# Sequence-controlled supramolecular copolymer constructed by self-sorting assembly of multiple noncovalent interactions

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## 1. Self-sorting binding investigation on model compounds 1-6

Fig. S1 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3$ /acetone- $d_6(3/1, v/v)$ , 293 K) of (a) 1, (b) an equimolar solution of 1+2+Zn(OTf)<sub>2</sub>, (c) 2



Fig. S2 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3$ /acetone- $d_6(3/1, v/v)$ , 293 K) of (a) 4, (b) an equimolar solution of 3 and 4, (c) 3.



Fig. S3 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3$ /acetone- $d_6(3/1, v/v)$ , 293 K) of (a) 5, (b) 6, (c) an equimolar solution of 5 and 6.



Fig. S4 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3$ /acetone- $d_6(3/1, v/v)$ , 293 K) of (a) 3, (b) an equimolar solution of 3 and 6, (c) 6.



Fig. S5 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3/acetone-d_6(3/1, v/v)$ , 293 K) of (a) 5, (b) an equimolar solution of 4 and 5, (c) 4.



Fig. S6 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3$ /acetone- $d_6(3/1, v/v)$ , 293 K) of (a) 3+4, (b)1+2+Zn(OTf)<sub>2</sub>+3+4, (c) 1+2+Zn(OTf)<sub>2</sub>.



Fig. S7 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3$ /acetone- $d_6(3/1, v/v)$ , 293 K) of (a) 5+6, (b)1+2+Zn(OTf)<sub>2</sub>+5+6, (c) 1+2+Zn(OTf)<sub>2</sub>.



Fig. S8 <sup>1</sup>H NMR spectra (400 MHz, chloroform-*d*<sub>3</sub>/acetone-*d*<sub>6</sub>(3/1, *v*/*v*), 293 K) of (a) 3+4, (b)3+4+5+6, (c) 5+6.



Fig. S9 <sup>1</sup>H NMR spectra (400 MHz, chloroform- $d_3/acetone-d_6(3/1, \nu/\nu)$ , 293 K) of (a) 5+6, (b) 3+4, (c) 1+2+3+4+5+6+Zn(OTf)<sub>2</sub>, (d) 1+2+Zn(OTf)<sub>2</sub>.

## 2. <sup>1</sup>H-<sup>1</sup>H COSY NMR



**Fig. S10** <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum (400 MHz, CDCl<sub>3</sub>-CD<sub>3</sub>COCD<sub>3</sub> = 3/1, v/v, 293 K, 30mM) of M1+M2+M3+M4+Zn(OTf)<sub>2</sub>. The strong correlations between the protons H<sub>1</sub> and H<sub>2</sub> and between H<sub>3</sub> and H<sub>4</sub> on M1 were observed, the correlations between H<sub>28</sub> and H<sub>29</sub> and between H<sub>29</sub> and H<sub>30</sub> on M4 were also observed at the same time. By means of the <sup>1</sup>H–<sup>1</sup>H COSY experiment, the complex <sup>1</sup>H NMR spectrum of M1+M2+M3+M4+Zn(OTf)<sub>2</sub> was identified.

## 3. Concentration-dependent <sup>1</sup>H NMR spectra



**Fig. S11** <sup>1</sup>H NMR spectra (400 MHz,  $CDCl_3-CD_3COCD_3 = 3/1$ , v/v, 298 K) of M1+M2+M3+M4+Zn(OTf)<sub>2</sub> at different concentrations (a) 4 mM, (b) 8 mM, (c) 20 mM, (d) 50 mM, (e) 90 mM, (f) 260 mM.

## 4. 2D DOSY NMR spectrum



**Fig. S12** Representative DOSY NMR spectrum (600 MHz,  $CDCl_3-CD_3COCD_3 = 3/1$ , v/v, 293 K) of M1+M2+M3+M4+Zn(OTf)<sub>2</sub>, the concentration of M1 is 130 mM.

#### 5. The discussion of binding constants

### (1) tpy-Zn<sup>2+</sup>-tay binding constant

To determine the association constant tpy- $Zn^{2+}$ -tay, UV-vis titration experiment (Job plot method) was performed according to the reported method.<sup>S1</sup> Model compounds **1** and **2** were chosen as the ligands. The samples were prepared so that the total molar concentration of ligands ([1] + [2]

 $^2$  ) and zinc ion was  $2 \times 10^{-5}$ M in each sample: only the ratios of zinc ion to ligands were altered. The absorbance intensity at 410 nm was plotted (Fig. S13) against the mole fraction of Zn<sup>2+</sup>. The Job plot indicates a 1:1:1 binding among Zn<sup>2+</sup>, 1 and 2.



