

Electronic Supporting Information

Efficient access to 3'-*O*-phosphoramidite derivatives of tRNA related *N*⁶-threonylcarbamoyladenine (**t⁶A**) and 2-methylthio-*N*⁶-threonylcarbamoyladenine (**ms²t⁶A**)

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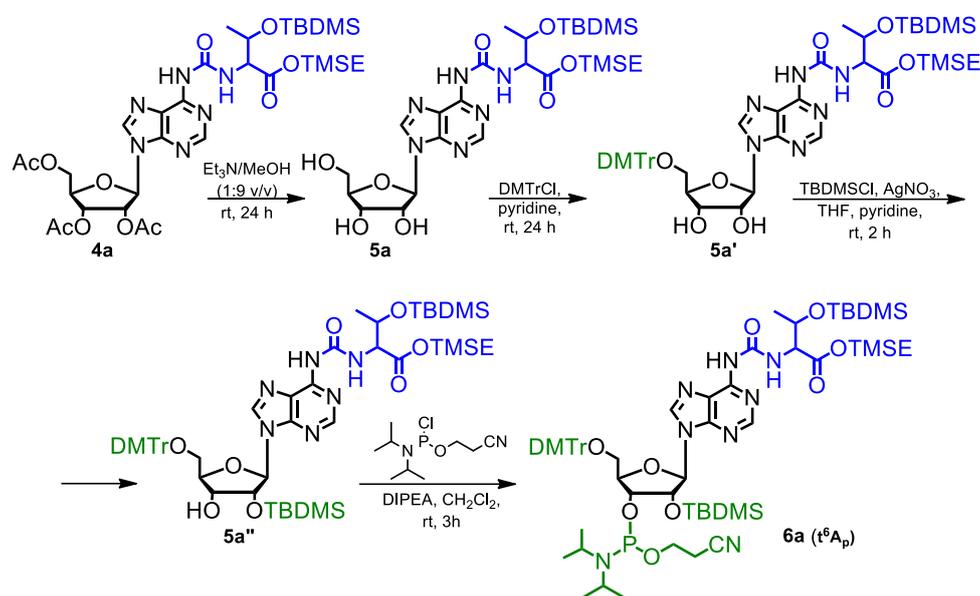
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I. General remarks

Commercial reagents and analytical grade solvents were used without additional purification unless otherwise stated. Analytical thin layer chromatography (TLC) was done on silica gel coated plates (60 F254, Supelco) with UV light (254 nm) or the ninhydrin test (for amino acids) detection. The products were purified by column chromatography on a silica gel 60 (mesh 230 – 400, Fluka) eluted with the indicated solvent mixtures. NMR spectra were recorded using a 700 MHz (for ^1H) instrument, 176 MHz for ^{13}C and 101 MHz for ^{31}P . Chemical shifts (δ) are reported in ppm relative to residual solvent signals CDCl_3 : 7.26 ppm for ^1H NMR, 77.16 ppm for ^{13}C NMR; DMSO-d_6 : 2.50 ppm for ^1H NMR, 39.52 ppm for ^{13}C NMR. The signal multiplicities are described as s (singlet), d (doublet), dd (doublet of doublets), ddd (doublet of doublets of doublets), dq (doublet of quartets), t (triplet), td (triplet of doublets), q (quartet), qd (quartet of doublets), m (multiplet), and br s (broad singlet). High-resolution mass spectra were recorded on Synapt G2Si mass spectrometer (Waters) equipped with an ESI source and quadrupole-Time-of-flight mass analyser. IR data were recorded on an FT-IR ALPHA instrument (Bruker) equipped with a Platinum ATR QuickSnap™ module. Analytical HPLC of nucleosides were performed on a Shimadzu Prominence HPLC system equipped with an SPD-M20A spectral photodiode array detector using a Kinetex® column (RP, C18, 5 μm , 4.6 \times 250 mm, 100 Å, Phenomenex). Analyses were run at 30 °C and the elution profiles were UV monitored at $\lambda = 254$ nm.

II. Experimental details for synthesis 6a, 6b.

2.1 Synthesis of t⁶A-phosphoramidite 6a from 4a.



N-[[9-(β-D-Ribofuranosyl)-purin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**5a**)

Ref: K. Debiec, M. Matuszewski, K. Podkoczyj, G. Leszczyńska, E. Sochacka, *Chem. Eur. J.*, **2019**, *25*, 13309; F. Himmelsbach, B. S. Schulz, T. Trichtinger, R. Charubala, W. Pfeleiderer, *Tetrahedron*, **1984**, *40*, 59.

To protected adenosine **4a** (0.3 g, 0.40 mmol) the solution of distilled solvents: Et₃N/MeOH (4.5 mL, 1:9 v/v) was added. The reaction mixture was stirred for 24 h at room temperature and the solvents were removed under reduced pressure. The oily residue was co-evaporated with toluene (2 x 5 mL). The crude product was purified by column chromatography (silica gel, 0-5 % MeOH in CHCl₃) to obtain pure product **5a** in 90 % yield (0.23 g, 0.36 mmol). **TLC:** R_f = 0.21 (CHCl₃/MeOH, 95:5 v/v);

¹H NMR (700 MHz, DMSO-*d*₆) δ: 9.91-9.89 (m, 2H, NH-6, NH Thr), 8.69 (s, 1H, H-8), 8.41 (s, 1H, H-2), 6.00 (d, 1H, ³J=5.6 Hz, H-1'), 5.52 (br s, 1H, 2'OH), 5.24 (s, 1H, 3'OH), 5.11 (s, 1H, 5'OH), 4.59 (t, 1H, ³J=5.2 Hz, H-2'), 4.48 (qd, 1H, ³J=6.3 Hz, ³J=1.9 Hz, CH-β Thr), 4.42 (dd, 1H, ³J=9.0 Hz, ³J=1.9 Hz, CH-α Thr), 4.21-4.16 (m, 2H, H-3', O-CH TMSE), 4.15-4.07 (m, 1H, O-CH TMSE), 3.98 (q, 1H, ³J=3.9 Hz, H-4'), 3.70 (dd, 1H, ²J=12.0 Hz, ³J=4.0 Hz, H-5'), 3.58 (dd, 1H, ²J=12.0 Hz, ³J=4.0 Hz, H-5''), 1.19 (d, 3H, ³J= 6.3 Hz, CH₃ Thr), 0.99-0.93 (m, 2H, Si-CH₂ TMSE), 0.89 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.08 (s, 3H, ³J= 6.3 Hz, CH₃ Thr), 0.01 (s, 3H, Si-CH₃ TBDMS), -0.00 (s, 9H, Si(CH₃)₃ TMSE).

N-[[9-(5'-*O*-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl)-purin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**5a'**)

Ref: M. Sundaram, P. F. Crain, D. R. Davis, *J. Org. Chem.*, **2000**, *65*, 5609;

To a stirred solution of nucleoside **5a** (0.2 g, 0.32 mmol) in dry pyridine (3.0 mL) DMTrCl (0.13 g, 0.38 mmol) was added. The reaction was stirred for 20 h at room temperature. After this time the reaction mixture was cooled to 0°C in an ice bath and quenched with H₂O (4 mL) and stirred at 0°C for 15 min. The mixture was extracted with CHCl₃ (2 x 15 mL) and combined organic layers were dried over anhydrous MgSO₄, filtered and the solvent was evaporated. The oily residue was co-evaporated with toluene (2 x 8 mL). The crude product was purified by column chromatography (silica gel, 0-1 % MeOH in CHCl₃) to obtain **5a'** as a white solid in 90 % yield (0.27 g, 0.29 mmol).

TLC: R_f = 0.43 (CHCl₃/MeOH, 95:5 v/v).

¹H NMR (700 MHz, DMSO-*d*₆) δ : 9.93 (d, ³J=8.9 Hz, 1H, NH Thr), 9.90 (s, 1H, NH-6), 8.58 (s, 1H, H-8), 8.34 (s, 1H, H-2), 7.36-7.33 (m, 2H, H_{Ar} DMTr), 7.24-7.19 (m, 6H, H_{Ar} DMTr), 7.19-7.15 (m, 1H, H_{Ar} DMTr), 6.81-6.78 (m, 4H, H_{Ar} DMTr), 6.03 (d, 1H, ³J=4.5 Hz, H-1'), 5.61 (br s, 1H, 2'OH), 5.28 (br s, 1H, 3'OH), 4.81 (t, 1H, ³J= 4.8 Hz, H-2'), 4.50 (qd, 1H, ³J=6.2 Hz, ³J=1.9 Hz, CH- β Thr), 4.42 (dd, 1H, ³J=9.1 Hz, ³J=1.8 Hz, CH- α Thr), 4.36 (t, 1H, ³J=5.1 Hz, H-3'), 4.19 (ddd, 1H, ²J=11.0 Hz, ³J=10.0 Hz, ³J=6.8 Hz, O-CH TMSE), 4.15-4.08 (m, 2H, H-4', O-CH TMSE), 3.71 (s, 3H, O-CH₃ DMTr), 3.71 (s, 3H, O-CH₃ DMTr), 3.28-3.22 (m, 2H, H-5', H-5''), 1.21 (d, 3H, ³J=6.3 Hz, CH₃ Thr), 0.99-0.93 (m, 2H, Si-CH₂ TMSE), 0.89 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.08 (s, 3H, Si-CH₃ TBDMS), 0.01 (s, 3H, Si-CH₃ TBDMS), -0.01 (s, 9H, Si(CH₃)₃ TMSE).

N-[[9-(2'-*O*-*tert*-butyldimethylsilyl-5'-*O*-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl)-purin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**5a''**)

Ref: K. Onizuka, M.E. Hazemi, J.M. Thomas, L.R. Monteleone, K. Yamada, S. Imoto, P.A. Beal, F. Nagatsugi, *Bioorg Med Chem.*, **2017**, *25*, 2191; V. Boudou, J. Langridge, A. V. Aerschot, C. Hendrix, A. Millar, P. Weiss, P. Herdewijn, *Helv. Chim. Acta*, **2000**, *83*, 152;

5'-DMTr nucleoside **5a'** (0.2 mg, 0.22 mmol) was dissolved in freshly distilled THF (1.0 mL) then anhydrous pyridine (0.09 mL, 1.1 mmol) and AgNO₃ (56 mg, 0.33 mmol) was added. The reaction mixture was stirred at room temperature for 30 min in darkness. Then TBDMS-Cl (0.06 g, 0.4 mmol) was added and stirring was continued at room temperature for 2 h. After this time, the reaction mixture was diluted with CH₂Cl₂, filtered through Celite and washed with CH₂Cl₂. The solution was extracted with saturated NaHCO₃ (2 x 10 mL) and brine (10 mL). The organic layer was dried over MgSO₄, filtered and concentrated under reduced pressure. The oily residue was

co-evaporated with toluene (2 x 5 mL) and the crude product was purified by the column chromatography (silica gel, 0-10 % acetone in CHCl₃). The pure 2'-isomer **5a''** was obtained as a white solid in 57 % yield (0.13 g, 0.13 mmol). The 3'-regioisomer (0.06 g, 0.06 mmol) was dissolved in dry methanol with few drops of TEA, to give equimolar mixture of 2' and 3' TBDMS isomers from which 2'-TBDMS nucleoside was chromatographically isolated to give finally 0.16 g of **5a''** (0.15 mmol; 70 % yield). **TLC:** R_f= 0.42 (CHCl₃/MeOH, 98:2 v/v).

¹H NMR (700 MHz, DMSO-*d*₆) δ: 9.93 (br s, 1H, NH Thr), 9.92 (s, 1H, NH-6), 8.60 (s, 1H, H-8), 8.28 (s, 1H, H-2), 7.42-7.38 (m, 2H, H_{Ar} DMTr), 7.29-7.26 (m, 6H, H_{Ar} DMTr), 7.26-7.22 (m, 1H, H_{Ar} DMTr), 7.21-7.17 (m, 4H, H_{Ar} DMTr), 6.84-6.80 (m, 4H, H_{Ar} DMTr), 6.03 (d, 1H, ³J=5.3 Hz, H-1'), 5.17 (d, 1H, ³J=5.7 Hz, 3'OH), 5.01 (t, 1H, ³J=5.2 Hz, H-2'), 4.48 (qd, 1H, ³J=6.3 Hz, ³J=2.0 Hz, CH-β Thr), 4.42 (dd, 1H, ³J=9.0 Hz, ³J=1.9 Hz, CH-α Thr), 4.29-4.24 (m, 1H, H-3'), 4.18 (ddd, 1H, ³J=11.1 Hz, ³J=9.8 Hz, ³J=6.8 Hz, O-CH TMSE), 4.15-4.09 (m, 2H, H-4', O-CH TMSE), 3.71 (s, 6H, 2x O-CH₃ DMTr), 3.34-3.31 (m, 1H, H-5'), 3.26 (dd, 1H, ²J=10.5 Hz, ³J=5.2 Hz, H-5''), 1.19 (d, 3H, ³J=6.2 Hz, CH₃ Thr), 0.96-0.93 (m, 2H, Si-CH₂ TMSE), 0.85 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.73 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.06 (s, 3H, Si-CH₃ TBDMS), -0.00 (s, 3H, Si-CH₃ TBDMS), -0.02 (s, 9H, Si(CH₃)₃ TMSE), -0.05 (s, 3H, Si-CH₃ TBDMS), -0.16 (s, 3H, Si-CH₃ TBDMS).

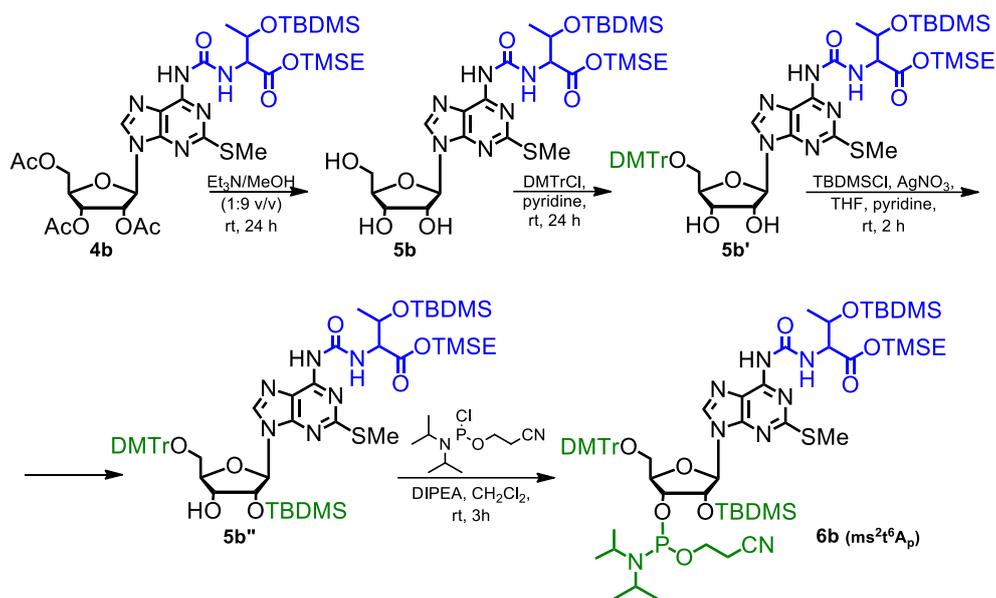
N-[[9-(2'-*O*-*tert*-butyldimethylsilyl)-3'-(2-cyanoethyl-*N,N*-diisopropylphosphoramidite)-5'-*O*-(4,4'-dimethoxytrityl)-β-*D*-ribofuranosyl)-purin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**6a**)

Ref: V. Boudou, J. Langridge, A. V. Aerschot, C. Hendrix, A. Millar, P. Weiss, P. Herdewijn, *Helv. Chim. Acta*, **2000**, *83*, 152;

To a stirred solution of 5'-DMTr, 2'-TBDMS-⁶A **5a''** (0.15 mg, 0.14 mmol) in freshly distilled CH₂Cl₂ (0.8 mL) anhydrous DIPEA was added (0.1 mL, 0.56 mmol) under argon atmosphere. Then 2-cyanoethyl-*N,N*-diisopropylchlorophosphoramidite (0.06 ml; 0.28 mmol) was added dropwise. The reaction was stirred for 3 h under argon atmosphere at room temperature. The reaction mixture was diluted with CH₂Cl₂ and washed with 5 % aq NaHCO₃ (2 x 10 mL) and H₂O (10 mL), then the organic layer was dried over anhydrous MgSO₄, filtered and evaporated under reduced pressure. The crude product was purified by the flash chromatography (silica gel, petroleum ether/acetone, 2:1 v/v) to obtain pure product **6a** as a white solid in 92 % yield (0.16 g; 0.13 mmol). **TLC:** R_f= 0.52 (CHCl₃/acetone, 95:5 v/v).

³¹P NMR: (176,03 MHz MHz, C₆H₆) δ: 149.89, 148.04.

2.2 Synthesis of $ms^{2t6}A$ -phosphoramidite **6b** from **4b**.



N-[[9-(β -D-Ribofuranosyl)-2-methylthiopurin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**5b**)

Ref: K. Debiec, M. Matuszewski, K. Podkoczyj, G. Leszczyńska, E. Sochacka, *Chem. Eur. J.*, **2019**, *25*, 13309; F. Himmelsbach, B. S. Schulz, T. Trichtinger, R. Charubala, W. Pfeleiderer, *Tetrahedron*, **1984**, *40*, 59.

