Electronic Supporting Information (ESI) for:

Pd-catalyzed stereoselective tandem ring-opening amination/cyclization of vinyl γ-lactones: Access to caprolactam diversity

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Contents:

- Page S2: General comments
- Page S2: Preparation of γ -oxobutanoic acids
- Page S3: Procedure for the preparation of vinyl γ -lactones
- Page S4: Procedure for the preparation of phosphoramidite ligands
- Page S6:Table S1: Preliminary evaluation of ligands
- Page S7: Table S2: Other ligands used to optimize the reaction conditions
- Page S8: Table S3: Further optimization towards caprolactam **3**
- Page S9: Procedure for the screening phase in the preparation of caprolactam **3**
- Page S10: Typical procedure for the preparation of caprolactams
- Page S11: Characterization data for non-reported phosphoramidite ligands
- Page S32: IR, NMR spectra, HRMS data for vinyl γ-lactones
- Page S87: Characterization data for the amino acid intermediate *E*-1
- Page S90: IR, NMR spectra and HRMS data for caprolactam products
- Page S188: Product diversification based on compounds **3** and **29**
- Page S211: Control reactions
- Page S219: Characterization of byproduct 2 from bis-allylation of aniline
- Page S222: X-ray structure of 3 and 53
- Page S223: References

S2. General comments

The amine reagents and solvents were purchased from Aldrich or TCI, and used without further purification. Phosphoramidite ligands L1,^{1a} L2,^{1b} L3,^{1c} L4-L5,^{1d} L6,^{1c} L7-L12,^{1d} L13^{1a} and L14-L19^{1d} were prepared according to previously reported protocols. Other ligands and palladium precursors were purchased from Aldrich or TCI. ¹H NMR, ¹³C{¹H} NMR, ³¹P{¹H} NMR and ¹⁹F{¹H} NMR spectra were recorded at room temperature on a Bruker AV-400 or AV-500 spectrometer and referenced to the residual deuterated solvent signals. All reported NMR values are given in parts per million (ppm). FT-IR measurements were carried out on a Bruker Optics FTIR Alpha spectrometer. Mass spectrometric analyses and X-ray diffraction studies were performed by the Research Support Group at ICIQ.

S2. Preparation of γ-oxobutanoic acids

The γ-oxobutanoic acids were synthesized following reported procedures if not commercially available. **H1-H9** were purchased from Aldrich or TCI, **H10-H14**,^{2a} **H15-H18**,^{2b} **H19**,^{2c} **H20**,^{2d} **H21**,^{2e} **H22**,^{2f} **H23**,^{2g} **H24**^{2h} and **H25**²ⁱ were prepared following previously reported procedures.



S3. Procedure for the preparation of vinyl γ -lactones

General Procedure A:



Under a N₂ atmosphere, to a separate flame-dried round-bottom flask equipped with a stirring bar was added the respective γ -oxobutanoic acid (10.0 mmol) and anhydrous THF (30 mL). The solution was cooled down to 0 °C (ice/water), followed by dropwise addition of vinyl magnesium bromide in THF (1.0 M, 30.0 mL, 30.0 mmol). The reaction mixture was allowed to warm to room temperature and stirred for 16 h. The reaction mixture was quenched with saturated aqueous NH₄Cl, then treated with HCl (4 M) until the pH was 3. The organic components were extracted with EtOAc (3 × 20 mL). Hereafter, the combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure. The residue was purified by flash chromatography to afford the corresponding lactone.

General Procedure B:



According to a previously reported procedure,³ to a solution of γ -oxobutanoic acid (10.0 mmol, 1.0 equiv) and *tert*-butyl alcohol (20.0 mmol, 2.0 equiv) in DCM (20 mL), DMAP (3.0 mmol, 0.3 equiv) was added. The resultant solution was cooled down to 0 °C and *N*,*N'*-dicyclohexylcarbodiimide (DCC, 12.0 mmol, 1.2 equiv) was added. The reaction mixture was stirred at room temperature for 12 h. The urea byproduct was filtered off and the organic layer was concentrated under vacuum. The crude residue was purified by a rapid flash chromatographic purification to give the corresponding ester product.

Under a N₂ atmosphere, to a separate flame-dried round-bottom flask equipped with a stirring bar was added the respective γ -oxobutanoic ester (5.0 mmol) and anhydrous THF (15 mL). The solution was cooled down to 0 °C (ice/water), followed by dropwise addition of vinyl magnesium bromide in THF (1.0 M, 7.5 mL, 7.5 mmol). The reaction mixture was allowed to warm to room temperature and stirred for 16 h. The reaction mixture was quenched with saturated aqueous NH₄Cl. The organic components were extracted with EtOAc (3 × 20 mL), and the combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure. The residue was purified by flash chromatography to afford the corresponding lactone.

The desired starting materials could be prepared by methods A or B in 30-70% isolated yields without further optimization.

S4. Procedure for the preparation of phosphoramidite ligands

The non-reported ligands L8-L12, L16-L17, L19 were prepared according to a reported procedure with slight modifications.^{2d}



To an oven-dried round bottom flask, distilled PCl₃ (180.0 μ L, 2.06 mmol) was added to a solution of anhydrous Et₃N (1.68 mL, 12.06 mmol) in DCM (15 mL) at 0 °C and the mixture stirred for 0.5 h at this temperature. Then, the respective amine (2.0 mmol) was added dropwise at 0 °C, and the reaction mixture stirred for 4 h at room temperature. The resultant solution was cooled to 0 °C, and then [1,1-biphenyl]-2,2-diol (372.4 mg, 2.0 mmol) or (±)-1,1'-binaphthalene-2,2'-diol (572.7 mg, 2.0 mmol) was added. The mixture was stirred for 16 h at room temperature, diluted with water and extracted with DCM (3 × 20 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure. The residue was purified by flash chromatography to afford the respective phosphoramidite ligand. All purified ligands were fully characterized by NMR (¹H, ¹³C, ³¹P), IR and HRMS. *[Note that, in some cases, it proved to be crucial to isolate the pure ligand by chromatography under a N₂ atmosphere]*

Dicyclopentylamine and dicycloheptylamine, being the starting materials of L9-L10 and L16-L17, were prepared following reported procedures.⁴



To a stirred solution of the cyclopentanone (420.6 mg, 5.0 mmol) and cyclopentylamine (425.8 mg, 5.0 mmol) in DCM (15 mL) were added sodium triacetoxy borohydride (1.4836 g, 7.0 mmol) and acetic acid (300.0 mg, 5.0 mmol). The reaction mixture was stirred for 12 h at room temperature, and hereafter, 1 N aqueous NaOH was added. The resultant mixture was extracted with ester (3×20 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure. The crude product could be directly used without further purification.



To a stirred solution of cycloheptanone (1.0 g, 8.9 mmol) and NH₄OAc (6.63 g, 86.0 mmol) in DCE (30 mL) were added sodium triacetoxy borohydride (2.65 g, 12.5 mmol) and Et₃N (2.5 mL, 17.9 mmol). The reaction mixture was stirred for 48 h at room temperature, followed by addition of saturated aqueous NaHCO₃. The resultant mixture was extracted with EtOAc (3×20 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure. The residue was purified by flash chromatography to afford the desired amine as a yellow liquid.

S6. Table S1: Preliminary evaluation of ligands



Reaction conditions: A (0.20 mmol), aniline (0.30 mmol, 1.5 equiv.), DCM (0.20 mL), $Pd_2(dba)_3 \cdot CHCl_3$ (2.0 mol%), monodentate ligand (8.0 mol%) or bidentate ligand (4.0 mol%), rt, 12 h; the yields and *E/Z* ratios were determined by ¹H NMR analysis of the crude reaction mixture, using CH₂Br₂ (1.0 equiv.) as an internal standard.



S7. Table S2: Other ligands used to optimize the reaction conditions

Reaction conditons: A (0.20 mmol), aniline (0.30 mmol, 1.5 equiv.), DCM (0.20 mL), $Pd_2(dba)_3 \cdot CHCl_3$ (2.0 mol%), ligand (8.0 mol%), rt, 12 h, then EDC (0.30 mmol), 1 h; the yields and *E/Z* ratios were determined by ¹H NMR analysis of the crude reaction mixture, using CH_2Br_2 (1.0 equiv.) as an internal standard.

S8. Table S3: Further optimization towards caprolactam 3



Entry ^[a]	Ligand	Solvent	<i>E:Z</i> -1 ^[c]	1/2 ^[c]	Yield of 3 [%] ^[b]
1	L8	DCM	98:2	>20:1	84
2	L8	DCE	-	-	0
3	L8	CHCl ₃	98:2	12:1	70
4	L8	MeOH	94:6	7:1	62
5	L8	EtOH	90:10	>20:1	76
6	L8	<i>i</i> -PrOH	96:4	7:1	70
7	L8	<i>t</i> -BuOH	98:2	7:1	68
8	L8	HFIP	92:8	7:1	52
9	L8	EtOH/DCM (1:1)	95:5	>20:1	84
10	L8	EtOH/DCM (2:1)	94:6	18:1	78

[a] Reaction conditons: A (0.20 mmol), aniline (0.40 mmol, 2.0 equiv.), solvent (0.30 mL), $Pd_2(dba)_3 \cdot CHCl_3$ (3.0 mol%), L8 (12.0 mol%), rt, 12 h, then EDC (0.30 mmol), 1 h; [b] Determined by ¹H NMR analysis in CDCl₃ using CH_2Br_2 as an internal standard. [c] Determined by ¹H NMR analysis.