Fig. S13 Job plot of the complex formed among zinc ion, 1 (ligand) and 2 (ligand) showing a 1:1:1 stoichiometry by plotting the absorbance intensity at 410 nm against the mole fraction of zinc ion. Concentration: [1]=[2], [1] + [2]

$$2 + [Zn(OTf)_2] = 20\mu M$$

Furthermore, the data of job plot were divided into two groups around  $X_m = 0.5$ . When  $X_m \le 0.5$ , the fitting equation is  $A = 0.1776X_m + 0.01311$ . When  $X_m \ge 0.5$ , the fitting equation is  $A = -0.20397X_m + 0.20349$ . The intersection point of the two fitting curves is taken ( $X_m = 0.4982$ , A=0.1023), and the experimental value is  $X_m = 0.5$ , A' = 0.0999. The degree of dissociation of complex [Zn12](OTf)<sub>2</sub> was calculated from Eq. 1. According to the formula,<sup>S5</sup> the dissociation degree( $\alpha$ ) of complex [Zn12](OTf)<sub>2</sub> was calculated to be 0.023.

$$\alpha = (A - A')/A, (Eq. 1)$$

The binding constant *K* was then calculated to be  $8.1 \times 10^{14}$  M<sup>-1</sup> based on Eq. 2.



$$\frac{[Zn12](OTf)_2}{K = \frac{[1][2][Zn(OTf)_2]}{[2][Zn(OTf)_2]} = \frac{C(1-\alpha)}{[C\alpha]^3} \quad (Eq.2)$$

Where C is the total concentration of the complex  $[Zn12](OTf)_2$  and  $\alpha$  is the degree of dissociation of complex  $[Zn12](OTf)_2$  when  $X_m$  value is 0.5, with the hypothesis that the ligands and zinc ion only form the complex  $[Zn12](OTf)_2$ . The C is  $1 \times 10^{-5}$  M and the  $\alpha$  is 0.023 when  $X_m$  is 0.5.

#### (2) B21C7-SEA binding constant:

Because B21C7-SEA is a slow exchange interaction, we used model compounds 5 and 6 to determine the binding constant  $K_a$  of the B21C7-SEA according to <sup>1</sup>H NMR single point method.<sup>S2</sup> It could be calculated from integrations of complexed and uncomplexed peaks in <sup>1</sup>H NMR spectrum. The Ka value was determined at 6.00 mM host and guest in CDCl<sub>3</sub>-CD<sub>3</sub>COCD<sub>3</sub>(3/1, v/v) solution. Using the reference method, <sup>S2</sup> Ka {[5•6]/[5][6] } = [(1.62/2.62) × 6 × 10<sup>-3</sup>]/[(1-1.62/2.62) × 6 × 10<sup>-3</sup>]<sup>2</sup> = 706 ± 56 M<sup>-1</sup> in chloroform/acetone solution(3/1, v/v).



Fig. S14 Partial <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>COCD<sub>3</sub> = 3/1, v/v, 298 K) of 6.00 mM 5 and 6.

#### (3) P5-TPN binding constant

P5-TPN binding interaction is a slow exchange interaction, the binding constant was referred by literature value <sup>S3</sup>: *K* is equal to  $(1.2 \pm 0.2) \times 10^4$  M<sup>-1</sup> in CDCl<sub>3</sub>-CD<sub>3</sub>COCD<sub>3</sub>.

#### **6.** Calculated value of maximum polymerization degree $n_{\text{max}}$ .

For the M1+M2+M3+M4+Zn(OTf)<sub>2</sub> system, the maximum possible polymerization degree  $(n_{max})$  could be estimated using a reported method by Gibson and coworkers.<sup>S4a</sup> Using the Carothers equation <sup>S4b</sup> and assuming that the same average association constant holds for each successive step (isodesmic), the maximum possible degree of polymerization( $n_{max}$ ) is related to the equilibrium constant *K* and the initial monomers concentration. It can be easily deduced as follows:

$$n_{\text{max}} = (2K \text{ [Host]}_0)^{1/2},$$
  
where [Host]\_0 = [M2].<sup>S4</sup>

The degree of polymerization  $(n_{\text{max}})$  for M1+M2+M3+M4+Zn(OTf)<sub>2</sub> system at a concentration of 260 mM (260 mM M1 + 260 mM M2 + 130mM M3+ 130mM M4 +260mM Zn(OTf)<sub>2</sub>) was calculated to be 19.2, with the hypothesis that the M2•Zn<sup>2+</sup>•M1•M3 dissociation was negligible in the solution because the association constant of P5-TPN and tpy-Zn<sup>2+</sup>-tay is much larger than that of B21C7-SEA. Thus, the repeat units of in the copolymer [M4•M2•Zn<sup>2+</sup>•M1•M3•M1•Zn<sup>2+</sup>•M2]<sub>n</sub> are about 19.2, and the molecular weight of supramolecular copolymer SCP is about 19.2\*(6280 Da)=120.6 kDa.

