

## Supporting Information

### Synthesis and Evaluation of (*S*)-5'-C-Aminopropyl and (*S*)-5'-C-Aminopropyl-2'-arabinofluoro Modified DNA Oligomers for Novel RNase H-dependent Antisense Oligonucleotides

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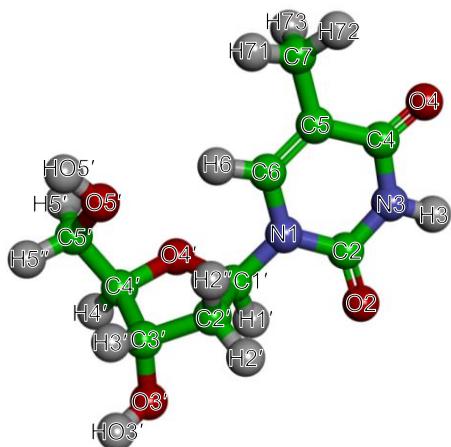
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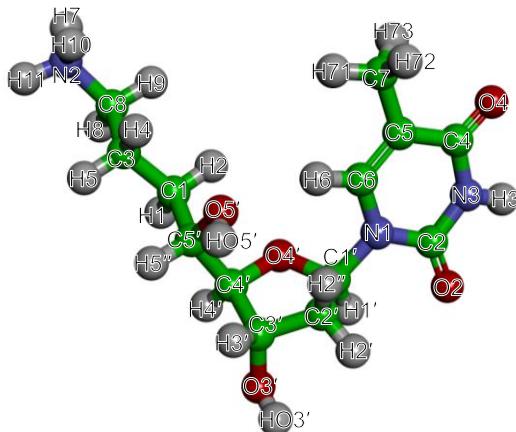
Reverse-phase HPLC spectra and MALDI-TOF/MS spectra of DNAs **1-3**, **5-13** and RNAs **1, 6**.

a.



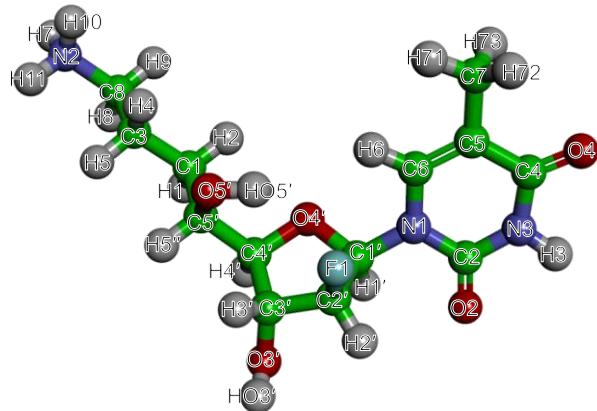
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O5'	oh	-0.642377	H1'	h2	0.183207
HO5'	ho	0.445969	N1	n	0.004881
C5'	c3	-0.026110	C6	cc	-0.258929
H5''	h1	0.085899	H6	h4	0.277171
H5'	h1	0.085899	C5	cd	-0.015884
C4'	c3	0.167490	C7	c3	-0.226761
C3'	c3	0.131625	H73	hc	0.078192
C2'	c3	-0.085285	H72	hc	0.078192
H2''	hc	0.056570	H71	hc	0.078192
H2'	hc	0.056570	C4	c	0.604130
H3'	h1	0.089949	O4	o	-0.561408
O3'	oh	-0.670856	N3	n	-0.505706
HO3'	ho	0.440276	H3	hn	0.345687
H4'	h1	0.110996	C2	c	0.606967
O4'	os	-0.394698	O2	o	-0.597377
C1'	c3	0.057527			

b.



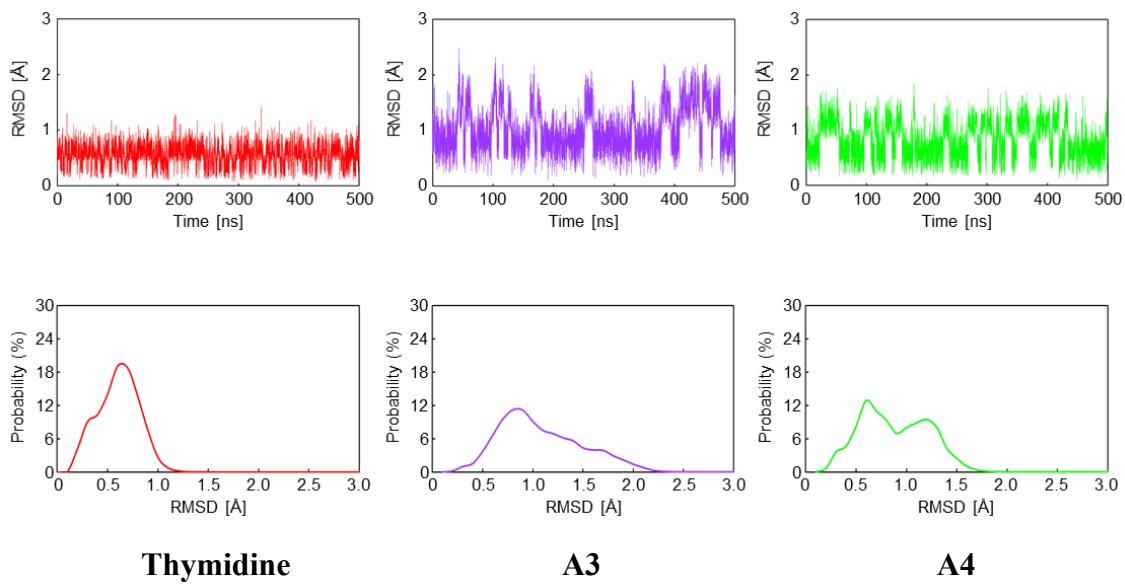
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N2	n4	-0.262310	H2"	hc	0.030480
H7	hn	0.296620	H2'	hc	0.030480
H10	hn	0.296620	H3'	h1	0.055066
H11	hn	0.296620	H4'	h1	0.141936
C8	c3	0.056368	O4'	os	-0.396941
H8	hx	0.087808	C1'	c3	0.108014
H9	hx	0.087808	H1'	h2	0.175389
C3	c3	0.031312	N1	n	-0.016848
H4	hc	0.035624	C6	cc	-0.231780
H5	hc	0.035624	H6	h4	0.266459
C1	c3	-0.116672	C5	cd	-0.020402
H1	hc	0.070456	C7	c3	-0.253122
H2	hc	0.070456	H73	hc	0.088682
C5'	c3	0.027808	H72	hc	0.088682
O5'	oh	-0.628030	H71	hc	0.088682
HO5'	ho	0.437222	C4	c	0.585963
H5"	h1	0.083525	O4	o	-0.537039
C4'	c3	0.098860	N3	n	-0.453277
C3'	c3	0.174422	H3	hn	0.344928
O3'	oh	-0.681593	C2	c	0.542066
HO3'	ho	0.455432	O2	o	-0.562935
C2'	c3	-0.028466			

c.

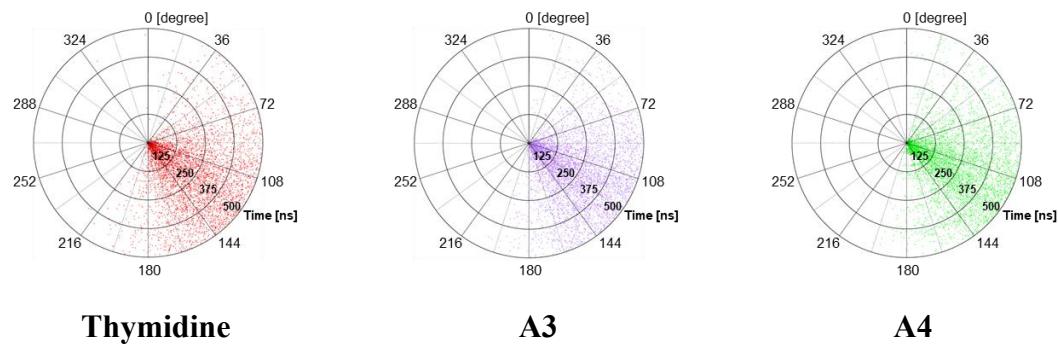


Atom name	Atom type	RESP charge	Atom name	Atom type	RESP charge
N2	n4	-0.320336	F1	f	-0.225659
H11	hn	0.312967	H2'	h1	0.146457
H10	hn	0.312967	H3'	h1	0.114231
H7	hn	0.312967	H4'	h1	0.164414
C8	c3	0.047498	O4'	os	-0.349118
H9	hx	0.093169	C1'	c3	0.041278
H8	hx	0.093169	H1'	h2	0.183224
C3	c3	0.002191	N1	n	0.018101
H5	hc	0.049590	C6	cc	-0.281532
H4	hc	0.049590	H6	h4	0.266824
C1	c3	-0.053607	C5	cd	0.004856
H2	hc	0.048251	C7	c3	-0.252306
H1	hc	0.048251	H73	hc	0.088667
C5'	c3	0.030046	H72	hc	0.088667
O5'	oh	-0.622083	H71	hc	0.088667
HO5'	ho	0.429055	C4	c	0.626134
H5"	h1	0.094626	O4	o	-0.542153
C4'	c3	0.057593	N3	n	-0.537816
C3'	c3	0.090972	H3	hn	0.365852
O3'	oh	-0.649012	C2	c	0.619609
HO3'	ho	0.454943	O2	o	-0.586622
C2'	c3	0.075419			

**Figure S1. Molecular Modeling: the reference structure, atom types and calculated RESP charges for each nucleoside.** The reference structures were obtained from an energy minimization calculation. (a) Thymidine. (b) (*S*)-5'-C-aminopropyl-thymidine (**A3**). (c) (*S*)-5'-C-aminopropyl-2'- $\beta$ -fluoro-thymidine (**A4**).



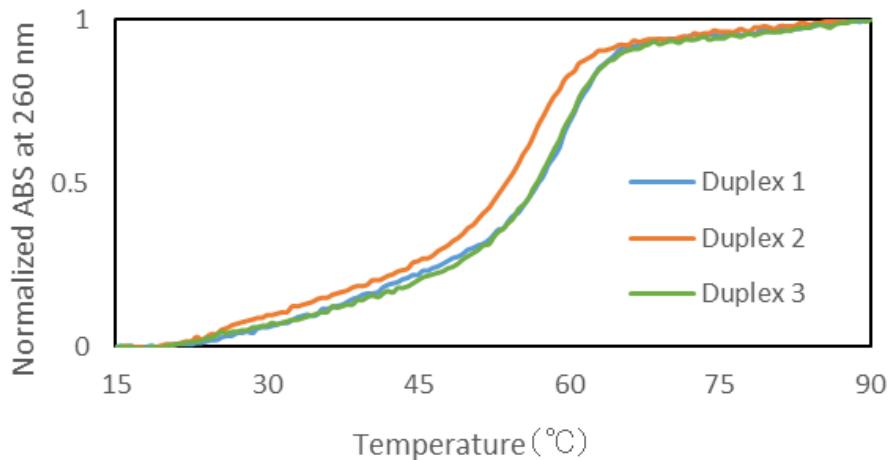
**Figure S2. Molecular Modeling: the changes (up) and the distributions (down) of time-dependent RMSD ( $\text{\AA}$ ) of each nucleoside**



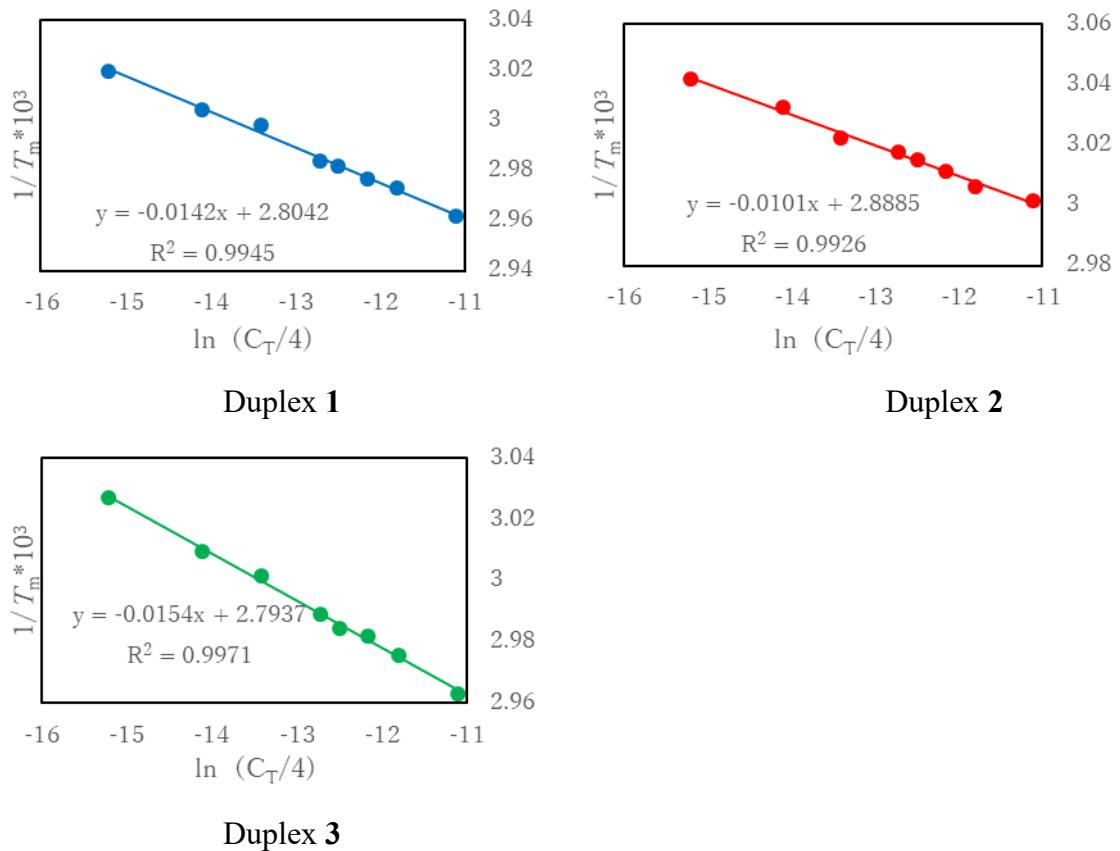
**Figure S3. Molecular Modeling: pseudorotational phase angle ( $P$ ).**

The radial axes are the times with the origin as 0 ns and increasing outward to 500 ns.

The angular axes represent the torsion angles with ranges from 0 to  $2\pi$  in clockwise direction.

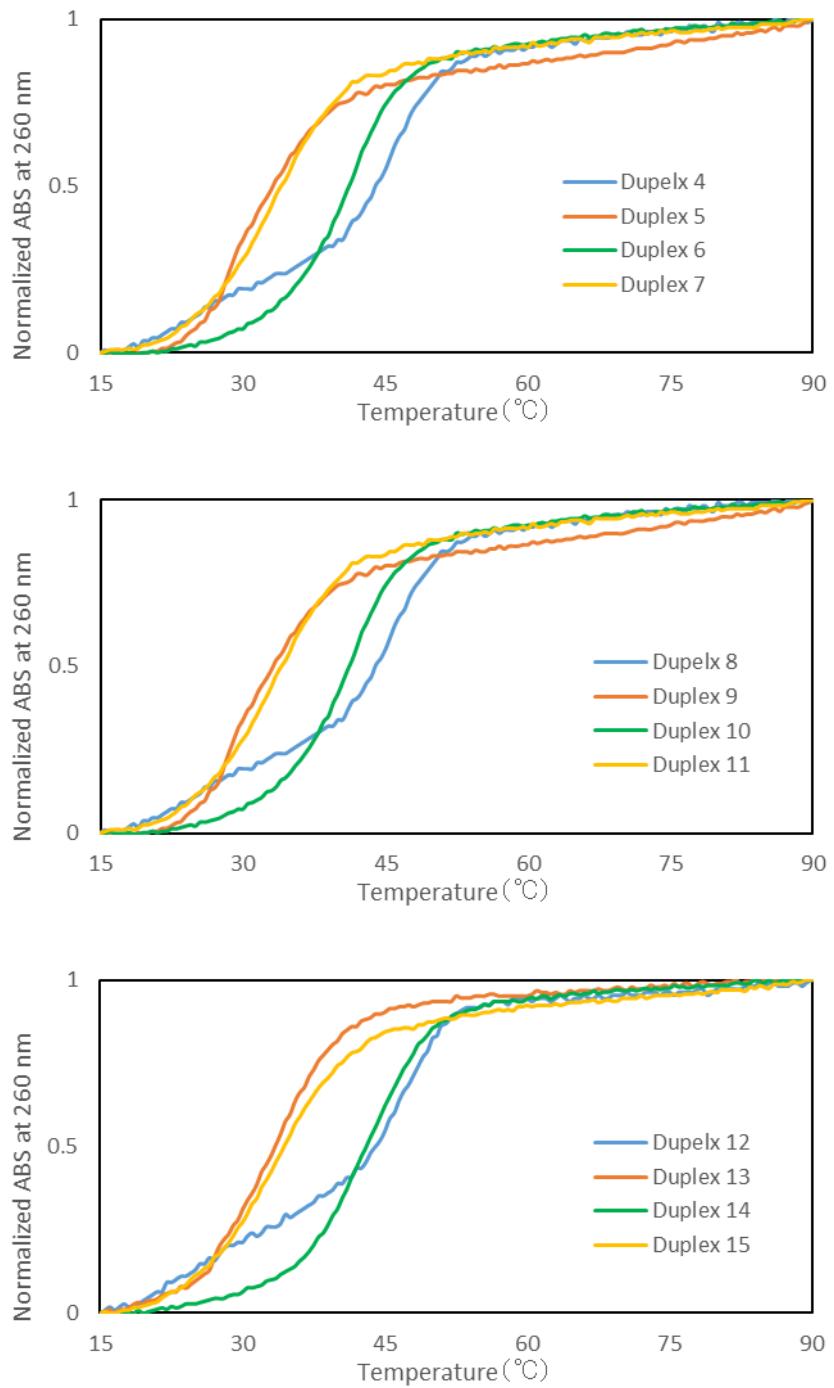


**Figure S4. Thermal Stability of Duplexes: the UV melting profiles of duplex 1-3.**

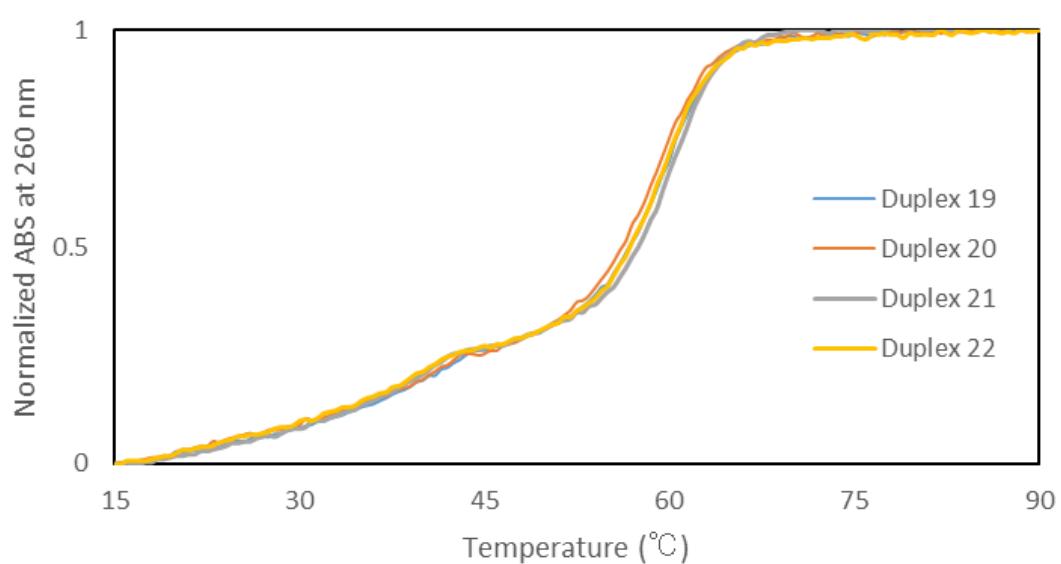


**Figure S5. Thermal Stability of Duplexes: the graphical data of  $1/T_m$  vs  $\ln(C_T/4)$**

## plots



**Figure S6. Thermal Stability of Duplexes: the UV melting profiles of duplex 4-15**



**Figure S7. Thermal Stability of Duplexes: the UV melting profiles of duplex 19-22.**

**Table S1. Thermal Stability of Duplexes: the sequences and  $T_m$  values of duplex 4-15.**

Abbreviation of DNA/RNA duplex	Abbreviation of DNA or RNA	Sequence <sup>a</sup>	$T_m$ (°C) <sup>b</sup>	$\Delta T_m$ (°C) <sup>c</sup>
Duplex 4	DNA 4	5' - d(GGCTATA <u>AATGTCG</u> ) - 3'	47.4	—
	RNA 2	5'- r(CGACAUU <u>UAAGCC</u> ) -3'		
Duplex 5	DNA 4	5' - d(GGCTATA <u>AATGTCG</u> ) - 3'	35.9	-11.5
	RNA 3	5'- r(CGACAUU <u>UUAGCC</u> ) -3'		
Duplex 6	DNA 4	5' - d(GGCTATA <u>AATGTCG</u> ) - 3'	42.1	-5.3
	RNA 4	5'- r(CGACAUU <u>GUAGCC</u> ) -3'		
Duplex 7	DNA 4	5' - d(GGCTATA <u>AATGTCG</u> ) - 3'	33.6	-13.8
	RNA 5	5'- r(CGACAUU <u>CUAGCC</u> ) -3'		
Duplex 8	DNA 5	5' – d(GGCTA <u>TAATGTCG</u> ) – 3'	47.1	—
	RNA 2	5'- r(CGACAUU <u>UAAGCC</u> ) -3'		
Duplex 9	DNA 5	5' – d(GGCTA <u>TAATGTCG</u> ) – 3'	31.3	-15.8
	RNA 3	5'- r(CGACAUU <u>UUAGCC</u> ) -3'		
Duplex 10	DNA 5	5' - d(GGCTA <u>TAATGTCG</u> ) - 3'	41.1	-6.0
	RNA 4	5'- r(CGACAUU <u>GUAGCC</u> ) -3'		
Duplex 11	DNA 5	5' - d(GGCTA <u>TAATGTCG</u> ) - 3'	32.4	-14.7
	RNA 5	5'- r(CGACAUU <u>CUAGCC</u> ) -3'		
Duplex 12	DNA 6	5' - d(GGCTA <u>TAATGTCG</u> ) - 3'	47.3	—
	RNA 2	5'- r(CGACAUU <u>UAAGCC</u> ) -3'		
Duplex 13	DNA 6	5' - d(GGCTA <u>TAATGTCG</u> ) - 3'	34.0	-13.3
	RNA 3	5'- r(CGACAUU <u>UUAGCC</u> ) -3'		
Duplex 14	DNA 6	5' - d(GGCTA <u>TAATGTCG</u> ) - 3'	43.3	-4.0
	RNA 4	5'- r(CGACAUU <u>GUAGCC</u> ) -3'		
Duplex 15	DNA 6	5' – d(GGCTA <u>TAATGTCG</u> ) – 3'	32.7	-14.6
	RNA 5	5'- r(CGACAUU <u>CUAGCC</u> ) -3'		

<sup>a</sup>Underlined letters denote mismatched base pairs. T(red) and T(green) denote (*S*)-5'-C-Aminopropyl-thymidine (**A3**) and (*S*)-5'-C-Aminopropyl-2'-β-fluoro-thymidine (**A4**), respectively. <sup>b</sup>The  $T_m$  value were

measured in 10 mM sodium phosphate buffer (pH 7.0) containing 100 mM NaCl. The concentrations the duplexes were 3  $\mu$ M.  $^c\Delta T_m$  represents [ $T_m$  (duplex fully matched)-  $T_m$  (duplex mismatched)].