To fully-protected nucleoside **4b** (156 mg, 0.20 mmol) solution of distilled solvents: TEA/MeOH (2.2 mL, 1:9 v/v) was added. The reaction mixture was stirred for 24 h at room temperature and the solvents were removed under reduced pressure. The oily residue was co-evaporated with toluene (2 x 5 mL). The crude product was purified by column chromatography (silica gel, 0-4 % MeOH in $CHCl_3$) to obtain pure product **5b** in 85 % yield (114 mg, 0.17 mmol). **TLC:** R_f = 0.18 ($CHCl_3/MeOH$, 95:5 v/v);

1H NMR (700 MHz, $DMSO-d_6$) δ : 9.89 (s, 1H, NH-6), 9.26 (d, 1H, $^3J=8.6$ Hz, NH Thr), 8.51 (s, 1H, H-8), 5.93 (d, 1H, $^3J=5.7$ Hz, H-1'), 5.48 (d, 1H, $^3J=6.0$ Hz, 2'OH), 5.21 (d, 1H, $^3J=5.0$ Hz, 3'OH), 4.99 (t, 1H, $^3J=5.6$ Hz, 5'OH), 4.63 (q, 1H, $^3J=5.6$ Hz, H-2'), 4.48-4.42 (m, 2H, CH- α Thr, CH- β Thr), 4.24-4.16 (m, 2H, H-3', O-CH TMSE), 4.13-4.10 (m, 1H, O-CH TMSE), 3.94-3.66 (q, 1H, $^3J=4.2$ Hz, H-4'), 3.66 (ddd, 1H, $^3J=11.9$ Hz, $^3J=5.5$ Hz, $^3J=4.3$ Hz, H-5''H), 3.56 (ddd, 1H, $^3J=11.8$ Hz, $^3J=5.7$ Hz, $^3J=4.4$ Hz, H-5''H), 2.56 (s, 3H, S- CH_3), 1.19 (d, 3H, $^3J=6.1$ Hz, CH_3 Thr), 1.04-0.91 (m, 2H, Si- CH_2 TMSE), 0.85 (s, 9H, Si- $C(CH_3)_3$ TBDMS), 0.08 (s, 3H, Si- CH_3 TBDMS), 0.03 (s, 3H, Si- CH_3 TBDMS), 0.01 (s, 9H, Si(CH_3) $_3$ TMSE).

N-[[9-(5'-*O*-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl)-2-methylthiopurin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**5b'**)

Ref: K. Onizuka, M.E. Hazemi, J.M. Thomas, L.R. Monteleone, K. Yamada, S. Imoto, P.A. Beal, F. Nagatsugi, *Bioorg Med Chem.*, **2017**, *25*, 2191; V. Boudou, J. Langridge, A. V. Aerschot, C. Hendrix, A. Millar, P. Weiss, P. Herdewijn, *Helv. Chim. Acta*, **2000**, *83*, 152;

To a stirred solution of free ribose nucleoside **5b** (105 mg, 0.16 mmol) in dry pyridine (1.2 mL) DMTrCl (80 mg, 0.24 mmol) was added. The reaction was stirred for 15 h at room temperature. After this time the mixture was cooled to 0°C in an ice bath and quenched with H₂O (2 mL) and stirred at 0°C for 15 min. The mixture was extracted with CH₂Cl₂ (2x 10 mL) and combined organic layers were dried over anhydrous MgSO₄, filtered and the solvent was evaporated. The oily residue was co-evaporated with toluene (2 x 5 mL). The crude product was purified by column chromatography (silica gel, 0-2 % MeOH in CHCl₃) to obtain **5b'** as a white solid in 91 % yield (143 mg, 0.14 mmol). **TLC:** R_f = 0.43 (CHCl₃/MeOH, 95:5 v/v).

¹H NMR (700 MHz, DMSO-*d*₆) δ : 9.91 (s, 1H, NH-6), 9.29 (d, 1H, ³J=8.4 Hz, NH Thr), 8.42 (s, 1H, H-8), 7.35-7.27 (m, 2H, H_{Ar} DMTr), 7.23-7.18 (m, 6H, H_{Ar} DMTr), 7.18-7.15 (m, 1H, H_{Ar} DMTr), 6.81-6.77 (m, 4H, H_{Ar} DMTr), 5.98 (d, 1H, ³J=4.2 Hz, H-1'), 5.58 (d, 1H, ³J=5.4 Hz, 2'OH), 5.24 (d, 1H, ³J=5.9 Hz, 3'OH), 4.77 (q, ³J=5.3 Hz, H-2'), 4.49-4.44 (m, 2H, CH- α Thr, CH- β Thr), 4.37 (q, 1H, ³J=5.5 Hz, H-3'), 4.22-4.18 (m, 1H, O-CH TMSE), 4.14-4.06 (m, 2H, H-4', O-CH TMSE), 3.71 (s, 3H, O-CH₃ DMTr), 3.71 (s, 3H, O-CH₃ DMTr), 3.25 (dd, 1H, ²J=10.3 Hz, ³J=5.9 Hz, H-5'), 3.18 (dd, 1H, ²J=10.3 Hz, ³J=3.4 Hz, H-5''), 2.43 (s, 3H, S-CH₃), 1.22 (d, 3H, ³J=6.3 Hz, CH₃ Thr), 1.02-0.95 (m, 2H, Si-CH₂ TMSE), 0.86 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.09 (s, 3H, Si-CH₃ TBDMS), 0.04 (s, 3H, Si-CH₃ TBDMS), 0.01 (s, 9H, Si(CH₃)₃ TMSE).

N-[[9-(2'-*O*-*tert*-butyldimethylsilyl-5'-*O*-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl)-2-methylthiopurin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**5b''**)

Ref: K. Onizuka, M.E. Hazemi, J.M. Thomas, L.R. Monteleone, K. Yamada, S. Imoto, P.A. Beal, F. Nagatsugi, *Bioorg Med Chem.*, **2017**, *25*, 2191; V. Boudou, J. Langridge, A. V. Aerschot, C. Hendrix, A. Millar, P. Weiss, P. Herdewijn, *Helv. Chim. Acta*, **2000**, *83*, 152;

To solution of 5'-DMTr nucleoside **5b'** (122 mg, 0.13 mmol) in freshly distilled THF (0.50 mL), anhydrous pyridine (50 μ L, 0.65 mmol) and AgNO₃ (32 mg, 0.19 mmol) were added. The reaction mixture was stirred at room temperature for 30 min in darkness. Then TBDMS-Cl (34 mg, 0.23 mmol) was added and stirring was continued at room temperature for 2 h. After this time, the reaction mixture was diluted with CH₂Cl₂ (5 mL), filtered through Celite and washed with CH₂Cl₂. The solution was extracted with saturated NaHCO₃ (2 x 5 mL) and brine (5 mL). The organic layer

was dried over MgSO₄, filtered and concentrated under reduced pressure. The oily residue was co-evaporated with toluene (2 x 5 mL) and the crude product was purified by the column chromatography (silica gel, 0-8 % acetone in CHCl₃). The pure 2'-isomer **5b''** was obtained as a white solid in 57 % yield (80 mg, 0.074 mmol). The 3'-regioisomer (38 mg, 0.035 mmol) was dissolved in methanol with few drops of TEA, to give equimolar mixture of 2' and 3' TBDMS isomers from which 2'-TBDMS nucleoside was chromatographically isolated to give finally 99 mg of **5b''** (0.091 mmol; 70 % yield). **TLC:** R_f = 0.42 (CHCl₃/MeOH, 98:2 v/v).

¹H NMR (700 MHz, DMSO-*d*₆) δ: 9.94 (s, 1H, NH-6), 9.27 (d, 1H, ³J=8.6 Hz, NH Thr), 8.41 (s, 1H, H-8), 7.36-7.32 (m, 2H, H_{Ar} DMTr), 7.26-7.21 (m, 6H, H_{Ar} DMTr), 7.21-7.16 (m, 1H, H_{Ar} DMTr), 6.85-6.80 (m, 4H, H_{Ar} DMTr), 5.99 (d, 1H, ³J=4.5 Hz, H-1'), 5.19 (d, 1H, ³J=6.1 Hz, 3'OH), 4.89 (t, 1H, ³J=4.7 Hz, H-2'), 4.48-4.42 (m, 2H, CH-α Thr, CH-β Thr), 4.30 (q, 1H, ³J=5.6 Hz, H-3'), 4.20-4.18 (m, 1H, O-CH TMSE), 4.15-4.06 (m, 2H, H-4', O-CH TMSE), 3.72 (s, 3H, O-CH₃DMTr), 3.71 (s, 3H, O-CH₃DMTr), 3.31-3.30 (m, 1H, H-5'), 3.23 (dd, 1H, ²J=10.3 Mz, ³J=3.4 Hz, H-5'), 2.39 (s, 3H, S-CH₃), 1.21 (d, 3H, ³J=6.2 Hz, CH₃ Thr), 0.99-0.95 (m, 2H, Si-CH₂ TMSE), 0.85 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.77 (s, 9H, Si-C(CH₃)₃ TBDMS), 0.08 (s, 3H, Si-CH₃ TBDMS), 0.03 (s, 3H, Si-CH₃ TBDMS), -0.00 (s, 9H, Si(CH₃)₃ TMSE), -0.02 (s, 3H, Si-CH₃ TBDMS), -0.11 (s, 3H, Si-CH₃ TBDMS).

N-[[9-(2'-*O*-*tert*-butyldimethylsilyl)-3'-(2-cyanoethyl-*N,N*-diisopropylphosphoramidite)-5'-*O*-(4,4'-dimethoxytrityl)-β-*D*-ribofuranosyl)-2-methylthiopurin-6-yl]carbamoyl]-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**6b**)

Ref: V. Boudou, J. Langridge, A. V. Aerschot, C. Hendrix, A. Millar, P. Weiss, P. Herdewijn, *Helv. Chim. Acta*, **2000**, *83*, 152;

To stirred solution of 5'-DMTr, 2'-TBDMS-ms²t⁶A **5b''** (80 mg, 0.08 mmol) in freshly distilled CH₂Cl₂ (0.4 mL) under argon atmosphere anhydrous DIPEA was added (0.05 mL, 0.29 mmol). Then 2-cyanoethyl-*N,N*-diisopropyl chlorophosphoramidite (0.04 ml; 0.14 mmol) were added dropwise. The reaction was stirred for 3 h under argon atmosphere at room temperature. The reaction mixture was diluted with CH₂Cl₂ (5 mL) and washed with 5 % aq NaHCO₃ (2 x 5 mL) and H₂O (5 mL), then the organic layer was dried over anhydrous MgSO₄, filtered and evaporated under reduced pressure. The crude product was purified by the flash chromatography (silica gel, petroleum ether/acetone, 2:1 v/v) to obtain pure product **6b** as a white solid in 90 % yield (85 mg; 0.07 mmol). **TLC:** R_f= 0.53 (CHCl₃/acetone, 95:5 v/v).

³¹P NMR: (283 MHz, C₆H₆,) δ: 150.90, 149.49.

III. Spectroscopic data.

3.1 NMR and IR spectra of threonine derivatives.

Figure S1 ^1H NMR spectrum of *N*-(*tert*-butoxycarbonyl)-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**1**)

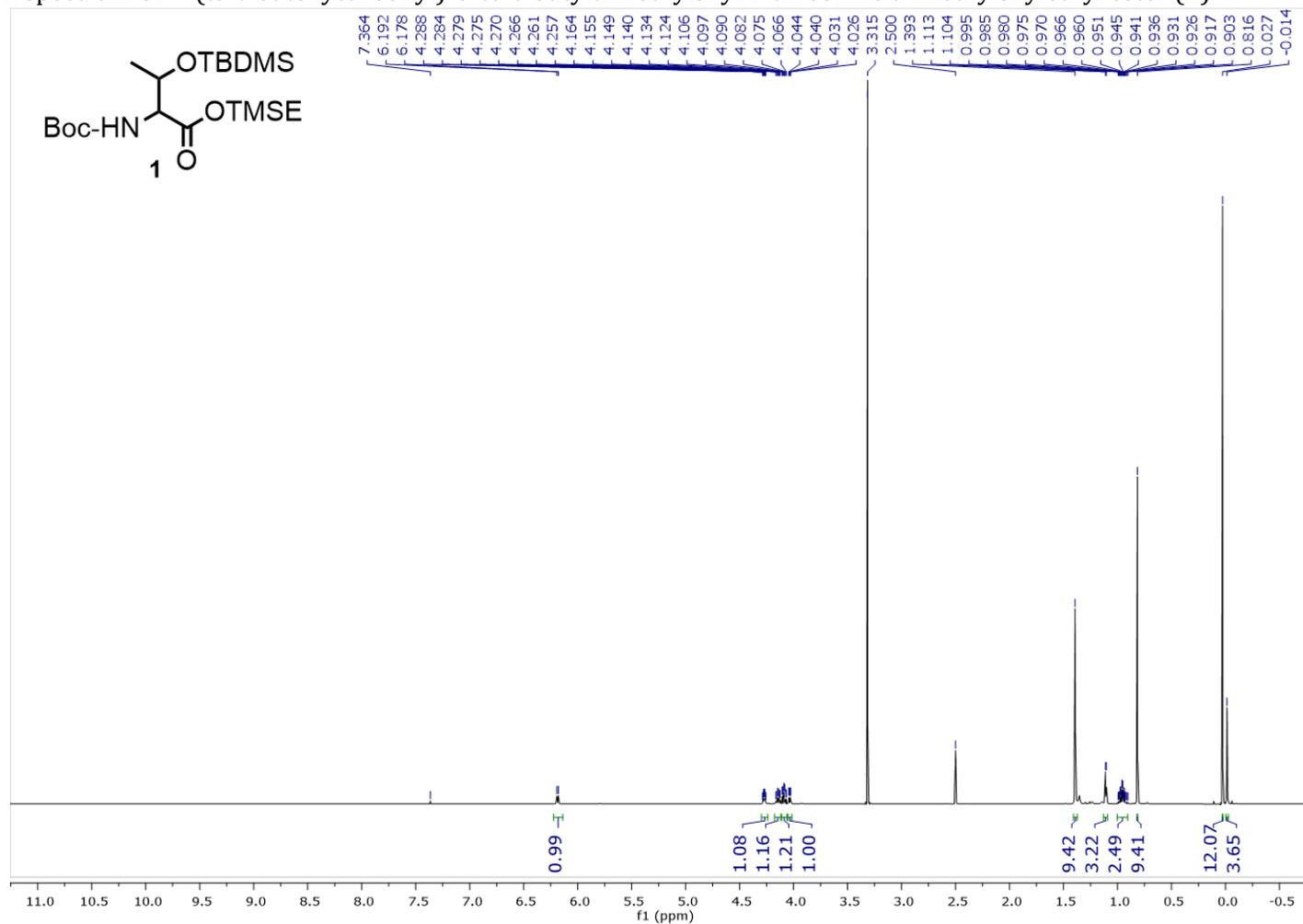
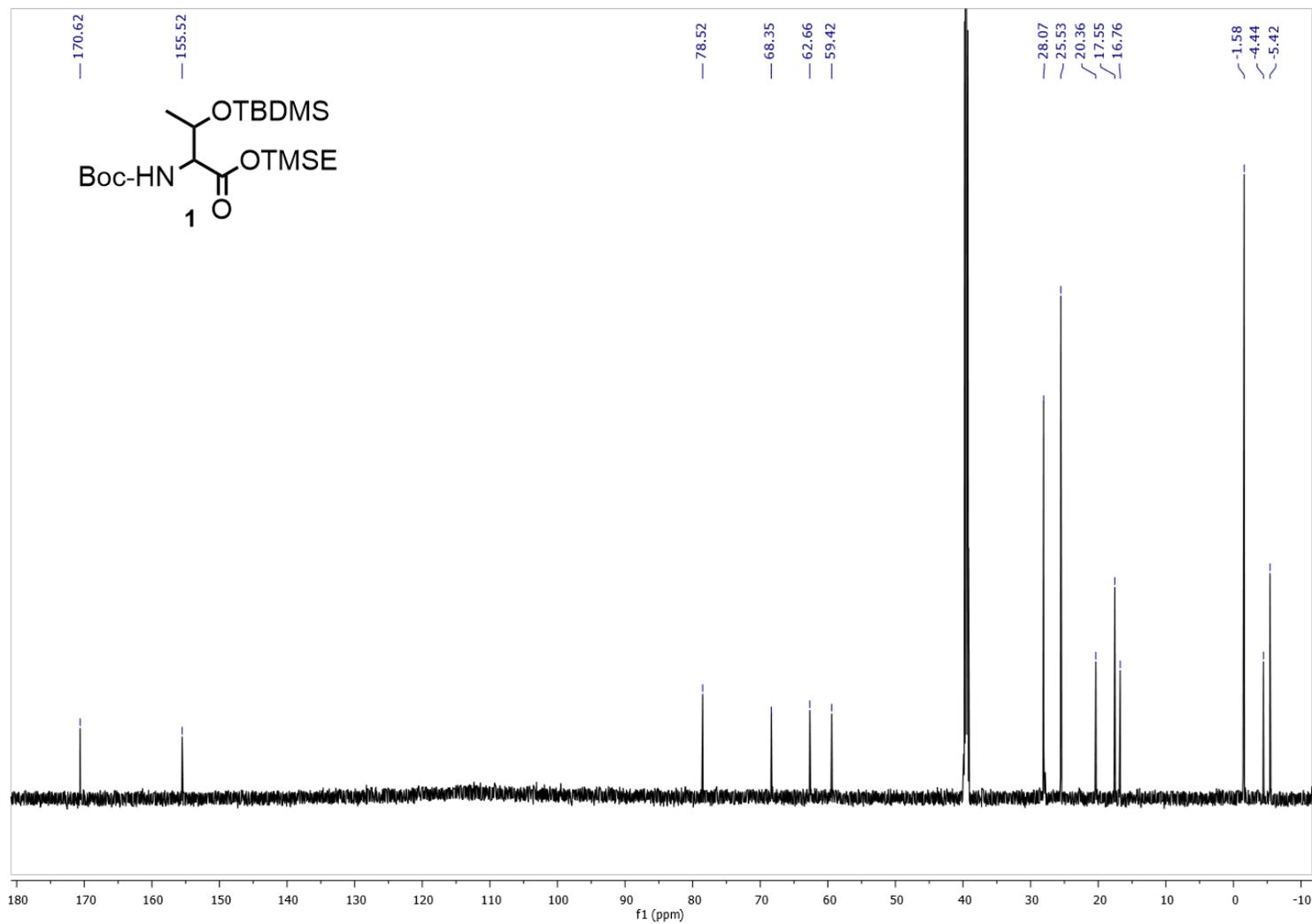
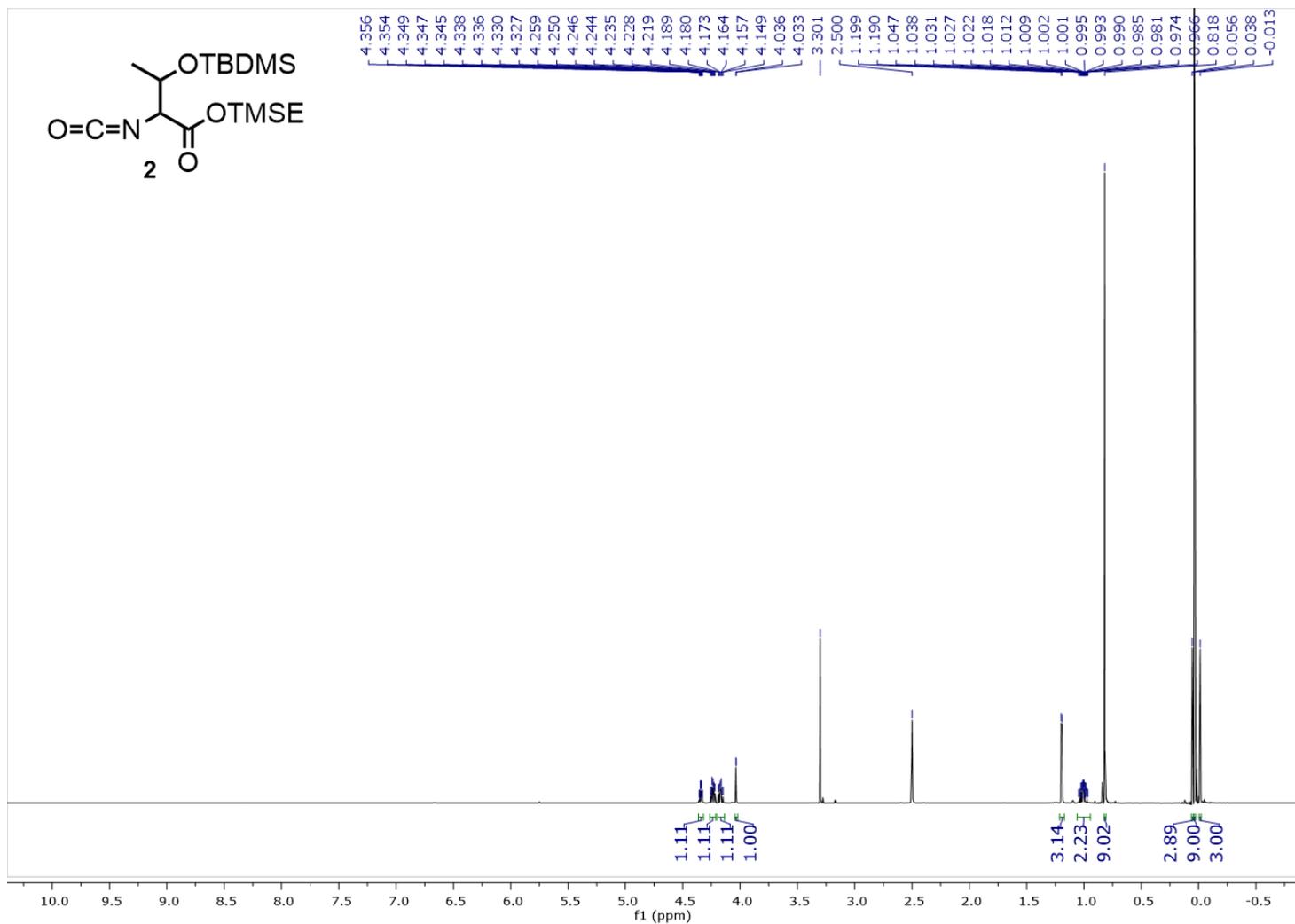


