Electronic Supplementary Material (ESI) for RSC Advances. This journal is © The Royal Society of Chemistry 2020

Supporting Information

Development of a horseradish peroxidase-nanobody fusion

protein for visual detection of ochratoxin A by dot

immunoassay

Qi Chen,^{ab‡} Yuanyuan Wang,^{ab‡} Fujing Mao,^{ab} Benchao Su,^{ab} Kunlu Bao,^{ab} Zeling Zhang,^{ab} Guifang Xie,^{ab} Xing Liu^{ab*}

^a College of Food Science and Engineering, Hainan University, Haikou 570228,

China.

^b Key Laboratory of Food Nutrition and Functional Food of Hainan Province, Haikou

570228, China.

qichen@hainanu.edu.cn; wangyuanyuan@hainanu.edu.cn; maofujing11@163.com;

benchao312@hainanu.edu.cn; baokunlu@hainanu.edu.cn;

zhangzeling@hainanu.edu.cn; xgf@hainanu.edu.cn;

[‡] These authors contributed equally to this work

*Corresponding Author: Tel./Fax: +86-898-66193581; e-mail: xliu@hainanu.edu.cn (X.L.).

Table of contents

s-3
s-4
s-5
s-6
s-7
s-8
s-9
s-10
s-11

Table	S 1	The	primers	for	constructing	the	recombinant	plasmids	pET25b-Nb28-HRP	and
pET25	b-H	RP-N	b28							

Name	The sequence of primers	Protein	Recombinant plasmid
HN-HF	GGAATTC <u>CATATG</u> CAGTTAACGCCGACTTTCTA CGATAAC	HRP	PET25b-HRP-Nb28
HN-HR	CCGCCAGAGCCACCTCCGCCTGAACCGCCTCC TCCTGAGTTCGAGTTTACGACTCGGC		
HN-NF	GTTCAGGCGGAGGTGGCTCTGGCGGTGGCGG ATCCATGGCCATGGCCCAGTTGC	Nb28	
HN-NR	ATAAGAAT <u>GCGGCCGC</u> TTGTGGTTTTGGTGTC TTGGGTTC		
NH-NF	GGAATTC <u>CATATG</u> ATGGCCATGGCCCAGTTGC	Nb28	PET25b-Nb28-HRP
NH-NR	CCGCCAGAGCCACCTCCGCCTGAACCGCCTCC TCCTTGTGGTTTTGGTGTCTTGGGTTC		
NH-HF	GTTCAGGCGGAGGTGGCTCTGGCGGTGGCGG ATCCCAGTTAACGCCGACTTTCTACGAT	HRP	
NH-HR	ATAAGAAT <u>GCGGCCGC</u> TGAGTTCGAGTTTACG ACTCG		



Fig.S1 Construction of the recombinant plasmids pET25b-HRP-Nb28 and pET25b-Nb28-HRP (A) and DNA agarose analysis of the gene fragments of HRP-Nb28 and Nb28-HRP (B). Lane 1: HRP-Nb28 fragment; Lane 2: Nb28-HRP fragment; M: D2000 DNA Ladder.