S9. Procedure for the screening phase in the preparation of caprolactam **3**



Vinyl γ -lactone **A** (37.6 mg, 0.20 mmol, 1.0 equiv) was combined with Pd₂(dba)₃·CHCl₃, the ligand, aniline and the solvent at room temperature under air. The internal standard CH₂Br₂ (1.0 equiv.) was added after the reaction mixture had been stirred at room temperature for 12 h, and then an aliquot of the mixture was taken for analysis allowing to determine the NMR yield of the amino acid intermediate, the *Z/E* ratio and the ratio 1/2 using signal integration. Hereafter, EDC (57.5 mg, 3.0 mmol, 1.5 equiv.) was added and the reaction mixture stirred for another 1 h, after which the yield of the targeted product **3** was determined by ¹H NMR spectroscopy.

S10. Typical procedure for the preparation of caprolactams



Representative case:

Vinyl γ -lactone **A** (37.6 mg, 0.20 mmol, 1.0 equiv) was combined with Pd₂(dba)₃·CHCl₃ (6.0 mg, 3.0 mol%), **L8** (9.6 mg, 12.0 mol%) and aniline (38.0 mg, 0.40 mmol) in DCM (0.30 mL) at room temperature under air. The reaction mixture was stirred at room temperature for 12 h, then an aliquot of the mixture was taken for NMR analysis, which provided the unsaturated amino acid intermediate with an *E/Z* ratio of 98:2. Then, EDC (57.5 mg, 3.0 mmol) was added and the reaction mixture stirred for another 1 h. The desired product was isolated by flash chromatography (43.2 mg, 82%, Hexane/EtOAc = 2:1, $R_f = 0.25$). Note that all purified caprolactam products were fully characterized by NMR (¹H, ¹³C; ¹⁹F where appropriate), IR and HRMS.

S11. Characterization data for non-reported phosphoramidite ligands



White solid; Column conditions: Hexane : EA = 50 : 1, $R_f = 0.30$ (Note: EA stands for ethyl acetate)



¹H NMR (400 MHz, CDCl₃) δ 7.45 (dd, J = 7.6, 1.7 Hz, 2H), 7.32 (td, J = 7.7, 1.7 Hz, 2H), 7.24 – 7.13 (m, 4H), 3.01 – 2.90 (m, 2H), 1.81 (d, J = 12.4 Hz, 4H), 1.73 – 1.65 (m, 4H), 1.62 – 1.49 (m, 6H), 1.07 – 0.96 (m, 6H).



³¹P NMR (162 MHz, CDCl₃) δ 155.25.



HRMS (ESI⁺, MeOH): *m/z* calcd. 396.2087 (M + H)⁺, found: 396.2090.



White solid; Column conditions: Hexane : $EA = 50 : 1, R_f = 0.30$



4H), 3.66 – 3.48 (m, 2H), 1.85 – 1.61 (m, 12H), 1.48 – 1.34 (m, 4H).



³¹P NMR (162 MHz, CDCl₃) δ 154.97.



HRMS (ESI⁺, MeOH): *m/z* calcd. 368.1774 (M + H)⁺, found: 368.1778.



White solid; Column conditions: Hexane : EA = 50 : 1, $R_f = 0.30$



¹H NMR (400 MHz, CDCl₃) δ 7.45 (dd, J = 7.6, 1.8 Hz, 2H), 7.32 (td, J = 7.6, 1.7 Hz, 2H), 7.25 – 7.10 (m, 4H), 3.15 (qt, J = 11.1, 3.9 Hz, 2H), 1.98 – 1.87 (m, 4H), 1.83 – 1.68 (m, 4H), 1.65 – 1.55 (m, 4H), 1.50 – 1.37 (m, 8H), 1.27 – 1.08 (m, 4H).



³¹P NMR (162 MHz, CDCl₃) δ 154.50.



HRMS (ESI⁺, MeOH): *m/z* calcd. 424.2400 (M + H)⁺, found: 424.2421.



White solid; Column conditions: Hexane : $EA = 50 : 1, R_f = 0.33$



2H), 7.25 - 7.11 (m, 4H), 2.78 (dd, J = 10.5, 7.3 Hz, 4H), 1.86 (dp, J = 13.6, 6.8 Hz, 2H), 0.87 (d, J = 6.6 Hz, 12H).



³¹P NMR (162 MHz, CDCl₃) δ 151.88.



HRMS (ESI⁺, MeOH): *m/z* calcd. 344.1774 (M + H)⁺, found: 344.1776.



White solid; Column conditions: Hexane : EA = 50 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 8.04 – 7.86 (m, 4H), 7.54 (dd, *J* = 8.7, 1.1 Hz, 1H), 7.49 – 7.37 (m, 4H), 7.37 – 7.31 (m, 1H), 7.30 – 7.22 (m, 3H), 3.54 – 3.38 (m, 2H), 1.86 – 1.58 (m, 12H), 1.43 – 1.27 (m, 4H).



34.07, 34.00, 33.57, 33.48, 24.34, 24.11.



HRMS (ESI⁺, MeOH): *m/z* calcd. 468.2087 (M + H)⁺, found: 468.2087.



White solid; Column conditions: Hexane : EA = 50 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 8.00 – 7.84 (m, 4H), 7.51 (d, *J* = 8.8 Hz, 1H), 7.46 – 7.35 (m, 4H), 7.32 (d, *J* = 7.7 Hz, 1H), 7.29 – 7.19 (m, 2H), 3.03 (qt, *J* = 10.8, 3.8 Hz, 2H), 2.06 – 1.87 (m, 4H), 1.86 – 1.66 (m, 4H), 1.66 – 1.50 (m, 4H), 1.47 – 1.28 (m, 8H), 1.25 – 0.97 (m, 4H).



27.23, 25.28, 25.23.



HRMS (ESI⁺, MeOH): *m/z* calcd. 524.2713 (M + H)⁺, found: 524.2720.



Colorless oil; Column conditions: Hexane : $EA = 50 : 1, R_f = 0.35$



(m, 6H), 0.87 - 0.74 (m, 6H).



21.95, 11.94.



HRMS (ESI⁺, MeOH): *m/z* calcd. 344.1774 (M + H)⁺, found: 344.1773.

S32. IR, NMR spectra, HRMS data for vinyl γ-lactones



General procedure A: Colorless oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.18$



-2.47 (m, 4H).





General procedure A: Colorless solid; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.18$



= 17.1, 10.7 Hz, 1H, 5.30 (dd, J = 17.1, 0.8 Hz, 1H), 5.20 (dd, J = 10.7, 0.8 Hz, 1H), 2.67 - 2.47 (m, 4H), 2.34 (s, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 203.1067 (M + H)⁺, found: 203.1068.



General procedure A: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.28 (m, 2H), 7.27 – 7.23 (m, 3H), 6.06 (dd, J = 17.1, 10.7 Hz, 1H), 5.31 (dd, J = 17.2, 0.7 Hz, 1H), 5.22 (dd, J = 10.7, 0.7 Hz, 1H), 2.66 – 2.45 (m, 7H).


HRMS (ESI⁺, MeOH): *m*/*z* calcd. 235.0787 (M + H)⁺, found: 235.0790.

Wavenumber cm-1



General procedure A: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



¹H NMR (300 MHz, CDCl₃) δ /.54 – /.45 (m, 2H), /.30 – /.23 (m, 2H), 6.04 (dd, J = 17.1, 10.7 Hz, 1H), 5.31 (d, J = 17.1 Hz, 1H), 5.24 (d, J = 10.7 Hz, 1H), 2.69 – 2.41 (m, 4H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 288.9835 (M + Na)⁺, found: 288.9835.



General procedure A: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.18$



– 2.43 (m, 4H).



S41



HRMS (ESI⁺, MeOH): *m/z* calcd. 207.0816 (M + H)⁺, found: 207.0818.



General procedure A: Yellow oil; Column conditions: Hexane : DCM = 1 : 1, $R_f = 0.20$



¹H NMR (500 MHz, CDCl₃) δ 7.33 – 7.26 (m, 2H), 6.91 – 6.86 (m, 2H), 6.06 (dd, J = 17.2, 10.7 Hz, 1H), 5.28 (dd, J = 17.2, 0.8 Hz, 1H), 5.20 (dd, J = 10.7, 0.7 Hz, 1H), 3.79 (s, 3H), 2.65 – 2.47 (m, 4H).





General procedure A: Slight yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.20$



¹H NMR (400 MHz, CDCl₃) $_{0}$ 7.42 – 7.36 (m, 2H), 7.35 – 7.27 (m, 2H), 6.09 (dd, J = 17.2, 10.7 Hz, 1H), 5.33 (dd, J = 17.2, 0.7 Hz, 1H), 5.21 (dd, J = 10.7, 0.7 Hz, 1H), 2.66 – 2.49 (m, 4H), 1.32 (s, 9H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 267.1356 (M + Na)⁺, found: 267.1359.