Figure S2 ^{13}C NMR spectrum of *N*-(*tert*-butoxycarbonyl)-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**1**)



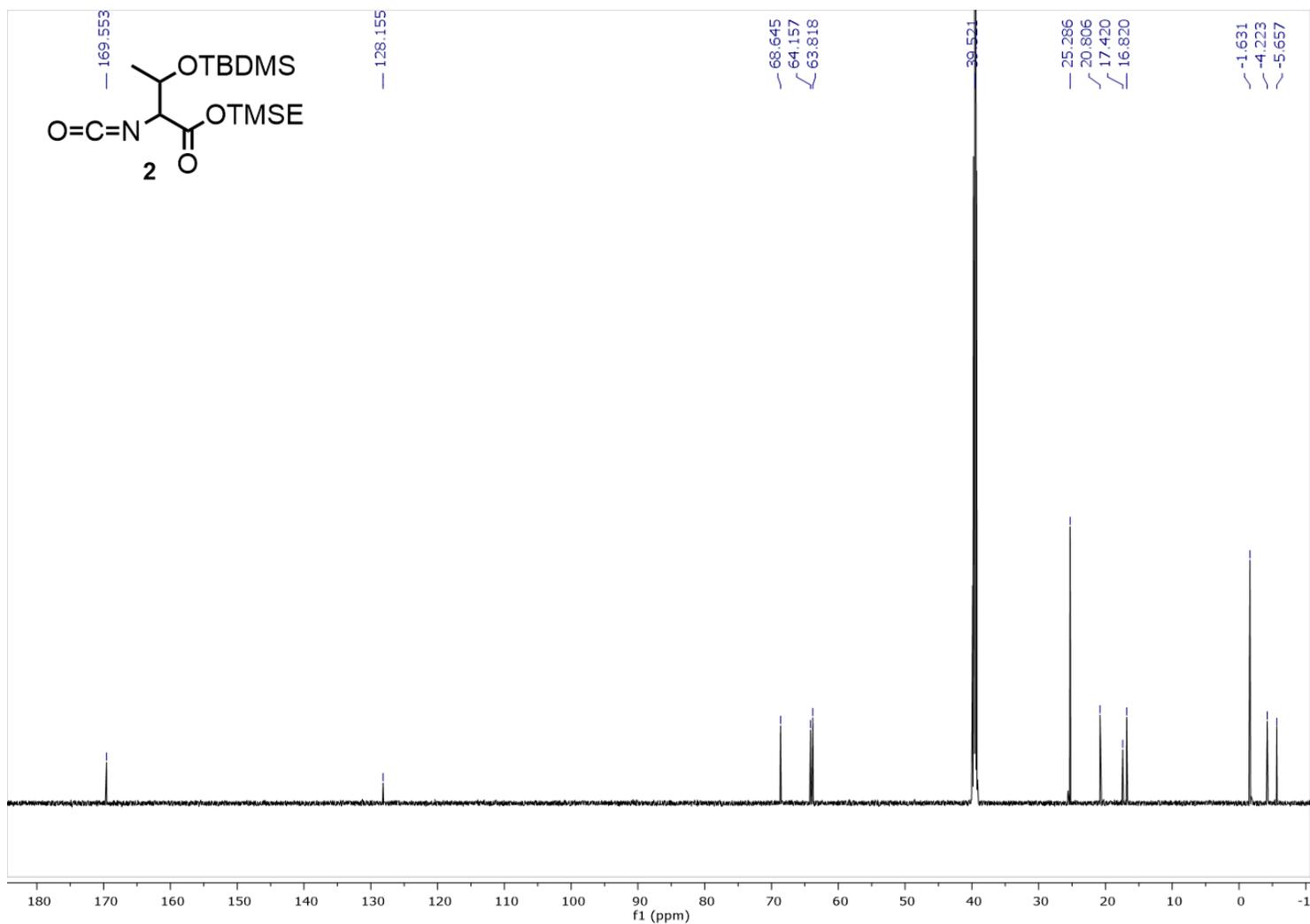
^{13}C NMR: (176,03 MHz DMSO- d_6) δ : 170.62 (C=O), 155.52 (C=O), 78.52 (C-(CH $_3$) $_3$ Boc), 68.35 (CH- β), 62.66 (O-CH $_2$ TMSE), 59.42 (CH- α), 28.07 (C-(CH $_3$) $_3$ Boc), 25.53 (Si-C(CH $_3$) $_3$ TBDMS), 20.36 (CH $_3$), 17.55 (Si-C-CH $_3$) TBDMS), 16.76 (Si-CH $_2$ TMSE), -1.58 (Si(CH $_3$) $_3$ TMSE), -4.44 (Si-CH $_3$ TBDMS), -5.42 (Si-CH $_3$ TBDMS).

Figure S3 ^1H NMR spectrum of 2-(S)-isocyane-3-(*O*-*tert*-butyldimethylsilyl)-3-hydroxybutanoic 2-trimethylsilylethyl ester (**2**)



^1H NMR (700 MHz, $\text{DMSO}-d_6$) δ : 4.34 (qd, 1H, $^3J=6.2$ Hz, $^3J=1.7$ Hz, CH- β), 4.24 (td, 1H, $^3J=10.9$ Hz, $^3J=6.4$ Hz, O-CH TMSE), 4.17 (td, 1H, $^3J=11.1$ Hz, $^3J=5.9$ Hz, O-CH TMSE), 4.03 (d, 1H, $^3J=1.7$ Hz, CH- α), 1.19 (d, 3H, $^3J=6.3$ Hz, CH_3), 1.07–0.94 (m, 2H, Si- CH_2 TMSE), 0.82 (s, 9H, Si-C-(CH_3) $_3$ TBDMS), 0.06 (s, 3H, Si- CH_3 TBDMS), 0.04 (s, 9H, Si-(CH_3) $_3$ TMSE), -0.01 (s, 3H, Si- CH_3 TBDMS).

Figure S4 ^{13}C NMR spectrum of 2-(S)-isocyane-3-(*O*-*tert*-butyldimethylsilyl)-3-hydroxybutanoic 2-trimethylsilylethyl ester (**2**)



^{13}C NMR (176,03 MHz DMSO- d_6) δ : 169.55 (C=O), 128.15 (N=C=O), 68.64 (CH- β), 64.16 (O-CH $_2$ TMSE), 63.82 (CH- α), 25.29 (Si-C(CH $_3$) $_3$ TBDMS), 20.81 (CH $_3$), 17.42 (Si-C(CH $_3$) $_3$ TBDMS), 16.82 (Si-CH $_2$ TMSE), -1.63 (Si(CH $_3$) $_3$ TMSE), -4.22 (Si-CH $_3$ TBDMS), -5.66 (Si-CH $_3$ TBDMS).

Figure S5 IR spectrum of *N*-(*tert*-butoxycarbonyl)-*O*-*tert*-butyldimethylsilyl-*L*-threonine trimethylsilylethyl ester (**1**)

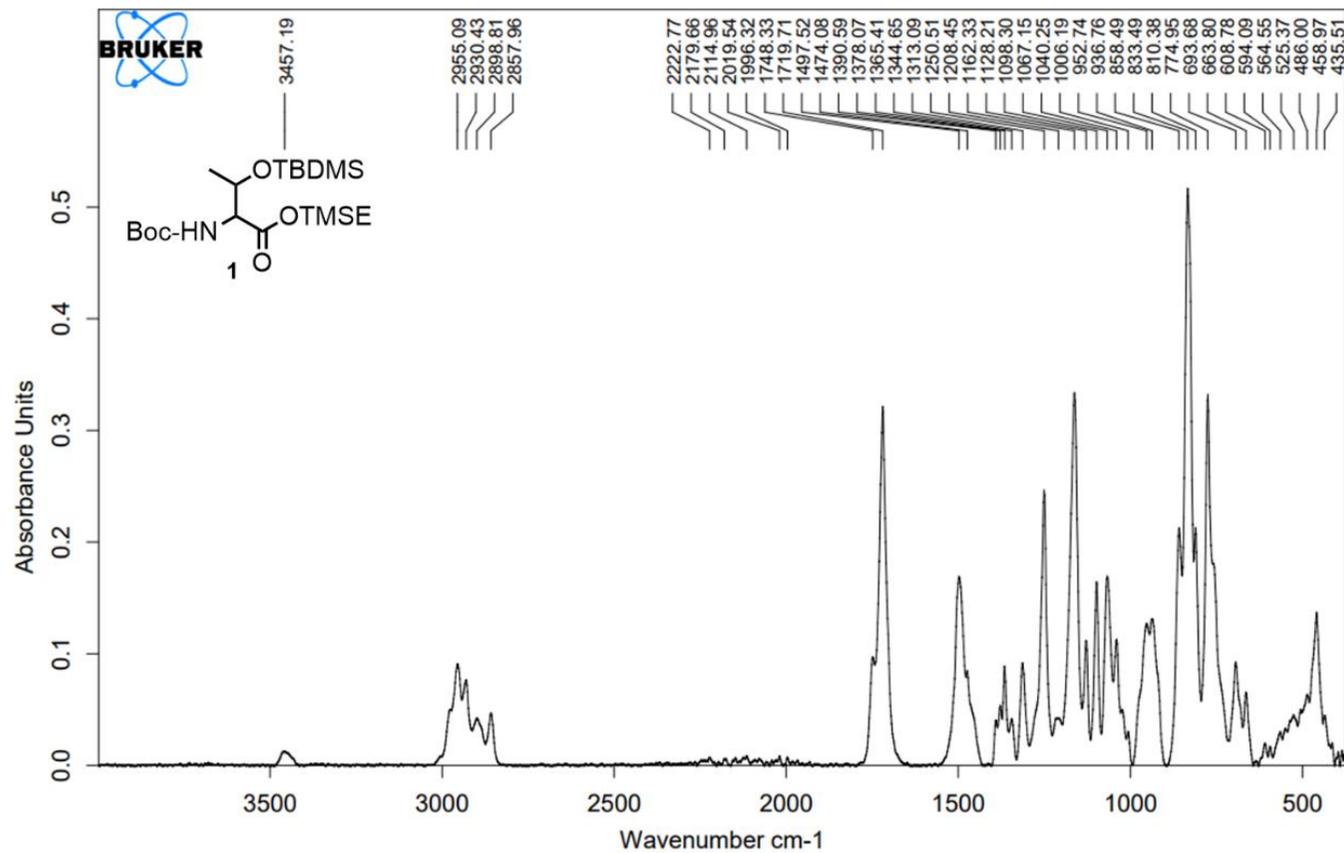
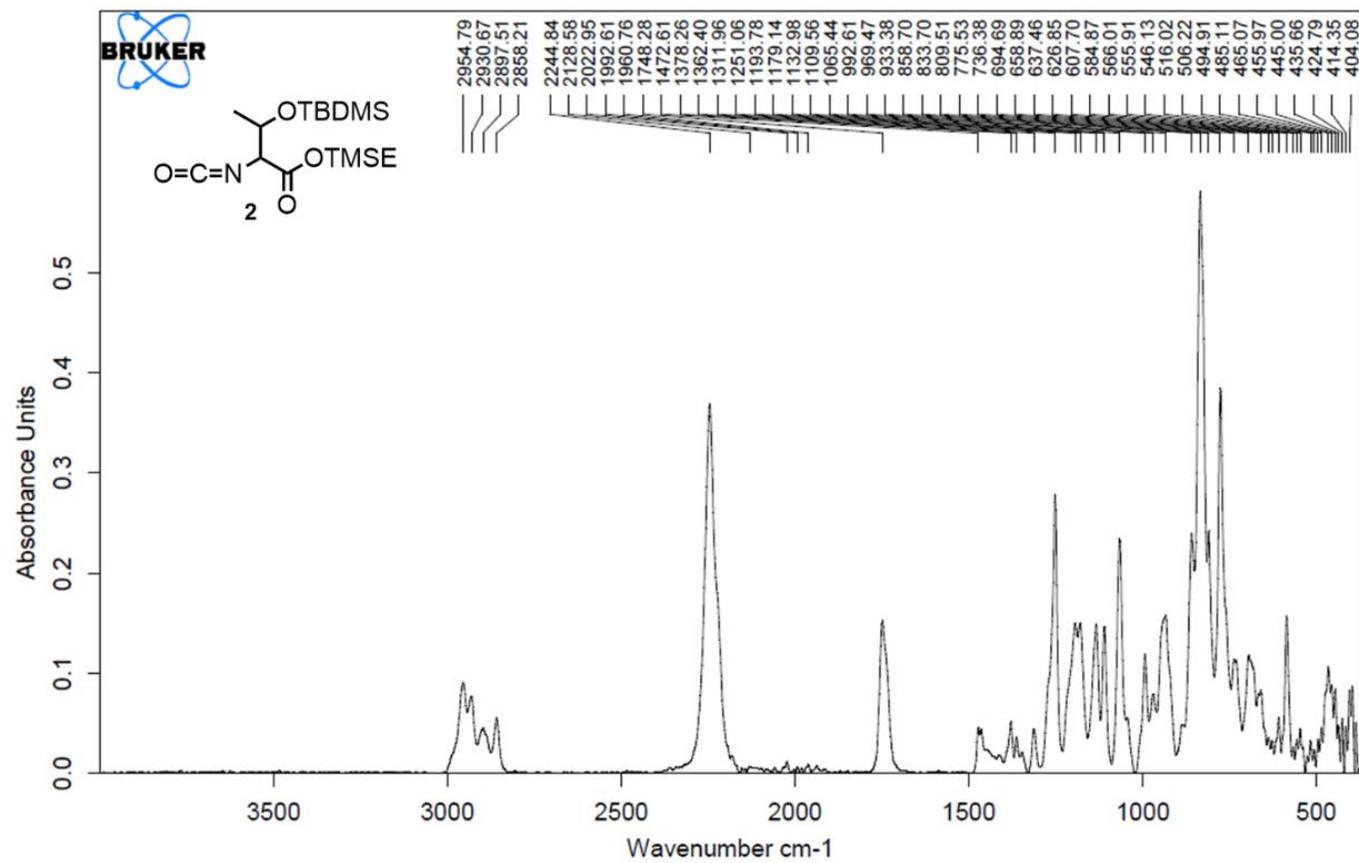
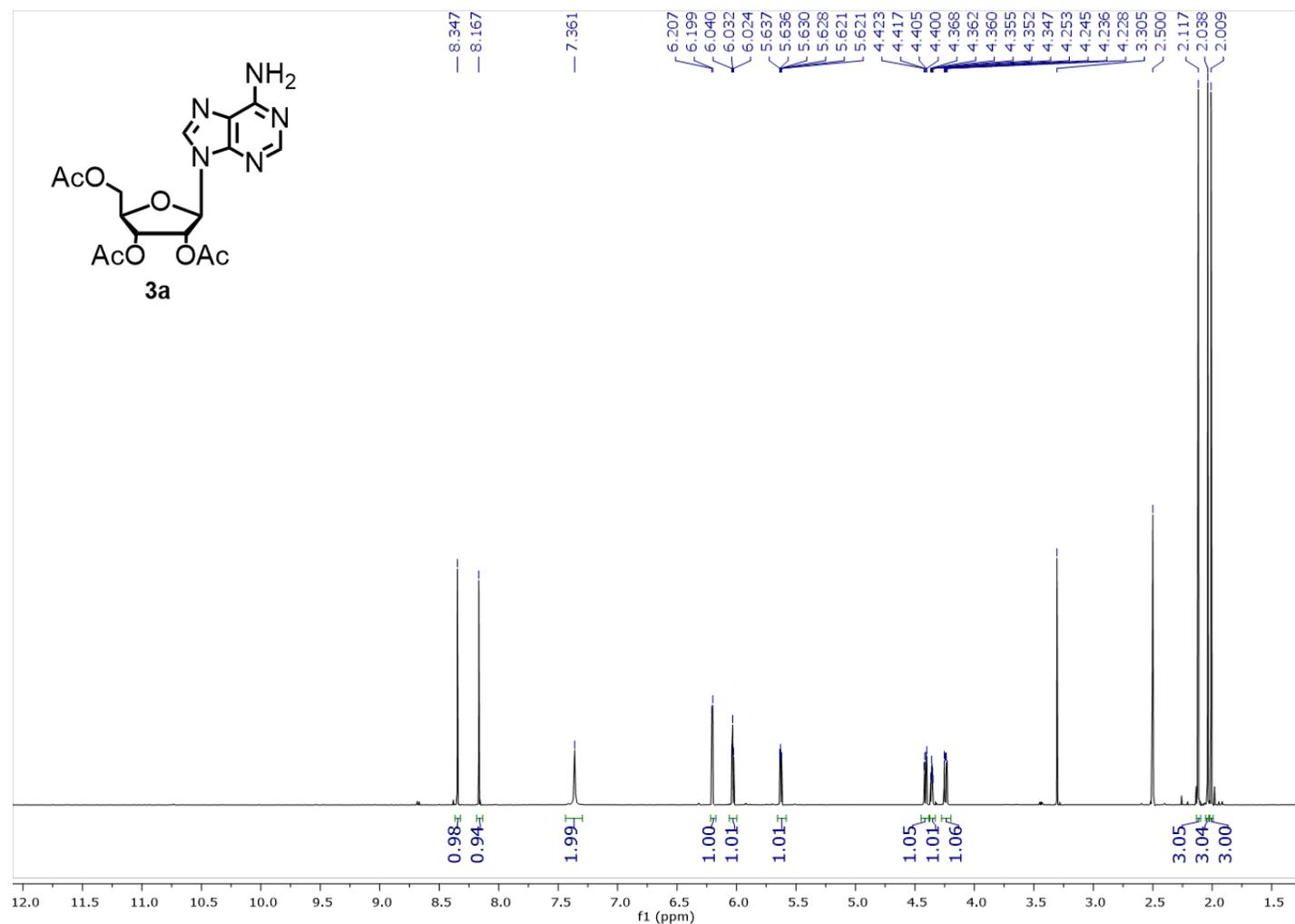