**Table S2. Thermal Stability of Duplexes: the sequences and  $T_m$  values of duplex 1, 16-19.**

Abbreviation of DNA/RNA duplex	Abbreviation of DNA or RNA	Sequence <sup>a</sup>	$T_m$ ( $^{\circ}$ C) <sup>b</sup>	$\Delta T_m$ ( $^{\circ}$ C) <sup>c</sup>
Duplex 1	DNA 1	5'- d(TCTTCTCTTCCCTT) -3'	60.0	—
	RNA 1	5'- r(AAGGGAAGAGAAAGA) -3'		
Duplex 19	DNA 10	5'- d(TCTTCTCTTCCCTT) -3'	60.1	+0.1
	RNA 1	5'- r(AAGGGAAGAGAAAGA) -3'		
Duplex 20	DNA 11	5'- d(TCTTCTCTTCCCTT) -3'	59.1	-0.9
	RNA 1	5'- r(AAGGGAAGAGAAAGA) -3'		
Duplex 21	DNA 12	5'- d(TCTTCTCTTCCCTT) -3'	60.3	+0.3
	RNA 1	5'- r(AAGGGAAGAGAAAGA) -3'		
Duplex 22	DNA 13	5'- d(TCTTCTCTTCCCTT) -3'	59.5	-0.5
	RNA 1	5'- r(AAGGGAAGAGAAAGA) -3'		

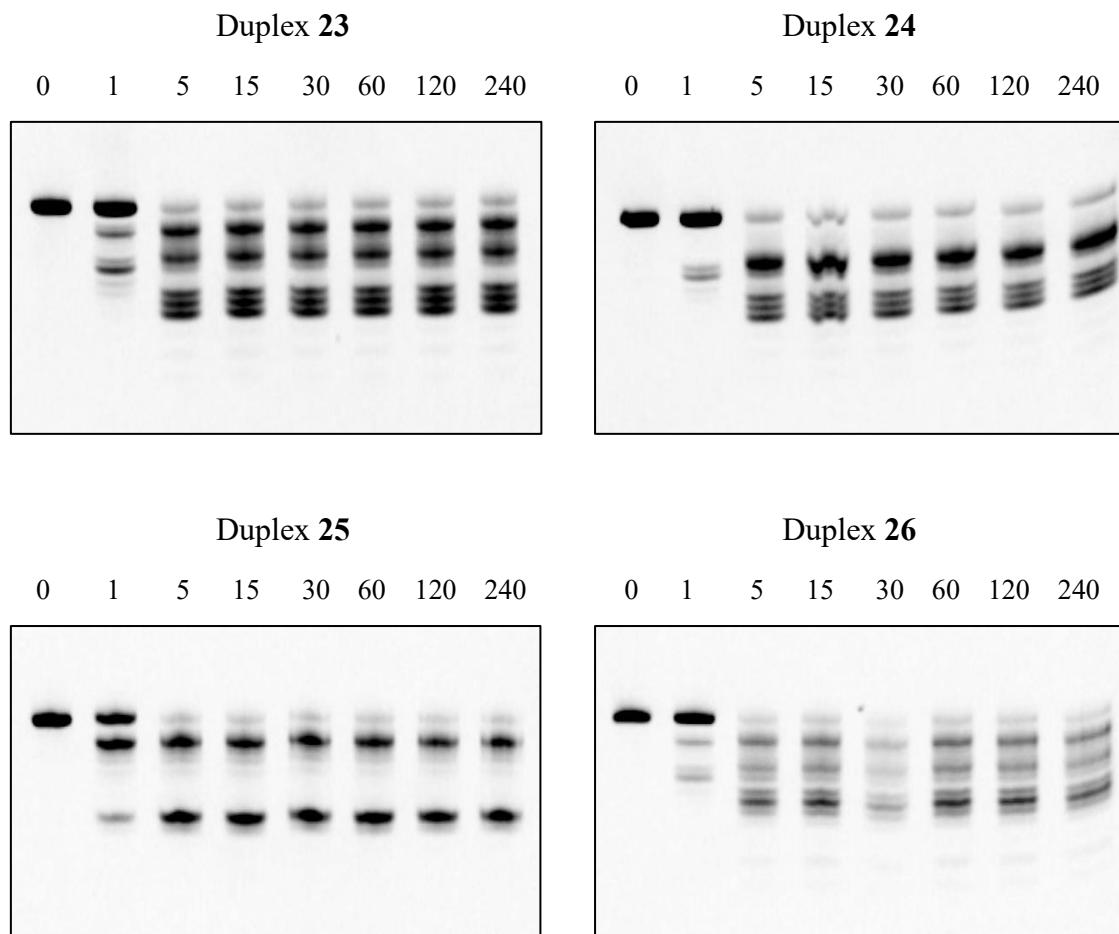
<sup>a</sup> T(green) denotes (S)-5'-C-Aminopropyl-2'- $\beta$ -fluoro-thymidine (**A4**). <sup>b</sup>The  $T_m$  value were measured in 10 mM sodium phosphate buffer (pH 7.0) containing 100 mM NaCl. The concentrations of the duplexes were 3  $\mu$ M. All measurements were carried out three times, and the data are shown as an average value. <sup>c</sup> $\Delta T_m$  represents [ $T_m$  (duplex 19-22)-  $T_m$  (duplex 1)].

(a)

Abbreviation of DNA/RNA duplex	Abbreviation of DNA or RNA	Sequence <sup>a</sup>
Duplex <b>23</b>	DNA <b>10</b>	5'- d(TCTTCTCTTCCCTT) -3'
	RNA <b>6</b>	5'- F - r(AAGGGAAGAGAAAGA) -3'
Duplex <b>24</b>	DNA <b>11</b>	5'- d(TCTTTCTCTTCCCTT) -3'
	RNA <b>6</b>	5'- F - r(AAGGGAAGAGAAAGA) -3'
Duplex <b>25</b>	DNA <b>12</b>	5'- d(TCTTTCTCTTCCCTT) -3'
	RNA <b>6</b>	5'- F - r(AAGGGAAGAGAAAGA) -3'
Duplex <b>26</b>	DNA <b>13</b>	5'- d(TCTTTCTCTTCCCTT) -3'
	RNA <b>6</b>	5'- F - r(AAGGGAAGAGAAAGA) -3'

<sup>a</sup>F(orange) and T(green) denote fluorescein and (S)-5'-C-Aminopropyl-2'-β-fluorothymidine (**A4**), respectively.

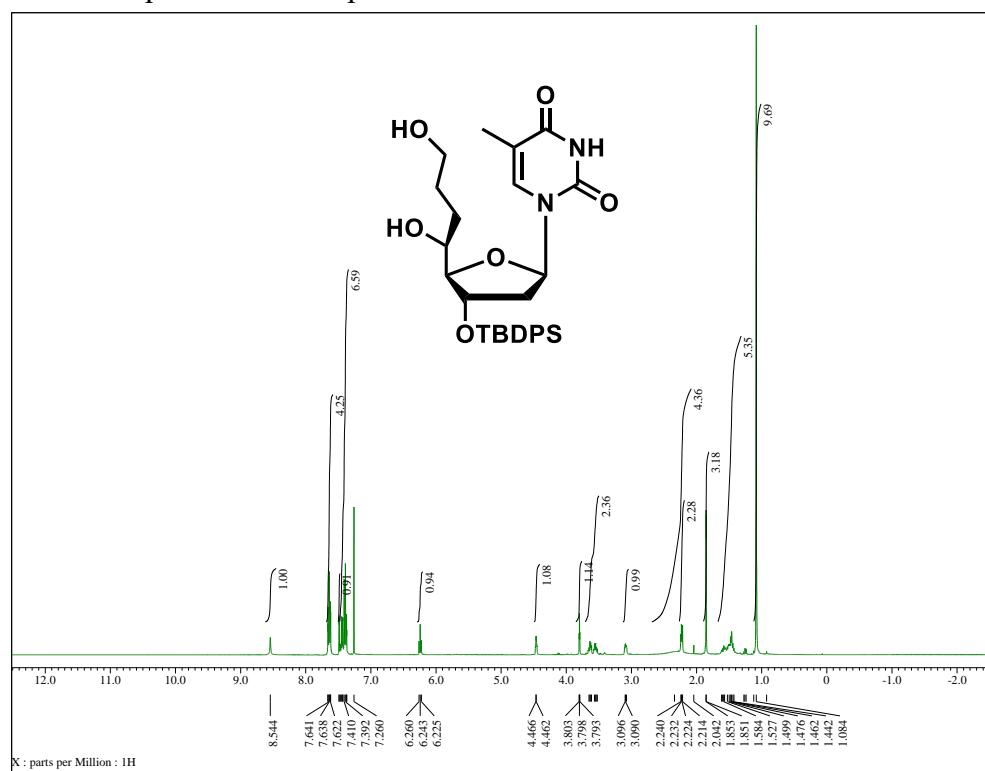
(b)



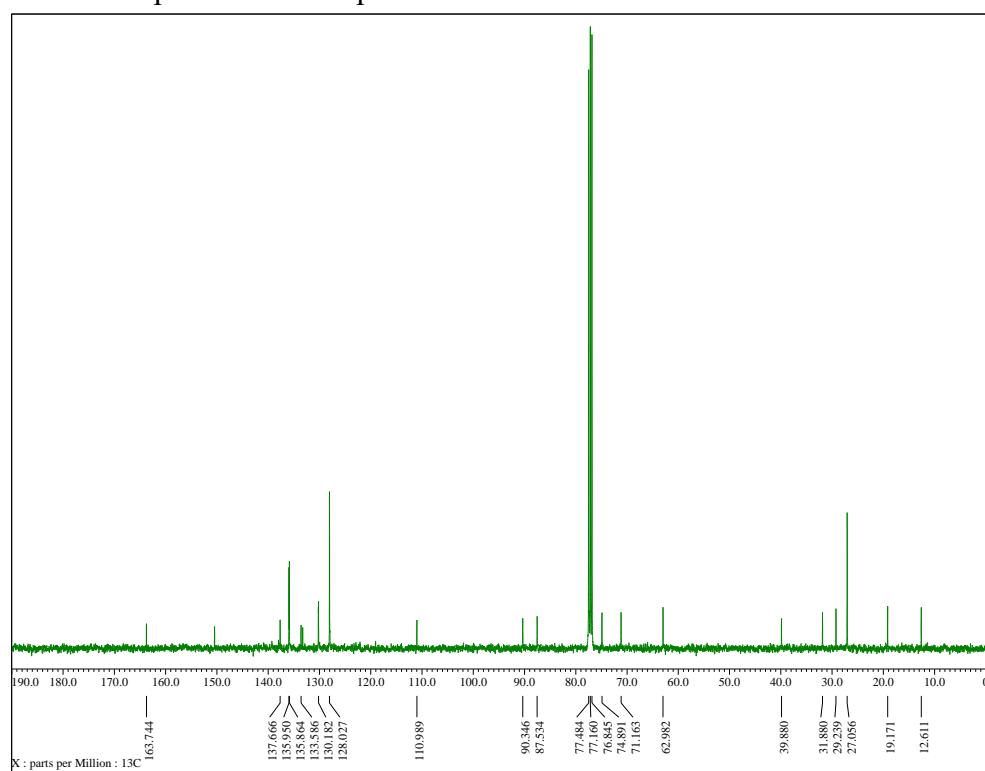
**Figure S8. RNase H-mediated cleavage assays: the sequences of duplex 20-23 (a) and the images of PAGE analysis (b).** PAGE analysis of DNA/RNA duplexes treated in a buffer containing 50 mM Tris-HCl (pH 8.0), 75 mM KCl, 3 mM MgCl<sub>2</sub>, 10 mM dithiothreitol, and RNase H from *E.coli*. The duplexes **23-26** were incubated in a buffer containing 50 mM Tris-HCl (pH 8.0), 75 mM KCl, 3 mM MgCl<sub>2</sub> and 10 mM dithiothreitol, and then diluted RNase H solution (60 unit/L in H<sub>2</sub>O) was added, and subsequently the mixture was incubated at 37°C for the required time. The reaction mixtures at various incubation time (0, 1, 5, 15, 30, 60, 120, 240 min) were analyzed by 20% PAGE.

**NMR spectra ( $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^{19}\text{F}$ ,  $^{31}\text{P}$  and NOESY)**

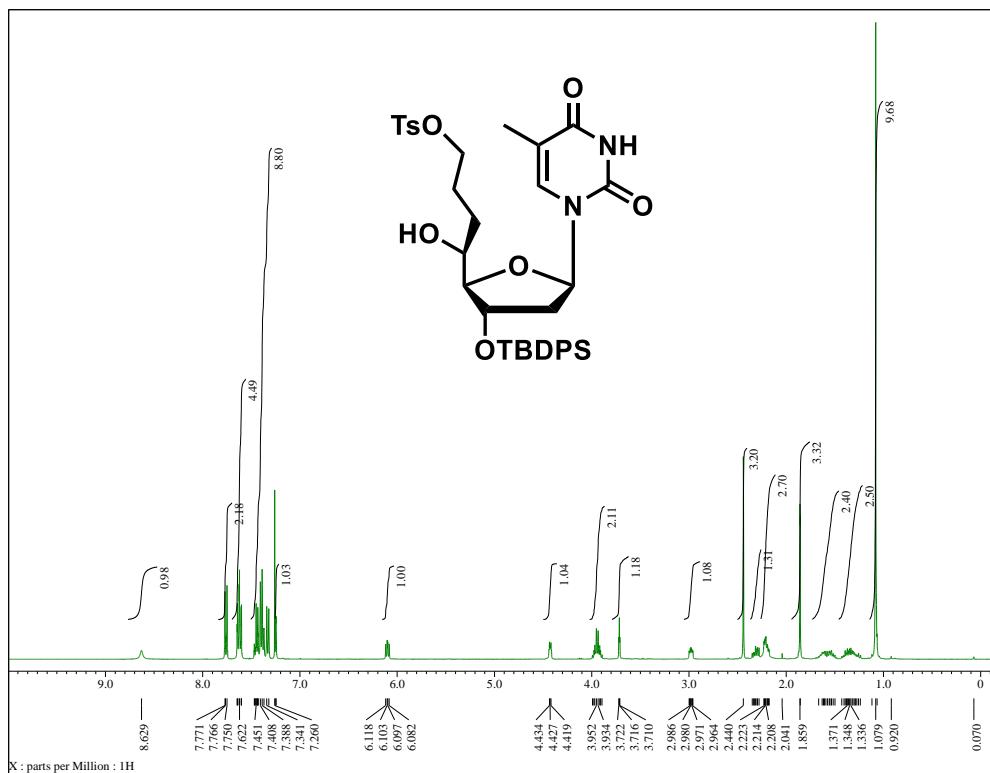
$^1\text{H}$  NMR spectrum of compound 2



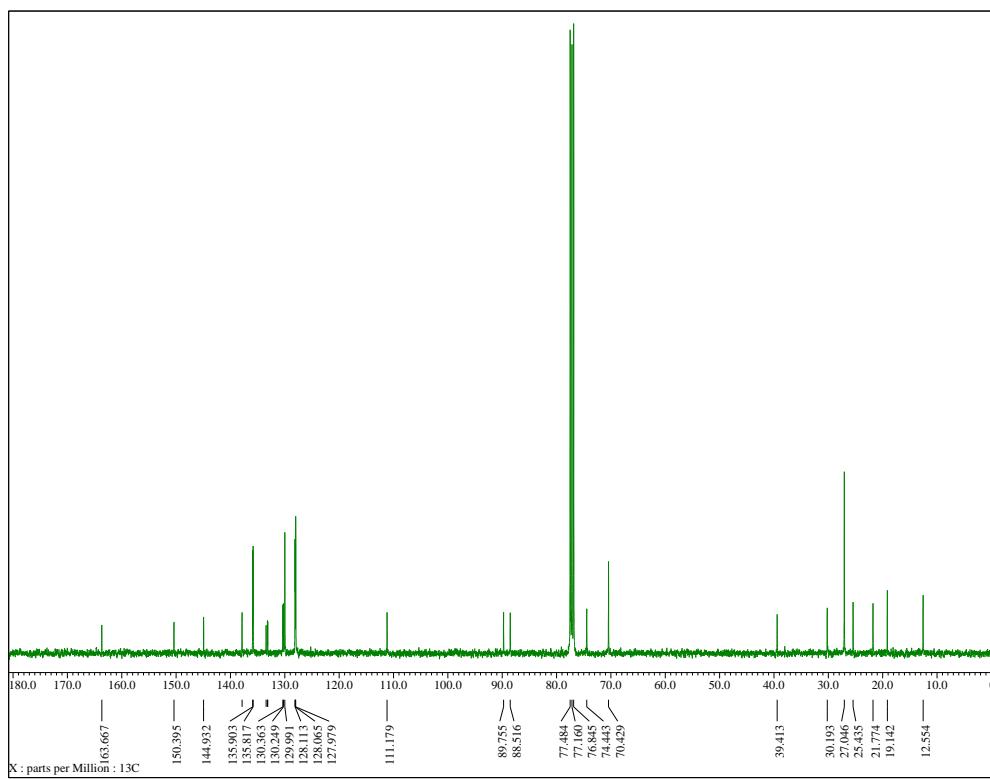
$^{13}\text{C}$  NMR spectrum of compound 2



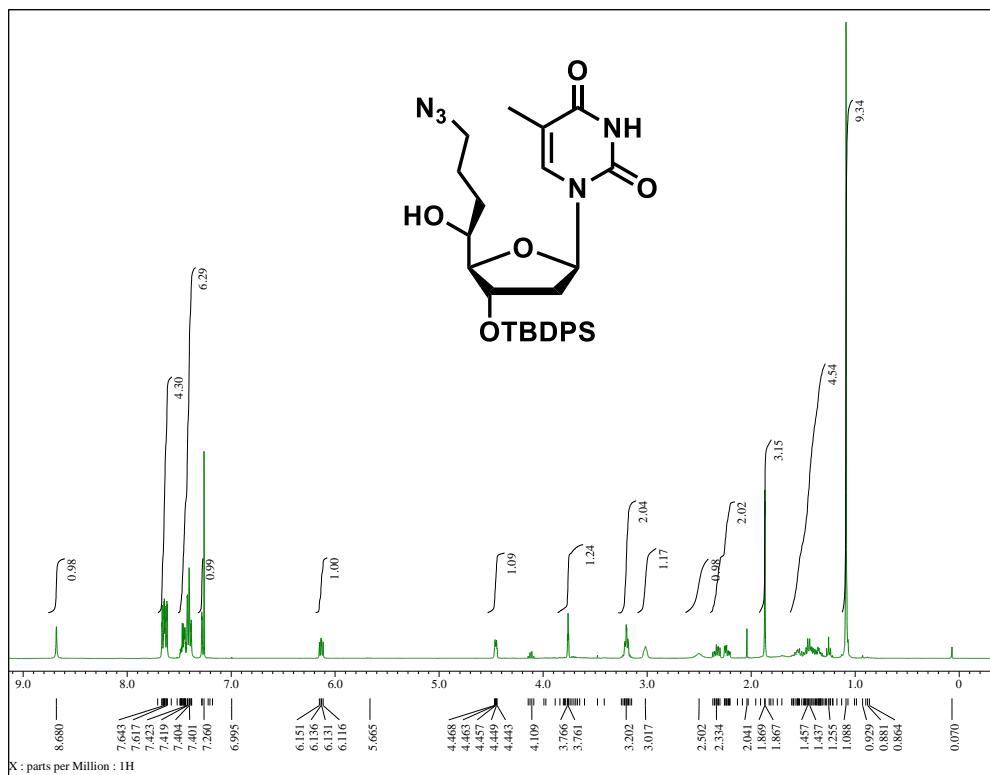
<sup>1</sup>H NMR spectrum of compound 3



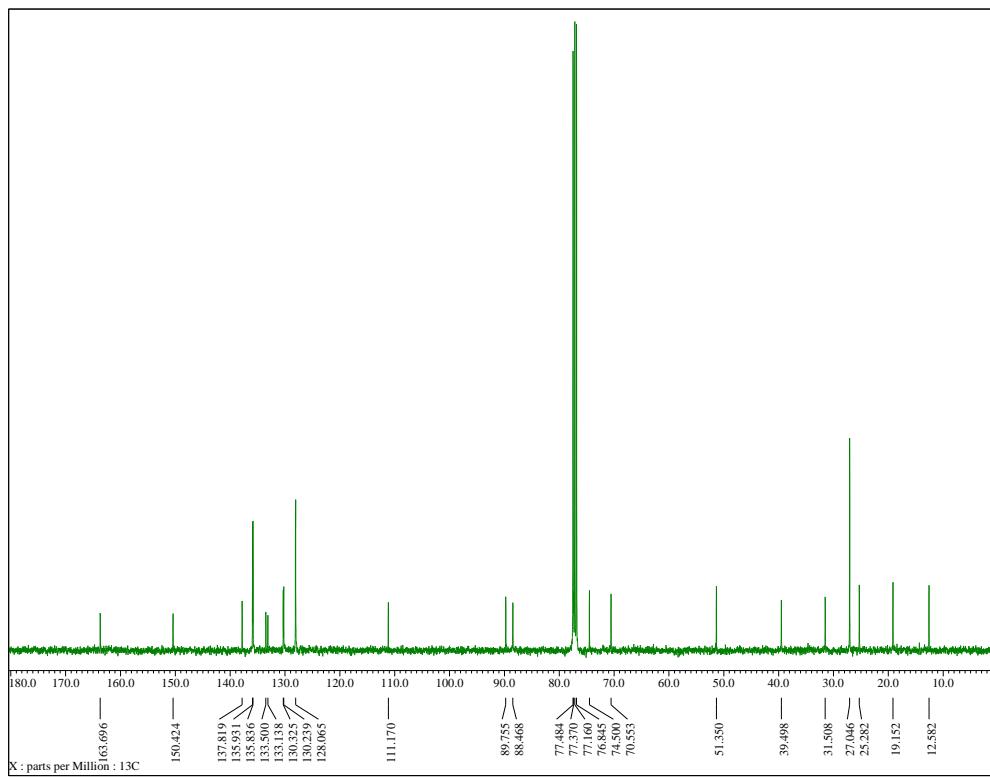
<sup>13</sup>C NMR spectrum of compound 3



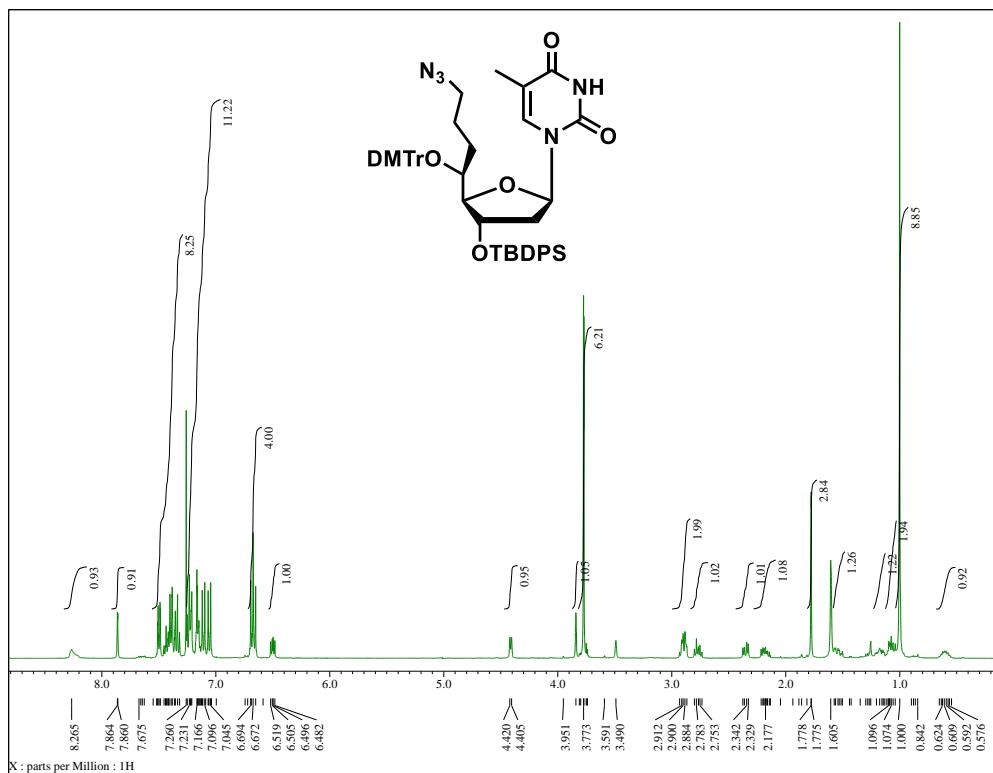
<sup>1</sup>H NMR spectrum of compound 4