General procedure A: White solid; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.10$



0.7 Hz, 1H), 2.71 – 2.51 (m, 4H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 265.1223 (M + H)⁺, found: 265.1226.



General procedure A: Colorless oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.20$



¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.38 (m, 1H), 7.36 – 7.28 (m, 2H), 7.21 – 7.13 (m, 1H), 6.09 (dd, J = 17.1, 10.7 Hz, 1H), 5.33 (dd, J = 17.2, 0.7 Hz, 1H), 5.22 (dd, J = 10.7, 0.7 Hz, 1H), 2.68 – 2.48 (m, 4H), 1.32 (s, 9H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 267.1356 (M + Na)⁺, found: 267.1357.



General procedure A: Yellow oil; Column conditions: Hexane : DCM = 1 : 1, $R_f = 0.18$



¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.36 (m, 1H), 7.36 – 7.29 (m, 2H), 7.29 – 7.25 (m, 1H), 6.05 (dd, J = 17.1, 10.7 Hz, 1H), 5.34 (dd, J = 17.1, 0.5 Hz, 1H), 5.26 (dd, J = 10.7, 0.5 Hz, 1H), 2.69 – 2.44 (m, 4H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 245.0340 (M + Na)⁺, found: 245.0344.



General procedure A: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



6.07 (dd, *J* = 17.2, 10.7 Hz, 1H), 5.36 (d, *J* = 17.1 Hz, 1H), 5.28 (d, *J* = 10.7 Hz, 1H), 2.72 - 2.61 (m, 2H), 2.59 - 2.45 (m, 2H).



¹⁹F NMR (376 MHz, CDCl₃) δ -62.34.



HRMS (ESI⁺, MeOH): *m/z* calcd. 279.0603 (M + Na)⁺, found: 279.0609.



General procedure A: Yellow oil; Column conditions: Hexane : DCM = 1 : 1, $R_f = 0.15$



(ddd, *J* = 17.1, 10.7, 1.0 HZ, 1H), 3.3. Hz, 1H), 2.76 – 2.48 (m, 4H).



Hz), 33.93 (d, J = 5.1 Hz), 28.04 (d, J = 1.8 Hz).



HRMS (ESI⁺, MeOH): *m/z* calcd. 247.0541 (M + Na)⁺, found: 247.0544.



General procedure A: Colorless oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.20$



¹H NMR (400 MHz, CDCl₃) δ 6.99 (s, 2H), 6.94 (s, 1H), 6.08 (dd, J = 17.2, 10.7 Hz, 1H), 5.32 (dd, J = 17.2, 0.8 Hz, 1H), 5.21 (dd, J = 10.7, 0.7 Hz, 1H), 2.66 – 2.46 (m, 4H), 2.32 (s, 6H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 239.1043 (M + Na)⁺, found: 239.1032.

0.2

Wavenumber cm-1



General procedure A: White solid; Column conditions: Hexane : $EA = 10 : 1, R_f = 0.11$



¹H NMR (400 MHz, CDCl₃) δ 7.80 – 7.72 (m, 3H), 7.42 (dd, J = 8.6, 1.9 Hz, 1H), 7.19 – 7.12 (m, 2H), 6.17 (dd, J = 17.2, 10.7 Hz, 1H), 5.36 (dd, J = 17.2, 0.7 Hz, 1H), 5.26 (dd, J = 10.7, 0.7 Hz, 1H), 3.92 (s, 3H), 2.71 – 2.52 (m, 4H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 291.0992 (M + Na)⁺, found: 291.0989.



General procedure A: White solid; Column conditions: Hexane : EA = 10 : 1, $R_f = 0.13$



-2.52 (m, 4H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 239.1067 (M + H)⁺, found: 239.1059.



General procedure A: Colorless oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 6.91 – 6.87 (m, 1H), 6.86 – 6.81 (m, 2H), 6.04 (dd, J = 17.2, 10.7 Hz, 1H), 5.30 (dd, J = 17.2, 0.6 Hz, 1H), 5.20 (dd, J = 10.7, 0.6 Hz, 1H), 4.25 (s, 4H), 2.66 – 2.45 (m, 4H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 247.0965 (M + H)⁺, found: 247.0955.



General procedure A: Colorless solid; Column conditions: Hexane : EA = 5 : 1, $R_f = 0.10$



¹H NMR (400 MHz, CDCl₃) δ 7.02 (dd, J = 1.7, 0.8 Hz, 1H), 6.99 – 6.93 (m, 2H), 6.05 (dd, J = 17.2, 10.7 Hz, 1H), 5.32 (dd, J = 17.1, 0.6 Hz, 1H), 5.24 (dd, J = 10.7, 0.7 Hz, 1H), 4.61 (s, 2H), 3.36 (s, 3H), 2.70 – 2.46 (m, 4H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 274.1074 (M + H)⁺, found: 274.1075.



General procedure **B**: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.32 (dd, J = 5.1, 1.3 Hz, 1H), 7.08 – 7.02 (m, 1H), 7.00 (dd, J = 5.0, 3.6 Hz, 1H), 6.15 (dd, J = 17.1, 10.7 Hz, 1H), 5.41 (dd, J = 17.1, 0.6 Hz, 1H), 5.30 (dd, J = 10.6, 0.6 Hz, 1H), 2.70 – 2.52 (m, 4H).





HRMS (ESI⁺, MeOH): *m/z* calcd. 195.0474 (M + H)⁺, found: 195.0471.



General procedure A: White solid; Column conditions: Hexane : EA = 10 : 1, $R_f = 0.10$



J = 17.2, 0.7 Hz, 1H), 5.26 (dd, J = 10.7, 0.7 Hz, 1H), 2.74 – 2.52 (m, 4H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 301.0835 (M + Na)⁺, found: 301.0841.


General procedure **B**: Colorless oil; Column conditions: Hexane : DCM = 1 : 1, $R_f = 0.13$



¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.37 (m, 2H), 6.37 (dd, J = 1.9, 1.0 Hz, 1H), 6.06 (dd, J = 17.2, 10.7 Hz, 1H), 5.35 (dd, J = 17.1, 0.7 Hz, 1H), 5.27 (dd, J = 10.7, 0.7 Hz, 1H), 2.68 – 2.53 (m, 2H), 2.50 – 2.38 (m, 2H).





HRMS (ESI⁺, MeOH): *m/z* calcd. 179.0703 (M + H)⁺, found: 179.0700.



General procedure **B**: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.18$

¹H NMR spectrum (CDCl₃)



¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.20 (m, 5H), 5.90 (dd, *J* = 17.2, 10.9 Hz, 1H), 5.28 (dd, *J* = 17.2, 0.9 Hz, 1H), 5.17 (dd, *J* = 10.9, 0.9 Hz, 1H), 3.10 (d, *J* = 14.0 Hz, 1H), 2.96 (d, *J* = 14.0 Hz, 1H), 2.46 – 2.35 (m, 1H), 2.24 – 2.04 (m, 3H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 225.0886 (M + Na)⁺, found: 225.0887.



General procedure **B**: Light yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.28 (m, 2H), 7.26 – 7.16 (m, 3H), 5.90 (dd, J = 17.3, 10.9 Hz, 1H), 5.39 (dd, J = 17.2, 0.9 Hz, 1H), 5.29 (dd, J = 10.9, 0.9 Hz, 1H), 2.81 – 2.50 (m, 4H), 2.25 – 2.01 (m, 4H).



¹³C NMR (101 MHz, CDCl₃) δ 176.76, 141.39, 138.63, 128.60, 128.37, 126.16, 115.02, 87.58, 42.00, 32.70, 30.20, 28.33.



HRMS (ESI⁺, MeOH): *m/z* calcd. 239.1043 (M + Na)⁺, found: 239.1052.



General procedure A: Light yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 5.81 (dd, J = 17.2, 10.9 Hz, 1H), 5.29 (dd, J = 17.2, 0.9 Hz, 1H), 5.19 (dd, J = 10.9, 0.9 Hz, 1H), 2.56 – 2.40 (m, 2H), 2.17 – 2.00 (m, 2H), 1.84 – 1.74 (m, 1H), 1.72 – 1.60 (m, 2H), 0.93 (dd, J = 9.6, 6.6 Hz, 6H).



¹³C NMR (101 MHz, CDCl₃) δ 177.10, 139.02, 114.31, 88.32, 48.92, 33.82, 28.11, 24.64, 24.33, 23.70.



HRMS (ESI⁺, MeOH): *m/z* calcd. 191.1043 (M + Na)⁺, found: 191.1036.



General procedure **B**: Slight yellow oil (39:61 *dr*); Column conditions: Hexane : DCM = $1 : 1, R_f = 0.20$

¹H NMR spectrum (CDCl₃)



¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.25 (m, 10H), 6.29 – 6.07 (m, 1H), 5.51 (d, J = 17.1 Hz, 0.39H), 5.34 – 5.23 (m, 2H), 4.03 (dd, J = 12.6, 8.3 Hz, 0.39H), 3.74 (dd, J = 12.5, 8.3 Hz, 0.61H), 3.17 – 3.08 (m, 1H), 2.78 – 2.62 (m, 1H).







General procedure A: Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.15$







HRMS (ESI⁺, MeOH): *m*/*z* calcd. 223.0730 (M + Na)⁺, found: 223.0731.



General procedure **B** (from 1-propenylmagnesium bromide): Yellow oil; Column conditions: Hexane : EA = 20 : 1, $R_f = 0.13$



¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.33 (m, 4H), 7.33 – 7.26 (m, 1H), 5.85 – 5.64 (m, 2H), 2.70 – 2.44 (m, 4H), 1.72 – 1.60 (m, 3H). The lactone is a mixture of two stereoisomers with a ratio of E/Z = 54:46.