Figure S6 IR spectrum of 2-(S)-isocyane-3-(*O*-*tert*-butyldimethylsilyl)-3-hydroxybutanoic 2-trimethylsilylethyl ester (**2**)



3.2 NMR spectra of adenosine derivatives

Figure S7 ^1H NMR spectrum of **3a**



^1H NMR: (700 MHz, DMSO- d_6) δ : 8.35 (s, 1H, H-8), 8.17 (s, 1H, H-2), 7.36 (br s, 2H, NH₂), 6.20 (d, 1H, $^3J=5.5$ Hz, H-1'), 6.03 (t, 1H, $^3J=5.7$ Hz, H-2'), 5.63 (dd, 1H, $^3J=5.8$ Hz, $^3J=5.1$ Hz, H-3'), 4.41 (dd, 1H, $^2J=11.9$ Hz, $^3J=3.8$ Hz, H-5'), 4.38-4.34 (m, 1H, H-4'), 4.24 (dd, 1H, $^2J=11.9$ Hz, $^3J=5.6$ Hz, H-5''), 2.12 (s, 3H, CH₃-CO Ac), 2.04 (s, 3H, CH₃-CO Ac), 2.01 (s, 3H, CH₃-CO Ac).

Figure S8 ¹H NMR spectrum of 4a

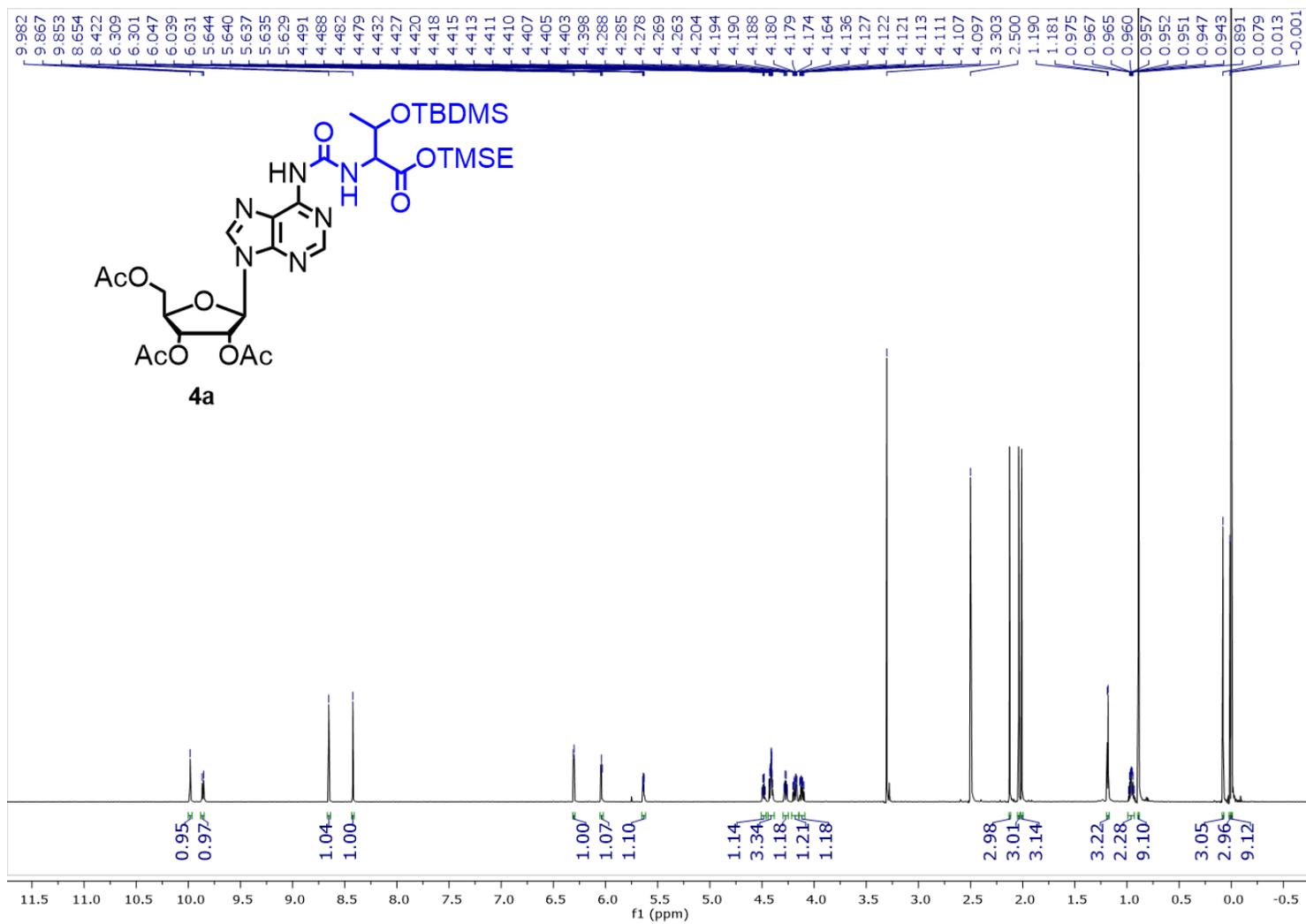


Figure S9 ¹H NMR spectrum of 5a

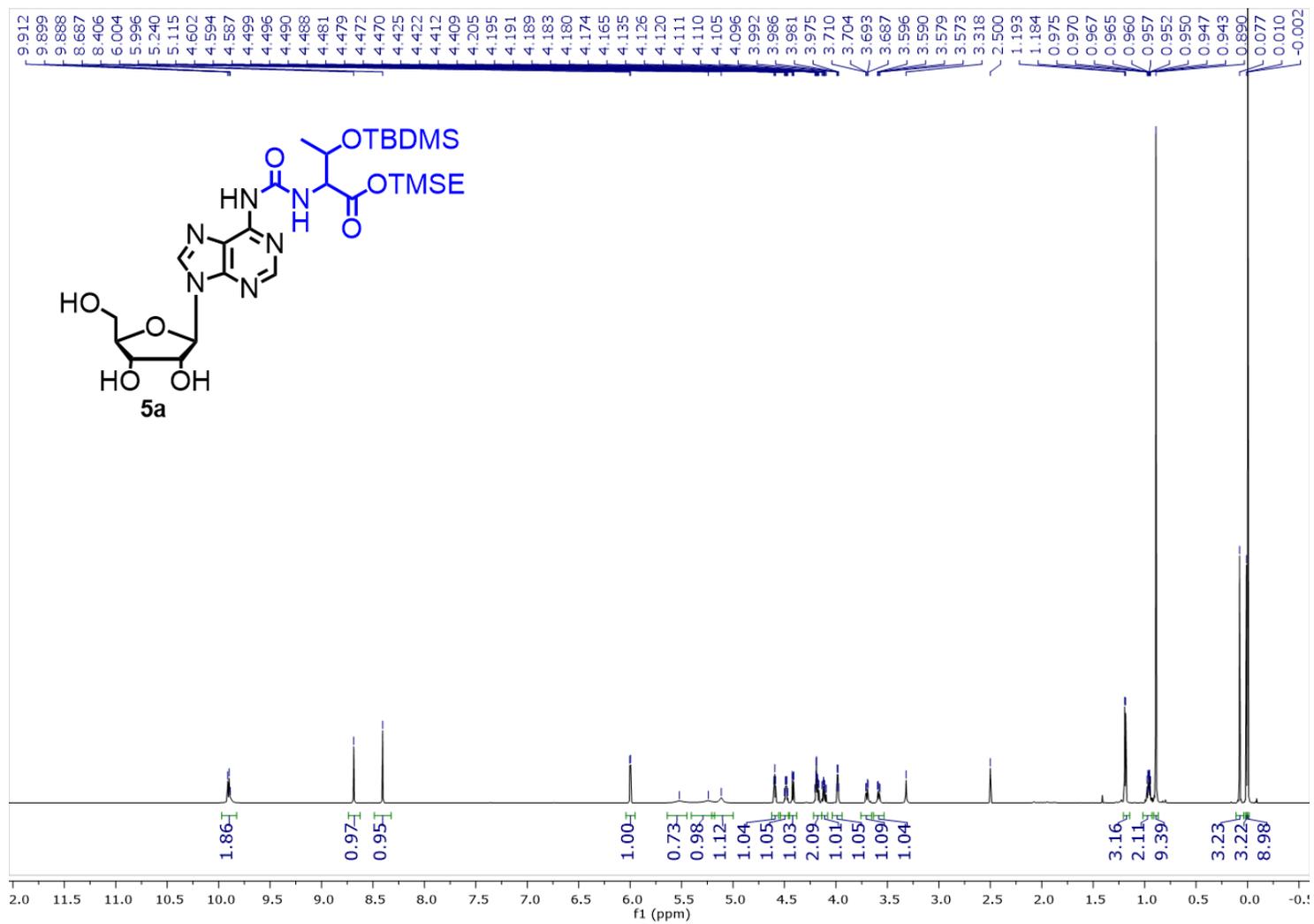


Figure S10 ¹H NMR spectrum of 5a'

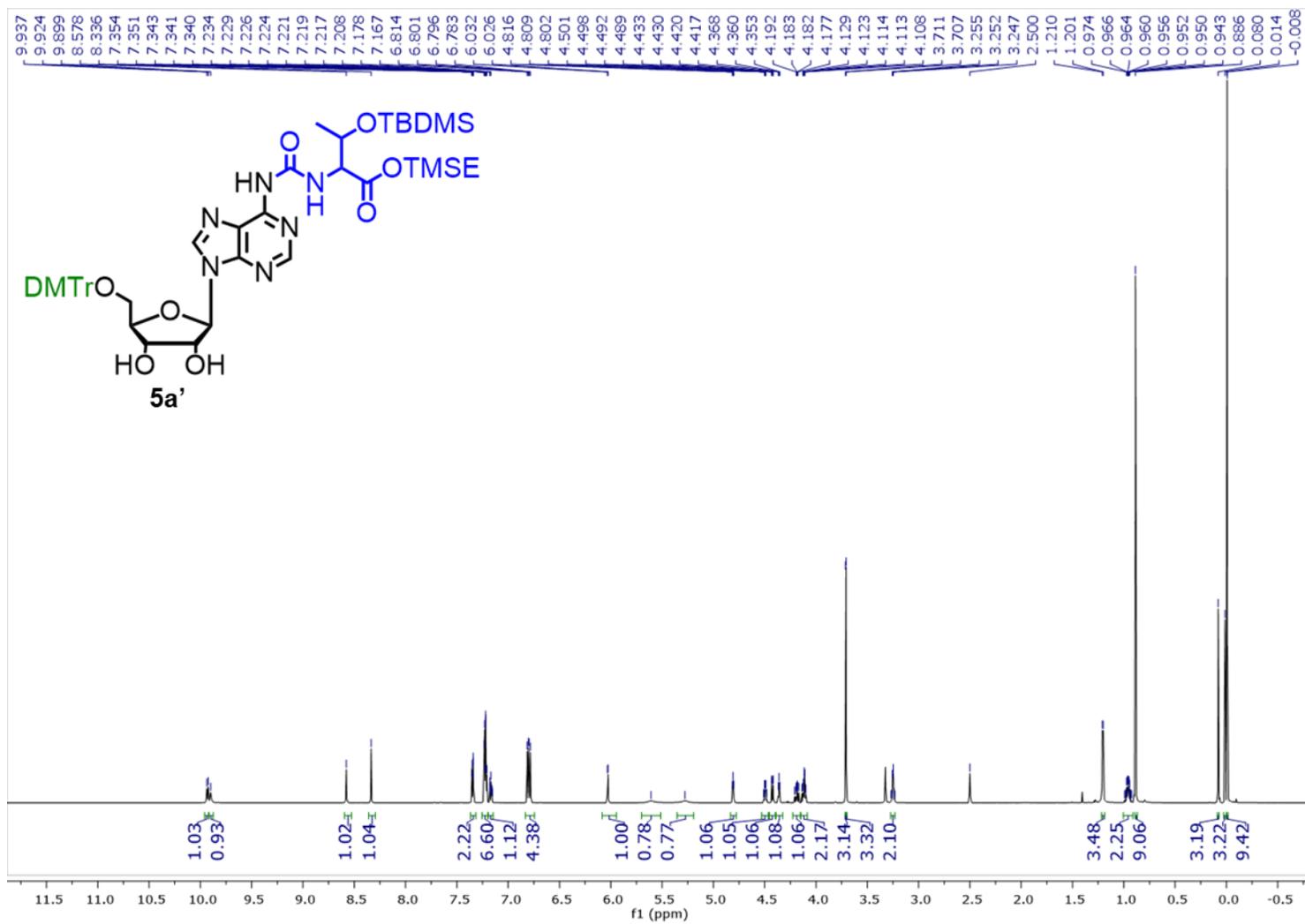


Figure S11 ¹H NMR spectrum of 5a''

