# Inside polyMOFs: Layered Structures in Polymer-Based Metal-Organic Frameworks

Kyle C. Bentz,<sup>1</sup> Karthikeyan Gnanesekaran.,<sup>2</sup> Jake B. Bailey,<sup>1</sup> Sergio Ayala Jr.,<sup>1</sup> F. Akif

Tezcan,<sup>1</sup> Nathan C. Gianneschi,<sup>2\*</sup> and Seth M. Cohen<sup>1\*</sup>

- 1. Department of Chemistry and Biochemistry, University of California, San Diego, La Jolla, California 92093, United States
  - Departments of Chemistry, Materials Science & Engineering, Biomedical Engineering, Pharmacology. International Institute for Nanotechnology, Simpson-Querrey Institute, Chemistry of Life Processes Institute, Lurie Cancer Center, Northwestern University, Evanston, Illinois, 60208, USA

## **SUPPORTING INFORMATION**

#### **Materials Synthesis**

All solvents and starting materials were purchased from chemical suppliers and used without further purification (Sigma Aldrich, Alfa Aesar, EMD, and TCI).

#### **Polymer and polyMOF Synthesis**

Monomers, polymers, and polyMOFs were synthesized according to a previously published report (S. Ayala Jr., K. C. Bentz, S. M. Cohen. *Chem. Sci.* **2019**, *10*, 1746). Briefly, the general procedure for polyUiO-66 preparation is described. The polymer ligand (0.03 mmol by monomer repeat unit), ZrCl<sub>4</sub> (0.037 mmol), and 2 mL of DEF were added to a 20 mL scintillation vial. After the solution became clear, 2 mL of formic acid was added. The vial was heated at 135 °C for 48 h. The resultant film was washed by exchanging solvent with copious amounts of DMF, followed by copious amounts of methanol.

### **Characterization Methods**

**Powder X-ray Diffraction (PXRD):** PXRD data were collected at room temperature on a Bruker D8 Advance diffractometer running at 40 kV, 40 mA for Cu K $\alpha$  ( $\lambda$  = 1.5418 Å), with a scan speed of 0.5 sec/step, a step size of 0.01° in 20, and a 20 range of 3-50° at room temperature.

**Scanning Electron Microscopy (SEM).** MOFs were placed on conductive carbon tape on a sample holder and coated using an Ir-sputter coating for 7 sec. A Zeiss Sigma 500 ESEM microscope was used for acquiring images using a 2-3 kV energy source under vacuum at a working distance of 5 mm.

**Small-angle X-ray scattering**. Crystals in capillaries were analyzed at beamline 5-ID-D of the Advanced Photon Source (Argonne National Laboratory). Data were collected using collimated X-ray radiation (0.7293 Å, 17 keV) calibrated with both a glassy carbon standard and a silicon diffraction grating. Scattered radiation was detected using a CCD area detector. One-dimensional scattering data were obtained through the azimuthal averaging of the two-dimensional data to produce plots of the scattering intensity as a function of the scattering vector length,  $q = 4\pi \sin(\theta/\lambda)$ , where  $\theta$  is one-half of the scattering angle and  $\lambda$  is the wavelength of the X-rays used. Analysis of the one-dimensional data was performed using the powder diffraction processing software JADE (MDI) or Origin (OriginLab). Scattering vectors, *q*, were converted to Bragg diffraction D-spacings through the relationship  $q = 2\pi/D$ .

Gel-permeation chromatography (GPC). Gel-permeation chromatography was performed in DMF (0.7 mL/min) using a Malvern GPC equipped with D4000 single-pore column and D-6000M general-purpose mixed-bed weight divinylbenzene column connected in series to determine molecular weights and molecular weight distributions, Mw/Mn, of our polymers. The solutions were filtered through 0.4 µm PTFE membrane before being injected into either GPC instrument. Narrow poly (methyl methacrylate) (PMMA) was used as the calibration standard. <sup>1</sup>**H NMR.** <sup>1</sup>H nuclear magnetic resonance (NMR) spectra were collected using a Varian spectrometer running at 400 MHz.

