

SUPPORTING INFORMATION

Iodine-mediated formal [3+2] annulation for synthesis of Furocoumarin from oxime esters

Quyên T. Pham¹, Ha V. Dang,¹ Hiep Q. Ha,² Huong T. D. Nguyen,³ Phong Q. Le^{4*}, Thanh Truong^{2*}, Tri Minh Le^{1*}

¹School of Medicine, VNU-HCM, Linh Trung Ward, Thu Duc District, Ho Chi Minh City, Viet Nam

²Department of Chemical Engineering, HCMC University of Technology, VNU-HCM, 268 Ly Thuong Kiet, District 10, Ho Chi Minh City, Viet Nam

³Department of Chemistry, HCMC University of Natural Science, VNU-HCM, 227 Nguyen Van Cu Street, District 5, Ho Chi Minh City, Viet Nam

⁴School of Biotechnology, International University, VNU-HCM, Quarter 6, Linh Trung Ward, Thu Duc District, Ho Chi Minh City, Viet Nam

*Email: leminhtri@ump.edu.vn

tvthanh@hcmut.edu.vn

lqphong@hcmiu.edu.vn

Table of Contents

Section S1	<i>Catalysis: Materials and Instrumentation</i>	S3
Section S2	<i>Synthesis of starting materials</i>	S4
Section S3	<i>GC yield measurement</i>	S4
Section S4	<i>Crystal data</i>	S6
Section S5	<i>NMR of products</i>	S8
Section S6	<i>References</i>	S55

Section S1. Catalysis: Materials and Instrumentation

Chemical used in this work. All chemicals were purchased from Sigma – Aldrich, Acros Organics, and Fisher and used without further purification unless otherwise noted.

Analytical techniques.

Single crystals suitable for X-ray analysis were obtained by re-crystallization from methanol. The single crystal data for a colorless plate-shaped crystal ($0.25 \times 0.25 \times 0.012$ mm³) was collected on a Bruker D8 QUEST diffractometer at 100 K with Mo K α radiation ($\lambda = 0.71076$ Å) using a TRIUMP monochromator, operated at 50 kV and 30.0 mA. The raw data was processed with the Bruker APEX3 software package¹ and then integrated with the Bruker SAINT package² using a narrow-frame algorithm – corrected for absorption using the SADABS procedure.³ The structures were solved by intrinsic phasing methods. The refinement was performed by full-matrix least squares on F^2 (SHELXL-2014)⁴ using the Olex2 software package.⁵

The products were indicated by GC-MS, analysis data were recorded on a Shimadzu GCMS-QP2010 Ultra with a ZB-5MS column (length = 30 m, inner diameter = 0.25 mm, film thickness = 0.25 μ m). The temperature program for GC-MS analysis heated samples at 50 °C for 2 min, from 50 °C to 280 °C at rate of 10 °C/min, then held at 280 °C for 5 min. MS spectra were compared with the spectra gathered in the NIST library.

Gas chromatography (GC) analysis were performed using a Shimadzu GC 2010-Plus equipped with a FID detector and a SPB-5 column (length = 30 m, inner diameter = 0.25 mm, film thickness = 0.25 μ m). The temperature program for GC analysis heated sample at 100 °C for 1 minute, from 100 °C to 120 °C at rate of 20 °C/minute, held at 120 °C for 2 minutes. And then from 120 °C to 280 °C at rate of 40 °C/minute, held 280 °C for 1 minute. 1,2-dichlorobenzene was used as internal standard.

The ¹H and ¹³C NMR spectra were recorded on a Bruker AV 500 MHz spectrometer operating at 500 MHz for ¹H and 125 MHz for ¹³C, respectively, using tetramethylsilane as standard. The chemical shifts (δ) are expressed as values in parts per million (ppm) and the coupling constant (J) is given in hertz (Hz). Spin multiplicities are described as *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), and *m* (multiplet).

Section S2. *Synthesis of starting materials*

Synthesis of ketoximes

In a typical procedure, the mixture of ketone derivatives (22 mmol), hydroxylamine hydrochloride $\text{NH}_2\text{OH}\cdot\text{HCl}$ (33 mmol) was stirred in 10 mL EtOH at 60 °C for 1h. During the reaction, K_2CO_3 (22 mmol) was added dropwise into this mixture. When the reaction was completed (TLC), the mixture was cooled to room temperature, and the desired product was extracted with EtOAc (20 mL), then washed the mixture with deionized water (3 x 10 mL) and dried the organic phase over anhydrous Na_2SO_4 before removed under reduced pressure to give corresponding ketoxime derivatives.

Synthesis of ketoxime carboxylates

In a typical procedure, the mixture of ketoxime (22 mmol), acetic anhydride (44.4 mmol, 2.0 eq.) was stirred in 10 mL EtOAc at room temperature for 1h. During the reaction, K_2CO_3 (22 mmol) was added dropwise into this mixture. When the reaction was completed (TLC), the mixture was cooled to room temperature, diluted with EtOAc (25 mL) and washed with H_2O (20 mL) and brine (10 mL). The organic layers were dried over anhydrous Na_2SO_4 and evaporated in vacuo. The residue was purified by column chromatography on silica gel to afford the ketoxime acetates with hexane/ethyl acetate as the eluent.

Section S3. *GC yield measurement*

After the transformations were completed, samples were withdrawn from reactors, quenched with water and then eluted with ethyl acetate. Subsequently, the organic layer was carefully shaken with anhydrous Na_2SO_4 before being analyzed by GC system. GC yields of the desired products was determined as following

$$\text{GC yield (\%)} = \frac{n_{\text{Pr}} \times 100\%}{n_{\text{Pr}'}} = \left(\frac{S_{\text{Pr}}}{S_{\text{IS}}} \times 0,823 + 0.0127 \right) \times n_{\text{IS}} \times \frac{100\%}{n_{\text{Pr}'}}$$

In which:

n_{Pr} (mg): Mole of 3-Phenyl-4H-furo[3,2-c]chromen-4-one product obtained

$n_{\text{Pr}'}$ (mg): Calculated mole of 3-Phenyl-4H-furo[3,2-c]chromen-4-one when yield = 100%

n_{IS} (mg): Mole of n-hexadecane in sample

S_{Pr} : Peak area of 2-(3-oxo-3-phenylpropyl) isoindoline-1,3-dione in sample

S_{IS} : Peak area of n-hexadecane in sample

This formula was determined, relied on the calibration curve achieved by the following process: diphenylether (42,3 mg) and 3-Phenyl-4H-furo[3,2-c]chromen-4-one (19,6 mg) were added to two distinct 8 mL volumetric flasks. After that, to dissolve these substances, the flasks were supplemented by dichlobenzene until the solvent masses reach 6494,8 mg and 1995,4 mg, respectively.

	Product	Internal standard
<i>Mass (mg)</i>	19,6	42,3
<i>Dichlobenzene (mg)</i>	1995,4	6494,8
<i>C% (w/w)</i>	0,9727	0.6471

Table **Error! No text of specified style in document.-1**: Calibration curve preparation for 3-Phenyl-4H-furo[3,2-c]chromen-4-one

Different fractions of the two solutions were then withdrawn and attributed to six 1,5 mL GC vials which were analyzed by GC method later. The areal ratio of 3-Phenyl-4H-furo[3,2-c]chromen-4-one to diphenylether were obtained via GC data. As a result, the calibration curve was illustrated in Figure 3-1.

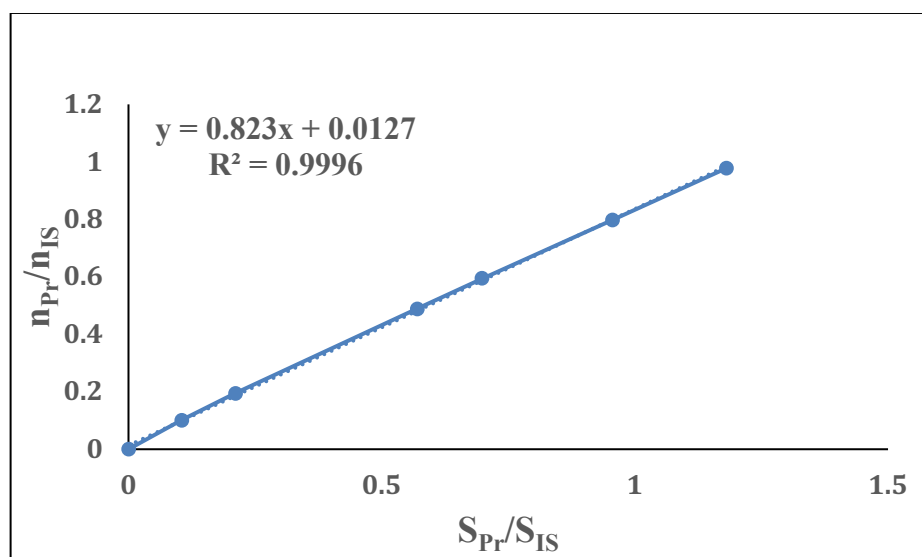


Figure **Error! No text of specified style in document.-1** Calibration curve of 3-Phenyl-4H-furo[3,2-c]chromen-4-one

Section S4. Crystal data

CIF deposit number: 2026801

Crystal data and structure refinement for 3a

Identification code	3a
Empirical formula	C ₁₇ H ₁₀ O ₃
Formula weight	262.25
Temperature/K	100
Crystal system	orthorhombic
Space group	Pbca
<i>a</i> /Å	13.2356(8)
<i>b</i> /Å	7.1744(4)
<i>c</i> /Å	25.3234(16)
α /°	90
β /°	90
γ /°	90
Volume/Å ³	2404.6(2)
<i>Z</i>	8
ρ_{calc} /cm ³	1.449
μ /mm ⁻¹	0.100
<i>F</i> (000)	1088.0
Crystal size/mm ³	0.25 × 0.25 × 0.012
Radiation	MoK α (λ = 0.71073)
2 θ range for data collection/°	6.156 to 52.772
Index ranges	-16 ≤ <i>h</i> ≤ 16, -8 ≤ <i>k</i> ≤ 8, -31 ≤ <i>l</i> ≤ 31
Reflections collected	25205
Independent reflections	2450 [<i>R</i> _{int} = 0.1303, <i>R</i> _{sigma} = 0.0499]
Data/restraints/parameters	2450/0/181
Goodness-of-fit on <i>F</i> ²	1.039
Final <i>R</i> indexes [<i>I</i> ≥ 2 σ (<i>I</i>)]	<i>R</i> ₁ = 0.0518, w <i>R</i> ₂ = 0.1000
Final <i>R</i> indexes [all data]	<i>R</i> ₁ = 0.0962, w <i>R</i> ₂ = 0.1151
Largest diff. peak/hole / e Å ⁻³	0.23/-0.21

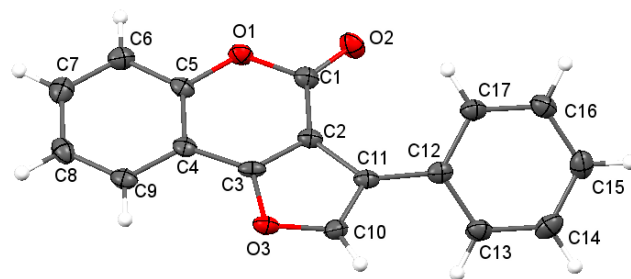
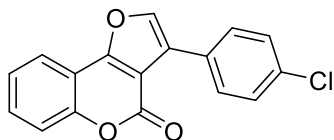


Figure S1. ORTEP representation of the asymmetric unit of compound 3a displayed with 50% probability. Atom colors: O, red; C, grey; H, white.

Section S5 NMR of products

Spectroscopic data for 3-(4-chlorophenyl)-4*H*-furo[3,2-*c*]coumarin (HP1)



^1H NMR (500 MHz, DMSO- d_6): δ 8.54 (d, $J = 1.0$ Hz, 1H), 7.99 (dt, $J = 7.8, 1.5$ Hz, 1H), 7.86 (dd, $J = 8.4, 2$ Hz, 2H), 7.67 (ddd, $J = 8.5, 7.3, 1.3$ Hz, 1H), 7.56 – 7.52 (m, 3H), 7.49 – 7.45 (t, $J = 7.5$ Hz, 1H).

^{13}C NMR (126 MHz, DMSO- d_6) δ 158.22, 157.10, 152.00, 143.49, 132.87, 131.47, 130.24, 128.41, 127.99, 124.95, 124.20, 120.98, 116.82, 112.07, 107.74.

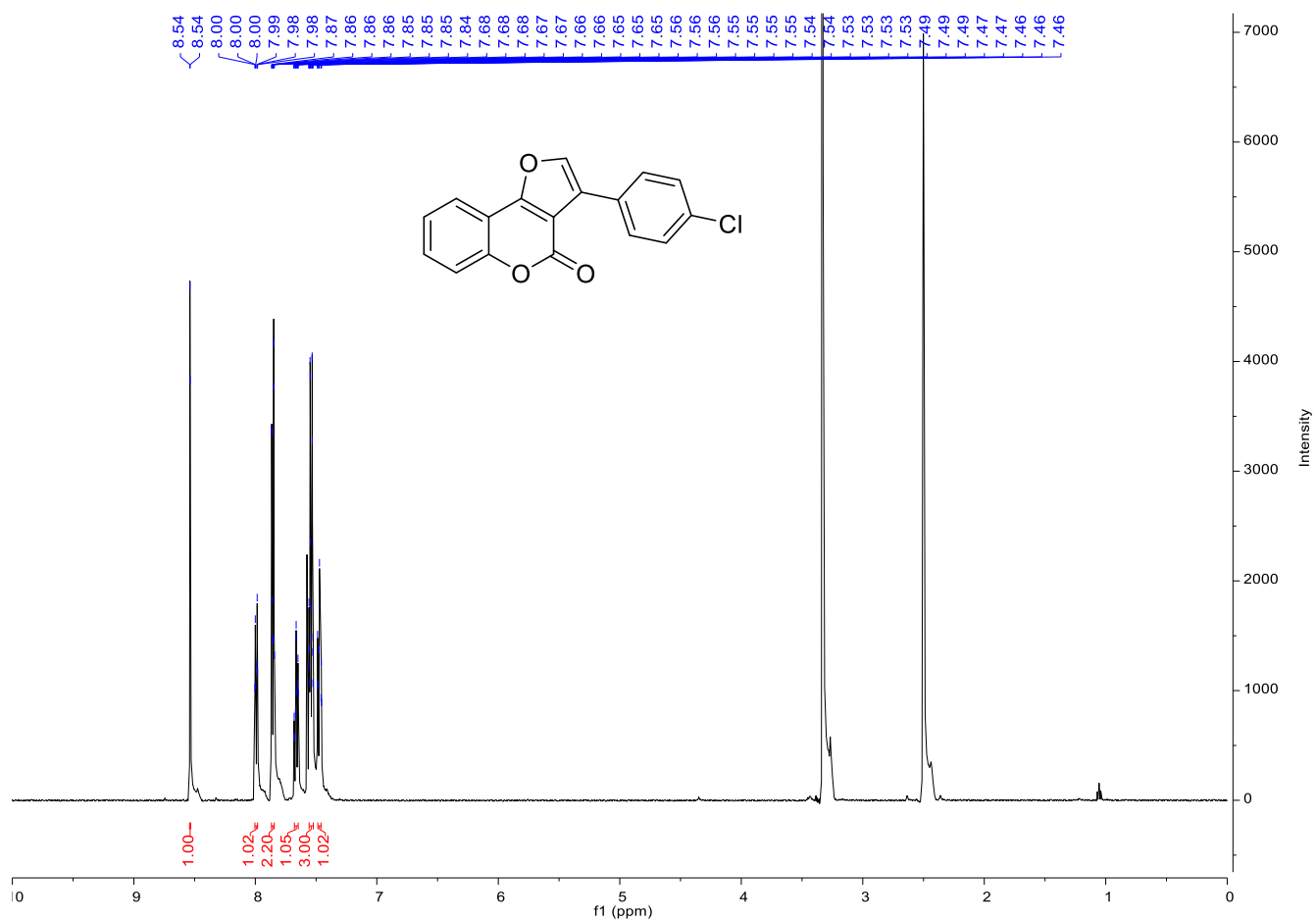


Fig. Sx. ^1H -NMR spectra of 3-(4-chlorophenyl)-4*H*-furo[3,2-*c*]coumarin.

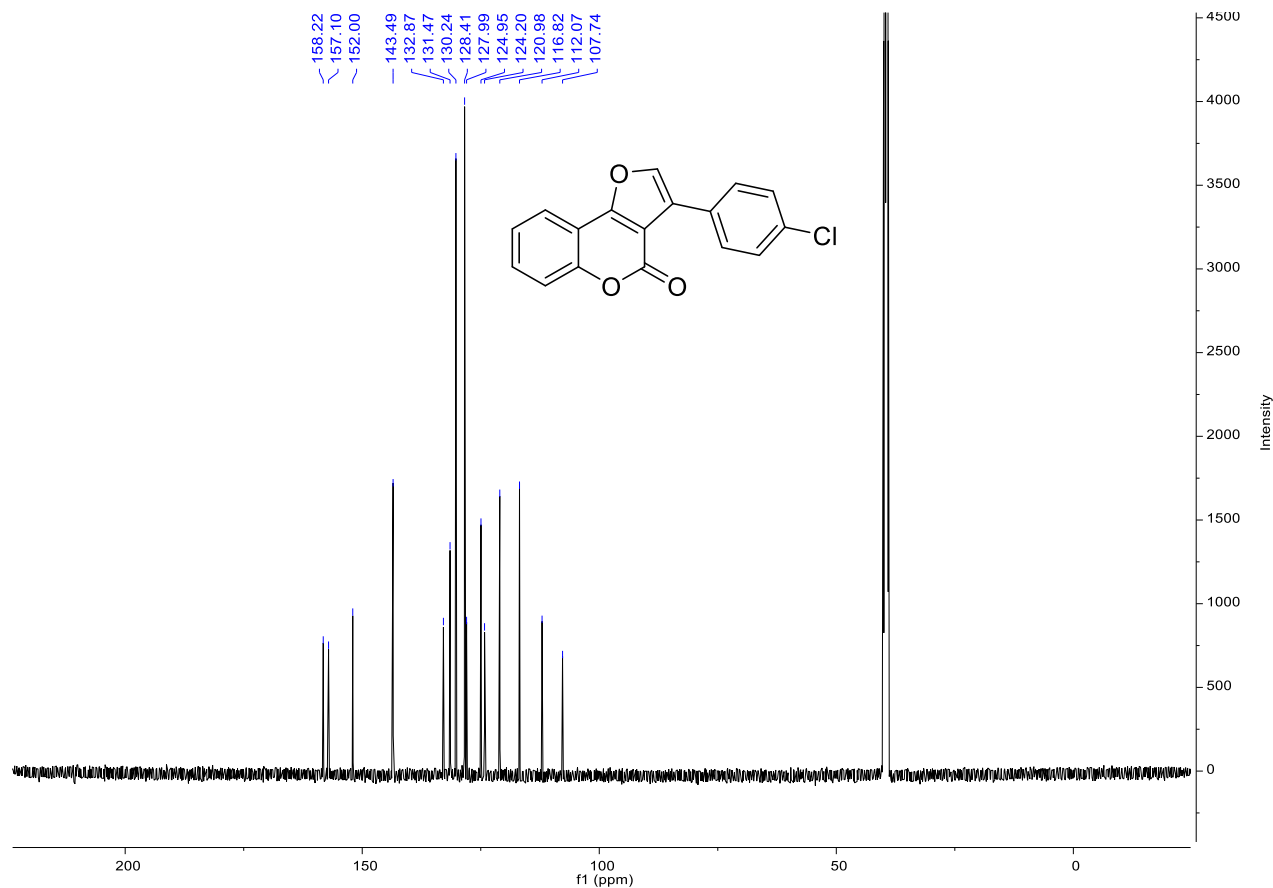
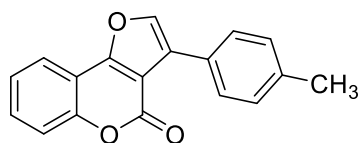


Fig. Sx. ^{13}C -NMR spectra of 3-(4-chlorophenyl)-4*H*-furo[3,2-*c*]coumarin.

Spectroscopic data for 3-(4-methylphenyl)-4*H*-furo[3,2-*c*]coumarin (HP2)



^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 8.39 (d, $J = 1.4$ Hz, 1H), 7.93 (dt, $J = 7.8, 1.8$ Hz, 1H), 7.69 – 7.66 (m, 2H), 7.62 (ddd, $J = 8.7, 7.3, 1.5$ Hz, 1H), 7.51 (dd, $J = 8.5, 1.1$ Hz, 1H), 7.43 (td, $J = 7.5, 1.1$ Hz, 1H), 7.24 (d, $J = 7.9$ Hz, 2H), 2.33 (s, 3H).

^{13}C NMR (126 MHz, DMSO) δ 158.01, 157.02, 151.92, 142.75, 137.46, 131.26, 128.89, 128.36, 126.08, 125.29, 124.82, 120.87, 116.73, 112.11, 107.81, 20.80.

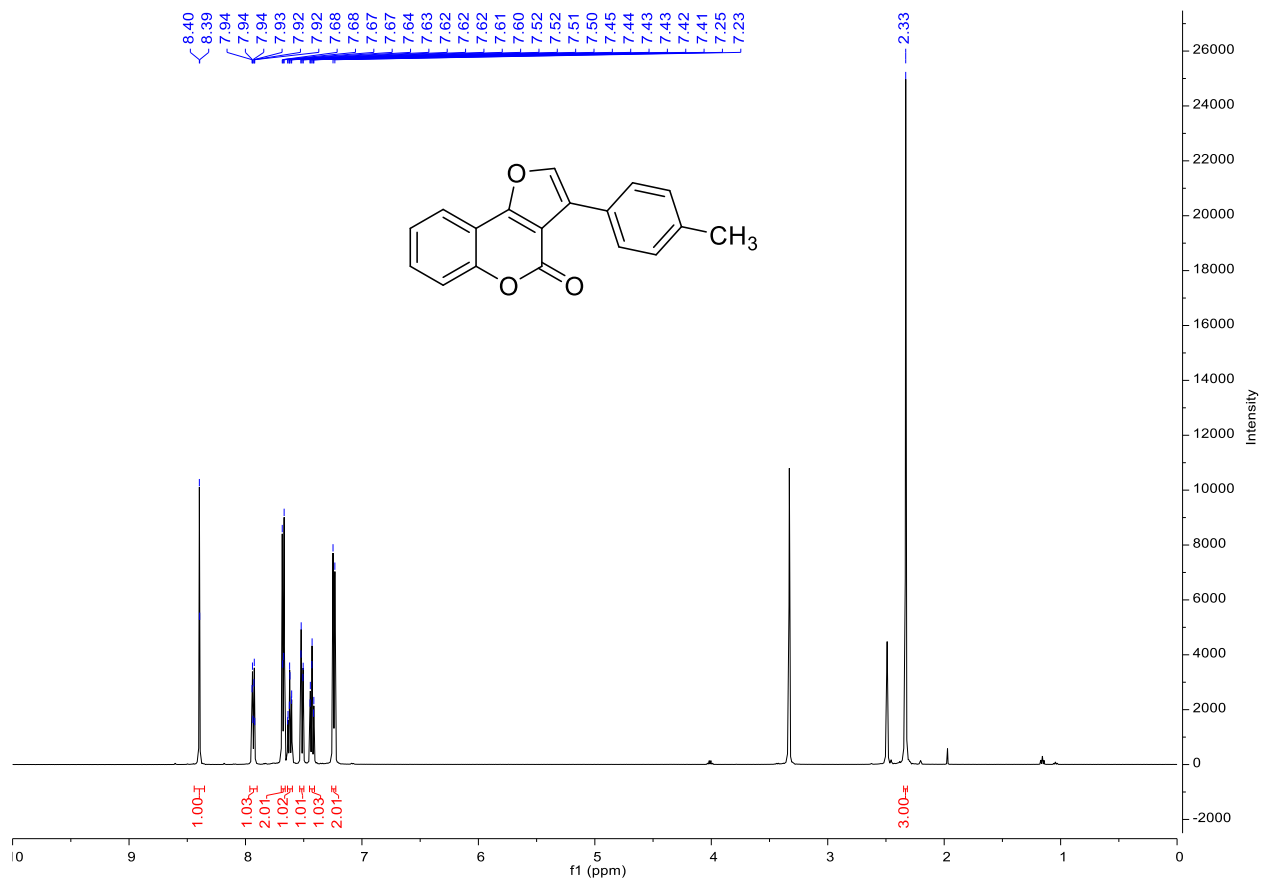


Fig. Sx. ¹H-NMR spectra of 3-(4-methylphenyl)-4H-furo[3,2-c]coumarin.

