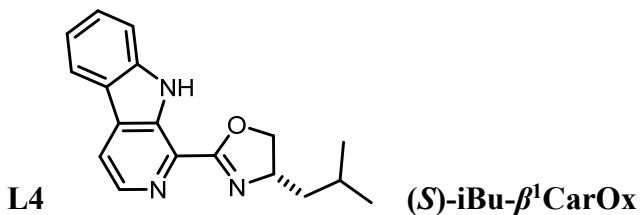


(S)-4-isopropyl-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

Pale yellow solid, ^1H -NMR (400 MHz, CDCl_3) δ 1.03 (d, $J = 6.72$ Hz, 3H, $\text{CH}(\text{CH}_3)_2$), 1.14 (d, $J = 6.72$ Hz, 3H, $\text{CH}(\text{CH}_3)_2$), 1.97 (m, 1H, $\text{CH}(\text{CH}_3)_2$), 4.28-4.35 (m, 2H, OCH_2CH), 4.59 (m, 1H, OCH_2CH), 7.32 (m, 1H, *H in Benzene Ring*), 7.58-7.64 (m, 2H, *H in Benzene Ring*), 8.07 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*), 8.16 (d, $J = 7.84$ Hz, 1H, *H in Benzene Ring*), 8.55 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*), 10.37 (s, br, 1H, *NH*).

^{13}C NMR (100 MHz, CDCl_3) δ 18.5, 19.1, 33.1, 70.2, 72.9, 112.0, 117.0, 120.3, 121.4, 121.9, 128.9, 129.0, 129.9, 135.9, 138.7, 140.6.

HRESI-MS: calcd for $\text{C}_{17}\text{H}_{18}\text{N}_3\text{O}$ [$\text{M} + \text{H}]^+$: 280.1483, found: 280.1450.



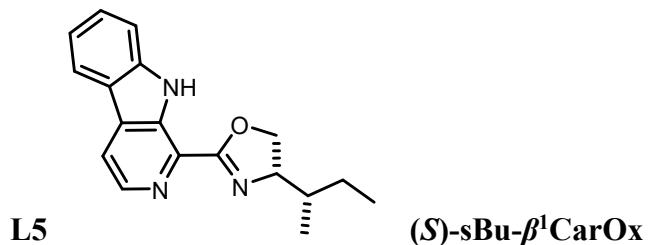
(S)-4-isobutyl-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

Pale yellow solid, ^1H -NMR (400 MHz, CDCl_3) δ 1.06 (s, 6H, $\text{CH}(\text{CH}_3)_2$), 1.51 (m, 1H, $\text{CH}(\text{CH}_3)_2$), 1.82 (m, 1H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 1.93 (m, 1H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 4.14 (m, 1H, OCH_2CH), 4.54 (m, 1H, OCH_2CH), 4.66 (m, 1H, OCH_2CH), 7.29 (m, 1H, *H in Benzene Ring*), 7.57-7.60 (m, 2H, *H in Benzene Ring*), 8.03 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*).

Ring), 8.13 (dd, $J_1 = 7.84$ Hz, $J_2 = 0.96$ Hz, 1H, H in Benzene Ring), 8.53 (d, $J = 5.08$ Hz, 1H, H in Pyridine Ring), 10.34 (s, br, 1H, NH).

¹³C NMR (100 MHz, CDCl₃) δ 22.8, 22.9, 25.8, 45.8, 65.4, 73.0, 112.0, 116.9, 120.3, 121.3, 121.9, 128.9, 129.1, 129.9, 135.9, 138.7, 140.6, 162.9.

HRESI-MS: calcd for C₁₈H₂₀N₃O [M+ H]⁺: 294.1606, found: 294.1610.

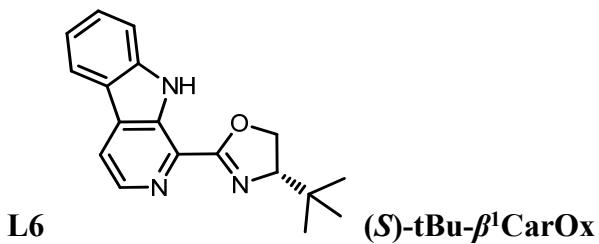


(*S*)-4-((*S*)-sec-butyl)-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

Yellow wax, $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 0.95 (d, $J = 6.72$ Hz, 3H, CH_2CH_3), 1.02 (t, $J = 7.40$ Hz, 3H, CH_2CH_3), 1.34 (m, 1H, CHCH_2CH_3), 1.72 (m, 1H, CHCH_2CH_3), 1.82 (m, 1H, CHCH_2CH_3), 4.29 (dd, $J_1 = 8.08$ Hz, $J_2 = 8.08$ Hz, 1H, OCH_2CH), 4.43 (m, 1H, OCH_2CH), 4.56 (dd, $J_1 = 9.48$ Hz, $J_2 = 8.08$ Hz, 1H, OCH_2CH), 7.31 (m, 1H, H in Benzene Ring), 7.56-7.61 (m, 2H, H in Benzene Ring), 8.04 (d, $J = 5.08$ Hz, 1H, H in Pyridine Ring), 8.14 (d, $J_1 = 7.20$ Hz, 1H, H in Benzene Ring), 8.53 (d, $J = 5.08$ Hz, 1H, H in Pyridine Ring), 10.36 (s, br, 1H, NH).

^{13}C NMR (100 MHz, CDCl_3) δ 11.7, 14.6, 26.3, 39.4, 69.7, 71.5, 112.0, 117.0, 120.3, 121.3, 121.9, 128.9, 129.0, 129.9, 135.9, 138.7, 140.6, 163.0.

HRESI-MS: calcd for C₁₈H₂₀N₃O [M+ H]⁺: 294.1606, found: 294.1611.

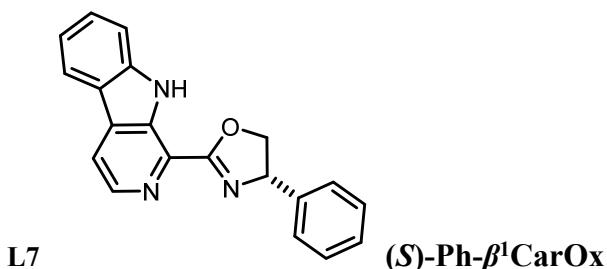


(S)-4-(tert-butyl)-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

White solid, ^1H -NMR (400 MHz, CDCl_3) δ 1.05 (s, 9H, $\text{C}(CH_3)_3$), 4.28 (dd, $J_1 = 10.00$ Hz, $J_2 = 7.92$ Hz, 1H, OCH_2CH), 4.40 (dd, $J_1 = 8.48$ Hz, $J_2 = 7.92$ Hz, 1H, OCH_2CH), 4.53 (dd, $J_1 = 10.00$ Hz, $J_2 = 8.48$ Hz, 1H, OCH_2CH), 7.32 (m, 1H, *H in Benzene Ring*), 7.59-7.60 (m, 2H, *H in Benzene Ring*), 8.07 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*), 8.16 (d, $J = 7.88$ Hz, 1H, *H in Benzene Ring*), 8.55 (d, $J = 5.08$ Hz, 1H, *H in Pyridine Ring*), 10.37 (s, br, 1H, *NH*).

^{13}C NMR (100 MHz, CDCl_3) δ 26.1 (3C), 34.1, 68.7, 76.4, 112.0, 117.0, 120.4, 121.4, 122.0, 128.9, 129.0, 129.9, 136.0, 138.7, 140.6, 163.1.

HRESI-MS: calcd for $\text{C}_{18}\text{H}_{20}\text{N}_3\text{O} [\text{M} + \text{H}]^+$: 294.1606, found: 294.1606.



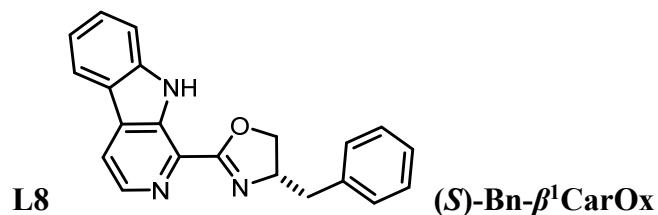
(S)-4-phenyl-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

Pale yellow solid, ^1H -NMR (400 MHz, CDCl_3) δ 4.43 (dd, $J_1 = 8.48$ Hz, $J_2 = 8.44$ Hz, 1H, OCH_2CH), 4.97 (dd, $J_1 = 10.08$ Hz, $J_2 = 8.44$ Hz, 1H, OCH_2CH), 5.61 ($J_1 = 10.08$ Hz, $J_2 = 8.48$ Hz, 1H, OCH_2CH), 7.28-7.38 (m, 2H, *H in Benzene Ring*), 7.38-7.48 (m, 4H, *H in Benzene Ring*), 7.55 (m, 2H, *H in Benzene Ring*), 8.07 (d, $J = 5.08$ Hz, 1H, *H*).

in Pyridine Ring), 8.16 (d, $J = 7.88$ Hz, 1H, H in Benzene Ring), 8.55 (d, $J = 5.08$ Hz, 1H, H in Pyridine Ring), 10.37 (s, br, 1H, NH).

^{13}C NMR (100 MHz, CDCl_3) δ 70.4, 74.8, 112.0, 117.3, 120.4, 121.3, 121.9, 126.9 (2C), 128.0, 128.6, 129.0 (2C), 129.1, 130.1, 136.1, 138.8, 140.6, 142.2, 164.5.

HRESI-MS: calcd for $\text{C}_{20}\text{H}_{16}\text{N}_3\text{O} [\text{M}+\text{H}]^+$: 314.1293, found: 314.1869.

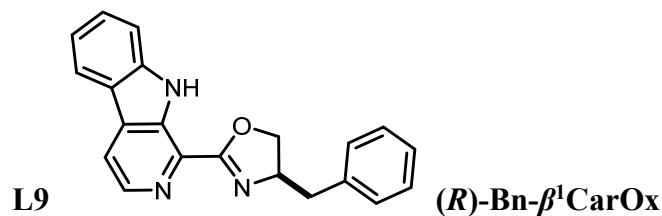


(S)-4-benzyl-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

Yellow oil, ^1H -NMR (400 MHz, CDCl_3) δ 2.92 (dd, $J_1 = 13.60$ Hz, $J_2 = 7.96$ Hz, 1H, $CH_2\text{Ph}$), 3.25 (dd, $J_1 = 13.60$ Hz, $J_2 = 6.56$ Hz, 1H, $CH_2\text{Ph}$), 4.32 (dd, $J_1 = 7.84$ Hz, $J_2 = 8.04$ Hz, 1H), 4.57 (dd, $J_1 = 9.12$ Hz, $J_2 = 8.04$ Hz, 1H), 4.79 (m, 1H), 7.29-7.35 (m, 4H, H in Benzene Ring), 7.35-7.39 (m, 2H, H in Benzene Ring), 7.55-7.62 (m, 2H, H in Benzene Ring), 8.07 (d, $J = 5.08$ Hz, 1H, H in Pyridine Ring), 8.16 (d, $J = 7.88$ Hz, 1H, H in Benzene Ring), 8.55 (d, $J = 5.08$ Hz, 1H, H in Pyridine Ring), 10.34 (s, br, 1H, NH).

^{13}C NMR (125 MHz, CDCl_3) δ 42.4, 68.2, 72.1, 111.9, 117.1, 120.4, 121.3, 122.0, 126.7, 128.7, 128.9, 129.0, 129.3, 130.0, 136.0, 138.1, 138.7, 140.7, 163.6.

HRESI-MS: calcd for $\text{C}_{21}\text{H}_{18}\text{N}_3\text{O} [\text{M}+\text{H}]^+$: 328.1450, found: 328.1448.

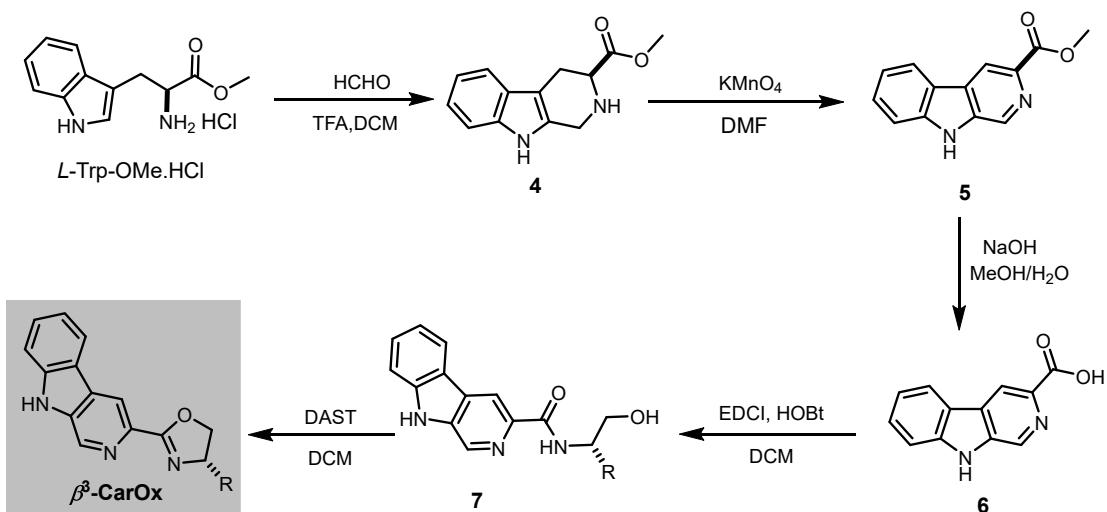


(R)-4-benzyl-2-(9H-pyrido[3,4-b]indol-1-yl)-4,5-dihydrooxazole

Yellow oil, ^1H -NMR (400 MHz, CDCl_3) δ 2.92 (dd, $J_1 = 13.60$ Hz, $J_2 = 7.96$ Hz, 1H, CH_2Ph), 3.25 (dd, $J_1 = 13.60$ Hz, $J_2 = 6.56$ Hz, 1H, CH_2Ph), 4.32 (dd, $J_1 = 7.84$ Hz, $J_2 = 8.04$ Hz, 1H), 4.56 (dd, $J_1 = 9.12$ Hz, $J_2 = 8.04$ Hz, 1H), 4.78 (m, 1H), 7.27-7.35 (m, 4H, *H* in Benzene Ring), 7.35-7.40 (m, 2H, *H* in Benzene Ring), 7.54-7.63 (m, 2H, *H* in Benzene Ring), 8.07 (d, $J = 5.08$ Hz, 1H, *H* in Pyridine Ring), 8.17 (dd, $J_1 = 7.08$ Hz, $J_1 = 1.08$ Hz, 1H, *H* in Benzene Ring), 8.55 (d, $J = 5.08$ Hz, 1H, *H* in Pyridine Ring), 10.34 (s, br, 1H, *NH*).
 ^{13}C NMR (100125 MHz, CDCl_3) δ 42.3, 68.2, 72.1, 111.9, 117.1, 120.4, 121.3, 122.0, 126.7, 128.7, 128.9, 129.0, 129.3, 130.0, 136.0, 138.1, 138.7, 140.7, 163.6.
HRESI-MS: calcd for $\text{C}_{21}\text{H}_{18}\text{N}_3\text{O} [\text{M}+\text{H}]^+$: 328.1450, found: 328.1448.

Synthesis and Structural Elucidation of β^3 -CarOx Ligands

Synthetic Route to the Chiral β^3 -CarOx Ligands



Step 1, To a stirred suspension of *L*-Tryptophan methyl ester hydrochloride (5.1 g, 20

mmol) and 37% formalin (40 mL) was added anhydrous dichloromethane. Trifluoroacetic acid (TFA) (3.26 mL) was added dropwise at 0 °C. The reaction mixture was stirred until the full consumption of the starting material detected by TLC. The mixture was quenched by the addition of 10% aqueous potassium carbonate solution (20 mL). The water phase was extracted with ethyl acetate (20 mL × 3), dried over anhydrous sodium sulfate, and concentrated under vacuum. This crude (*S*)- β -1,2,3,4-tetrahydrocarboline -3-carboxylic acid methyl ester **4** (91% yield) was used for the next step without further purification.

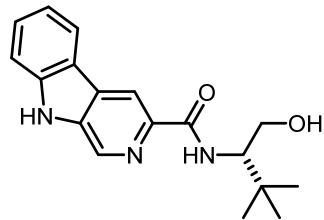
Step 2, To a stirred solution of (*S*)- β -1,2,3,4-tetrahydrocarboline-3-carboxylic acid methyl ester **4** (4.6 g, 20 mmol) in DMF (60 mL) was added potassium permanganate (KMnO₄) (2.3 g, 20 mmol) in portions while the mixture was in an ice bath. The reaction mixture was stirred vigorously and allowed to gradually warm to room temperature until the full consumption of the starting material was detected by TLC. The heterogeneous mixture was filtered and rinsed with distilled methanol. The β -carboline-3-carboxylic acid methyl ester, compound **5**, was collected as yellow solid in 62% yield (2.8 g). Compound **5** was used for the next step without further purification.

Step 3, To a stirred suspension of β -carboline-3-carboxylic acid methyl ester **5** (2.80 g, 12 mmol) was added methanol (30 mL). Aqueous sodium hydroxide solution (12 mL, 1 M) was added dropwise at 0 °C. The heterogeneous mixture was immersed in a preheated oil bath (50 °C) and stirred until the full consumption of the starting material was detected by TLC. The mixture was concentrated under vacuum to remove methanol and was adjusted to pH 6-7 with hydrochloric acid to precipitate the desired product, the heterogeneous mixture was filtered to give β -carboline-1-carboxylic acid intermediate **6** as yellow solid in 92% yield (2.34 g). This intermediate was used for the next step without further purification.

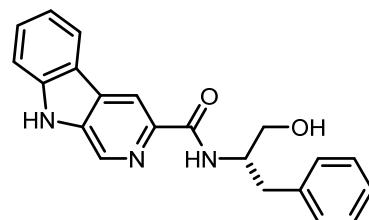
Step 4, General procedure for Steglich Condensation: To a dried Schlenk flask charged with the synthesized β -carboline-3-carboxylic acid, compound **6**, (1 mmol, 212 mg) and the specific chiral amino alcohol (1 mmol), was added anhydrous dichloromethane (5 mL) for dissolution. Hydroxybenzotriazole (HOEt) (175 mg, 1.3

mmol) and *N*-(3-(dimethyl amino)propyl)-*N'*-ethylcarbodiimide hydrochloride (EDCI-HCl) (0.25 g, 1.3 mmol) were then added while the reaction flask was in an ice bath. The mixture was allowed to gradually warm to room temperature, and it was stirred overnight until full consumption of the carboxylic acid was detected by thin layer chromatography (TLC). The mixture was quenched by the addition of a saturated aqueous solution of NaHCO₃ (20 mL) and separated. The water phase was extracted with dichloromethane (10 mL × 3), and the combined organic phase was sequentially washed with water (10 mL × 2) and saturated aqueous NaCl (10 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum. Purification by silica gel column chromatography on silica gel (200-300m) with hexane/EtOAc (2:1, v/v) as the eluent gave the amide intermediate 7.

Yields for the intermediate 7 are listed below,



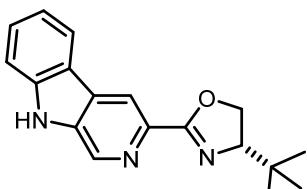
78%



73%

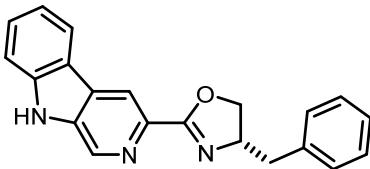
Step 4, General procedure for the DAST mediated cyclization to produce β^3 -CarOx Ligand: To a Schlenk tube charged the amide intermediate 7 (1.0 mmol) was added anhydrous DCM (5.0 mL) under N₂ atmosphere. Diethylaminosulfur trifluoride (DAST) (160 mg, 1mmol) was added dropwise at -78 °C until the full consumption of the starting material was detected by TLC. The mixture was quenched by the addition of a saturated aqueous solution of NaHCO₃ (10 mL) and separated, The water phase was extracted with dichloromethane (10 mL × 3), and the combined organic phase was sequentially washed with water (10 mL × 2) and saturated aqueous NaCl (10 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum. Purification by silica gel column chromatography on silica gel (200-300m) with hexane/EtOAc (2:1, v/v) as the eluent gave the product β^3 -CarOx Ligand.

Yields for β^3 -CarOx ligands are listed below,



L10: (S)-tBu- β^3 CarOx

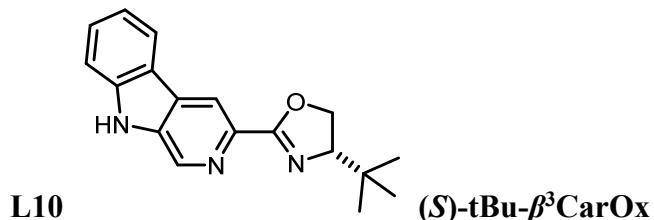
72%



L11: (S)-Bn- β^3 CarOx

67%

Characterization of the Synthesized Chiral β^3 -CarOx Ligands



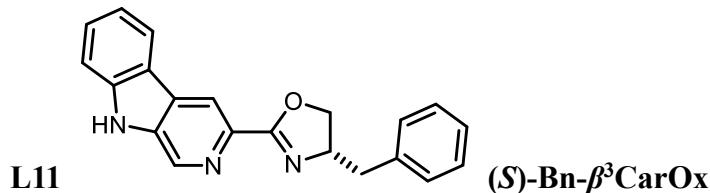
(S)-tBu- β^3 CarOx

(S)-4-(tert-butyl)-2-(9H-pyrido[3,4-b]indol-3-yl)-4,5-dihydrooxazole

White solid, 1 H-NMR (500 MHz, CDCl₃) δ 1.00 (s, 9H, C(CH₃)₃), 4.18 (dd, J_1 = 10.05 Hz, J_2 = 8.05 Hz, 1H, OCH₂CH), 4.36 (t, J = 8.25 Hz, 1H, OCH₂CH), 4.50 (dd, J_1 = 9.95 Hz, J_2 = 8.40 Hz, 1H, OCH₂CH), 7.31 (m, 1H, *H* in Benzene Ring), 7.53-7.56 (m, 2H, *H* in Benzene Ring), 8.14 (d, J = 9.90 Hz, 1H, *H* in Benzene Ring), 8.78 (s, 1H, *H* in Pyridine Ring), 8.94 (s, 1H, *H* in Pyridine Ring), 9.75 (s, br, 1H, NH).

13 H-NMR (125 MHz, CDCl₃) δ 26.0, 34.1, 63.3, 76.4, 112.0, 116.2, 120.6, 121.5, 122.0, 128.9, 129.1, 133.3, 136.1, 136.9, 140.9, 163.3.

HRESI-MS: calcd for C₁₈H₂₀N₃O [M+ H]⁺: 294.1606, found: 294.1606.



(S)-Bn- β^3 CarOx

(S)-4- Benzyl-2-(9H-pyrido[3,4-b]indol-3-yl)-4,5-dihydrooxazole

White solid, 1 H-NMR (500 MHz, CDCl₃) δ 2.80 (dd, J_1 = 9.15 Hz, J_2 = 13, 85 Hz, 1H,

*CH₂Ph), 3.34 (dd, *J*₁ = 5.05 Hz, *J*₂ = 13.80 Hz, 1H, *CH₂Ph*), 4.28 (t, *J* = 7.95Hz, 1H), 4.49 (t, *J* = 9.25 Hz, 1H), 4.73 (m, 1H), 7.17-7.20 (s, 1H, *H* in Benzene Ring), 7.25-7.32 (m, 5H, *H* in Benzene Ring), 7.52-7.56 (m, 2H, *H* in Benzene Ring), 8.12 (d, *J* = 7.90 Hz, 1H, *H* in Benzene Ring), 8.78 (s, 1H, *H* in Pyridine Ring), 8.98 (s, 1H, *H* in Pyridine Ring), 10.20 (s, br, 1H, *NH*).*

¹³H-NMR (125 MHz, CDCl₃) δ 42.0, 68.0, 72.5, 112.1, 116.2, 120.5, 121.5, 121.9, 126.6, 128.6, 128.8, 129.0, 129.2, 133.6, 135.7, 137.1, 137.9, 141.4, 164.4.

HRESI-MS: calcd for C₂₁H₁₈N₃O [M+H]⁺: 328.1451, found: 328.1349.

Enantioselective Michael Addition of Arylboronic Acids to Nitroalkenes:

General procedure



To a Schlenk tube charged Pd(TFA)₂ (4.15 mg, 0.0125 mmol) and the specific chiral *β*-CarOx ligand (0.01875 mmol) was added MeOH (1.0 mL) under N₂ atmosphere. The mixture was stirred at 40 °C for 0.5 h to afford the catalyst solution.

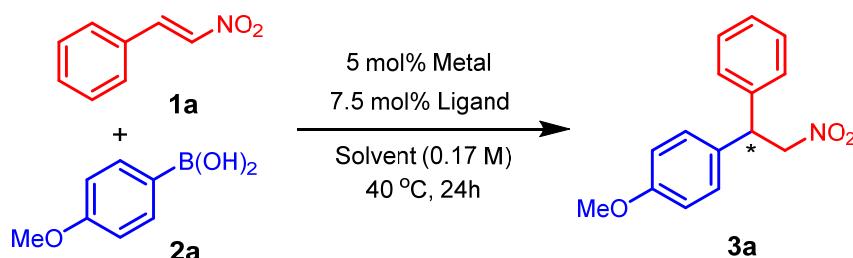
To the above solution was added nitrostyrene **1** (0.25 mmol) and aryl boronic acid **2** (0.375 mmol). The wall of the tube was rinsed with MeOH (0.5 mL) or some oil substrate was dissolved in MeOH (0.5 mL) (The volume of solvent is 1.5 mL). The tube was placed in the modules of the reactor which was set at 40 °C. After stirring for 24 h, the reaction mixture was cooled to room temperature, and the solvent was removed by

rotary evaporation. The residue was purified by column chromatography (petroleum/ether/EtOAc = 20/1, v/v) to give the product.

2-(isoquinolin-3-yl)-4,5-dihydrooxazole was utilized as a ligand for the preparation of the racemic products.

Optimization of the Reaction Conditions

Table 1. Optimization of the Reaction Conditions ^a

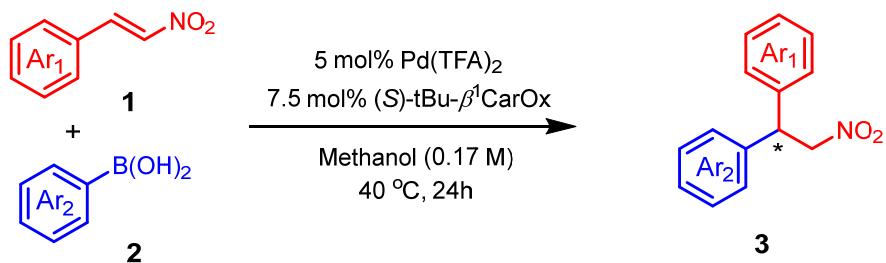


Entry	Metal	Ligand	Solvent	Isolated Yield (%) ^b	ee (%) ^c
1	Pd(TFA) ₂	L₁	MeOH	81	80 (<i>S</i>)
2	Pd(TFA) ₂	L₂	MeOH	82	86 (<i>S</i>)
3	Pd(TFA) ₂	L₃	MeOH	88	89 (<i>S</i>)
4	Pd(TFA) ₂	L₄	MeOH	78	87 (<i>S</i>)
5	Pd(TFA) ₂	L₅	MeOH	78	87 (<i>S</i>)
6	Pd(TFA) ₂	L₆	MeOH	92	95 (<i>S</i>)
7	Pd(TFA) ₂	L₇	MeOH	86	85 (<i>S</i>)
8	Pd(TFA) ₂	L₈	MeOH	84	89 (<i>S</i>)
9	Pd(TFA) ₂	L₉	MeOH	81	88 (<i>R</i>)
10	Pd(TFA) ₂	L₁₀	MeOH	89	80 (<i>S</i>)
11	Pd(TFA) ₂	L₁₁	MeOH	85	73 (<i>S</i>)
12	Pd(OAc) ₂	L₆	MeOH	49	83 (<i>S</i>)
13	PdCl ₂	L₆	MeOH	<5	n.d.
14	Pd(TFA) ₂	L₆	EtOH	82	92 (<i>S</i>)
15	Pd(TFA) ₂	L₆	ⁱ Pr-OH	51	73 (<i>S</i>)
16	Pd(TFA) ₂	L₆	^t Bu-OH	64	75 (<i>S</i>)
17	Pd(TFA) ₂	L₆	2-Methyl-2-butanol	58	66 (<i>S</i>)
18	Pd(TFA) ₂	L₆	TFE	51	71 (<i>S</i>)

^a, unless otherwise mentioned, reactions were carried out on a 0.25 mmol of **2a**, 0.375 mmol of *para*-MeO-C₆H₄B(OH)₂ using 5 mol % Pd(TFA)₂ and 7.5 mol % ligand in 1.5 mL solvent at 40 °C for 24 h under N₂ atmosphere.

^b, Isolated yield. ^c, Determined by HPLC using a Daicel column (OD-H). The absolute configuration was assigned by comparing the retention time of **3aa** with that reported in the literature.^[3] n.d. = not determined.

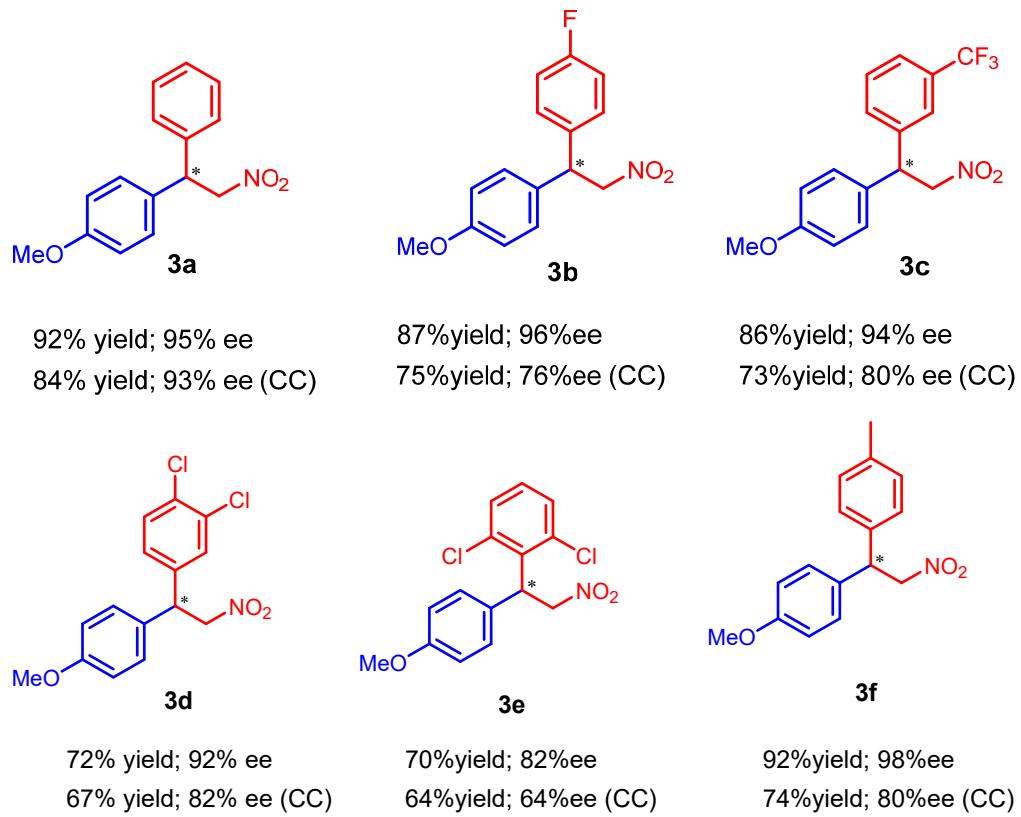
Substrate Scope

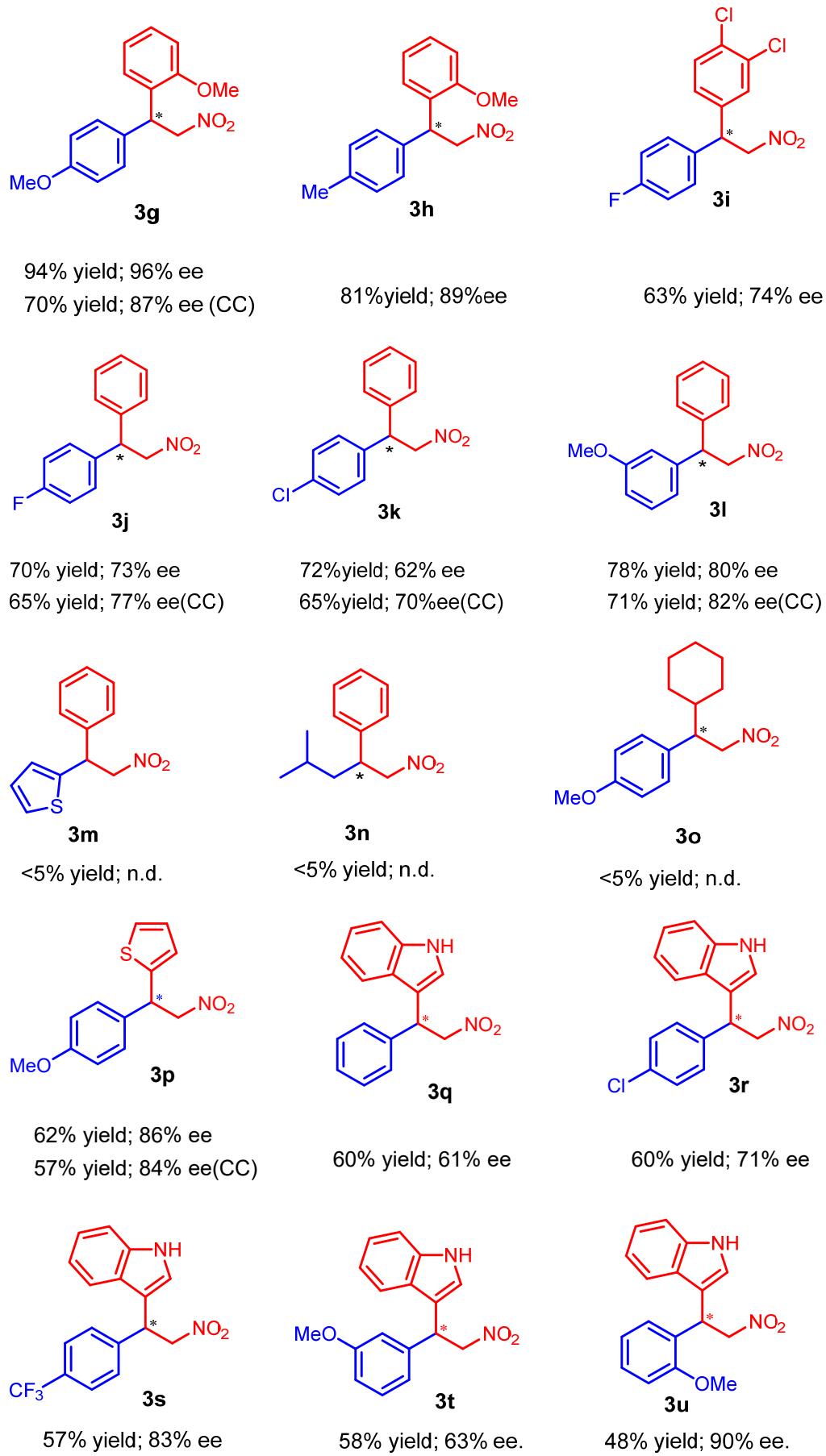


NOTE:

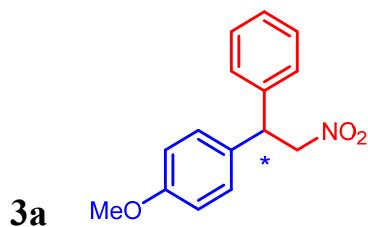
The *nitroalkenes* and *arylboronic acids* were highlighted by red and blue, respectively.

Data marked (CC) means that they were reported in “*Chem. Commun.* 2019, 55, 5902–5905.”^[4]





HPLC traces of the Enantioenriched β -aryl nitroethanes



Colorless oil, 92% yield.

The NMR data is in accordance with that of previous publications.^[4-7]

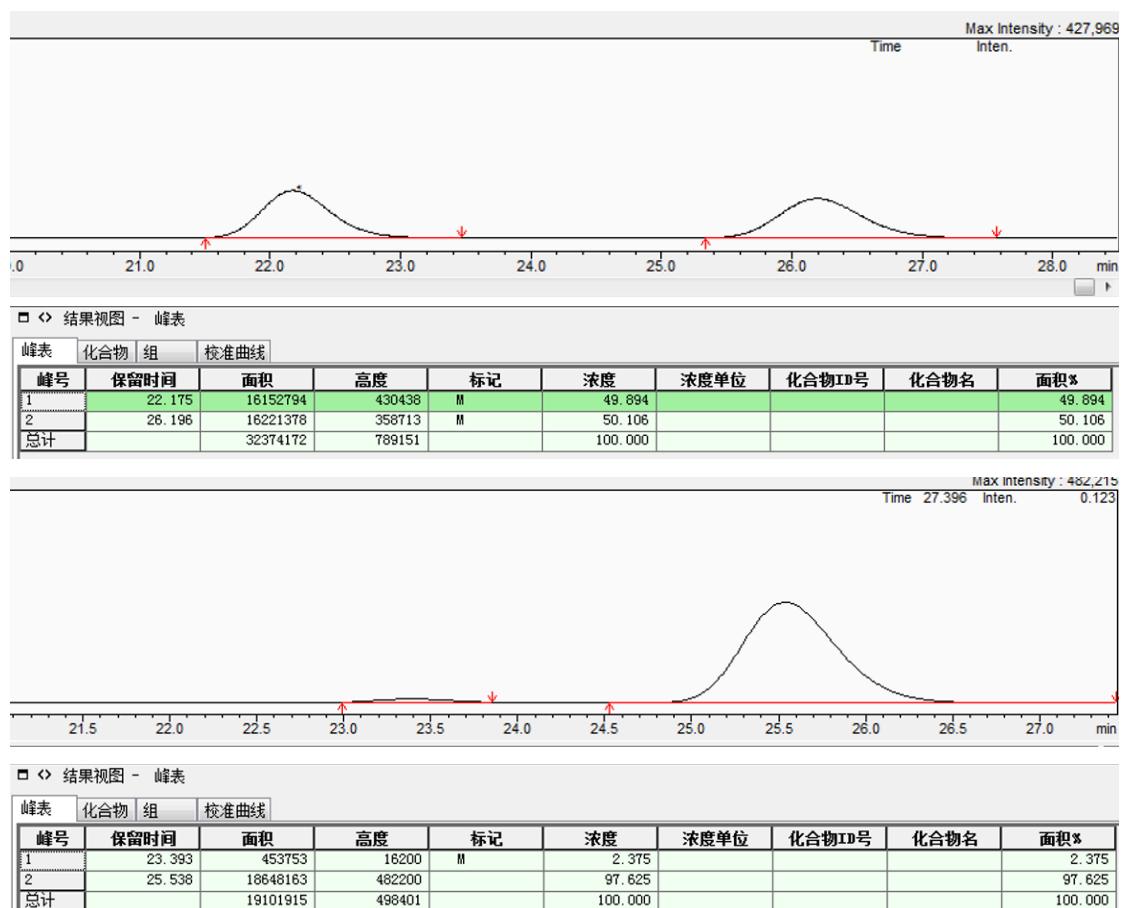
^1H NMR (400 MHz, CDCl_3), δ 3.78 (s, 3H, OCH_3), 4.86 (m, 1H), 4.94-4.96 (m, 2H), 6.84-6.87(m, 2H, Aromatic H), 7.13-7.18 (m, 2H, Aromatic H), 7.20-7.24 (m, 2H, Aromatic H), 7.26 (m, 1H, Aromatic H), 7.29-7.36 (m, 2H, Aromatic H).

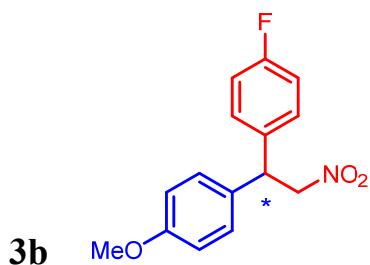
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 80/20, 220 nm, 1.0 mL/min.

$t_{\text{R}1}$ = 23.4 min (major), $t_{\text{R}2}$ = 25.5 min (minor)];

ee = 95.3%.





Colorless oil, 87% yield.

The NMR data is in accordance with that of previous publications.^[4-5]

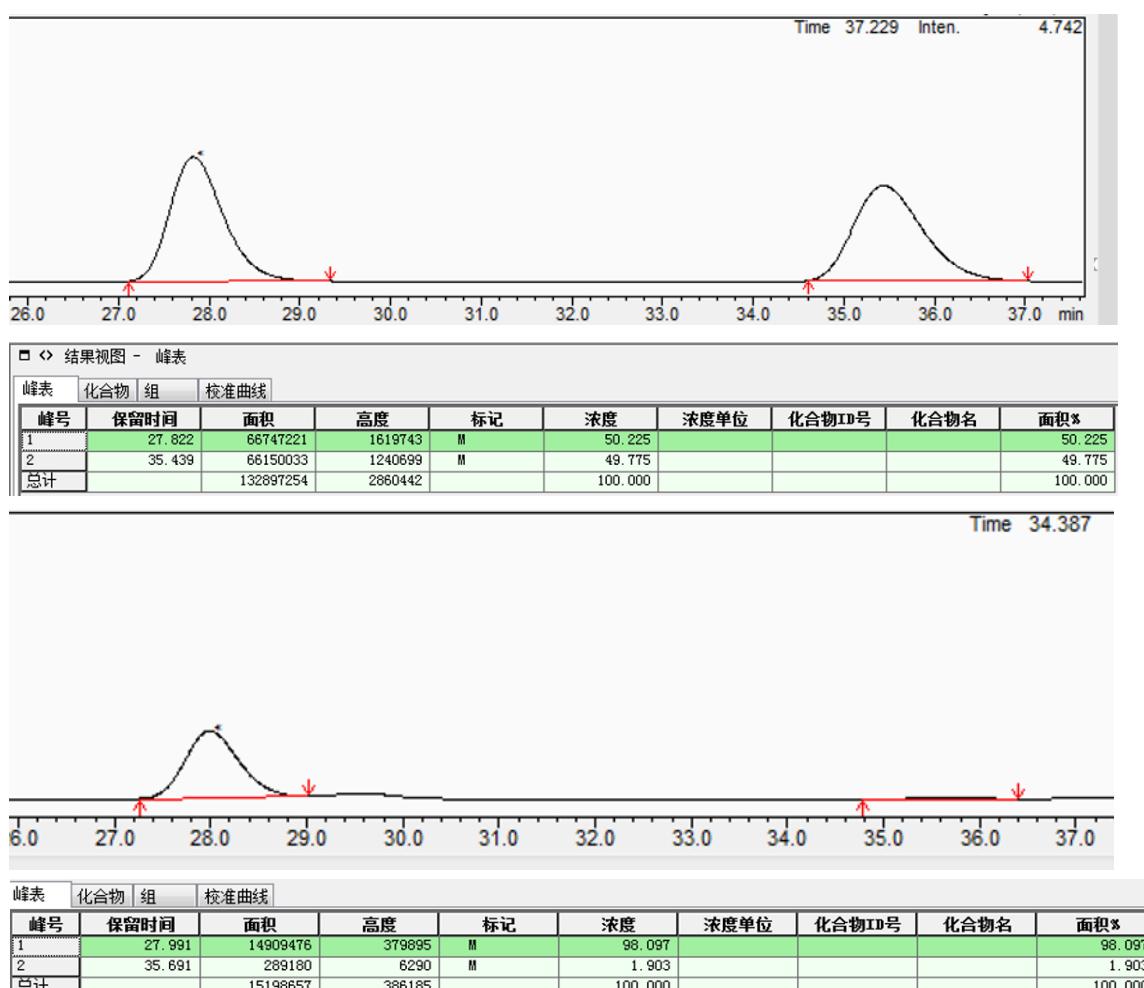
¹H NMR (400 MHz, CDCl₃), δ 3.78 (s, 3H, OCH₃), 4.84 (m, 1H), 4.89-4.94 (m, 2H), 6.84-6.89 (m, 2H, Aromatic H), 6.99-7.04 (m, 2H, Aromatic H), 7.10-7.15 (m, 2H, Aromatic H), 7.17-7.21 (m, 2H, Aromatic H).

