

Supporting Information

Chiral Phosphoric Acid Catalyzed Atroposelective and Diastereoselective Synthesis of 9-Aryltetrahydroacridines

You-Dong Shao, Dan-Dan Han, Wen-Yue Ma and Dao-Juan Cheng*

School of Chemistry and Chemical Engineering, Heze University, Heze 274015, People's Republic of
China

E-mail: chengdaojuan0614@163.com

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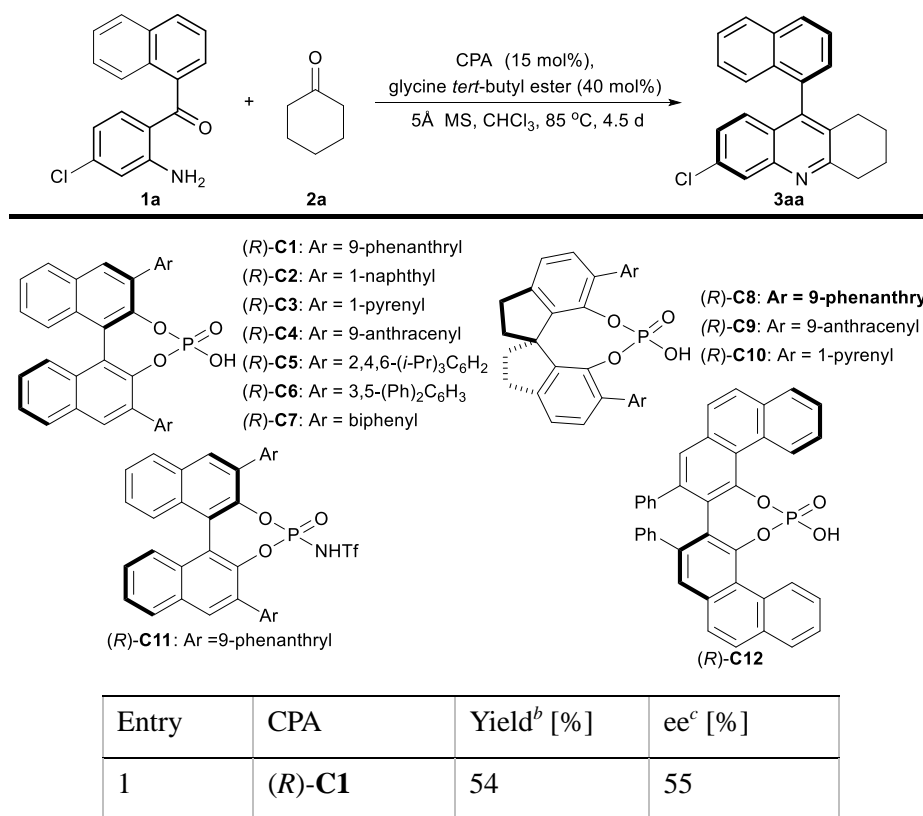
General Information

^1H and ^{13}C NMR spectra were recorded on a Bruker AC-400 FT (400 MHz for ^1H NMR and 100 MHz for ^{13}C NMR, respectively) using tetramethylsilane as an internal reference. Chemical shifts (δ) and coupling constants (J) were expressed in ppm and Hz, respectively. High resolution mass spectra (HRMS) were recorded on a LC-TOF spectrometer (Micromass). ESI-HRMS data were acquired using a Thermo LTQ Orbitrap XL Instrument equipped with an ESI source and controlled by Xcalibur software. High pressure liquid chromatography (HPLC) analyses were performed on a Thermo Scientific UltiMate 3000 instrument equipped with an isostatic pump, using a chiral stationary phase column (Daicel Co. CHIRALPAK). The chiral HPLC methods were calibrated with the corresponding racemic mixtures. Optical Rotation was measured on an Anton Paar MCP 100/150 polarimeter.

Chloroform was distilled over calcium hydride. Other solvents and chemicals were purchased from the Sinopharm Chemical Reagent Co., Adamas, Acros, Alfa Aesar, and TCI, and used as received. Catalysts (*R*)-**C1-C7** were prepared according to the literatures.¹ Catalysts (*R*)-**C8-C12** were purchased from Daicel Chiral Technologies (China) CO., LTD. and used directly. 2-Aminoaryl ketones **1** were prepared in accordance with literature methods.²⁻⁴

Screening of Catalysts and Condition Optimization

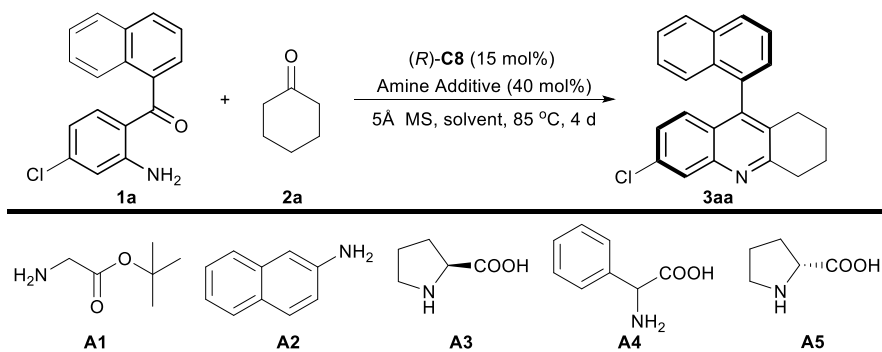
Table S1. Screening of catalysts.^a



2	(<i>R</i>)- C2	58	23
3	(<i>R</i>)- C3	56	49
4	(<i>R</i>)- C4	60	55
5	(<i>R</i>)- C5	trace	15
6	(<i>R</i>)- C6	43	1
7	(<i>R</i>)- C7	50	3
8	(<i>R</i>)- C8	67	89
9	(<i>R</i>)- C9	61	82
10	(<i>R</i>)- C10	63	84
11	(<i>R</i>)- C11	trace	30
12	(<i>R</i>)- C12	52	24

^a All reactions were carried out with (2-amino-4-chlorophenyl)(naphthalen-1-yl)methanone **1a** (28.1 mg, 0.10 mmol), cyclohexanone **2a** (29.4 mg, 31.0 μ L, 0.30 mmol), catalyst CPA (15 mol%), glycine *tert*-butyl ester (40 mol%) and CHCl_3 (1.0 mL) in sealed tube for 4.5 d. ^b Isolated yield. ^c Determined by chiral stationary phase HPLC analysis.

Table S2. Screening of additives, solvents, temperatures *et al.*^a

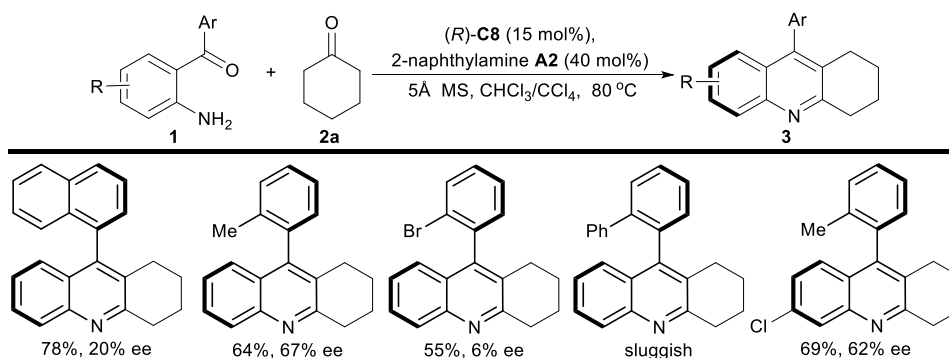


Entry	Solvent	Amine Additive	Yield ^b [%]	ee ^c [%]
1	CHCl_3	A1	67	89
2	CHCl_3	A2	72	90
3	CHCl_3	A3	61	87
4	CHCl_3	A4	49	86
5	CHCl_3	A5	60	86
6	Toluene	A2	trace	ND
7	CH_3CN	A2	NP	ND

8	EtOAc	A2	trace	ND
9	CCl ₄	A2	81	82
10	DCE	A2	trace	ND
11	<i>n</i> -hexane	A2	80	66
12	MTBE	A2	73	81
13 ^d	CHCl₃:CCl₄=4:1	A2	77	92
14 ^d	CHCl ₃ :CCl ₄ =3:2	A2	79	90
15 ^{d,e}	CHCl ₃ :CCl ₄ =4:1	A2	73	90
16 ^{d,f}	CHCl ₃ :CCl ₄ =4:1	A2	67	87
17 ^{d,g}	CHCl ₃ :CCl ₄ =4:1	A2	74	91
18 ^{d,h}	CHCl ₃ :CCl ₄ =4:1	A2	trace	ND

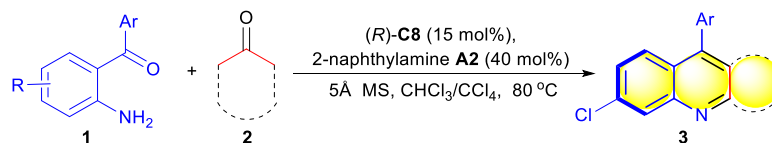
^a Unless otherwise stated, all reactions were carried out with (2-amino-4-chlorophenyl)(naphthalen-1-yl)methanone **1a** (28.1 mg, 0.10 mmol), cyclohexanone **2a** (29.4 mg, 31.0 μL, 0.30 mmol), catalyst (*R*)-**C8** (10.0 mg, 15 mol%), amine additive (40 mol%) and solvent (1.0 mL) in sealed tube at 85 °C for 4.5 d. ^b Isolated yield. ^c Determined by chiral stationary phase HPLC analysis. ^d The reaction was run at 80 °C for 4 d. ^e The reaction was run with 0.70 mmol **2a**. ^f The reaction was run with 0.15 mmol **2a**. ^g The reaction was run with 60 mol% of **A2**. ^h The reaction was run with 100 mol% of **A2**.

Table S3. Preliminary investigations on the different combinations of substrates.^a



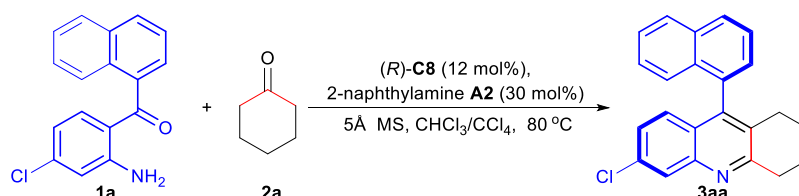
^a All reactions were carried out with **1** (0.10 mmol), cyclohexanone **2a** (0.30 mmol), (*R*)-**C8** (15 mol%), 2-naphthylamine **A2** (40 mol%) and 5 Å molecular sieves (100 mg) in CHCl₃ (0.8 mL) and CCl₄ (0.2 mL) at 80 °C in sealed tube for 4 d. Yields refer to isolated pure compounds. The ee values were determined by chiral stationary phase HPLC analysis.

General Procedure for the Atroposelective and Diastereoselective Synthesis of 9-Aryltetrahydroacridines



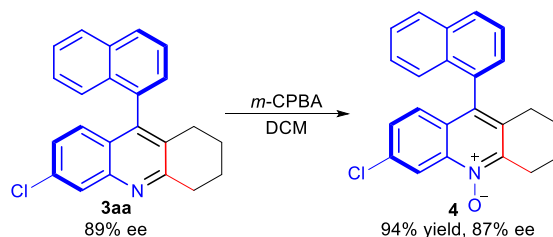
To a flame dried sealed tube equipped with a magnetic stirring bar were added powdered 5 Å molecular sieves (100 mg), chiral phosphoric acid (*R*)-**C8** (10.0 mg, 0.015 mmol), 2-naphthylamine **A2** (5.7 mg, 0.040 mmol), 2-aminoaryl ketone **1** (0.10 mmol), carbon tetrachloride (0.2 mL), anhydrous chloroform (0.8 mL) and alicyclic ketone **2** (0.30 mmol) successively. The resulting mixture was stirred at 80 °C for 4-5 d, and directly charged onto silica gel. Product **3** was isolated using petroleum ether/ethyl acetate (30:1 to 5:1) as eluent.

Procedure for Scale-up Experiment



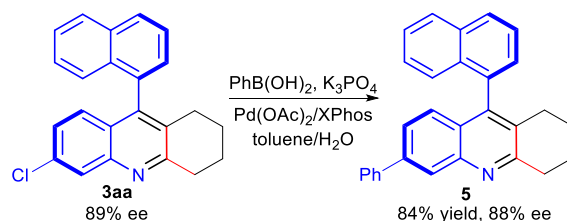
To a flame dried sealed tube equipped with a magnetic stirring bar were added powdered 5 Å molecular sieves (1.00 g), chiral phosphoric acid (*R*)-**C8** (80.1 mg, 0.12 mmol), 2-naphthylamine **A2** (43.0 mg, 0.30 mmol), **1a** (281.1 mg, 1.0 mmol), carbon tetrachloride (4.0 mL), anhydrous chloroform (6.0 mL) and **2a** (245.4 mg, 2.5 mmol) successively. After stirring at 80 °C for 4.5 d, the reaction was quenched with saturated sodium bicarbonate solution (10.0 mL). After being extracted with dichloromethane (3 × 15 mL), the organic phases were combined and washed with brine, dried over anhydrous sodium sulfate, filtered and concentrated in vacuo to give a crude residue which was purified by flash column chromatography (petroleum ether/ethylacetate = 20:1) to give **3aa** (240.2 mg, 70% yield, 89% ee) as a yellow solid.

Procedure for the Transformation of Product 3aa



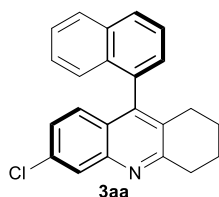
m-CPBA (37.0 mg, 0.15 mmol) was added to a solution of (*S*)-**3aa** (34.3 mg, 0.10 mmol) in dichloromethane (5.0 mL) at 0 °C. After stirring for 2 h, the reaction mixture was adjusted to pH = 10 by 1 N NaOH. After being extracted with dichloromethane (3 × 10 mL), the organic phases were combined and washed with brine, dried over anhydrous sodium sulfate, filtered and concentrated in vacuo to give a crude residue which was purified by flash column

chromatography (petroleum ether/ethylacetate = 70:30) to give **4** (33.8 mg, 94% yield) as a white solid.⁴

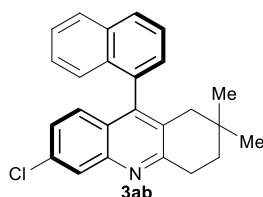


To a flame dried Schlenk flask were added **3aa** (34.3 mg, 0.10 mmol), Pd(OAc)₂ (0.50 mg, 0.002 mmol), XPhos (2.0 mg, 0.004 mmol), phenylboronic acid (24.4 mg, 0.20 mmol) and K₃PO₄ (85.0 mg, 0.40 mmol) successively. The flask was evacuated and backfilled with nitrogen three times, then toluene (0.80 mL) and H₂O (0.20 mL) were added. After stirring at 70 °C for 16 hours, the reaction mixture was cooled to room temperature, diluted with ethyl acetate (10 mL) and washed with aqueous NaOH (1 N, 10 mL). After being extracted with ethyl acetate (2 × 10 mL), the combined organic layers were washed with brine (10 mL), dried over anhydrous sodium sulfate, filtered and concentrated in vacuo to give a crude residue which was purified by flash column chromatography (petroleum ether/ethylacetate = 30:1–5:1) to give **5** (32.4 mg, 84% yield) as a yellow oil.⁵

Analytic Data for the Products

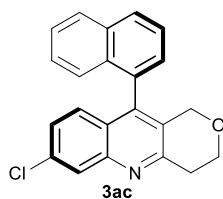


6-Chloro-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3aa** was obtained as a white solid in 77% yield (26.4 mg) and 92% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 7.63 min, t_r (major) = 8.06 min]. m.p. 133-134 °C; $[\alpha]_D^{25} = +27.74$ ($c = 1.0$, EtOAc); ¹H NMR (400 MHz, CDCl₃): δ 8.07 (d, $J = 2.0$ Hz, 1H), 7.97 (d, $J = 8.4$ Hz, 2H), 7.61 (t, $J = 7.6$ Hz, 1H), 7.50 (t, $J = 7.6$ Hz, 1H), 7.35-7.27 (m, 2H), 7.17-7.09 (m, 2H), 7.03 (d, $J = 8.8$ Hz, 1H), 3.23 (t, $J = 6.8$ Hz, 2H), 2.62-2.47 (m, 1H), 2.38-2.26 (m, 1H), 2.01-1.90 (m, 2H), 1.78-1.65 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 160.5, 146.7, 145.0, 134.3, 134.2, 133.7, 131.3, 130.0, 128.6, 128.5, 127.4, 127.3, 126.9, 126.7, 126.5, 126.3, 125.7, 125.6, 125.2, 34.3, 27.5, 22.8, 22.8; HRMS (ESI) calcd for C₂₃H₁₉ClN (M+H)⁺ 344.1201, found 344.1200.

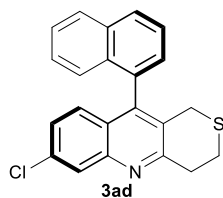


6-Chloro-2,2-dimethyl-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3ab** was obtained as a

white solid in 69% yield (25.6 mg) and 91% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 9.12 min, t_r (minor) = 9.93 min]. m.p. >240 °C; $[\alpha]_D^{25} = +16.67$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.07 (s, 1H), 7.99 (t, $J = 8.8$ Hz, 2H), 7.63 (t, $J = 7.6$ Hz, 1H), 7.51 (t, $J = 7.6$ Hz, 1H), 7.36-7.22 (m, 2H), 7.18-7.05 (m, 2H), 6.99 (d, $J = 8.8$ Hz, 1H), 3.27 (t, $J = 6.8$ Hz, 2H), 2.34 (d, $J = 16.8$ Hz, 1H), 2.10 (d, $J = 16.8$ Hz, 1H), 1.77 (t, $J = 6.8$ Hz, 2H), 0.90 (s, 3H), 0.86 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 159.7, 146.8, 145.5, 134.3, 134.2, 133.7, 131.4, 129.4, 128.5, 128.5, 127.4, 126.9, 126.7, 126.5, 126.3, 125.7, 125.6, 125.2, 41.2, 35.6, 31.0, 29.5, 28.4, 27.4; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{23}\text{ClN}$ (M+H) $^+$ 372.1514, found 372.1513.

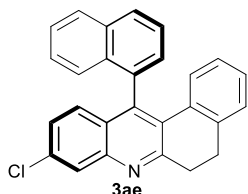


7-Chloro-10-(naphthalen-1-yl)-3,4-dihydro-1H-pyrano[4,3-*b*]quinoline **3ac** was obtained as a yellow solid in 70% yield (24.2 mg) and 90% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (10:90), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 8.10 min, t_r (major) = 9.53 min]. m.p. 141-142 °C; $[\alpha]_D^{25} = +14.74$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.16 (d, $J = 2.0$ Hz, 1H), 8.05-7.95 (m, 2H), 7.66-7.59 (m, 1H), 7.56-7.49 (m, 1H), 7.38-7.29 (m, 2H), 7.24 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.12 (d, $J = 9.2$ Hz, 2H), 4.56 (d, $J = 16.0$ Hz, 1H), 4.31 (d, $J = 16.0$ Hz, 1H), 4.18-4.05 (m, 2H), 3.42-3.28 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 155.3, 146.2, 141.5, 134.0, 132.6, 131.0, 129.9, 128.1, 127.6, 126.6, 126.4, 126.1, 126.1, 126.0, 125.8, 125.5, 124.5, 124.4, 123.9, 65.6, 64.5, 32.0; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClNO}$ (M+H) $^+$ 346.0993, found 346.0992.

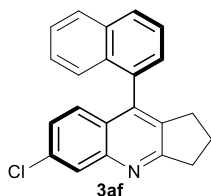


7-Chloro-10-(naphthalen-1-yl)-3,4-dihydro-1H-thiopyrano[4,3-*b*]quinoline **3ad** was obtained as a yellow solid in 74% yield (26.7 mg) and 89% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (07:93), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 6.96 min, t_r (major) = 7.54 min]. m.p. 78-79 °C; $[\alpha]_D^{25} = +15.63$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.11 (d, $J = 2.0$ Hz, 1H), 8.03-7.94 (m, 2H), 7.67-7.58 (m, 1H), 7.55-7.48 (m, 1H), 7.38-7.31 (m, 2H), 7.22 (dd, $J = 8.8, 2.0$ Hz,

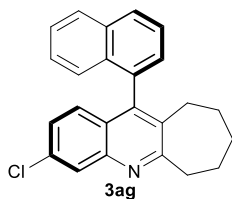
1H), 7.16 (d, $J = 8.8$ Hz, 1H), 7.10 (d, $J = 9.2$ Hz, 1H), 3.56-3.45 (m, 3H), 3.39 (d, $J = 5.2$ Hz, 1H), 3.16-3.03 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 159.4, 147.0, 143.3, 135.0, 133.6, 132.9, 131.6, 129.1, 128.7, 128.6, 127.9, 127.6, 127.3, 127.2, 127.0, 126.5, 125.8, 125.5, 125.3, 34.7, 27.0, 25.8; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClNS}$ ($\text{M}+\text{H}$) $^+$ 362.0765, found 362.0764.



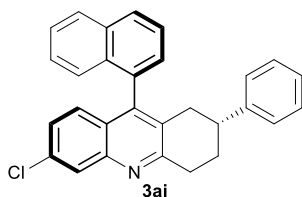
9-Chloro-12-(naphthalen-1-yl)-5,6-dihydrobenzo[*a*]acridine **3ae** was obtained as a pink solid in 80% yield (31.3 mg) and 81% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 8.49 min, t_r (major) = 9.03 min]. m.p. 97-98 °C; $[\alpha]_D^{25} = +12.44$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.11 (d, $J = 2.4$ Hz, 1H), 7.98 (d, $J = 8.0$ Hz, 2H), 7.55-7.46 (m, 2H), 7.35-7.30 (m, 2H), 7.28-7.23 (m, 1H), 7.22-7.18 (m, 1H), 7.16 (dd, $J = 9.2, 2.4$ Hz, 1H), 7.08 (d, $J = 9.2$ Hz, 1H), 7.04-6.98 (m, 1H), 6.61-6.55 (m, 2H), 3.41-3.32 (m, 1H), 3.31-3.22 (m, 1H), 3.11-2.97 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 162.5, 146.7, 142.2, 139.6, 135.4, 135.0, 133.6, 132.5, 128.8, 128.8, 128.7, 128.4, 127.8, 127.8, 127.7, 127.7, 127.6, 127.0, 126.9, 126.8, 126.4, 126.0, 126.0, 125.8, 34.8, 29.3; HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{19}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 392.1201, found 392.1199.



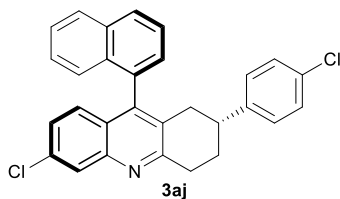
6-Chloro-9-(naphthalen-1-yl)-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline **3af** was obtained as a white solid in 81% yield (26.7 mg) and 80% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 8.82 min, t_r (major) = 9.42 min]. m.p. 85-86 °C; $[\alpha]_D^{25} = +13.57$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.12-8.08 (m, 1H), 7.97 (t, $J = 8.4$ Hz, 2H), 7.64-7.57 (m, 1H), 7.53-7.48 (m, 1H), 7.39-7.31 (m, 2H), 7.24-7.19 (m, 3H), 3.31-3.24 (m, 2H), 2.80-2.70 (m, 1H), 2.66-2.55 (m, 1H), 2.19-2.09 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 168.8, 148.3, 141.4, 135.5, 134.2, 133.9, 133.7, 131.2, 128.7, 128.6, 127.8, 127.3, 126.9, 126.6, 126.5, 126.3, 125.6, 125.5, 125.4, 35.2, 30.0, 23.2; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 330.1044, found 330.1044.



3-Chloro-11-(naphthalen-1-yl)-7,8,9,10-tetrahydro-6H-cyclohepta[*b*]quinoline **3ag** was obtained as a white solid in 70% yield (25.0 mg) and 80% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 7.27 min, t_r (major) = 7.74 min]. m.p. 99-100 °C; $[\alpha]_D^{25} = +12.86$ ($c = 1.0$, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.07 (d, $J = 2.0$ Hz, 1H), 7.96 (t, $J = 9.2$ Hz, 2H), 7.59 (t, $J = 7.6$ Hz, 1H), 7.49 (t, $J = 7.6$ Hz, 1H), 7.34-7.26 (m, 2H), 7.18-7.09 (m, 2H), 6.94 (d, $J = 8.8$ Hz, 1H), 3.37-3.25 (m, 2H), 2.60-2.49 (m, 2H), 1.95-1.75 (m, 4H), 1.56-1.39 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 166.1, 146.3, 143.8, 135.4, 134.7, 134.1, 133.5, 132.0, 128.5, 128.4, 127.9, 127.6, 127.1, 126.6, 126.6, 126.3, 125.9, 125.7, 125.4, 40.3, 31.9, 30.9, 28.3, 27.0; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{21}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 358.1357, found 358.1356.

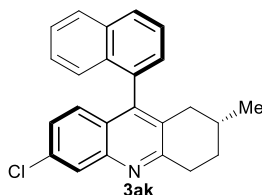


6-Chloro-9-(naphthalen-1-yl)-2-phenyl-1,2,3,4-tetrahydroacridine **3ai** was obtained as a yellow solid in 71% yield (29.8 mg), 90:10 dr and 92% ee for the major diastereoisomer. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 11.15 min, t_r (minor) = 12.06 min]. m.p. 217-218 °C; $[\alpha]_D^{25} = +115.78$ ($c = 1.0$, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.10 (d, $J = 2.4$ Hz, 1H), 7.97-7.89 (m, 2H), 7.64-7.55 (m, 1H), 7.50-7.43 (m, 1H), 7.35-7.27 (m, 2H), 7.22-7.07 (m, 7H), 7.03 (d, $J = 8.8$ Hz, 1H), 3.51-3.27 (m, 2H), 3.08-2.93 (m, 1H), 2.86-2.76 (m, 1H), 2.49-2.38 (m, 1H), 2.32-2.22 (m, 1H), 2.19-2.09 (m, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 159.5, 146.8, 145.5, 145.4, 134.6, 133.9, 133.8, 131.4, 129.4, 128.7, 128.7, 128.5, 128.5, 127.5, 127.4, 126.8, 126.8, 126.7, 126.6, 126.4, 126.3, 125.7, 125.7, 125.6, 125.1, 40.4, 35.1, 34.3, 30.1; HRMS (ESI) calcd for $\text{C}_{29}\text{H}_{23}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 420.1514, found 420.1512.

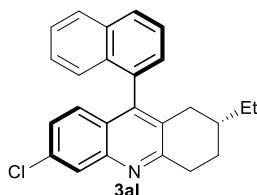


6-Chloro-2-(4-chlorophenyl)-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3aj** was obtained

as a yellow solid in 74% yield (33.5 mg), 88:12 dr and 92% ee for the major diastereoisomer. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 10.08 min, t_r (minor) = 10.95 min]. m.p. 214-215 °C; $[\alpha]_D^{25} = +128.30$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.09 (d, $J = 2.4$ Hz, 1H), 7.95 (t, $J = 7.6$ Hz, 2H), 7.60 (t, $J = 7.6$ Hz, 1H), 7.49 (t, $J = 7.6$ Hz, 1H), 7.37-7.28 (m, 2H), 7.21-7.13 (m, 3H), 7.10 (d, $J = 8.4$ Hz, 1H), 7.03 (t, $J = 8.0$ Hz, 3H), 3.51-3.27 (m, 2H), 3.08-2.93 (m, 1H), 2.86-2.76 (m, 1H), 2.49-2.38 (m, 1H), 2.32-2.22 (m, 1H), 2.19-2.09 (m, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 159.2, 146.9, 145.4, 143.7, 134.6, 133.7, 133.7, 132.0, 131.2, 128.9, 128.7, 128.6, 128.5, 128.1, 127.4, 127.4, 126.8, 126.7, 126.6, 126.3, 125.6, 125.5, 125.0, 39.7, 35.0, 34.0, 29.9; HRMS (ESI) calcd for $\text{C}_{29}\text{H}_{22}\text{Cl}_2\text{N}$ (M+H) $^+$ 454.1124, found 454.1123.

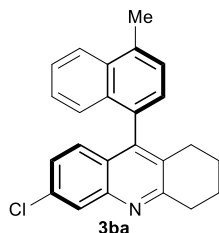


6-Chloro-2-methyl-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3ak** was obtained as a yellow solid in 68% yield (24.3 mg), 75:25 dr and 87% ee for the major diastereoisomer. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak OD-H, isopropanol/hexane (04:96), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 5.09 min, t_r (minor) = 5.46 min]. m.p. 117-118 °C; $[\alpha]_D^{25} = +68.14$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.07 (d, $J = 2.0$ Hz, 1H), 7.97 (t, $J = 8.8$ Hz, 2H), 7.64-7.58 (m, 1H), 7.52-7.46 (m, 1H), 7.33-7.27 (m, 2H), 7.16-7.08 (m, 2H), 7.02 (d, $J = 8.8$ Hz, 1H), 3.39-3.30 (m, 1H), 3.27-3.15 (m, 1H), 2.65-2.55 (m, 1H), 2.08-2.00 (m, 1H), 1.92-1.80 (m, 1H), 1.66-1.53 (m, 2H), 0.87 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 160.1, 146.8, 145.0, 134.3, 134.2, 133.7, 131.4, 129.6, 128.6, 128.5, 127.4, 127.4, 126.7, 126.7, 126.5, 126.3, 125.7, 125.6, 125.3, 35.5, 33.8, 31.0, 29.0, 21.5; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{21}\text{ClN}$ (M+H) $^+$ 358.1357, found 358.1356.

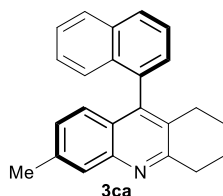


6-Chloro-2-ethyl-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3al** was obtained as a white solid in 75% yield (27.8 mg), 88:12 dr and 91% ee for the major diastereoisomer. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 8.47 min, t_r (minor) = 9.70 min]. m.p. 135-136 °C; $[\alpha]_D^{25} = +80.71$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.07

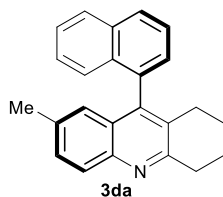
(d, $J = 2.0$ Hz, 1H), 7.99 (t, $J = 8.4$ Hz, 2H), 7.67-7.60 (m, 1H), 7.54-7.47 (m, 1H), 7.35-7.27 (m, 2H), 7.16 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.10 (d, $J = 8.4$ Hz, 1H), 7.02 (d, $J = 8.8$ Hz, 1H), 3.39-3.29 (m, 1H), 3.25-3.11 (m, 1H), 2.68-2.56 (m, 1H), 2.18-2.06 (m, 1H), 2.04-1.95 (m, 1H), 1.68-1.51 (m, 2H), 1.25-1.15 (m, 2H), 0.78 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.5, 146.8, 145.1, 134.3, 134.2, 133.7, 131.4, 129.7, 128.6, 128.5, 128.5, 127.4, 127.4, 126.7, 126.5, 126.3, 125.7, 125.6, 125.3, 35.5, 33.7, 33.5, 28.4, 28.2, 11.5; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{23}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 372.1514, found 372.1512.



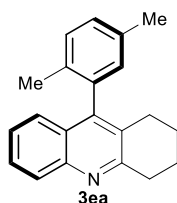
6-Chloro-9-(4-methylnaphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3ba** was obtained as a white solid in 79% yield (28.2 mg) and 92% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 7.24 min, t_r (major) = 7.68 min]. m.p. 95-96 °C; $[\alpha]_{\text{D}}^{25} = +30.05$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.19-7.94 (m, 2H), 7.58-7.40 (m, 2H), 7.35-7.26 (m, 1H), 7.21-6.98 (m, 4H), 3.33-3.14 (m, 2H), 2.80 (s, 3H), 2.60-2.25 (m, 2H), 2.01-1.61 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.4, 146.7, 145.4, 135.0, 134.2, 132.8, 132.4, 131.4, 130.2, 127.4, 127.4, 126.6, 126.4, 126.4, 126.3, 126.1, 125.9, 125.8, 124.7, 34.4, 27.5, 22.9, 22.8, 19.6; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{21}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 358.1357, found 358.1356.



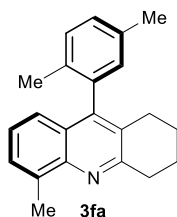
6-Methyl-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3ca** was obtained as a yellow solid in 82% yield (26.5 mg) and 90% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AS-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 7.23 min, t_r (minor) = 8.07 min]. m.p. 118-119 °C; $[\alpha]_{\text{D}}^{25} = +15.17$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.00-7.90 (m, 2H), 7.87 (s, 1H), 7.59 (t, $J = 7.6$ Hz, 1H), 7.47 (t, $J = 7.6$ Hz, 1H), 7.34-7.23 (m, 2H), 7.16 (d, $J = 8.4$ Hz, 1H), 7.07-6.94 (m, 2H), 3.24 (t, $J = 6.8$ Hz, 2H), 2.59-2.42 (m, 4H), 2.36-2.24 (m, 1H), 2.00-1.86 (m, 2H), 1.79-1.60 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 159.0, 146.5, 144.9, 138.7, 135.0, 133.6, 131.5, 128.8, 128.5, 128.2, 127.8, 127.3, 126.8, 126.5, 126.2, 125.6, 125.6, 125.4, 125.3, 34.2, 27.4, 23.0, 23.0, 21.8; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{22}\text{N}$ ($\text{M}+\text{H}$) $^+$ 324.1747, found 324.1745.



7-Methyl-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine **3da** was obtained as a white solid in 77% yield (24.9 mg) and 70% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (08:92), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 8.09 min, t_r (minor) = 8.96 min]. m.p. 179-180 °C; $[\alpha]_D^{25} = +9.85$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.02-7.88 (m, 3H), 7.59 (t, $J = 8.0$ Hz, 1H), 7.50-7.38 (m, 2H), 7.31-7.25 (m, 2H), 7.19 (d, $J = 8.4$ Hz, 1H), 6.86 (s, 1H), 3.23 (t, $J = 6.8$ Hz, 2H), 2.55-2.44 (m, 1H), 2.35-2.25 (m, 1H), 2.21 (s, 3H), 1.99-1.89 (m, 2H), 1.76-1.61 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 158.0, 145.0, 144.3, 135.3, 135.1, 133.7, 131.5, 130.9, 129.6, 128.5, 128.2, 127.2, 126.9, 126.6, 126.2, 125.7, 125.5, 124.6, 34.3, 27.5, 23.1, 23.0, 21.7; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{22}\text{N}$ ($\text{M}+\text{H}$) $^+$ 324.1747, found 324.1746.

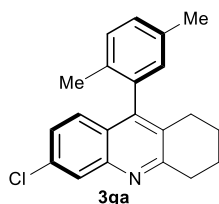


9-(2,5-Dimethylphenyl)-1,2,3,4-tetrahydroacridine **3ea** was obtained as a white solid in 73% yield (21.0 mg) and 76% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (10:90), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 4.44 min, t_r (major) = 5.77 min]. m.p. 134-135 °C; $[\alpha]_D^{25} = -5.86$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.03 (d, $J = 8.4$ Hz, 1H), 7.59 (t, $J = 7.6$ Hz, 1H), 7.35-7.12 (m, 4H), 6.87 (s, 1H), 3.21 (t, $J = 6.8$ Hz, 2H), 2.66-2.50 (m, 1H), 2.45-2.38 (m, 1H), 2.36 (s, 3H), 2.04-1.91 (m, 2H), 1.86 (s, 3H), 1.83-1.73 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 159.2, 146.5, 146.3, 136.5, 135.6, 132.6, 130.1, 129.4, 128.7, 128.5, 128.4, 126.4, 125.5, 34.3, 27.7, 23.0, 23.0, 21.0, 19.0; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{22}\text{N}$ ($\text{M}+\text{H}$) $^+$ 288.1747, found 288.1746.

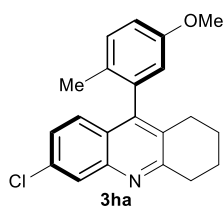


9-(2,5-Dimethylphenyl)-5-methyl-1,2,3,4-tetrahydroacridine **3fa** was obtained as a white

solid in 82% yield (24.7 mg) and 94% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 3.37 min, t_r (major) = 3.95 min]. m.p. 130-131 °C; $[\alpha]_D^{25} = +12.43$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.43 (d, $J = 7.2$ Hz, 1H), 7.23 (d, $J = 8.0$ Hz, 1H), 7.20-7.13 (m, 2H), 7.03 (d, $J = 8.4$ Hz, 1H), 6.85 (s, 1H), 3.21 (t, $J = 6.8$ Hz, 2H), 2.83 (s, 3H), 2.63-2.51 (m, 1H), 2.42-2.30 (m, 4H), 2.00-1.91 (m, 2H), 1.87-1.71 (m, 5H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 158.0, 146.4, 146.3, 137.0, 136.2, 135.5, 132.6, 130.1, 129.4, 128.6, 128.5, 128.0, 126.3, 125.1, 123.5, 34.7, 27.7, 23.2, 23.2, 21.1, 19.1, 18.2; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{24}\text{N}$ (M+H) $^+$ 302.1903, found 302.1902.

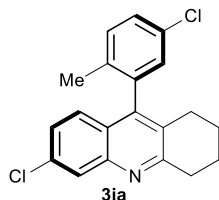


6-Chloro-9-(2,5-dimethylphenyl)-1,2,3,4-tetrahydroacridine **3ga** was obtained as a yellow solid in 78% yield (25.1 mg) and 95% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 7.24 min, t_r (major) = 9.42 min]. m.p. 129-130 °C; $[\alpha]_D^{25} = +22.86$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.02 (d, $J = 2.0$ Hz, 1H), 7.28-7.21 (m, 2H), 7.20-7.16 (m, 1H), 7.14 (d, $J = 8.8$ Hz, 1H), 6.85 (s, 1H), 3.18 (t, $J = 6.8$ Hz, 2H), 2.64-2.53 (m, 1H), 2.43-2.37 (m, 1H), 2.36 (s, 3H), 2.00-1.92 (m, 2H), 1.85 (s, 3H), 1.83-1.72 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 160.5, 146.7, 146.5, 136.9, 135.7, 134.2, 132.5, 130.3, 129.2, 129.0, 128.8, 127.4, 126.9, 126.4, 124.9, 34.3, 27.6, 22.9, 21.0, 19.0; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{21}\text{ClN}$ (M+H) $^+$ 322.1357, found 322.1356.

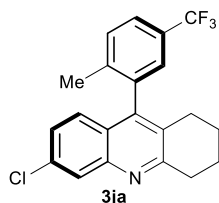


6-Chloro-9-(5-methoxy-2-methylphenyl)-1,2,3,4-tetrahydroacridine **3ha** was obtained as a yellow solid in 64% yield (21.6 mg) and 94% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (10:90), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 5.66 min, t_r (major) = 8.51 min]. m.p. 131-132 °C; $[\alpha]_D^{25} = +33.33$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.02 (d, $J = 2.0$ Hz, 1H), 7.31-7.22 (m, 2H), 7.17 (d, $J = 9.2$ Hz, 1H), 6.92 (dd, $J = 8.4, 2.8$ Hz, 1H), 6.62 (d, $J = 2.8$ Hz, 1H),

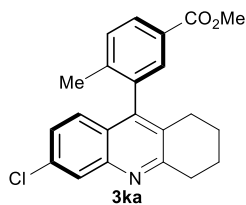
3.79 (s, 3H), 3.18 (t, $J = 6.8$ Hz, 2H), 2.69-2.55 (m, 1H), 2.48-2.35 (m, 1H), 2.04-1.91 (m, 2H), 1.88-1.72 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.5, 158.0, 146.7, 146.1, 137.0, 134.2, 131.4, 128.8, 127.5, 127.4, 126.9, 126.5, 124.7, 114.1, 113.7, 55.3, 34.2, 27.5, 22.9, 22.8, 18.5; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{21}\text{ClNO}$ ($\text{M}+\text{H}$) $^+$ 338.1306, found 338.1305.



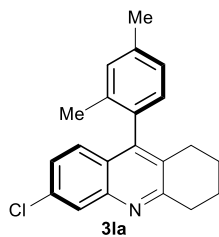
6-Chloro-9-(5-chloro-2-methylphenyl)-1,2,3,4-tetrahydroacridine **3ia** was obtained as a white solid in 80% yield (27.3 mg) and 94% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 7.05 min, t_r (major) = 8.42 min]. m.p. 107-108 °C; $[\alpha]_{\text{D}}^{25} = +27.75$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.03 (d, $J = 2.0$ Hz, 1H), 7.38-7.34 (m, 1H), 7.33-7.24 (m, 2H), 7.14-7.03 (m, 2H), 3.18 (t, $J = 6.8$ Hz, 2H), 2.65-2.53 (m, 1H), 2.42-2.30 (m, 1H), 2.01-1.93 (m, 2H), 1.86 (s, 3H), 1.85-1.74 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.6, 146.7, 144.7, 137.8, 134.4, 134.3, 132.0, 131.8, 128.8, 128.6, 128.4, 127.6, 126.8, 126.4, 124.4, 34.2, 27.6, 22.8, 18.9; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{18}\text{Cl}_2\text{N}$ ($\text{M}+\text{H}$) $^+$ 342.0811, found 342.0810.



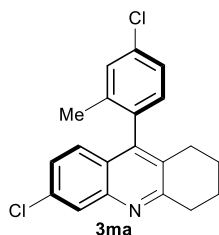
6-Chloro-9-(2-methyl-5-(trifluoromethyl)phenyl)-1,2,3,4-tetrahydroacridine **3ja** was obtained as a yellow solid in 76% yield (28.5 mg) and 91% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (08:92), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 4.63 min, t_r (minor) = 5.04 min]. m.p. 149-150 °C; $[\alpha]_{\text{D}}^{25} = -15.57$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.04 (d, $J = 2.0$ Hz, 1H), 7.65 (d, $J = 8.0$ Hz, 1H), 7.51 (d, $J = 8.0$ Hz, 1H), 7.35 (s, 1H), 7.28 (dd, $J = 8.4, 2.0$ Hz, 1H), 7.04 (d, $J = 9.2$ Hz, 1H), 3.20 (t, $J = 6.8$ Hz, 2H), 2.62-2.50 (m, 1H), 2.41-2.28 (m, 1H), 2.04-1.93 (m, 5H), 1.90-1.72 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.7, 146.7, 144.5, 140.2, 136.8, 134.5, 131.0, 129.0 (q, $J = 32.4$ Hz), 128.8, 127.7, 126.9, 126.2, 125.7 (q, $J = 3.7$ Hz), 125.2 (q, $J = 3.7$ Hz), 124.3, 124.1 (q, $J = 270.4$ Hz), 34.2, 27.6, 22.8, 22.7, 19.5; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{18}\text{ClF}_3\text{N}$ ($\text{M}+\text{H}$) $^+$ 376.1074, found 376.1073.



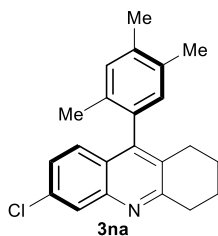
Methyl 3-(6-chloro-1,2,3,4-tetrahydroacridin-9-yl)-4-methylbenzoate **3ka** was obtained as a yellow solid in 62% yield (22.6 mg) and 95% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (15:85), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 7.69 min, t_r (major) = 13.74 min]. m.p. 162-163 °C; $[\alpha]_D^{25} = +35.16$ ($c = 1.0$, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.11-8.00 (m, 2H), 7.77 (d, $J = 2.0$ Hz, 1H), 7.46 (d, $J = 8.0$ Hz, 1H), 7.26 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.06 (d, $J = 8.8$ Hz, 1H), 3.90 (s, 3H), 3.20 (t, $J = 6.8$ Hz, 2H), 2.65-2.51 (m, 1H), 2.41-2.29 (m, 1H), 2.04-1.93 (m, 5H), 1.90-1.72 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 166.7, 160.6, 146.7, 145.0, 141.5, 136.4, 134.4, 130.7, 130.1, 129.5, 128.9, 128.5, 127.6, 126.8, 126.4, 124.5, 52.2, 34.2, 27.6, 22.8, 22.8, 19.7; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{21}\text{ClNO}_2$ ($\text{M}+\text{H}$) $^+$ 366.1255, found 366.1254.



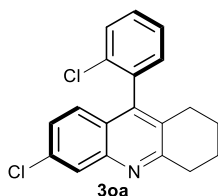
6-Chloro-9-(2,4-dimethylphenyl)-1,2,3,4-tetrahydroacridine **3la** was obtained as a yellow solid in 66% yield (21.2 mg) and 94% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak IC, isopropanol/hexane (07:93), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 5.51 min, t_r (major) = 6.04 min]. m.p. 95-96 °C; $[\alpha]_D^{25} = +20.14$ ($c = 1.0$, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.02 (d, $J = 2.0$ Hz, 1H), 7.22 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.18 (s, 1H), 7.14-7.10 (m, 2H), 6.92 (d, $J = 7.6$ Hz, 1H), 3.18 (t, $J = 6.8$ Hz, 2H), 2.64-2.53 (m, 1H), 2.42 (s, 3H), 2.41-2.34 (m, 1H), 2.00-1.92 (m, 2H), 1.86 (s, 3H), 1.84-1.72 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 160.5, 146.7, 146.4, 137.9, 135.4, 134.2, 133.0, 131.1, 129.1, 128.7, 127.4, 127.0, 126.9, 126.4, 125.1, 34.3, 27.6, 22.9, 21.3, 19.4; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{21}\text{ClN}$ ($\text{M}+\text{H}$) $^+$ 322.1357, found 322.1356.



6-Chloro-9-(4-chloro-2-methylphenyl)-1,2,3,4-tetrahydroacridine **3ma** was obtained as a yellow solid in 70% yield (23.9 mg) and 94% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 4.82 min, t_r (major) = 6.70 min]. m.p. 115-116 °C; $[\alpha]_D^{25} = +27.50$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.02 (d, $J = 2.0$ Hz, 1H), 7.38 (d, $J = 2.4$ Hz, 1H), 7.32 (dd, $J = 8.0, 2.0$ Hz, 1H), 7.26 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.10 (d, $J = 8.8$ Hz, 1H), 6.99 (d, $J = 8.0$ Hz, 1H), 3.18 (t, $J = 6.8$ Hz, 2H), 2.61-2.50 (m, 1H), 2.41-2.31 (m, 1H), 2.02-1.93 (m, 2H), 1.89 (s, 3H), 1.86-1.74 (m, 2H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 160.6, 146.7, 145.0, 137.8, 134.5, 134.4, 134.0, 130.4, 130.1, 129.0, 127.6, 126.7, 126.6, 126.4, 124.6, 34.2, 27.6, 22.8, 19.4; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{18}\text{Cl}_2\text{N}$ (M+H) $^+$ 342.0811, found 342.0810.

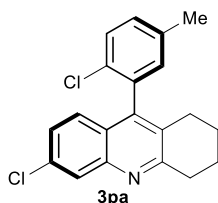


6-Chloro-9-(2,4,5-trimethylphenyl)-1,2,3,4-tetrahydroacridine **3na** was obtained as a yellow solid in 81% yield (27.2 mg) and 93% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 4.58 min, t_r (major) = 6.43 min]. m.p. 129-130 °C; $[\alpha]_D^{25} = +33.75$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.01 (d, $J = 2.0$ Hz, 1H), 7.23 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.18-7.10 (m, 2H), 6.80 (s, 1H), 3.18 (t, $J = 6.8$ Hz, 2H), 2.65-2.54 (m, 1H), 2.44-2.35 (m, 1H), 2.33 (s, 3H), 2.27 (s, 3H), 2.00-1.91 (m, 2H), 1.85-1.71 (m, 5H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 160.5, 146.7, 146.6, 136.5, 134.3, 134.1, 133.3, 132.8, 131.6, 129.8, 129.0, 127.3, 127.1, 126.3, 125.1, 34.3, 27.7, 22.9, 19.6, 19.4, 18.9; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{23}\text{ClN}$ (M+H) $^+$ 336.1514, found 336.1512.