### 7. Stimuli-responsiveness study by adding/removing K<sup>+</sup> or adding

#### butanedinitrile

Because B21C7 can capture  $K^+$ ,<sup>S2</sup> the adding-removing  $K^+$  was expected to realize the reversible disassembly-reassembly of SCP, when adding 1 equiv. KPF<sub>6</sub> to the solution of SCP, the complicated <sup>1</sup>H NMR became relatively simpler (Fig. S15b), the sharp peaks corresponding to the uncomplexed protons H<sub>28</sub>, H<sub>30</sub>, and H<sub>EO</sub> (denoted as H<sub>28uc</sub>, H<sub>30uc</sub>, and H<sub>EOuc</sub>) were observed, indicating the disassembly of SCP. It should be noted that K<sup>+</sup> only destroyed the binding of B21C7-SEA, the host-guest interaction of P5-TPN and metal coordination tpy-Zn<sup>2+</sup>-tay were not affected by the observation of <sup>1</sup>H NMR (H<sub>1c-4c</sub> and H<sub>13c-14c</sub> were still observed). After adding another smaller crown ether B18C6 to the solution, the <sup>1</sup>H NMR became complicated again as the B18C6 can capture K<sup>+</sup> tighter (Fig. S15c), suggesting the reformation of SCP. In addition, viscosity measurement also provided important evidence of disassembly-reassembly of SCP. When 1 equiv. KPF<sub>6</sub> was added into the solution of SCP, the specific viscosity of the solution of SCP decreased remarkably (Fig. S16), implying the disassembly of SCP. After adding 1.1 equiv. B18C6 into the solution, the SCP decreased the original value, indicating the reformation of SCP.

On the other hand, the host-guest interaction P5-TPN may also be adjusted by adding a

competitive guest molecule. As shown in Fig. S17, when 1 equiv. butanedinitrile was added into the solution of  $M1+M2+M3+M4+Zn(OTf)_2$ , the complexed protons  $H_{1-4}$  disappeared, new complexed proton  $H_{ac}$  was observed in the highfield region (-1.3 ppm), indicating TPN moiety inside the P5 cavity was replaced by the competitive butanedinitrile and the SCP disassembled into low molecular weight species. From the observation of <sup>1</sup>H NMR, the addition of butanedinitrile only destroyed the host-guest interaction of P5-TPN and did not affect the binding of B21C7-SEA and tpy-Zn<sup>2+</sup>-tay.



**Fig. S15** <sup>1</sup>H NMR spectra (400 MHz,  $CDCl_3-CD_3COCD_3= 3/1$ , v/v, 298 K, 30 mM) of (a) M1+M2+M3+M4+Zn(OTf)<sub>2</sub>, (b) after the addition of 1 equiv. KPF<sub>6</sub>, and (c) after the addition of 1.1 equiv. B18C6. Peaks of complexed monomers and uncomplexed monomers were designated as c and uc, respectively.



**Fig. S16** The specific viscosity (CHCl<sub>3</sub>-CH<sub>3</sub>COCH<sub>3</sub>= 3/1, v/v, 298 K, 30 mM) of (a) M1+M2+M3+M4+Zn(OTf)<sub>2</sub>, (b) after the addition of 1 equiv. KPF<sub>6</sub>, and (c) after the addition of 1.1 equiv. B18C6.



**Fig. S17** <sup>1</sup>H NMR spectra (400 MHz,  $CDCl_3-CD_3COCD_3= 3/1$ , v/v, 298 K, 20 mM) of (a) M1+M2+M3+M4+Zn(OTf)<sub>2</sub>, (b) after the addition of 1 equiv. butanedinitrile. Peaks of complexed monomers and uncomplexed monomers were designated as c and uc, respectively.

#### (1) Destroy/Recover the B21C7-SEA binding





Fig. S18 Graphical representation of stimuli-responsiveness by adding/removing K<sup>+</sup> or adding butanedinitrile.