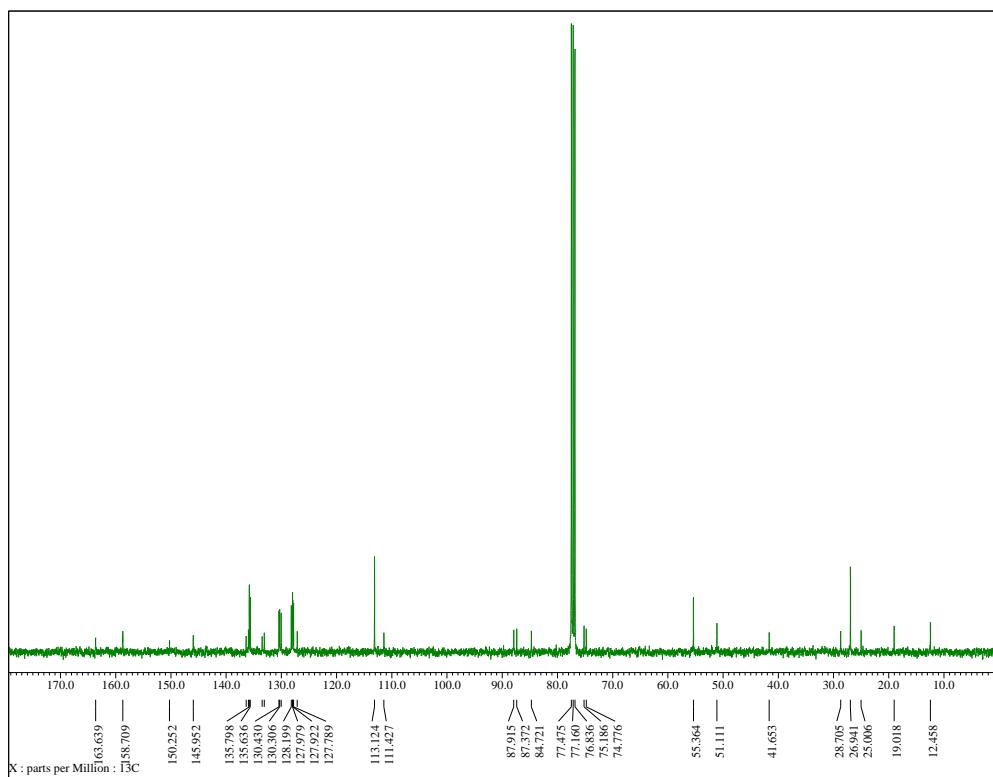
<sup>13</sup>C NMR spectrum of compound 4



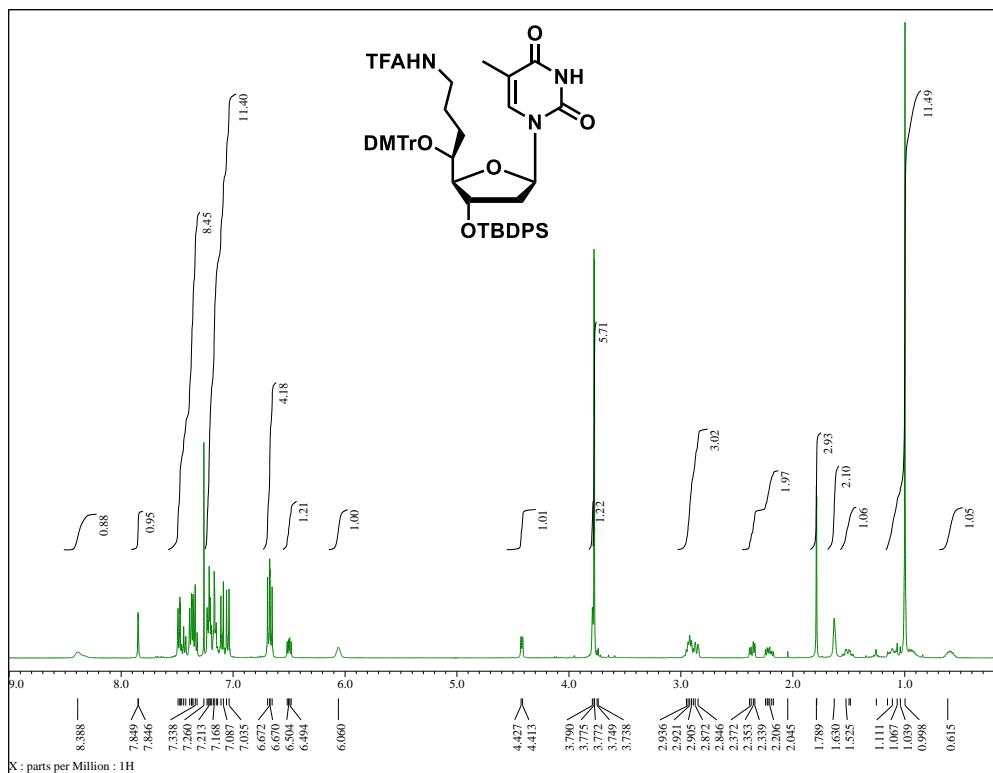
<sup>1</sup>H NMR spectrum of compound 5



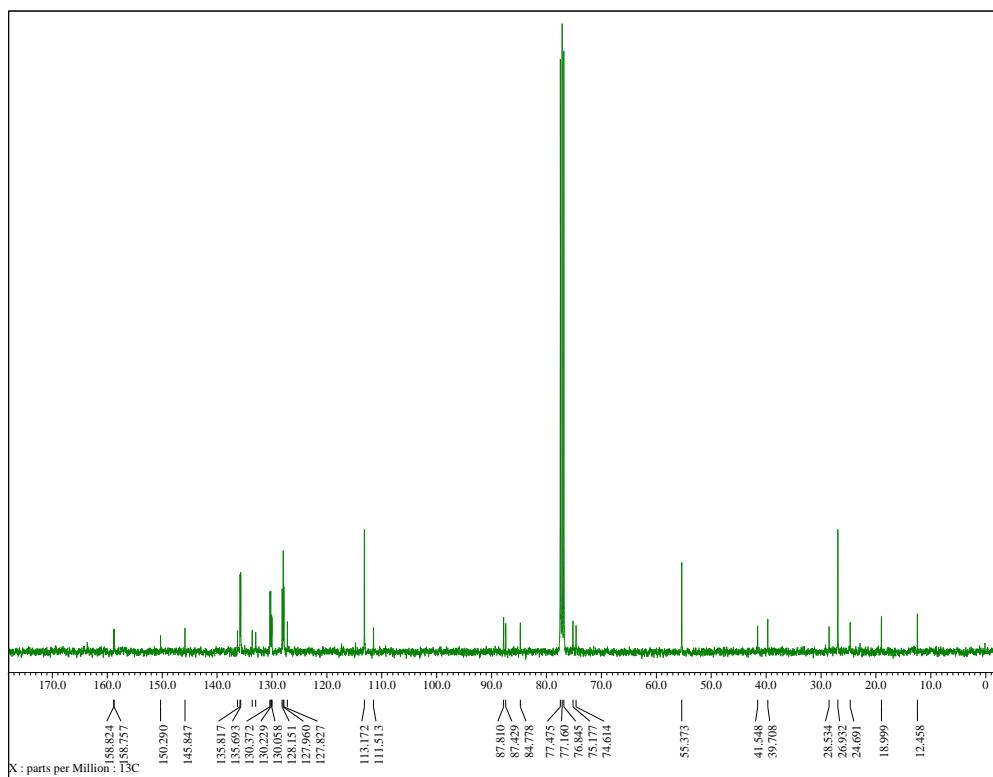
<sup>13</sup>C NMR spectrum of compound 5



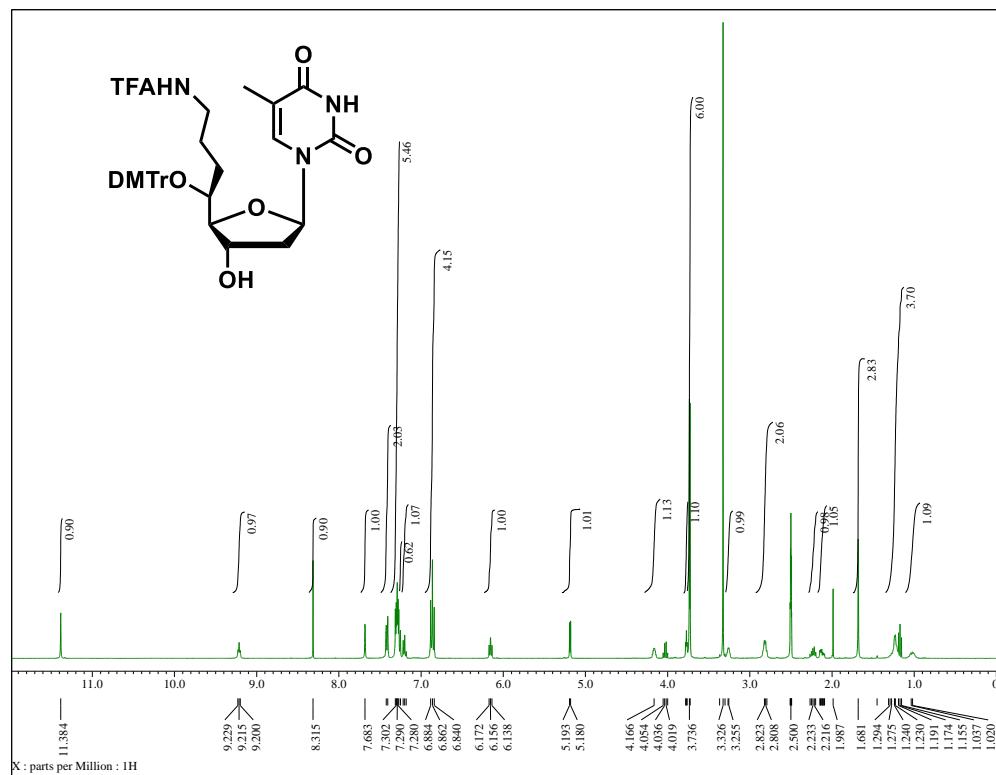
<sup>1</sup>H NMR spectrum of compound 6



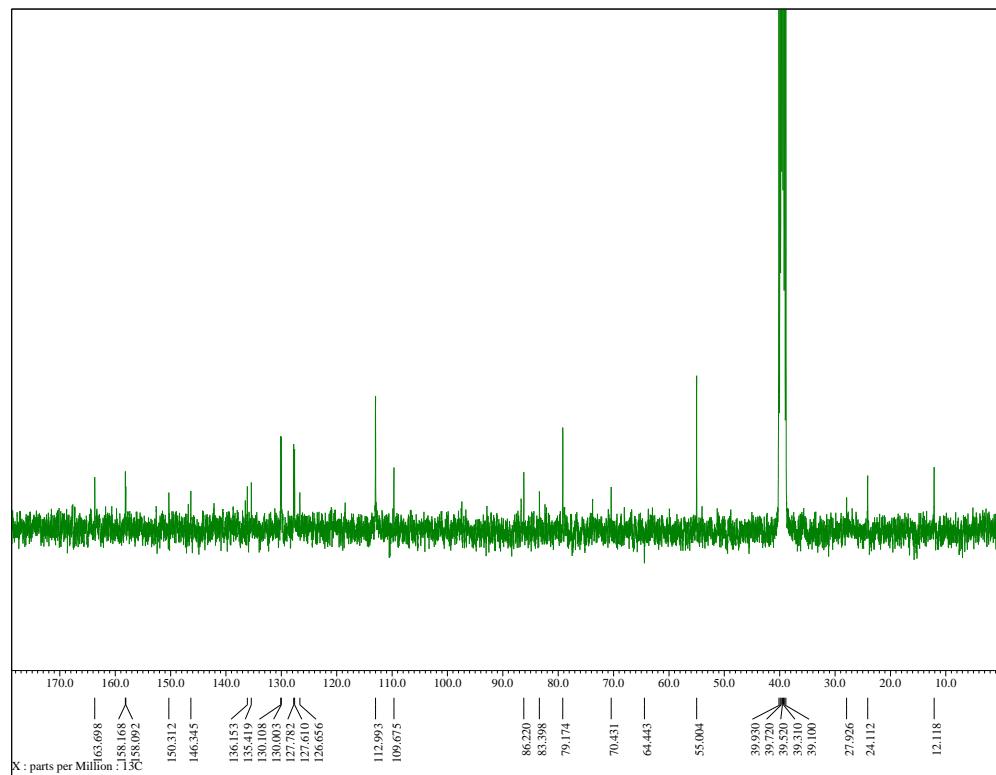
<sup>13</sup>C NMR spectrum of compound 6



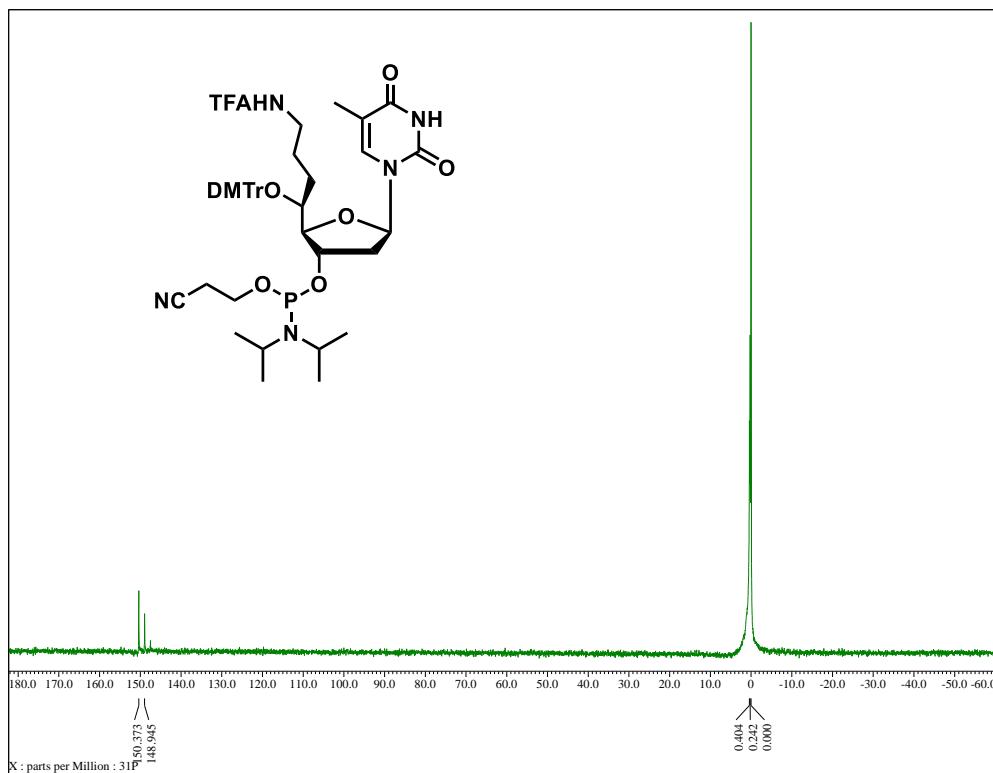
<sup>1</sup>H NMR spectrum of compound 7



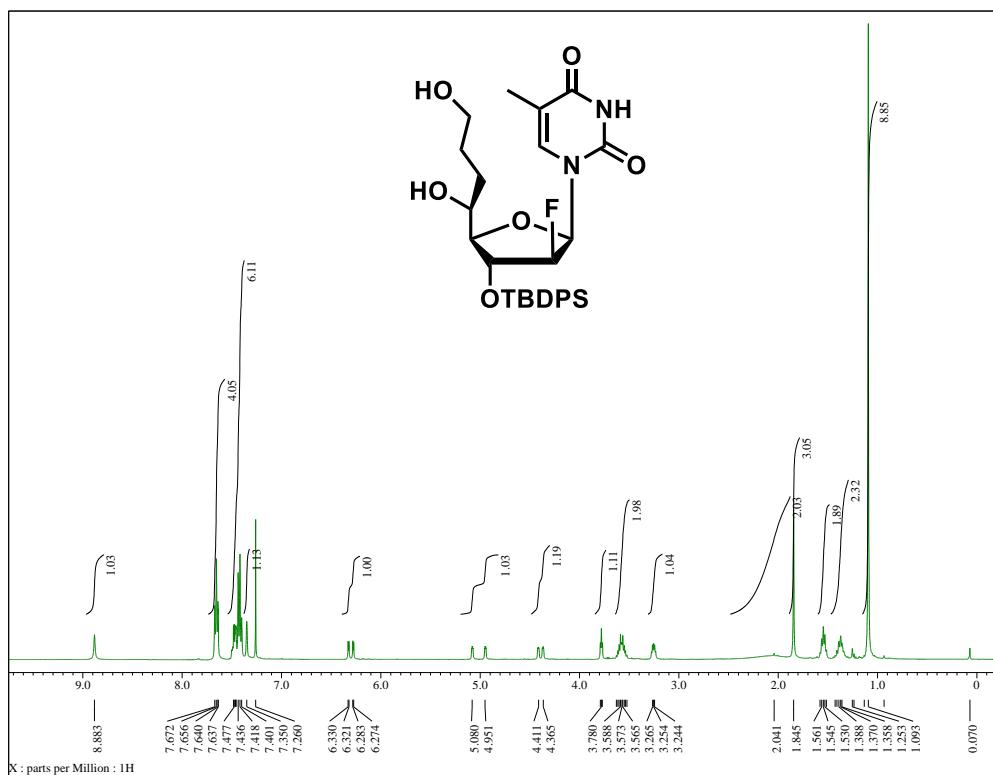
<sup>13</sup>C NMR spectrum of compound 7



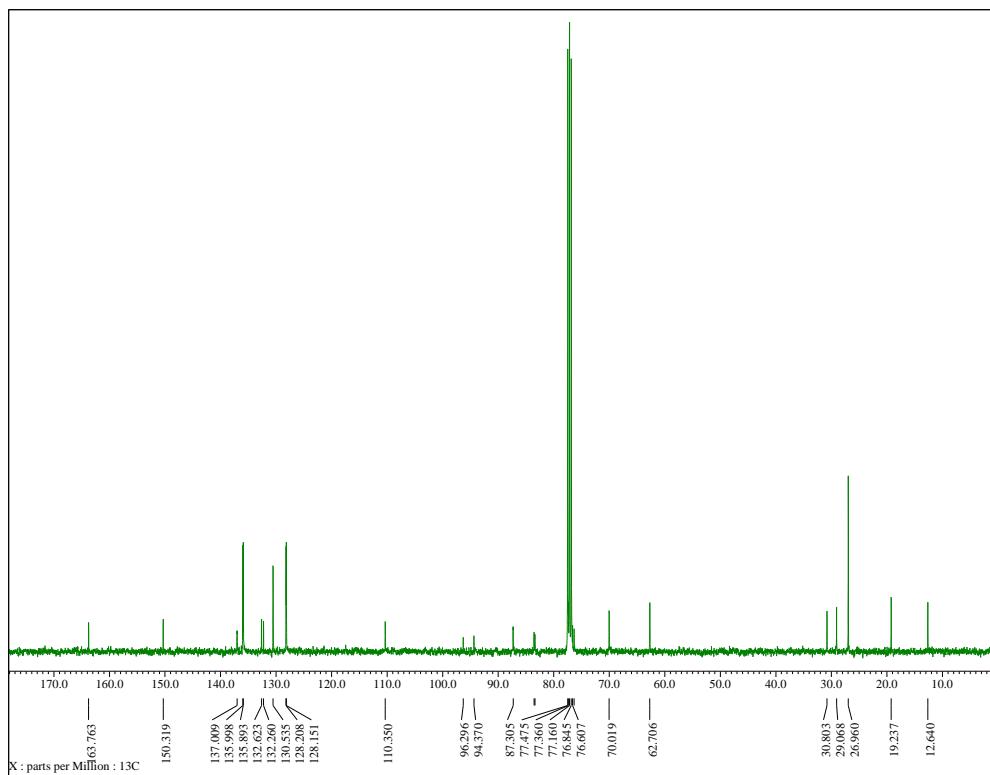
$^{31}\text{P}$  NMR spectrum of compound 8



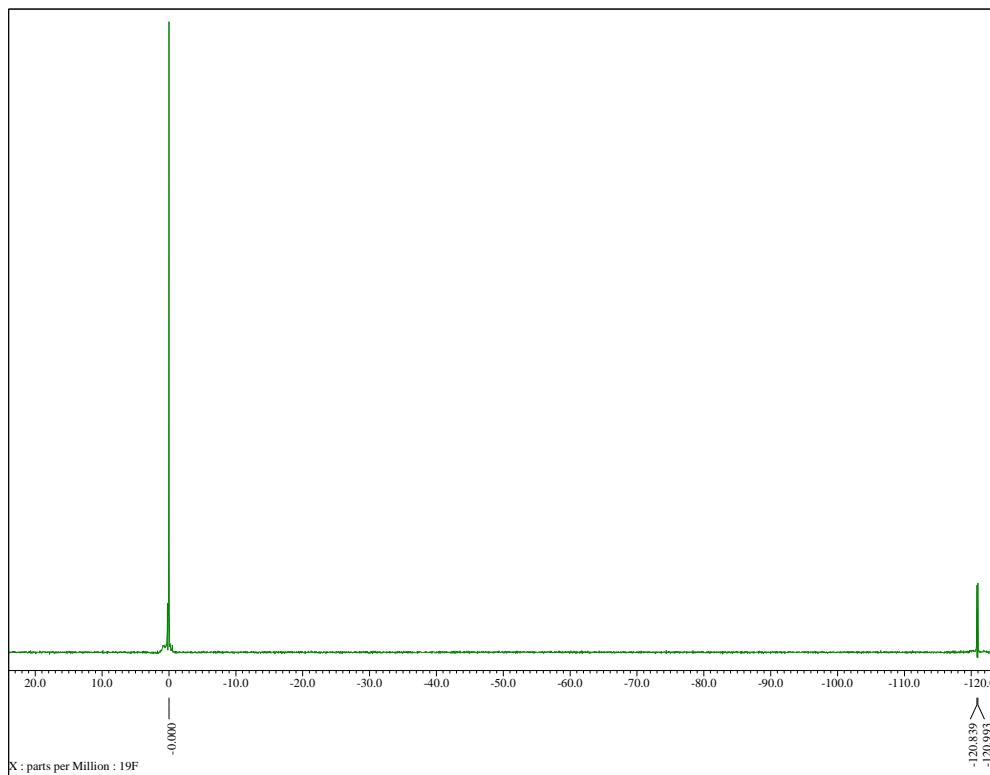
$^1\text{H}$  NMR spectrum of compound 13