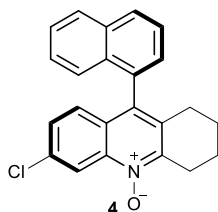


6-Chloro-9-(2-chlorophenyl)-1,2,3,4-tetrahydroacridine **3oa** was obtained as a yellow solid in 71% yield (23.2 mg) and 80% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (10:90), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 5.61 min, t_r (major) = 8.46 min]. m.p. 139-140 °C; $[\alpha]_D^{25} = +13.17$ (c = 1.0, EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.03 (d, $J = 2.0$ Hz, 1H), 7.59-7.53 (m, 1H),

7.47-7.38 (m, 2H), 7.26 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.19-7.14 (m, 1H), 7.11 (d, $J = 8.8$ Hz, 1H), 3.27-3.10 (m, 2H), 2.63-2.45 (m, 2H), 2.05-1.91 (m, 2H), 1.88-1.75 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.5, 146.7, 143.7, 135.5, 134.3, 133.2, 130.6, 130.0, 129.7, 129.3, 127.5, 127.2, 126.7, 126.5, 124.5, 34.2, 27.4, 22.8, 22.7; HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{16}\text{Cl}_2\text{N}(\text{M}+\text{H})^+$ 328.0654, found 328.0653.

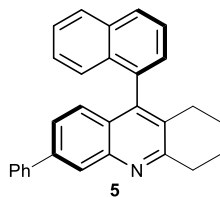


6-Chloro-9-(2-chloro-5-methylphenyl)-1,2,3,4-tetrahydroacridine **3pa** was obtained as a yellow solid in 72% yield (24.6 mg) and 86% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (05:95), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 8.45 min, t_r (major) = 10.16 min]. m.p. 150-151 °C; $[\alpha]_{\text{D}}^{25} = +38.42$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.03 (d, $J = 2.0$ Hz, 1H), 7.44 (d, $J = 8.4$ Hz, 1H), 7.31-7.20 (m, 2H), 7.13 (d, $J = 8.8$ Hz, 1H), 6.97 (d, $J = 2.4$ Hz, 1H), 3.25-3.11 (m, 2H), 2.54 (t, $J = 6.4$ Hz, 2H), 2.39 (s, 3H), 2.03-1.92 (m, 2H), 1.86-1.76 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.5, 146.6, 143.9, 137.2, 135.1, 134.3, 131.0, 130.5, 130.1, 129.6, 129.3, 127.5, 126.6, 126.6, 124.6, 34.2, 27.4, 22.8, 22.7, 20.9; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{18}\text{Cl}_2\text{N}(\text{M}+\text{H})^+$ 342.0811, found 342.0810.

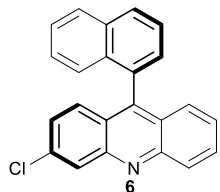


6-Chloro-9-(naphthalen-1-yl)-1,2,3,4-tetrahydroacridine 10-oxide **4** was obtained as a white solid in 94% yield (33.8 mg) and 87% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (30:70), 1.0 mL/min, $\lambda = 254$ nm, t_r (minor) = 14.87 min, t_r (major) = 18.22 min]. m.p. 91-92 °C; $[\alpha]_{\text{D}}^{25} = +17.78$ ($c = 1.0$, EtOAc); ^1H NMR (400 MHz, CDCl_3): δ 8.88 (d, $J = 2.4$ Hz, 1H), 8.05-7.94 (m, 2H), 7.64 (t, $J = 7.2$ Hz, 1H), 7.53 (t, $J = 7.2$ Hz, 1H), 7.38-7.31 (m, 2H), 7.27 (dd, $J = 8.8, 2.0$ Hz, 1H), 7.17 (d, $J = 8.4$ Hz, 1H), 7.10 (d, $J = 8.8$ Hz, 1H), 3.34 (t, $J = 6.8$ Hz, 2H), 2.59-2.47 (m, 1H), 2.36-2.25 (m, 1H), 2.01-1.90 (m, 2H), 1.75-1.60 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 147.3, 138.9, 135.1, 134.4, 132.7, 132.0, 130.7, 130.5, 128.0, 127.7, 127.6, 127.0, 126.6, 126.0, 125.9, 125.4, 124.6, 123.9, 117.8, 26.6, 25.7, 20.8, 20.6; HRMS (ESI) calcd for

$C_{23}H_{19}ClNO$ ($M+H$)⁺ 360.1150, found 360.1149.



9-(Naphthalen-1-yl)-6-phenyl-1,2,3,4-tetrahydroacridine **5** was obtained as a yellow oil in 84% yield (32.4 mg) and 88% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak OD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 11.11 min, t_r (minor) = 12.35 min]. $[\alpha]_D^{25} = -41.50$ ($c = 1.0$, EtOAc); 1H NMR (400 MHz, $CDCl_3$): δ 8.35 (s, 1H), 7.97 (t, $J = 7.6$ Hz, 2H), 7.71 (d, $J = 7.6$ Hz, 2H), 7.62 (t, $J = 7.6$ Hz, 1H), 7.53-7.40 (m, 4H), 7.38-7.27 (m, 3H), 7.23-7.11 (m, 2H), 3.29 (t, $J = 6.8$ Hz, 2H), 2.64-2.50 (m, 1H), 2.41-2.29 (m, 1H), 2.03-1.90 (m, 2H), 1.81-1.64 (m, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 159.55, 146.41, 145.04, 141.17, 140.38, 134.64, 133.65, 131.41, 129.68, 128.88, 128.48, 128.34, 127.63, 127.31, 126.85, 126.61, 126.37, 126.21, 125.83, 125.57, 125.35, 125.19, 34.16, 27.48, 22.89, 22.86; HRMS (ESI) calcd for $C_{29}H_{24}N$ ($M+H$)⁺ 386.1903, found 386.1925.

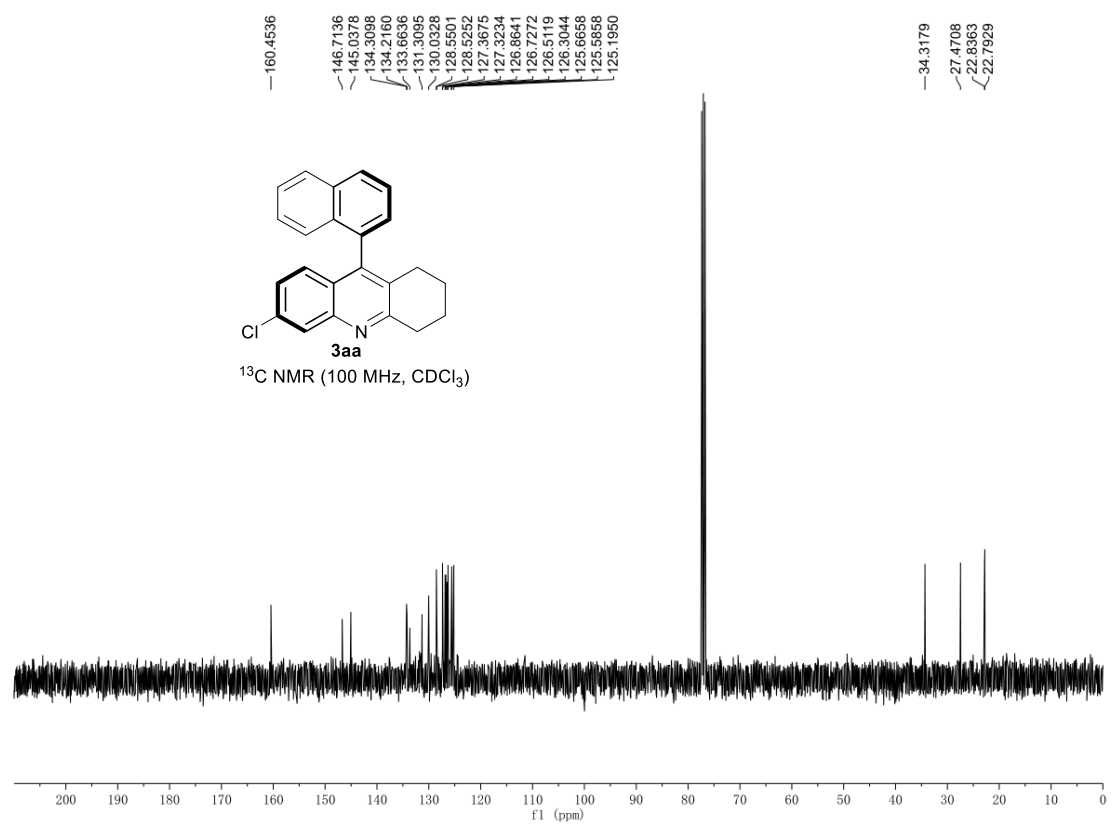
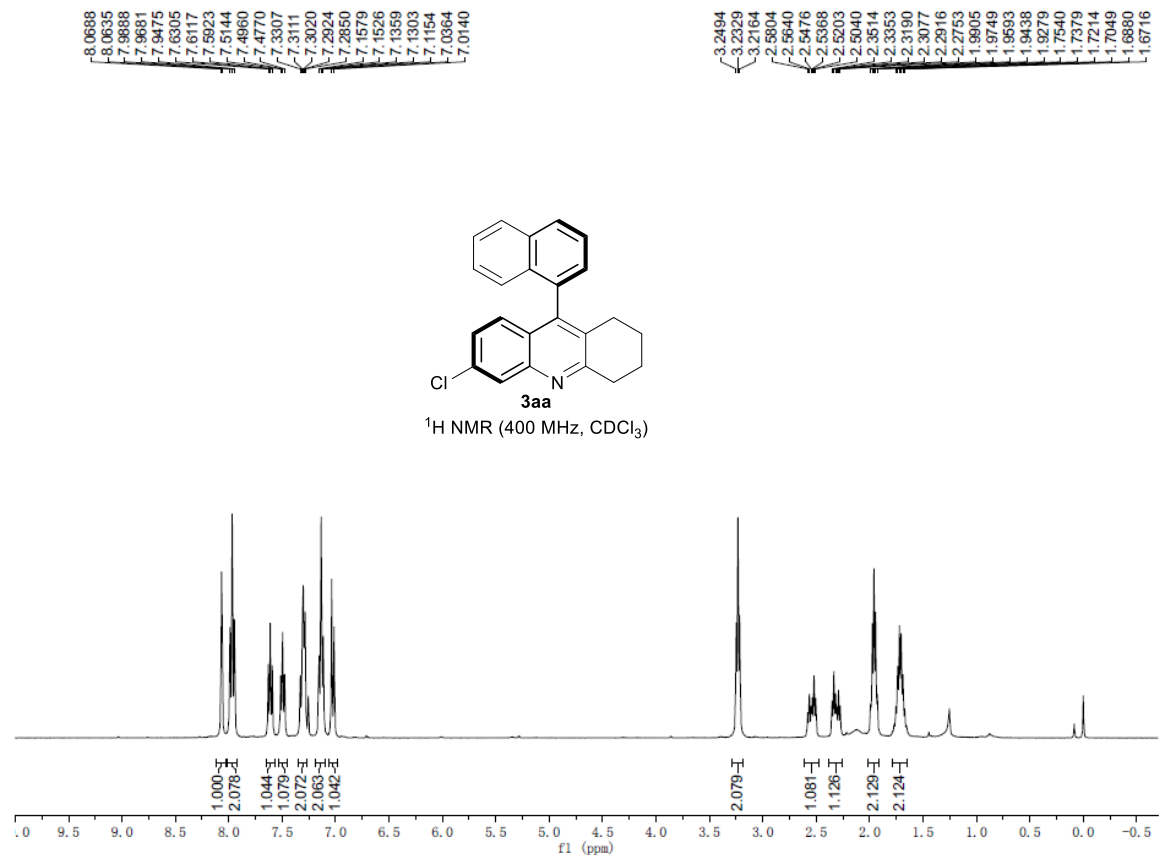


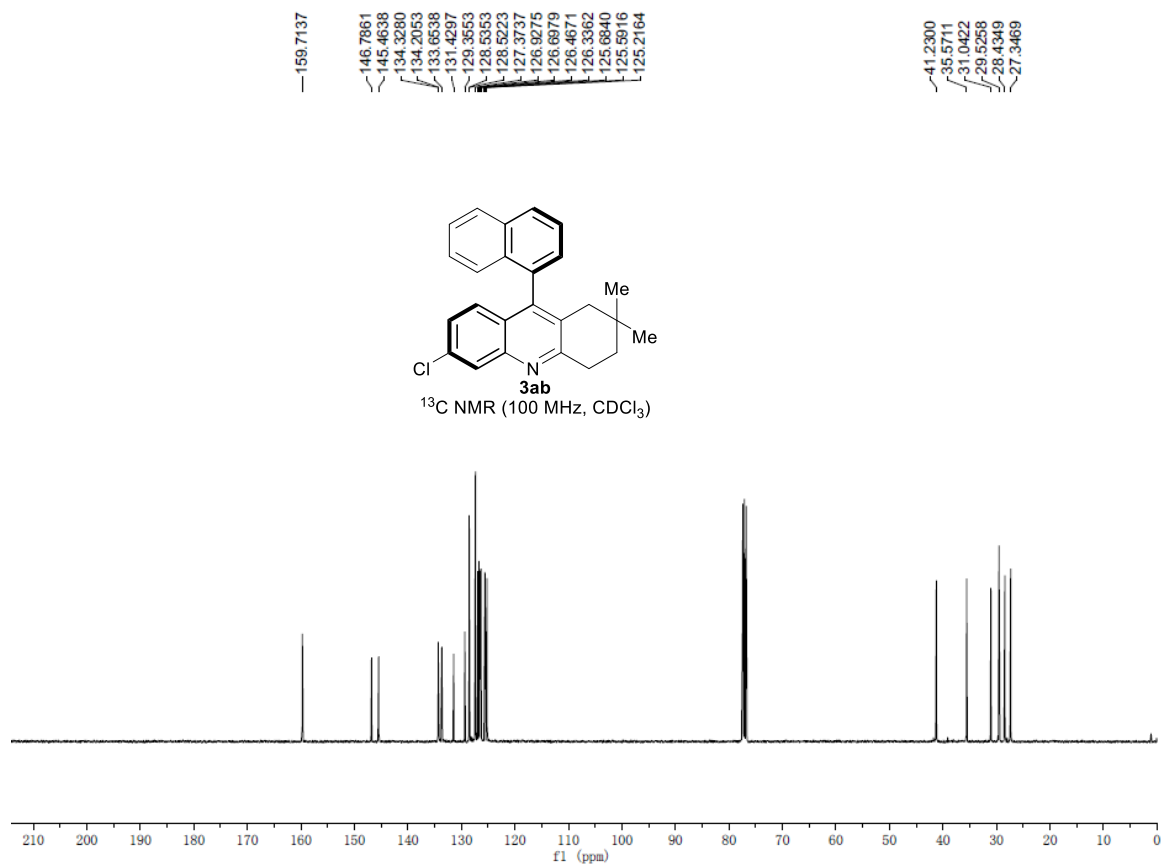
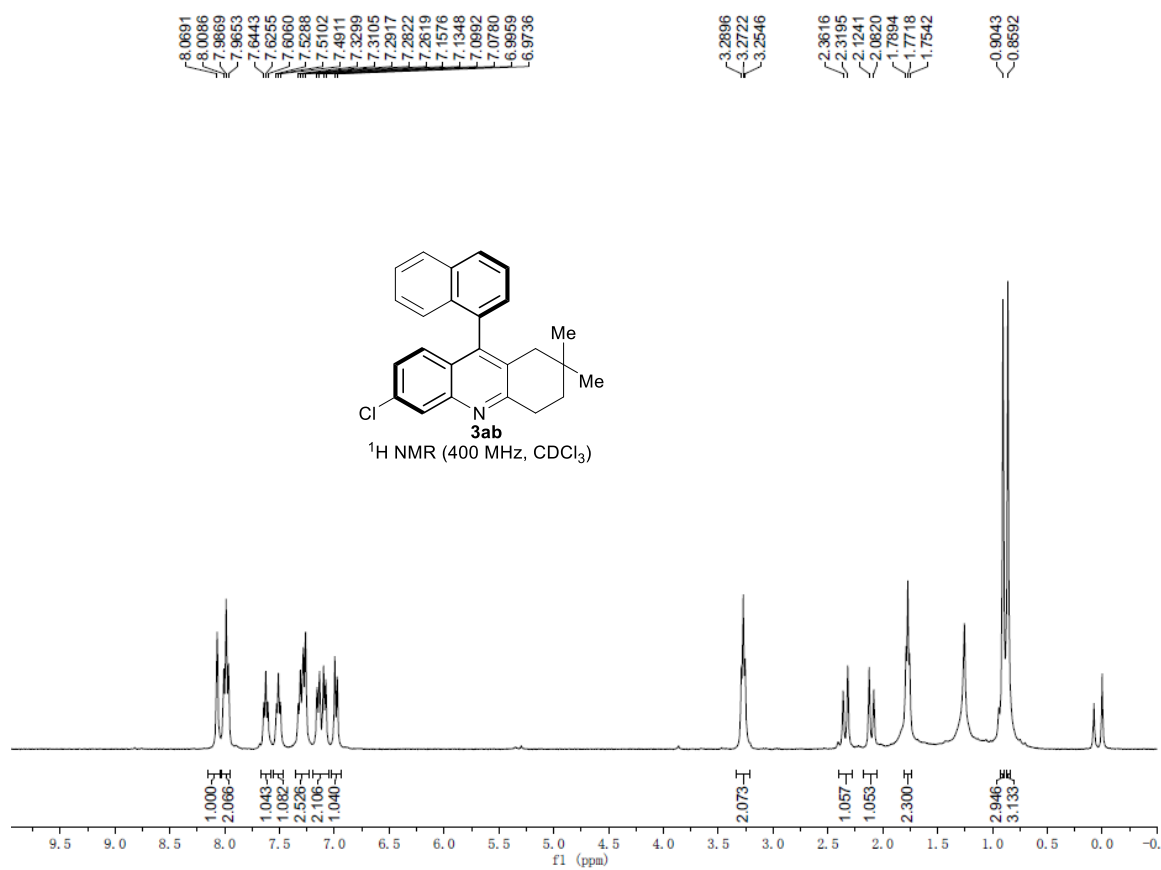
3-Chloro-9-(naphthalen-1-yl)acridine **6** was obtained as a yellow solid in 34% yield (11.5 mg) and 84% ee. The ee value was determined by chiral stationary phase HPLC analysis [Daicel Chiralpak AD-H, isopropanol/hexane (03:97), 1.0 mL/min, $\lambda = 254$ nm, t_r (major) = 6.74 min, t_r (minor) = 7.14 min]. m.p. 213-214 °C; $[\alpha]_D^{25} = +30.04$ ($c = 1.0$, EtOAc); 1H NMR (400 MHz, $CDCl_3$): δ 8.38-8.24 (m, 2H), 8.10 (d, $J = 8.4$ Hz, 1H), 8.03 (d, $J = 8.4$ Hz, 1H), 7.79 (t, $J = 7.6$ Hz, 1H), 7.70 (t, $J = 7.6$ Hz, 1H), 7.56-7.47 (m, 2H), 7.47-7.31 (m, 3H), 7.30-7.19 (m, 2H), 6.99 (d, $J = 8.4$ Hz, 1H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 148.3, 147.7, 145.2, 135.2, 132.5, 132.0, 131.4, 129.9, 129.7, 128.5, 128.1, 127.8, 127.4, 127.0, 126.1, 126.0, 125.8, 125.4, 125.1, 125.0, 124.9, 124.3, 123.3; HRMS (ESI) calcd for $C_{23}H_{15}ClN$ ($M+H$)⁺ 340.0888, found 340.0887.

References

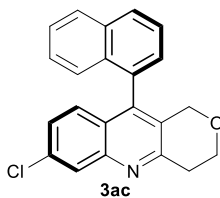
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¹H and ¹³C NMR Spectra

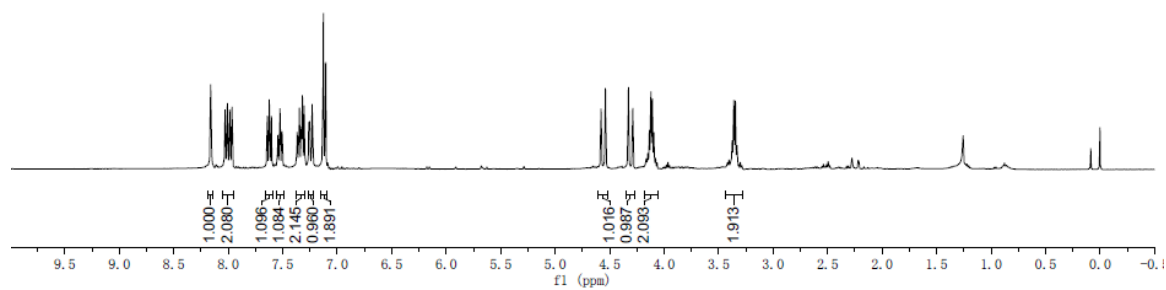




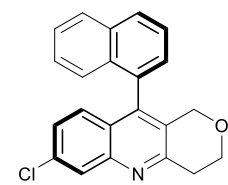
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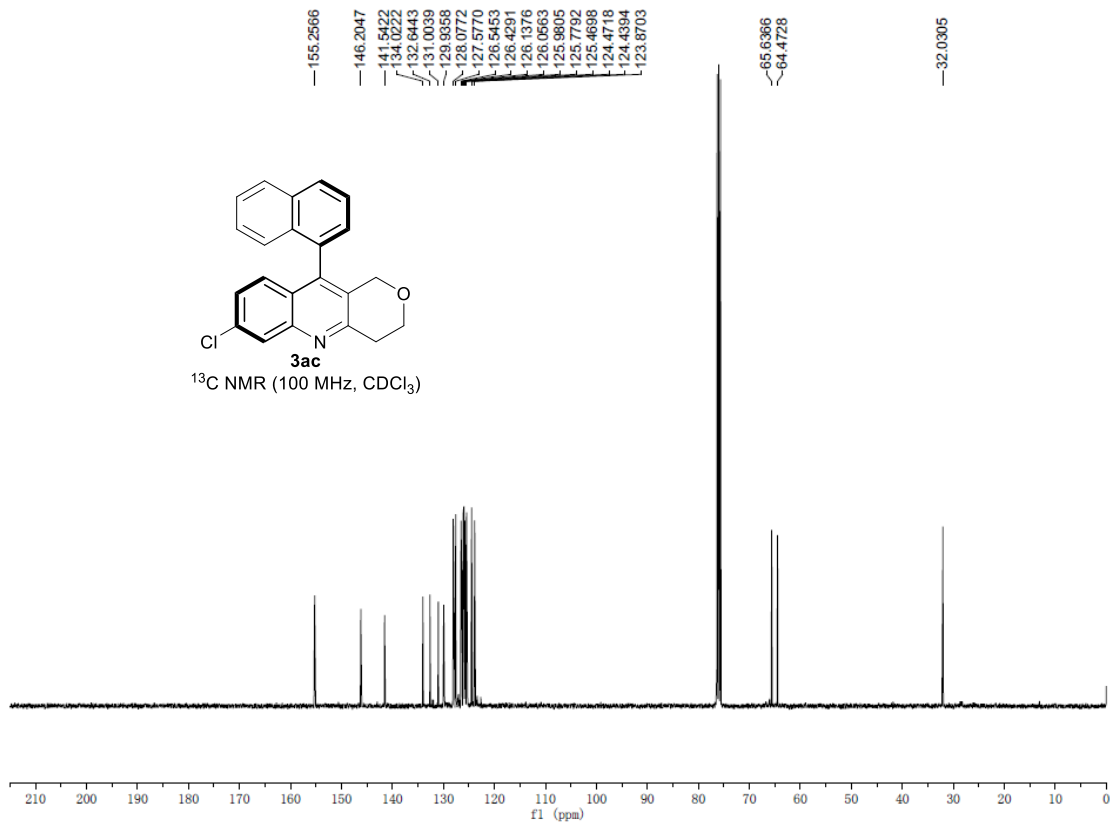
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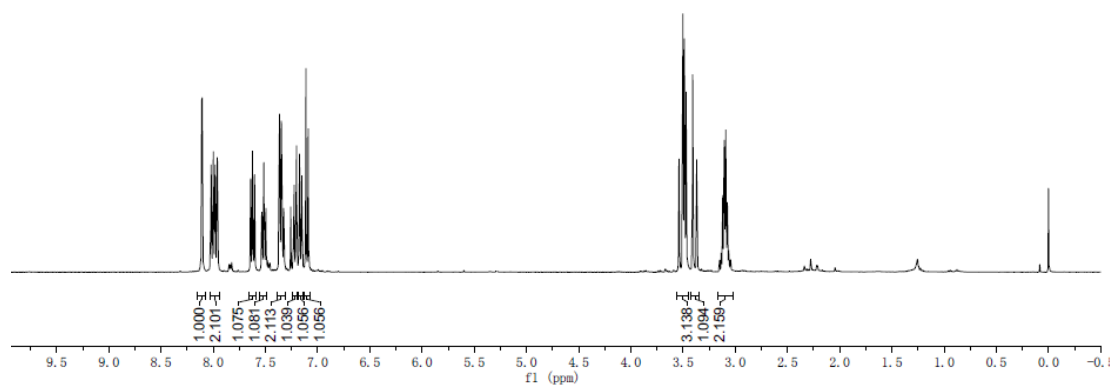
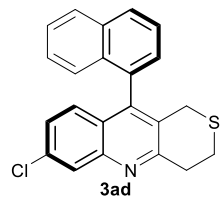
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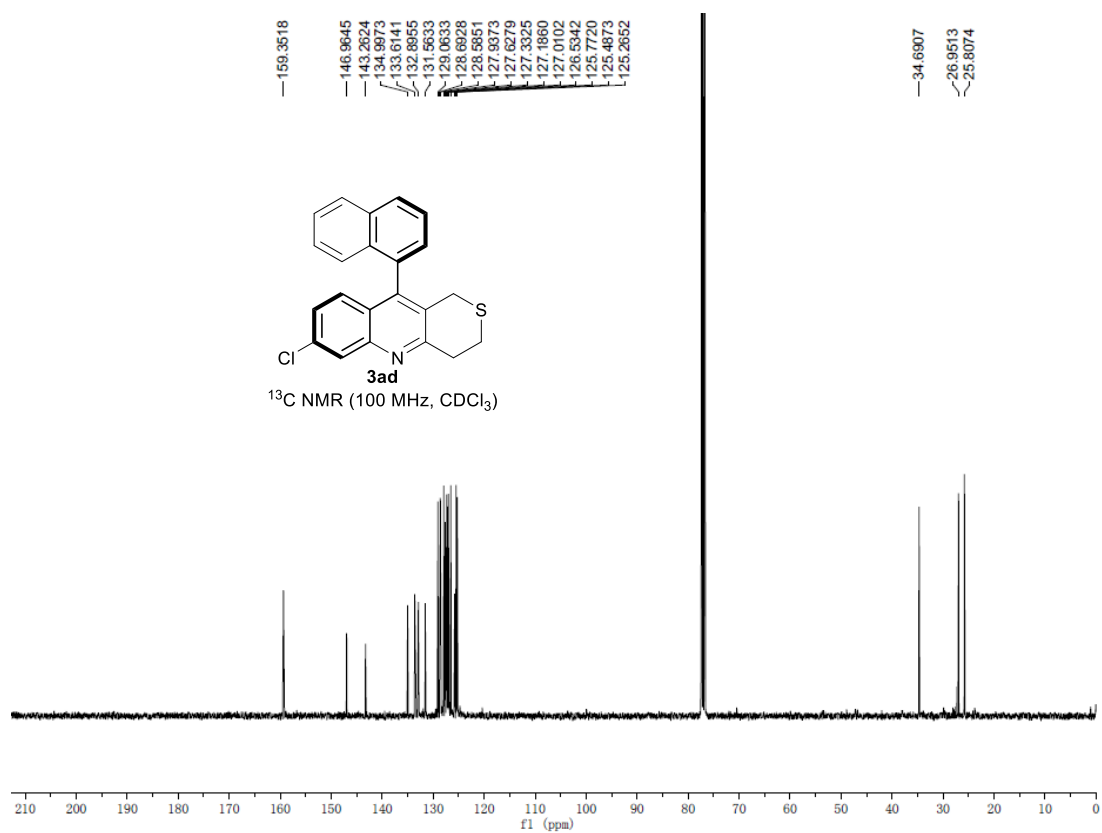
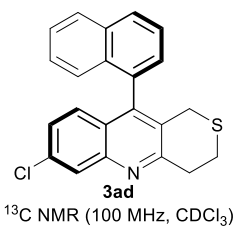
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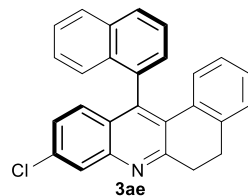
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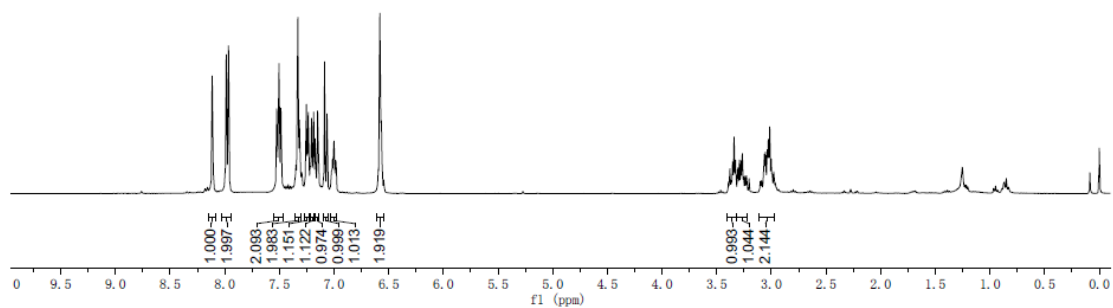
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7.1454
7.0867
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7.0146
7.0067
7.0004
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3.3414
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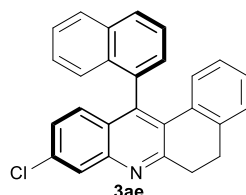


$^1\text{H NMR}$ (400 MHz, CDCl_3)

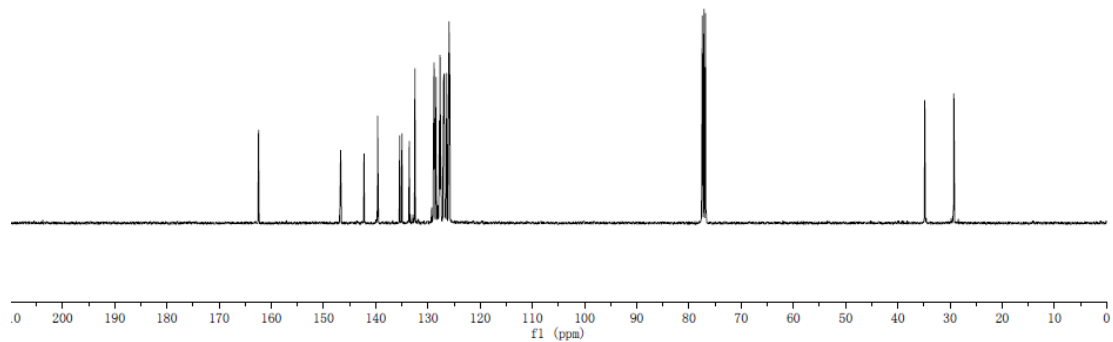


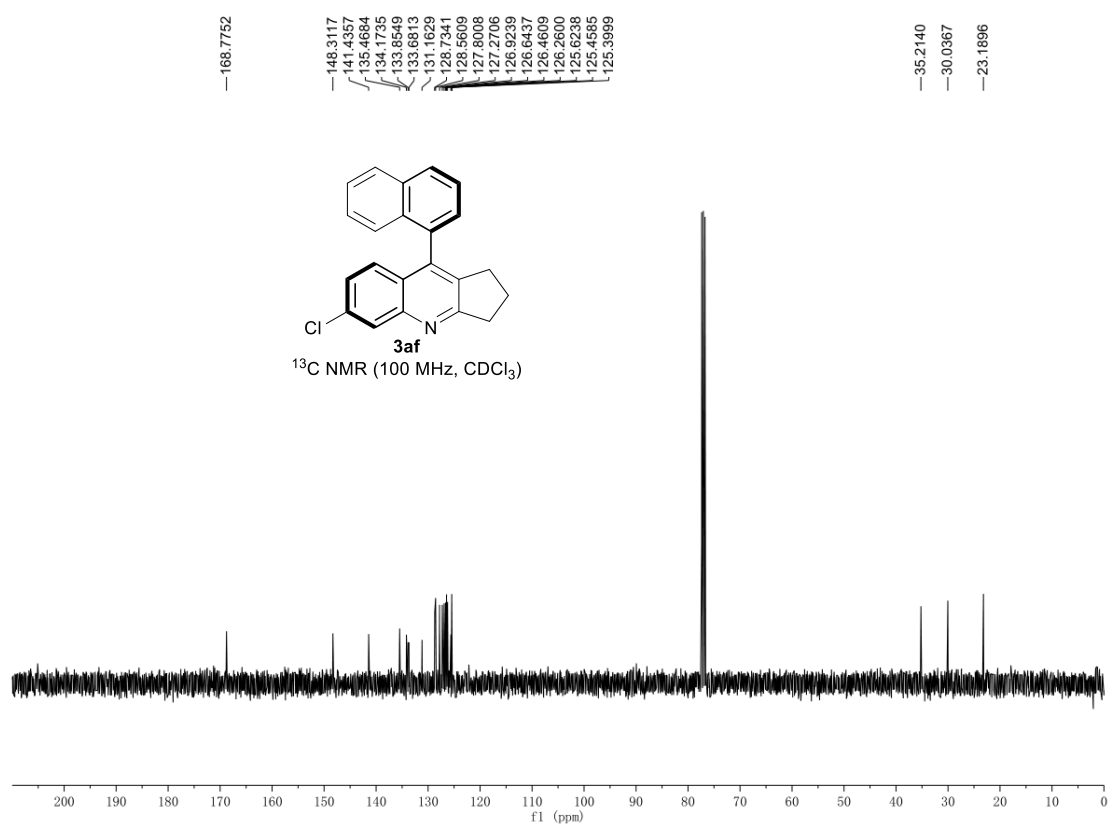
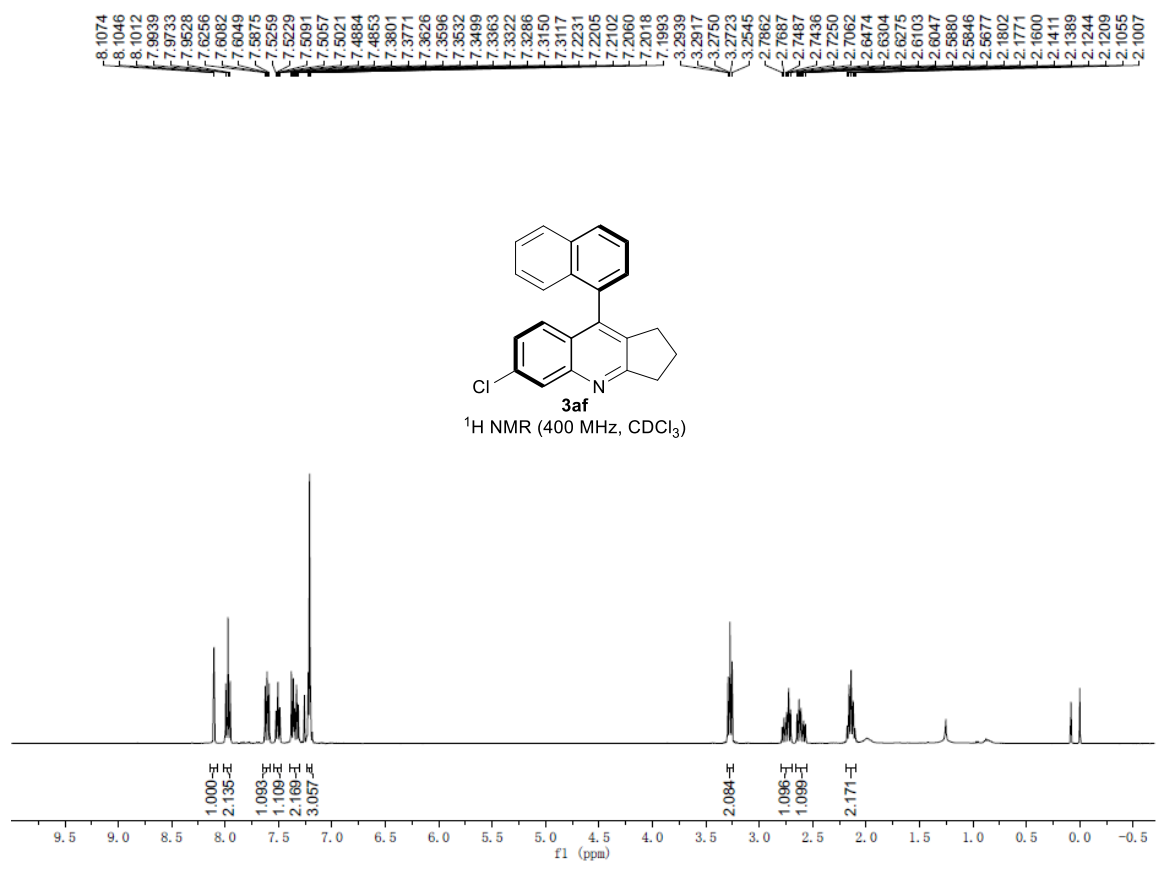
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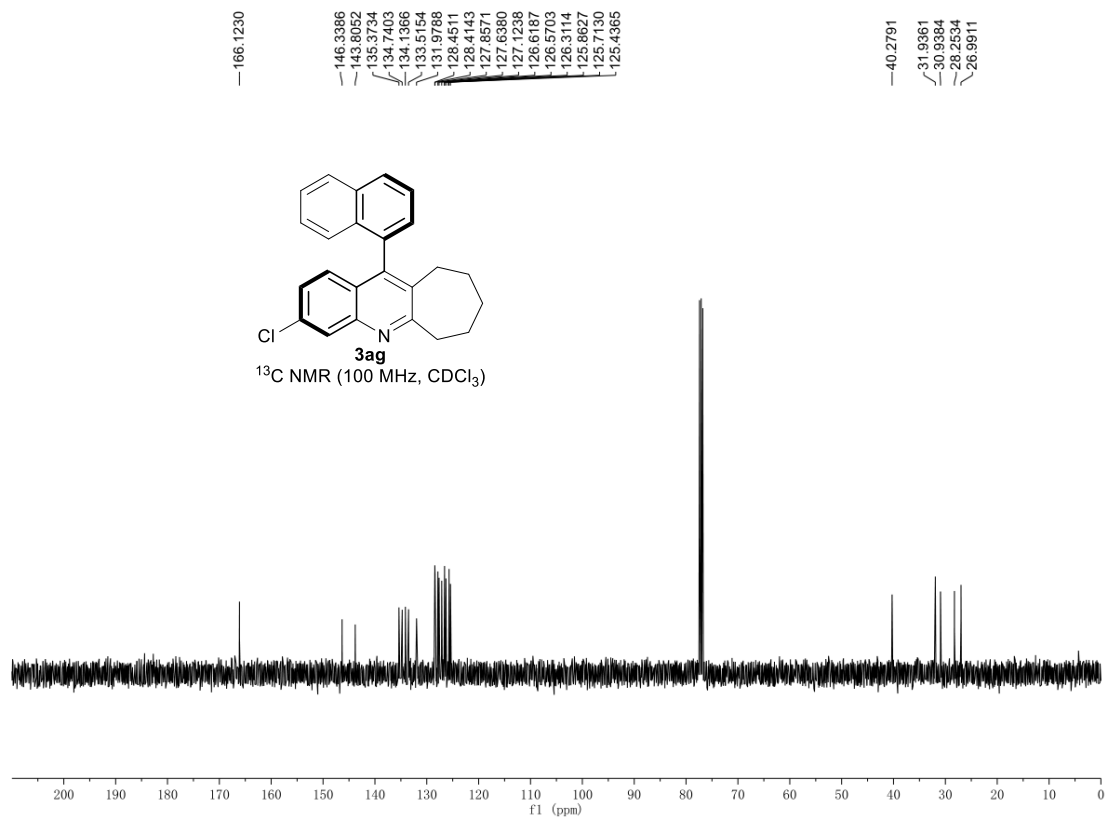
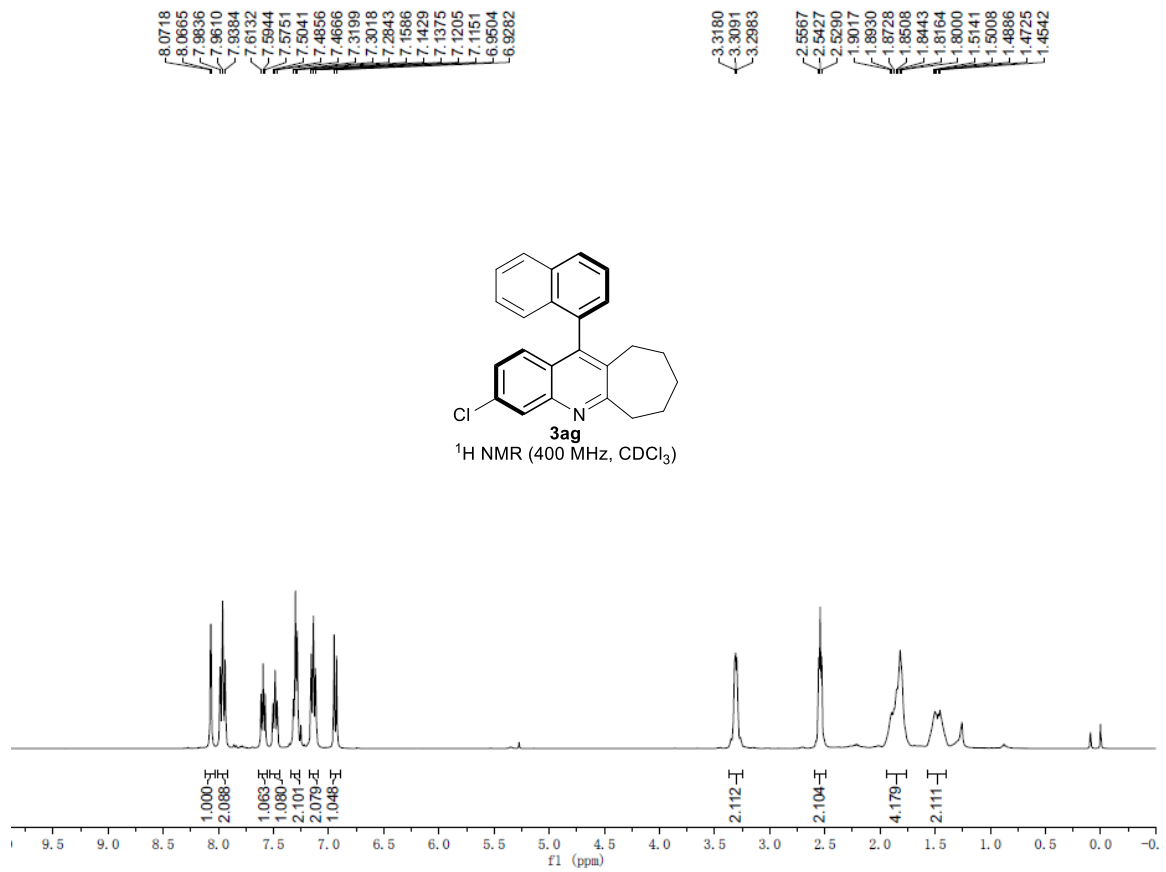
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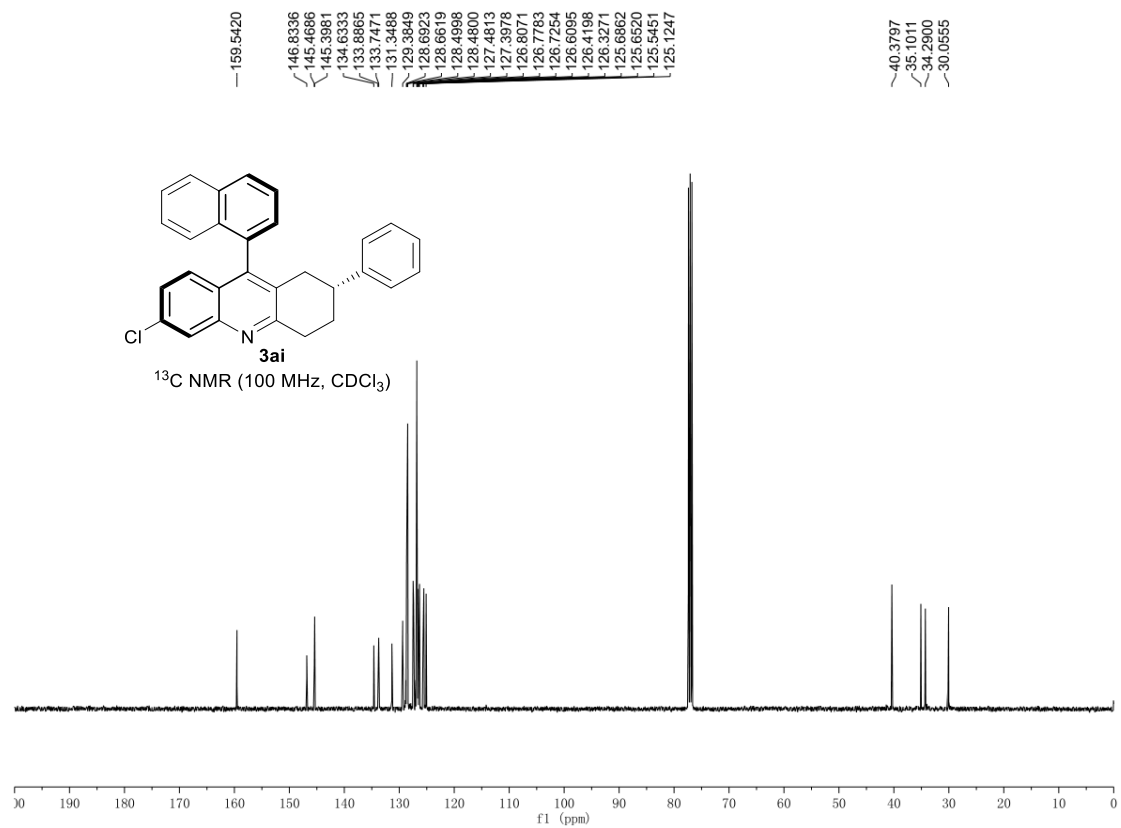
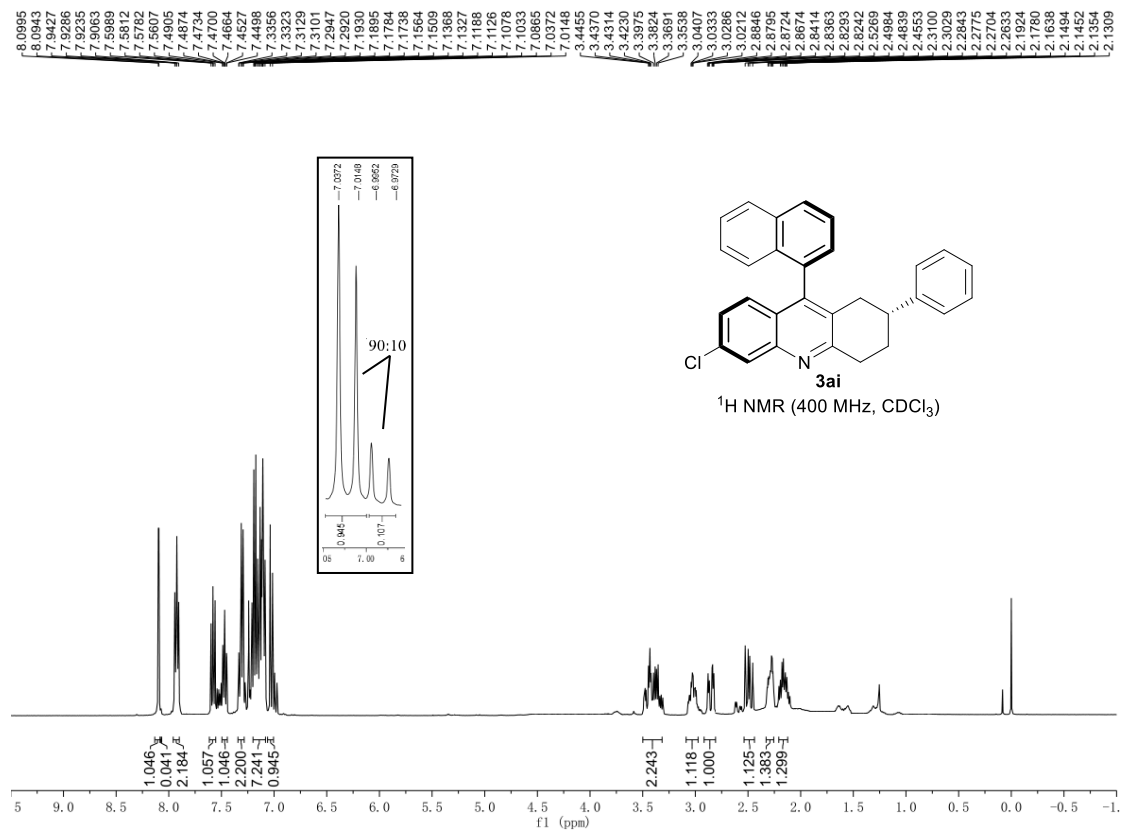