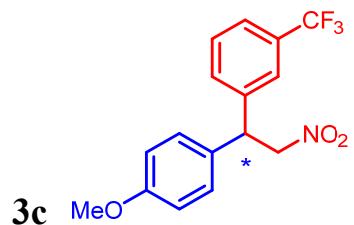
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 60/40, 220 nm, 1.0 mL/min.

t_{R1} = 28.0 min (major), t_{R2} = 35.7 min (minor)

ee = 96.2%.





Colorless oil, 86% yield.

The NMR data is in accordance with that of previous publication.^[4]

¹H NMR (400 MHz, CDCl₃), δ 3.79 (s, 3H, OCH₃), 4.92 (m, 1H), 4.95-5.00 (m, 2H), 6.86-6.91 (m, 2H, Aromatic H), 7.12-7.16 (m, 2H, Aromatic H), 7.41-7.50 (m, 3H, Aromatic H), 7.53 (d, *J* = 7.16 Hz, 1H, Aromatic H).

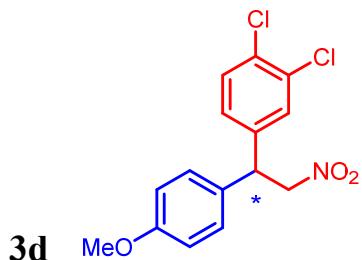
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 60/40, 220 nm, 1.0 mL/min.

t_{R1} = 19.7 min (major), t_{R2} = 23.3 min (minor)

ee = 94.4%.





Colorless oil, 72% yield.

The NMR data is in accordance with that of previous publication.^[4]

¹H NMR (400 MHz, CDCl₃), δ 3.79 (s, 3H, OCH₃), 4.82 (dd, *J*₁ = 8.76 Hz, *J*₂ = 7.20 Hz, 1H), 4.92-4.95 (m, 2H), 6.85-6.89 (m, 2H, Aromatic H), 7.08 (dd, *J*₁ = 8.36 Hz, *J*₂ = 2.24 Hz, 1H), 7.09-7.13 (m, 2H, Aromatic H), 7.31 (d, *J* = 2.2 Hz, 1H, Aromatic H), 7.40 (d, *J* = 8.28 Hz, 1H, Aromatic H).

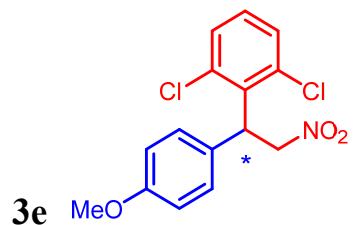
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 0.8 mL/min.

t_{R1} = 35.2 min (major), t_{R2} = 37.1 min (minor);

ee = 92.4%.





Colorless oil, 70% yield.

The NMR data is in accordance with that of previous publication.^[4]

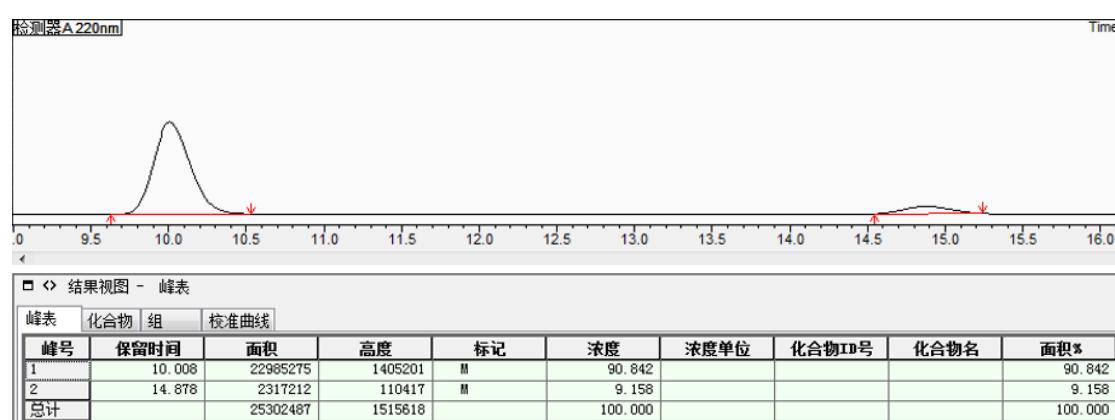
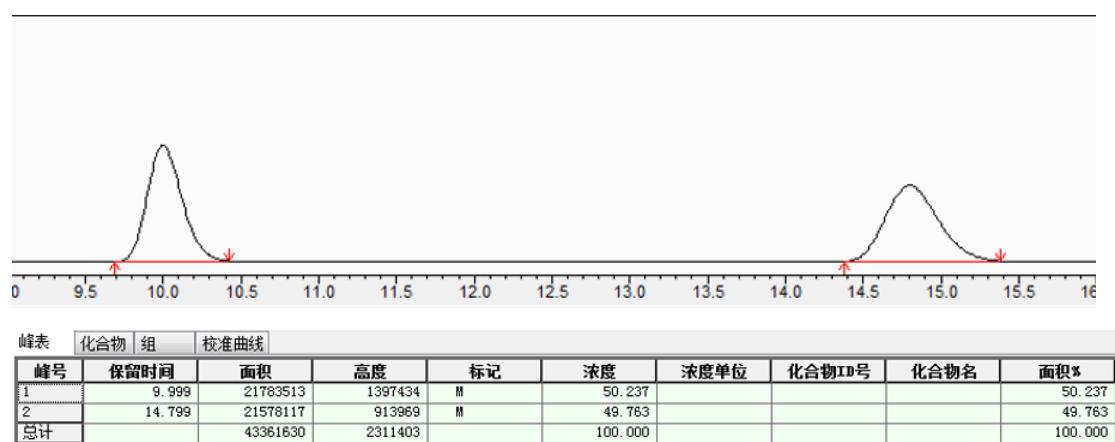
¹H NMR (400 MHz, CDCl₃), δ 3.78 (s, 3H, OCH₃), 5.26 (dd, *J*₁ = 13.28 Hz, *J*₂ = 7.44 Hz, 1H), 5.42 (dd, *J*₁ = 13.28 Hz, *J*₂ = 7.44 Hz, 1H), 5.95 (dd, *J*₁ = *J*₂ = 7.44 Hz, 1H), 6.82-6.86 (m, 2H, Aromatic H), 7.08-7.14 (m, 2H, Aromatic H), 7.18 (dd, *J*₁ = *J*₂ = 8.28 Hz, 1H, Aromatic H), 7.28-7.40 (m, 2H, Aromatic H).

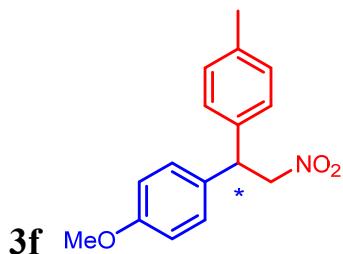
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 80/20, 220 nm, 1.0 mL/min.

t_{R1} = 10.0 min (major), t_{R2} = 14.9 min (minor);

ee = 81.7%.





Colorless oil, 92% yield.

The NMR data is in accordance with that of previous publications.^[4, 6]

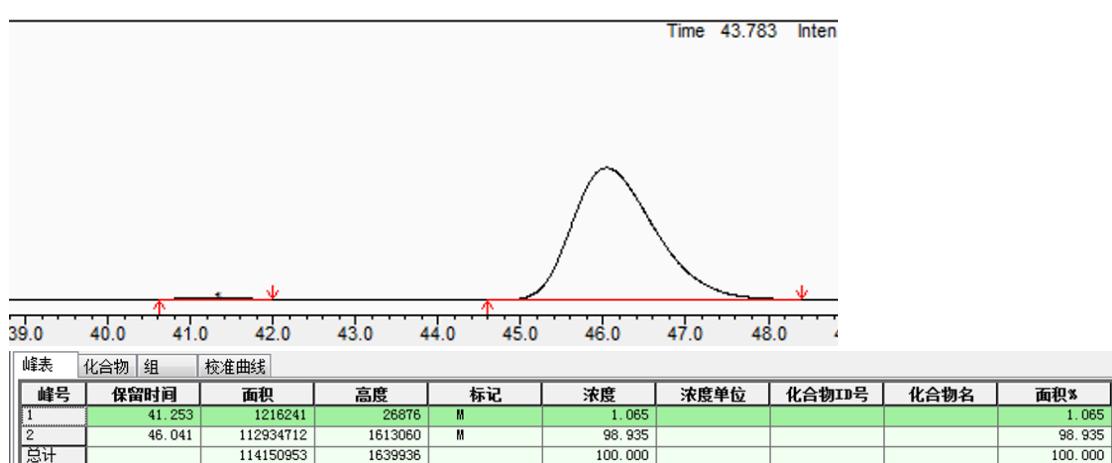
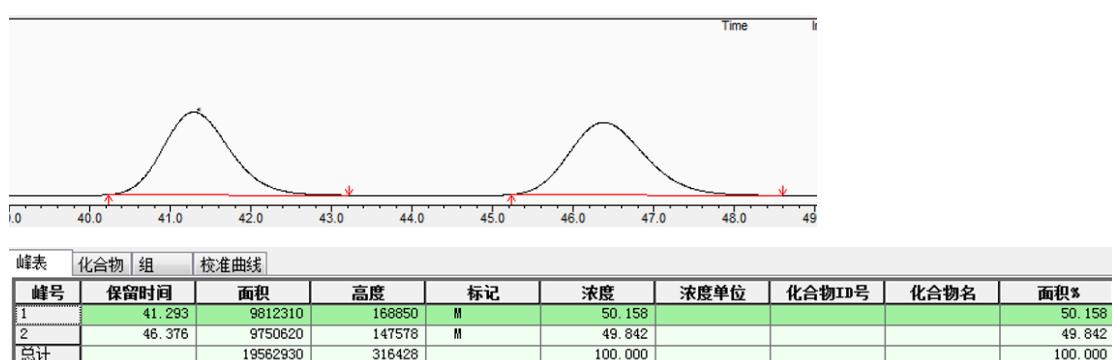
¹H NMR (400 MHz, CDCl₃), δ 2.31 (s, 3H, CH₃), 3.77 (s, 3H, OCH₃), 4.82 (dd, *J*₁ = *J*₂ = 8.16 Hz, 1H), 4.92-4.94 (m, 2H), 6.82-6.87 (m, 2H, Aromatic H), 7.09-7.17 (m, 6H, Aromatic H).

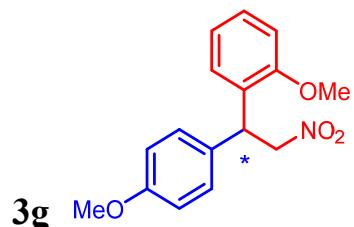
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 80/20, 220 nm, 1.0 mL/min.

t_{R1} = 41.3 min (major), t_{R2} = 46.0 min (minor)];

ee = 97.9%.





Colorless oil, 94% yield.

The NMR data is in accordance with that of previous publication.^[4]

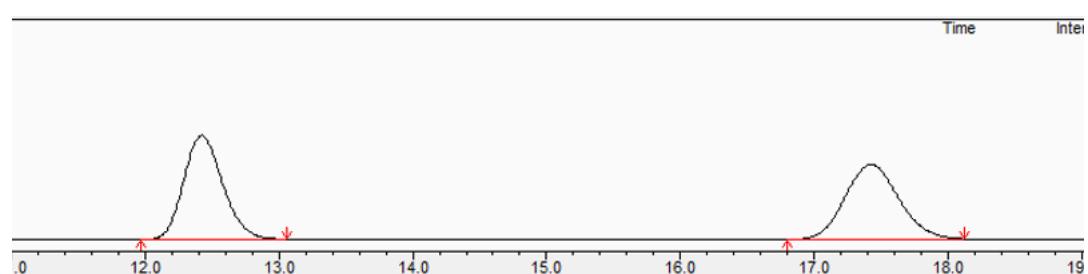
¹H NMR (400 MHz, CDCl₃), δ 3.77 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 4.91 (dd, *J*₁ = 12.76 Hz, *J*₂ = 9.32 Hz, 1H), 5.01 (dd, *J*₁ = 12.76 Hz, *J*₂ = 6.84 Hz, 1H), 5.21 (dd, *J*₁ = 9.32 Hz, *J*₂ = 6.84 Hz, 1H), 6.83-6.87 (m, 2H, Aromatic H), 6.87-6.92 (m, 2H, Aromatic H), 7.05 (dd, *J*₁ = 7.44 Hz, *J*₂ = 1.64 Hz, 1H, Aromatic H), 7.17-7.21 (m, 2H, Aromatic H), 7.22 (dd, *J*₁ = 7.72 Hz, *J*₂ = 1.60 Hz, 1H, Aromatic H).

HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 80/20, 220 nm, 1.0 mL/min.

t_{R1} = 12.5 min (major), t_{R2} = 17.5 min (minor)];

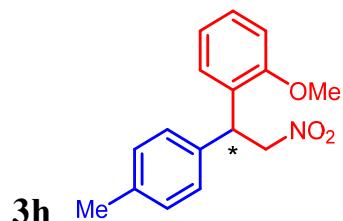
ee = 95.7%.



峰表	化合物	组	校准曲线						
峰号	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1	12.424	33190704	1638671	M	50.169				50.169
2	17.424	32966899	1178456	M	49.831				49.831
总计		66157602	2817126		100.000				100.000



峰表	化合物	组	校准曲线						
峰号	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1	12.451	66673023	3240052	M	97.897				97.897
2	17.444	1432411	64390	M	2.103				2.103
总计		68105435	3304442		100.000				100.000



Colorless oil, 81% yield.

The NMR data is in accordance with that of previous publications.^[6, 8]

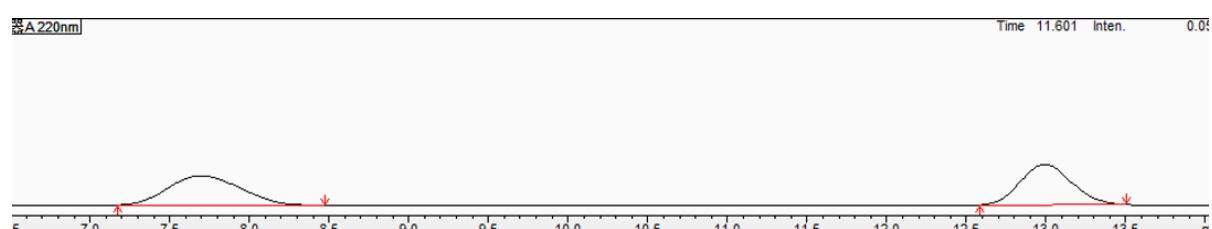
¹H NMR (400 MHz, CDCl₃), δ 2.31 (s, 3H, CH₃), 3.83 (s, 3H, OCH₃), 4.90-5.04 (m, 2H), 5.23(m, 1H), 6.88 (t, *J*=8.60 Hz, 2H, Aromatic H), 7.05 (m, 1H, Aromatic H), 7.11(d, *J*=8.12 Hz, 2H, Aromatic H), 7.16 (d, *J*=8.16 Hz, 2H, Aromatic H), 7.23(m, 1H, Aromatic H).

HPLC trace:

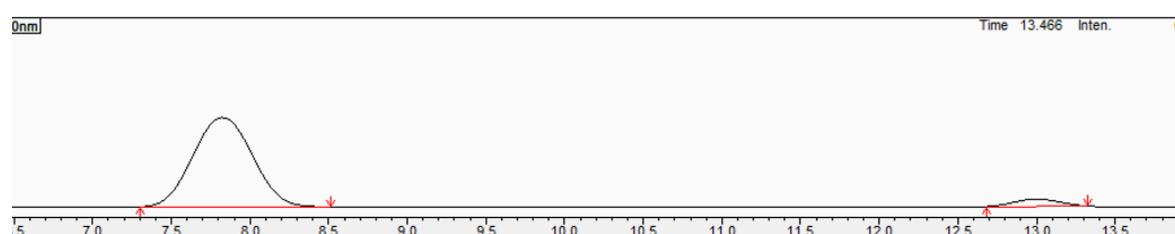
Daicel chiralcel OD-H, hexane/i-PrOH = 90/10, 220 nm, 1.5 mL/min.

t_{R1} = 7.83 min (major), t_{R2} = 13.0 min (minor)

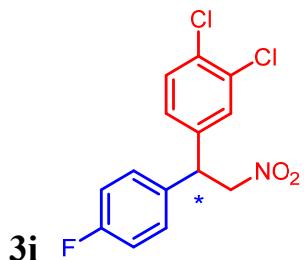
ee = 88.8%.



峰表	化合物	组	校准曲线						
峰号	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1	7.896	5910017	187268	M	50.460				50.460
2	12.991	5802165	258236	M	49.540				49.540
总计		11712182	445524		100.000				100.000



峰表	化合物	组	校准曲线						
峰号	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1	7.825	8958936	342260	M	94.374				94.374
2	13.001	534097	27046	M	5.626				5.626
总计		9493032	369306		100.000				100.000



Colorless oil, 63% yield.

¹H NMR (500 MHz, CDCl₃), δ 4.85 (t, *J*=7.75 Hz, 1H), 4.91-4.93 (m, 2H), 7.03-7.08 (m, 3H, Aromatic H), 7.16-9.19 (m, 2H, Aromatic H), 7.30 (s, 1H, Aromatic H), 7.41 (d, *J*=8.15 Hz, 1H, Aromatic H).

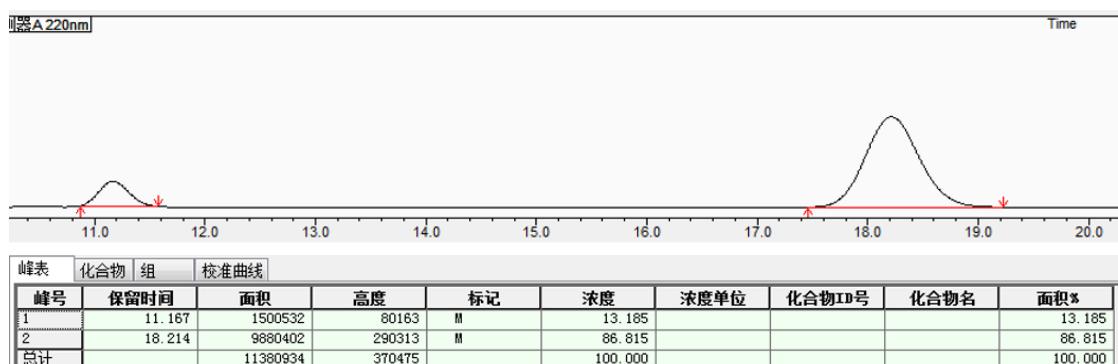
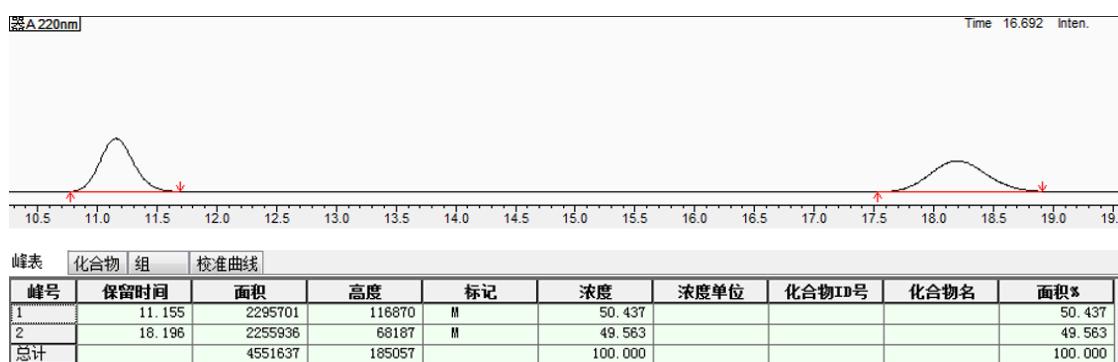
¹³C NMR (125 MHz, CDCl₃), δ 47.3, 78.7, 116.3, 116.4, 126.8, 129.2, 129.3, 129.7, 131.1, 132.9, 133.3, 139.2, 161.3, 163.3.

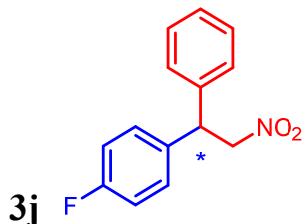
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.2 mL/min.

t_{R1} = 18.2 min (major), t_{R2} = 11.2 min (minor)

ee = 73.6%.





Colorless oil, 70% yield.

The NMR data is in accordance with that of previous publications.^[4, 8]

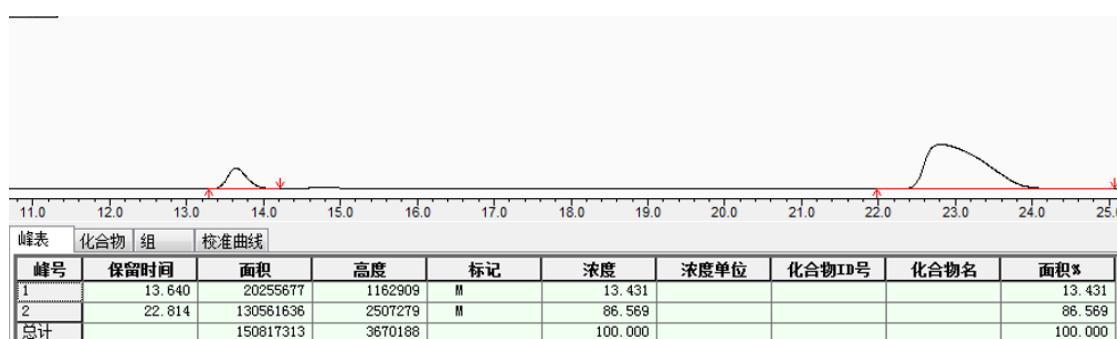
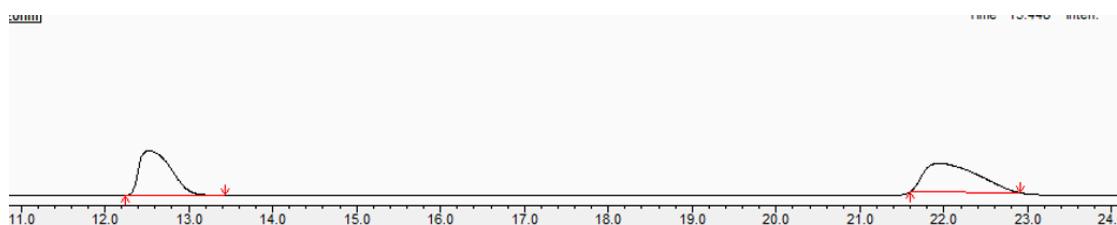
¹H NMR (400 MHz, CDCl₃), δ 4.88 (m, 1H), 4.91-4.96 (m, 2H), 6.96-7.04 (m, 2H, Aromatic H), 7.16-7.23 (m, 4H, Aromatic H), 7.27 (m, 1H, Aromatic H), 7.29-7.36 (m, 2H, Aromatic H).

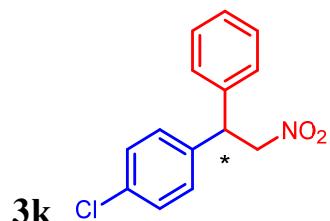
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

t_{R1} = 13.6min (minor), t_{R2} = 22.8 min (major)

ee = 73.1%.





Colorless oil, 72% yield.

The NMR data is in accordance with that of previous publications.^[4, 6, 8]

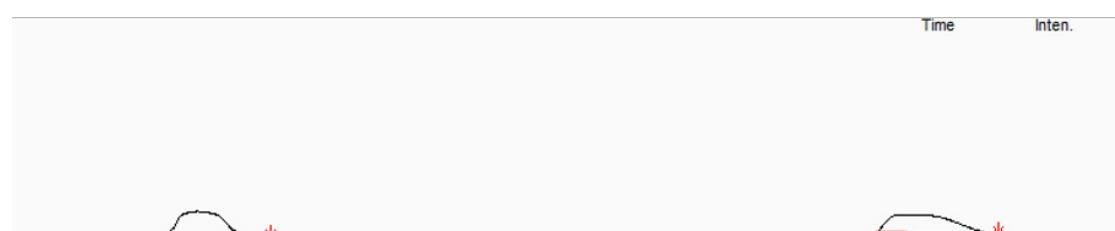
¹H NMR (400 MHz, CDCl₃), δ 4.88 (m, 1H), 4.92-4.98 (m, 2H), 7.16-7.23 (m, 4H, Aromatic H), 7.27-7.36 (m, 5H, Aromatic H).

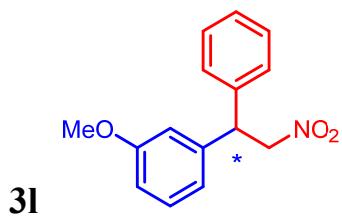
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

t_{R1} = 17.6 min (minor), t_{R2} = 28.0 min (major)

ee = 61.8%.





Colorless oil, 78% yield.

The NMR data is in accordance with that of previous publications.^[4, 6]

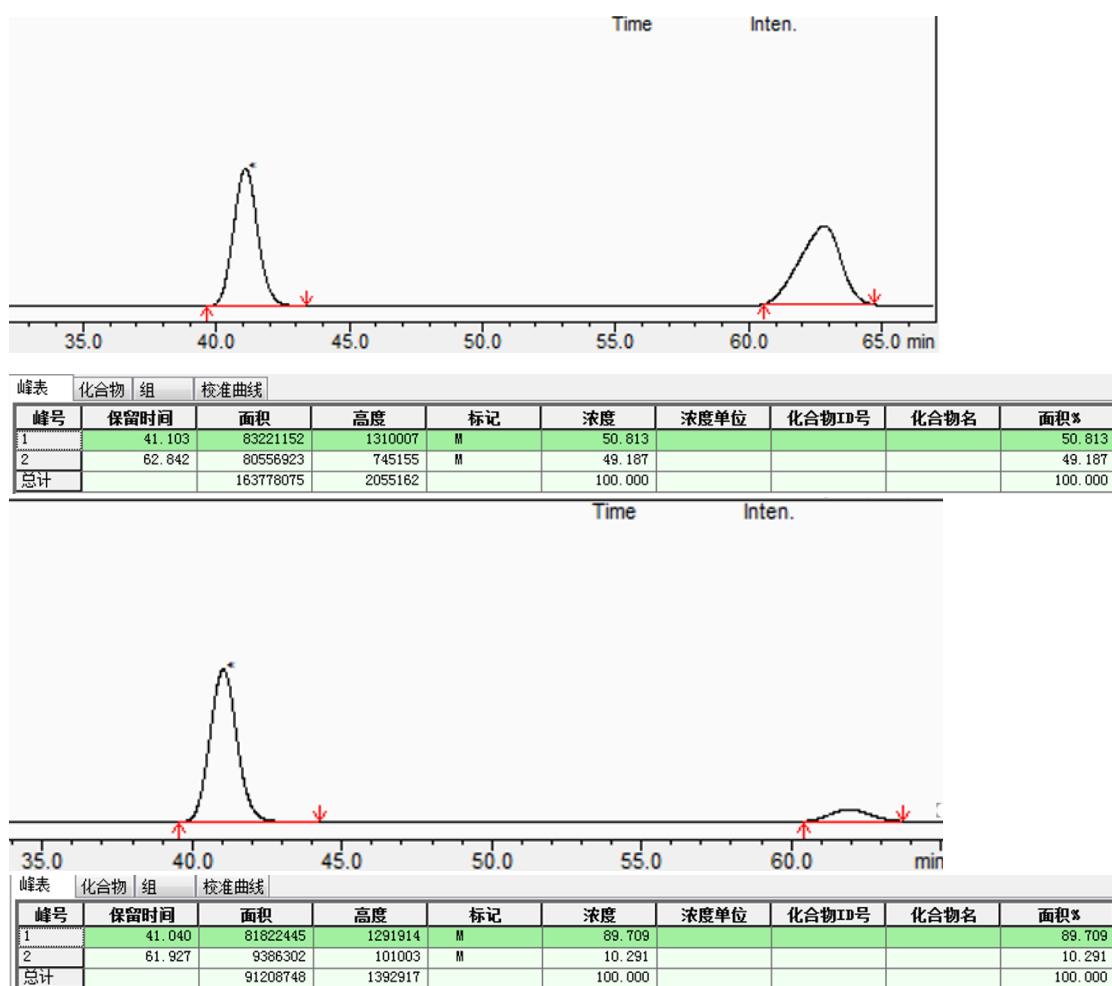
¹H NMR (400 MHz, CDCl₃), δ 3.77 (s, 3H, OCH₃), 4.87 (m, 1H), 4.94-4.99 (m, 2H), 6.77 (m, 1H, Aromatic H), 6.79 (m, 1H, Aromatic H), 6.84 (d, *J* = 7.88 Hz, 1H, Aromatic H), 7.22-7.27 (m, 4H, Aromatic H), 7.30-7.35 (m, 2H, Aromatic H).

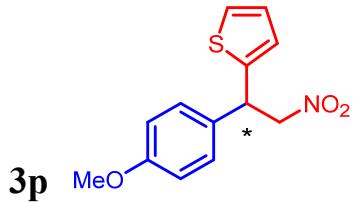
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 80/20, 220 nm, 1.0 mL/min.

t_{R1} = 41.0 min (minor), t_{R2} = 61.9 min (major)

ee = 79.5%.





Colorless oil, 62% yield.

The NMR data is in accordance with that of previous publications.^[7-9]

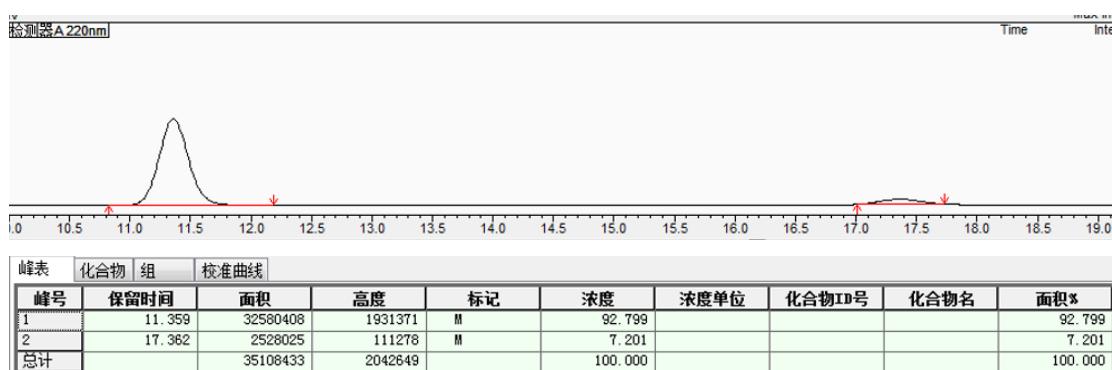
¹H NMR (400 MHz, CDCl₃), δ 3.79 (s, 3H, OCH₃), 4.89 (dd, *J*₁ = 12.60 Hz, *J*₂ = 8.56 Hz, 1H), 4.97 (dd, *J*₁ = 12.60 Hz, *J*₂ = 7.52 Hz, 1H), 5.08 (dd, *J*₁ = 8.56 Hz, *J*₂ = 7.52 Hz, 1H), 6.86-6.90 (m, 3H, Aromatic H), 6.95 (dd, *J*₁ = 5.12 Hz, *J*₂ = 3.52 Hz, 1H, Aromatic H), 7.19-7.24 (m, 3H, Aromatic H).

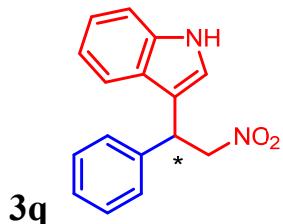
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 60/40, 220 nm, 1.0 mL/min.

t_{R1} = 11.4 min (major), t_{R2} = 17.4 min (minor)

ee = 85.5%.





Colorless solid, 62% yield.

The NMR data is in accordance with that of previous publications.^[10-11]

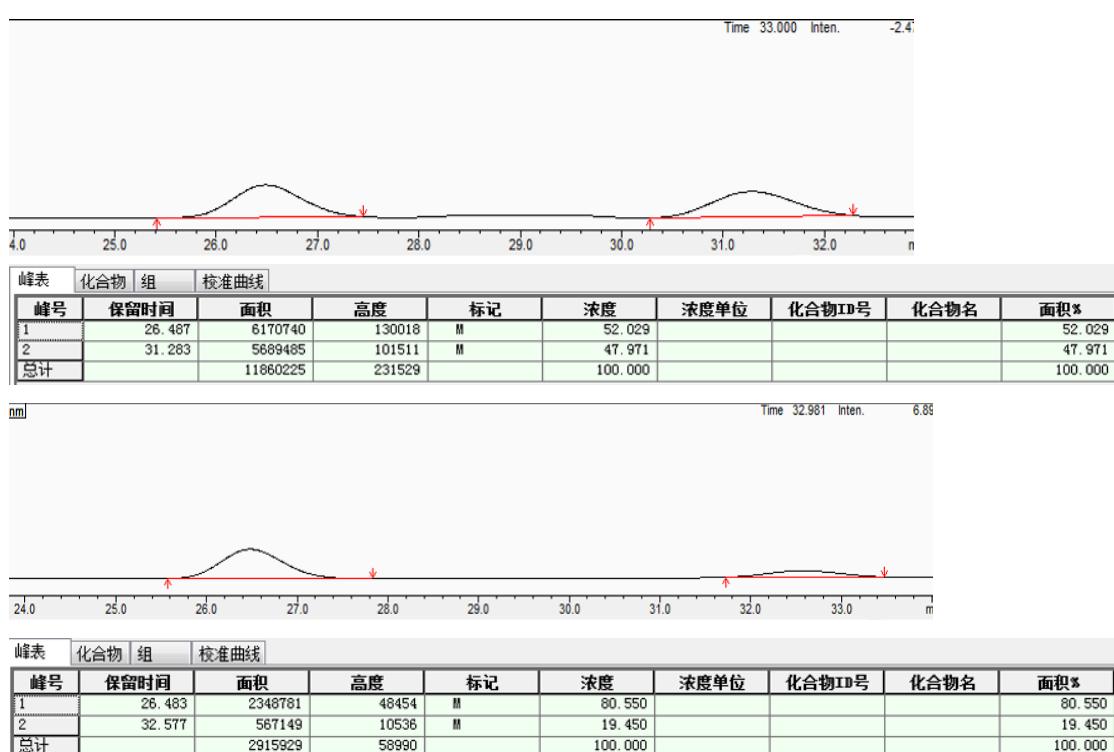
¹H NMR (400 MHz, CDCl₃), δ 4.97 (dd, *J*₁ = 12.48 Hz, *J*₂ = 8.36 Hz, 1H), 4.99 (dd, *J*₁ = 12.48 Hz, *J*₂ = 7.68 Hz, 1H), 5.22 (t, *J* = 8.04 Hz, 1H), 7.05 (d, *J* = 2.20 Hz, 1H, Aromatic H), 7.11 (m, 1H, Aromatic H), 7.24 (m, 1H, Aromatic H), 7.29 (m, 1H, Aromatic H), 7.33-7.39 (m, 5H, Aromatic H), 7.48 (d, *J* = 8.04 Hz, 1H, Aromatic H), 8.12 (s, 1H).

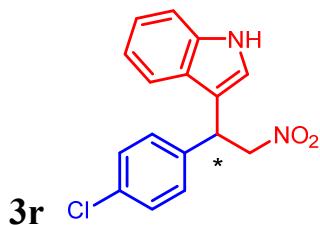
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

t_{R1} = 26.5 min (major), t_{R2} = 32.6 min (minor)

ee = 61.1%





Colorless solid, 60% yield.

The NMR data is in accordance with that of previous publications.^[10-11]

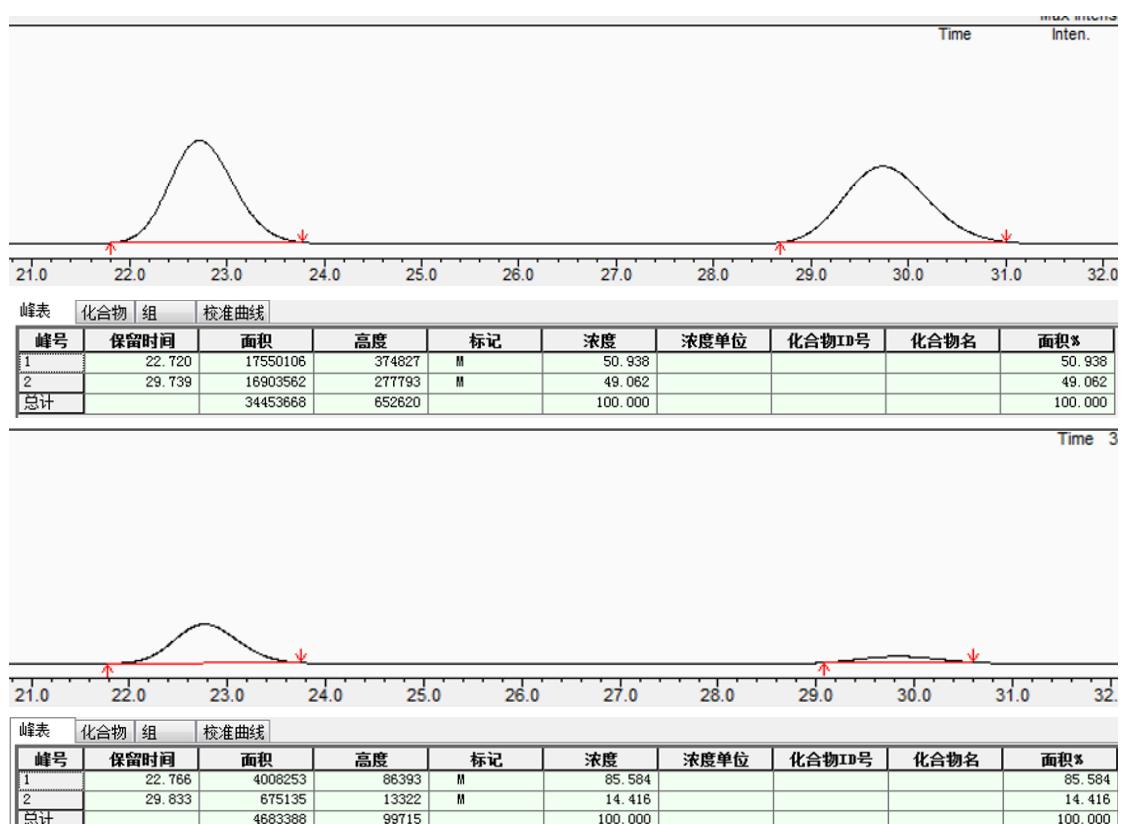
¹H NMR (500 MHz, CDCl₃), δ 4.85 (t, *J* = 110.25 Hz, 1H), 5.00 (dd, *J₁* = 10.25 Hz, *J₂* = 7.55 Hz, 1H), 5.12 (t, *J* = 7.60 Hz, 1H), 6.697-7.01 (m, 3H, Aromatic H), 7.04 (t, *J* = 7.30 Hz, 1H, Aromatic H), 7.16 (t, *J* = 7.20 Hz, 1H, Aromatic H), 77.27-7.30 (m, 2H, Aromatic H), 7.31 (d, *J* = 8.00 Hz, 1H, Aromatic H) 7.36 (d, *J* = 7.85 Hz, 1H, Aromatic H), 8.08 (s, 1H).

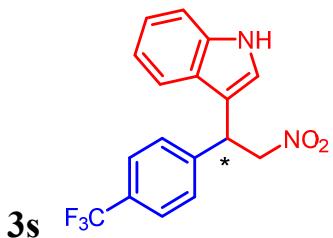
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

t_{R1} = 22.8 min (major), t_{R2} = 29.8 min (minor)

ee = 71.1%.





Colorless solid, 57% yield.

The NMR data is in accordance with that of previous publications.^[12-13]

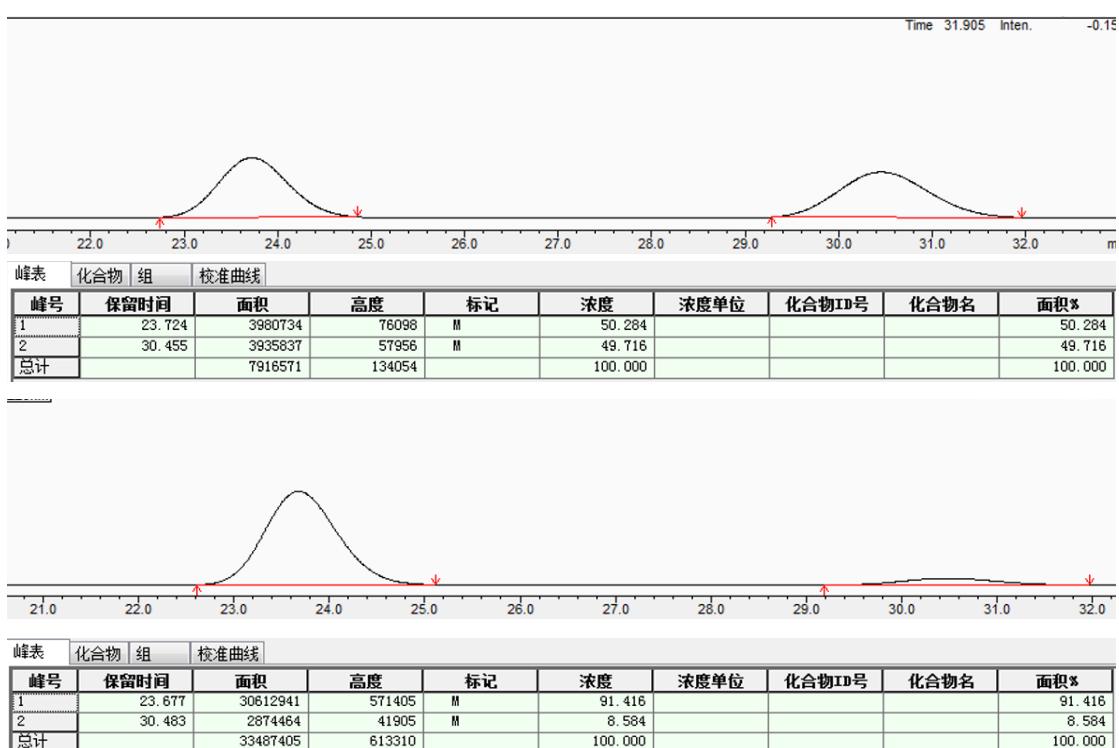
¹H NMR (500 MHz, CDCl₃), δ 4.97 (dd, *J*₁ = 11.75 Hz, *J*₂ = 8.85 Hz, 1H), 5.09 (dd, *J*₁ = 12.10 Hz, *J*₂ = 7.20 Hz, 1H), 5.26 (t, *J* = 8.05 Hz, 1H), 7.04 (m, 1H, Aromatic H), 7.09 (m, 1H, Aromatic H), 7.22 (t, *J* = 7.80 Hz, 1H, Aromatic H), 7.35-7.42 (m, 2H, Aromatic H), 7.47 (d, *J* = 7.90 Hz, 2H, Aromatic H), 7.58 (d, *J* = 7.95 Hz, 2H, Aromatic H), 8.17 (s, 1H).