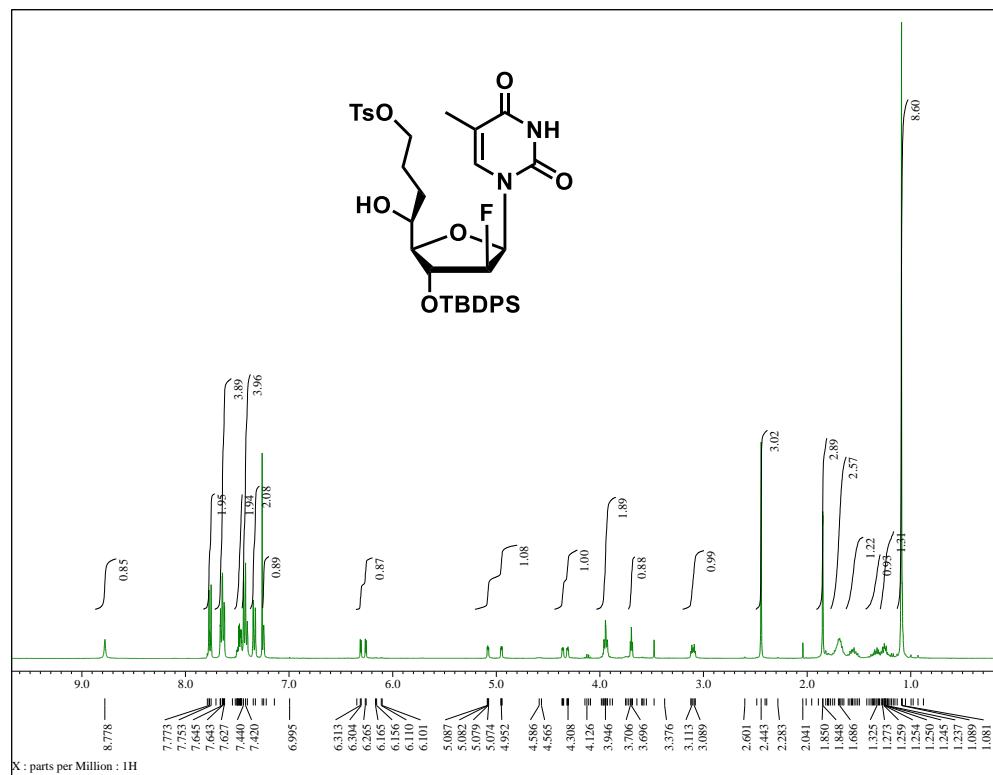
$^{13}\text{C}$  NMR spectrum of compound **13**



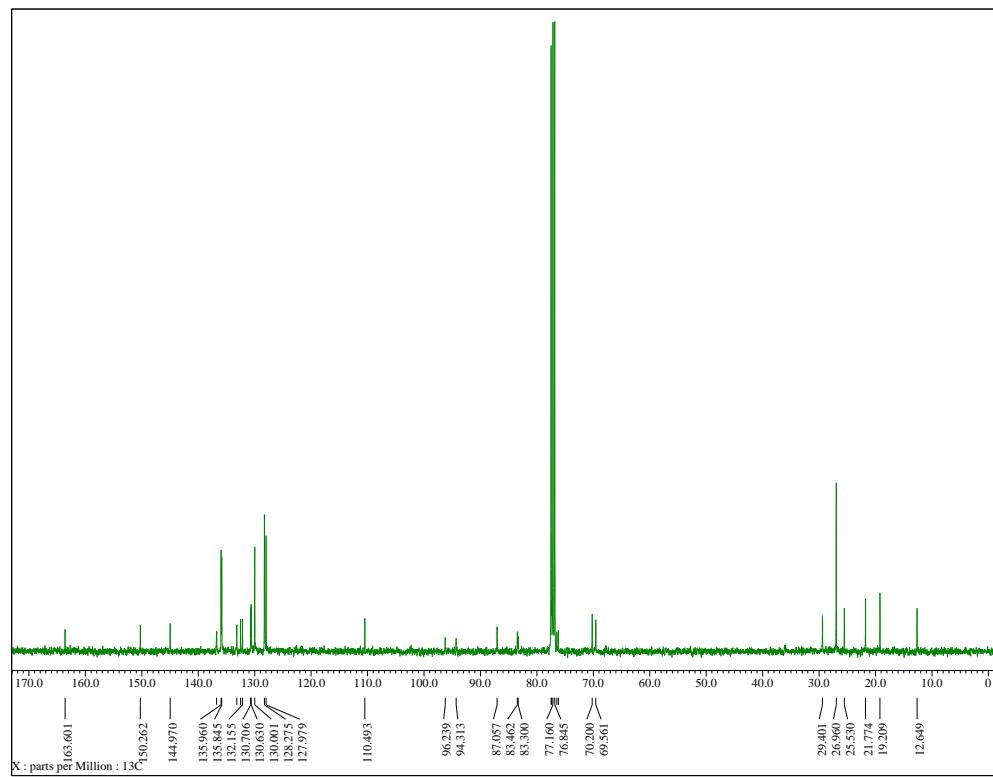
$^{19}\text{F}$  NMR spectrum of compound **13**



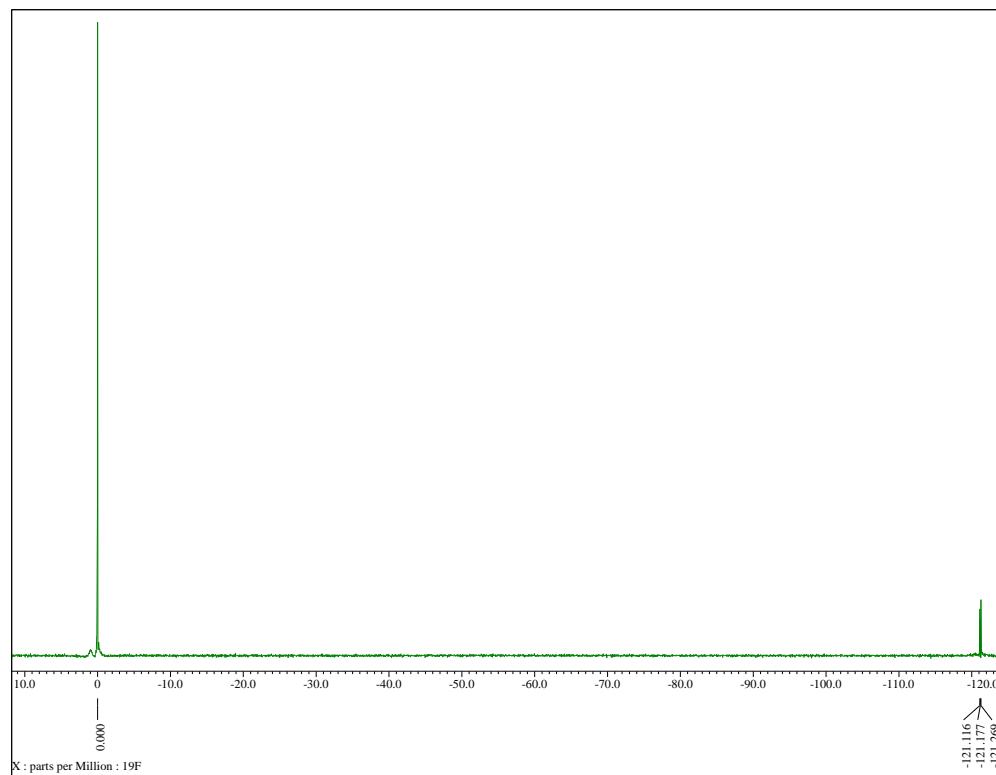
<sup>1</sup>H NMR spectrum of compound 14



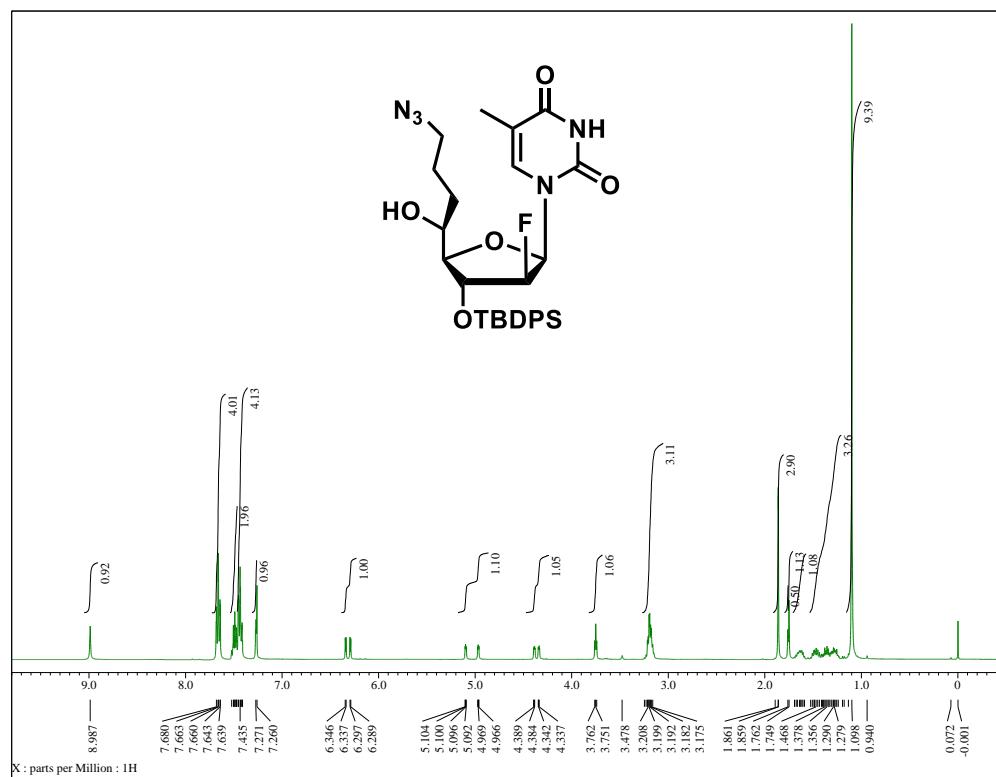
<sup>13</sup>C NMR spectrum of compound 14



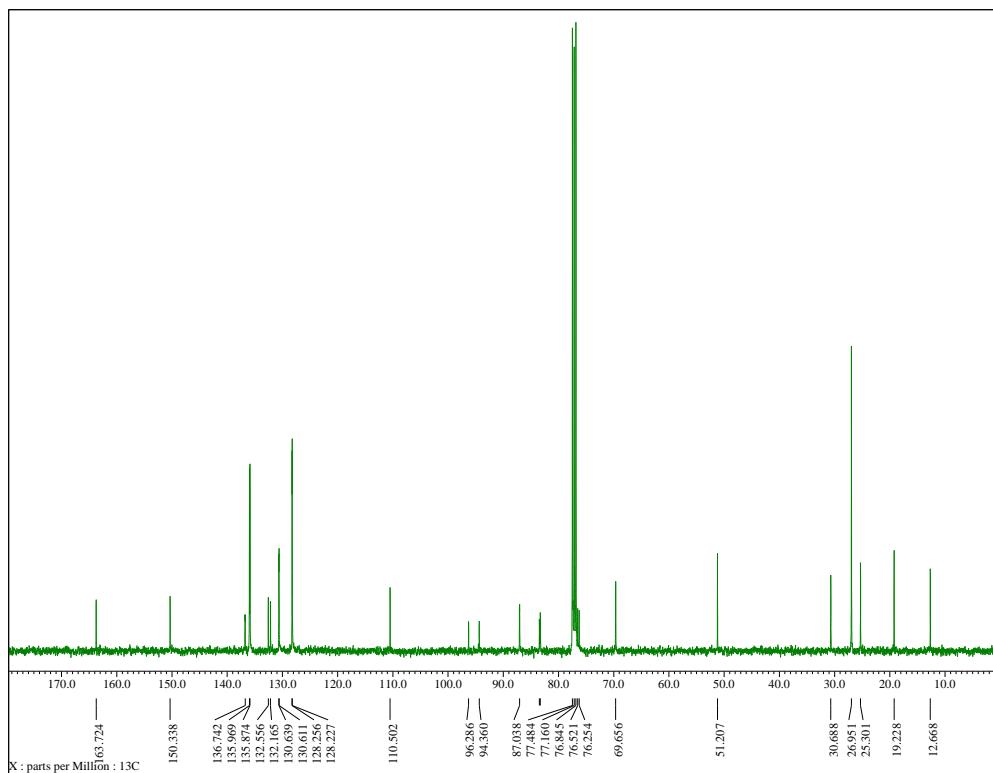
<sup>19</sup>F NMR spectrum of compound 14



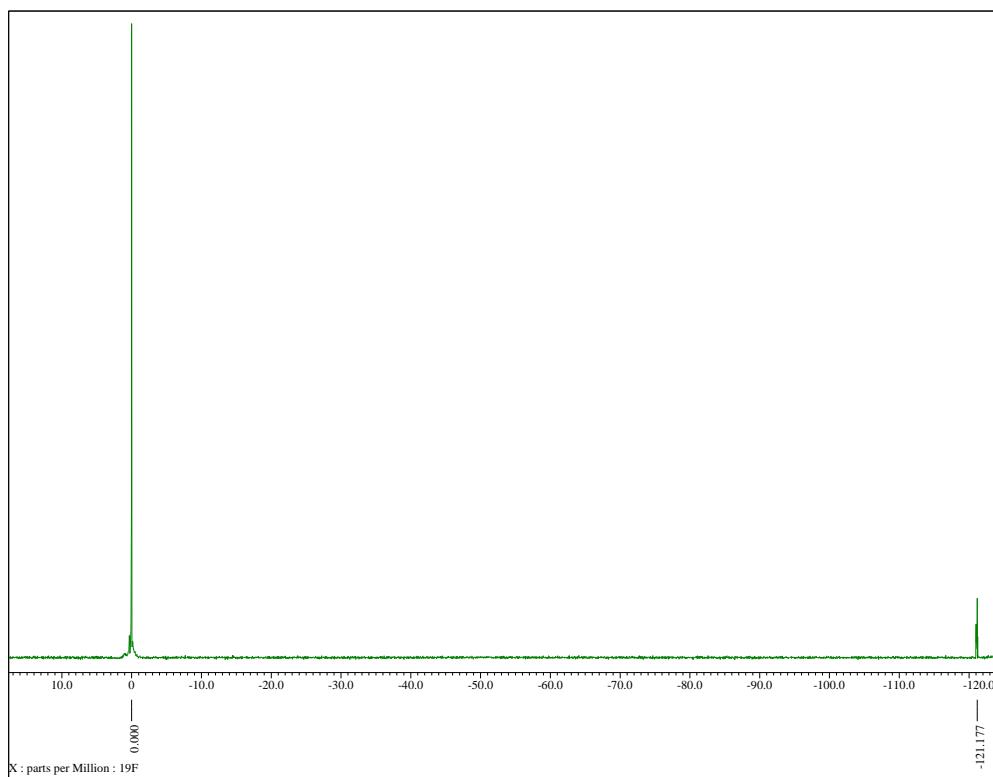
<sup>1</sup>H NMR spectrum of compound 15



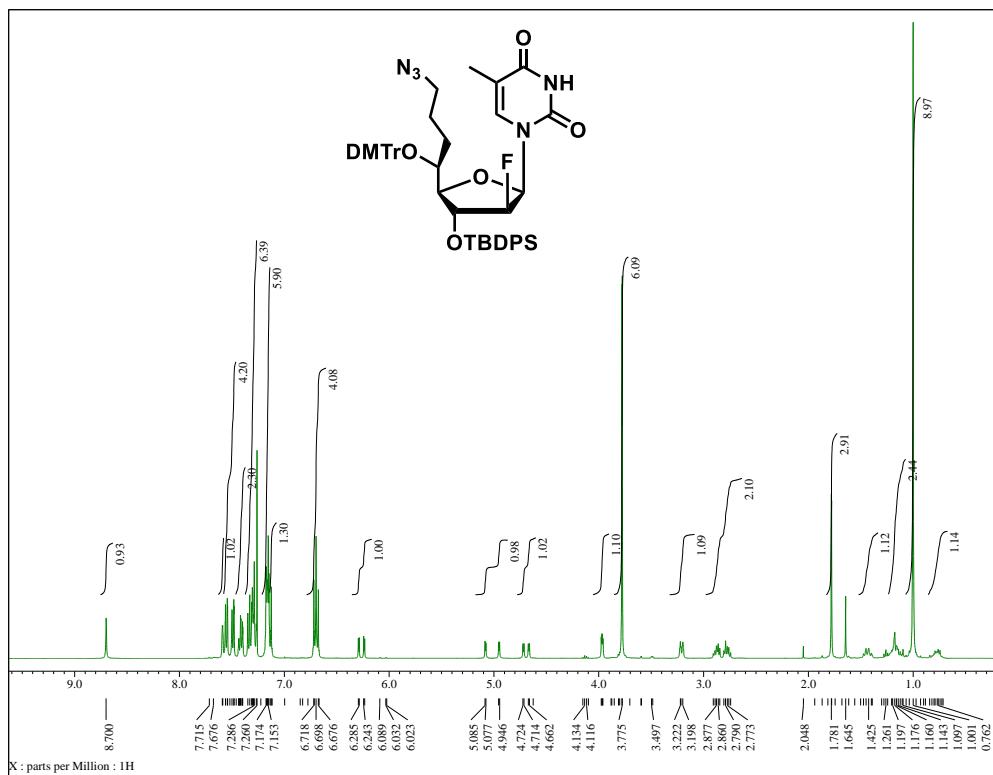
$^{13}\text{C}$  NMR spectrum of compound **15**



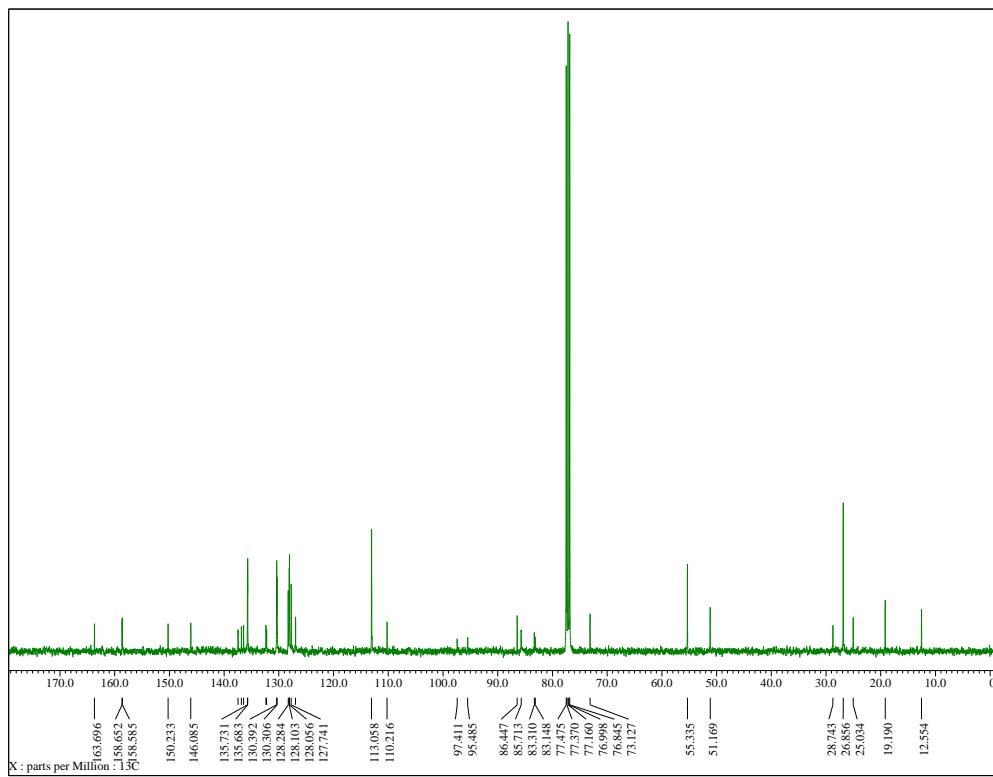
$^{19}\text{F}$  NMR spectrum of compound **15**



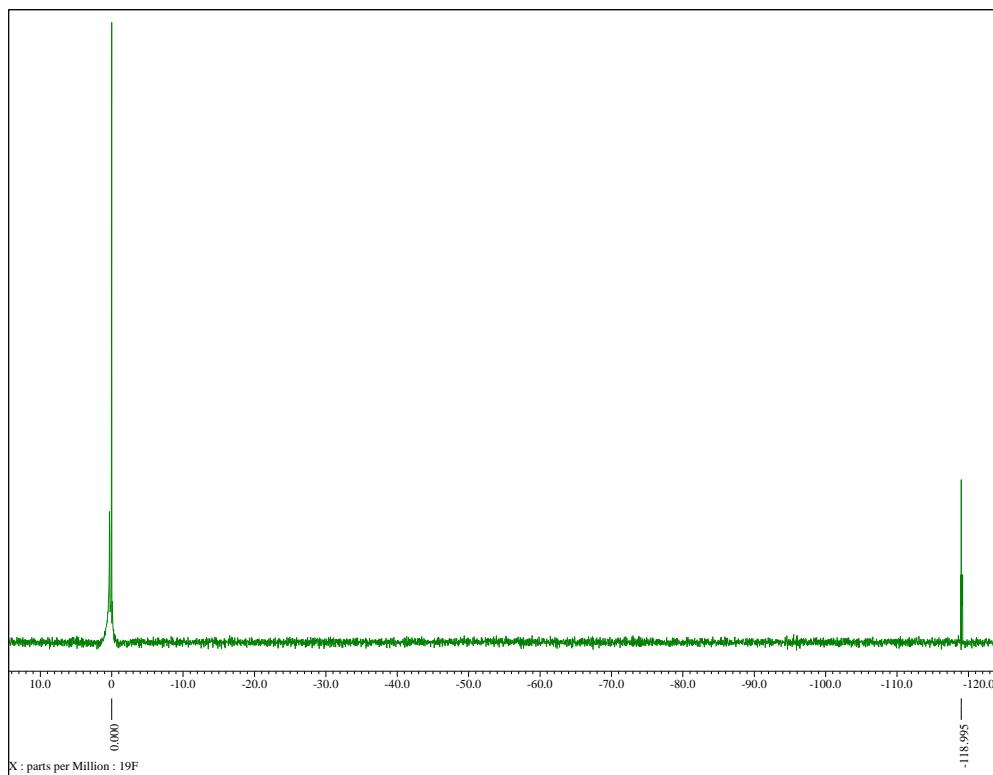
<sup>1</sup>H NMR spectrum of compound 16



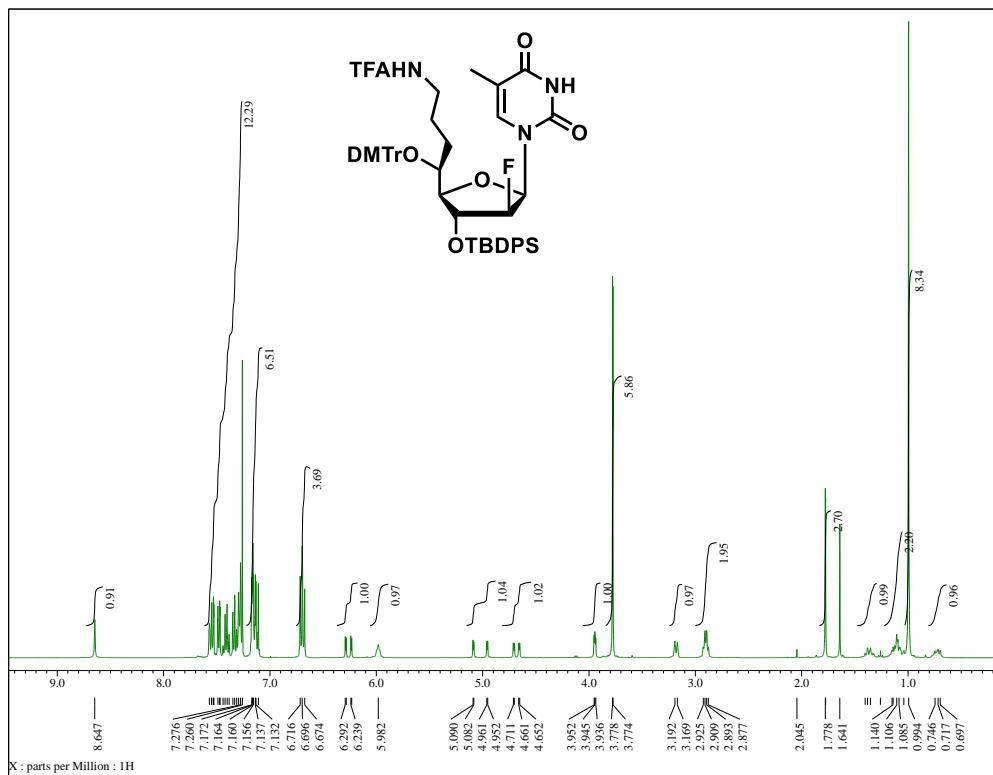
<sup>13</sup>C NMR spectrum of compound 16