¹³C NMR (101 MHz, CDCl₃) δ 176.66, 176.53, 143.54, 142.45, 133.09, 132.22, 130.69, 128.69, 128.64, 127.87, 127.78, 126.74, 125.11, 124.93, 88.39, 88.27, 38.00, 34.96, 28.92, 28.90, 17.79, 14.60.



HRMS (ESI⁺, MeOH): *m/z* calcd. 225.0886 (M + Na)⁺, found: 225.0877.

S87. Characterization data for the amino acid intermediate E-1



1.1 Hz, 1H), 6.72 - 6.66 (m, 2H), 5.86 (t, J = 6.6 Hz, 1H), 3.96 (d, J = 6.7 Hz, 2H), 2.93 (t, J = 7.7 Hz, 2H), 2.44 - 2.38 (m, 2H).



HRMS (ESI⁻, MeOH): *m/z* calcd. 280.1343 (M - H)⁻, found: 280.1349.



2D ¹H-¹H NOESY NMR spectrum (CDCl₃) for the mixture of Z-1 and E-1



S90. IR, NMR spectra and HRMS data for caprolactam products



Scale: 0.2 mmol; E/Z = 98:2, isolated 43.2 mg (82% yield), slight yellow solid, Hexane : $EA = 2: 1, R_f = 0.25$







HRMS (ESI⁺, MeOH): *m/z* calcd. 264.1383 (M + H)⁺, found: 264.1379.



Scale: 0.2 mmol; E/Z = 96:4, isolated 44.9 mg (81% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.35 (m, 2H), 7.30 – 7.21 (m, 5H), 7.19 – 7.13 (m, 2H), 6.16 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.10 – 3.03 (m, 2H), 2.94 – 2.87 (m, 2H), 2.36 (s, 3H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 300.1359 (M + Na)⁺, found: 300.1360.



Scale: 0.2 mmol; E/Z = 90:10, isolated 50.7 mg (82% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.34 (m, 2H), 7.32 – 7.19 (m, 8H), 6.17 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.12 – 3.01 (m, 2H), 2.94 – 2.86 (m, 2H), 2.49 (s, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 310.1260 (M + H)⁺, found: 310.1267.



Scale: 0.2 mmol; E/Z = 97:3, isolated 57.5 mg (84% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.44 (m, 2H), 7.40 – 7.34 (m, 2H), 7.28 – 7.19 (m, 5H), 6.17 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.2 Hz, 2H), 3.08 – 3.04 (m, 2H), 2.90 – 2.86 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 342.0488 (M + H)⁺, found: 342.0499.



Scale: 0.2 mmol; E/Z = 97:3, isolated 50.6 mg (90% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.30 (m, 4H), 7.30 – 7.20 (m, 3H), 7.09 – 6.97 (m, 2H), 6.13 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.11 – 3.02 (m, 2H), 2.92 – 2.83 (m, 2H).



⁻¹⁰⁰ f1 (ppm) -10 -20 -40 -50 -120 -30 -60 -70 -80 -90 -110 -130 -140 -150 -160 -170 -180 -190 ¹⁹F NMR (376 MHz, CDCl₃) δ -114.64.



HRMS (ESI⁺, MeOH): *m/z* calcd. 304.1108 (M + Na)⁺, found: 304.1105.



Scale: 0.2 mmol; E/Z = 93:7, isolated 41.1 mg (70% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.20 (m, 7H), 6.93 – 6.84 (m, 2H), 6.12 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.43 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.82 (s, 3H), 3.08 – 3.04 (m, 2H), 2.95 – 2.87 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 294.1489 (M + H)⁺, found: 294.1502.



Scale: 0.2 mmol; E/Z>99:1, isolated 58.1 mg (91% yield), white solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.31 (m, 6H), 7.29 – 7.21 (m, 3H), 6.19 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.45 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.10 – 3.03 (m, 2H), 2.96 – 2.90 (m, 2H), 1.34 (s, 9H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 320.2009 (M + H)⁺, found: 320.2008.



Scale: 0.2 mmol; E/Z = 97:3, isolated 54.3 mg (80% yield), white solid, Hexane : EA = 2 : 1, $R_f = 0.20$



¹H NMR (400 MHz, CDCl₃) δ 7.66 – 7.60 (m, 4H), 7.51 – 7.46 (m, 4H), 7.44 – 7.36 (m, 3H), 7.34 – 7.25 (m, 3H), 6.28 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.50 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.18 – 3.09 (m, 2H), 3.03 – 2.95 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 340.1696 (M + H)⁺, found: 340.1696.


Scale: 0.2 mmol; E/Z = 97:3, isolated 47.3 mg (74% yield), white solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.33 (m, 4H), 7.32 – 7.21 (m, 4H), 7.20 – 7.16 (m, 1H), 6.16 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.46 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.18 – 3.03 (m, 2H), 2.97 – 2.87 (m, 2H), 1.35 (s, 9H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 320.2009 (M + H)⁺, found: 320.2013.



Scale: 0.2 mmol; E/Z = 96:4, isolated 53.6 mg (90% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) $_{0}$ /.41 – /.35 (m, 3H), /.30 – /.22 (m, 6H), 6.18 (tt, J = 6.0 1.9 Hz, 1H), 4.45 (dt, J = 6.1, 2.2 Hz, 2H), 3.11 – 3.04 (m, 2H), 2.93 – 2.85 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 298.0993 (M + H)⁺, found: 298.0993.



Scale: 0.2 mmol; E/Z = 97:3, isolated 56.3 mg (85% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.62 (s, 1H), 7.58 – 7.53 (m, 2H), 7.49 – 7.44 (m, 1H), 7.41 – 7.35 (m, 2H), 7.30 – 7.21 (m, 3H), 6.23 (tt, *J* = 6.0, 1.8 Hz, 1H), 4.48 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.15 – 3.04 (m, 2H), 2.97 – 2.87 (m, 2H).





HRMS (ESI⁺, MeOH): *m/z* calcd. 332.1257 (M + H)⁺, found: 332.1259.



Scale: 0.2 mmol; E/Z = 99:1, isolated 46.1 mg (77% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.38 (m, 2H), 7.32 – 7.18 (m, 4H), 6.95 – 6.78 (m, 2H), 6.01 (tt, *J* = 5.9, 2.2 Hz, 1H), 4.46 (dt, *J* = 5.9, 2.2 Hz, 2H), 3.11 – 3.04 (m, 2H), 2.89 – 2.81 (m, 2H).



¹⁹F NMR (376 MHz, CDCl₃) δ -111.12 (d, J = 7.5 Hz), -111.50 (d, J = 7.5 Hz).



HRMS (ESI⁺, MeOH): *m/z* calcd. 322.1013 (M + Na)⁺, found: 322.1014.



Scale: 0.2 mmol; E/Z>99:1, isolated 41.4 mg (71% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.36 (m, 2H), 7.30 – 7.22 (m, 3H), 7.00 – 6.95 (m, 3H), 6.14 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.11– 3.02 (m, 2H), 2.95 – 2.87 (m, 2H), 2.34 (s, 6H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 292.1698 (M + H)⁺, found: 292.1696.



Scale: 0.2 mmol; E/Z>99:1, isolated 44.6 mg (65% yield), white solid, Hexane : EA = 2 : 1, $R_{\rm f} = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.76 – 7.69 (m, 3H), 7.50 (dd, J = 8.6, 2.0 Hz, 1H), 7.41 – 7.36 (m, 2H), 7.32 – 7.27 (m, 2H), 7.27 – 7.22 (m, 1H), 7.18 – 7.12 (m, 2H), 6.30 (tt, J = 6.1, 1.8 Hz, 1H), 4.49 (dt, J = 6.1, 2.0 Hz, 2H), 3.93 (s, 3H), 3.13 – 3.09 (m, 2H), 3.07 – 3.00 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 344.1645 (M + H)⁺, found: 344.1646.



Scale: 0.2 mmol; E/Z = 99:1, isolated 48.3 mg (77% yield), white solid, Hexane : EA = 2 : 1, $R_f = 0.18$



¹H NMR (500 MHz, CDCl₃) δ 7.89 – 7.79 (m, 4H), 7.55 – 7.46 (m, 3H), 7.42 – 7.35 (m, 2H), 7.31 (d, J = 7.3 Hz, 2H), 7.27 – 7.22 (m, 1H), 6.33 (tt, J = 6.1, 1.7 Hz, 1H), 4.50 (dt, J = 6.1, 2.1 Hz, 2H), 3.14 – 3.10 (m, 2H), 3.08 – 3.03 (m, 2H).





HRMS (ESI⁺, MeOH): *m/z* calcd. 314.1539 (M + H)⁺, found: 314.1545.



Scale: 0.2 mmol; E/Z = 98:2, isolated 54.6 mg (85% yield), white solid, Hexane : EA = 2 : 1, $R_f = 0.18$



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.34 (m, 2H), 7.27 – 7.26 (m, 1H), 7.26 – 7.19 (m, 2H), 6.90 – 6.81 (m, 3H), 6.11 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.42 (dt, *J* = 6.1, 2.1 Hz, 2H), 4.26 (s, 4H), 3.08 – 3.00 (m, 2H), 2.90 – 2.82 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 322.1438 (M + H)⁺, found: 322.1443.