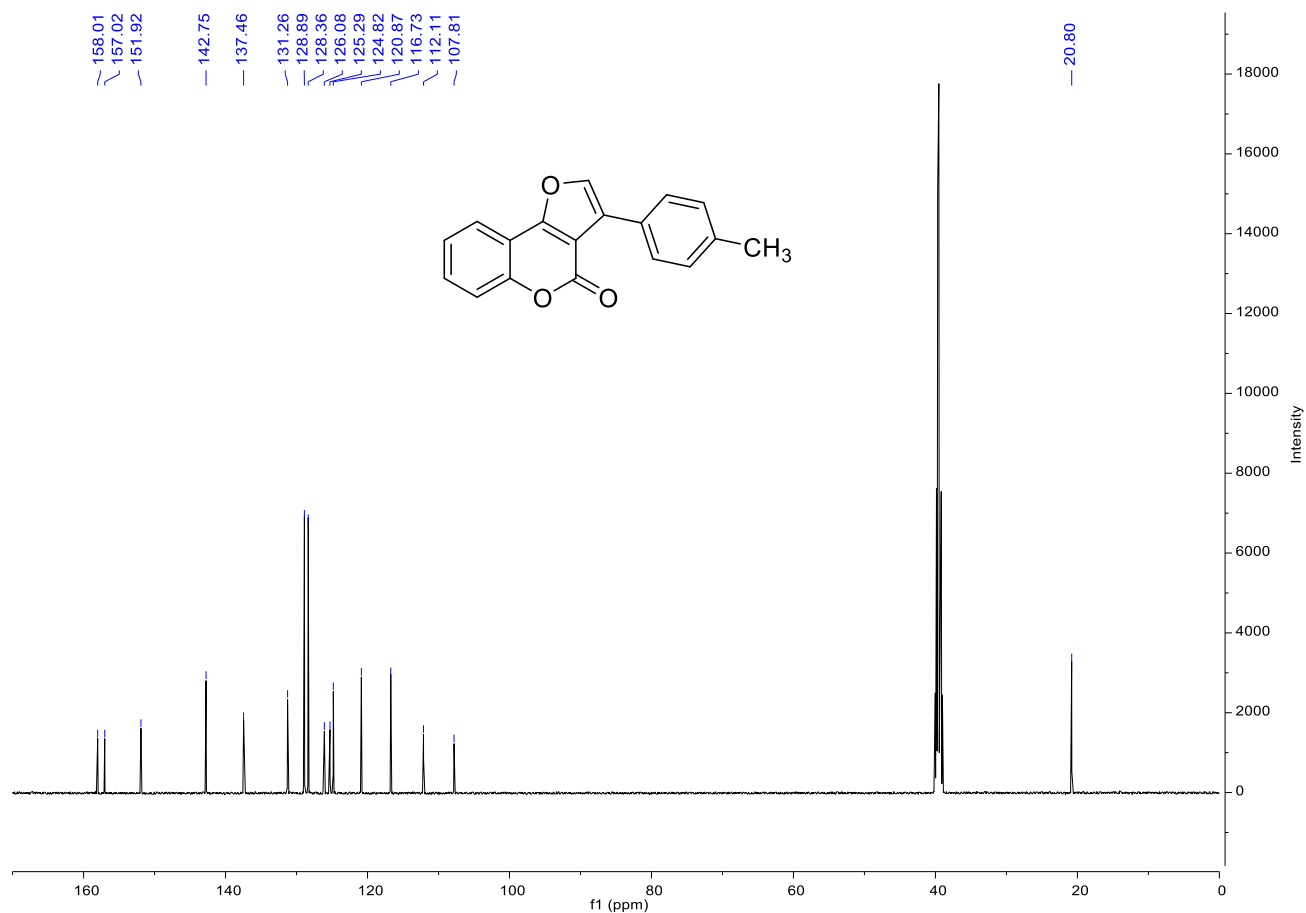
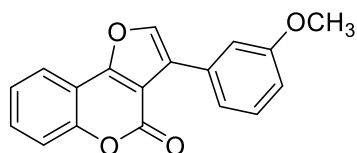


Fig. Sx. ¹³C-NMR spectra of 3-(4-methylphenyl)-4*H*-furo[3,2-*c*]coumarin.

Spectroscopic data for 3-(4-methoxyphenyl)-4*H*-furo[3,2-*c*]coumarin (HP3)



¹H NMR (500 MHz, DMSO-*d*₆) δ 8.52 (s, 1H), 8.00 (dd, *J* = 7.8, 1.6 Hz, 1H), 7.66 (ddd, *J* = 8.6, 7.3, 1.6 Hz, 1H), 7.56 (dd, *J* = 8.4, 1.0 Hz, 1H), 7.49 – 7.45 (m, 2H), 7.39 – 7.35 (m, 2H), 6.97 (dt, *J* = 7.4, 2.2 Hz, 1H), 3.81 (s, 3H).

¹³C NMR (126 MHz, DMSO) δ 159.17, 158.18, 157.06, 151.98, 143.38, 131.41, 130.27, 129.44, 125.25, 124.91, 120.98, 120.67, 116.79, 114.29, 113.61, 112.12, 107.83, 55.12.

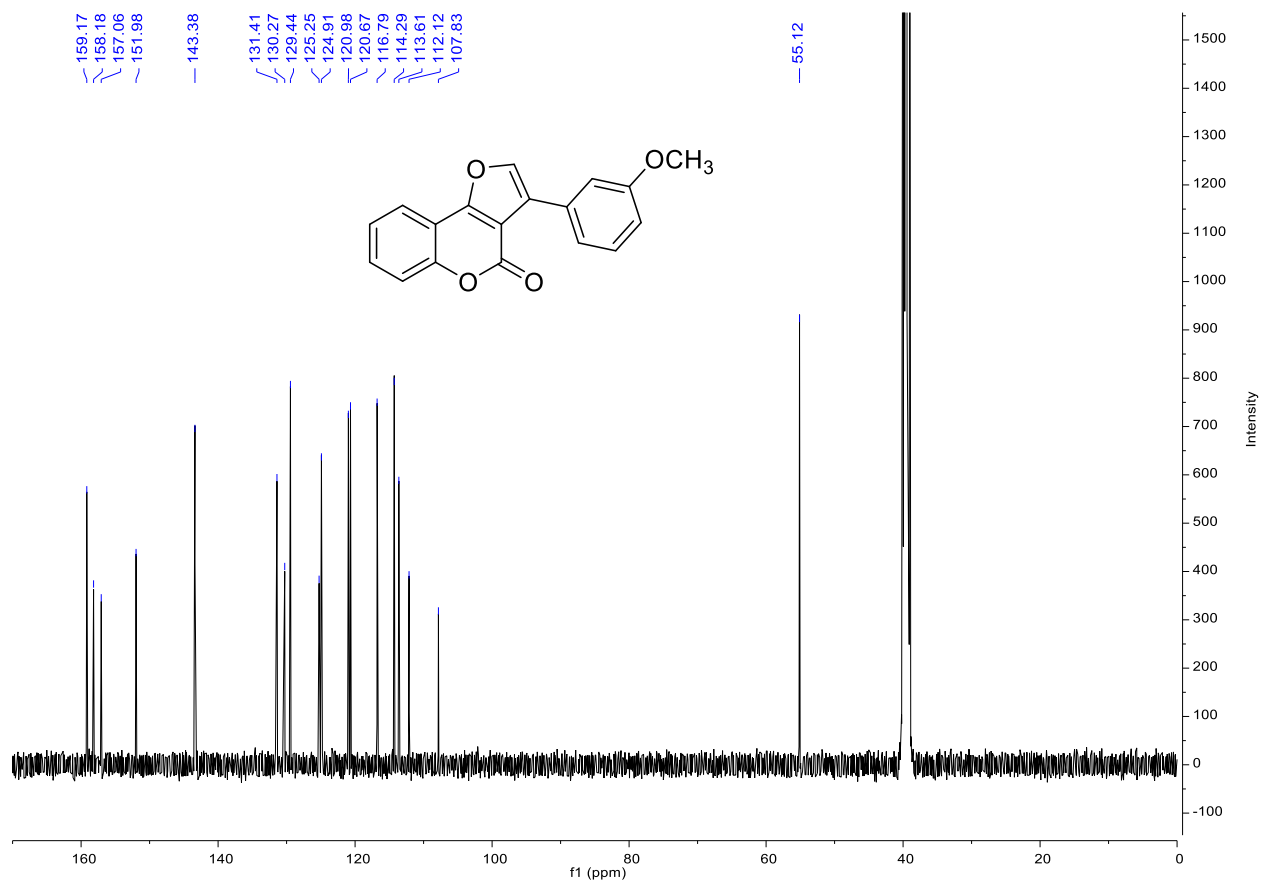
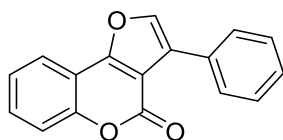


Fig. Sx. ¹³C-NMR spectra of 3-(4-methoxyphenyl)-4*H*-furo[3,2-*c*]coumarin.

Spectroscopic data for 3-phenyl-4*H*-furo[3,2-*c*]coumarin (HP4)



¹H NMR (500 MHz, CDCl₃) δ 7.92 (dd, *J* = 7.8, 1.8 Hz, 1H), 7.80 – 7.75 (m, 3H), 7.54 (ddd, *J* = 8.7, 7.2, 1.6 Hz, 1H), 7.48 – 7.44 (m, 3H), 7.38 (dt, *J* = 14.8, 7.4 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 158.96, 152.81, 141.38, 131.08, 129.24, 128.83, 128.71, 128.52, 126.94, 124.62, 121.13, 117.30, 112.97, 108.68.

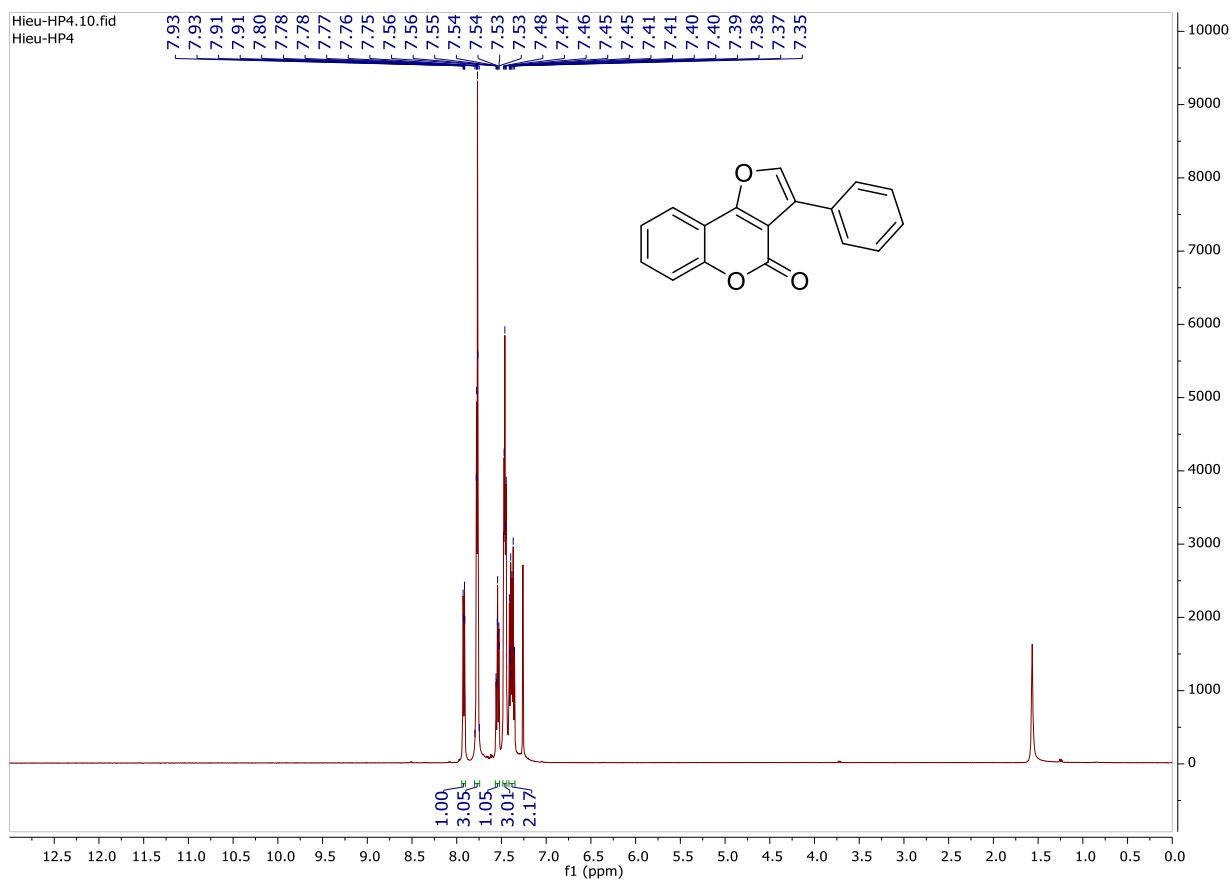


Fig. Sx. $^1\text{H-NMR}$ spectra of 3-phenyl-4*H*-furo[3,2-*c*]coumarin.

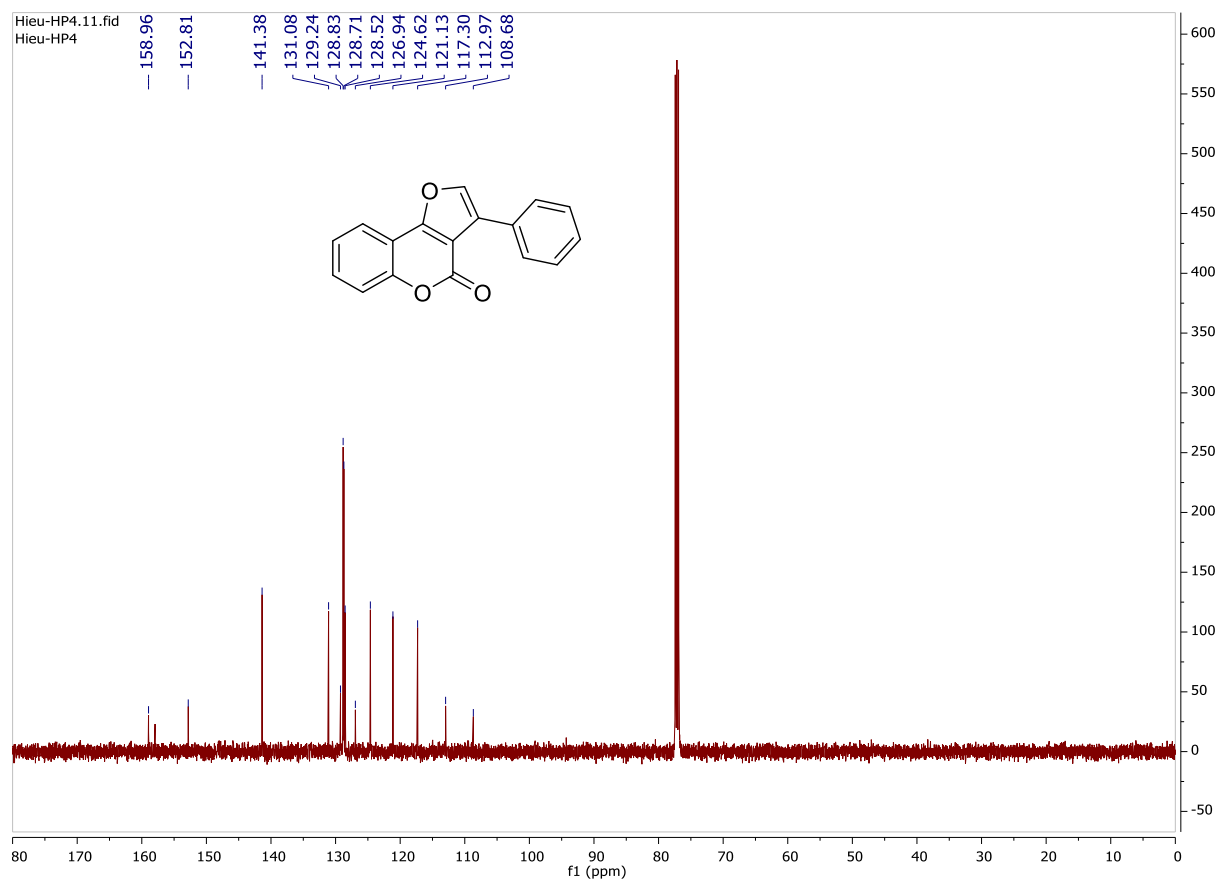
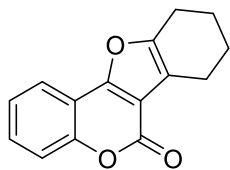
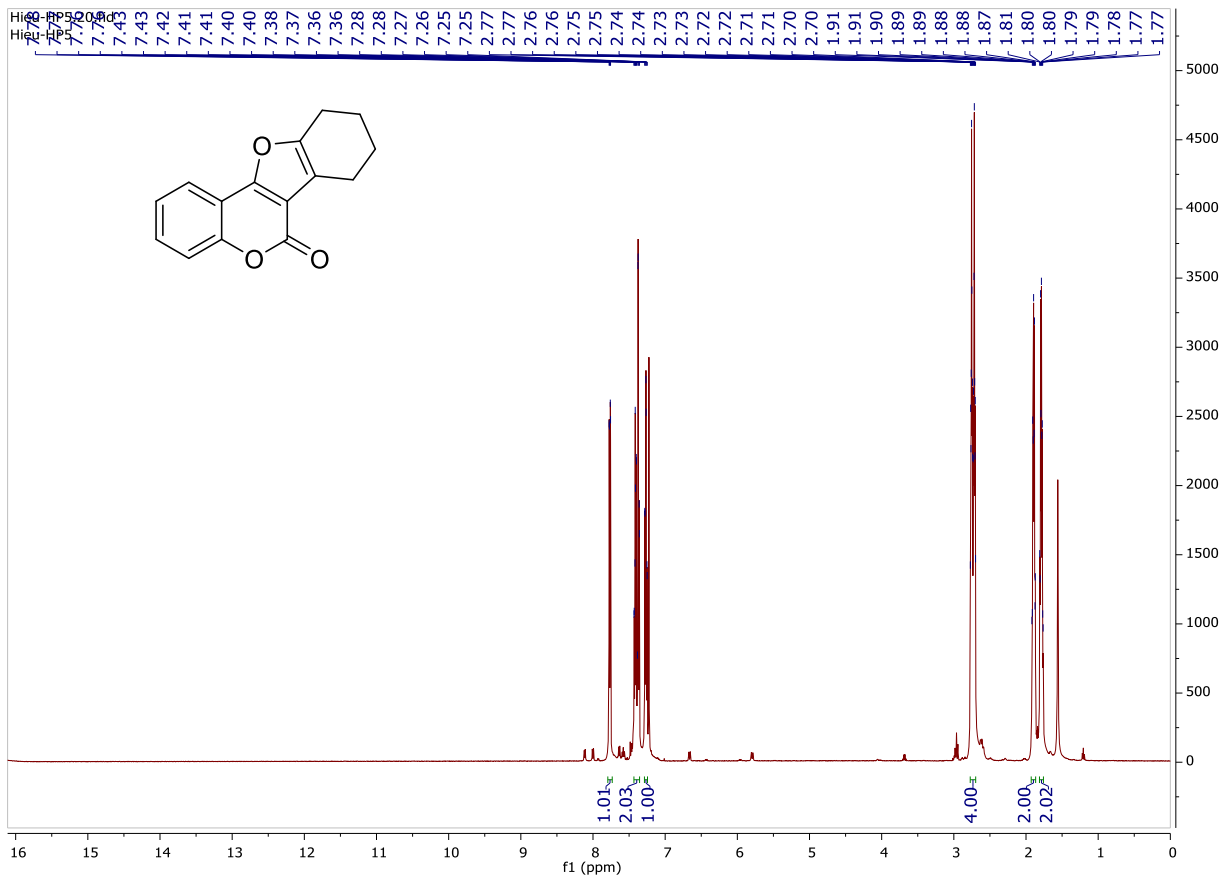


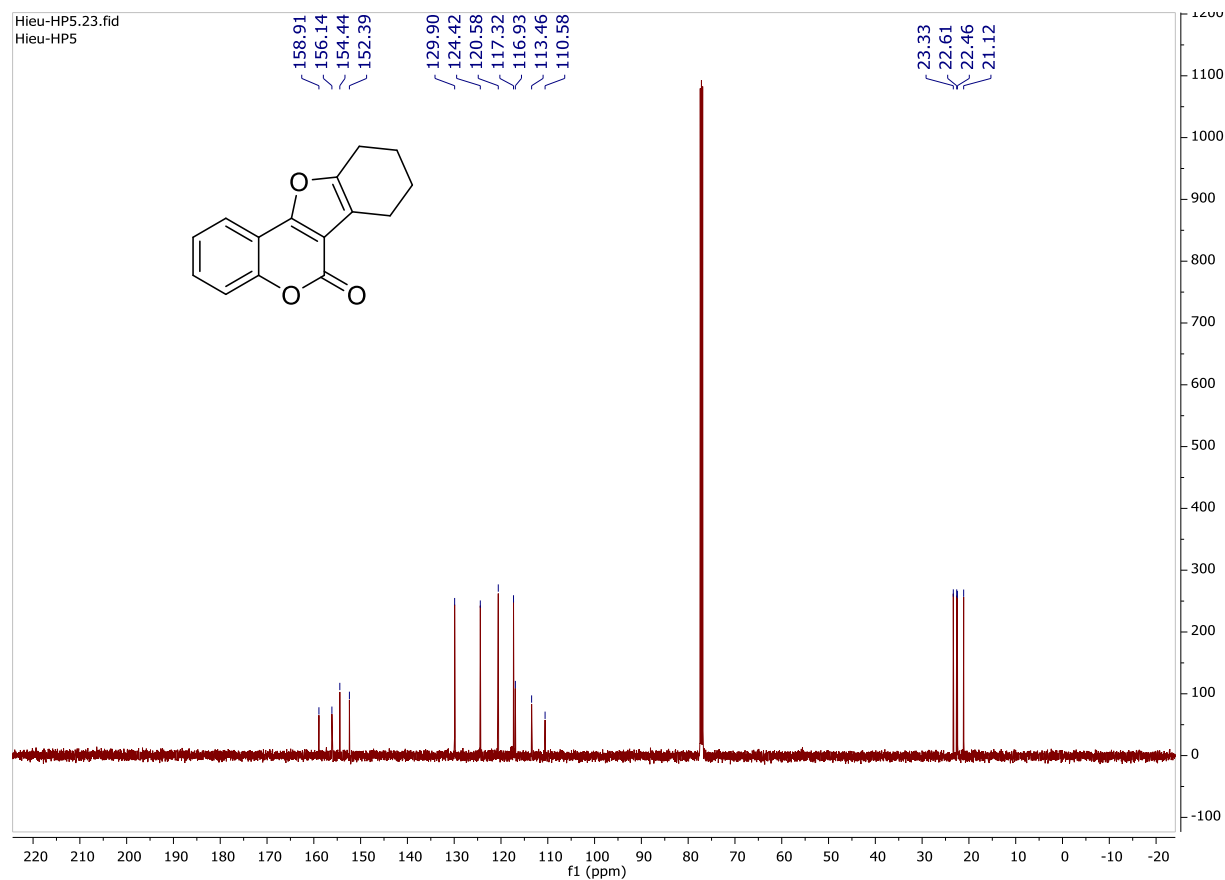
Fig. Sx. ^{13}C -NMR spectra of 3-phenyl-4*H*-furo[3,2-*c*]coumarin.

Spectroscopic data for 7,8,9,10-tetrahydro-6*H*-benzofuro[3,2-*c*]coumarin (HP5)

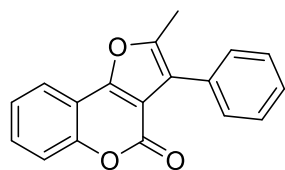


^1H NMR (500 MHz, CDCl_3) δ 7.77 (dd, $J = 7.8, 1.6$ Hz, 1H), 7.43 – 7.35 (m, 2H), 7.27 (td, $J = 7.6, 1.3$ Hz, 1H), 2.77 – 2.70 (m, 4H), 1.93 – 1.87 (m, 2H), 1.81 – 1.76 (m, 2H).
 ^{13}C NMR (126 MHz, CDCl_3) δ 158.91, 156.14, 154.44, 152.39, 129.90, 124.42, 120.58, 117.32, 116.93, 113.46, 110.58, 23.33, 22.61, 22.46, 21.12.