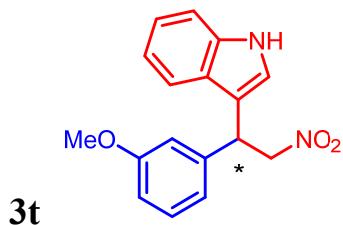
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

t_{R1} = 23.7 min (major), t_{R2} = 30.5 min (minor)

ee = 82.9%.





Colorless solid, 58% yield.

The NMR data is in accordance with that of previous publications.^[10-11]

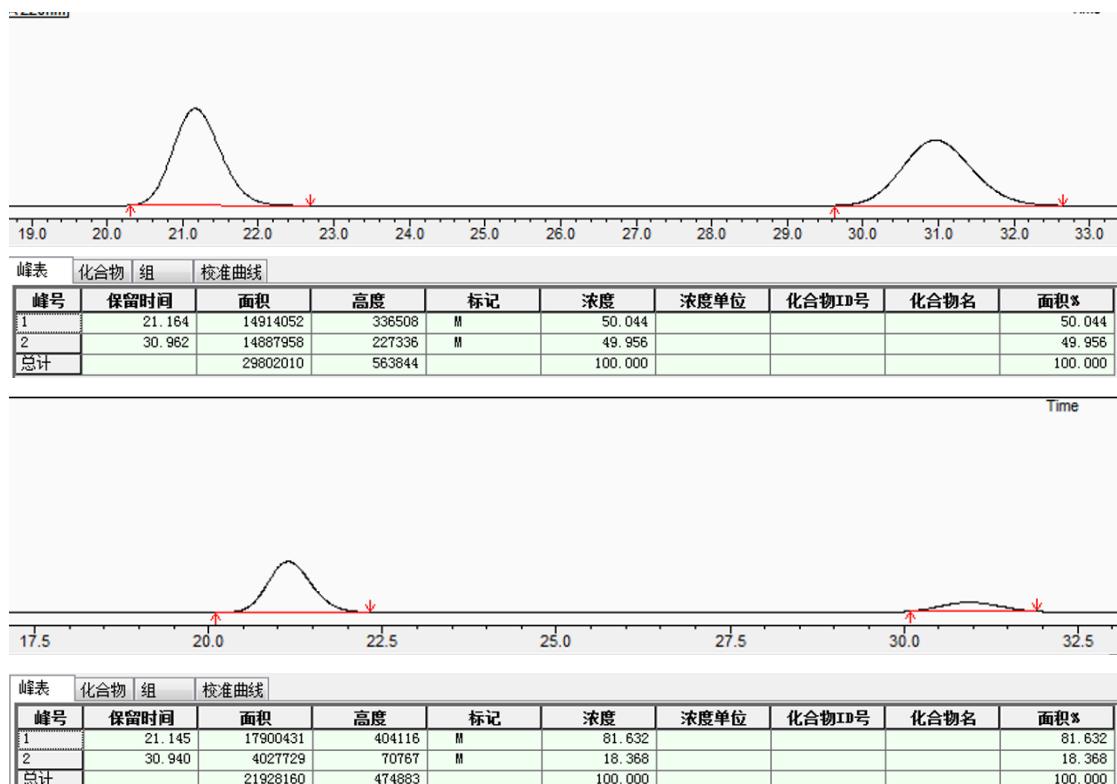
¹H NMR (500 MHz, CDCl₃), δ 3.76 (s, 3H, OMe), 4.93 (dd, *J*₁=9.90 Hz, *J*₂=7.15 Hz, 1H), 5.04 t, *J*= 10.30 Hz, 1H), 5.16 (t, *J*=8.15 Hz, 1H), 6.79 (d, *J*= **8.40** Hz, 1H, Aromatic H), 6.86 (s, 1H, Aromatic H), 6.93 (d, *J*= 7.80 Hz, 1H, Aromatic H), 7.04 (s, 1H, Aromatic H), 7.08 (t, *J*= 7.70 Hz, 1H, Aromatic H), 7.18-7.24 (m, 2H, Aromatic H), 7.35 (d, *J*= 8.25 Hz, 1H, Aromatic H), 7.47 (d, *J*= 8.00 Hz, 1H, Aromatic H), 8.10 (s, 1H).

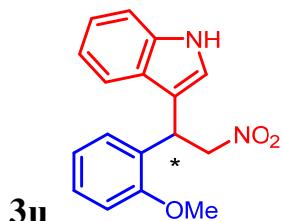
HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

t_{R1} = 21.1min (major), t_{R2} = 30.9 min (minor)

ee = 63.3%.





Colorless solid, 48% yield.

The NMR data is in accordance with that of previous publications.^[10-11]

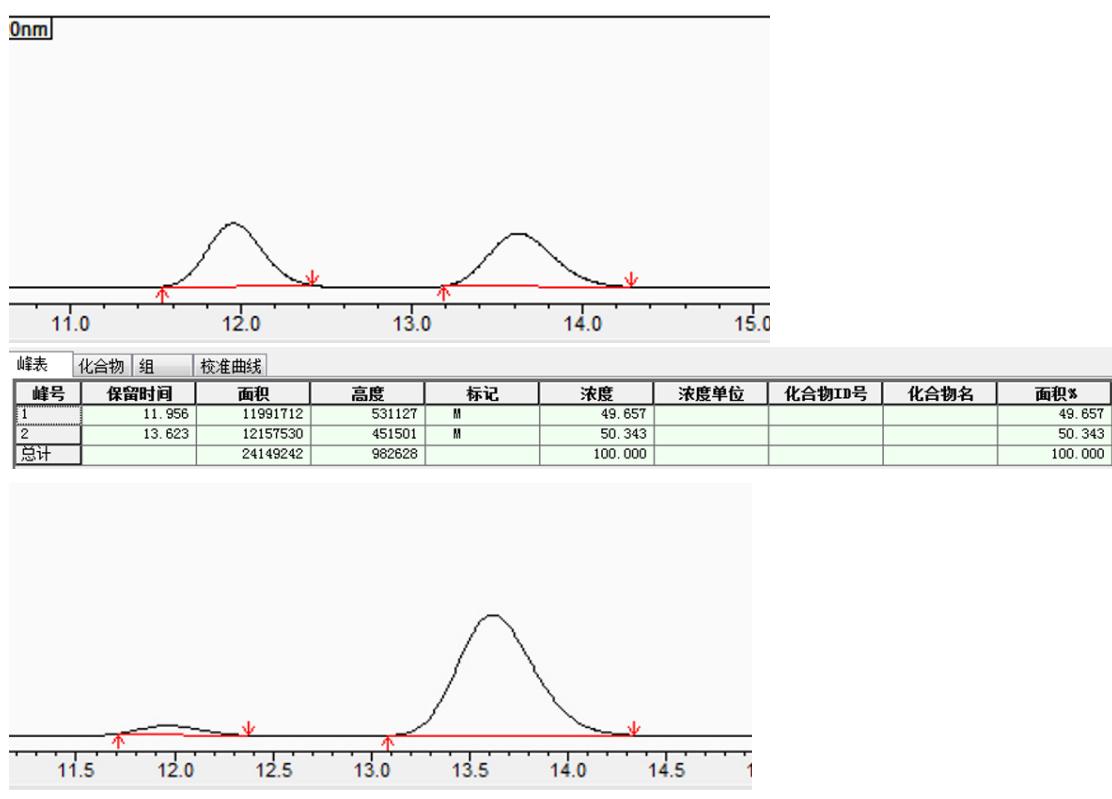
¹H NMR (500 MHz, CDCl₃), δ 3.87 (s, 3H, OMe), 4.92 -5.03 (m, 2H), 5.57 (t, *J* = 7.50 Hz, 1H), 6.80 (t, *J* = 7.10 Hz, 1H, Aromatic H), 6.89 (d, *J* = 8.10 Hz 1H, Aromatic H), 7.03 -7.07 (m, 3H, Aromatic H), 7.15 (t, *J* = 7.10 Hz, 1H, Aromatic H), 7.20 (t, *J* = 6.40 Hz, 1H, Aromatic H), 7.28 (d, *J* = 8.05 Hz, 1H, Aromatic H), 7.44 (d, *J* = 7.75 Hz, 1H, Aromatic H), 8.03 (s, 1H).

HPLC trace:

Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.0 mL/min.

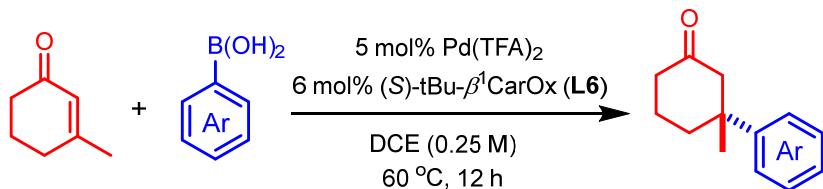
t_{R1} = 12.0 min (minor), t_{R2} = 13.6 min (minor)

ee = 90.3%.



Enantioselective Addition of Arylboronic Acids to β -substituted Cyclic Enones

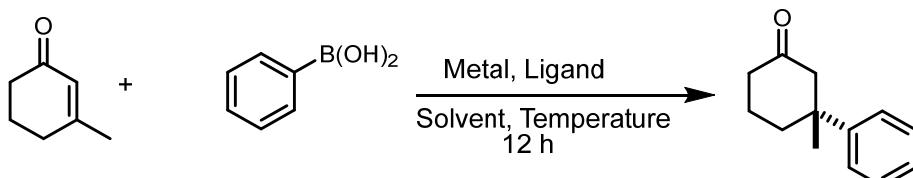
General procedure



To a Schlenk tube charged with $\text{Pd}(\text{TFA})_2$ (4.15 mg, 0.0125 mmol), (*S*)-tBu- $\beta^1\text{CarOx}$ (4.4 mg, 0.015 mmol) and aryl boronic acid (0.5 mmol), was added dichloroethane (0.5 mL) for dissolution and then β -substituted cyclic enones (0.25 mmol) was added. The walls of the vial were rinsed with an additional portion of dichloroethane (0.5 mL). The vial was capped with a Teflon/silicone septum and stirred in a 60 °C oil bath for 12 h. The reaction mixture was cooled to room temperature, and the solvent was removed by rotary evaporation. The residue was purified by column chromatography (petroleumether/EtOAc = 20/1, v/v) to give the product.

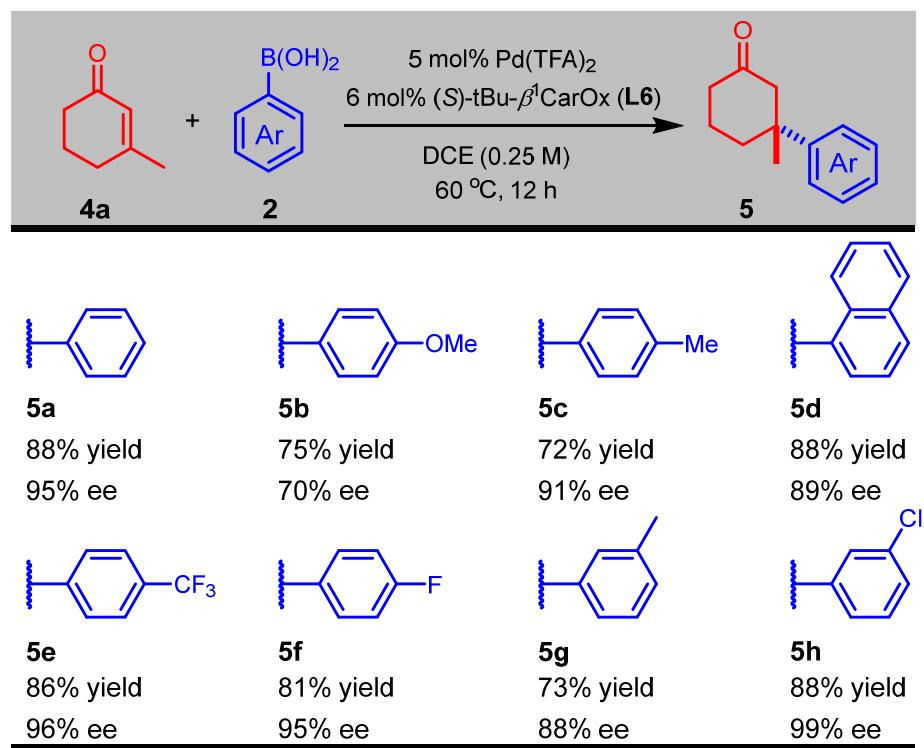
Racemic products were synthesized in a manner analogous to the general procedure using bipyridine (2.1 mg, 0.015 mmol, 6 mol%) as an achiral ligand.

Concise optimization

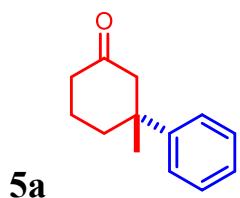


Entry	Metal	β -CarOx	Solvent	Temperature	Yield(%)	ee(%)
1	$\text{Pd}(\text{TFA})_2$	L3	DCE	60 °C	78	50
2	$\text{Pd}(\text{TFA})_2$	L6	DCE	60 °C	88	94
3	$\text{Pd}(\text{TFA})_2$	L7	DCE	60 °C	76	91
4	$\text{Pd}(\text{TFA})_2$	L8	DCE	60 °C	73	32
5	$\text{Pd}(\text{TFA})_2$	L10	DCE	60 °C	56	93
6	PdCl_2	L6	DCE	60 °C	<5	n.d.
7	$\text{Pd}(\text{OAc})_2$	L6	DCE	60 °C	67	89
8	$\text{Pd}(\text{TFA})_2$	L6	DCM	40 °C	49	90
9	$\text{Pd}(\text{TFA})_2$	L6	MeOH	60 °C	<5	n.d.
10	$\text{Pd}(\text{TFA})_2$	L6	DCE	40 °C	72	90
11	$\text{Pd}(\text{TFA})_2$	L6	DCE	80 °C	86	87

Substrate Scope



HPLC traces of the Enantioenriched β -aryl ketones



Colorless oil, 88% yield.

The NMR data is in accordance with that of previous publication.^[14]

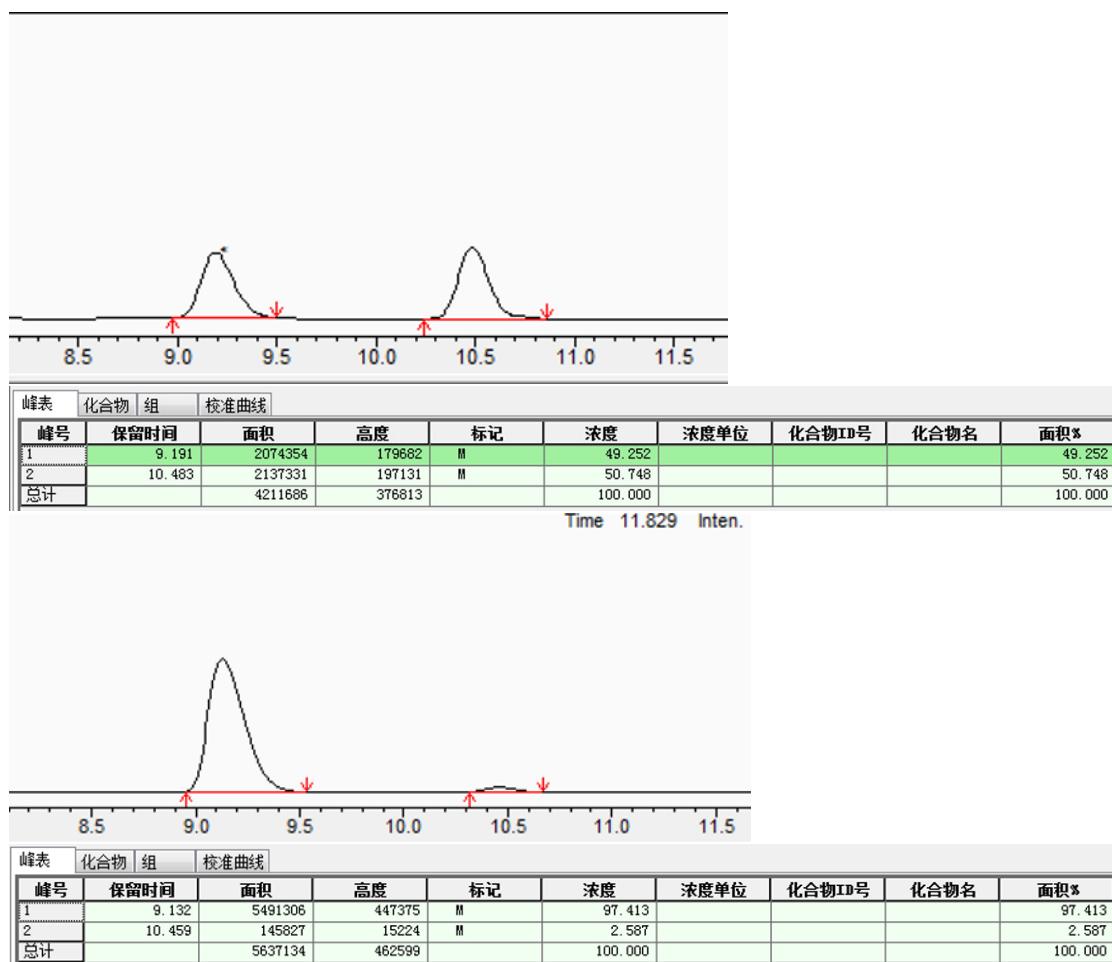
^1H NMR (400 MHz, CDCl_3), δ 1.35(s, 3H, CH_3), 1.67 (m, 1H), 1.87-1.98 (m, 2H), 2.21 (m, 1H), 2.231-2.42 (m, 2H), 2.46 (d, $J=14.12$ Hz, 1H), 2.91 (d, $J=14.20$ Hz, 1H), 7.20-7.25 (m, 1H, Aromatic H), 7.32-7.37 (m, 4H, Aromatic H).

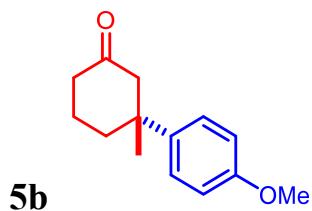
HPLC trace:

Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 1.0 mL/min.

$t_{\text{R}1}$ = 9.1 min (major), $t_{\text{R}2}$ = 10.5 min (minor)

ee = 94.9%.





Colorless oil, 75% yield.

The NMR data is in accordance with that of previous publications.^[14-16]

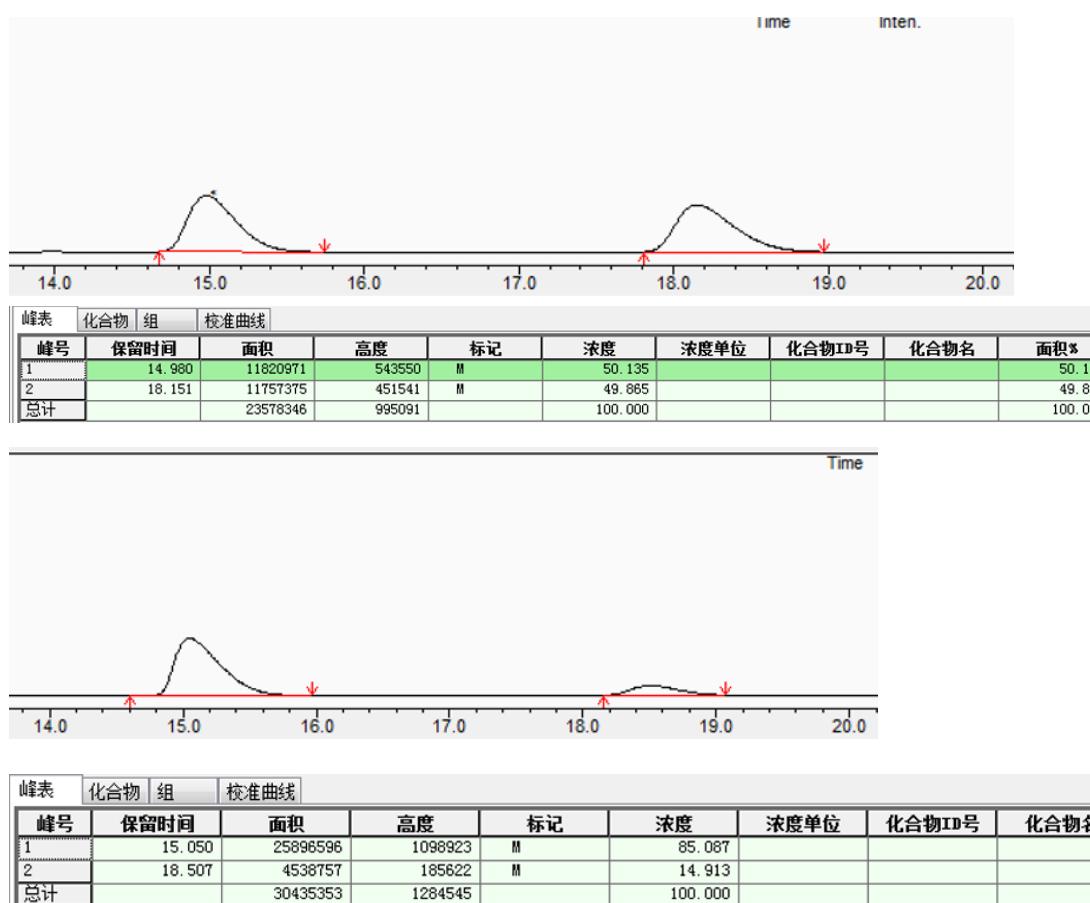
¹H NMR (400 MHz, CDCl₃), δ 1.31(s, 3H, CH₃), 1.68 (m, 1H), 1.85-1.94 (m, 2H), 2.2.21 (m, 1H), 2.30-2.32 (m, 2H), 2.43 (d, *J*=14.12 Hz, 1H), 2.87 (d, *J*=14.16 Hz, 1H), 3.79 (s, 3H), 6.85-6.89 (m, 2H, Aromatic H), 7.23-7.27 (m, 2H, Aromatic H).

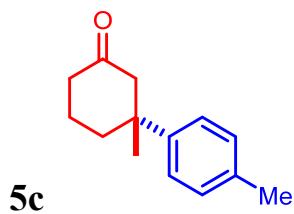
HPLC trace:

Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 1.0 mL/min.

t_{R1} = 15.0 min (major), t_{R2} = 18.5 min (minor)

ee = 70.2%.





Colorless oil, 72% yield.

The NMR data is in accordance with that of previous publications.^[14-16]

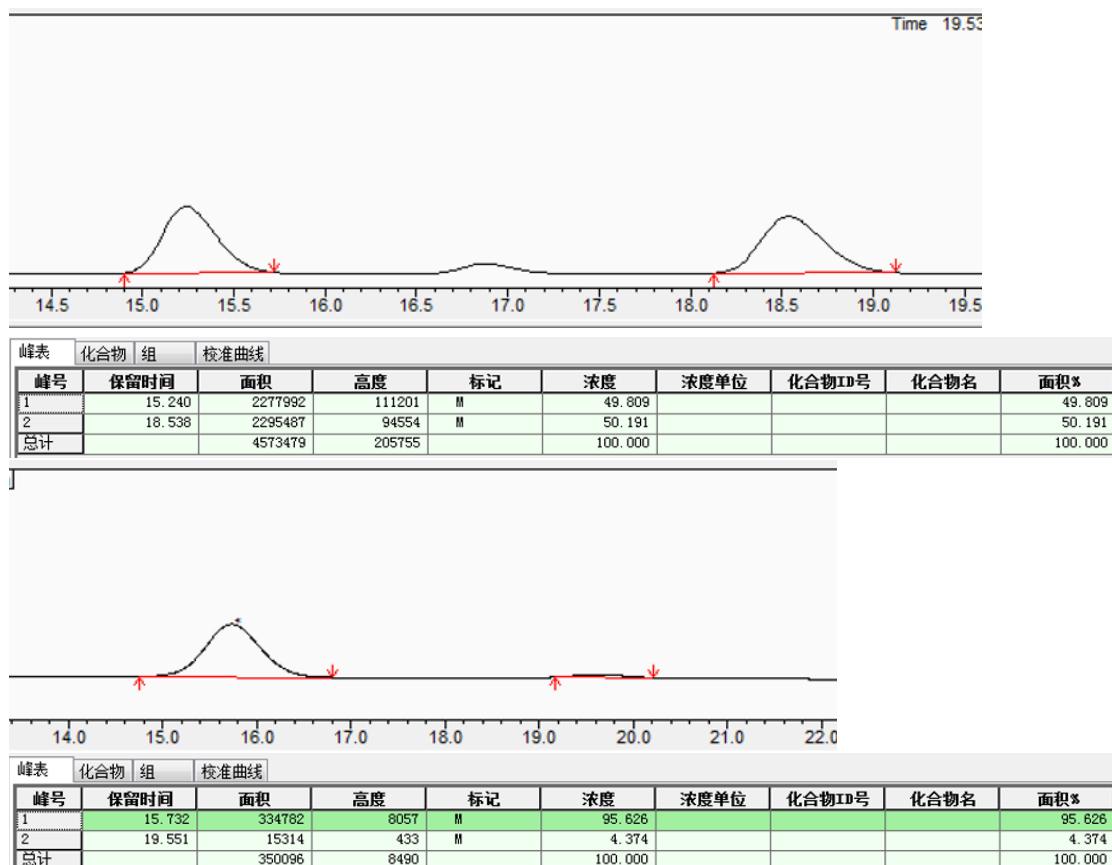
¹H NMR (400 MHz, CDCl₃), δ 1.33(s, 3H, CH₃), 1.1.70 (m, 1H), 1.86-1.95 (m, 2H), 2.12.19 (m, 1H), 2.28-2.32 (m, 2H), 2.34 (s, 3H, CH₃), 2.44 (d, *J*=8.12 Hz, 1H), 2.89 (d, *J*=9.80 Hz, 1H), 7.16 (d, *J*=8.16 Hz, 2H, Aromatic H), 7.23 (d, *J*=8.28 Hz, 2H, Aromatic H).

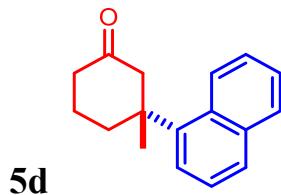
HPLC trace:

Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 1.0 mL/min.

t_{R1} = 15.7 min (major), t_{R2} = 19.5 min (minor)

ee = 91.2%.





Colorless oil, 88% yield.

The NMR data is in accordance with that of previous publication.^[17]

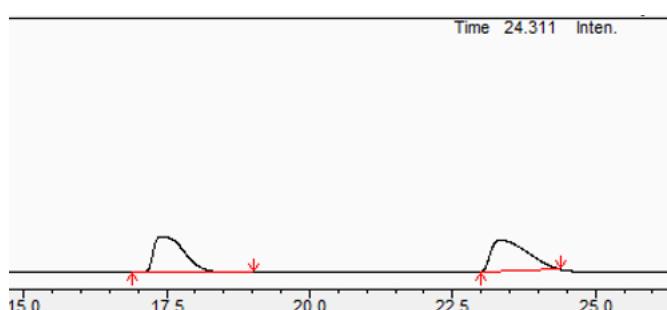
¹H NMR (400 MHz, CDCl₃), δ 1.44 (s, 3H, CH₃), 1.67 (m, 1H), 1.89-2.05 (m, 2H), 2.31-2.38 (m, 3H), 2.55 (d, *J*=14.24 Hz, 1H), 3.05 (d, *J*=14.32 Hz, 1H), 7.46-7.52 (m, 3H, Aromatic H), 7.73 (d, *J*=1.64 Hz, 1H, Aromatic H), 7.82-7.86 (m, 3H, Aromatic H).

HPLC trace:

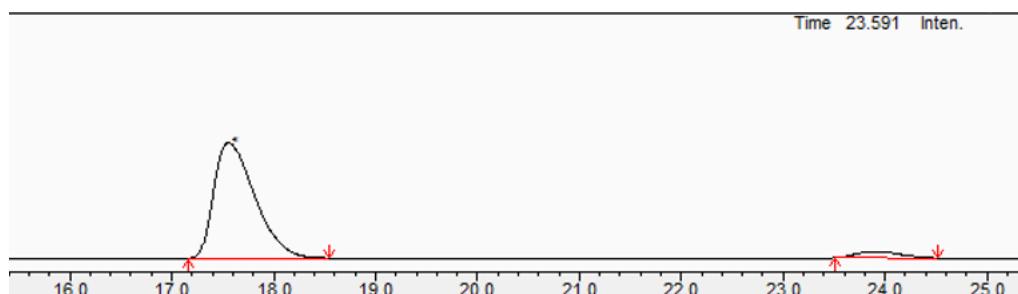
Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 1.0 mL/min.

t_{R1} = 17.5 min (major), t_{R2} = 23.9 min (minor)

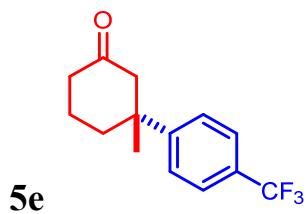
ee = 88.8%.



峰表	化合物	组	校准曲线	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1				17.428	108316047	2955125	M	49.015				49.015
2				23.347	112667848	2565653	M	50.985				50.985
总计					220983896	5520776		100.000				100.000



峰表	化合物	组	校准曲线	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1				17.559	51900871	1855385	M	94.409				94.409
2				23.908	3073584	104045	M	5.591				5.591
总计					54974455	1959430		100.000				100.000



Colorless oil, 86% yield.

The NMR data is in accordance with that of previous publications.^[14, 16]

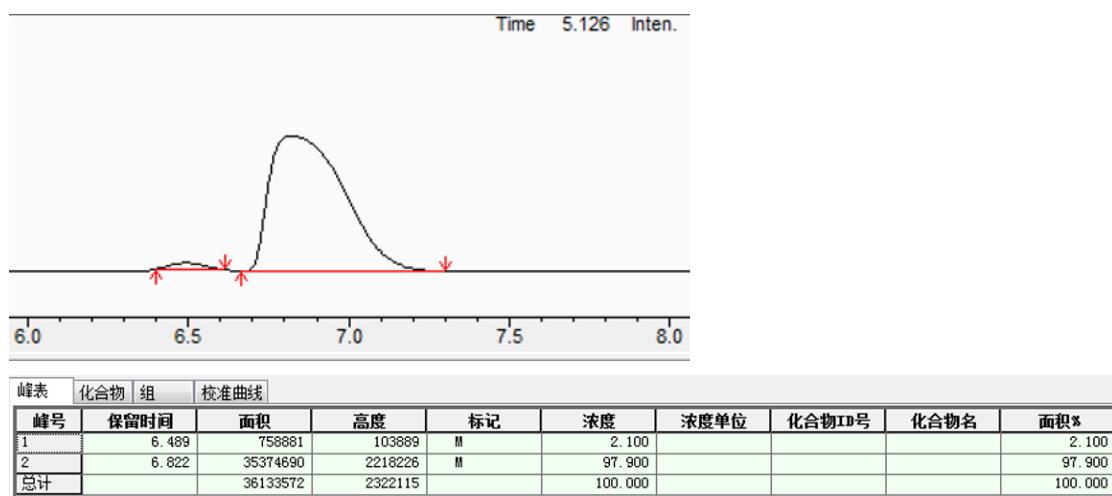
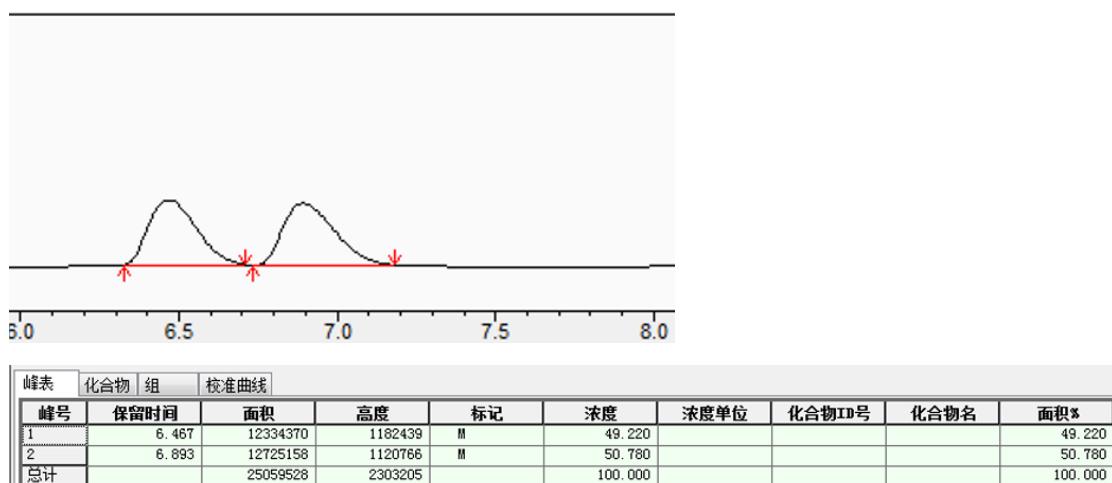
¹H NMR (500 MHz, CDCl₃), δ 1.34 (s, 3H, CH₃), 1.62-1.67 (m, 2H), 1.89-1.98 (m, 2H), 2.19 (m, 1H), 2.32-2.36 (m, 2H), 2.48 (d, *J*=14.20 Hz, 1H), 2.88 (d, *J*=14.20 Hz, 1H), 7.44 (d, *J*=8.10 Hz, 2H, Aromatic H), 7.58 (d, *J*=7.75 Hz, 2H, Aromatic H).

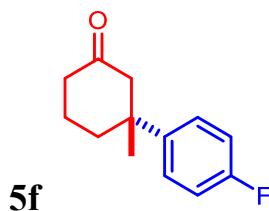
HPLC trace:

Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 1.0 mL/min.

t_{R1} = 6.82 min (major), t_{R2} = 6.49 min (minor)];

ee = 95.8%.





Colorless oil, 81% yield.

The NMR data is in accordance with that of previous publications.^[14, 18]

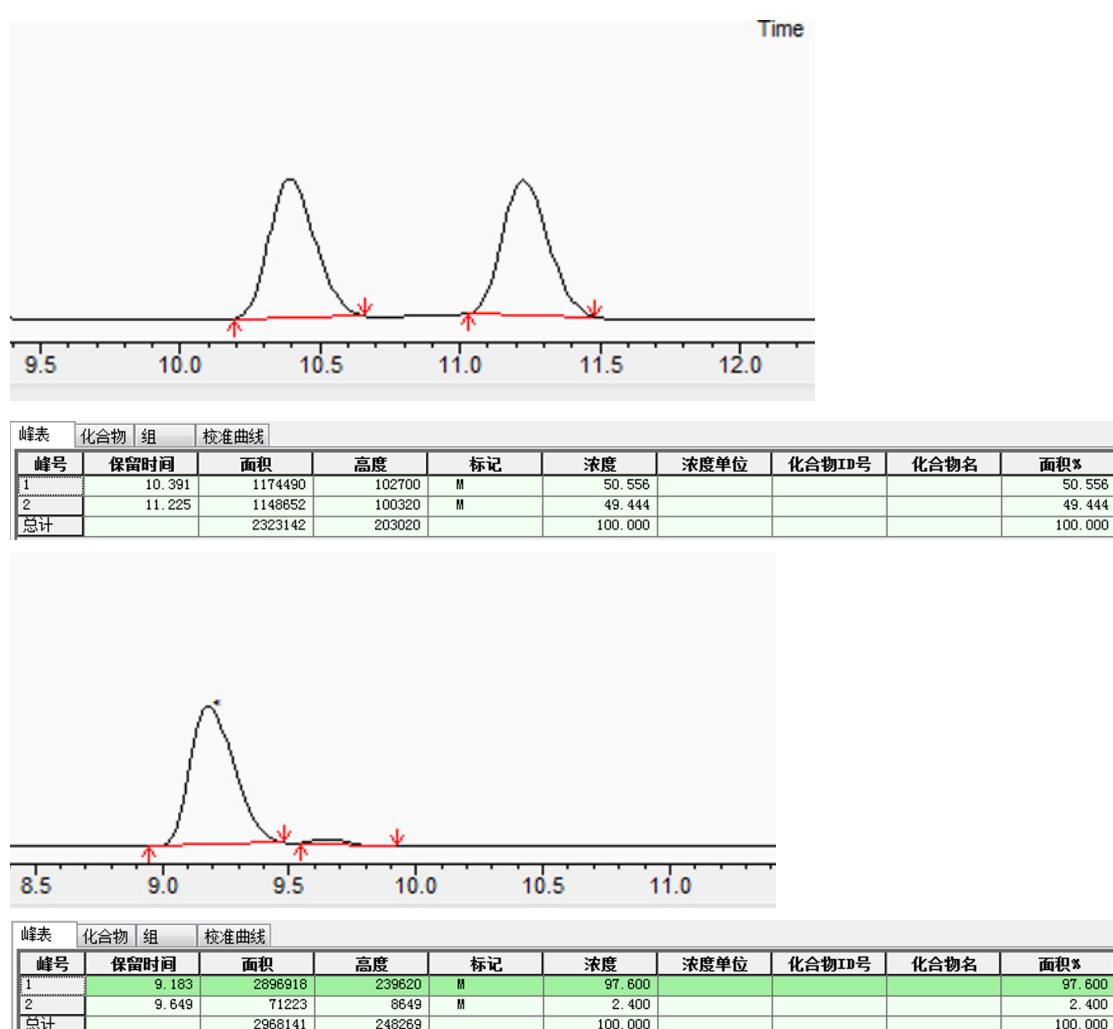
¹H NMR (400 MHz, CDCl₃), δ 1.32 (s, 3H, CH₃), 1.61-1.69 (m, 1H), 1.84-1.95 (m, 2H), 2.13-2.19 (m, 1H), 2.32 (t, J₁=13.48 Hz, J₂=6.74 Hz, 2H), 2.44 (d, J=10.12 Hz, 1H), 2.85 (d, J=14.20 Hz, 1H), 7.23-7.31 (m, 4H, Aromatic H).

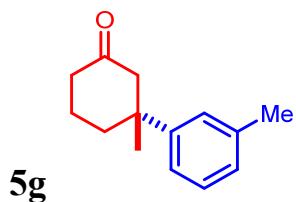
HPLC trace:

Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 0.9 mL/min.

t_{R1} = 9.18 min (major), t_{R2} = 10.8 min (minor)];

ee = 95.2%.





Colorless oil, 73% yield.

The NMR data is in accordance with that of previous publication.^[14]

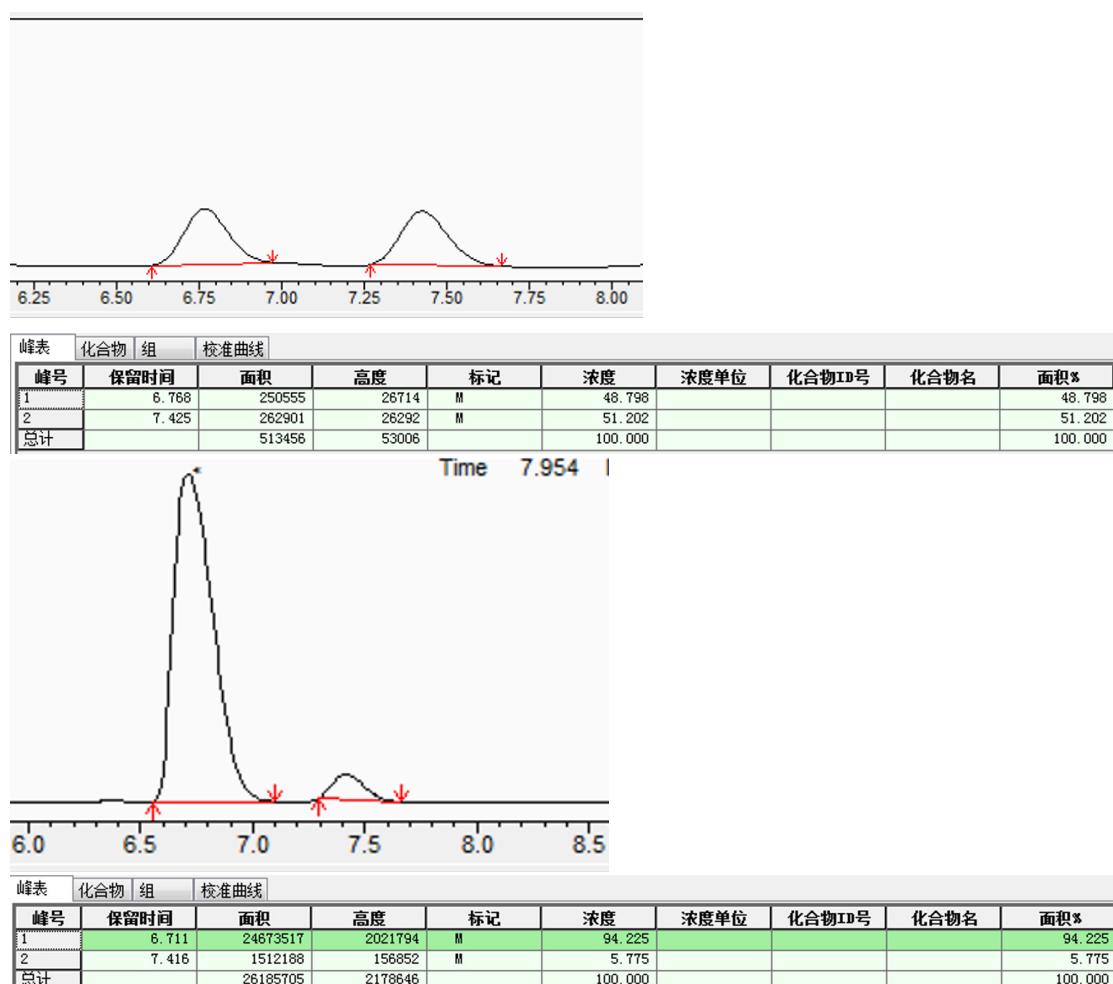
¹H NMR (500 MHz, CDCl₃), δ 1.31 (s, 3H, CH₃), 1.70 (m, 1H), 1.85-1.93 (m, 2H), 2.18 (m, 1H), 2.30-2.34 (m, 5H), 2.42 (d, *J*=14.15 Hz, 1H), 2.87 (d, *J*=14.20 Hz, 1H), 7.02 (d, *J*=7.60 Hz, 1H, Aromatic H), 7.11 (d, *J*=8.80 Hz, 2H, Aromatic H), 7.21 (m, 1H, Aromatic H).

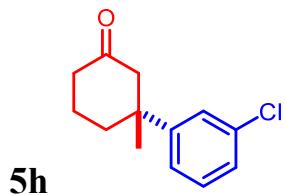
HPLC trace:

Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 1.0 mL/min.

t_{R1} = 6.71 min (major), t_{R2} = 7.42 min (minor)];

ee = 88.4%.





Colorless oil, 88% yield.