Scale: 0.2 mmol; E/Z = 93:7, isolated 50.9 mg (73% yield), white solid, Hexane : EA = 1 : 1, $R_f = 0.10$



¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.34 (m, 2H), 7.28 (s, 1H), 7.26 – 7.21 (m, 2H), 7.04 – 6.99 (m, 1H), 6.98 – 6.92 (m, 2H), 6.16 – 6.09 (m, 1H), 4.62 (s, 2H), 4.46 (d, *J* = 6.1 Hz, 2H), 3.39 (s, 3H), 3.11 – 3.04 (m, 2H), 2.92 – 2.87 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 349.1547 (M + H)⁺, found: 349.1549.



Scale: 0.2 mmol; E/Z = 78:22, isolated 36.1 mg (67% yield), yellow solid, Hexane : EA = 3 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.33 (m, 2H), 7.27 – 7.21 (m, 3H), 7.20 – 7.15 (m, 1H), 7.07 – 7.04 (m, 1H), 7.03 – 6.96 (m, 1H), 6.37 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.10 – 3.03 (m, 2H), 3.01 – 2.92 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 270.0947 (M + H)⁺, found: 270.0943.



Scale: 0.2 mmol; E/Z = 99:1, isolated 56.5 mg (80% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.99 – 7.93 (m, 2H), 7.60 – 7.52 (m, 2H), 7.50 – 7.44 (m, 2H), 7.43 – 7.35 (m, 3H), 7.35 – 7.30 (m, 2H), 7.28 – 7.23 (m, 1H), 6.23 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.50 (dt, *J* = 6.1, 2.2 Hz, 2H), 3.16 – 3.09 (m, 2H), 3.06 – 2.98 (m, 2H).





HRMS (ESI⁺, MeOH): *m/z* calcd. 376.1308 (M + Na)⁺, found: 376.1305.



Scale: 0.2 mmol; E/Z = 87:13, isolated 36.5 mg (72% yield), yellow oil, Hexane : EA = $4:1, R_f = 0.15$



6.53 - 6.48 (m, 1H), 6.18 (tt, J = 6.1, 1.8 Hz, 1H), 4.42 (dt, J = 6.1, 2.0 Hz, 2H), 3.06 - 3.00 (m, 2H), 2.83 - 2.77 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 276.0995 (M + Na)⁺, found: 276.0996.



Scale: 0.2 mmol; E/Z>99:1, isolated 41.6 mg (75% yield), slight yellow solid, Hexane : EA = 2 : 1, $R_f = 0.2$



¹H NMR (300 MHz, CDCl₃) $_{0}$ $_{.52}$ $_{-2.27}$ (iii, 2H), $_{.23}$ $_{-2.21}$ (iii, 2H), $_{.18}$ $_{-2.08}$ (iii, 6H), 5.65 (t, J = 6.1 Hz, 1H), 4.21 (d, J = 5.9 Hz, 2H), 3.25 (s, 2H), 2.85 $_{-2.76}$ (m, 2H), 2.36 $_{-2.29}$ (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 278.1539 (M + H)⁺, found: 278.1542.



Scale: 0.2 mmol; E/Z = 26:74, isolated 29.7 mg (51% yield), yellow oil, Hexane : EA = $2:1, R_f = 0.20$



¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.27 (m, 4H), 7.24 – 7.20 (m, 2H), 7.19 – 7.14 (m, 4H), 5.62 (tt, *J* = 6.0, 1.5 Hz, 1H), 4.21 (d, *J* = 5.9 Hz, 2H), 2.96 – 2.89 (m, 2H), 2.80 – 2.74 (m, 2H), 2.54 – 2.47 (m, 2H), 2.37 – 2.31 (m, 2H).



¹³C NMR (101 MHz, CDCl₃) δ 173.96, 143.84, 142.53, 141.51, 129.04, 128.53, 128.51, 126.52, 126.07, 125.90, 119.96, 49.19, 40.66, 34.37, 34.01, 28.69.



HRMS (ESI⁺, MeOH): *m/z* calcd. 292.1696 (M + H)⁺, found: 292.1693.



Scale: 0.2 mmol; E/Z = 62:38, isolated 26.3 mg (54% yield), yellow oil, Hexane : EA = $2:1, R_f = 0.25$



¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.33 (m, 2H), 7.24 – 7.18 (m, 3H), 5.62 (tt, *J* = 6.0, 1.0 Hz, 1H), 4.24 (d, *J* = 5.9 Hz, 2H), 2.93 – 2.89 (m, 2H), 2.44 – 2.40 (m, 2H), 1.88 (d, *J* = 7.4 Hz, 2H), 1.82 – 1.74 (m, 1H), 0.88 (d, *J* = 6.5 Hz, 6H).



¹³C NMR (126 MHz, CDCl₃) δ 174.06, 143.94, 142.61, 129.08, 126.50, 125.89, 120.33, 49.31, 48.85, 34.06, 28.57, 26.18, 22.46.



HRMS (ESI⁺, MeOH): *m/z* calcd. 266.1515 (M + Na)⁺, found: 266.1519.



Scale: 0.2 mmol; E/Z = 97:3, isolated 55.0 mg (81% yield), yellow oil, Hexane : EA = 6 : 1, $R_f = 0.15$



¹H NMR (500 MHz, CDCl₃) δ 7.48 – 7.44 (m, 2H), 7.43 – 7.35 (m, 8H), 7.34 – 7.28 (m, 4H), 7.24 (t, *J* = 7.3 Hz, 1H), 6.25 (ddt, *J* = 7.8, 4.1, 1.9 Hz, 1H), 4.98 – 4.88 (m, 1H), 4.70 (dd, *J* = 11.7, 3.6 Hz, 1H), 4.08 (dd, *J* = 18.0, 7.5 Hz, 1H), 3.35 – 3.27 (m, 1H), 3.10 – 3.04 (m, 1H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 340.1696 (M + H)⁺, found: 340.1699.



Scale: 0.2 mmol; E/Z>99:1, isolated 28.6 mg (52% yield), yellow oil, Hexane : EA = 5 : 1, $R_{\rm f} = 0.15$



¹H NMR (400 MHz, CDCl₃) $_{0}$ 7.43 – 7.31 (m, 9H), 7.27 – 7.22 (m, 1H), 6.28 (tt, J = 6.5, 1.8 Hz, 1H), 5.63 (d, J = 1.2 Hz, 1H), 5.44 (d, J = 1.3 Hz, 1H), 4.39 (d, J = 6.5 Hz, 2H), 3.64 (d, J = 1.5 Hz, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 276.1383 (M + H)⁺, found: 276.1380.


Scale: 0.2 mmol; E/Z>99:1, isolated 34.9 mg (63% yield), colorless oil, Hexane : EA = 1 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.43 – 7.28 (m, 8H), 7.21 – 7.14 (m, 2H), 5.95 (dt, J = 6.0, 1.7 Hz, 1H), 4.88 – 4.77 (m, 1H), 3.21 (td, J = 12.4, 11.8, 5.2 Hz, 1H), 3.01 – 2.83 (m, 3H), 1.36 (d, J = 7.2 Hz, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 278.1539 (M + H)⁺, found: 278.1542.



Scale: 0.2 mmol; E/Z = 97:3, isolated 46.9 mg (80% yield), slight yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.27 (m, 5H), 7.20 – 7.15 (m, 2H), 6.92 – 6.86 (m, 2H), 6.16 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.41 (dt, *J* = 6.1, 2.2 Hz, 2H), 3.80 (s, 3H), 3.08 – 3.02 (m, 2H), 2.95 – 2.89 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 294.1489 (M + H)⁺, found: 294.1489.



Scale: 0.2 mmol; E/Z>99:1, isolated 53.4 mg (91% yield), slight yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



11 NMR (400 MHz, CDC13) 6 7.40 – 7.52 (iii, 411), 7.51 – 7.28 (iii, 111), 7.27 – 7.28 (iii, 111), 7.28 (ii



HRMS (ESI⁺, MeOH): *m/z* calcd. 294.1489 (M + H)⁺, found: 294.1490.



Scale: 0.2 mmol; E/Z = 99:1, isolated 50.5 mg (86% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.27 (m, 6H), 6.88 – 6.77 (m, 3H), 6.17 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.44 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.80 (s, 3H), 3.10 – 3.04 (m, 2H), 2.96 – 2.89 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 294.1489 (M + H)⁺, found: 294.1489.



Scale: 0.2 mmol; E/Z = 92:8, isolated 41.1 mg (74% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.25$



1H), 7.09 - 7.04 (m, 3H), 6.17 (tt, J = 6.1, 1.8 Hz, 1H), 4.43 (dt, J = 6.1, 2.1 Hz, 2H), 3.09 - 3.03 (m, 2H), 2.95 - 2.89 (m, 2H), 2.35 (s, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 278.1539 (M + H)⁺, found: 278.1546.



Scale: 0.2 mmol; E/Z = 98:2, isolated 52.3 mg (93% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.28 (m, 5H), 7.26 – 7.20 (m, 2H), 7.09 – 7.02 (m, 2H), 6.17 (tt, *J* = 6.0, 1.8 Hz, 1H), 4.42 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.09 – 3.03 (m, 2H), 2.95 – 2.89 (m, 2H).