$^{13}\text{C NMR}$ (100 MHz, CDCl_3)

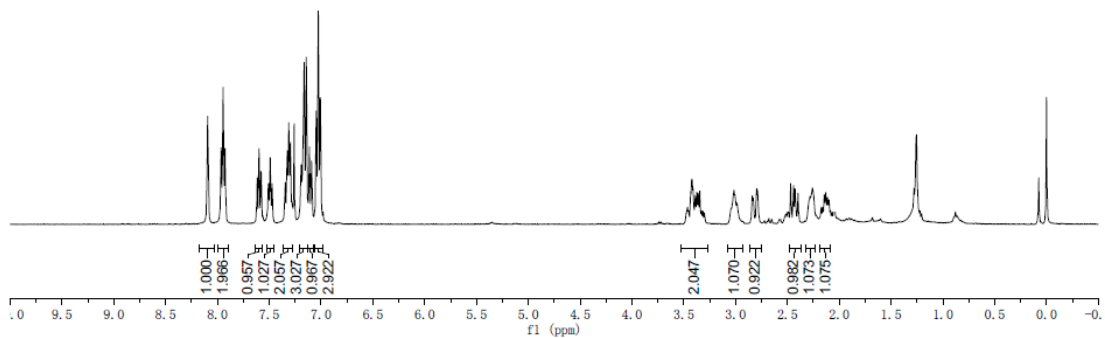
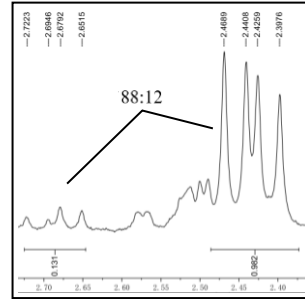
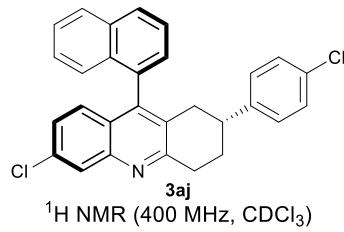




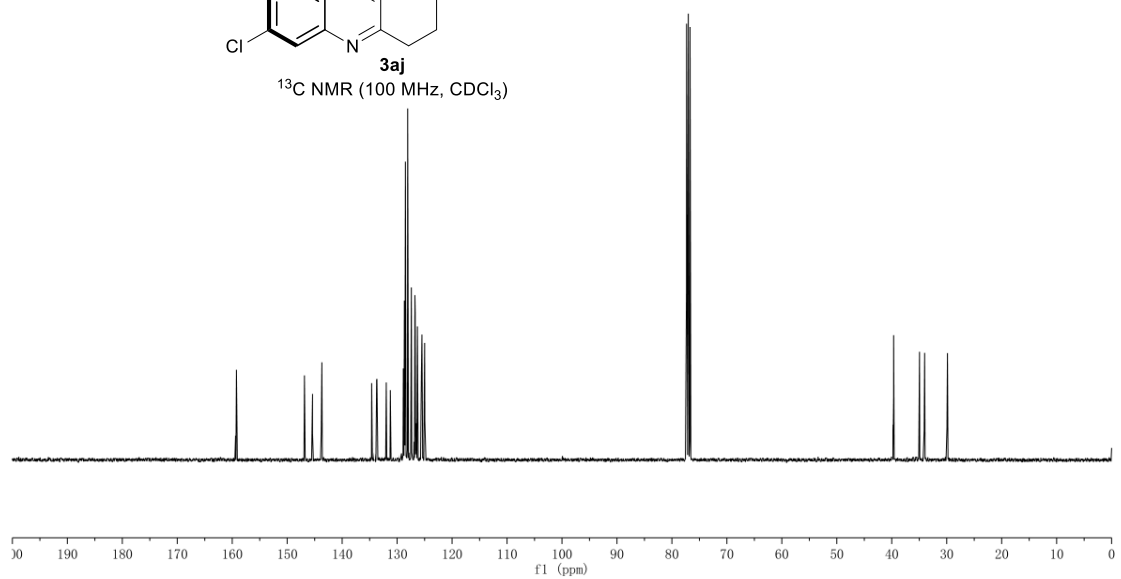
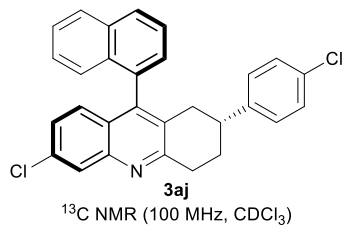


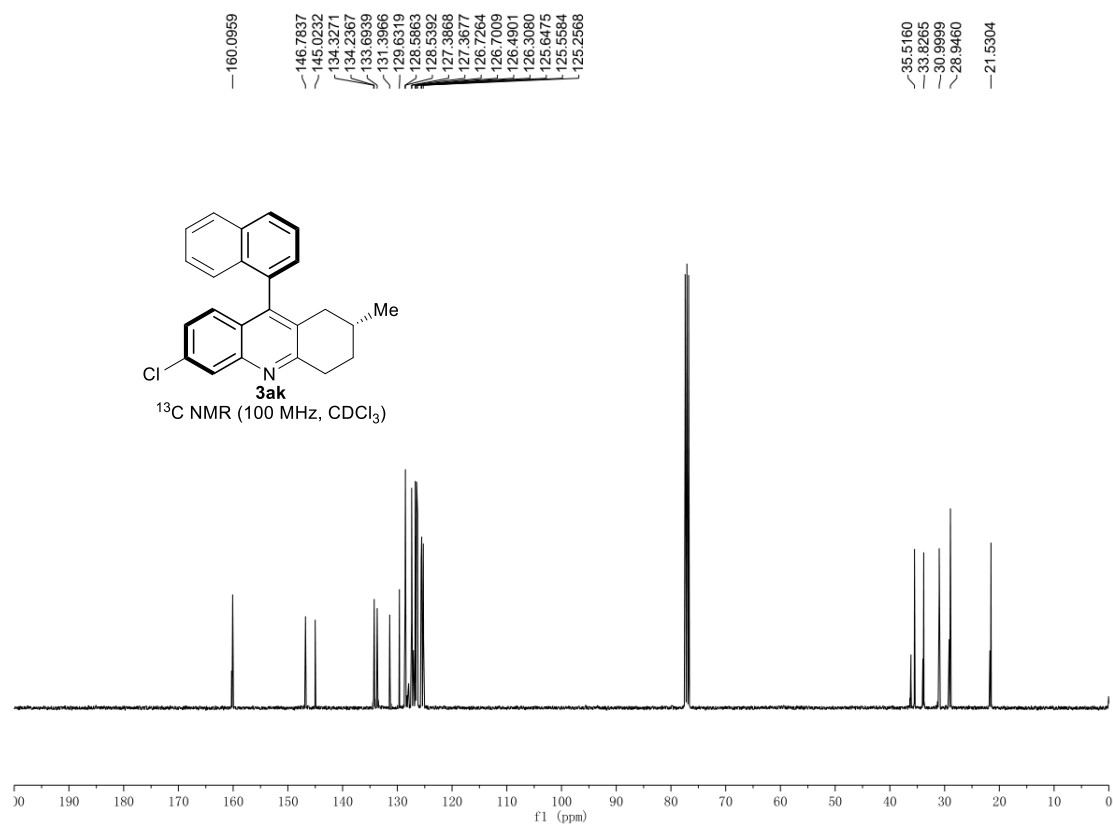
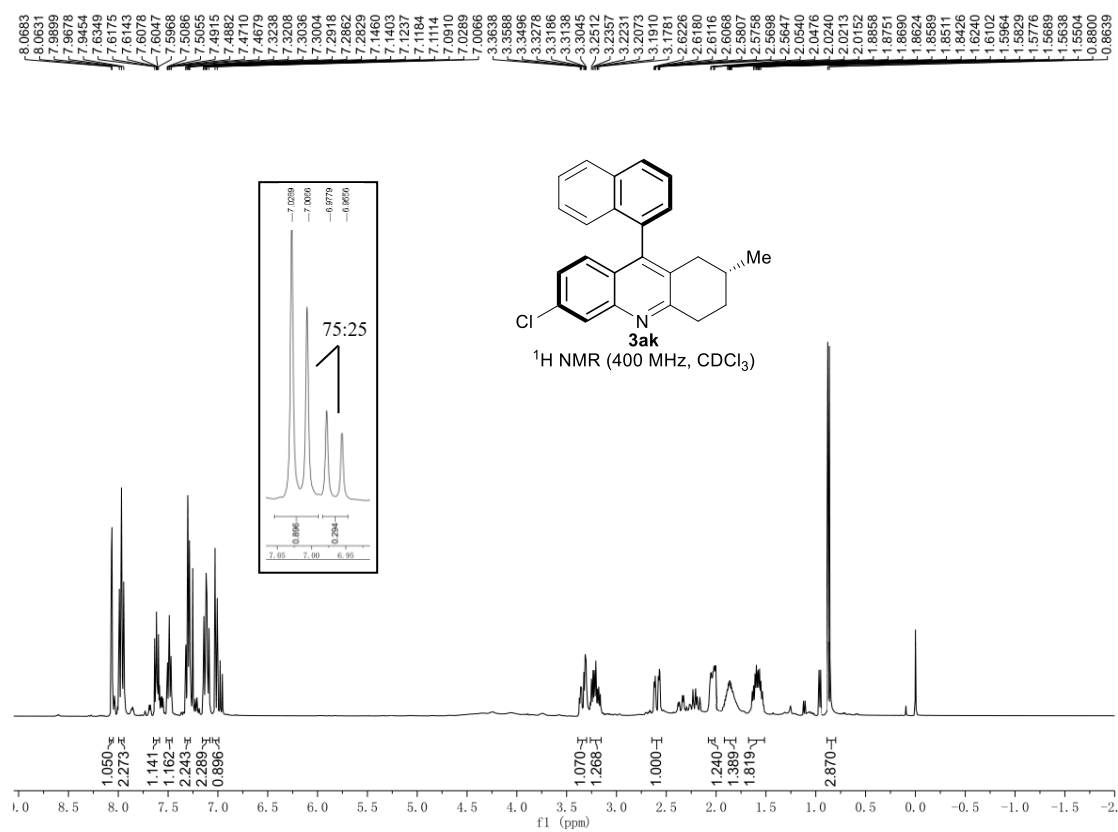


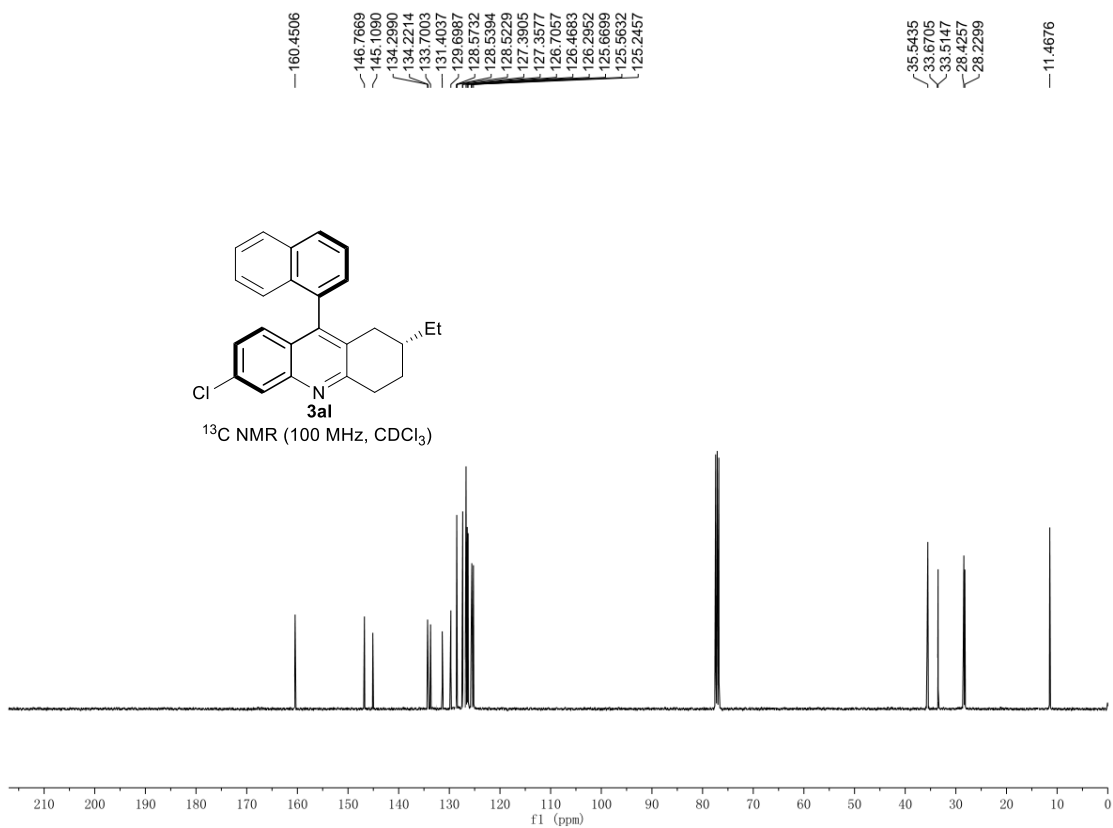
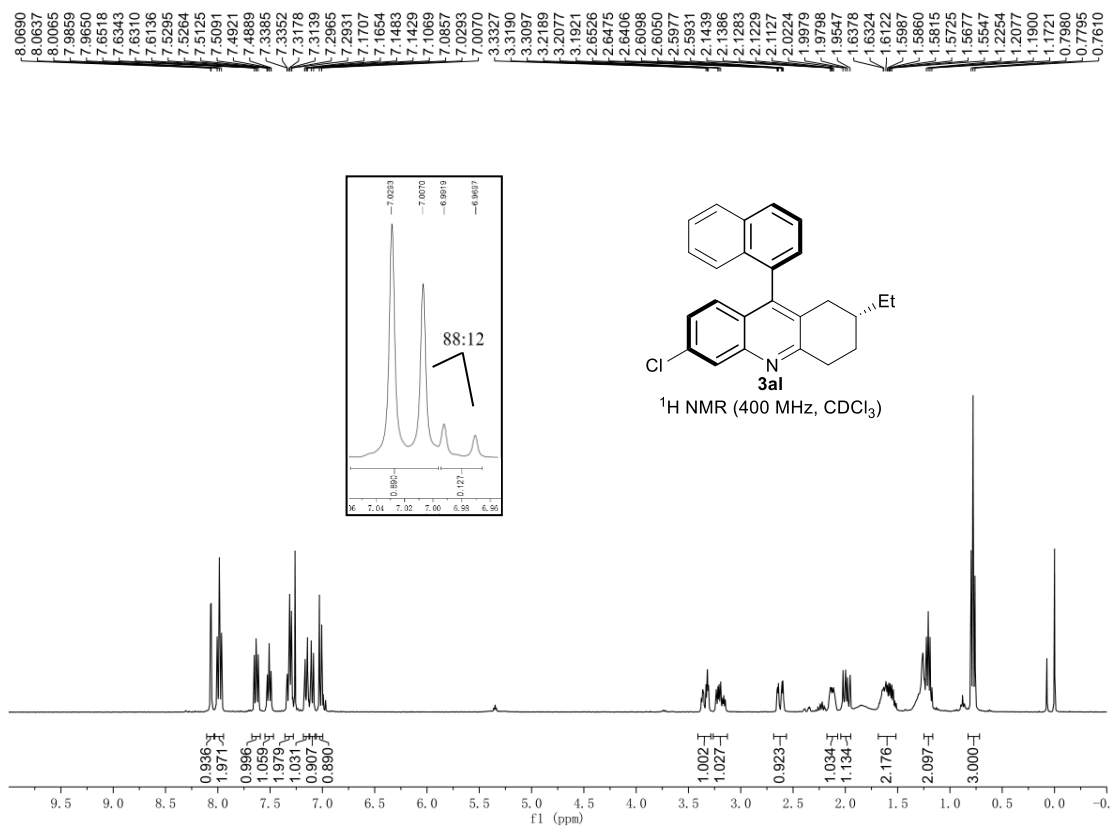
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 -2.0879



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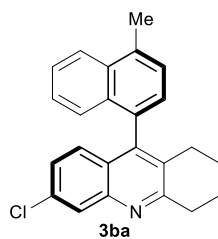




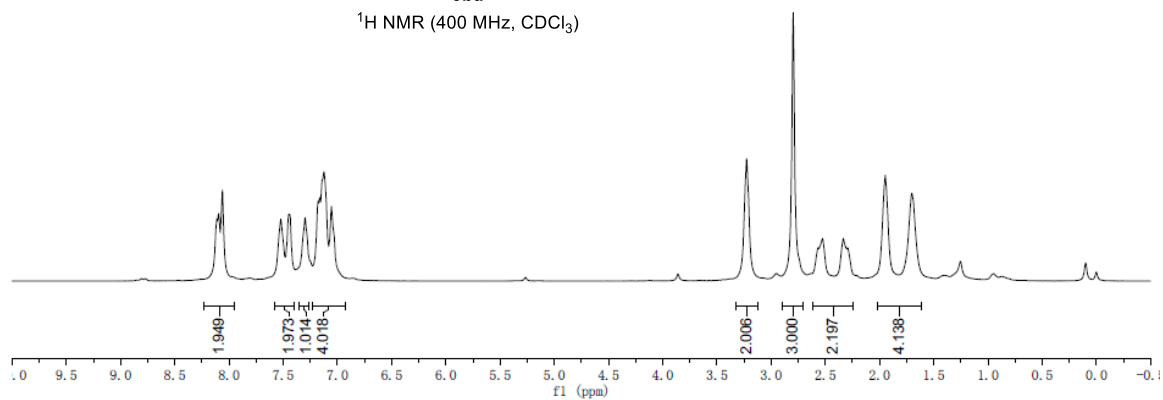


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7.1421
7.1235
7.1061
7.0546
7.0317

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1.6892

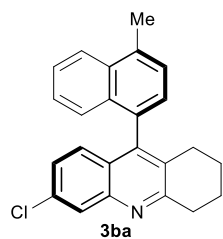


¹H NMR (400 MHz, CDCl₃)

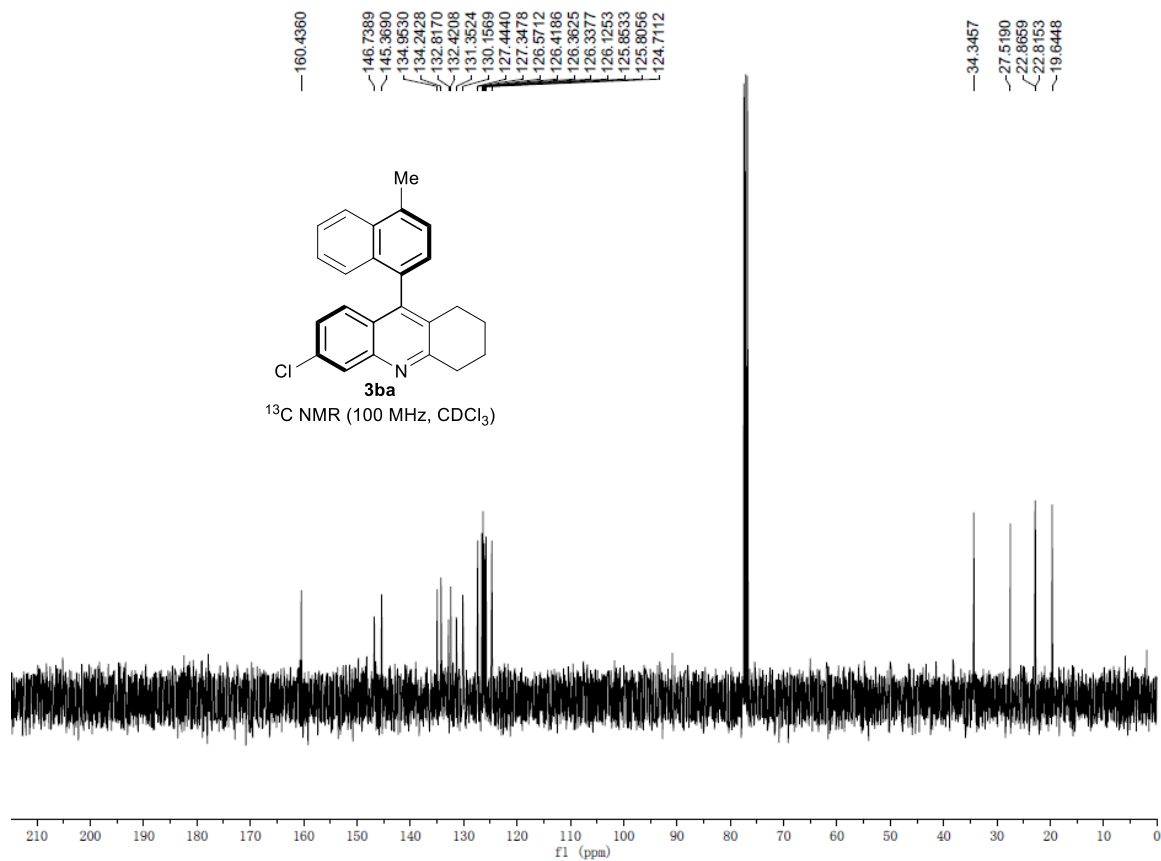


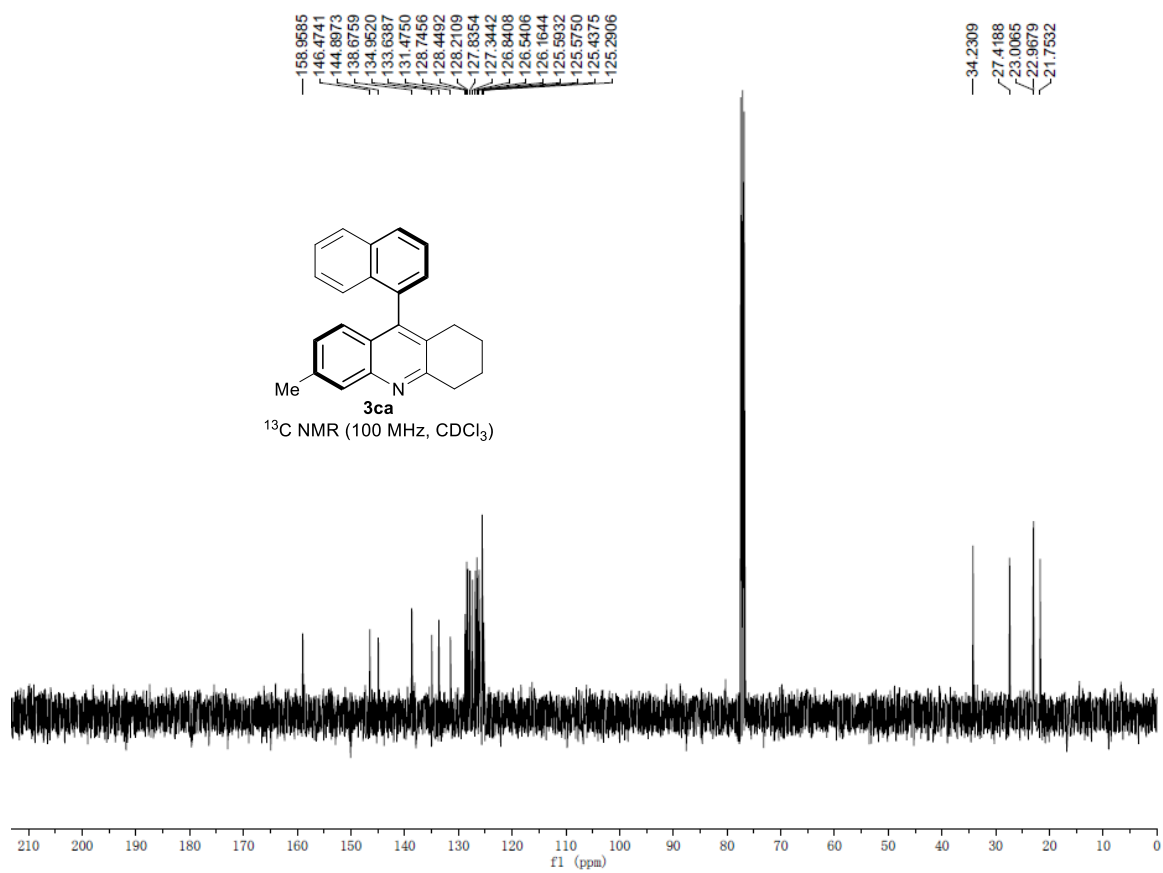
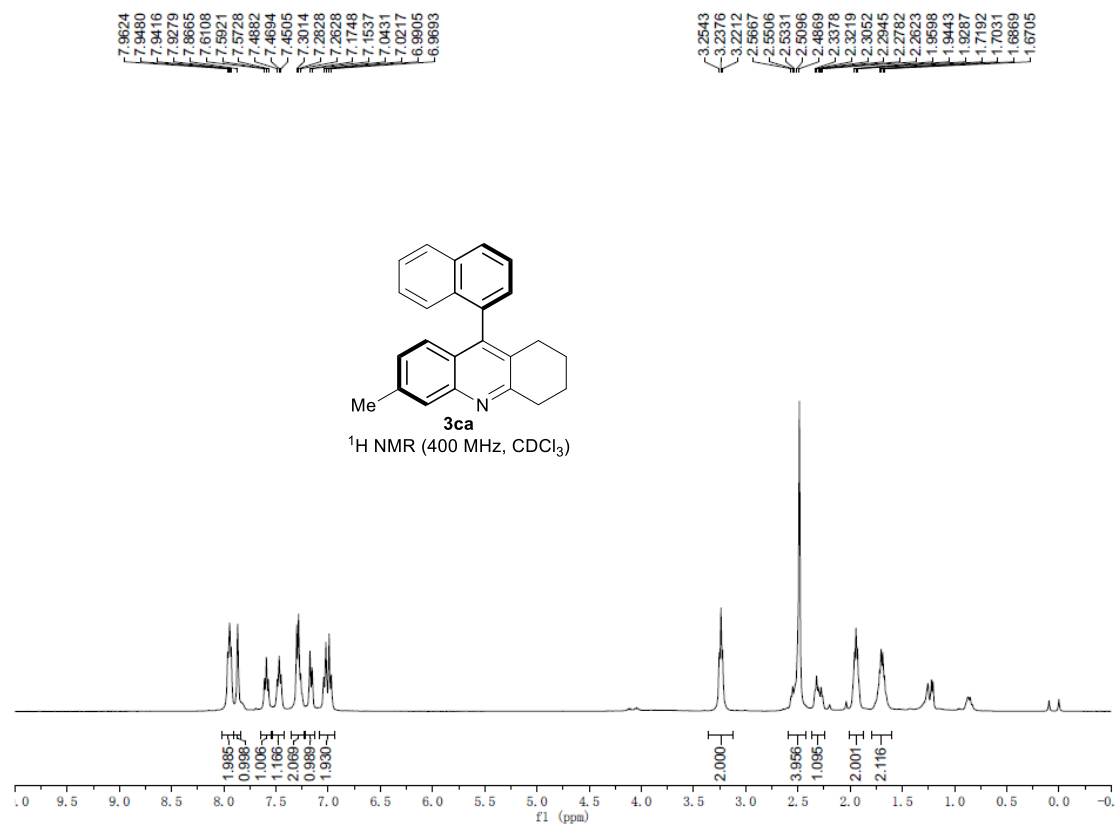
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125.8056
124.7112

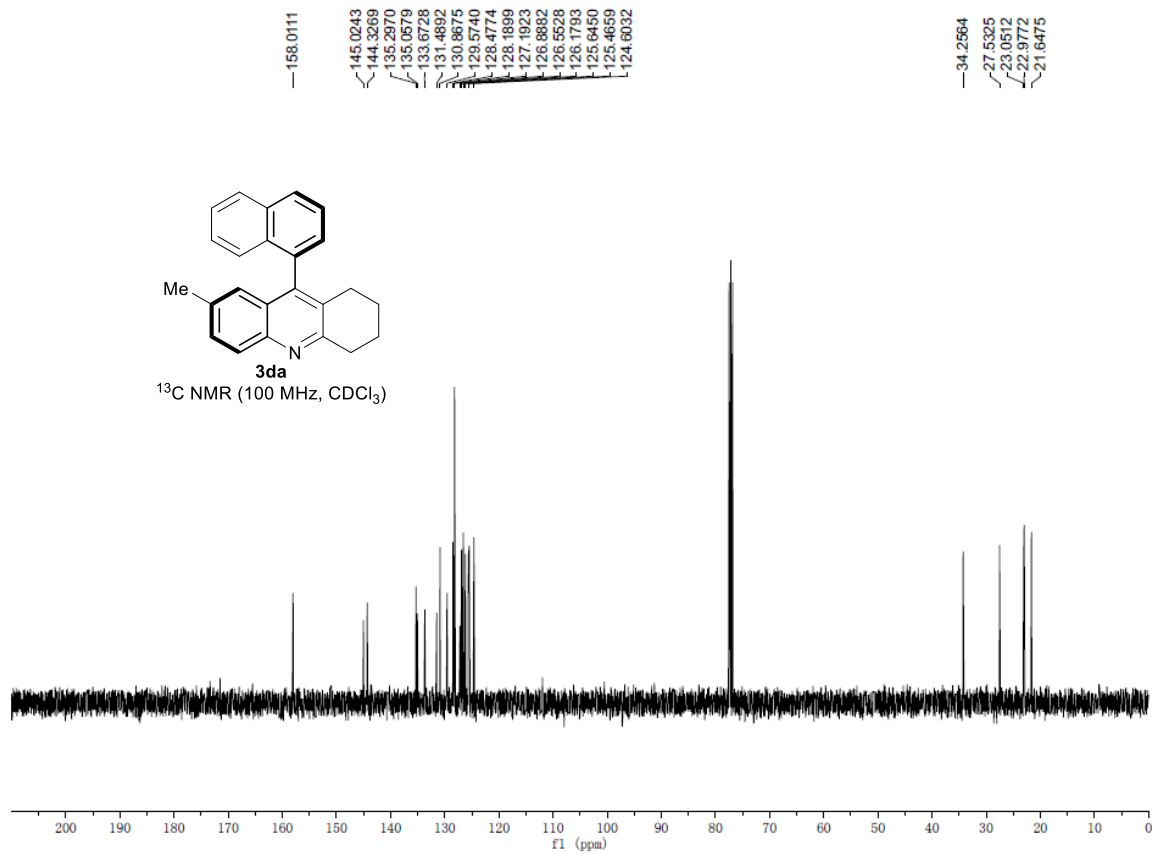
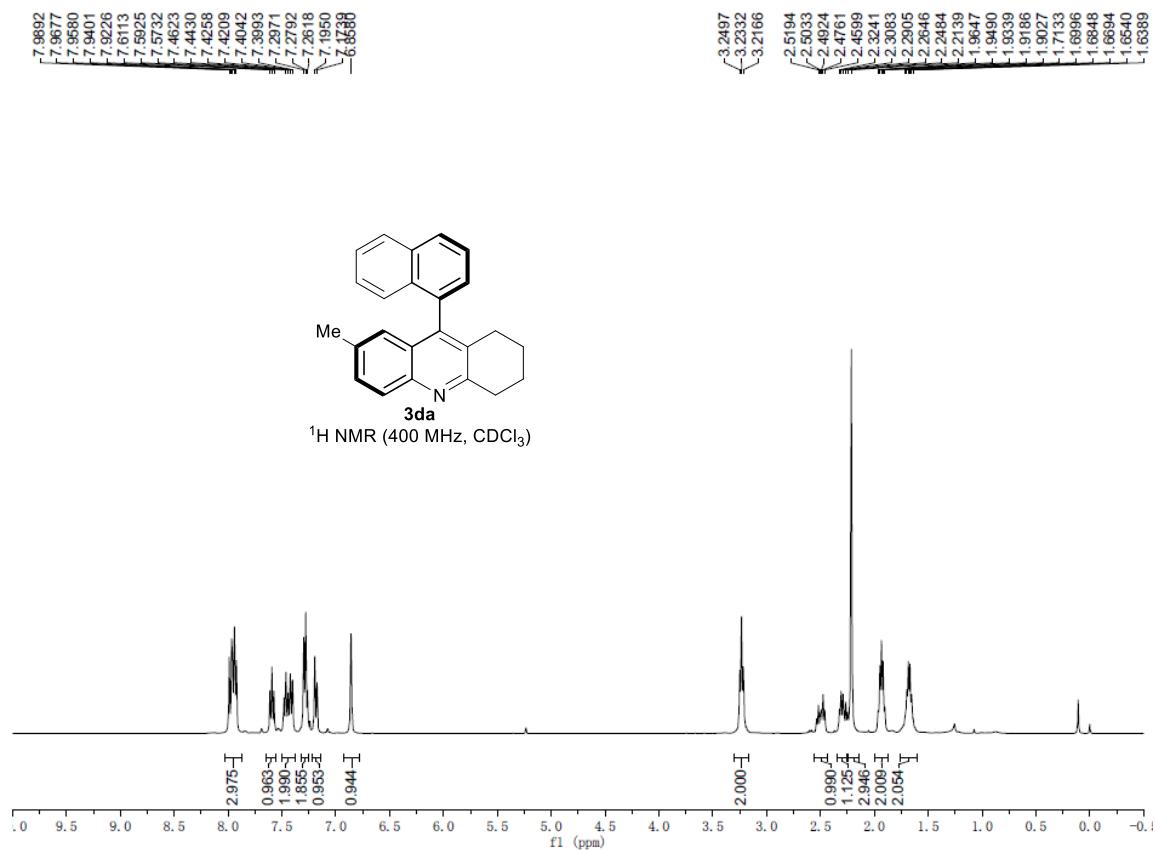
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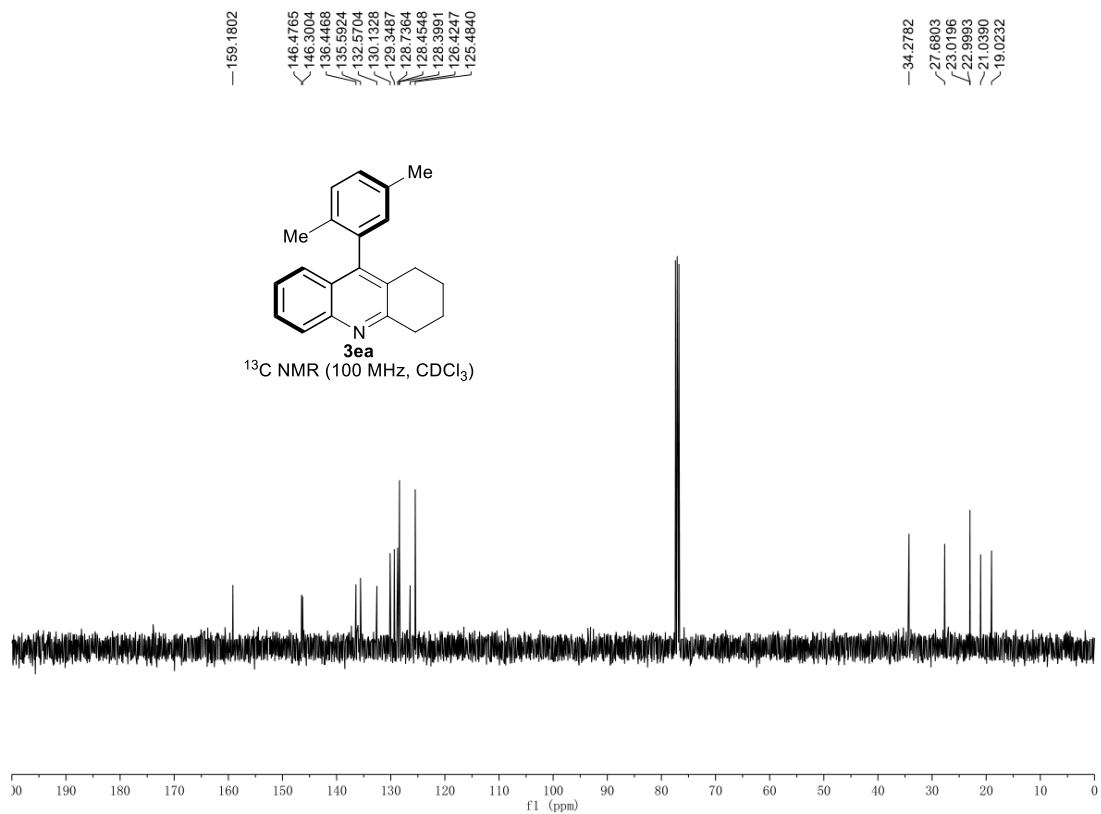
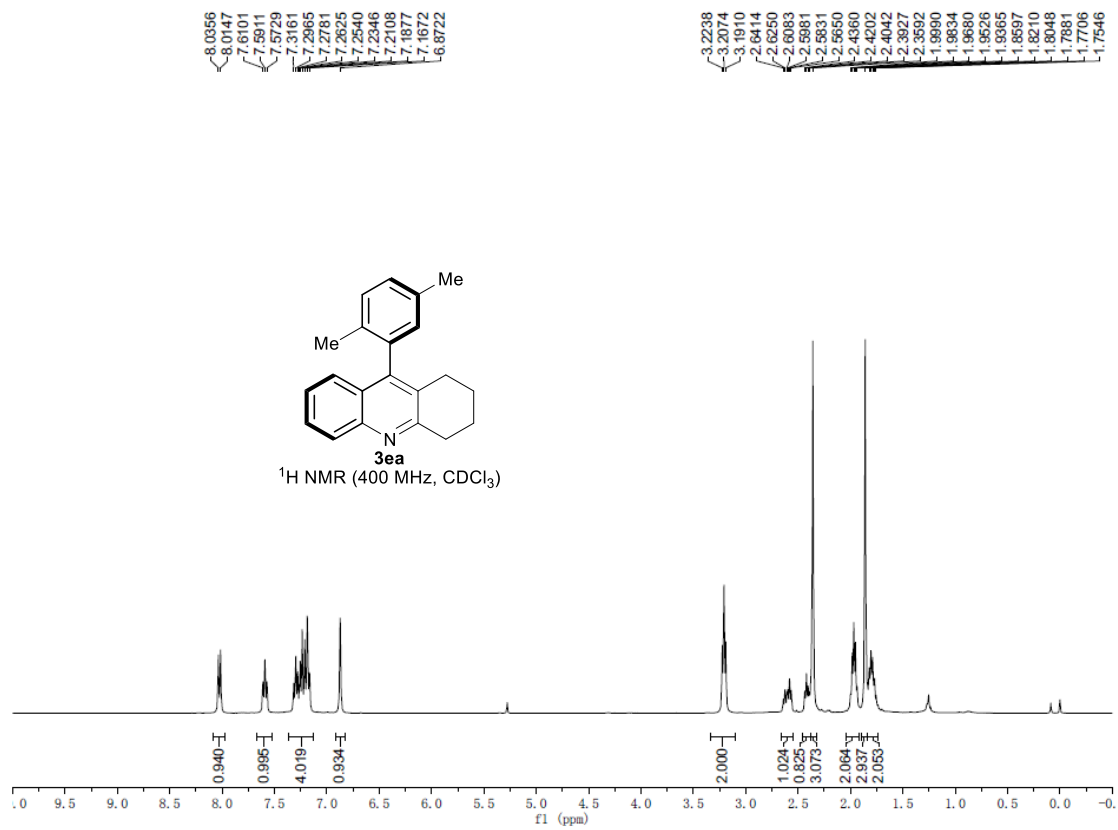


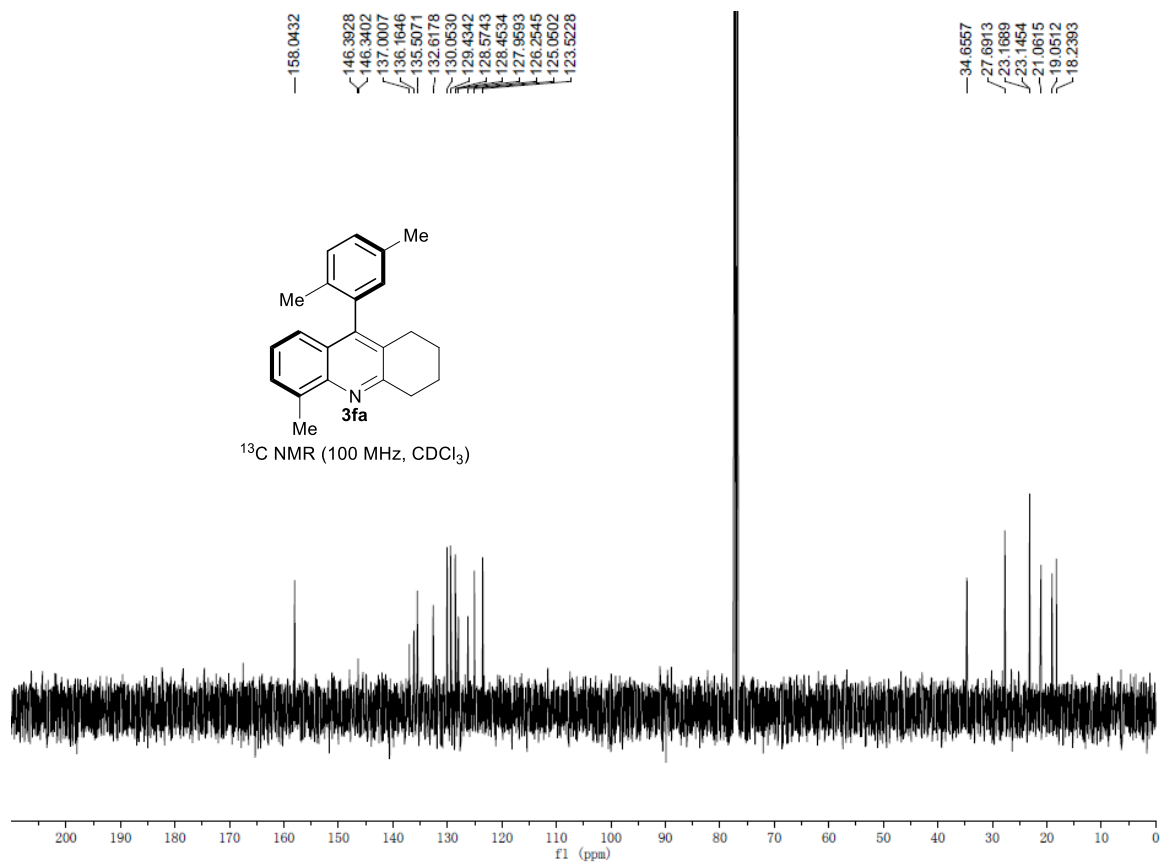
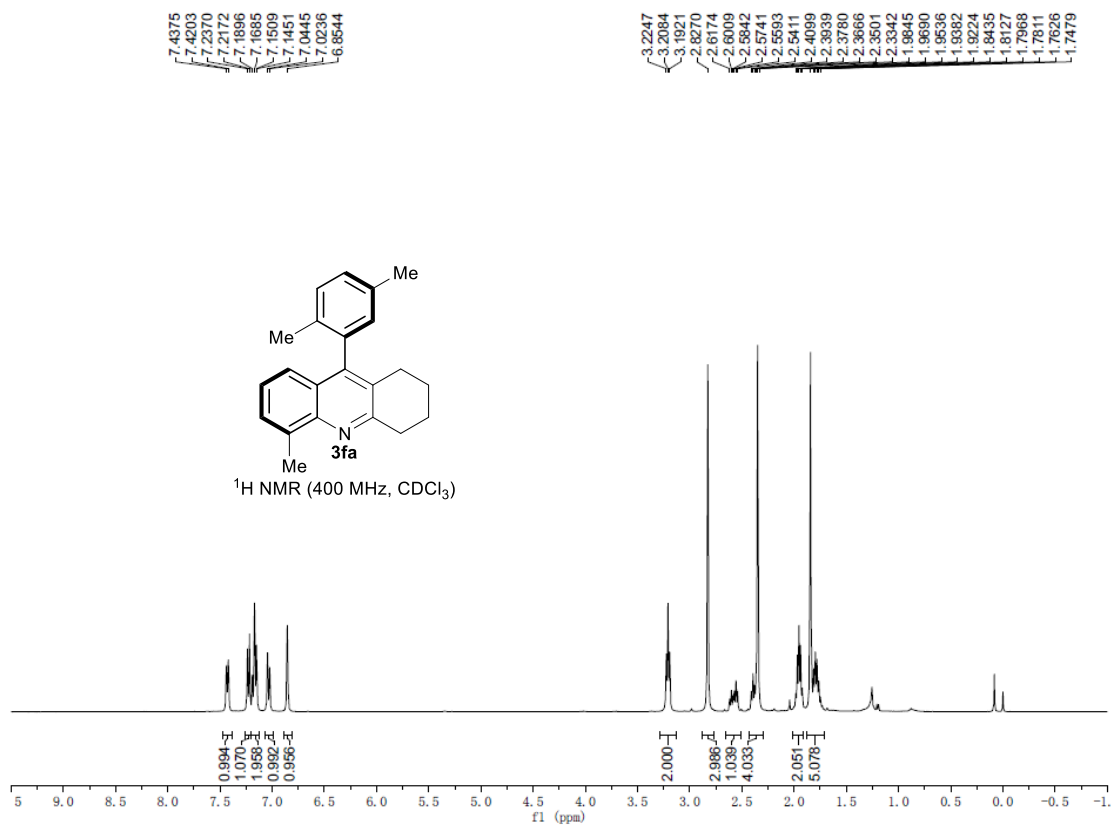
¹³C NMR (100 MHz, CDCl₃)