The NMR data is in accordance with that of previous publications.^[14-15]

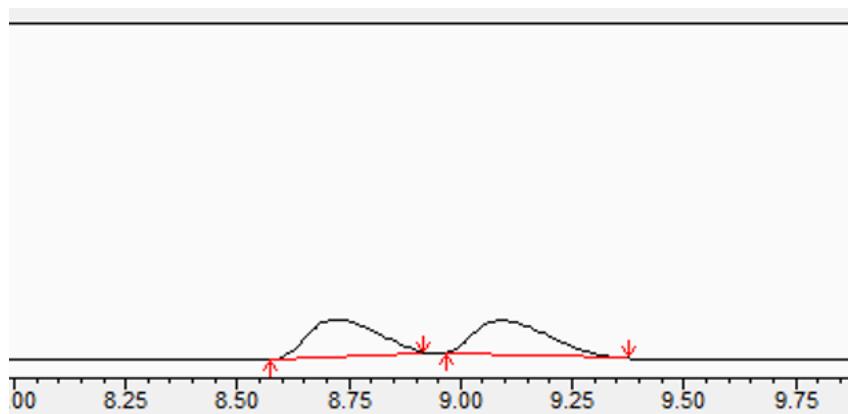
¹H NMR (500 MHz, CDCl₃), δ 1.31 (s, 3H, CH₃), 1.69 (m, 1H), 1.87-1.94 (m, 2H), 2.15 (m, 1H), 2.31-2.34 (m, 2H), 2.43 (d, *J*=14.15 Hz, 1H), 2.84 (d, *J*=14.15 Hz, 1H), 7.18-7.20 (m, 2H, Aromatic H), 7.25 (m, 1H, Aromatic H) 7.30 (s, 1H, Aromatic H).

HPLC trace:

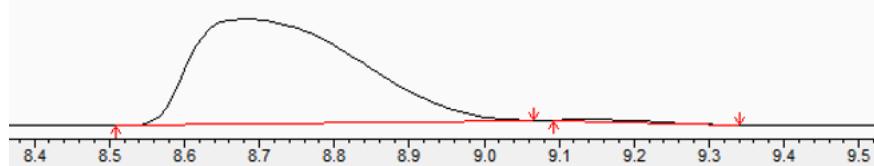
Daicel chiralcel OJ-H, hexane/i-PrOH = 95/5, 220 nm, 0.9 mL/min.

t_{R1} = 8.68 min (major), t_{R2} = 9.14 min (minor)];

ee = 98.6%.



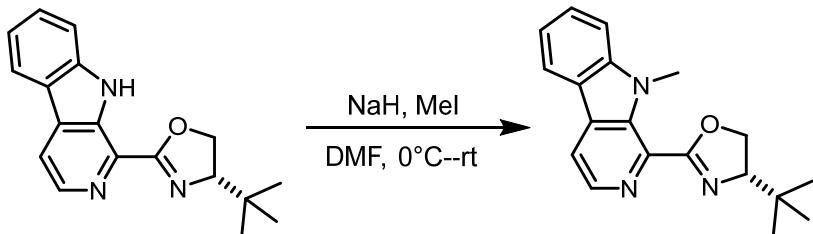
峰号	化合物	组	校准曲线	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1				8.722	17233770	1622842	M	49.360				49.360
2				9.094	17680500	1522708	M	50.640				50.640
总计					34914270	3145550		100.000				100.000



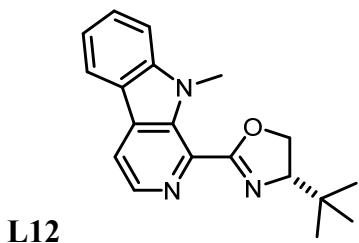
峰号	化合物	组	校准曲线	保留时间	面积	高度	标记	浓度	浓度单位	化合物ID号	化合物名	面积%
1				8.681	36838698	2402542	M	99.300				99.300
2				9.143	259805	36243	M	0.700				0.700
总计					37098503	2438785		100.000				100.000

Synthesis and Application of Chiral Ligands L12 and L13

Synthesis of *N*-Methylated (*S*)-tBu- β^1 -CarOx L12



L12 was synthesized following the reported procedure.^[19] To a suspended solution of NaH (4.8 mg, 0.2 mmol) in DMF (1 mL), (*S*)-tBu- β^1 -CarOx (29.3 mg, 0.1 mmol) in DMF (1 mL) was added dropwise at 0 °C. The heterogeneous mixture was stirred at 0 °C for 15 min and 1 h at room temperature. The mixture was then cooled to 0 °C, treated with iodomethane (10 µL, 0.15 mmol), and allowed to warm to room temperature. After 6 h, the reaction mixture was cooled to 0 °C, quenched with saturated aqueous NH₄Cl (3 mL), and extracted with ethyl acetate (3×5 mL). The organic layers were combined, washed with brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The resulting oil was purified by column chromatography on silica gel to give product *N*-Methylated (*S*)-tBu- β^1 -CarOx **L12** as a white solid (56% yield).



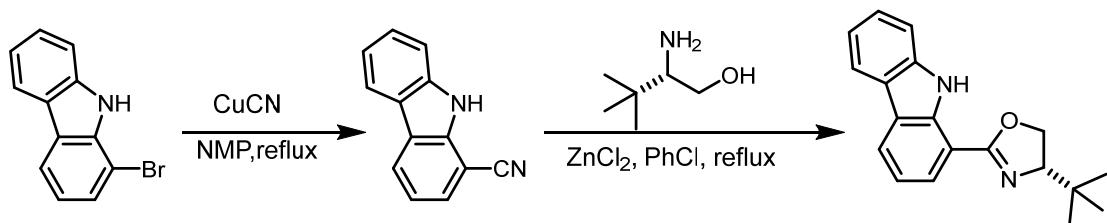
(*S*)-4-(tert-butyl)-2-(9-methyl-9H-pyrido[3,4-b]indol-3-yl)-4,5-dihydrooxazole

¹H NMR (500 MHz, CDCl₃), δ 1.07 (s, 9H, 3 × CH₃), 4.09 (s, 3H, N-CH₃), 4.28 (dd, J₁ = 9.95 Hz, J₂ = 9.95 Hz, 1H), 4.35 (dd, J₁ = 9.95 Hz, J₂ = 8.7 Hz, 1H), 4.57 (dd, J₁ = 9.95 Hz, J₂ = 8.7 Hz, 1H), 7.32 (m, 1H), 7.48 (d, J = 8.20 Hz, 1H) 7.62 (dd, J₁ = 7.85 Hz, J₂ = 6.90 Hz, 1H), 8.06 (s, 1H), 8.14 (d, J = 7.90 Hz 1H), 8.54 (s, 1H).

¹³C NMR (125 MHz, CDCl₃), δ 26.3, 32.9, 33.9, 68.9, 77.4, 109.8, 116.2, 120.2, 120.8, 121.5, 129.0, 131.1, 131.3, 135.8, 138.3, 143.1, 162.3.

HRESI-MS: calcd for C₁₉H₂₂N₃O [M+H]⁺: 308.1764, found: 308.1658.

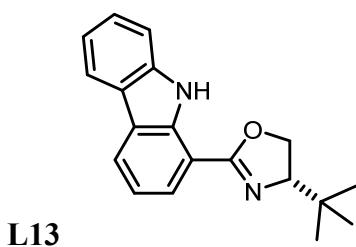
Synthesis of Carbazole-Oxazoline Ligand L13



L13 was synthesized following the reported procedure. [6], [20]

Step 1, To a suspended solution of CuCN (182 mg, 2.0 mmol) in NMP (3 mL), 1-Bromocarbazole (246 mg, 1.0 mmol) in 2 ml NMP was added dropwise , the reaction mixture was heated to reflux for 2h. The reaction mixture was then cooled to ambient temperature, quenched with H₂O (3 mL), and extracted with ethyl acetate (3×5 mL). The organic layers were combined, washed with brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The resulting oil was purified by column chromatography on silica gel to give product 1-cyanide carbazole (yield: 68%, as white solid).

Step 2, To a suspended solution of S-tert-Leucinol (140 mg, 1.2 mmol) and ZnCl₂(8.3 mg, 0.06 mmol) in PhCl (3 mL), 1-Cyanide carbazole (115 mg, 0.6 mmol) in 2 ml in PhCl was added dropwise , the reaction mixture was heated to reflux for 12h. The reaction mixture was then cooled to ambient temperature, quenched with H₂O (3 mL), and extracted with ethyl acetate (3×5 mL). The organic layers were combined, washed with brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The resulting oil was purified by column chromatography on silica gel to give product C₁-S-'Bu-Carbazole-Oxazoline Ligand (**L13**) (yield: 48 %, as white solid).



(*S*)-4-(tert-butyl)-2-(9H-carbazol-3-yl)-4,5-dihydrooxazole

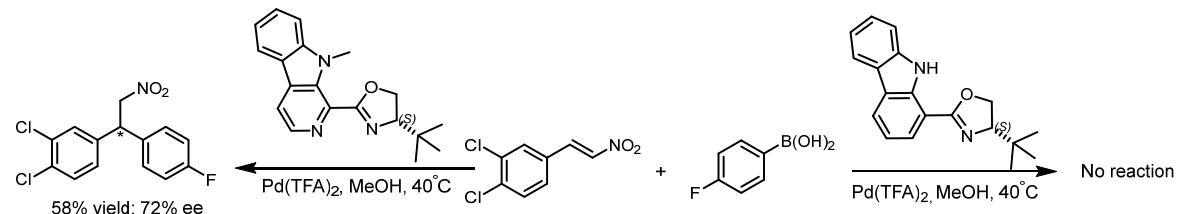
¹H NMR (500 MHz, CDCl₃), δ 1.03 (s, 9H, 3 × CH₃), 4.20 (dd, J₁ = 9.95 Hz, J₂ = 7.75 Hz, 1H), 4.28 (t, J₁ = 7.75 Hz, J₂ = 8.25 Hz, 1H), 4.40 (dd, J₁ = 9.95 Hz, J₂ = 8.25 Hz, 1H), 7.23 -7.27 (m, 2H), 7.46 (m, 1H), 7.54 (d, J = 8.05 Hz, 1H), 7.86 (d, dd, J₁ =

7.60 Hz, J_2 = 1.10 Hz, 1H), 8.10 (d, J = 7.75 Hz, 1H), 8.19 (d, J = 7.75 Hz, 1H), 10.55 (s, 1H).

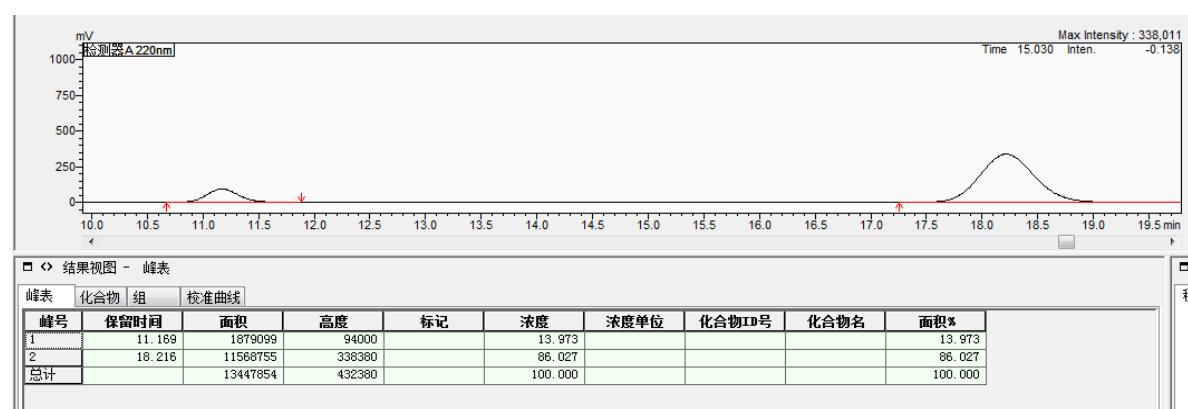
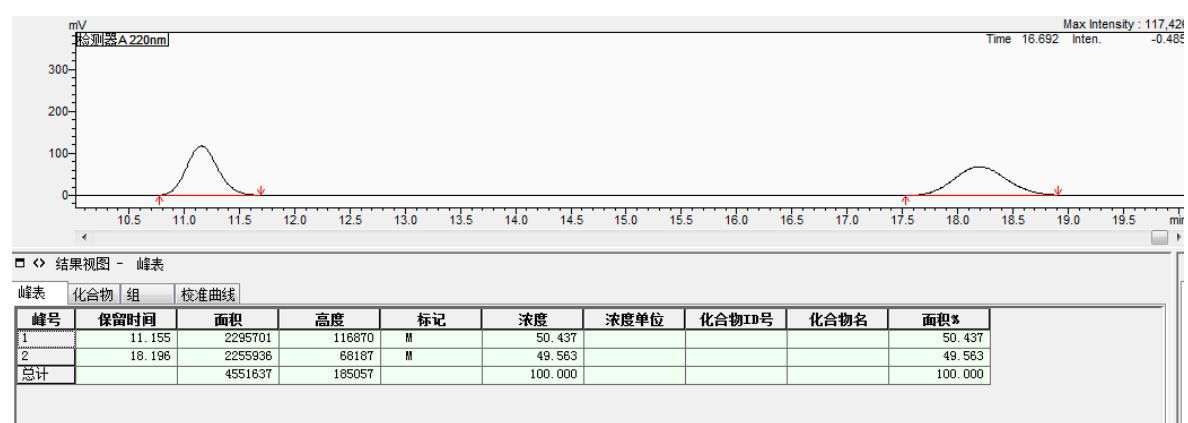
^{13}C NMR (125 MHz, CDCl_3), δ 26.0, 34.1, 68.1, 76.2, 109.5, 111.3, 118.3, 119.6, 120.5, 122.9, 123.4, 123.6, 125.3, 126.2, 139.2, 139.6, 163.1.

HRESI-MS: calcd for $\text{C}_{19}\text{H}_{21}\text{N}_2\text{O} [\text{M}+\text{H}]^+$: 293.1655, found: 293.1658.

Application of **L12** and **L13** in the enantioselective synthesis of compound **3i**



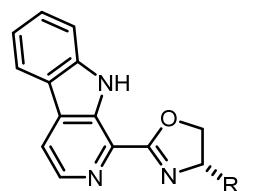
HPLC traces for the chiral ligand **L12**,



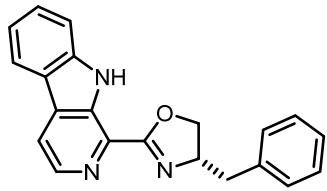
HPLC [Daicel chiralcel OD-H, hexane/i-PrOH = 70/30, 220 nm, 1.2 mL/min. t_{R1} = 11.17 min (major), t_{R2} = 18.22 min (minor)]; ee = 72.0%.

Antifungal Bioassay of CarOx Ligands

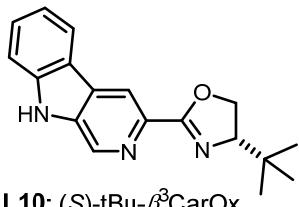
Compounds Selected for Biotest



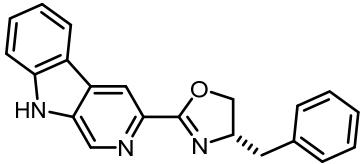
- L1: (S)-Me- β^1 CarOx
L2: (S)-Et- β^1 CarOx
L3: (S)-iPr- β^1 CarOx
L4: (S)-iBu- β^1 CarOx
L5: (S)-sBu- β^1 CarOx
L6: (S)-tBu- β^1 CarOx
L7: (S)-Ph- β^1 CarOx
L8: (S)-Bn- β^1 CarOx



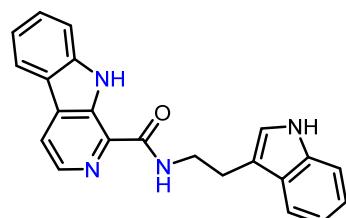
L9: (R)-Bn- β^1 CarOx



L10: (S)-tBu- β^3 CarOx



L11: (S)-Bn- β^3 CarOx



alangiolbussinine

Initial Screening

The antifungal activity of the target compounds was tested in *vitro* against the plant pathogenic fungi using the mycelium growth rate test. All the tested compounds were dissolved in DMSO at a concentration of $\mu\text{g/mL}$. The media containing compounds at a concentration of $50 \mu\text{g/mL}$ were then poured into Petri dishes for initial screening.

$$\text{Inhibition rate (\%)} = (C - T) / (C - 5 \text{ mm}) \times 100\%$$

Where C : The average diameter (in mm) of mycelia in the blank test, T : The average diameter (in mm) of mycelia on treated PDA with tested compounds.

Compd.	Inhibitory Rate at $50 \mu\text{g/mL}$ (%)					
	<i>Rhizoctonia solani</i>	<i>Sclerotinia sclerotiorum</i>	<i>Botrytis cinerea</i>	<i>Fusarium graminearum</i>	<i>Phytophthora capsici</i>	<i>Magnaporthe oryzae</i>
L1	39.2	63.5	57.8	56.8	65.1	64.7
L2	66.3	86.5	83.1	71.4	80.8	87.2
L3	70.4	86.5	98.0	69.1	87.8	82.1
L4	94.9	70.2	78.2	45.5	52.9	57.1
L5	86.8	74.0	71.7	67.0	63.9	66.7
L6	69.4	92.3	72.3	66.7	77.3	70.5
L7	75.5	90.9	59.4	53.8	55.8	65.8
L8	61.8	54.5	31.5	52.9	56.4	51.9
L9	71.5	88.5	61.2	54.8	74.4	83.3
L10	64.2	47.8	37.9	60.0	46.5	63.5
L11	78.1	92.5	63.7	60.0	62.1	61.5
alangiobussinine	33.5	48.9	52.6	52.3	34.5	52.5

Precise Antifungal Test

In the precision antifungal test, the 20 mg/mL stock solution was diluted to 50, 25, 12.5, 6.25, 3.125, 1.5625, 0.78125 µg/mL and the above experiments were repeated for three times, the inhibition rates were calculated separately. The statistical analyses were performed by SPSS software version 20.0. Inhibition rate was calculated as follows,

$$\text{Inhibition rate (\%)} = (C-T) / (C-5 \text{ mm}) \times 100\%$$

Where C: The average diameter (in mm) of mycelia in the blank test, T: The average diameter (in mm) of mycelia on treated PDA with tested compounds.

EC₅₀ values (µM) of the Selected Antifungal β-CarOx Ligands

Compd.	<i>Rhizoctonia solani</i>	<i>Sclerotinia sclerotiorum</i>	<i>Botrytis cinerea</i>	<i>Fusarium graminearum</i>	<i>Phytophthora capsici</i>	<i>Magnaporthe oryzae</i>
L2	57.7	22.4	72.5	66.4	55.4	55.5
L3	50.3	15.5	37.7	60.9	40.5	54.1
L5	28.7	17.7	56.2	67.3	92.9	47.5
L6	35.7	13.0	45.7	92.9	26.9	42.7
L7	57.5	17.3	65.3	104.8	109.1	38.6
L9	16.4	12.0	75.5	84.5	17.4	36.6
alangiolobussinine	>141.6	>141.6	>141.6	>141.6	>141.6	>141.6
<i>Boscalid</i>	4.6	0.9	4.9	165.3	7.4	3.3

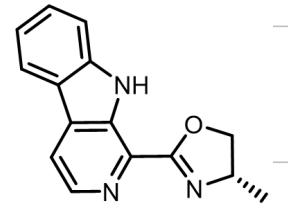
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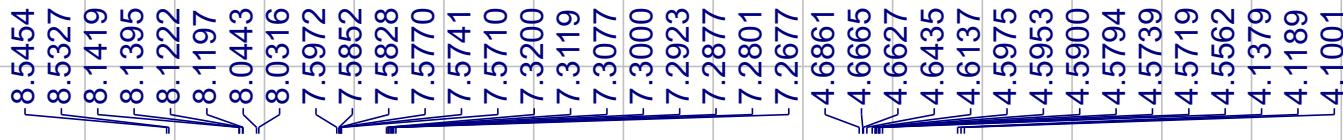
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NMR Spectra Traces

L1/L1H NMR
K67



-10.3599



1.4831

1.4670

11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5

f1 (ppm)

L1/L1 CNMR
K67



-163.0771

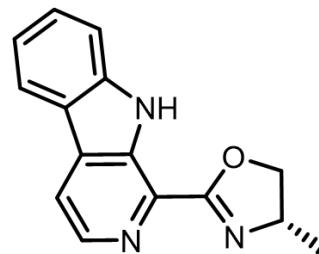
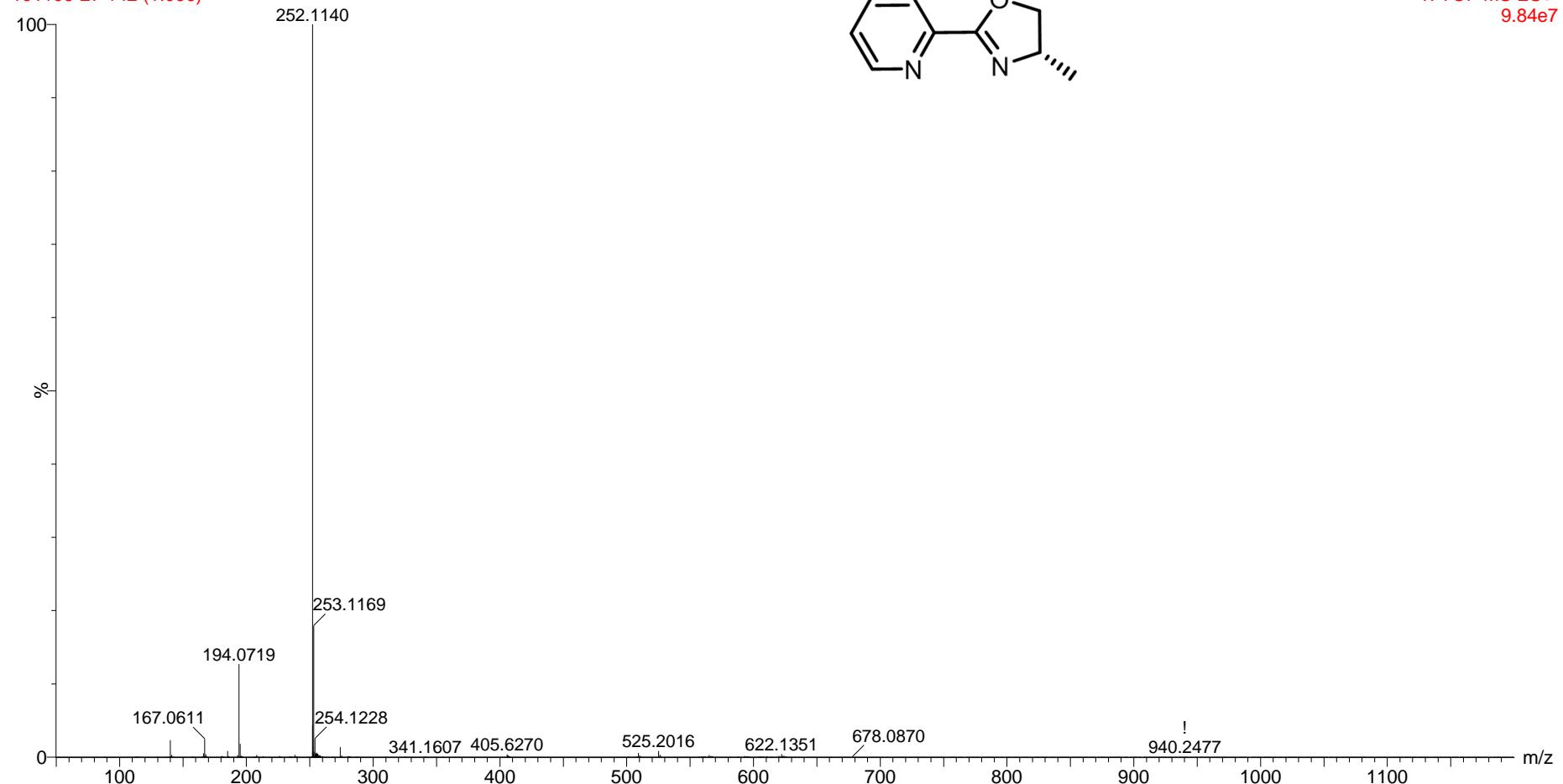
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111.9442
77.4025
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73.9123
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-21.7961

210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

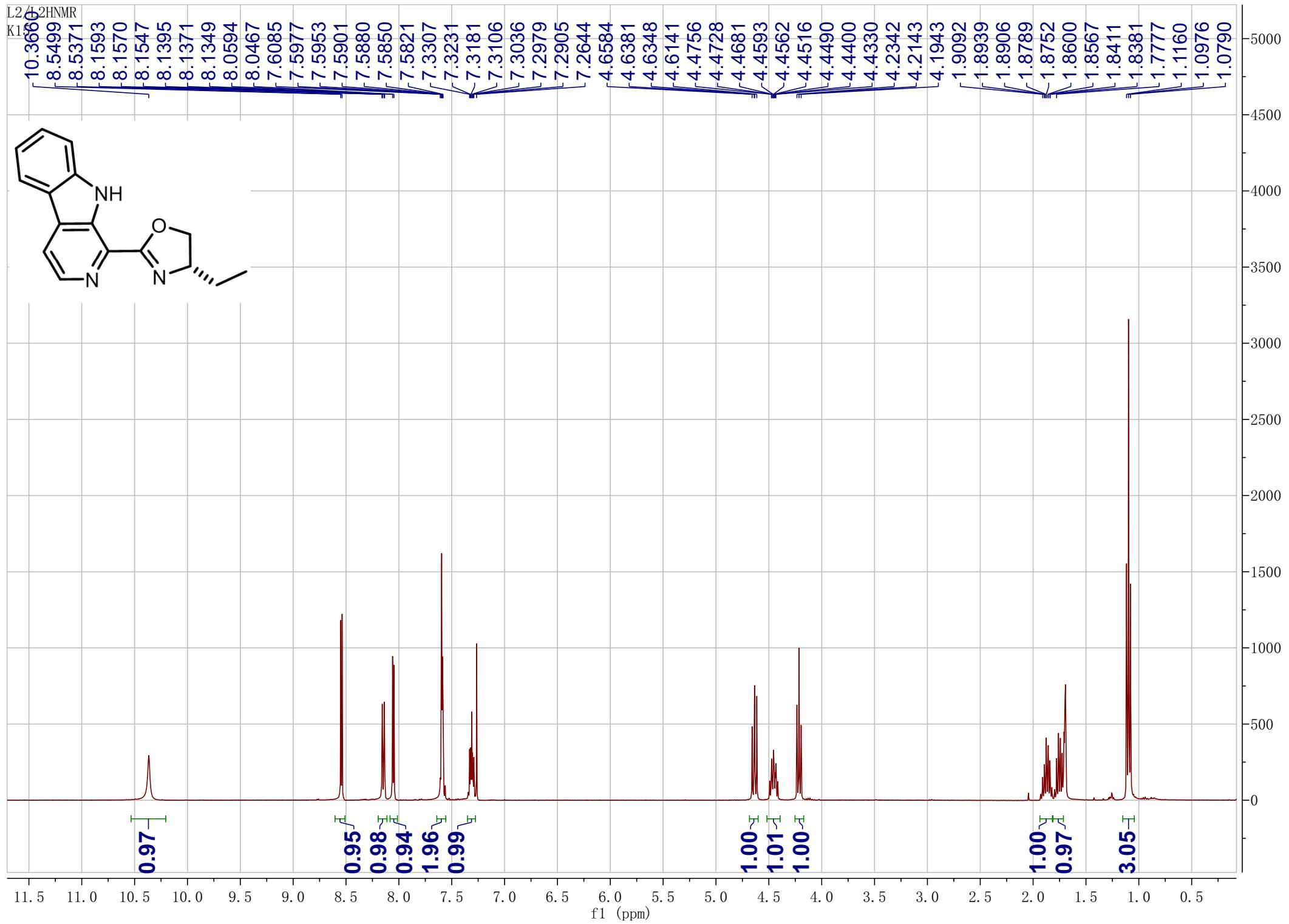
f1 (ppm)

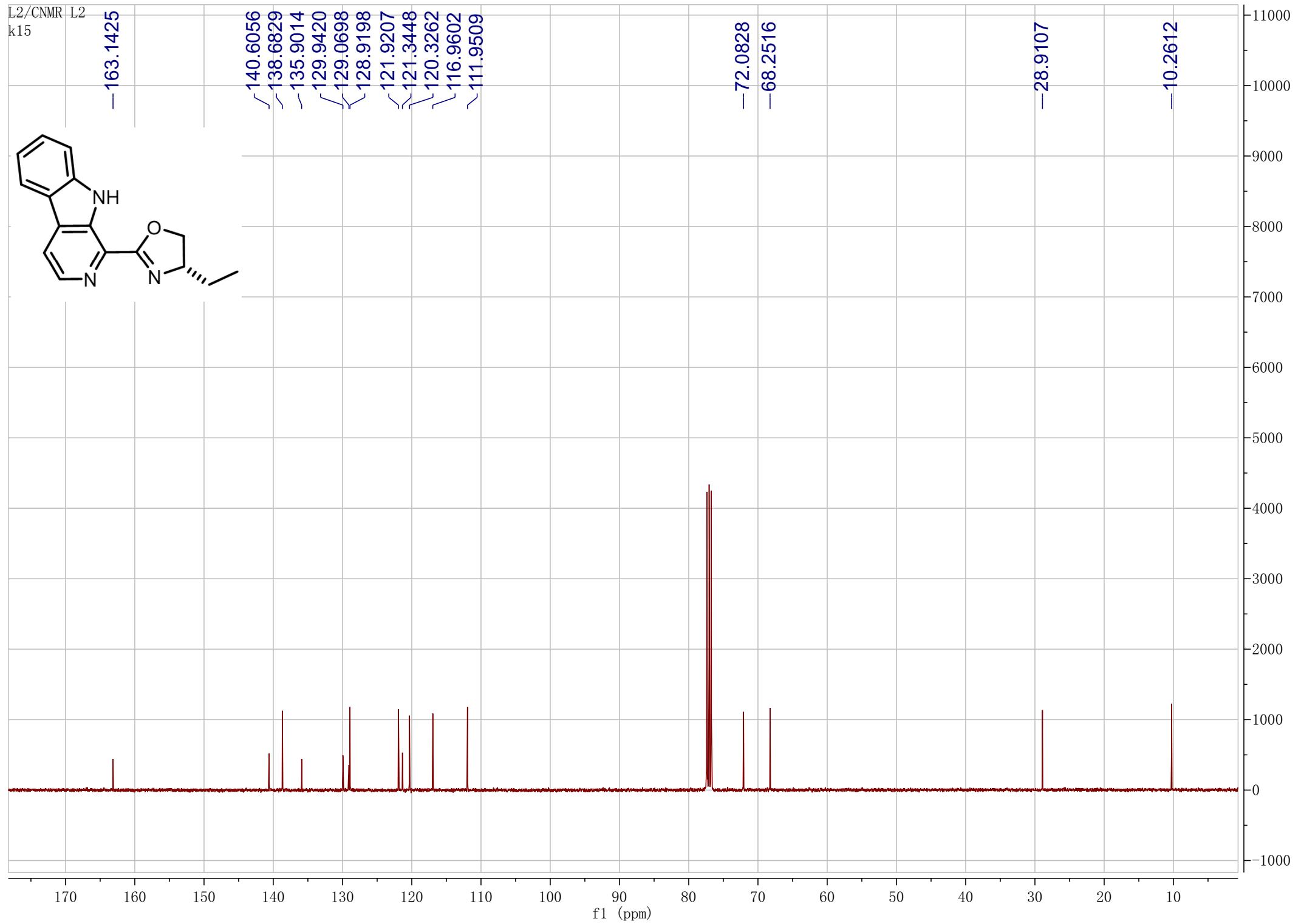
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0
-50

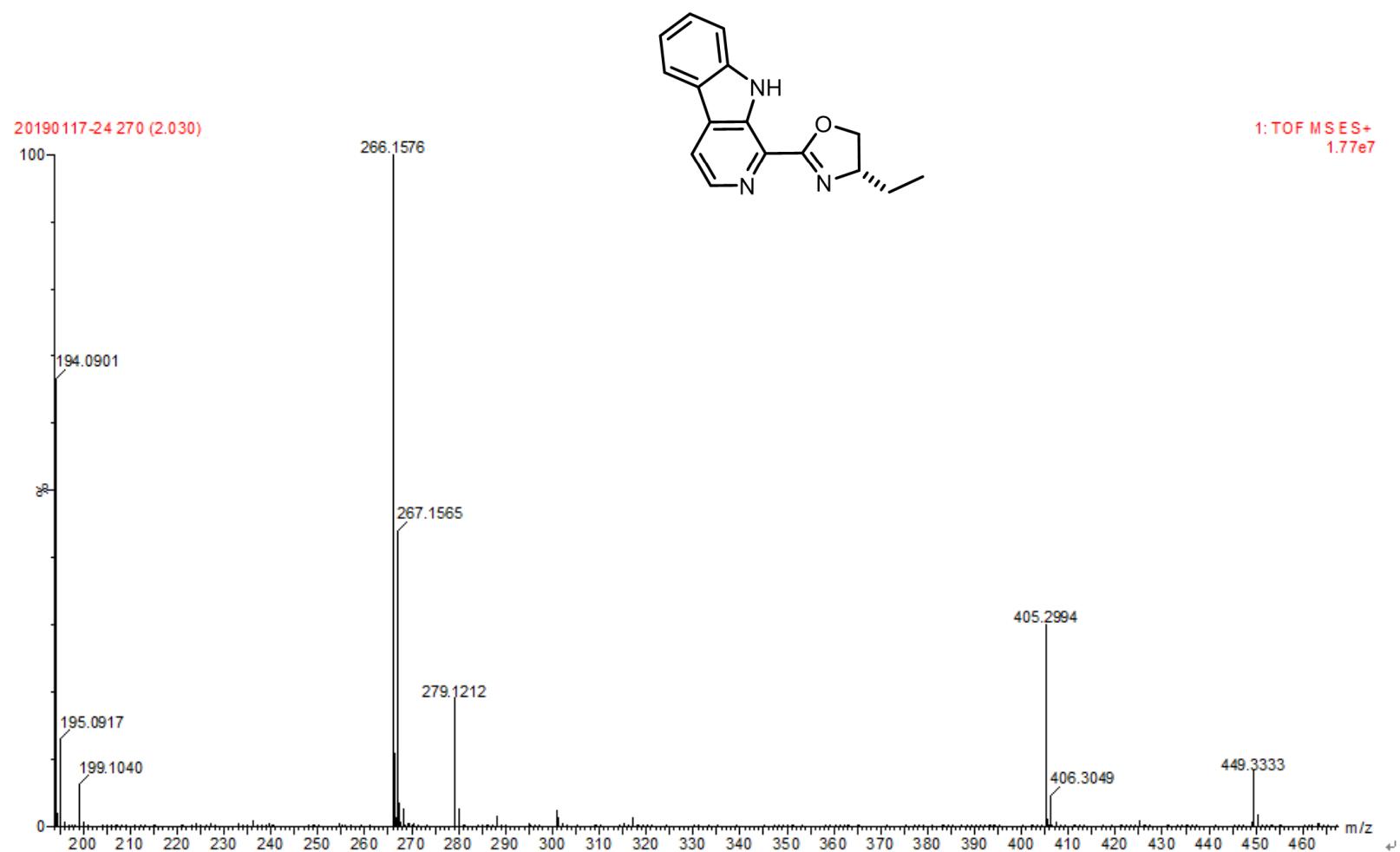
191106-27 142 (1.096)



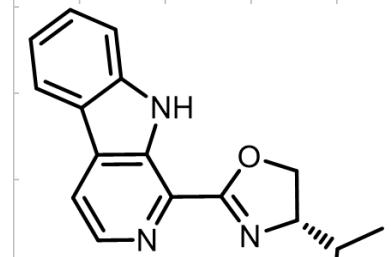
1: TOF MS ES+
9.84e7



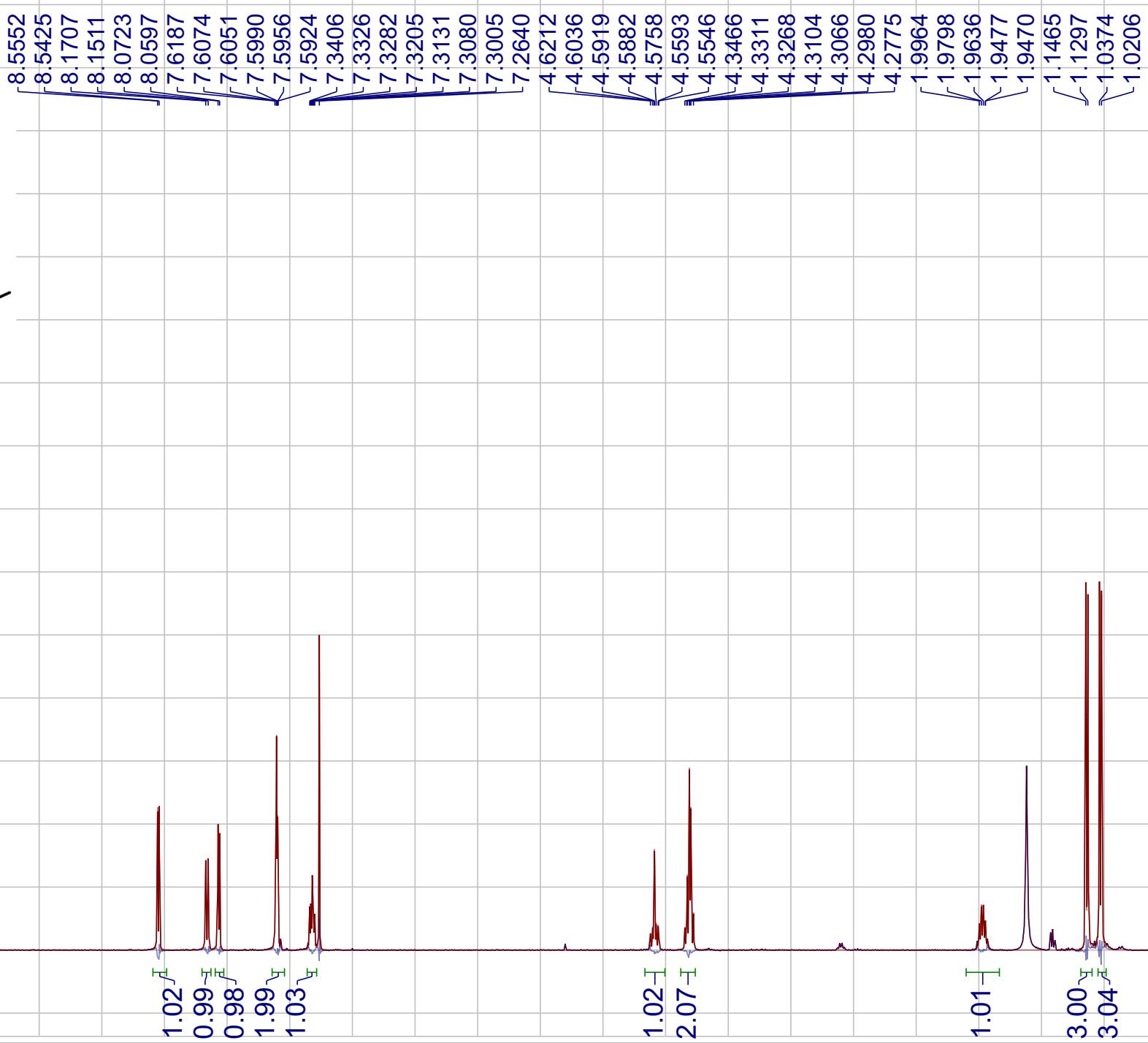




L2/L2-HNMR
K34



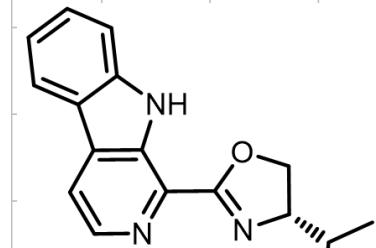
-10.3742



11.5 11.0 10.5 9.5 9.0 8.0 7.0 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5

f1 (ppm)

L2/CNMR L2
K35



-163.0912

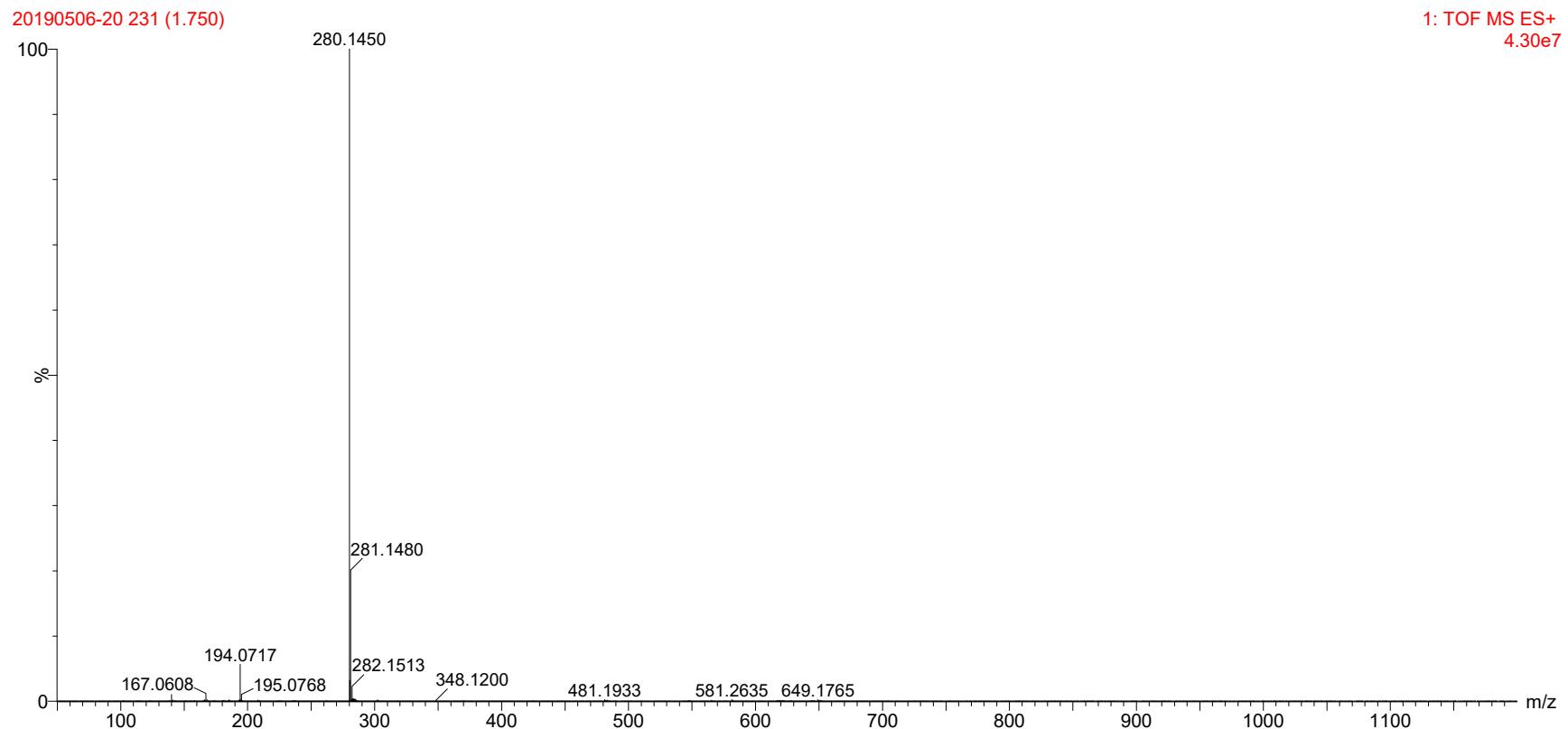
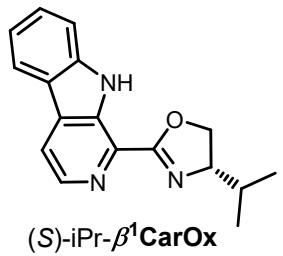
-72.8910
-70.2338