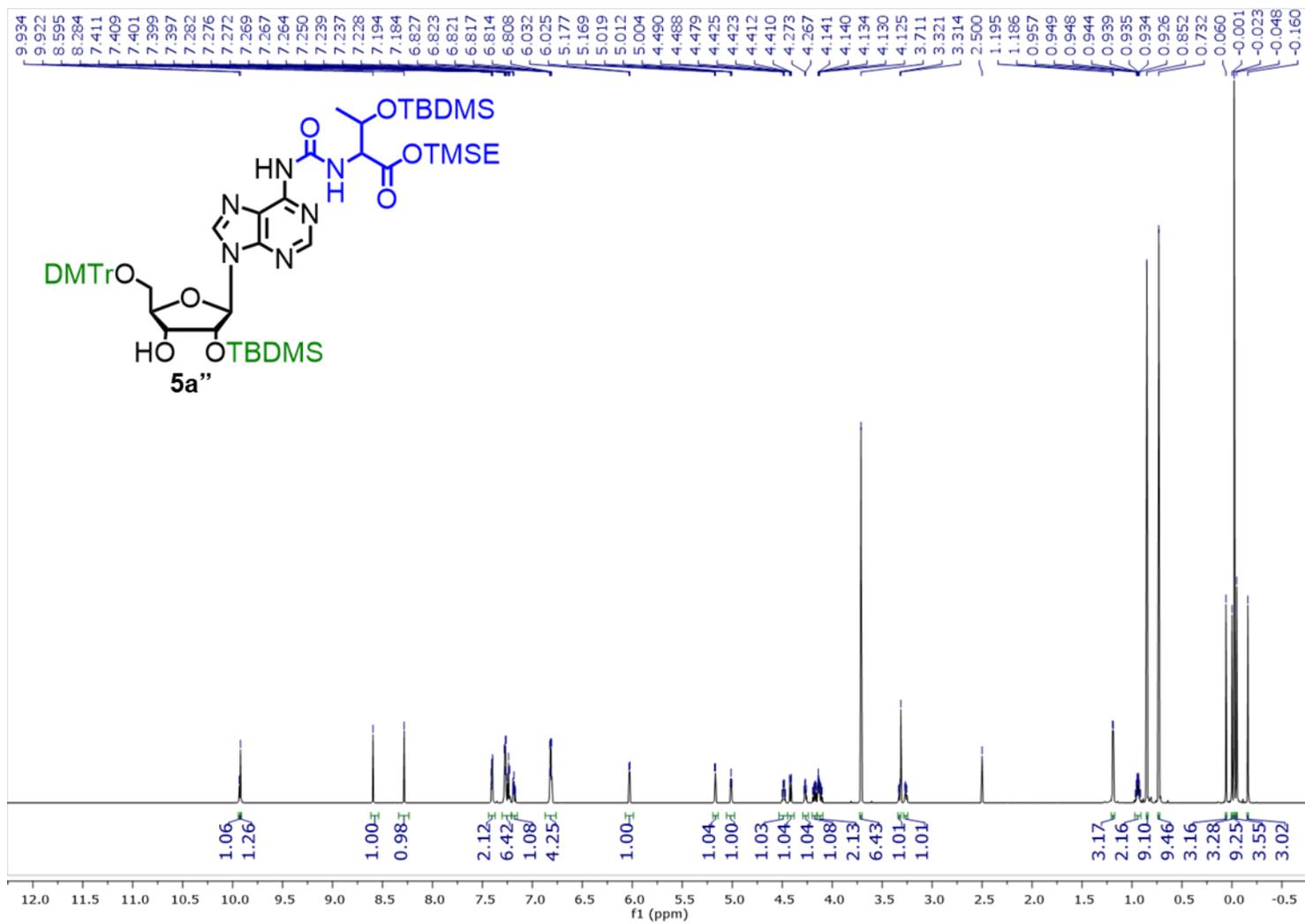


Figure S12 ^{31}P NMR spectrum of **6a**

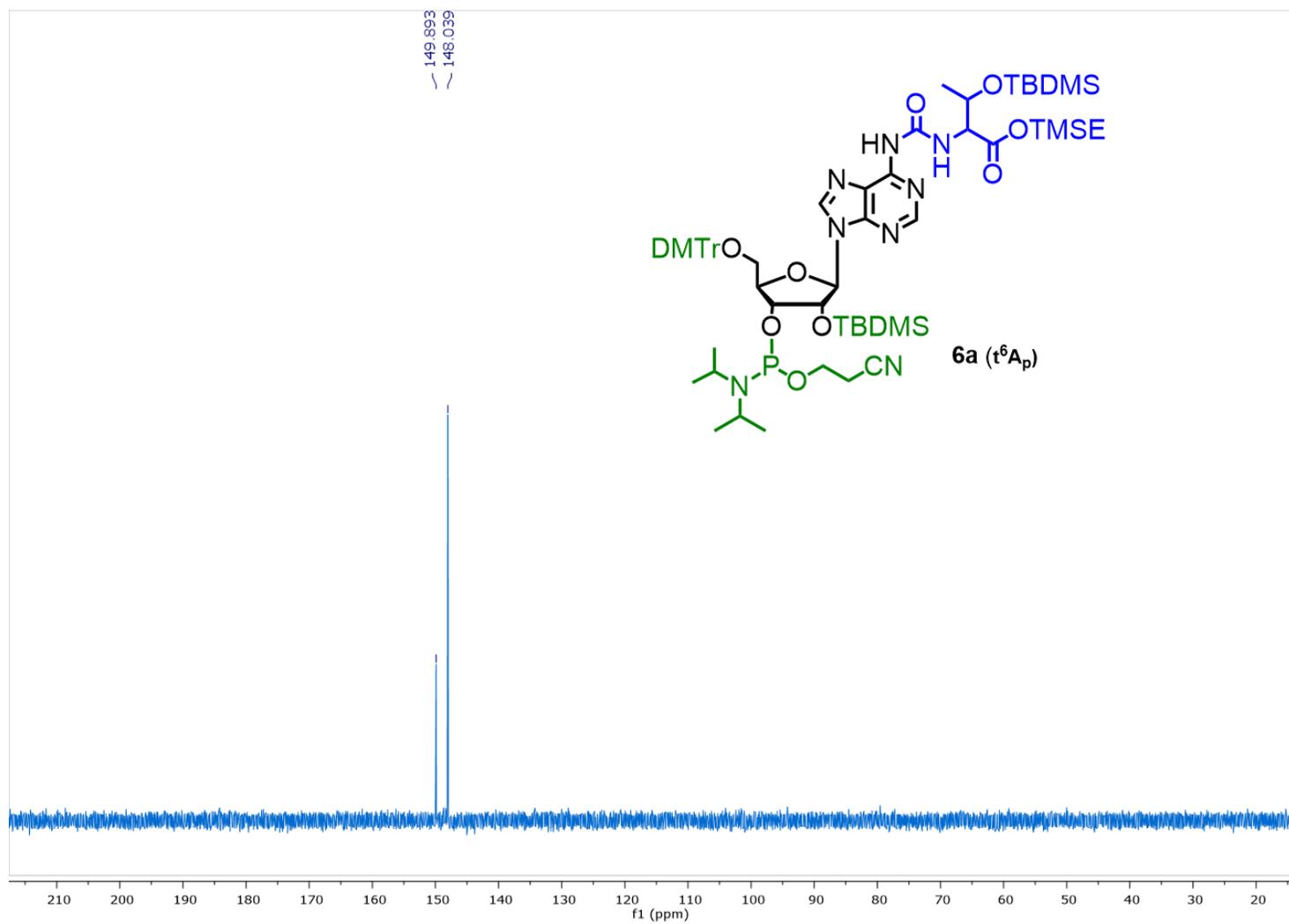
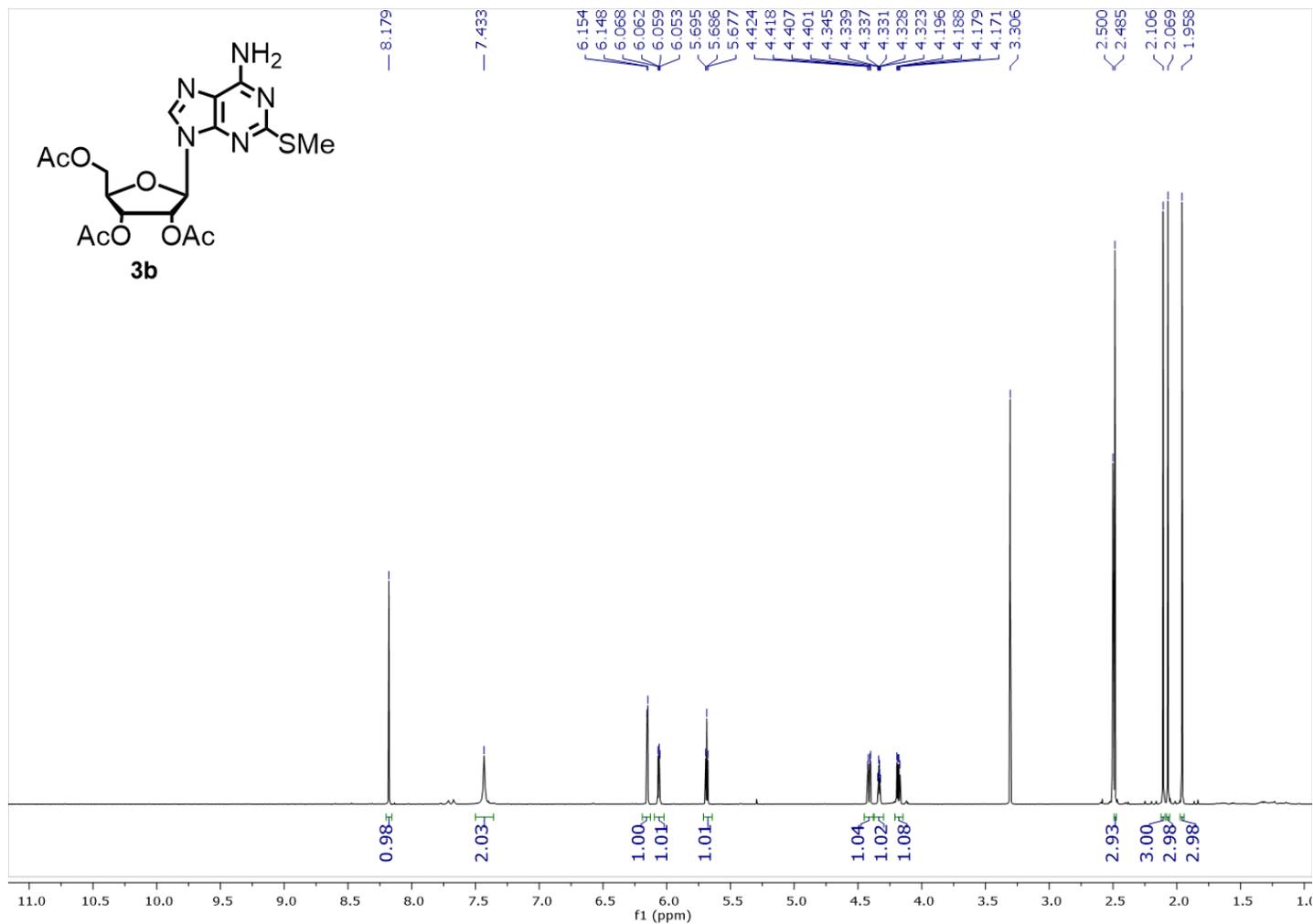


Figure S13 ^1H NMR spectrum of **3b**



^1H NMR: (700 MHz, DMSO- d_6) δ : 8.18 (s, 1H, H-2), 7.43 (br s, 2H, NH_2), 6.15 (d, 1H, $^3J=4.3$ Hz, H-1'), 6.06 (dd, 1H, $^3J=6.1$ Hz, $^3J=4.3$ Hz, H-2'), 5.69 (t, 1H, $^3J=6.1$ Hz, H-3'), 4.41 (dd, 1H, $^2J=12.0$ Hz, $^3J=3.8$ Hz, H-5'), 4.41 (td, 1H, $^3J=5.8$ Hz, $^3J=3.7$ Hz, H-4'), 4.18 (dd, 1H, $^2J=12.0$ Hz, $^3J=5.6$ Hz, H-5''), 2.48 (s, 3H, -SCH₃), 2.11 (s, 3H, CH₃-CO Ac), 2.07 (s, 3H, CH₃-CO Ac), 1.96 (s, 3H, CH₃-CO Ac).