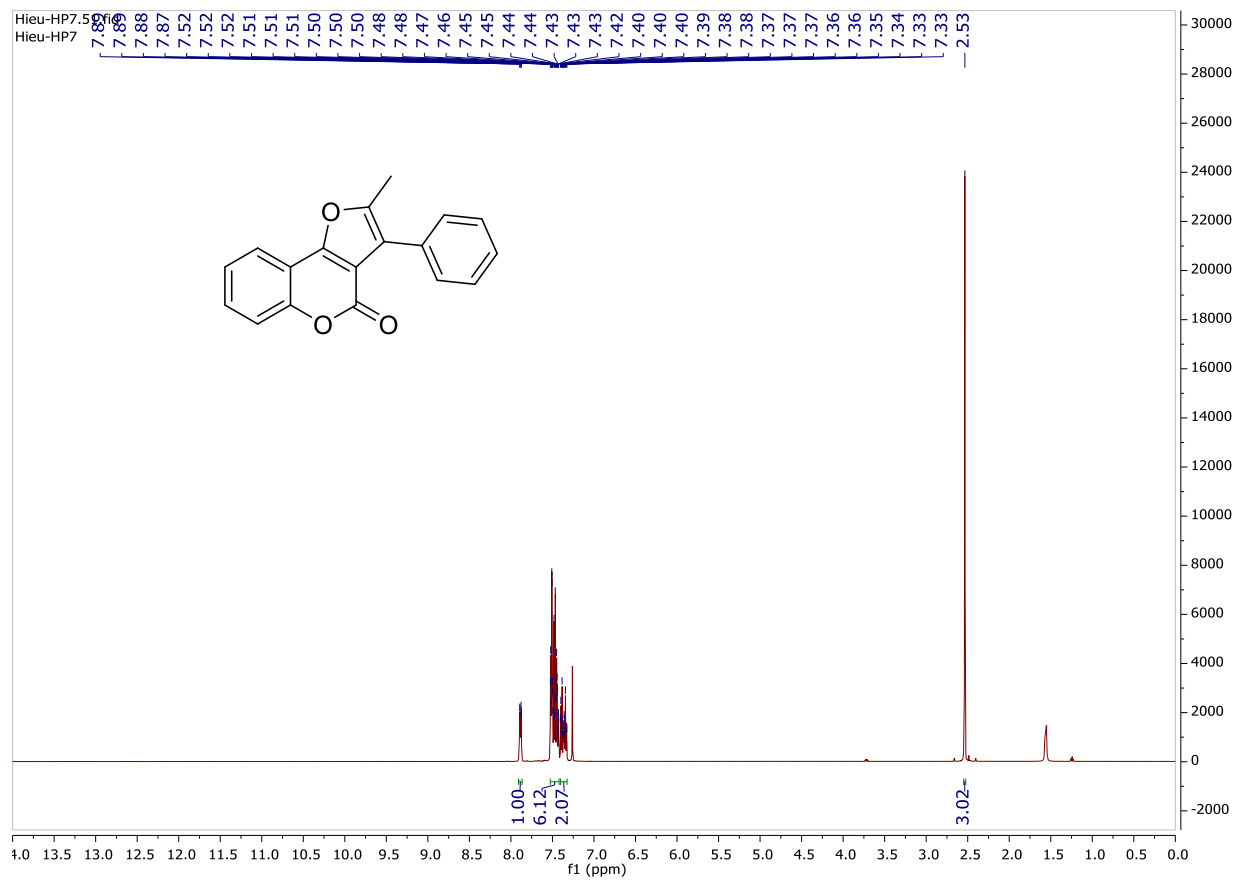


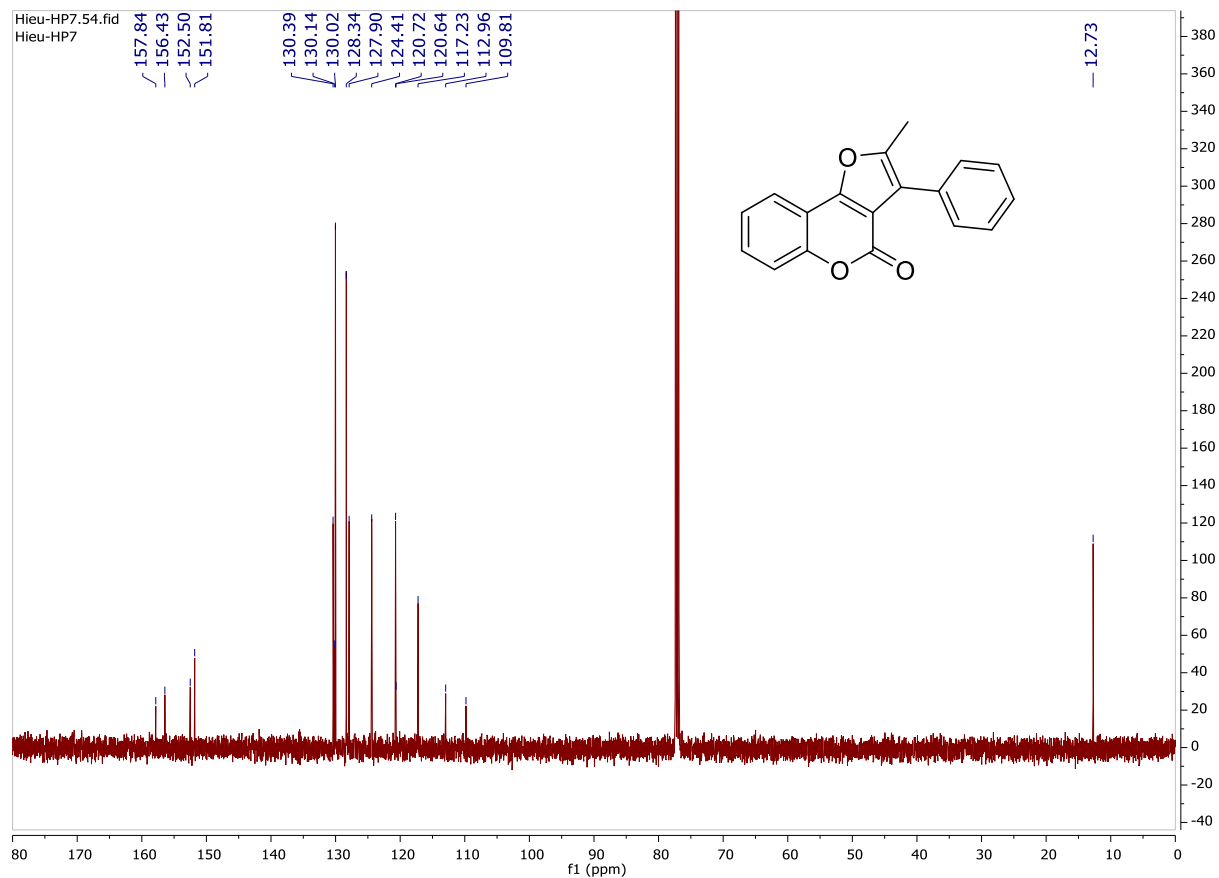
Spectroscopic data for 2-methyl-3-phenyl-4H-furo[3,2-c]coumarin (HP7)



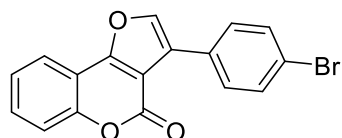
^1H NMR (500 MHz, CDCl_3) δ 7.88 (ddd, $J = 7.8, 1.6 \text{ Hz}, 0.5 \text{ Hz}$, 1H), 7.53 – 7.33 (m, 8H), 2.53 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 157.84, 156.43, 152.50, 151.81, 130.39, 130.14, 130.02, 128.34, 127.90, 124.41, 120.72, 120.64, 117.23, 112.96, 109.81, 12.73.





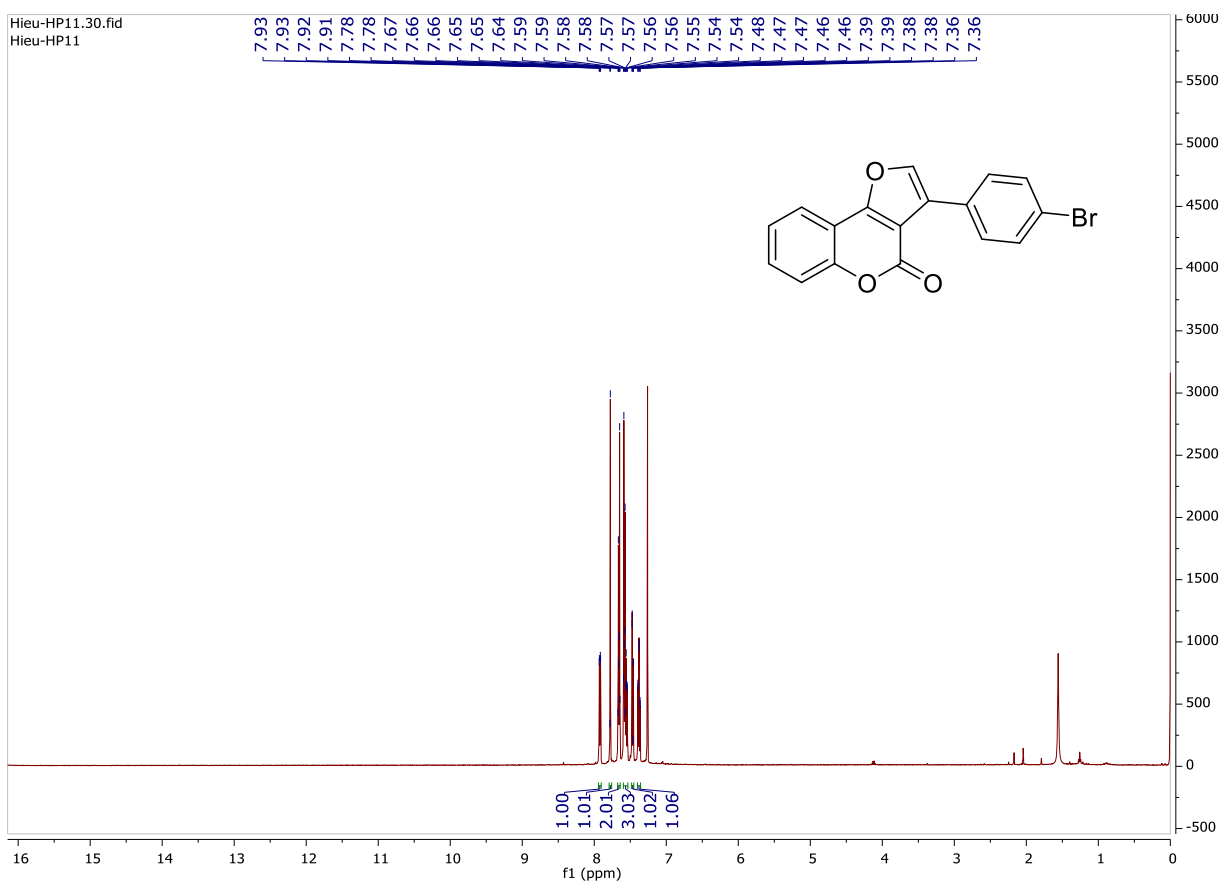
Spectroscopic data for 3-(4-bromophenyl)-4H-furo[3,2-c]coumarin (HP11)

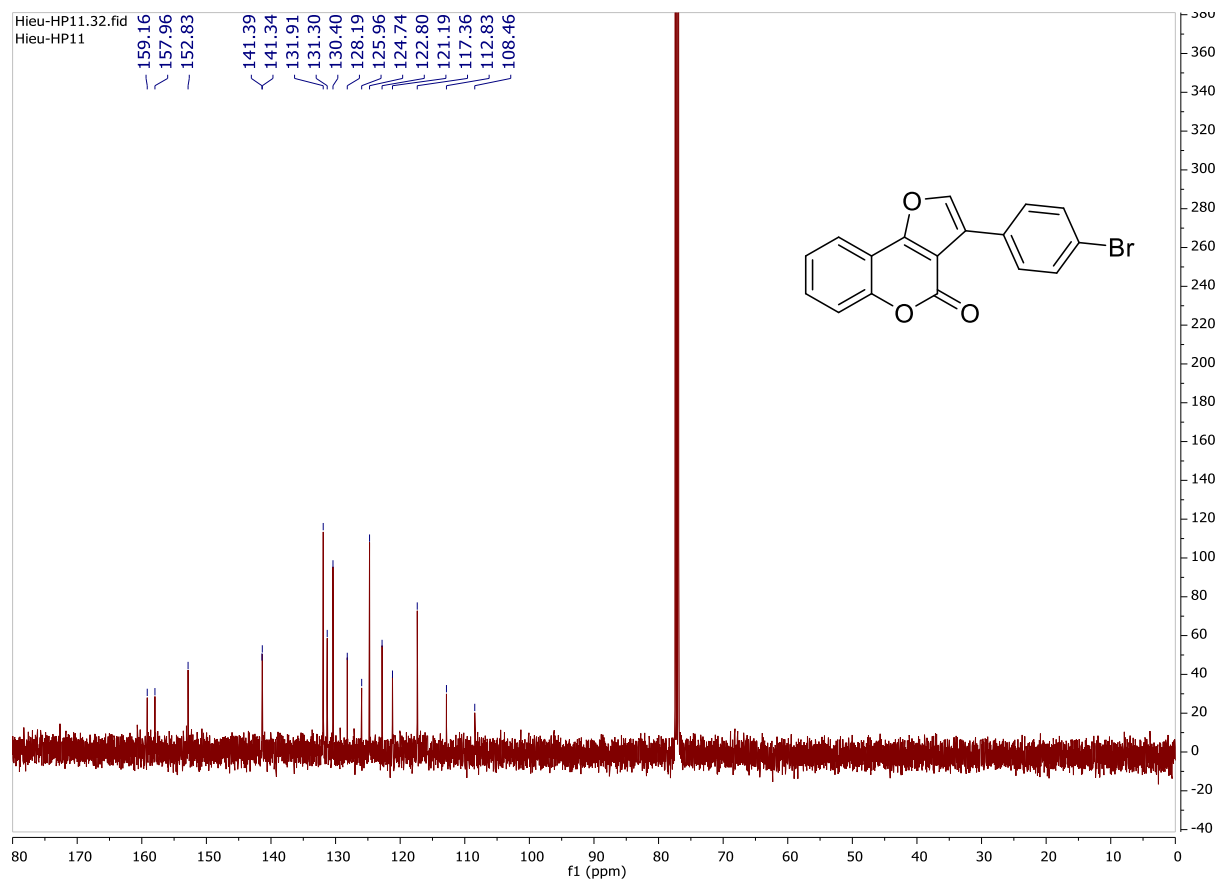


^1H NMR (500 MHz CDCl_3) δ 7.92 (dd, $J = 7.9, 1.6$ Hz, 1H), 7.78 (s, 1H), 7.67 – 7.53 (m, 5H), 7.47 (dd, $J = 8.4, 1.1$ Hz, 1H), 7.38 (td, $J = 7.5, 1.1$ Hz, 1H).

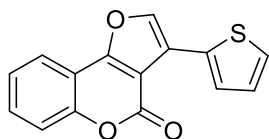
^{13}C NMR (126 MHz, CDCl_3) δ 159.16, 157.96, 152.83, 141.39, 141.34, 131.91, 131.30, 130.40, 128.19, 125.96, 124.74, 122.80, 121.19, 117.36, 112.83, 108.46.

Hieu-HP11.30.fid
Hieu-HP11



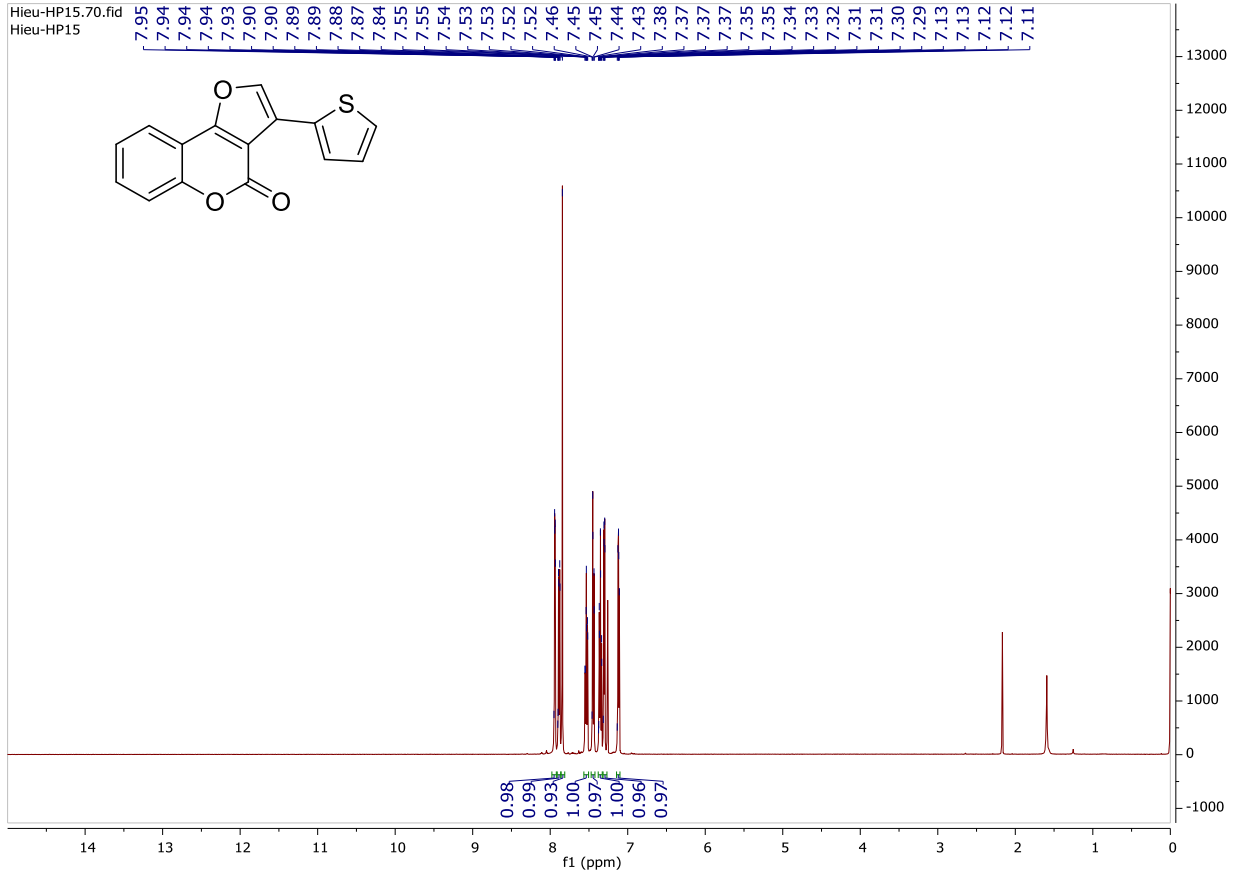


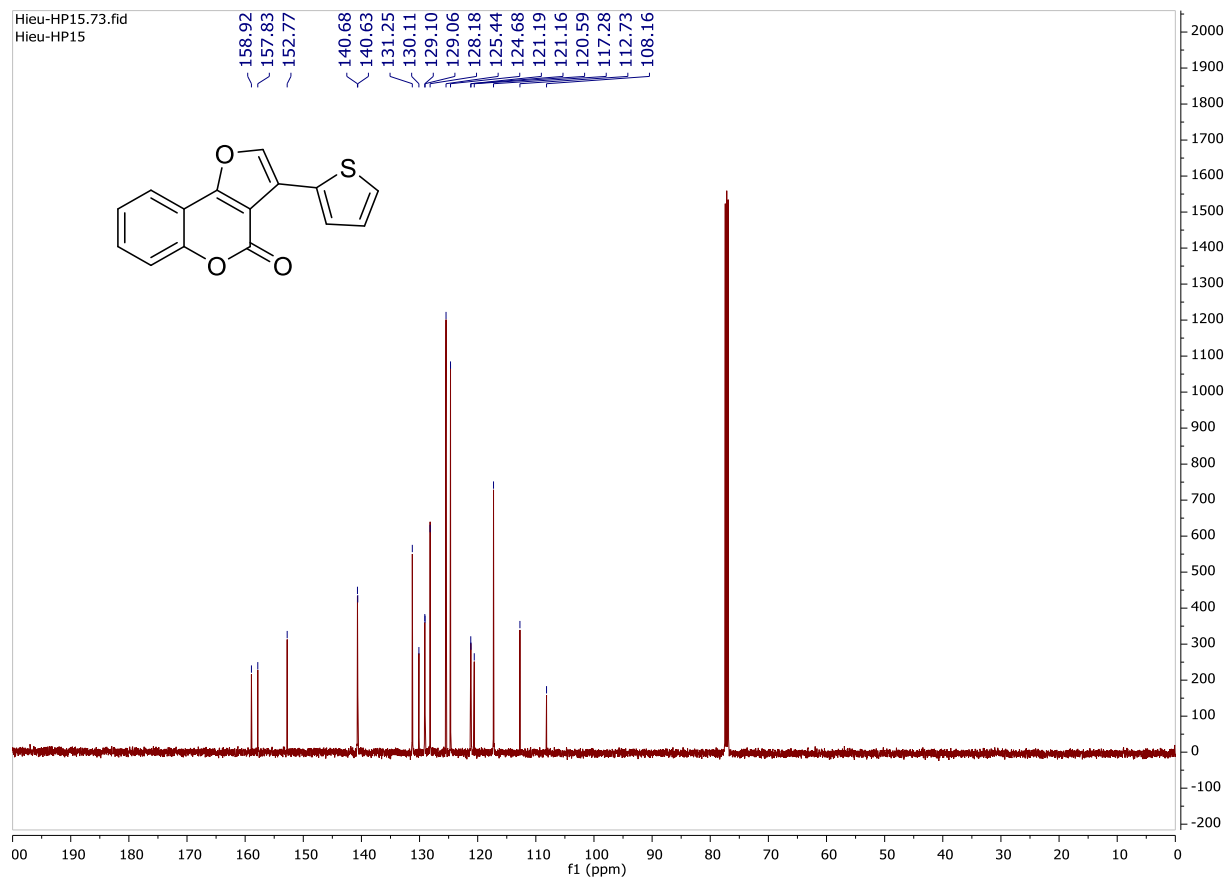
Spectroscopic data for 3-(thiophen-2-yl)-4H-furo[3,2-c]coumarin (HP15)



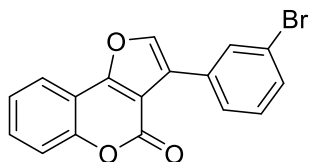
^1H NMR (500 MHz, CDCl_3) δ 7.94 (dd, $J = 3.7, 1.1$ Hz, 1H), 7.88 (dd, $J = 7.9, 1.6$ Hz, 1H), 7.84 (s, 1H), 7.53 (ddd, $J = 8.7, 7.3, 1.6$ Hz, 1H), 7.44 (dd, $J = 8.4, 1.1$ Hz, 1H), 7.35 (td, $J = 7.5, 1.1$ Hz, 1H), 7.30 (dd, $J = 5.1, 1.1$ Hz, 1H), 7.12 (dd, $J = 5.1, 3.6$ Hz, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 158.92, 157.83, 152.77, 140.68, 140.63, 131.25, 130.11, 129.10, 129.06, 128.18, 125.44, 124.68, 121.19, 121.16, 120.59, 117.28, 112.73, 108.16.



