(Supporting Information)

GaN Nanowire as a Reusable Photoredox Catalyst for Radical Coupling of Carbonyl under Blacklight-Irradiation

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Materials and Methods

1. Growth of GaN Nanowires.

The nanowires are grown on a Si (111) wafer using radio frequency plasma-assisted molecular beam epitaxy (MBE) in nitrogen rich conditions. The Si substrates were cleaned in clean room in sequence by absolute methanol, acetone, and hydrofluoric acid prior of loading into the MBE system. Growth conditions: temperature ~750 °C, nitrogen flow rate 1 sccm, forward plasma power ~400 W. The typical growth time for each catalyst is three hours, giving the equivalence of 2 mg GaN NW on a 2-inches wafer. The as-synthesized nanowires can be doped with tetravalent (Si⁴⁺) or divalent (Mg²⁺) ions for making n- and p- type semiconductors, respectively. The doping density is controlled by tuning the effusion cell temperatures of Si and Mg. For n-type doping, the Si effusion cell temperature is 1100 °C. For p-type doping, the Mg effusion cell temperature is 265 °C. The electron and hole concentrations for the Si-doped n-type and Mg-doped p-type GaN NWs were estimated to be on the order of $n = 5 \times 10^{18}$ cm⁻³ and $p = 1 \times 10^{18}$ cm⁻³, respectively. Other growth parameters were kept constant.

2. Photo-driven pinacol coupling reaction.

A slice (3.5 cm²) of GaN NW (equivalent to 0.35 mg GaN) grown on Si(111) wafer was placed at the bottom of a glass flange equipped with a venting hose and a quartz window. The flange was then evacuated by oil pump for 30 min before the injection of ketone reactant (0.2 mmol) dissolved in 2 mL methanol. The flange was kept under a UV box equipped with HITACHI FL8BL-B black light bulb under room temperature for 12 h. After reaction, the crude reaction mixture can be characterized either by taking a drop of the methanol solution into NMR solvent for NMR study (solvent suppression mode can be applied to eliminate the huge methanol signal) or directly stripped of methanol to give the desired product, which can be further purified by flash chromatography to obtain a higher purity grade.

3. Identification of product.



¹H-NMR (CDCl₃, ppm): rac-, 7.27 (m, 6H), 7.24 (m, 4H), 1.54 (s, 6H); meso-, 7.27 (m, 6H),

7.24 (m, 4H), 1.62 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 143.4, 127.4, 127.3, 126.9, 78.6, 25.0; meso-, 143.8, 127.4, 127.3,

126.9, 78.9, 25.1

 $2b^{31}$ \xrightarrow{HO} OH (iso. yield > 99%, calc. based on ketone)

¹H-NMR (CDCl₃, ppm): 2.17 (br, 2H), 1.24 (s, 12H)

¹³C-NMR (CDCl₃, ppm): 75.1, 24.9



¹³C-NMR (CDCl₃, ppm): rac-, 140.3, 128.3, 127.1, 126.6, 81.9, 27.7, 7.5, 7.5; meso-, 141.3,

128.3, 127.1, 126.8, 81.9, 28.1, 7.5, 7.5



¹H-NMR (CDCl₃, ppm): rac-, 7.45 (m, 2H), 7.26-7.10 (m, 4H), 6.95 (m, 2H), 3.09 (br, 2H), 1.64(s, 6H); meso-, 7.45 (m, 2H), 7.26-7.10 (m, 4H), 6.95 (m, 2H), 3.18 (br, 2H), 1.76 (s, 6H) ¹³C-NMR (CDCl₃, ppm): rac-, 160.9 ($^{1}J_{C-F} = 201$ Hz), 130.2, 129.3, 123.6, 123.2, 116.0 ($^{2}J_{C-F} = 45$ Hz), 79.6, 24.0; meso-, 160.9 ($^{1}J_{C-F} = 201$ Hz), 130.2, 129.3, 123.6, 123.2, 116.0 ($^{2}J_{C-F} = 45$ Hz), 79.7, 24.5