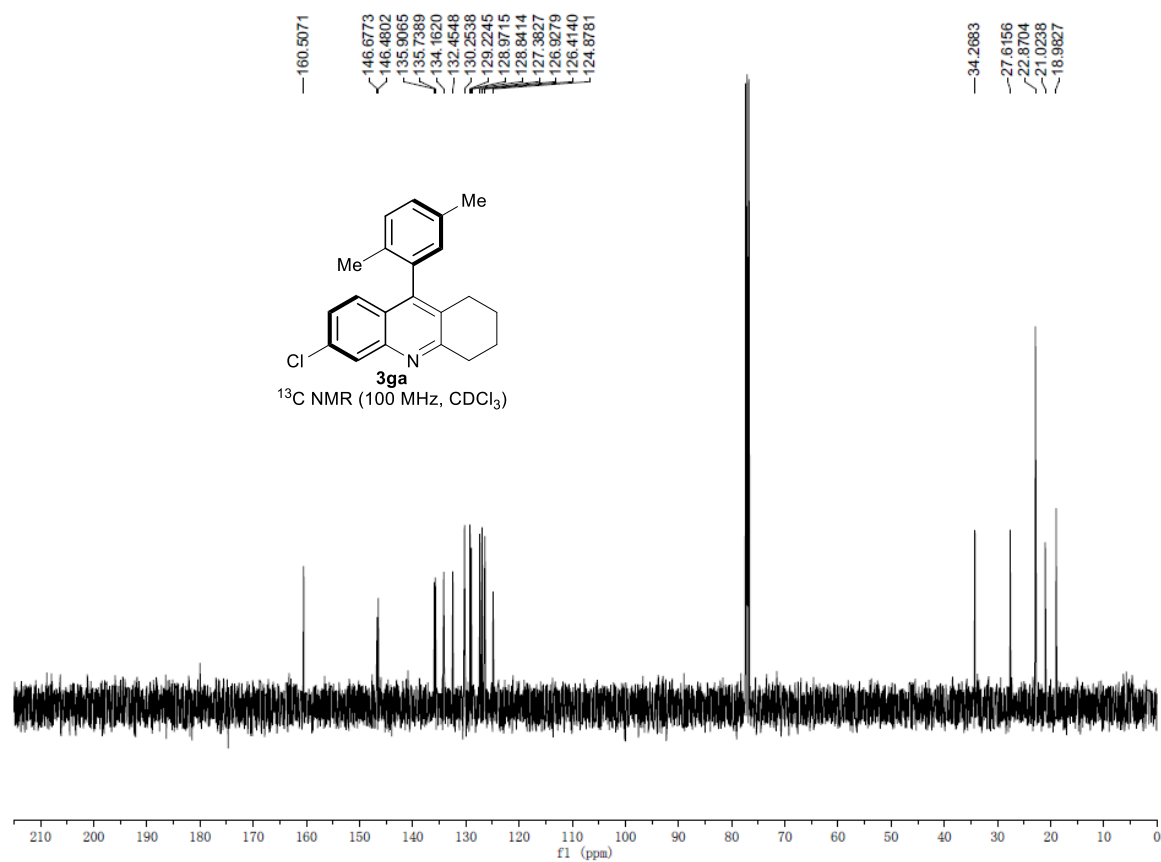
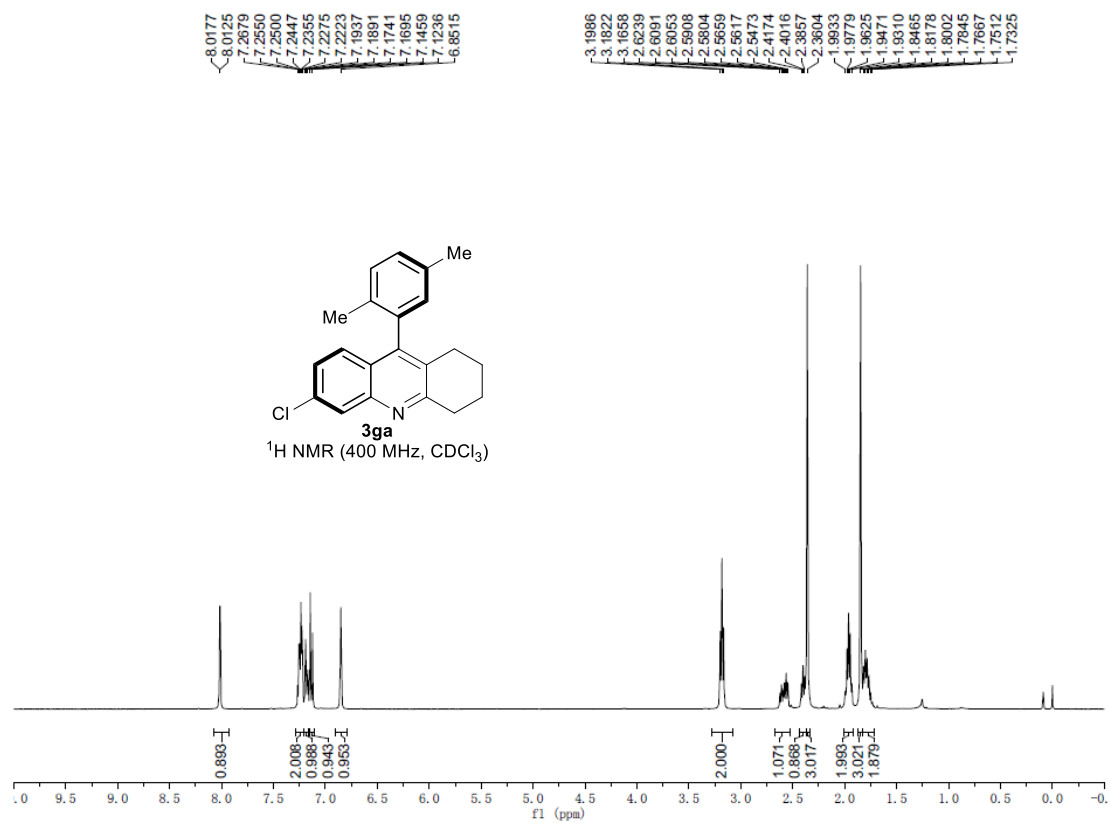


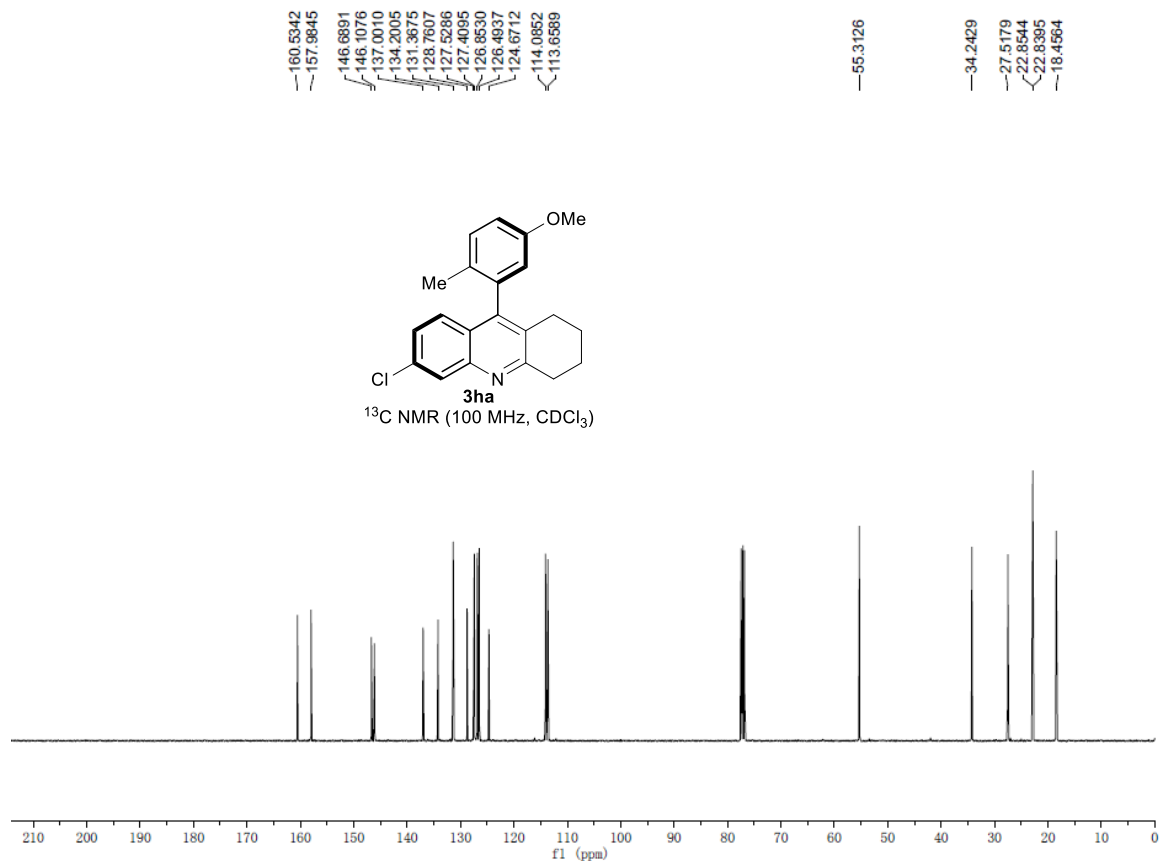
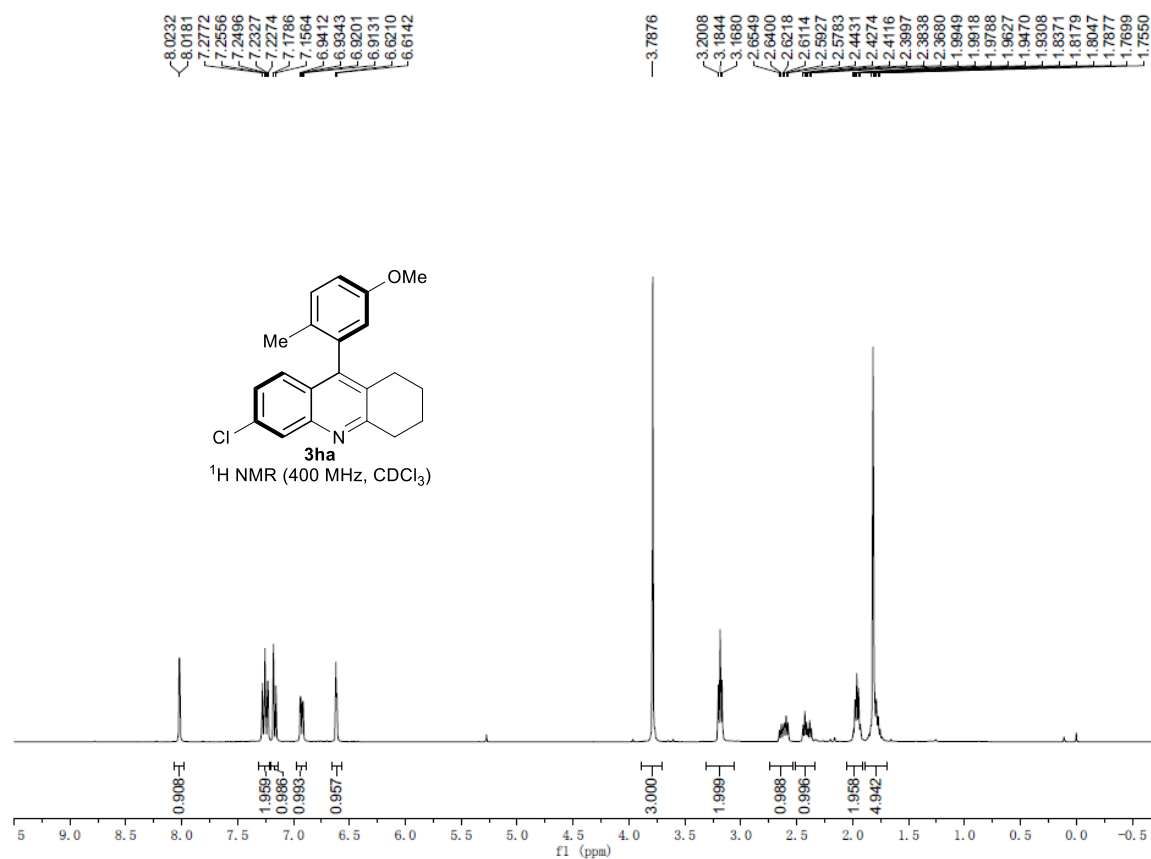


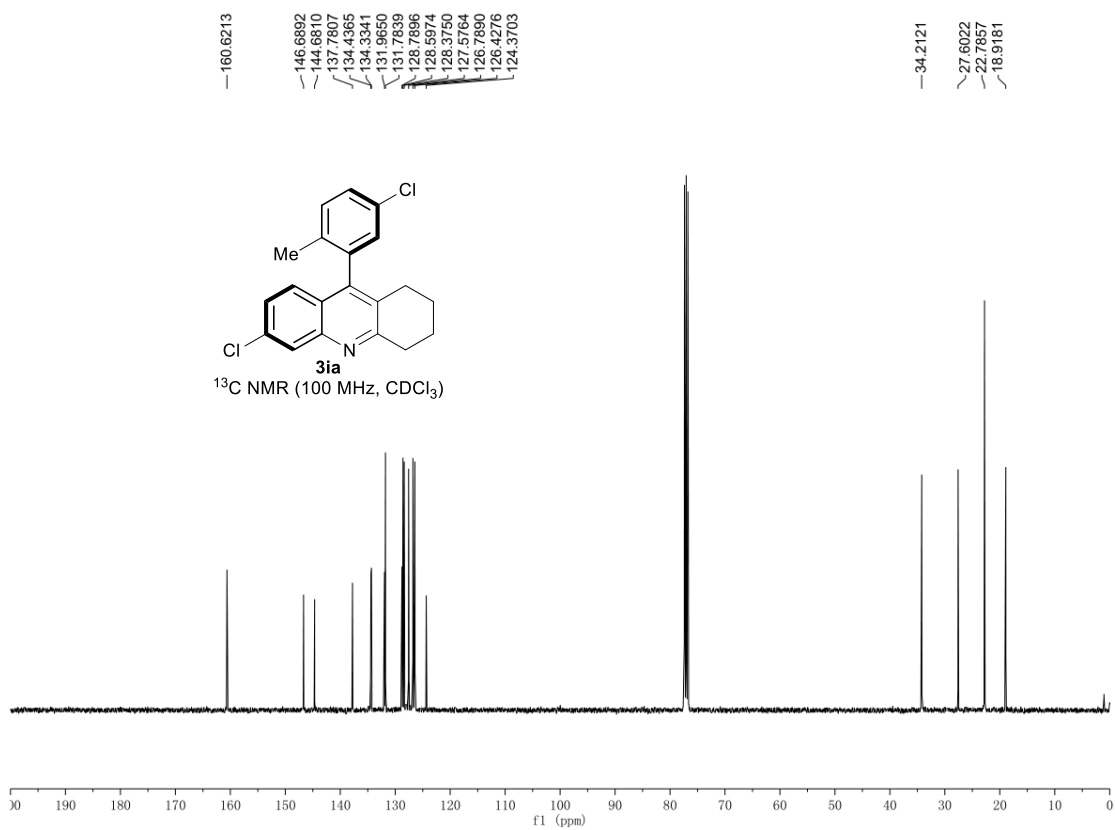
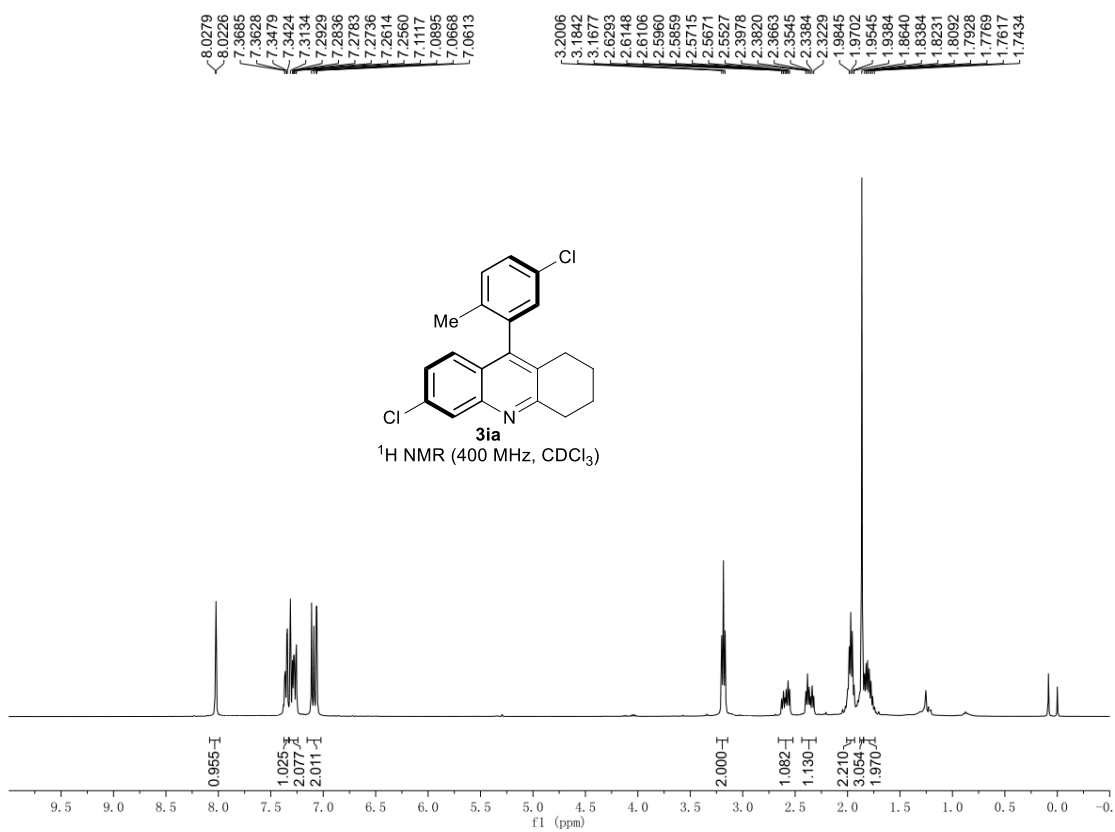


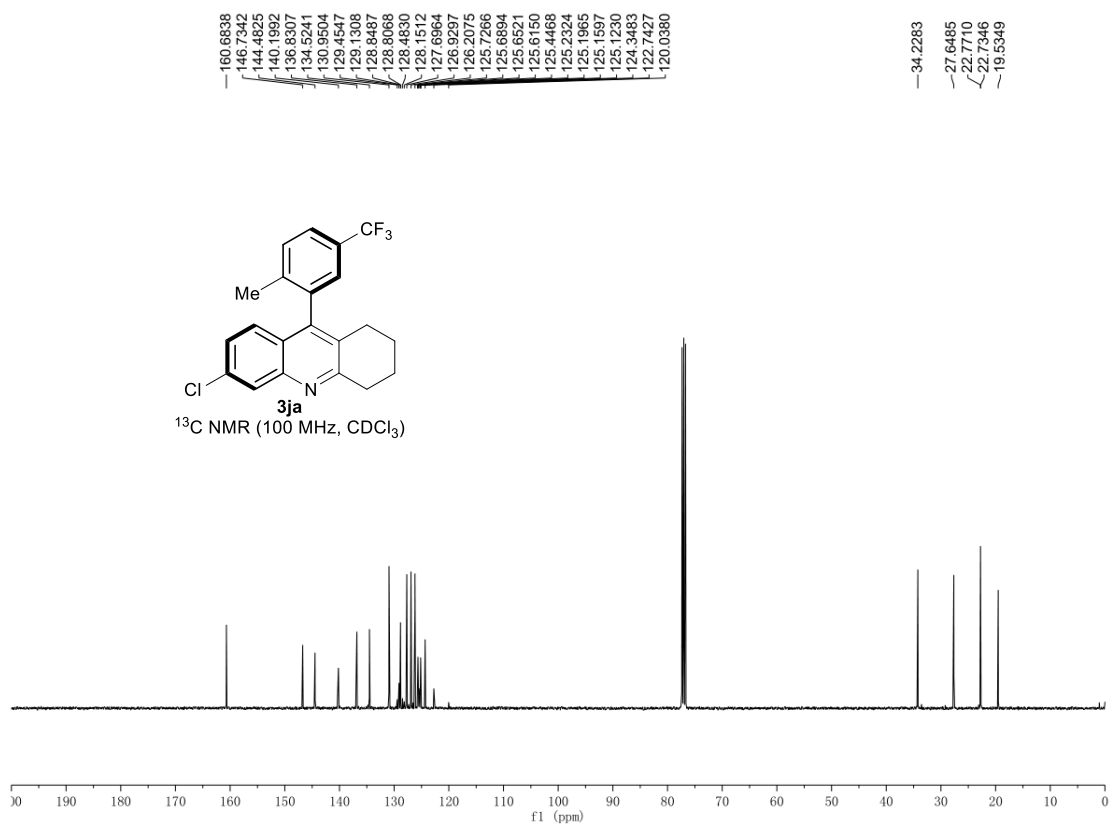
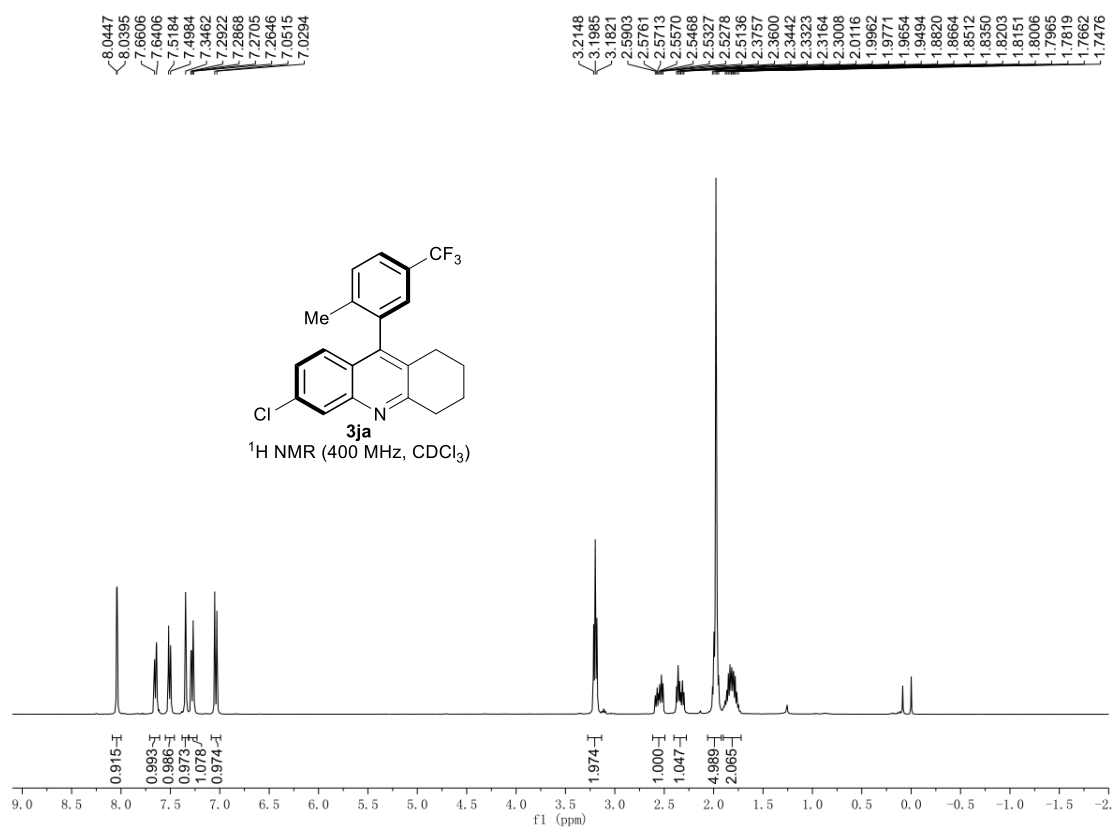


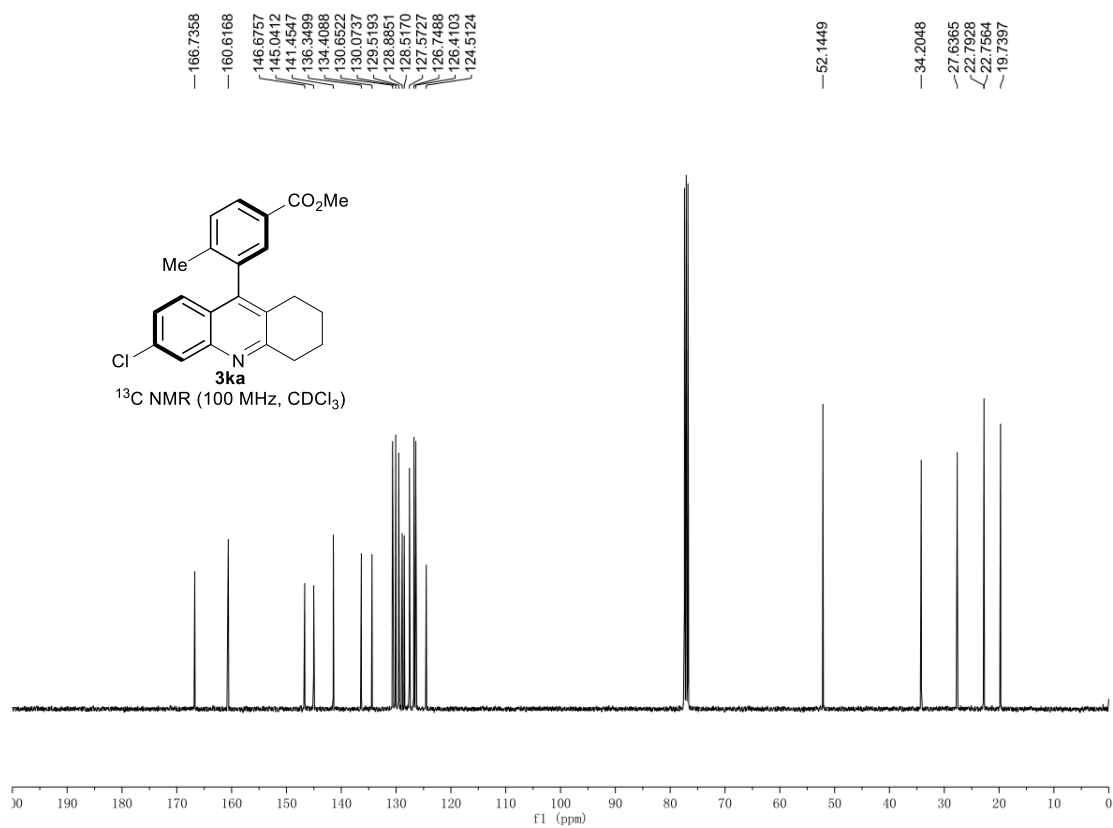
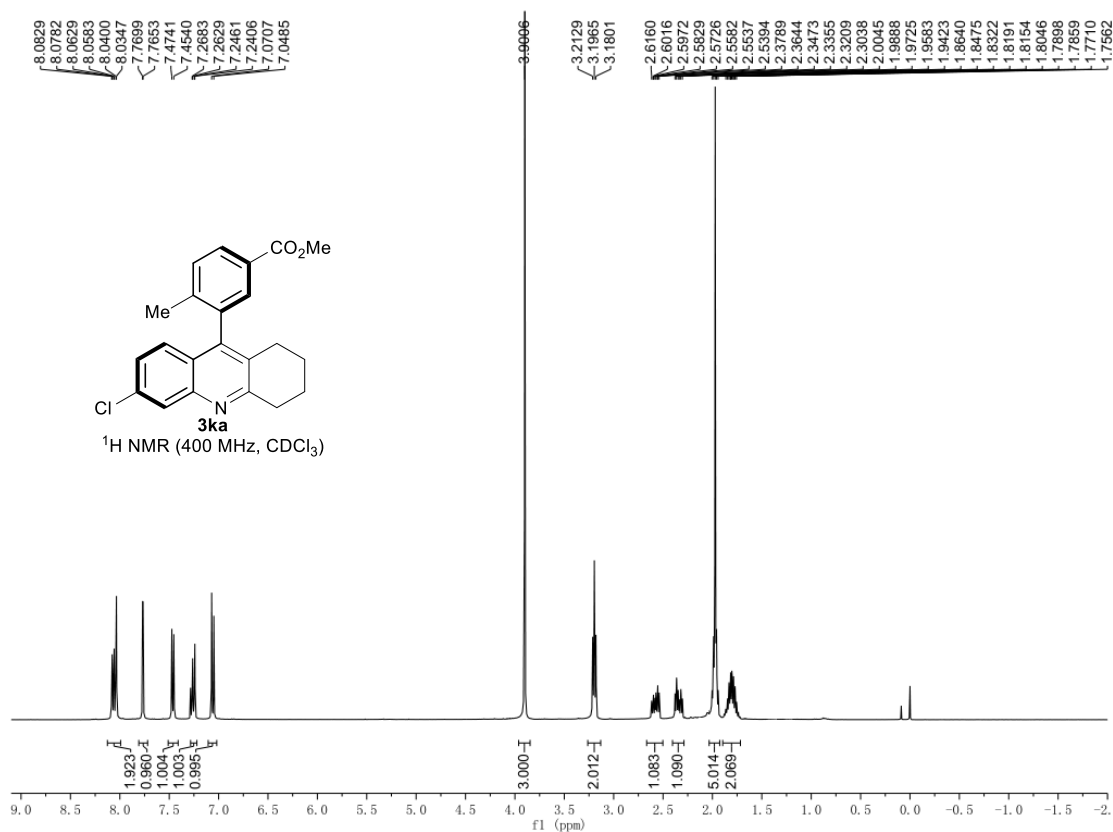


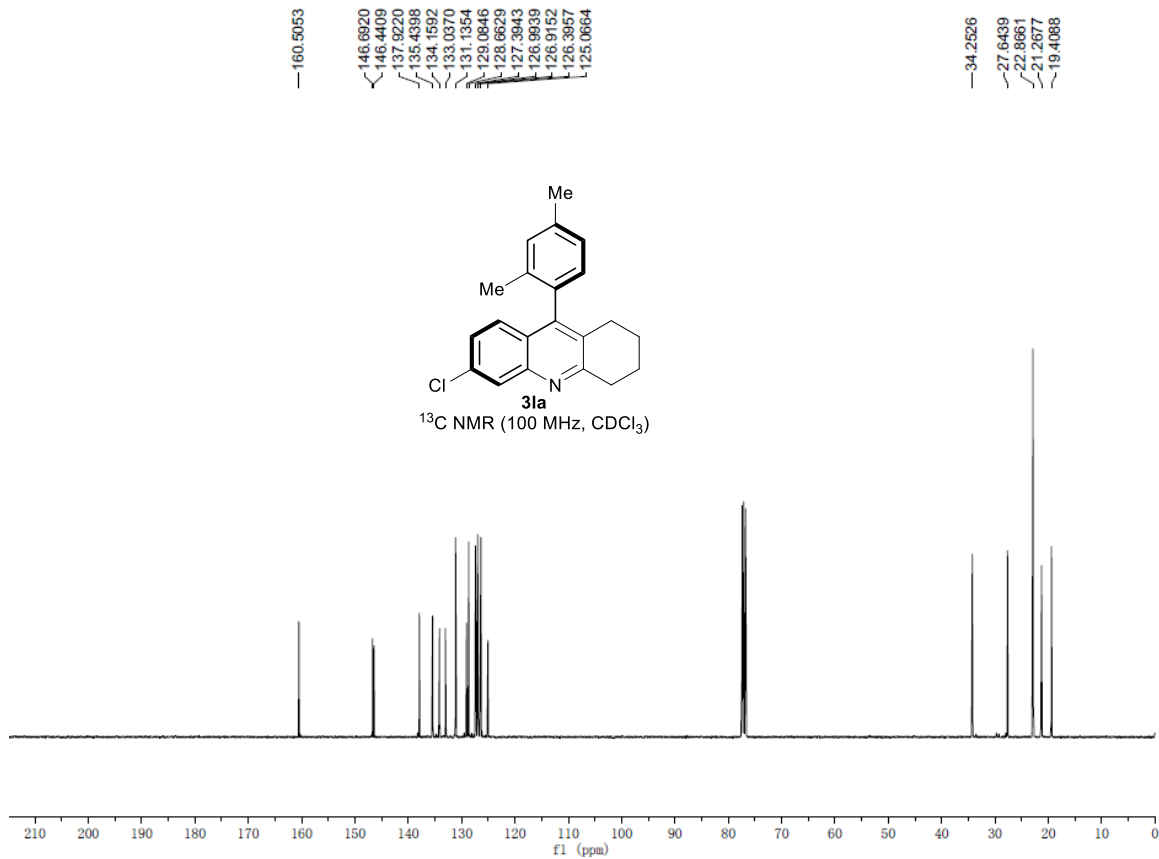
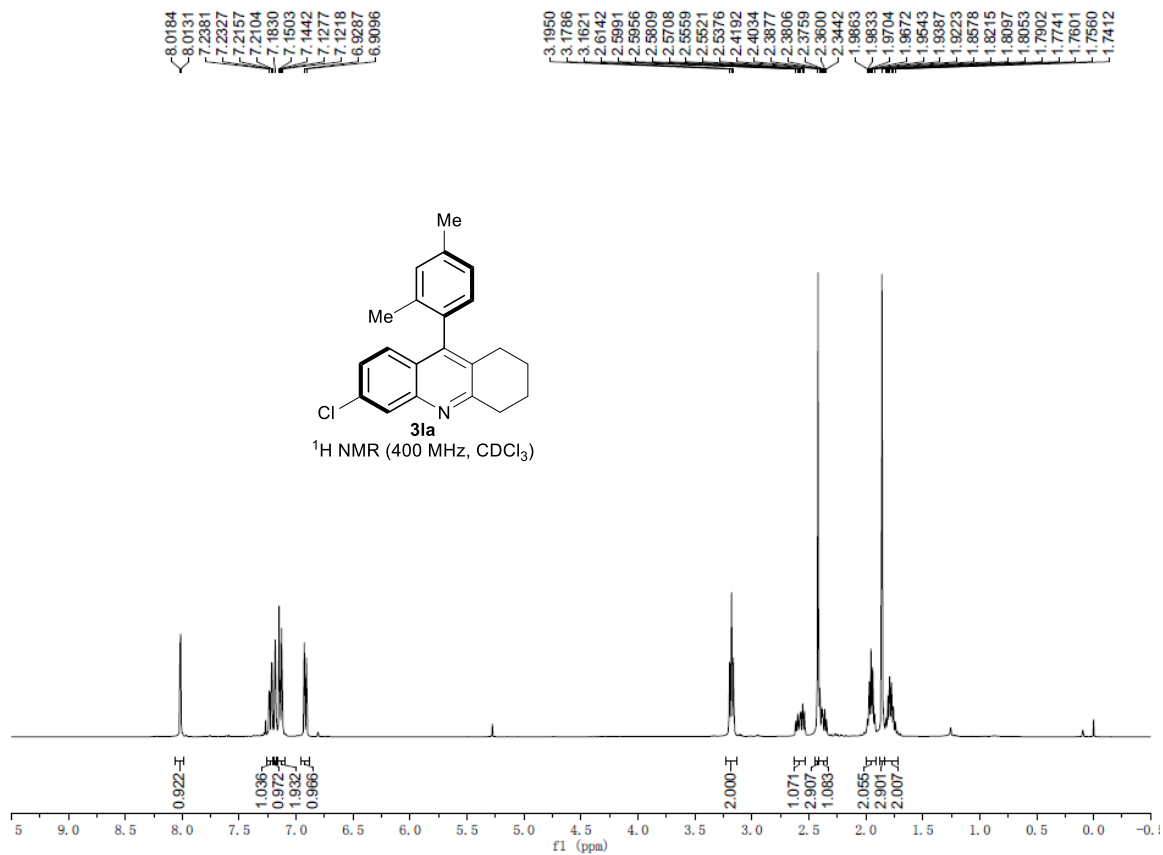


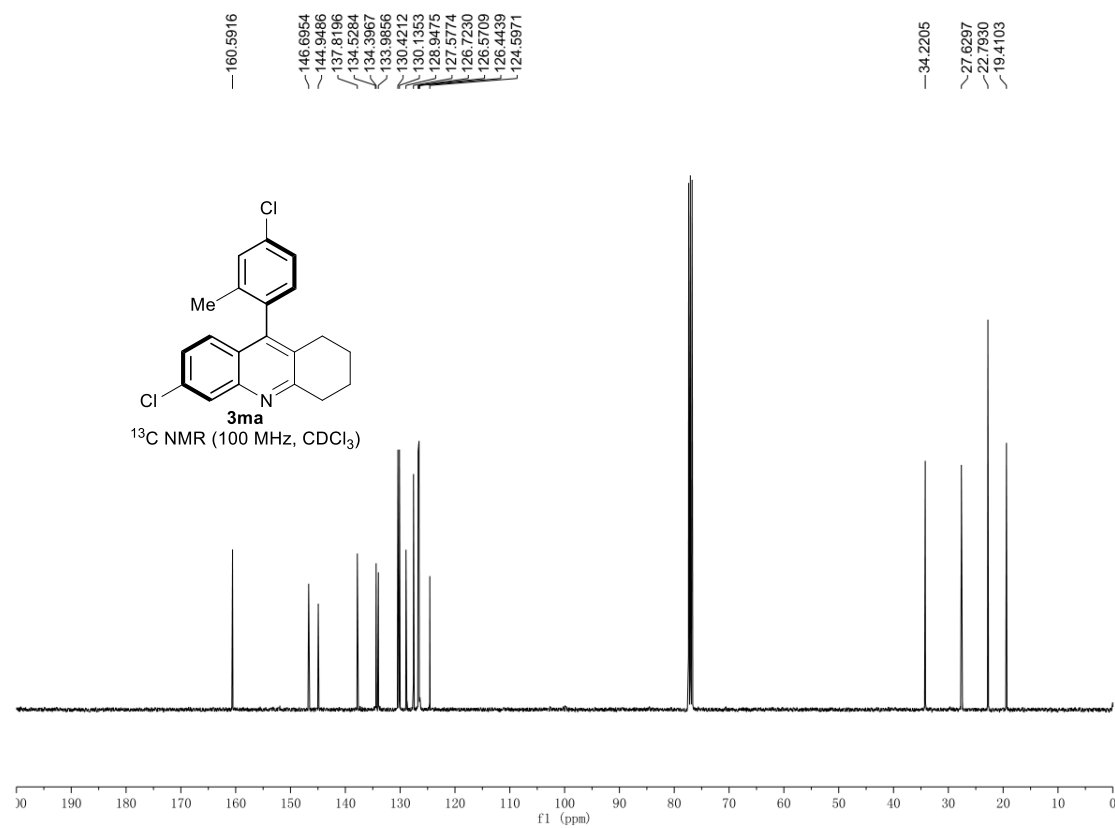
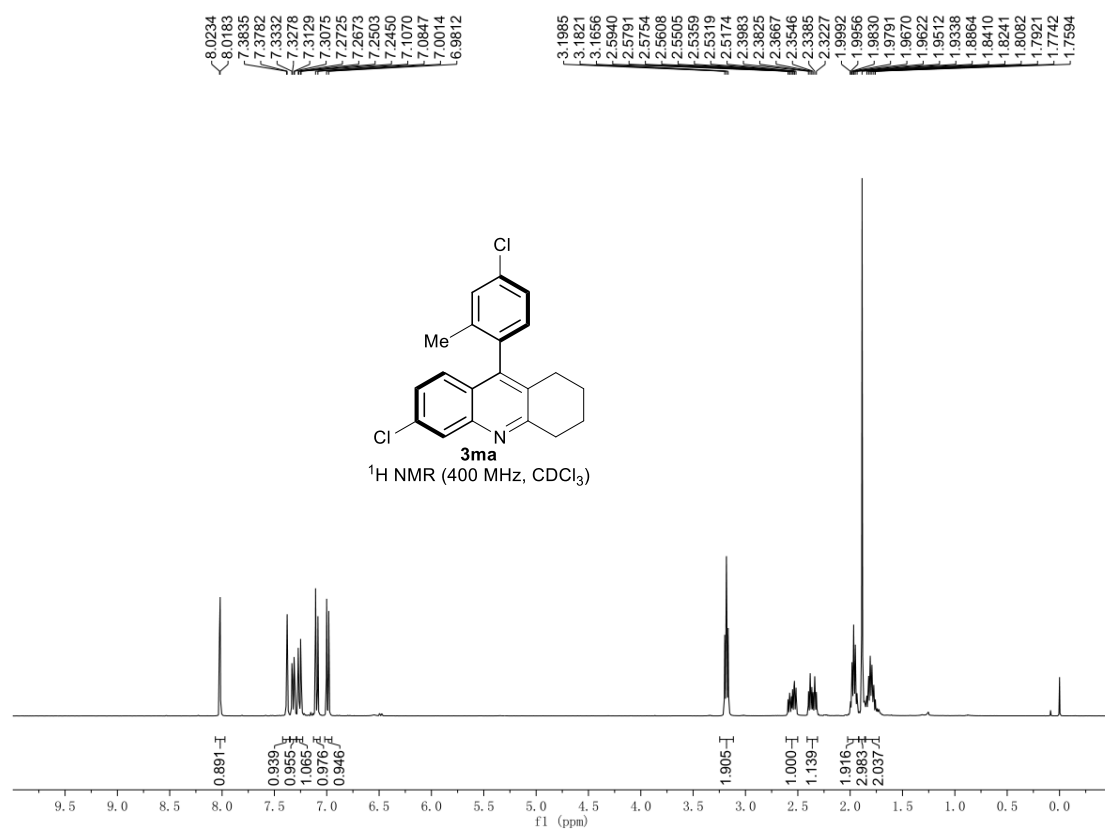


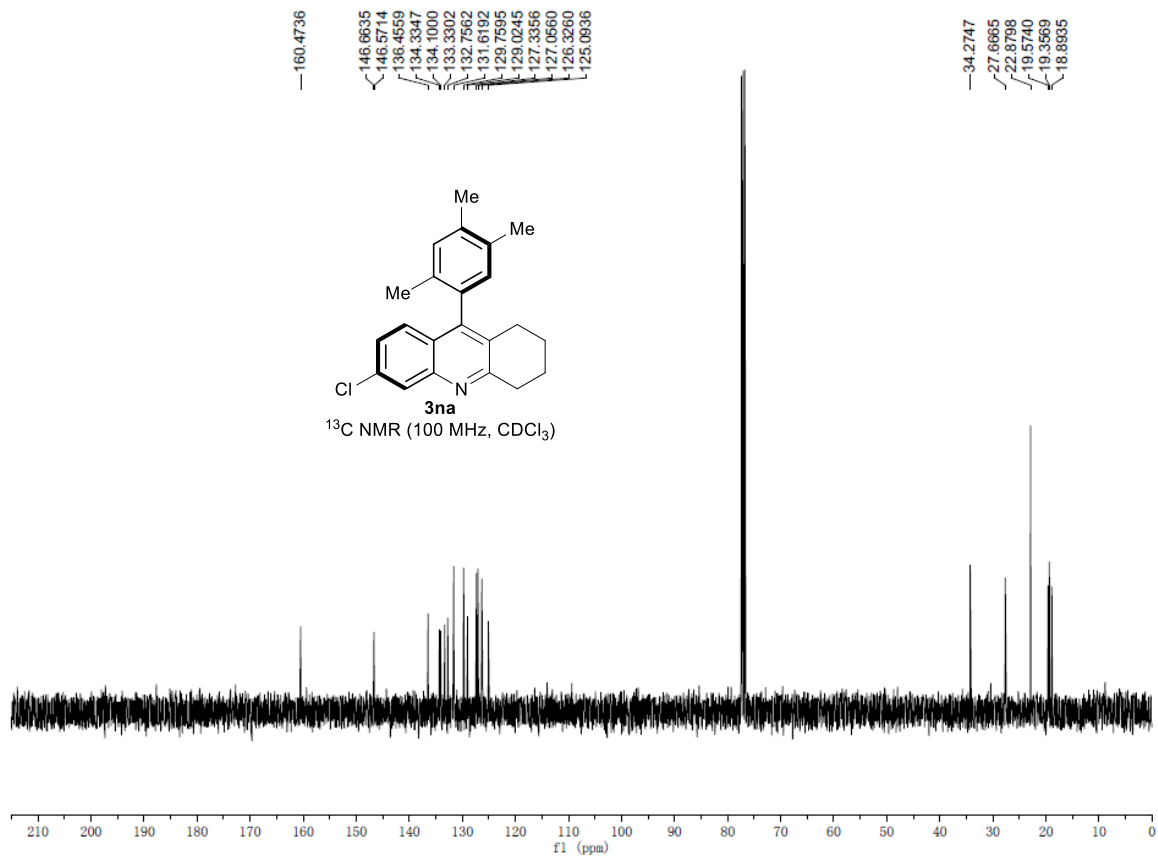
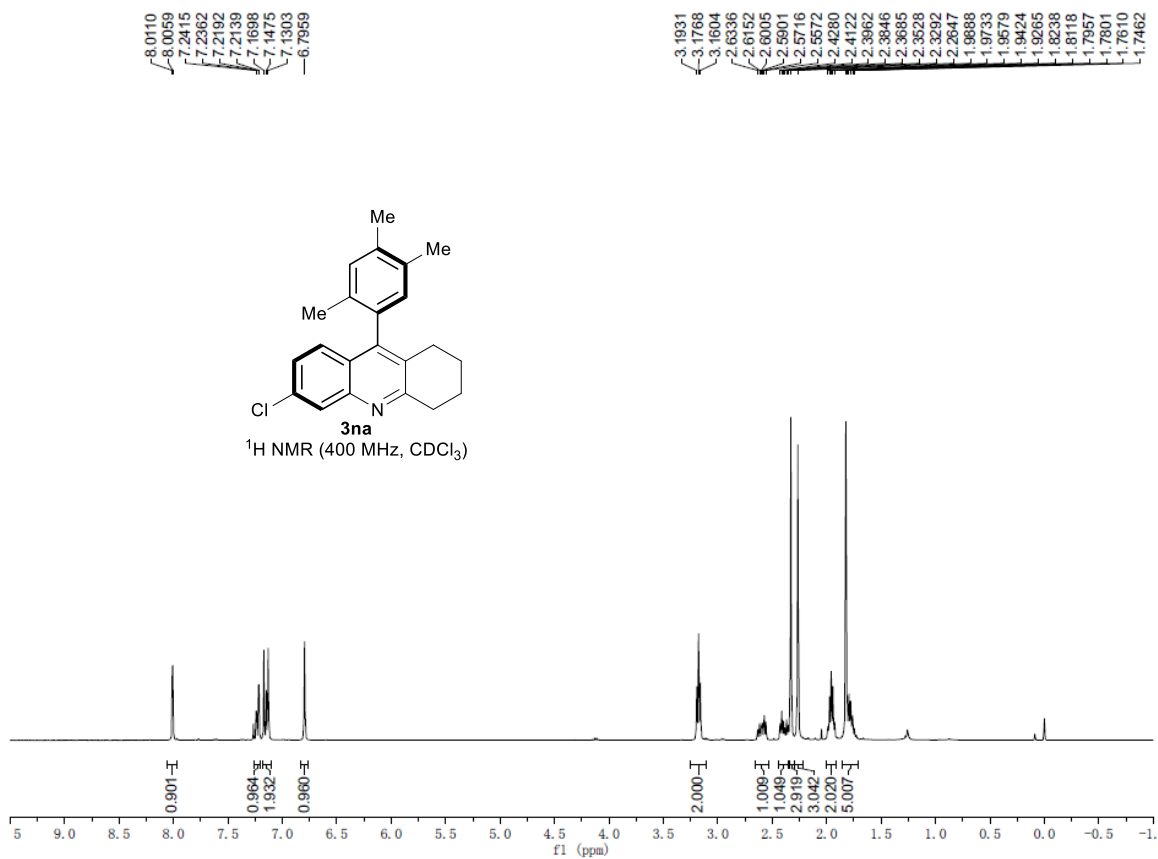