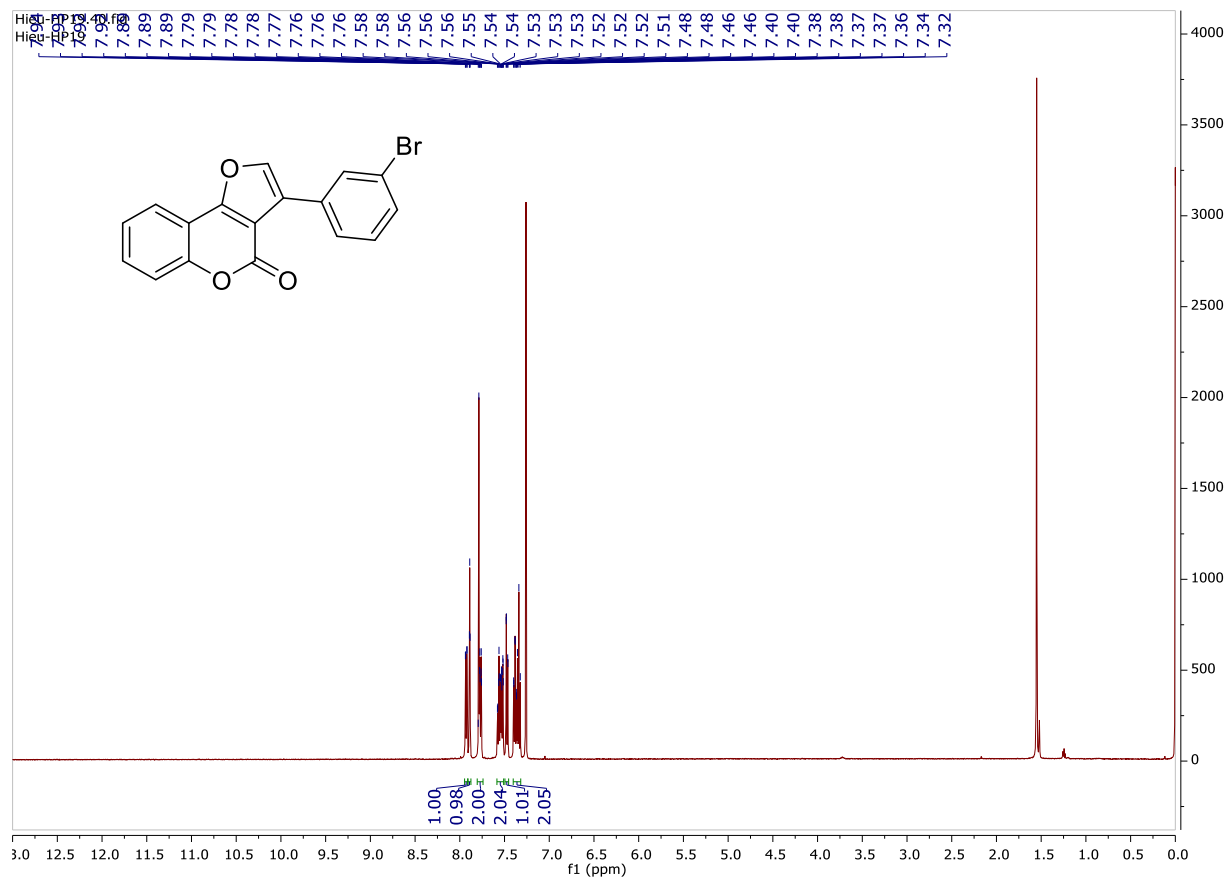


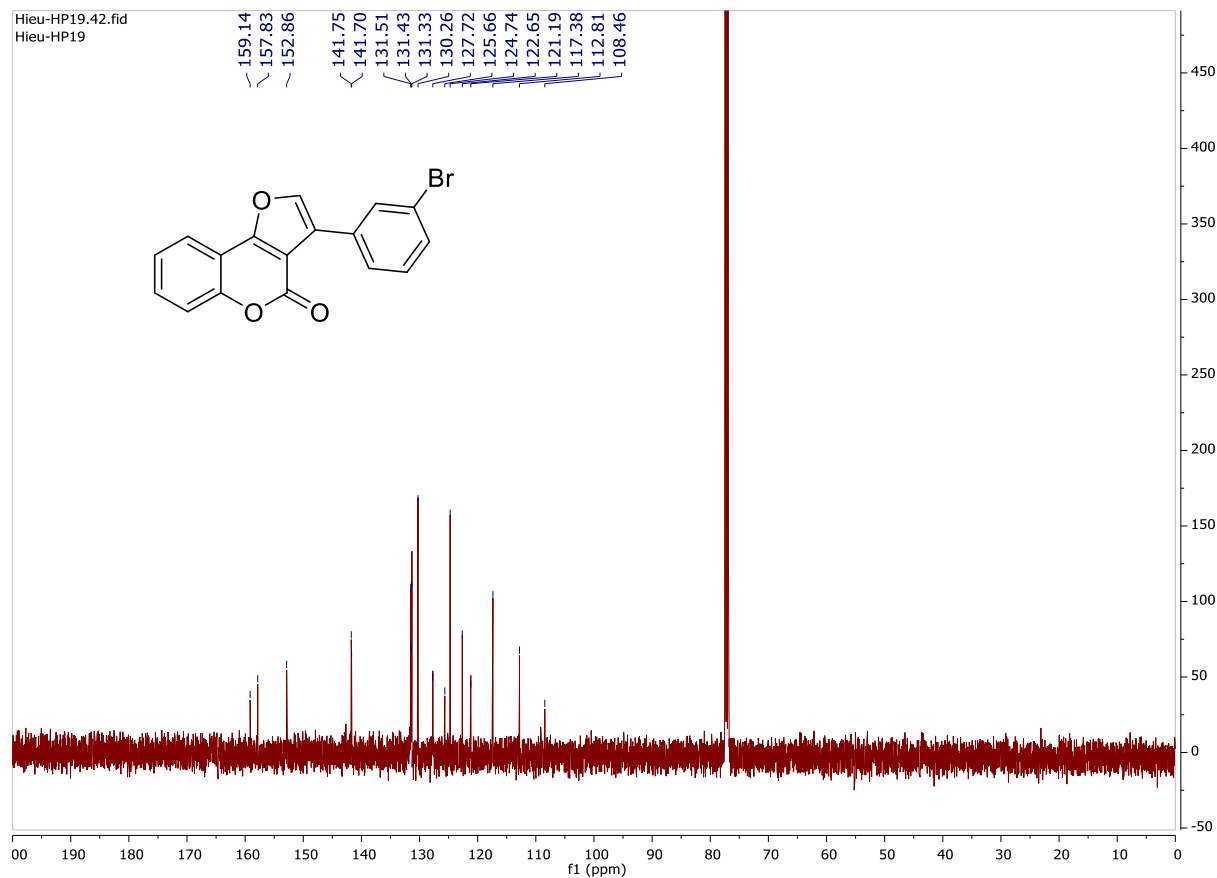
Spectroscopic data for 3-(3-bromophenyl)-4H-furo[3,2-c]coumarin (HP19)



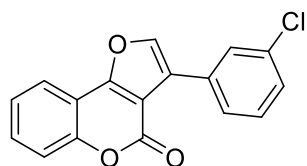
^1H NMR (500 MHz, CDCl_3) δ 7.93 (dd, $J = 7.7, 1.6$ Hz, 1H), 7.89 (t, $J = 1.8$ Hz, 1H), 7.80 – 7.74 (m, 2H), 7.58 – 7.51 (m, 2H), 7.47 (dd, $J = 8.4, 1.1$ Hz, 1H), 7.40 – 7.32 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 159.14, 157.83, 152.86, 141.75, 141.70, 131.51, 131.43, 131.33, 130.26, 127.72, 125.66, 124.74, 122.65, 121.19, 117.38, 112.81, 108.46.





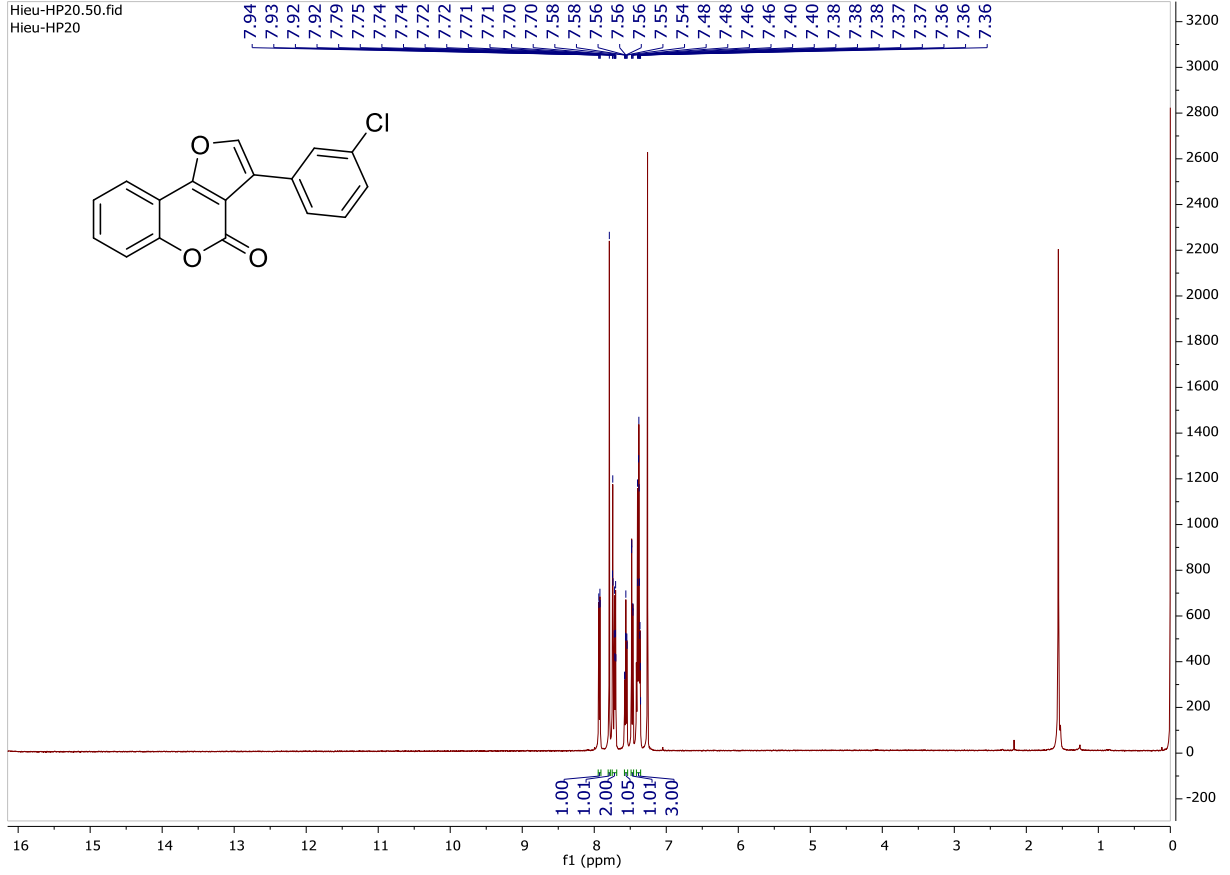
Spectroscopic data for 3-(3-chlorophenyl)-4H-furo[3,2-c]coumarin (HP20)

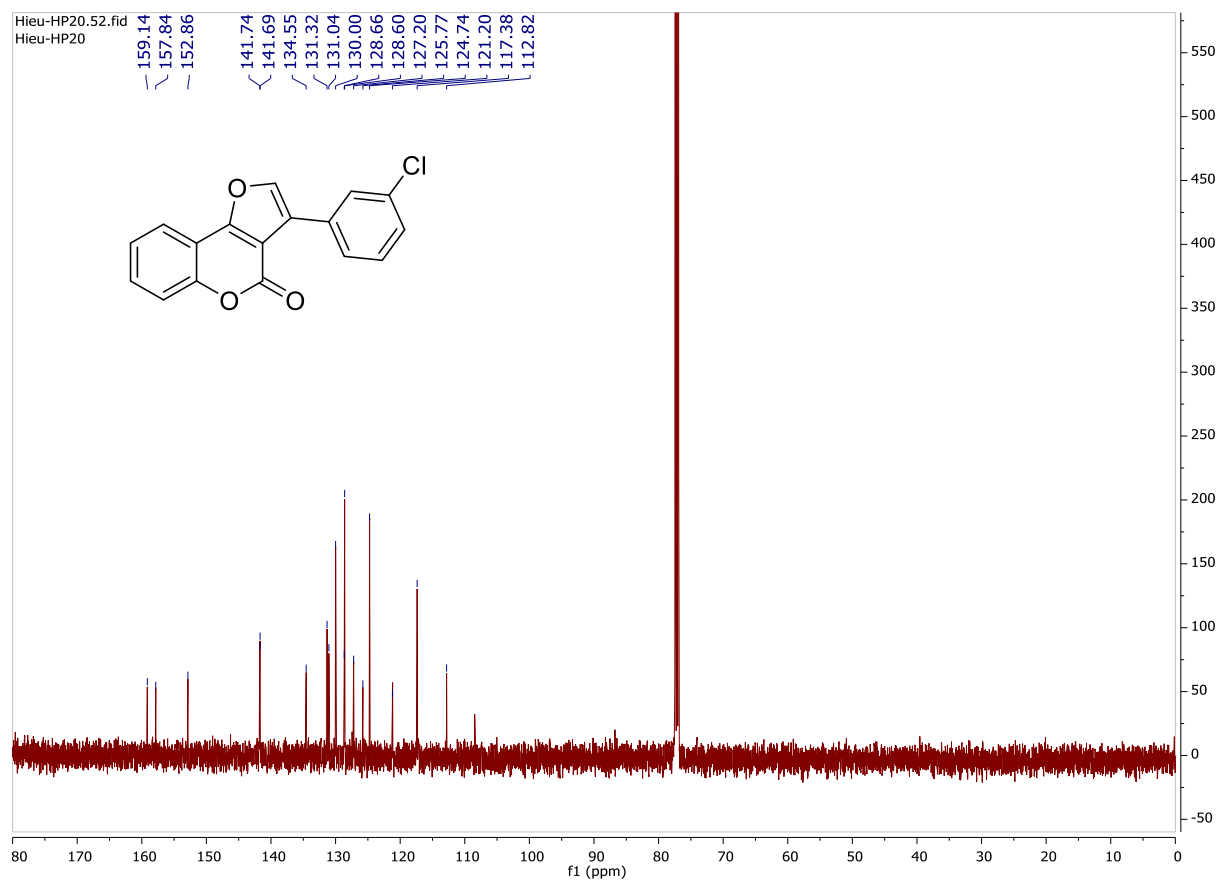


^1H NMR (500 MHz, CDCl_3) δ 7.93 (dd, $J = 7.8, 1.6$ Hz, 1H), 7.79 (s, 1H), 7.75 – 7.69 (m, 2H), 7.56 (ddd, $J = 8.7, 7.3, 1.7$ Hz, 1H), 7.47 (dd, $J = 8.4, 1.1$ Hz, 1H), 7.41 – 7.36 (m, 3H).

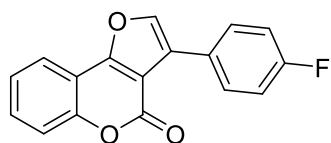
^{13}C NMR (126 MHz, CDCl_3) δ 159.14, 157.84, 152.86, 141.74, 141.69, 134.55, 131.32, 131.04, 130.00, 128.66, 128.60, 127.20, 125.77, 124.74, 121.20, 117.38, 112.82.

Hieu-HP20.50.fid
Hieu-HP20





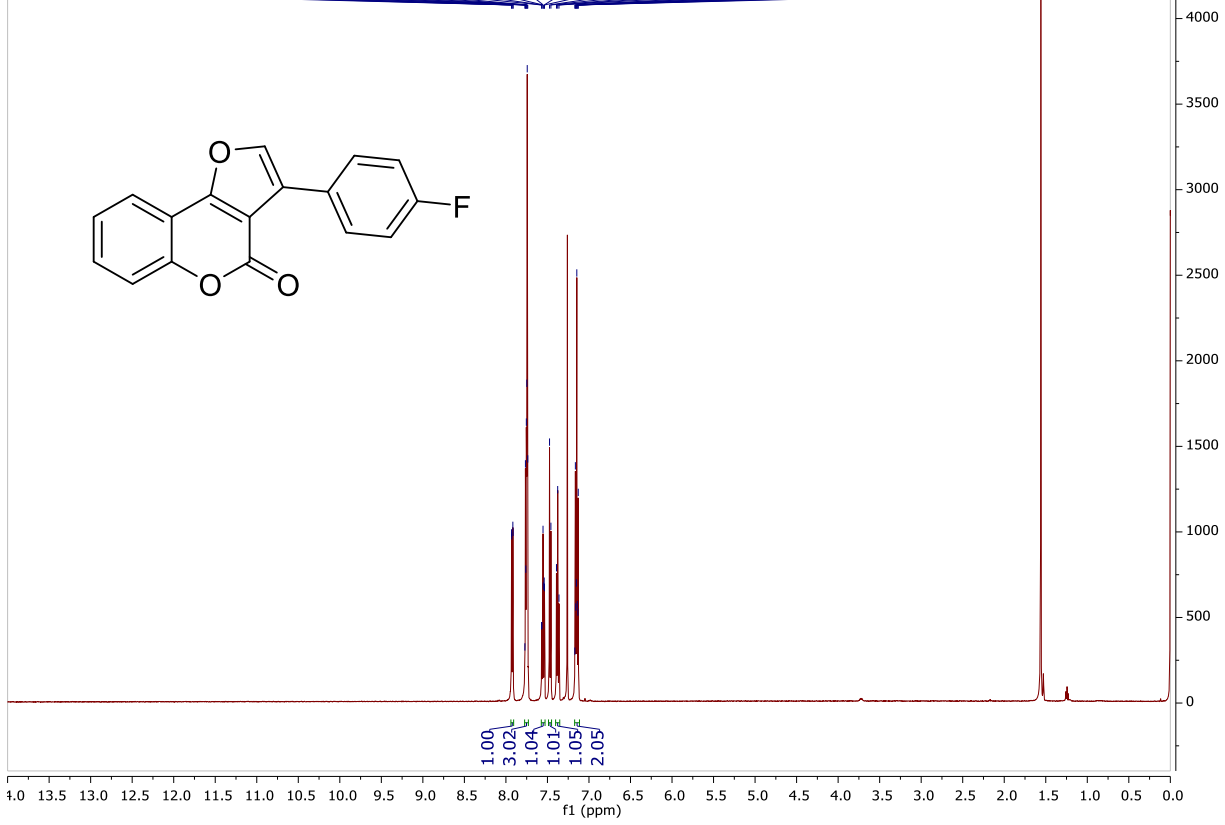
Spectroscopic data for 3-(3-fluorophenyl)-4H-furo[3,2-c]coumarin (HP21)

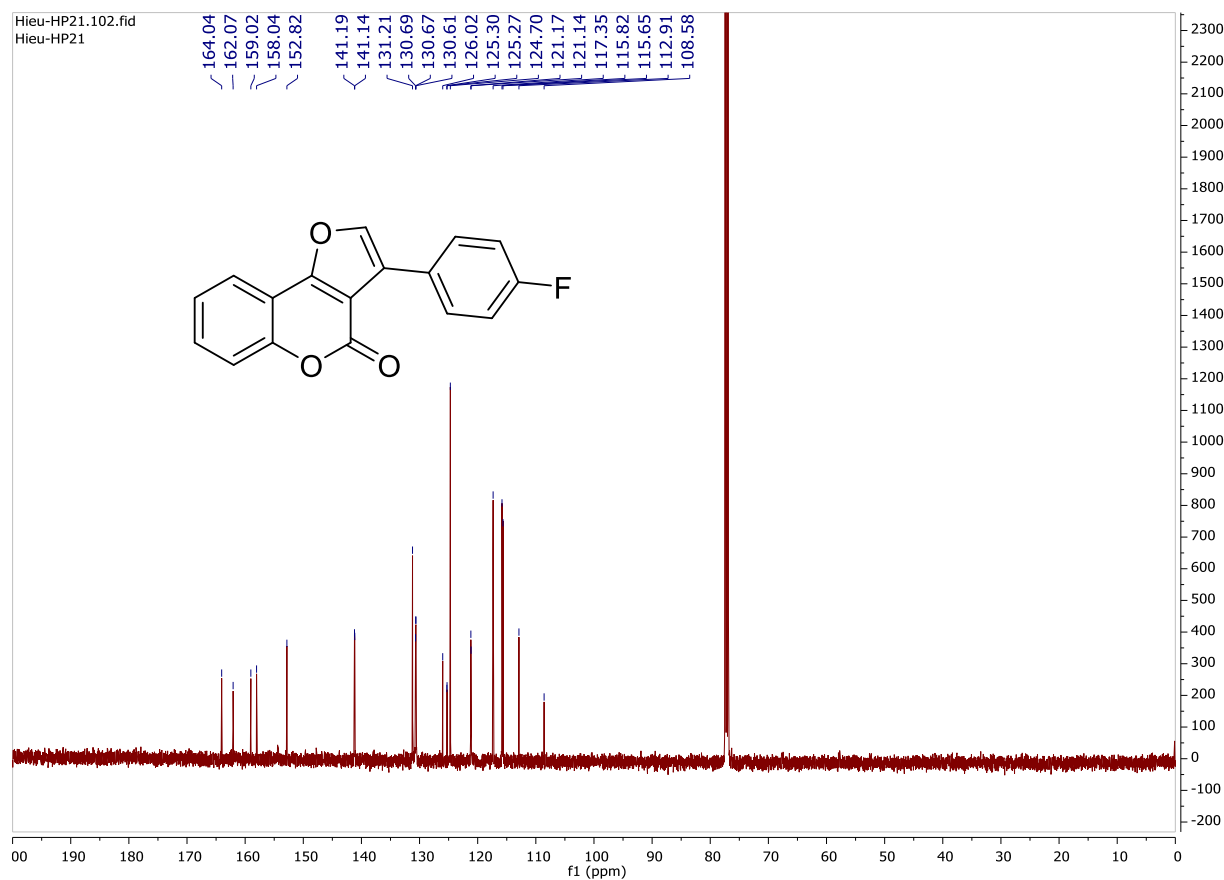


^1H NMR (500 MHz, CDCl_3) δ 7.92 (dd, $J = 7.8, 1.6$ Hz, 1H), 7.78 – 7.73 (m, 3H), 7.55 (td, $J = 7.8, 7.1, 1.6$ Hz, 1H), 7.47 (d, $J = 8.3$ Hz, 1H), 7.38 (t, $J = 7.5$ Hz, 1H), 7.17 – 7.12 (m, 2H).

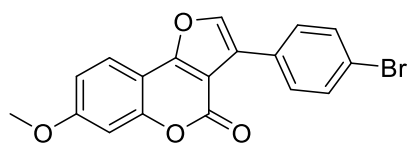
^{13}C NMR (126 MHz, CDCl_3) δ 164.04, 162.07, 159.02, 158.04, 152.82, 141.19, 141.14, 131.21, 130.69, 130.67, 130.61, 126.02, 125.30, 125.27, 124.70, 121.17, 121.14, 117.35, 115.82, 115.65, 112.91, 108.58.