2e³⁴

F(iso. yield 98%, calc. based on ketone; rac- : meso- = 1:1)

¹H-NMR (CDCl₃, ppm): rac-, 7.20 (m, 2H), 7.14 (m, 2H), 6.94 (m, 4H), 1.52 (s, 6H); meso-,

7.20 (m, 2H), 7.14 (m, 2H), 6.94 (m, 4H), 1.60 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 162.0 (¹J_{C-F} = 245 Hz), 139.1, 128.8 (²J_{C-F} = 38 Hz), 114.0, 78.3, 24.9; meso-, 162.0 (¹J_{C-F} = 245 Hz), 139.5, 128.8 (²J_{C-F} = 38 Hz), 114.0, 78.6, 25.2



Cl(iso. yield 93%, calc. based on ketone; rac- : meso- = 1:1.1)

¹H-NMR (CDCl₃, ppm): rac-, 7.23-7.08 (m, 8H), 1.47 (s, 6H); meso-, 7.23-7.08 (m, 8H), 1.55 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 141.7, 133.0, 128.8, 127.3, 78.2, 24.8; meso-, 141.7, 133.0, 128.8, 127.3, 78.2, 25.1



 CF_3 (iso. yield 99%, calc. based on ketone; rac- : meso- = 1.2:1)

¹H-NMR (CDCl₃, ppm): rac-, 7.50 (m, 6H), 7.29 (m, 2H), 1.53 (s, 6H); meso-, 7.50 (m, 6H), 7.29 (m, 2H), 1.57 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 147.7, 147.1, 127.5, 127.4, 125.8 (¹J_{C-F} = 240 Hz), 78.2, 24.8; meso-, 147.7, 147.1, 127.5, 127.4, 125.8 (¹J_{C-F} = 240 Hz), 78.6, 25.2



CN(iso. yield 94%, calc. based on ketone; rac- : meso- = 1:1.3)

¹H-NMR (acetone-d6, ppm): rac-, 7.68 (m, 4H), 7.41 (m, 4H), 4.59 (br, 2H), 1.48 (s, 6H); meso-, 7.75 (m, 4H), 7.49 (m, 4H), 4.77 (br, 2H), 1.75 (s, 6H)

¹³C-NMR (DMSO-d6, ppm): rac-, 152.5, 131.0, 128.4, 119.6, 108.9, 77.4, 24.7; meso-, 152.5, 130.9, 129.1, 119.6, 109.3, 77.4, 24.7



¹H-NMR (CDCl₃, ppm): rac-, 7.48 (m, 2H), 7.33 (m, 8H), 3.51 (br, 2H), meso-, 7.48 (m, 2H),

7.33 (m, 8H), 3.58 (br, 2H)

¹³C-NMR (CDCl₃, ppm): rac-, 133.5, 129.0, 127.8, 127.1 (${}^{1}J_{C-F} = 201 \text{ Hz}$), 126.9, 80.2; meso-, 133.6, 129.0, 127.8, 127.1 (${}^{1}J_{C-F} = 201 \text{ Hz}$), 126.9, 80.4



¹H-NMR (CDCl₃, ppm): 7.28 (m, 8H), 7.18 (m, 12H), 3.02 (br, 2H)

¹³C-NMR (CDCl₃, ppm): 144.1, 128.6, 127.3, 126.9, 83.0



¹H-NMR (CDCl₃, ppm): 7.15 (m, 8H), 6.97 (m, 8H), 2.28 (s, 12H)

¹³C-NMR (CDCl₃, ppm): 141.5, 136.3, 128.5, 127.9, 82.8, 21.0



F(iso. yield 96%, calc. based on ketone)

¹H-NMR (CDCl₃, ppm): 7.25 (m, 8H), 6.90 (m, 8H), 2.88 (br, 2H)

 13 C-NMR (CDCl₃, ppm): 161.8 (1 J_{C-F} = 250 Hz), 139.7, 130.3, 114.3 (2 J_{C-F} = 21 Hz), 82.6



Cl(iso. yield 97%, calc. based on ketone)