 Table S2 Amino acid sequences of the fragments of HRP-Nb28 and Nb28-HRP

DNA fragment	Amino acid sequence (5'-3')
Nb28-HRP	MAMAQLQLVESGGGLVQAGGSLRLSCAASGSTVGVNAMDMGWYRQ APGKQRELVAAIINGGGSTNLADSVKGRFTISRDGAKRTLYLQMNSLK PEDTAVYYCYVRSGVGLVYWGQGTQVTVSSEPKTPKPQGGGGSGGG GSGGGGSQLTPTFYDNSCPNVSNIVRDTIVNELRSDPRIAASILRLHFHD CFVNGCDASILLDNTTSFRTEKDAFGNANSARGFPVIDRMKAAVESAC PRTVSCADLLTIAAQQSVTLAGGPSWRVPLGRRDSLQAFLDLANANLP APFFTLPQLKDSFRNVGLNRSSDLVALSGGHTFGKNQCRFIMDRLYNFS NTGLPDPTLNTTYLQTLRGLCPLNGNLSALVDFDLRTPTIFDNKYYVNL EEQKGLIQSDQELFSSPNATDTIPLVRSFANSTQTFFNAFVEAMDRMGN ITPLTGTQGQIRLNCRVVNSNS
HRP-Nb28	QLTPTFYDNSCPNVSNIVRDTIVNELRSDPRIAASILRLHFHDCFVNGCD ASILLDNTTSFRTEKDAFGNANSARGFPVIDRMKAAVESACPRTVSCA DLLTIAAQQSVTLAGGPSWRVPLGRRDSLQAFLDLANANLPAPFFTLP QLKDSFRNVGLNRSSDLVALSGGHTFGKNQCRFIMDRLYNFSNTGLPD PTLNTTYLQTLRGLCPLNGNLSALVDFDLRTPTIFDNKYYVNLEEQKGL IQSDQELFSSPNATDTIPLVRSFANSTQTFFNAFVEAMDRMGNITPLTGT QGQIRLNCRVVNSNSGGGGSGGGGSGGGGSMAMAQLQLVESGGGLV QAGGSLRLSCAASGSTVGVNAMDMGWYRQAPGKQRELVAAIINGGG STNLADSVKGRFTISRDGAKRTLYLQMNSLKPEDTAVYYCYVRSGVG LVYWGQGTQVTVSSEPKTPKPQ



Fig.S2 Western blot analysis of the auto-induction expression of fusion proteins HRP-Nb28 (left) and Nb28-HRP (right). Lane 1 and 6: The precipitated protein of the induced *E.coli* cell after sonication; Lane 2 and 5: Total protein of the induced *E.coli* cell; lane 3 and 4: The supernatant protein of the induced *E.coli* cell after sonication; lane M: Prestained protein ladder. The red arrows point to the target proteins.



Fig. S3 SDS-PAGE analysis of the purification of HRP-Nb28 inclusion body. Lane M: Prestained protein ladder; Lane 1: The purified inclusion body; Lane 2-4: Supernatants collected after washing the inclusion body with wash buffer I; Lane 5-7: Supernatants collected after washing the inclusion body with wash buffer II.



Fig. S4 The performance analysis of HRP-Nb28 fusion protein

No.	Methods	Cut-off value	Detection time	References
1	Dot ELISA	10 ng/mL	10 min	This work
2	Test strip	16 ng/mL	10 min	1
3	Test strip	5 ng/mL	5-10 min	2
4	Dot ELISA	0.625 ng/mL	21 min	3
5	Dot ELISA	0.3125 ng/mL	< 6 min	4

Table S3 Comparison of the proposed method with other reported OTA detection methods



Fig.S5 Evaluation of the matrix effect of rice (A) and oats (B) samples at different dilutions on the performance of HN-DIA.

References

(1) Cheng, Y.; Liu, L.; Liu, H.; Xu, L.; Kuang, H. Rapid and sensitive detection of ochratoxin A in rice flour using a fluorescent microsphere immunochromatographic test strip assay. *Food and Agricultural Immunology*. **2020**, *31*, 563-574.

(2) Liu, L.; Xu, L.; Suryoprabowo, S.; Song, S.; Kuang, H. Development of an immunochromatographic test strip for the detection of ochratoxin A in red wine. *Food and Agricultural Immunology*. **2017**, 1-11.

(3) Sun, Z.; Duan, Z.; Liu, X.; Deng, X.; Tang, Z. Development of a nanobody-based competitive dot ELISA for visual screening of ochratoxin A in cereals. *Food Analytical Methods*. **2017**, *10*, 3558-3564.

(4) Tang, Z.; Wang, X.; Lv, J.; Hu, X.; Liu, X. One-step detection of ochratoxin A in cereal by dot immunoassay using a nanobody-alkaline phosphatase fusion protein. *Food Control.* **2018**, *92*, 430-436.