Figure S14 ¹H NMR spectrum of **4b**

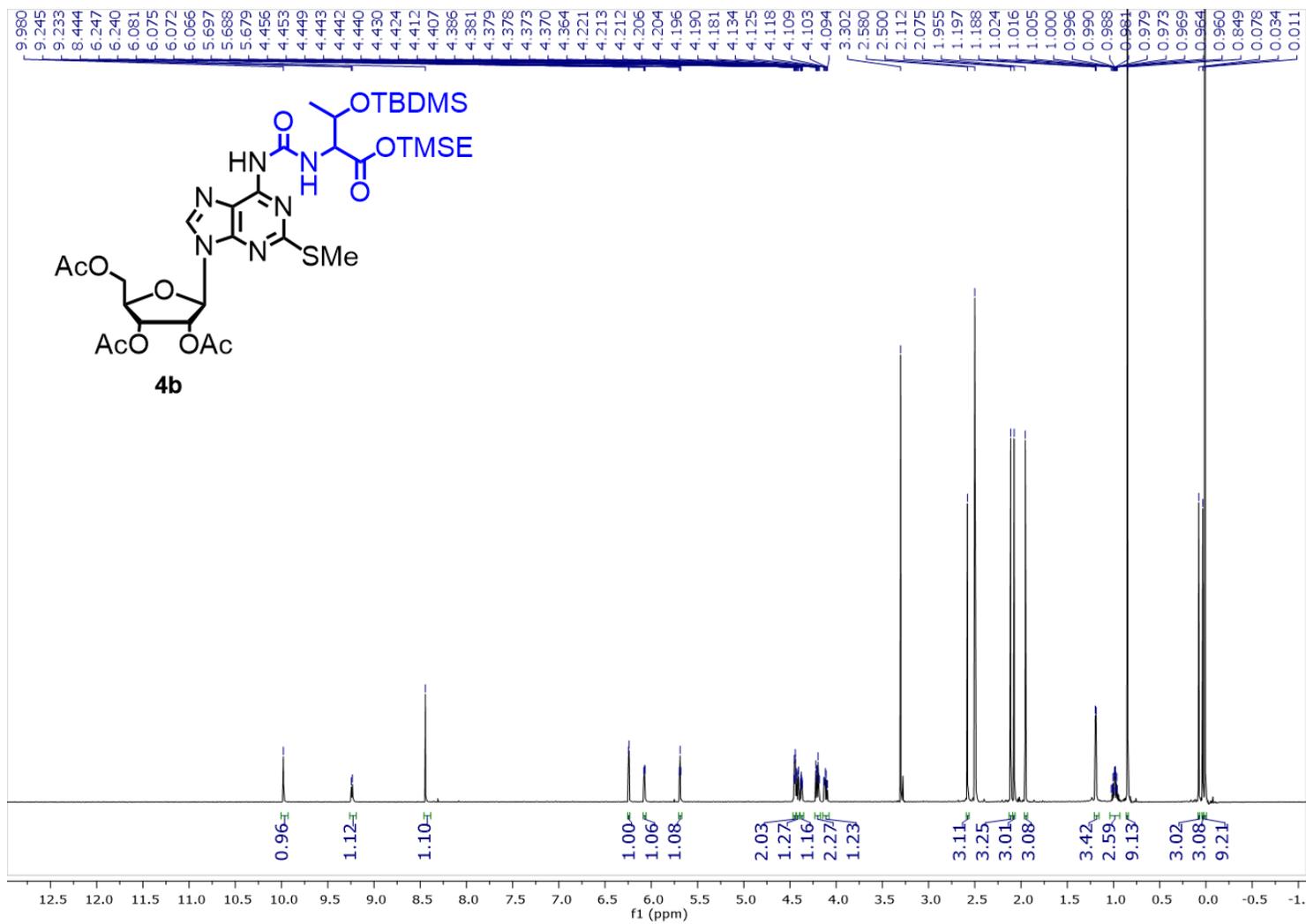


Figure S15 ¹H NMR spectrum of **5b**

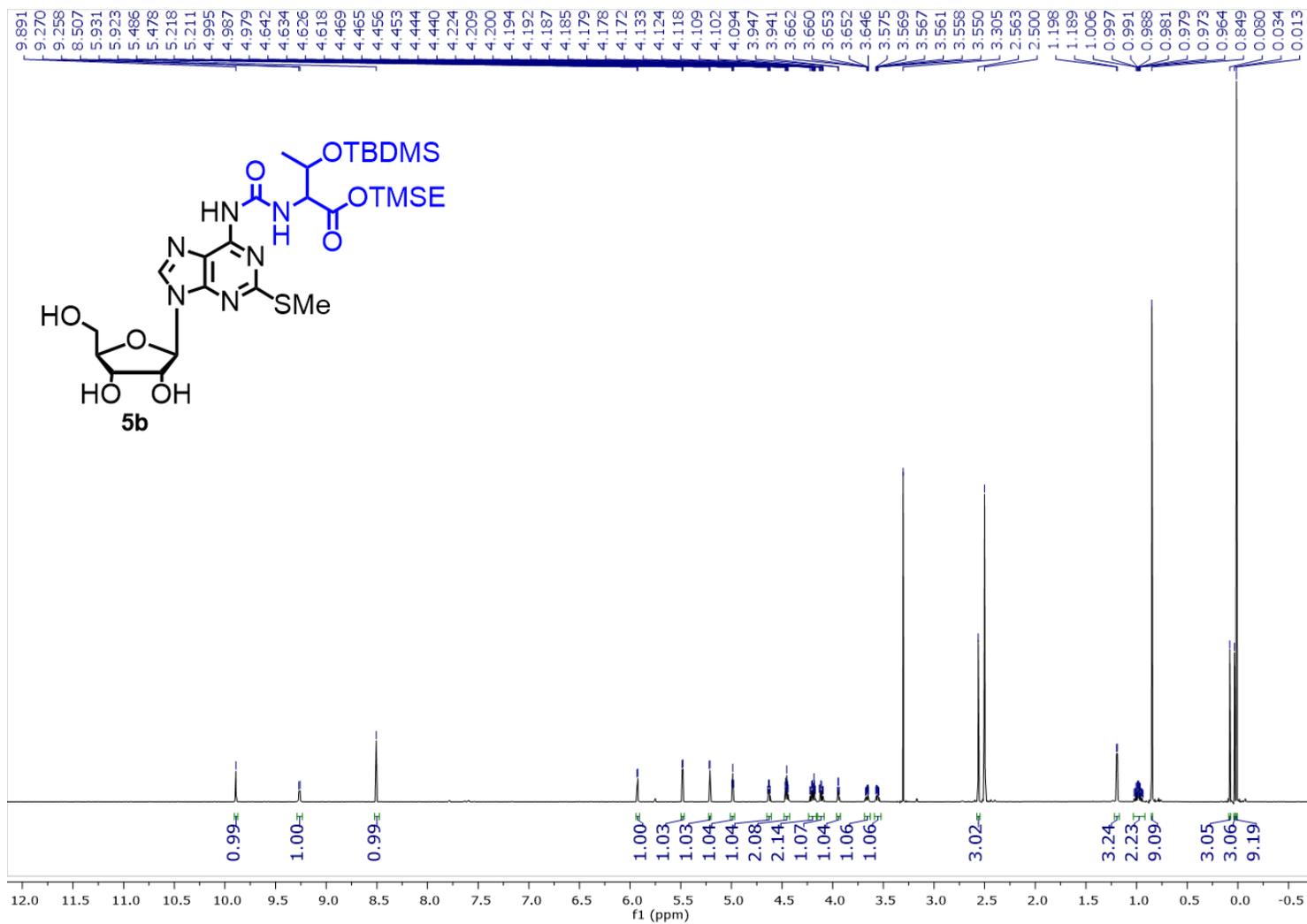


Figure S16 ¹H NMR spectrum of **5b'**

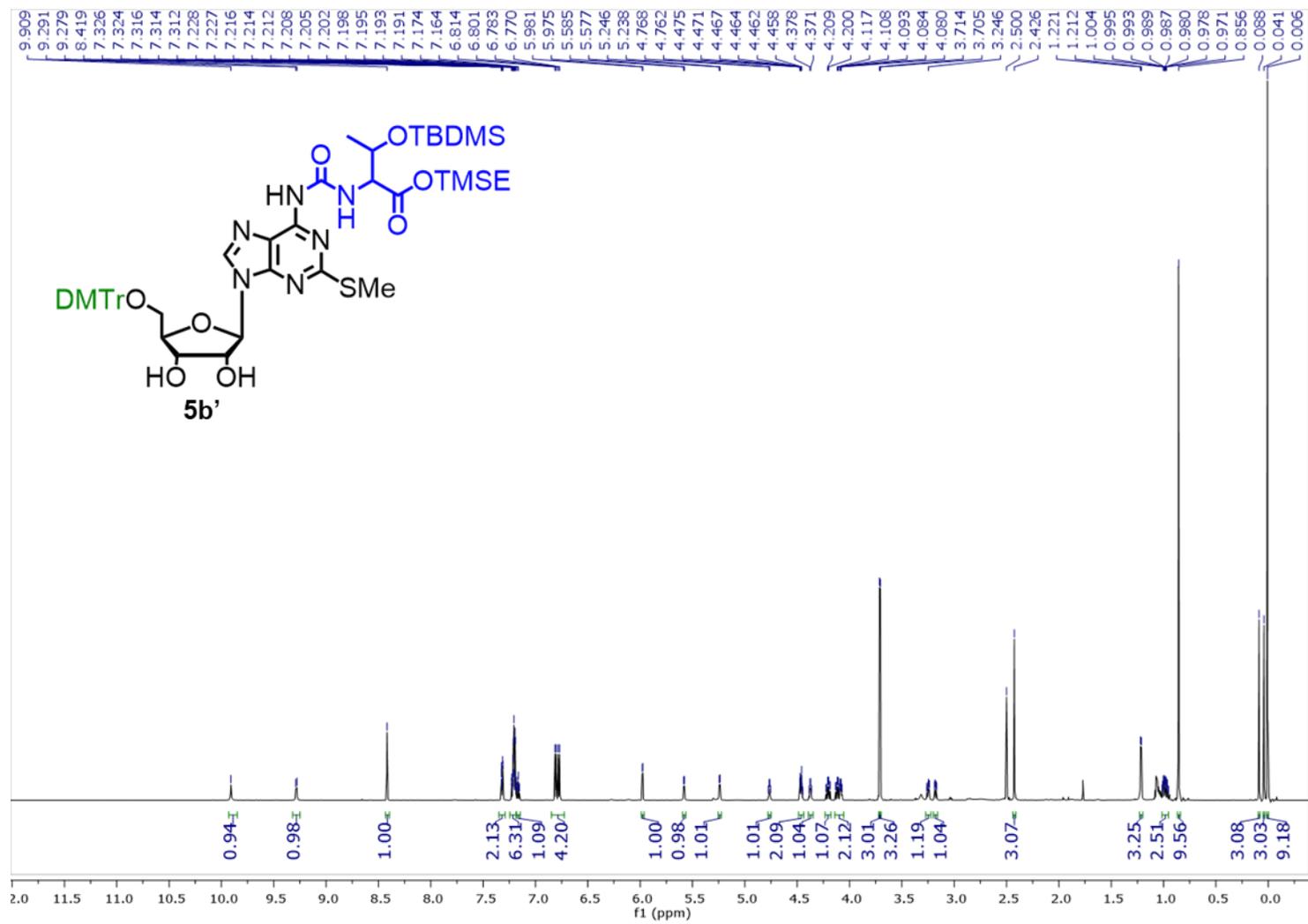


Figure S17 ¹H NMR spectrum of 5b''

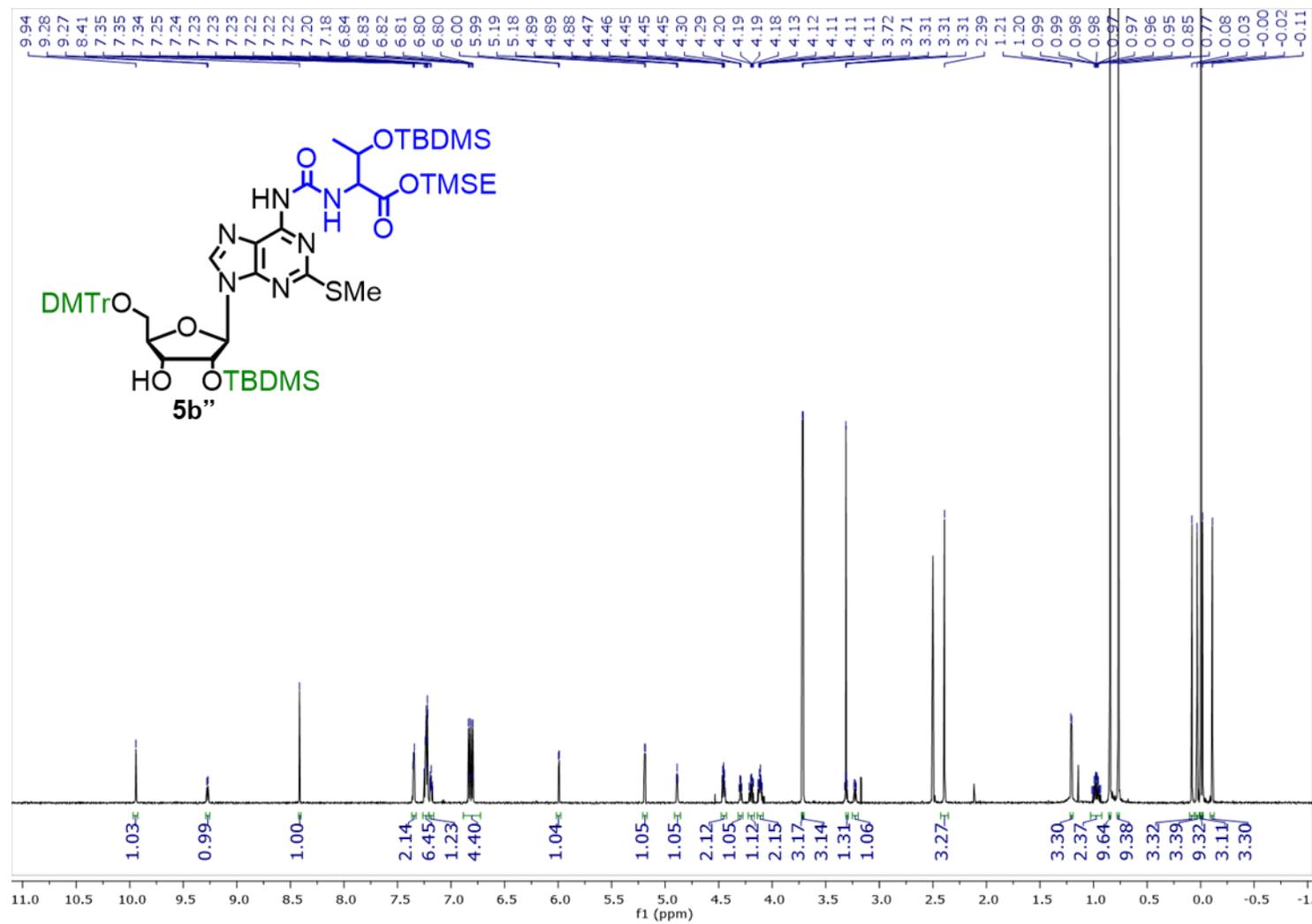


Figure S18 ^{31}P NMR spectrum of **6b**

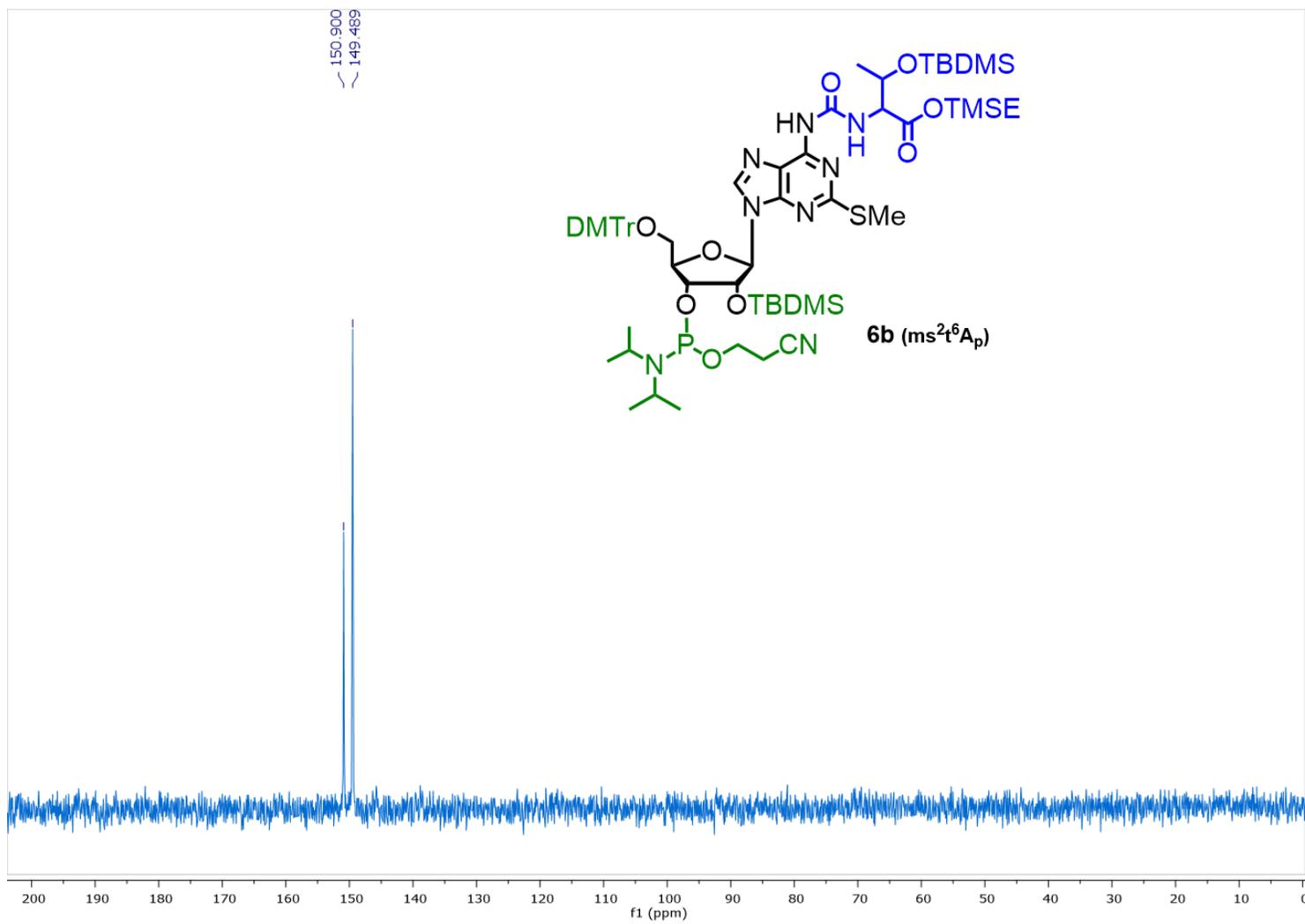
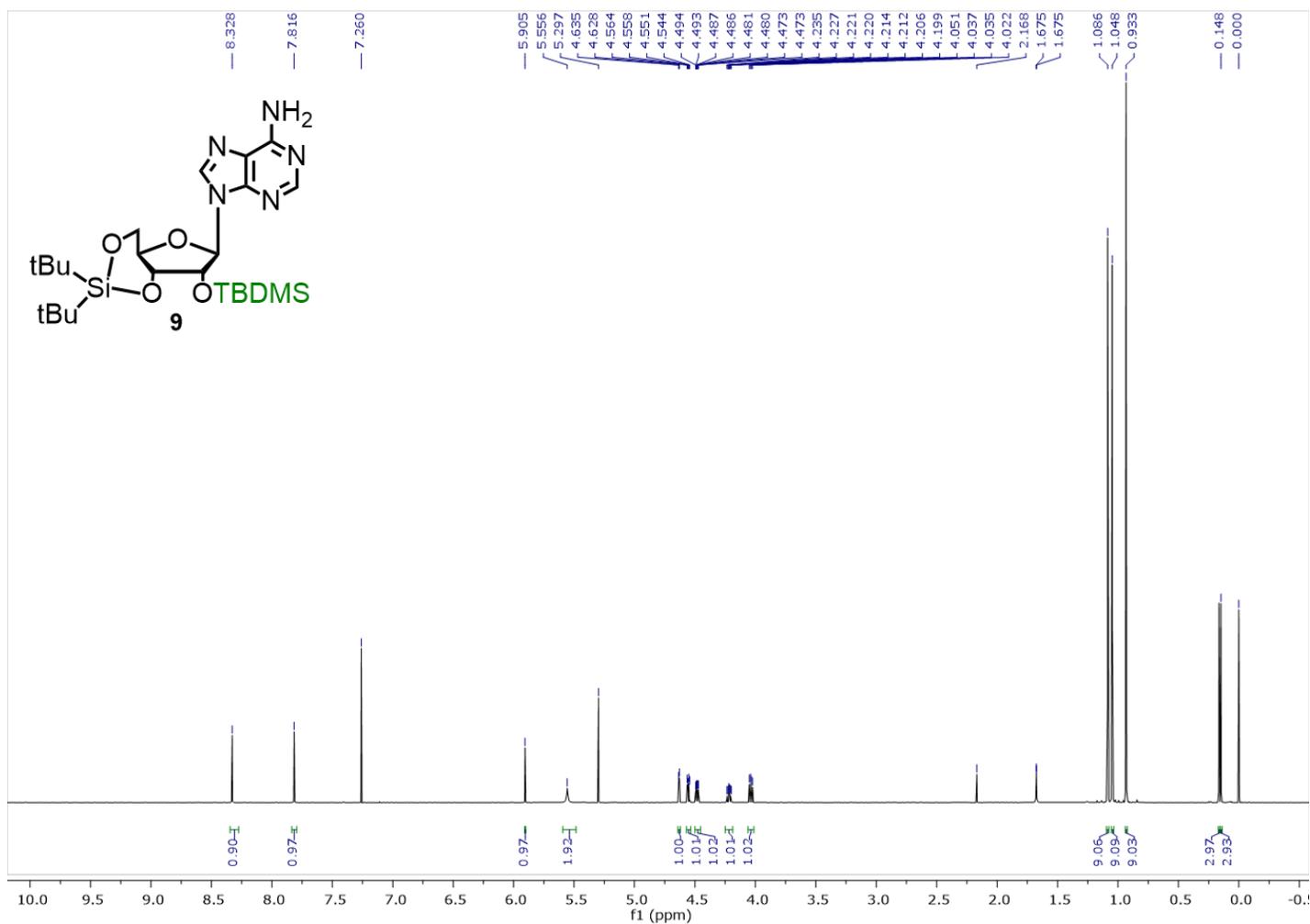


Figure S19 ^1H NMR spectrum of **9**



^1H NMR (700 MHz, CDCl_3) δ : 8.33 (s, 1H H-8), 7.82 (s, 1H, H-2), 5.91 (br s, 1H, H-1'), 5.56 (br s, 2H, NH_2), 4.63 (br d, 1H, $^3J=4.7$ Hz, H-2'), 4.55 (dd, 1H, $^3J=9.7$ Hz, $^3J=4.7$ Hz, H-3'), 4.48 (dd, 1H, $^2J=9.3$ Hz, $^3J=5.2$ Hz, H-5'), 4.22 (ddd, 1H, $^3J=10.7$ Hz, $^3J=9.6$ Hz, $^3J=5.2$ Hz, H-4'), 4.04 (dd, 1H, $^3J=10.6$ Hz, $^2J=9.2$ Hz, H-5''), 1.09 (s, 9H, $\text{Si-C}(\text{CH}_3)_3$ tBu_2Si), 1.05 (s, 9H, $\text{Si-C}(\text{CH}_3)_3$ tBu_2Si), 0.93 (s, 9H, $\text{Si-C}(\text{CH}_3)_3$ TBDMS), 0.16 (s, 3H, Si-CH_3 TBDMS), 0.15 (s, 3H, Si-CH_3 TBDMS).