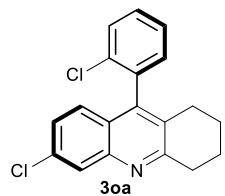




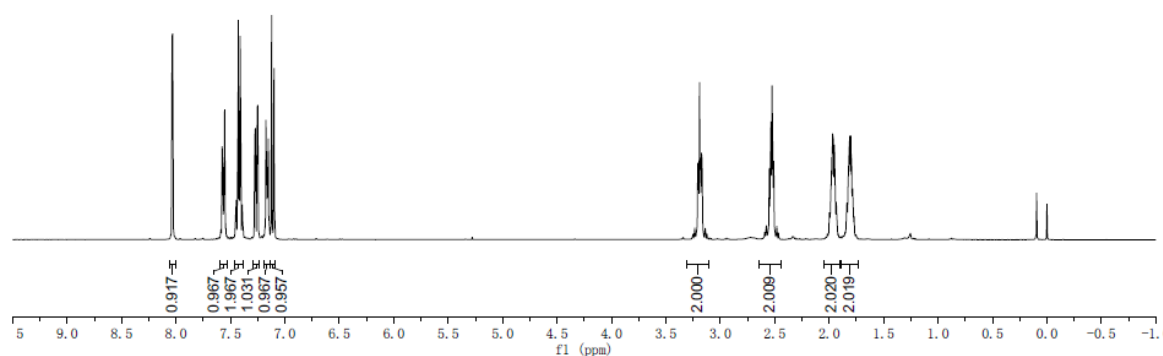




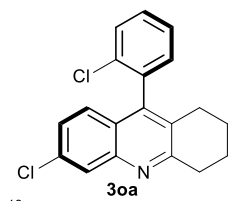
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$^1\text{H NMR}$ (400 MHz, CDCl_3)

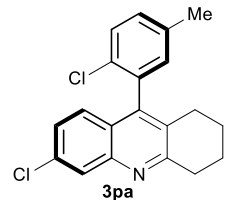
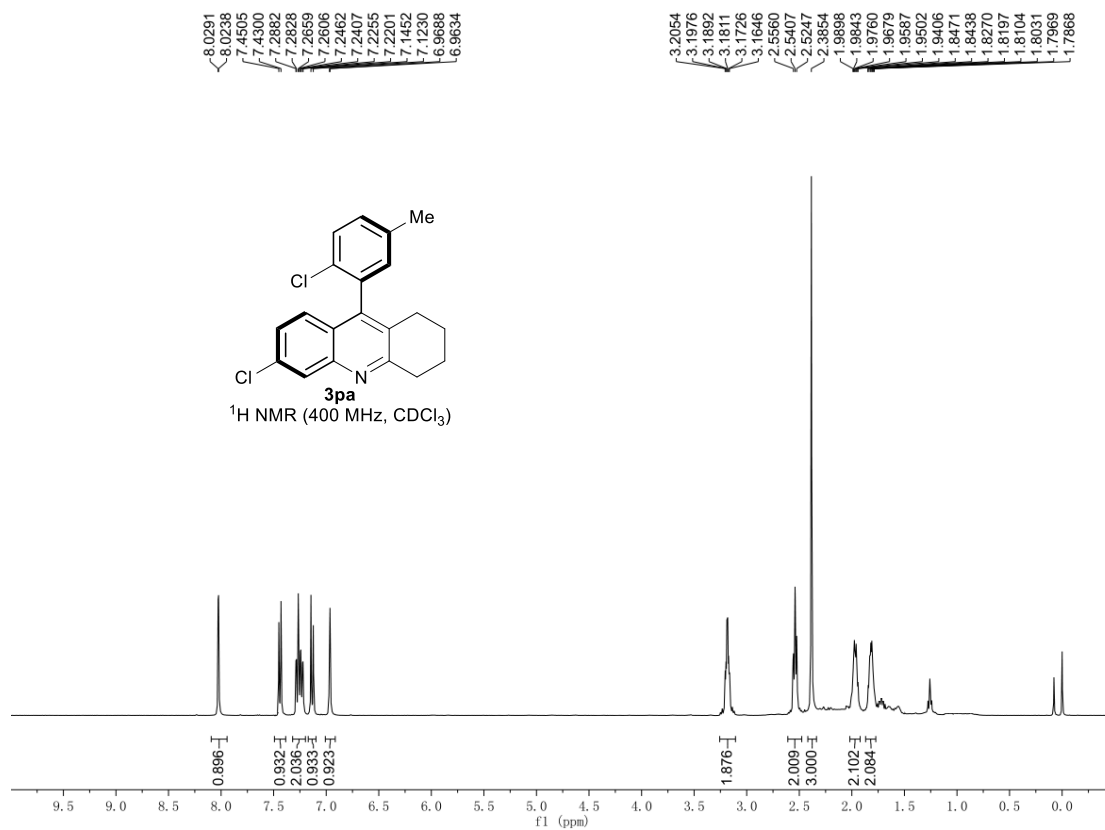


160.4837
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135.4492
134.2901
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129.7351
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127.2446
126.6656
126.4799
124.4854
34.2299
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22.7646
22.7146

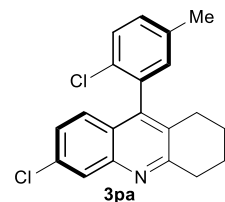
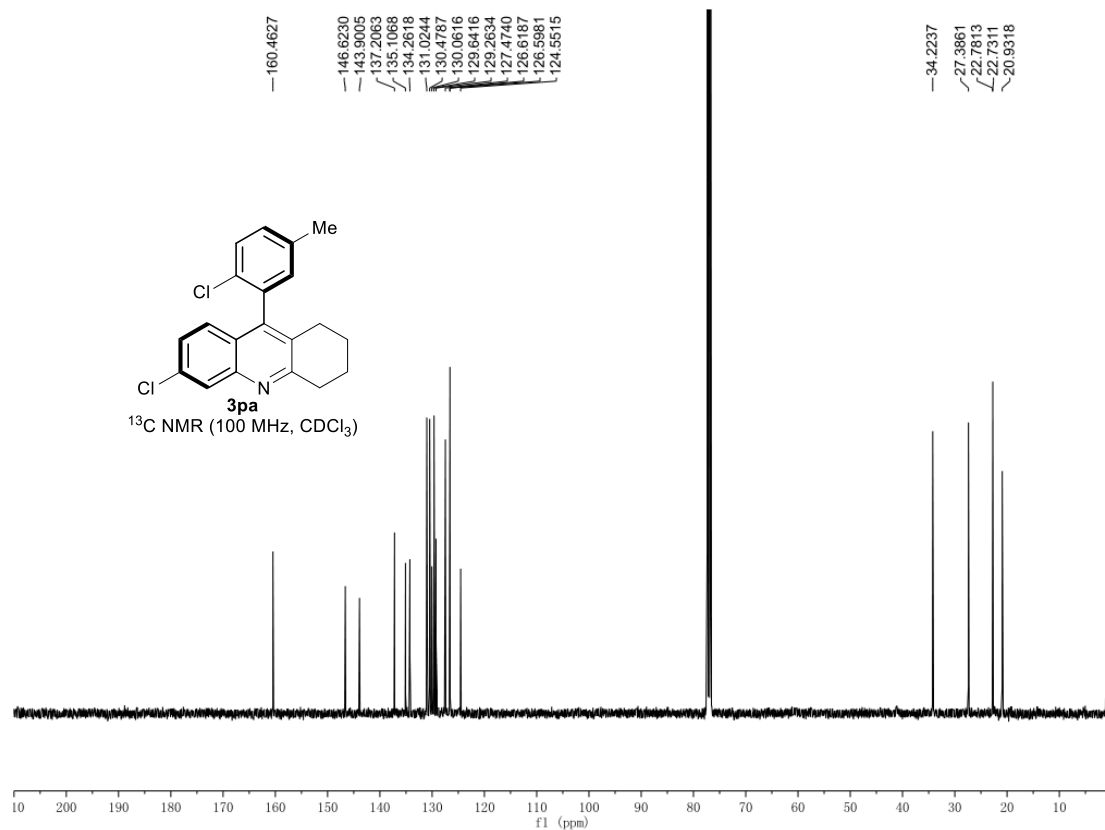


$^{13}\text{C NMR}$ (100 MHz, CDCl_3)

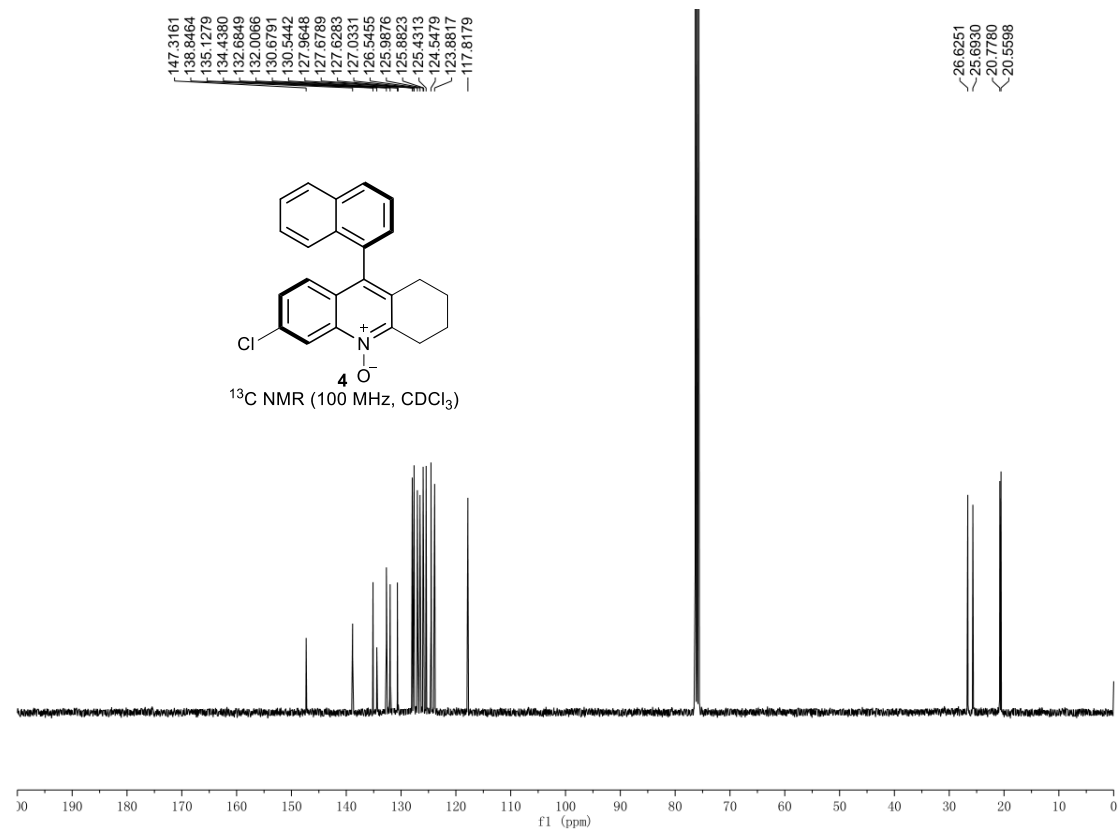
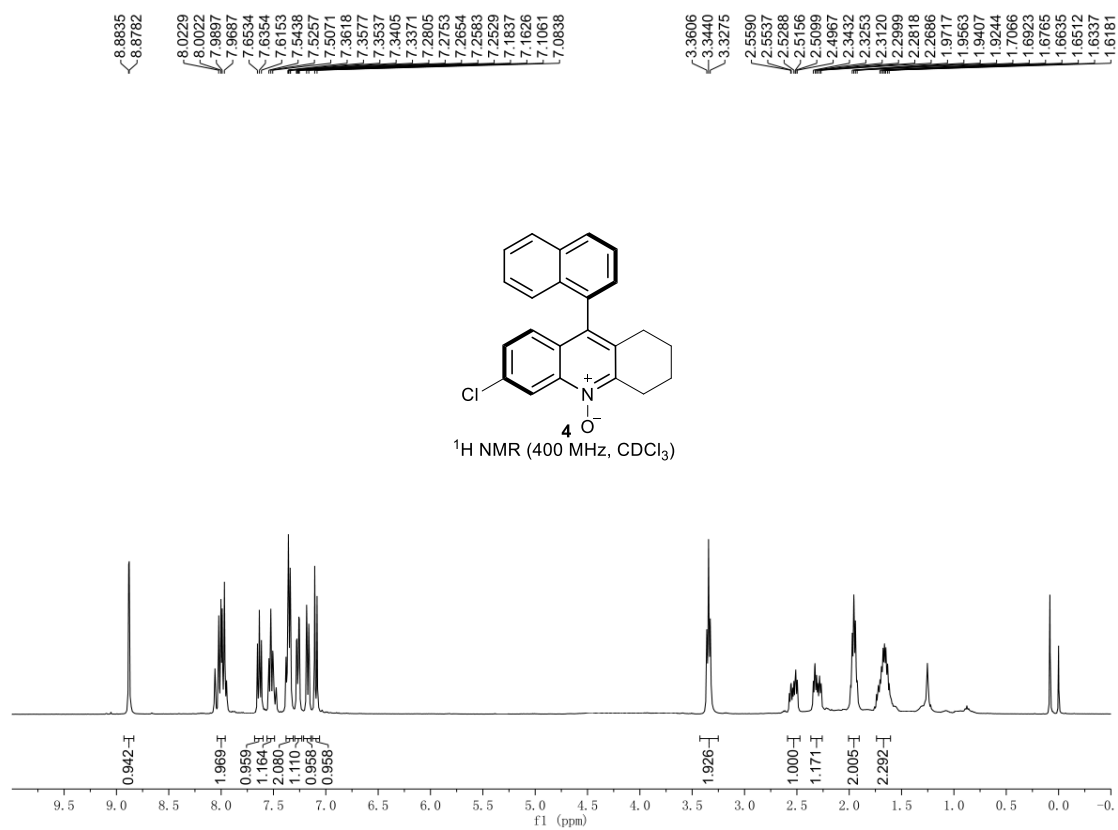


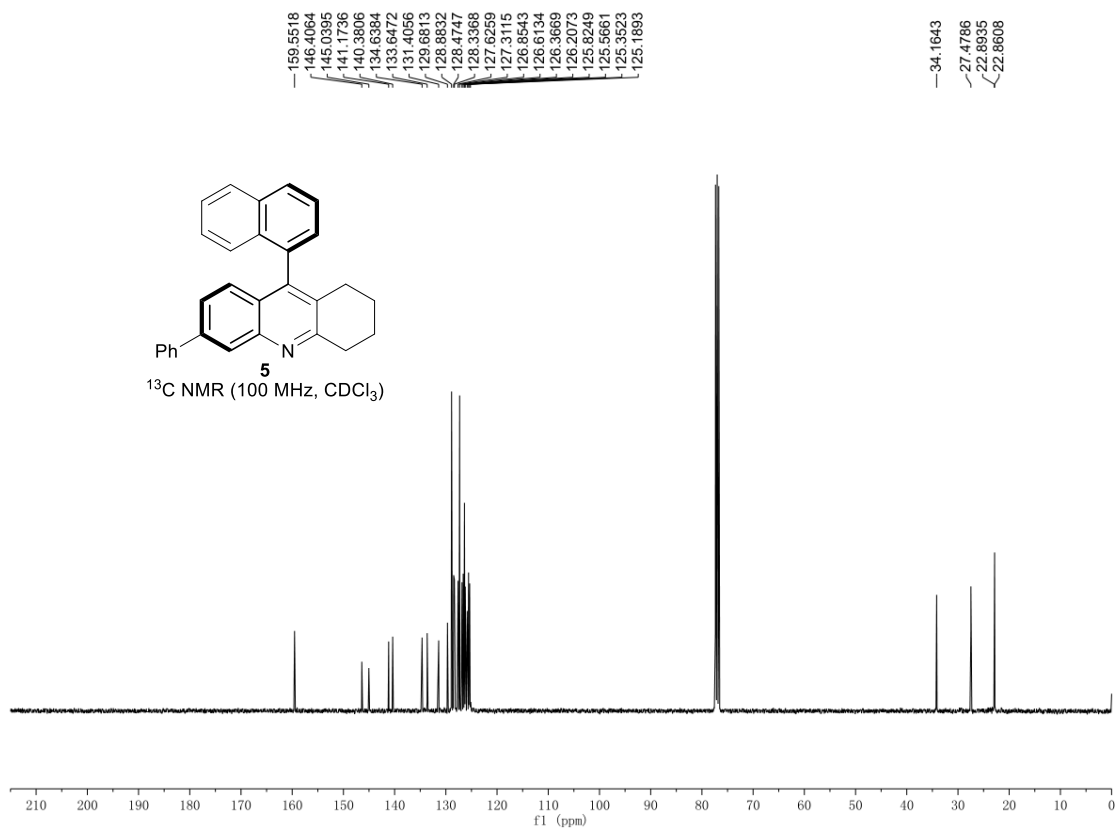
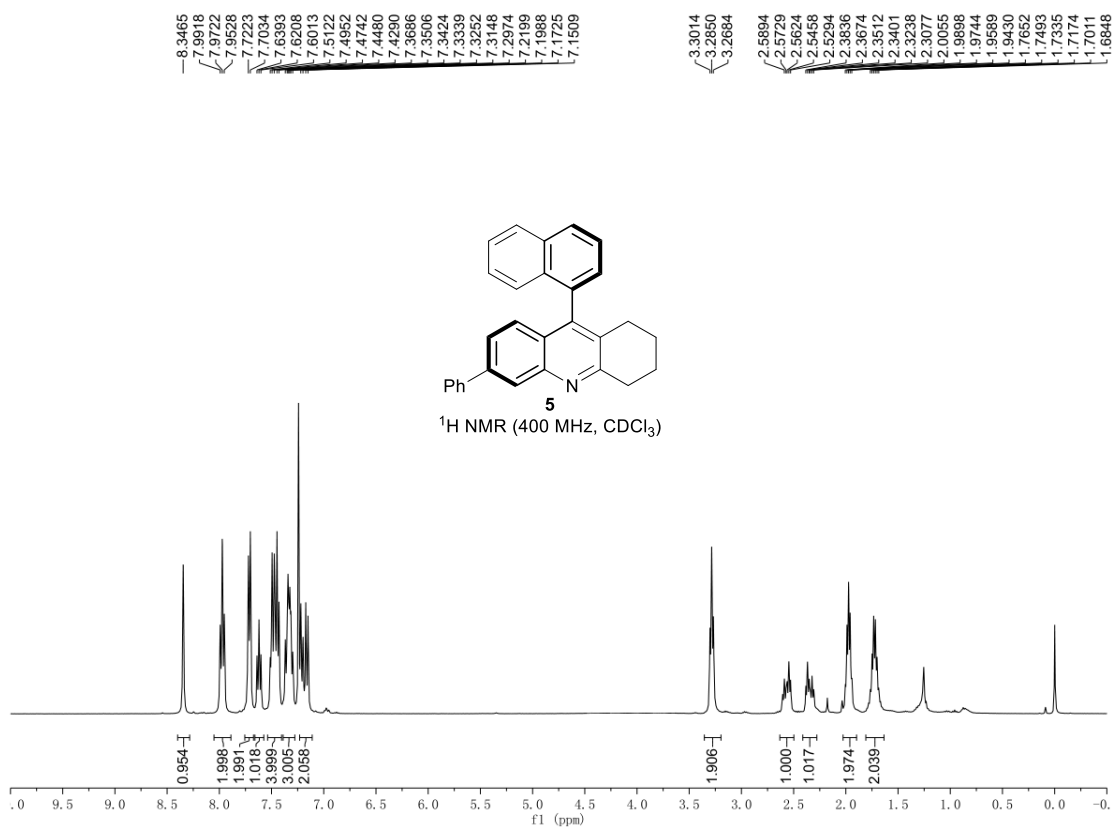


¹H NMR (400 MHz, CDCl₃)

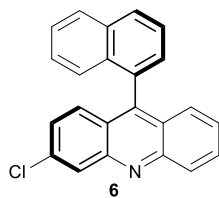


¹³C NMR (100 MHz, CDCl₃)

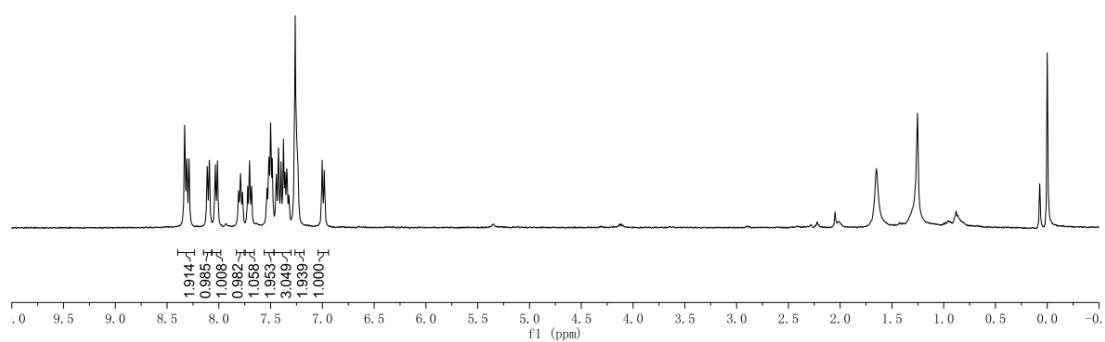




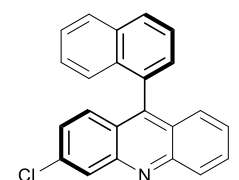
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7.7033
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7.4839
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7.4001
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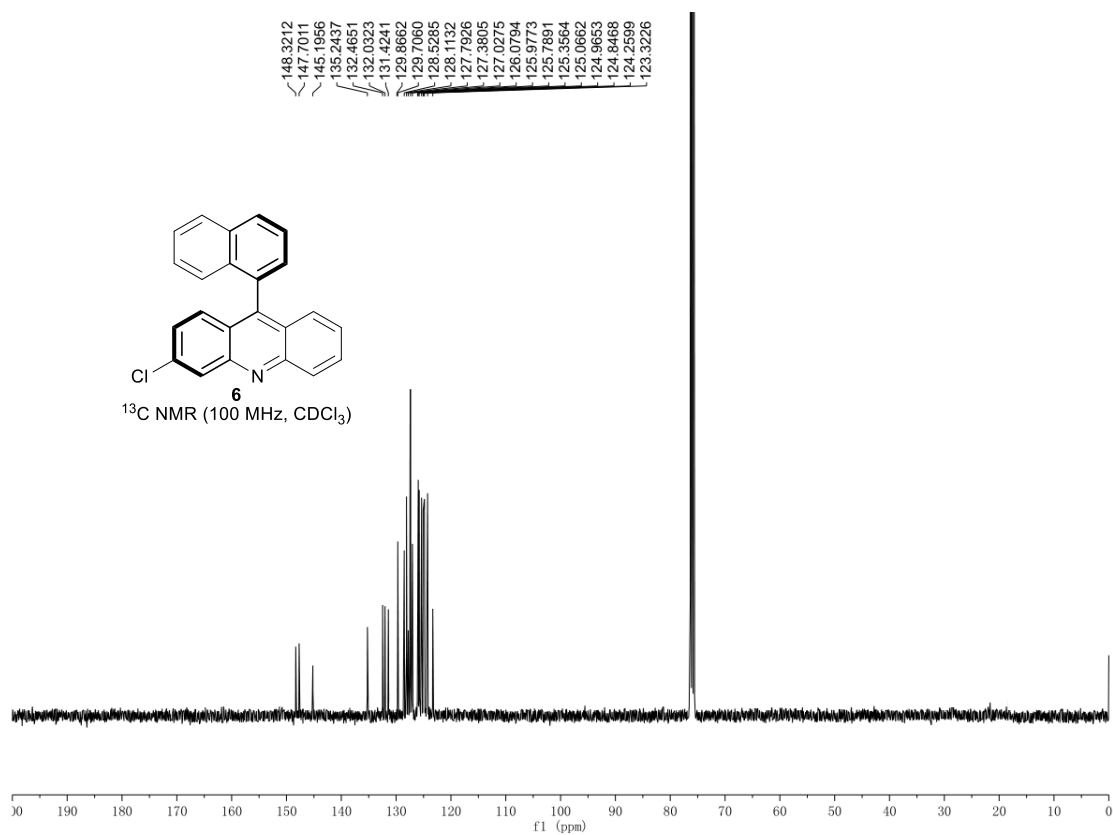
$^1\text{H NMR}$ (400 MHz, CDCl_3)



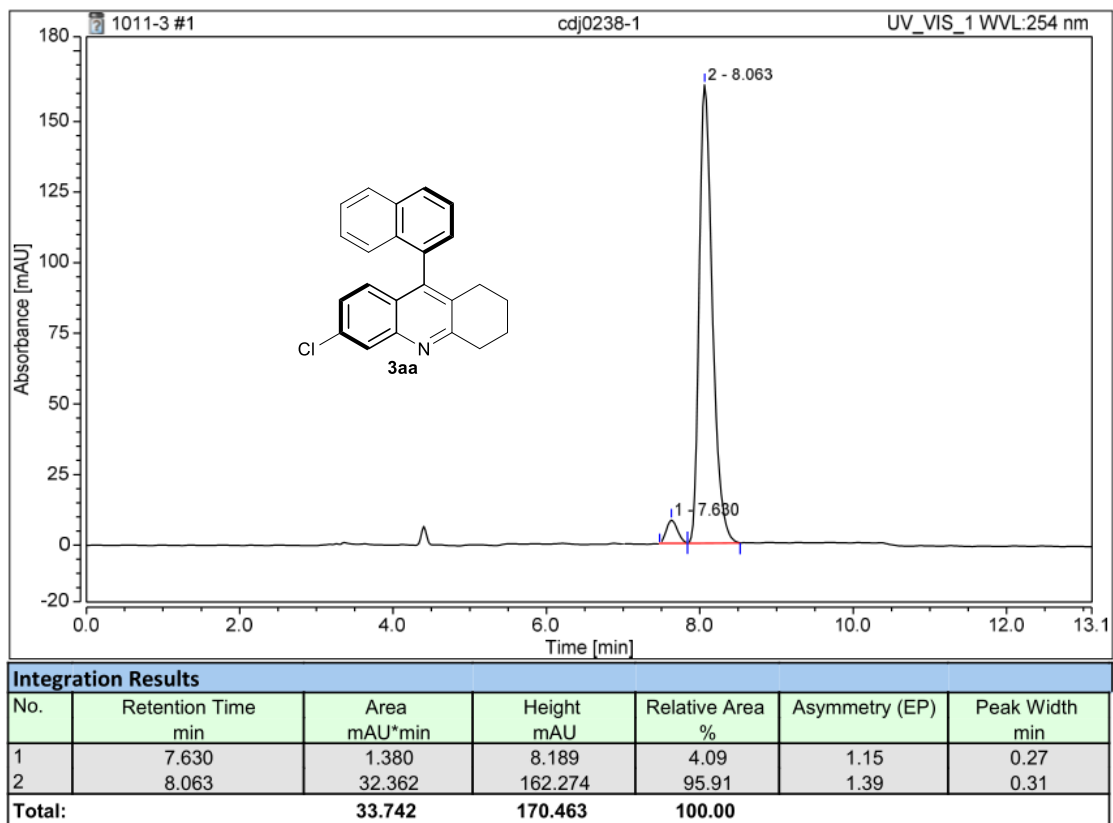
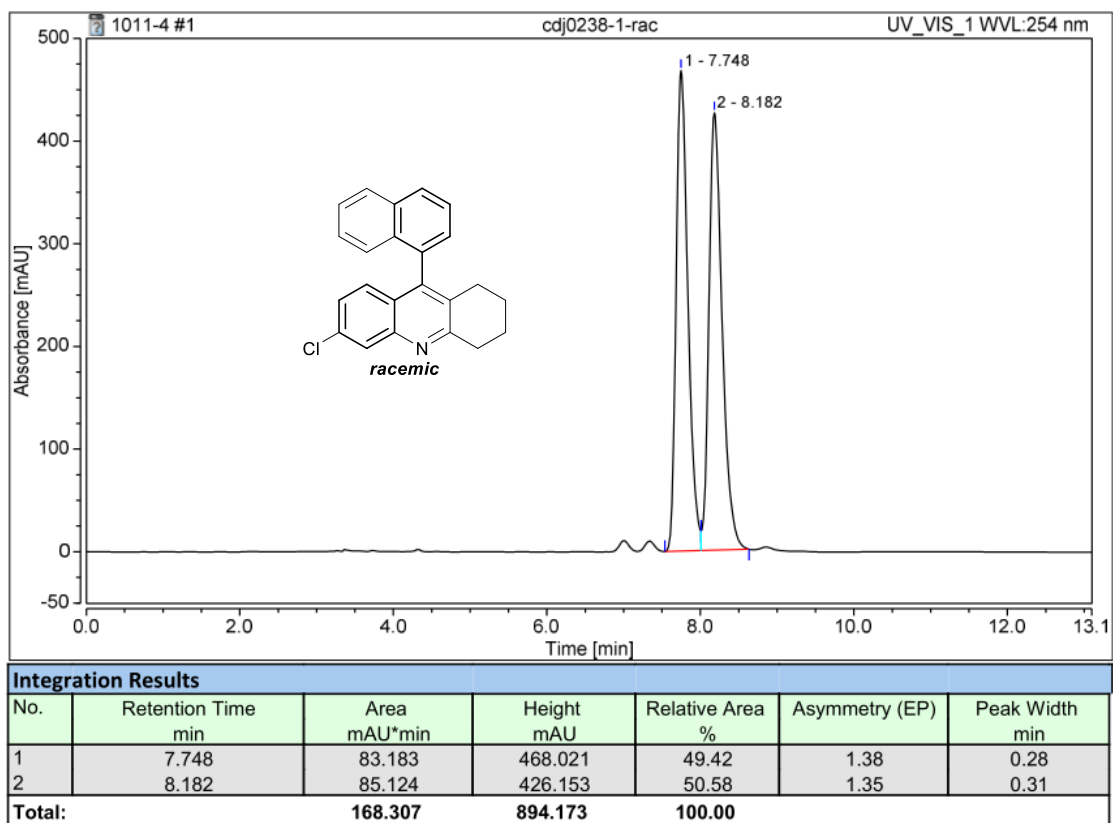
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127.3805
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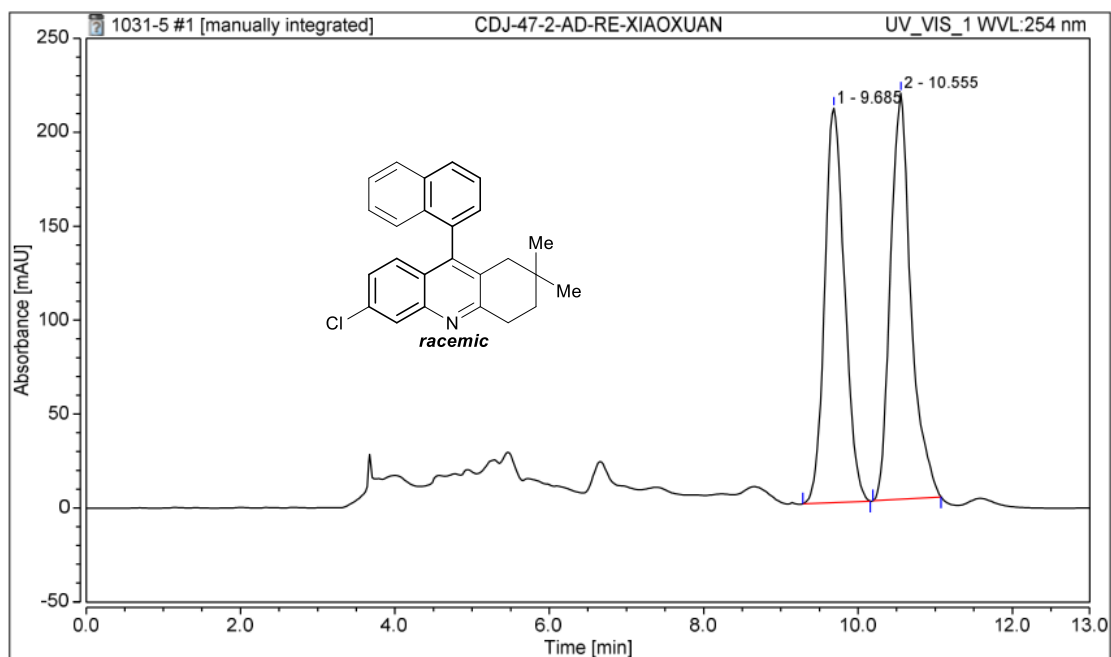


$^{13}\text{C NMR}$ (100 MHz, CDCl_3)

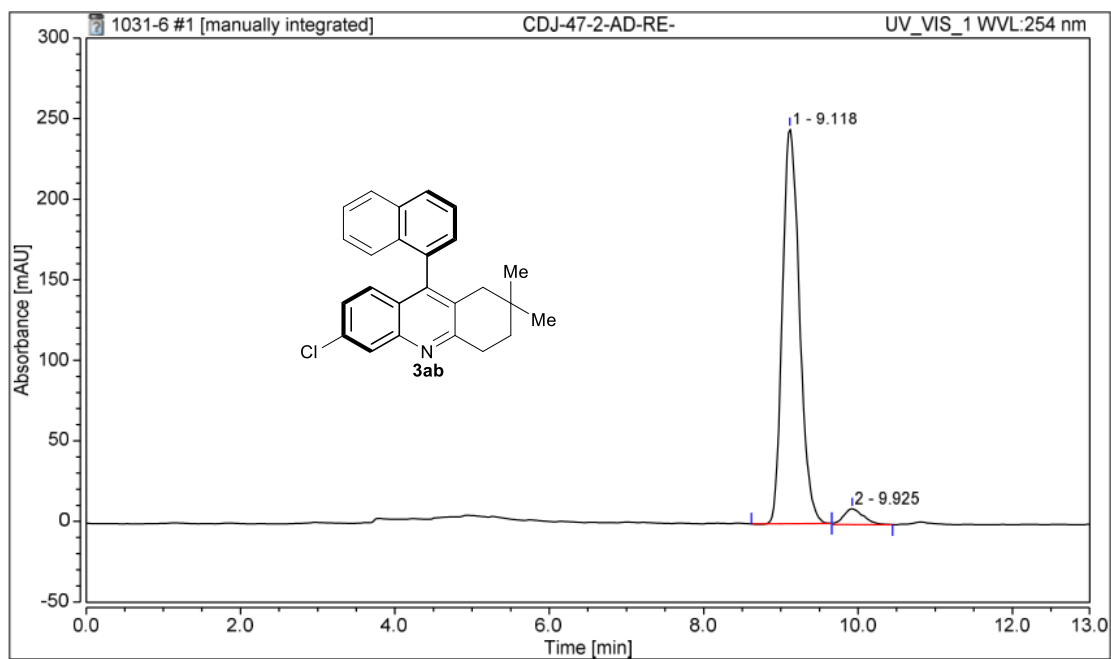


HPLC Traces

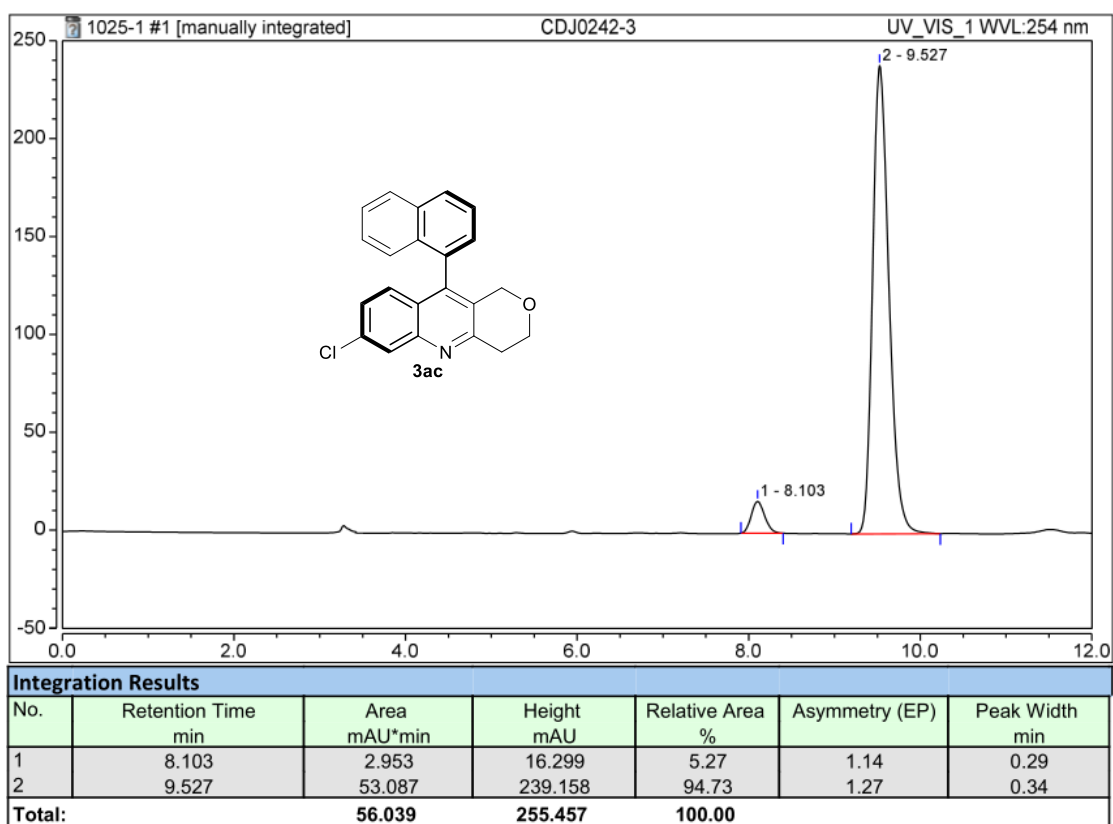
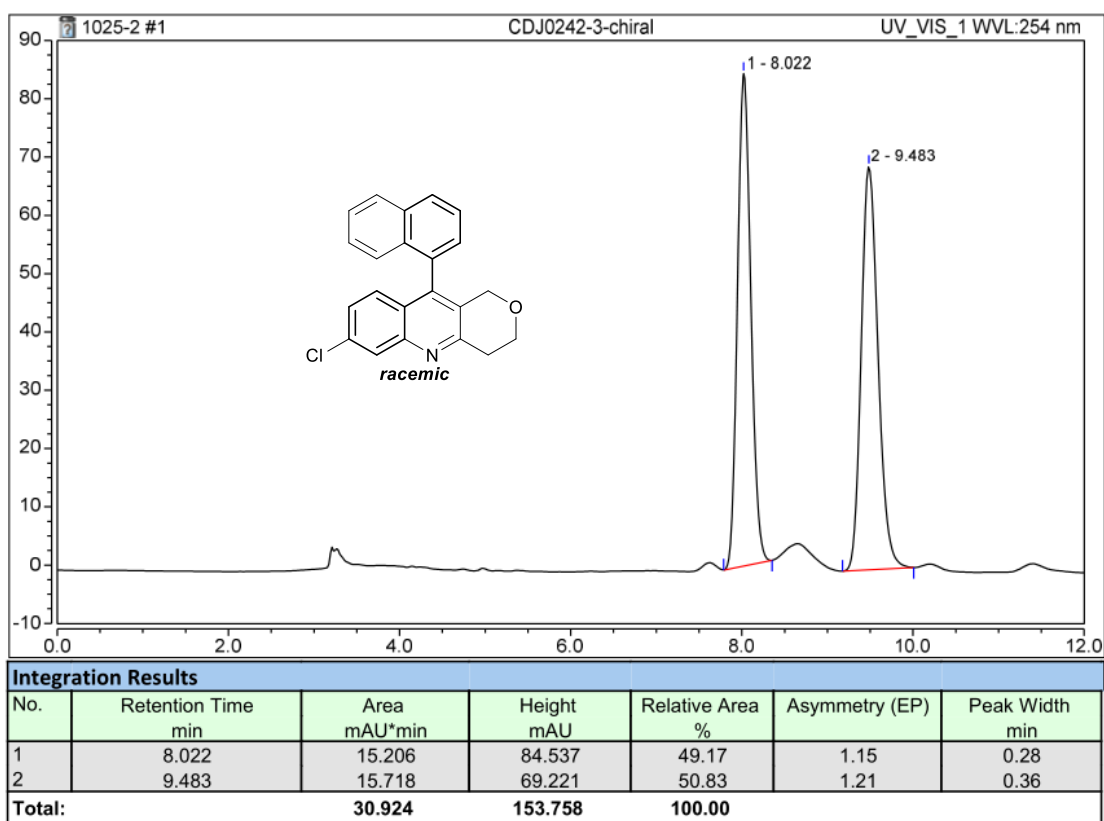


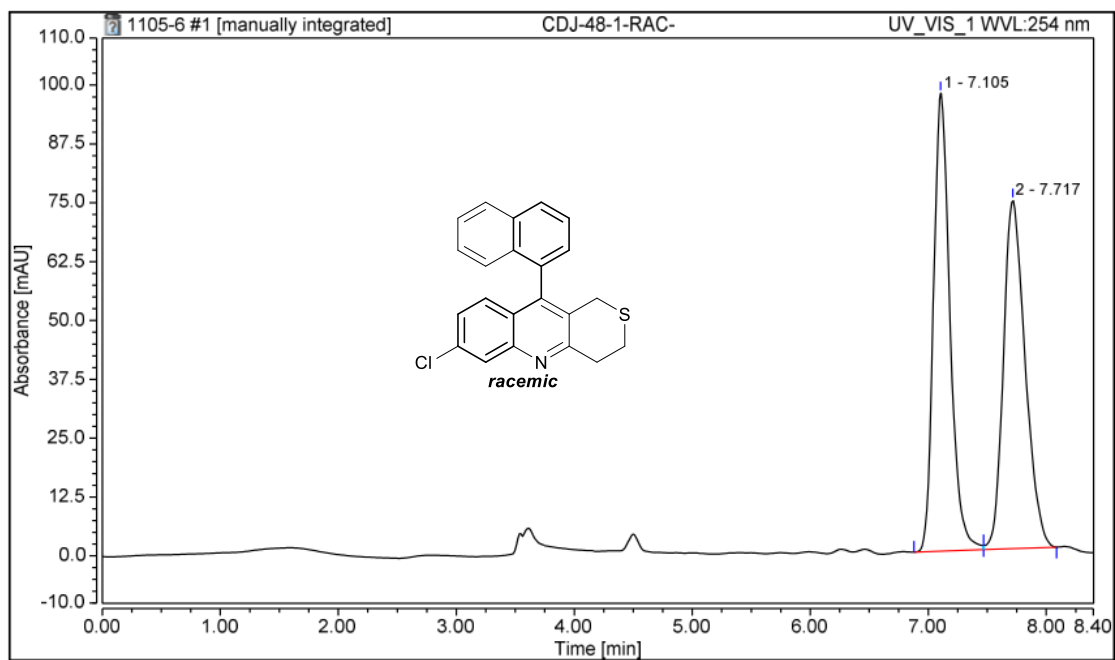


Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	9.685	62.749	210.040	48.46	1.13	0.45
2	10.555	66.745	216.242	51.54	1.25	0.44
Total:		129.495	426.282	100.00		

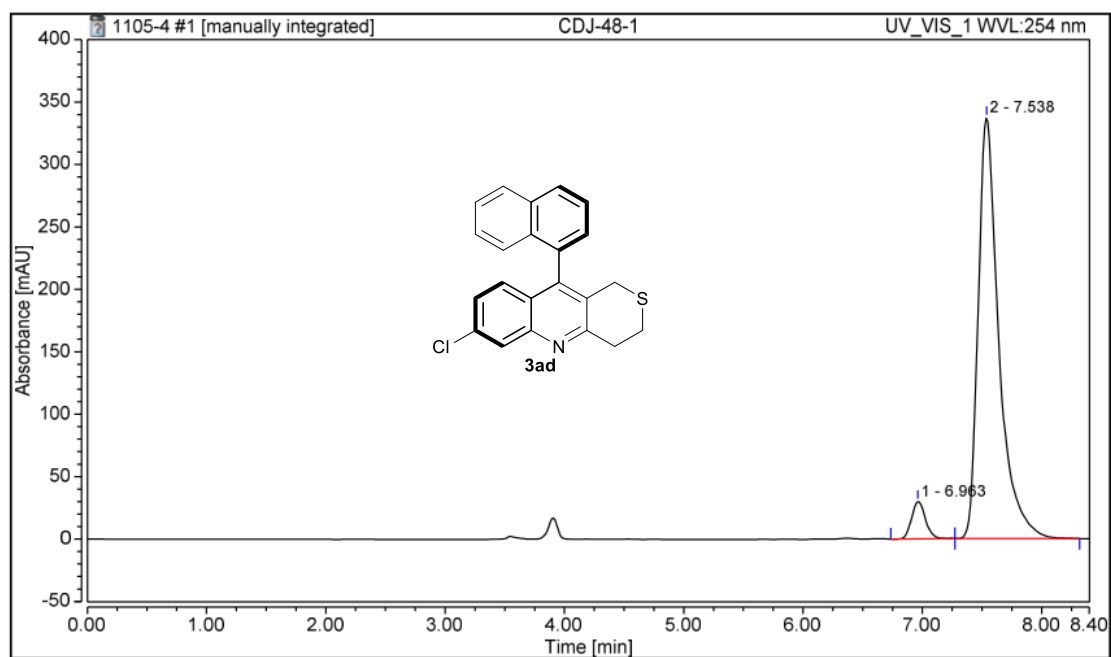


Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	9.118	62.733	245.102	95.59	1.23	0.50
2	9.925	2.892	9.725	4.41	n.a.	0.43
Total:		65.626	254.827	100.00		