Hieu-HP21.101.fid
Hieu-HP21



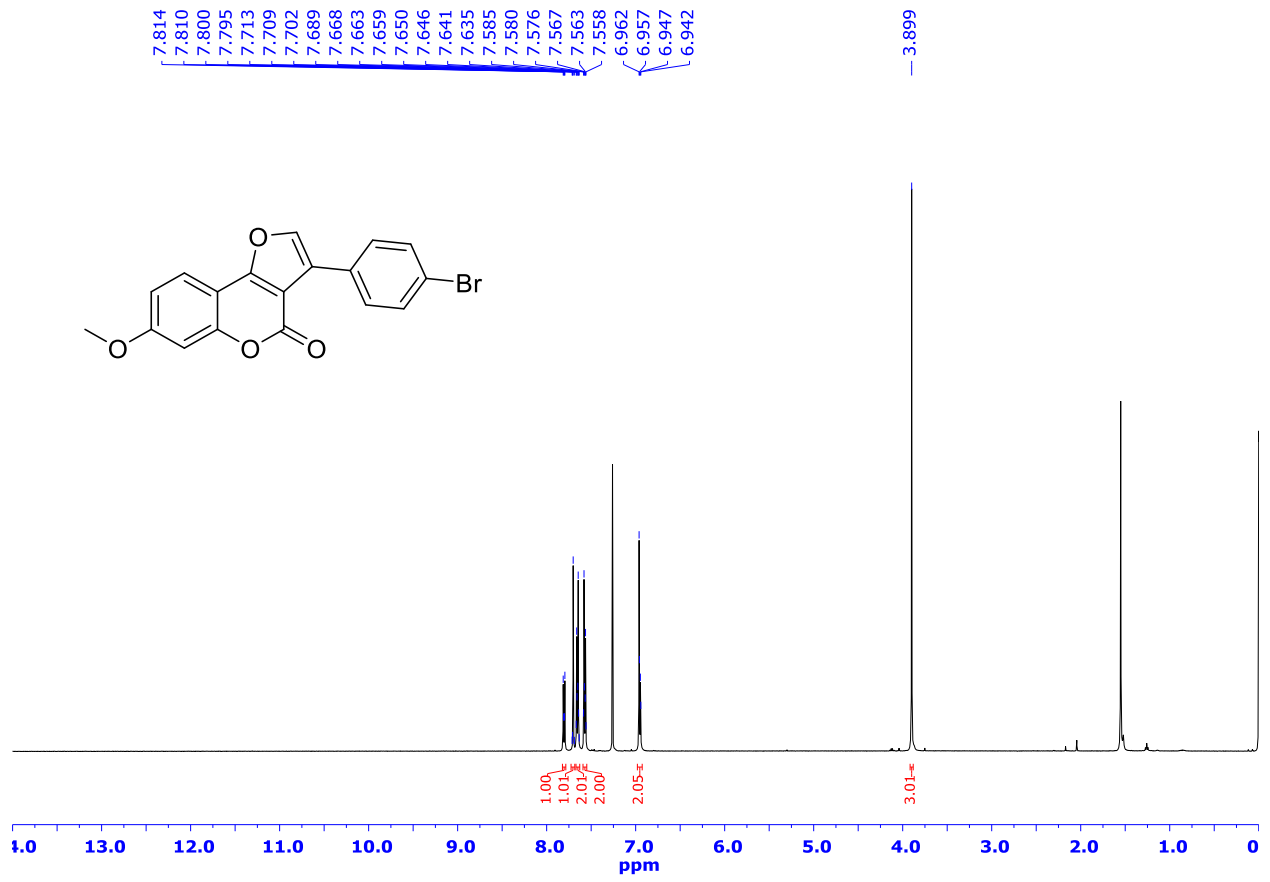


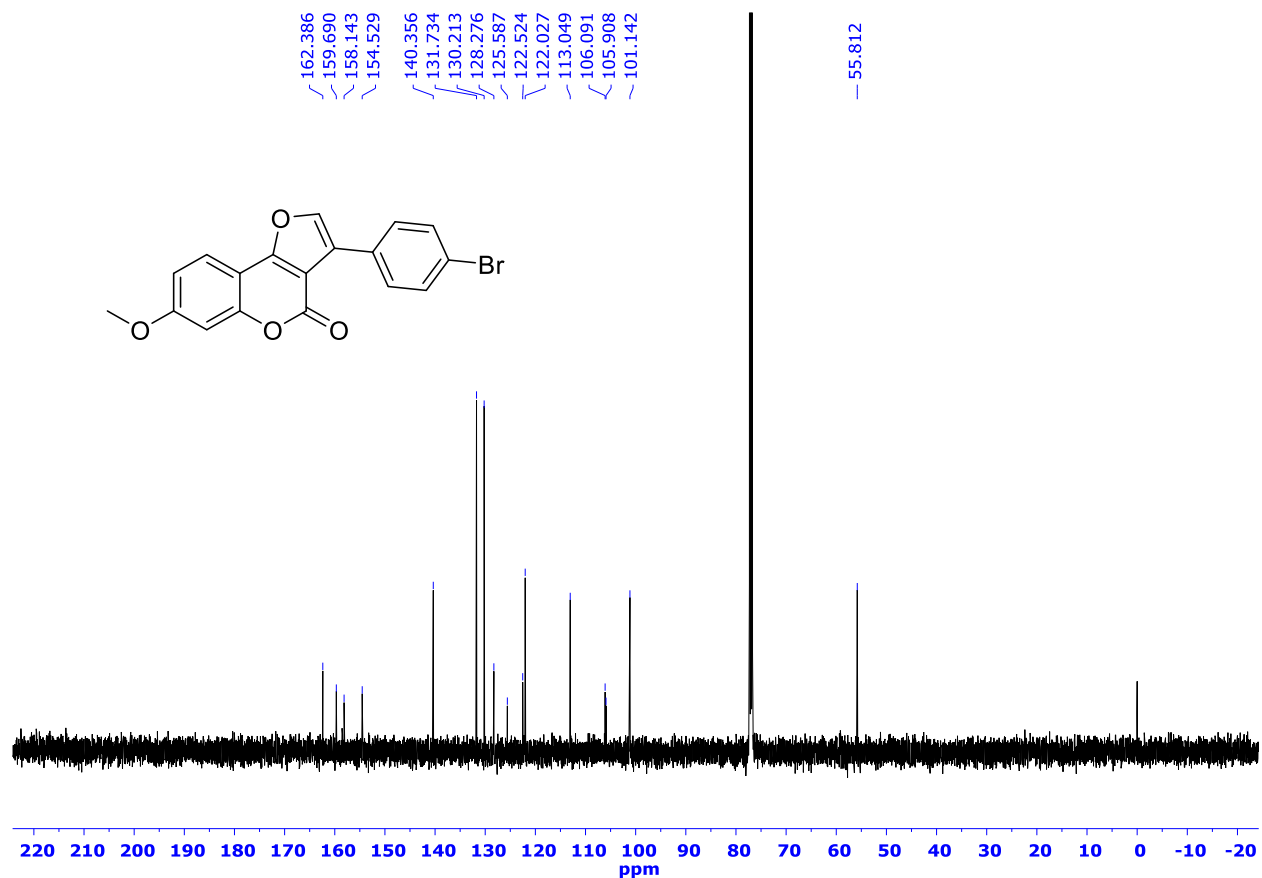
Spectroscopic data for 3-(4-bromophenyl)-7-methoxy-4H-furo[3,2-c]coumarin (HP24)



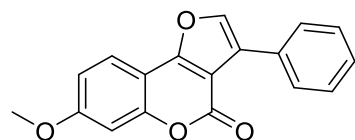
^1H NMR (500 MHz, CDCl_3) δ 7.82 – 7.78 (m, 1H), 7.70 (s, 1H), 7.67 – 7.63 (m, 2H), 7.59 – 7.55 (m, 2H), 6.98 – 6.93 (m, 2H), 3.90 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 162.39, 159.69, 158.14, 154.53, 140.36, 131.73, 130.21, 128.28, 125.59, 122.52, 122.03, 113.05, 106.09, 105.91, 101.14, 55.81.



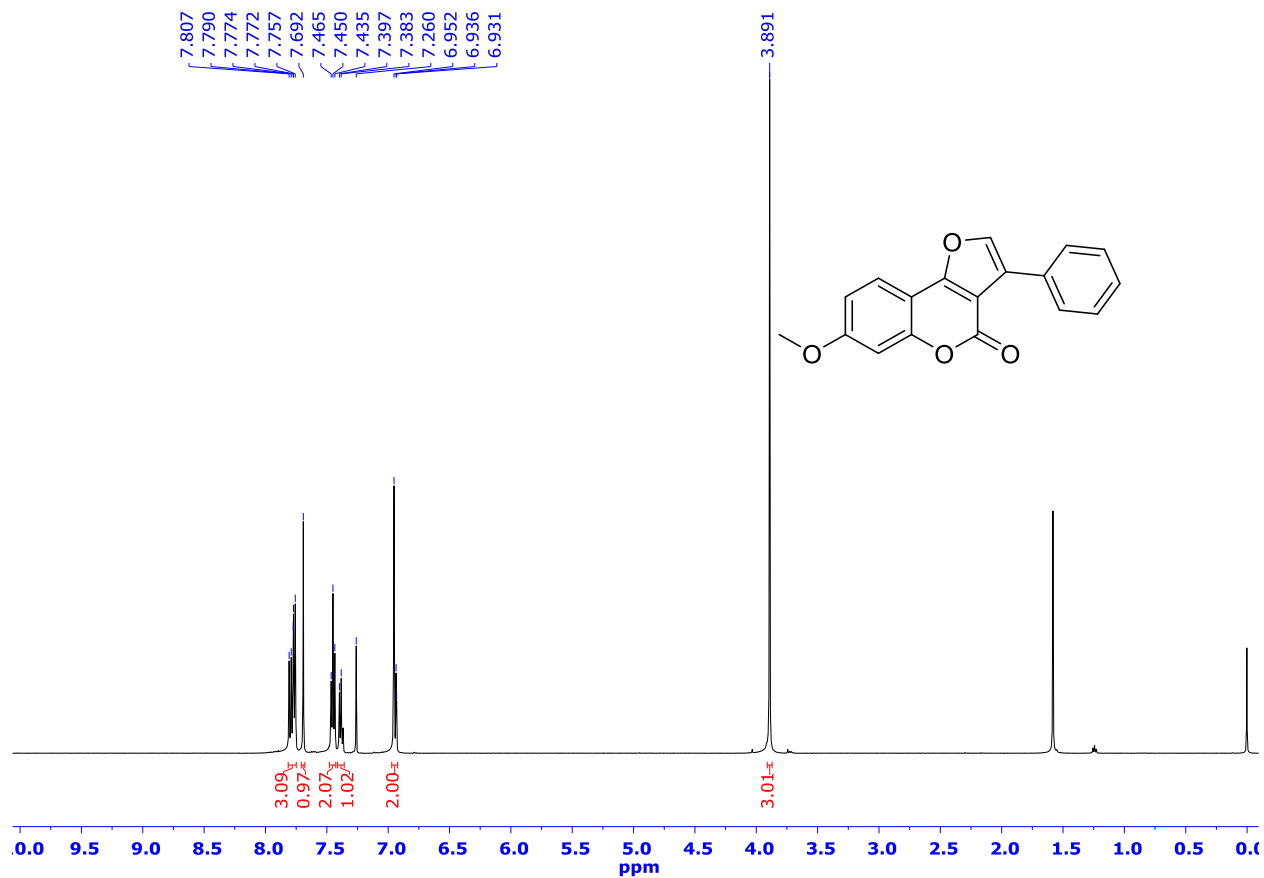


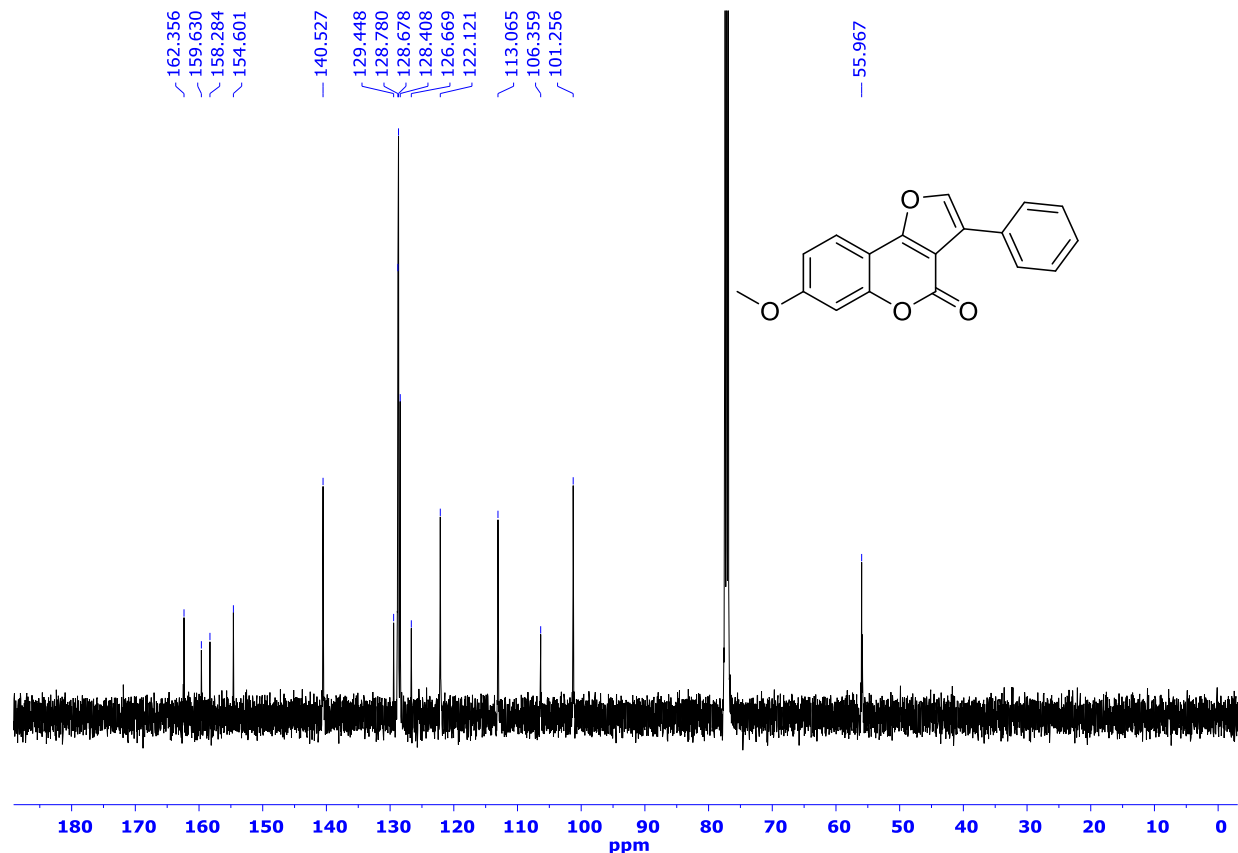
Spectroscopic data for 3-phenyl-7-methoxy-4H-furo[3,2-c]coumarin (HP25)



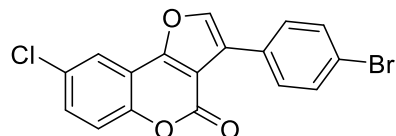
^1H NMR (500 MHz, CDCl_3) δ 7.81 – 7.75 (m, 3H), 7.69 (s, 1H), 7.45 (t, $J = 7.5$ Hz, 2H), 7.39 (d, $J = 7.3$ Hz, 1H), 6.97 – 6.92 (m, 2H), 3.89 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 162.36, 159.63, 158.28, 154.60, 140.53, 129.45, 128.78, 128.68, 128.41, 126.67, 122.12, 113.07, 106.36, 101.26, 55.97.



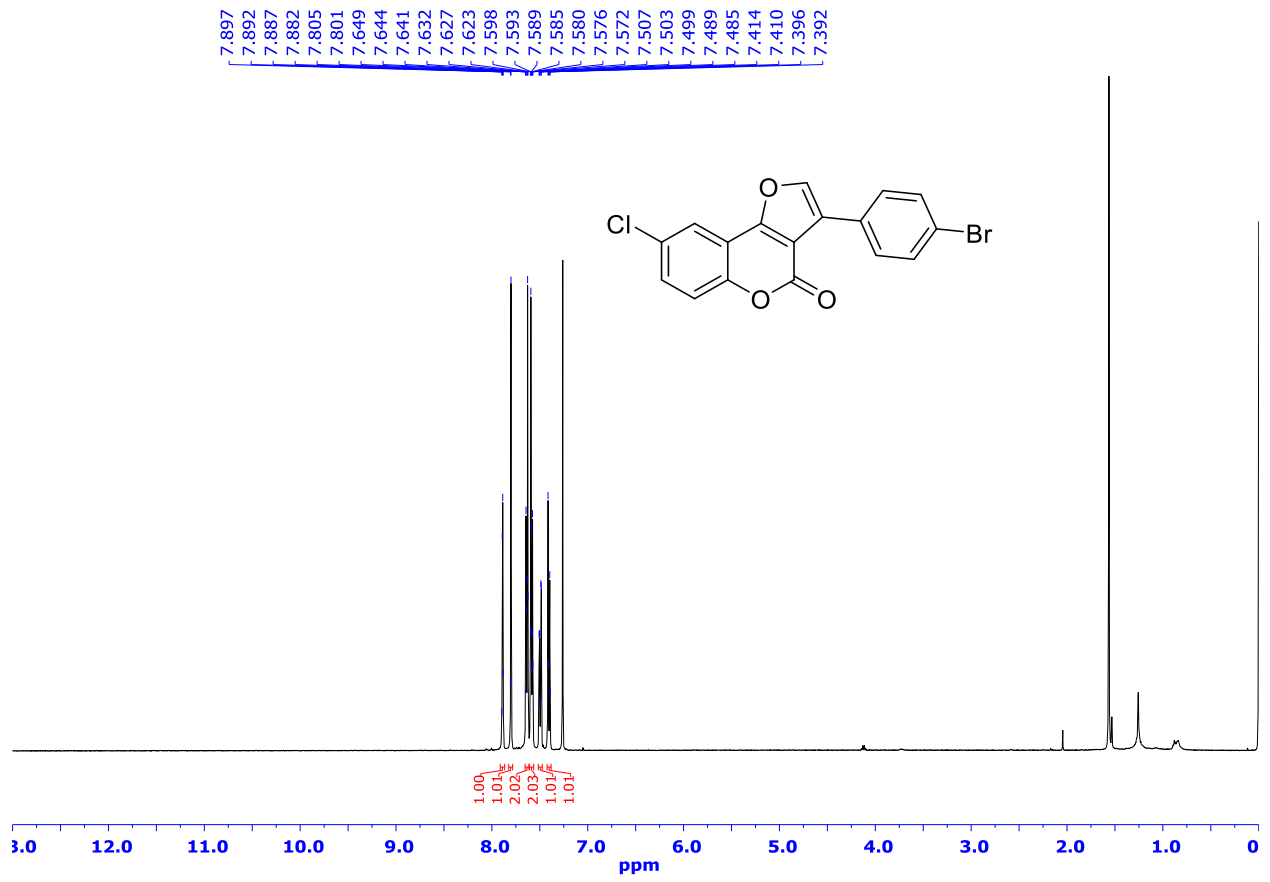


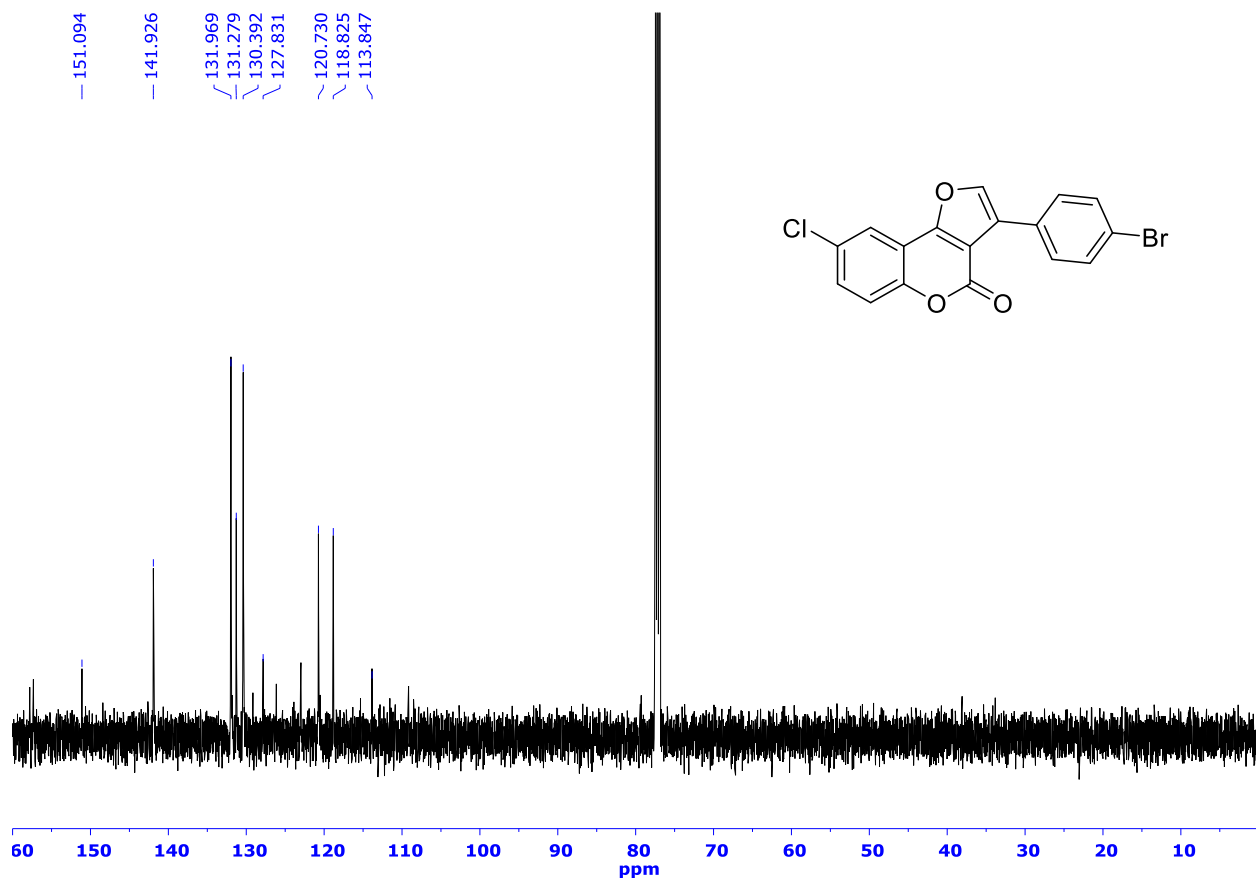
Spectroscopic data for 3-(4-bromophenyl)-8-chloro-4H-furo[3,2-c]coumarin (HP26)



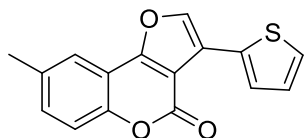
¹H NMR (500 MHz, CDCl₃) δ 7.89 (t, *J* = 2.5 Hz, 1H), 7.80 (s, 1H), 7.66 – 7.61 (m, 2H), 7.61 – 7.56 (m, 2H), 7.50 (dd, *J* = 9.0, 2.5 Hz, 1H), 7.41 (d, *J* = 8.9 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 151.09, 141.93, 131.97, 131.28, 130.39, 127.83, 120.73, 118.82, 113.85.



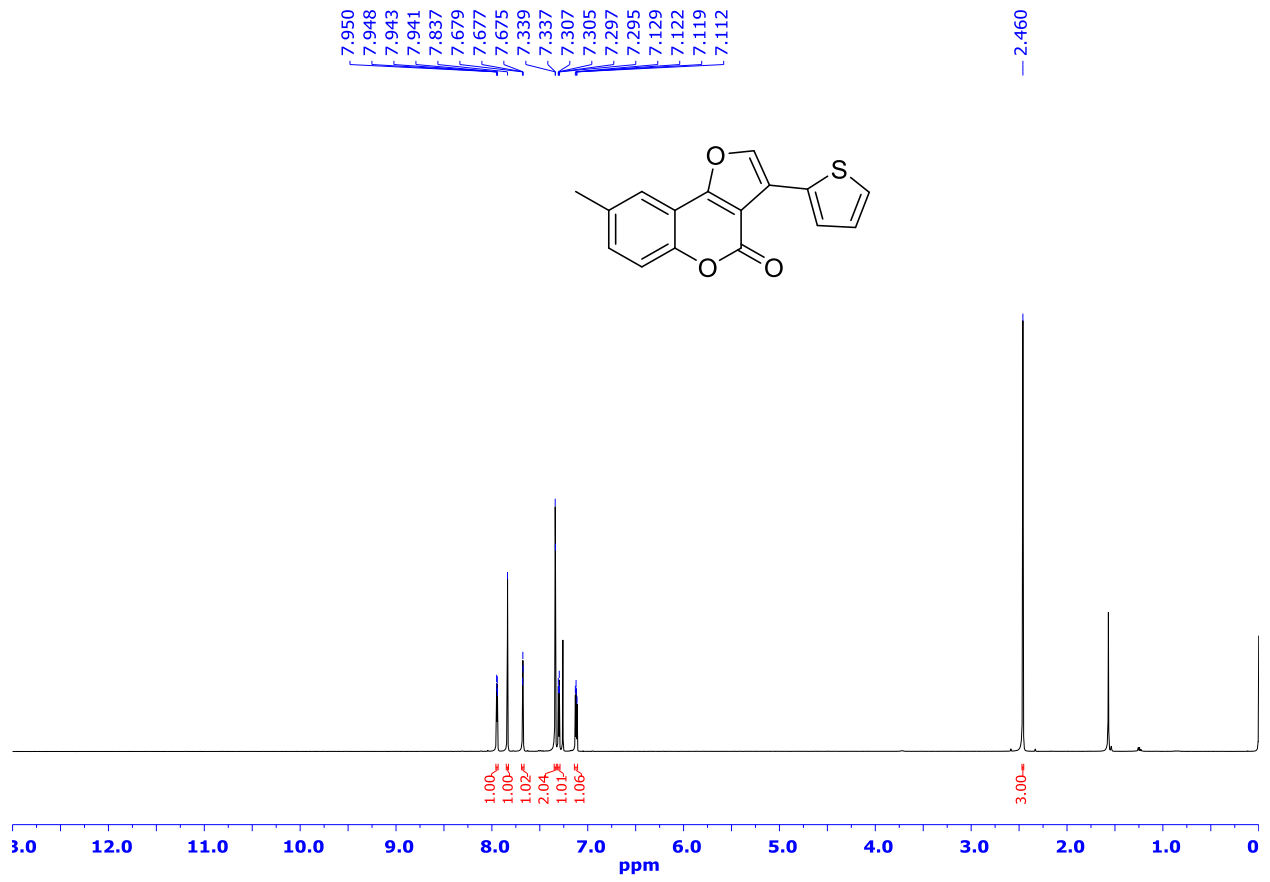


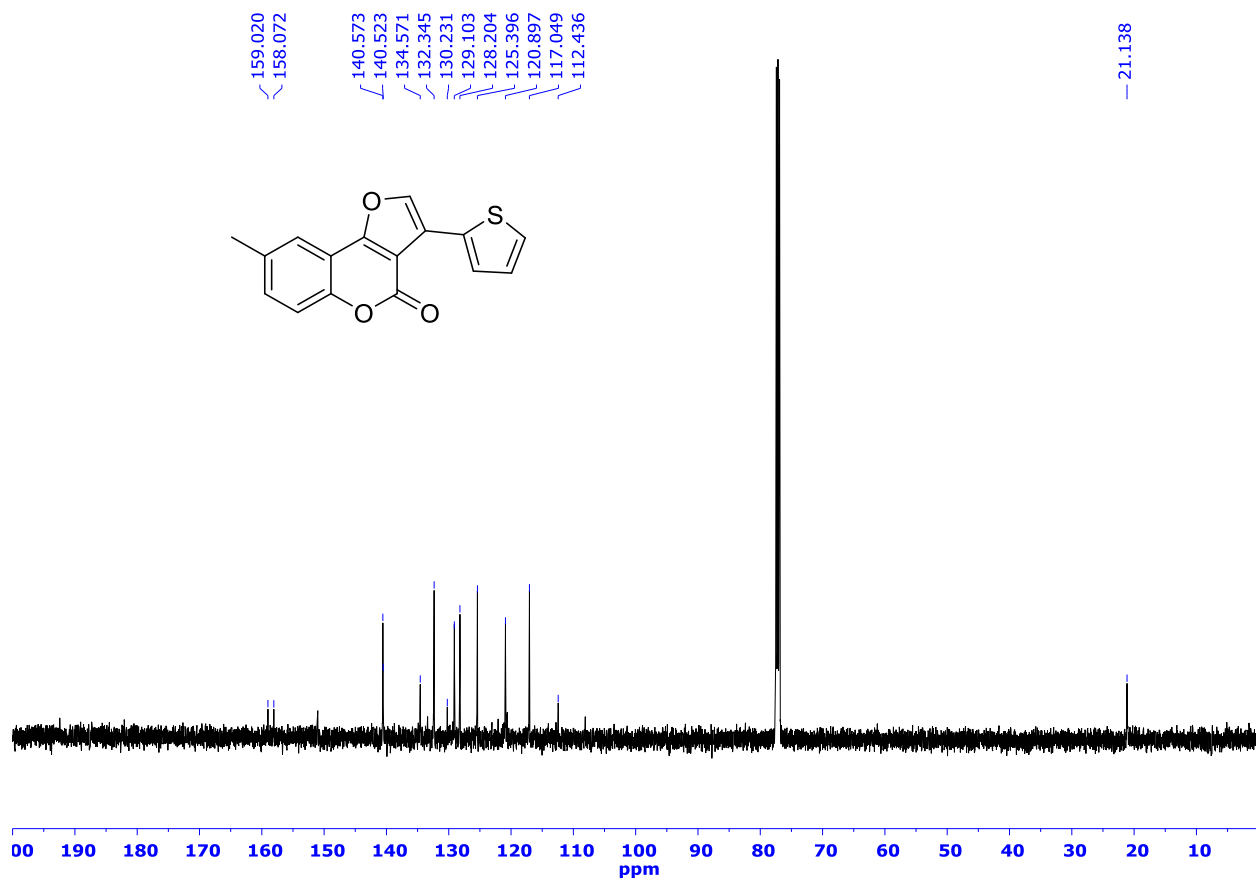
Spectroscopic data for 8-methyl-(3-thiophen-2-yl)-4H-furo[3,2-c]coumarin (HP27)



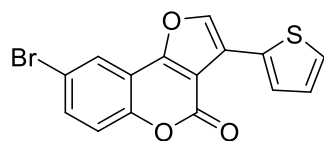
¹H NMR (500 MHz, CDCl₃) δ 7.95 (dd, *J* = 3.6, 1.1 Hz, 1H), 7.84 (s, 1H), 7.69 – 7.67 (m, 1H), 7.34 (d, *J* = 1.2 Hz, 2H), 7.30 (dd, *J* = 5.2, 1.2 Hz, 1H), 7.12 (dd, *J* = 5.1, 3.6 Hz, 1H), 2.46 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 159.02, 158.07, 140.57, 140.52, 134.57, 132.35, 130.23, 129.10, 128.20, 125.40, 120.90, 117.05, 112.44, 21.14.