¹H-NMR (CDCl₃, ppm): 7.19 (m, 16H), 2.84 (br, 2H)

¹³C-NMR (CDCl₃, ppm): 142.0, 133.5, 129.8, 127.7, 82.5



OPh (iso. yield > 99%, calc. based on ketone; rac- : meso- = 1.1:1)

¹H-NMR (CDCl₃, ppm): rac-, 7.36-6.92 (m, 18H), 1.55 (s, 6H); meso-, 7.36-6.92 (m, 18H), 1.64 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 157.1, 156.3, 138.3, 129.7, 128.4, 123.3, 118.9, 117.4, 78.7, 25.0;
meso-, 157.1, 156.3, 138.3, 129.7, 128.8, 123.3, 118.9, 117.4, 78.7, 25.0

HR-MS: ESI [C₂₈H₂₅O₄Na]⁺ calc.: 448.1651, found: 448.1610



OMe(iso. yield > 99%, calc. based on ketone; rac- : meso- = 1.3:1)

¹H-NMR (CDCl₃, ppm): rac-, 7.09 (m, 4H), 6.79 (m, 4H), 3.79 (s, 6H), 1.46 (s, 6H); meso-, 7.09 (m, 4H), 6.79 (m, 4H), 3.81 (s, 6H), 1.55 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 158.5, 135.6, 128.5, 112.4, 78.7, 55.1, 25.0; meso-, 158.5, 135.6, 128.5, 112.4, 78.7, 55.1, 25.0



OMe(iso. yield > 99%, calc. based on ketone; rac- : meso- = 1:1)

¹H-NMR (CDCl₃, ppm): rac-, 7.35 (t, 1H), 7.04 (t, 1H), 6.67 (m, 1H), 6.56 (m, 2H), 6.44 (m,

1H), 3.79 (s, 6H), 2.99 (br, 2H), 1.62 (s, 6H); meso-, 7.35 (t, 1H), 7.04 (t, 1H), 6.67 (m, 1H),

6.56 (m, 2H), 6.44 (m, 1H), 3.82 (s, 6H), 3.09 (br, 2H), 1.74 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 161.0 (¹J_{C-F} = 245 Hz), 160.0, 130.5, 122.0, 109.2, 101.8 (²J_{C-F} = 57 Hz), 79.4, 55.5, 24.1; meso-, 161.1 (¹J_{C-F} = 245 Hz), 160.1, 130.6, 122.2, 109.3, 101.8 (²J_{C-F} = 57 Hz), 79.4, 55.5, 24.5

HR-MS: ESI [C₁₈H₁₉O₄F₂Na]⁺ calc.: 360.1149, found: 360.1111



¹³C-NMR (CDCl₃, ppm, rac- and meso-): 139.8, 128.3, 128.1, 127.1, 78.1



(iso. yield 75%, calc. based on aldehyde; rac- : meso- = 1:1.3)

¹H-NMR (CDCl₃, ppm): rac-, 7.17-7.03 (m, 8H), 4.66 (s, 2H) 2.29 (s, 6H); meso-, 7.17-7.03 (m, 8H), 4.73 (s, 2H), 2.33 (s, 6H)

¹³C-NMR (CDCl₃, ppm): rac-, 137.8, 136.9, 128.9, 127.0, 78.0, 21.1; meso-, 137.8, 136.9, 128.9, 127.0, 78.7, 21.1



Cl(iso. yield 69%, calc. based on aldehyde; rac- : meso- = 1.2:1)

¹H-NMR (CDCl₃, ppm): rac-, 7.25-7.01 (m, 8H), 4.59 (s, 2H) 2.61 (br, 2H); meso-, 7.25-7.01 (m, 8H), 4.81 (s, 2H) 2.61 (br, 2H)

¹³C-NMR (CDCl₃, ppm, rac- and meso-): 137.9, 133.8, 128.4, 128.3, 78.5



OMe (iso. yield 61%, calc. based on aldehyde; rac- : meso- = 1:1)