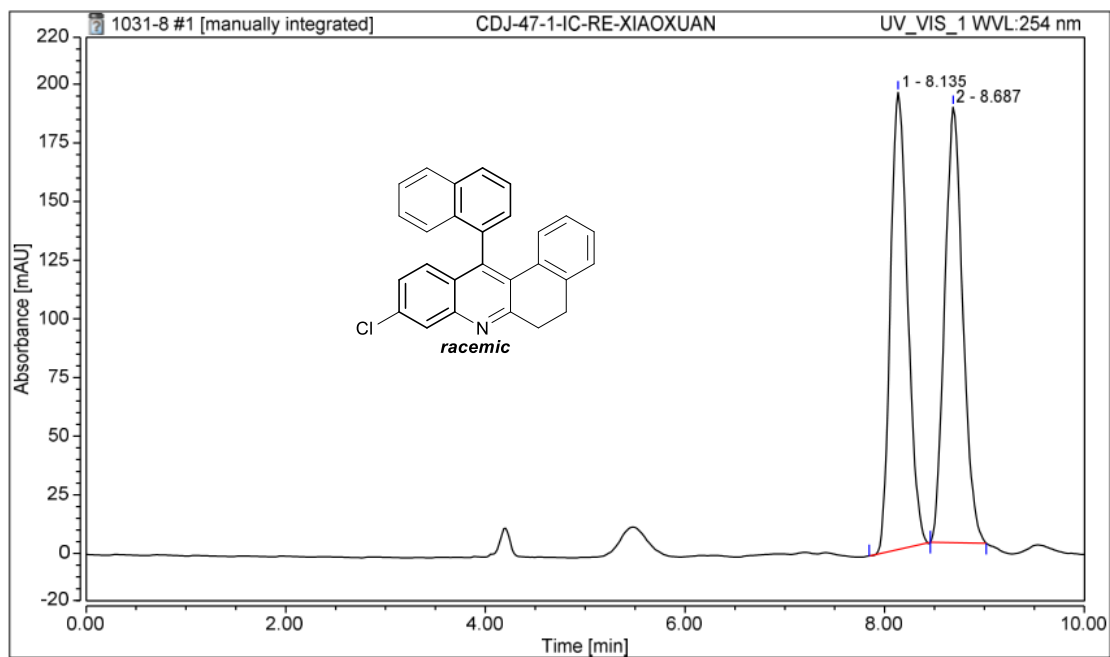




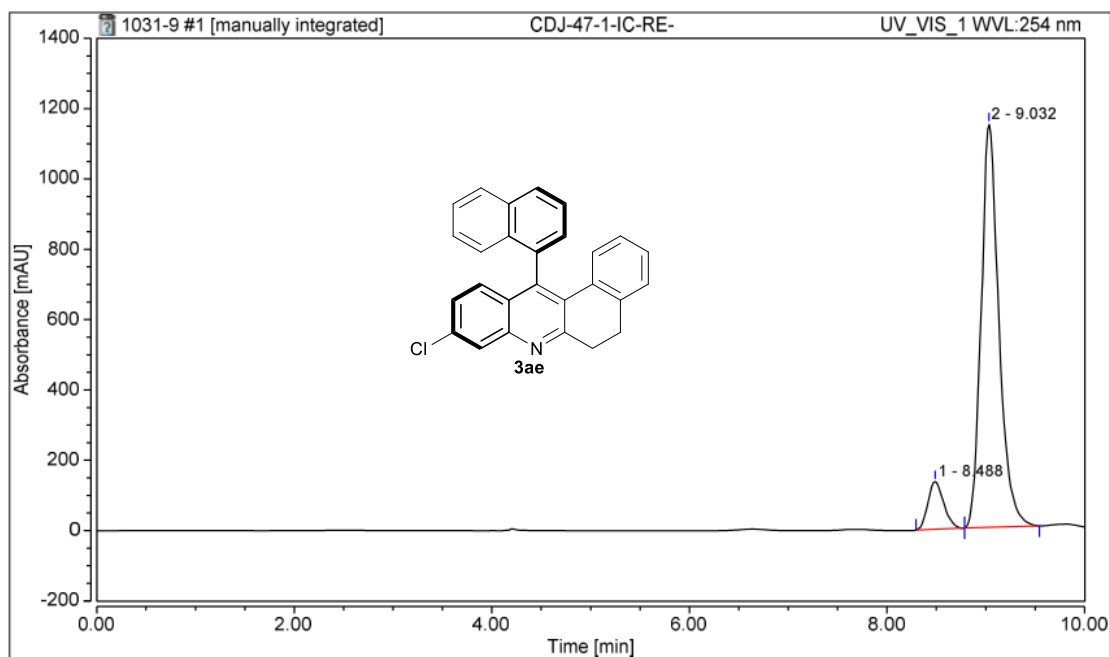
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.105	15.659	97.336	49.77	1.29	0.24
2	7.717	15.806	74.080	50.23	1.25	0.32
Total:		31.465	171.416	100.00		



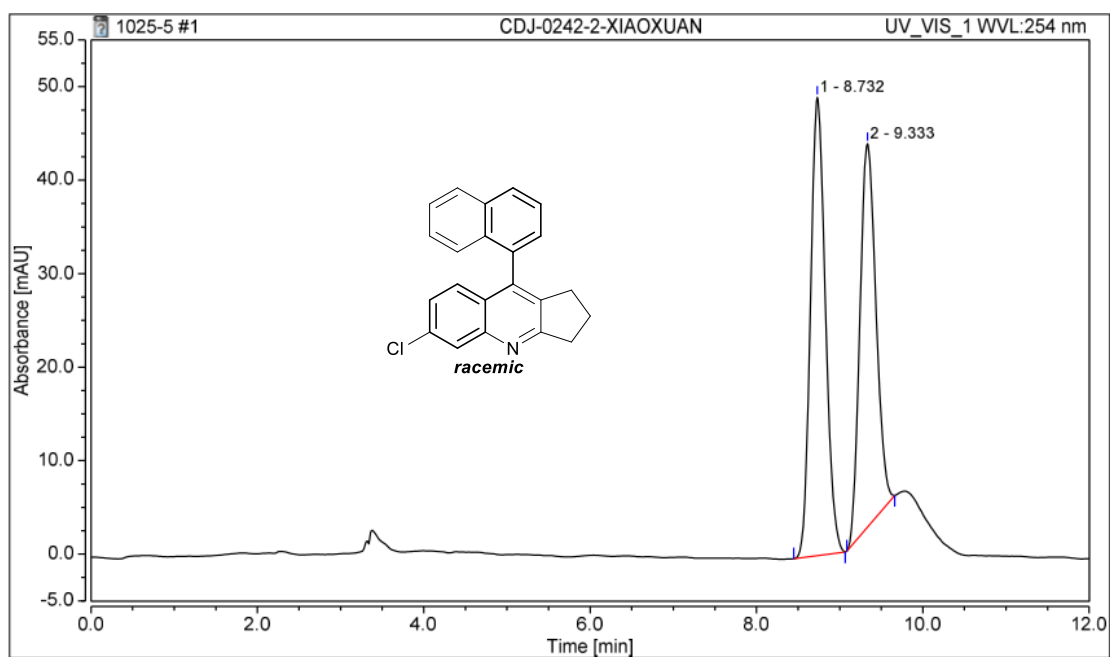
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	6.963	3.994	29.872	5.58	1.14	0.21
2	7.538	67.542	337.078	94.42	1.60	0.29
Total:		71.536	366.950	100.00		



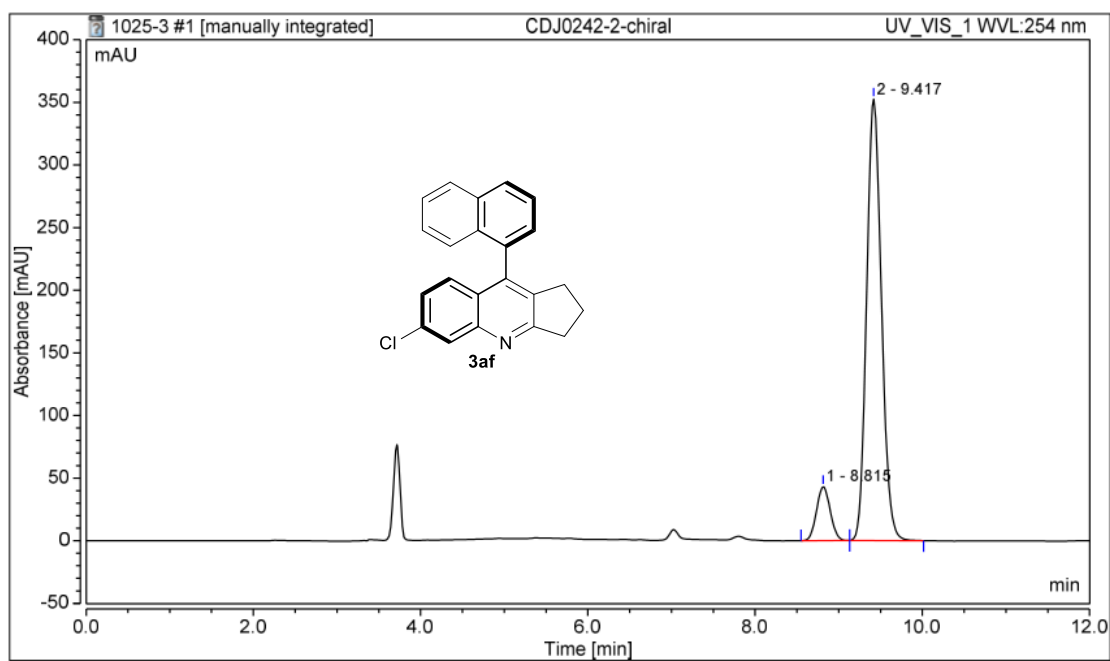
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.135	38.689	194.888	49.82	1.20	0.30
2	8.687	38.962	185.792	50.18	1.15	0.35
Total:		77.651	380.680	100.00		



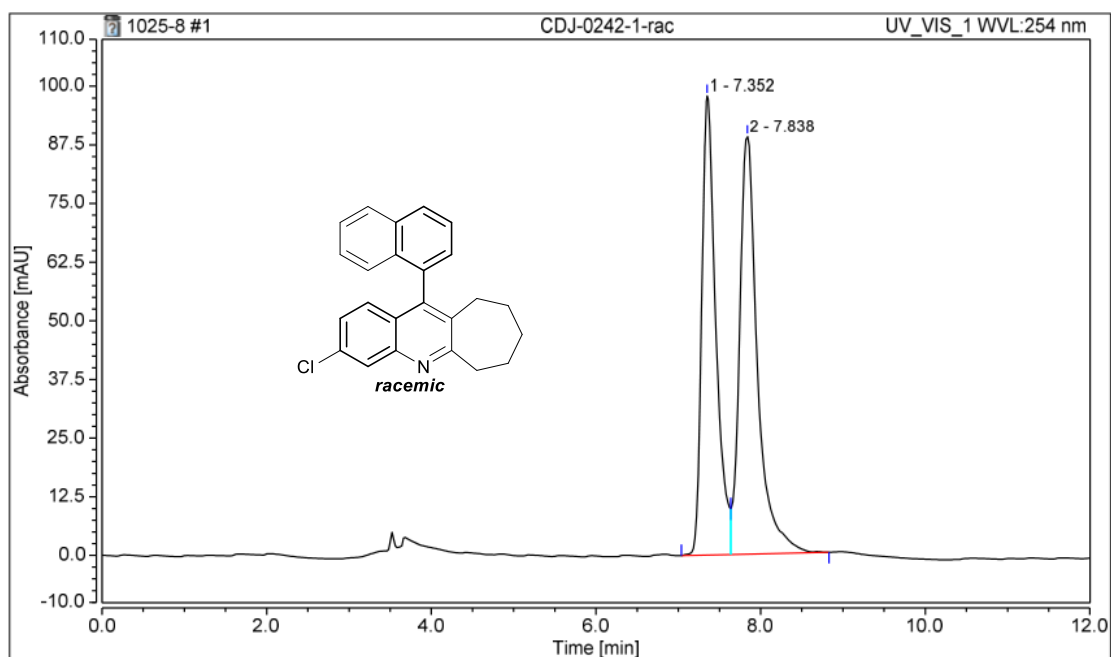
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.488	24.874	134.761	9.46	1.18	0.29
2	9.032	238.190	1146.390	90.54	1.26	0.30
Total:		263.064	1281.151	100.00		



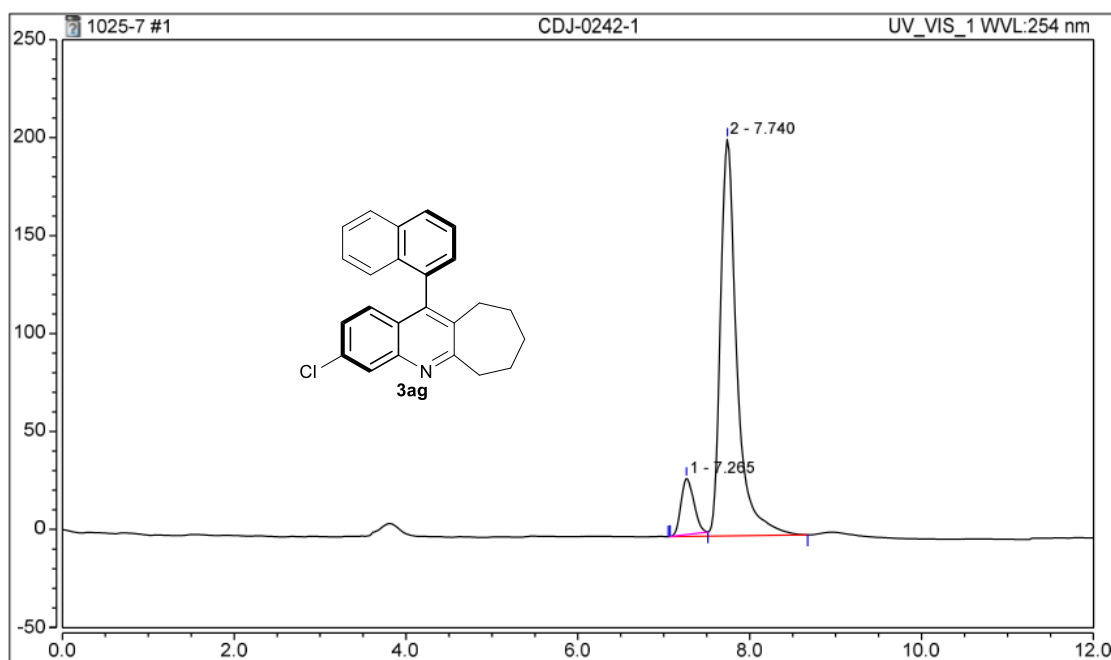
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.732	10.153	49.049	52.53	1.15	0.32
2	9.333	9.175	41.025	47.47	1.15	0.36
Total:		19.328	90.074	100.00		



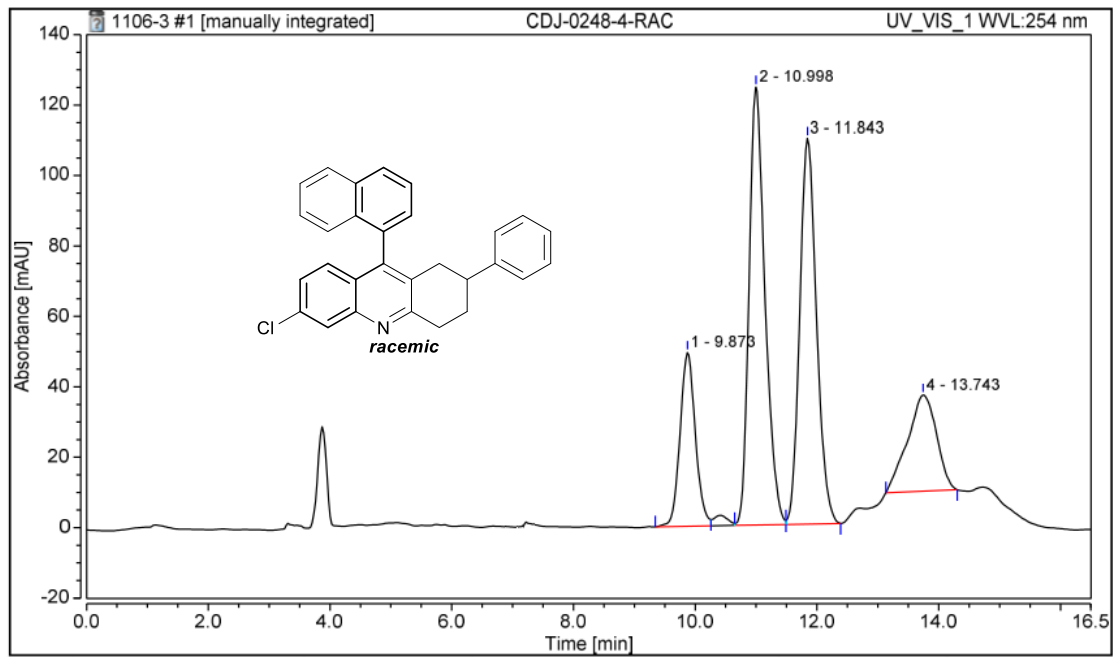
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.815	8.136	43.043	10.11	1.06	0.30
2	9.417	72.347	352.439	89.89	1.13	0.33
Total:		80.483	395.482	100.00		



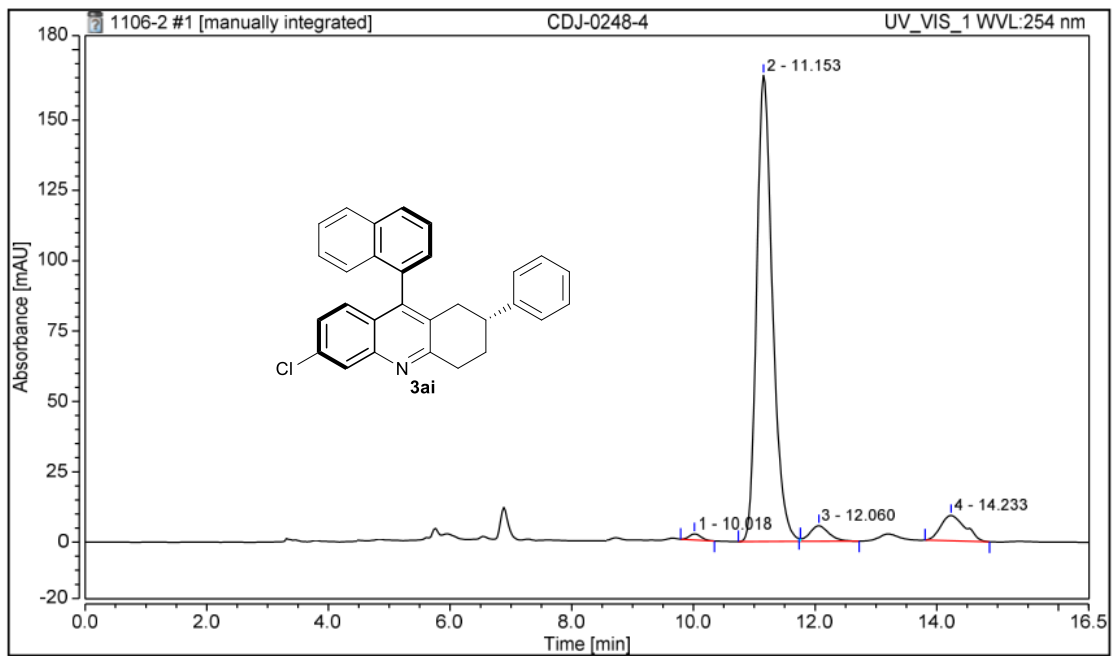
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.352	19.385	97.895	46.28	n.a.	0.29
2	7.838	22.501	89.051	53.72	n.a.	0.35
Total:		41.885	186.947	100.00		



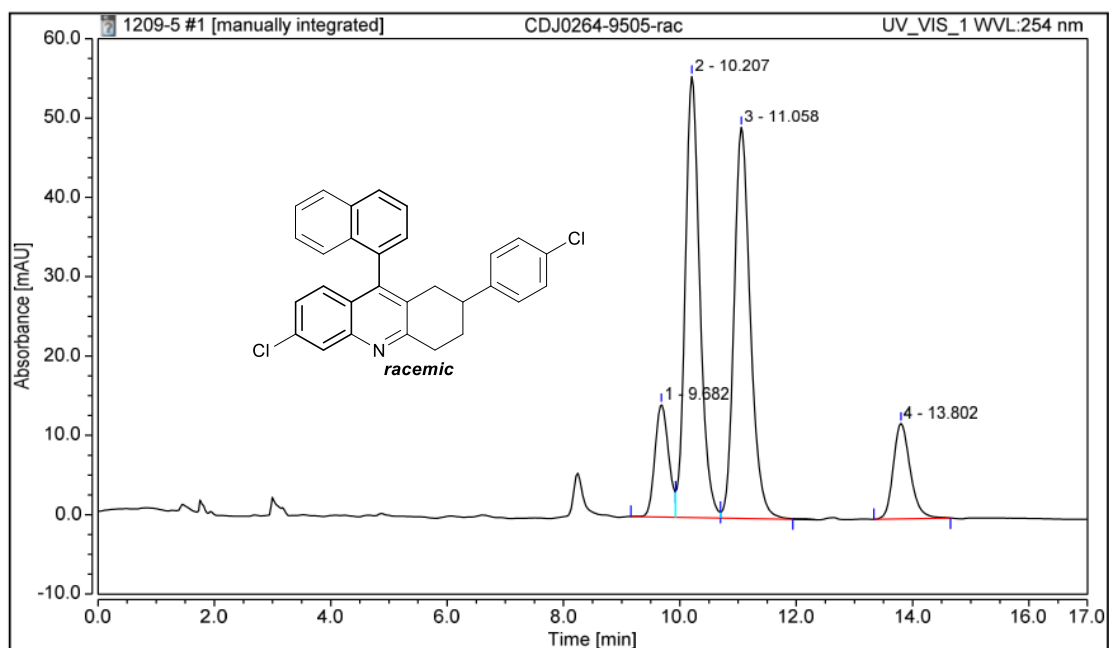
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.265	4.923	28.669	9.84	n.a.	n.a.
2	7.740	45.128	202.541	90.16	1.55	0.31
Total:		50.051	231.210	100.00		



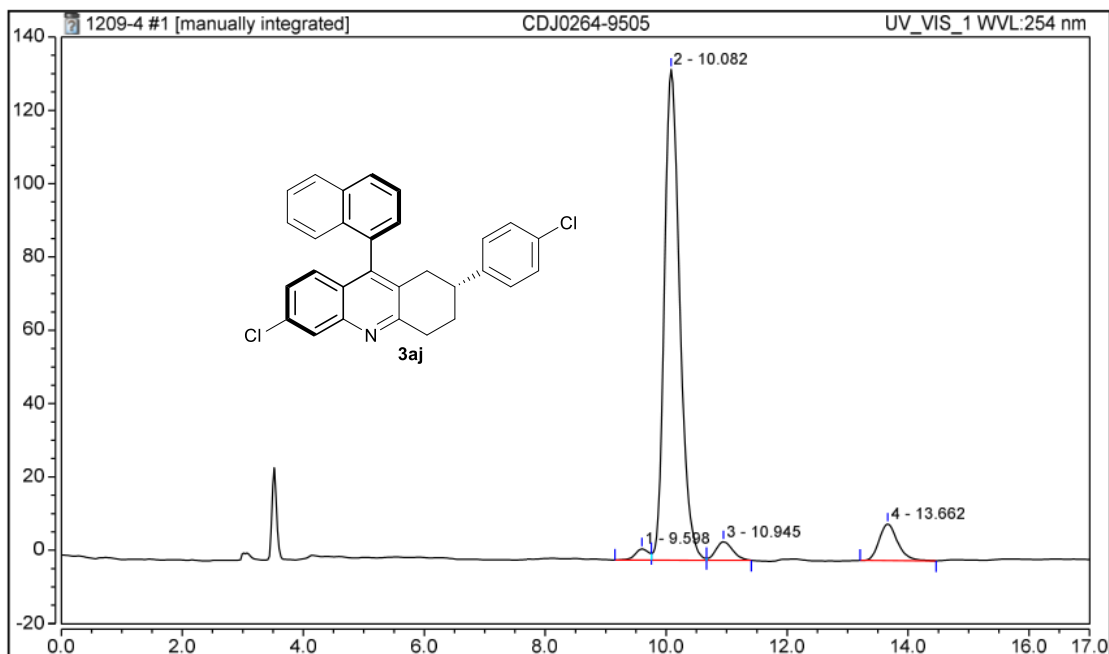
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	9.873	14.415	49.400	14.11	1.11	0.45
2	10.998	37.722	124.412	36.93	1.28	0.46
3	11.843	34.946	109.596	34.22	1.17	0.52
4	13.743	15.052	27.353	14.74	0.91	0.92
Total:		102.136	310.762	100.00		



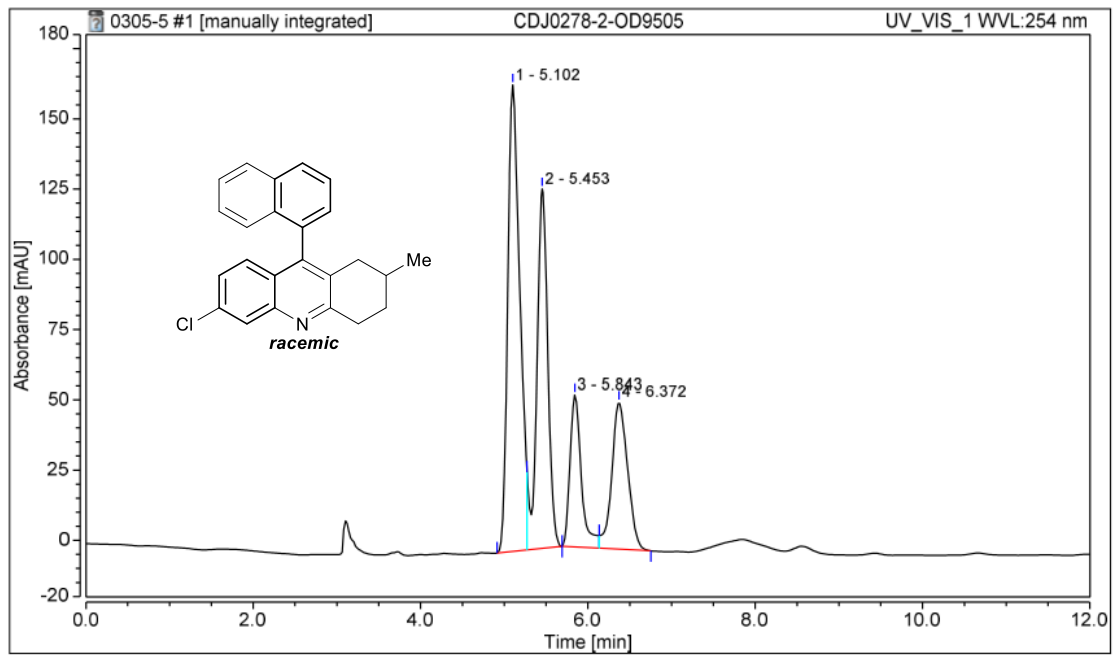
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	10.018	0.468	2.131	0.83	1.13	0.34
2	11.153	49.944	165.678	88.24	1.28	0.45
3	12.060	2.004	5.513	3.54	n.a.	0.58
4	14.233	4.184	8.950	7.39	1.21	0.74
Total:		56.600	182.271	100.00		



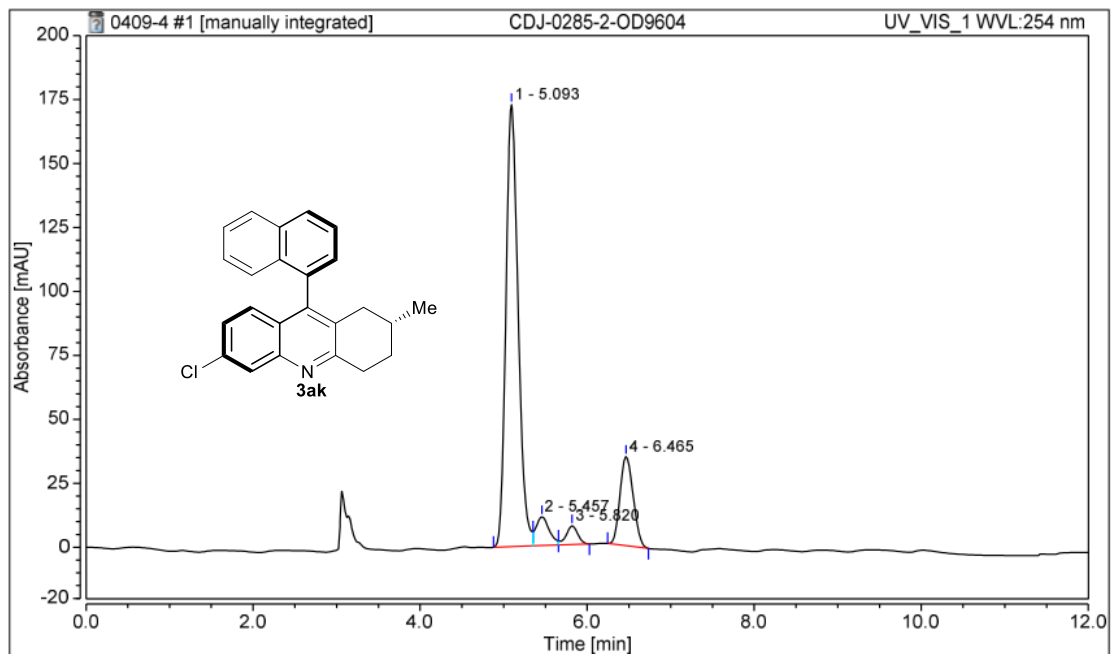
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	9.682	3.858	14.188	9.64	n.a.	0.44
2	10.207	16.127	55.607	40.29	n.a.	0.45
3	11.058	15.968	49.304	39.89	1.23	0.50
4	13.802	4.077	12.057	10.19	1.21	0.52
Total:		40.030	131.156	100.00		



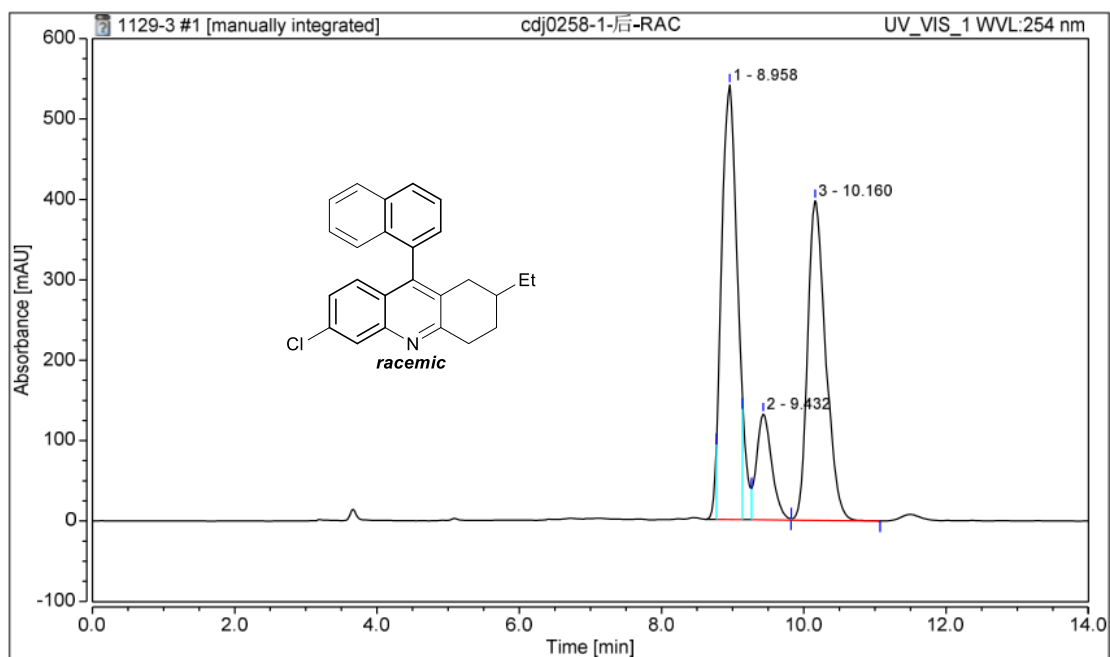
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	9.598	0.793	3.055	1.75	n.a.	0.49
2	10.082	39.503	133.931	86.90	1.27	0.46
3	10.945	1.635	5.024	3.60	n.a.	0.51
4	13.662	3.526	9.967	7.76	1.22	0.55
Total:		45.457	151.977	100.00		



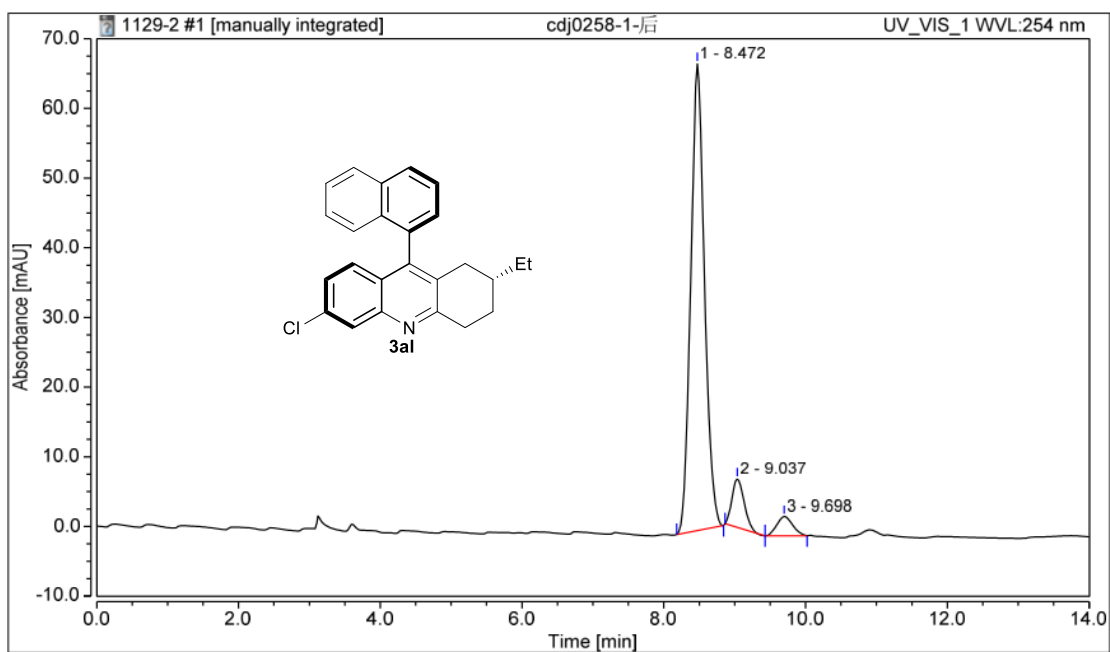
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.102	23.542	165.999	35.27	n.a.	0.27
2	5.453	23.243	127.965	34.82	n.a.	0.23
3	5.843	9.893	54.133	14.82	n.a.	0.23
4	6.372	10.065	52.259	15.08	n.a.	0.34
Total:		66.744	400.356	100.00		



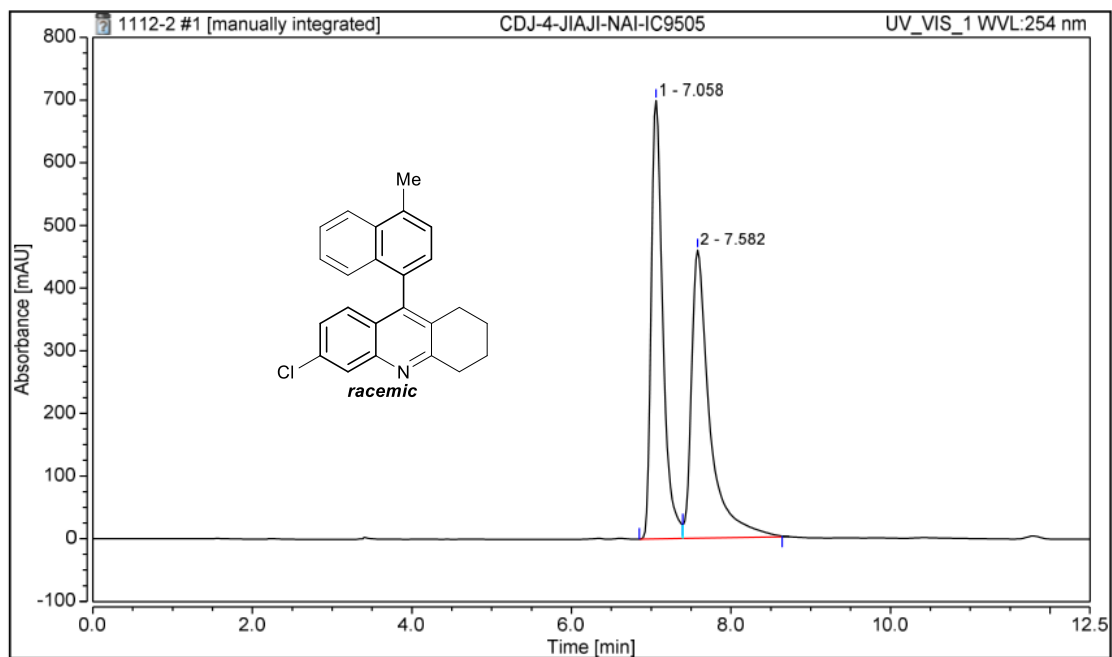
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.093	29.232	172.834	75.25	1.22	0.27
2	5.457	2.054	11.279	5.29	n.a.	0.31
3	5.820	1.177	7.302	3.03	n.a.	0.25
4	6.465	6.383	34.882	16.43	1.13	0.30
Total:		38.846	226.296	100.00		



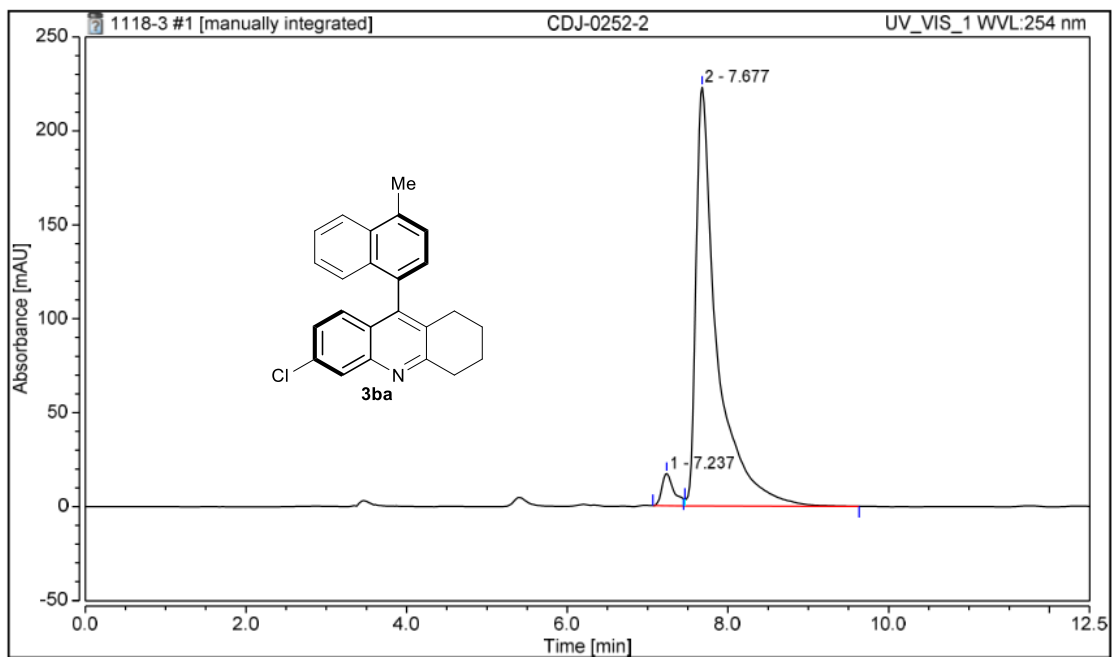
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.958	128.773	540.529	46.90	n.a.	0.43
2	9.432	128.773	131.718	11.60	n.a.	0.39
3	10.160	128.773	397.550	41.50	1.30	0.46
Total:		386.319	1069.797	100.00		



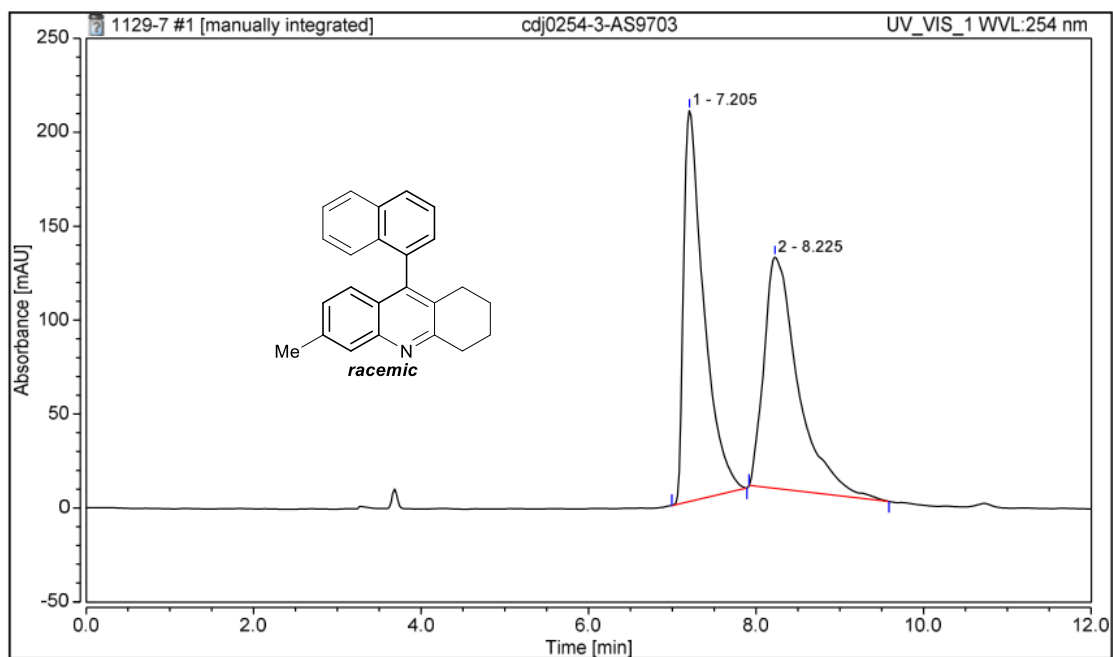
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.472	14.988	67.026	87.68	1.12	0.37
2	9.037	1.388	6.945	8.12	1.22	0.32
3	9.698	0.717	2.763	4.20	1.15	0.42
Total:		17.093	76.734	100.00		



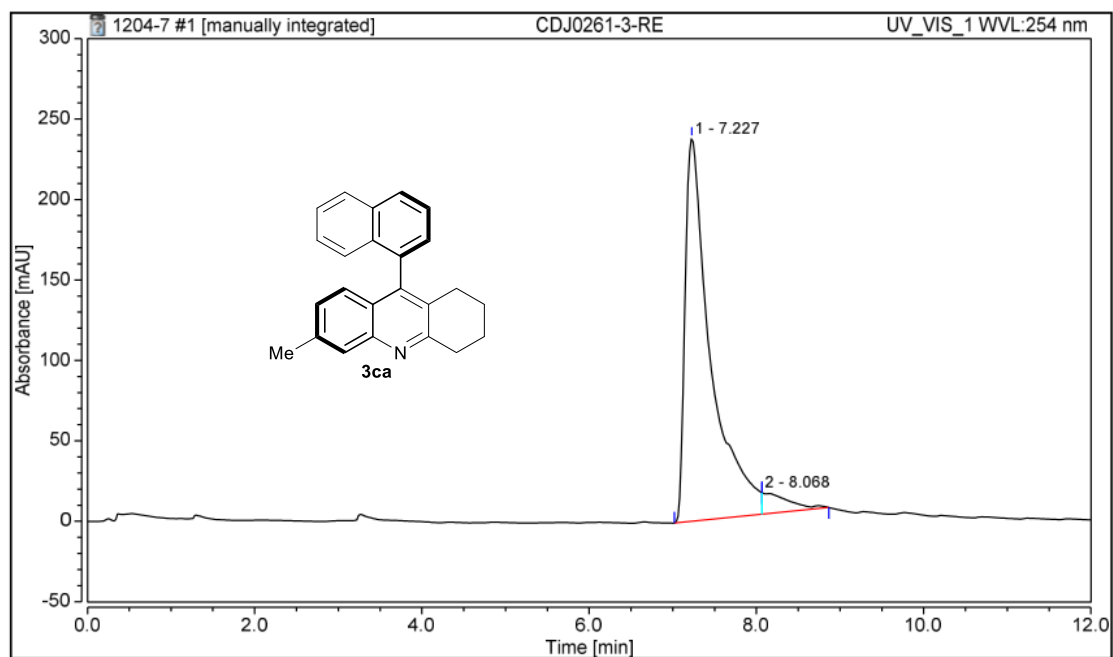
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.058	118.124	699.523	49.37	1.62	0.25
2	7.582	121.148	460.024	50.63	2.09	0.34
Total:		239.272	1159.548	100.00		



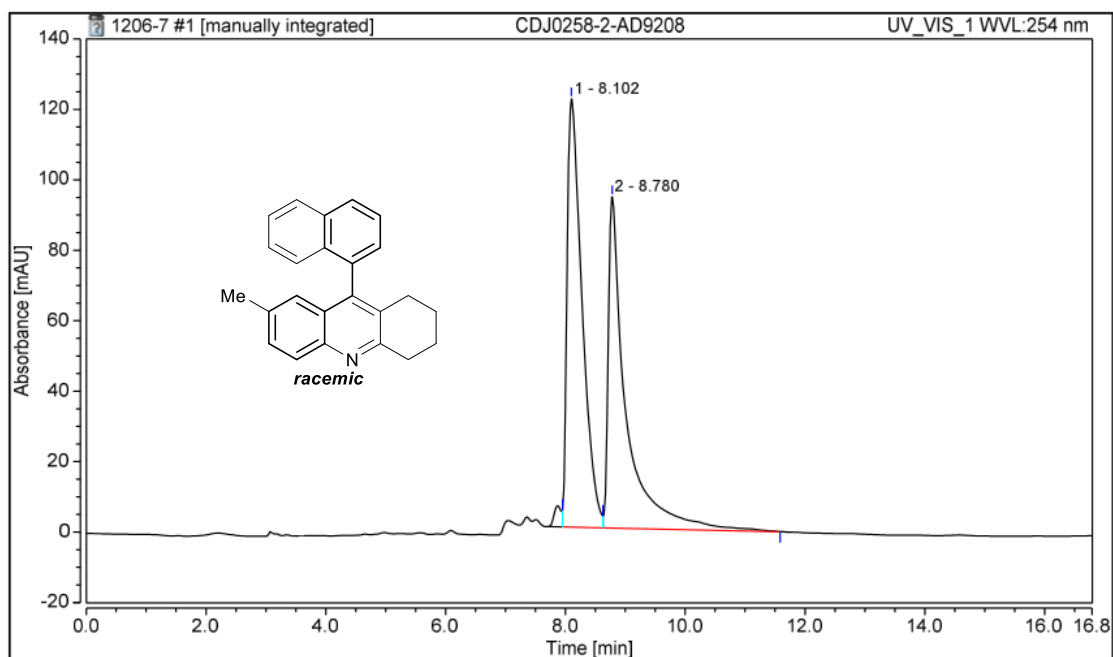
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.237	2.943	17.145	4.20	n.a.	0.25
2	7.677	67.049	222.881	95.80	2.75	0.37
Total:		69.992	240.026	100.00		



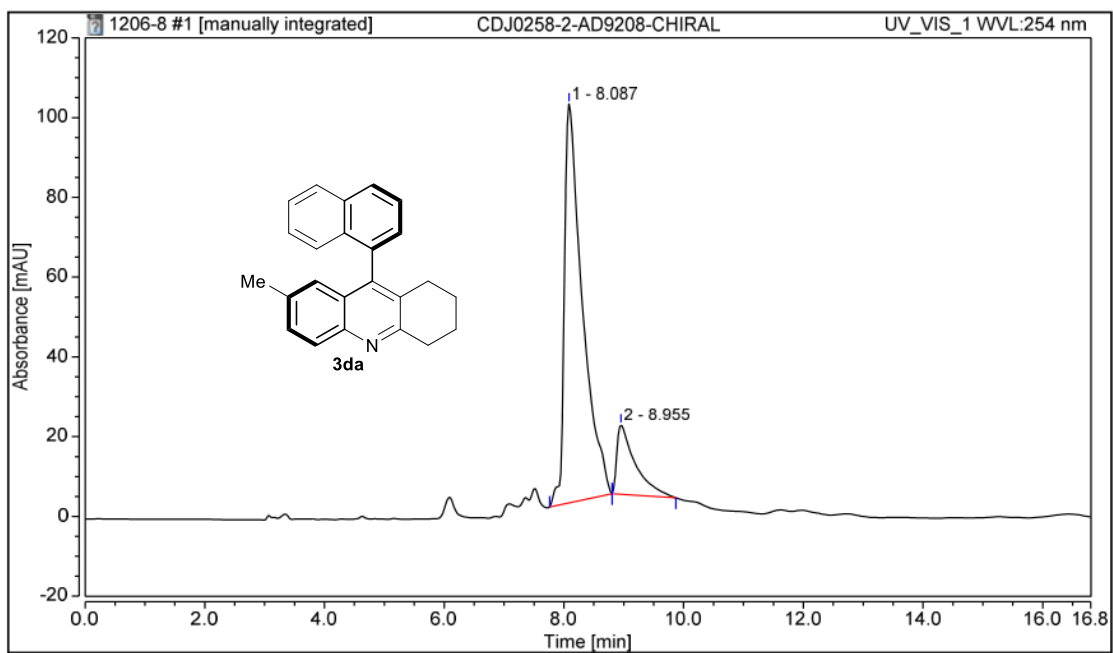
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.205	58.289	208.278	50.00	2.38	0.41
2	8.225	58.285	123.108	50.00	1.98	0.64
Total:		116.574	331.386	100.00		