-100 f1 (ppm) -10 -20 -30 -40 -50 -60 -70 -90 -110 -120 -140 -150 -160 -170 -190 -80 -130 -180 ¹⁹F NMR (376 MHz, CDCl₃) δ -115.48.



HRMS (ESI⁺, MeOH): *m/z* calcd. 282.1289 (M + H)⁺, found: 282.1295.



Scale: 0.2 mmol; E/Z = 97:3, isolated 50.9 mg (75% yield), white solid, Hexane : EA = 2 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.62 – 7.56 (m, 4H), 7.47 – 7.42 (m, 2H), 7.41 – 7.32 (m, 8H), 6.21 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.50 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.13 – 3.07 (m, 2H), 2.98 – 2.91 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 340.1696 (M + H)⁺, found: 340.1705.



Scale: 0.2 mmol; E/Z>99:1, isolated 32.4 mg (58% yield), white solid, DCM : MeOH = 50 : 1, $R_f = 0.13$



¹H NMR (400 MHz, CDCl₃) δ 7.73 (s, 1H), 7.39 – 7.27 (m, 5H), 6.98 – 6.88 (m, 2H), 6.63 – 6.57 (m, 2H), 6.12 (tt, *J* = 6.0, 1.8 Hz, 1H), 4.37 (dt, *J* = 6.1, 2.2 Hz, 2H), 3.12 – 3.00 (m, 2H), 2.96 – 2.86 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 280.1332 (M + H)⁺, found: 280.1330.



Scale: 0.2 mmol; E/Z = 94:6, isolated 38.1 mg (62% yield), yellow solid, Hexane : EA = 1 : 3, $R_f = 0.2$



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.27 (m, 5H), 7.24 – 7.18 (m, 4H), 6.16 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.43 (dt, *J* = 6.1, 2.2 Hz, 2H), 3.79 (t, *J* = 6.7 Hz, 2H), 3.10 – 3.02 (m, 2H), 2.95 – 2.88 (m, 2H), 2.83 (t, *J* = 6.6 Hz, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 308.1645 (M + H)⁺, found: 308.1652.



Scale: 0.2 mmol; E/Z = 96:4, isolated 45.6 mg (68% yield), yellow solid, Hexane : EA = $1:1, R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.94 – 7.90 (m, 2H), 7.52 – 7.46 (m, 1H), 7.47 – 7.40 (m, 1H), 7.40 – 7.30 (m, 5H), 6.18 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.47 (dt, *J* = 6.1, 2.1 Hz, 2H), 4.37 (q, *J* = 7.1 Hz, 2H), 3.14 – 3.03 (m, 2H), 2.97 – 2.89 (m, 2H), 1.38 (t, *J* = 7.2 Hz, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 336.1594 (M + H)⁺, found: 336.1595.



Scale: 0.2 mmol; E/Z = 94:6, isolated 48.4 mg (83% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (500 MHz, CDCl₃) δ 7.40 – 7.28 (m, 5H), 6.92 – 6.82 (m, 3H), 6.16 (tt, *J* = 6.1, 1.8 Hz, 1H), 4.41 (dt, *J* = 6.1, 2.1 Hz, 2H), 3.08 – 3.04 (m, 2H), 2.95 – 2.90 (m, 2H), 2.31 (s, 6H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 292.1696 (M + H)⁺, found: 292.1704.



Scale: 0.2 mmol; E/Z = 99:1, isolated 54.4 mg (73% yield), yellow solid, Hexane : EA = $2:1, R_f = 0.18$







HRMS (ESI⁺, MeOH): *m/z* calcd. 372.0594 (M + H)⁺, found: 372.0599.



Scale: 0.2 mmol; E/Z>99:1, isolated 45.5 mg (71% yield), yellow solid, Hexane : EA = 1 : 1, $R_f = 0.15$



¹H NMR (500 MHz, CDCl₃) δ 7.99 (s, 1H), 7.40 (t, J = 2.1 Hz, 1H), 7.39 – 7.33 (m, 4H), 7.32 – 7.27 (m, 2H), 7.24 (t, J = 7.9 Hz, 1H), 6.91 (dt, J = 7.6, 1.6 Hz, 1H), 6.16 (tt, J = 6.1, 2.0 Hz, 1H), 4.46 – 4.40 (m, 2H), 3.09 – 3.01 (m, 2H), 2.94 – 2.86 (m, 2H), 2.02 (s, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 321.1598 (M + H)⁺, found: 321.1604.



Scale: 0.2 mmol; E/Z>99:1, isolated 36.3 mg (60% yield), yellow solid, Hexane : EA = 1 : 2, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 8.58 (s, 1H), 7.47 (d, J = 2.1 Hz, 1H), 7.42 – 7.34 (m, 4H), 7.33 – 7.28 (m, 1H), 7.24 – 7.19 (m, 1H), 7.13 (t, J = 2.8 Hz, 1H), 7.02 – 6.96 (m, 1H), 6.47 (t, J = 2.8 Hz, 1H), 6.19 (tt, J = 6.0, 1.9 Hz, 1H), 4.48 (dt, J = 6.1, 2.1 Hz, 2H), 3.14 – 3.07 (m, 2H), 3.00 – 2.92 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 303.1492 (M + H)⁺, found: 303.1494.



Scale: 0.2 mmol; E/Z = 95:5, isolated 45.5 mg (74% yield), yellow solid, Hexane : EA = 1 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.30 – 7.20 (m, 5H), 6.73 – 6.66 (m, 2H), 6.63 – 6.60 (m, 1H), 6.07 (tt, *J* = 6.0, 1.8 Hz, 1H), 5.88 (s, 2H), 4.30 (dt, *J* = 6.1, 2.1 Hz, 2H), 2.99 – 2.92 (m, 2H), 2.86 – 2.78 (m, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 308.1281 (M + H)⁺, found: 308.1273.



Scale: 0.2 mmol; E/Z = 95:5, isolated 60.8 mg (81% yield), yellow solid, Hexane : EA = 1 : 1, $R_f = 0.15$



¹H NMR (400 MHz, CDCl₃) δ 7.59 – 7.54 (m, 2H), 7.48 (d, J = 0.9 Hz, 1H), 7.40 – 7.28 (m, 6H), 6.19 (tt, J = 6.1, 1.8 Hz, 1H), 4.53 – 4.47 (m, 2H), 4.44 (q, J = 7.1 Hz, 2H), 3.16 – 3.02 (m, 2H), 3.00 – 2.88 (m, 2H), 1.43 (t, J = 7.1 Hz, 3H).





HRMS (ESI⁺, MeOH): *m*/*z* calcd. 398.1363 (M + Na)⁺, found: 398.1363.

S177



Scale: 0.2 mmol; E/Z>99:1, isolated 44.5 mg (71% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.20$



(m, 2H), 7.36 - 7.31 (m, 3H), 7.30 - 7.27 (m, 2H), 7.26 - 7.22 (m, 1H), 6.16 (tt, J = 6.1, 1.8 Hz, 1H), 4.47 (dt, J = 6.1, 2.1 Hz, 2H), 3.07 - 3.02 (m, 2H), 2.93 - 2.86 (m, 2H).





HRMS (ESI⁺, MeOH): *m/z* calcd. 314.1539 (M + H)⁺, found: 314.1531.



Scale: 0.1 mmol *p*-phenylenediamine, 0.2 mmol vinyl lactone **A**; isolated 31.0 mg (69% yield), yellow solid, DCM : MeOH = $50 : 1, R_f = 0.10$



(dt, J = 6.1, 2.1 Hz, 4H), 3.13 - 3.00 (m, 4H), 2.97 - 2.84 (m, 4H).


HRMS (ESI⁺, MeOH): *m/z* calcd. 449.2224 (M + H)⁺, found: 449.2225.



Scale: 0.2 mmol; E/Z>99:1, isolated 32.1 mg (55% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.23 (m, 10H), 6.08 (q, *J* = 7.0 Hz, 1H), 5.80 (tt, *J* = 6.0, 1.8 Hz, 1H), 3.74 (dt, *J* = 6.0, 2.2 Hz, 2H), 3.06 – 2.76 (m, 4H), 1.53 (d, *J* = 7.0 Hz, 3H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 292.1696 (M + H)⁺, found: 292.1700.



Scale: 0.2 mmol; E/Z>99:1, isolated 26.6 mg (48% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$





HRMS (ESI⁺, MeOH): *m/z* calcd. 278.1539 (M + H)⁺, found: 278.1544.



Scale: 0.2 mmol; E/Z>99:1, isolated 19.9 mg (37% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.25$



¹H NMR (500 MHz, CDCl₃) δ 7.32 – 7.23 (m, 5H), 6.02 (tt, *J* = 6.1, 1.9 Hz, 1H), 4.47 (tt, *J* = 11.8, 3.8 Hz, 1H), 3.91 (dt, *J* = 6.1, 2.1 Hz, 2H), 2.89 – 2.85 (m, 2H), 2.78 – 2.73 (m, 2H), 1.79 – 1.73 (m, 2H), 1.68 – 1.64 (m, 3H), 1.43 – 1.34 (m, 2H), 1.33 – 1.25 (m, 2H), 1.10 – 1.01 (m, 1H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 270.1852 (M + H)⁺, found: 270.1855.