¹H-NMR (CDCl₃, ppm, rac- and meso-): 7.20 (d, 4H), 6.84 (d, 4H), 4.72 (s, 2H), 3.79 (s, 6H), 2.14 (br, 2H)

¹³C-NMR (DMSO-d6, ppm, rac- and meso-): 158.4, 134.8, 128.8, 113.4, 76.8, 55.3



¹³C-NMR (CDCl₃, ppm): rac-, 158.2, 133.2, 133.1, 128.1, 117.7, 114.3, 78.8, 68.8; meso-, 158.2, 133.2, 133.1, 128.1, 117.7, 114.5, 77.8, 68.8



¹H-NMR (CDCl₃, ppm): 7.38-7.23 (m, 5H), 3.65-3.53 (m, 2H), 3.35 (br, 1H), 2.91 (br, 1H), 1.45 (s, 3H)

¹³C-NMR (CDCl₃, ppm): 145.1, 128.3, 127.5, 125.1, 74.9, 70.8, 25.9



¹H-NMR (CDCl₃, ppm): 7.42-7.24 (m, 10H), 4.16 (d, 2H), 3.22 (br, 1H), 1.94 (br, 1H)

¹³C-NMR (CDCl₃, ppm): 143.8, 128.4, 127.4, 126.4, 78.5, 69.4



¹H-NMR (CDCl₃, ppm): 7.26 (d, 2H), 6.87 (d, 2H), 4.75 (m, 1H), 3.78 (s, 3H), 3.62 (m, 2H), 2.80 (br, 1H), 2.42 (br, 1H)

¹³C-NMR (CDCl₃, ppm): 159.4, 132.6, 127.3, 113.9, 74.3, 68.0, 55.3



¹H-NMR (CDCl₃, ppm): 6.90-6.83 (m, 3H), 4.75 (m, 1H), 3.87 (s, 3H), 3.86 (s, 3H), 3.67 (m,

2H), 2.64 (br, 1H), 2.20 (br, 1H)

¹³C-NMR (CDCl₃, ppm): 148.8, 148.7, 133.11, 118.3, 111.1, 109.1, 74.4, 68.1, 55.9



(iso. yield 82%)

¹H-NMR (CDCl₃, ppm): 7.60 (d, 2H), 7.50 (d, 2H), 4.86 (m, 1H), 3.79-3.63 (m, 2H), 2.81 (br, 1H), 2.20 (br, 1H)

¹³C-NMR (CDCl₃, ppm): 144.4, 129.9 (${}^{2}J_{C-F} = 130 \text{ Hz}$), 126.4, 125.4, 124.1 (${}^{1}J_{C-F} = 272 \text{ Hz}$)

74.0, 67.9



¹H-NMR (CDCl₃, ppm): 7.38 (d, 2H), 7.28 (d, 2H), 4.80 (m, 1H), 3.69 (m, 2H), 2.58 (br, 1H), 2.21 (br, 1H), 1.30 (s, 9H) ¹³C-NMR (CDCl₃, ppm): 151.0, 137.5, 125.8, 125.5, 74.5, 68.0, 34.5, 31.3



(iso. yield 89%)

¹H-NMR (CDCl₃, ppm): 7.30 (m, 2H), 6.92 (m, 3H), 4.01 (m, 2H), 3.80 (m, 1H), 3.68-3.50 (m, 2H), 3.05 (br, 1H), 2.76 (br, 1H), 1.99-1.88 (m, 2H), 1.62 (m, 2H)