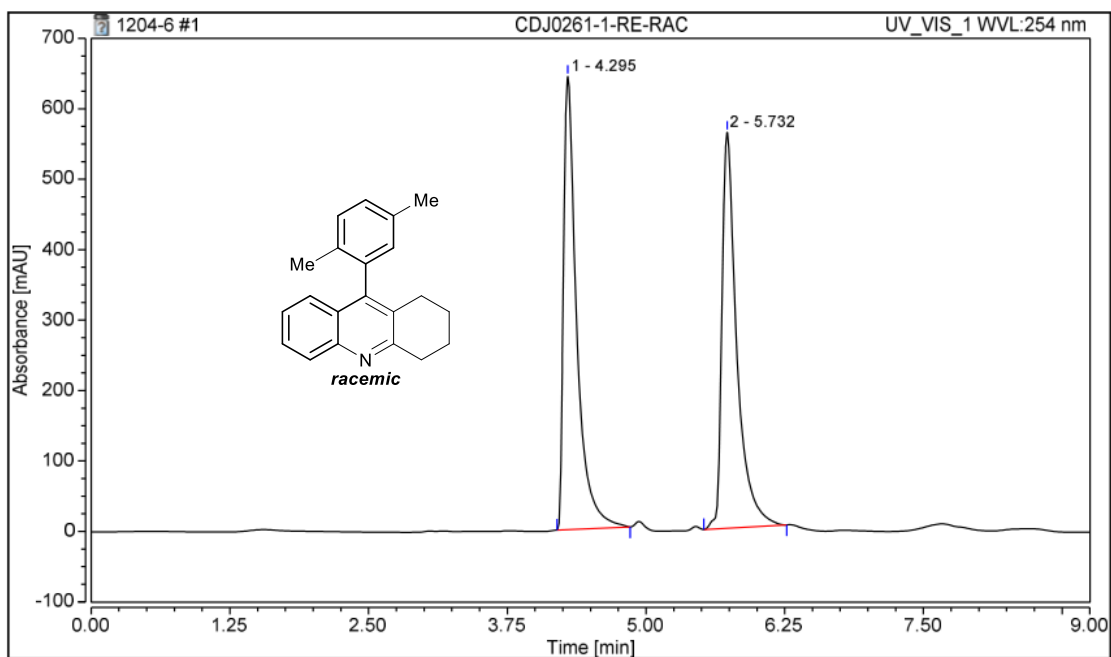
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.227	83.757	238.103	94.78	n.a.	0.45
2	8.068	4.615	13.263	5.22	n.a.	n.a.
Total:		88.372	251.366	100.00		



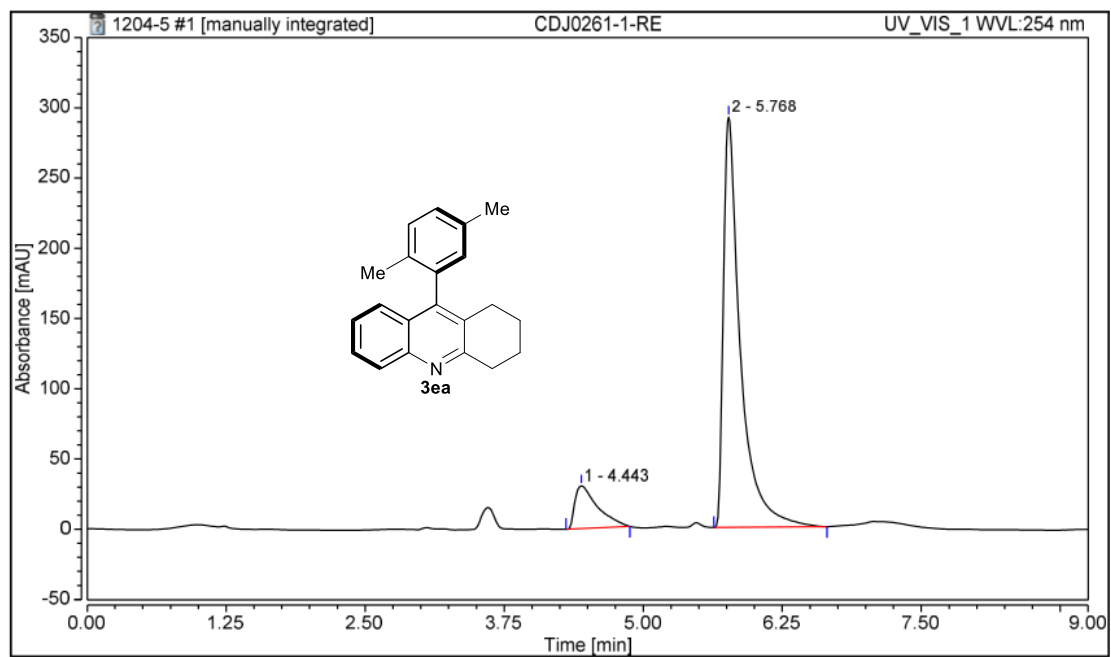
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.102	35.186	121.625	52.22	2.13	0.45
2	8.780	32.191	94.053	47.78	4.17	0.37
Total:		67.376	215.678	100.00		



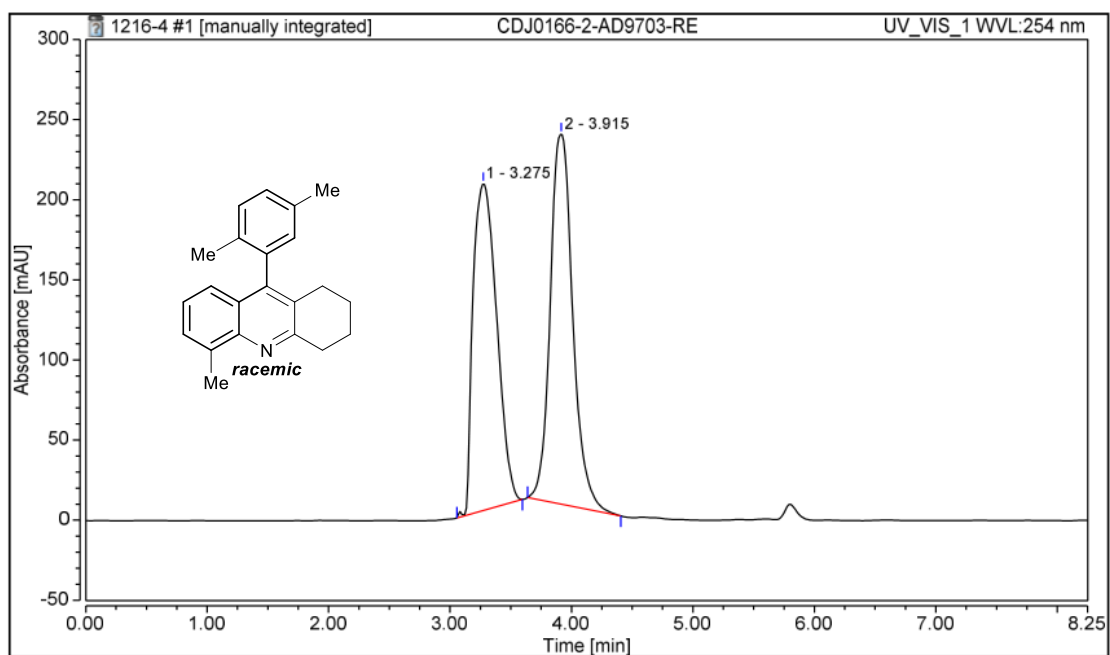
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.087	34.936	100.024	84.95	2.43	0.51
2	8.955	6.189	17.344	15.05	3.45	0.48
Total:		41.125	117.368	100.00		



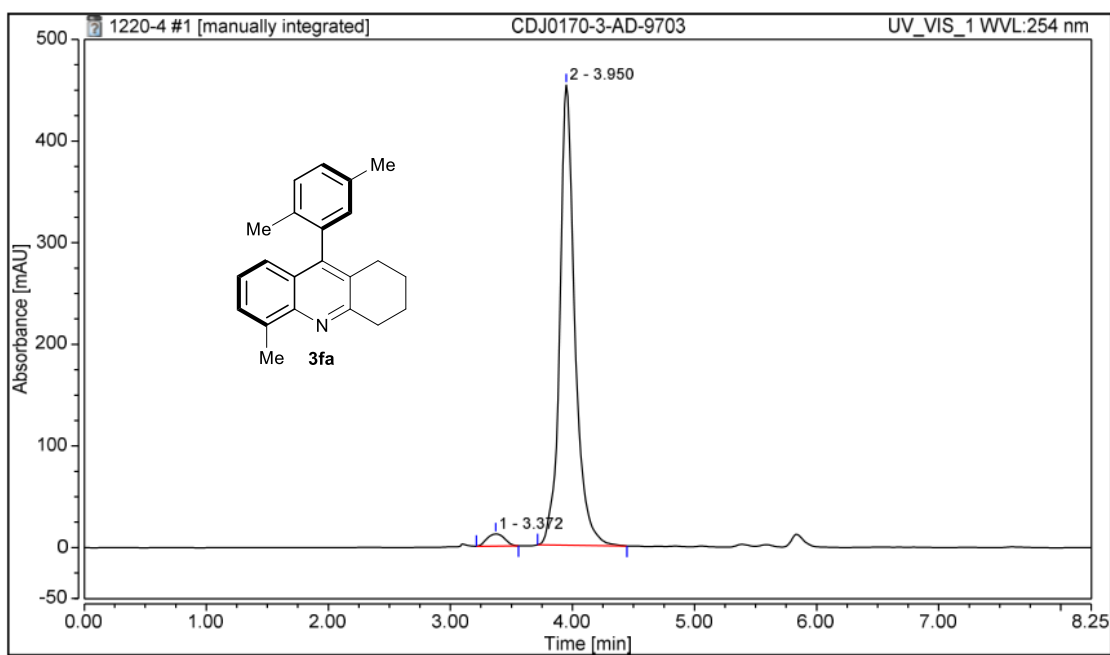
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.295	88.719	643.701	50.47	2.27	0.19
2	5.732	87.073	562.053	49.53	1.97	0.22
Total:		175.791	1205.754	100.00		



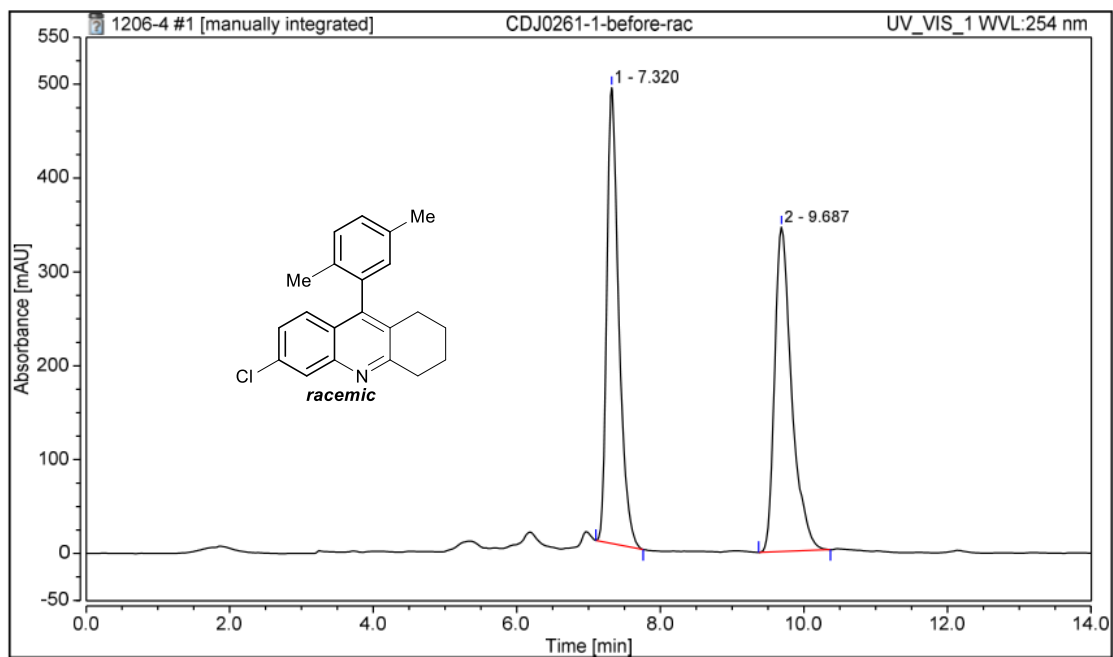
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.443	7.191	30.427	12.22	2.41	0.34
2	5.768	51.648	292.015	87.78	2.42	0.24
Total:		58.839	322.442	100.00		



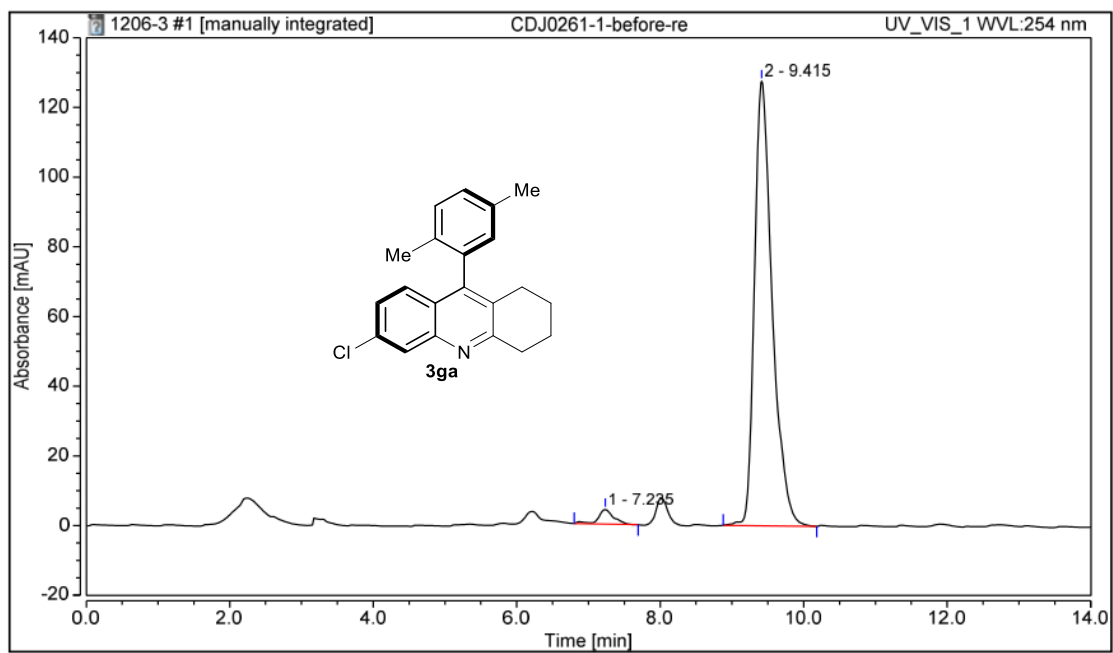
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	3.275	45.714	203.998	47.98	1.41	0.35
2	3.915	49.560	231.041	52.02	1.15	0.30
Total:		95.274	435.039	100.00		



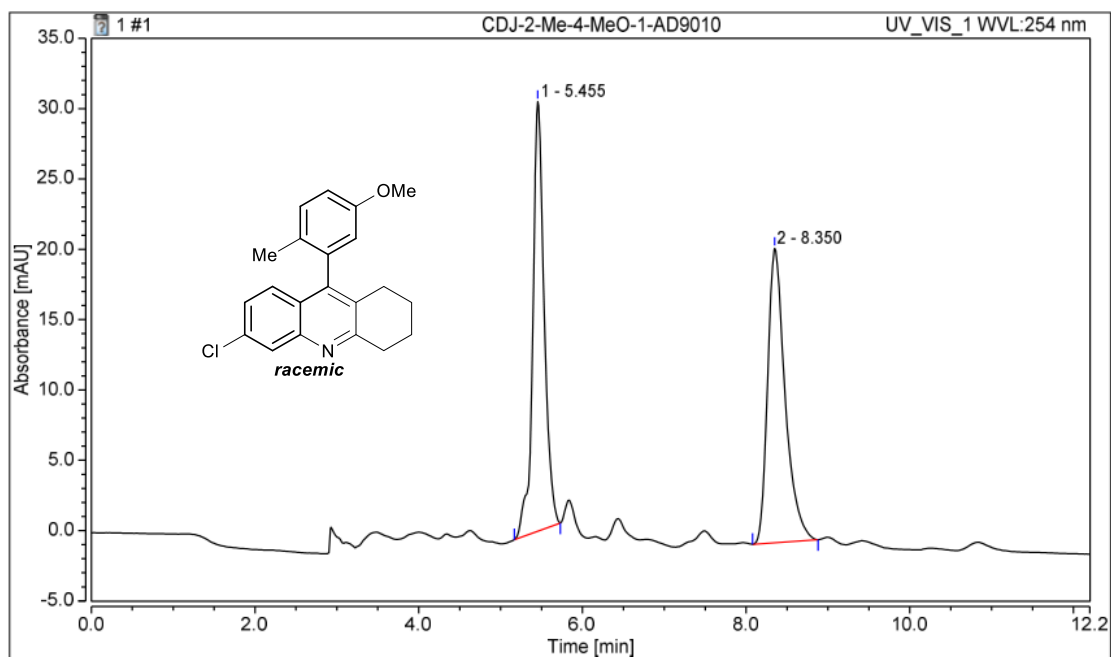
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	3.372	1.972	11.767	2.95	1.05	0.26
2	3.950	64.805	452.912	97.05	1.15	0.20
Total:		66.777	464.679	100.00		



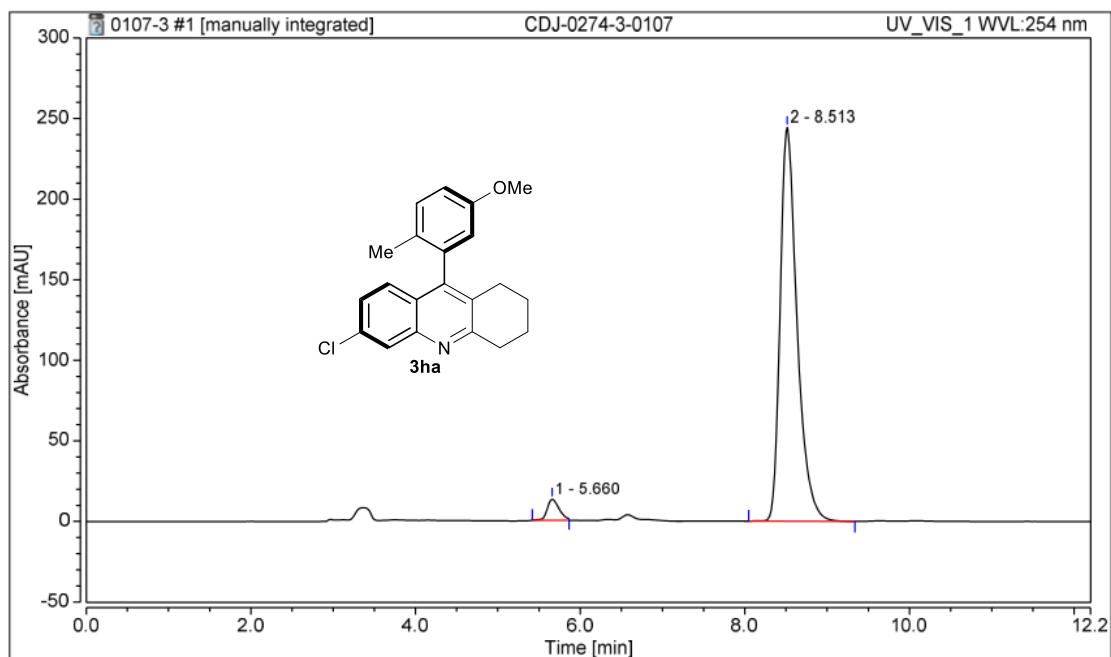
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.320	93.921	485.959	49.60	1.46	0.29
2	9.687	95.419	345.866	50.40	1.50	0.41
Total:		189.340	831.826	100.00		



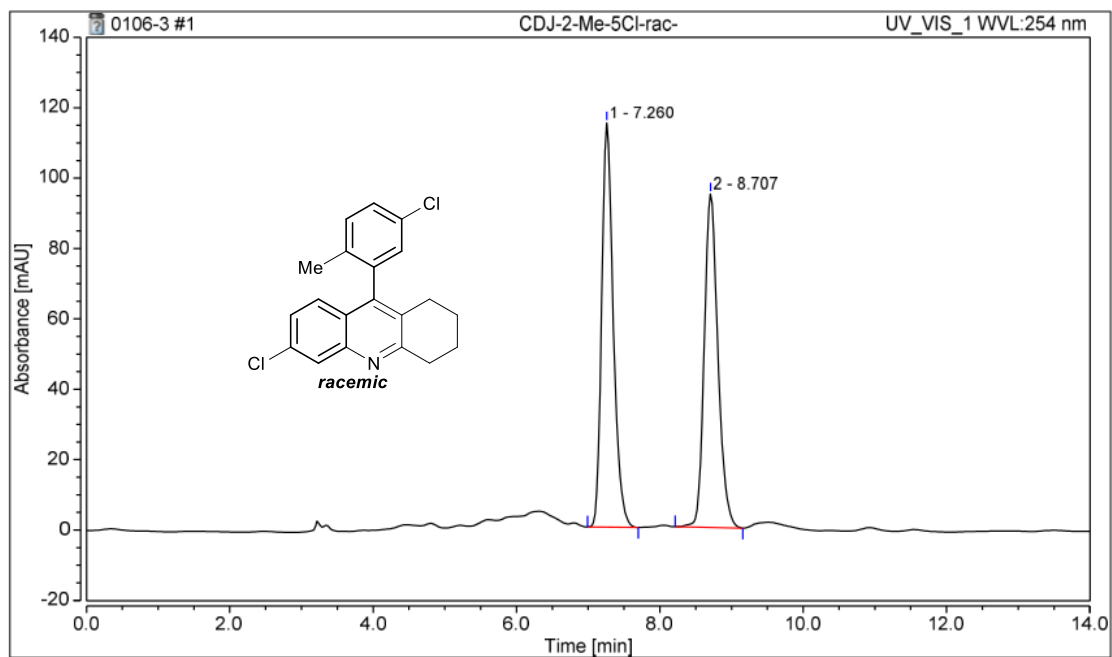
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.235	1.045	4.189	2.77	0.87	0.32
2	9.415	36.639	127.629	97.23	1.43	0.43
Total:		37.684	131.819	100.00		



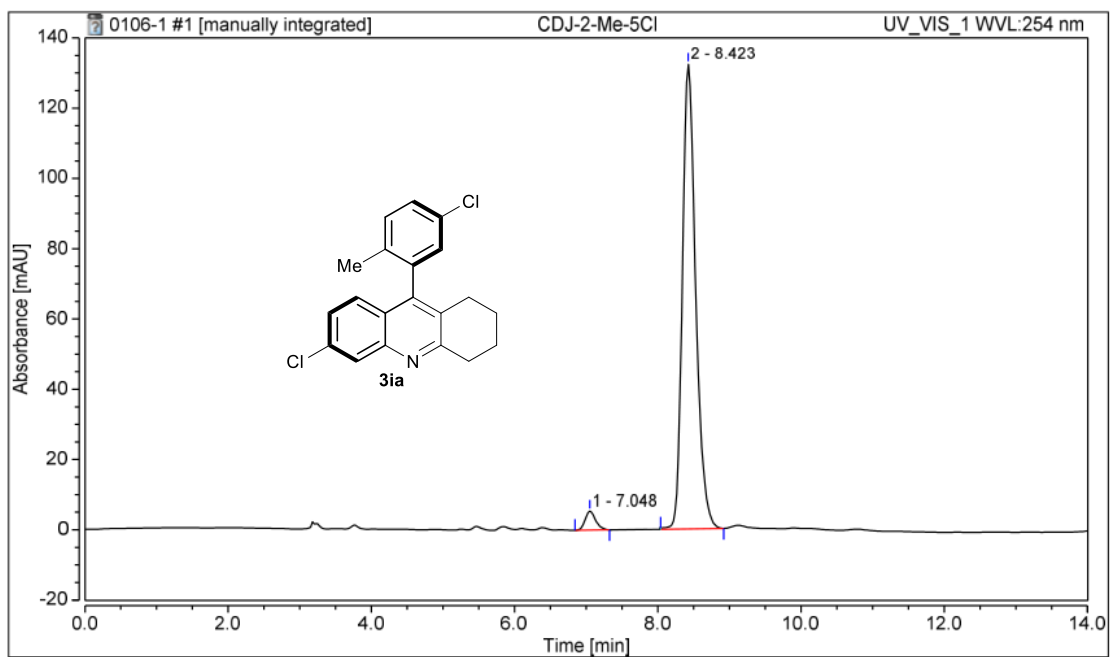
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.455	4.922	30.554	48.71	1.00	0.24
2	8.350	5.184	20.929	51.29	1.46	0.37
Total:		10.106	51.483	100.00		



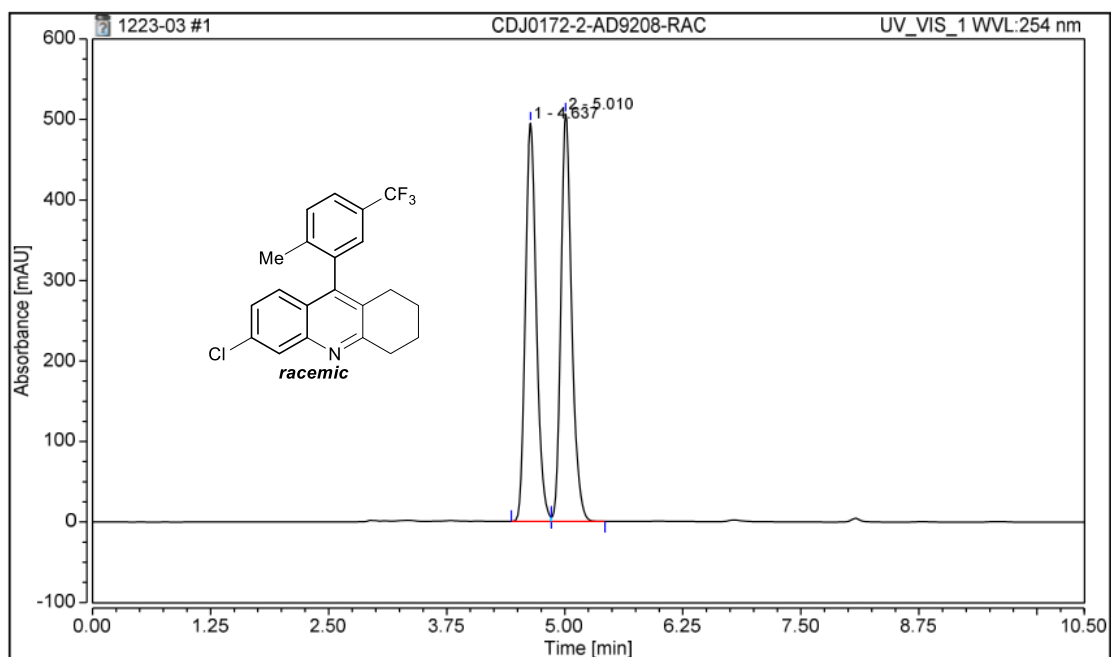
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.660	2.209	13.171	3.65	n.a.	0.26
2	8.513	58.265	244.437	96.35	1.43	0.36
Total:		60.474	257.608	100.00		



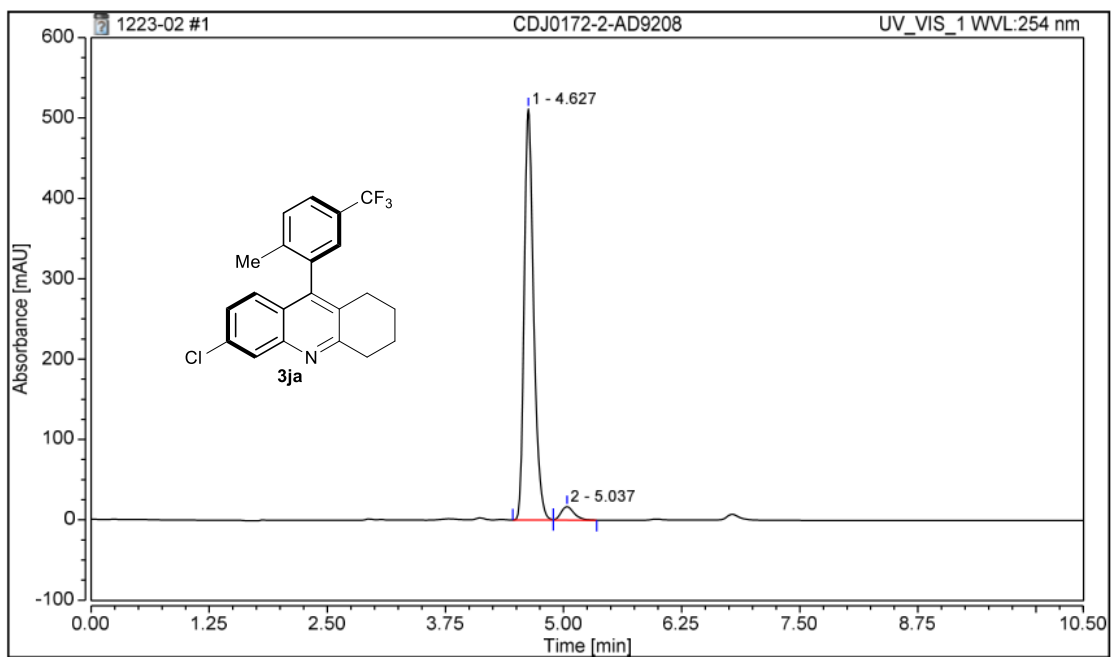
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.260	21.529	114.848	50.61	1.27	0.28
2	8.707	21.009	94.805	49.39	1.21	0.35
Total:		42.537	209.652	100.00		



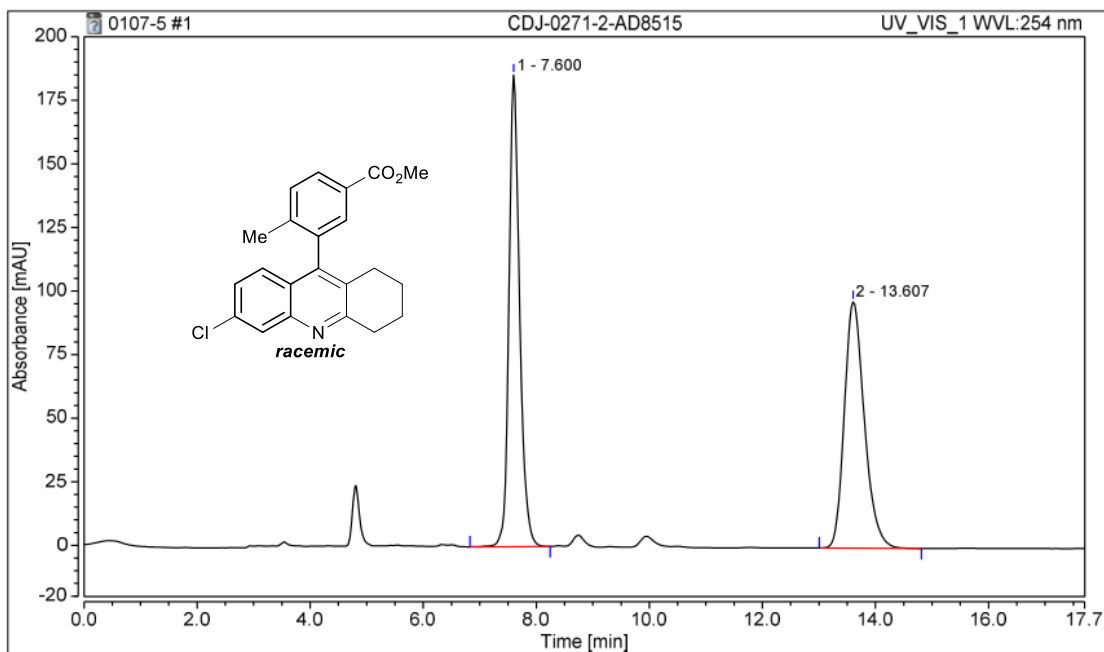
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.048	0.917	5.403	3.12	1.23	0.26
2	8.423	28.428	132.281	96.88	1.28	0.32
Total:		29.344	137.683	100.00		



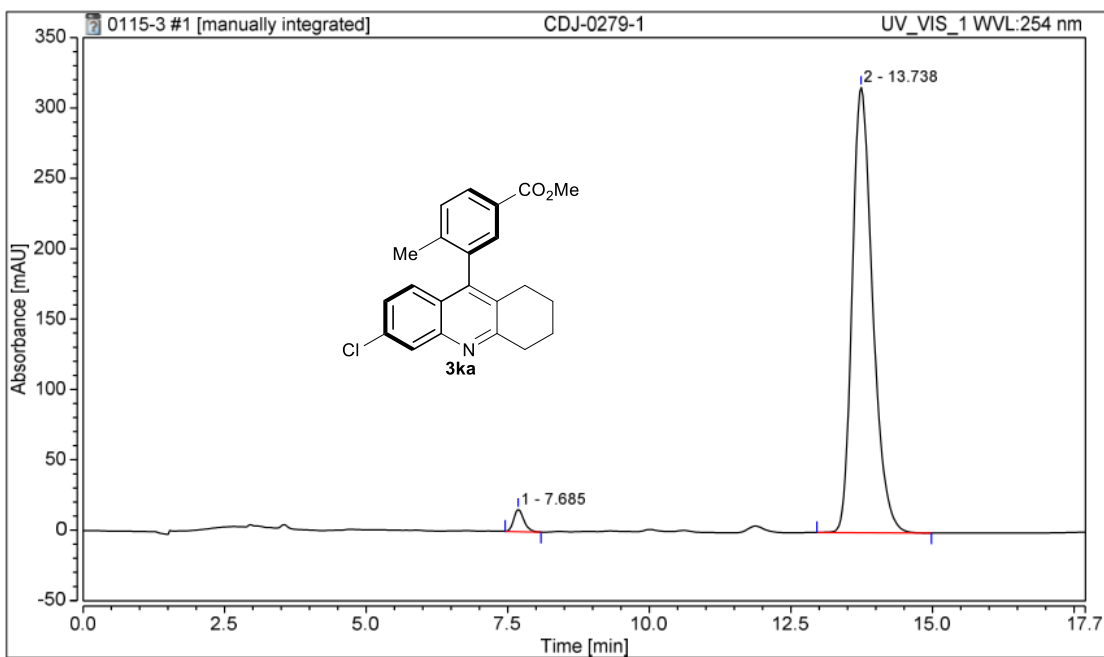
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.637	67.054	494.745	49.94	1.19	0.21
2	5.010	67.225	505.984	50.06	1.25	0.20
Total:		134.279	1000.729	100.00		



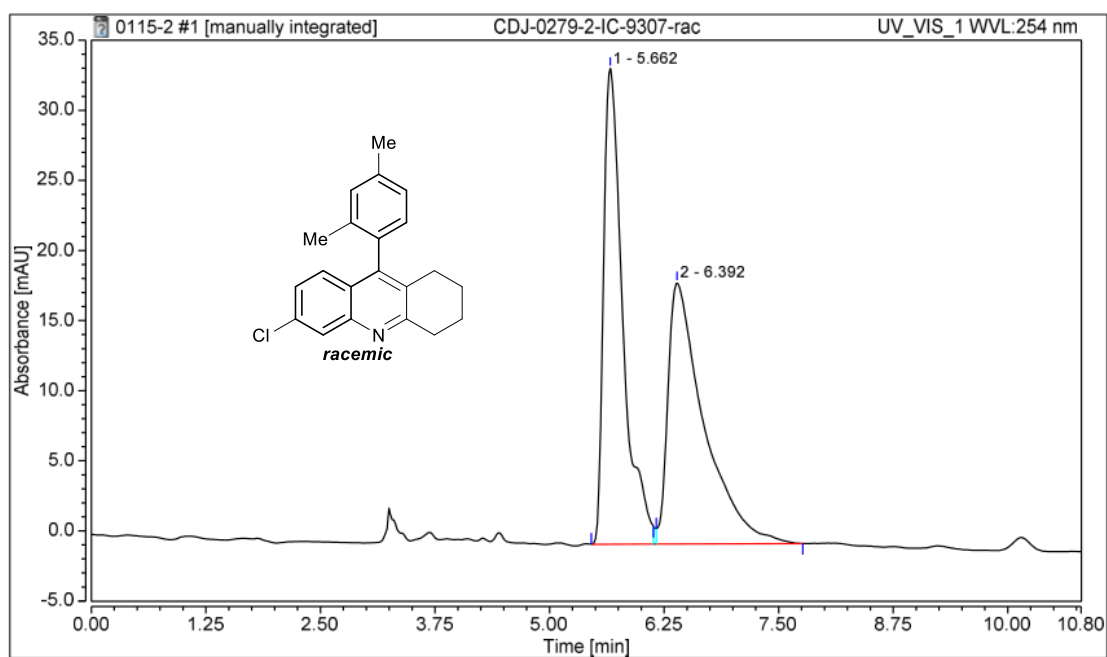
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.627	58.631	511.665	95.58	1.32	0.18
2	5.037	2.714	16.719	4.42	1.27	0.25
Total:		61.346	528.384	100.00		



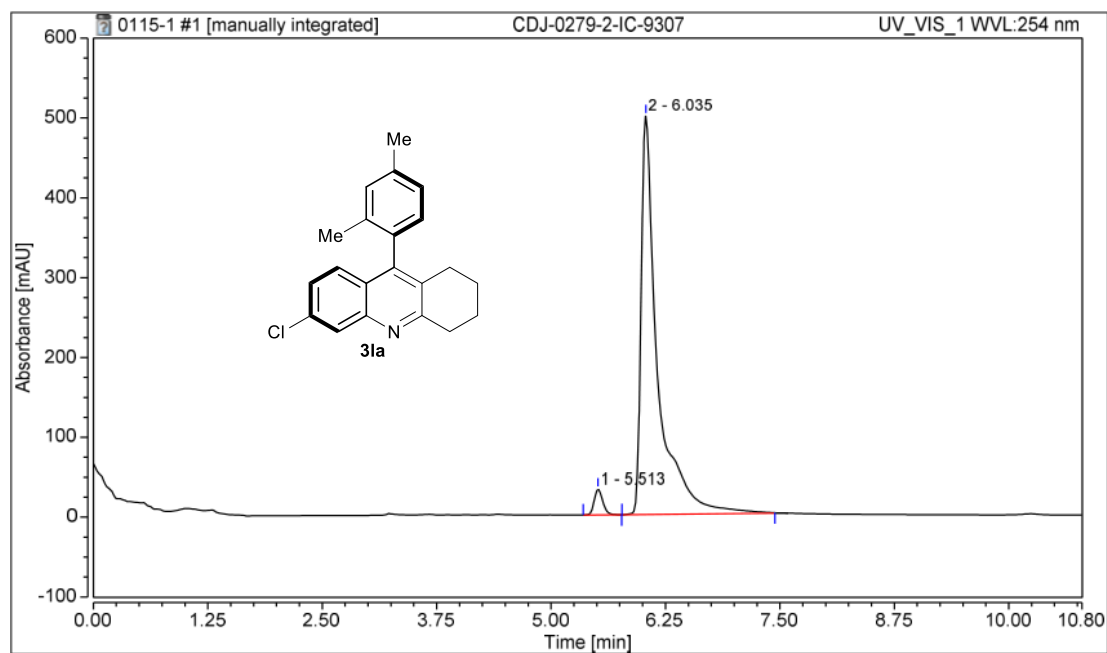
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.600	40.320	185.495	50.34	1.22	0.33
2	13.607	39.783	96.897	49.66	1.25	0.63
Total:		80.103	282.393	100.00		



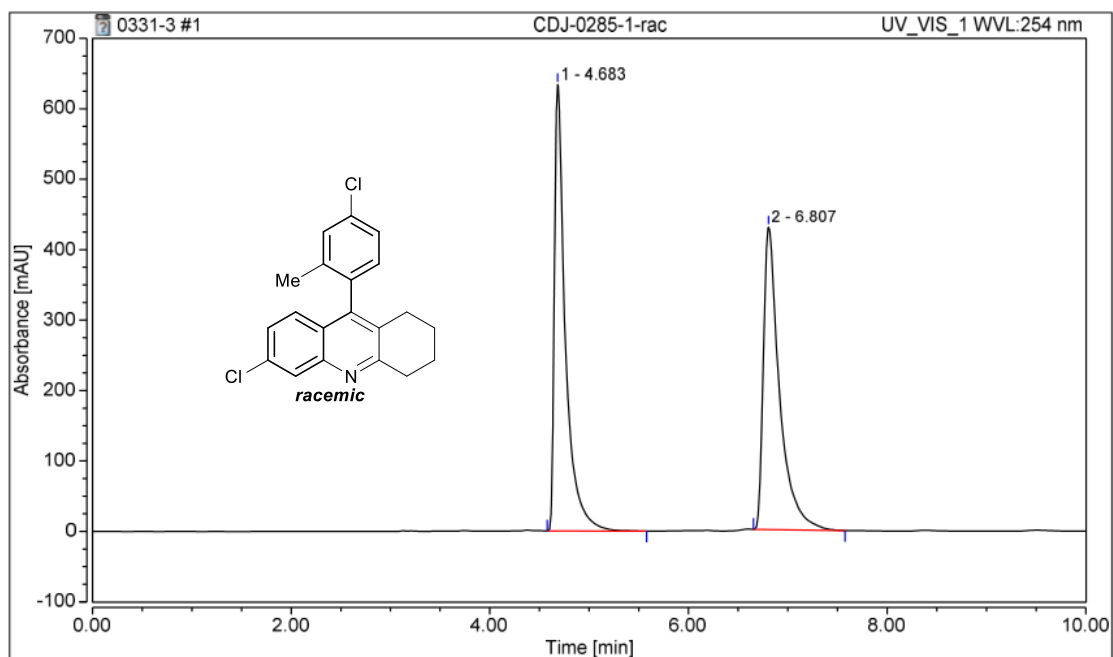
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	7.685	3.255	15.742	2.40	1.26	0.33
2	13.738	132.278	316.524	97.60	1.29	0.65
Total:		135.533	332.266	100.00		



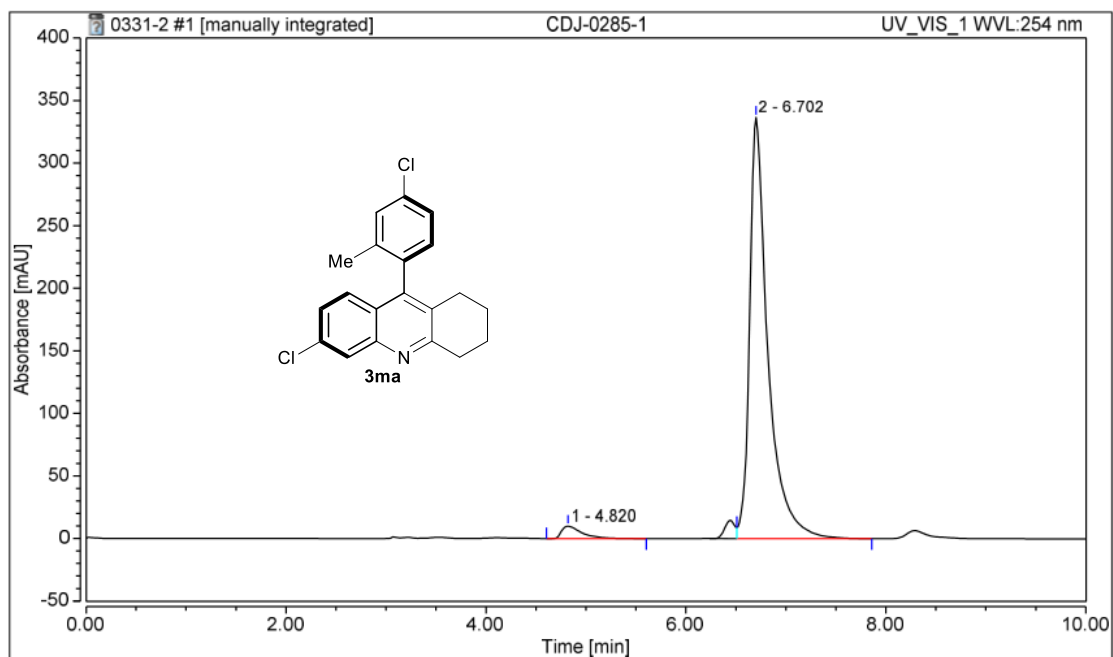
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.662	8.268	33.929	49.46	2.07	0.35
2	6.392	8.447	18.631	50.54	n.a.	0.63
Total:		16.715	52.560	100.00		



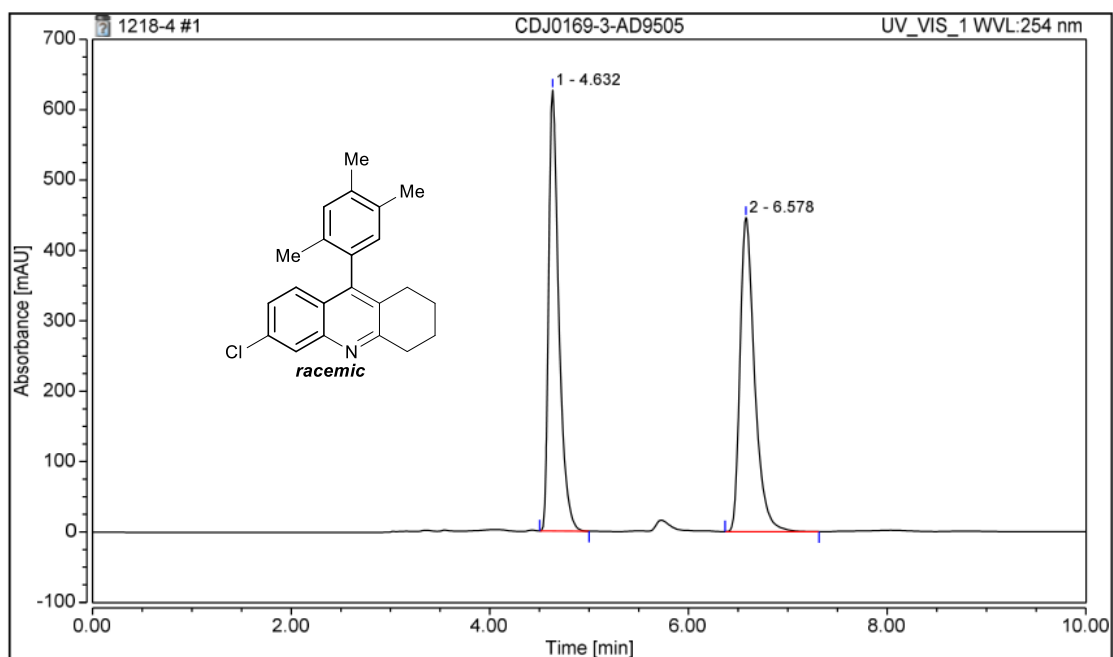
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.513	3.451	32.121	3.19	1.21	0.17
2	6.035	104.768	499.225	96.81	2.88	0.25
Total:		108.220	531.346	100.00		