#### 8. Fluorescence emission spectra

**Fig. S19** Fluorescence emission spectra of the M1+M2+M3+M4 and M1+M2+M3+M4+Zn(OTf)<sub>2</sub> upon an excitation at 320 nm in CHCl<sub>3</sub>-CH<sub>3</sub>COCH<sub>3</sub> (v/v = 3/1, 0.1 mM). Inset: visual fluorescence emission images of M1+M2+M3+M4 and M1+M2+M3+M4+Zn(OTf)<sub>2</sub> using 365 nm UV lamp irradiation.

#### 9. Synthesis of monomers

#### Synthesis of monomer M2



Scheme S1. Synthesis of the monomer M2.

In a 250 mL round–bottom flask, compound 7 (4.00g, 5.9mmol), CS<sub>2</sub>CO<sub>3</sub> (5.82 g, 18mmol), compound 8 (3.32 g, 5.9mmol), and DMF (150mL) were added. The reaction mixture was stirred at 80 °C for 14 hours. After the solid was filtered off, the solvent was removed under reduced pressure. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (150 mL) and washed twice with H<sub>2</sub>O (200 mL). The organic phase was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated to afford the crude product, which was purified by flash column chromatography (dichloromethane/methanol=70:1). The fractions containing the product were combined and concentrated under vacuum to give M2 (4.10 g, 60 %) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) 8.95(d, J = 10.4 Hz, 2H), 8.68(s, 2H), 8.59 (s, 2H), 8.16 (t, J = 10.2 Hz, 2H), 8.09 (d, J = 11.2 Hz, 4H), 7.77 (d, J = 11.6 Hz, 4H), 7.58-7.64 (m, 5H), 7.47-7.53 (m, 5H), 7.36-7.43 (m, 4H), 6.84 (d, J = 11.2 Hz, 1H), 6.75 (d, J = 11.6Hz, 2H), 4.26 (t, J = 8.6 Hz, 2H), 4.16-4.21 (m, 4H), 3.89-3.97 (m, 4H), 3.85 (t, J = 8.8 Hz, 2H), 3.76-3.83 (m, 4H), 3.70-3.75 (m, 4H), 3.62-3.69 (m, 8H), 1.72-1.79 (m, 4H), 1.41-1.47 (m, 4H). <sup>13</sup>C NMR(100MHz, CDCl<sub>3</sub>): δ (ppm) = 166.5, 159.9, 157.7, 156.6, 155.9, 152.9, 150.1, 148.4, 137.2, 135.6, 131.6, 130.3, 128.6, 127.6, 126.4, 125.9, 125.3, 123.9, 123.3, 120.2, 119.1, 114.7, 112.3, 71.4, 71.3, 71.1, 71.0, 70.7, 69.8, 69.6, 69.4, 69.2, 67.9, 64.9, 29.1, 28.8, 25.9, 25.8. High-resolution MALDI-TOF-MS (C74H69N3O10): m/z calcd for [M]<sup>+</sup> =1159.4983, found =1159.4958, error 2.1 ppm.



Fig. S21 <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, room temperature) of monomer M2.



## Synthesis of monomer M3



Scheme S2. Synthesis of the monomer M3.

A solution of compound 9 (2.00g, 2.7mmol), 1.6-dibromohexane (0.33g, 1.35mmol), Cs<sub>2</sub>CO<sub>3</sub> (2.64g, 8.1mmol) in DMF (120 mL) was stirred for 14 h at 75 °C. After the reaction mixture was cooled to ambient temperature, the solvent was evaporated under reduced pressure and the residue was partitioned between dichloromethane (70 mL) and water (70 mL). The aqueous layer was further washed with dichloromethane ( $2 \times 30$  mL). The organic phases were combined and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. After the solvent was removed, the resulting residue was subjected to column chromatography (CH<sub>2</sub>Cl<sub>2</sub> as eluent), to give M3 (1.25 g, 58 %) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 298 K): ppm = 6.79-6.73 (m, 20H), 3.87 (t, J = 6.8 Hz, 4H), 3.72-3.79 (m, 20H), 3.61-3.69 (m, 54H), 1.82-1.87 (m, 4H), 1.60-1.66 (m, 4H). <sup>13</sup>C NMR (100MHz, CDCl<sub>3</sub>):  $\delta$ 



(ppm) =150.9, 150.8, 128.3, 128.2, 115.1, 114.2, 114.1, 68.5, 55.9, 30.0, 29.4, 26.4. HR-ESI-MS ( $C_{94}H_{106}O_{20}$ ): m/z calcd for [M]<sup>+</sup> =1555.7311, found =1555.7302, error 0.6 ppm.

Fig. S24 <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, room temperature) of compound M3.



Fig. S25 High-resolution electrospray ionization mass spectrum of compound M3.

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