¹³C-NMR (CDCl₃, ppm): 158.8, 129.5, 120.8, 114.5, 72.0, 67.7, 66.8, 29.9, 25.5



¹H-NMR (CDCl₃, ppm): 7.18 (d, 1H), 6.71 (dd, J = 8.5, 2.1 Hz, 1H), 6.62 (s, 1H), 3.79 (d, 1H), 3.76 (s, 3H), 3.61 (d, 1H), 2.84 (dd, J = 22.0, 11.3 Hz, 2H), 2.35-2.20 (m, 2H), 2.20 (s, 1H), 2.01-1.76 (m, 6H), 1.75-1.63 (m, 2H), 1.57-1.39 (m, 3H), 1.31 (dt, J = 17.5, 8.2 Hz, 1H), 0.78 (s, 3H)

¹³C-NMR (CDCl₃, ppm): 157.4, 138.0, 132.7, 126.3, 113.8, 111.5, 83.9, 66.9, 55.2, 49.6, 46.6,
43.6, 38.8, 34.6, 31.7, 29.9, 27.8, 26.2, 23.6, 15.1



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Figure S1. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2a



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| Owner | | mcgillnmr | Points Count | 32768 | Pulse Sequence | zgpg30 | Receiver Gain | 192.72 |
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Figure S2. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2b**



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Figure S3. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2c**



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Figure S4. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2d





Figure S5. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2e



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Figure S6. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2f



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Figure S7. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2g



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Figure S8. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2h**



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Figure S9. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2i



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Figure S10. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2j



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Figure S11. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2k





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| No. | (ppm) | (Hz) | Height | | | | | | | | | | | | | | | | | | | | | | |
| 1 | 82.58 | 10388.5 | 0.0561 | _ | | | | | | | | | | | | | | | | | | | | | |
| 2 | 114.24 | 14371.1 | 0.4347 | _ | | | | | | | | | | | | | | | | | | | | | |
| 3 | 114.40 | 14392.1 | 0.5676 | _ | | | | | | | | | | | | | | | | | | | | | |
| 4 | 130.32 | 16394.4 | 0.5076 | | | | | | | | | | | | | | | | | | | | | | |
| 5 | 139.66 | 17569.0 | 0.0651 | _ | | | | | | | | | | | | | | | | | | | | | |
| 6 | 160.83 | 20232.3 | 0.1396 | | | | | | | | | | | | | | | | | | | | | | |
| 7 | 162.79 | 20479.5 | 0.1236 | | | | | | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | | | | | | | | | |

Figure S12. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2**I





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Figure S13. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2m**





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| | | | | | | | | | | | | 5/8/2019 9: | 30:54 AM |
|--|------------------|------------------------------------|-------------------|---------------|----------|--|---|-----------------------------------|--------------|-------------------|--------------|-----------------|----------|
| Acquisition Time (| sec) 1.0 | 923 | Comment | | Li 1d (| C13 CDCI3 | E:\\ mingxin 8 | | | Date | 28 Ma | r 2019 03:13:04 | 1 |
| Date Stamp | 28 | Mar 2019 03:13:0 | 04 | | | | | | | | | | |
| File Name | C:\ | Users\Admin\One | Drive - McGi | II University | PostdocN | lcGill\Rese | arch\pp10\Pure NMR | h\pp10\Pure NMR\Imxpp10-scope-PhO | | fid | | | |
| Frequency (MHz) | 125 | 5.81 | Nucleus | | 13C | | Number of Trans | Number of Transients | | Origin | AVIII5 | AVIII500HD | |
| Original Points Co | ount 327 | 768 | Owner | | mcgillr | ımr | Points Count | | 32768 | Pulse Sequence | zgpg3 | 0 | |
| Receiver Gain | 192 | 2.72 | SW(cyclical) (Hz) | | 30000 | 00 | Solvent | | CHLOROFORM-d | Spectrum Offset (| 12578 | .9238 | |
| Spectrum Type | ST | ANDARD | Sweep Wid | lth (Hz) | 29999 | .08 | Temperature (de | gree C) | 24.999 | | | | |
| Spectrum Type 1.0 Imxxp10-s 0.9 0.9 0.8 0.7 Also 0.6 0.9 0.8 0.7 0.9 0.6 0.7 0.6 0.8 0.7 0.8 0.7 0.8 0.7 0.8 0.7 0.8 0.7 0.8 0.7 0.3 0.3 | ST. cope-PhOA | andard c ^{ph} viiteisc | Sweep Wid | e | 29999 | 128.41 128.79 20 20 20 20 20 20 20 20 20 20 20 20 20 | Temporature (de | gree C) | 24.999 | | 2 | | |
| 0.2 | | | | 156.30 | | | ette skriften en e | 78 73 | 2 | | | | |
| | | | | | | | | | | | | | |
| 220 2 | 210 200 | 190 180 | 170 160 |) 150 | 140 13 | 30 120 | 110 100 90 | 0 80 | 70 60 | 50 40 30 | 20 10 | 0 -10 | -20 |
| | | | | | | | Chemical Shift (pp | m) | | | | | |
| No. (ppm) | (Hz) | Height | | | | | | | | | | | |
| 1 25.03 | 3149.5 | 0.1419 | | | | | | | | | | | |
| 2 78.73 | 9905.1 | 0.0991 | | | | | | | | | | | |
| 3 117.36 | 14765.7 | 0.3276 | | | | | | | | | | | |
| 4 118.90 | 14958.8 | 0.5219 | | | | | | | | | | | |
| 6 129.44 | 10012.7 | 0.1788 | | | | | | | | | | | |
| 7 128.41 | 16204.0 | 0.2011 | | | | | | | | | | | |
| / 128.79 | 16204.0 | 0.3442 | | | | | | | | | | | |
| 0 129.73 | 17401 5 | 0.4353 | | | | | | | | | | | |
| 9 138.31 | 1/401.5 | 0.1210 | | | | | | | | | | | |
| 10 150.30 | 10765.4 | 0.0727 | | | | | | | | | | | |
| 11 157.10 | 19705.4 | 0.1297 | | | | | | | | | | | |