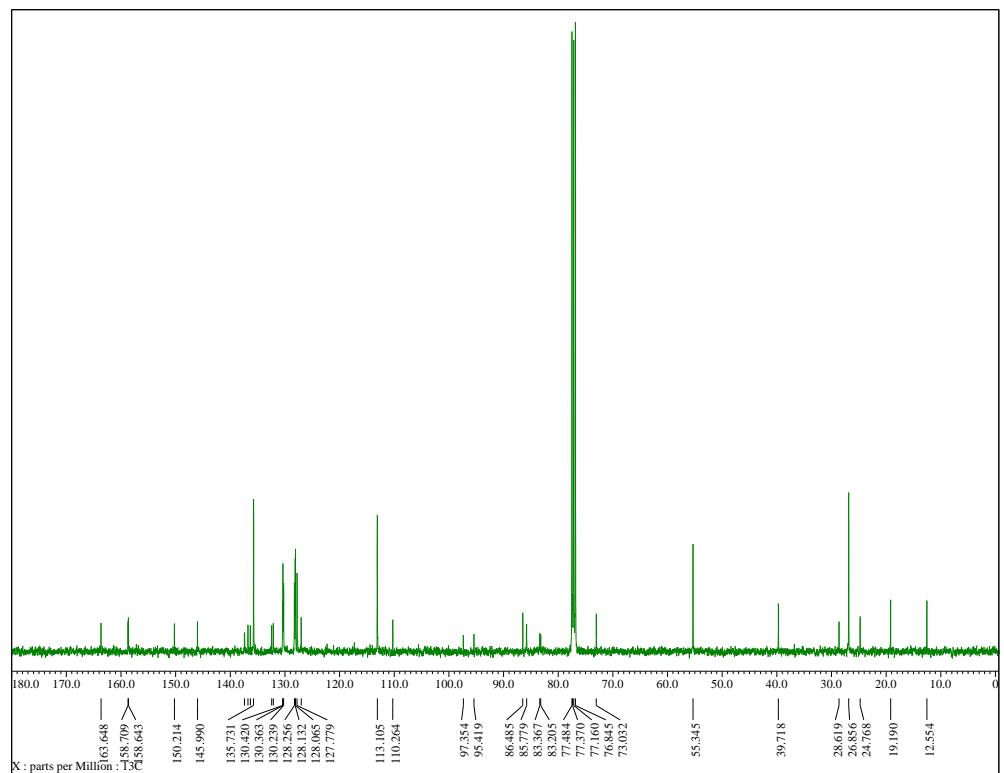
<sup>19</sup>F NMR spectrum of compound **16**



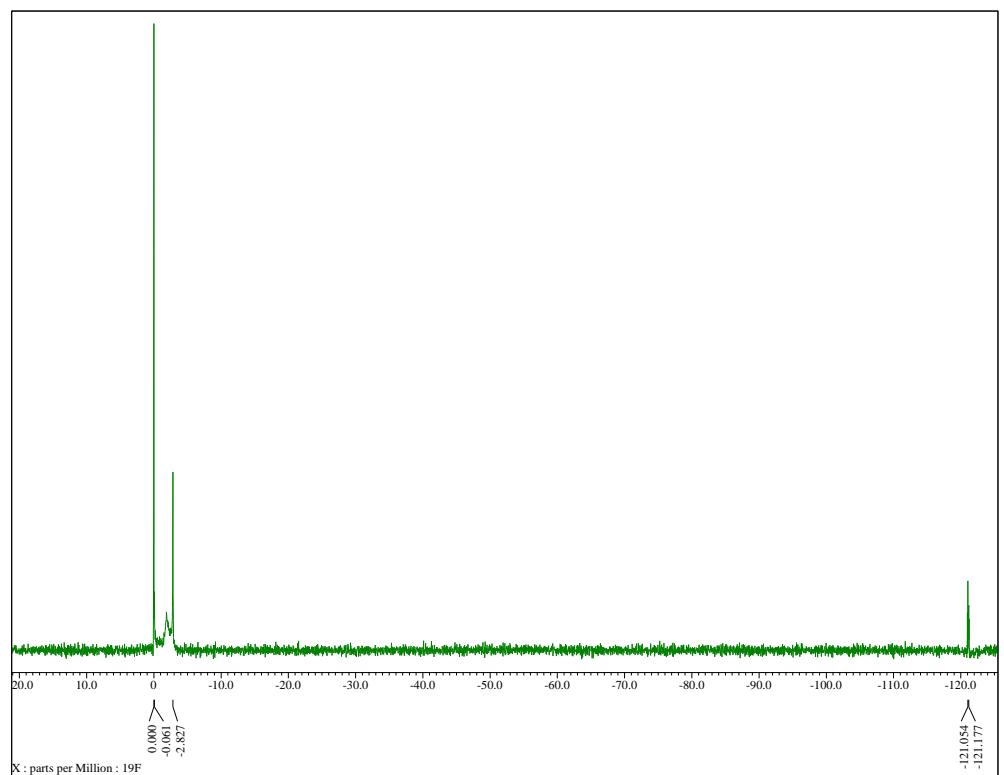
<sup>1</sup>H NMR spectrum of compound **17**



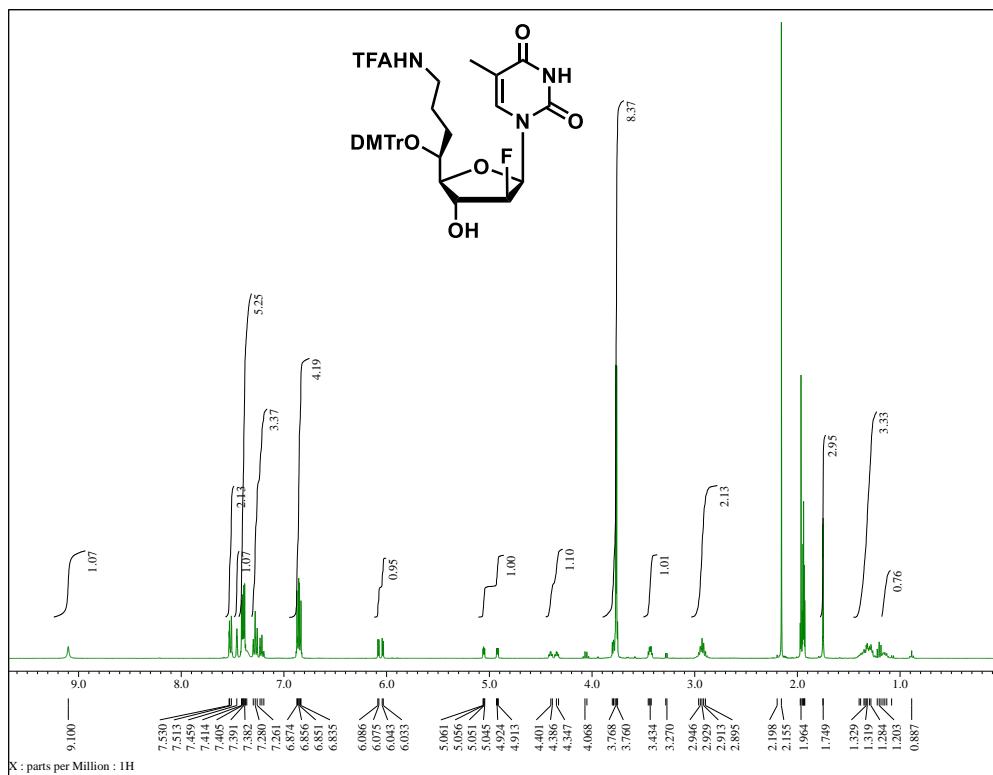
$^{13}\text{C}$  NMR spectrum of compound **17**



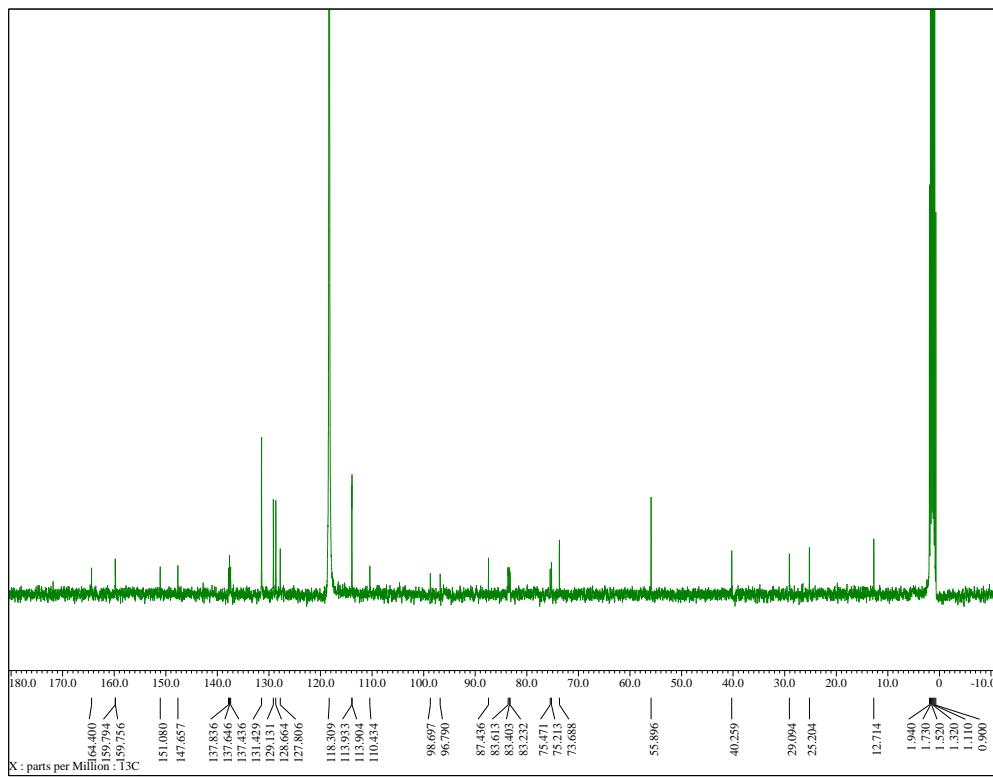
$^{19}\text{F}$  NMR spectrum of compound **17**



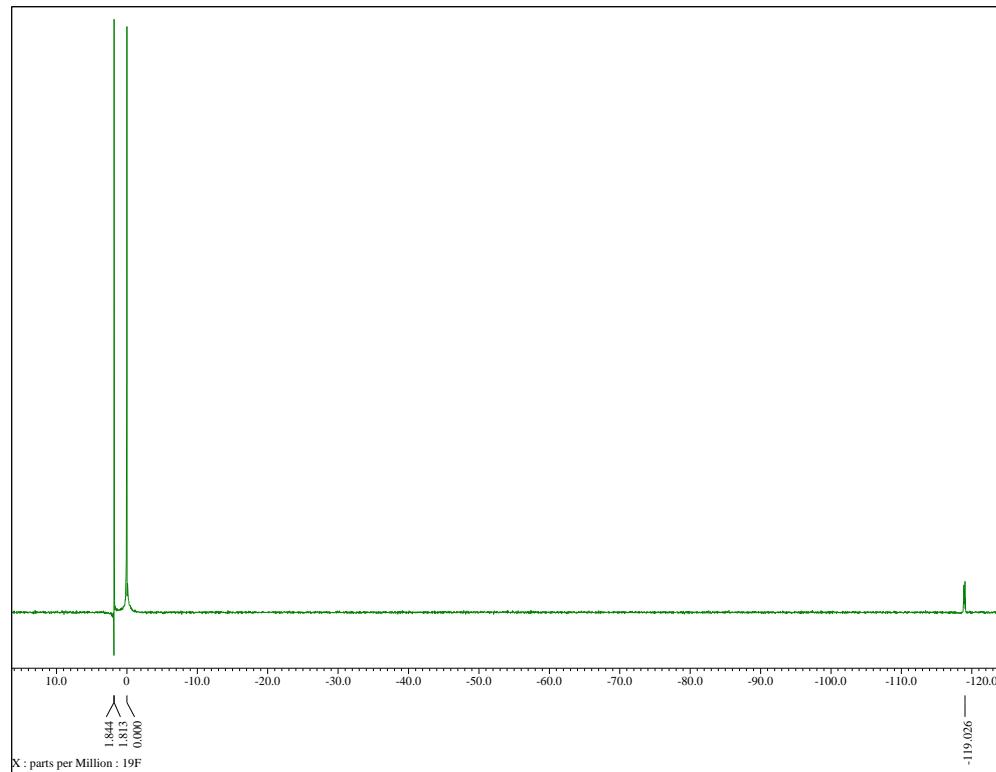
<sup>1</sup>H NMR spectrum of compound 18



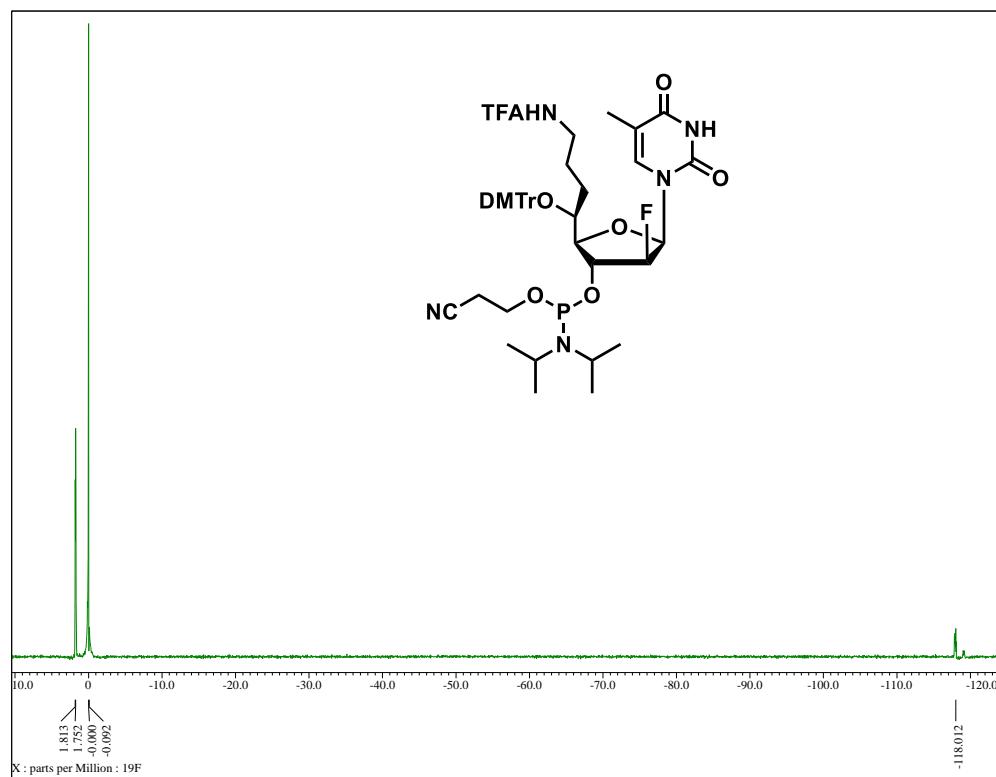
<sup>13</sup>C NMR spectrum of compound 18



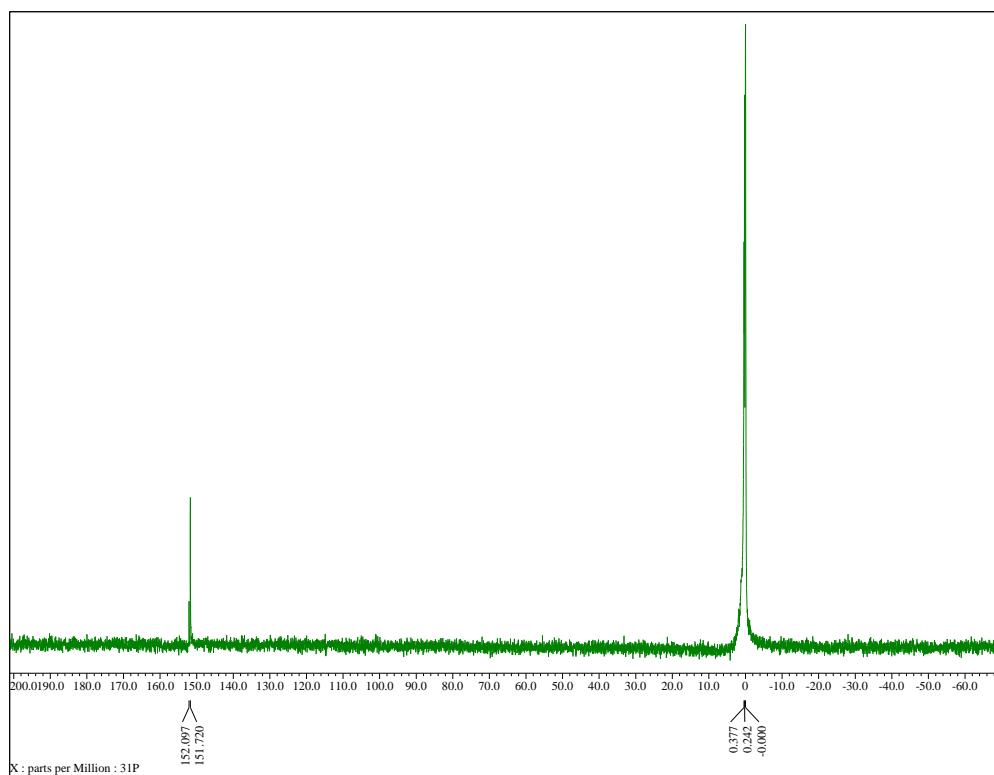
<sup>19</sup>F NMR spectrum of compound **18**



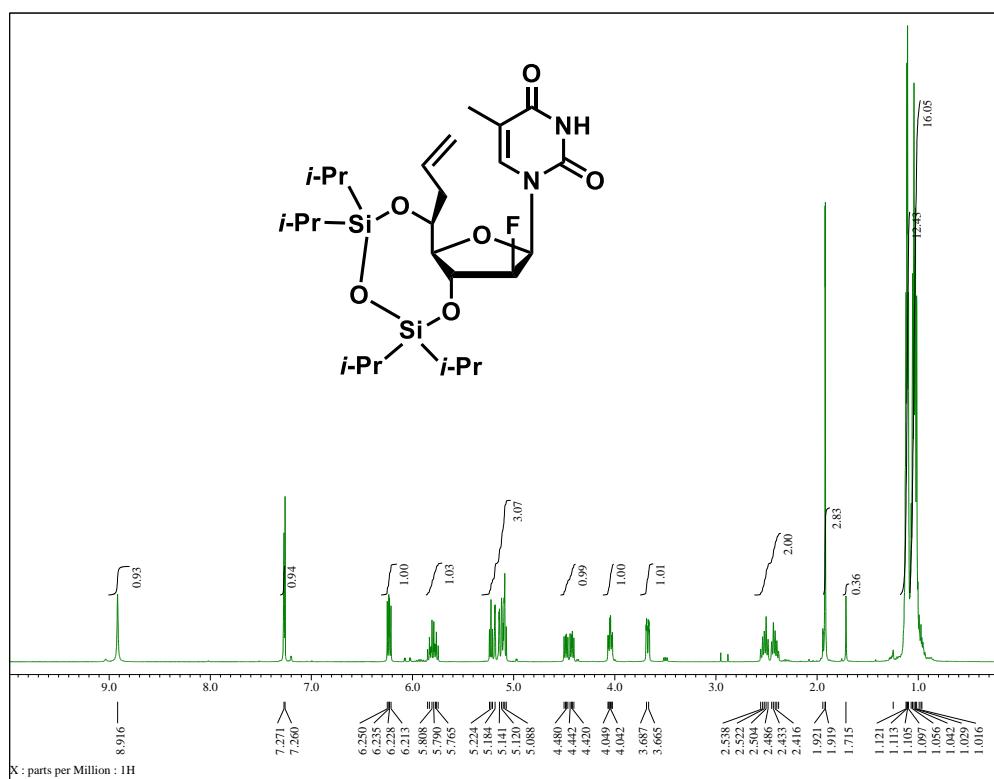
<sup>19</sup>F NMR spectrum of compound **19**



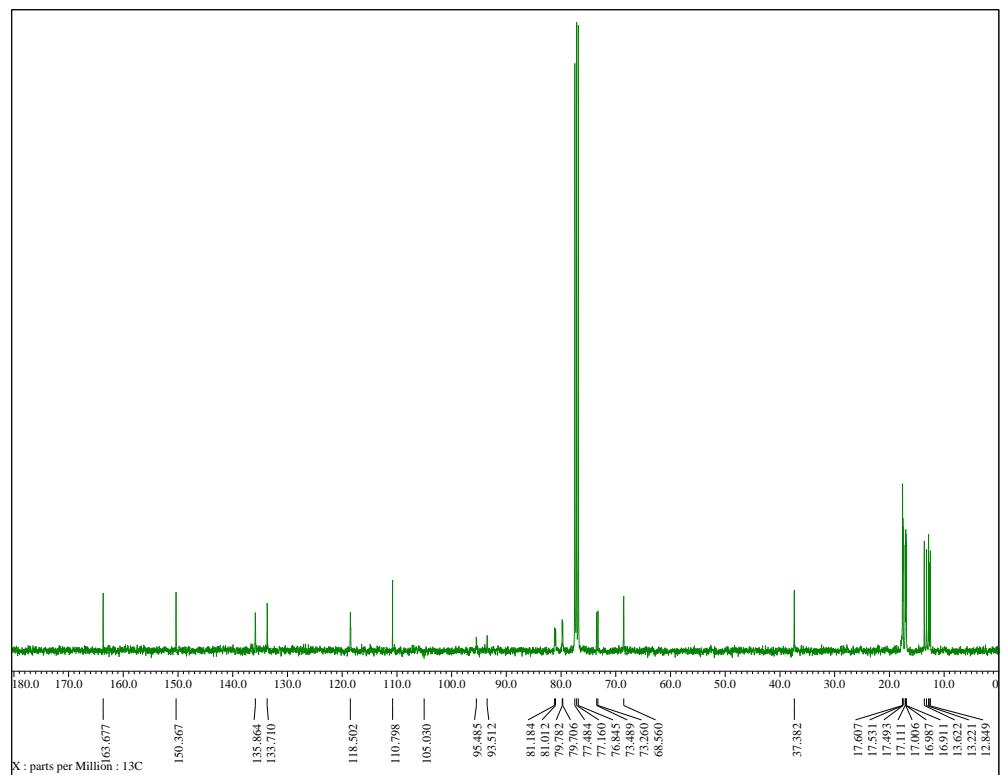
$^{31}\text{P}$  NMR spectrum of compound **19**



