Regio- and diastereoselective synthesis of spiropyrroloquinoxaline grafted indole hetercyclic hybrids and evaluation of their anti-Mycobacterium tuberculosis activity

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S. No	List of Figures	
		No
1	¹ H NMR spectrum of 6k	6
2	Expanded ¹ H NMR spectrum of 6k	6
3	¹³ C NMR spectrum of 6 k	7
4	DEPT-135 spectrum of 6k	8
5	¹ H, ¹ H-COSY spectrum of 6 k	9
6	Expanded ¹ H, ¹ H-COSY spectrum of 6 k	10
7	¹³ C, ¹ H-COSY(HMQC) spectrum of 6 k	11
8	Expanded ¹³ C, ¹ H-COSY(HMQC) spectrum of 6 k	12
9	HMBC spectrum of 6k	13
10	Expanded HMBC spectrum of 6k	14

Crystal Data			
Chemical Formula	$C_{34}H_{27}N_5O_2$		
Formula weight (a.k.b.)	537.60		
Temperature (K)	296		
Crystal system	Triclinic		
Space group	P-1		
Unit cell parameters			
a, b, c (Å)	11.5002 (7), 11.7543 (8), 14.3642 (10)		
α , β, γ (°)	97.266 (5), 106.616 (5), 111.900 (5)		
Crystal size (mm)	$0.39 \times 0.29 \times 0.23$		
Volume, V (Å ³)	1665.9 (2)		
Z	2		
μ (mm ⁻¹)	0.07		
F ₀₀₀	564		
Calculated density (Mg/m ³)	1.072		
Data collection			
Diffractometer	STOE IPDS 2		
Wavelength (Å)	0.71073		
θ range for data collection (°)	$2.1 \le \theta \le 26.0$		
Index ranges			
$h_{min}, h_{max}; k_{min}, k_{max}; l_{min}, l_{max}$	-13, 14; -14, 14; -17, 17		
Measurement method	$\omega_{\rm scan}$		
Reflections collected	16765		
Independent reflections	6550		
Observed reflections $[I > 2\sigma(I)]$	3270		
Absorption correction	Integration		
T _{min} , T _{max}	0.9732, 0.9866		
R _{int}	0.051		
Refinement			
Refinement method	SHELXL17/1		
Parameters	375		
$R[F^2 > 2\sigma(F^2)]$	0.057		
$wR(F^2)$	0.152		
GooF = S	0.94		
$\Delta \rho_{\min}, \Delta \rho_{\max} (e/Å^3)$	-0.10, 0.17		

 Table 1. Crystal data and structure refinement parameters for 6e.

<i>D</i> —Н…А	<i>D</i> —Н	Н…А	<i>D</i> ····A	<i>D</i> —Н…А
C18–H18…N2	0.93	2.61	3.338 (3)	135
C24– $H24$ ···O2 ⁱ	0.98	2.56	3.433 (3)	148
N3–H3…N1 ⁱⁱ	0.99(2)	2.35(3)	3.260 (3)	153.5 (2)

 Table 2. Hydrogen bonding geometry for 6e.

Symmetry codes: (*i*) -x+2, -y, -z+1; (*ii*) -x+1, -y-1, -z+1.

General Method

Chemistry

¹H and ¹³C NMR spectra were recorded on a Varian Mercury JEOL-400/500 NMR spectrometers in DMSO-d₆ using TMS as internal standard. Chemical shifts are given in parts per million (δ -scale) and coupling constants are given in hertz. Elemental analyses were performed on a Perkin Elmer 2400 Series II Elemental CHNS analyzer.

Biology

In vitro anti tubercular activity

Briefly, the *Mycobacterium tuberculosis* H37Rv inoculum was prepared from fresh LJ medium re-suspended in 7H9-S medium (7H9 broth, 0.1% casitone, 0.5% glycerol, supplemented oleic acid, albumin, dextrose, and catalase [OADC]), adjusted to a OD_{590} 1.0, and diluted 1:20; 100 µl was used as inoculum. Each drug stock solution was thawed and diluted in 7H9-S at four-fold the final highest concentration tested. Serial two-fold dilutions of each drug were prepared directly in a sterile 96-well microtiter plate using 100 µl 7H9-S. A growth control containing no antibiotic and a sterile control were also prepared on each plate. Sterile water was added to all perimetre wells to avoid evaporation during the incubation. The plate was covered, sealed in plastic bags and incubated at 37°C in normal atmosphere. After 7 days incubation, 30 µl of alamar blue solution was added to each well, and the plate was re-incubated overnight. A change in colour from blue (oxidised state) to pink (reduced) indicated the growth of bacteria, and the MIC was defined as the lowest concentration of drug that prevented this change in colour ^{1,2}.

In vitro cytotoxicity screening:

The *in vitro* cytotoxicity of the privileged antitubercular active analogues with lower MIC value were assessed by 3-(4,5- dimethylthiazol-2- yl)-2,5- diphenyltetrazolium bromide (MTT) assay against growth inhibition of RAW 264.7 cells at 50 μ g/mL concentration.³ Cell lines were maintained at 37°C in a humidified 5% CO₂ incubator (Thermo scientific). Detached the adhered cells and followed by centrifugation to get cell pellet. Fresh media was added to the pellet to make a cell count using haemocytometer and plate 100 μ l of

media with cells ranging from 5,000 - 6,000 per well in a 96-well plate. The plate was incubated overnight in CO₂ incubator for the cells to adhere and regain its shape. After 24hr cells were treated with the test compounds at 50 µg/mL diluted using the media to deduce the percentage inhibition on normal cells. The cells were incubated for 48 hr to assay the effect of the test compounds on different cell lines. Zero hour reading was noted down with untreated cells and also control with 1% DMSO to subtract further from the 48hr reading. After 48 hr incubation, cells were treated by MTT (4, 5-dimethylthiazol- 2-yl)- 2, 5-diphenyltetrazolium bromide) dissolved in PBS (5mg/ml) and incubated for 3-4 hr at 37°C. The formazan crystals thus formed were dissolved in 100µl of DMSO and the viability was measured at 540nm on a multimode reader (Spectra max). The values were further calculated for percentage inhibition which in turn helps us to know the cytotoxicity of the test compounds.

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Figure S1. Selected ¹H and ¹³C NMR Chemical shifts of 6k



Figure S2. ¹H NMR spectrum of 6k



Figure S3. Expanded ¹H NMR spectrum of 6k



Figure S4. ¹³C NMR spectrum of 6k



Figure S5. DEPT-135 spectrum of 6k



Figure S6. ¹H,¹H-COSY spectrum of 6k



Figure S7. Expanded ¹H,¹H-COSY spectrum of 6k



Figure S8. ¹³C, ¹H,-COSY spectrum of 6k



Figure S9. Expanded ¹³C, ¹H-COSY spectrum of 6k



Figure S10. HMBC spectrum of 6k



Figure S11. Expanded HMBC spectrum of 6k