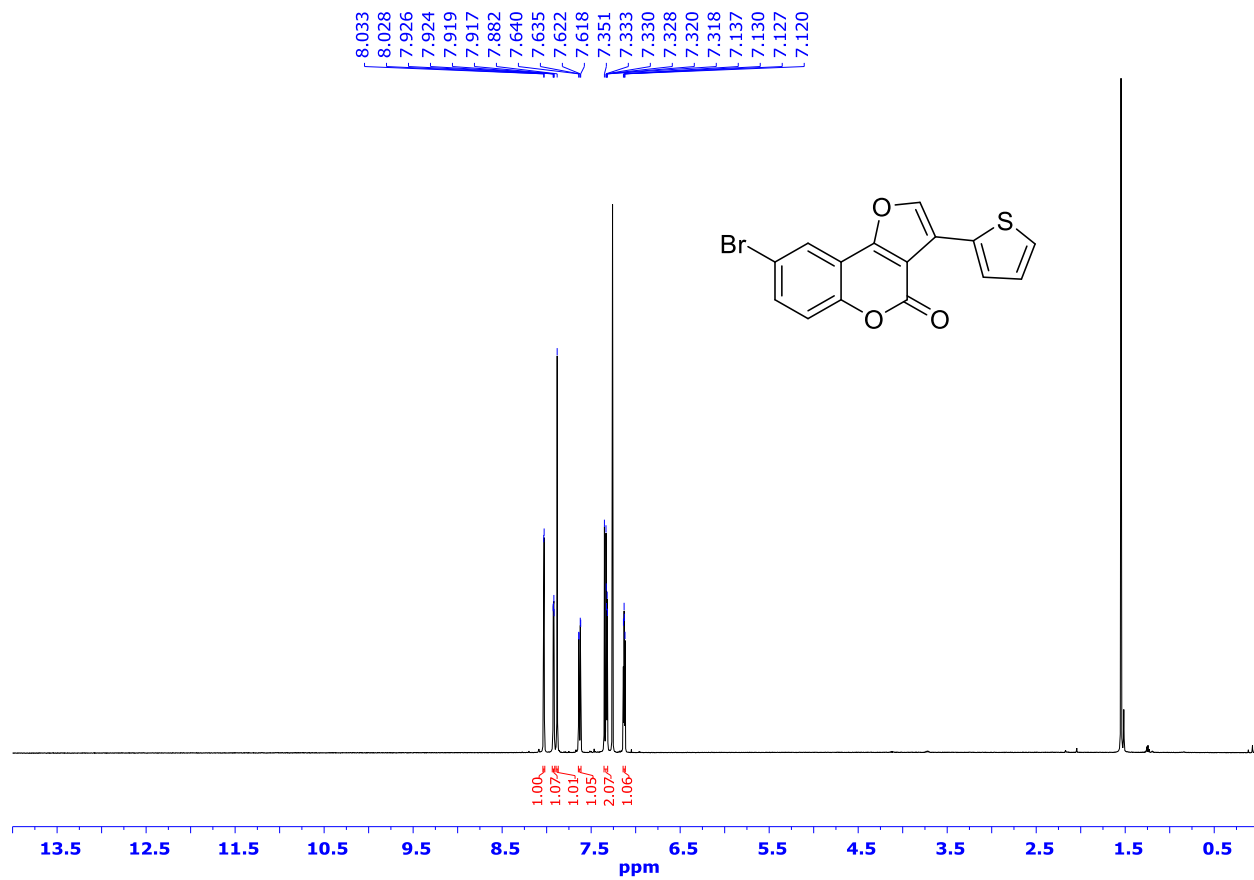


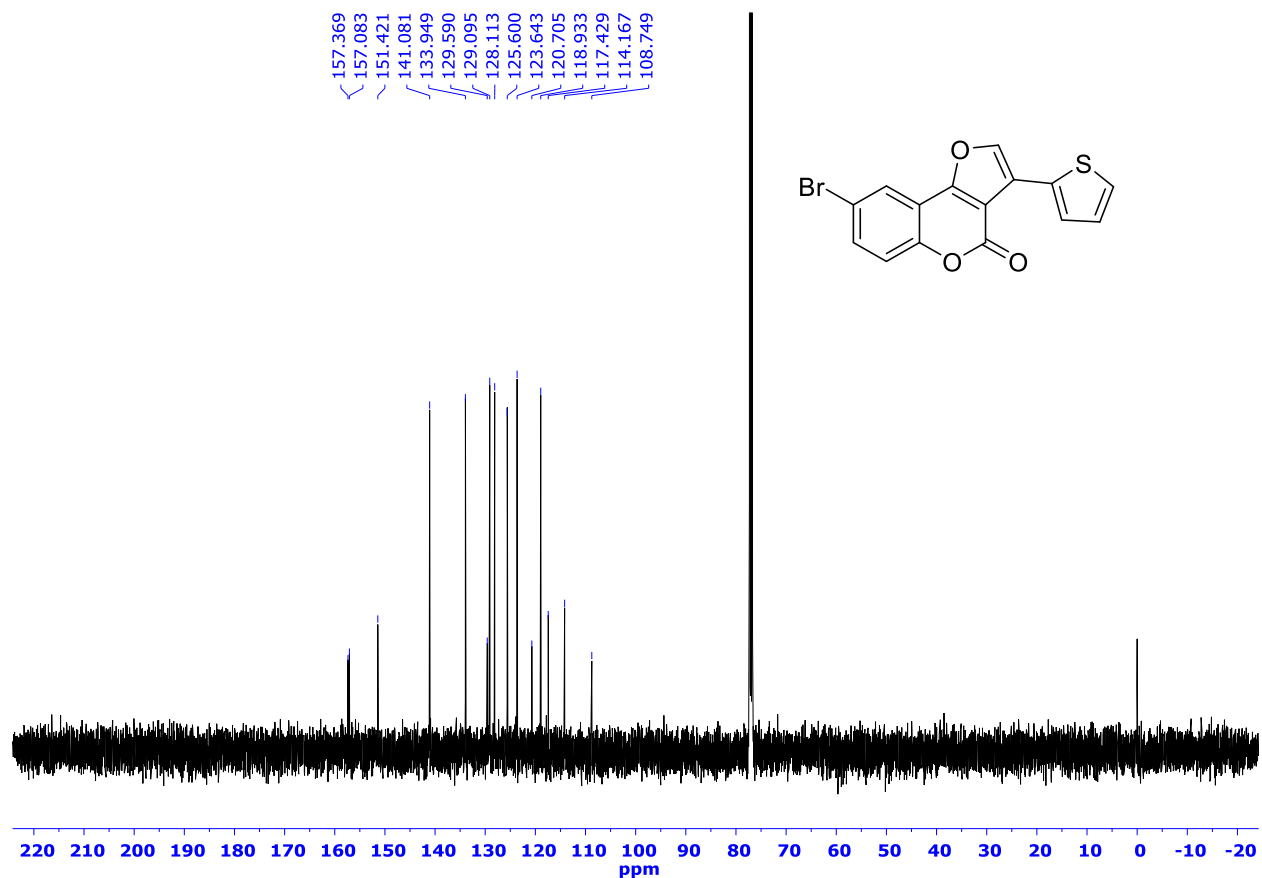
Spectroscopic data for 8-bromo-(3-thiophen-2-yl)-4H-furo[3,2-c]coumarin (HP28)



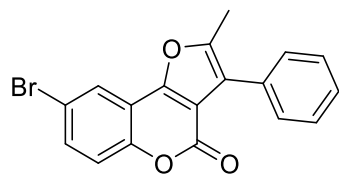
¹H NMR (500 MHz, CDCl₃) δ 8.03 (d, *J* = 2.3 Hz, 1H), 7.92 (dd, *J* = 3.7, 1.2 Hz, 1H), 7.88 (s, 1H), 7.63 (dd, *J* = 8.9, 2.3 Hz, 1H), 7.36 – 7.31 (m, 2H), 7.13 (dd, *J* = 5.1, 3.6 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 157.37, 157.08, 151.42, 141.08, 133.95, 129.59, 129.10, 128.11, 125.60, 123.64, 120.70, 118.93, 117.43, 114.17, 108.75.



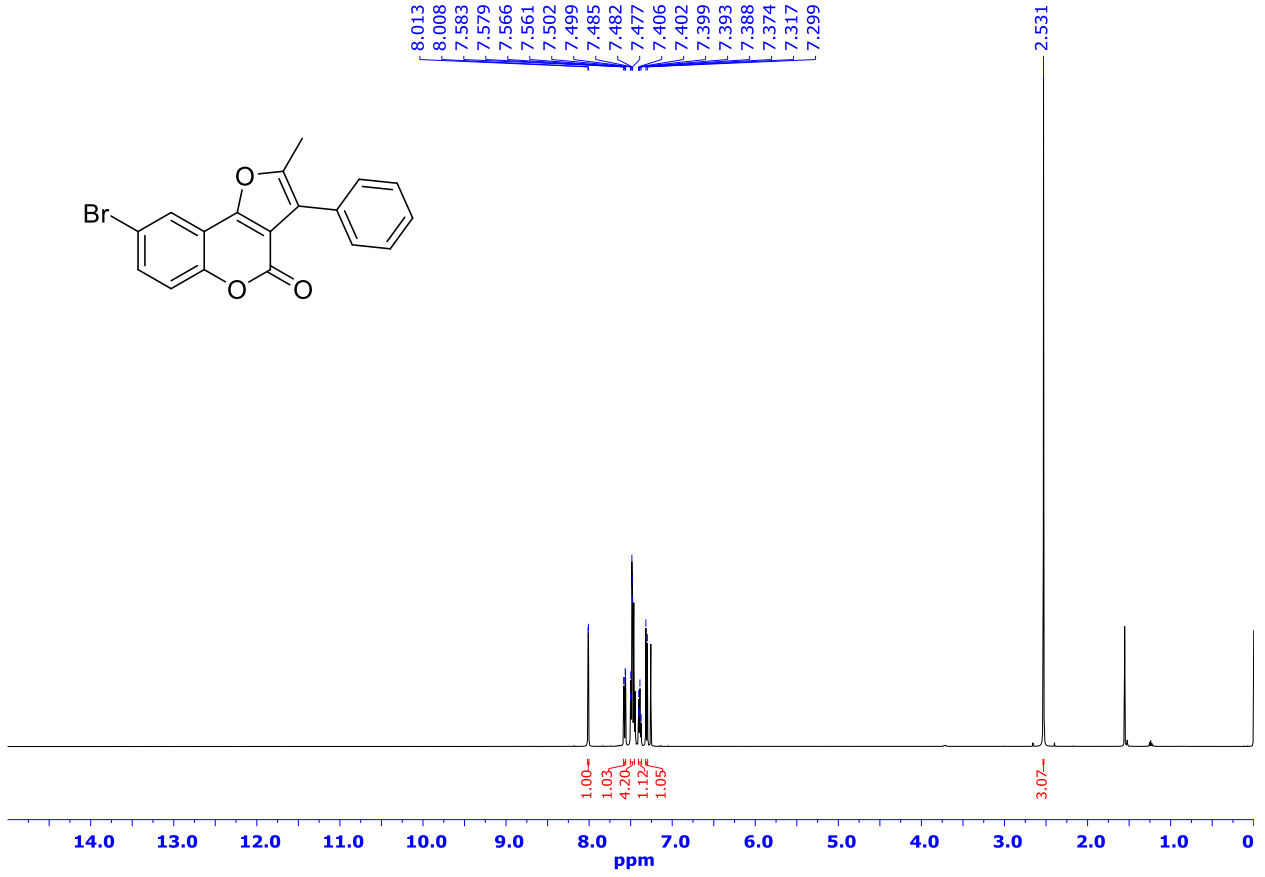
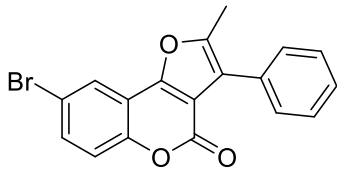


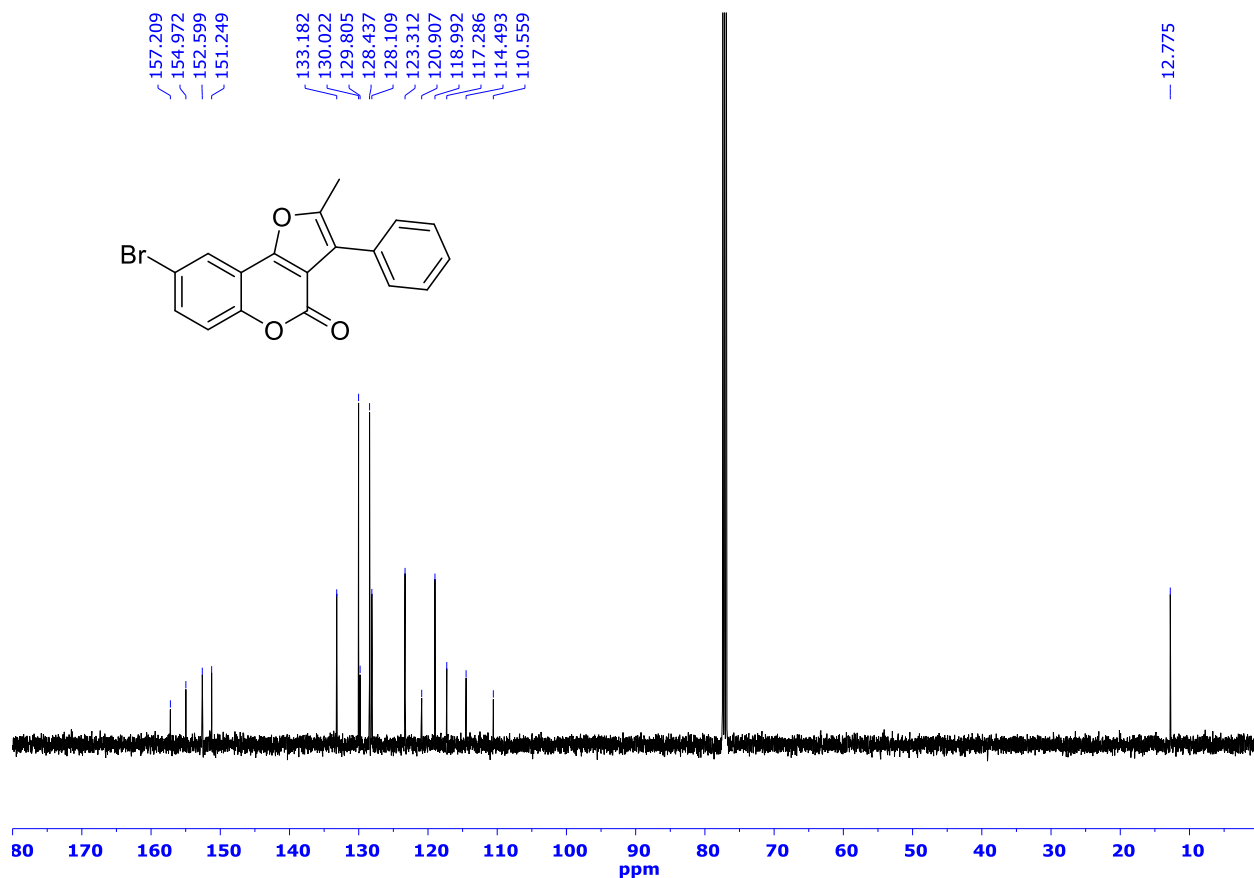
Spectroscopic data for 8-bromo-2-methyl-3-phenyl-4H-furo[3,2-c]coumarin (HP29)



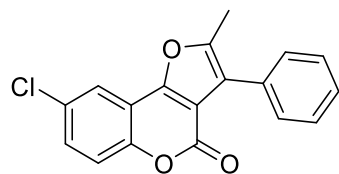
¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 2.4 Hz, 1H), 7.57 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.51 – 7.44 (m, 4H), 7.42 – 7.36 (m, 1H), 7.31 (d, *J* = 8.8 Hz, 1H), 2.53 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 157.21, 154.97, 152.60, 151.25, 133.18, 130.02, 129.81, 128.44, 128.11, 123.31, 120.91, 118.99, 117.29, 114.49, 110.56, 12.77.





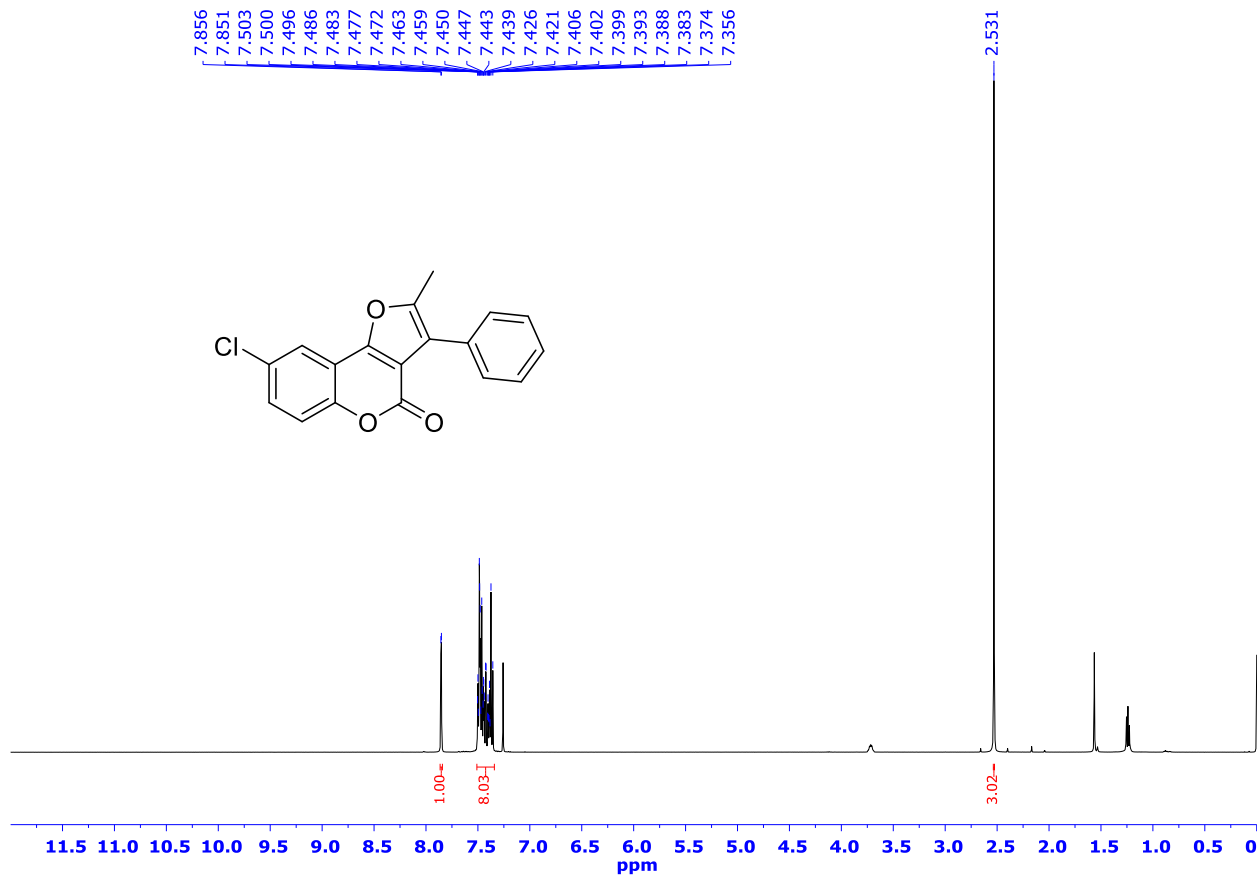
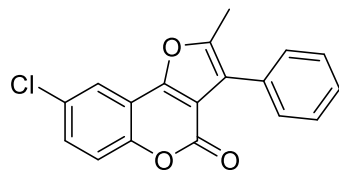
Spectroscopic data for 8-chloro-2-methyl-3-phenyl-4H-furo[3,2-c]coumarin (HP30)

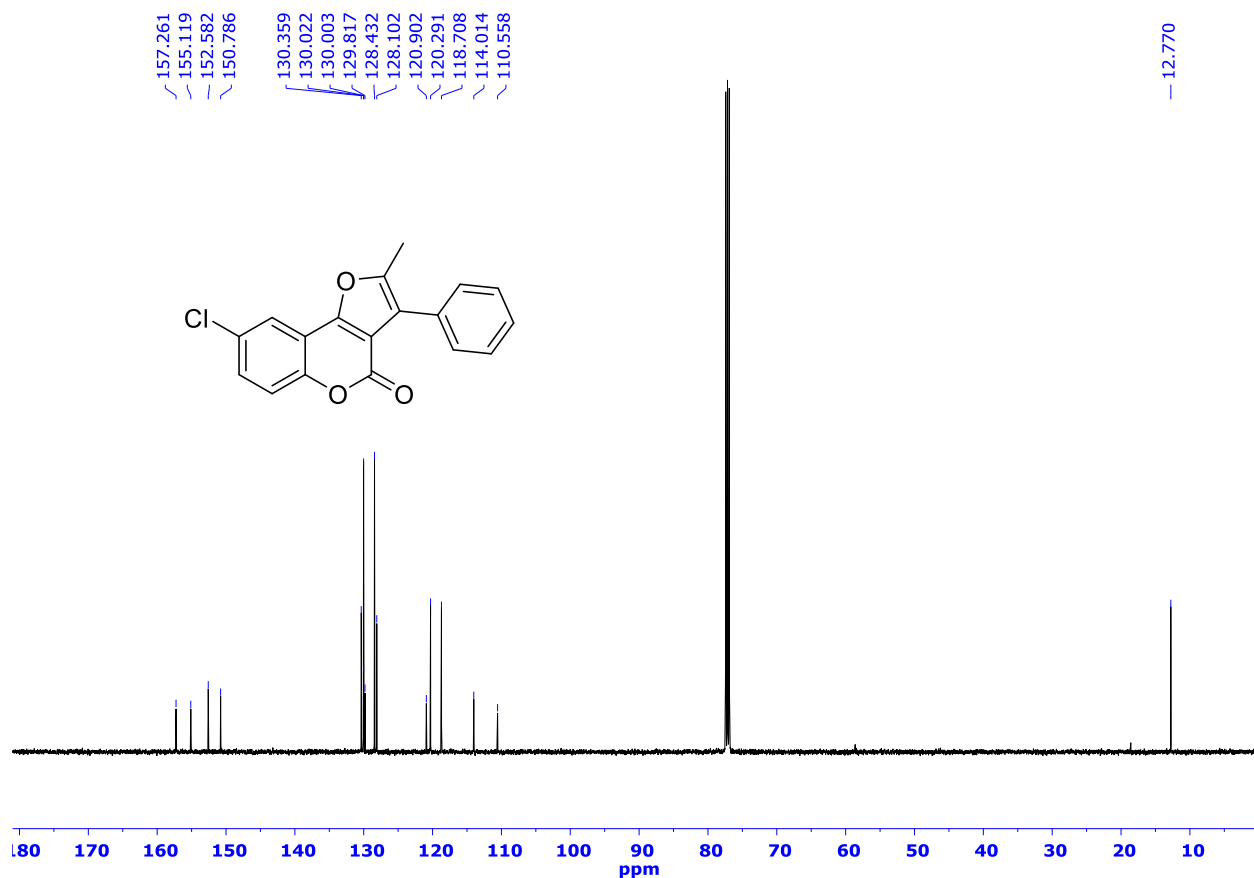


^1H NMR (500 MHz, CDCl_3) δ 7.85 (d, $J = 2.4$ Hz, 1H), 7.52 – 7.34 (m, 8H), 2.53 (s, 3H).