Figure S20 ¹H NMR spectrum of 10

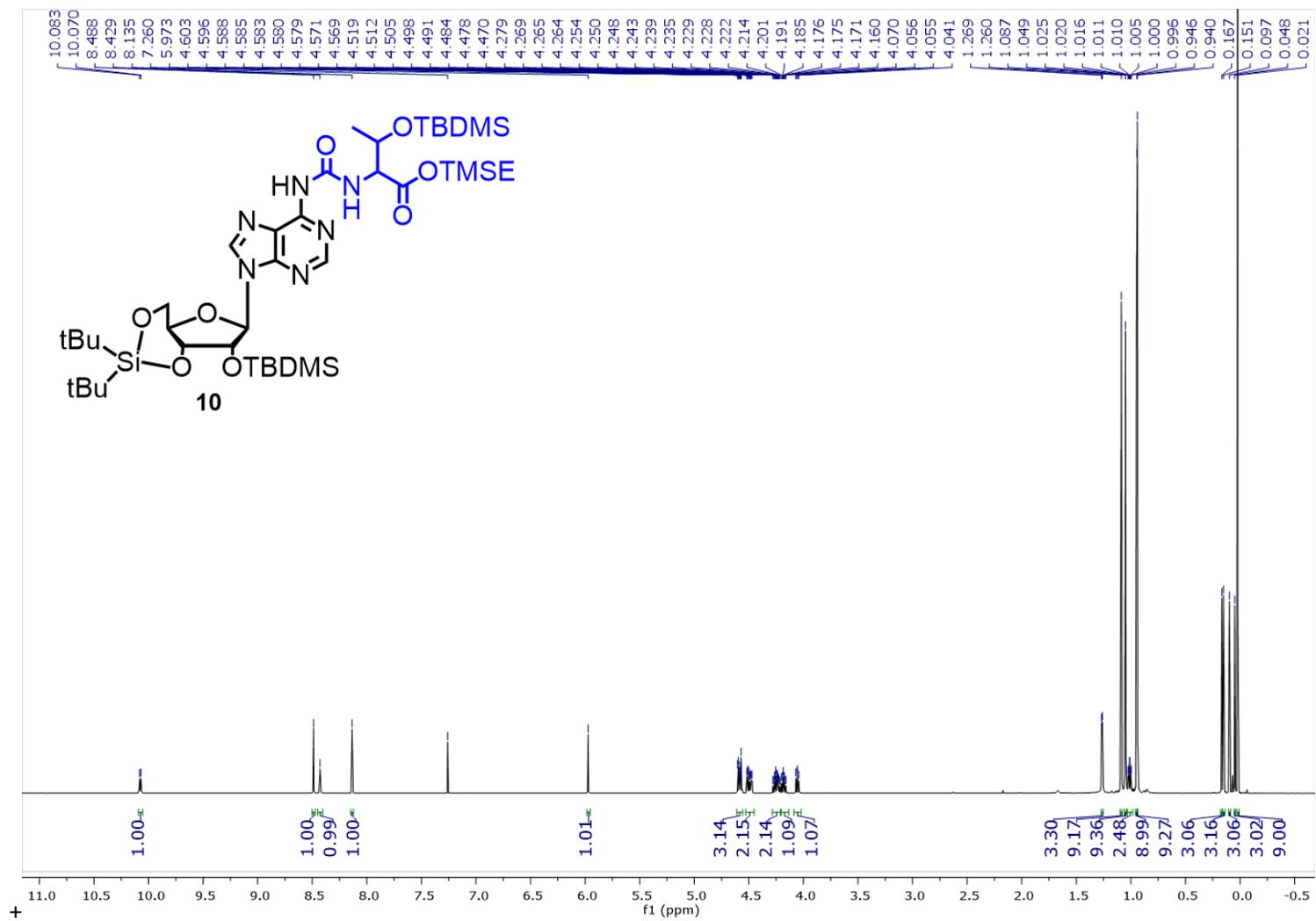


Figure S21 ¹³C NMR spectrum of **10**

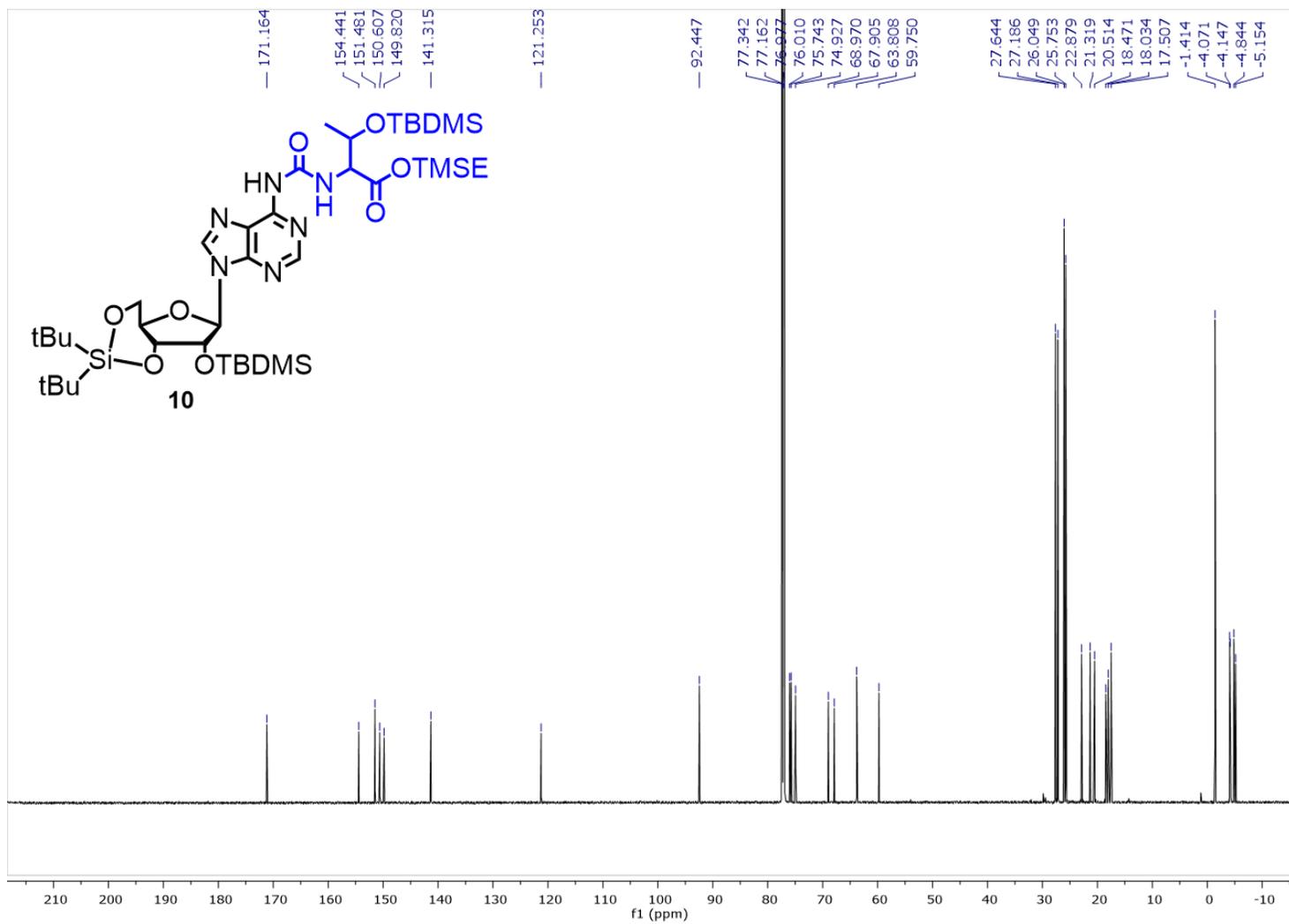


Figure S22 ¹H NMR spectrum of **11**

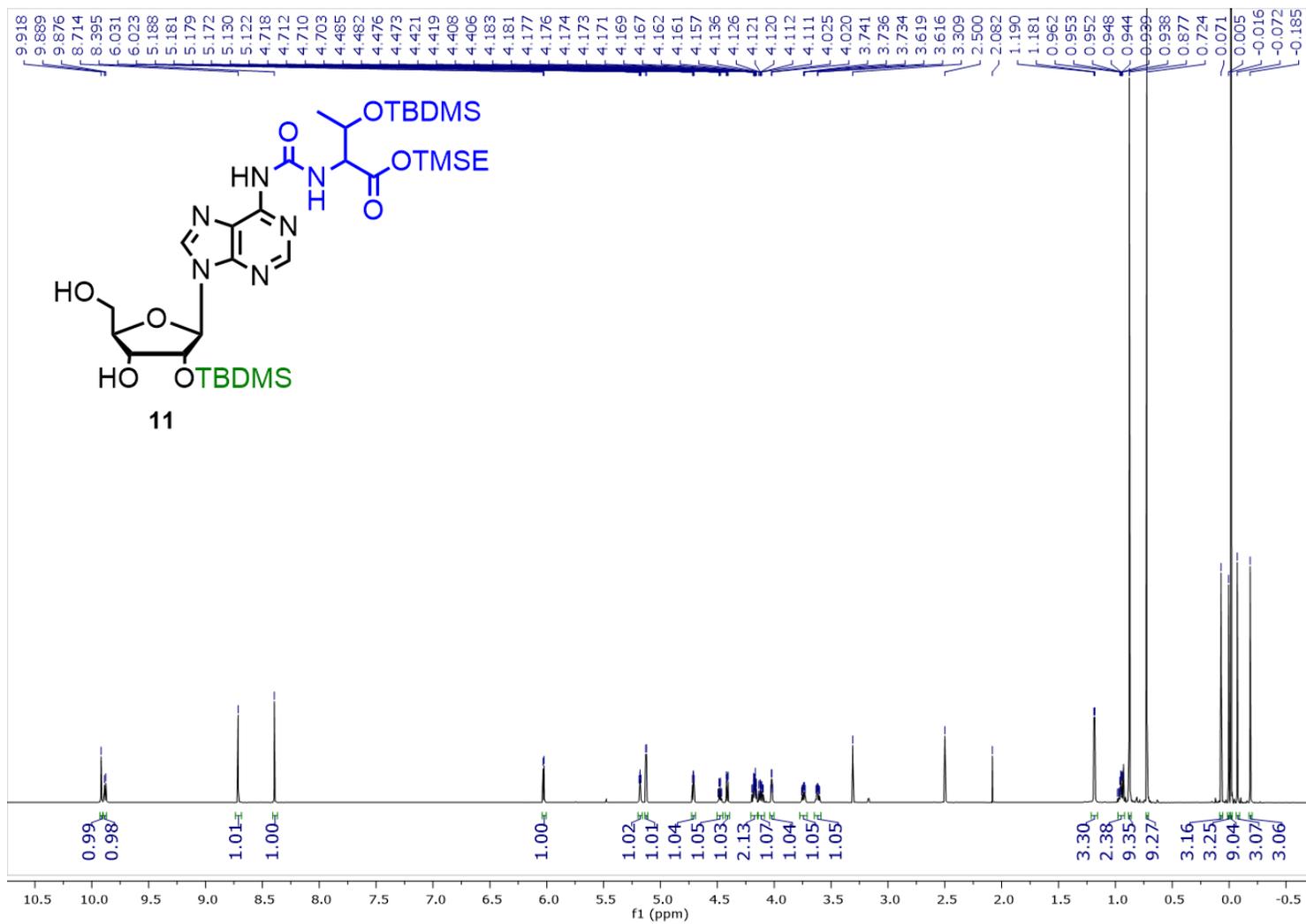


Figure S23 ^{13}C NMR spectrum of **11**

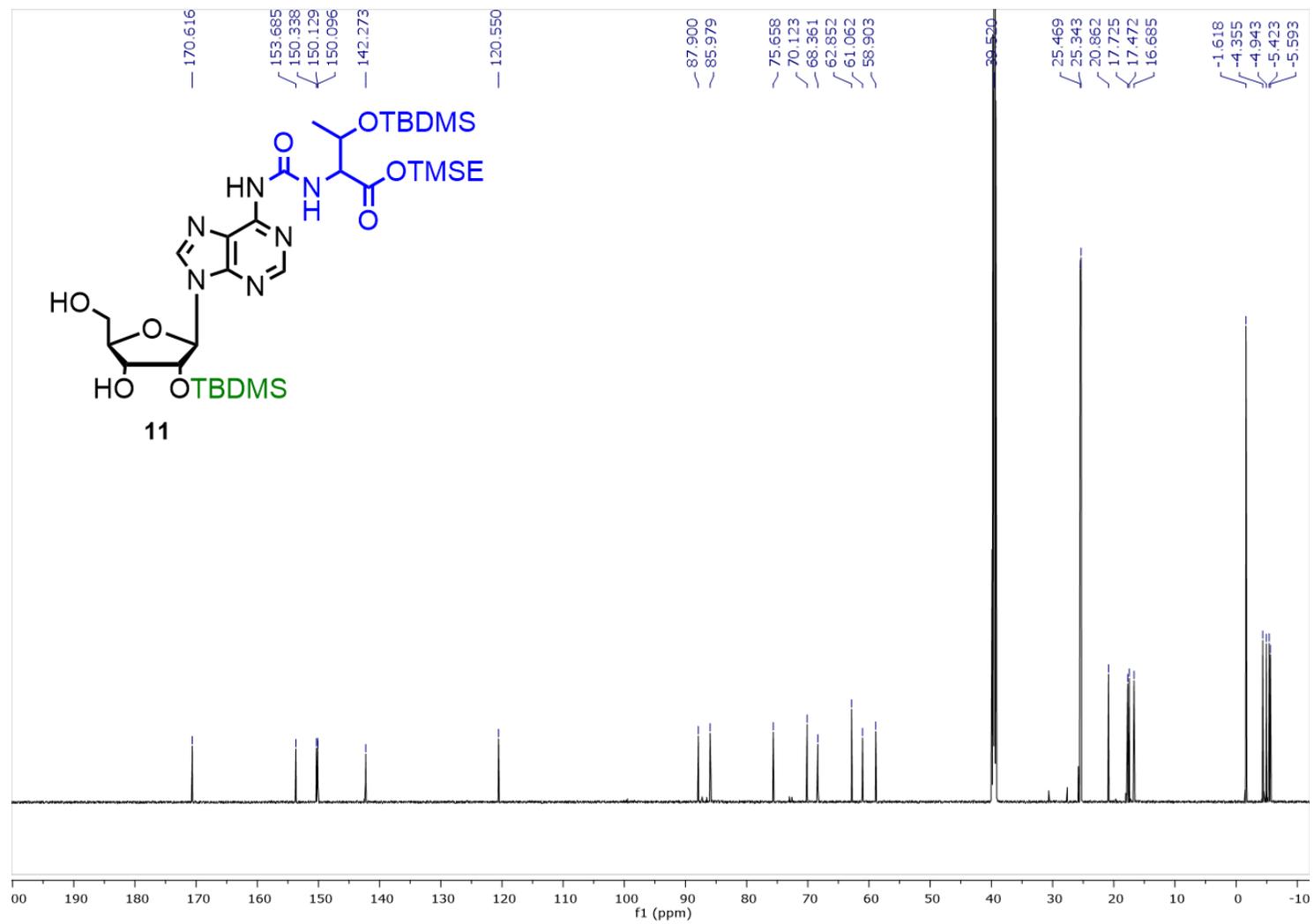


Figure S24 ¹H NMR spectrum of 12

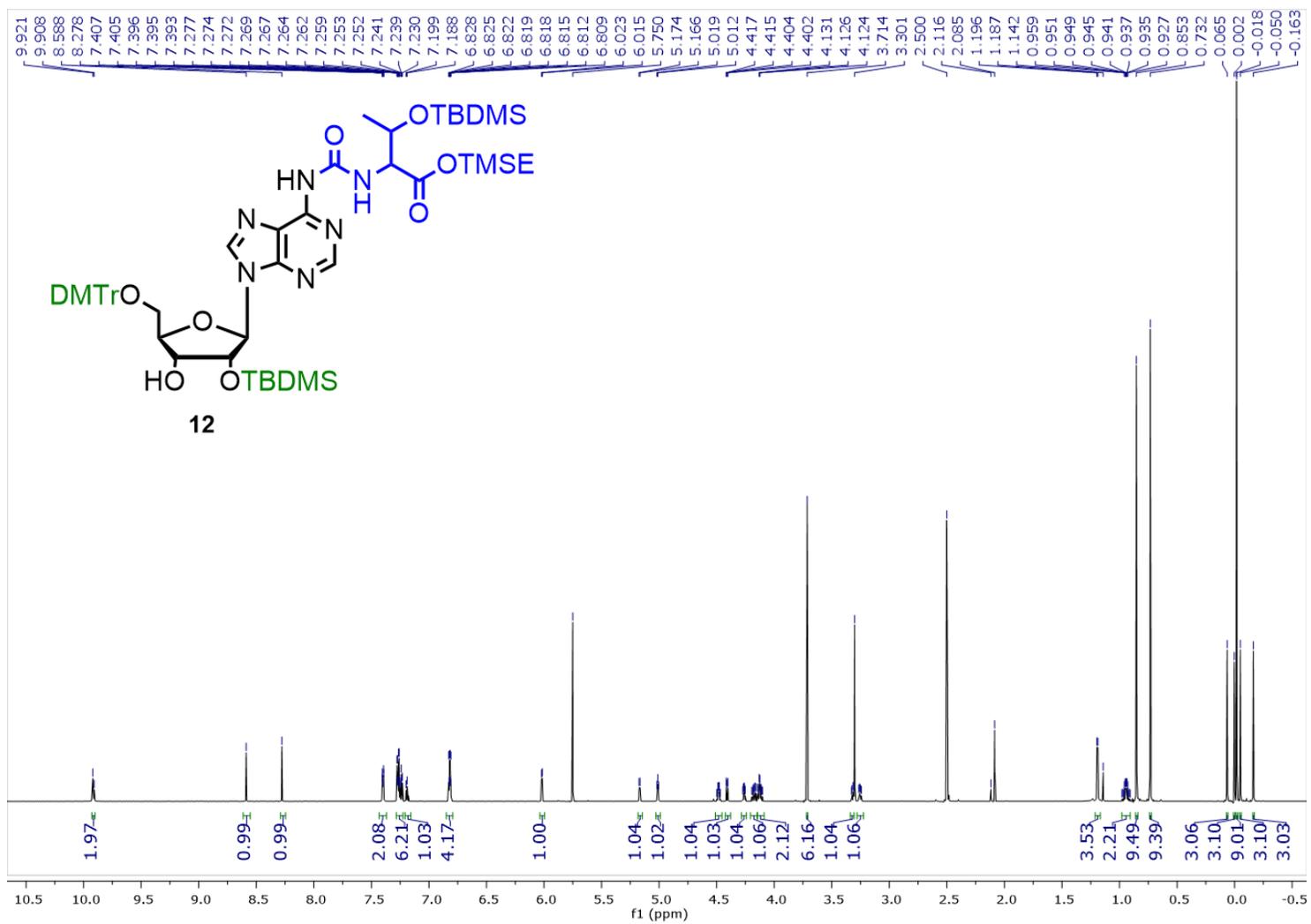
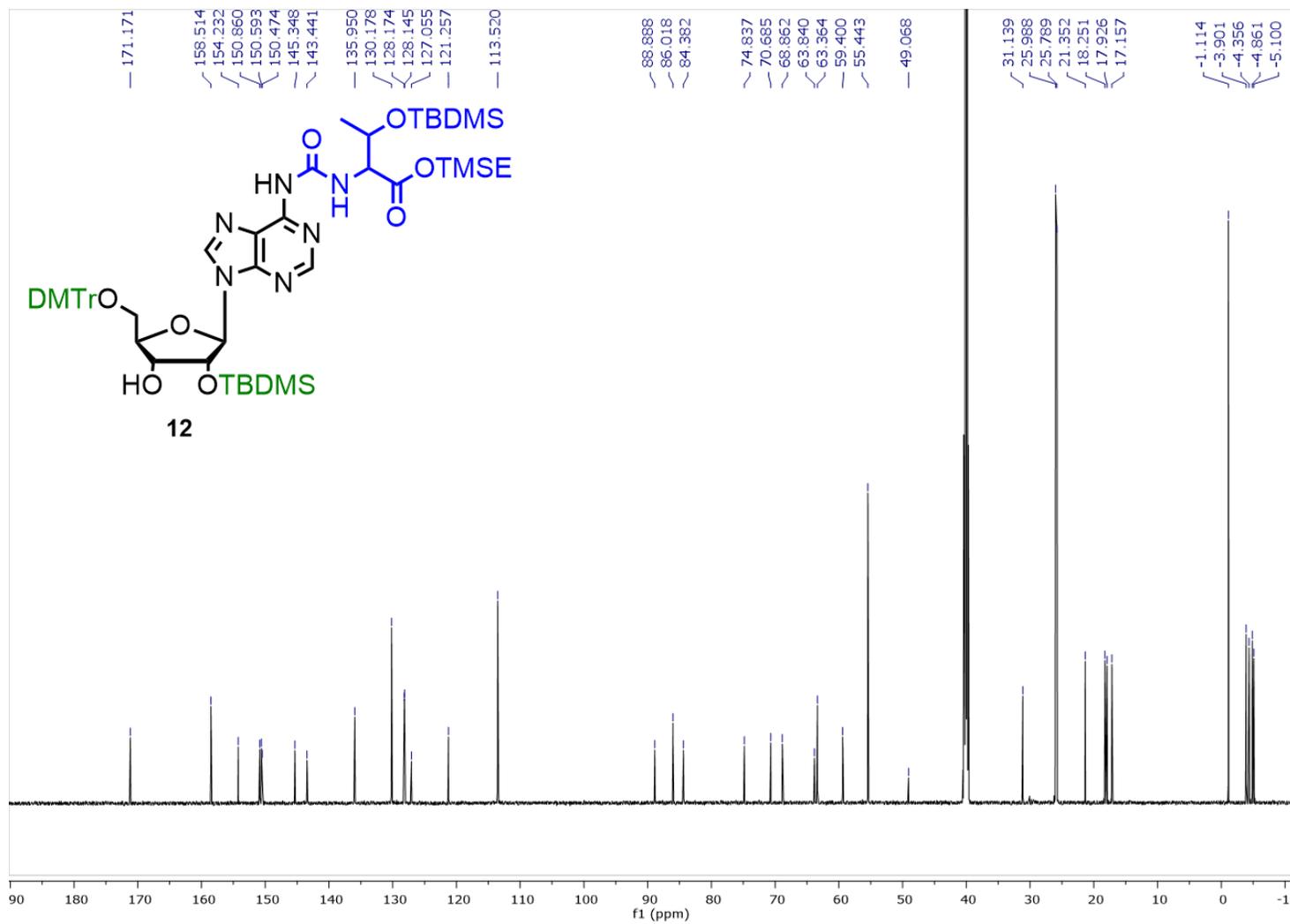