Figure S14. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2n**



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Figure S15. ¹H-(upper), and ¹³C-(lower)-NMR of compound **20**



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| | | | | | | | | | | | | | | | | | 5/8/ | 2019 9:3 | 31:32 AI |
|----------|----------------|------------------|----------------------|----------|------------------|--------------------|----------------|--|------------------|---|----------|------------|-------------|-----------|-------------------------|--------|----------|----------|----------|
| Acquisit | ion Time (| (sec) 1.0 |)923 | | Comment | | Li 1d_0 | C13 CDCI | 3 E:\\ ming | gxin 8 | | | | Date | | 13 Ap | r 2019 1 | 4:42:08 | |
| Date Sta | amp | 13 | Apr 2019 | 14:42:08 | 8 | | | | | | | | | | | | | | |
| File Nar | ne | C: | Users\Adm | nin\One | Drive - McGi | II University | /\PostdocM | lcGill\Res | earch\pp1 | 0\Pure NMR | Nmxpp10 |)-scope-FC | MeAcP 2 | 2\2\fid | | | | | |
| Frequer | ncy (MHz) | 12 | 5.81 | | Nucleus | | 13C | | Num | ber of Tran | sients | 3400 | | Origin | | AVII15 | 00HD | | |
| Original | Points Co | ount 32 | 768 | | Owner | | mcgilln | ımr | Poin | ts Count | | 32768 | | Pulse Seq | uence | zgpg3 | 0 | | |
| Receive | r Gain | 19 | 2.72 | | SW(cyclica | l) (Hz) | 30000. | 30000.00 | | ent | | CHLORO | FORM-d | Spectrum | Offset (Hz) | 12578 | .9238 | | |
| Spectru | т Туре | ST | ANDARD | | Sweep Wid | th (Hz) | 29999. | 08 | Tem | perature (de | egree C) | 25.000 | | | | | | | |
| 1.0 | mxpp10-s | 210 200 | eAc R & P | 2419ca | 170 16C | - 1 | 2000 140 13 | 00 130.47 05 172.17 172.08 | | 20 20 20 20 20 20 20 20 20 20 20 20 20 2 | 08 08 | 70 | 60 55.48 | 50 40 | 20 20 30 20 30 20 | 10 | 0 | -10 | -20 |
| | | | | | | | | | Chemi | cal Shift (pp | m) | | | | | | | | |
| No. | (ppm) | (Hz) | Height | No. | (ppm) | (Hz) | Height | No. | (ppm) | (Hz) | Heigh | nt No. | (ppm |) (Hz) | Height | | | | |
| 1 | 24.08 | 3029.5 | 0.0557 | 5 | 101.50 | 12769.8 | 0.1255 | 9 | 122.17 | 15370.8 | 0.041 | 0 13 | 160.1 | 5 20149.0 | 0.0571 | | | | |
| | 21.00 | | | | | | | | | - | | | | | | 1 | | | |
| 2 | 24.47 | 3079.0 | 0.0558 | 6 | 102.01 | 12834.8 | 0.1352 | 10 | 130.47 | 16414.5 | 0.112 | 1 14 | 161.9 | 1 20370.5 | 0.0447 | | | | |
| 2 3 | 24.47 55.48 | 3079.0 6980.0 | 0.0558 | 6 | 102.01 109.33 | 12834.8 13754.9 | 0.1352 | 10 | 130.47 130.62 | 16414.5 16433.8 | 0.112 | 1 14 1 | 161.9 | 1 20370.5 | 0.0447 | | | | |