-33.0660

19.1231
18.4774

180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -50

f1 (ppm)



L4/L4 HNMR

K6
 10.335
 8.5274
 8.1404
 8.1379
 8.1289
 8.1206
 8.1182
 8.0394
 8.0268

3800

3600

3400

3200

3000

2800

2600

2400

2200

2000

1800

1600

1400

1200

1000

800

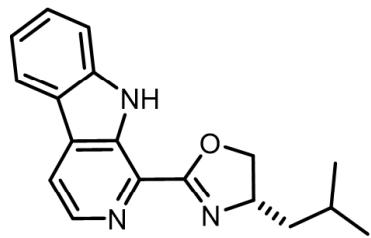
600

400

200

0

-200



2.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5

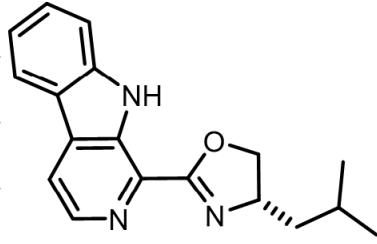
f1 (ppm)

0.96
 1.04
 0.98
 2.07
 1.04

1.03
 1.05
 1.00

1.12
 1.08
 1.05
 6.11

-162.9845



140.5821
138.6762
135.8604
129.9198
129.0944
128.9072
121.9204
121.3300
120.3215
116.9444
111.9558

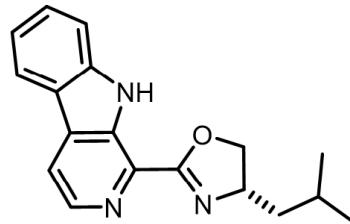
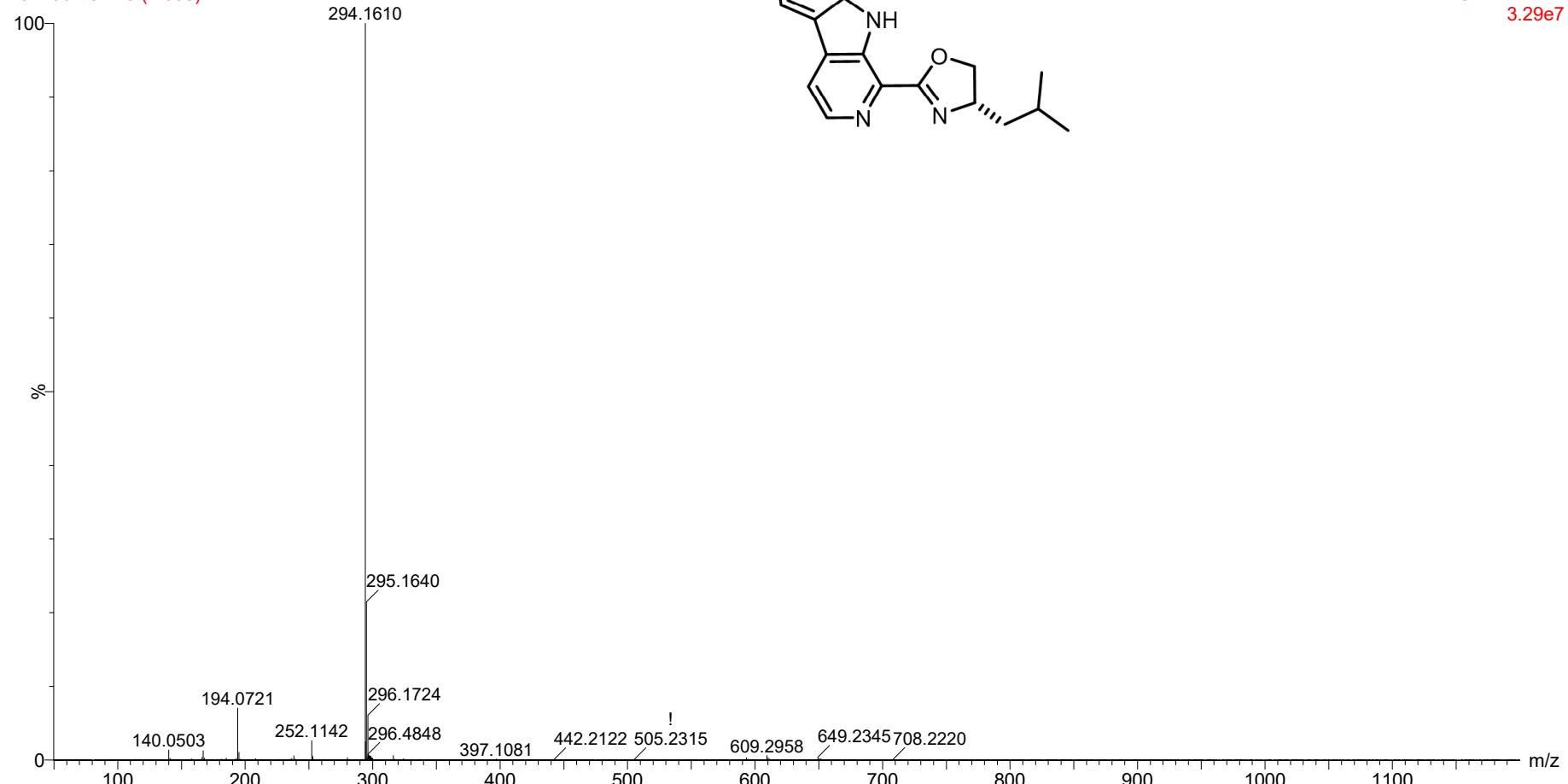
-73.0227

-65.3853

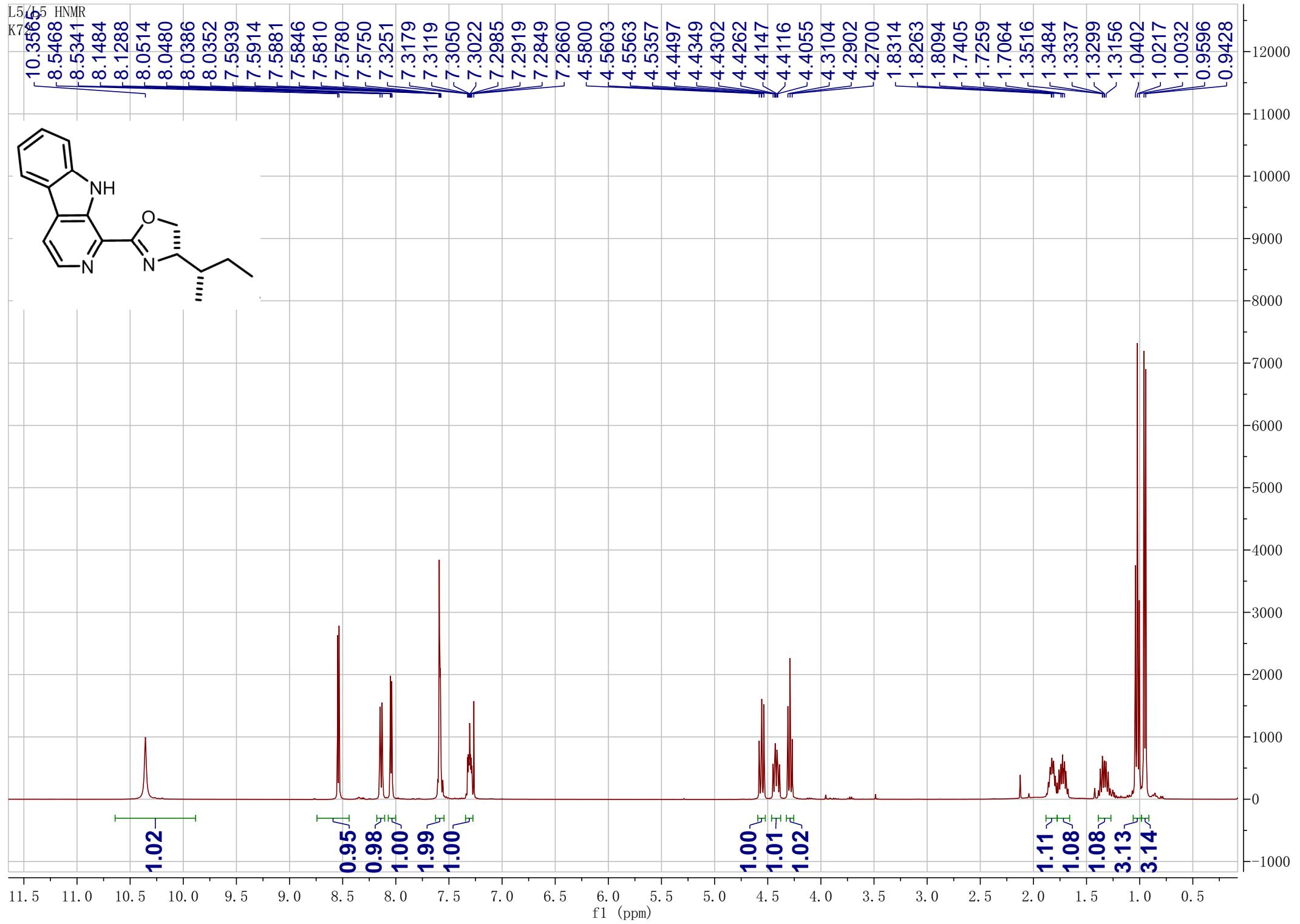
-45.7738

25.7504
22.9152
22.8244

191106-28 215 (1.638)

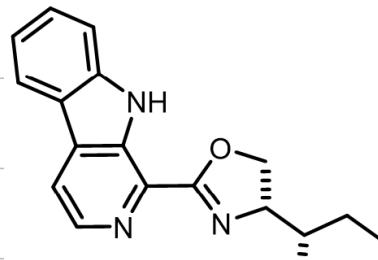


L5/15 HNMR
K7



L5/L5CNMR
K72

-163.0070



140.5785
138.6786
135.9135
129.9167
129.0440
128.9183
121.9420
121.3437
120.3347
116.9675
111.9766

~71.5033
~69.7236

-39.3594

-26.2938

-14.6165
-11.6667

180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

1400
1300
1200
1100
1000
900
800
700
600
500
400
300
200
100
0
-100

191106-29 228 (1.740)

294.1611

100

80

60

0

140.0503

194.0721

195.0749

295.1640

296.1653

382.2133

!

468.6967

!

505.2346

!

609.2955

!

649.2357

!

706.2260

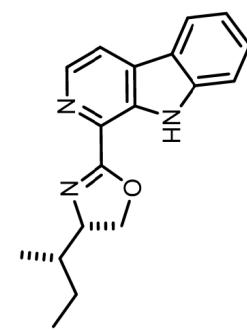
!

779.2517

!

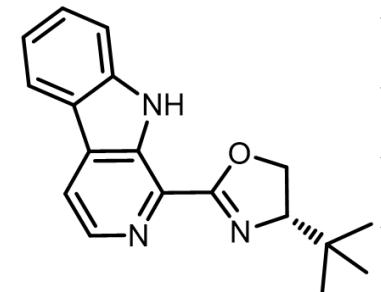
1066.3732

m/z

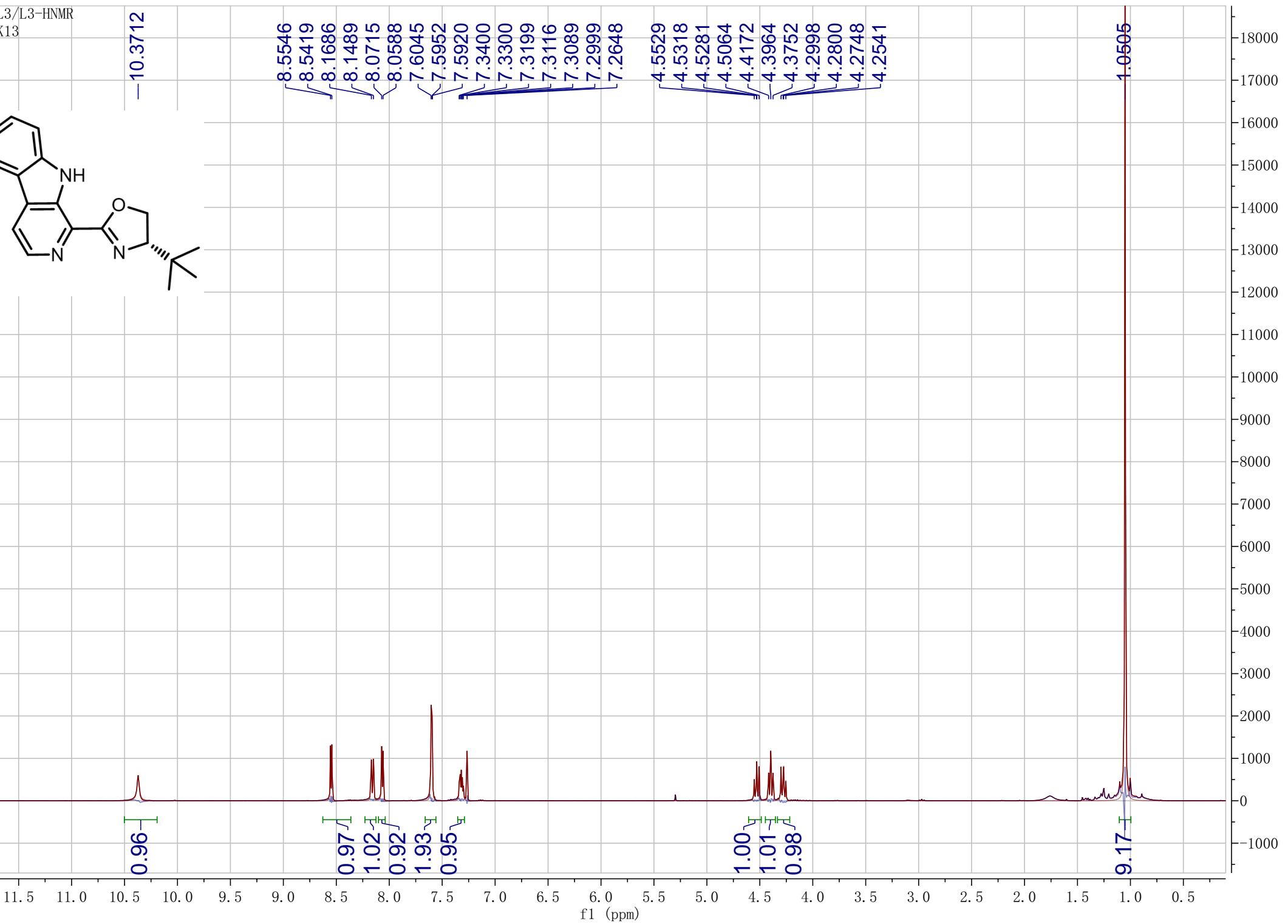


1: TOF MS ES+
3.20e7

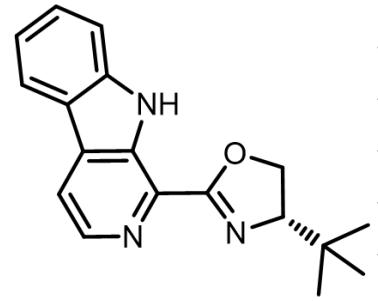
L3/L3-HNMR
K13



-10.3712



L3/L3-CNMR
K36



-163.0793

140.5675
138.7020
136.0016
129.9225
129.0324
128.9273
121.9570
121.3810
120.3590
116.9735
111.9766

-76.4154

-68.6983

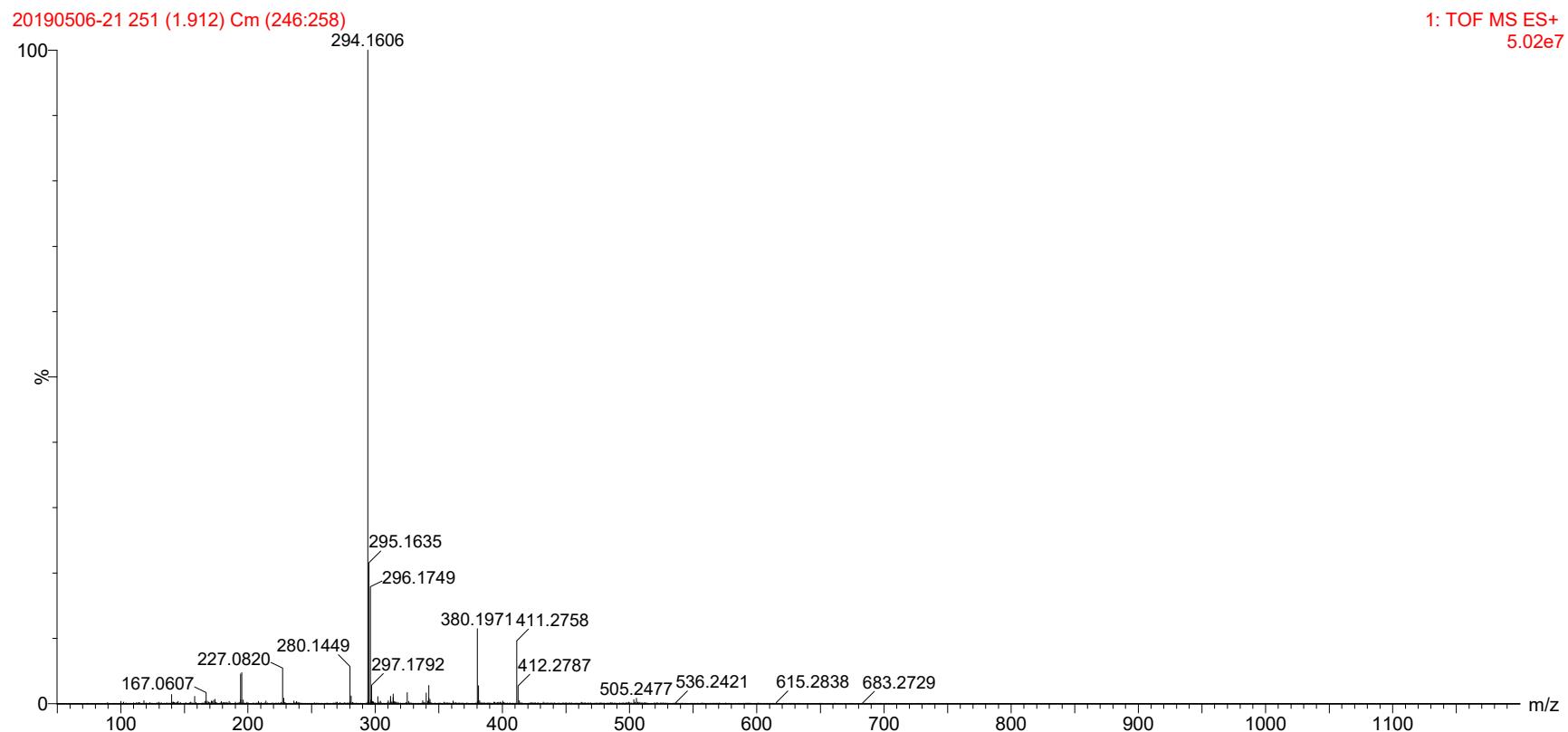
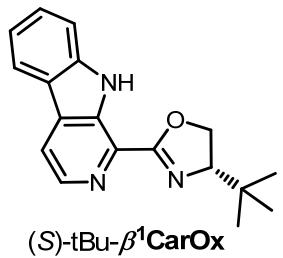
-34.0663

-26.0718

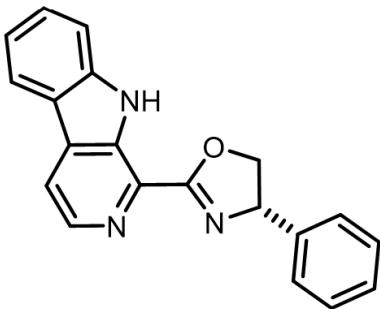
180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

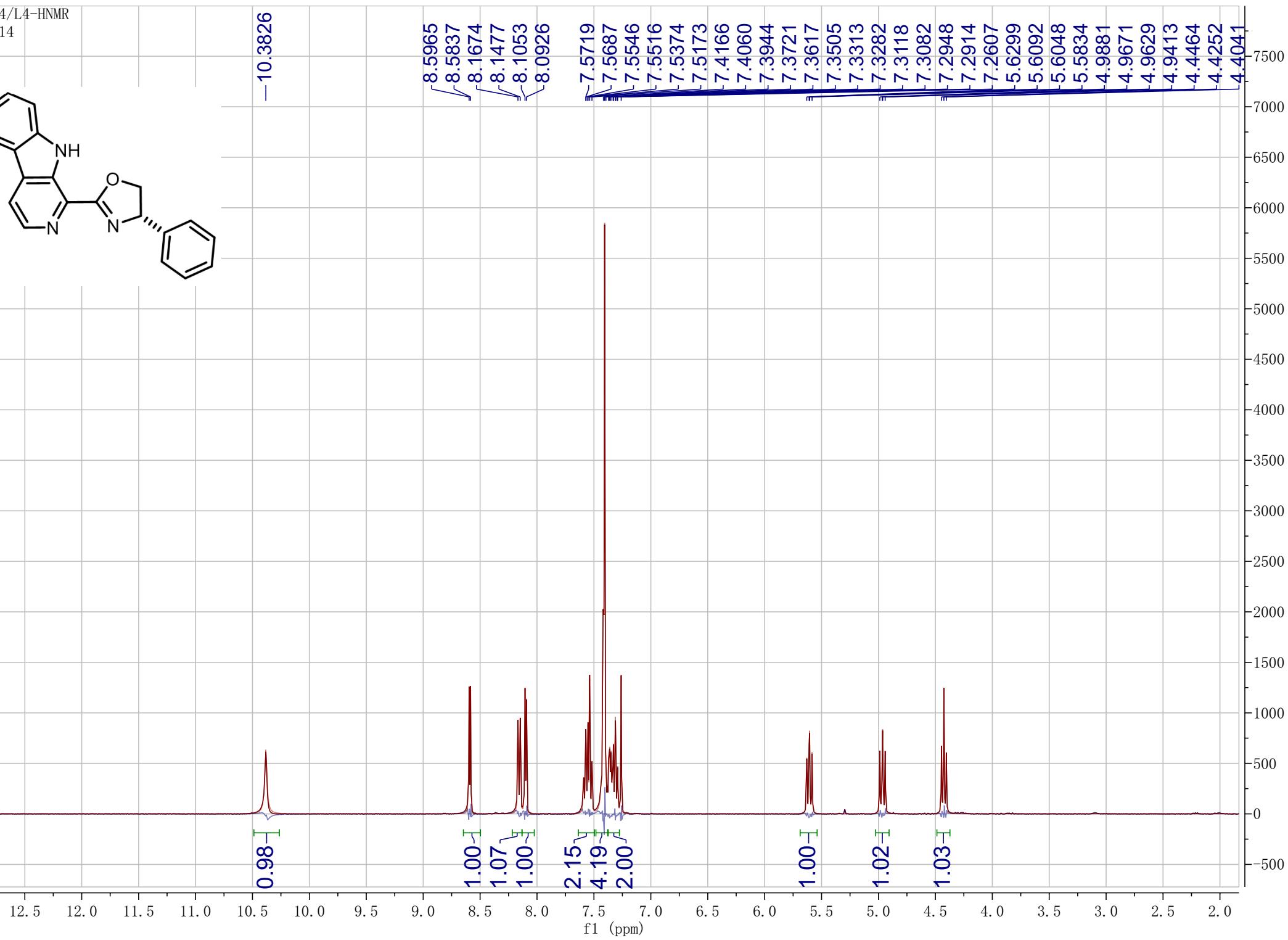
2400
2300
2200
2100
2000
1900
1800
1700
1600
1500
1400
1300
1200
1100
1000
900
800
700
600
500
400
300
200
100
0
-100
-200

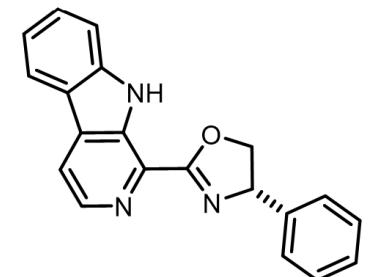


L4/L4-HNMR
K14



-10.3826



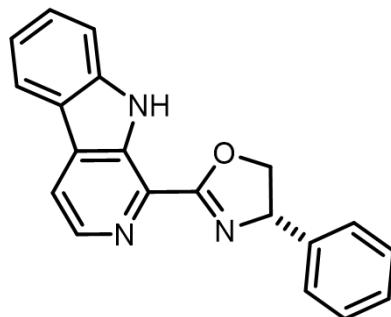
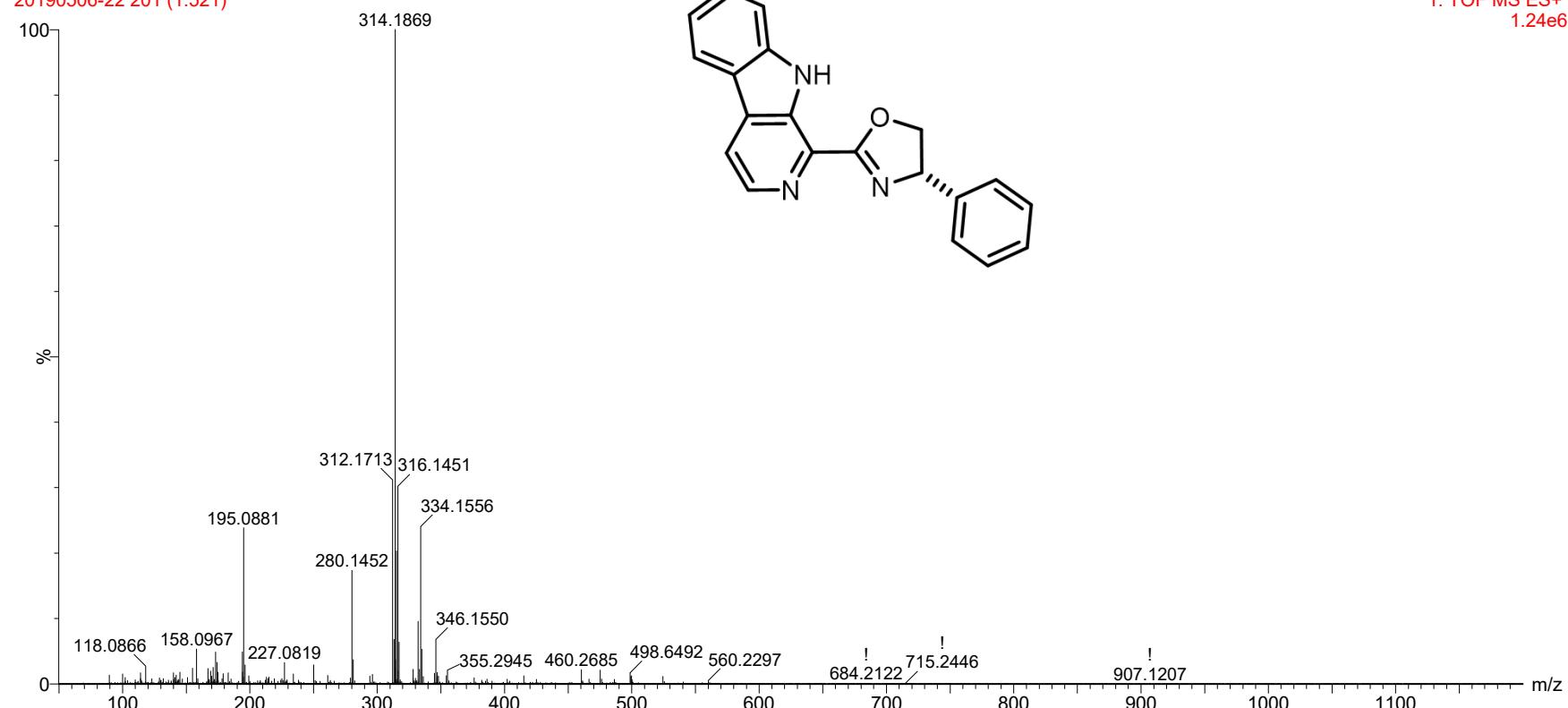


-164.5194

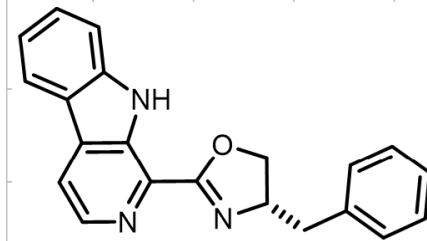
142.1747
140.6450
138.7635
136.1156
130.1164
129.0132
128.9539
128.6423
127.9762
126.9113
121.9080
121.2789
120.3967
117.2657
112.0251

-74.8059
-70.4367

20190506-22 201 (1.521)

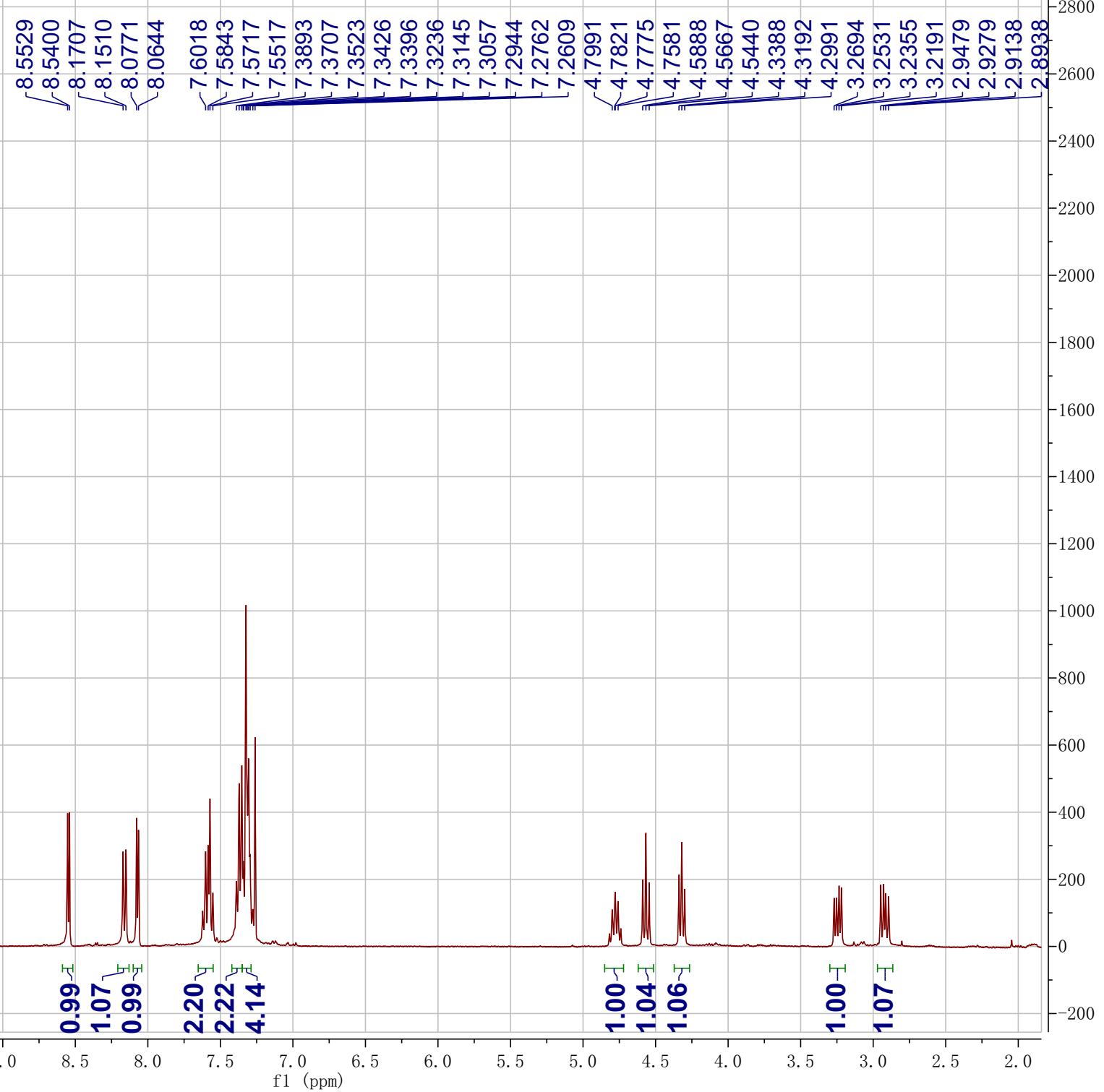


L8/HNMR-L8
K38

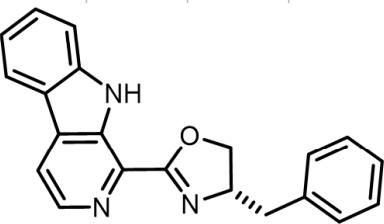


-10.3461

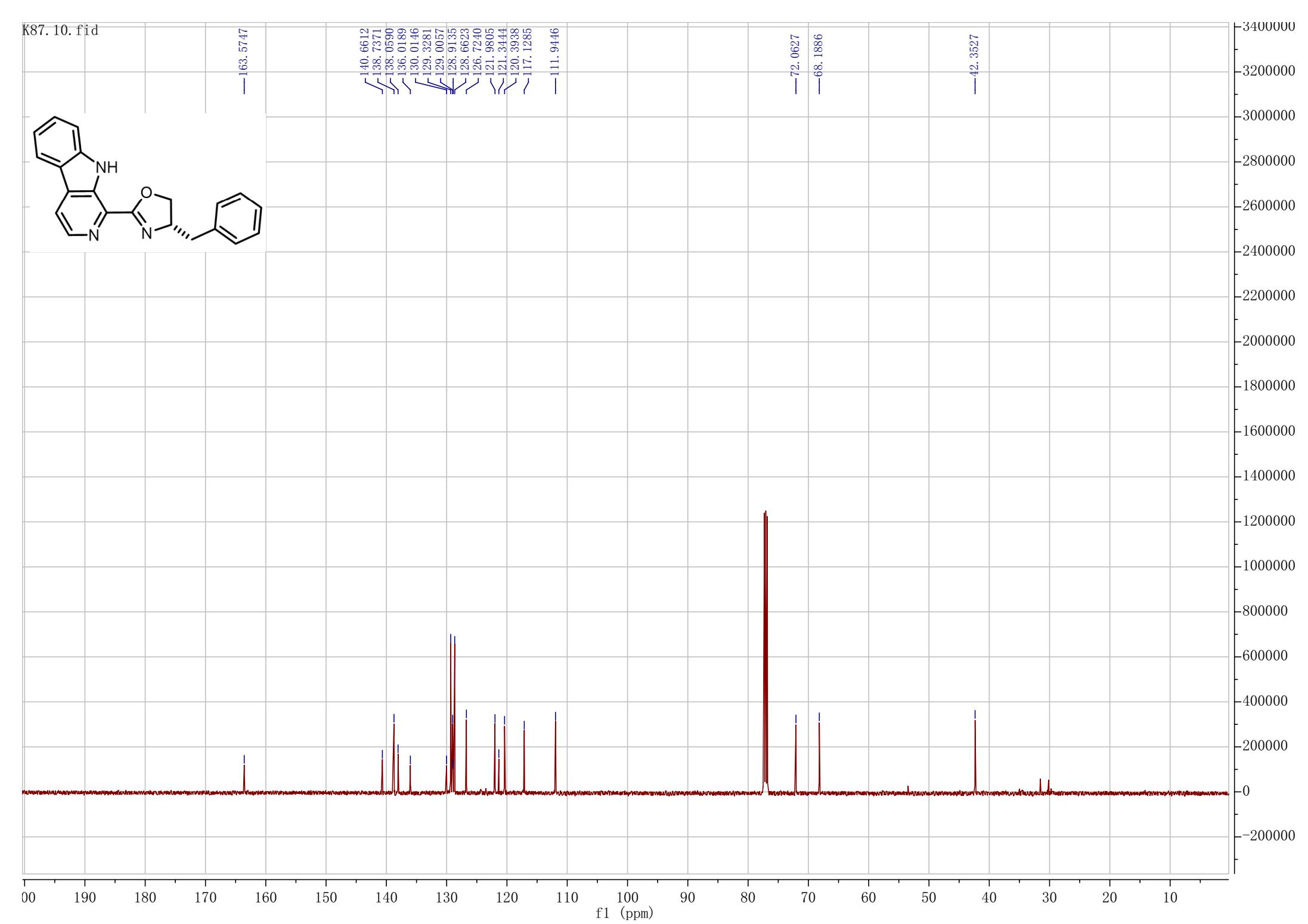
1.09



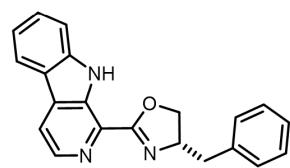
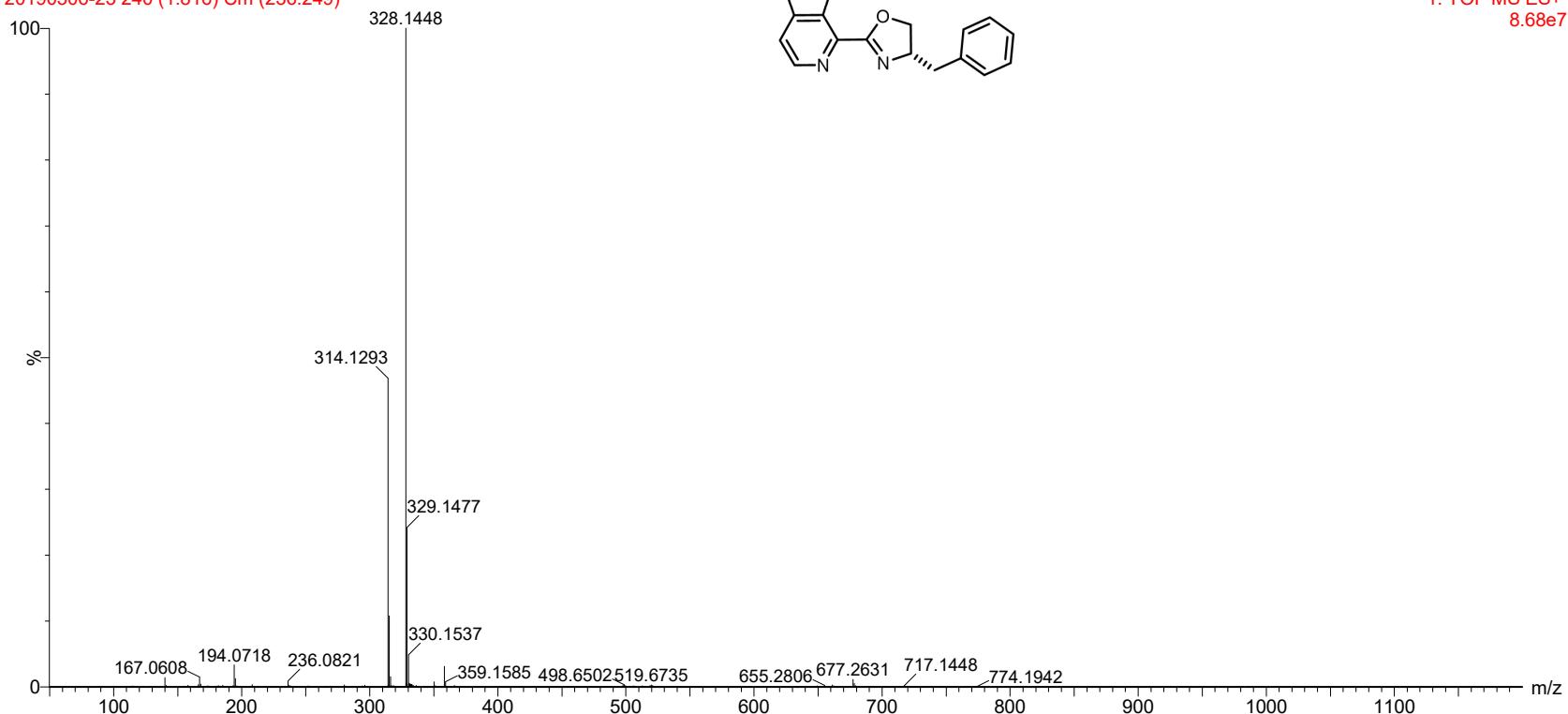
K87. 10. fid



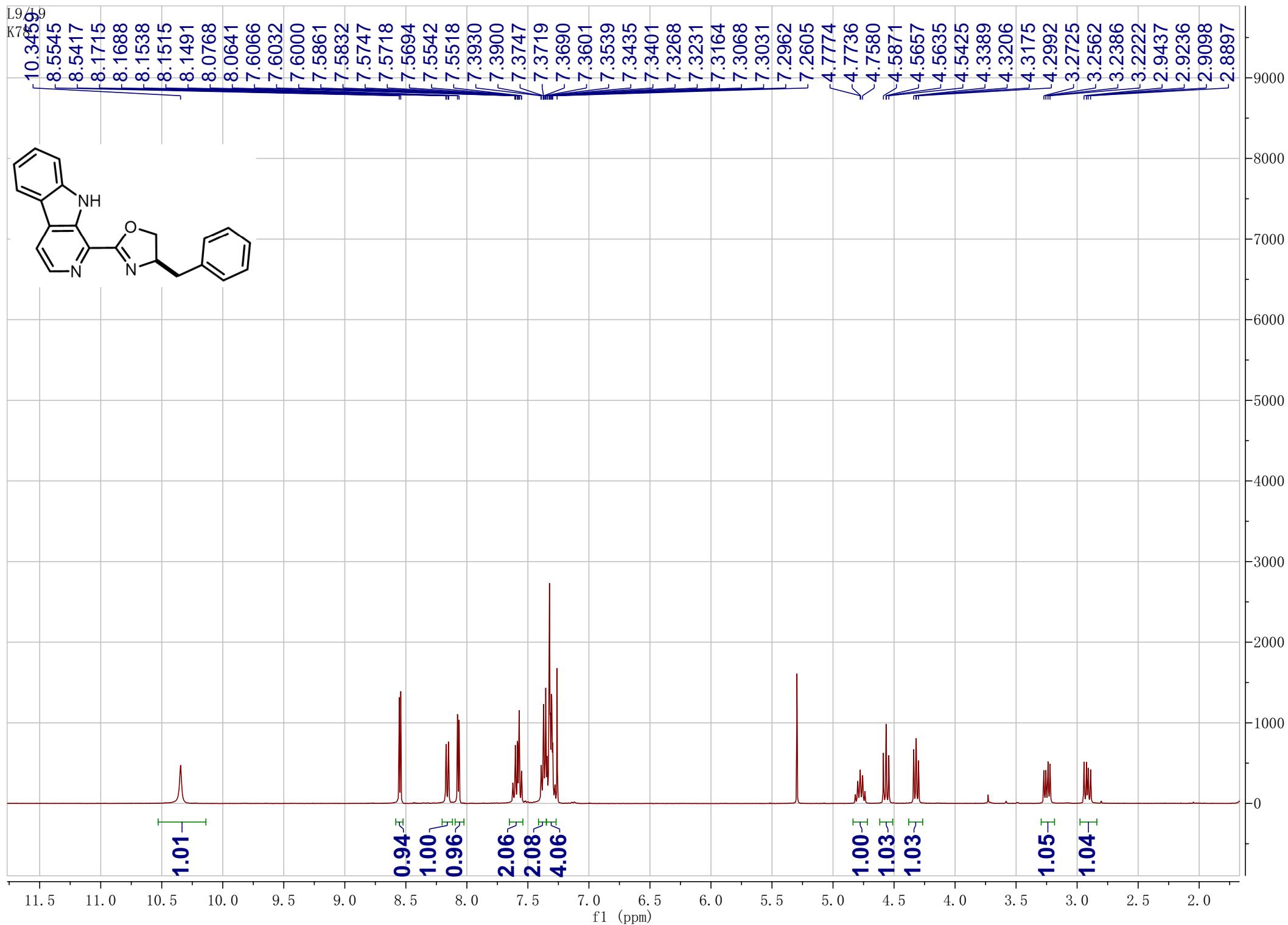
— 163. 5747

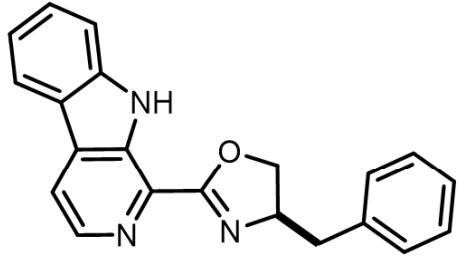


20190506-23 240 (1.816) Cm (236:249)