Figure S25 ¹³C NMR spectrum of 12



3.3 HR-MS spectra of adenosine derivatives.

Figure S26 ESI HR-MS spectrum of **4a**

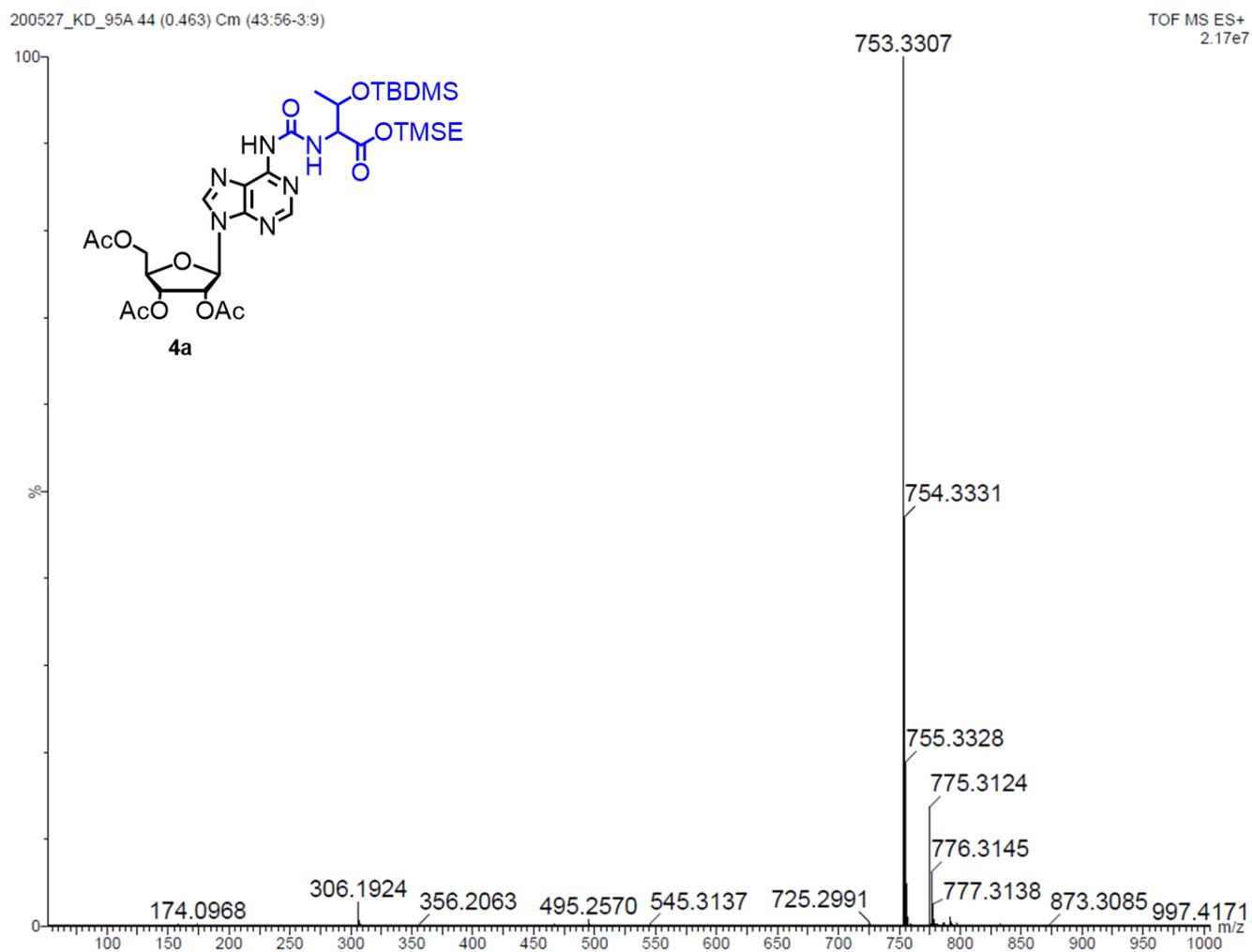


Figure S27 ESI HR-MS spectrum of **4b**

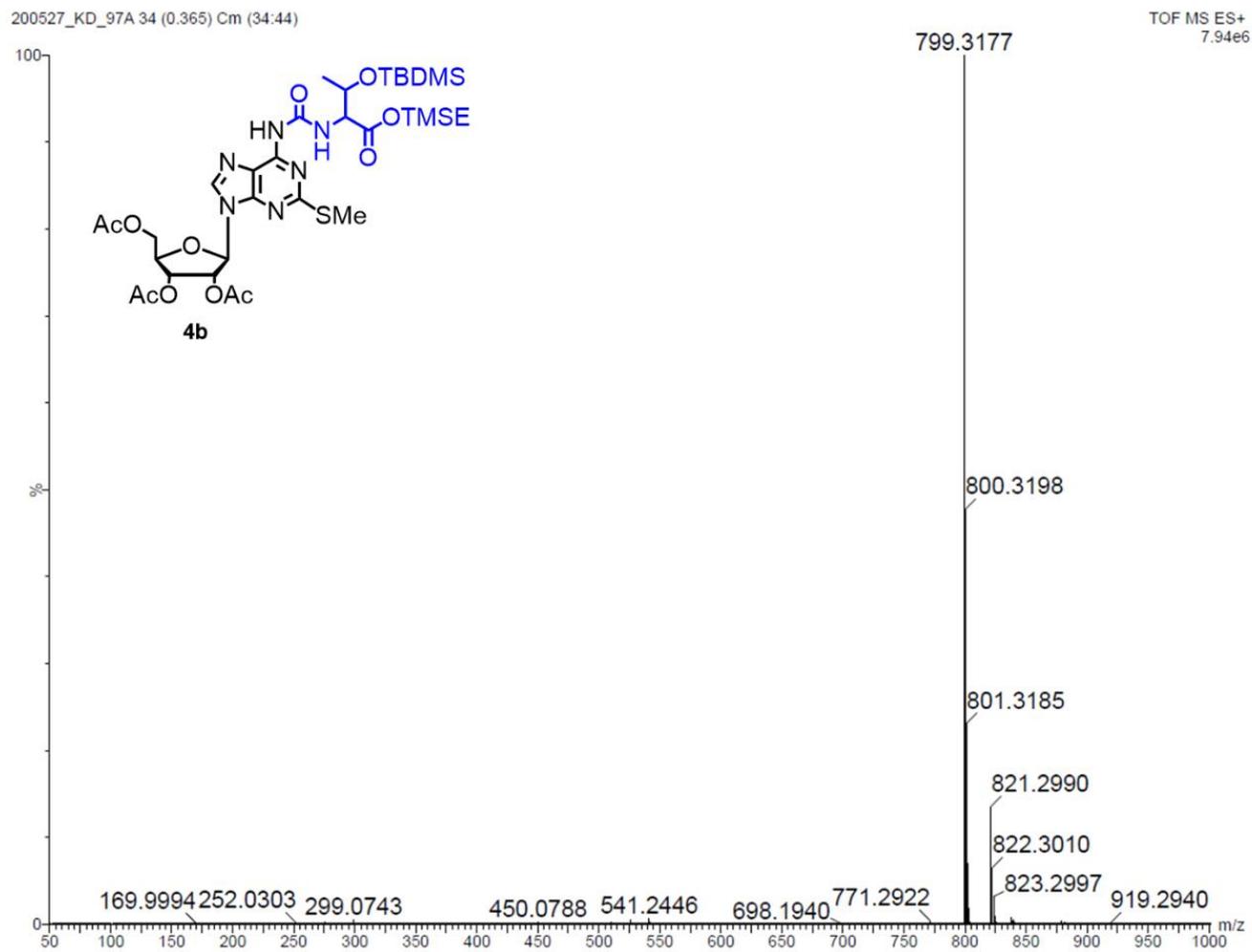


Figure S28 ESI HR-MS spectrum of **10**

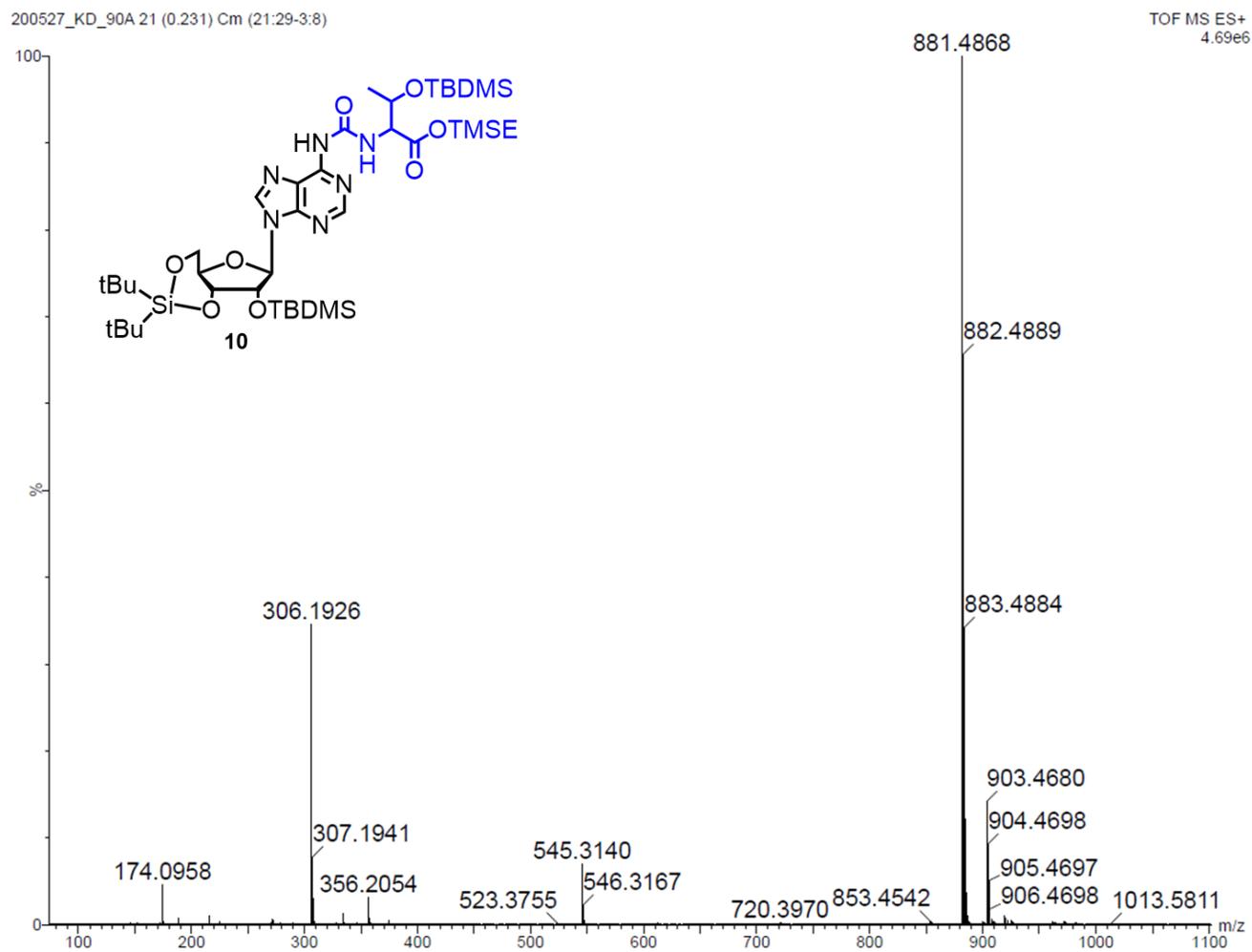


Figure S29 ESI HR-MS spectrum of **11**

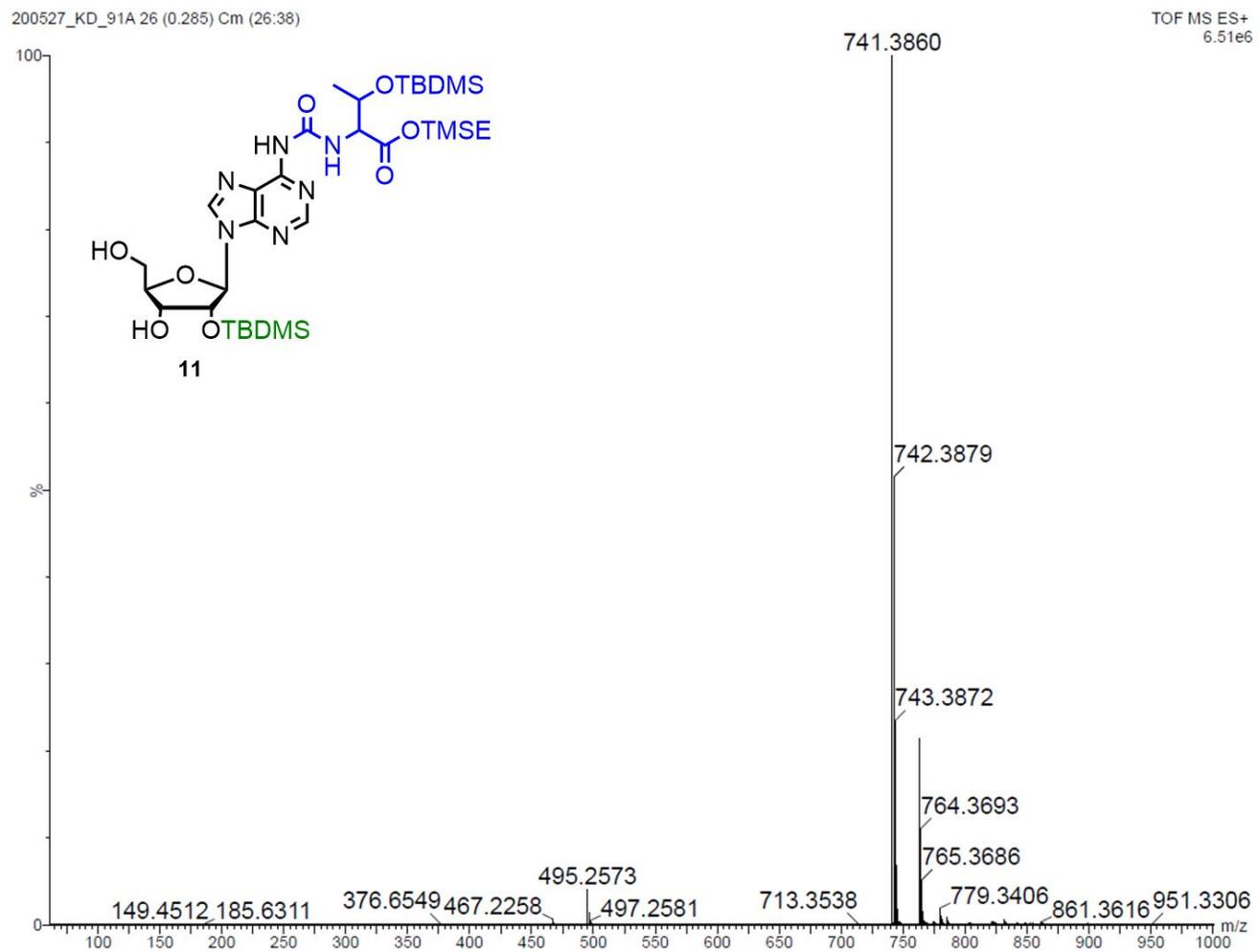


Figure S30 ESI HR-MS spectrum of **12**