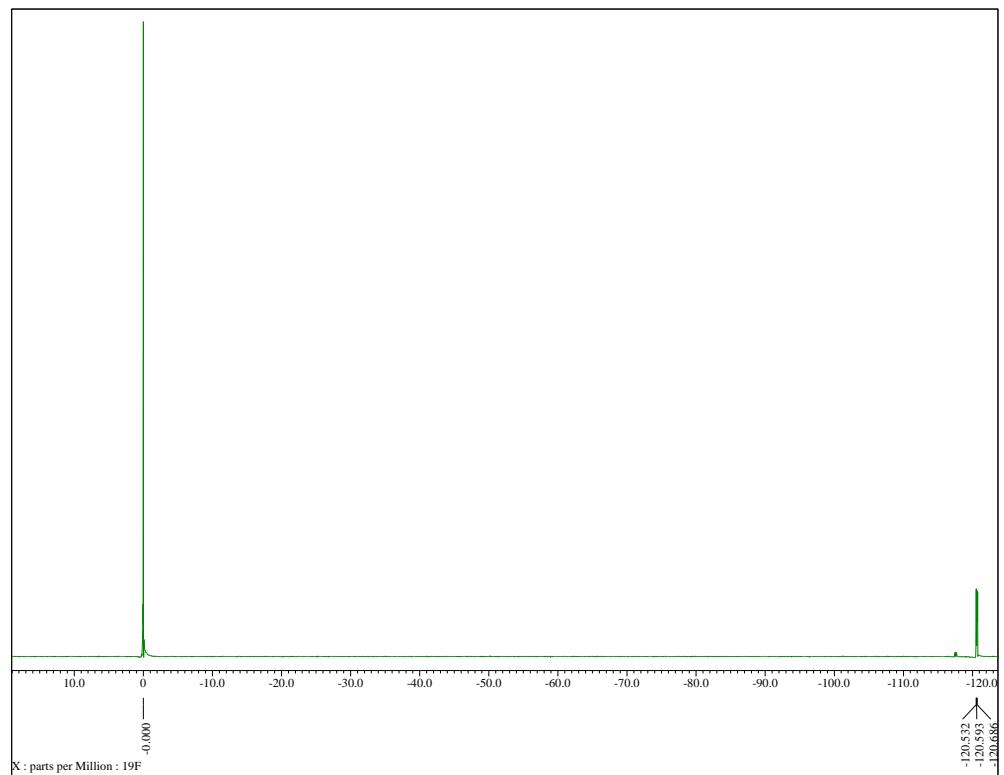
$^1\text{H}$  NMR spectrum of compound **20**



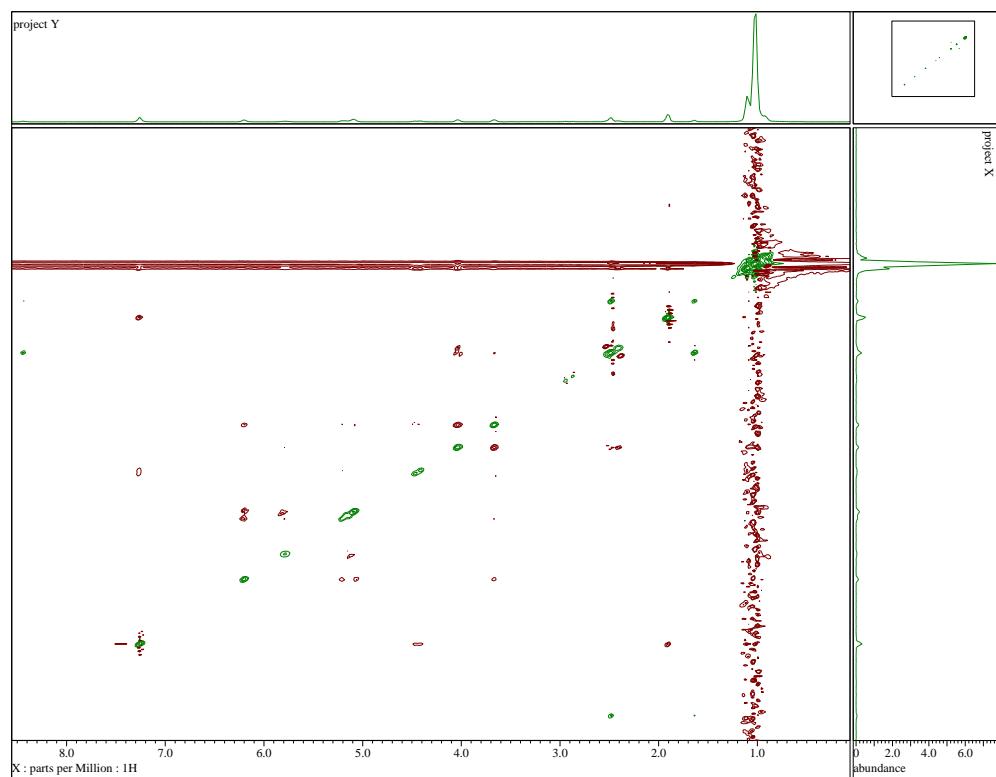
$^{13}\text{C}$  NMR spectrum of compound **20**



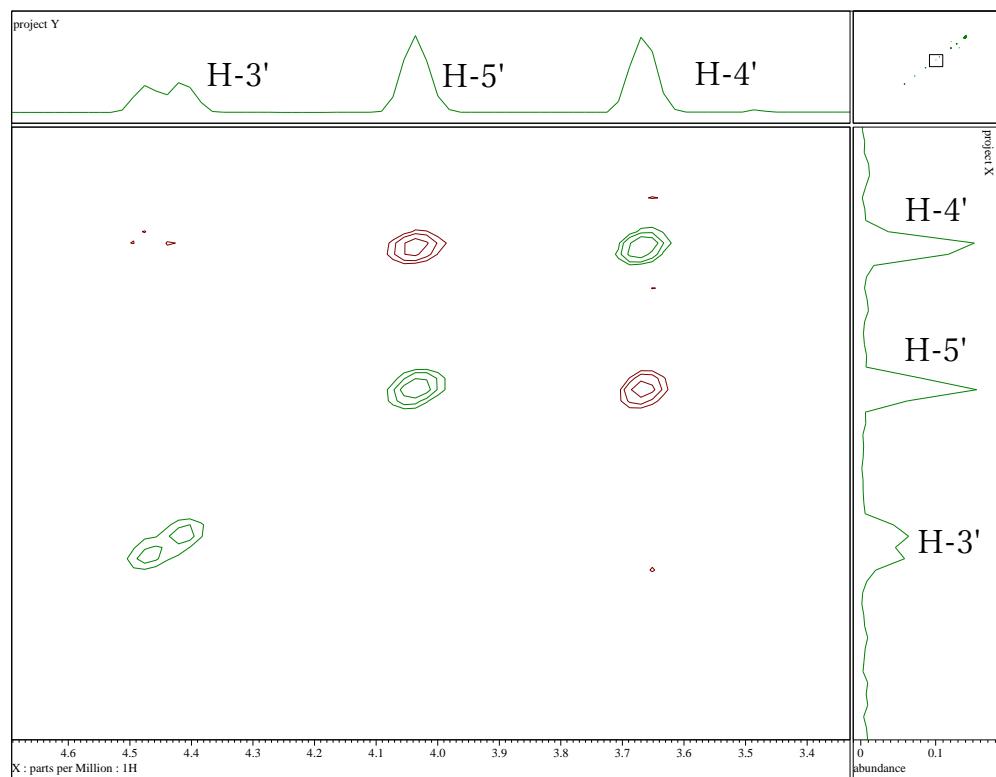
$^{19}\text{F}$  NMR spectrum of compound **20**



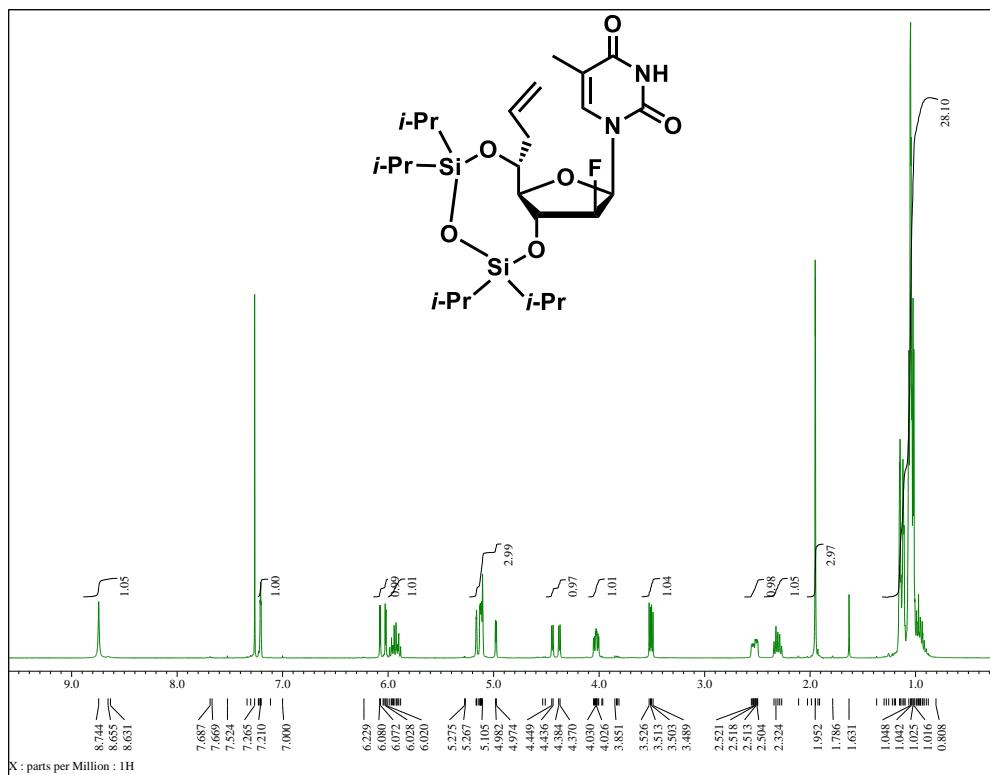
<sup>1</sup>H NOESY spectrum of compound 20



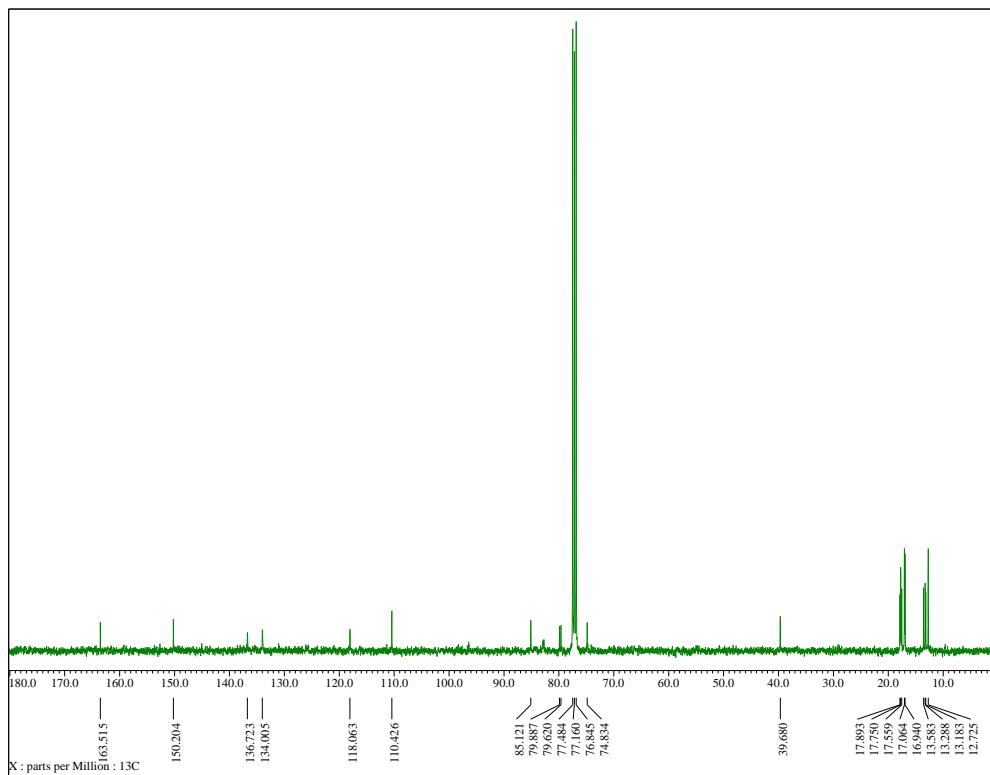
(Enlargement of H-3', H-4' and H-5')



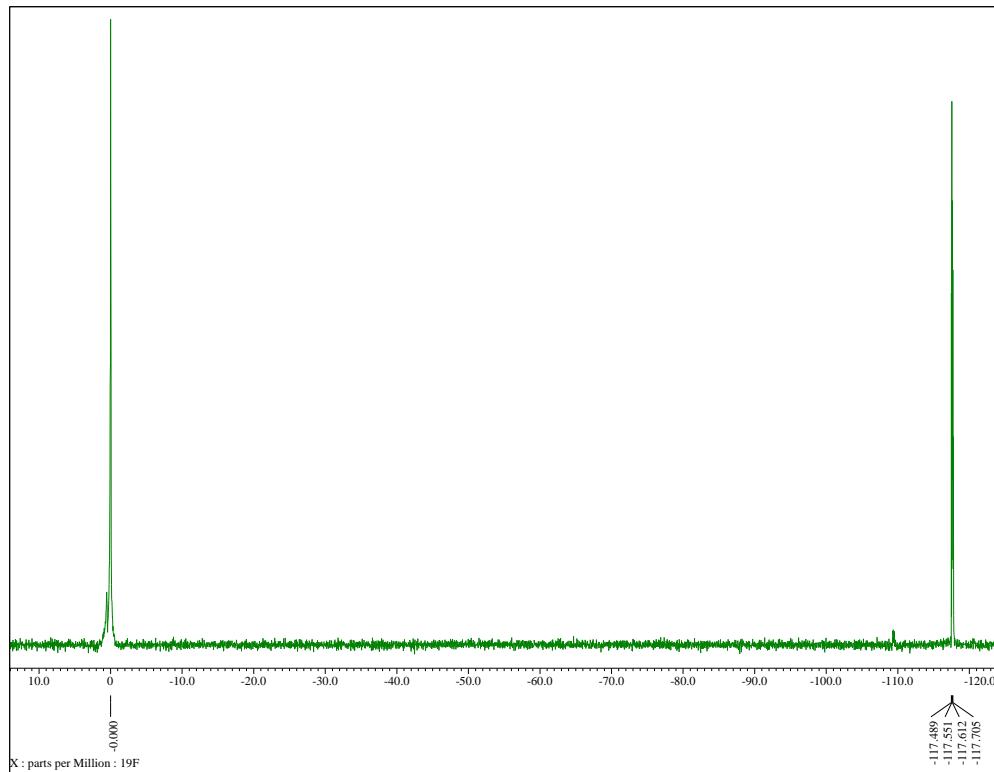
<sup>1</sup>H NMR spectrum of compound 21



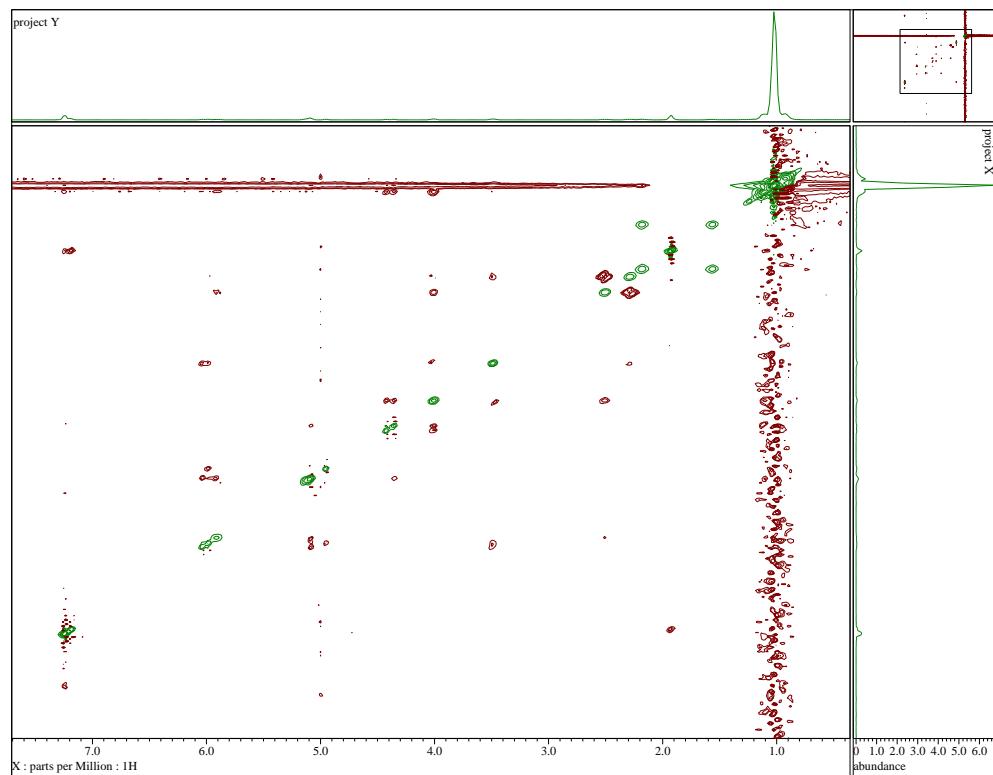
<sup>13</sup>C NMR spectrum of compound 21



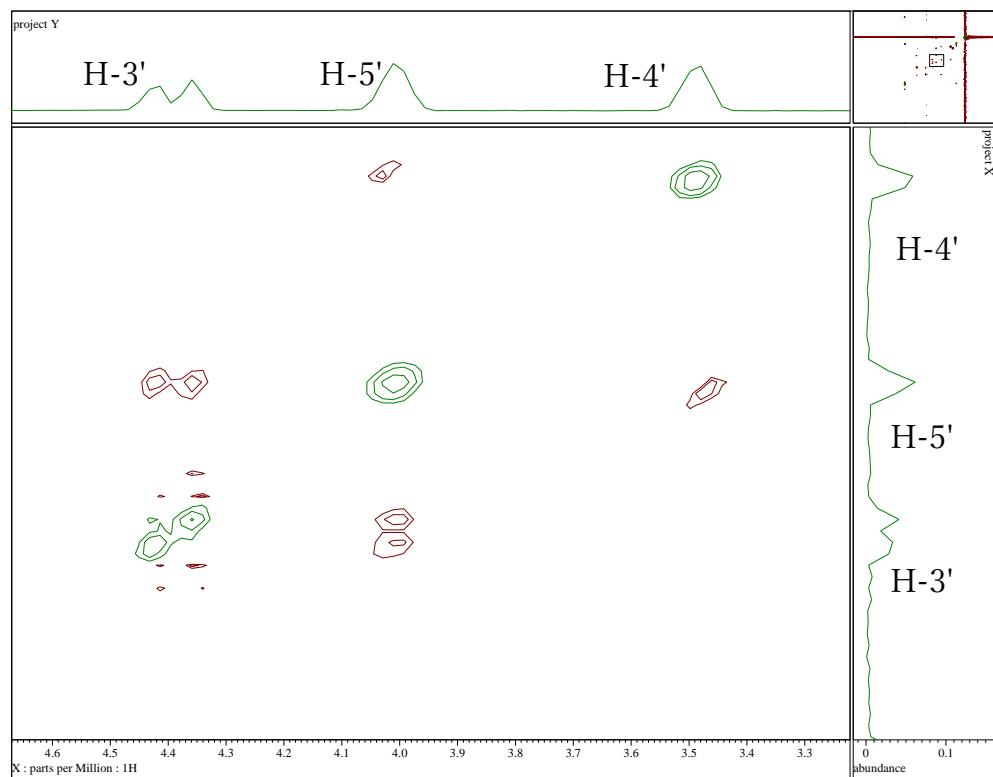
$^{19}\text{F}$  NMR spectrum of compound **21**



<sup>1</sup>H NOESY spectrum of compound 24

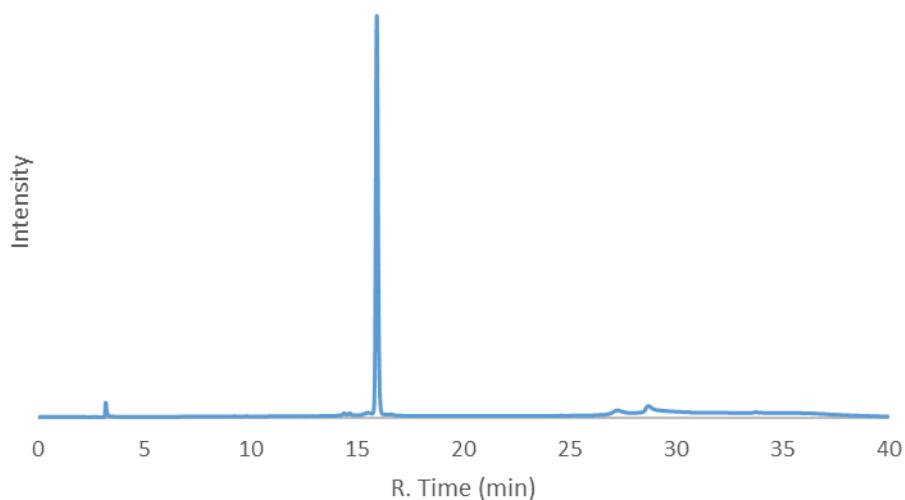


(Enlargement of H-3', H-4' and H-5')