1: TOF MS ES+
8.68e7





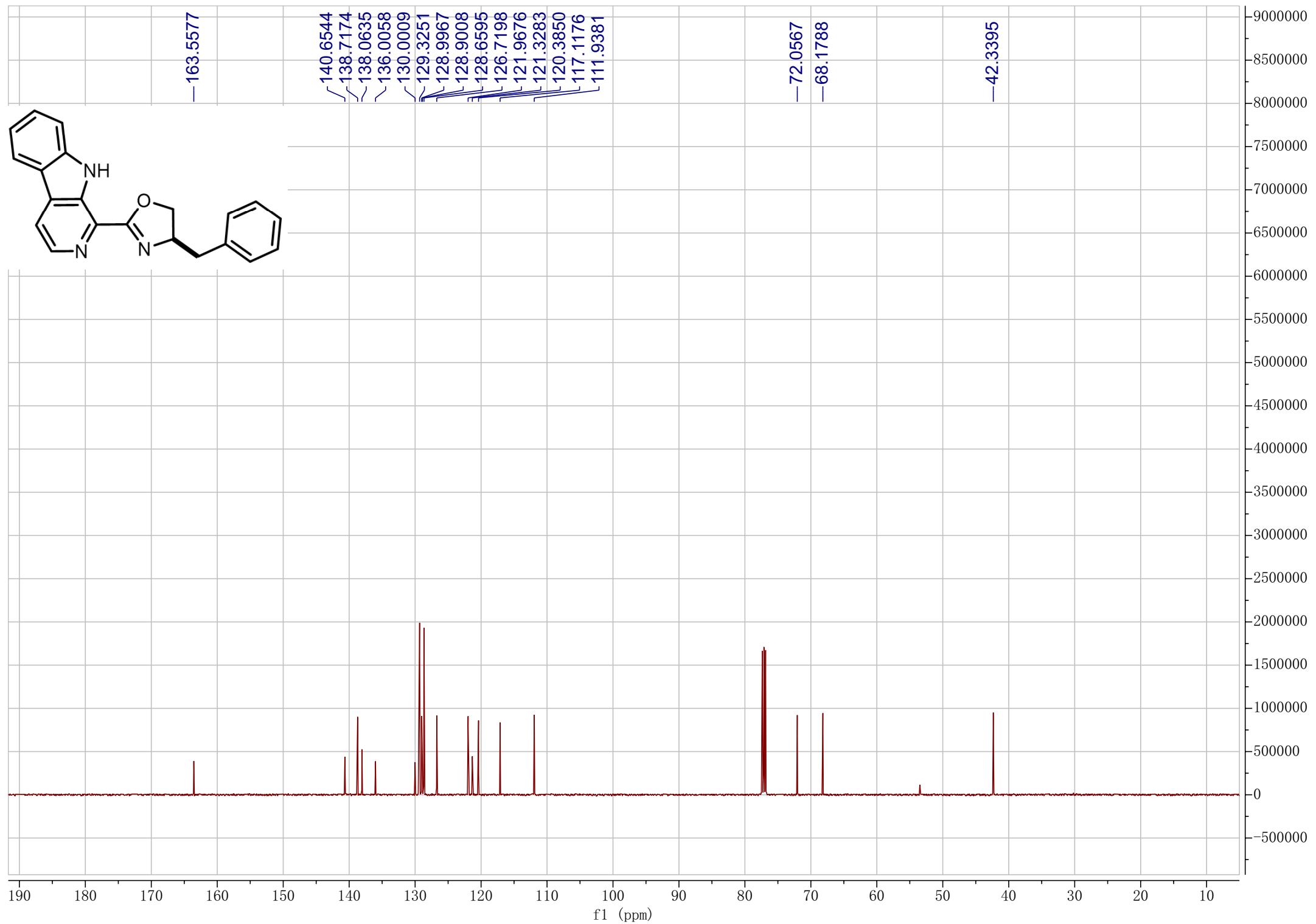
-163.5577

140.6544
138.7174
138.0635
136.0058
130.0009
129.3251
128.9967
128.9008
128.6595
126.7198
121.9676
121.3283
120.3850
117.1176
111.9381

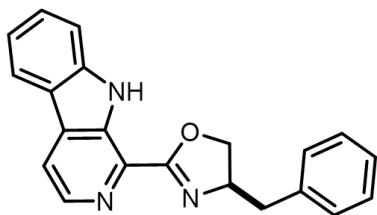
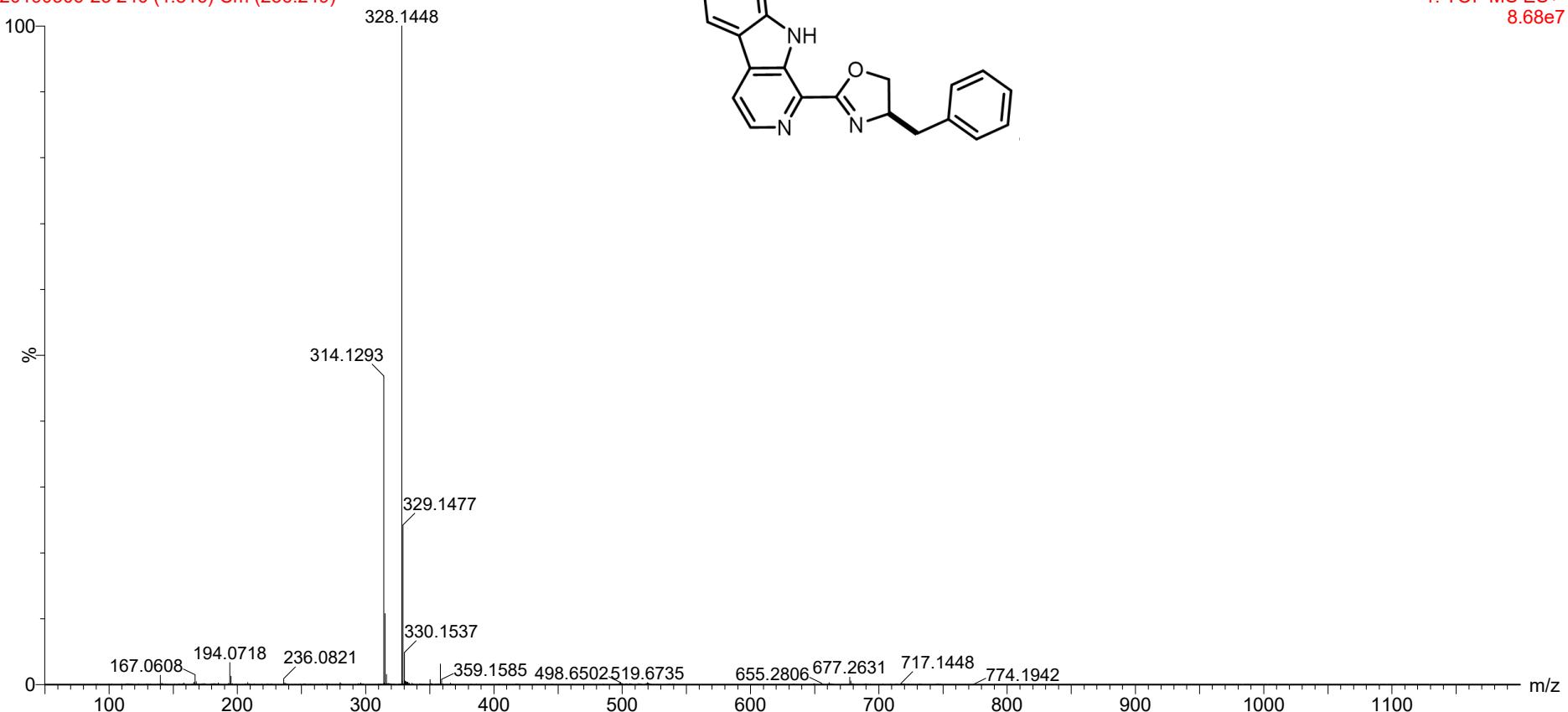
-72.0567

-68.1788

-42.3395

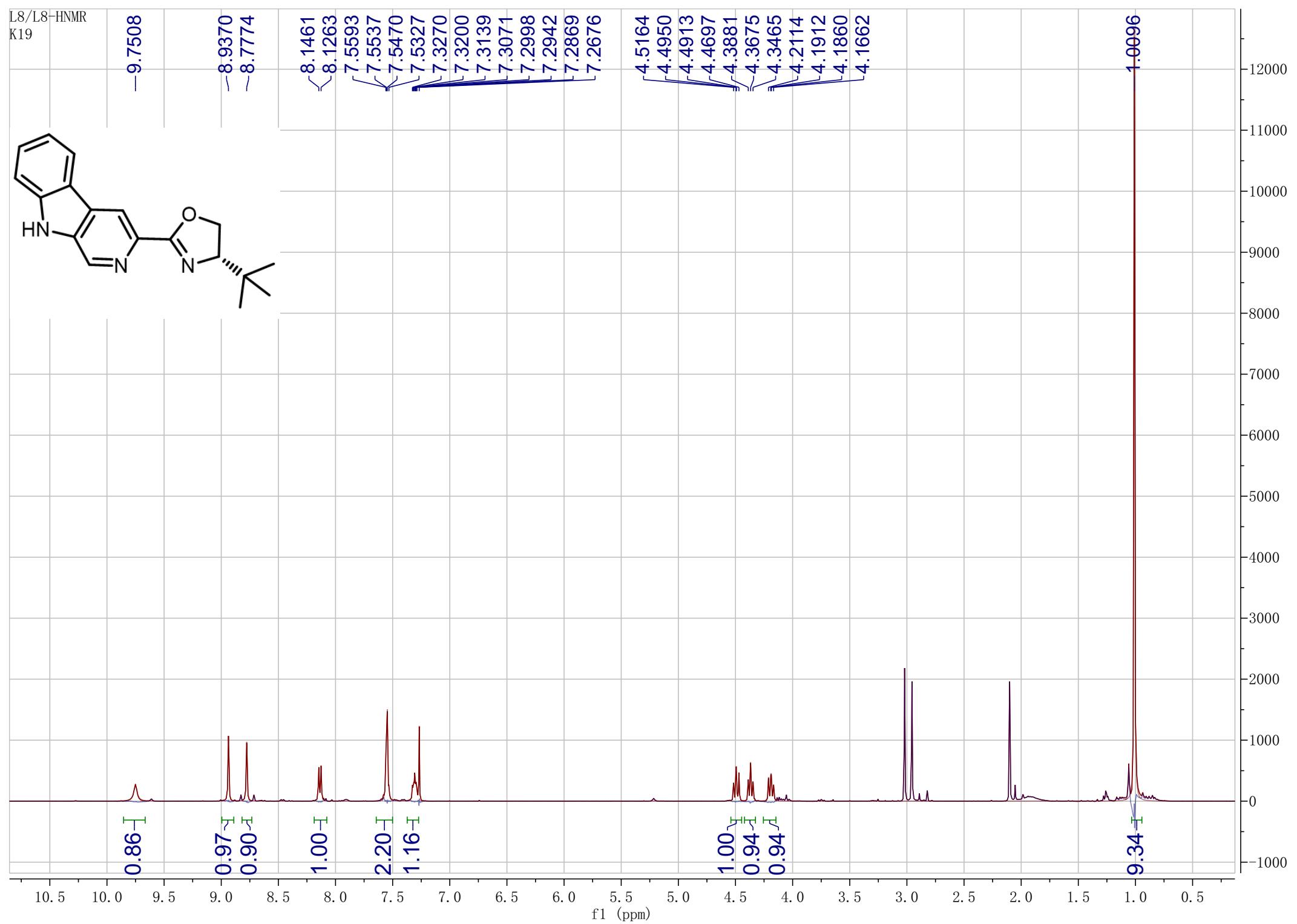
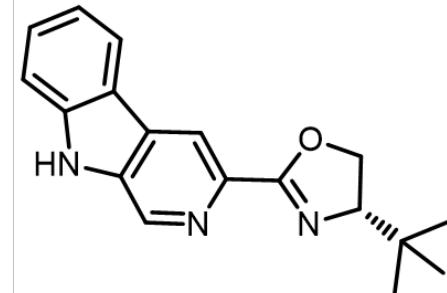


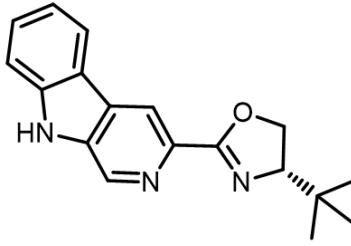
20190506-23 240 (1.816) Cm (236:249)



1: TOF MS ES+
8.68e7

L8/L8-HNMR
K19





-163.6157

✓140.9838
✓136.8566
✓136.0727
-133.3482
✓129.0621
✓128.8705
✓122.0086
✓121.5207
✓120.6455
✓116.1772
~112.0195

-76.3482

-69.3060

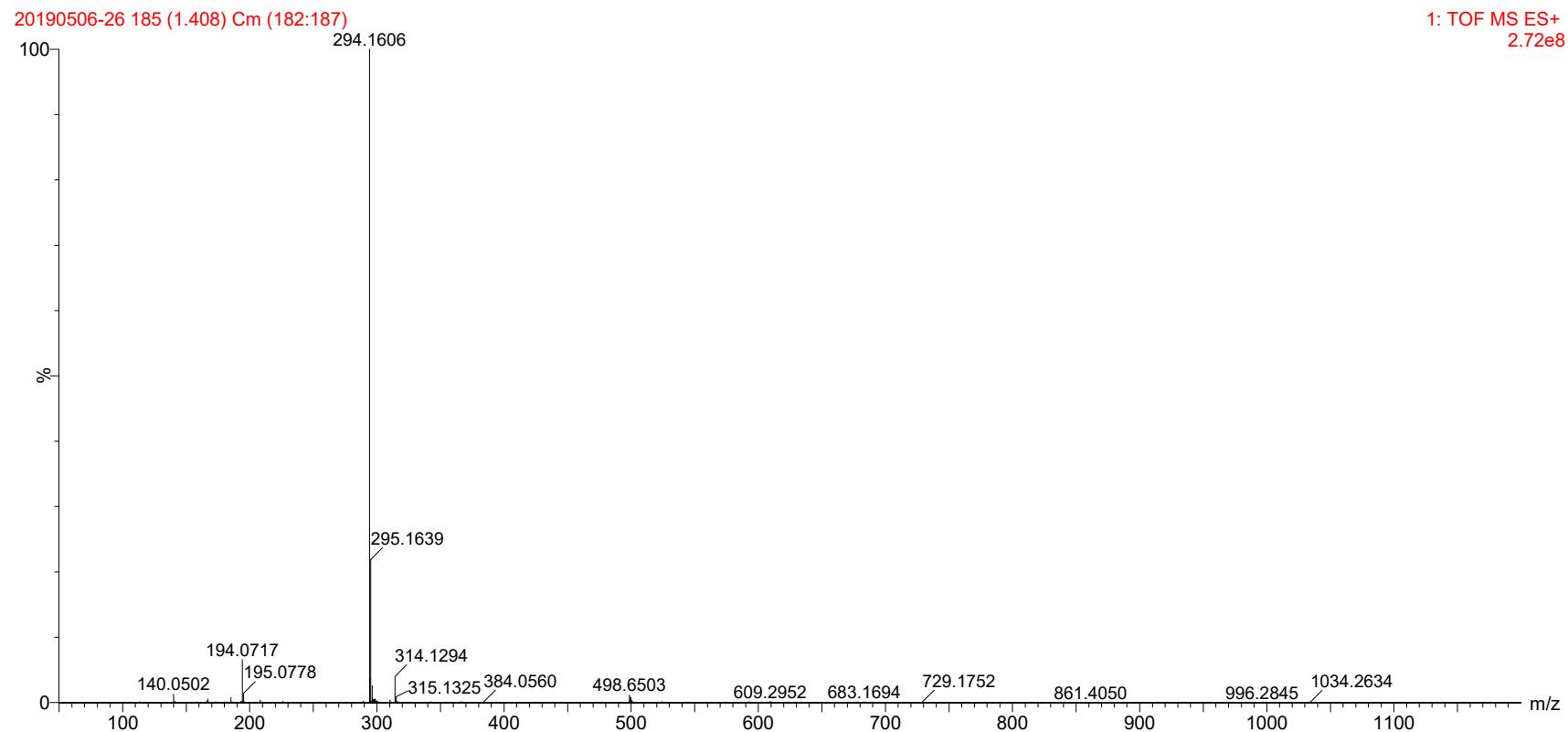
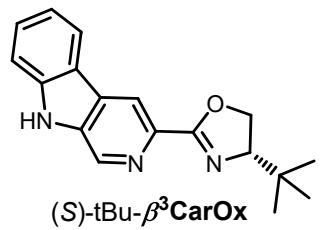
-34.0790

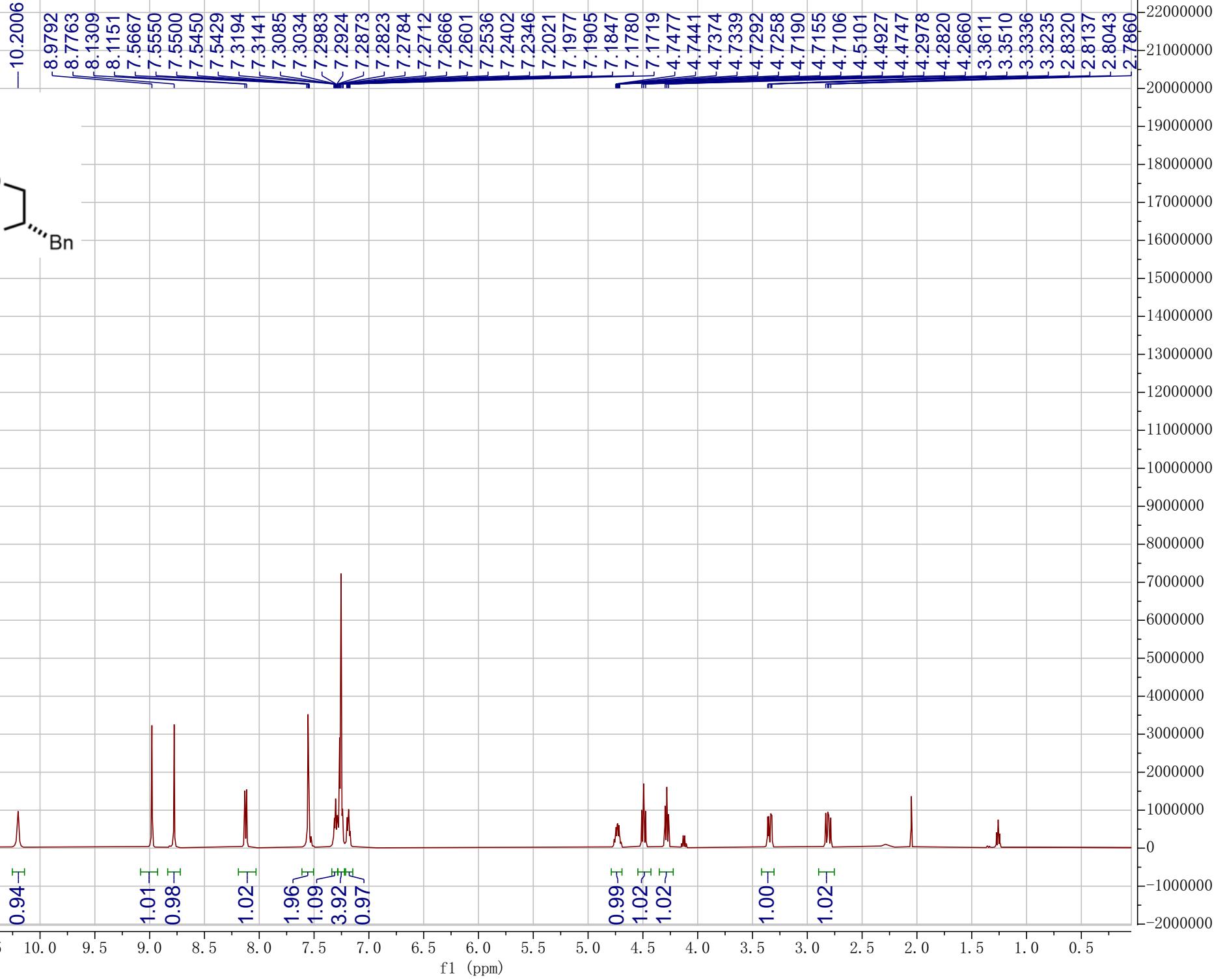
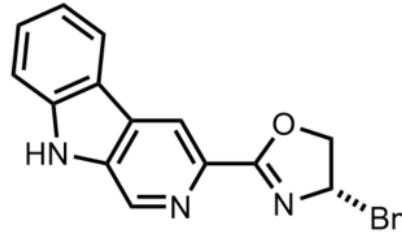
-26.0122

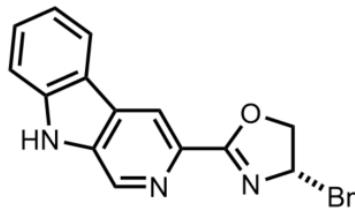
180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

2500000
2200000
2100000
2000000
1900000
1800000
1700000
1600000
1500000
1400000
1300000
1200000
1100000
1000000
900000
800000
700000
600000
500000
400000
300000
200000
100000
0
-100000
-200000







-164.4229

141.1389
137.9103
137.0980
135.7017
133.5921
129.2301
128.9589
128.8387
128.6048
126.5508
121.9164
121.4749
120.5824
116.2400
112.1412

-72.5099

-67.9847

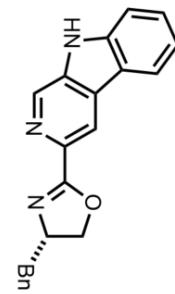
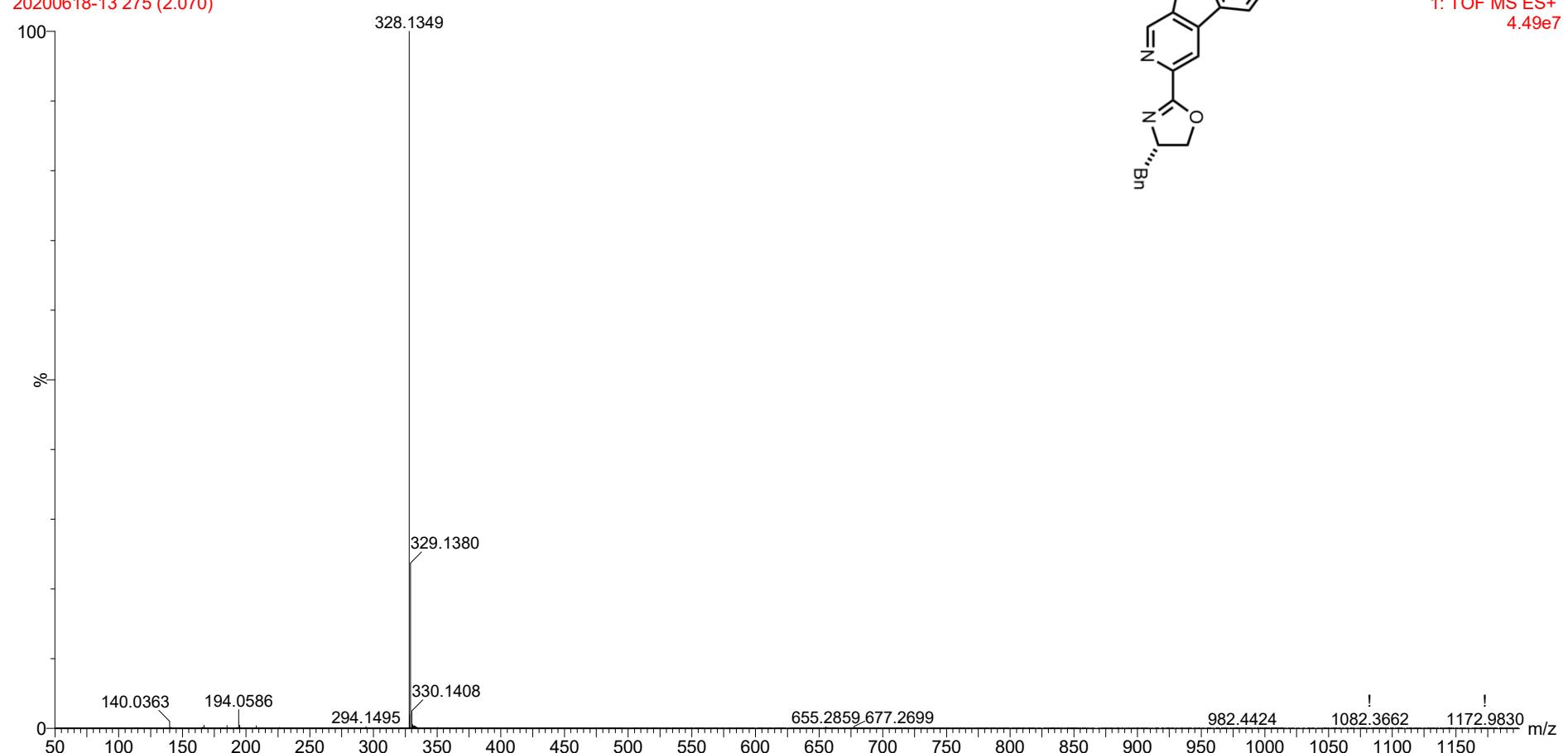
-41.9661

190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -500000

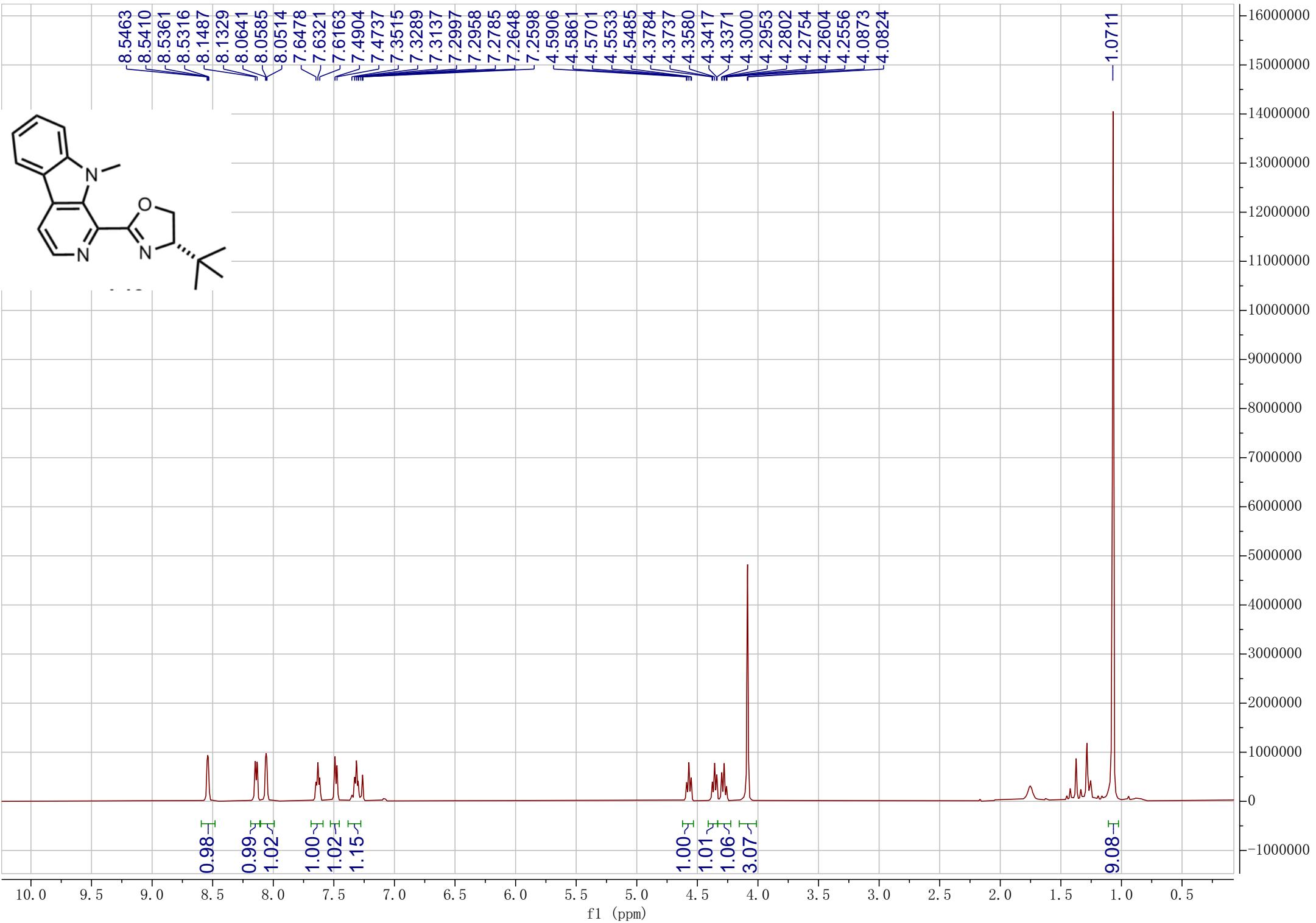
f1 (ppm)

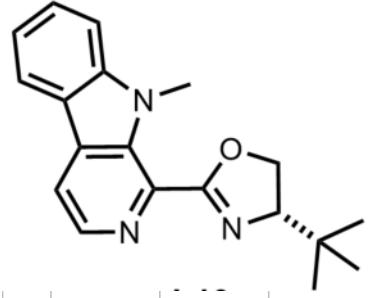
7500000
7000000
6500000
6000000
5500000
5000000
4500000
4000000
3500000
3000000
2500000
2000000
1500000
1000000
500000
0 -500000

20200618-13 275 (2.070)



1: TOF MS ES+
4.49e7





-162.3166

-143.0603
✓ 138.3044
✓ 135.7897
✓ 131.3426
✓ 131.1376
-128.9995
✓ 121.4610
✓ 120.8174
✓ 120.1580
✓ 116.2352
-109.8608

-77.3819

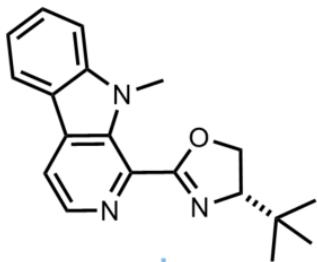
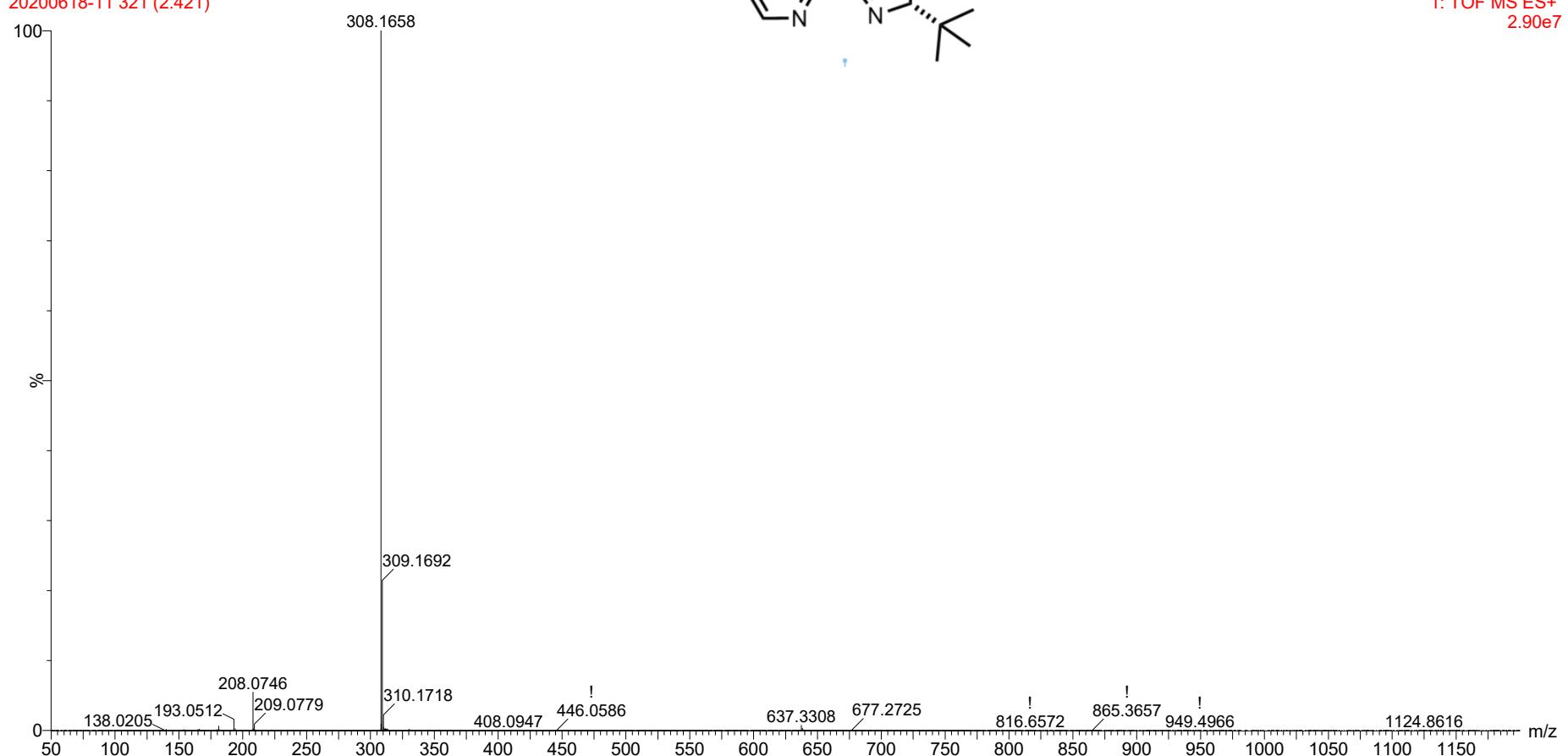
-68.9142

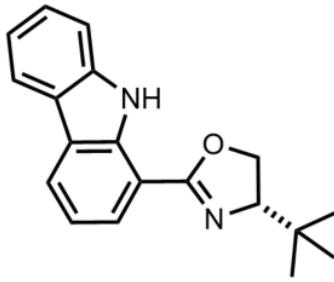
✓ 33.8958
✓ 32.9351
-26.3446

180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

20200618-11 321 (2.421)





-10.5459

8.2008
8.1857
8.1104
8.0948
7.8684
7.8662
7.8532
7.8512
7.5521
7.5360
7.4724
7.4747
7.4582
7.4558
7.4442
7.4418
7.2729
7.2749
7.2612
7.2586
7.2529
7.2461
7.2433
7.2310
4.4247
4.4078
4.4047
4.3880
4.3015
4.2857
4.2692
4.2186
4.2031
4.1987
4.1833

1.0281

0.98

0.93
0.94
0.96
0.99
0.97
2.07

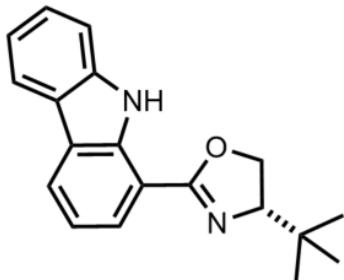
1.00
1.03
1.01

9.02

12.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5

f1 (ppm)

2.8 × 10⁷
2.6 × 10⁷
2.4 × 10⁷
2.2 × 10⁷
2.0 × 10⁷
1.8 × 10⁷
1.6 × 10⁷
1.4 × 10⁷
1.2 × 10⁷
1.0 × 10⁷
8.0 × 10⁶
6.0 × 10⁶
4.0 × 10⁶
2.0 × 10⁶
0.0
-2.0 × 10⁶



-163.1049

139.5738
139.1566
126.1670
125.2715
123.6459
123.4039
122.9284
120.4651
119.5950
118.3310
111.2512
109.5320

-76.1612

-68.1163

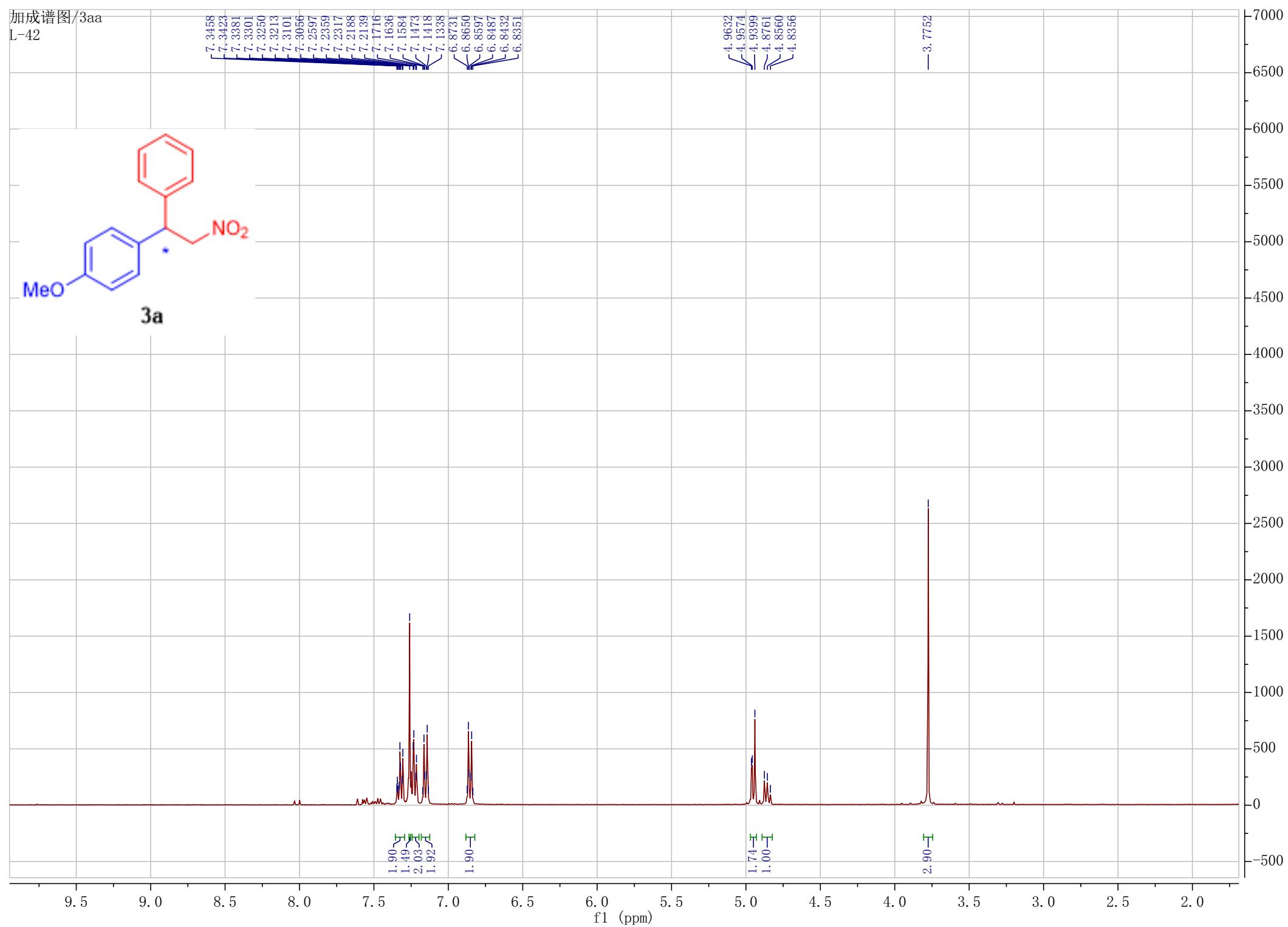
-34.0769

26.0350

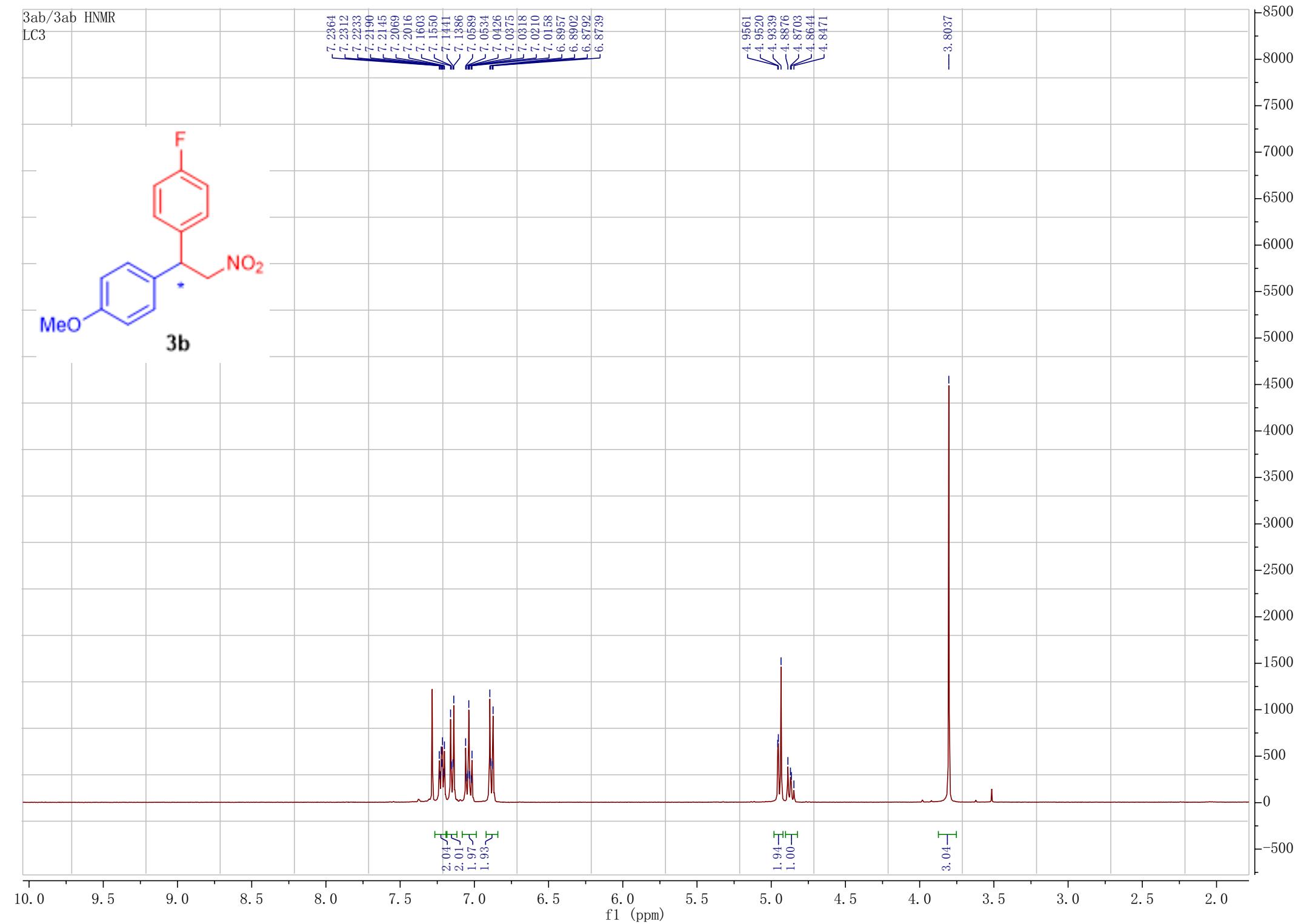
180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