**TEM Imaging.** TEM was performed using a JEOL ARM300F GrandARM TEM (operated at 300 kV) with Gatan OneView-IS camera and Gatan K3 in situ direct electron detector. An electron flux of 0.5 to 2 e<sup>-</sup> Å<sup>-2</sup> s<sup>-1</sup> (with a beam current of 1.52 nA) such that the cumulative dose not exceeding ~10 e<sup>-</sup> Å<sup>-2</sup> was used over the course of acquisition. HAADF-STEM and elemental mapping were performed using a JEOL ARM200CF Aberration-Corrected TEM (operated at 200 kV). X-ray signals were collected using a Dual SDD EDS detector. HAADF-STEM data were collected with a probe semi-convergence angle of 10 mrad and at a camera length of 20 cm. A beam current of 0.3 nA and pixel dwell times between 1 and 5  $\mu$ s. TEM samples were prepared by pipetting ~3  $\mu$ L of polyMOF sample dispersed in methanol onto a 200-mesh copper TEM grids with lacey carbon support layer. Prior to drop cast, TEM grids were glow discharged for 60 sec with a 25 mA current. Thin sections of ~100 to ~200 nm polyMOF samples were prepared by microtoming using a Leica EM UC7/FC7 Cryo-Ultramicrotome.



Figure S1. GPC chromatogram of pbdc-8e, methyl ester protected polymer ligand.



**Figure S2**. GPC chromatogram of pbdc-8e-PEG<sub>4k</sub>-10%, methyl ester protected polymer ligand.



**Figure S3**. GPC chromatogram of pbdc-8e-PEG<sub>2k</sub>-OMe, methyl ester protected polymer ligand.



Figure S4. GPC chromatogram of pbdc-8e-COD<sub>1:1</sub>, methyl ester protected polymer ligand.



Figure S5. HR-TEM images of thin films (100 nm) of polyUiO-66 derived from pbdc-8a-

PEG<sub>4k</sub>-10%.



**Figure S6**. PXRD patterns of (top to bottom): 4,000 g/mol semi-crystalline PEG; pbdc-8a-PEG<sub>4k</sub>-10% block polymer (free acid); pbdc-8e-PEG<sub>4k</sub>-10% block polymer (methyl ester protected); pbdc-8e homopolymer (methyl ester protected); pbdc-8a homopolymer (free acid, with trapped potassium chloride); potassium chloride; polyUiO-66 derived from pbdc-8a-PEG<sub>4k</sub>-10% block polymer; polyUiO-66 derived from pbdc-8a homopolymer physically blended with 4,000 g/mol PEG at the same weight percent in the pbdc-8a-PEG<sub>4k</sub>-10% block polymer; simulated powder pattern for UiO-66. Small amounts of potassium chloride are present in the free acid polymers due to deprotection conditions from the methyl ester precursor.



**Figure S7.** HR-TEM images of thin films of polyUiO-66 derived from pbdc-8a-PEG<sub>2k</sub>-OMe. Green arrow denotes the x-y plane of a crystal where the lattice spacing of 1.2 to 1.4 nm represents the distance between neighboring Zr clusters; orange arrow denotes the crystal in x-z axis where the 2D stacked architecture of UiO-66 and polymer are evident by 4.2 nm spacing.



**Figure S8**. Top-down view of polyUiO-66 derived from pbdc-8a-PEG<sub>2k</sub>-OMe showing the presence of nanochannels: a) HRTEM of microtomed section of bulk crystal, b) high magnification image of white inset in (a).



**Figure S9.** HR-TEM images of thin films (100 nm) of polyUiO-66 derived from pbdc-8a- $COD_{1:1}$ .



**Figure S10.** HR-TEM images of thin films (100 nm) of polyUiO-66 derived from homopolymer pbdc-8a (inset: electron diffraction pattern).



Figure S11. TEM images of bulk crystals of polyUiO-66 derived from pbdc-8a-PEG<sub>2k</sub>-OMe.



Figure S12. TEM images of bulk crystals of polyUiO-66 derived from pbdc-8a-COD<sub>1:1</sub>.



Figure S13. SAXS scattering profile of polyUiO-66 derived from homopolymer pbdc-8a.



Figure S14. SAXS scattering profile of polyUiO-66 derived from polyBDC-*b*-PEG<sub>4k</sub>-10%



**Figure S15**. SAXS scattering profile of polyUiO-66 derived from  $polyBDC-b-PEG_{2k}$ -OMe.



Figure S16. SAXS scattering profile of polyUiO-66 derived from polyBDC-co- $COD_{1:1}$ .



Figure S17. <sup>1</sup>H NMR spectrum of pbdc-8a ligand.



Figure S18. <sup>1</sup>H NMR spectrum of pbdc-8a-PEG<sub>4k</sub>-10%, ligand.



Figure S19. <sup>1</sup>H NMR spectrum of pbdc-8a-PEG<sub>2k</sub>-OMe ligand.



Figure S20. <sup>1</sup>H NMR spectrum of pbdc-8a-COD<sub>1:1</sub> ligand.