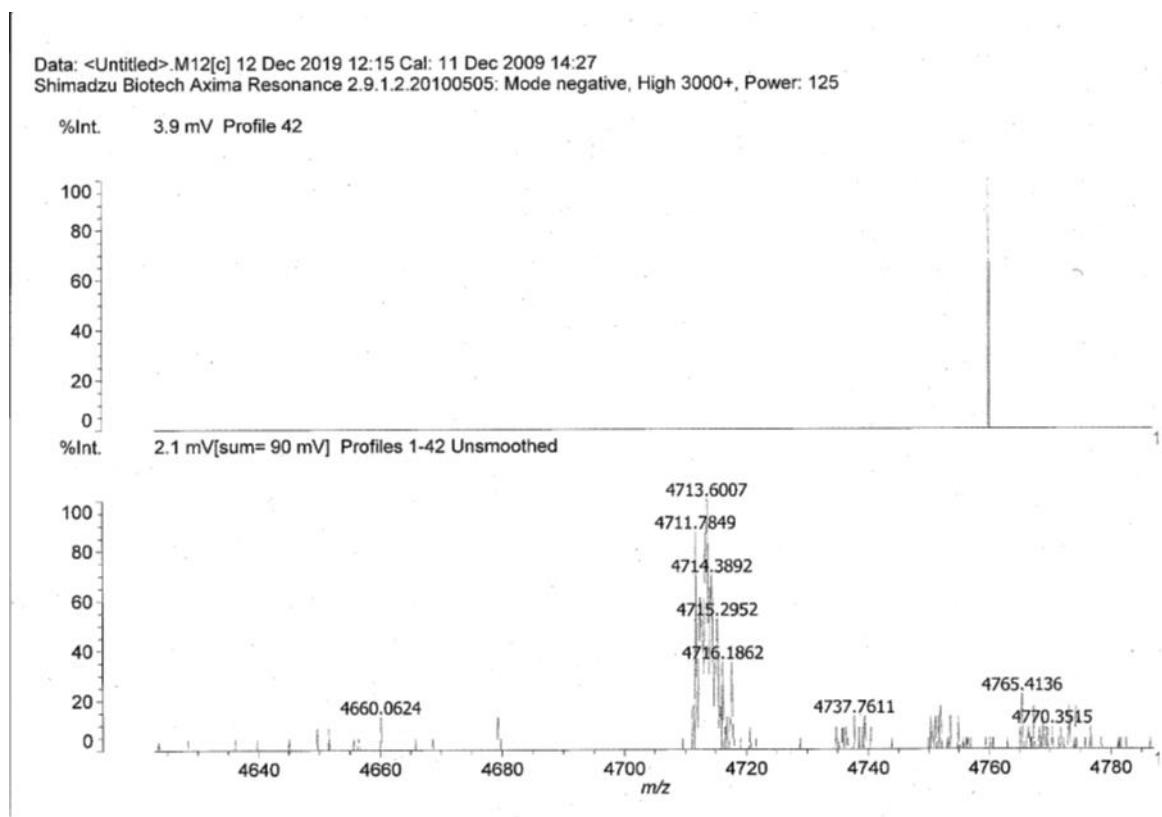


## Reverse-phase HPLC spectra and MALDI-TOF/MS spectra

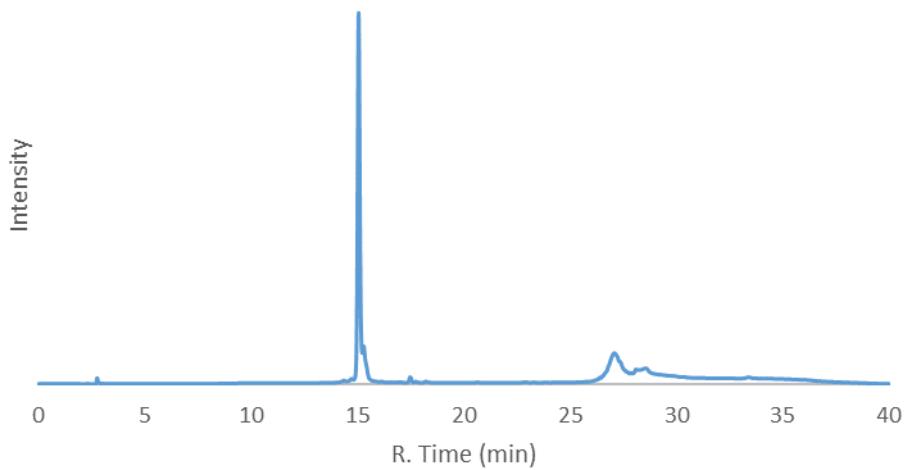
### Reverse-phase HPLC spectra of DNA 1



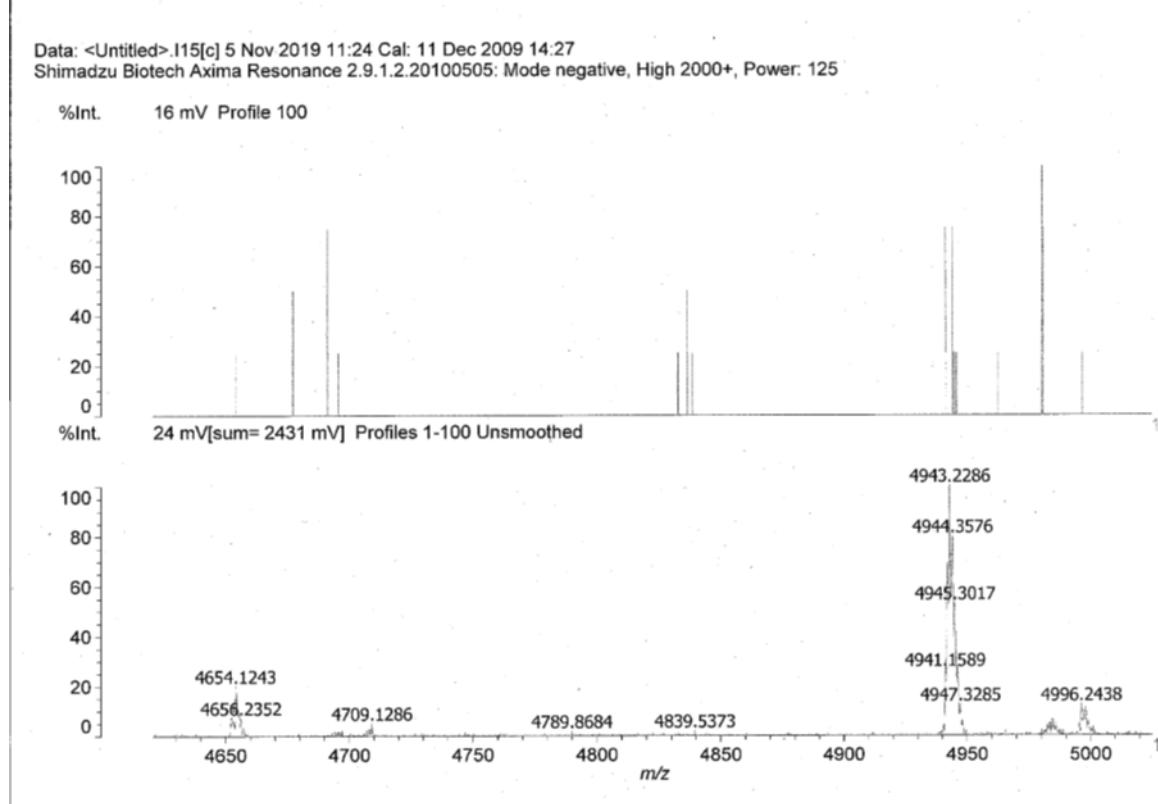
### MALDI-TOF/MS spectra of DNA 1



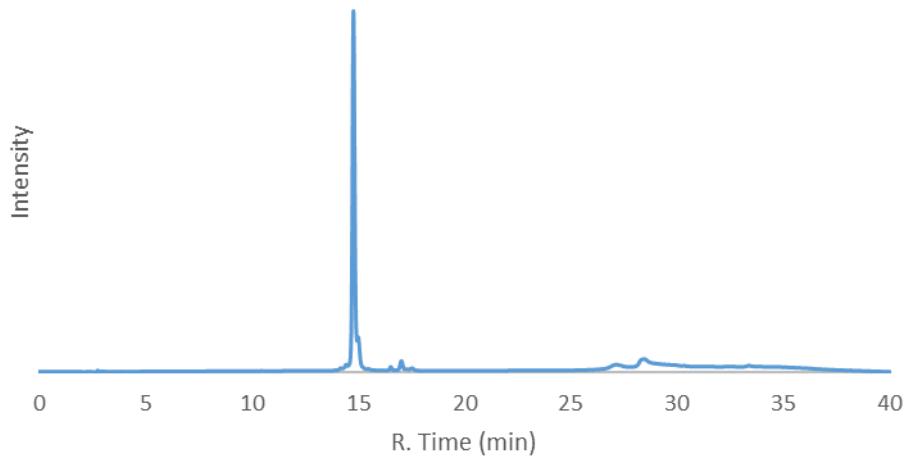
### Reverse-phase HPLC spectra of DNA 2



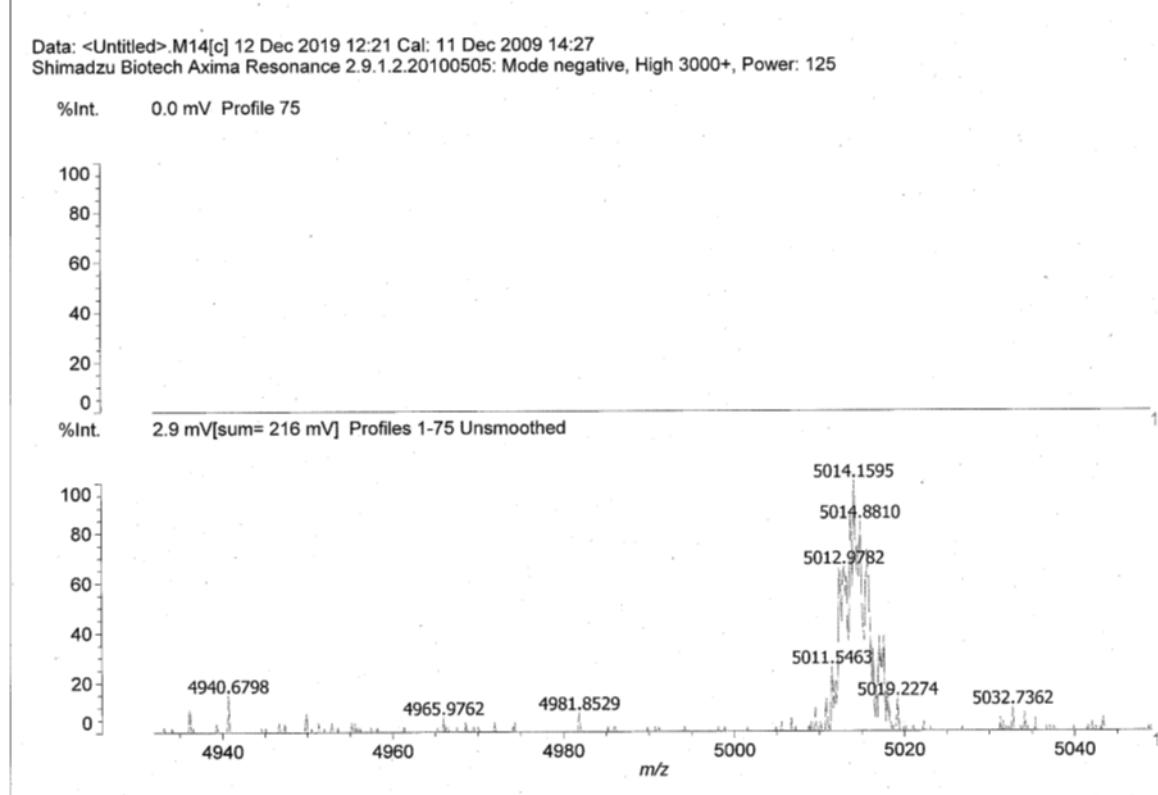
### MALDI-TOF/MS spectra of DNA 2



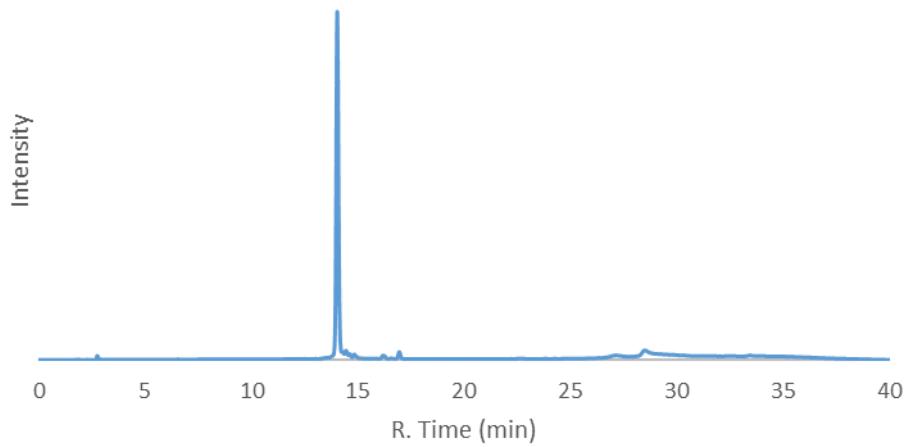
### Reverse-phase HPLC spectra of DNA 3



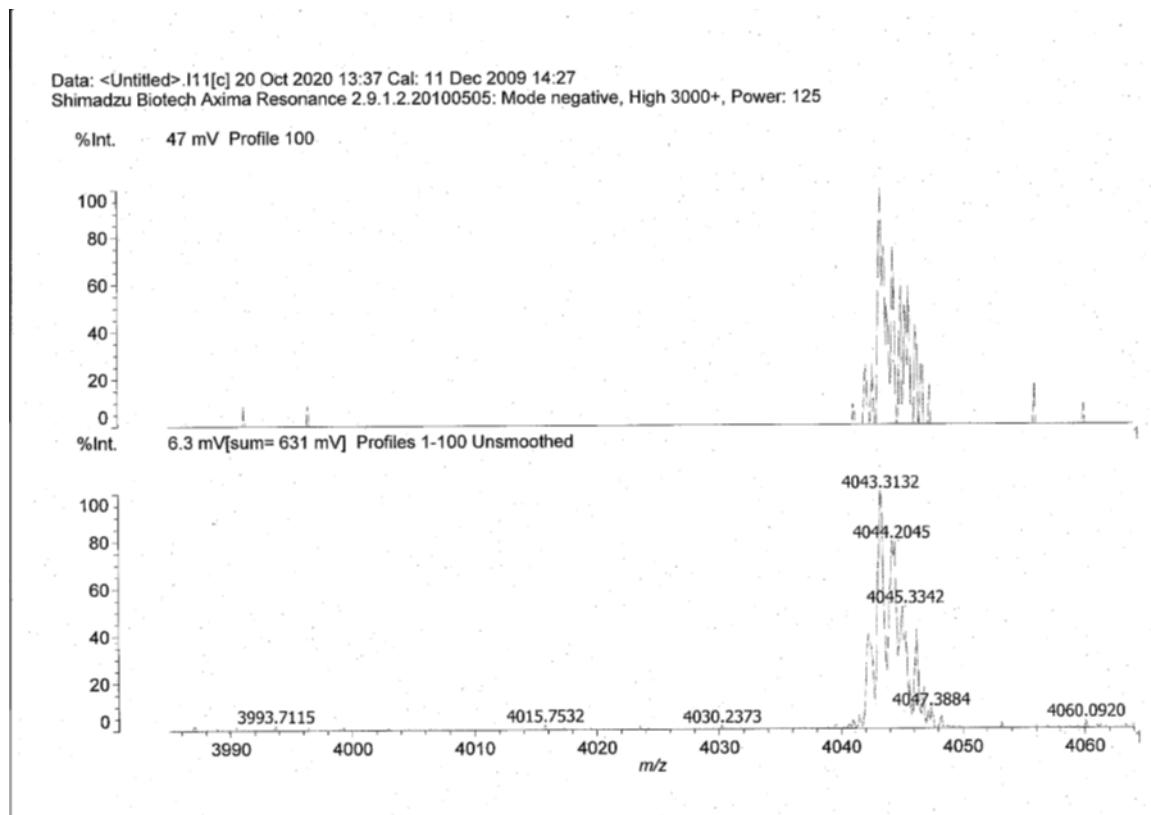
### MALDI-TOF/MS spectra of DNA 3



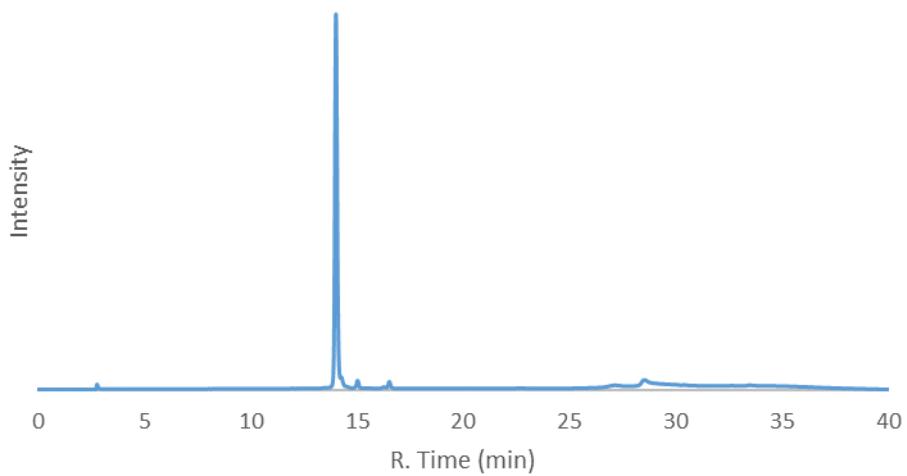
### Reverse-phase HPLC spectra of DNA 5



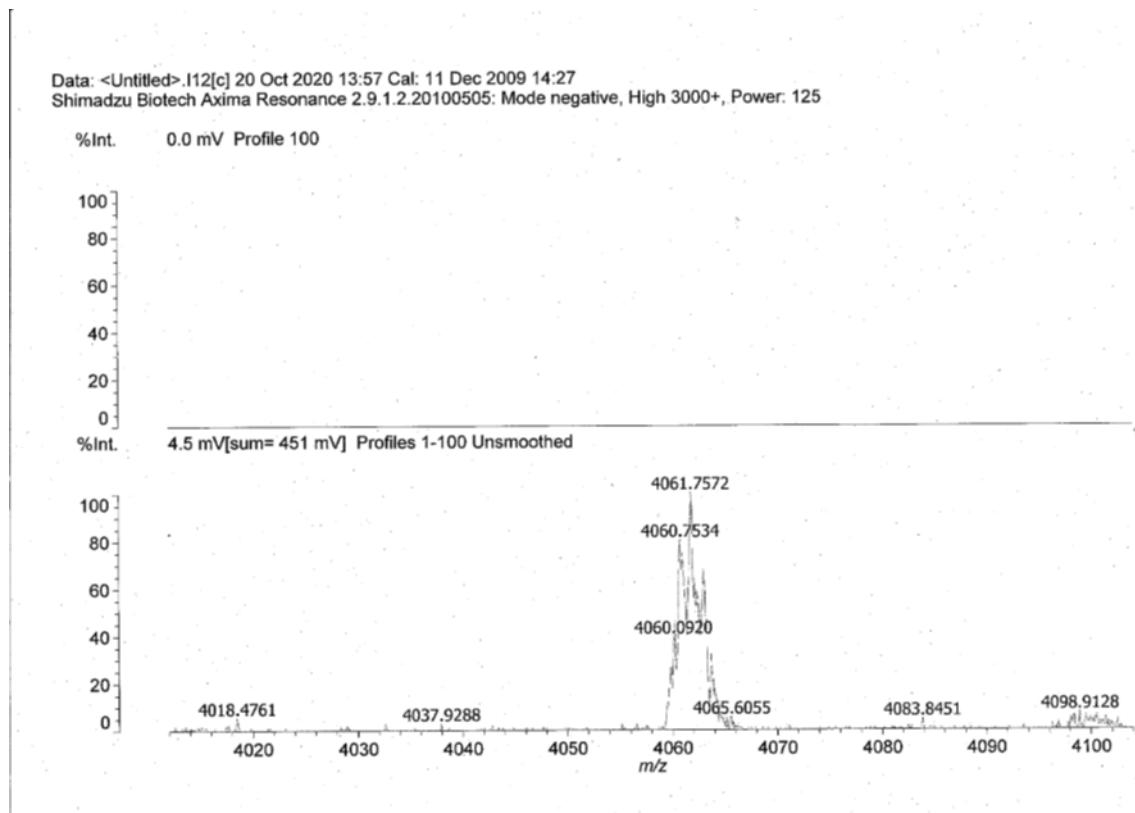
### MALDI-TOF/MS spectra of DNA 5



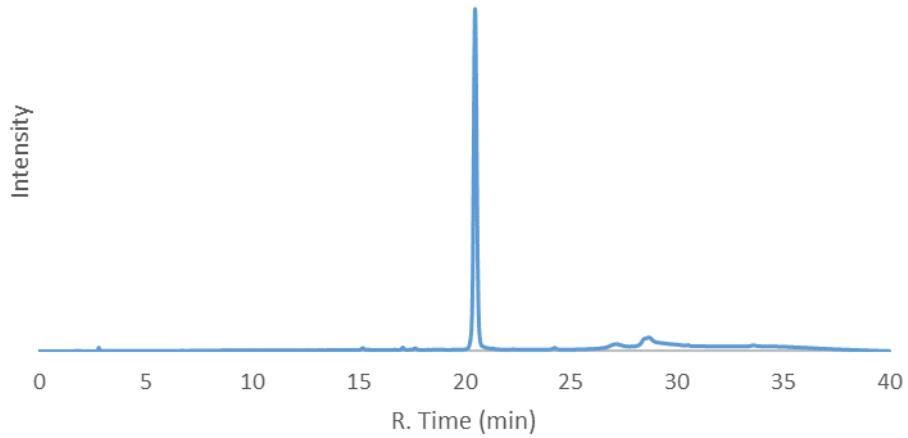
### Reverse-phase HPLC spectra of DNA 6



### MALDI-TOF/MS spectra of DNA 6

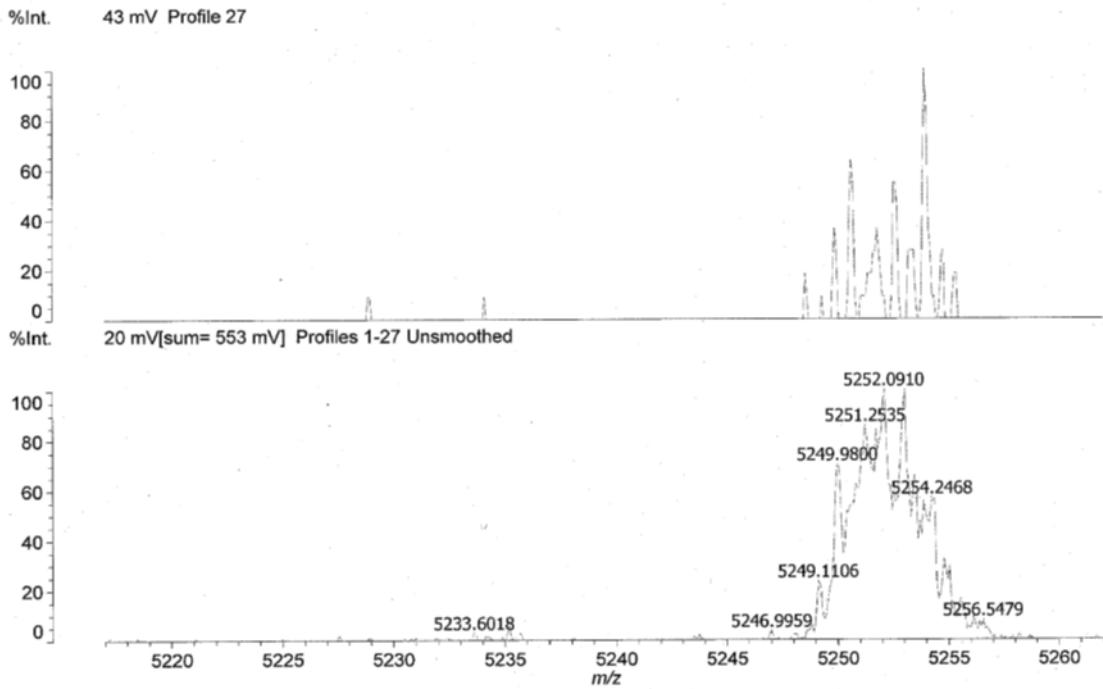


### Reverse-phase HPLC spectra of DNA 7

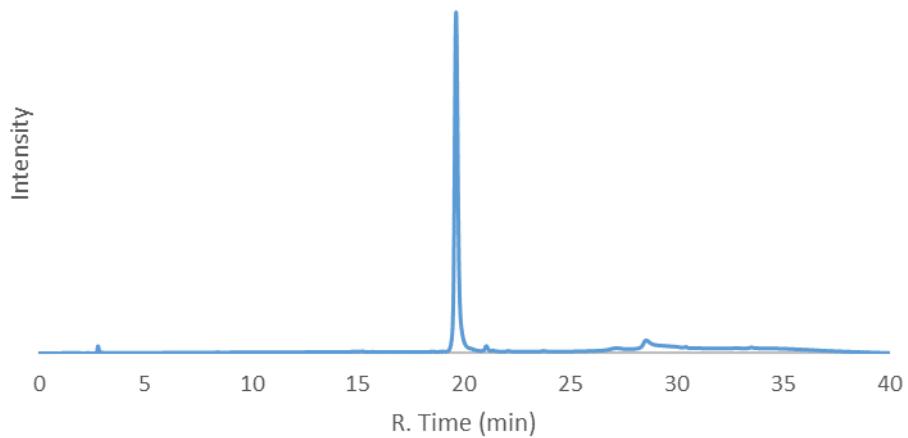


### MALDI-TOF/MS spectra of DNA 7

Data: <Untitled>.M15[c] 12 Dec 2019 12:22 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125