Scale: 0.2 mmol; E/Z>99:1, isolated 41.0 mg (58% yield), yellow solid, Hexane : EA = 2 : 1, $R_f = 0.15$



2.75 (m, 2H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 376.1672 (M + Na)⁺, found: 376.1669.

S188. Product diversification based on compounds 3 and 29



In a vial equipped with a magnetic stirring bar, caprolactam **3** (52.7 mg, 0.20 mmol, 1.0 equiv.) was dissolved into DCM (0.20 mL), and then trifluoromethanesulfonic acid (0.035 mL, 0.40 mmol, 2.0 equiv.) was added with a syringe. The resultant solution was stirred for 8 h at room temperature. Hereafter, saturated aqueous NaHCO₃ was slowly added and the organic components were extracted with DCM (3×10 mL). Then, the combined organic layers were dried over Na₂SO₄, filtered, and concentrated under vacuum. The crude product was purified by flash chromatography (Hexane/EtOAc = 2:1) to afford the isomerized product **50** (40.6 mg, 77%) as a light yellow solid.



To a solution of caprolactam **3** (52.7 mg, 0.20 mmol, 1.0 equiv.) in DCM (0.40 mL) was added *m*CPBA (0.40 mmol, 69.0 mg, 2.0 equiv.) at 0 °C under air. Then the reaction mixture was allowed to warm to room temperature, and stirred for 12 h. The reaction mixture was filtered through Celite. The solvent in the filtrate was evaporated under reduced pressure, and the crude product was purified by flash chromatography (Hexane/EtOAc = 1:1) to afford the pure epoxide product **51** (47.5 mg, 85%, >99:1 *dr*) as a yellow solid.



A Schlenk tube charged with **51** (83.8 mg, 0.30 mmol, 1.0 equiv) and Pd/C catalyst (6.4 mg, 10.0 wt% palladium on carbon) was evacuated and filled with H₂ (balloon) for three times. After that, MeOH (1.0 mL) was added and the reaction mixture was stirred under a H₂ atmosphere (balloon) for 2 h at room temperature and monitored by NMR. The reaction mixture was filtered through Celite. The solvent in the filtrate was evaporated under reduced pressure. The residue was purified by flash chromatography (Hexane/EtOAc = 1:2) to afford the pure product **52** (77.7 mg, 92%, >99:1 *dr*) as a white solid.



To an oven-dried Schlenk tube was added caprolactam **3** (52.7 mg, 0.20 mmol, 1.0 equiv) and anhydrous DCM (0.40 mL), and the solution was cooled to 0 °C. A solution of Br₂ (10.2 μ L, 0.40 mmol 1.0 equiv) in DCM (0.20 mL) was added dropwise under N₂ and the reaction mixture stirred at 0 °C. After completion (about 1 h, followed by TLC), the reaction mixture was diluted with DCM and washed with 5% NaHCO₃ solution. The organic layer was then dried over MgSO₄, filtered, and then concentrated. The residue was purified by flash chromatography (Hexane/EtOAc = 5:1) to afford the dibrominated product **53** (70.2 mg, 83%, >99:1 *dr*) as a colorless solid. The X-ray molecular structure was also determined.



Under N₂, to a magnetically stirred solution of caprolactam **3** (52.7 mg, 0.20 mmol, 1.0 equiv) in DCM (1.0 mL) was added pyridinium chlorochromate (PCC, 86.2 mg, 0.40 mmol, 2.0 equiv.). The mixture was stirred at 40 °C for 18 h, then cooled, diluted with water (10 mL), and extracted with EtOAc (3×10 mL). The combined organic layers were washed with 5% aqueous NaHCO₃ (10 mL), and dried with sodium sulfate, filtered, and then concentrated. Purification of the residue by flash chromatography (Hexane/EtOAc = 10:1) to afford the desired product **54** (29.3 mg, 56%) as a colorless oil.



In a vial equipped with a magnetic stirring bar, caprolactam **29** (44.0 mg, 0.15 mmol, 1.0 equiv.) was dissolved into ACN (0.9 mL) and stirred at 0 °C. A solution of CAN (205.6 mg, 0.375 mmol, 2.5 equiv.) in H₂O (1.3 mL) was added dropwise. The resultant solution was stirred at 0 °C until no starting materials could be detected by TLC (about 40 min). Hereafter, saturated aqueous NaHCO₃ was added and the organic components were extracted with DCM (3×10 mL). The combined organic layers were then dried over MgSO₄, filtered, and then concentrated, the crude product was purified by flash chromatography (DCM/MeOH = 30:1) to afford the deprotected lactam **55** (11.2 mg, 40%) as a light yellow solid.



A Schlenk tube was charged with **29** (176.0 mg, 0.6 mmol, 1.0 equiv) and Pd/C as catalyst (12.8 mg, 10.0 wt % palladium on carbon), and then the reactor was evacuated/filled with H_2 (balloon) for three times. After that, MeOH (2.0 mL) was added and the reaction mixture was stirred under H_2 atmosphere (balloon) for 2 h at room temperature, and monitored by NMR. The reaction mixture was filtered through Celite. The solvent in the filtrate was evaporated under reduced pressure. The residue was purified by flash chromatography (Hex/EtOAc = 3:1) to afford the pure saturated lactam **56** (159.5 mg, 90%) as a yellow solid.

In a vial equipped with a magnetic stirring bar, caprolactam **56** (44.3 mg, 0.15 mmol, 1.0 equiv.) was dissolved into ACN (0.9 mL) and stirred at 0 °C. A solution of CAN (205.6 mg, 0.375 mmol, 2.5 equiv.) in H₂O (1.3 mL) was added dropwise. The resultant solution was stirred at 0 °C for 4 h. Hereafter, saturated aqueous NaHCO₃ was added and the organic components were extracted with DCM (3×10 mL). The combined organic layers were then dried over MgSO₄, filtered, and then concentrated, the crude product was purified by flash chromatography (DCM/MeOH = 30:1) to afford the deprotected lactam **57** (21.3 mg, 75%) as a light brown solid.



Six-membered vinyl lactone **58** (40.5 mg, 0.20 mmol, 1.0 equiv) was combined with $Pd_2(dba)_3 \cdot CHCl_3$ (6.0 mg, 3.0 mol%), **L8** (9.6 mg, 12.0 mol%) and aniline (38.0 mg, 0.40 mmol) in DCM (0.30 mL) at room temperature in air. The reaction mixture was stirred at room temperature for 12 h, then an aliquot of the mixture was taken for NMR analysis, which showed the unsaturated amino acid intermediate to have an E/Z ratio of >99:1. Hereafter, the mixture was diluted to 0.01 M and EDC (57.5 mg, 3.0 mmol) was added and the reaction mixture stirred for another 1 h. The desired eight-membered lactam **59** (30.0 mg, 54%) was isolated by flash chromatography (Hexane/EtOAc = 2:1).



Scale: 0.2 mmol; isolated 40.6 mg (77% yield), light yellow solid, Hexane : EA = 2 : 1, $R_{\rm f} = 0.25$



2.2 Hz, 2H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 264.1383 (M + H)⁺, found: 264.1386.



Scale: 0.2 mmol; isolated 47.5 mg (85% yield, >99:1 dr), yellow solid, Hexane : EA = 1 : 1, $R_{\rm f} = 0.15$





HRMS (ESI⁺, MeOH): *m/z* calcd. 280.1332 (M + H)⁺, found: 280.1328.



Scale: 0.3 mmol; isolated 77.7 mg (92% yield, >99:1 dr), white solid, Hexane : EA = 1 : 2, $R_f = 0.15$



¹H NMR (400 MHz, DMSO) 8 7.37 – 7.32 (m, 4H), 7.32 – 7.26 (m, 4H), 7.22 – 7.15 (m, 2H), 4.85 (d, J = 6.8 Hz, 1H), 4.19 (d, J = 15.1 Hz, 1H), 3.90 (t, J = 7.3 Hz, 1H), 3.65 (dd, J = 15.2, 6.7 Hz, 1H), 2.97 – 2.91 (m, 1H), 2.88 – 2.80 (m, 1H), 2.57 – 2.52 (m, 1H), 2.33 – 2.21 (m, 1H), 1.73 – 1.66 (m, 1H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 282.1489 (M + H)⁺, found: 282.1495.



Scale: 0.2 mmol; isolated 70.2 mg (83% yield, >99:1 dr), colorless solid, Hexane : EA = 5 : 1, $R_f = 0.10$



¹H NMR (400 MHz, CDCl₃) δ 7.60 – 7.54 (m, 2H), 7.46 – 7.35 (m, 7H), 7.29 – 7.24 (m, 1H), 5.25 (d, *J* = 15.9 Hz, 1H), 5.10 (dd, *J* = 5.8, 2.1 Hz, 1H), 4.00 (dd, *J* = 15.9, 6.1 Hz, 1H), 3.55 – 3.46 (m, 1H), 3.09 – 2.86 (m, 2H), 2.81 – 2.68 (m, 1H).



HRMS (ESI⁺, MeOH): *m/z* calcd. 421.9750 (M + H)⁺, found: 421.9755.



Scale: 0.2 mmol; isolated 29.3 mg (56% yield), colorless oil, Hexane : EA = 10 : 1, $R_{\rm f} = 0.15$





HRMS (ESI⁺, MeOH): *m/z* calcd. 262.1226 (M + H)⁺, found: 262.1225.