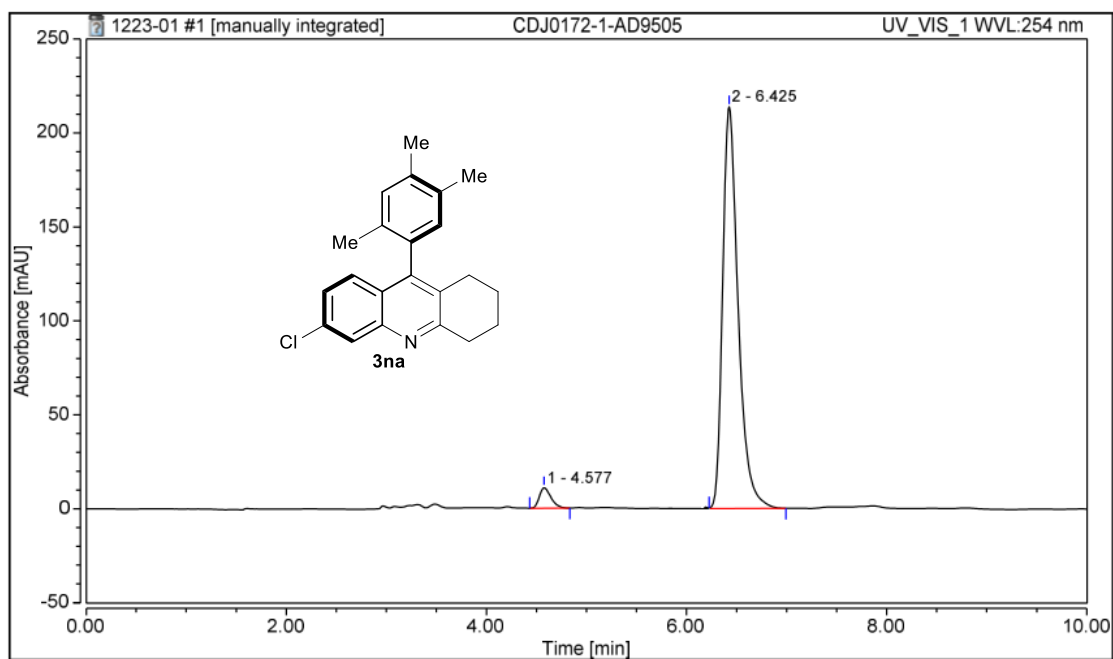
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.683	81.016	633.637	50.06	2.47	0.17
2	6.807	80.834	429.489	49.94	2.14	0.26
Total:		161.849	1063.126	100.00		



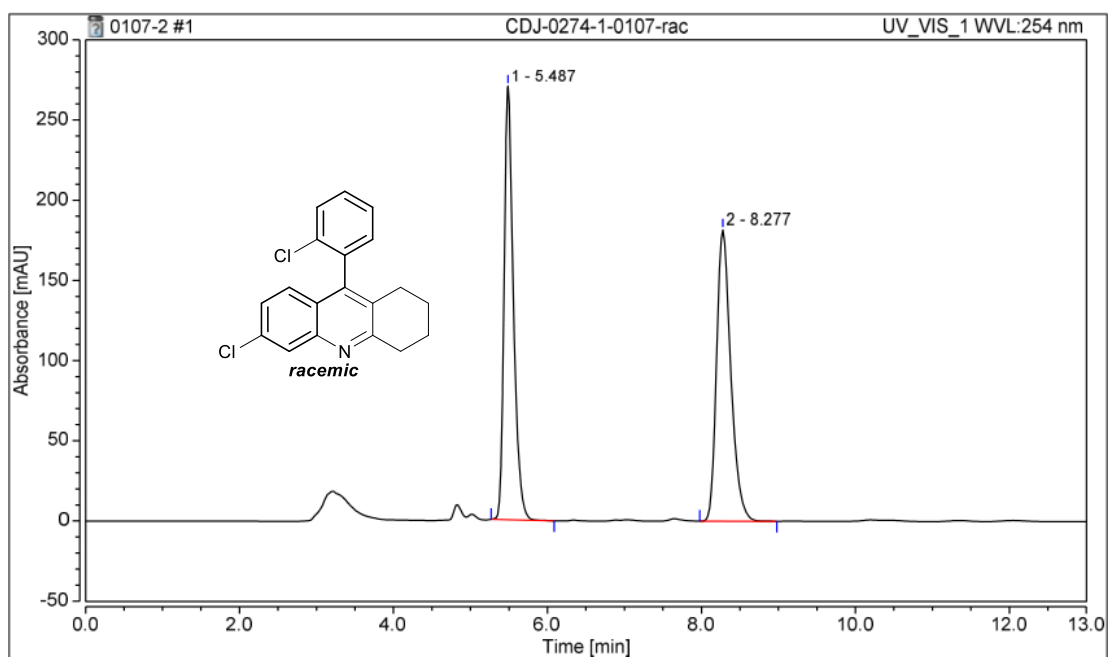
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.820	2.365	9.936	3.20	2.24	0.33
2	6.702	71.474	336.617	96.80	1.71	0.29
Total:		73.839	346.552	100.00		



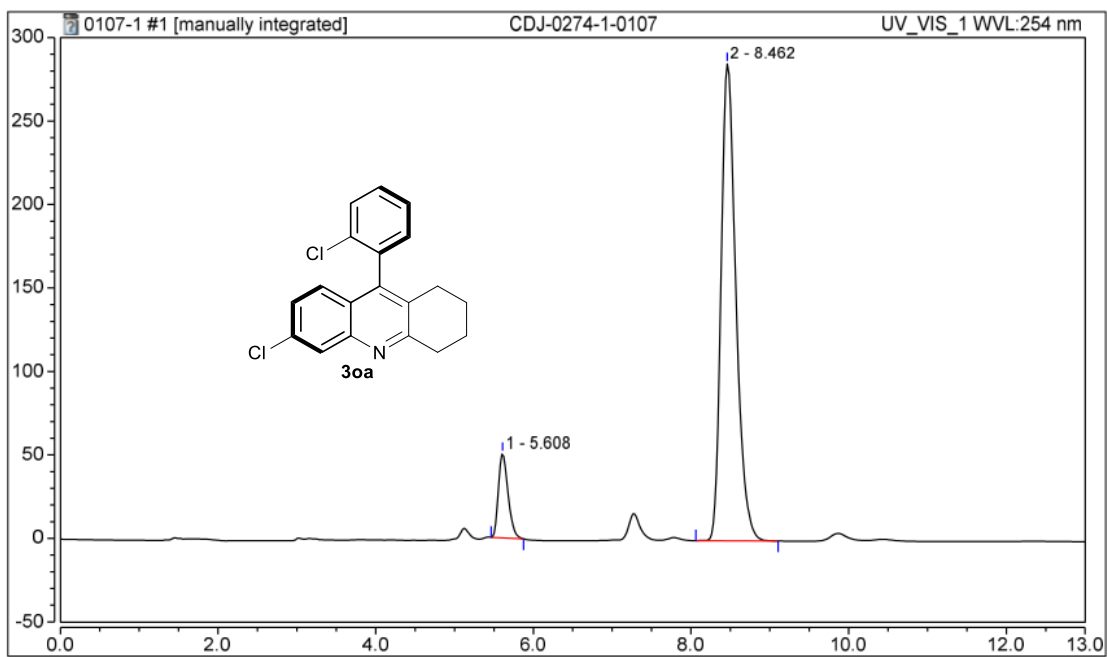
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.632	74.335	626.715	49.46	1.59	0.18
2	6.578	75.968	446.470	50.54	1.46	0.26
Total:		150.303	1073.184	100.00		



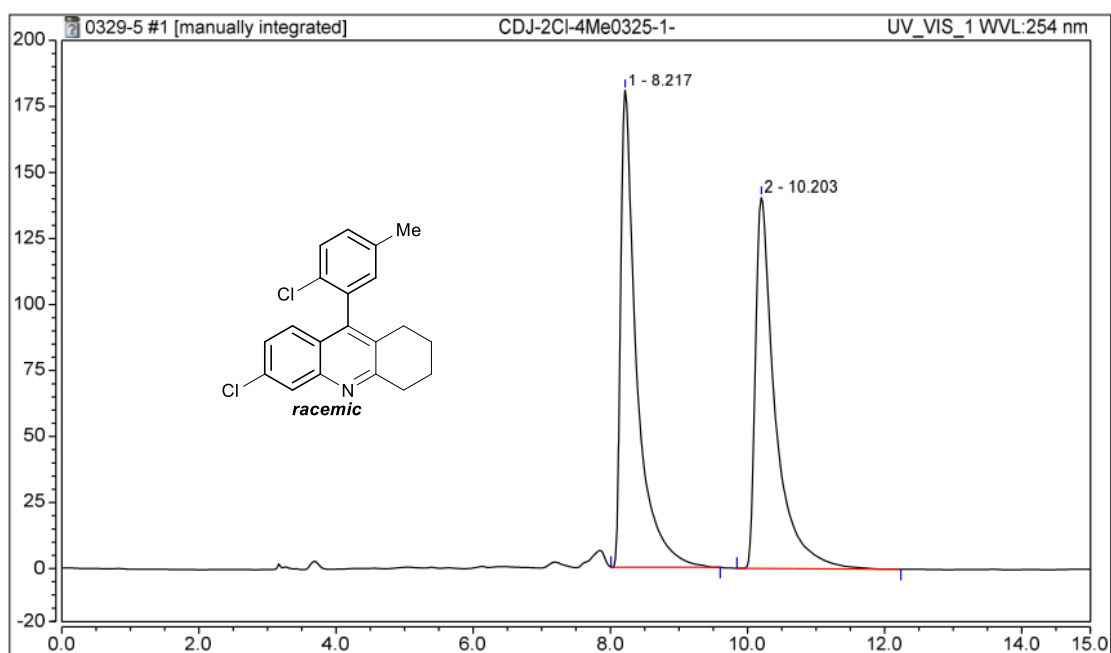
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	4.577	1.458	10.870	3.67	1.36	0.21
2	6.425	38.250	213.767	96.33	1.43	0.27
Total:		39.709	224.637	100.00		



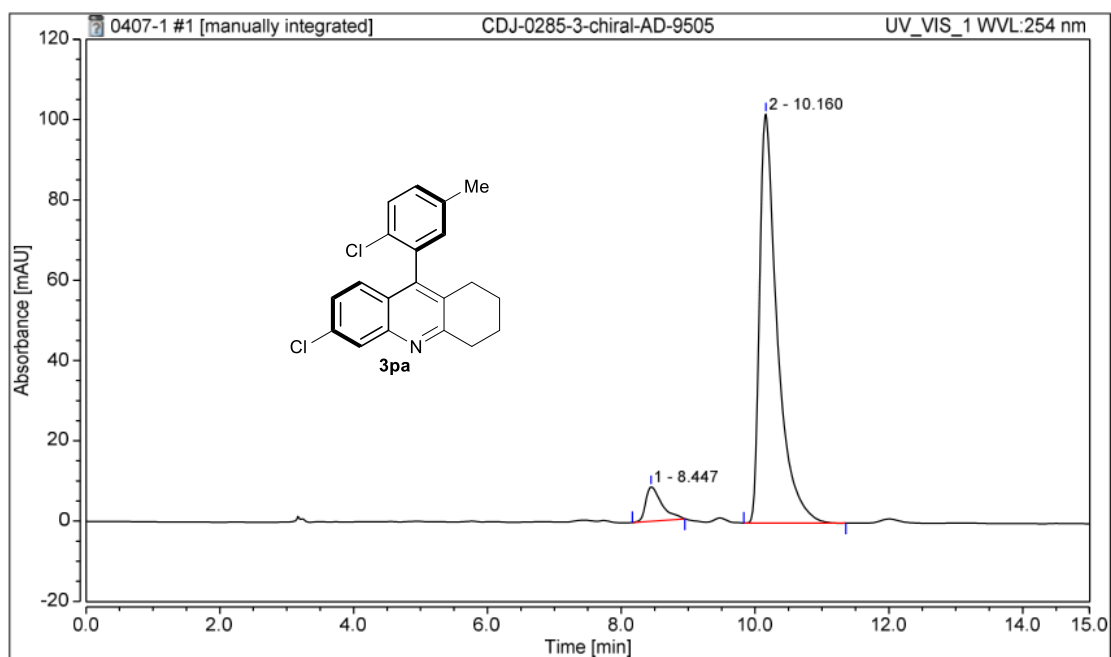
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.487	37.944	270.432	49.40	1.38	0.21
2	8.277	38.867	181.674	50.60	1.32	0.33
Total:		76.810	452.106	100.00		



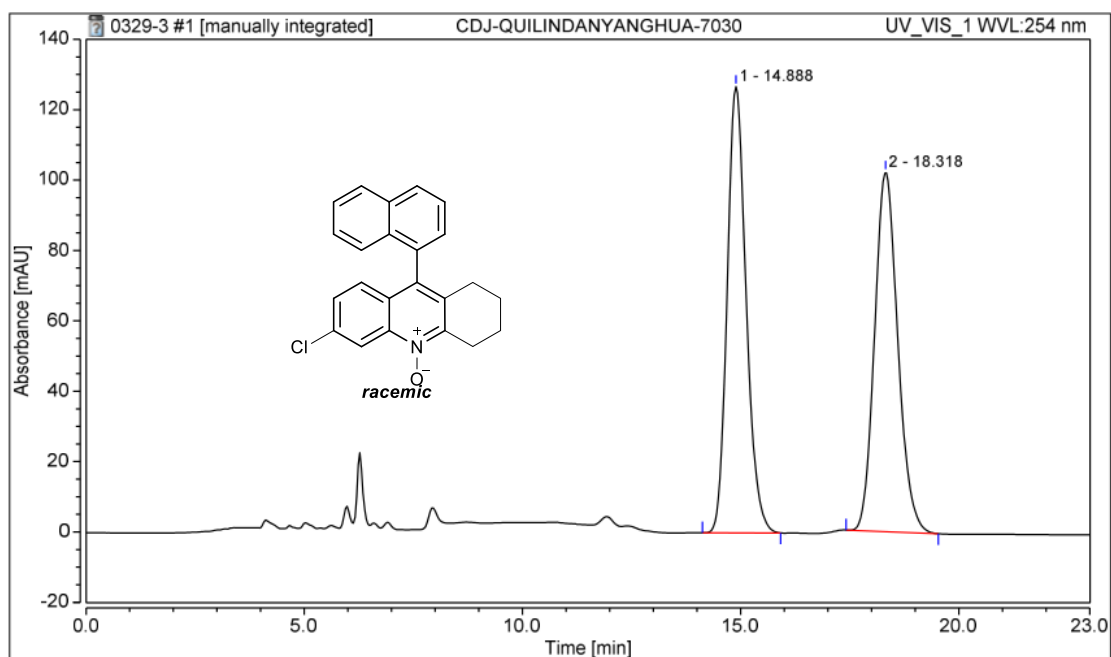
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	5.608	7.093	50.527	10.24	1.33	0.22
2	8.462	62.173	285.716	89.76	1.32	0.34
Total:		69.266	336.244	100.00		



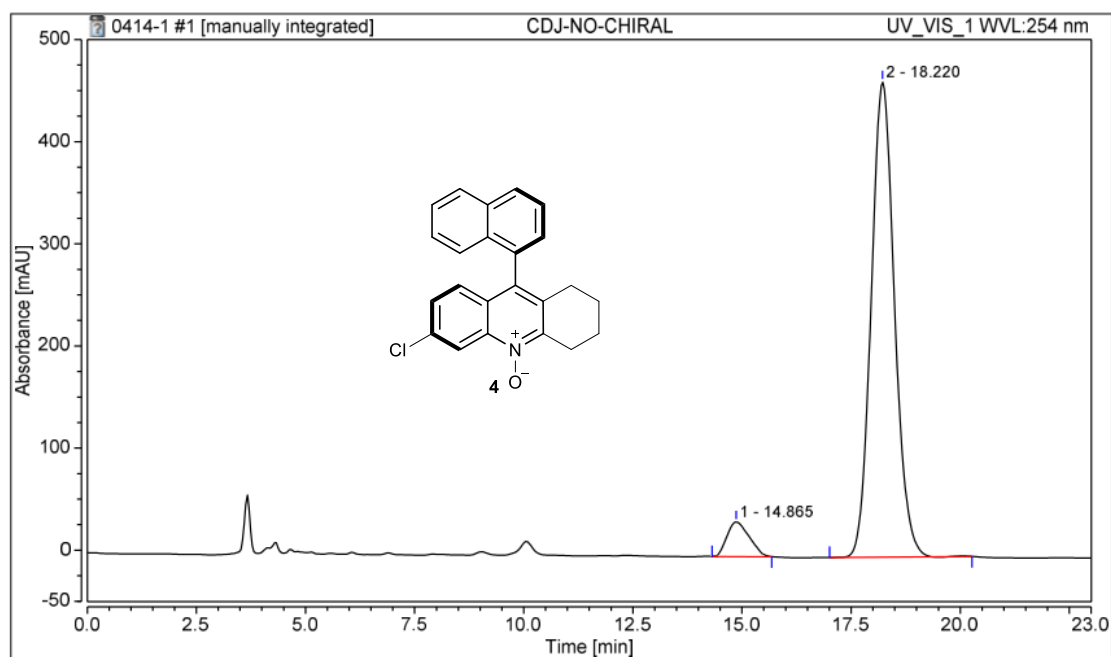
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.217	48.834	180.556	50.96	2.81	0.35
2	10.203	46.991	140.620	49.04	2.50	0.46
Total:		95.825	321.176	100.00		



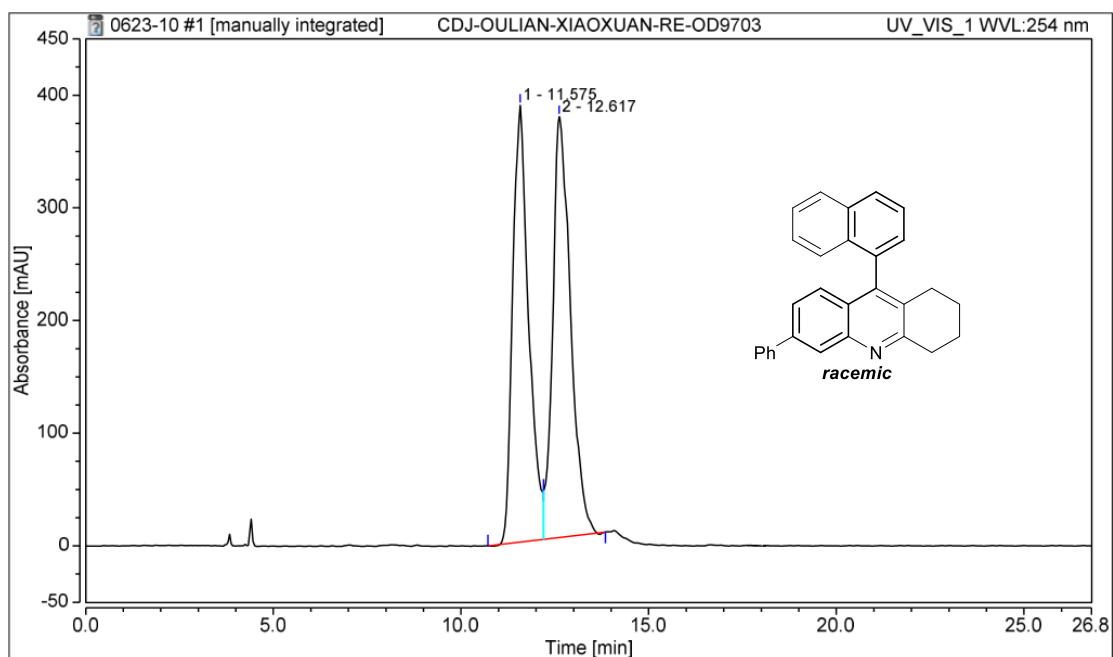
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	8.447	2.391	8.554	7.14	1.79	0.41
2	10.160	31.074	101.889	92.86	1.98	0.43
Total:		33.465	110.443	100.00		



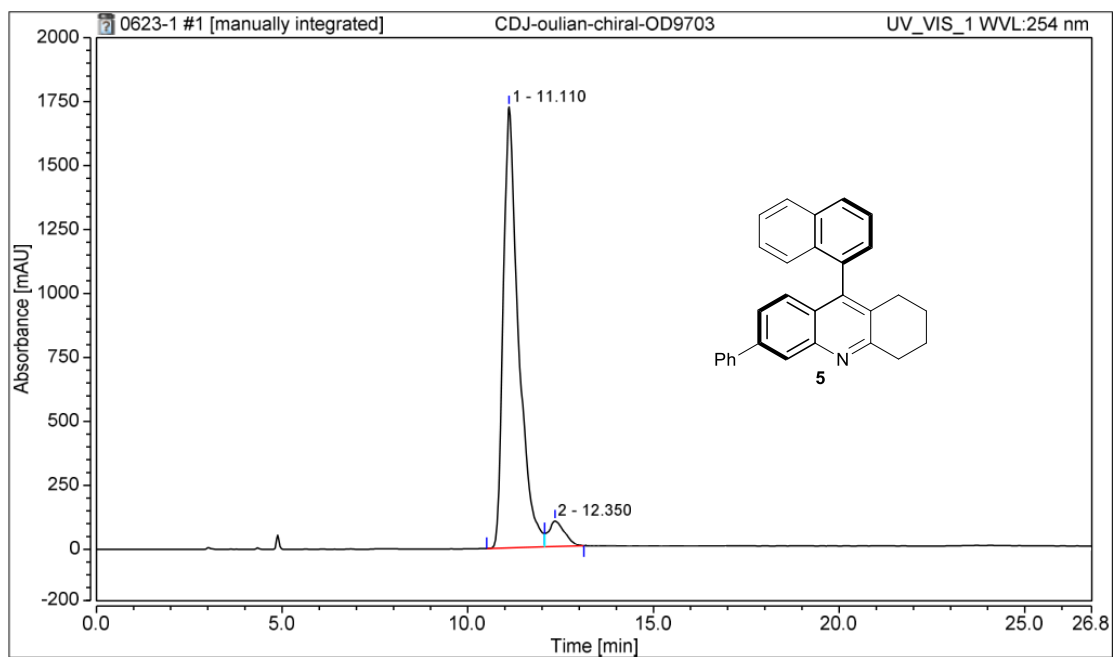
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	14.888	63.720	126.870	50.33	1.19	0.78
2	18.318	62.878	102.223	49.67	1.15	0.98
Total:		126.597	229.093	100.00		



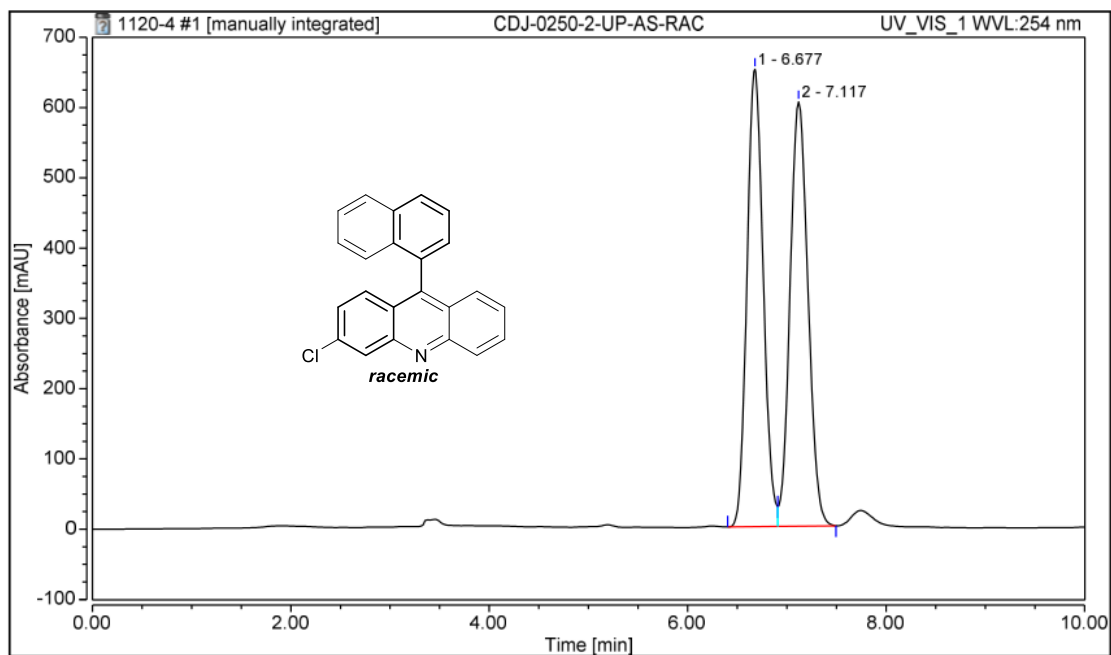
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	14.865	20.545	34.391	6.68	1.27	1.02
2	18.220	286.874	465.264	93.32	1.10	0.97
Total:		307.419	499.656	100.00		



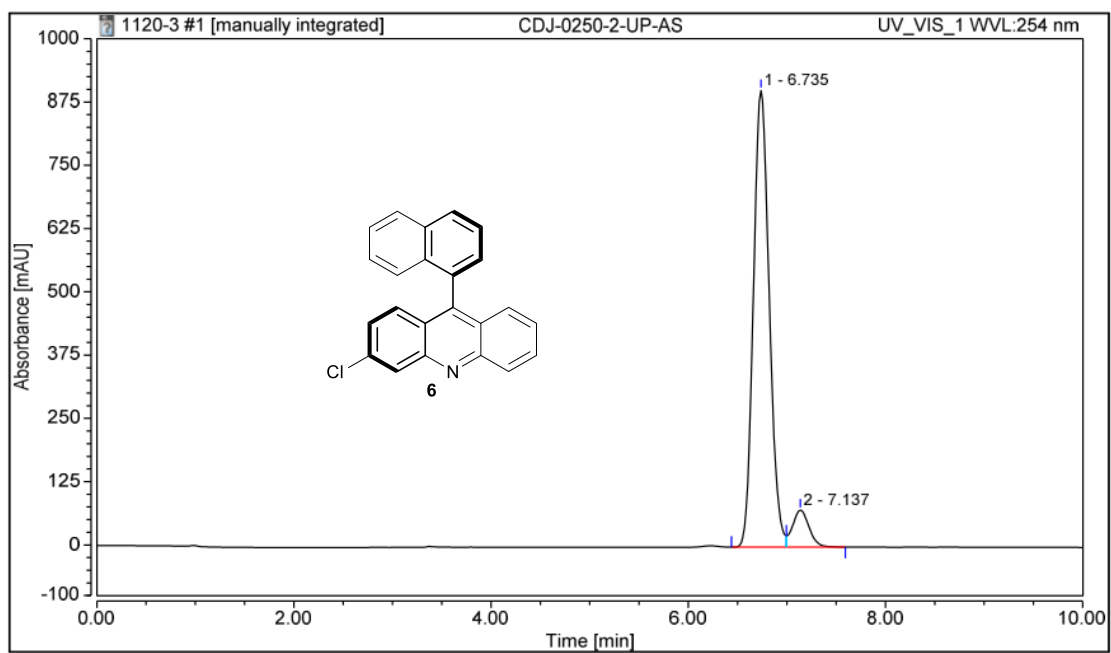
Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	11.575	191.004	387.659	48.17	n.a.	0.99
2	12.617	205.530	373.531	51.83	n.a.	0.87
Total:		396.534	761.190	100.00		



Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	11.110	820.365	1723.729	94.13	1.70	0.73
2	12.350	51.184	99.650	5.87	n.a.	0.76
Total:		871.549	1823.379	100.00		



Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	6.677	122.921	650.959	49.25	1.13	0.30
2	7.117	126.641	604.088	50.75	1.07	0.34
Total:		249.562	1255.046	100.00		



Integration Results						
No.	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Asymmetry (EP)	Peak Width min
1	6.735	167.938	902.390	92.18	1.14	0.30
2	7.137	14.248	73.226	7.82	n.a.	0.31
Total:		182.187	975.616	100.00		

X-ray Crystallographic Information

CCDC 1998526 (**3aa**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

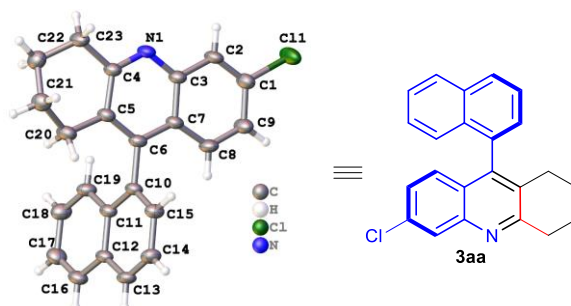


Table S4. Crystal data and structure refinement

Identification code	ndj-hz-3-300k	
Empirical formula	C ₂₃ H ₁₈ ClN	
Formula weight	343.83	
Temperature	299.61(10) K	
Wavelength	1.54184 Å	
Crystal system	Monoclinic	
Space group	P 1 21 1	
Unit cell dimensions	a = 9.4300(2) Å b = 8.4784(2) Å c = 11.9996(2) Å	α = 90°. β = 110.650(2)°. γ = 90°.
Volume	897.74(3) Å ³	
Z	2	
Density (calculated)	1.272 Mg/m ³	
Absorption coefficient	1.892 mm ⁻¹	
F(000)	360	
Crystal size	? x ? x ? mm ³	
Theta range for data collection	3.937 to 76.831°.	
Index ranges	-11 ≤ h ≤ 11, -10 ≤ k ≤ 10, -14 ≤ l ≤ 9	
Reflections collected	9452	
Independent reflections	3392 [R(int) = 0.0334]	
Completeness to theta = 67.684°	99.8%	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.70534	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	3392 / 31 / 245	
Goodness-of-fit on F ²	1.313	
Final R indices [I > 2σ(I)]	R1 = 0.0425, wR2 = 0.1525	
R indices (all data)	R1 = 0.0467, wR2 = 0.1547	
Absolute structure parameter	0.040(15)	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.247 and -0.216 e.Å ⁻³	

Table S5. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$). $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
Cl(1)	192(1)	4424(3)	1109(1)	102(1)
N(1)	5411(3)	4407(5)	4265(2)	54(1)
C(21)	8601(12)	3720(20)	7682(13)	70(4)
C(16)	3020(5)	6157(6)	9535(4)	62(1)
C(11)	3620(4)	5116(5)	7859(3)	49(1)
C(12)	3096(4)	4850(5)	8823(3)	50(1)
C(7)	3288(4)	4139(5)	4967(3)	50(1)
C(19)	4004(5)	6668(5)	7647(4)	58(1)
C(6)	4324(4)	4037(5)	6152(3)	50(1)
C(4)	6345(4)	4314(6)	5372(3)	52(1)
C(5)	5861(4)	4124(5)	6372(3)	52(1)
C(10)	3739(4)	3803(5)	7156(3)	50(1)
C(3)	3896(4)	4317(6)	4046(3)	52(1)
C(15)	3334(5)	2334(5)	7397(4)	58(1)
C(14)	2791(5)	2086(6)	8342(4)	61(1)
C(8)	1694(4)	4073(6)	4639(3)	63(1)
C(13)	2679(5)	3312(5)	9031(3)	57(1)
C(2)	2909(4)	4396(7)	2853(3)	60(1)
C(23)	8009(4)	4398(8)	5552(3)	71(1)
C(20)	7014(5)	4029(7)	7613(3)	70(1)
C(18)	3926(6)	7887(6)	8366(4)	67(1)
C(9)	754(4)	4168(8)	3477(3)	70(1)
C(17)	3441(6)	7621(6)	9325(4)	69(1)
C(1)	1393(4)	4318(7)	2590(3)	67(1)
C(22)	9006(18)	4740(20)	6813(9)	70(4)
C(22A)	9054(13)	3900(20)	6742(7)	76(4)
C(21A)	8568(10)	4620(20)	7704(9)	72(3)

Table S6. Bond lengths [Å] and angles [°].

Cl(1)-C(1)	1.740(4)
N(1)-C(4)	1.311(4)
N(1)-C(3)	1.361(4)
C(21)-C(20)	1.493(10)
C(21)-C(22)	1.506(12)
C(16)-C(12)	1.416(6)
C(16)-C(17)	1.354(7)
C(11)-C(12)	1.428(4)
C(11)-C(19)	1.412(6)
C(11)-C(10)	1.424(5)
C(12)-C(13)	1.409(6)
C(7)-C(6)	1.415(5)
C(7)-C(3)	1.422(4)
C(7)-C(8)	1.415(5)
C(19)-C(18)	1.365(6)
C(6)-C(5)	1.381(5)
C(6)-C(10)	1.505(4)
C(4)-C(5)	1.435(4)
C(4)-C(23)	1.509(5)
C(5)-C(20)	1.505(5)
C(10)-C(15)	1.363(6)
C(3)-C(2)	1.407(5)
C(15)-C(14)	1.415(5)
C(14)-C(13)	1.355(6)
C(8)-C(9)	1.365(6)
C(2)-C(1)	1.353(5)
C(23)-C(22)	1.501(10)
C(23)-C(22A)	1.483(9)
C(20)-C(21A)	1.518(9)
C(18)-C(17)	1.399(6)
C(9)-C(1)	1.402(5)
C(22A)-C(21A)	1.515(11)
C(4)-N(1)-C(3)	118.5(3)
C(20)-C(21)-C(22)	110.8(12)
C(17)-C(16)-C(12)	121.6(4)
C(19)-C(11)-C(12)	118.5(3)
C(19)-C(11)-C(10)	122.9(3)
C(10)-C(11)-C(12)	118.6(3)
C(16)-C(12)-C(11)	118.1(4)
C(13)-C(12)-C(16)	122.7(3)
C(13)-C(12)-C(11)	119.1(3)

C(6)-C(7)-C(3)	117.6(3)
C(8)-C(7)-C(6)	124.5(3)
C(8)-C(7)-C(3)	117.9(3)
C(18)-C(19)-C(11)	121.2(4)
C(7)-C(6)-C(10)	119.6(3)
C(5)-C(6)-C(7)	119.7(3)
C(5)-C(6)-C(10)	120.7(3)
N(1)-C(4)-C(5)	123.8(3)
N(1)-C(4)-C(23)	115.8(3)
C(5)-C(4)-C(23)	120.4(3)
C(6)-C(5)-C(4)	117.9(3)
C(6)-C(5)-C(20)	122.0(3)
C(4)-C(5)-C(20)	120.2(3)
C(11)-C(10)-C(6)	119.9(3)
C(15)-C(10)-C(11)	120.1(3)
C(15)-C(10)-C(6)	120.0(3)
N(1)-C(3)-C(7)	122.6(3)
N(1)-C(3)-C(2)	117.8(3)
C(2)-C(3)-C(7)	119.5(3)
C(10)-C(15)-C(14)	120.9(4)
C(13)-C(14)-C(15)	120.2(4)
C(9)-C(8)-C(7)	121.7(3)
C(14)-C(13)-C(12)	121.0(3)
C(1)-C(2)-C(3)	120.0(3)
C(22)-C(23)-C(4)	113.8(7)
C(22A)-C(23)-C(4)	115.4(6)
C(21)-C(20)-C(5)	115.1(6)
C(5)-C(20)-C(21A)	113.3(5)
C(19)-C(18)-C(17)	120.3(4)
C(8)-C(9)-C(1)	118.9(3)
C(16)-C(17)-C(18)	120.2(4)
C(2)-C(1)-C(1)	119.3(3)
C(2)-C(1)-C(9)	121.9(3)
C(9)-C(1)-C(1)	118.7(3)
C(23)-C(22)-C(21)	111.4(11)
C(23)-C(22A)-C(21A)	110.0(9)
C(22A)-C(21A)-C(20)	110.8(10)

Table S7. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$). The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2hka^* b^* U^{12}]$

	U^{11}	U^{22}	U^{33}	U^{23}	U^{13}	U^{12}
Cl(1)	57(1)	205(2)	43(1)	2(1)	15(1)	6(1)
N(1)	49(1)	84(2)	38(1)	4(2)	25(1)	4(2)
C(21)	61(7)	92(9)	54(6)	11(7)	19(5)	9(6)
C(16)	62(2)	88(3)	44(2)	-1(2)	30(2)	7(2)
C(11)	45(2)	71(2)	37(2)	5(1)	21(1)	6(2)
C(12)	43(2)	78(3)	36(2)	5(2)	21(1)	6(2)
C(7)	53(2)	68(2)	38(2)	1(2)	26(1)	3(2)
C(19)	61(2)	74(3)	50(2)	6(2)	35(2)	4(2)
C(6)	56(2)	64(2)	41(2)	3(2)	29(1)	4(2)
C(4)	49(2)	73(2)	41(2)	3(2)	24(1)	4(2)
C(5)	53(2)	71(2)	38(2)	3(2)	24(1)	6(2)
C(10)	49(2)	72(3)	35(2)	4(2)	23(1)	3(2)
C(3)	50(2)	73(2)	42(2)	2(2)	26(1)	4(2)
C(15)	64(2)	67(2)	51(2)	0(2)	33(2)	1(2)
C(14)	65(2)	71(3)	58(2)	11(2)	34(2)	0(2)
C(8)	53(2)	96(3)	49(2)	5(2)	31(2)	1(2)
C(13)	54(2)	84(3)	42(2)	12(2)	29(2)	4(2)
C(2)	56(2)	97(3)	36(2)	5(2)	26(1)	7(2)
C(23)	51(2)	120(4)	48(2)	6(3)	26(2)	3(3)
C(20)	61(2)	110(4)	42(2)	7(2)	24(2)	7(2)
C(18)	73(3)	70(3)	72(3)	-2(2)	41(2)	0(2)
C(9)	49(2)	119(4)	49(2)	3(3)	23(2)	5(3)
C(17)	73(3)	81(3)	61(2)	-13(2)	35(2)	4(2)
C(1)	54(2)	106(3)	42(2)	4(2)	18(1)	4(2)
C(22)	54(6)	87(8)	67(7)	4(6)	20(5)	-3(6)
C(22A)	56(5)	119(9)	58(5)	14(5)	26(4)	17(6)
C(21A)	60(5)	111(8)	43(4)	-1(5)	17(3)	-2(6)

Table S8. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$).

	x	y	z	U(eq)
H(21A)	8703	2616	7505	84
H(21B)	9294	3929	8483	84
H(16)	2672	6005	10163	74
H(19)	4316	6864	7006	69
H(15)	3416	1484	6933	69
H(14)	2510	1079	8490	74
H(8)	1275	3961	5230	75
H(13)	2322	3135	9650	68
H(2)	3300	4502	2245	72
H(23A)	8317	3402	5310	85
H(23B)	8161	5214	5038	85
H(23C)	8194	3742	4955	85
H(23D)	8251	5476	5414	85
H(20A)	6998	5015	8019	84
H(20B)	6712	3199	8040	84
H(20C)	7102	2941	7879	84
H(20D)	6657	4644	8144	84
H(18)	4197	8898	8217	81
H(9)	-292	4134	3277	85
H(17)	3407	8453	9820	82
H(22A)	8901	5841	6990	84
H(22B)	10056	4558	6901	84
H(22C)	10078	4232	6849	92
H(22D)	9048	2759	6802	92
H(21C)	8542	5764	7625	86
H(21D)	9300	4359	8480	86

Table S9. Torsion angles [°].

N(1)-C(4)-C(5)-C(6)	0.0(7)
N(1)-C(4)-C(5)-C(20)	-179.9(4)
N(1)-C(4)-C(23)-C(22)	163.8(9)
N(1)-C(4)-C(23)-C(22A)	-165.4(9)
N(1)-C(3)-C(2)-C(1)	-179.7(5)
C(16)-C(12)-C(13)-C(14)	-178.9(4)
C(11)-C(12)-C(13)-C(14)	1.0(6)
C(11)-C(19)-C(18)-C(17)	-0.8(7)
C(11)-C(10)-C(15)-C(14)	0.0(6)
C(12)-C(16)-C(17)-C(18)	1.8(7)
C(12)-C(11)-C(19)-C(18)	2.1(6)
C(12)-C(11)-C(10)-C(6)	-178.5(3)
C(12)-C(11)-C(10)-C(15)	1.1(5)
C(7)-C(6)-C(5)-C(4)	-0.1(6)
C(7)-C(6)-C(5)-C(20)	179.8(4)
C(7)-C(6)-C(10)-C(11)	-99.9(4)
C(7)-C(6)-C(10)-C(15)	80.5(5)
C(7)-C(3)-C(2)-C(1)	0.6(8)
C(7)-C(8)-C(9)-C(1)	0.7(8)
C(19)-C(11)-C(12)-C(16)	-1.4(5)
C(19)-C(11)-C(12)-C(13)	178.7(4)
C(19)-C(11)-C(10)-C(6)	1.2(6)
C(19)-C(11)-C(10)-C(15)	-179.2(4)
C(19)-C(18)-C(17)-C(16)	-1.1(7)
C(6)-C(7)-C(3)-N(1)	-0.7(6)
C(6)-C(7)-C(3)-C(2)	178.9(4)
C(6)-C(7)-C(8)-C(9)	-179.6(5)
C(6)-C(5)-C(20)-C(21)	164.6(9)
C(6)-C(5)-C(20)-C(21A)	-162.9(7)
C(6)-C(10)-C(15)-C(14)	179.6(4)
C(4)-N(1)-C(3)-C(7)	0.5(7)
C(4)-N(1)-C(3)-C(2)	-179.1(5)
C(4)-C(5)-C(20)-C(21)	-15.5(11)
C(4)-C(5)-C(20)-C(21A)	17.0(9)
C(4)-C(23)-C(22)-C(21)	47.0(18)
C(4)-C(23)-C(22A)-C(21A)	-45.3(16)
C(5)-C(6)-C(10)-C(11)	81.3(5)
C(5)-C(6)-C(10)-C(15)	-98.3(5)
C(5)-C(4)-C(23)-C(22)	-16.8(10)
C(5)-C(4)-C(23)-C(22A)	14.0(10)
C(5)-C(20)-C(21A)-C(22A)	-48.8(14)
C(10)-C(11)-C(12)-C(16)	178.3(3)

C(10)-C(11)-C(12)-C(13)	-1.6(5)
C(10)-C(11)-C(19)-C(18)	-177.6(4)
C(10)-C(6)-C(5)-C(4)	178.6(4)
C(10)-C(6)-C(5)-C(20)	-1.5(7)
C(10)-C(15)-C(14)-C(13)	-0.6(7)
C(3)-N(1)-C(4)-C(5)	-0.1(7)
C(3)-N(1)-C(4)-C(23)	179.2(4)
C(3)-C(7)-C(6)-C(5)	0.5(6)
C(3)-C(7)-C(6)-C(10)	-178.3(4)
C(3)-C(7)-C(8)-C(9)	0.3(7)
C(3)-C(2)-C(1)-Cl(1)	179.8(4)
C(3)-C(2)-C(1)-C(9)	0.4(9)
C(15)-C(14)-C(13)-C(12)	0.1(6)
C(8)-C(7)-C(6)-C(5)	-179.6(4)
C(8)-C(7)-C(6)-C(10)	1.6(6)
C(8)-C(7)-C(3)-N(1)	179.4(4)
C(8)-C(7)-C(3)-C(2)	-1.0(7)
C(8)-C(9)-C(1)-Cl(1)	179.5(5)
C(8)-C(9)-C(1)-C(2)	-1.1(9)
C(23)-C(4)-C(5)-C(6)	-179.3(4)
C(23)-C(4)-C(5)-C(20)	0.8(7)
C(23)-C(22A)-C(21A)-C(20)	63.5(18)
C(20)-C(21)-C(22)-C(23)	-62(2)
C(17)-C(16)-C(12)-C(11)	-0.6(6)
C(17)-C(16)-C(12)-C(13)	179.3(4)
C(22)-C(21)-C(20)-C(5)	45.3(17)

CCDC 1998519 (**3ac**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

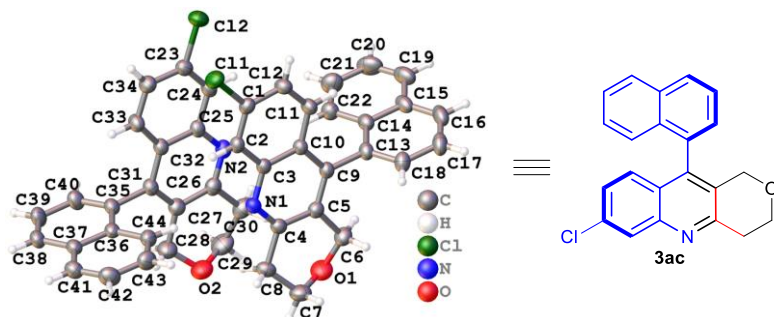


Table S10. Crystal data and structure refinement

Identification code	ndj-hz-2-300k	
Empirical formula	C ₂₂ H ₁₆ ClNO	
Formula weight	345.81	
Temperature	299.69(10) K	
Wavelength	1.54184 Å	
Crystal system	Orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit cell dimensions	a = 9.35760(10) Å	α = 90°.
	b = 18.24240(10) Å	β = 90°.
	c = 20.28210(10) Å	γ = 90°.
Volume	3462.26(4) Å ³	
Z	8	
Density (calculated)	1.327 Mg/m ³	
Absorption coefficient	2.011 mm ⁻¹	
F(000)	1440	
Crystal size	? x ? x ? mm ³	
Theta range for data collection	3.258 to 77.281°.	
Index ranges	-11 ≤ h ≤ 11, -23 ≤ k ≤ 20, -23 ≤ l ≤ 25	
Reflections collected	46005	
Independent reflections	7109 [R(int) = 0.0370]	
Completeness to theta = 67.684°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.83498	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	7109 / 0 / 451	
Goodness-of-fit on F ²	1.051	
Final R indices [I > 2σ(I)]	R1 = 0.0385, wR2 = 0.1143	
R indices (all data)	R1 = 0.0403, wR2 = 0.1162	
Absolute structure parameter	-0.002(5)	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.503 and -0.300 e.Å ⁻³	

CCDC 1998518 (**3aj**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

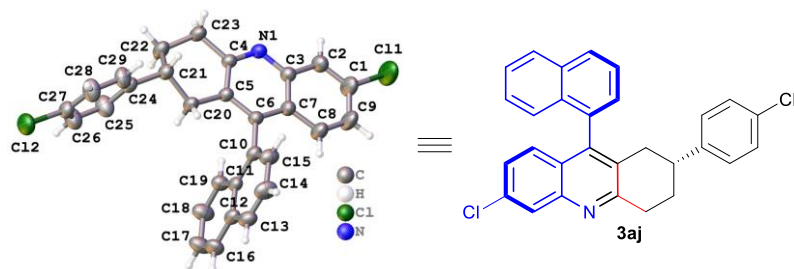


Table S11. Crystal data and structure refinement

Identification code	ndj-hz-1-300k	
Empirical formula	C ₂₉ H ₂₁ Cl ₂ N	
Formula weight	454.37	
Temperature	299.60(10) K	
Wavelength	1.54184 Å	
Crystal system	Monoclinic	
Space group	P 1 21 1	
Unit cell dimensions	a = 10.8865(3) Å	α = 90°.
	b = 8.8581(3) Å	β = 97.528(2)°.
	c = 12.0837(4) Å	γ = 90°.
Volume	1155.23(6) Å ³	
Z	2	
Density (calculated)	1.306 Mg/m ³	
Absorption coefficient	2.644 mm ⁻¹	
F(000)	472	
Crystal size	? x ? x ? mm ³	
Theta range for data collection	3.690 to 76.873°.	
Index ranges	-12 ≤ h ≤ 13, -11 ≤ k ≤ 10, -14 ≤ l ≤ 15	
Reflections collected	12833	
Independent reflections	4168 [R(int) = 0.0348]	
Completeness to theta = 67.684°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.74959	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4168 / 1 / 289	
Goodness-of-fit on F ²	1.079	
Final R indices [I > 2σ(I)]	R1 = 0.0611, wR2 = 0.1885	
R indices (all data)	R1 = 0.0646, wR2 = 0.1934	
Absolute structure parameter	0.05(3)	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.262 and -0.196 e.Å ⁻³	