Figure S16. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2p**



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| | | | | | | | 2019/10/9 20:03: | |
|---|--|--------------------------|---------------------|------------------------|--------------|----------------------|----------------------|--|
| Acquisition Time (sec) | 1.0923 | Comment | Li 1d C13 CDCI3 E:\ | \ mingxin 1 | | Date | 15 Apr 2019 23:52:32 | |
| Date Stamp | 15 Apr 2019 23:52:3 | 2 | | | | | | |
| File Name | C:\Users\Admin\One | Drive - McGill Universit | -scope-hydrobenzoin | pin 2\2\fid | | | | |
| Frequency (MHz) | 125.81 | Nucleus | 13C | Number of Transients | 3400 | Origin | AVIII500HD | |
| Original Points Count | 32768 | Owner | mcgillnmr | Points Count | 32768 | Pulse Sequence | zgpg30 | |
| Receiver Gain | 192.72 | SW(cyclical) (Hz) | 30000.00 | Solvent | CHLOROFORM-d | Spectrum Offset (Hz) | 12578.9238 | |
| Spectrum Type | STANDARD | Sweep Width (Hz) | 29999.08 | Temperature (degree C) | 24.998 | | | |
| I .0 0.9 0.8 0.7 0.8 0.7 0.6 0.5 0.5 0.5 0.5 0.5 0.7 0.5 0.5 0.5 0.7 0.2 0.1 0.7 0.2 0.2 0.7 0.7 0.7 0.7 0.7 0.7 0.7 0.7 0.7 0.7 | hydroben ჯაც ჩტმცხ ე 200 190 180 | ∦eFactor = 1 | | 2 8 10 10 90 80 | 70 60 5 | 0 40 30 20 | 10 0 -10 -20 | |
| | | | C | hemical Shift (ppm) | | | | |
| No. (ppm) (H | z) Height | | | | | | | |
| 1 78.12 982 | 9.1 0.1146 | | | | | | | |
| 2 127.09 1598 | 9.7 0.9048 | | | | | | | |
| 3 128.13 1612 | 0.6 0.4205 | | | | | | | |
| 4 128.26 1613 | 6.2 1.0000 | | | | | | | |
| 5 139 77 1758 | 4.6 0.0625 | | | | | | | |

Figure S17. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2q**



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Figure S18. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2r**



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Figure S19. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2s