Scale: 0.15 mmol; isolated 11.2 mg (40% yield), light yellow solid, DCM : MeOH = 30 : 1, $R_{\rm f} = 0.15$





HRMS (ESI⁺, MeOH): *m*/*z* calcd. 210.0889 (M + Na)⁺, found: 210.0884.



Scale: 0.6 mmol; isolated 159.5 mg (90% yield), yellow solid, Hexane : EA = 3 : 1, $R_{\rm f} = 0.18$



¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.30 (m, 2H), 7.26 – 7.20 (m, 3H), 7.20 – 7.13 (m, 2H), 6.96 – 6.88 (m, 2H), 4.02 (dd, J = 15.2, 10.7 Hz, 1H), 3.81 (s, 3H), 3.67 (ddd, J = 15.4, 6.4, 1.7 Hz, 1H), 2.92 – 2.75 (m, 3H), 2.17 – 2.05 (m, 2H), 2.00 – 1.88 (m, 2H).



¹³C NMR (101 MHz, CDCl₃) δ 175.32, 158.16, 146.14, 137.49, 128.83, 127.43, 126.87, 126.77, 114.63, 55.63, 52.64, 48.37, 37.09, 36.74, 31.05.



HRMS (ESI⁺, MeOH): *m/z* calcd. 296.1645 (M + H)⁺, found: 296.1640.



Scale: 0.15 mmol; isolated 21.3 mg (75% yield), light brown solid, DCM : MeOH = 30 : 1, $R_{\rm f} = 0.14$



¹H NMR (500 MHz, CDCl₃) δ 7.33 – 7.28 (m, 2H), 7.24 – 7.15 (m, 3H), 6.66 (s, 1H), 3.47 – 3.21 (m, 2H), 2.76 (tt, *J* = 12.1, 3.4 Hz, 1H), 2.67 – 2.53 (m, 2H), 2.05 – 1.96 (m, 2H), 1.85 – 1.69 (m, 2H).





¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.34 (m, 4H), 7.32 – 7.27 (m, 1H), 6.05 (dd, J = 17.2, 10.7 Hz, 1H), 5.32 (dd, J = 17.2, 0.8 Hz, 1H), 5.24 (dd, J = 10.8, 0.8 Hz, 1H), 2.62 – 2.53 (m, 1H), 2.51 – 2.42 (m, 1H), 2.31 – 2.15 (m, 2H), 1.96 – 1.85 (m, 1H), 1.80 – 1.69 (m, 1H).





Scale: 0.2 mmol; isolated 30.0 mg (54% yield), slight yellow solid, Hexane : EA = 2 : 1, $R_{\rm f} = 0.25$



Hz, 1H), 4.45 (d, J = 6.2 Hz, 2H), 2.89 – 2.83 (m, 4H), 2.14 – 2.07 (m, 2H).



HRMS (ESI⁺, MeOH): *m*/*z* calcd. 300.1359 (M + Na)⁺, found: 300.1347.

S211. Control reactions

The nucleophilic attack at the less hindered, terminal carbon is typically favored in Pdcatalyzed allylic substitution reactions. However, this preference can be modulated by the sterics and electronics of the π -allylmetals. For electronically related π -acceptor ligands (see: *B. L. Feringa, J. F. Teichert, Angew. Chem. Int. Ed.* **2010**, 49, 2486) such as phosphoramidites and phosphites can increase the cationic character of the allyl unit, with an increased cationic character being more stable at the internal carbon center of the allyl group and thus directs (electronically) the nucleophilic addition to that position (see also: *B. M. Trost, M. R. Machacek, A. Aponick, Acc. Chem. Res.* **2006**, 39, 747). According to our previous research based on Pd/phosphoramidite mediated regioselective allylic substitution (see refs. 14a and 17 of the main text), it showed significant potential for "branched amination" and thus providing a possible regio-selectivity issue when using

phosphoramidite ligands (such as **L8**) in the formation of caprolactams in our tandem process, and potentially lowering the yield of the target caprolactam. Indeed, when we used vinyl cyclic carbonate as an allylic surrogate instead of a γ -vinyl lactone under the standard conditions, a mixture of regio-isomers were detected with a ratio of branched/(*Z*)-linear of 15:85 as determined by ¹H



NMR integration. <u>Note</u>: branched allylic amine product (double doublet at 6.45 ppm), linear allylic amine product with a triplet at 6.18 ppm.



The use of a vinyl lactone having a secondary carbon center ($R^1 = H$, see below: this is a known compound) was also attempted after its preparation through the following procedure⁷:



Amination (after ring-opening) of this vinyl γ -lactone substrate (with R¹ = H) proceeded efficiently under standard conditions (main text, Table 1, entry 20) with 100% conversion and producing a 71% NMR yield of the *ɛ*-amino acid intermediate. However, the stereocontrol was significantly decreased with a *Z/E* ratio of 11:89 (<u>note</u>: for this compound the *E/Z* priority assignment is different and here the "Z" isomer is warranted for cyclization). As a result, only trace amount of the desired caprolactam could be achieved. This reasoning follows the general observation that preparation of (*Z*)-configured 1,2-disubstituted alkenes by allylic substitution is very challenging because of steric effects.



Analytical data for this unsubstituted vinyl lactone:



Using general procedure **B**: Light yellow oil; Column conditions: Hexane : EA = 10 : 1, $R_f = 0.15$



- 2.34 (m, 1H), 2.05 - 1.94 (m, 1H).


The preparation of a vinyl ε -lactone substrate failed by using the standard method used to get access to the γ -lactones. After some optimization, it could be prepared though a two-step method from 5-benzoyl pentanoic acid:



60% yield of two steps

Procedure: under a N₂ atmosphere, to a separate flame-dried round-bottom flask equipped with a stirring bar was added the respective 5-benzoyl pentanoic acid (1.03 g, 5.0 mmol) and anhydrous THF (15 mL). The solution was cooled down to 0 °C (ice/water), followed by dropwise addition of vinyl magnesium bromide in THF (1.0 M, 15.0 mL, 15.0 mmol). The reaction mixture was allowed to warm to room temperature and stirred for 12 h. The reaction mixture was quenched with saturated aqueous NH₄Cl, then treated with HCl (4 M) until the pH was 3. The organic components were extracted with EtOAc (3×20 mL). Hereafter, the combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure. The residue could be used directly for next step. To a solution of the 5-hydroxyl acid (1.0 equiv.) in DCM (15 mL) was added DMAP (0.1833, 0.3 equiv) and EDC (1.4375 g, 1.5 equiv). The reaction was stirred at room temperature for 4 h. The reaction mixture was then concentrated under reduced pressure.



The allylic amination step could be realized using modified conditions (55% NMR yield, E/Z = 94:6), while low yield (< 10%) of nine-membered lactam was detected from the crude reaction mixture by ¹H NMR after attempted cyclization.

Procedure: the vinyl ε -lactone (43.3 mg, 0.20 mmol, 1.0 equiv) was combined with Pd₂(dba)₃·CHCl₃ (4.0 mg, 2.0 mol%), **L8** (6.4 mg, 8.0 mol%) and aniline (28.5 mg, 0.30

mmol) in DCM (0.30 mL) at rt in air. The reaction mixture was stirred at rt for 12 h, then an aliquot of the mixture was taken for NMR analysis, which showed the unsaturated amino acid intermediate (55% NMR yield) to have an E/Z ratio of 96:4. Hereafter, the mixture was diluted to 0.01 M and EDC (57.5 mg, 3.0 mmol) was added, and the reaction mixture stirred for another 1 h. Hereafter, only a trace amount of target product could be detected by ¹H NMR of the crude mixture.

The lower yield of the nine-membered lactam can be explained by the increasing entropic cost to pre-organize the substrate for intramolecular cyclization versus intermolecular "amide" bond formation.

Analytical data for the seven-membered vinyl lactone:



¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.33 (m, 4H), 7.31 – 7.27 (m, 1H), 6.03 (dd, J = 17.3, 10.6 Hz, 1H), 5.15 – 5.08 (m, 2H), 2.67 – 2.54 (m, 2H), 2.25 – 2.13 (m, 2H), 1.95 – 1.81 (m, 2H), 1.76 – 1.58 (m, 2H).



¹³C NMR (101 MHz, CDCl₃) δ 175.25, 143.31, 141.81, 129.01, 127.63, 125.95, 114.03, 85.94, 38.24, 37.13, 24.57, 23.14.



HRMS (ESI⁺, MeOH): *m/z* calcd. 239.1043 (M + Na)⁺, found: 239.1036.

S219. Characterization of byproduct 2 from bis-allylation of aniline

The isolation of completely pure byproduct 2 was unsuccessful. The characterization by ¹H NMR, IR and MS data of the purest sample of 2 is here provided to support its proposed structure.



Yellow solid, Hexane : EA = 1 : 3, $R_f = 0.1$, or DCM:MeOH = 30 : 1, $R_f = 0.12$





HPLC-MS conditions: Zorbax C18 100×4.6 mm, 3.5μ m, $H_2O/MeOH = 30 : 70$. 70% MeOH up to 100% in 10'; hold 5', 1 mL/min. APCI +/-; Sample: 1 mg/ml in MeOH.





S222. X-ray structures of 3 and 53



Molecular structure for **3** (CCDC-1998235):

Molecular structure for **53** (CCDC-2001102):



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