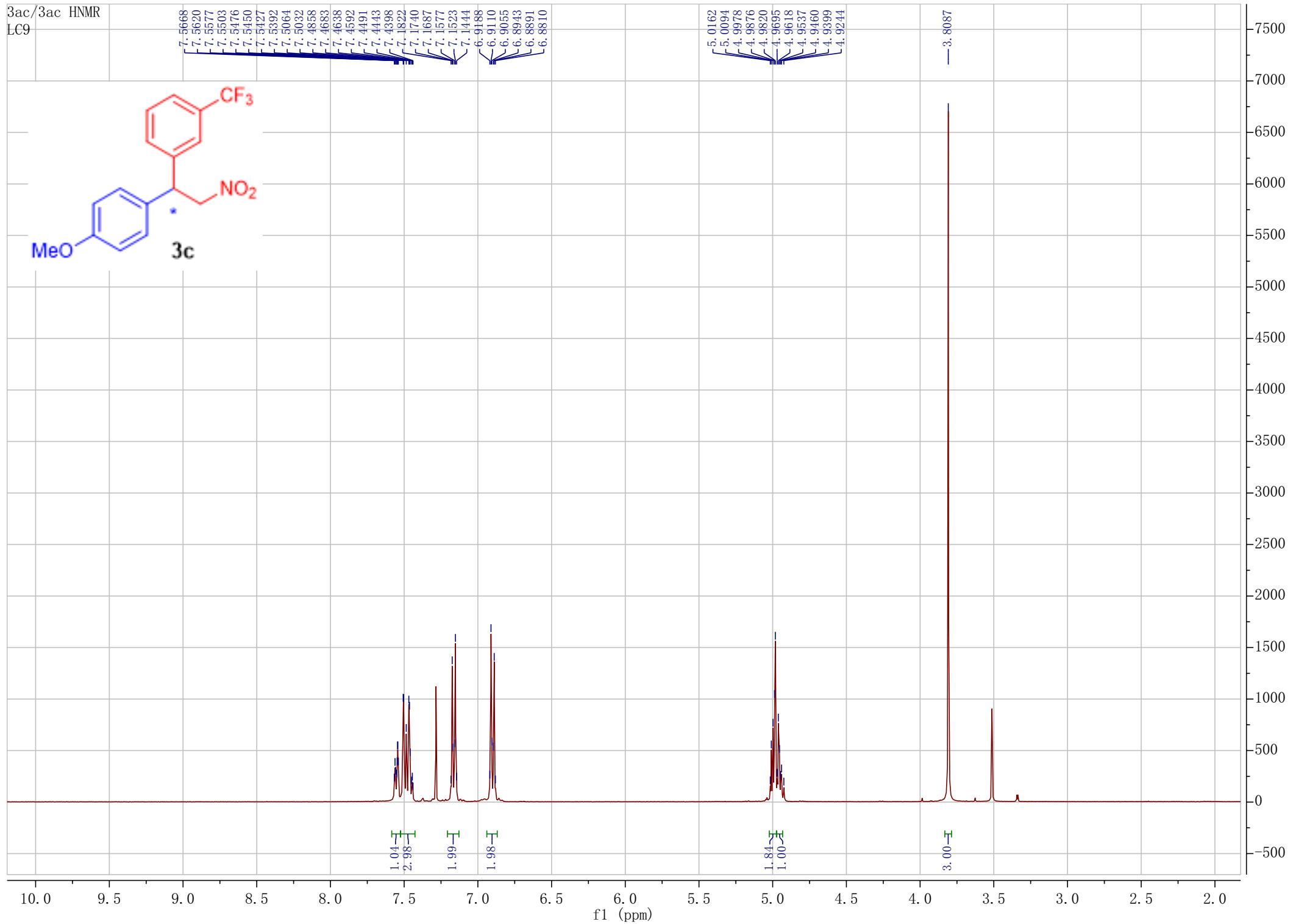
f1 (ppm)

1200000
1100000
1000000
900000
800000
700000
600000
500000
400000
300000
200000
100000
0
-100000

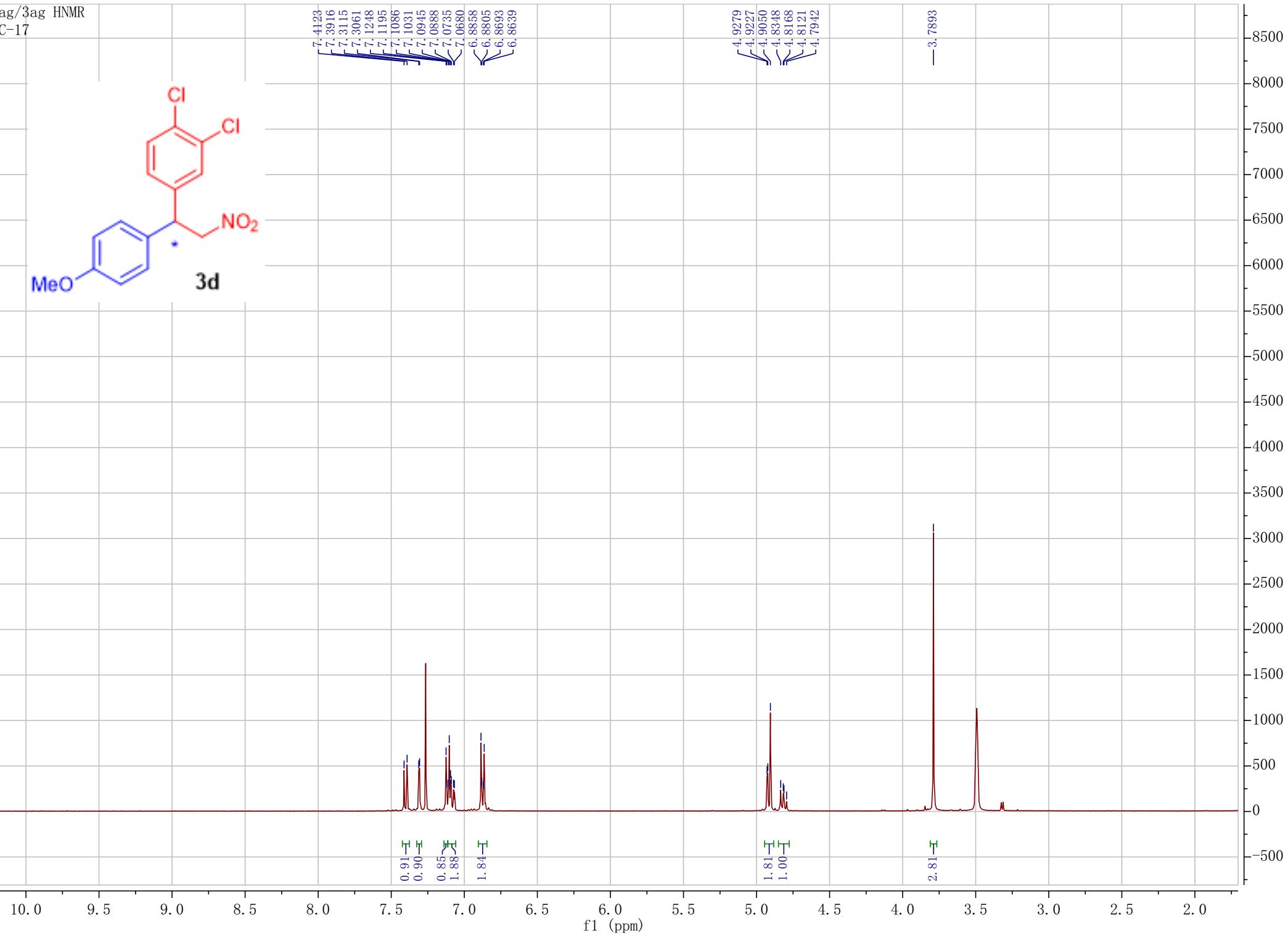
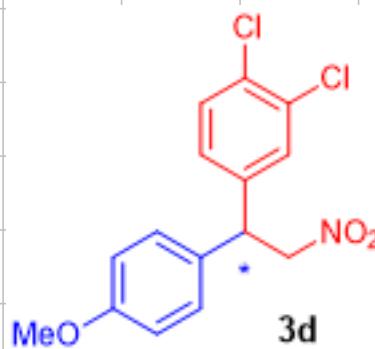


3ab/3ab HNMR
LC3

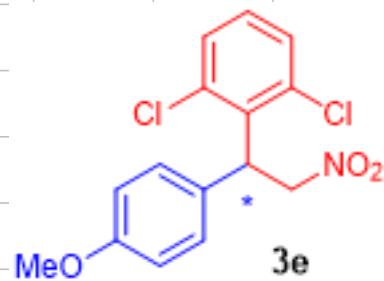




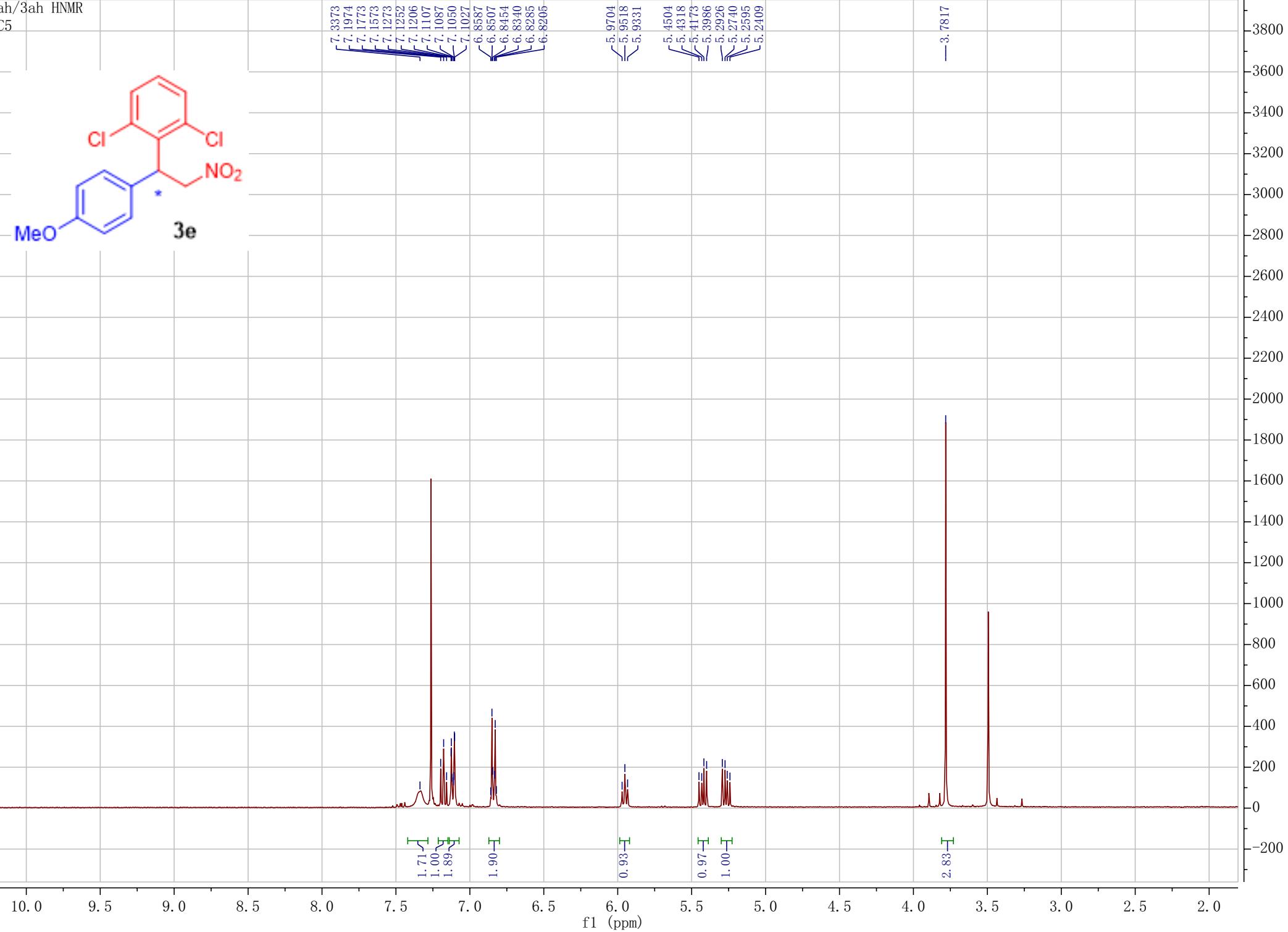
3ag/3ag HNMR
LC-17



3ah/3ah HNMR
LC5

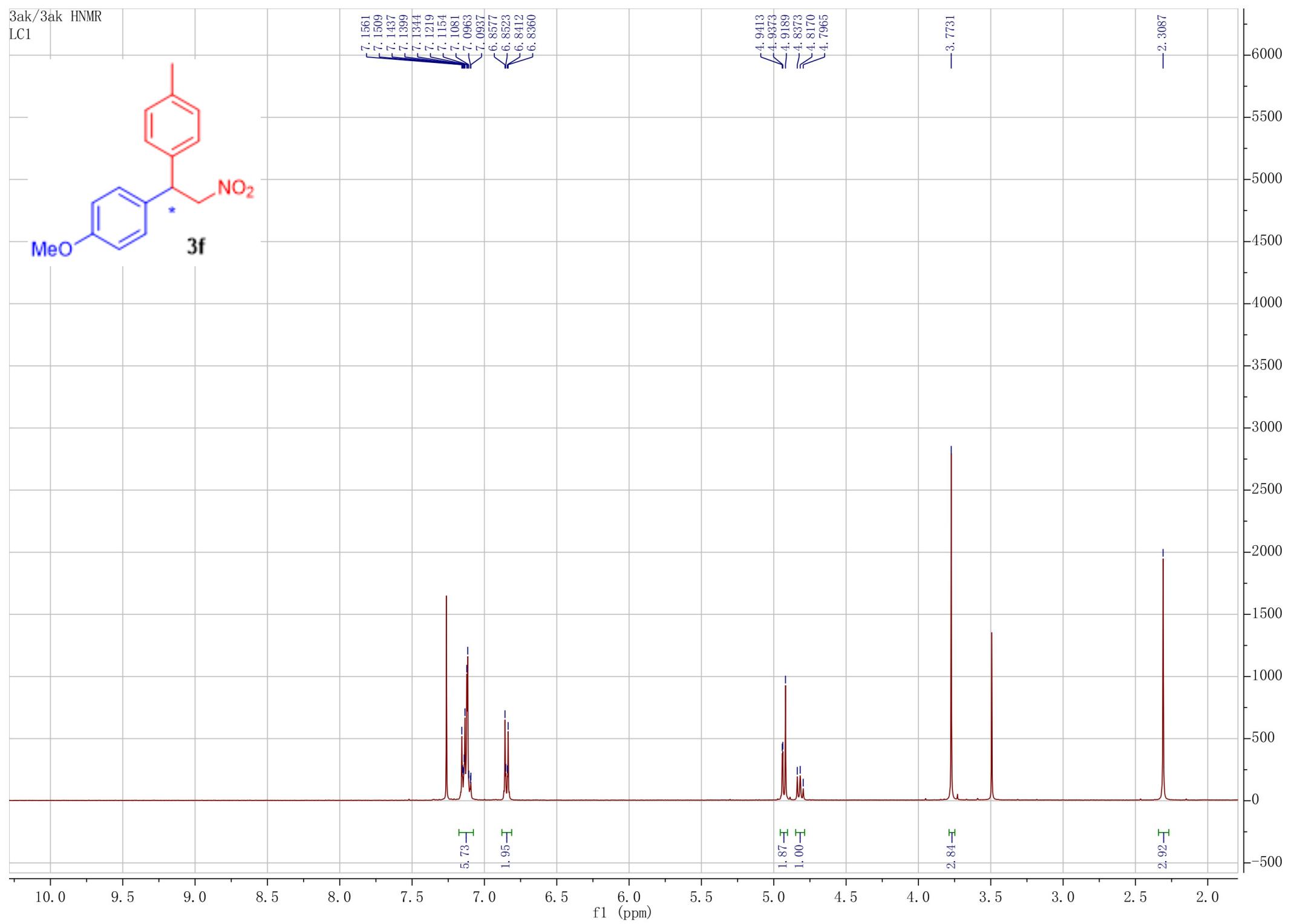
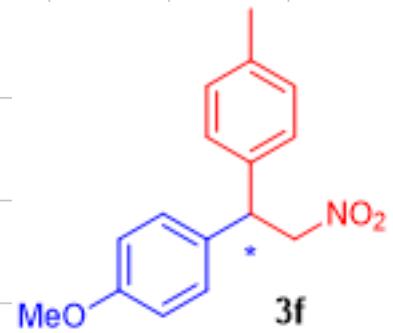


3e

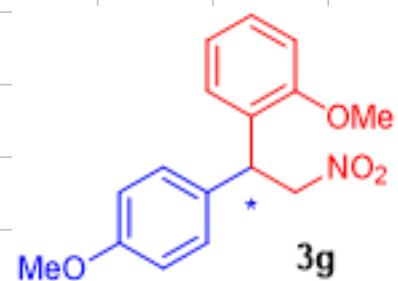


3ak/3ak HNMR

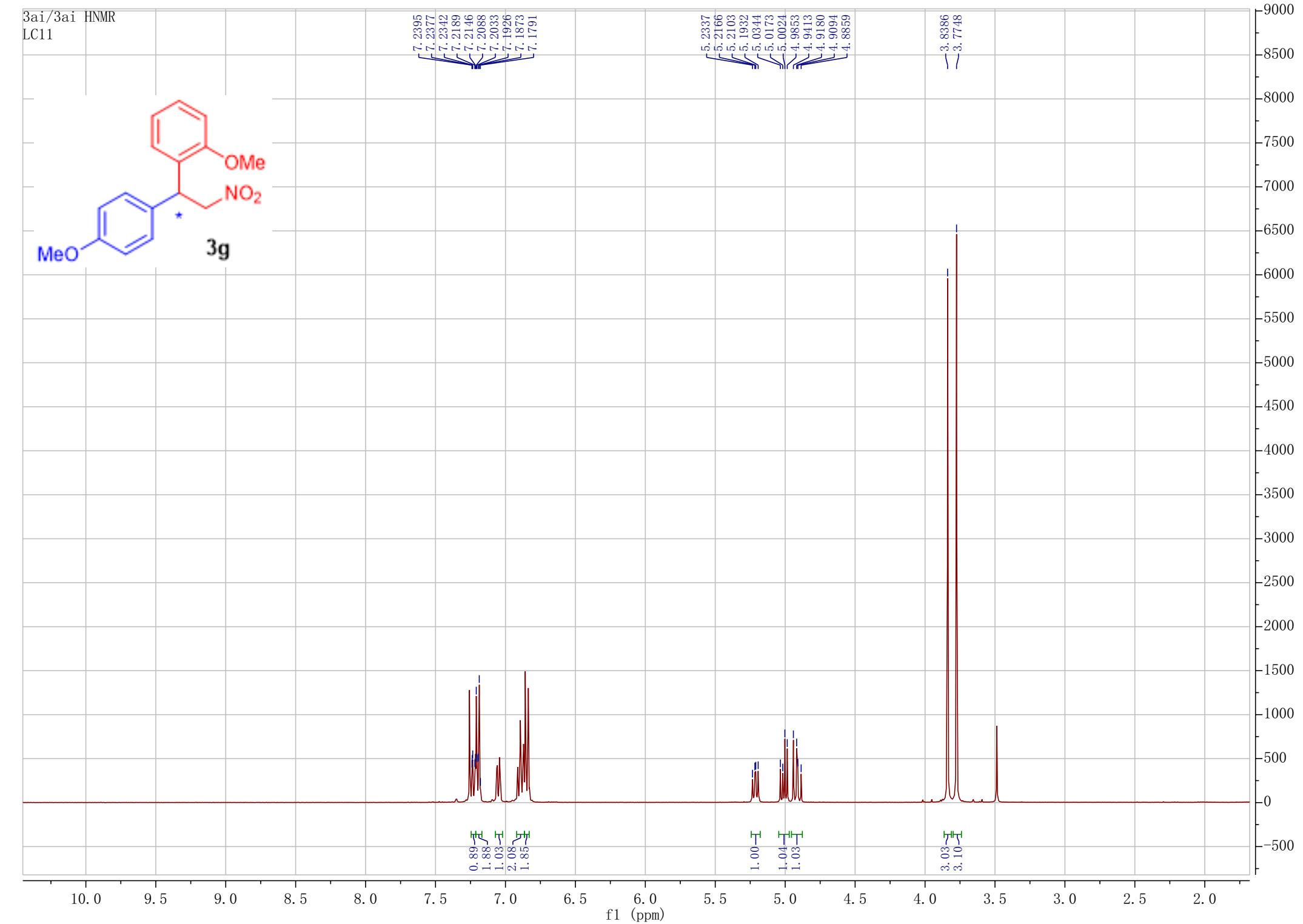
LC1



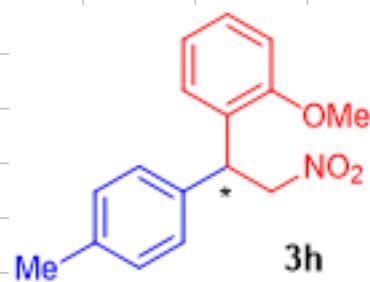
3ai/3ai HNMR
LC11



3g

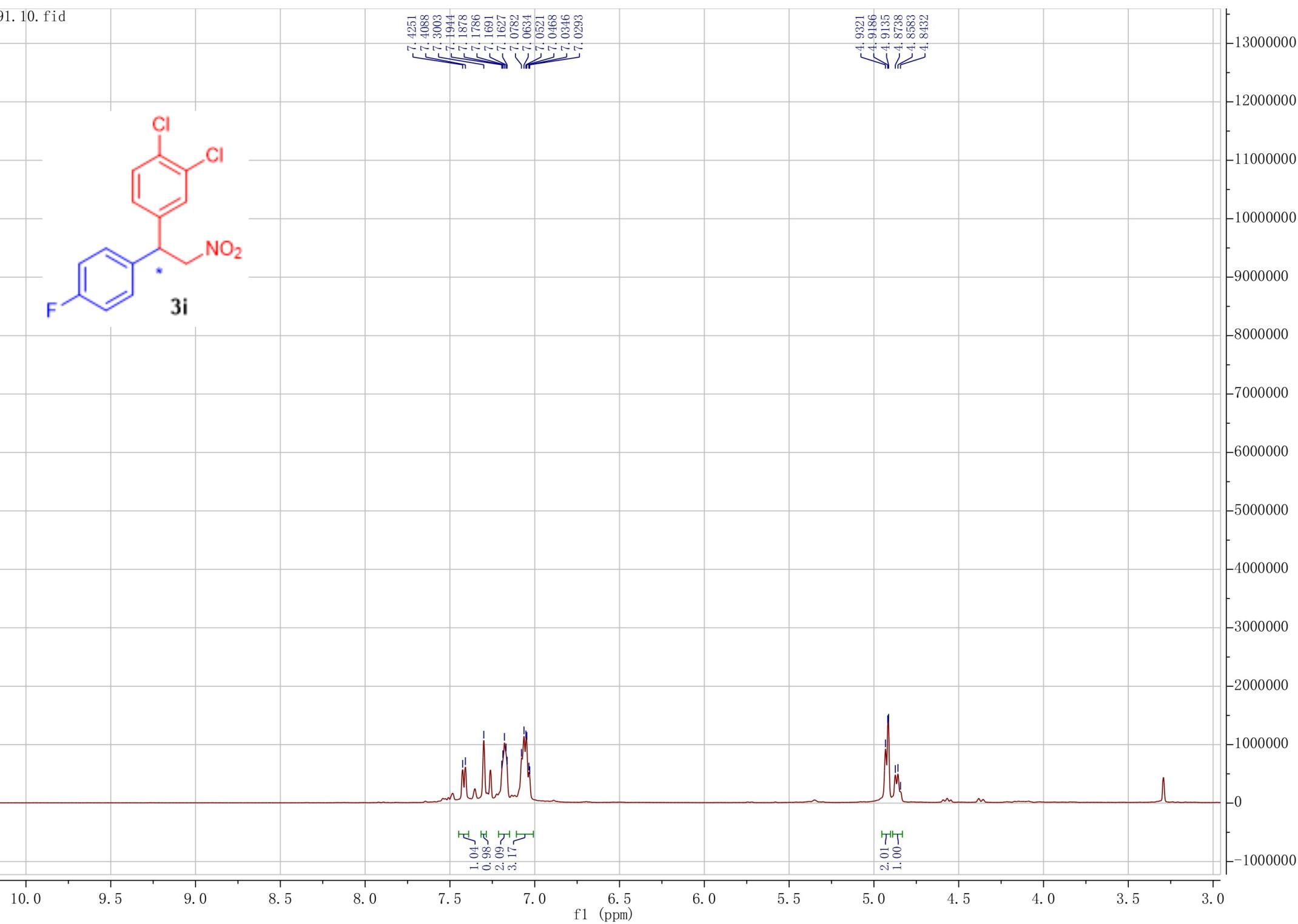
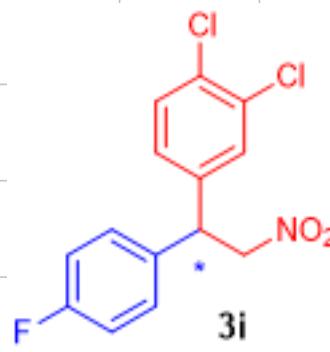


3ae/3ae-HNMR
K45

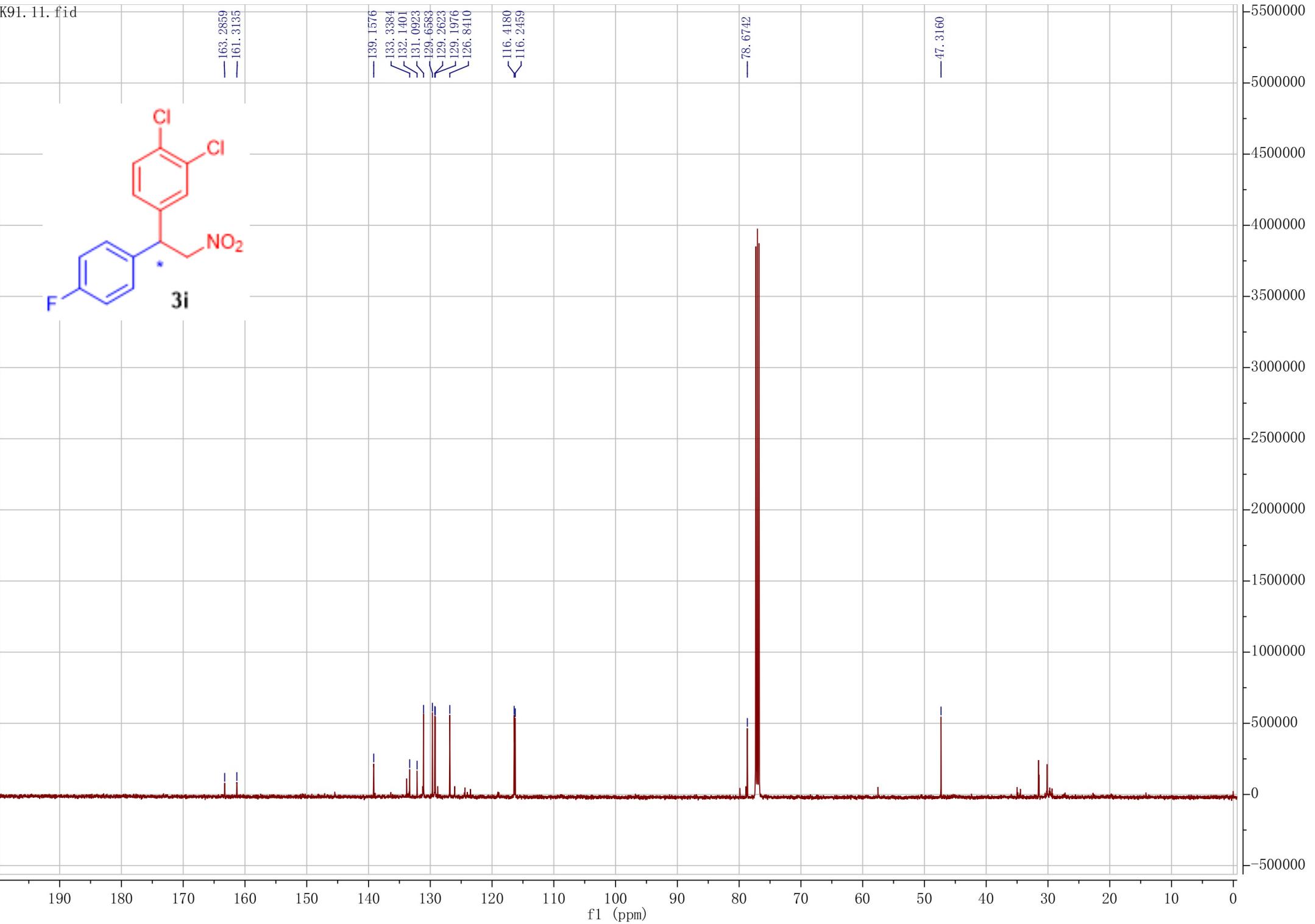
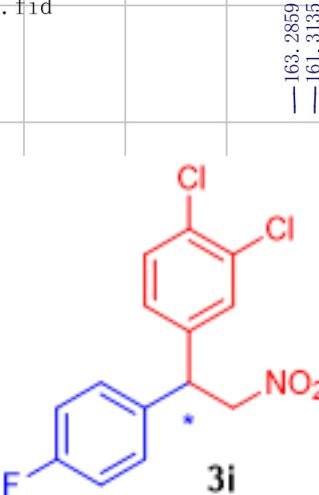


3h

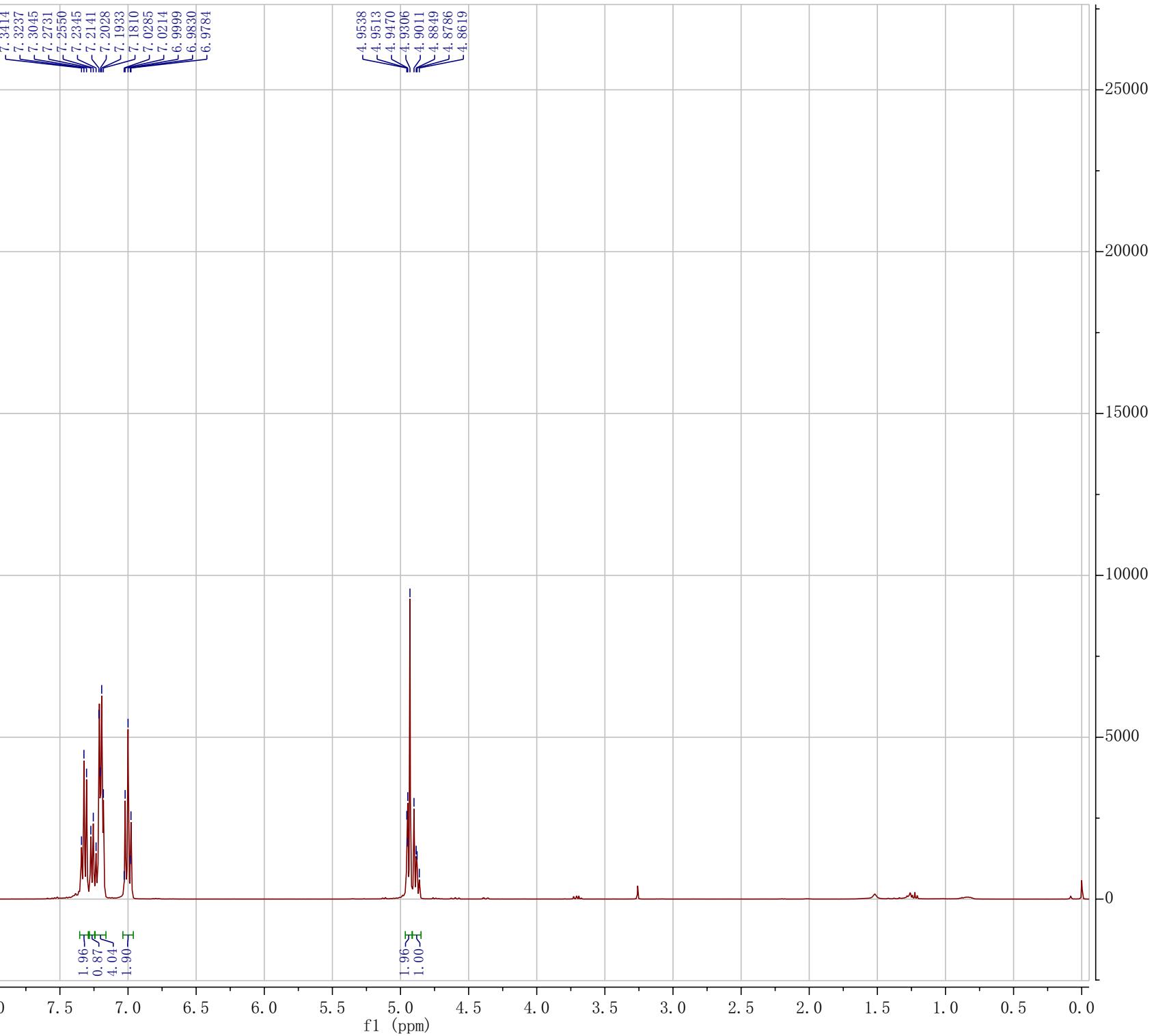
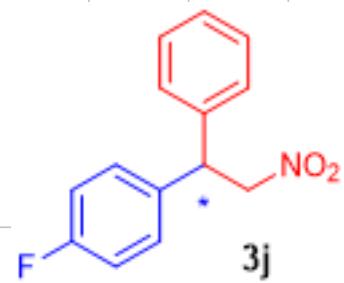




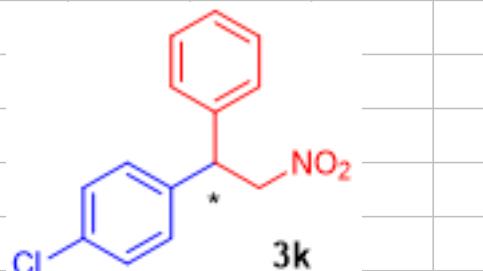
K91.11.fid



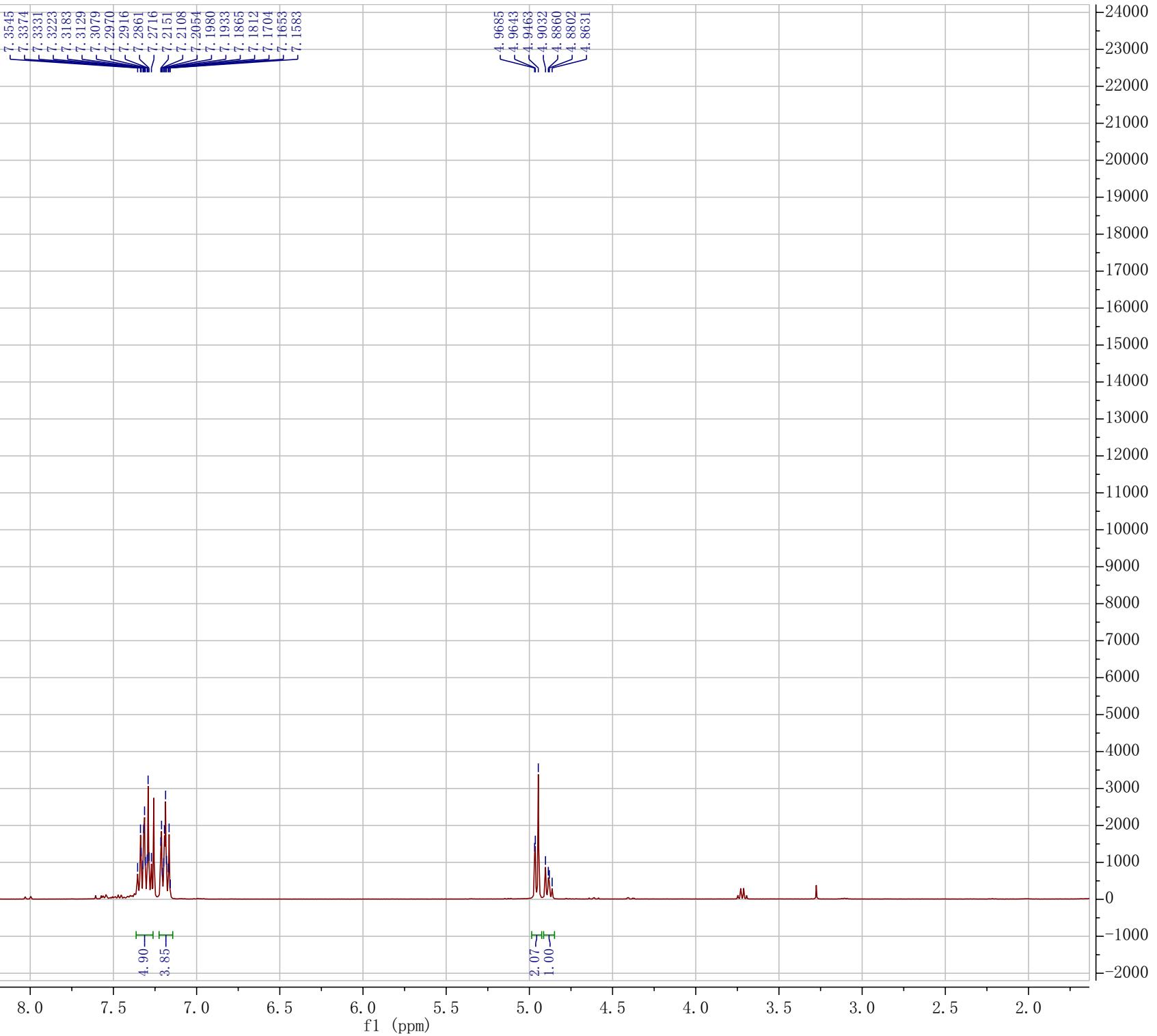
3ea/3ea HNMR
LD7



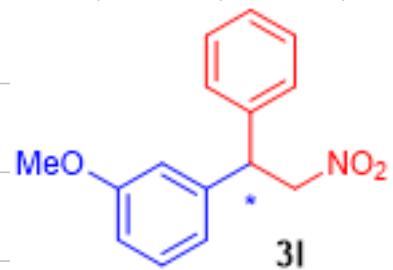
3fa/3fa HNMR
LD10



3k



3ba/3ba HNMR
LD5



3l

9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0

f1 (ppm)