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Acquisition Time (sec) Imxpp10-oct2019-OMeAI-PCR-500 Oct 9 2019 1.0486 Comment Date C:\Users\Admin\Desktop\Imxpp10-oct2019-OMeAI-PCR-500_CARBON_01.fid\fid Date Stamp Oct 9 2019 File Name Frequency (MHz) 125.71 Nucleus Number of Transients 3500 Original Points Count 32768 13C Points Count 32768 Pulse Sequence s2pul **Receiver Gain** 30.00 Solvent DMSO-d6 Spectrum Offset (Hz) 13826.9600 Spectrum Type STANDARD Sweep Width (Hz) Temperature (degree C) 25.000 31250.0 Imxpp10-oct2019-OMeA₩PerRcanscatePatton = 1 0.13 0.12 13 38 128.83 0.11 0.10 55.34 0.09 158.42 0.08 76.82 0.07 hariler 134.75 0.06 0.05 0.04 0.03 0.02 0.01 -20 220 200 120 100 Chemical Shift (ppm) 40 20 6 180 160 140 80 60 No. (ppm) (Hz) Height
 55.34
 6957.0

 76.82
 9656.9

 113.38
 14253.7
 1 2 6957.00.08369656.90.0654 3 0.0982 128.83 16195.5 0.0964
 5
 134.75
 16940.3
 0.0473

 6
 158.42
 19914.9
 0.0662

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Figure S20. ¹H-(upper), and ¹³C-(lower)-NMR of compound 2t







Figure S21. ¹H-(upper), and ¹³C-(lower)-NMR of compound **2u**

10 158.17 11918.3 0.0516



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Figure S22. ¹H-(upper), and ¹³C-(lower)-NMR of compound 5a



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Figure S23. ¹H-(upper), and ¹³C-(lower)-NMR of compound **5b**



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Figure S24. ¹H-(upper), and ¹³C-(lower)-NMR of compound 5c



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Figure S25. ¹H-(upper), and ¹³C-(lower)-NMR of compound 5d

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Figure S26. ¹H-(upper), and ¹³C-(lower)-NMR of compound 5e

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Figure S27. ¹H-(upper), and ¹³C-(lower)-NMR of compound **5**f

Chemical Shift (ppm)

Ó -10 -20

Figure S28. ¹H-(upper), and ¹³C-(lower)-NMR of compound 5g

71.96

25.52 29.89

No. (ppm)

210 200

66.79 8403.7 0.2177

67.73 8520.8 0.3890

 120.80
 15197.8
 0.4356

 129.49
 16291.8
 0.5354

 158.78
 19976.8
 0.0581

(Hz) Height

3210.8 0.1684 3761.0 0.4193

9053.7 0.3430 114.49 14404.9 1.0000

This report was created by ACD/NMR Processor Academic Edition. For more information go to www.acdlabs.com/nmrproc/ 2019/10/4 11:32:55

Figure S29. ¹H-(upper), and ¹³C-(lower)-NMR of compound **5h**

Figure S30. HR-ESI spectrum of the reaction mixture of Table 4, entry 3.

Figure S31. Photo-induced reaction setup.

Figure S32. EPR signal of p-GaN in dark.

Figure S33. UV-Vis absorption spectrum for GaN NW catalysts. The glitches at 350 nm on the spectra were system error caused by the spectrometer's lamp switching.

Figure S34. SEM (left) and TEM (right) of the GaN NW after 13 consecutive catalyses.

| Table S1. Band edge energy | of various | photosensitizing | semiconductors | and their PCR | reactivity |
|----------------------------|-------------|------------------|----------------|---------------|------------|
| Tuele STI Buna euge energy | 01 (411040) | photosenshizing | Senneenaaetens | | 1000011109 |

| semiconductor ^a | band gap / eV | reference | PCR yield of 2a (same conditions as Table 1) / % |
|----------------------------|---------------|-----------|---|
| GaN (commercial powder) | 3.4 | (13) | 21 |
| TiO ₂ | 3.05 | (40) | 44 |
| CdS | 2.42 | (41) | 89 |
| C_3N_4 | 2.7 | (42) | 62 |
| ZnO | 3.3 | (43) | not detected |

^aAll reagents used were commercially available. Catalysts were introduced as 1 mg powder into 0.5 mmol MeOH.

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