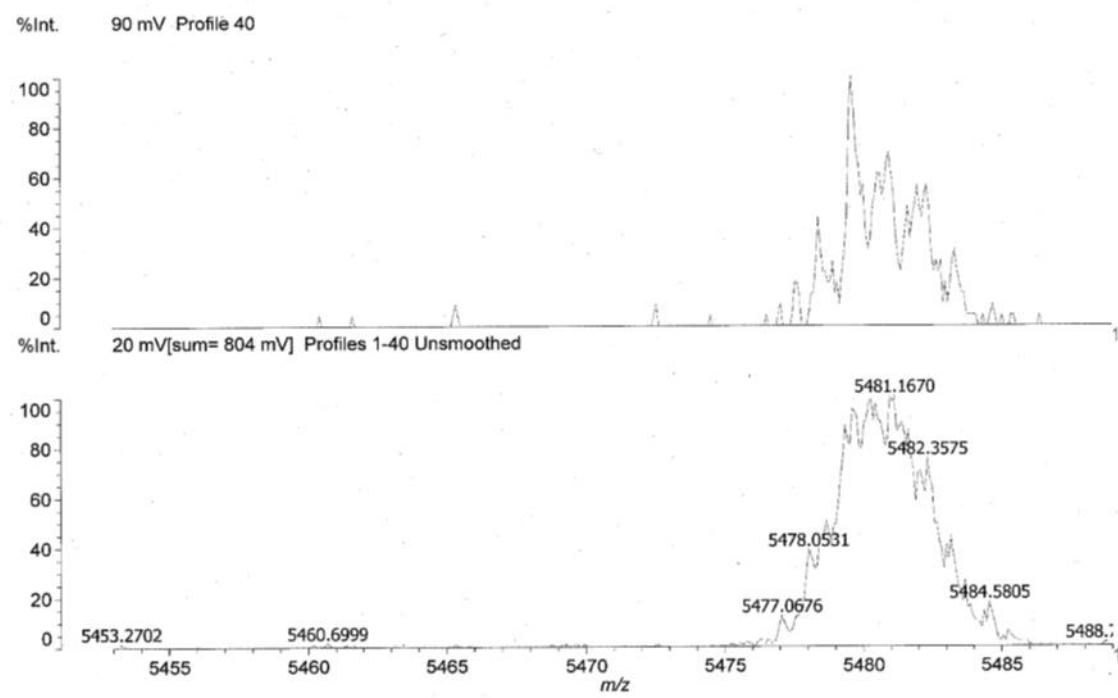


### Reverse-phase HPLC spectra of DNA 8

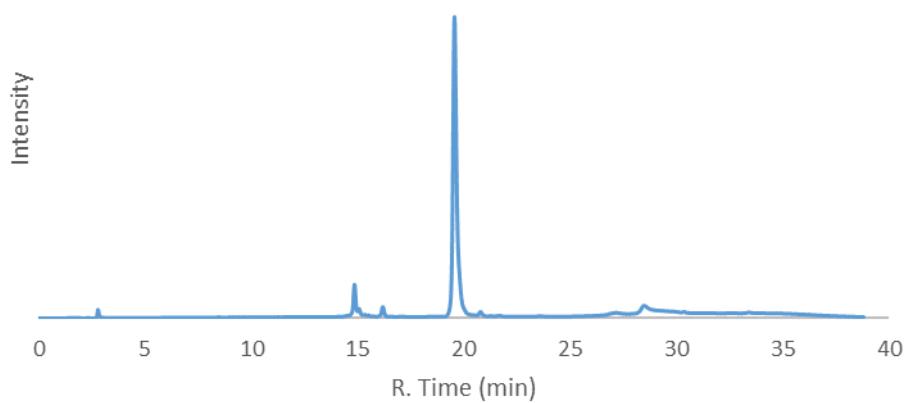


### MALDI-TOF/MS spectra of DNA 8

Data: <Untitled>.M16[c] 12 Dec 2019 12:23 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125



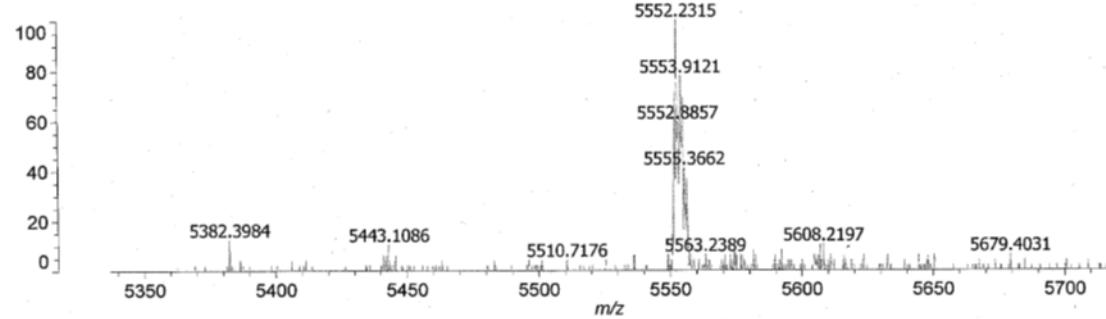
### Reverse-phase HPLC spectra of DNA 9



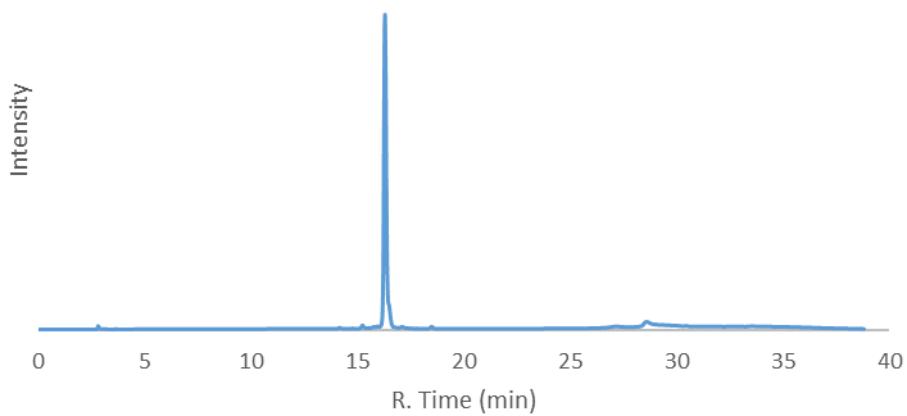
### MALDI-TOF/MS spectra of DNA 9

Data: <Untitled>.A21[c] 7 Jan 2020 10:19 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125

%Int. 3.9 mV Profile 57

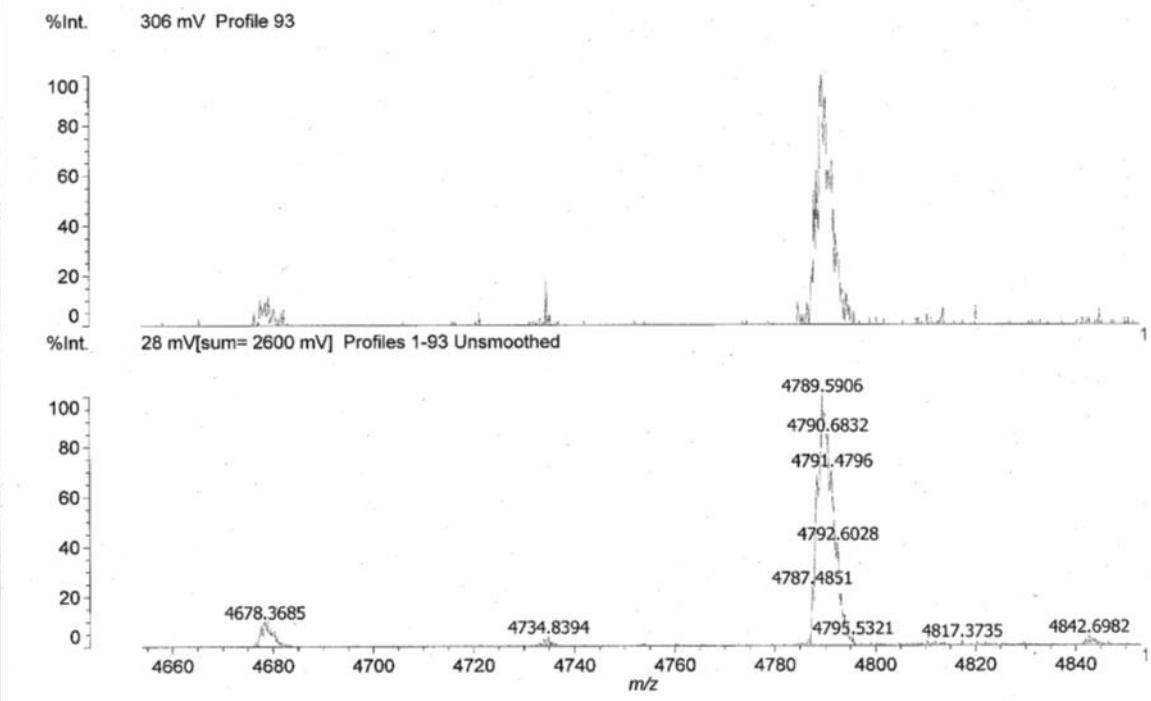


### Reverse-phase HPLC spectra of DNA **10**

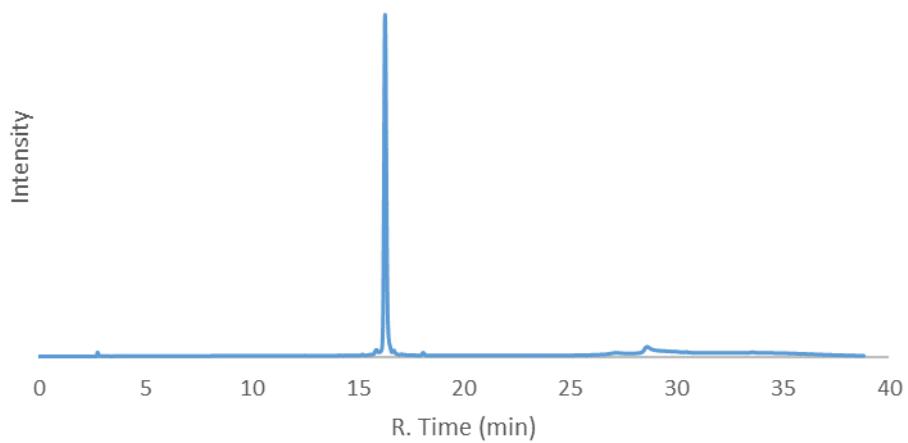


### MALDI-TOF/MS spectra of DNA **10**

Data: <Untitled>.C3[c] 7 Jan 2020 10:27 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125

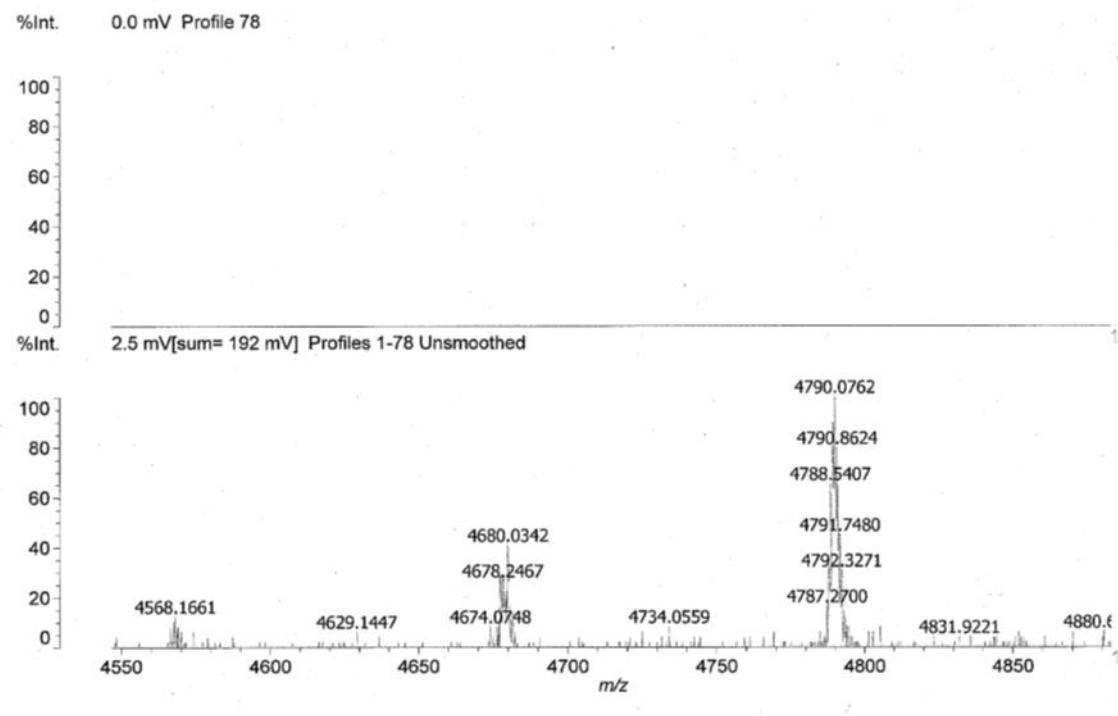


### Reverse-phase HPLC spectra of DNA 11

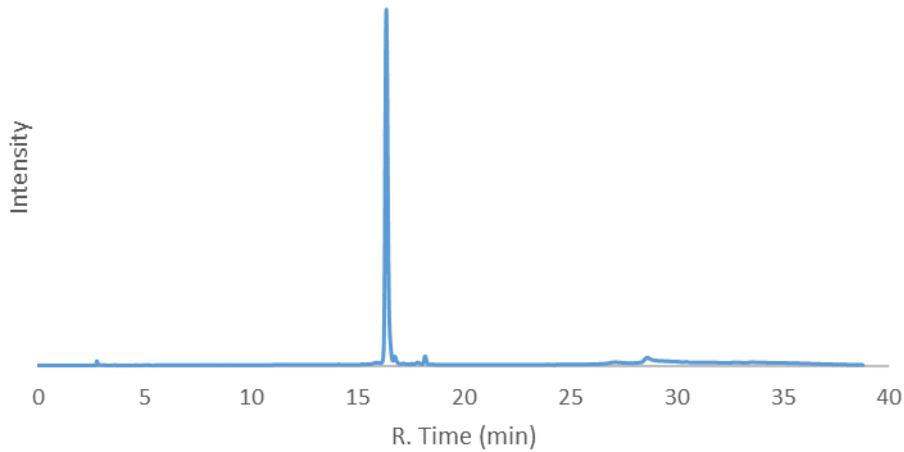


### MALDI-TOF/MS spectra of DNA 11

Data: <Untitled>.C4[c] 7 Jan 2020 10:28 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125

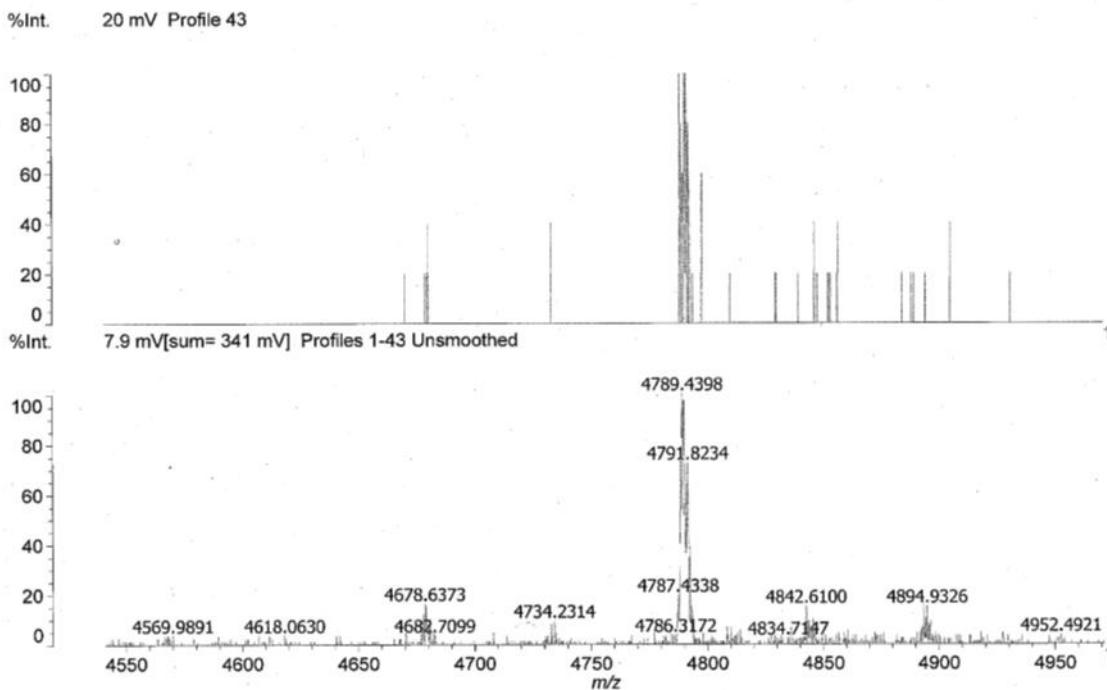


## Reverse-phase HPLC spectra of DNA 12

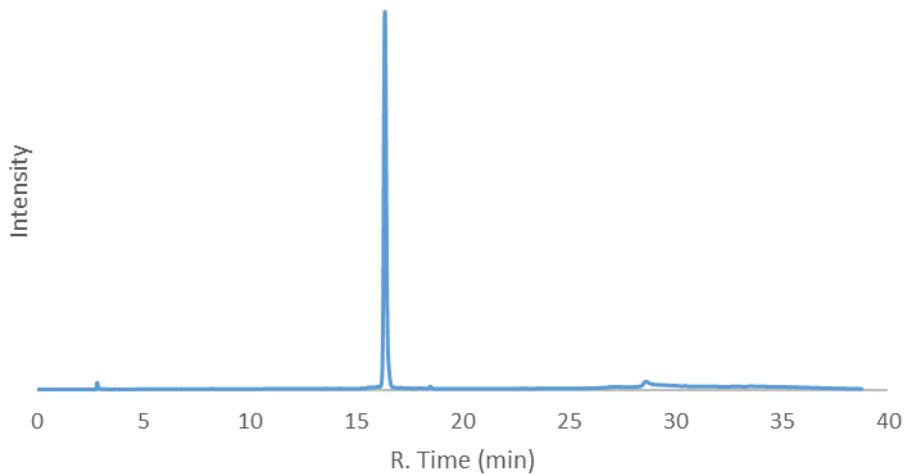


## MALDI-TOF/MS spectra of DNA 12

Data: <Untitled>.D5[c] 7 Jan 2020 10:30 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125



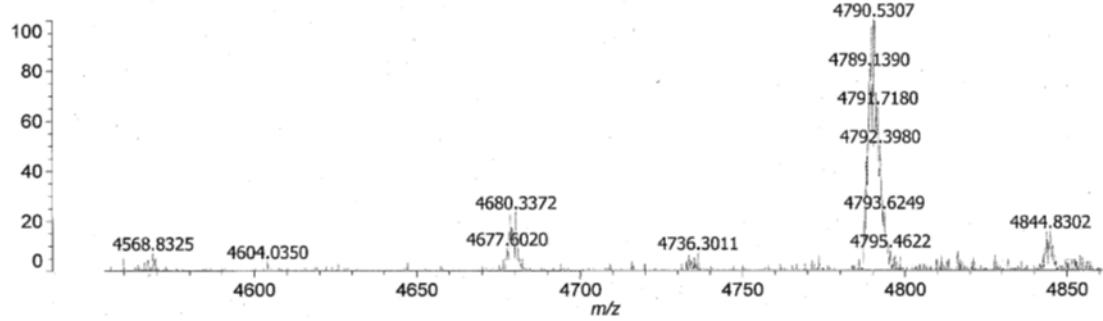
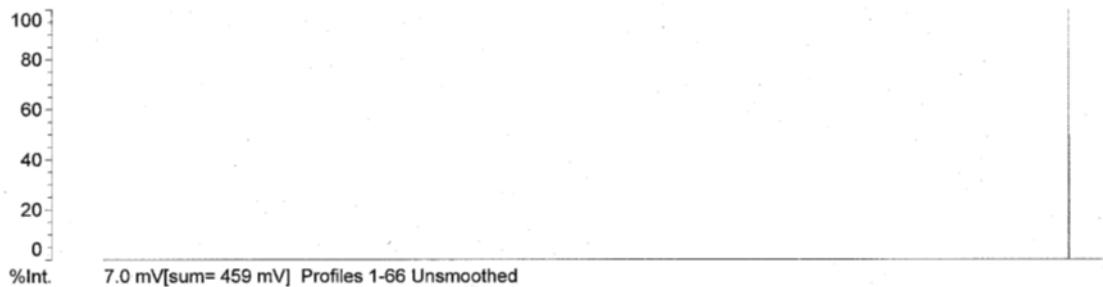
### Reverse-phase HPLC spectra of DNA 13



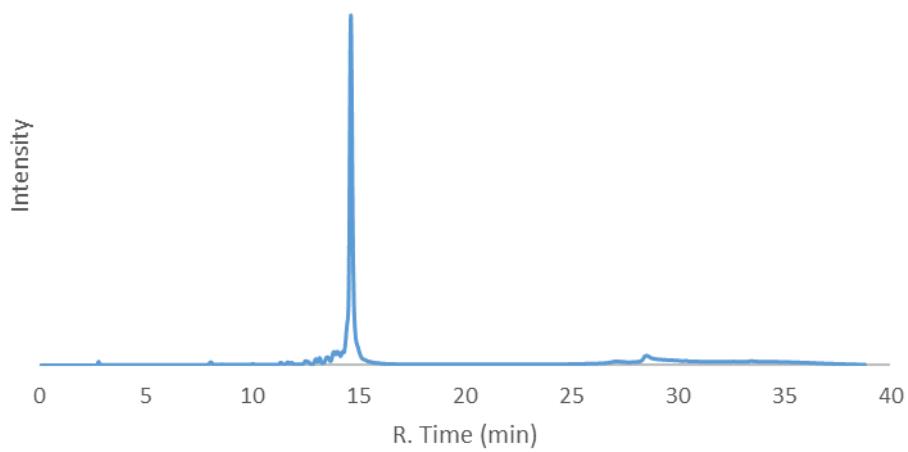
### MALDI-TOF/MS spectra of DNA 13

Data: <Untitled>.C6[c] 7 Jan 2020 10:31 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125

%Int. 3.9 mV Profile 66



## Reverse-phase HPLC spectra of RNA 1



## MALDI-TOF/MS spectra of RNA 1

Data: <Untitled>.O13[c] 12 Dec 2019 12:20 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 3000+, Power: 125

%Int. 0.0 mV Profile 96

100  
80  
60  
40  
20  
0  
%Int.

0.7 mV[sum= 67 mV] Profiles 1-96 Unsmoothed

100  
80  
60  
40  
20  
0  
%Int.

5300.3951

5301.6995

5297.4345

5255.5913

5268.0883

5286.0344

5303.9274

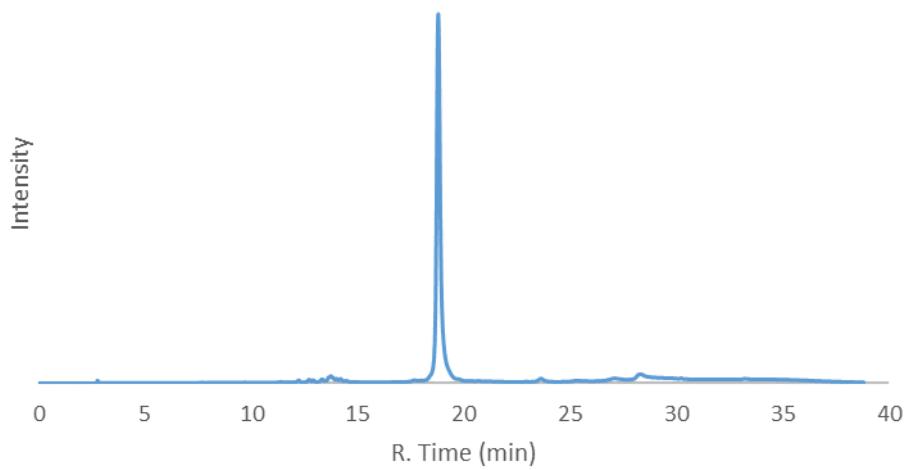
5316.4398

5335.6132

5250 5260 5270 5280 5290 5300 5310 5320 5330 5340

m/z

### Reverse-phase HPLC spectra of RNA 6



### MALDI-TOF/MS spectra of RNA 6

Data: <Untitled>.I22[c] 18 Nov 2019 11:44 Cal: 11 Dec 2009 14:27  
Shimadzu Biotech Axima Resonance 2.9.1.2.20100505: Mode negative, High 2000+, Power: 125

%Int. 0.0 mV Profile 72