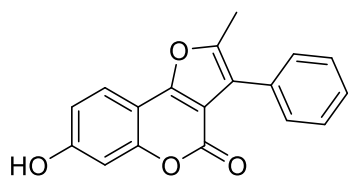
^{13}C NMR (126 MHz, CDCl_3) δ 157.26, 155.12, 152.58, 150.79, 130.36, 130.02, 130.00, 129.82, 128.43, 128.10, 120.90, 120.29, 118.71, 114.01, 110.56, 12.77.

7.856
7.851
7.503
7.500
7.496
7.486
7.483
7.477
7.472
7.463
7.459
7.450
7.447
7.443
7.439
7.426
7.421
7.406
7.402
7.399
7.393
7.388
7.383
7.374
7.356



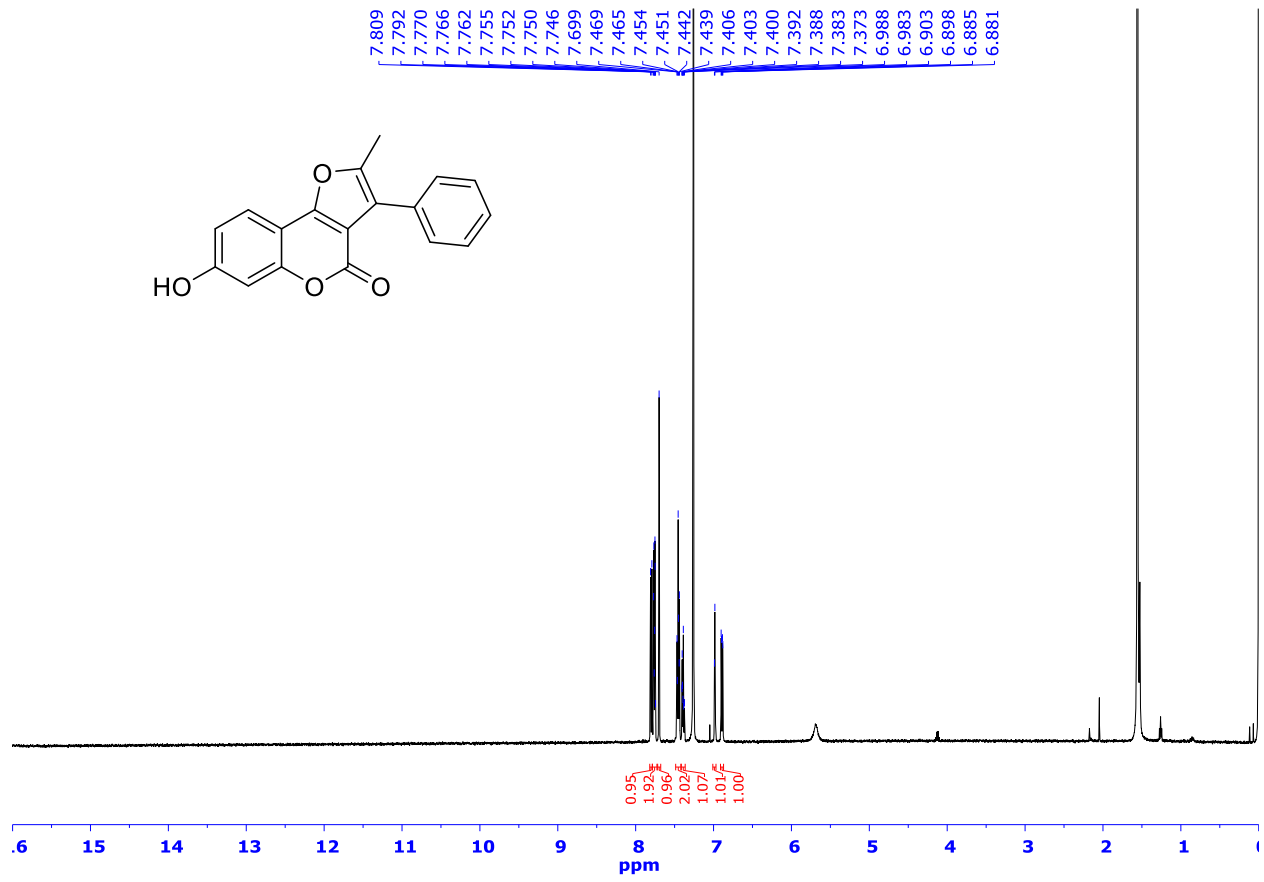


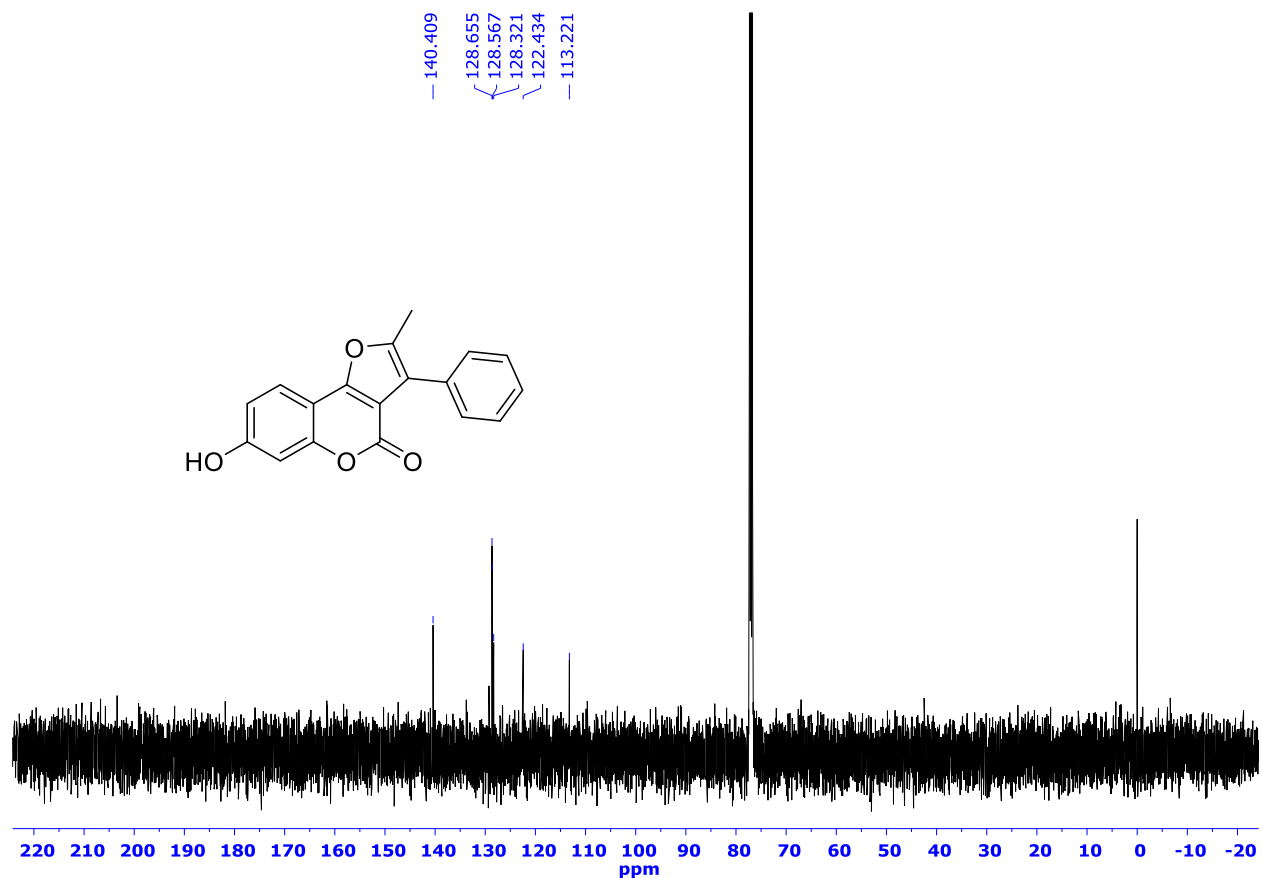
Spectroscopic data for 7-hydroxy-2-methyl-3-phenyl-4H-furo[3,2-c]coumarin (HP31)



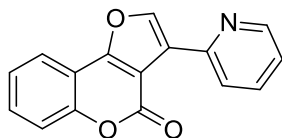
¹H NMR (500 MHz, CDCl₃) δ 7.80 (d, *J* = 8.6 Hz, 1H), 7.79 – 7.74 (m, 2H), 7.70 (s, 1H), 7.45 (dd, *J* = 8.4, 6.8 Hz, 2H), 7.41 – 7.37 (m, 1H), 6.99 (d, *J* = 2.6 Hz, 1H), 6.89 (dd, *J* = 8.6, 2.3 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 140.41, 128.65, 128.57, 128.32, 122.43, 113.22.



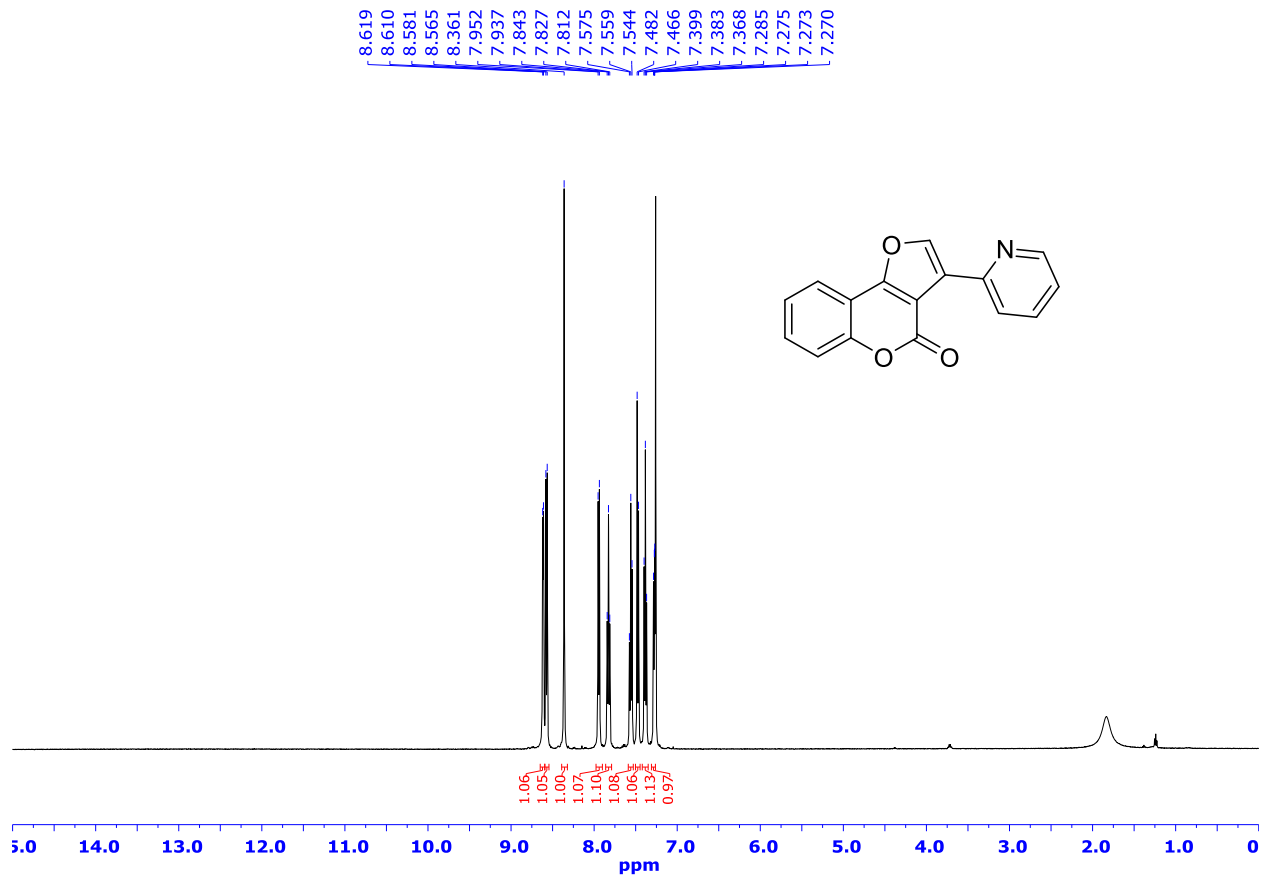


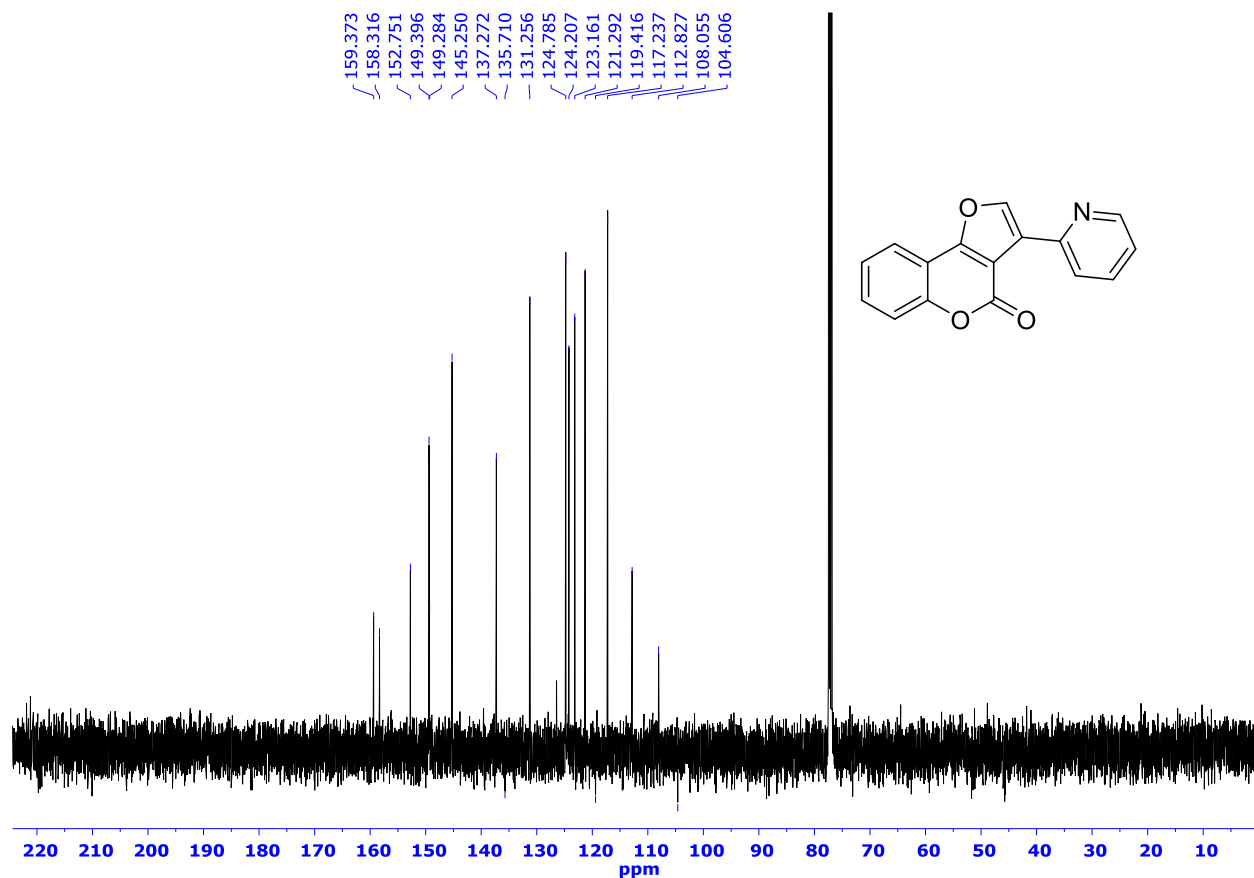
Spectroscopic data for 3-(pyridin-2-yl)-4H-furo[3,2-c]coumarin (HP8)



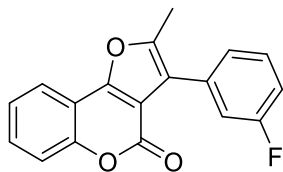
^1H NMR (500 MHz, CDCl_3) δ 8.61 (d, $J = 4.6$ Hz, 1H), 8.57 (d, $J = 8.0$ Hz, 1H), 8.36 (s, 1H), 7.94 (d, $J = 7.8$ Hz, 1H), 7.83 (t, $J = 7.7$ Hz, 1H), 7.56 (t, $J = 7.8$ Hz, 1H), 7.47 (d, $J = 8.3$ Hz, 1H), 7.38 (t, $J = 7.5$ Hz, 1H), 7.28 (dd, $J = 4.1, 3.3$ Hz, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 159.37, 158.32, 152.75, 149.40, 149.28, 145.25, 137.27, 135.71, 131.26, 124.78, 124.21, 123.16, 121.29, 119.42, 117.24, 112.83, 108.06, 104.61.



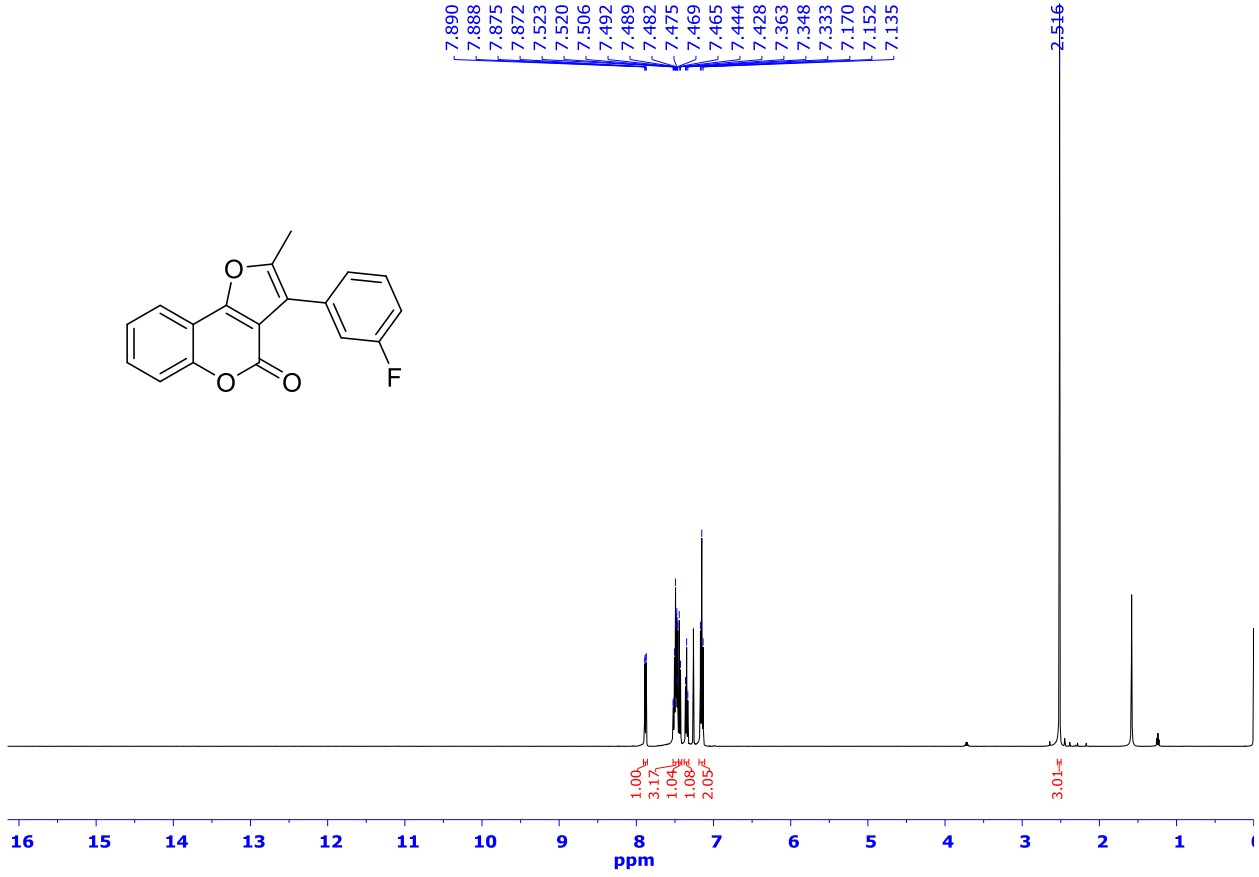
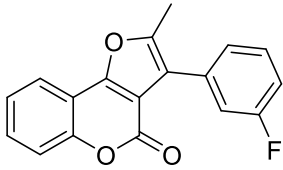


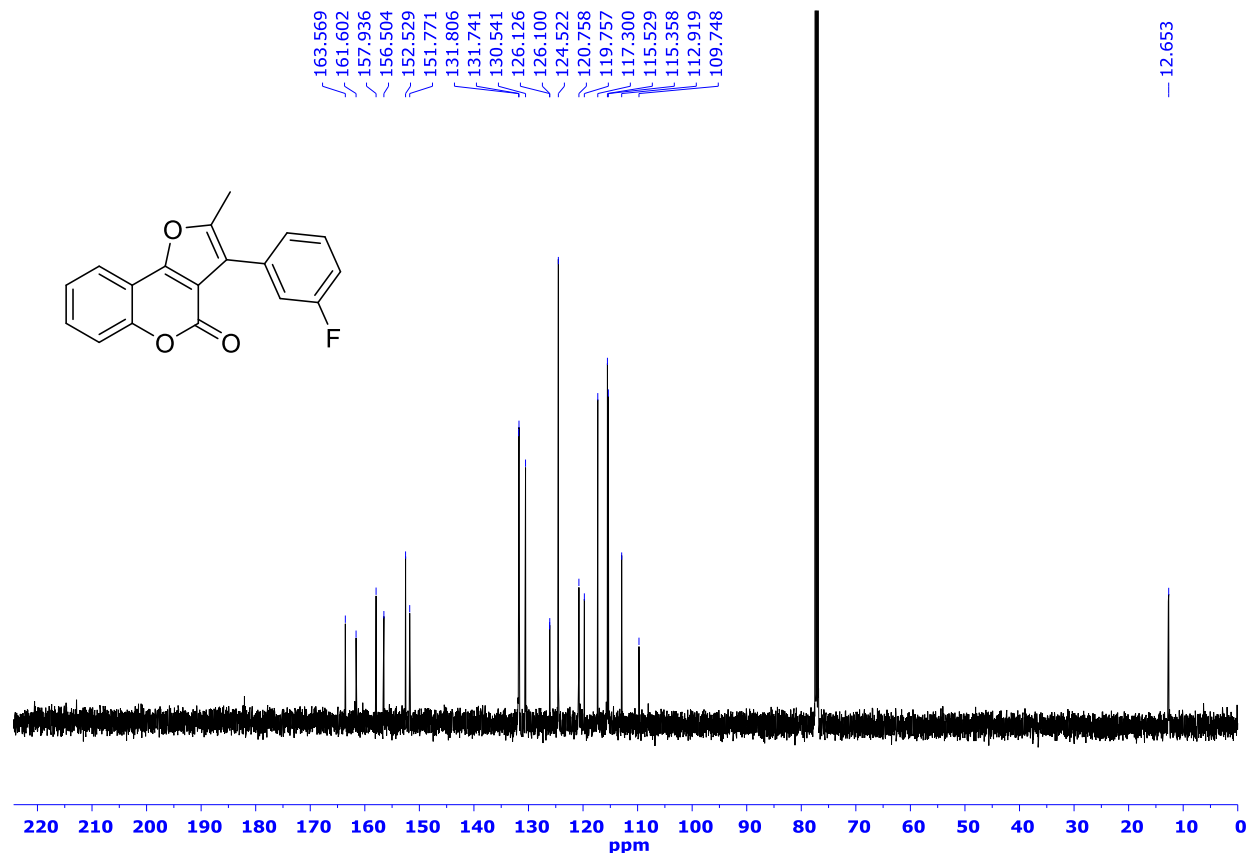
Spectroscopic data for 3-(3-fluorophenyl)-2-methyl-4H-furo[3,2-c]coumarin (HP8)



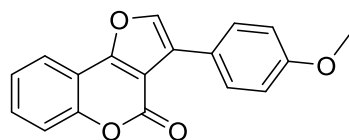
^1H NMR (500 MHz, CDCl_3) δ 7.88 (dd, $J = 7.8, 1.1$ Hz, 1H), 7.52 – 7.45 (m, 3H), 7.44 (d, $J = 8.2$ Hz, 1H), 7.35 (t, $J = 7.5$ Hz, 1H), 7.15 (t, $J = 8.7$ Hz, 2H), 2.52 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 163.57, 161.60, 157.94, 156.50, 152.53, 151.77, 131.81, 131.74, 130.54, 126.13, 126.10, 124.52, 120.76, 119.76, 117.30, 115.53, 115.36, 112.92, 109.75, 12.65.