7000
6500
6000
5500
5000
4500
4000
3500
3000
2500
2000
1500
1000
500
0
-500

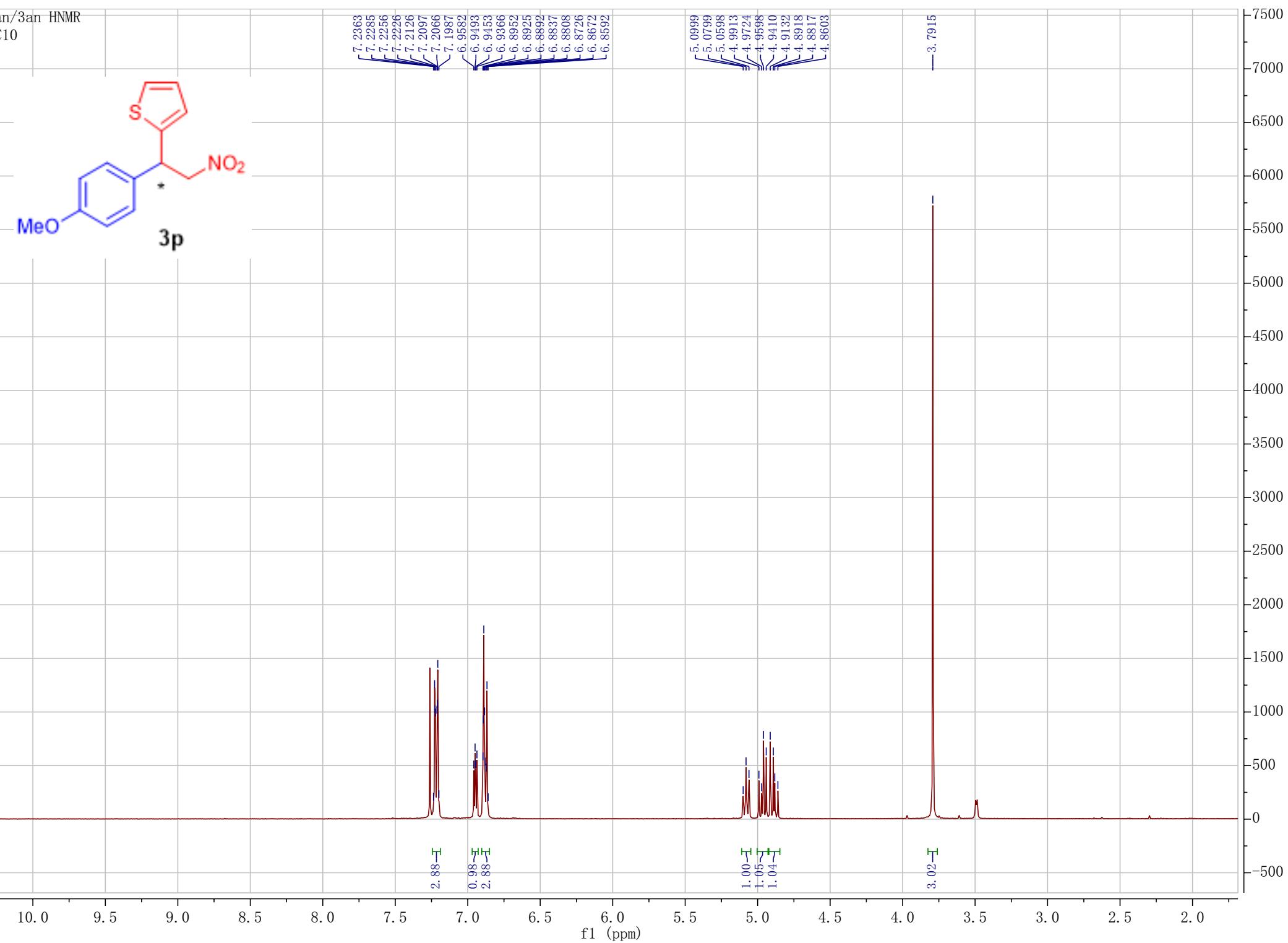
2.01
4.51
0.97
1.93

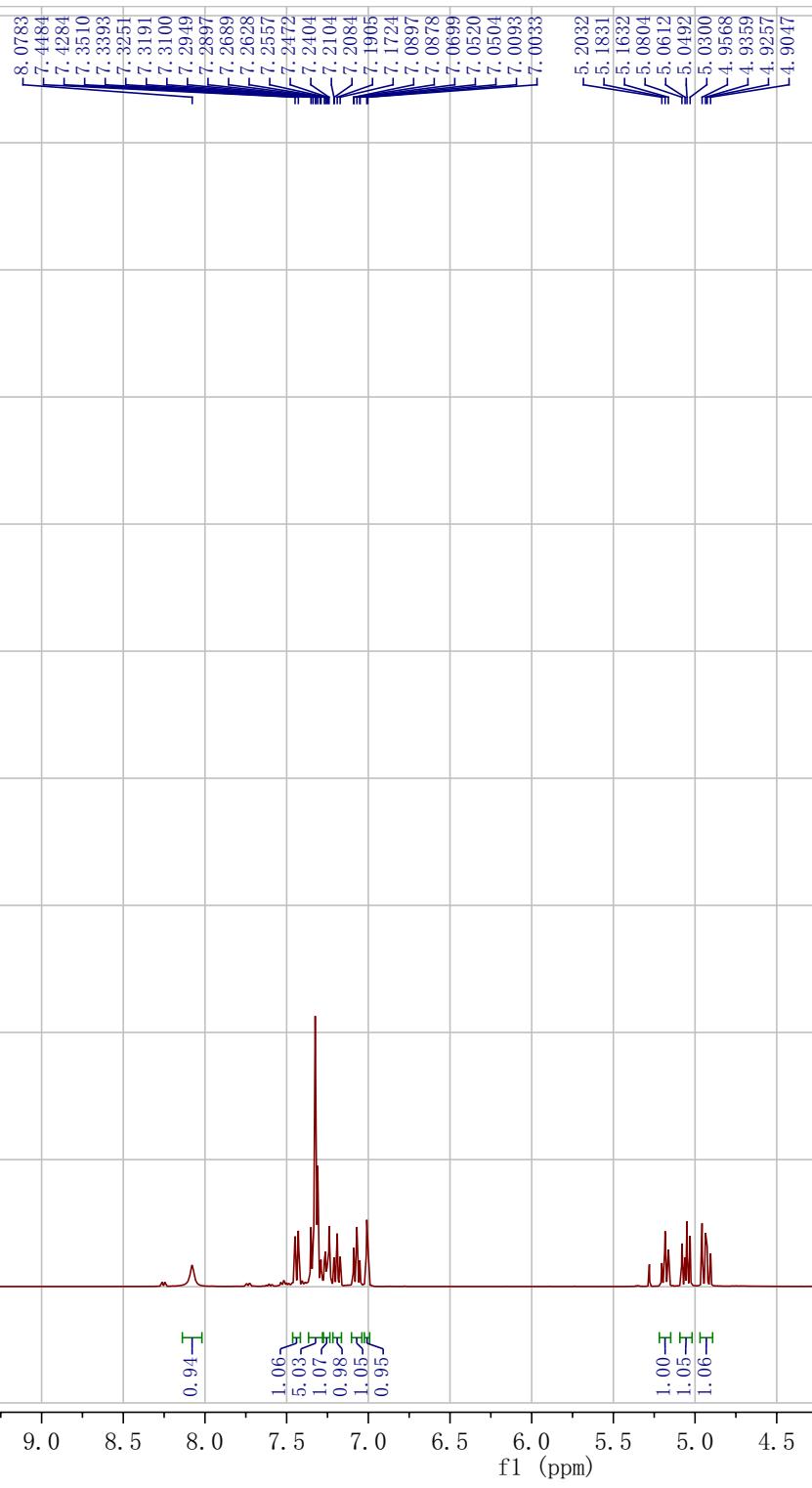
1.95
1.00

2.96

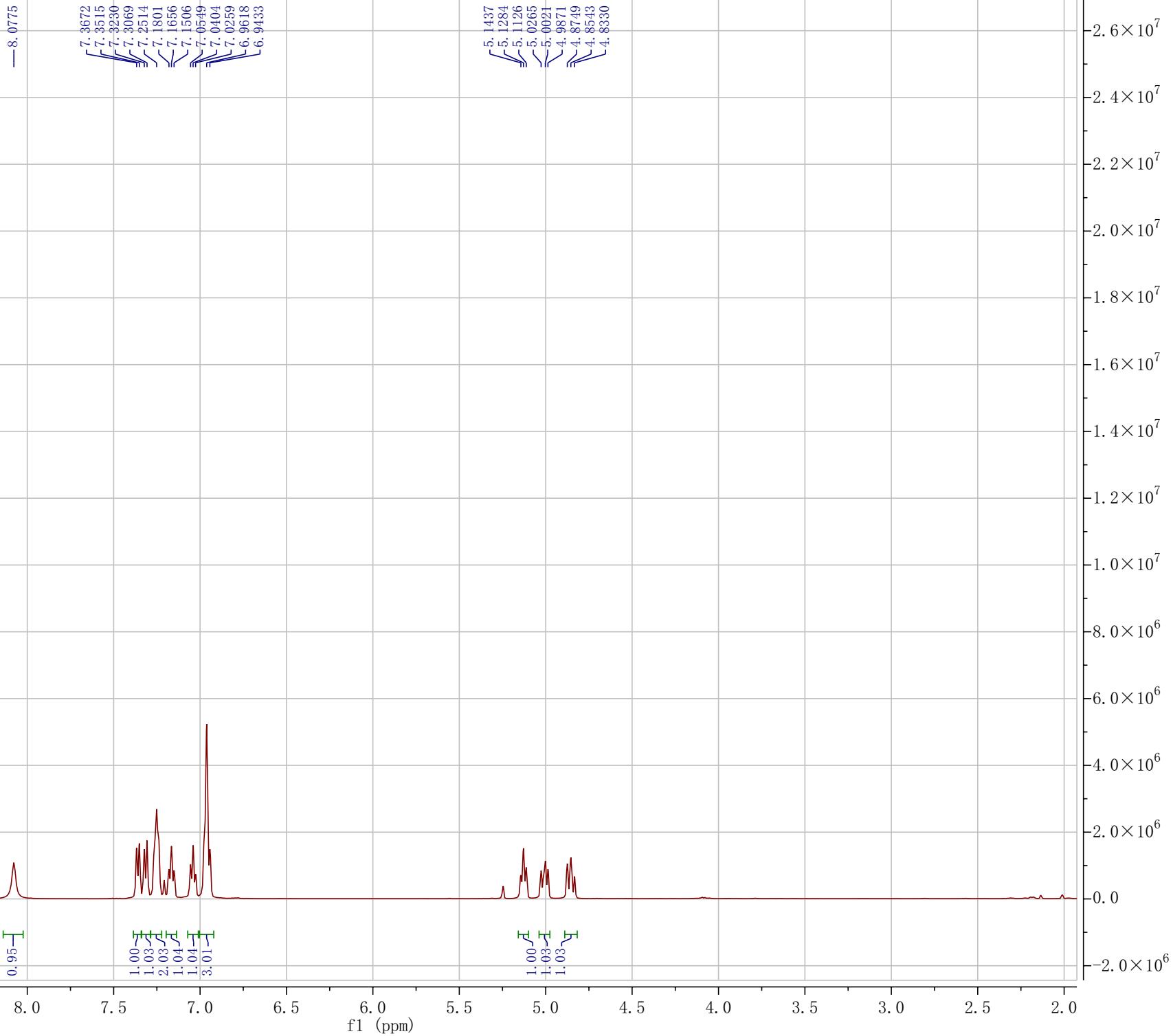
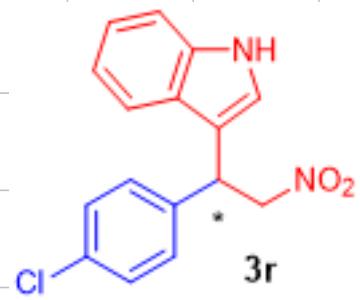
4.9824
4.9792
4.9606
4.8929
4.8749
4.8706
4.8525

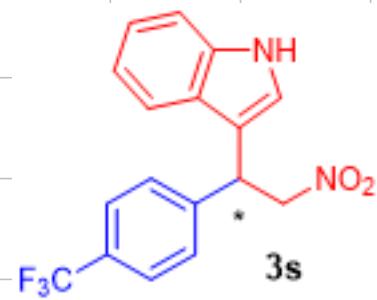
3.7707

**3p**

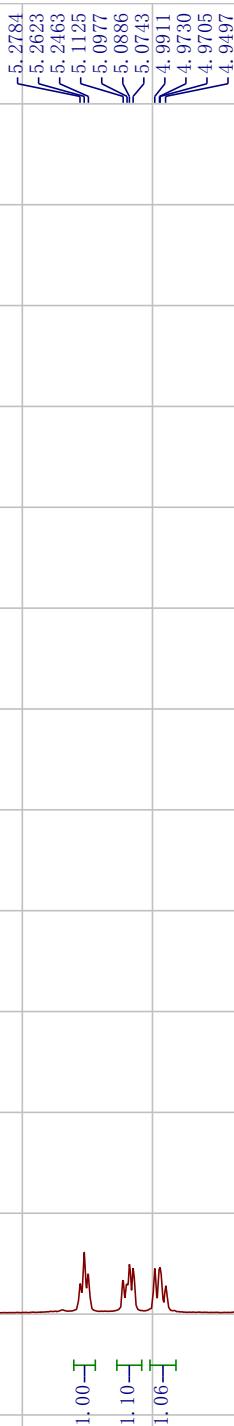
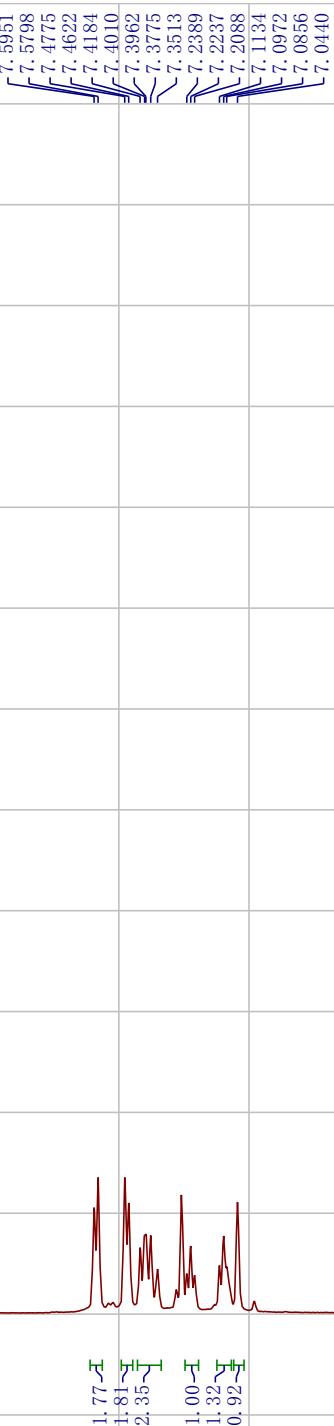
3cf
K79**3q**

K51-3.10.fid



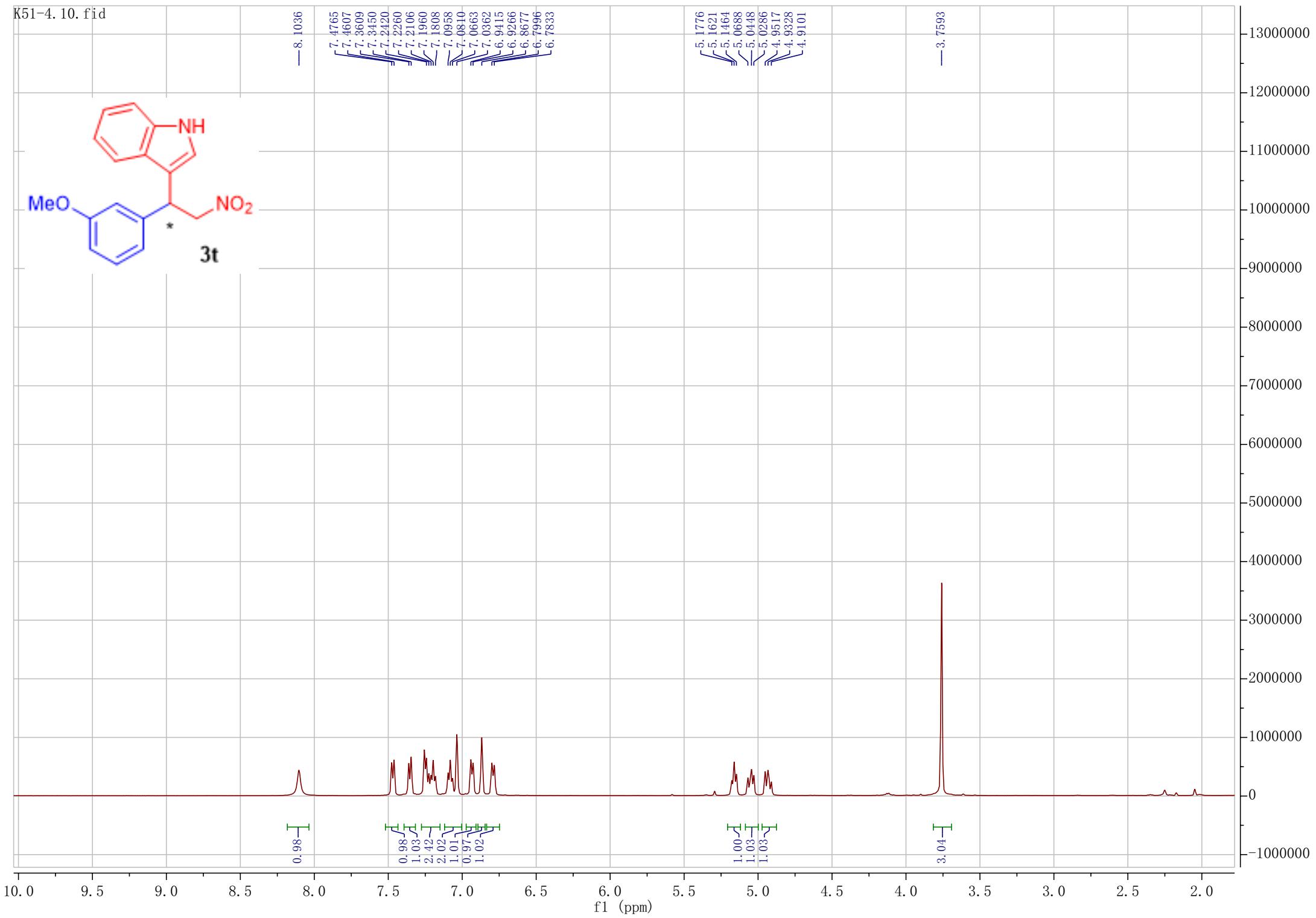


8.1744

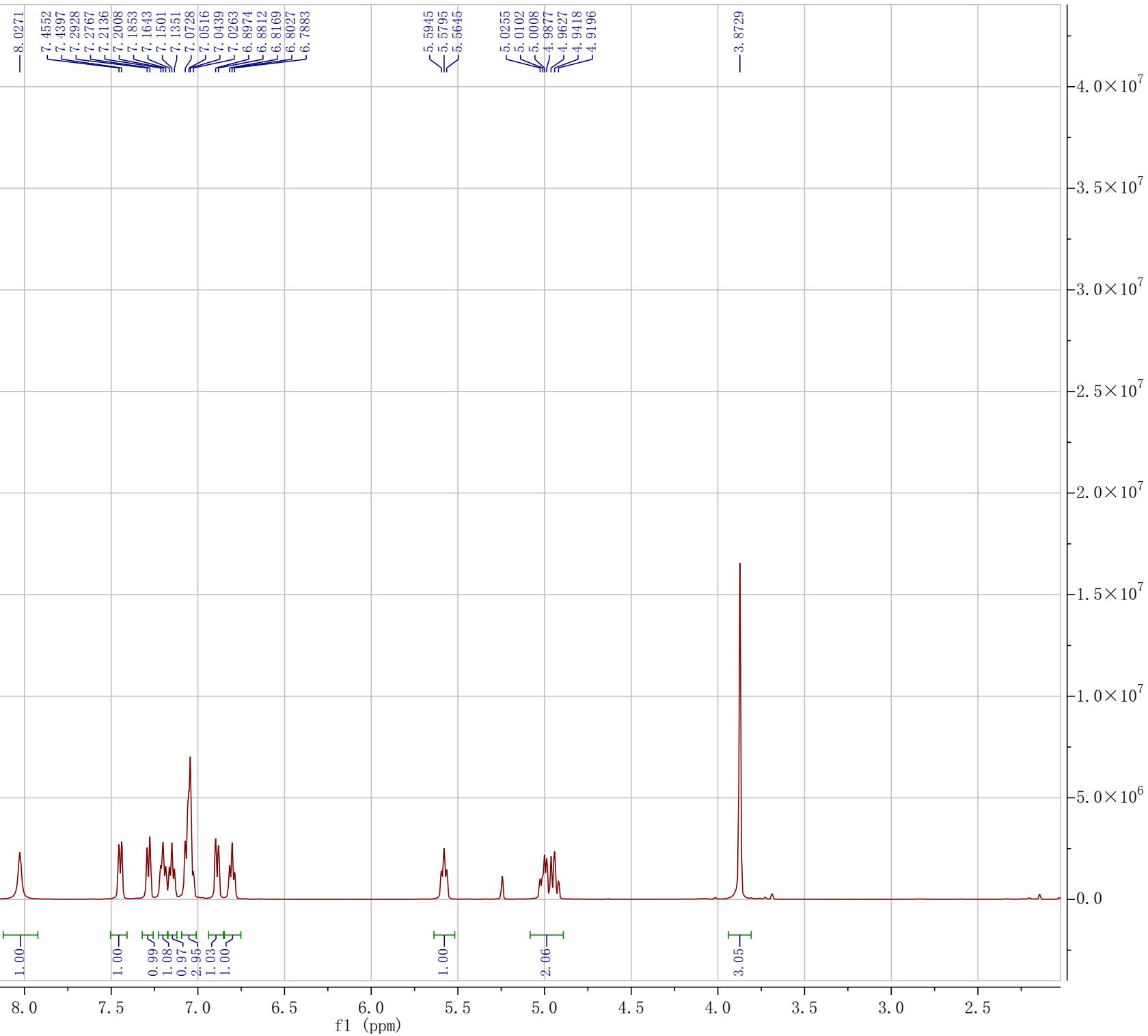


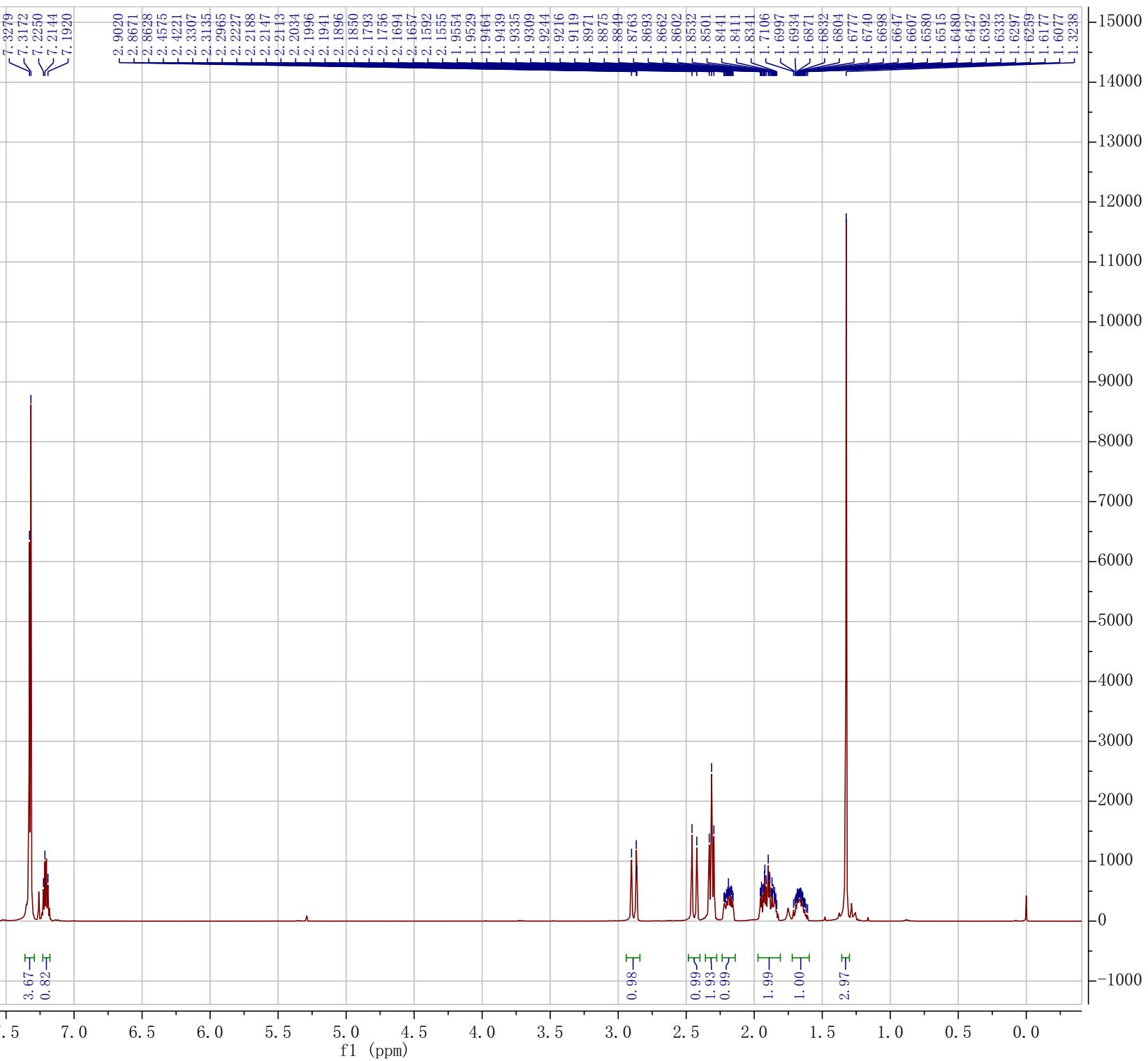
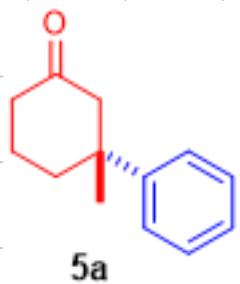
10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0 -1.5 -2.0 f1 (ppm)

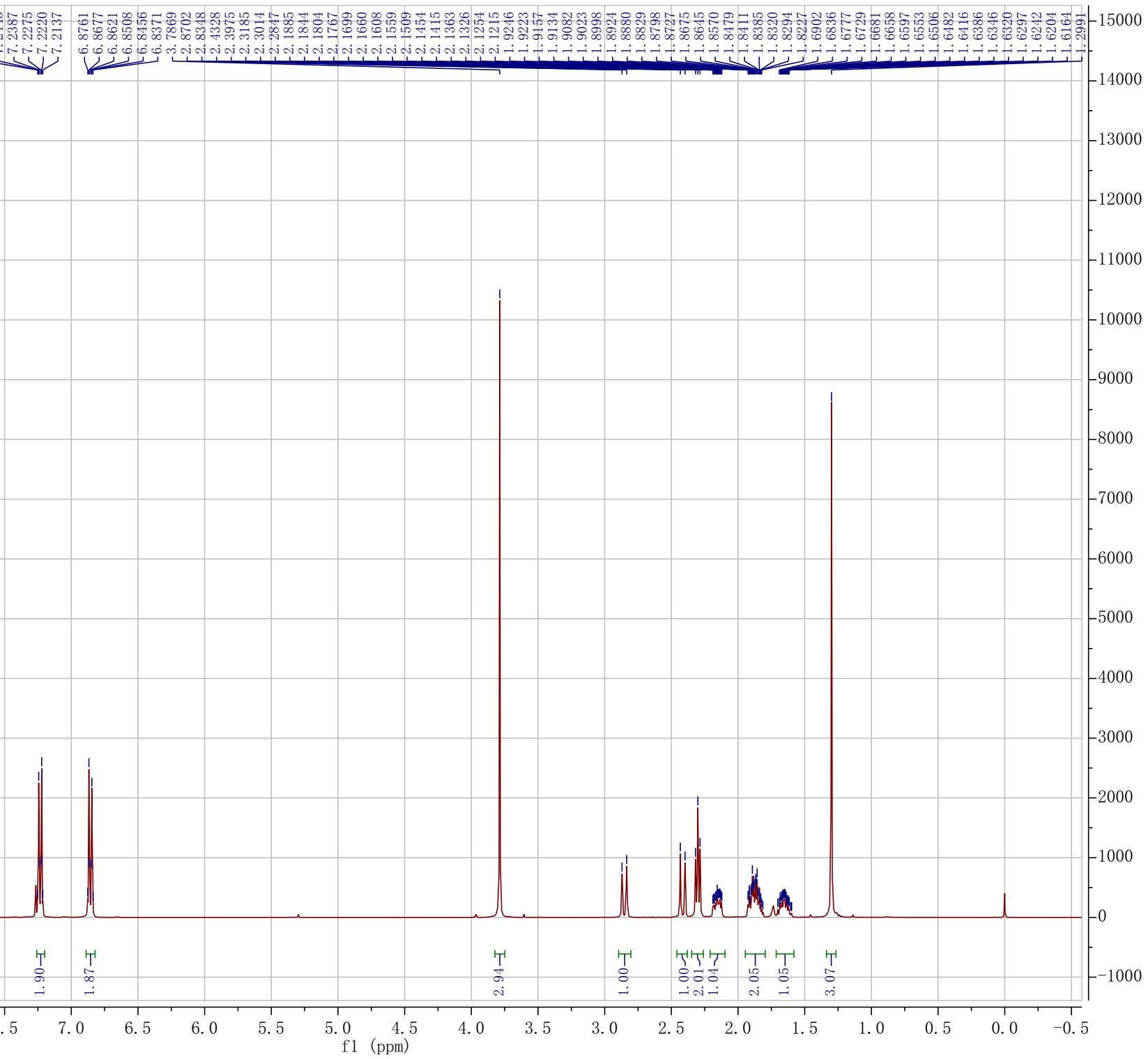
K51-4.10.fid

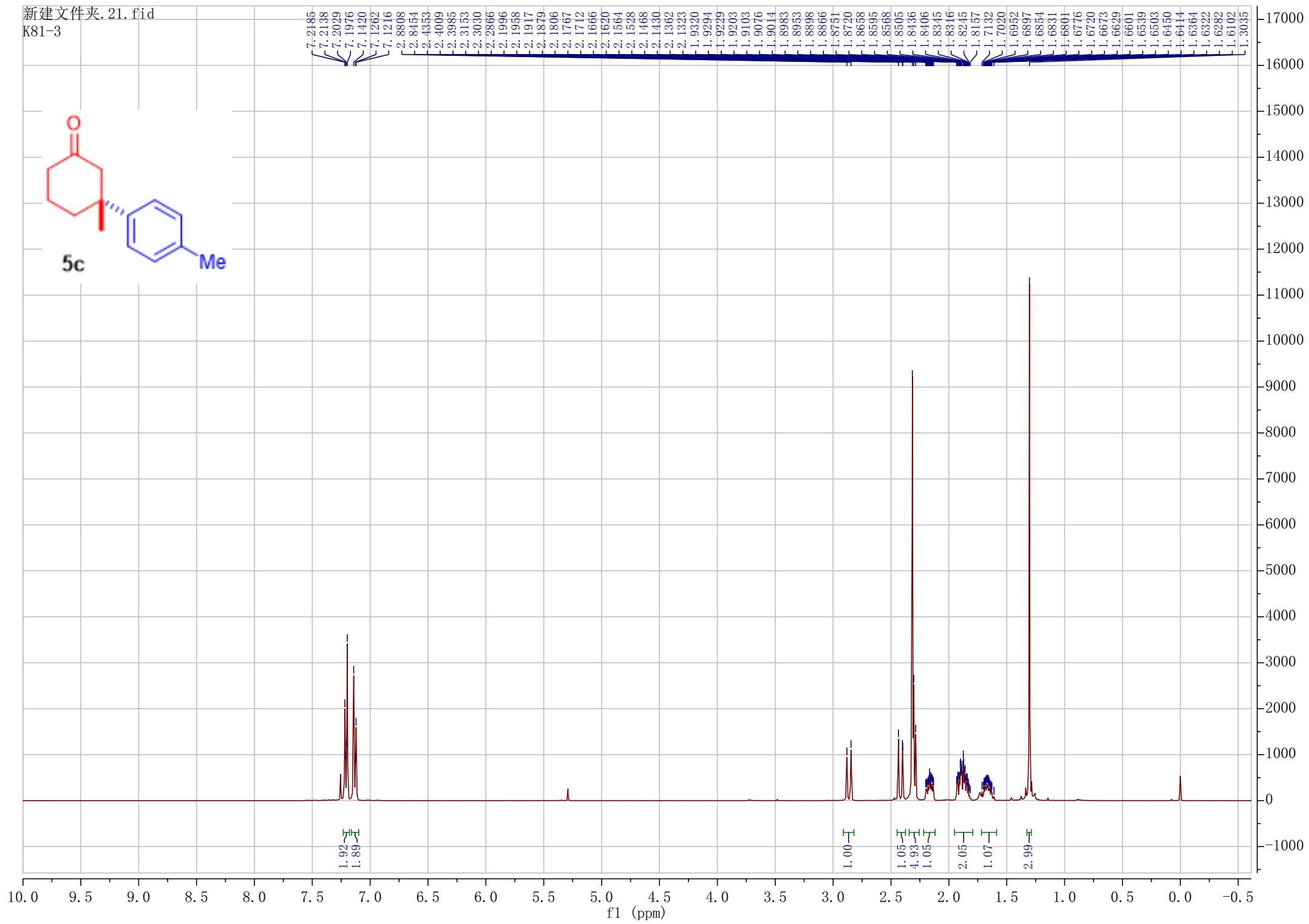
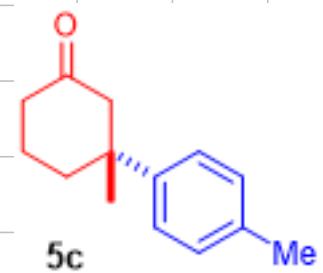


K51-5.10.fid

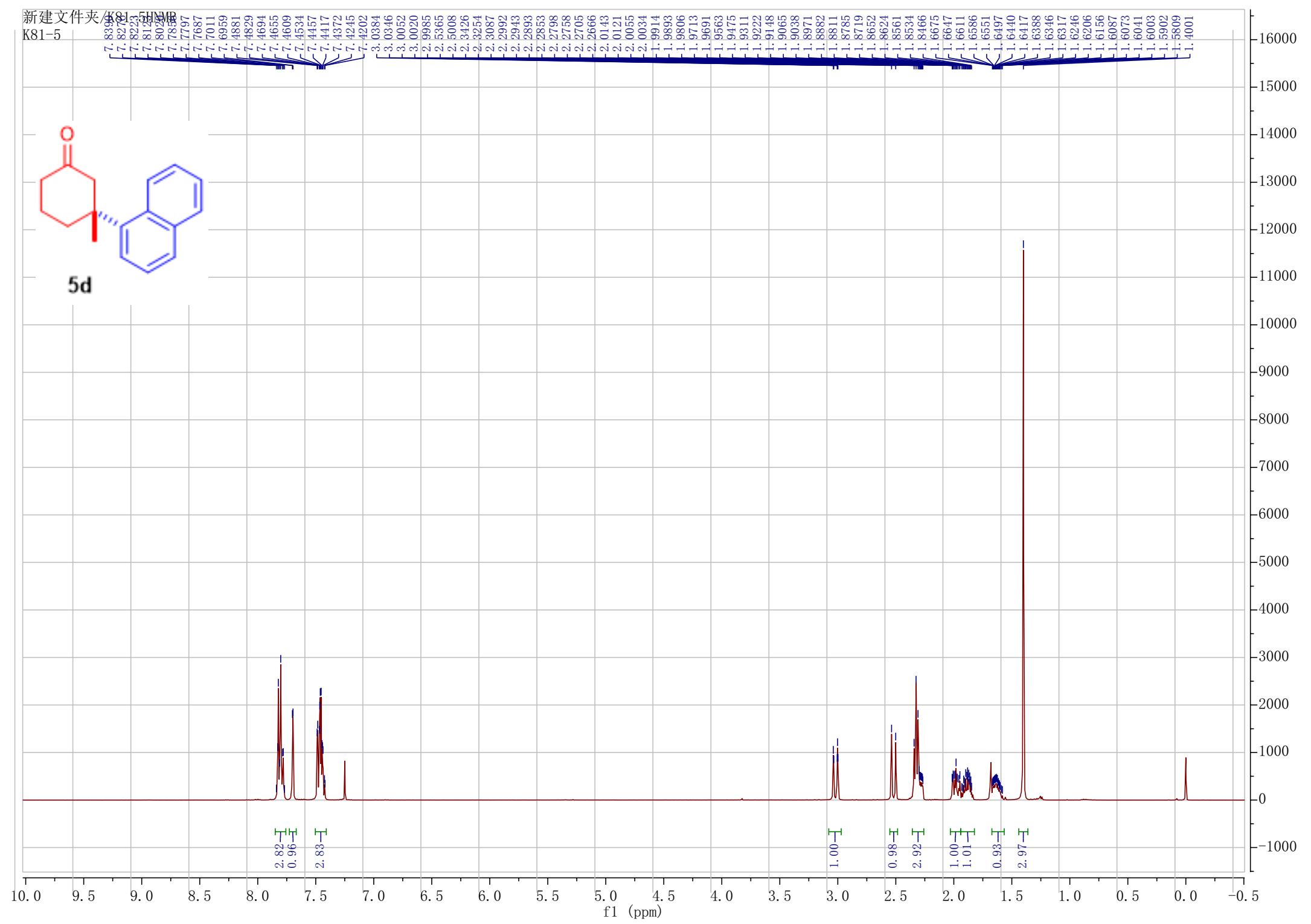
**3u**



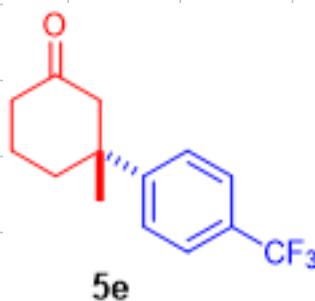




新建文件夹/K81-5HNMP
K81-5



7.5893
7.5737
7.4495
7.4338



2.9002
2.8719
2.4925
2.4642
2.3807
2.3640
2.3332
2.3020
2.2105
2.1954
2.1760
1.9842
1.9641
1.9380
1.9128
1.9048
1.8994
1.8924
1.6168
1.3431

1.90
1.98

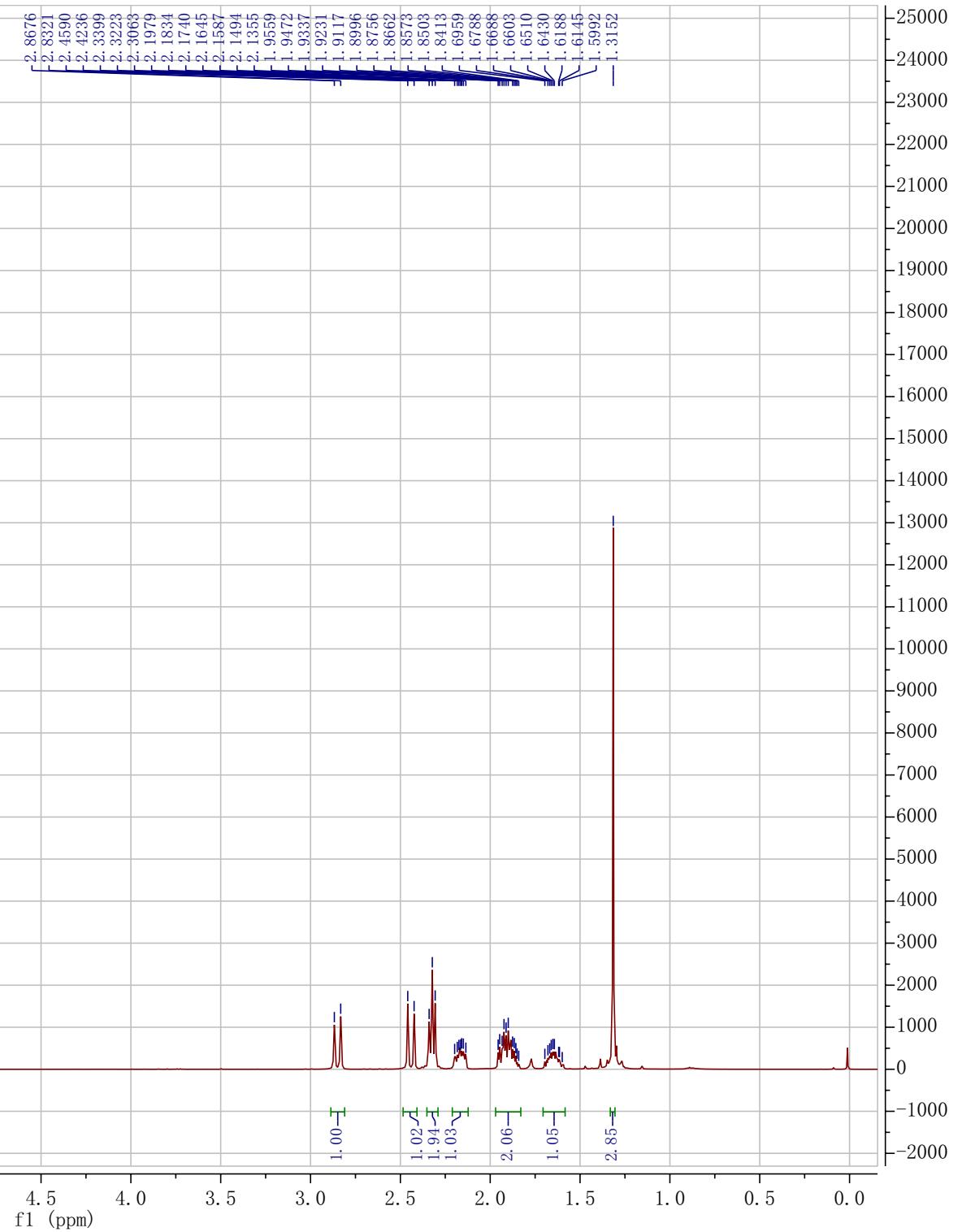
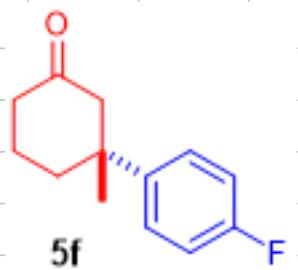
1.00
1.02
2.01
1.31
2.11
2.21
3.20

8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0

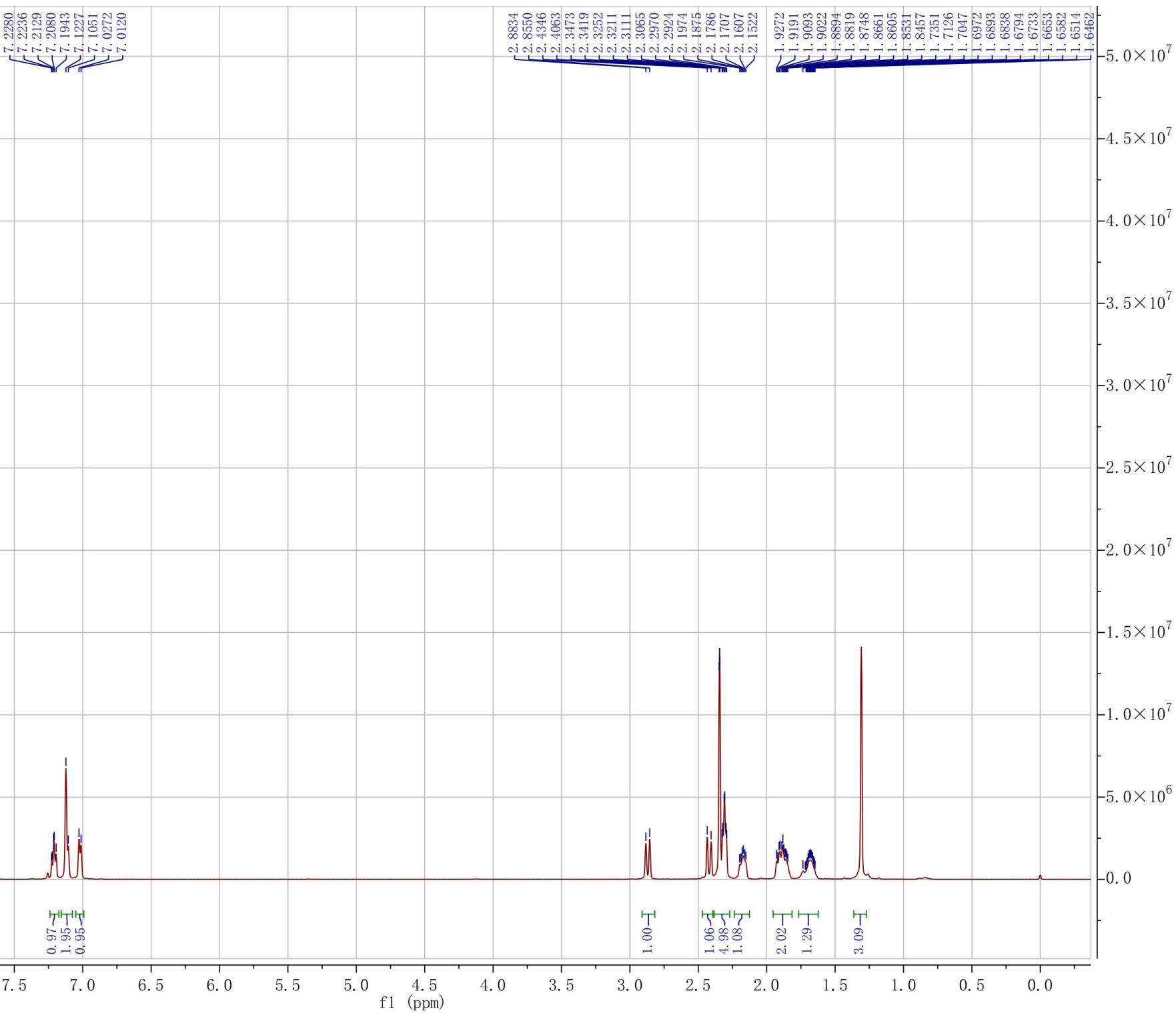
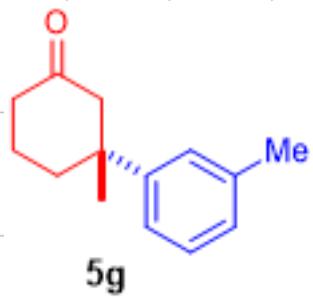
f1 (ppm)

17000000
16000000
15000000
14000000
13000000
12000000
11000000
10000000
9000000
8000000
7000000
6000000
5000000
4000000
3000000
2000000
1000000
0
-1000000

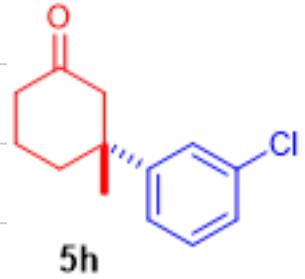
7.3085
7.3024
7.2922
7.2862
7.2679
7.2618
7.2514
7.2456



K81-8.10.fid



K81-12. 10. fid

**5h**