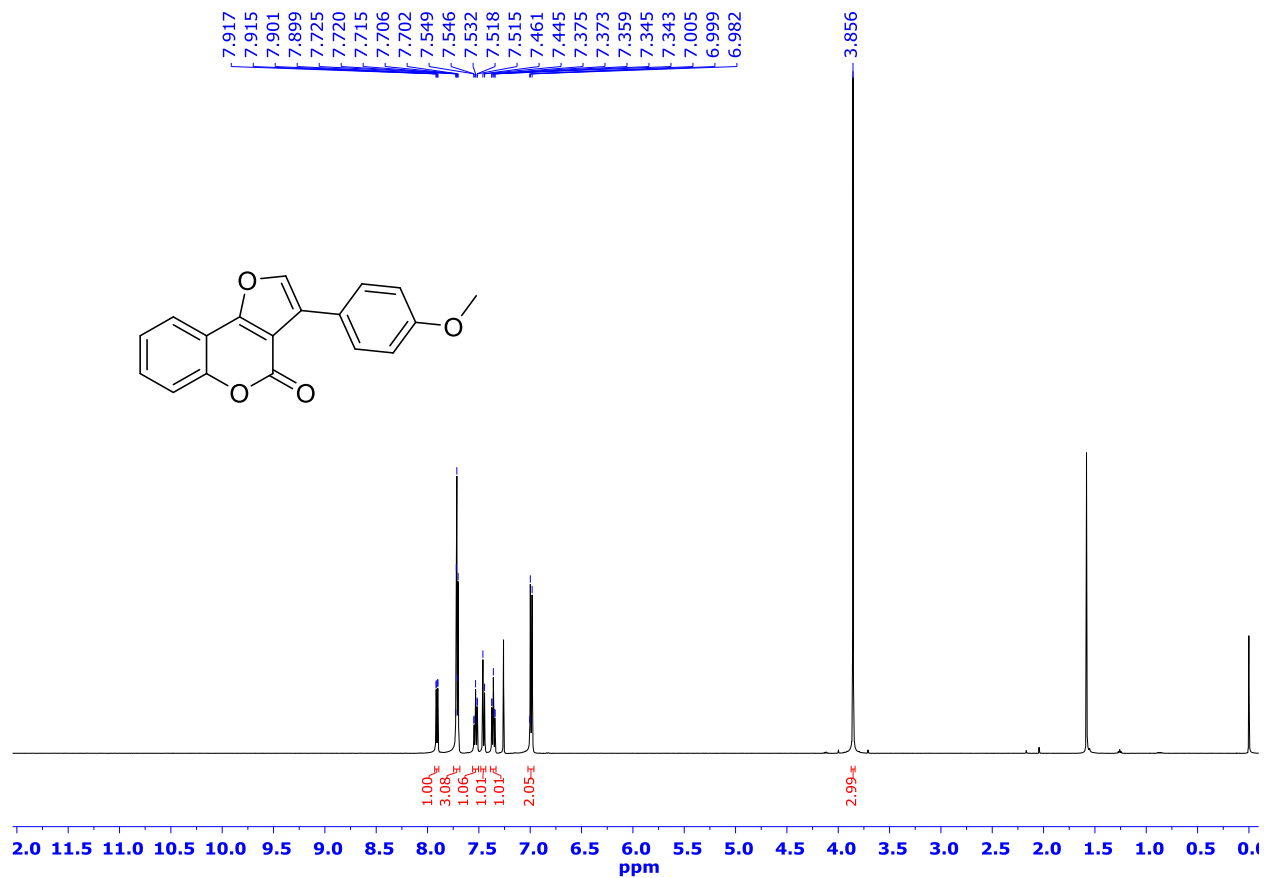


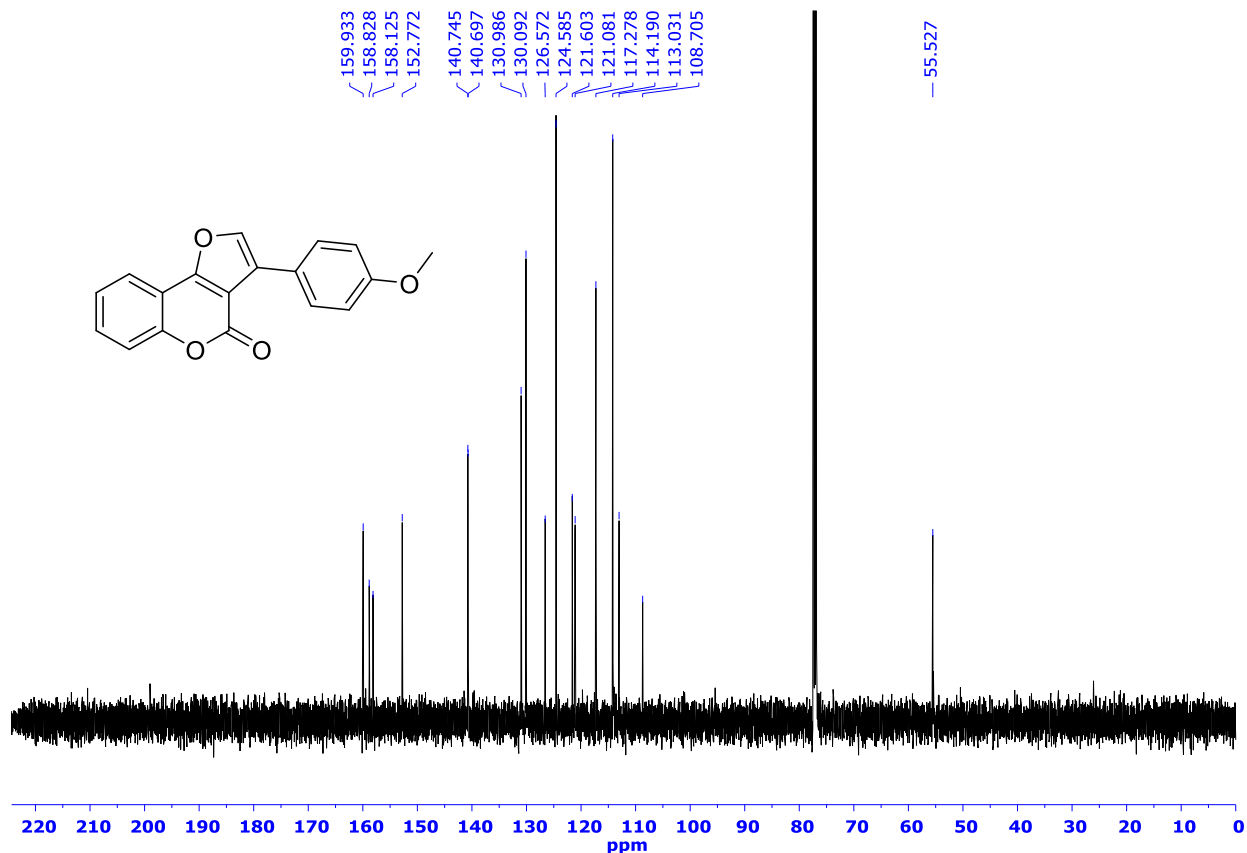
Spectroscopic data for 3-(4-methoxyphenyl)-4H-furo[3,2-c]coumarin (HP8)



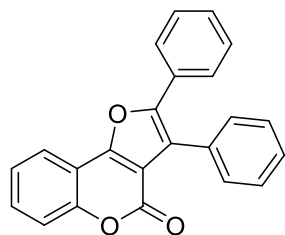
¹H NMR (500 MHz, CDCl₃) δ 7.91 (dd, *J* = 7.8, 1.2 Hz, 1H), 7.75 – 7.69 (m, 3H), 7.56 – 7.50 (m, 1H), 7.45 (d, *J* = 8.0 Hz, 1H), 7.39 – 7.33 (m, 1H), 7.00 (t, *J* = 5.8 Hz, 2H), 3.86 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 159.93, 158.83, 158.13, 152.77, 140.74, 140.70, 130.99, 130.09, 126.57, 124.59, 121.60, 121.08, 117.28, 114.19, 113.03, 108.70, 55.53.



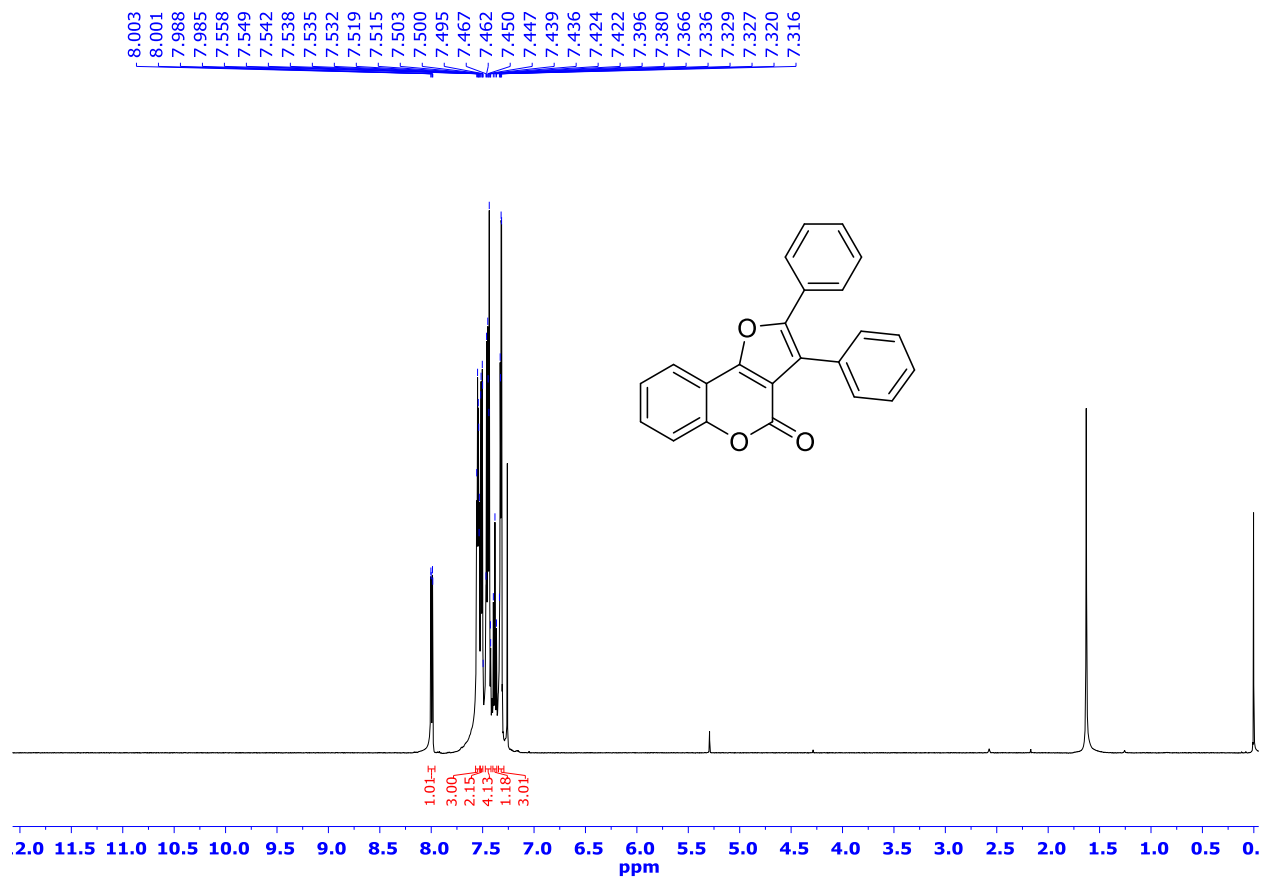


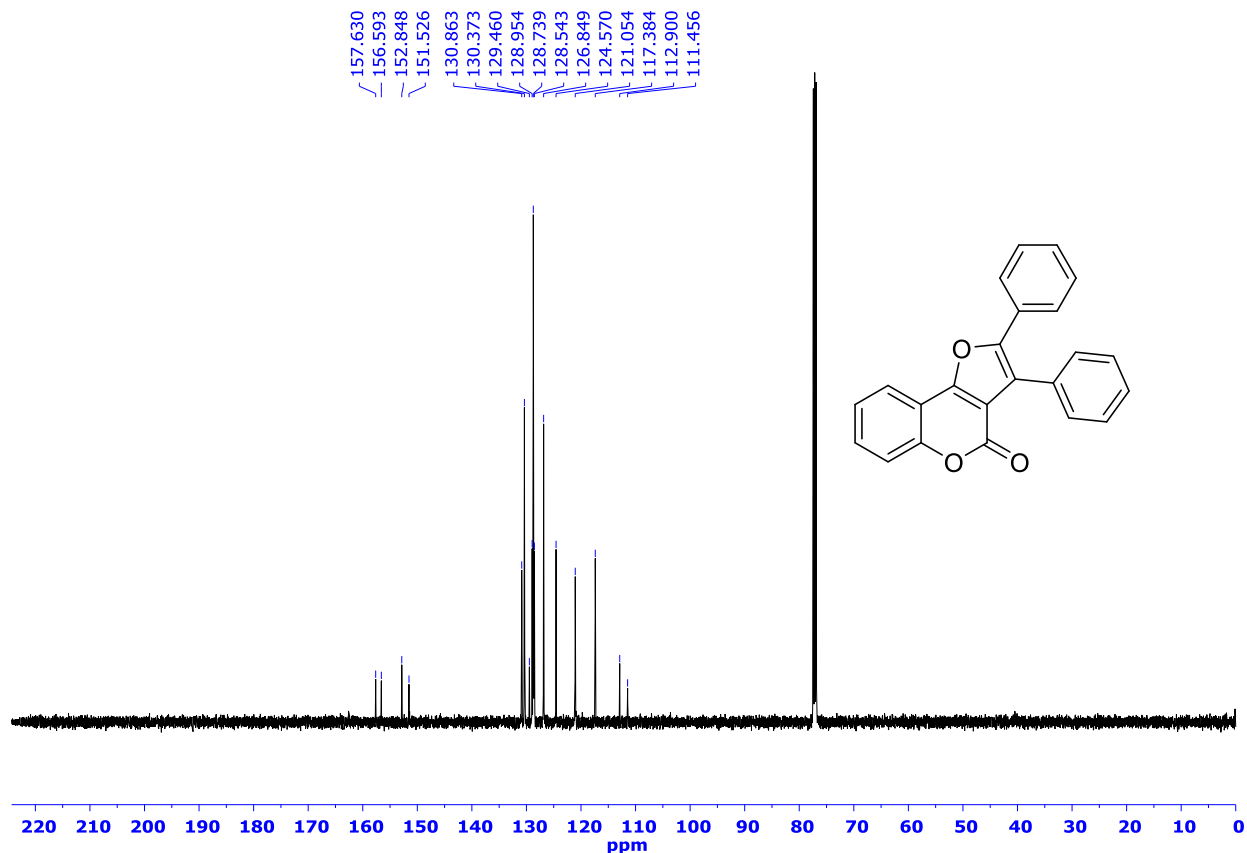
Spectroscopic data for 2,3-diphenyl-4H-furo[3,2-c]coumarin (HP8)



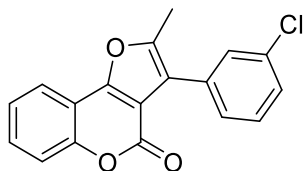
^1H NMR (500 MHz, CDCl_3) δ 7.99 (dd, $J = 7.8, 1.4$ Hz, 1H), 7.57 – 7.53 (m, 3H), 7.51 (dd, $J = 7.6, 1.6$ Hz, 2H), 7.47 – 7.42 (m, 4H), 7.38 (t, $J = 7.5$ Hz, 1H), 7.35 – 7.29 (m, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 157.63, 156.59, 152.85, 151.53, 130.86, 130.37, 129.46, 128.95, 128.74, 128.54, 126.85, 124.57, 121.05, 117.38, 112.90, 111.46



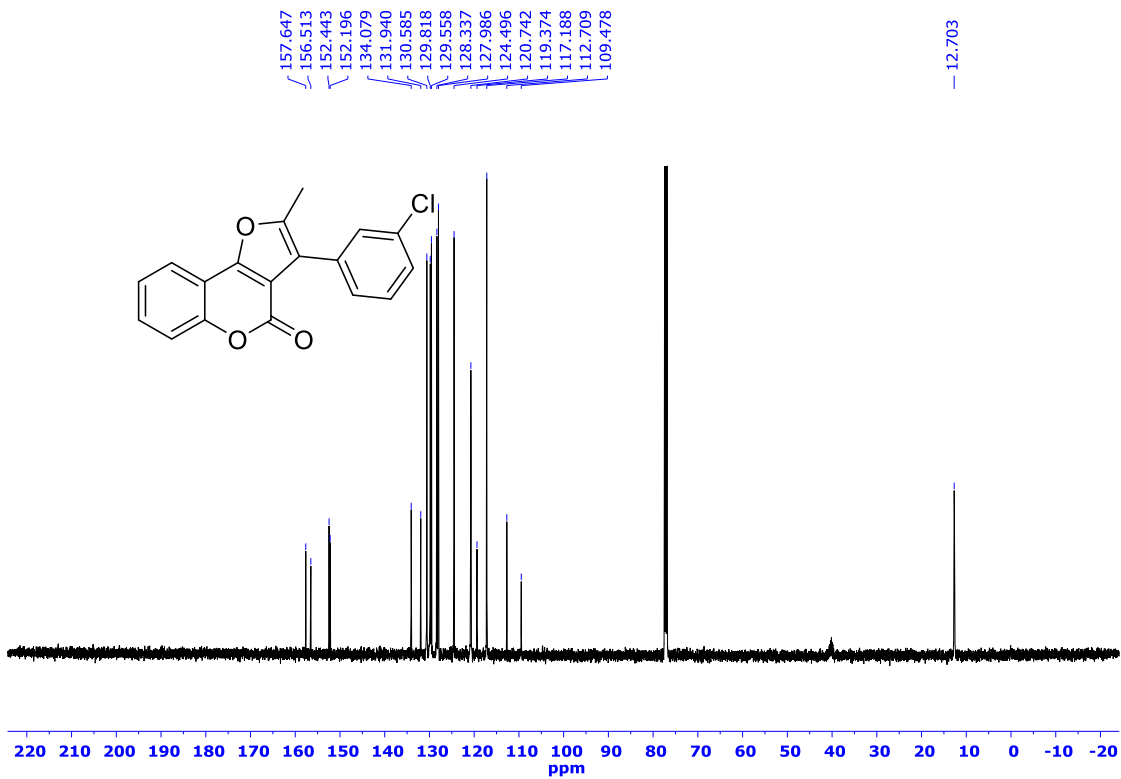
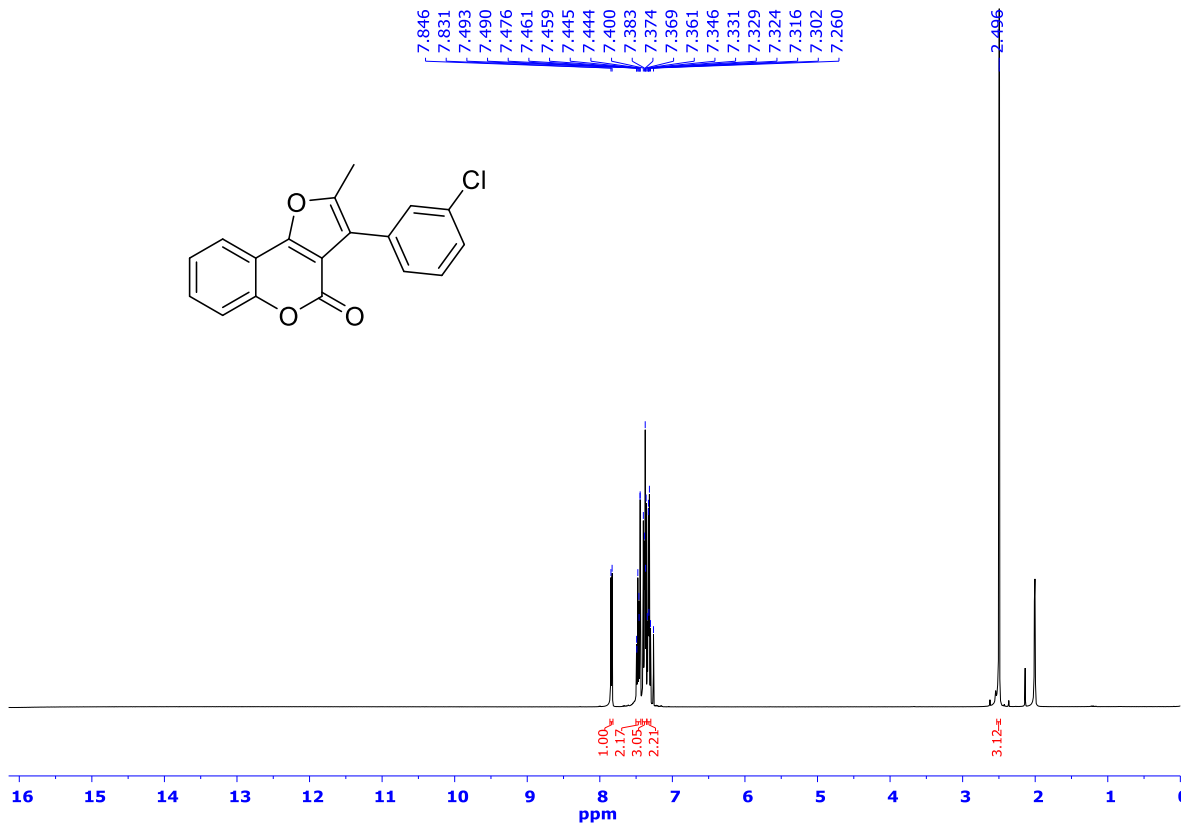


Spectroscopic data for 3-(3-chlorophenyl)-2-methyl-4H-furo[3,2-c]coumarin (HP17)



¹H NMR (500 MHz, CDCl₃) δ 7.84 (d, *J* = 7.8 Hz, 1H), 7.50 – 7.43 (m, 2H), 7.41 – 7.35 (m, 3H), 7.32 (dt, *J* = 11.1, 7.4 Hz, 2H), 2.50 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 157.65, 156.51, 152.44, 152.20, 134.08, 131.94, 130.58, 129.82, 129.56, 128.34, 127.99, 124.50, 120.74, 119.37, 117.19, 112.71, 109.48, 12.70.



Section S7. Reference

- (1) Bruker. *APEX3* v2015.5-2. (Bruker AXS Inc., Madison, Wisconsin, U.S.A. 2015).
- (2) Bruker. *SAINTE* v8.37A. (Bruker AXS Inc., Madison, Wisconsin, U.S.A. 2015).
- (3) Bruker. *SADABS* -2016/2. (Bruker AXS Inc., Madison, Wisconsin, U.S.A. 2016).
- (4) Sheldrick, G. A Short History of Shelx. *Acta Cryst.* **2008**, *A64*, 112-122.
- (5) Dolomanov, O.V., Bourhis, L.J., Gildea, R.J, Howard, J.A.K. & Puschmann, H. Olex2: A Complete Structure Solution, Refinement and Analysis Program. *J. Appl. Cryst.* **2009**, *42*, 339-341.